



Multiple Ecological Parameters Affect Living Benthic Foraminifera in the River-Influenced West-Central Bay of Bengal

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The huge riverine influx and associated processes decrease the ambient salinity, stratify the water column, modulate the oxygen-deficient zone, and are also responsible for the recent acidification in the Bay of Bengal. Here, we have studied the effect of these riverine influx-dominated ecological parameters on living benthic foraminifera in the west-central Bay of Bengal. We report that the pH below 7.6 in front of the Krishna river, reduces the diversity and the richness of living benthic foraminifera on the adjacent shelf and the slope. A similar decreased diversity and richness is also observed in front of the Godavari River. We delineate three prominent assemblages, representing different depth zones with associated distinct physico-chemical conditions. The shallow water assemblage (~27-100 m) is represented by Nonionella labradorica, Hanzawaia nipponica, Brizalina dilatata, Ammonia tepida, and Nonionella limbato-striata. These species are adapted to relatively warmer temperatures and more oxygenated waters. The deepwater assemblage (~1,940-2,494 m) includes Bulimina cf. delreyensis, Bulimina marginata, Hormosinella guttifera, Cassidulina laevigata, and Gyroidinoides subzelandica and can tolerate a relatively colder temperature. The intermediatedepth assemblage (~145-1,500 m) dominated by Eubuliminella exilis, Bolivinellina earlandi, Fursenkoina spinosa, Bolivinellina lucidopunctata, Globobulimina globosa, Fursenkoina spinosa, Eubuliminella cassandrae, Uvigerina peregrina, Rotaliatinopsis semiinvoluta, and Cassidulina laevigata, represents oxygen-deficient and organic carbon-rich environment. Besides the pH, temperature, dissolved oxygen and organic matter, we also report a strong influence of bathymetry, coarse fraction (CF) and the type of organic matter on a few living benthic foraminifera. The ecological preferences of 40 such dominant living benthic foraminifera, each representing a specific environment, have also been reported for site-specific proxy. We conclude that although the huge riverine influx affects living benthic foraminifera on the shelf, the dissolved oxygen and organic carbon mostly control benthic foraminiferal distribution in the deeper west-central Bay of Bengal.

Keywords: foraminifera, ecology, organic matter, pH, Bay of Bengal, riverine influx

INTRODUCTION

The huge riverine influx creates a unique environment in the Bay of Bengal. A few of the world's largest snow and rainfall-fed rivers (Irrawaddy, Ganga, Brahmaputra, Mahanadi, Krishna, and Godavari) drain freshwater and sediments into the Bay of Bengal, creating a specific environment with associated physical and chemical parameters. The riverine influx reduces the salinity, warms the seawater and creates strong stratification (Sengupta et al., 2006). The associated sediment influx contributes enormous nutrients, sustaining the primary productivity (Prasanna Kumar et al., 2002). In addition to the riverine influx, the anthropogenic input (sulfate and nitrogen aerosols) to the Bay of Bengal significantly affects its biogeochemistry (Sarma et al., 2016). Therefore, the riverine influx primarily modulates the ambient physicochemical conditions in the northern Bay of Bengal. Additionally, the dissolved oxygen, a critical requirement of a large fraction of marine biota, is also perennially low at the Bay of Bengal's intermediate depths (Sarma et al., 2013). As suggested by the acidification and a possibility of the bay turning into the foremost nitrogen contributor soon (Bristow et al., 2017), the increasing stress implies that it is essential to understand the effect of these critical ecological parameters on the marine biota.

Benthic foraminifera are the dominant marine biota in the continental margin sediments and are the primary component of the global carbon cycling. The large abundance, combined with the sensitivity to the ambient conditions and long-term preservation in sediments, makes benthic foraminifera a widely used proxy to reconstruct paleoceanographic changes (Saraswat, 2015; Saraswat et al., 2017). The precise knowledge of the influence of ambient environmental parameters on abundance and distribution, is a prerequisite to use benthic foraminiferal characteristics for paleoceanographic and paleoclimatic interpretation (Murray, 2001; Gooday, 2003; Jorissen et al., 2007; Gooday and Jorissen, 2012; Saraswat and Nigam, 2013). Various environmental parameters, including food, oxygen concentration, salinity, pH, temperature, substrate, water depth and others, influence the diversity and abundance of benthic foraminifera (Lutze and Coulbourn, 1984; Kaminski et al., 1988; Altenbach, 1992; Mackensen et al., 1995; Gooday and Rathburn, 1999; Van der Zwaan et al., 1999; Ernst et al., 2002; Altenbach et al., 2003; Lei et al., 2017, 2019). The relative influence of various factors on faunal distribution, however, varies from region to region. The salinity and temperature are the major factors in riverine influx dominated continental shelves (Manasa et al., 2016). However, the deeper depth continental slope assemblages are mainly controlled by the availability of food and dissolved oxygen concentration (Gooday, 1986; Corliss and Chen, 1988; Mackensen and Douglas, 1989; Corliss and Emerson, 1990; Barmawidjaja et al., 1992; Jorissen et al., 1992; Rosoff and Corliss, 1992; Rathburn and Corliss, 1994; Nisha and Singh, 2012; Singh et al., 2015a, 2018; Verma et al., 2018). Benthic foraminiferal abundance, diversity, species richness and evenness also depend on ambient conditions (Gibson and Buzas, 1973; Buzas and Culver, 1991; Murray, 2006; Singh et al., 2015a,b). The overall benthic foraminiferal abundance and diversity decreases, whereas the abundance of abnormal or stress-tolerant specimen increases, under both naturally and anthropogenically stressed environment (Alve, 1995; Saraswat et al., 2004; Nigam et al., 2007). As the extent of stress varies regionally, the benthic foraminiferal response has to be assessed from different regions.

The fossil benthic foraminiferal assemblage is often biased by the biological and taphonomical processes, including transport, test disintegration or dissolution (Murray, 1991; Jorissen and Wittling, 1999; Berkeley et al., 2014). Therefore, living benthic foraminifera (rose-Bengal stained) provide a comparatively more reliable response to the ecological conditions. The previous studies on the distribution and ecology of living benthic foraminifera (rose-Bengal stained) from the northern Bay of Bengal are rare (Boltovskoy, 1978). The few studies documenting living benthic foraminifera from the Bay of Bengal cover a limited spatial extent, concentrating only on nearshorebeaches and estuaries (maximum 19 m water depth) (Gandhi et al., 2007; Gandhi and Solai, 2010). The influence of deep water ecological conditions on benthic foraminifera from the Bay of Bengal is rarely documented. Recently, Suokhrie et al. (2020), compared the living benthic foraminifera of the western Bay of Bengal's oxygen-deficient zone with that from a similar region in the Arabian Sea. A distinctly different benthic foraminiferal assemblage was reported from the Bay of Bengal oxygen deficient waters. However, the effect of other stressors, including salinity, temperature, pH, riverine influx on benthic foraminifera, both above and below the oxygen-deficient zone of the western Bay of Bengal, was not addressed. Therefore, we study the combined effect of multiple ambient parameters on the living (stained) benthic foraminiferal assemblage from the shallow inner shelf to the slope and further deeper region of the west-central Bay of Bengal. The main objective of this study is to establish representative living (rose-Bengal stained) benthic foraminifera assemblages characterizing a particular set of ecological niches on the shallow continental shelf to the abyssal depth in the westcentral Bay of Bengal, where the freshwater influx and associated processes primarily influence the physico-chemical parameters. The baseline information will be beneficial to understand the foraminiferal response to climatic changes in the future. The precise information about benthic foraminifera's ecological preferences, will also be useful for a better reconstruction of the past environmental conditions.

OCEANOGRAPHIC SETTING

The west-central Bay of Bengal is strongly influenced by the riverine influx. Some of the world's largest rivers drain into the western Bay of Bengal, with an average runoff being Ganga-11,892 m³s⁻¹, Brahmaputra-16,186 m³s⁻¹, Godavari-3,180 m³s⁻¹, Krishna-1,730 m³s⁻¹, Pennar-95 m³s⁻¹, and Cauveri-664 m³s⁻¹ (Rao, 1975; Subramanian, 1979; Dai and Trenberth, 2002). The lithogenic components' contribution is \sim 39% in the northern bay and \sim 12.6% in the southern bay (Unger et al., 2003). The riverine influx is most intense during

the summer monsoon and is also responsible for the salinitycontrolled stratification (Gomes et al., 2000; Unger et al., 2003; Sarma et al., 2016) and surface circulation in the Bay of Bengal. The southwestern Bay of Bengal receives significant precipitation during the winter monsoon. The surface circulation (East India Coastal Current) reverses seasonally along the bay's western boundary, flowing southward during the winter season and northward during the summer season (Vinayachandran et al., 1999; Shankar et al., 2002). The precipitation resulting from the seasonal reversal of winds in the Bay of Bengal is maximum during the summer monsoon (318 mm/month) as compared to the winter monsoon (88 mm/month) (Ramesh Kumar and Prasad, 1997). The summer precipitation causes substantial spatial variation in the salinity and temperature in the bay. The sea surface salinity varies from 29.1 psu closer to the continental margin on the inner shelf to 33.7 psu toward the open ocean. The warmer water input from the rivers is evident by the relatively higher sea surface temperature (29-30°C) in the river mouths' vicinity (Durand et al., 2011; Locarnini et al., 2013). The dissolved oxygen is low at intermediate depths (~100-1,000 m), with concentration being as low as 5 μ M (Wyrtki, 1971; Rao et al., 1994; Sardessai et al., 2007; Bristow et al., 2017). The shallower inner shelf water in the vicinity of the river has a higher dissolved oxygen. The chlorophyll-a concentration varies from $\sim 0.3 \text{ mg/m}^3$ to 2.0 mg/m³ in the western Bay of Bengal, with higher concentration closer to the river mouth (NASA Goddard Space Flight Center, Ocean Ecology Laboratory, Ocean Biology Processing Group, 2014). A strong wind-induced upwelling spread over \sim 40 km wide band, all along the eastern margin of India, is observed in the western Bay of Bengal, during the summer season (Shetye et al., 1991). As a consequence of this upwelling, salinity increases coastward (Shetye et al., 1991). The modern-day lysocline depth is at 2,000-2,600 m, shallowing from south to north in the Bay of Bengal (Cullen and Prell, 1984).

MATERIALS AND METHODS

Field Sampling

A total of 46 surface sediment samples (35 multi-core and 11 spade-core) were collected in 2014 and 2012, during the 308th cruise of ORV Sagar Kanya (SK 308) and RV Sindhu Sankalp (SSK 35) from the west-central Bay of Bengal, respectively (Figure 1 and Supplementary Table 1). The samples were collected along the coast perpendicular transects at regular depth intervals, starting from the continental shelf to the slope (27-2,494 m), during the pre-summer monsoon season of 2012 and 2014 (more details in Suokhrie et al., 2020). After retrieval, the top 10 cm (0-10 cm) of the sediment collected by using Ocean Science Industries Limited (OSIL) - Maxi multi-corer was sub-sampled at 1 cm interval. Half of the sediment from each section was immediately preserved in ethanol rose-Bengal solution (2 g rose-Bengal per liter of ethanol), to stain living benthic foraminifera. The top two sections (0-2 cm) were analyzed, as they contain the majority of the living benthic foraminifera in the northern Indian Ocean (Singh et al., 2018). The use of rose-Bengal stained living benthic foraminifera has been widely discussed since its

introduction by Walton (1952). A study by Walker et al. (1974) compared rose-Bengal and Sudan Black B stains and reported that rose-Bengal stain identified 70% of the specimens containing protoplasm. The concern also remains in differentiating the protoplasm living at the time of sample collection with the recently dead protoplasm (Bernhard, 1988). However, a majority of the tests lying in the sediments are usually devoid of protoplasm because of reproduction, predation, or growth stages (Murray and Bowser, 2000). Also, the living protoplasm (those living at the time of staining) tend to acquire deeper and denser stain up to one additional chamber. So, if the pattern of staining can be understood thoroughly, the error of over-estimating the stained living fauna can be reduced (Linshy, 2010).

The overlying bottom water in the multi-cores was collected at each station, and its salinity, pH and temperature were measured onboard by using a hand-held multimeter (S47-K/"SevenMulti") (**Supplementary Table 2**). The bottom water sample was not retrieved at a few multi-core stations. For these stations, salinity, temperature, dissolved oxygen, and also all the environmental data for the 11 spade core stations were downloaded from the World Ocean Atlas 2013 (Garcia et al., 2013; Locarnini et al., 2013; Zweng et al., 2013) by using the Ocean Data View software (Schlitzer, 2016, Ocean Data View)¹ (**Figure 2**).

Laboratory Analysis

The stained sediments were stored in cold-storage (4°C) for a minimum of 2 weeks and were then processed following the standard procedure (Manasa et al., 2016; Singh et al., 2018; Suokhrie et al., 2020). The excess ethanol rose-Bengal was removed, and the sediments were freeze-dried. The dried samples were then weighed, and ~ 2 g of the sediment was finely powdered for total carbon, nitrogen and inorganic carbon analysis. The rest of the sample was wet sieved by using a 63 μ m sieve. The >63 μ m material coarse fraction (CF) was dried, weighed and then transferred into plastic vials. A representative aliquot of the CF was weighed to pick a minimum of 300 living benthic foraminifera, wherever possible. All available living benthic foraminifera were picked from a fixed 0.1 g CF, in samples where a limited number of specimens were found. The picked specimens were mounted on the micropaleontological slides for counting and identification. Only those specimens with precise pink coloration with more than one chamber completely stained were picked and counted for living benthic foraminifera. All specimens were identified by using the previously published literature. The foraminiferal treatise (Loeblich and Tappan, 1988) was followed for genus confirmation, and for species confirmation, we have referred to the foraminiferal catalog (Ellis and Messina, 1940-2015). The genus of a few species have been shifted, as per the taxonomic shifts. Bolivinellina has been shifted from Bolivina following Saidova (1975). One species (Eubuliminella cassandrae) was also shifted from genus Bulimina to Eubuliminella following Revets (1993). Only those references mentioned in the main text of the paper have been added to the reference list. All other species identification references are mentioned in Supplementary Table 4 for further reading.

¹http://odv.awi.de



spade core stations. The major rivers draining in the area are marked by thin black lines. The bathymetry is marked by colored contours.

The sedimentary characteristics, namely total inorganic carbon (IC), total nitrogen (TN), and total carbon (TC), were analyzed by using dried and finely powdered sediment (\sim 10 mg weight). TIC was analyzed by using UIC CM 5015 CO₂ Coulometer and TC, TN by CNS elemental analyzer (Thermo Fisher Scientific). Calcium carbonate (CaCO₃) was calculated from IC and

organic carbon (C_{org}) was estimated by subtracting TIC from TC.

Statistical Analysis

To draw meaningful inferences from the large dataset of the relative abundance of all benthic foraminifera species identified from the western Bay of Bengal, 54 dominant species (\geq 3%









(d'Orbigny 1846); (7) Ammoglobigerina globigeriniformis (Parker and Jones, 1865); (8) Trochamminopsis quadriloba (Hoglund, 1947); (a) Ventral view (b) Dorsal view; (9) Eggerella propinqua (Brady, 1884); (10) Textularia oceanica Cushman, 1932; (11) Spiroloculina convexa Said, 1949, (a–c); (12) Quinqueloculina bicarinata (d'Orbigny, 1878); (a) 4-chambered view (b) Apertural view (c) 3-chambered view; (13, 14) Bolivinellina earlandi (Parr, 1950); (15) Bolivinellina lucidopunctata (Conato, 1964); (16) Bolivinellina pacifica (Cushman and McCulloch, 1942); (17) Bolivinellina translucens (Phleger and Parker, 1951); (18, 19) Brizalina dilatata (Reuss, 1850); (20) Bolivina mekranensis (Haque, 1970); (21) Brizalina spathulata (Williamson, 1858); (22) Latibolivina lepida; (23) Latibolivina persiensis (Lutze, 1974); (24) Cassidulina carinata Silvestri, 1896; (a) Ventral view showing aperture (b) Dorsal view; (25) Cassidulina laevigata (d'Orbigny, 1826), (a) Dorsal view, (b) Ventral view showing aperture. Scale bar is 100 μm.

abundance at a minimum of two stations) (**Figures 3–6**), were used for the Cluster analysis. Bray Curtis similarity index was applied through Q-mode cluster analysis by using Primer 6.1.10 software. The major clusters were identified with similar species having a particular percentage contribution in each cluster. The dominant assemblage representing a cluster and the relative contribution of each species to the respective cluster was identified by using the Simple Percentage (SIMPER) program at 100%. The canonical correspondence analysis (CCA) was done to understand the influence of multiple parameters on the dominant species. The multivariate statistical package version 3.1 (Kovach, 1998) was used for CCA. The CCA provides a better insight



FIGURE 4 | (1) Globocassidulina oblonga (Reuss, 1850); (a) Ventral view (b) Dorsal view; (2, 3) Islandiella cushmani (Stewart and Stewart, 1930); (a, b) Side views (c) Apertural view; (3) Eubuliminella cassandrae (Revets, 1993); (4, 5) Eubuliminella exilis (Brady, 1884); (6) Hopkinsinella glabra (Millett, 1903); (7) Bulimina cf. delreyensis (Cushman and Galliher, 1934); (8) Bulimina marginata (d'Orbigny, 1826); (9) Globobulimina globosa (Leroy, 1944) (10); Protoglobobulimina pupoides (d'Orbigny 1846); (11) Uvigerina globulosa (Egger, 1895); (12, 13) Neouvigerina ampullacea (Brady, 1884); (14) Uvigerina peregrina (Cushman, 1923); (15) Fursenkoina pauciloculata (Brady, 1884); (16) Fursenkoina spinosa (Heron-Allen and Earland, 1932); (17) Fursenkoina schreibersiana (Czjzek, 1848); (18) Rutherfordoides rotundiformis (McCulloch, 1977); (19) Suggrunda alata (Seguenza 1862); (a) Side view (b) Lateral view showing aperture; (20) Suggrunda semiclara (McCulloch 1977); (a) Side view (b) Lateral view showing aperture; (21) Baggina irregularis (McCulloch, 1977); (a) Ventral view (b) Dorsal view. Scale bar is 100 μm.

into the cumulative effect of a set of ambient environmental parameters on the abundance of species.

RESULTS

Bottom Water Parameters (Salinity, Temperature, Dissolved Oxygen, and pH)

The salinity at the sediment-water interface was higher in the shallow regions than at the deeper regions. The salinity ranged from \sim 32.15 psu to 35.2 psu (**Figure 2A**). The dissolved oxygen varies from \sim 0.3 ml/l to a maximum of \sim 4.9 ml/l. The dissolved oxygen is low within the depth range of \sim 100–1,000 m (**Figure 2B**). The previous reports suggest excessively



view (b) Dorsal view; (3) *Gavelinopsis mira* (Cushman, 1922); (a) Ventral view (b) Lateral view (c) Dorsal view; (4) *Pseudoeponides japonicum* (Uchio, 1950); (a) Dorsal view (b) Ventral view; (5) *Nonionella auris* (d'Orbigny, 1839); (a) Side view (b) Apertural view (c) Side view showing the last chamber flap covering the umbilicus; (6) *Nonionella limbato-striata* (Cushman 1931); (7) *Nonionellina labradorica* (Dawson, 1860); (a,c) Side views (b) Apertural view; (8) *Melonis* (?) *chathamensis* (McCulloch, 1977); (a,c) Side views (b) Apertural view; (9) *Pullenia bulloides* (d'Orbigny, 1846); (a,b) Side views (c) Apertural view. Scale bar is 100 μm.

low oxygen concentration (as low as 18 nM) at ~150–450 m (Bristow et al., 2017). The temperature at the sediment-water interface was relatively warmer (>23°C) in the shallow coastal zone as compared to the deeper region (<17°C) (**Figure 2C**). The seawater pH at the spade core stations was not available, and hence the data from the multicore stations only are plotted. The measured pH at the sediment-water interface in the study area ranges from 7.5 to 8.1. The region off Godavari and Pennar rivers has higher pH than off the Krishna river (**Figure 2D**).

Sediment Characteristics

The fraction retained on the sieve (>63 μ m) is considered as the CF and comprises both biogenic remains (shells and tests), and terrigenous material (quartz, mica, feldspars, and others). A relatively more CF is observed in the southern segment and at the shallower and deeper stations (**Figure 2E**). Overall, the upper slope stations (~250–1,000 m) have a comparatively higher finer fraction. CF varied from ~0.5% to an enormously high percentage upto ~90% at the southern stations. The influence of riverine influx on CF, by contributing terrigenous



material, is evident from the increase in CF in front of the river mouths, especially in the southern part of the study area (SC42, SC43, and SC44). Interestingly, the increased CF is also observed in the deeper region and is attributed to increased contribution from biogenic remains, including foraminiferal tests (**Figure 2E**).

The organic carbon varied from 0.3 to 1.8% in the west-central Bay of Bengal (Supplementary Table 2). The maximum %Corg was at the deeper depths. The influence of riverine discharge by contributing terrigenous organic matter and the fine fraction is also evident (Figure 2F). The organic carbon comes both from the terrigenous influx as well as marine productivity. These two sources' relative contribution is delineated by the organic carbon to nitrogen ratio of the organic matter (Corg/TN). Generally, $C_{org}/TN > 6.0$ is considered to be of terrestrial input, as vascular plants have nitrogen depleted organic matter (Müller and Mathesius, 1999; Ramaswamy et al., 2008), and values < 6.0 are used to infer marine input. A majority of the Corg/TN values are >6.0 and hence suggests that the organic matter is mainly contributed by terrestrial sources rather than by marine productivity. Expectedly, the maximum Corg/TN is infront of the Godavari, Krishna, and Cauvery rivers (SC37, SC38, MC30, MC31, SC40, SC39, MC27, and MC28) (Figure 2G).

The biogenic skeletal remains mainly contribute $CaCO_3$ to the sediments. $CaCO_3$ concentration in the west-central Bay of Bengal ranged from 1.3 to 87.2%. $CaCO_3$ concentration increases toward the southern part of the west-central Bay of Bengal. Additionally, $CaCO_3$ is low at a few stations on the lower slope (depth >1000 m), likely due to a reduction in the total for a miniferal abundance, which mainly contributes to the CaCO₃ in the sediments at these depths (**Figure 2H**).

Cluster Analysis

A total of 263 living benthic foraminifera have been identified (**Supplementary Tables 3, 4**). The cluster analysis groups similar objects together to draw meaningful inferences from a large dataset like the number of species and their relative abundance in the western Bay of Bengal (Kaufman and Rousseeuw, 2009; Khare et al., 2017). Based on the distribution at different stations, 54 dominant species were selected for the cluster analysis. We delineate four major clusters at 20% similarity index, other than a few insignificant ones (**Figure 7**). The clusters were named as Cluster I, II, III, and IV. The dominant assemblage representing a cluster and each species' relative contribution to the respective cluster is described below.

Cluster I

This cluster is represented by six stations, namely SC45, MC36, MC32, SC36, MC24, and MC09. The stations in this cluster are from a shallow depth (107–274 m) except for one station MC36 (1,511 m). The similarity percentage (SIMPER) analysis shows that this cluster has an average similarity of 20.59% consisting of 12 species. The dominant species are *Eubuliminella exilis* (24.88%), *Bolivinellina earlandi* (19.89%), *Fursenkoina spinosa* (12.54%), *Bolivinellina lucidopunctata* (8.76%), and *Globobulimina globosa* (6.48%).

Cluster II

Fursenkoina spinosa (15.64%), Eubuliminella cassandrae (11.41%), Rotaliatinopsis semiinvoluta (11.07%), Uvigerina

peregrina (10.4%), and *Cassidulina laevigata* (8.26%) are the dominant species in this cluster having an average similarity of 51.31%. This cluster represents a depth range of 500–1,518 m.

Cluster III

This cluster is represented by five stations (MC15, MC2, MC37, MC18, and MC38) representing a depth range of \sim 1,940–2,494 m with a similarity of 47.62%. Twelve species comprise of this cluster and the five most dominant ones are *Bulimina* cf. *delreyensis* (18.59%), *Bulimina marginata* (18.14%), *Hormosinella guttifera* (9.05%), *Cassidulina laevigata* (8.02%), and *Gyroidinoides subzelandica* (6.18%).

Cluster IV

The SIMPER analysis shows that the average similarity of this cluster is 34.53%. The representative benthic foraminiferal assemblage of this cluster includes nine species of which the five most dominant ones are listed here, *Nonionellina labradorica* (37.85%), *Hanzawaia nipponica* (17.58%), *Brizalina dilatata* (17.31%), *Ammonia tepida* (4.71%), and *Nonionella limbato-striata* (4.08%).

From the spatial distribution of various clusters, it is clear that Cluster I, Cluster II and Cluster III, and Cluster IV follow a depth zonation (**Figure 8**). Cluster IV is confined to the shallow water depth. Cluster I represents a shallow to intermediate depth zone covering the upper slope region. Cluster II mainly comprising of angular asymmetrical foraminifera, represents a wide range of intermediate water depth to deep water depth locations. The assemblages in this cluster consist of species that are indicative of an oxygen-deficient region. The species in Cluster III represent the deep-water species.





Canonical Correspondence Analysis

The dominant effect of one or a combination of several parameters, out of a set of multiple parameters in the field, on living benthic foraminiferal species, can be identified from CCA. Out of the 54 dominant species, only 40 species showed a significant relationship with atleast one of the ambient environmental parameters (**Figure 9**). The rest of the species do not show a significant relationship with any particular parameter, as evident from the close proximity of these species to the centroid in the CCA biplot (Ter Braak, 1986). From CCA, it is evident that the dissolved oxygen, CF, and temperature, are the most significant parameters controlling the living benthic foraminifera species distribution in the west-central Bay of Bengal. Besides these dominant factors, C_{org},

salinity, and $C_{\rm org}/TN$ also influence the distribution of living benthic foraminifera.

Species Representing Specific Set of Parameter

Species Preferring Oxic, Colder Water With Organic Matter Rich Sediments

Ammobaculites cf. agglutinans, Quinqueloculina bicarinata, Bolivinellina pacifica, Brizalina dilatata, Latibolivina lepida, Latibolivina persiensis, Fursenkoina pauciloculata, Eponides cribrorepandus, Nonionella auris, Nonionella limbato-striata, Nonionellina labradorica, Hanzawaia nipponica, and Ammonia tepida were positively correlated with dissolved oxygen and organic carbon, whereas negatively correlated with temperature



triangles represent the species (modified from Suokhrie et al., 2020).

(Figure 9). Ammobaculites cf. agglutinans, an agglutinated benthic foraminifer, was present only at four stations, all shallower than ~60 m. Quinqueloculina bicarinata was present at three stations, all shallower than 56 m. Even Bolivinellina pacifica was present only upto 76 m. Brizalina dilatata was found at stations mostly within depths of \sim 300 m, beyond which <1% abundance was observed at two stations. Latibolivina lepida was found only at two stations with abundance varying from 4.2% (53 m) to 6.2% (52 m). Likewise, Latibolivina persiensis was also present at shallow depths between 40 and 76 m, beyond which it was absent. Fursenkoina pauciloculata was abundant over a wider depth range (~30-1,000 m) but the highest abundance was at the shallower depths. Eponides cribrorepandus was present only at three stations with relative abundance varying from 0.15% (500 m water depth) to 25% (107 m water depth). It was totally absent at stations deeper than \sim 500 m. Nonionella auris was present at most of the stations and abundant between ${\sim}200$ m and \sim 1,000 m. Nonionellina labradorica was present upto a depth of ~750 m. Hanzawaia nipponica, Ammonia tepida and Nonionella *limbato-striata* were abundant at stations shallower than \sim 150 m.

Species Preferring Oxygen Deficient, Warmer, and Hypersaline Water

The species having a negative correlation with dissolved oxygen and a positive correlation with temperature and salinity were *Reophax scottii*, *Eggerella propinqua*, *B. earlandii*, *B. lucidopunctata*, *Bolivinellina translucens*, *Brizalina spathulata*, *Eubuliminella cassandrae*, *E. exilis*, *Hopkinsinella glabra*, *Protoglobobulimina pupoides*, *Uvigerina globulosa*, *Uvigerina peregrina*, *Fursenkoina spinosa*, *Rutherfordoides rotundiformis*, *Suggrunda semiclara*, *Baggina irregularis*, *Cancris oblongus*, *Pullenia bulloides*, *Chilostomella oolina*, and *Rotaliatinopsis semiinvoluta* (**Figure 9**).

Agglutinated benthic foraminifera, *Reophax scottii* was abundant only at four stations with abundance ranging from 0.14% (1,069 m) to 40.96% (259 m). Other agglutinated foraminifer, *Eggerella propinqua* was abundant at depths varying from \sim 200 m to \sim 700 m except for two shallow stations at 27 m and 31 m. The species of genus *Bolivinellina* reported in the west-central Bay of Bengal include *B. translucens*, *B. earlandi*, and *B. lucidopunctata*. The abundance of these

three species was influenced by bathymetry, with B. translucens and B. lucidopunctata being the most abundant between 76 m and \sim 500 m, while *B. earlandi* was abundant from 76 m upto \sim 2,000 m. Brizalina spathulata was the most abundant between \sim 100 m and 270 m. Eubuliminella cassandrae was abundant at depths $> \sim 150$ up to $\sim 1,000$ m, whereas *E. exilis* was present only upto a depth of 274 m. Hopkinsinella glabra was present only at 11 stations, with most of the stations being within \sim 200– 900 m. Protoglobobulimina pupoides was present only at depth $> \sim 250$ m. Uvigerina globulosa was present at depths $> \sim 750$ m except at one shallow station (55 m). Uvigerina peregrina was also dominant at deeper than 190 m with a maximum abundance of 21%. Fursenkoina spinosa was also abundant at stations >200 m deep, with a relative abundance ranging from 0 to a maximum of 36%. Rutherfordoides rotundiformis was present at stations deeper than ~ 100 m, except for one station at \sim 40 m. Suggrunda semiclara was present at seven stations, of which four stations were on the upper slope, within a depth range of ~390-500 m. Baggina irregularis, Pullenia bulloides and Rotaliatinopsis semiinvoluta were abundant at stations deeper than 500 m. Cancris oblongus was present only at nine stations with the highest abundance of 4.3% at 500 m depth. Similarly, Chilostomella oolina with a maximum relative abundance of 6.2% was present at stations deeper than \sim 200 m. Interestingly the maximum abundance of all the species listed above was within the oxygen deficient zone ($\sim 100-1,000$ m).

Species Preferring Coarse Sediments With High Corg/TN

Ammoglobigerina globigeriniformis, Trochammina quadriloba, Islandiella cushmani, Bulimina marginata, Bulimina delreyensis, Gavelinopsis mira, and Gyroidinoides subzelandica were positively correlated with CF and Corg/TN (Figure 9). Ammoglobigerina globigeriniformis, an agglutinated foraminifer was present at almost all the stations with the highest abundance (\sim 19%) at ~1,980 m. Trochammina quadriloba, another agglutinated for a minifer, was abundant at stations deeper than ~250 m, except for three shallower stations with abundance $\leq 2\%$. Islandiella cushmani was present at most of the stations in the study area but mostly abundant at stations deeper than ~ 200 m. Bulimina marginata, Bulimina delreyensis, Gavelinopsis mira, and Gyroidinoides subzelandica were abundant at the deeper stations $(>\sim 250 \text{ m})$ in the west-central Bay of Bengal. Although the CCA biplot shows influence of both CF as well as Corg/TN on these species, the influence of CF was more significant. Interestingly, Hormosina guttifera shows a positive correlation only with CF.

DISCUSSION

Differential Foraminiferal Abundance on the Shelf and Slope

Interestingly, the abundance of living (stained) benthic foraminifera between ~ 200 m and $\sim 1,000$ m was higher ($\sim 1,300$ specimen/g sediment), as compared to the continental shelf (~ 300 specimen/g sediment upto ~ 200 m) as well as at depths deeper than $\sim 1,500$ m (~ 46 specimen/g sediment)

(Suokhrie et al., 2020). We suggest that the lower dissolved oxygen concentration with an average of ~0.5 ml/l within the depth range of ~100–1,000 m and corresponding high organic matter content, dominantly control the living benthic foraminiferal population at these depths in the western Bay of Bengal. The high %C_{org} concentration between ~250 m and 1,500 m water depth is attributed to the relatively finer sediments at these depths. The finer sediments can preserve more C_{org} (Mayer, 1994). Additionally, there is an enormous influx of both freshwater and sediments in the northern Bay of Bengal. The riverine influx is also a source of organic carbon. The immense terrestrial organic carbon contribution is further confirmed by a clear pattern of very high C_{org}/TN (~20.0) close to the river mouth as compared to low C_{org}/TN, indicating marine organic carbon, away from the river mouth (>6.0).

The foraminiferal abundance is low at stations shallower than \sim 100 m water depth with sediments dominated by terrestrially sourced organic matter. This low abundance is attributed to the taphonomic processes, including more disturbance or turbulence from the riverine influx leading to less proliferation and diversification of benthic foraminifera (Berkeley et al., 2014). Besides, the %Corg at the shallow depths is also lower than the intermediate water depth, except in front of the Godavari river mouth. The enormous freshwater runoff in the bay results in ocean surface stratification and an oligotrophic condition and thus the low organic matter on the shelf. This is also a factor for the low living benthic foraminiferal abundance at the shallow depth stations as the organic carbon is the food for benthic foraminifera. The lack or low organic carbon content in sediments is detrimental to benthic foraminifera (Duffield et al., 2014). The coarser sediments close to the river mouth are one reason for the lower %Corg despite the high primary productivity at these depths, as evident from the chlorophyll-a content (NASA Goddard Space Flight Center, Ocean Ecology Laboratory, Ocean Biology Processing Group, 2014). The intermediate depths within ~100-1,000 m have abundant organic matter, which leads to reduced bottom water DO, and hence the abundance of total living abundance is a positive response to the organic matter supply. The increased foraminiferal abundance at these depths is also due to the significantly reduced predatory pressure as the macrofauna are comparatively more susceptible to oxygen-deficient environments. These depths are dominated by opportunistic species that thrive well in low DO concentrations. With a decrease in organic matter, the DO concentration increases beyond ~1,000 m water depth. A drastic decrease in foraminiferal abundance is seen below ~1,000 m water depth in the study area, and is attributed to the increase in predatory pressure. Additionally, the environment is more food limiting than oxygen and leads to a significant reduction in the total living fauna.

Drivers of Benthic Foraminiferal Diversity and Richness

The low diversity and species richness in front of the major rivers suggests a strong riverine influx. Both the diversity and species richness are low in the southern part, suggesting a restricted environment facilitating a select few species to survive (Suokhrie et al., 2020). Incidentally, the seawater pH is very low in front of the major river mouths. Although benthic foraminifera can tolerate a wide range of pH, comparatively less alkaline pH is detrimental to calcareous tests (Saraswat et al., 2015). Therefore, the low diversity in low pH regions is attributed to the thin-walled benthic foraminifera's inability to survive in adverse conditions. In contrast, more diverse microenvironments in the northern part are inferred from the high diversity and richness. From our earlier work based on CCA between benthic foraminiferal abundance and ambient parameters in the oxygen-deficient zone of this region, we inferred that the increased abundance of a few angular benthic foraminifera thriving as opportunistic species (F. spinosa, E. cassandrae, U. peregrina, R. semiinvoluta, S. semiclara, E. propingua, and B. irregularis) contributes to the overall more population of living benthic foraminifera in the oxygen-deficient zones on the slope. The dominance of a few opportunistic species is supported by low diversity and richness at these depths (Suokhrie et al., 2020). The present study's cluster analysis groupings also indicate the abundance of angular forms at the depths where dissolved oxygen is low. Such a decrease in diversity of living benthic foraminifera in a low dissolved oxygen environment has also been reported from the oxygen minimum zone (Jannink et al., 1998; Gooday et al., 2000; Schumacher et al., 2007; Suokhrie et al., 2020).

Living Benthic Foraminiferal Assemblages

From cluster analysis, we delineate three faunal assemblages based on significant correlation of the assemblages to a similar set of environmental parameters in the study area. Incidentally, the assemblages dominate different depth zones as well. The details of the representative assemblages are discussed below.

Assemblage I (27-107 m)

This assemblage comprises of Cluster IV. The dominant species are N. labradorica, H. nipponica, B. dilatata, A. tepida, and N. limbato-striata with minor contributions from *F. pauciloculata*, *B. pacifica*, *L. persiensis*, and *N. auris* (Figure 10). These species are abundant on the continental shelf ($<\sim$ 100 m water depth). The Assemblage I includes only calcareous benthic foraminifera. Nonionellina labradorica is a dominant species in this assemblage and has also been reported from the Corg rich fjords in the Arctic (Shetye et al., 2011). This species has been referred to as Arctic-boreo species (Seidenkrantz, 1993). Its high abundance is also reported from the oxygen-depleted environments where it sequesters chloroplast as it enables it to survive even in the anoxic conditions (Cedhagen, 1991; Bernhard and Bowser, 1999). In our study area, however, the abundant presence of N. labradorica is associated with the shallow inner shelf warm well-oxygenated waters with low organic carbon. We report another species belonging to the Assemblage I, namely H. nipponica, with its preference for shallow, well-oxygenated waters. From the Bay of Bengal, this species has been reported as a 'turbidity sensitive species' (Jayaraju et al., 2010).

Similarly, *B. dilatata* is also abundant on the shallow shelf, with a rare presence at a few stations upto \sim 700 m. Its abundance in

shallow to intermediate water depth is also reported by Gooday (1993) and Bernhard and Gupta (1999). Brizalina dilatata is also commonly reported from low oxygenated and high organic carbon-rich environments (Gupta and Machain-Castillo, 1993; Rouchy et al., 1998; Kuhnt et al., 2007). In the present study, this species correlates well with the shallow and warmer waters. The characteristic shallow-water species A. tepida is also abundant on the shelf of the present study. Ammonia tepida is widely distributed over estuaries, shelf and salt marshes (Murray, 1991; Nisha and Singh, 2012). Most workers have reported A. tepida as a pollution-sensitive species (Bergin et al., 2006; Burone et al., 2007; Frontalini and Coccioni, 2008; Elshanawany, 2011; Debenay et al., 2015; Schintu et al., 2016; Saalim et al., 2017). Ammonia tepida is also reported as a dominant species in the modern intertidal and estuarine areas (Nigam, 1984; Adarsh and Rajeshwara Rao, 2010). We also report A. tepida being abundant in shallow and warm water with a moderate organic matter supply in the study area. Nonionella limbato-striata is another dominant species in this assemblage, preferring the shallow and well-oxygenated waters.

Fursenkoina pauciloculata is a minor species in this assemblage. *Bolivinellina pacifica*, another dominant species of this assemblage, was earlier reported from the shallow water depths (Saidova, 2010), same as that in this study area. *Latibolivina persiensis* is reported from shallow depth (Sliter, 1969, 1970; Panchang and Nigam, 2014). It has been found to prefer oligotrophic and well-oxygenated water (Abu-Zied, 2013), similar to its distribution in this study, where this species is present only in shallow, well-oxygenated water (upto 76 m). Another minor species is *N. auris* in this assemblage.

The species belonging to Assemblage I representing the shallow locations indicate that they are positively correlated with dissolved oxygen, temperature and C_{org} . The dissolved oxygen concentration is high (>3.0 ml/l) and the organic matter content is low (~0.9%) with warmer temperature on the shelf. Thus, Assemblage I represents a shallow, well-oxygenated, warm environment with relatively high organic matter in the sediments.

Assemblage II (~145–1,500 m)

The dominant species in this assemblage are *E. exilis, B. earlandi, F. spinosa, B. lucidopunctata, G. globosa* from Cluster I and *F. spinosa, E. cassandrae, U. peregrina, R. semiinvoluta, C. laevigata* from Cluster II (**Figure 11**).

Eubuliminella exilis has been reported as deep infaunal taxa (Jorissen, 1999) and an indicator of low oxygenated environments and unchanged supply of organic matter (Caralp, 1989; Jannink et al., 1998; Schumacher et al., 2007). Amongst the dominant species belonging to Assemblage II, *F. spinosa* is a shallow infaunal species inhabiting waters upto a depth of \sim 120 m (Phleger, 1963). In the present study, *F. spinosa* is abundant at depths deeper than \sim 200 m, indicating less depth dependence and a large influence of dissolved oxygen. The relative abundance of infaunal foraminifera is controlled by the availability of both the metabolizable organic matter and dissolved oxygen. The infaunal foraminifera are abundant in organic matter rich but oxygen-deficient environments (Jorissen et al., 1995; Van der Zwaan et al., 1999). The higher relative



abundance of F. spinosa at depths >200 m within the low DO concentration in the western Bay of Bengal, is more in line with its infaunal nature. The species belonging to Bolivinellina have been reported from shallow regions (Nigam and Chaturvedi, 2006; Nagendra et al., 2011; Panchang and Nigam, 2014; Gandhi et al., 2016). Both B. earlandi and B. lucidopunctata have been recorded from the upper slope region in the study area. G. globosa is another dominant species in this assemblage. Generally, rectilinear foraminifera comprising of genera like Bolivina, Brizalina, Bulimina, Fursenkoina, and Uvigerina have previously been reported as a common indicator of low oxygenated environments by several workers (Gupta and Machain-Castillo, 1993; Kaiho, 1994; Murray, 2006; Nigam et al., 2009; Mazumder and Nigam, 2014; Das et al., 2017; Verma et al., 2018). In the study area, these species are abundant at the stations falling within a depth range of $\sim >100$ to $\sim 1,500$ m where DO concentration is minimum (0.5 ml/l). All the major species of this assemblage are significantly negatively correlated with the dissolved oxygen, suggesting that they are well adapted to low dissolved oxygen (Suokhrie et al., 2020). The type species of E. cassandrae is E. exilis (Revets, 1993). The ecology of E. cassandrae has been rarely described. However, like its type species, E. cassandrae is also abundant at the intermediate depth locations with the low oxygenated environment. Uvigerina peregrina is also reported

from this assemblage, suggesting its preference to low dissolved oxygen at these depths, which is in line with its reported dominance within OMZ (Hermelin and Shimmield, 1990). It has also been described as a shallow dysoxic-infaunal species (Das et al., 2017). Rotaliatinopsis semiinvoluta has been well documented as a low oxygen tolerant species reported from the Arabian Sea OMZ at intermediate water depths (den Dulk et al., 1998; Jannink et al., 1998; Caulle et al., 2015), and a similar preference to low dissolved oxygen is also found in our study. Cassidulina laevigata is another intermediate to deep-water fauna and is a dominant species in this assemblage. Amongst the other minor species of this assemblage, R. scottii is predominantly infaunal (Kaminski et al., 1988) and a characteristic fauna of muddy substrate, sometimes present in low oxygenated fjords (Murray, 1970; Filipsson and Nordberg, 2004). The substrate seems to be a less influential factor for this species in the west-central Bay of Bengal. Here, R. scottii is more indicative of low oxygenated conditions as its abundance increases at the stations with low DO values. Bolivina spathulata has been described as a deep infaunal and dysoxic species occurring in high organic flux environments (Bernhard and Gupta, 1999; Das et al., 2017). Furthermore, B. spathulata has been associated with a more saline environment (Eris et al., 2011), as is also seen in the study area. Brizalina spathulata is often the last



black inverted triangles. The thin black lines mark the bathymetric contours.

species to disappear before oxygen deficiency starts to inhibit any benthic life (Jonkers, 1984). In the west-central Bay of Bengal, *B. spathulata* is most common at stations between ~120 m and ~250 m. A significant drop in DO is observed at these depths. *Uvigerina peregrina* and *E. cassandrae* are minor species in this assemblage. *Trochamminopsis quadriloba* is more abundant at ~2,000 m. Alve et al. (2011) reported the abundant presence of *T. quadriloba* at ~100 m. However, the depth does not seem to significantly control its abundance (Buzas et al., 1993).

The ecology of *S. semiclara* is rarely described. Here, its abundance in the intermediate waters suggests its preference for low oxygen and relatively more saline environment. *Pullenia*

bulloides was earlier reported as a part of the assemblage representing the Antarctic Bottom Water (AABW) (Corliss, 1979). In the present study, *P. bulloides* prefers lower levels of oxygen, high food supply and deeper depth, as also reported by other workers [north Atlantic-Schnitker, 1979; southwestern Indian Ocean- Corliss, 1983; Gooday, 1994; south Atlantic-Mackensen et al., 1995; Off Pakistan (Arabian Sea)-Schumacher et al., 2007; southeastern Indian Ocean-Verma et al., 2013; Arabian Sea-Caulle et al., 2015]. Although the type and preservation status of the organic matter also matters (Nomaki et al., 2005; Duffield et al., 2014), the organic carbon (C_{org}) is a good indicator of the food availability for foraminifera.

Some foraminifera prefer the "easily metabolizable food particles" while the infaunal forms mainly tolerate more refractory organic matter (Gooday, 1993; Jorissen et al., 1995). Rutherfordoides rotundiformis was found in extremely low oxygen conditions and in sapropelic sequences (Rohling et al., 1997; Mercone et al., 2001; Abu-Zied et al., 2008). Rutherfordoides rotundiformis is the most abundant at the stations with minimum DO concentration in the present study. Chilostomella oolina is a deep infaunal species abundant in low oxygenated environments similar to its generic preferences (Corliss, 1991; Gooday and Rathburn, 1999; Jorissen, 1999; Schmiedl et al., 2003; Schumacher et al., 2007; Kuhnt et al., 2013). The survival and abundance of C. oolina in low oxygenated environment is attributed to its nitrate respiration strategy (Kuhnt et al., 2013). Cassidulina carinata and G. globosa are also part of this intermediate assemblage. Cassidulina carinata is part of the assemblage preferring high food supply (Gupta and Thomas, 1999), while G. globosa is often described as an abyssal species (Takata et al., 2013). Bolivina earlandi have been described as a deep infaunal and dysoxic species occurring in high organic flux environments (Bernhard and Gupta, 1999; Das et al., 2017). Genus Chilostomella with the characteristic unornamented and thin-walled tests was an abundant genus among organic carbonrich and less oxygenated sediments (Kaiho and Nishimura, 1992). Trochamminopsis quadriloba is another minor species in this assemblage as well as the previous assemblage. Other

minor species also include F. pauciloculata abundant in the low oxygenated regions of the Indian waters (Mazumder and Nigam, 2014) and N. auris is an indicator of high organic carbon rich environments (Wefer et al., 1994; Bhaumik and Gupta, 2007). In the west-central Bay of Bengal, B. marginata is a deep infaunal species (Jorissen, 1999). Both B. marginata and B. cf. delreyensis, are abundant at depths deeper than \sim 2,000 m, on the slope. Although B. marginata is often used as oxygen-deficient water proxy, we report a strong negative influence of temperature on the abundance of both B. marginata and B. cf. delreyensis, in the study area. Baggina irregularis is also part of the same assemblage that prefers less oxygenated waters. This species' ecology is not well known, although they are reported from the Bay of Bengal (Panchang and Nigam, 2014). In short, all the species described above are abundant in more saline, organic matter rich and poorly oxygenated environment.

Assemblage III (~1,940-2,494 m)

A set of 12 species from Cluster III (B. cf. delreyensis, B. marginata, H. guttifera, C. laevigata, G. subzelandica, C. carinata, G. mira, N. ampullacea, P. bulloides, U. peregrina, A. globigeriniformis, and T. quadriloba) has been clubbed together in Assemblage III. The dominant species of this assemblage are B. cf. delreyensis, B. marginata, H. guttifera, C. laevigata, and G. subzelandica (Figure 12). Bulimina marginata is a deep



infaunal species (Jorissen, 1999). Both B. marginata and B. cf. delreyensis, are abundant at depths deeper than ~2,000 m, on the slope. Although, B. marginata is often used as oxygen deficient water proxy, the abundance of these two species at deeper colder waters (>1,500 m) is suggestive of a strong negative influence of temperature on the abundance of both B. marginata and B. cf. delreyensis. The dominant agglutinating species in this assemblage is H. guttifera, which represents deep-water assemblages (Bernhard et al., 2009; Enge et al., 2012). C. laevigata is a shallow to deep infaunal species (Singh et al., 2012). Gyroidinoides subzelandica is a part of the same assemblage. All these species are significantly negatively correlated with the ambient temperature as the deeper waters have colder temperature. Additionally, the species in Assemblage III are also influenced by the type of organic matter (marine origin) and sediment texture, evidenced from the relatively lower Corg content in the samples at these depths.

The other minor species in this assemblage like *C. carinata*, *U. peregrina* and *P. bulloides* also represent Assemblage III indicating that these species have a wide range of abundance from the intermediate to the deep waters in the study area. *U. peregrina* is a dominant species in Assemblage III which suggests its preference for the environmental conditions prevalent at the intermediate waters rather than the deep waters. *Gavelinopsis mira*, *N. ampullacea* in Assemblage III, also have a significant preference for colder water with lower CF abundance and marine organic matter.

CONCLUSION

The spatial distribution of living benthic foraminifera and its comparison with the ambient parameters in the riverine influx dominated west-central Bay of Bengal, suggests a strong influence of multiple ecological parameters, mainly the dissolved oxygen, CF, and temperature to some extent. The organic carbon content is high (\sim 1.4% on average) at intermediate depths (\sim 100–1,000 m) where bottom water dissolved oxygen is at its lowest (~0.5 ml/l on average). The maximum living benthic foraminiferal abundance at these depths with limited diversity indicates high abundance of a few opportunistic species. The high Corg/TN suggests that Corg% is mostly of terrestrial origin, especially closer to the river mouths. Three significant faunal assemblages delineated from the statistical analysis give more insight on the ecological preferences of foraminifera in the west-central Bay of Bengal. The shallow water (~27-100 m) assemblage comprising of N. labradorica, H. nipponica, B. dilatata, A. tepida, and N. limbato-striata represents well oxygenated warmer waters with low Corg% and coarser sediment. The intermediate water (\sim 145–1,500 m) assemblage comprising of E. exilis, B. earlandi, F. spinosa, B. lucidopunctata, G. globosa, F. spinosa, E. cassandrae, U. peregrina, R. semiinvoluta, and C. laevigata assemblage represents poorly oxygenated, colder waters with high Corg%. The deep water (~1,940-2,494 m water depth) assemblage comprising of B. cf. delreyensis, B. marginata, H. guttifera, C. laevigata, and G. subzelandica represents high Corg% but relatively higher bottom water dissolved oxygen

than the intermediate water depth. We also report the specific preference of 40 dominant species each influenced by a set of environmental parameters. The ecological preferences of foraminiferal species and foraminiferal assemblages delineated from this study can be utilized as proxies in sub-surface samples to understand paleo-ecology and paleoclimatic changes from the west-central Bay of Bengal.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/**Supplementary Material**, further inquiries can be directed to the corresponding author/s.

AUTHOR CONTRIBUTIONS

TS curated the data and wrote the first draft of the manuscript. RS conceptualized and investigated the research work. RN helped in the funding acquisition needed to carry out the work. All authors contributed to the article and approved the submitted version.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fmars.2021. 656757/full#supplementary-material

Supplementary Table 1 | The details of the sampling locations for multicore as well as spadecore.

Supplementary Table 2 | The details of stations and ambient physico-chemical parameters at each station. Also given are the measured sedimentary characteristics (CF, C_{org}, C_{org}/TN, and CaCO₃). The salinity and pH at the sediment water interface were measured from the water retrieved in multi-core tops. The DO and other data for the spade core station were retrieved from the World Ocean Atlas 2013 (Garcia et al., 2013; Locarnini et al., 2013; Zweng et al., 2013).

Supplementary Table 3 | The relative abundance of the dominant species having atleast >3% abundance at a minimum of two stations.

Supplementary Table 4 | Check-list of living benthic foraminiferal species found in the surface sediments of the western Bay of Bengal. The maximum relative abundance of the species and the depth of the respective station is also provided.

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Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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