



Trichome Lengths of the Heterocystous N₂-Fixing Cyanobacteria in the Tropical Marginal Seas of the Western North Pacific

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Calothrix rhizosoleniae and *Richelia intracellularis* are heterocystous cyanobacteria found in the tropical oceans. *C. rhizosoleniae* commonly live epiphytically on diatom genera *Chaetoceros* (C-C) and *Bacteriastrium* (B-C) while *R. intracellularis* live endosymbiotically within *Rhizosolenia* (R-R), *Guinardia* (G-R), and *Hemiaulus* (H-R); although, they occasionally live freely (FL-C and FL-R). Both species have much shorter trichomes than the other marine filamentous cyanobacteria such as *Trichodesmium* spp. and *Anabaena gerdii*. We investigated the trichome lengths of *C. rhizosoleniae* and *R. intracellularis* in the South China Sea (SCS) and the Philippine Sea (PS) between 2006 and 2014. On average, H-R had the shortest trichome lengths (3.5 cells/trichome), followed by B-C and C-C (4.9–5.2 cells/trichome) and FL-C (5.9 cells/trichome), and R-R, G-R, and FL-R had the longest trichome lengths (7.4–8.3 cells/trichome). Field results showed the trichome lengths of C-C and B-C did not vary seasonally or regionally. However, FL-C and H-R from the SCS and during the cool season had longer trichomes, where/when the ambient nutrient concentrations were higher. R-R, G-R, and FL-R also showed regional and seasonal variations in trichome length. Ultrastructural analysis found no gas vesicles within the *C. rhizosoleniae* cells to assist in buoyancy regulation. Results suggest that the trichome lengths of *C. rhizosoleniae* and *R. intracellularis* might be regulated by their diatom hosts' symbiotic styles and by ambient nutrients. Short trichome length might help *C. rhizosoleniae* and *R. intracellularis* to stay in the euphotic zone regardless as to whether they are free-living or symbiotic.

Keywords: *Calothrix rhizosoleniae*, *Richelia intracellularis*, diatom diazotroph associations, free-living *Richelia* and *Calothrix*, South China Sea, Philippine Sea, gas vesicles

INTRODUCTION

Marine cyanobacterial dinitrogen (N₂) fixation plays a substantial role in global biogeochemical nitrogen (N) and carbon cycling. Nitrogen fixers (diazotrophs) transform N₂ into bioavailable ammonium (NH₄⁺), which can promote primary and secondary production (Carpenter et al., 1999; Sohm et al., 2011) and support large non-diazotrophic diatoms in oligotrophic seawater

(Chen et al., 2011). Diazotrophic cyanobacteria are photoautotrophic, or symbiotic with photoautotrophs, and reside in the upper euphotic zone of the ocean (Carpenter and Capone, 2008). They are classified into several phylogenetic groups that include heterocystous (heterocyst-forming) filamentous (e.g., *Richelia intracellularis*, *Calothrix rhizosoleniae*, and *Anabaena gerdii*), non-heterocystous filamentous (*Trichodesmium* spp.), and unicellular cyanobacteria (*Candidatus Atelocyanobacterium thalassa*, *Crocospaera watsonii*, and *Cyanothece* spp.) (Carpenter and Janson, 2001; Capone et al., 2005; Foster et al., 2007). While the unicellular representatives can be abundant in both coastal and oligotrophic regions, filamentous groups often occur primarily in oligotrophic oceanic regions and riverine plume-influenced mesohaline waters (Foster et al., 2007; Subramaniam et al., 2008; Grosse et al., 2010; Turk et al., 2011; Chen et al., 2014; Shiozaki et al., 2018, 2020; Mulholland et al., 2019).

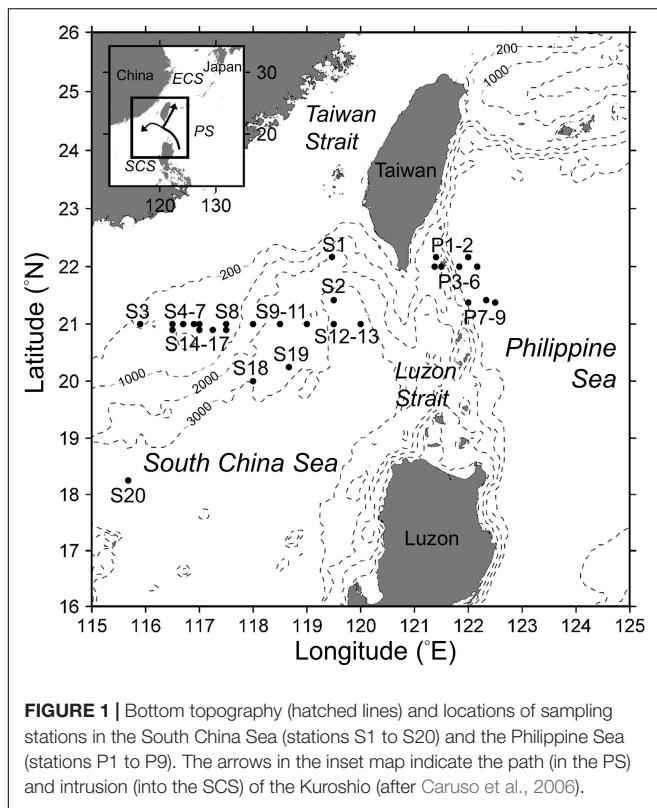
Calothrix rhizosoleniae (hereafter *Calothrix*, unless noted) and *Richelia intracellularis* (hereafter *Richelia*) are commonly observed in tropical and subtropical ocean regions and marginal seas (Kimor et al., 1992; Carpenter et al., 1999; Gómez et al., 2005; Bar Zeev et al., 2008; Tuo et al., 2017). Both *Calothrix* and *Richelia* are most often observed in symbiotic relationships with certain diatom species and only occasionally observed free-living (FL) outside of a host (Karsten, 1907; Gómez et al., 2005; White et al., 2007; Foster et al., 2011; Tuo et al., 2017). These diatom diazotroph associations/assemblages (DDAs) are thought to contribute substantial fixed N to oceanic regions where they occur (Mague et al., 1974; Carpenter et al., 1999; Bar Zeev et al., 2008; Grosse et al., 2010).

Calothrix and *Richelia* have different styles of symbiosis. *Calothrix* is often epiphytic with chain-forming *Chaetoceros compressus* and *Bacteriastrum* spp. (both belong to Family Chaetocerotaceae) while *Richelia* most often occurs as an endosymbiont with *Rhizosolenia clevei*, *Guinardia cylindrus* (both belong to Family Rhizosoleniaceae), and *Hemiaulus* spp. (Family Hemiaulaceae) (Sundström, 1984; Gómez et al., 2005; Tuo et al., 2017). More precisely, symbiotic *Richelia* filaments dwell inside the cytosol of *Hemiaulus* hosts or dwell in the periplasmic space between the plasmalemma and frustule of *Rhizosolenia* hosts (Villareal, 1990; Caputo et al., 2019; Nieves-Morióń et al., 2020). The actual location of symbiotic *Richelia* inside the *Guinardia* hosts is still unknown. It is also unknown whether the *Richelia* associated with different host diatoms are distinct species or strains (Sundström, 1984; Janson et al., 1999; Foster and Zehr, 2006). *Hemiaulus*-associated *Richelia* (*H-R*) is considered an obligate symbiont due to the fact that its smaller genome (3.2 Mbp) lacks many N-associated metabolic pathways, e.g., glutamine 2-oxoglutarate aminotransferase (GOGAT) and transporters for NH_4^+ , nitrate (NO_3^-), nitrite (NO_2^-), and urea (Hilton et al., 2013; Hilton, 2014). The larger genome (5.4 Mbp) of *Rhizosolenia*-associated *Richelia* (*R-R*) also lacks the combined-N transporters (Hilton, 2014; Nieves-Morióń et al., 2020). Both the *nifH* gene analysis and morphological examination of FL-*Richelia* (FL-*R*) from field samples suggest that FL filaments originate from *Rhizosolenia* rather than *Hemiaulus* (Zhang et al., 2011; Tuo et al., 2017). However, little is known about the genome of FL-*R* or *Guinardia*-associated *Richelia*

(*G-R*). In contrast, *Calothrix* is considered a facultative symbiont or an opportunist. Its genome (6.0 Mbp) is more similar to those of FL heterocystous cyanobacterial species (Hilton et al., 2013; Nieves-Morióń et al., 2020), suggesting its ability to live freely (FL-*C*) in nature. In the marginal seas of the low-latitude western North Pacific, FL-*C* comprised about 20–40% of overall *Calothrix* abundance [FL-*C* plus *Chaetoceros*- (*C-C*) and *Bacteriastrum*-associated *Calothrix* (*B-C*), $48\text{--}698 \times 10^3$ filaments m^{-2}] (Tuo et al., 2017). FL-*R* were occasionally found in the upper water column of the tropical western North Pacific and their abundance ($1\text{--}29 \times 10^3$ filaments m^{-2}) was much lower than symbiotic *Richelia* ($403\text{--}1,703 \times 10^3$ filaments m^{-2}) (Tuo et al., 2017). It has been suggested that FL-*R* are not viable due to their lack of gas vesicles to help them regulate their buoyancy (Janson et al., 1995). However, to date, little is known about how cellular ultrastructure affects the buoyancy regulation of *Calothrix* or whether their symbiotic behavior offer these organisms the ability to stay in the upper water column in the absence of cellular architecture facilitating buoyancy.

It is interesting that both *Calothrix* and *Richelia* have much shorter trichome lengths than other marine filamentous cyanobacteria. According to previous studies from different regions, symbiotic *Calothrix* filaments have no more than 14 cells per trichome (vegetative cells plus one terminal heterocyst) whereas symbiotic *Richelia* have no more than 21 cells per trichome (Schmidt, 1901; Foster and Zehr, 2006; Tuo et al., 2017). Little is known about the trichome lengths of FL-*C* and FL-*R*. In contrast, a filament of *Trichodesmium* may contain more than 100 cells per trichome (Capone et al., 1997; Luo et al., 2012) and a filament of *A. gerdii*, a sparsely distributed heterocystous species, may contain more than 300 cells per trichome (Carpenter and Janson, 2001). These field observations give rise to a question about the trichome length of *Calothrix* and *Richelia*: were the trichome lengths different among the symbionts associated with distinct diatom hosts?

The northern South China Sea (SCS) and the western Philippine Sea (PS), two large and connected marginal seas of the western North Pacific Ocean, harbor diverse N_2 -fixing cyanobacteria (Chen et al., 2014, 2019; Shiozaki et al., 2014; Tuo et al., 2017). Both regions are influenced by the Kuroshio western boundary current, in which *Trichodesmium*, *Calothrix*, and *Richelia* are prevalent (Chen et al., 2008; Shiozaki et al., 2014, 2015; Tuo et al., 2014). The Kuroshio originates along the eastern coast of Luzon and flows north across the Luzon Strait where it exchanges waters with the SCS before meandering along the eastern coast of Taiwan and meeting the continental slope of the East China Sea (Figure 1; Caruso et al., 2006; Jan et al., 2015). The monsoonal circulation of the SCS results in seasonal variations in the velocity, width, and path of the Kuroshio at the northwestern boundary of the PS (Jan et al., 2015), as well as stronger Kuroshio intrusion from the PS into the SCS during the northeast monsoon (NEM) (Liang et al., 2003). During the NEM, the SCS has cool surface seawater temperatures (SST) and the circulation in the basin is cyclonic, while during the southwest monsoon (SWM), SST is warm and the circulation is anticyclonic in the SCS (Liu et al., 2002). Although monsoon-driven seasonal changes in SST, surface photosynthetically active



radiation (SPAR), and stratification index (SI) are observed, the SCS and the PS are both oligotrophic year-round. However, the nutrient distributions in the upper water column differ somewhat between the regions; the SCS generally has shallower nitracline (D_N) and phosphocline depths (D_P) than the PS, suggesting a potential for higher fluxes of nitrate and phosphate from the depth (Chen et al., 2008; Tuo et al., 2017). The two marginal seas provide opportunities to examine whether the trichome length of each distinct host-associated or FL *Calothrix* or *Richelia* was different as the filaments habited in specific regions and seasons.

In this study, we evaluated how trichome length and intracellular characteristics contribute to the distribution of *Calothrix* and *Richelia* in the field. Specifically, we (1) examined whether the trichome lengths were different among the symbionts associated with distinct diatom hosts (and their FL counterparts) and (2) examined whether the trichome length for each *Calothrix*/*Richelia* symbiont was different between the regions and between the seasons. We also used laboratory cultures of an FL-C strain isolated from the SCS seawater to examine whether *C. rhizosoleniae* has gas vesicles and other intracellular structures.

MATERIALS AND METHODS

Sampling Strategy

Field samples were collected during 20 cruises between 2006 and 2014 aboard the R/V *Ocean Researcher I* (OR1) and the

R/V *Ocean Researcher III* (OR3) (Table 1). The cruises were conducted seasonally during the warm SWM (from April to October, 14 cruises) and the cool NEM (from mid-December to January, 6 cruises) seasons (Table 1). Samples were collected from 1 to 11 stations per cruise to investigate trichome lengths of *Calothrix* and *Richelia* and environmental parameters (Figure 1 and Table 1). In total, 83 stations were sampled from 29 discrete locations (some stations were revisited) over the 9-year period, and these were categorized into four region–season combinations: SCS-SWM, SCS-NEM, PS-SWM, and PS-NEM. At each station, water samples were collected from seven to nine discrete depths between 5 and 200 m using 20-L Go-Flo bottles attached to a rosette equipped with a Seabird SBE 9 conductivity–temperature–depth sensor and an underwater photosynthetically active radiation (PAR) sensor (OSP200L, Biospherical).

Trichome Length Examination on *Richelia* and *Calothrix*

Samples collected to identify *Richelia* and *Calothrix* and to measure their trichome lengths were prepared as described in Tuo et al. (2017). *Richelia* and *Calothrix*, showing an orange-yellow fluorescence, were identified at $\times 400$ magnification with Zeiss epi-fluorescence microscopes equipped filter set of blue excitation and long pass emission (BP 450–490, FT 510, LP 520). Bright field was also used to distinguish symbionts from FL and to identify their host diatom species.

In total, 12,254 filaments of *Richelia* and 2,697 filaments of *Calothrix*, in free-living (176 filaments of FL-R and 1,206 filaments of FL-C) or symbiotic forms (12,078 filaments of *Richelia* and 1,491 filaments of *Calothrix*), were examined and their trichome lengths recorded. The morphological examination of 1,382 FL filaments (FL-R and FL-C) was found at the 83 stations sampled during the 20 cruises. A total of 13,569 symbiont filaments (H-R, R-R, G-R, C-C, and B-C) were examined from 43 of the 83 stations (from seven of the 20 cruises, conducted between December 2010 and October 2014) (Table 1). The general morphological descriptions and the abundances of each DDA and FL were published in Tuo et al. (2017).

Environmental Parameters

At each station, the euphotic depth (D_E , m), surface PAR influx (SPAR, $E\ m^{-2}\ day^{-1}$), stratification index (SI, $kg\ m^{-4}$), mixed-layer depth (D_M , m), NO_3^- plus NO_2^- (N+N), soluble reactive phosphorus (phosphate, P), and chlorophyll *a* ($mg\ m^{-3}$) concentrations were calculated or measured following Tuo et al. (2017). To better assess the total nutrient inventory in the upper water column at each station, in addition to surface concentrations (N_S and P_S , nM), depth-integrated nutrient inventories (N_I and P_I) and the nutricline depths (D_N and D_P) were calculated for each nutrient (N+N or P). Depth-integrated inventories (N_I and P_I , $mmol\ m^{-2}$) were calculated by trapezoidal integration from the surface to 100 m depth. The nutricline is defined as the seawater layer where the greatest change in the nutrient concentration occurs with depth. Based on the definition, the nutricline depth for each nutrient (D_N and D_P) was found from the sampling depths as the depth at which

TABLE 1 | Sampling stations occupied during 20 cruises in the SCS and the PS (see **Figure 1** for station locations).

Season	Cruise	Date	SCS stations	PS stations
SWM	OR3-1217*	April 21–25, 2007		1, 2, 3
	OR3-1234*	July 7–10, 2007		3, 5
	OR1-843*	September 27–October 8, 2007	1, 2	3
	OR3-1284*	April 14–19, 2008	2	3
	OR3-1310*	August 2–7, 2008		3, 6
	OR3-1316*	September 5–10, 2008		3, 6
	OR1-899*	May 17–20, 2009		3
	OR1-910*	August 14–20, 2009	2, 12, 13	4, 7, 9
	OR3-1455*	May 12–17, 2010	4, 7, 8, 9, 12, 13	7, 9
	OR1-973	August 19–28, 2011	3, 4, 7, 8, 9, 11, 12, 13	7, 9
	OR3-1603	May 3–8, 2012	7, 8, 9, 11, 12	
	OR3-1683	May 6–11, 2013	7, 8, 9, 11, 12, 13	
	OR1-1074	May 17–21, 2014	10, 18	
	OR1-1091	October 12–18, 2014	4, 5, 6, 7, 8, 9, 11, 14, 15, 16, 17	
NEM	OR1-819*	December 18–30, 2006	2, 19	8
	OR1-821*	January 12–19, 2007	20	
	OR1-852*	December 10–15, 2007		3, 5, 6
	OR1-886*	December 8–14, 2008	2	3, 5, 6
	OR1-950	December 2–10, 2010	7, 8, 9, 11, 12, 13	9
	OR1-1022	December 7–12, 2012	7	7

*Examinations of free-living filaments only.

the nutrient gradient increased significantly ($57 \pm 52 \text{ nM m}^{-1}$ for N+N and $4 \pm 5 \text{ nM m}^{-1}$ for P) in the upper water column. The data for some environmental parameters, including SPAR, SST, SI, D_M , surface salinity (Sal), N_S , P_S , surface chlorophyll *a* (Chl), and depth of chlorophyll *a* maximum (D_{CM}) were previously published in Tuo et al. (2017).

Isolation and Identification of FL-C Strain

A unicyanobacterial strain of FL *C. rhizosoleniae* was isolated from the surface water in the SCS collected at $21^{\circ}25'N$, $119^{\circ}28'E$ on December 10, 2007 (cruise OR1-852) and enriched with nitrate ($10 \mu\text{M}$ final concentration). After a 1-week incubation in the nitrate-enriched natural seawater, cells were transferred into new polycarbonate (PC) bottles and placed near the window at room temperature (around $19\text{--}28^{\circ}\text{C}$) in the laboratory. Brownish-yellow clumps appeared at the bottom of the bottles after 3 weeks, and these were transferred into a new PC bottle containing YBCII culture medium (Chen et al., 1996) using a pneumatic micropipette under a Zeiss inverted microscope. Solitary filaments were visible in the bottle a few days later, and these were isolated individually, rinsed several times with YBCII medium, and then transferred to a sterilized Pyrex flask with a cotton stopper containing YBCII medium. A stir bar was placed inside the flask, and the flask was placed on a magnetic stir plate to keep the filaments in suspension. The unicyanobacterial culture was then incubated in a light and temperature-controlled incubator at an illumination intensity of $100\text{--}120 \mu\text{mol quanta m}^{-2} \text{ s}^{-1}$ on a 12:12-h light:dark cycle at 28°C .

Both molecular phylogenetic and morphological examinations were done to identify the strain of isolated FL-C. Phylogenetic analysis based on *nifH* gene sequence (Foster

and Zehr, 2006) was applied with several minor modifications (Taniuchi et al., 2012). The *nifH* primers and nested polymerase chain reaction protocol followed Zehr and Turner (2001). Cloned deoxyribonucleic acid (DNA) fragments were sequenced by Tri-I Biotech (Taipei, Taiwan). The sequences were aligned by ClustalW program of DDBJ¹. A phylogenetic tree was then constructed *via* neighbor-joining by ClustalW. Distance matrix of deduced amino acid sequences of *nifH* were calculated with Kimura's model. The bootstrapped value was based on 1,000 iterations to test for robustness. For the phylogenetic analysis, the *nifH* genes of *Trichodesmium* sp. IMS 101 and *Trichodesmium thiebautii* were used as the outgroups. The identity with close strains was calculated by the identity matrix of BioEdit 4.0 (Hall, 1999). The *nifH* sequences determined in the study have been submitted in the GenBank database² under Accession No.: KF284162. Morphological observations included size of heterocysts, the size of heterocysts relative to adjacent vegetative cells, and the tapering of filaments, to distinguish *Calothrix* from *Richelia* (Tuo et al., 2017).

Ultrastructural Analysis on Cultured FL-C

To verify whether gas vesicles were present in cells of *Calothrix*, ultrathin sections of cells were examined using a transmission electron microscope (TEM) on April 7, 2015. The examined filaments, harvested from the laboratory cultures in March 2015, were first transferred to 2.5% glutaraldehyde fixative (in YBCII), then postfixed using 1% osmium tetroxide, dehydrated in an acetone series, embedded in Spurr's resin (Spurr, 1969), and

¹<http://clustalw.ddbj.nig.ac.jp/top-e.html>

²<http://www.ncbi.nlm.nih.gov/genbank/>

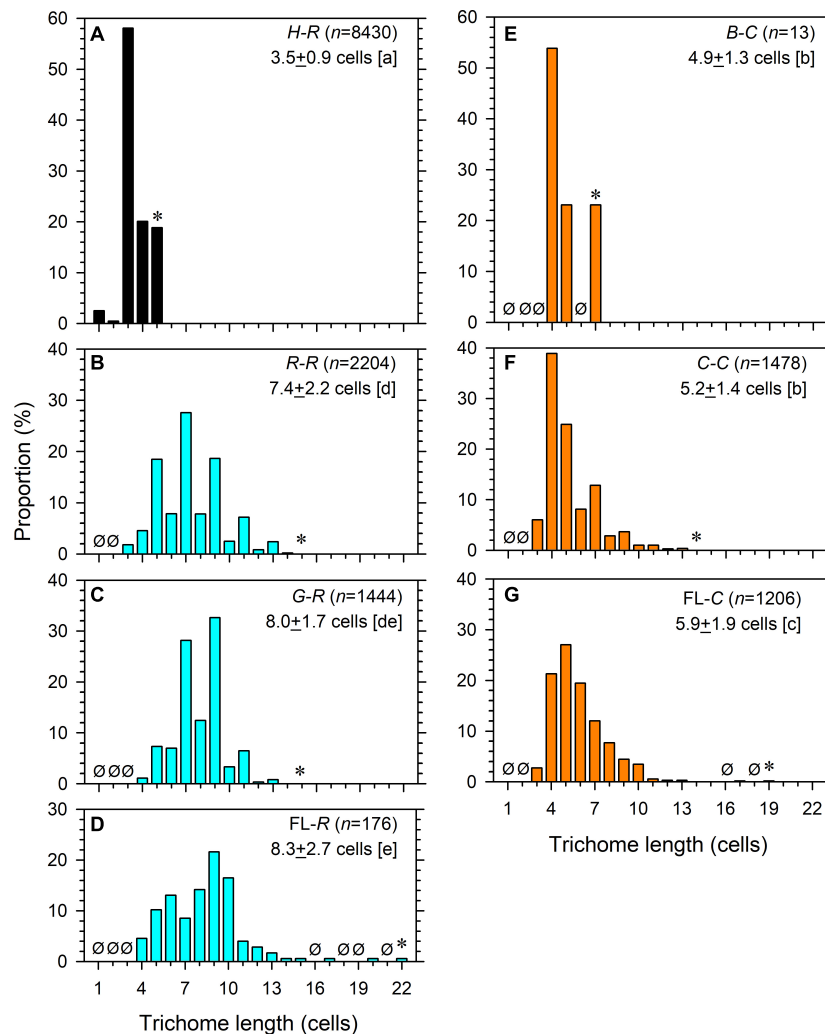


FIGURE 2 | Trichome lengths of (A) internal and (B,C) partial symbionts *Richelia* and (D) free-living *Richelia*, as well as (E,F) external symbionts *Calothrix* and (G) free-living *Calothrix*. ∅ indicates none observed at the length. Asterisk indicates the maximal length observed. *n*, number of filaments examined. The mean ± 1 standard deviation of length is noted. Different lowercase letters in the brackets refer to significant difference ($p < 0.05$) of the mean after ANOVA ($F_{6,14,944} = 3,645$, $p < 0.0001$) and Tukey-Kramer test.

sectioned with an ultramicrotome (EM UC7, Leica). Ultrathin sections were double stained with uranyl acetate and lead citrate (Reynolds, 1963) and observed under TEM (Tecnai G2 Spirit TWIN, FEI) at 80 kV.

Statistical Analysis

To compare the trichome lengths among the seven types of *Richelia* and *Calothrix* (H-R, R-R, G-R, FL-R, B-C, C-C, and FL-C), one-way analysis of variance (ANOVA) and a *post hoc* Tukey-Kramer test with 0.05 of significance level were used. Before the ANOVA, data from the four region-season classifications were combined for each type, respectively. Differences in trichome lengths of *Calothrix* and *Richelia* and in hydrographic parameters and biogeochemical parameters between regions (SCS vs. PS) and seasons (NEM vs. SWM) were compared using two-way ANOVA. For the unbalanced datasets, a Type II sum-of-square method was

applied for the ANOVA. If the interaction effect between region and season was significant ($p < 0.05$), then a *post hoc* Tukey-Kramer test was performed to investigate the difference among the four region-season classifications (SCS-NEM, SCS-SWM, PS-NEM, and PS-SWM). All the data were presented as mean ± 1 standard deviation. SAS program (SAS Institute) was used for all statistical procedures.

RESULTS

Trichome Lengths of *Richelia* and *Calothrix* in the Field

The combined field data showed that trichome lengths were different among the different symbiotic styles of *Calothrix* and *Richelia* (Figures 2A–G). On average, H-R had the shortest

TABLE 2 | Trichome length of *Richelia* and *Calothrix* in the SCS and the PS during SWM and NEM.

Types	SCS		PS		F values in two-way ANOVA		
	SWM	NEM	SWM	NEM	Region × season	Region	Season
Richelia							
H-R	3.5 ± 0.9 ^b (n = 7,453)	3.8 ± 0.8 ^c (n = 687)	2.6 ± 1.2 ^a (n = 241)	3.2 ± 0.4 ^b (n = 49)	9.91** (df = 1, 8,426)		
R-R	7.6 ± 2.1 ^b (n = 1,656)	6.7 ± 2.1 ^{ab} (n = 39)	6.7 ± 2.5 ^a (n = 460)	7.9 ± 2.2 ^b (n = 49)	18.1** (df = 1, 2,200)		
G-R†	8.0 ± 1.7 (n = 1,397)	11.5 ± 0.7 (n = 2)	7.4 ± 1.6 (n = 45)	ND	–	–	–
FL-R	9.0 ± 1.8 ^b (n = 69)	9.3 ± 2.8 ^b (n = 4)	7.6 ± 2.7 ^a (n = 87)	8.8 ± 4.5 ^a (n = 16)	0.35 (df = 1, 172)	10.8**	2.25
Calothrix							
C-C	5.2 ± 1.7 ^a (n = 1,157)	5.0 ± 2.5 ^a (n = 22)	5.3 ± 1.7 ^a (n = 285)	6.0 ± 2.0 ^a (n = 14)	2.05 (df = 1, 1,474)	1.07	0.35
B-C	5.0 ± 1.3 (n = 12)	4 (n = 1)	ND	ND	–	–	–
FL-C	6.0 ± 1.9 ^b (n = 512)	8.7 ± 3.3 ^d (n = 45)	5.5 ± 1.5 ^a (n = 531)	6.5 ± 2.0 ^c (n = 118)	22.8** (df = 1, 1,202)	–	–

n, number of filaments examined; ND, no data; df, degree of freedom. **p < 0.01. Different superscript letters indicate significant differences (p < 0.05) after Tukey-Kramer tests. †Comparisons for G-R: SCS vs. PS during the SWM, $t_{df=1,440} = -2.32$, p < 0.05 (two-tailed).

TABLE 3 | Hydrographic and biogeochemical properties in the SCS and the PS during SWM and NEM.

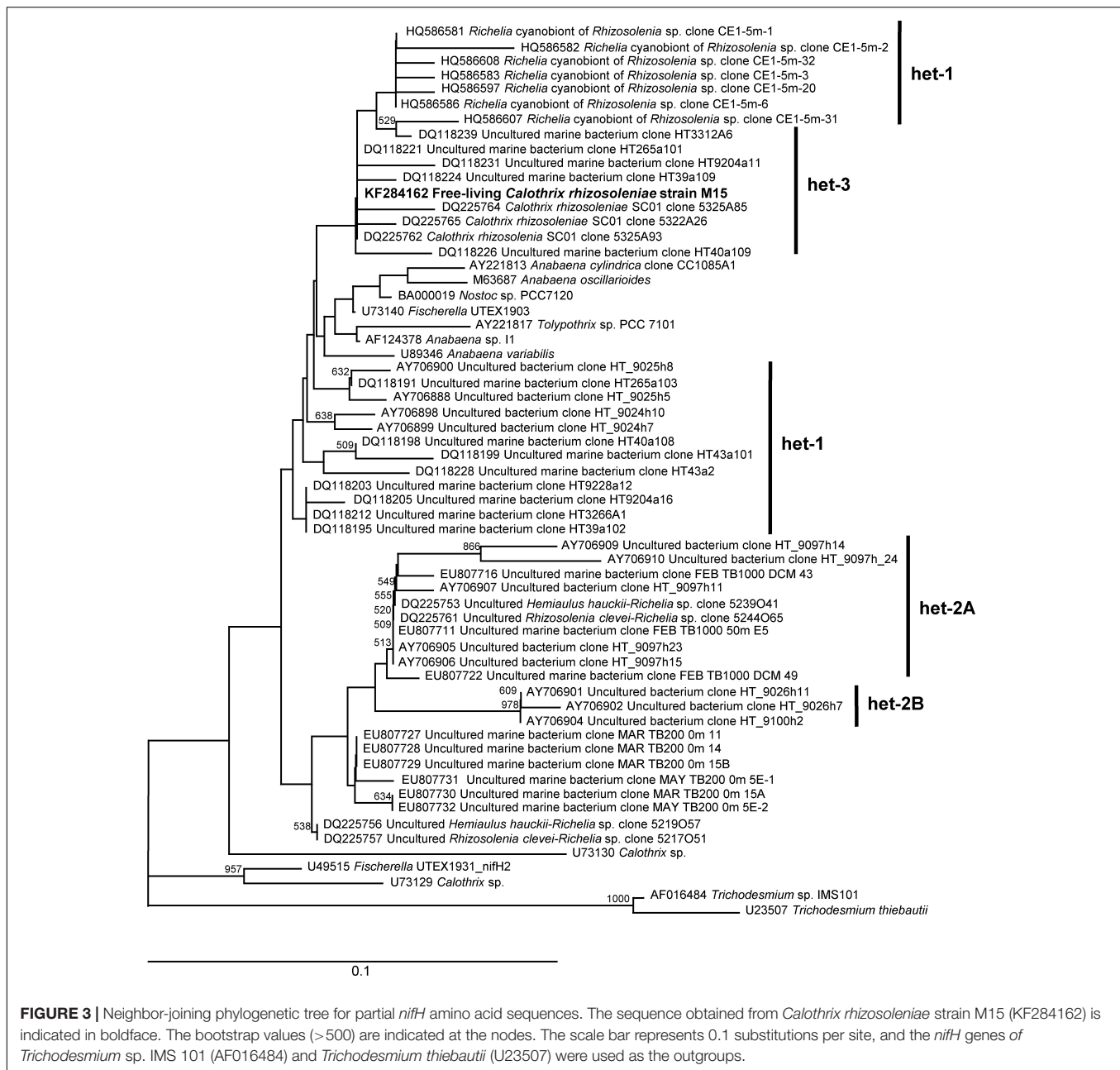
Parameters	SCS		PS		F _{1,179} in two-way ANOVA		
	SWM (n = 44)	NEM (n = 11)	SWM (n = 19)	NEM (n = 9)	Region × season	Region	Season
SPAR (E m ⁻² day ⁻¹)	62.8 ± 12.5 ^b (n = 40)	27.1 ± 17.1 ^a	63.5 ± 17.6 ^b (n = 14)	31.5 ± 16.0 ^a	0.22 (F _{1,170})	0.25	77.2**
D _E (m)	83 ± 13 ^a (n = 31)	78 ± 4 ^a (n = 5)	113 ± 19 ^b (n = 14)	107 ± 11 ^b (n = 6)	0.01 (F _{1,62})	52.0**	1.14
SST (°C)	28.2 ± 1.2 ^c	25.7 ± 1.0 ^a	28.9 ± 1.0 ^d	26.4 ± 0.9 ^b	0.00	6.37*	76.5**
SI (×10 ⁻² kg m ⁻⁴)	2.6 ± 0.9 ^c	0.9 ± 0.4 ^{ab}	1.6 ± 0.5 ^b	0.8 ± 0.6 ^a	5.57*	–	–
D _M (m)	56 ± 31 ^a	100 ± 30 ^c	89 ± 46 ^b	125 ± 37 ^d	0.17	14.2**	19.8**
Sal	33.7 ± 0.3 ^a	34.0 ± 0.2 ^b	34.2 ± 0.3 ^c	34.4 ± 0.2 ^d	0.48	53.0**	8.25**
N _S (nM)	24 ± 23 ^a	360 ± 729 ^b	24 ± 21 ^a	27 ± 18 ^a	5.89*	–	–
P _S (nM)	30 ± 15 ^a	69 ± 40 ^b	30 ± 17 ^a	41 ± 28 ^a	6.19*	–	–
N _S :P _S	0.9 ± 0.7 ^a	3.5 ± 3.8 ^b	1.0 ± 1.2 ^a	0.7 ± 0.5 ^a	11.7**	–	–
N _I (mmol m ⁻²)	142 ± 99 ^b	107 ± 77 ^b	15 ± 20 ^a	26 ± 28 ^a	1.20	38.1**	0.58
P _I (mmol m ⁻²)	15.0 ± 7.6 ^b	12.4 ± 5.8 ^b	5.5 ± 4.3 ^a	5.9 ± 2.6 ^a	0.83	34.2**	0.70
N _I :P _I	8.7 ± 2.9 ^b	7.9 ± 2.3 ^b	2.8 ± 2.4 ^a	4.1 ± 2.8 ^a	2.04	71.9**	0.01
D _N (m)	45 ± 15 ^a	43 ± 24 ^a	87 ± 26 ^b	73 ± 26 ^b	1.47	67.4**	1.74
D _P (m)	38 ± 17 ^a	54 ± 30 ^a	75 ± 42 ^b	69 ± 27 ^b	2.35	24.1**	1.08
Chl (mg m ⁻³)	0.16 ± 0.12 ^a	0.45 ± 0.12 ^b	0.10 ± 0.05 ^a	0.21 ± 0.12 ^a	11.6**	–	–
D _{CM} (m)	63 ± 19 ^c	15 ± 24 ^a	89 ± 22 ^d	60 ± 30 ^b	2.75	35.8**	50.7**

n, number of stations; df, degree of freedom. *p < 0.05; **p < 0.01. Different superscript letters indicate significant differences (p < 0.05) after Tukey-Kramer tests.

trichome length (3.5 ± 0.9 cells), followed by B-C (4.9 ± 1.3 cells) and C-C (5.2 ± 1.4 cells), and R-R (7.4 ± 2.2 cells) and G-R (8.0 ± 1.7 cells) had the longest trichome length (p < 0.05). The differences between B-C and C-C and between R-R and G-R were not significant (p > 0.05). FL-C showed longer average trichome length (5.9 ± 1.9 cells) than its symbiotic counterparts B-C and C-C (p < 0.05). FL-R also showed longer trichome length (8.3 ± 2.7 cells) than its symbiotic counterpart R-R (p < 0.05); however, it showed comparable (p > 0.05) trichome lengths with G-R. The FL cyanobacteria were able to develop longer trichomes than their symbiotic counterparts, according to the maximum observed cells per trichome (22 cells for FL-R and 19 cells for FL-C) (Figures 2B–G).

Among the host-associated and FL filaments, some of them showed significant seasonal and/or regional differences in their average trichome length, although the difference was only around 1 cell per trichome (Table 2). H-R had longer trichomes during

the NEM than the SWM in both regions and had longer trichomes in the SCS than in the PS during both seasons (p < 0.05) (Table 2 and Supplementary Figure 1). The trichome length of R-R was longer in the SCS than the PS during the SWM and was longer during the NEM than during the SWM in the PS (p < 0.05) (Table 2 and Supplementary Figure 2). Similarly, the trichome length of G-R was also longer in the SCS than the PS during the SWM (p < 0.05) (Table 2 and Supplementary Figure 3). However, FL-R had longer trichomes in the SCS than the PS during both seasons (p < 0.05), but the seasonal differences were not significant (p > 0.05) (Table 2 and Supplementary Figure 4). On the other hand, the symbiotic *Calothrix* samples did not show regional and seasonal differences. Specifically, the trichome length of C-C was comparable (p > 0.05) between the regions and/or between the seasons (Table 2 and Supplementary Figure 5). The data of B-C were too few for statistical tests (Table 2 and Supplementary Figure 6). However, FL-C had



longer trichomes during the NEM than the SWM in both regions and had longer trichomes in the SCS than the PS during both seasons ($p < 0.05$) (Table 2 and Supplementary Figure 7).

Hydrographic and Biogeochemical Properties in the Study Areas

Hydrographic characteristics, including SPAR, SST, SI, and D_M , differed significantly ($p < 0.05$) between seasons in both the SCS and the PS. The first three parameters were higher during the SWM than during the NEM. SST was also higher in the PS than the SCS during both seasons ($p < 0.05$). On the other hand, D_M was deeper during the NEM than during the SWM in both

regions ($p < 0.01$) and was deeper in the PS than the SCS during both seasons ($p < 0.05$). The SI was higher in the SCS than the PS during the SWM ($p < 0.05$), but during the NEM, it was comparable between the regions ($p > 0.05$) (Table 3).

The SCS had significantly higher N_I and P_I ($p < 0.01$) than the PS during both seasons. The SCS also had significantly ($p < 0.05$) higher N_S and P_S than the PS during the NEM. However, there were no differences in N_S and P_S between the two regions during the SWM (Table 3). Both D_N and D_P were deeper in the PS than the SCS during both seasons ($p < 0.01$). In the SCS, N_S and P_S were significantly ($p < 0.05$) higher during the NEM than the SWM. In contrast, seasonal variations in N_S and P_S in the PS were not significant ($p > 0.05$) (Table 3). Significant differences

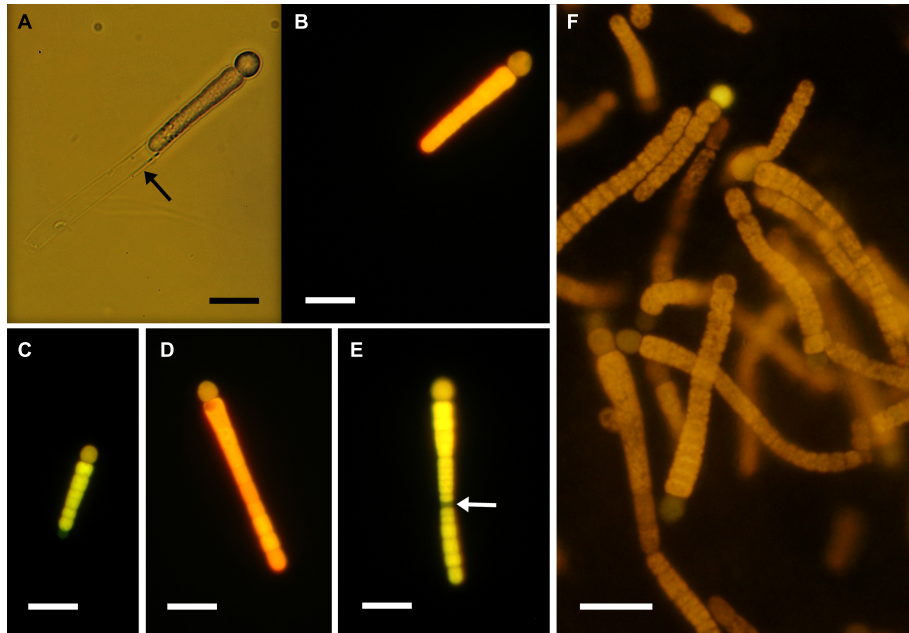


FIGURE 4 | (A–E) Solitary or **(F)** clumping filaments of *Calothrix rhizosoleniae* strain M15 under **(A)** bright field and **(B–F)** epi-fluorescence microscopy. Black and white arrows indicate empty sheath and necridium, respectively. Scale bar: 10 μm .

in D_{CM} were observed seasonally or regionally ($p < 0.01$). Seasonal differences in Chl were also observed in the SCS but not in the PS. Ratios of N:P ($N_S:P_S$ and $N_I:P_I$) were consistently below the Redfield ratio of 16 (Redfield, 1958), suggesting that phytoplankton growth was limited by N in both regions (Table 3).

In summary, the SCS-NEM stations had the lowest SST, the highest N_S , P_S , and Chl, and the shallowest D_{CM} . On the other hand, there were significant regional differences in nutrient concentrations; i.e., higher N_I and P_I and shallower D_N and D_P in the SCS than the PS (Table 3).

Identification of the Cultured FL-C

The isolated FL-C strain, M15, was positively identified as *C. rhizosoleniae* according to phylogenetic (Figure 3) and morphological examinations (Figure 4). Phylogenetic trees reconstructed from *nifH*-deduced amino acid sequences revealed that the isolated strain was included in the het-3 clade (Figure 3) that contained *C. rhizosoleniae* strain SC01, which was isolated from the North Pacific (Foster and Zehr, 2006). In the analysis of *nifH* DNA sequence, the strain showed high identity to *C. rhizosoleniae* SC01 (het-3) (99.6–100%). In contrast, its identity to *Richelia* associated with *Rhizosolenia* (het-1) or *Hemiaulus* (het-2) was $\leq 91.3\%$ (Supplementary Table 1). Morphologically, a filament of the isolate consisted of several vegetative cells followed by one basal heterocyst covered with thin hyaline sheath and lacked hair formation (Figures 4A,B). The trichome length in maximum and in minimum recorded two and 114 cells, respectively. The heterocyst was often smaller (Figures 4C,D) or equal in diameter to its adjacent vegetative cell (Figures 4A,B,E,F). Most of the newly divided filaments had slightly tapering ends and sometimes empty sheaths near

their ends (Figures 4A–C). Some longer filaments tapered in the center and had a developing rounded end-cell at its opposite end suggesting the filament was getting ready to divide (Figures 4D,E). The narrowest cell at the middle part would become a necridium (Figure 4E) from which a filament would divide into the longer mother filament the shorter hormogonium.

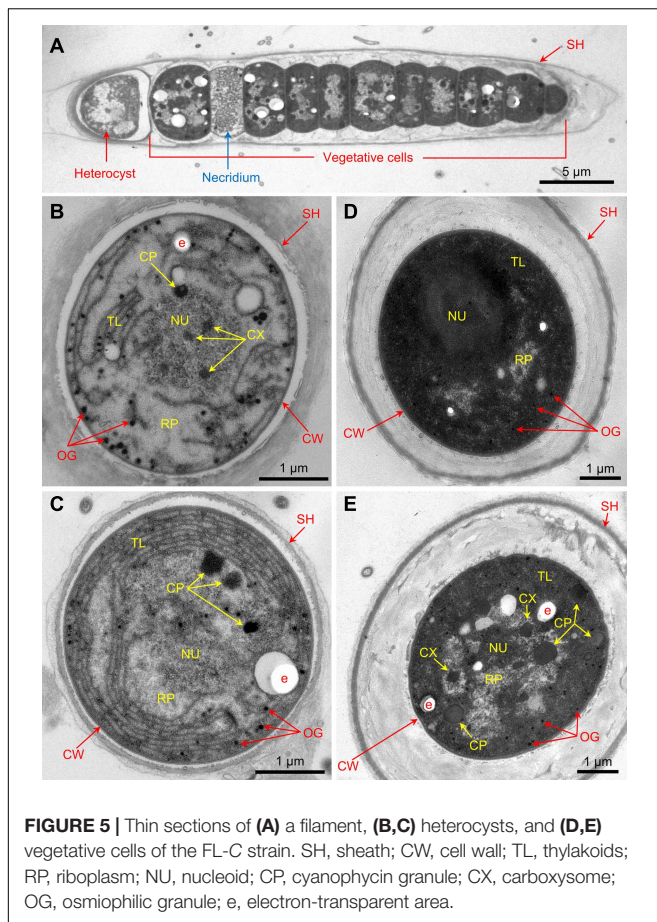
Ultrastructure of Cultured FL-C

The images of thin sections showed no gas vesicles in either vegetative cells or heterocysts in the cultured FL-C (Figure 5). The photosynthetic thylakoids were more abundant and denser in vegetative cells (Figures 5D,E) but distributed sparsely in heterocysts (Figures 5B,C) and necridia (Figure 5A and Supplementary Figure 8). The difference in thylakoid abundance among vegetative cells, heterocysts, and necridia was apparent from epifluorescence microscopy, i.e., vegetative cells often had brighter fluorescence than heterocysts and necridia (Figures 4B–F). Carboxysomes, cyanophycin granules, osmiophilic granules, and electron-transparent areas were found in both heterocysts and vegetative cells (Figure 5).

DISCUSSION

Different Trichome Lengths Among Host-Specific Symbionts

The field results showed that the trichome lengths were different among the three types of *Richelia/Calothrix* symbionts, classified by Nieves-Mori3n et al. (2020). The internal symbiont (*H-R*) had the shortest trichome lengths, followed by the external symbiont (*Calothrix*). The partial/periplasmic symbiont (*R-R*, *FL-R*, and



perhaps *G-R*) had the longest trichome lengths (Figure 2). According to a coarse-grained cell flux model of the *Hemiaulus-Richelia* symbiosis (CFM-DDA), the *Hemiaulus* host provided its symbiont with two-thirds of its fixed carbon (C) requirement (Inomura et al., 2020). Because the *Hemiaulus* host can provide such a large proportion of their fixed C requirement, an *H-R* symbiont may not need to maintain as many vegetative cells and so form shorter trichomes (Sundström, 1984; Villareal, 1994; Foster and Zehr, 2006; Foster et al., 2007; Bar Zeev et al., 2008; Lyimo, 2011; Hilton et al., 2013), consistent with our field observations (Table 4). In addition, a reduction in *H-R* symbiont vegetative cells/trichome length may facilitate their dwelling in the compact cytoplasm of their *Hemiaulus* hosts (Caputo et al., 2019). On the other hand, external symbionts (*C-C* and *B-C*) (Lemmermann, 1905; Janson et al., 1999; Foster and Zehr, 2006) and periplasmic symbionts (*R-R*) (Schmidt, 1901; Janson et al., 1999; Zehr et al., 2007; Bar Zeev et al., 2008; Kulkarni et al., 2010; Padmakumar et al., 2010; Lyimo, 2011; Jabir et al., 2013; Madhu et al., 2013; Table 4) may require more vegetative cells to satisfy their fixed C demand as their symbiotic locations lead to less-efficient exchange of molecules (e.g., fixed C and N) than internal symbionts. The comparable trichome lengths between *R-R* and *G-R* (Figures 2B,C) suggest that *G-R* is also a periplasmic symbiont. The fact that both FL-*R* and FL-*C* showed longer trichomes (both average and maximum

trichome lengths) than their symbiotic counterparts (*R-R* and *C-C*) (Figure 2) was also predicted by the CFM-DDA (Inomura et al., 2020). This study reports the first quantification of *G-R*, FL-*R*, *B-C*, and FL-*C* trichome lengths from natural waters. For *H-R*, *R-R*, and *C-C*, our observations of trichome length from the SCS and PS were comparable with those reported from other oceanic regions (Table 4). In a previous global model study, the C biomass of *Calothrix* and *Richelia* were calculated by using a fixed number of vegetative cells per trichome (trichome length minus 1) without considering variations in trichome length for symbionts associated with different diatom hosts (Luo et al., 2012). Our results suggest that trichome length varies based on the nature of the DDA, and this should be considered in future models.

The exceptionally short trichome length of *H-R* observed in this study (Figure 2A) was similar to an earlier study (Table 4; Foster et al., 2007). The short trichomes of *H-R* are thought to be strongly manipulated or restricted by their hosts; they have an extremely slimmed genome and are located in the host's cytosol (Hilton et al., 2013; Caputo et al., 2019). Unlike *Calothrix*, which is considered a facultative symbiont or an opportunist, *H-R* is unable to live alone without its host. The limited field data from this study suggests that all of the observed FL-*R* filaments were similar in length to those associated with *Rhizosolenia* and *Guinardia*. In a previous study, it was found that none of the FL-*R* filaments were derived from *Hemiaulus* (Tuo et al., 2017). *Rhizosolenia*-associated *Richelia* may be able to live outside of its host for at least a short time because its genome is larger than that of *Hemiaulus*-associated *Richelia*, probably of metabolic necessity since its symbiotic location is just outside the host's cytosol (Villareal, 1990; Hilton, 2014; Caputo et al., 2019). Microscopic evidence from field samples have shown *Richelia* filaments escaping from moribund or dead *Rhizosolenia* hosts by direct penetration through the frustule or attached on the plasmalemma of *Rhizosolenia* lacking a frustule (Villareal, 1989; Gómez et al., 2005) supporting this assertion.

Although the average trichome lengths between FL and symbiotic *Calothrix* were comparable, the maximum lengths of the FL-*C* trichomes were much longer than *B-C* and *C-C* (Figures 2E-G), implying that *Bacteriastrum* and *Chaetoceros* hosts prefer their epiphytic symbionts to be short. In addition, with only a single heterocyst, a long filament might not be as beneficial to their host in providing fixed N (Foster et al., 2011). As an epiphyte, a long filament might also increase N loss to the surrounding seawater due to the inefficiency of a long-trichome N transport. In addition, longer filaments have more vegetative cells thereby increasing the N demand of the cyanobacterial symbiont itself. Overall, shorter *Calothrix* seemed to benefit their *Chaetoceros* and *Bacteriastrum* hosts by not weighing them down and providing better transfer efficiency for the recently fixed N.

Spatiotemporal Variations of Trichome Lengths

Field results showed significant regional and seasonal differences in the trichome lengths of *H-R* and FL-*C*; they were longer in the SCS than the PS and longer during the NEM than the SWM.

TABLE 4 | Trichome lengths of *Calothrix* and *Richelia* symbionts by region.

Type	Study area	Trichome length (cells/trichome)			References
		Minimal	Maximal	Average	
<i>Richelia</i>	Subtropical North Pacific Ocean	2	7	–	Zehr et al. (2007) [†]
<i>H-R</i>	Western Philippine Sea	1	5	3*	This study
	Northern South China Sea	1	5	4*	This study
	Malacca Strait	2	6	–	Sundström (1984)
	Western Indian Ocean	–	–	5	Lyimo (2011)
	Eastern Mediterranean Sea	4	7	–	Bar Zeev et al. (2008)
	Western North Atlantic Ocean and Caribbean Sea	6	11	–	Villareal (1994)
	Western tropical North Atlantic Ocean	1	5	4	Foster et al. (2007)
	Tropical North Atlantic Ocean	2	3	–	Foster and Zehr (2006)
	Laboratory cultures (Gulf of Mexico)	4	6	–	Hilton et al. (2013)
<i>R-R</i>	Subtropical North Pacific Ocean	4	5	–	Foster and Zehr (2006)
	Western Philippine Sea	3	15	7*	This study
	Northern South China Sea	3	15	8*	This study
	Malacca Strait and Gulf of Thailand	7	20	–	Schmidt (1901)
	Bay of Bengal	9	11	–	Kulkarni et al. (2010)
	Palk Bay	15	24	–	Madhu et al. (2013)
	Southeastern Arabian Sea	13	20	–	Jabir et al. (2013)
	Northern Arabian Sea	10	15	–	Padmakumar et al. (2010)
	Western Indian Ocean	5	15	–	Lyimo (2011)
	Eastern Mediterranean Sea	–	11	–	Bar Zeev et al. (2008)
	Western tropical North Atlantic Ocean	–	–	5	Foster et al. (2007)
	The Bahamas and northern Caribbean Sea	5	11	–	Janson et al. (1999)
<i>G-R</i>	Western Philippine Sea	5	11	7*	This study
	Northern South China Sea	4	15	8*	This study
<i>FL-R</i>	Western Philippine Sea	4	22	8*	This study
	Northern South China Sea	5	15	9*	This study
<i>Calothrix</i>	Subtropical North Pacific Ocean	4	8	–	Lemmermann (1905) [‡]
<i>C-C</i>	Subtropical North Pacific Ocean	4	5	–	Foster and Zehr (2006)
	Western Philippine Sea	3	12	5*	This study
	Northern South China Sea	3	14	5*	This study
	The Bahamas and northern Caribbean Sea	5	11	–	Janson et al. (1999)
<i>B-C</i>	Northern South China Sea	4	7	5*	This study
<i>FL-C</i>	Laboratory cultures (North Pacific strain)	–	>11	–	Foster et al., 2010
	Laboratory cultures (North Pacific strain)	2	4	–	Nieves-Mori6n et al. (2020)
	Western Philippine Sea	3	17	6*	This study
	Northern South China Sea	3	19	6*	This study
	Laboratory cultures (South China Sea strain)	2	114	–	This study

*SWM and NEM data combined, see **Table 3** for the seasonal differences. [†]Not host specific. [‡]Reported as epiphytes on *Rhizosolenia* and *Hemiaulus*.

Similarly, FL-R trichomes were longer in the SCS than in the PS during both seasons, whereas R-R and G-R trichomes were longer in the SCS than the PS only during the SWM. The trichomes of R-R and G-R were also longer during the NEM than the SWM in the PS and the SCS, respectively. Such spatiotemporal differences in trichome lengths were accompanied by variations in nutrient supplies among regions and seasons. Both higher nutrient inventories (N_I and P_I) and shallower nutriclines (D_N and D_P) were observed in the SCS than in the PS during both seasons. In the SCS, higher surface nutrient concentrations (N_S and P_S) seemed related to longer trichome lengths during the NEM than the SWM. In the PS, although N_S and P_S were seasonally comparable, relatively high nutrient fluxes, reflected

in the lower SI and deeper D_M , may have contributed to the longer trichome lengths during the NEM. Furthermore, higher Chl and shallower D_{CM} also reflected the relatively high nutrient supplies in the SCS and during the NEM. The correlation matrix also showed positive relationships between trichome length of H-R and nutrients (N+N or P); however, the correlation coefficients were very low ($r < 0.10$) (**Supplementary Table 2**). Like nutrients, certain physical variables including SST and SPAR also showed significant regional and seasonal differences (**Table 2**). The correlation matrix also showed negative relationships between trichome lengths of H-R, R-R, and FL-C and temperature (**Supplementary Table 2**). However, temperature (or SST) was usually highly correlated with nutrients

TABLE 5 | Similarities of gas vesicle structural proteins (Gvp) between cyanobacterial diazotrophs and *Dolichospermum flos-aquae* CCAP 1403/13F (GenBank: U17109.1) by BLASTp.

Strain [†]	GvpA	GvpC	GvpN	GvpA/J	GvpK	GvpF/L	GvpG	GvpV	GvpW	GvpP
<i>Aphanizomenon flos-aquae</i> MDT14a	96.8	88.1	93.9	71.9	93.1	91.1	88.3	84.9	90.2	88.4
<i>Aphanizomenon flos-aquae</i> NIES-81	95.5	72.3	75.3	72.4	74.0	84.3	91.9	81.8	79.0	88.7
<i>Aphanizomenon flos-aquae</i> WA102	98.7	99.5	99.4	94.6	99.2	98.8	99.3	99.1	98.7	88.7
<i>Calothrix</i> sp. NIES-2098	81.2	29.4	50.2	32.6	63.2	47.7	45.8	48.4	50.9	69.9
<i>Calothrix</i> sp. NIES-2100	92.4	64.0	74.5	43.2	81.1	76.7	55.4	65.1	69.3	69.1
<i>Calothrix</i> sp. NIES-3974	na	na	na	na	na	na	na	na	na	62.6
<i>Calothrix</i> sp. NIES-4071	na	na	na	na	na	na	na	na	na	58.3
<i>Calothrix</i> sp. NIES-4105	na	na	na	na	na	na	na	na	na	58.3
<i>Calothrix brevissima</i> NIES-22	88.5	67.5	76.6	40.7	77.6	73.8	60.1	56.2	32.6	68.0
<i>Calothrix desertica</i> PCC 7102	na	na	na	na	na	na	na	na	na	59.9
<i>Calothrix elsteri</i> CCALA 953	na	na	na	na	na	na	na	na	na	62.9
<i>Calothrix parasitica</i> NIES-267	na	na	na	na	na	na	na	na	na	59.4
<i>Calothrix rhizosoleniae</i> SC01	na	na	na	na	na	na	na	na	na	69.3
<i>Nostoc punctiforme</i> PCC73102	82.2	28.0	49.4	33.5	63.0	60.8	48.2	49.1	51.1	65.9
<i>Nostoc punctiforme</i> NIES-2108	89.5	48.1	59.3	41.6	78.8	73.9	56.6	57.5	67.4	69.1
<i>Richelia intracellularis</i> RC01	na	na	na	na	na	na	na	na	na	60.2
<i>Tolypothrix</i> sp. PCC 7601	91.4	67.5	73.4	68.9	76.7	76.0	57.8	65.4	69.9	68.5
<i>Tolypothrix</i> sp. NIES-4075	79.9	na	63.1	38.6	70.7	70.3	50.1	56.0	na	68.5
<i>Tolypothrix bouteillei</i> VB521301	81.8	na	63.8	34.5	68.8	68.0	51.6	55.7	57.0	67.5
<i>Tolypothrix tenuis</i> PCC 7101	91.7	67.1	73.4	41.1	77.5	75.4	55.2	65.1	70.5	68.5
<i>Trichodesmium erythraeum</i> IMS101	31.2	na	64.0	35.1	63.0	56.7	50.1	na	50.8	49.7

[†]See **Supplementary Tables 3, 4** for genome information and Gvp accession numbers, respectively.

(**Supplementary Table 3**; Tuo et al., 2017). Nutrient stress has been considered a cause of short trichome length in cyanobacteria (Smith and Gilbert, 1995). For example, when a freshwater cyanobacterium *Oscillatoria agardhii* was under a period of P limitation, its cell phosphorus content declined and its trichomes became progressively shorter (Gibson and Stevens, 1979). Our results imply that higher nutrient concentrations (especially P) might support the production of longer trichomes of both *Richelia* and *Calothrix*.

In contrast to the other associations and free-living *Richelia* and *Calothrix*, C-C had comparable trichome lengths regardless of the region or season. Unlike endosymbiotic *Richelia*, which inhabit space inside the host cells, C-C attach to the hosts' apertures and are oriented transversely to the long axis of the chains (Gómez et al., 2005; Tuo et al., 2014). As a facultative symbiont or an opportunist, a filament of C-C could easily detach from its host and lengthen, especially under replete nutrient conditions (Tuo et al., 2017). The clear spatiotemporal length variations of the FL-C trichome relative to C-C and B-C are reflected in **Table 4**.

It should be noticed that regional and/or seasonal differences in the mean trichome length was around one vegetative cell for most types of *Calothrix* (~1.0 cells) and *Richelia* (0.3–1.7 cells), except FL-C and G-R (**Table 2**). The larger difference for G-R (4.1 cells) was likely due to the very small sample size at the SCS-NEM stations ($n = 2$). When ignoring the SCS-NEM sample, the regional difference during the SWM for G-R was also around one cell. On the other hand, FL-C showed larger spatiotemporal differences (0.5–3.2 cells), implying stronger influences from ambient nutrient availability. However, the small variation in

symbiotic *Calothrix* and *Richelia* trichome length (i.e., one cell) implies that nutrient availability in this study area was not a primary control on trichome length in the SCS and PS.

Lack of Gas Vesicles in *C. rhizosoleniae* Cells

Gas vesicles are common cellular components in freshwater species of heterocystous cyanobacteria such as *Dolichospermum* spp., *Aphanizomenon* spp., *Nostoc* spp., and *Calothrix epiphytica*, which usually have elongated trichomes with tens to hundreds of cells per trichome (West and West, 1897; Walsby, 1994; Wacklin et al., 2009). We found that *C. rhizosoleniae* lacks gas vesicles (**Figure 5**), similar to *Richelia*, suggesting poor buoyancy regulation or an inclination to sink (Janson et al., 1995). The lack of gas vesicles in cultured *C. rhizosoleniae* might have been the result of acclimation to cultivation; cultures were isolated in December 2007 and harvested for TEM in March 2015. To support our ultrathin section results (**Figure 5**), we also examined gas vesicle structural proteins (Gvp) in the genomes of *C. rhizosoleniae*, *R. intracellularis*, and other representative filamentous cyanobacteria to those of *Dolichospermum flos-aquae* CCAP 1403/13F (GenBank: U17109.1). The similarity comparisons showed that *C. rhizosoleniae* (SC01, a FL strain isolated from *Chaetoceros* hosts), *R. intracellularis* (RC01, associated with *Rhizosolenia* hosts), and certain species of *Calothrix* lacked the sequences of the structural protein (GvpA), large outer surface protein (GvpC), and putative gas vesicle protein (GvpN, GvpA/J, GvpK, GvpF/L, GvpG, GvpV, and GvpW) (**Table 5** and **Supplementary Tables 4, 5**). While

GvpP was found and is also a putative protein related to gas vesicles, it showed no obvious amino acid identity to the other *gvp* gene products (especially GvpA) (Li and Cannon, 1998). Thus, the genomic information supported the observation that *C. rhizosoleniae* and *R. intracellularis* cells lacked gas vesicles. Without the ability to produce gas vesicles, cyanobacterial species must regulate their buoyancy by turgor collapse and accumulation of condensed carbohydrate (Reynolds et al., 1987), and/or they might decrease their filament volume by shortening trichome length to control their buoyancy/sinking, if the Stokes' Law worked for them.

SUMMARY

The trichome length of diatom host-associated symbionts may be controlled by their hosts/symbiotic styles. On average, the internal *Richelia* symbiont (*H-R*) produced the shortest trichomes, followed by the external *Calothrix* symbionts (*C-C* and *B-C*); the periplasmic *Richelia* symbionts (*R-R* and perhaps *G-R*) produced the longest trichomes. The trichome lengths of *FL-C* and *FL-R* were comparable with their symbiotic counterparts *C-C* and *R-R*, respectively. Regional and/or seasonal differences in trichome length were observed for various types of *Richelia* and *FL-C*. However, the difference in length was only about one cell per trichome. Both *Richelia* and *Calothrix* (*rhizosoleniae*) may have limited buoyancy regulation due to their lack of gas vesicles and the ability to produce them. This may require them to maintain shorter trichome lengths unlike other filamentous cyanobacterial species that have gas vacuoles.

DATA AVAILABILITY STATEMENT

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article/**Supplementary Material**.

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AUTHOR CONTRIBUTIONS

ST and YLLC conceived the idea. ST, MRM, and YLLC wrote the manuscript with contributions from YT. ST, YT, WNJ, and YHL performed the experiments and created the figures and tables. ST, MRM, YLLC, YT, and HYC revised the manuscript. All authors reviewed the manuscript.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fmars.2021.678607/full#supplementary-material>

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Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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