



Size Matters: Analyses of Benthic Foraminiferal Assemblages Across Differing Size Fractions

Elena Lo Giudice Cappelli^{1*} and William E. N. Austin^{1,2}

¹ School of Geography and Sustainable Development, University of St Andrews, St Andrews, United Kingdom, ² Scottish Association for Marine Science, Scottish Marine Institute, Oban, United Kingdom

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*Correspondence:

Elena Lo Giudice Cappelli
elena.logiudice.cappelli@gmail.com

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Benthic foraminiferal assemblages are the object of numerous studies spanning from (palaeo)environmental reconstructions to biomonitoring; however, the establishment of a procedure to standardize these studies remains a recent achievement. Not all studies based on benthic foraminiferal assemblages adopt the same methodology, which potentially hinders the use and comparison of samples prepared prior to the creation of a standard protocol or, indeed, without the knowledge of it. One of the main issues is to understand and possibly quantify the influence of different size fractions on foraminiferal biodiversity and richness. In this study, we analyzed benthic foraminiferal assemblages from the west coast of Shetland (Scotland), which were deliberately prepared without following the standard procedure, and were instead picked from the size fractions 63–150 μm and $>150 \mu\text{m}$. Based on assemblage composition, biodiversity indices and multivariate analyses of the data, we assessed the quality and precision of the environmental information that could be extrapolated from these samples. We found that general biodiversity trends remain the same regardless of size fraction, whereas the assemblage internal composition is significantly different between size fractions, with the small fraction retaining a greater degree of environmental sensitivity. We recommend compiling the two sample sets to produce a more holistic and detailed picture of environmental change and generate high-resolution environmental reconstructions. Nevertheless, we conclude that benthic foraminiferal assemblages picked from the large size fraction ($>150 \mu\text{m}$) still provide useful information on prevailing environmental conditions and remain useful for an overview of environmental change in these coastal settings.

Keywords: benthic foraminifera, shell size, assemblages, environmental reconstructions, standard protocol

INTRODUCTION

In the past few decades, benthic foraminiferal assemblages have been extensively used to reconstruct current and past environmental conditions based on their composition, species diversity and richness, using a range of indices, statistical tests and transfer functions (e.g., Buzas, 1969, 1970; Gibson and Buzas, 1973; Schroeder et al., 1987; Poole et al., 1994; Austin and Kroon, 1996; Lei et al., 2017; Weinkauff and Milker, 2018). The literature is peppered with records discussing the influence of size fraction on benthic foraminiferal analyses and to what extent this may affect

environmental reconstructions (Jonkers, 1984; Schroeder et al., 1987; Jennings and Helgadottir, 1994; Knudsen and Austin, 1995; Bouchet et al., 2012; Weinkauff and Milker, 2018). On one hand, studies report that focusing on the larger size fraction only ($>150\ \mu\text{m}$) might result in a loss of information on species richness, biodiversity and species composition compared to studies based on the smaller size fraction (e.g., $>63\ \mu\text{m}$). On the other hand, it is argued that analyzing the smaller size fraction is much more labor intense and requires specialist taxonomic skills, with higher chances of misidentification of specimens and/or a higher proportion of undetermined specimens than obtained from the larger fraction. Recently, a protocol has been designed to standardize the study of benthic foraminiferal assemblages in biomonitoring (Schönfeld et al., 2012), which resulted in the suggestion of the routine use of the $>125\ \mu\text{m}$ size fraction, with the recommendation of analyzing the smaller size fraction ($63\text{--}125\ \mu\text{m}$) only to cases where the $>125\ \mu\text{m}$ fraction is insufficiently informative. Applying this size “threshold” at $125\ \mu\text{m}$ has a twofold effect; first, it allows, where necessary, for smaller species that are absent in the larger size fractions ($125\text{--}150\ \mu\text{m}$) to be accounted (Weinkauff and Milker, 2018), thus improving the representation of the entire assemblage. Second, in most cases, it optimizes the time spent at the microscope by avoiding the need to pick and identifying very small specimens of benthic foraminifera, often juvenile and propagule stages, which are common below $125\ \mu\text{m}$. Therefore, despite this protocol being primarily intended for biomonitoring studies, it can readily be adopted for its consideration of size threshold when benthic foraminiferal assemblages are applied more widely. However, the question remains regarding how best to treat valuable samples that were processed either prior to the establishment of this protocol or without the knowledge of the protocol to enable their use; for example in (palaeo)environmental reconstructions the $125\ \mu\text{m}$ mesh size is by no means standard (Jian et al., 1999; Fontanier et al., 2002; Goineau et al., 2011; Lei et al., 2017).

In this brief report, we use assemblage composition, biodiversity and multivariate analyses of foraminiferal data to compare benthic foraminiferal assemblages from two size fractions $63\text{--}150\ \mu\text{m}$ and $>150\ \mu\text{m}$ to understand the similarities between the two sample sets and assess what type of environmental information can be extracted from each size fraction. Samples were collected from six fjords (voes) on the west coast of Shetland (Scotland, United Kingdom), providing valuable new information on the biogeography of benthic foraminifera in Scotland, which is currently limited to a handful of studies (e.g., Klitgaard-Kristensen et al., 2002; Murray, 2003a,b; Nørgaard-Pedersen et al., 2006). Additionally, these samples help to shed light on the response of benthic foraminifera to environmental stressors such as organic carbon enrichment in coastal sediments (e.g., Lo Giudice Cappelli et al., 2019).

MATERIALS AND METHODS

Sampling and Preparation

The MV *Moder Dy* cruise in west Shetland surveyed six voes in August 2015: Clift Sound, Sand Sound, Olna Firth, Aith

Voe, Busta Voe, and Vaila Sound. Marine surface sediments were sampled using a Duncan and Associates Van Veen grab with a sampling area of $0.1\ \text{m}^2$; 23 surface samples were obtained by sub-sampling the top layer of each grab ($\sim 1\ \text{cm}$ thick) and samples were then stored in a cold-box in sealed plastic bottles. At two sites, MD15-01 and MD15-05, replicate samples were taken (-01A and -01B, -05A and -05B, respectively) to check measurement reproducibility and seafloor heterogeneity [for more detailed information on sample locations and regional setting please refer to Lo Giudice Cappelli et al. (2019)].

For foraminiferal analysis, aliquots of surface sediments were stained with Rose Bengal to allow the identification of living foraminifera. These samples were then wet sieved over a $63\ \mu\text{m}$ mesh, the residues oven-dried at $<60^\circ\text{C}$, weighed and sieved into $63\text{--}150\ \mu\text{m}$ and $>150\ \mu\text{m}$ size fractions. In an early unpublished study, only the larger size fraction was analyzed, and depending on sample volume, samples were subdivided into a number of splits using a standard splitter and ~ 100 specimens were picked and counted. In this study, we built on this unpublished work and analyzed both size fractions, picked at least 300 specimens from each sample and size fraction to ensure statistical significance when discussing benthic foraminiferal assemblages' diversity and composition. Please note that picking 300 specimens from each size fraction does not equate to picking the same proportion of specimens from each size fraction; the latter would depend on the composition of the sediment sample, namely its percentage of sand, silt, and clays. Our overall aim was to follow the standard protocol described in Schönfeld et al. (2012) in its parts that more generally apply to benthic foraminiferal assemblage studies and thus provide an opportunity for data comparisons. We acknowledge that oven-drying our samples may have had a negative effect on the preservation of more fragile agglutinated species (Scott and Vilks, 1991); however, it has been exclusively adopted in previous studies of Scottish fauna (e.g., Klitgaard-Kristensen et al., 2002; Murray, 2003a,b; Nørgaard-Pedersen et al., 2006).

Benthic Foraminiferal Counts

Foraminiferal counts are “total” (live + dead) because the sampling technique may lead to underrepresentation of “live” foraminifera. Additionally, using total assemblages provides a more integrated picture of population dynamics and thus prevailing (long-term) environmental conditions than focusing only on living fauna which is more sensitive to short-term and seasonal variability (e.g., Conradsen, 1993). When possible, we identified specimens to species level following Austin (1991). However, in a few instances, we combined species under their genus to avoid misrepresentation and/or misidentification of rare species especially from the small size fraction. We also grouped under the name *Elphidium excavatum* both forma *selseyense* and forma *clavata* despite their recognition as genetically and morphologically distinct species (Darling et al., 2016), as we were unable to consistently identify the two species. Relative benthic foraminiferal abundances were calculated for both sample sets.

Biodiversity Indices

The Palaeontological Statistics software package PAST (version 3.16, Hammer et al., 2001) was used to analyze relationships between benthic foraminiferal assemblages through a comparison of diversity indices. To discuss biodiversity, we used the Shannon-Wiener index (H') calculated as

$$H' = - \sum_i \frac{n_i}{n} \ln \frac{n_i}{n}$$

where n is the total number of specimens in the sample, and n_i is the number of specimens of taxon i . Therefore, the Shannon-Wiener index took into account both the number of specimens and the number of taxa, varying from 0 for assemblages with only a single taxon to high values for assemblages with many taxa. We compared biodiversity based on the Shannon-Wiener index with the Simpson (S) index calculated as

$$S = 1 - \sum_i \left(\frac{n_i}{n} \right)^2$$

which is a measure of the “evenness” of the assemblage ranging from 0 (one taxon dominates the assemblage) to 1 (taxa are equally represented). Additionally, we evaluated species richness based on the Margalef index (M) calculated as

$$M = \frac{S - 1}{\ln(n)}$$

where S is the total number of taxa. Assemblages similarity was also assessed using the Morisita-Horn index (M-H) that calculates the compositional similarity of paired assemblages (same site-different size fraction) based on the relative abundance of each species (Morisita, 1959; Horn, 1966). M-H values range from 0 in paired samples that are completely different to 1 in paired samples that are identical.

Multivariate Analyses

PAST was also used to run multivariate analyses of the two sample sets (small and large size fraction) based on the Bray-Curtis similarity index and non-metric multidimensional scaling (MDS), ANOSIM, Mantel and SIMPER tests. Non-metric MDS (Taguchi and Oono, 2005) was used to illustrate (dis)similarities in the internal composition of benthic foraminiferal assemblages between the two sample sets. Additionally, we compared the MDS solutions for both size fractions after applying Procrustes transformations (translation, scaling, and rotation of the ordinations) to remove all arbitrary differences in the spatial distribution of samples. This means that any observed difference between paired samples that remains after these transformations is true and can be quantified to compare the relative positions of the 23 paired samples (Rohlf and Slice, 1990; Rudemo, 2000). The ANOSIM test was used to discuss similarities between paired samples (same site – different size fraction), where large positive R -values (up to 1) meant significant dissimilarities between assemblages (Clarke, 1993). The Mantel test reflects how taxonomically similar paired samples are, and the resulting R -value is the Pearson’s correlation coefficient ranging from -1

to $+1$ (Mantel, 1967; Mantel and Valand, 1970). The SIMPER test is a method to assess which taxa are primarily responsible for the observed difference in assemblage composition between sample sets and provides an estimate of the overall average dissimilarity (Clarke, 1993).

RESULTS AND DISCUSSION

Assemblages’ Composition

Together, the two sample sets contain 57 taxa of benthic foraminifera, with 46 taxa present in the larger size fraction and 48 in the smaller fraction. In the small size fraction, benthic foraminiferal assemblages are on average composed of 86% hyaline species, 9% agglutinated, and 1% porcelaneous species. In the large size fraction, benthic foraminiferal assemblages comprise on average 69% hyaline, 26% agglutinated, and 1% porcelaneous species, indicating that the proportions of agglutinated species increase while hyaline species decrease from the smaller to the larger size fraction.

Based on relative abundance data, taxa must appear in at least one size fraction of one sample with relative abundances greater than 10% to be scored as abundant, between 5 and 10% as common and below 5% as rare. Additionally, nine taxa are absent in the small size fraction but present in the large fraction: *Ammoscalaria runiana*, *Connemarella rudis*, *Elphidium crispum*, *Paromalina*, *Planorbulina distoma*, *Quinqueloculina bicornis*, *Leptohalysis fusiformis*, *Spirillina vivipara*, and *Spiroloculina rotunda*. However, all these taxa are rare and sparse in the large size fraction, suggesting that their absence in the smaller fraction is probably a matter of chance. The only exception is *E. crispum*, which is common at one site (MD15-21) while rare and sparse in all other samples. Conversely, ten taxa were absent in the large size fraction, but present in the small size fraction: *Astrononion*, *Bolivina skagerrakensis*, *Bolivina spathulata*, *Cornuspira*, *Epistominella*, *Fissurina*, *Globobulimina affinis*, *Lamarckina haliotidea*, *Leptohalysis scottii*, and *Trifarina angulosa*. Of these, *B. skagerrakensis* and *Epistominella* are common taxa and *L. scottii* is an abundant species in the small size fraction, highlighting that their absence in the large size fraction cannot be attributed solely to chance. However, *L. scottii* specimens in the small size fraction are rather long and narrow, and they could have easily fallen through the larger mesh size along their long axis so that their relative abundance calculations in the small size fraction might be biased. The remaining taxa are rare and sparse in the small size fraction meaning that their absence in the large fraction is negligible.

Overall, 12 taxa of benthic foraminifera are deemed abundant (Table 1). Two of these taxa are abundant only in the larger size fraction: *Ammonia* and *Bulimina marginata*. Five are abundant only in the smaller size fraction: *Buliminella elegantissima*, *Cassidulina laevigata*, *Rosalina*, *L. scottii*, and *Stainforthia fusiformis*, while five are abundant in both size fractions: *Cibicoides*, *Elphidium excavatum*, *Elphidium gerthi*, *Elphidium margaritaceum*, and *Eggerelloides scaber*. Altogether, these 12 taxa account for more than 83% of the total assemblage in the large size fraction and more than 60% in the small size fraction

TABLE 1 | Relative abundance of the 12 abundant taxa and, in italic, output of the SIMPER test.

Sample ID	<i>Ammonia</i>		<i>B. marginata</i>		<i>B. elegantissima</i>		<i>C. laevigata</i>		<i>Cibicoides</i>		<i>E. scaber</i>		<i>E. gerthi</i>		<i>E. margaritaceum</i>		<i>E. excavatum</i>		<i>L. scottii</i>		<i>Rosalina</i>		<i>S. fusiformis</i>		Total %	
	S	L	S	L	S	L	S	L	S	L	S	L	S	L	S	L	S	L	S	L	S	L	S	L	S	L
MD15-01A	-	0.3	-	0.8	4.6	-	2.3	-	11.8	16.8	10.2	32.1	7.9	13.1	17.4	22.3	6.6	8.9	0.7	-	8.2	-	6.6	-	76.1	94.4
MD15-01B	-	0.3	1.0	0.3	3.2	0.9	1.6	-	10.7	19.1	6.8	32.4	11.0	10.8	21.7	26.9	0.3	4.0	0.3	-	6.1	-	9.1	-	71.8	94.8
MD15-02	-	-	0.3	1.1	0.7	-	2.4	-	33.9	61.9	2.1	19.6	4.5	6.8	3.4	2.8	0.3	2.5	-	-	12.7	0.4	7.5	-	67.8	95.0
MD15-03	-	0.3	0.7	1.6	1.3	-	5.7	0.6	38.4	68.2	0.7	5.8	7.4	13.5	0.7	1.0	0.3	1.9	-	-	9.8	0.6	10.1	-	75.1	93.6
MD15-04	0.6	-	0.3	1.6	0.6	-	6.5	0.6	40.4	58.5	0.3	8.5	6.8	21.5	-	0.3	-	-	-	-	8.9	0.9	5.6	-	70.0	92.1
MD15-05A	-	-	0.7	-	-	-	9.7	1.4	36.0	75.7	-	3.8	0.3	0.3	-	0.3	-	-	-	-	20.7	7.4	1.7	-	69.0	88.8
MD15-05B	-	-	0.3	0.9	-	-	12.4	-	40.6	72.2	-	4.2	0.3	-	-	-	0.9	-	-	-	20.3	6.0	2.7	0.3	77.6	83.5
MD15-06	1.3	8.1	-	1.0	9.1	-	0.3	-	2.6	1.3	10.7	39.6	21.5	9.4	6.5	5.5	8.1	22.1	6.2	-	-	-	2.6	-	69.1	87.0
MD15-07	0.4	9.8	2.0	0.2	13.3	-	0.8	-	1.6	7.6	11.0	36.7	9.0	10.9	9.0	6.7	27.5	20.2	7.8	-	1.6	0.2	-	-	83.9	92.4
MD15-08	0.6	5.1	1.0	2.4	23.1	0.6	0.6	-	4.2	0.3	13.0	44.8	3.6	11.0	1.9	0.9	16.2	27.8	2.9	-	0.3	-	13.0	-	80.5	92.8
MD15-09	0.7	9.6	1.6	0.3	17.8	0.6	1.0	-	1.6	2.1	8.2	29.9	3.6	15.3	11.2	9.0	25.3	25.1	0.3	-	0.3	-	3.3	-	75.0	91.9
MD15-10	-	1.8	1.6	5.7	3.6	0.3	3.3	-	15.1	2.1	9.5	72.9	4.9	4.5	1.6	0.9	2.6	7.4	10.5	-	3.0	-	23.0	0.6	78.9	96.1
MD15-11	-	-	3.3	24.6	1.6	-	-	-	17.7	5.0	3.0	56.4	6.9	2.2	1.6	2.0	-	1.4	-	-	0.7	0.3	43.6	5.6	78.4	97.5
MD15-12	0.3	16.2	2.0	14.8	1.4	-	0.3	-	15.1	11.7	3.1	41.3	2.0	2.2	0.9	0.8	-	4.7	0.6	-	0.6	-	46.2	5.3	72.4	97.2
MD15-13	-	6.2	2.0	9.4	21.7	3.2	0.3	0.6	16.6	11.8	3.4	20.6	5.1	13.8	3.4	4.7	2.8	17.6	3.7	-	0.3	-	12.4	2.1	71.5	90.0
MD15-14	1.0	22.3	2.9	5.1	9.9	-	0.6	0.3	20.4	30.6	1.3	17.2	8.3	8.1	1.3	0.5	-	7.8	0.6	-	0.3	0.3	16.2	1.3	62.7	93.5
MD15-15	0.3	1.6	1.7	3.8	3.8	0.3	-	-	19.8	39.7	0.7	24.3	3.8	10.1	5.1	5.0	6.5	4.4	0.3	-	1.7	-	16.0	0.9	59.7	90.2
MD15-16	0.3	10.5	2.8	13.4	1.9	-	-	-	8.7	6.1	1.9	47.7	2.8	6.7	2.2	0.3	2.8	7.0	0.6	-	0.3	-	42.1	3.8	66.3	95.3
MD15-17	0.3	3.8	2.1	0.5	0.3	-	-	-	54.7	82.2	0.3	4.8	2.1	0.8	1.5	0.3	3.0	0.3	-	-	2.4	-	10.4	-	76.9	92.6
MD15-18	0.5	12.5	-	1.9	21.7	0.3	-	-	18.3	31.7	1.9	18.2	2.4	13.6	11.1	6.8	9.6	8.9	2.2	-	4.3	-	7.2	0.5	79.3	94.3
MD15-19	-	21.1	-	4.3	3.5	-	-	0.5	32.8	25.7	1.3	16.8	5.8	10.8	3.3	3.5	7.3	11.1	-	-	5.6	0.3	7.6	0.8	67.2	94.9
MD15-20	-	-	0.3	0.6	0.3	-	-	-	50.8	88.4	0.3	0.6	1.0	1.4	1.0	0.9	2.3	-	0.5	-	20.6	2.9	1.0	-	77.9	94.8
MD15-21	-	0.6	0.5	2.2	0.5	-	-	0.3	46.2	66.6	0.8	3.0	-	3.9	0.8	1.1	2.3	3.3	-	-	12.8	6.4	2.6	-	66.6	87.3
SIMPER test (dissimilarity contribution)	4.1		2.7		4.5		1.5		20.7		16.4		4.6		4.2		6.1		1.2		4.3		8.8		79.1	

S = small size fraction (63–150 μm); L = large size fraction (>150 μm).

(Table 1), which points to a potentially significant difference in the biodiversity between the two sample sets. Additionally, eleven taxa are identified as common. Nine of these taxa are common only in the small size fraction while rare or absent in the larger one: *B. skagerrakensis*, *Bolivina pseudopunctata*, *Bolivina variabilis*, *Buccella frigida*, *Buccella tenerrima*, *Cassidulina obtusa*, *Elphidium albiumbelicatum*, *Epistominella*, and *Trochammina*. Conversely, *Spiroplectammina wrightii* is common in the large size fraction, while rare in the small fraction; *Haynesina germanica* is common in both size fractions. Altogether, these eleven taxa on average make up 16% of the total assemblage in the small size fraction and only 2.5% in the large size fraction. Therefore, common species have a greater impact on the composition of benthic assemblages in the small size fraction than in the large size fraction.

Biodiversity Analyses

Based on the Shannon-Wiener index, biodiversity in benthic foraminiferal assemblages is higher in the smaller than in the larger size fraction (Figure 1A). However, both size fractions show a similar trend with high values in samples MD15-06 to 10 (Sand Sound) and MD15-13 and -14 (Aith Voe), as well as at sites proximal to land like samples MD15-15 (Busta Voe) and MD15-18 and 19 (Vaila Sound). The average value of the Shannon-Wiener index for the small size fraction is $H' = 2.4$, ranging from 1.7 to 2.8, while the average value for the large size fraction is $H' = 1.6$, varying between 0.6 and 2.4. In the large size fraction, minima correspond to assemblages dominated by one taxon: *Cibicidoides* accounts for 73% of the total assemblage in MD15-05, ~82% in MD1-17 and ~88% MD15-20, whereas *E. scaber* dominates at station MD15-10 at ~73%. We further assessed the biodiversity of benthic foraminiferal assemblages using the Simpson index and found a pattern like that obtained with the Shannon-Wiener index (Figure 1B). Based on the Margalef index, species richness is higher in the small size fraction than in the larger one (Figure 1C). The average value of the Margalef index for the small fraction is 4.4, varying between 3.8 and 5.1, while the average value for the large size fraction is 2.7, ranging between 1.9 and 3.7. Two maxima are evident in the large size fraction, which correspond to the only two samples that do not have a clear taxon dominating their assemblages (Figure 1C). The most abundant taxon in these two samples accounts for ~23% of total variability, whereas in other samples the most abundant taxon usually accounts for more than 50% of total variability.

In general, diversity indices reveal that both diversity (Shannon-Wiener and Simpson indices) and richness (Margalef index) are higher in the smaller size fraction than in the large size fraction and that differences in biodiversities and richness between sample sets are statistically significant (Figure 1D). This confirms observations of the assemblages' composition, in agreement with previous studies (Schroeder et al., 1987; Jennings and Helgadottir, 1994; Schönfeld et al., 2012; Weinkauf and Milker, 2018). Nevertheless, both sample sets show similar diversity trends despite the considerable differences in absolute values, as shown by the positive correlation ($R^2 = 0.5$) between the Shannon-Wiener index values calculated for both size fractions (Figure 1E). Additionally, compositional similarity based on the

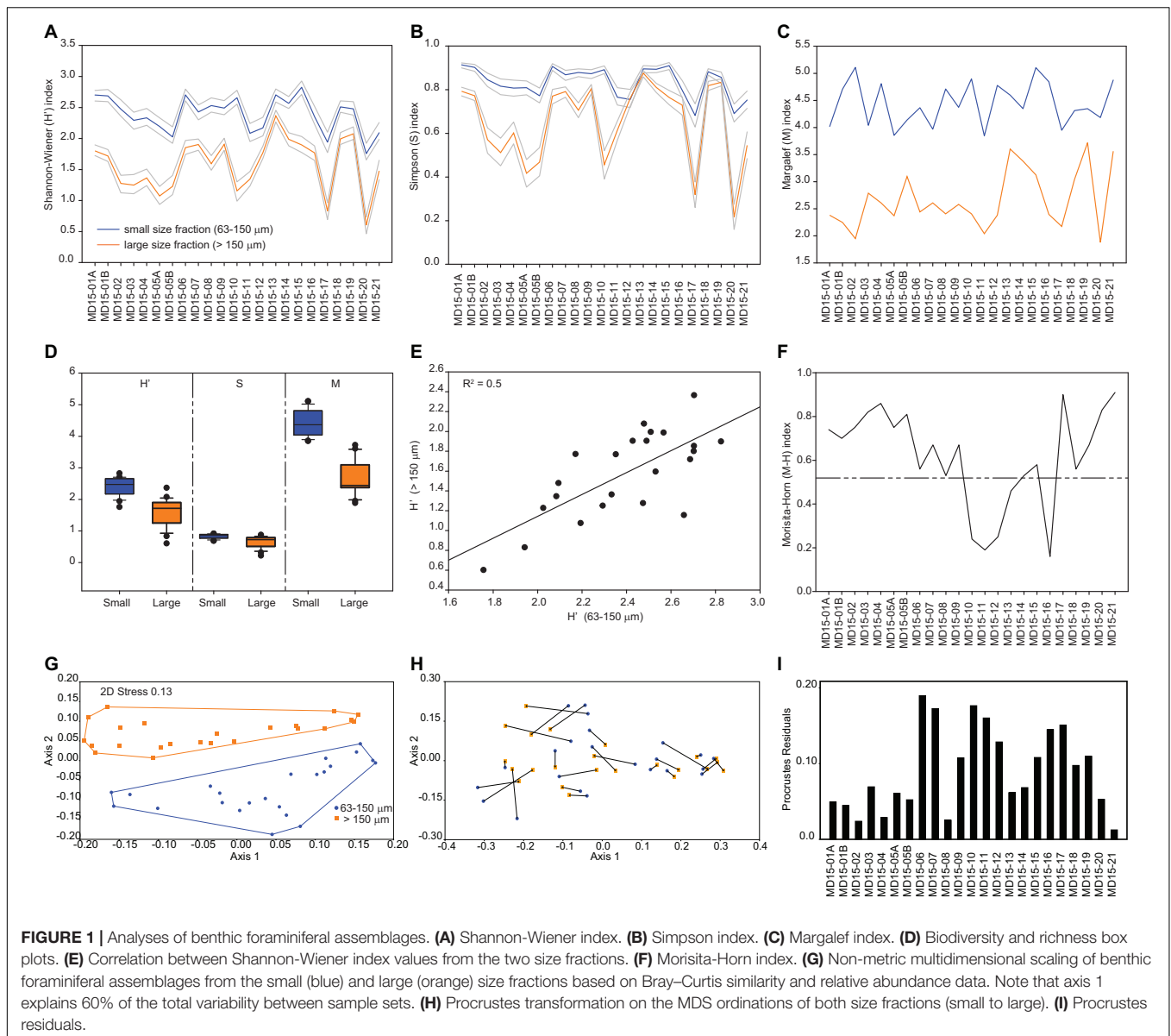
Morisita-Horn index is >0.75 in 8 samples, between 0.5 and 0.75 in 10 samples, and <0.5 in 5 samples, which means that 18 out of 23 paired samples share at least 50% similarity in their assemblage composition (Figure 1F). The main differences in species composition (lowest values of the Morisita-Horn index) are recorded in assemblages dominated by *E. scaber*, with *B. marginata* and *S. fusiformis* as associated species, as *E. scaber* and *B. marginata* only occur in high numbers in the large size fraction (Figure 1F and Table 1).

Multivariate Analyses

Significant differences in the assemblages' internal composition are evident in the MDS ordination plot, which shows no overlapping between the two size fractions (Figure 1G); notably, the first axis explains 60% of the total variability between sample sets. This is better illustrated after Procrustes transformations are applied to the MDS ordinations of each size fraction (Figure 1H). The Procrustes residuals, the distances between paired samples, indicate the true differences between size fractions. Residuals are highly variable, ranging between 0.01 and 0.19 with an average value of 0.09, which results in an error of 5.9% in the ordination of samples depending on the size fraction used. Additionally, residuals are unevenly distributed between paired samples, with a standard deviation of ± 0.05 , hindering the development of a size fraction correction-factor (Figure 1H). The SIMPER test indicates an overall average dissimilarity of 79.1 with ten out of the 12 abundant taxa explaining about 75% of dissimilarity between size fractions (Table 1). Among the abundant taxa, *C. laevigata* and *L. scottii* play a minor role in contributing to the average dissimilarity between the two samples sets ($<1.5\%$) owing their overall patchy distribution within and between size fractions. Conversely, ANOSIM and Mantel tests detect significant similarities between the two sample sets. The ANOSIM test has an $R = 0.4$ and $p = 0.0001$ which means that the two size fractions share a certain level of similarity between paired samples, albeit small, considering that an R close to 1 would indicate dissimilarity between samples. The Mantel test resulted in an $R = 0.7$ and $p = 0.0001$, pointing to a strong positive correlation between paired samples despite the significant differences in the internal composition of benthic foraminiferal assemblages. In summary, multivariate analyses reflect significant differences in the internal composition of benthic foraminiferal assemblage between the two size fractions (non-metric MDS and SIMPER); however, diversity trends stay the same between the two size fractions (ANOSIM and Mantel tests), as also confirmed by the Morisita-Horn index and the positive correlation between Shannon-Wiener indices (Figure 1).

Data Interpretation in an Ecological-Environmental Context

It is crucial to have a clear understanding of the relative abundance of ecologically diagnostic species within and between the assemblages of the two size fractions to reconstruct environmental conditions reliably. Based on benthic foraminiferal relative abundance data, we found that five of the abundant taxa are present in the small size



fraction but generally rare in the large size fraction. Specifically, *B. elegantissima* favors organic rich sediments; *C. laevigata* thrives in marine conditions as well as *Rosalina*, which is also often found in high-energy environments; *L. scottii* is an opportunistic species with an affinity for labile organic matter; and *S. fusiformis* can live in low oxygen concentrations following high organic matter gradients (Jonkers, 1984; Alve, 1994; Klitgaard-Kristensen et al., 2002; Murray, 2003b; Scott et al., 2005). Environmental reconstructions based only on the large size fraction would result in the loss of this information and in lower-precision environmental reconstructions. Additionally, nine of the eleven common species are only present in the small size fraction; once again, negatively influencing environmental reconstructions based only on the large size fraction and resulting in underestimated assemblage diversity. **Figure 1H** illustrates these differences quite clearly, after Procrustes transformations

were applied to the MDS ordinations of both size fractions. None of the paired samples overlap and only a few fall relatively close to each other (residues <0.02), whereas most samples plot in significantly different positions depending on the chosen size fraction. Therefore, we recommend that size fractions should be combined (>63 μm) to achieve high-resolution, precise and accurate environmental reconstructions. Further details concerning the interpretation of the compiled dataset to reconstruct local environmental gradients in west Shetland are available in Lo Giudice Cappelli et al. (2019). Alternatively, it is possible to focus only on the large size fraction, but we suggest that the abundant taxa category should be redefined to include species with relative abundances >5%, in agreement with Lei et al. (2017). In this way, more diagnostic taxa would be included in the discussion of environmental and ecological trends minimizing the loss of information, while in turn avoiding

the labor-intensive challenge of identifying small specimens. In our sample sets, by lowering the abundant taxa “threshold” from 10 to 5% in the large size fraction, *Rosalina* and *S. fusiformis* become included in the list of abundant taxa, thus enabling more statistically robust environmental reconstructions. Conversely, *C. laevigata*, and *L. scottii*, are not included in the abundant taxa list after this “threshold adjustment”; however, based on the SIMPER test results, these two taxa play a minor contribution to the dissimilarity between size fractions (Table 1). In general, care should be taken not to over interpret changes in benthic foraminiferal assemblages, especially when the distribution of certain taxa are patchy or sparse, and instead we recommend a focus on the prevailing (long-term) environmental forcing that could be responsible for the overall observed variability.

CONCLUSION

Our results indicate that the small size fraction (63–150 μm) provides the largest range of diagnostic species and contains the highest species diversity and richness, which provide a more robust statistical base for high-resolution environmental reconstructions, despite the inherent difficulties to define the taxonomic identity of small specimens. We also show that benthic foraminiferal assemblages picked from the large size fraction (>150 μm) represent prevailing, long-term environmental trends as long as they contain enough ecologically diagnostic taxa (relative abundance higher than 5%) consistently distributed across sampling sites. In general, our results highlight the importance of following a standardized methodology for benthic foraminiferal assemblage studies to enable the comparison of different data sets, extending the recommendations of Schönfeld et al. (2012) in this regard beyond biomonitoring studies. Nevertheless, we conclude that it is still possible to extrapolate important environmental information from benthic foraminiferal assemblages of historical samples or samples that did not follow a protocol. For high-resolution environmental reconstructions, data from different size fractions should be combined together to optimize the number of diagnostic

species (>63 μm) and thus provide statistically more robust reconstructions. Alternatively, we suggest focusing on the large size fraction (>150 μm) and making a considered choice in the definition and characterization of abundant taxa, and the identification of diagnostic species in the assemblages as these will significantly affect the quality, precision and accuracy of the resulting environmental reconstructions.

DATA AVAILABILITY STATEMENT

The datasets generated for this study can be found in PANGAEA at <https://doi.pangaea.de/10.1594/PANGAEA.909438>.

AUTHOR CONTRIBUTIONS

WA collected the samples, reviewed, and edited the manuscript. EL processed the sediment samples, designed and performed the analysis with input from WA, and wrote the initial draft. EL and WA identified all specimens.

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Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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