



High-Resolution Habitat Suitability Models for the Conservation and Management of Vulnerable Marine Ecosystems on the Louisville Seamount Chain, South Pacific Ocean

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Vulnerable marine ecosystems (VMEs) are ecosystems at risk from the effects of fishing or other kinds of disturbance, as determined by the vulnerability of their components (e.g., habitats, communities, or species). Habitat suitability modeling is being used increasingly to predict distribution patterns of VME indicator taxa in the deep sea, where data are particularly sparse, and the models are considered useful for marine ecosystem management. The Louisville Seamount Chain is located within the South Pacific Regional Fishery Management Organization (SPRFMO) Convention Area, and some seamounts are the subject of bottom trawling for orange roughy by the New Zealand fishery. The aim of the present study was to produce high-resolution habitat suitability maps for VME indicator taxa and VME habitat on these seamounts, in order to evaluate the feasibility of designing within-seamount spatial closures to protect VMEs. We used a multi-model habitat suitability mapping approach, based on bathymetric and backscatter data collected by multibeam echo sounder survey, and data collected by towed underwater camera for the stony coral and habitat-forming VME indicator species *Solenosmilia variabilis*, as well as two taxa indicative of stony coral habitat (Brisingida, Crinoidea). Model performance varied among the different model types used (Boosted Regression Tree, Random Forest, Generalized Additive Models), but abundance-based models consistently out-performed models based on presence-absence data. Uncertainty for ensemble models (combination of all models) was lower overall compared to the other models. Maps resulting from our models showed that suitable habitat for *S. variabilis* is distributed around the summit-slope break of seamounts, and along ridges that extend down the seamount flanks. Only the flat, soft sediment summits are predicted to be unsuitable habitat for this stony coral species. We translated a definition for stony coral-reef habitat into a *S. variabilis* abundance-based threshold in order to use our models to map this VME habitat. These maps showed that coral-reef occurred in small and isolated patches, and that most of the seabed on these seamounts is predicted to be unsuitable

habitat for this VME. We discuss the implications of these results for spatial management closures on the Louisville Seamount Chain seamounts and the wider SPRFMO area, and future modeling improvements that could aid efforts to use habitat suitability maps for managing the impact of fishing on VMEs.

Keywords: vulnerable marine ecosystems, habitat suitability modeling, *Solenosmilia variabilis*, fishery management, Louisville Seamount Chain, SPRFMO, seamount

INTRODUCTION

Vulnerable marine ecosystems (VMEs) are ecosystems at potential risk from the effects of fishing or other kinds of disturbance, as determined by the vulnerability of their components (e.g., habitats, communities, or species) (FAO, 2009). A number of United Nations General Assembly resolutions have been passed that require nation states and fishery management organizations to identify VMEs within their jurisdiction as one of the steps toward their protection (see Ardron et al., 2014 for details). Species or taxonomic groups have been identified that can be used as indicators of the presence of VMEs in particular ocean regions, in order to assist agencies responsible for their protection [e.g., the South Pacific Regional Fisheries Management Organization (SPRFMO) in the South Pacific region—Parker et al., 2009]. Such taxa possess characteristics that make them particularly vulnerable to disturbance (such as slow growth rates, longevity, late maturity, and fragility), and include species that form structurally complex features, like coral reefs and sponge aggregations, which provide three-dimensional structure associated with diverse communities, and discrete areas of functional significance (e.g., necessary habitat for rare, threatened or endangered species of the habitat, and/or the survival, function, spawning/reproduction or recovery of fish stocks and particular life-history stages) (FAO, 2009).

Habitat suitability modeling (sometimes called species distribution modeling) is a method for predicting the suitability of a location for a species, or group of species, based on their observed relationship with environmental conditions. Habitat suitability modeling is being used increasingly to predict distribution patterns of VME indicator taxa in the deep sea, where data are particularly sparse, and such models are considered useful for marine ecosystem management (Ross and Howell, 2013; Reiss et al., 2014). Habitat suitability models have been produced for numerous deep-sea taxa (see review by Vierod et al., 2014), but the predicted distributions are dependent on how the models are constructed. The quantity, quality, and distribution of species presence records, the availability of true absence records, and the environmental predictor variables used can all influence the reliability of the models (Araújo and Guisan, 2006; Guisan et al., 2006). Recent efforts to improve the accuracy of habitat suitability models, and thus their usefulness for the management of the impact of fishing on VMEs, have included use of abundance data (as opposed to relying only on presence-absence data), ground-truth model validation, ensemble modeling, and estimates of model uncertainty (Rooper et al., 2014, 2016; Anderson et al., 2016a,b; Robert et al., 2016).

Despite these improvements, models are still sometimes found to be unsuitable for management purposes because they are either at spatial resolutions that are too coarse, or because they predict the presence of VME indicator taxa but not the VME itself. These issues can be reduced if high-resolution multibeam data and seafloor images are used in building habitat suitability models (e.g., Howell et al., 2011; Rengstorf et al., 2012, 2013, 2014).

The aim of the present study was to produce high-resolution habitat suitability models for VME indicator taxa, and VME habitat, on seamounts of the Louisville Seamount Chain at the scale of individual seamounts. These seamounts are located within the SPRFMO Convention Area, and some are the subject of bottom trawling for orange roughy by the New Zealand fishery. SPRFMO has protection measures in place for VMEs, but the efficacy of these measures has been questioned (e.g., Penny and Guinotte, 2013), and additional and alternative measures are being sought and considered by stakeholders. Currently, large 20-min latitude/longitude spatial closures are implemented on some seamounts for the New Zealand fishery, based on past fishing history or the presence of a VME detected by the bycatch of VME indicator taxa exceeding a particular threshold (Parker et al., 2009; Penney et al., 2009). One possible alternative management measure is to close small areas on individual seamounts that have, or are likely to have, VMEs, and allow fishing elsewhere on the seamount. High resolution, seamount-scale habitat suitability models for VME indicator taxa would aid the design of such within-seamount spatial closure measures.

In the present study we built such models for a species of reef-forming stony coral, *Solenosmilia variabilis* and two taxa that are considered useful indicators for the occurrence of coral reef habitat; brisingid starfish and crinoids (Parker et al., 2009). The present study builds on previous habitat suitability modeling work in the region (Anderson et al., 2016a,b), and collectively these models can be used to inform spatial management planning for protecting VMEs in the seas around New Zealand, including key bottom-trawling regions within the SPRFMO Convention Area.

METHODS

Study Area and Seamounts

The Louisville Seamount Chain lies to the east of New Zealand in the SPRFMO Convention Area, the region of the South Pacific beyond areas of national jurisdiction. It is made up of more than 80 seamounts and extends over 4,000 km from the junction of the Pacific and Indo-Australian Plates (at latitude $\sim 27^\circ$ S) southeastwards into the central southwest

Pacific (to latitude $\sim 47^\circ$ S) (**Figure 1**). Many of the seamounts in the chain are large guyots; flat-topped seamounts formed by erosion and subsequent submersion of islands (Lonsdale, 1988). Parts of the Louisville Seamount Chain are subject to bottom trawling, primarily for orange roughy (*Hoplostethus atlanticus*) by the New Zealand deepwater fishing fleet (Clark et al., 2016).

Seven large seamounts in the Louisville Seamount Chain were selected for survey (Forde, CenSeam, Anvil, JCM, 39 South, Ghost, Valerie) (**Figure 1**) with the aim to represent as wide a geographic range as possible given the available survey time. All seven seamounts have a vertical elevation of more than 1 km from the surrounding seafloor, and summary details for each are given in **Table 1**. Among the chosen seamounts there is a general gradient of increasing historical bottom trawling from north to south. Bottom trawling has taken place on all these seamounts and much of the area (excluding Forde and CenSeam) remains open to fishing by New Zealand bottom trawlers under SPRFMO regulations. These regulations control fishing in designated 20-min latitude/longitude blocks by vessels flagged to SPRFMO Member States. Regulations are based on a “bottom fishing footprint” established during the 2002-06 SPRFMO reference period (SPRFMO CMM2.03, <https://www.sprfmo.int/>), and for New Zealand vessels these are subject to open, move-on, and closed rules (Penney et al., 2009) (**Figure 1**).

Sampling

A random-stratified survey of benthic fauna and habitats using a towed camera system and an epibenthic sledge was undertaken on each study seamount to test the abilities of two broad-scale models to predict the presence of reef-forming corals (Anderson et al., 2016a). Data collected during this survey were also intended to enable subsequent development of high-resolution habitat suitability models at the scale of individual seamounts in the present study. Clark et al. (2015) and Anderson et al. (2016a) describe the survey design and sampling methodology in detail, but some of this information is repeated here for completeness.

Prior to running photographic transects, a multibeam echosounder (MBES) survey was conducted on each seamount (except Anvil where existing data were available) using a 30 KHz Kongsberg EM302, producing a data resolution of 25×25 m. Each photographic transect was ~ 1.4 km long (mean \pm SD, 1.43 ± 0.27) and 2 m wide, and some targeted epibenthic sled deployments were made along the video transects to collect physical samples for verification of visual identifications and other research. Photographic transects were carried out using NIWA's deep towed imaging system (DTIS, Hill, 2009), which incorporates high definition video (Sony HD 1080i format) and still camera (Canon 10MP single lens reflex) systems, with an ultra-short baseline positioning system (USBL, Kongsberg HiPAP) for tracking and recording the precise seabed position of the equipment (accurate under ideal conditions to within 1 m). In total, 118 photographic transects were completed, with 20, 22, 13, 1, 17, 29, and 16 on Forde, CenSeam, Anvil, JCM, 39 South, Ghost and Valerie seamounts, respectively (**Figure 2**).

Data and Data Processing

Identifications and counts of fauna observed in video transects were made in real-time via a low-resolution video link to the ship, and from later analysis of full-resolution video for selected transects in which VME indicator taxa (primarily stony corals) occurred. *S. variabilis* was the only significant habitat-forming coral observed in the study area. Because much of the *S. variabilis* matrix of colonies observed was evidently not alive (dark colouration with no feeding polyps visible in high resolution still imagery), occurrence of this taxon was recorded in two ways: as percent cover of the seabed by intact coral matrix (“intact coral”), and counts of distinct coral colonies or “heads” with live polyps (“live heads”). The occurrence of non-intact coral matrix (broken up coral colonies that can occur around intact coral) was not recorded. Abundance data were then extracted for the three study taxa: live heads of *S. variabilis*; brisingid starfishes (Order Brisingida), and sea-lilies and feather stars (Class Crinoidea). The first species belongs to the VME indicator taxon Order Scleractinia, while the latter two are VME habitat indicator taxa for coral reefs (*sensu* Parker et al., 2009). Abundances were recorded as the number of individuals (or live heads) per 25 m^2 segment of video transect (~ 2 m transect image width \times 12 m along the transect track).

A working definition of VME coral reef habitat, as opposed to individual occurrence of *S. variabilis*, was derived from video observations in the context of: (a) guidelines on identifying “sensitive environments” in the New Zealand region (schedule 6, Exclusive Economic Zone and Continental Shelf (Environmental Effects—Permitted Activities) Regulations 2013 <http://www.mfe.govt.nz/marine/legislation/regulations-under-eez-act>) and (b) our intuitive evaluation of VME status from observation of the seabed video imagery during the voyage and post-voyage confirmation from high-resolution imagery. According to the guidelines, “stony coral thicket or reef” habitat (**Figure 3A**) exists where either live or dead colonies of a structure-forming coral taxon cover more than 15% of the seabed over areas of $> 100 \text{ m}^2$. Using this as a starting point, for each 25 m^2 video segment, we calculated the proportion of the seabed identified as intact coral. To approximate to $\sim 100 \text{ m}^2$, we then identified areas along each transect where any 3 or more segments in a 5-segment moving window had more than 15% intact coral. We then compared a number of possible formulations for VME definition (excluding extensive areas of dead coral matrix), each incorporating different measures of the numbers of live *S. variabilis* heads, against our intuitive evaluation of VME status. The definition (“rule”) that best matched our intuitive evaluation combined only two criteria: (1) more than 15% intact coral present in 3 or more segments of a 5-segment moving window, and (2) more than one live *S. variabilis* head present in 3 or more segments of the 5-segment moving window. Having arrived at this definition of VME habitat status, we then calculated the mean number of live *S. variabilis* heads per VME habitat segment per transect for each transect in which VME habitat occurred, and then took the lowest mean value obtained (2.78 *S. variabilis* live heads per VME habitat segment, transect #135) as a threshold indicator of VME coral reef habitat (**Figure 3B**). This abundance threshold value was applied to the outputs from the suitability

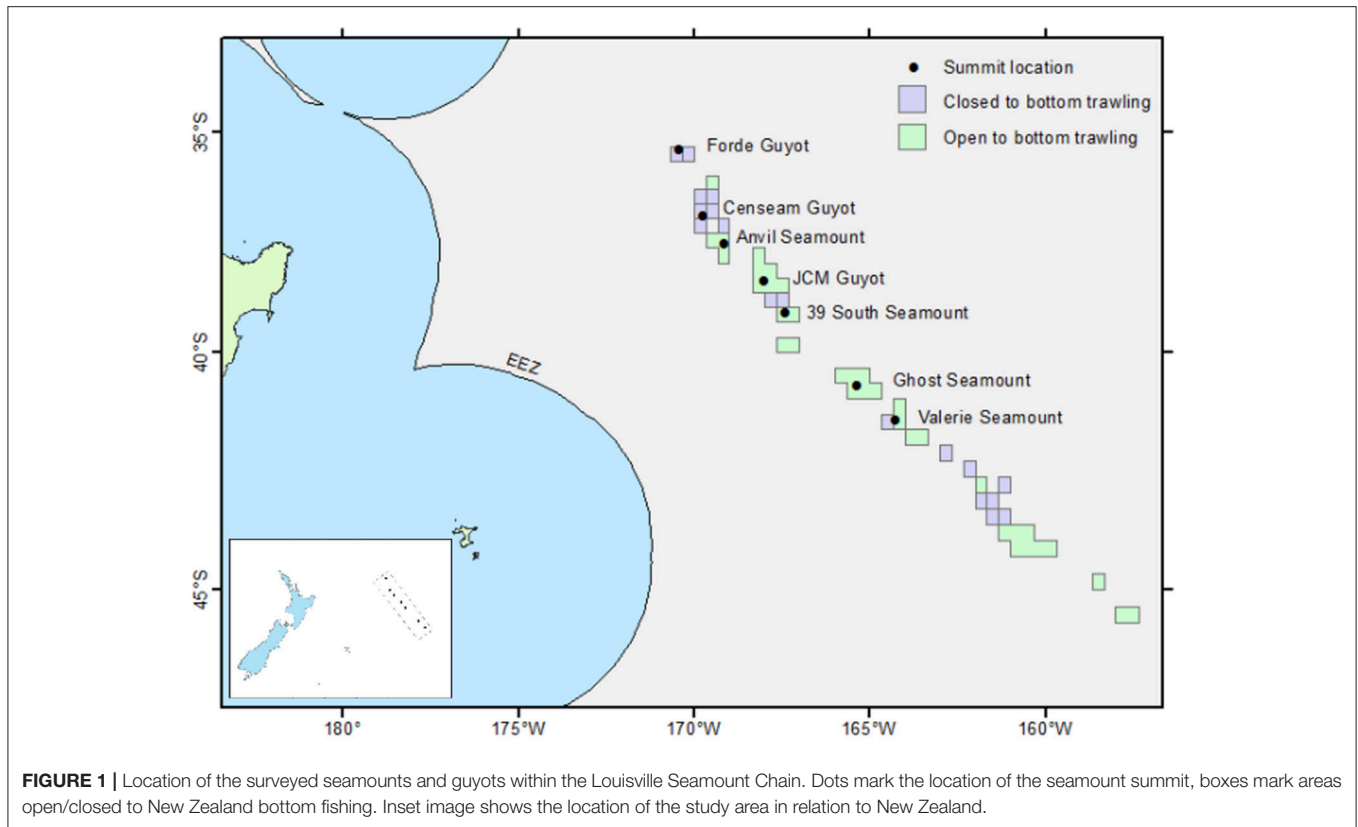


TABLE 1 | Summary details of the surveyed seamounts.

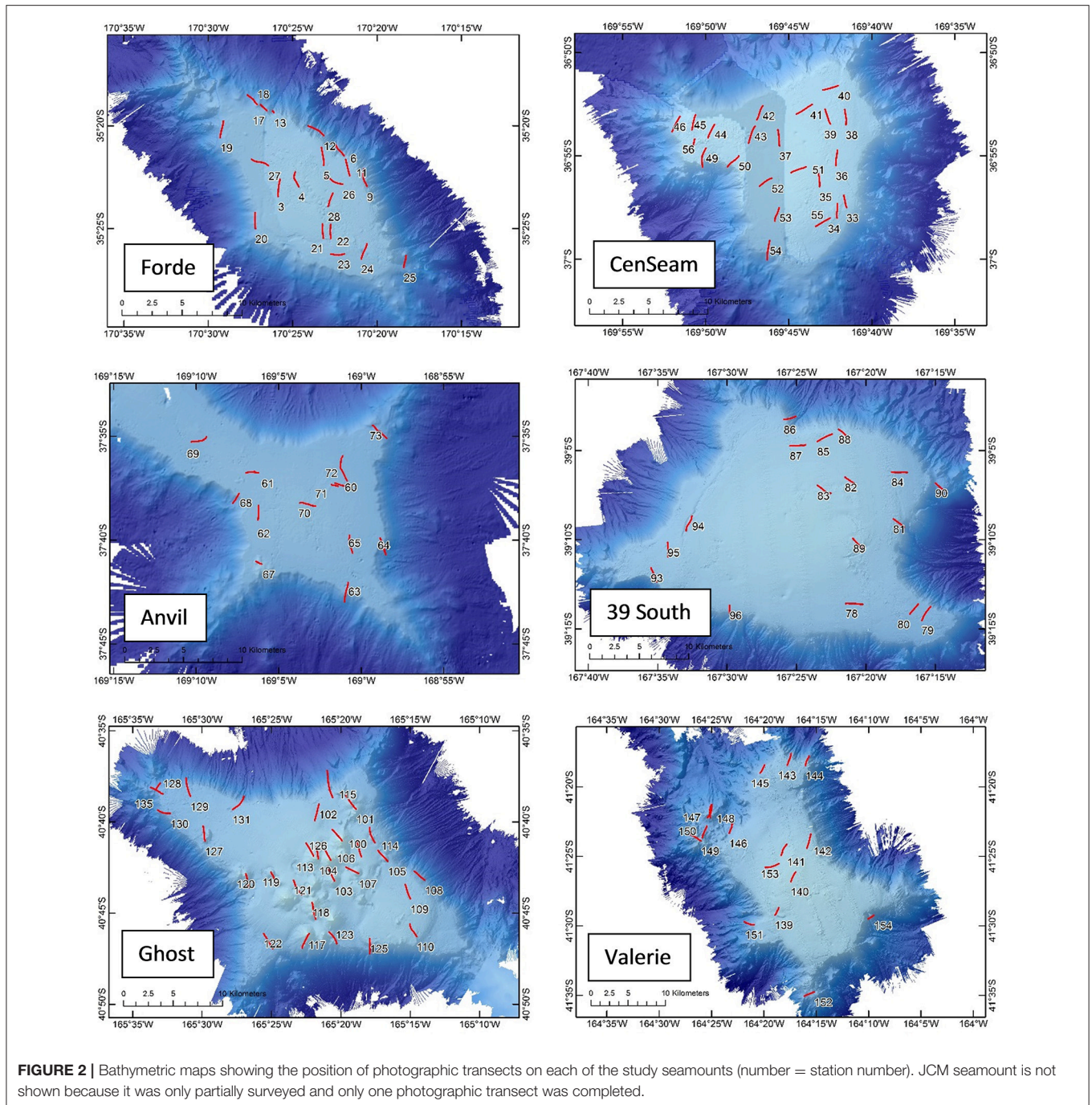
Seamount name	Latitude (S decimal degrees)	Longitude (W decimal degrees)	Depth range (m)	Fishing effort (#tows)	Open/Closed to fishing (year last fished)
Forde Guyot	35.40	170.40	980–4,500	370	Closed (2005)
CenSeam Guyot	36.92	169.73	955–4,823	672	Closed (2007)
Anvil Seamount	37.56	169.15	1,036–4,648	205	Open (2007)
JCM Guyot	38.41	167.99	265–4,772	1,819	Open (2013)
39 South Seamount	39.10	167.40	878–3,750	1,134	Open (2014)
Ghost Seamount	40.70	165.35	620–4,500	3,865	Open (2014)
Valerie Guyot	41.45	164.25	750–4,500	1,826	Open (2013)

Fishing effort is for bottom trawling by New Zealand vessels for orange roughy (*Hoplostethus atlanticus*) targeted or caught, and covers the years 1992–2014. Open/closed refers to New Zealand bottom fishing measures.

modeling (see below) to map the predicted distribution of VME habitat.

A set of more than 100 topographical terrain variables was derived from bathymetric data collected by the MBES survey using Benthic Terrain Modeler in ArcGIS 10.3.1 (ESRI, 2015). Terrain variables included those that have been shown previously to be useful for predicting suitable habitat for VME indicator taxa (e.g., Rengstorf et al., 2012). The continuous variables were: depth; slope (steepest gradient to any neighboring cell); aspect (direction of slope); curvature (change of slope); plan curvature (curvature of the surface perpendicular to the slope direction); and profile curvature (curvature of the surface in the direction of slope). In addition, a further set of derivative

continuous variables were calculated using focal mean analysis in ArcGIS for the standard deviation of depth, depth range, standard deviation of the slope, and terrain ruggedness. Focal mean analysis finds the average value of a variable within a specified neighborhood centered on a given cell and outputs these data to the corresponding cell location. Four different neighborhood sizes (3×3 , 5×5 , 7×7 , and 15×15 grid cells) were computed to produce more generalized datasets. The different neighborhood cell sizes were selected to mimic a range of spatial scales at which seafloor bathymetry and topography may affect the distribution of benthic communities (e.g., 3×3 cells = 75×75 m; 5×5 = 125×125 m; 7×7 = 175×175 m; 15×15 = 375×375 m). Aspect, measured continuously in degrees,

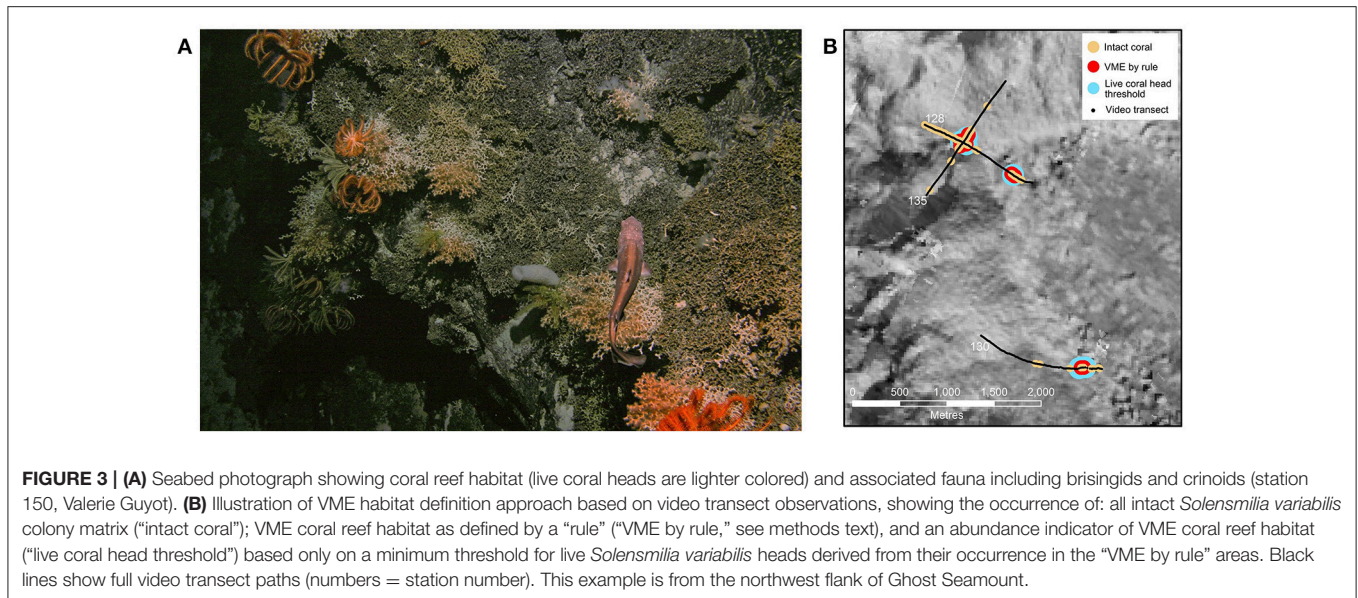


was transformed into a categorical variable representing the 16 sectors of the compass (i.e., N, NNE, NE, etc).

Backscatter data from the MBES survey were processed using SonarScope (Augustin and Lurton, 2005). Processing consisted of statistical compensation of the signal as a function of its incidence angle on the seafloor, to attenuate the strong signal from specular reflection at the nadir and the rapid decrease of the signal strength with increasing incidence angle (Fonseca et al., 2009). Backscatter strength (a continuous variable), which measures the acoustic reflectivity of the seafloor, has been found

to be a useful surrogate for substratum type (Lamarche et al., 2011). Substratum type influences the distribution of benthic organisms, including the study taxa which typically occupy hard substratum (high backscatter seafloor), and backscatter strength has been related to the distribution of deepwater coral species in previous studies (e.g., Georgian et al., 2014).

Because backscatter data were not available from the earlier MBES survey of Anvil Seamount, this seamount was excluded from modeling. JCM Seamount was also excluded because only a part of the seamount was surveyed using MBES, and only one



photographic transect was completed on the feature. That is, only models for five out of the original seven seamounts surveyed were made. Some biological data from the two excluded seamounts is presented later.

Model Types and Selected Model Settings

We selected three statistical modeling methods in common usage for habitat suitability modeling; Boosted Regression Trees, General Additive Models, and Random Forests. Model predictions of habitat suitability across all seamounts were restricted to a maximum depth of 2,000 m (the depth limit of the video survey) and a minimum depth of 265 m (the summit of the shallowest seamount).

Boosted Regression Tree (BRT)

Boosted Regression Tree (BRT) modeling is an approach which can utilize both presence-absence data and abundance data, and has been used previously to predict distributions of deep-sea taxa in the study region (Anderson et al., 2016a,b). BRT models incorporate recursive binary splits within a regression tree structure to explain the relationship between the response and predictor variables (Elith et al., 2008). BRT models were fitted in R (R Development Core Team, 2016) using a standard approach, including optimization of the learning rate and number of trees by internal cross validation (Elith et al., 2008), and setting tree-complexity to 3 to allow a level of interactions between terms. The minimum number of trees for each model was set at 700. For construction of presence-absence models a Bernoulli (=binomial) distribution was assumed and for abundance models (count data) a Poisson distribution was assumed. Other model settings were left at default values.

A Generalized Additive Model (GAM)

A Generalized Additive Model (GAM) is a specific form of a generalized linear model that estimates smooth, non-parametric functions for each predictor variable (Hastie and Tibshirani,

1986). The distributions of a number of deep-sea species have been successfully modeled using a GAM approach, including corals (Rooper et al., 2014; Murillo et al., 2016), sponges (Rooper et al., 2014; Murillo et al., 2016), and fish (Leathwick et al., 2006). GAMs were developed in R using the "mgcv" package (Wood, 2006). For presence-absence models, a binomial distribution was assumed during model construction. For abundance models, a zero-inflated Poisson distribution with no data transformation was selected after exploring several alternative distributions (Gaussian, Poisson, quasi-Poisson, and Tweedie) and transformations (log+1, log+1% of mean abundance, square root, and fourth root) through analysis of residuals and model deviance. For all models, predictors were fitted with smooth terms allowing up to 4 degrees of freedom.

Random Forest (RF)

Random Forest (RF) modeling (Breiman, 2001) is a non-parametric approach which builds classification trees (presence-absence data) or regression trees (abundance data) using random subsets of the input data. One potential benefit of random forest over modeling approaches such as GAMs is the lack of any underlying assumption of the distribution of the response variable. RF models were constructed using the "randomForest" package (Liaw and Wiener, 2002) in R. For all models, 501 trees were run, which was always sufficient to allow the error rate to stabilize. Different values of "mtry," the number of variables used in each tree node, were explored during initial model construction using the "tuneRF" function in the "randomForest" package. Default values (square root of the number of environmental variables for classification models, 1/3 the number of environmental variables for regression models), consistently produced the best results and were used in all models. Classification trees are highly sensitive to imbalances in the response variable, as the model will minimize the overall error rate at the expense of the error rate of the minority class

(Chen et al., 2004; Evans et al., 2011). As our taxa data were skewed (>10:1 ratio of absences to presences for all species), we set the “cutoff” parameter in “randomForest” equal to species prevalence for each presence-absence model (Guo et al., 2004), which greatly reduced omission errors without affecting overall model performance.

Model Performance and Precision

Model performance was assessed using a cross-validation procedure in which models were trained using a random partition of data (70%) and tested against the remaining portion (30%). This cross-validation was carried out for 10 test datasets derived from different combinations of extracted data. For presence-absence models the area under the curve (AUC) metric was assessed for each of the ten test datasets, and an average calculated. In general, AUC values of ≥ 0.5 indicate better than random performance, ≥ 0.7 indicate adequate performance, and ≥ 0.8 indicate excellent performance (Hosmer et al., 2013). For abundance models, correlations (R^2) between predicted and observed values for the 10 test datasets were calculated, and averaged. In addition, for both presence-absence and abundance models (and ensemble models—see below), the correlation (R^2) between predicted and observed values was calculated for the whole dataset. For presence-absence models this was achieved by first converting model probabilities into a binomial form comparable with the observed data using cut-offs which provided an equivalent number of presences and absences. In all cases the final models were constructed using all data.

In order to assess the relative confidence in predictions across the model extent, we used a bootstrap technique for each model type to produce spatially explicit uncertainty measures, after Anderson et al. (2016b). Random samples of the input data, in which presence and absence records were in equal proportion and number to the original (for both presence-absence and abundance models) were drawn with replacement, and models constructed with the same settings as the original. Predictions were then made for each cell of the model extent. This process was repeated 500 times for BRT models and 200 times for GAM and RF models (limited by processing capacity) resulting in 500 or 200 estimates for each cell. We then calculated model uncertainties as the coefficient of variation (CV) of the bootstrap output.

Ensemble Models

Fundamental differences in model structure among the three models will inevitably result in different patterns of predicted habitat suitability. Evaluations of different types of models are often unable to demonstrate the unequivocal superiority of any single type, and studies have shown that predictions by alternative models can be so variable that they compromise their use for guiding policy or decision making; a solution to this issue is to use “ensemble” models (Araújo and New, 2007). Hence, for incorporation into decision-making tools for spatial management planning (e.g., Zonation, Moilanen, 2007) an ensemble model is a practical way to avoid dependence on a single model type and better describe in general the predicted spatial variation and uncertainties (Robert et al., 2016). To incorporate the predictions and underlying assumptions of each model into a single output grid we produced ensemble models

of presence-absence and abundance, by calculating weighted averages of the relevant BRT, GAM, and RF models (after Oppel et al., 2012 and Anderson et al., 2016b).

$$W_B = \frac{PS_B}{PS_B + PS_G + PS_R}, W_G = \frac{PS_G}{PS_B + PS_G + PS_R}, \text{ and}$$

$$W_R = \frac{PS_R}{PS_B + PS_G + PS_R}$$

$$X_E = X_B * W_B + X_G * W_G + X_R * W_R$$

$$CV_E = \sqrt{\frac{(CV_B * X_B)^2 * W_B^2 + (CV_G * X_G)^2 * W_G^2 + (CV_R * X_R)^2 * W_R^2}{X_E^2}}$$

where PS_B , PS_G , and PS_R are the model performance statistics (AUC for presence-absence models and R^2 values for abundance models); X_B , X_G , X_R , and X_E the model predictions; and CV_B , CV_M , CV_G , and CV_E the bootstrap CVs from the BRT, GAM, and RF, and ensemble models respectively.

Precision estimates for the ensemble models were calculated as the correlation (R^2) between predicted and observed values calculated for the whole dataset, as described above.

Predictor Variable Selection

The topographical terrain and backscatter variables derived from the MBES survey formed the basis of the model predictor variable set. Chemical and other water property variables, including biological productivity variables available from various global or regional climatologies, were not considered as none were available with a native resolution similar to that of the MBES-derived variables. Several of these variables (e.g., temperature, salinity, surface-derived production, and aragonite saturation) are correlated with depth to varying degrees, and so a depth variable in this case can act as a surrogate for the changes in these parameters (Leathwick et al., 2006; Thresher et al., 2014). The categorical variable seamount, identifying the individual seamounts, was however added to MBES-derived candidate variables. Seamount was used rather than latitude and longitude to predict potential geographic-related environmental differences in habitat suitability, because this variable could also be used to reflect variables that may be specific to a particular seamount that are not related simply to geography.

The full set of candidate variables was initially reduced by eliminating highly correlated variables, based on Pearson product-moment correlation coefficients. Values for the remaining variables were then determined for the midpoint locations of each of the 25 m² taxa presence/absence video segments and used to build a set of single-variable presence-absence GAM models for each of the three taxa, as well as models using all of the variables together. The predictive value of each variable was then assessed according to its chi-square score in the all-variable models, and its AUC score in the single-variable models. In order to further assess the relative utility of the remaining candidate variables, we considered relationships among them using a cluster dendrogram, again using Pearson product-moment correlation coefficients. In this way priority was given to variables that had low correlations with other variables and occurred

in unique clusters, so as to avoid losing information that was not also provided by other variables, and to avoid including only high-performing variables that provided similar information.

Moran's index was used, with the "spdep" package for R (Bivand, 2011), to measure spatial autocorrelation in model residuals compared with the raw presence-absence or abundance data. This index measures the correlation between observations as a function of the distance separating them, with values between -1 (highly dispersed) and 1 (highly clustered).

In order to account for the inherent spatial autocorrelation in the model data, where relationships between observations are correlated with the geographic distances between them, we created an additional predictor, the residual autocovariate (RAC), representing the similarity between the residual from initial models at a location compared with those of neighboring locations. This method can account for spatial autocorrelation without compromising model performance, and can easily be implemented in most modeling approaches (Crase et al., 2012).

Model Outputs

All model results [predicted probability of occurrence/suitable habitat $[0-1]$ and predicted abundance [number 25m^{-2}] for each model and taxon, and the corresponding coefficients of variation (CV)] were mapped onto the newly derived bathymetry for the seamounts using ArcMap 10.3.1 (ESRI, 2015) at the same spatial resolution as the model environmental input data ($25 \times 25\text{m}$). Maps were drawn with a constant probability/abundance scale between models and taxa so that visual comparisons can be easily made.

As a measure of the utility of the two VME habitat indicator taxa Brisingida and Crinoidea for identifying the location of stony coral VME habitat, correlations between the predicted abundance of *S. variabilis* and the predicted abundance of these taxa were calculated from the ensemble-abundance model outputs.

RESULTS

Photographic Transects

The stony coral *S. variabilis* and brisingids were present in less than 500 of the 12,504 25m^2 video segments observed. Crinoids were two–three times more common across all seamounts, but this taxon was represented by a greater number of species than the other VME habitat indicator taxon (Table 2, Figure S1). During the survey, at least 11 species of crinoids were identified from samples collected using the epibenthic sled (the comatulids *Aglaometra incerta*, *Charitometra basicurva*, *Florometra novaezealandiae*, *Glyptometra inaequalis*, *Antedon* sp., *Crotalometra* sp., *Cyclometra* sp., *Porphycrinus* sp., *Strotometra* sp., and the hyocrinid *Thalassocrinus*, sp. plus several unidentified comatulids), compared with three species of brisingids (*Novodinia novaezealandiae*, *Freyella* sp., and *Hymenodiscus* sp.).

Variable Selection

Related variables based on a single focal mean neighborhood size (e.g., the range and standard deviation of curvature, curvature aspect, and plan curvature) were often highly correlated ($>75\%$). Correlations between these variables and all remaining variables were examined graphically to eliminate all but the least correlated of them. Similarly, ruggedness was highly correlated with standard deviation of slope, and the latter variable was eliminated due to its higher correlation with other retained variables. This logic was followed for all variable types and focal mean neighborhood sizes to eliminate the most correlated variables, reducing the set of more than 100 MBES-derived variables to 63.

Chi-square values for the GAM models based on all 63 remaining variables, and AUC scores for 63 single-variable models (for each taxon) revealed a consistent set of eight variables which showed a high level of explanatory power while covering a range of the variable types derivable from MBES data. Cluster dendrograms and correlation coefficients showed clustering of these variables and no two variables were more than 70% correlated (Figures S2, S3).

Separate RAC variables were constructed for each model type (BRT, GAM, and RF), prediction type (presence-absence and abundance), and taxon (*S. variabilis*, Brisingida, and Crinoidea), based on the residuals from initial models constructed from the eight MBES-derived variables. The final set of nine variables used in each of the models is shown in Table 3.

Model Performance

Cross-validated AUC values were very similar for each of the presence-absence models, ranging between 0.90 and 0.93 in all cases except for the Brisingida GAM model which was lower (0.87) but still considered excellent (Hosmer et al., 2013) (Table 4). The generally similar AUC values among the three model types for each taxon meant that there was a similar weighting, and therefore a similar influence, for each model in the ensemble models. Overall, and consistently across the three taxa, RF models performed slightly better than BRT models, and GAM models performed worst according to cross validated AUC. Correlations (R^2) between predicted and observed presence-absence for all model data showed broad agreement with the AUC performance measures, although they were consistently slightly higher for BRT models (26.6–37.8%) than for RF models (23.7–33.7%), and considerably lower for GAM models (7.5–16.0%). Among the three taxa, the model performance was highest for *S. variabilis* and Crinoidea, and lowest for Brisingida, according to both the AUC and correlation statistics. The performance measures (correlation only) for the ensemble models reflect the mean performance of the three contributing models, and do not directly assess their ability to predict outside observed cells.

For the abundance models, cross validated correlations (Mean CV R^2) were lowest overall for Brisingida but similar among the three model types (15.1–15.7%); they were more variable for *S. variabilis* (from 12.7% for BRT to 24.2% for RF) and consistently highest for Crinoidea (22.4–33.5%). By this measure GAM and RF models performed similarly well, and consistently better than BRT models. Correlations (R^2) between predicted

TABLE 2 | VME indicator and habitat indicator taxa, the number of presence and absence records, and for presence records, the minimum and maximum number of records per video transect segment (25 m²).

Seamount	<i>Solenosmilia variabilis</i>		Brisingida		Crinoidea	
	Presence records (range)	Absence records	Presence records (range)	Absence records	Presence records (range)	Absence records
Forde	92 (1–33)	2,110	30 (1–11)	2,172	444 (1–40)	1,758
CenSeam	36 (1–2)	2,452	15 (1–2)	2,473	41 (1–4)	2,447
Anvil	75 (1–8)	1,155	12 (1–4)	1,218	85 (1–11)	1,145
JCM	4 (1–6)	135	5 (1–2)	134	7 (1–5)	132
39 South	95 (1–10)	2,060	37 (1–4)	2,118	96 (1–18)	2,059
Ghost	96 (1–18)	3,465	174 (1–17)	3,387	338 (1–33)	3,223
Valerie	146 (1–16)	1,813	68 (1–9)	1,891	165 (1–19)	1,794
Total	544 (1–33)	13,190	341 (1–17)	13,393	1,176 (1–40)	12,558

These data for all seven seamounts surveyed.

TABLE 3 | Final set of predictor variables used in the habitat suitability models.

Variable	Description
Seamount	Categorical variable indexing the six seamounts modeled
Slope	Slope (degrees)
Aspect	Aspect of slope based on a focal mean neighborhood size of 15 cells, in categorical form (equivalent to the 16 points of the compass)
Depth	Depth of the grid cell (m)
Slope SD	Standard deviation of slope values within a 7 × 7 grid of cells, based on a focal mean neighbourhood size of 7 grid cells
Curvature	Mean curvature based on a focal mean neighbourhood size of 15 grid cells
Ruggedness	Mean Vector Ruggedness Measure within a 5 × 5 grid of cells, based on a focal mean neighbourhood size of 15 grid cells
Backscatter	Backscatter values (dB)
Residual Autocorrelation (RAC)	Variable derived from residuals of initial models to account for spatial autocorrelation

and observed presence-absence for all model data were highest for BRT models (56.9–67.1%) and similar for RF and GAM models (18.7–36.4%). By this measure the models for Crinoidea again consistently performed better than those for the other two taxa. The all-data correlation values were considerably higher for abundance models compared with presence-absence models for BRT and GAM models (24.5–67.1%), but similar for RF models. By this measure BRT models (and the ensemble models) performed better than RF and GAM models.

Predictor Influence

For all model types, autocorrelation in the residuals (measured using Moran's index) was substantially reduced with the inclusion of the RAC variable. The RAC variable had the greatest influence in all models except for the abundance model for Crinoidea (Table 5). The most influential environmental variable in all the BRT models was Aspect, with a tendency for higher abundance and presence on westerly sector slopes. In contrast, for the GAM and RF models there was no single consistently important environmental variable for all indicator taxa and the two types of models. The variable SlopeSD was the most important for GAM presence-absence and abundance models for *S. variabilis*, while Curvature and Slope were the most important variables for Brisngida and Crinoidea GAM

models, respectively. Slope SD and Depth were the two most important environmental variables for RF models of *S. variabilis*, while Backscatter was more important for the abundance model of this species. Slope SD and Curvature were the most important environmental variables for RF models of Brisngida presence-absence and abundance, respectively. For RF models of Crinoidea, Slope was the most important variable. The least important environmental variables also varied among BRT, GAM, and RF models for VME indicator taxa and the two model types, but Seamount, Backscatter and Aspect (with the exception of the BRT models) were most often the least important variables (Table 5). See Figure S4 for details of the influence of the predictor variables across their full range in the model data.

Habitat Suitability Predictions

Generally across all seamounts, the distribution of suitable habitat (≥ 0.5 probability of occurrence for presence-absence models and > 0.5 live coral heads per 25×25 m grid cell for abundance models; thresholds = more likely to occur than not, and relatively high abundance, respectively) was predicted to be more extensive by the BRT models than either the GAM or RF models, although the basic distribution patterns were similar. Differences between the models were more pronounced in the presence-absence models than the abundance models. The

TABLE 4 | Comparison of performance statistics for four types of presence-absence and abundance models for *Solenosmilia variabilis*, *Brisingida*, and *Crinoidea* on seamounts of the Louisville Seamount Chain.

Model	Taxon	Presence-absence		Abundance	
		Mean AUC	R ² (%)	Mean CV R ² (%)	R ² (%)
BRT	<i>Solenosmilia variabilis</i>	0.923	32.0	12.7	57.7
	<i>Brisingida</i>	0.904	26.6	15.1	56.9
	<i>Crinoidea</i>	0.922	37.8	22.4	67.1
GAM	<i>Solenosmilia variabilis</i>	0.905	7.5	20.8	28.4
	<i>Brisingida</i>	0.868	8.3	15.3	24.5
	<i>Crinoidea</i>	0.913	16.0	32.1	34.3
RF	<i>Solenosmilia variabilis</i>	0.930	27.1	24.2	27.9
	<i>Brisingida</i>	0.926	23.7	15.7	18.7
	<i>Crinoidea</i>	0.924	33.7	33.5	36.4
Ensemble	<i>Solenosmilia variabilis</i>	–	19.2	–	35.9
	<i>Brisingida</i>	–	14.9	–	38.1
	<i>Crinoidea</i>	–	29.1	–	50.1

Performance measures for presence-absence models are the mean test AUC from a 10-fold cross validation (Mean AUC) and the correlation (R²) between predicted and observed values for all model data; performance measures for abundance models are the mean correlation (Mean CV R²) between predicted and observed values from a 10-fold cross validation procedure and the correlation (R²) between predicted and observed values for all model data (see text for more details). BRT, Boosted Regression Tree; GAM, General Additive Model; RF, Random Forest; –, not calculated.

distribution of suitable habitat predicted by the ensemble models for all taxa reflected the underlying averaging principle of this approach (e.g., **Figures 4A,B** for Forde Guyot; see Figure S5 for comparison among model types for other seamounts).

Because the abundance models performed better than presence-absence models, and ensemble models are practical for spatial management planning purposes (to avoid dependence on single model types), only the habitat suitability results of the ensemble-abundance models are described in detail hereafter. All the results of the other models are provided in Figure S5.

Suitable habitat for *S. variabilis* was generally predicted to occur at the summit-slope break, and on ridge-like features that extended down the flanks of the seamounts. Some areas of the seamount flanks were predicted to have concentrations of suitable habitat for this coral. Generally, these areas were on the north and north-west flanks of the seamounts. Where they occurred (particularly on Ghost Seamount), isolated knoll-like features on the flat summits of seamounts were also sometimes predicted to have suitable habitat for *S. variabilis* (**Figure 5**).

The distribution patterns of suitable habitat predicted by ensemble-abundance models for crinoids and brisingids were generally similar to those for *S. variabilis* across all seamounts (i.e., summit-slope break, ridges on flanks). However, highly suitable habitat for crinoids was more extensive than for brisingids on the seamount flanks (particularly on Forde Guyot where the mean number of crinoid observations per cell were over three times higher than for other seamounts) (**Figure 6**).

Correlations varied between the ensemble-abundance models for the VME indicator taxa *S. variabilis* and the two taxa that are supposedly indicators of the habitat formed by the stony coral. The correlation between the models for *Crinoidea* was relatively high ($R = 0.53$), compared to that for *Brisingida* ($R = 0.25$).

Uncertainty

Spatial patterns of model uncertainty varied among BRT, GAM, and RF models, but these patterns were generally similar between presence-absence and abundance models. Highest levels of uncertainty were generally associated with areas of highest habitat suitability for BRT models (i.e., seamount flanks), low habitat suitability at the deepest depths for GAM models (i.e., base of seamount flanks), and low habitat suitability at the shallowest depths for RF models (i.e., the flat seamount summit). Uncertainty for ensemble models was lower overall compared to the other models, and where areas of relatively high uncertainty occurred for ensemble models they were limited to small patches on the flanks of the seamounts (e.g., **Figure 7** abundance models for *S. variabilis* on Forde Guyot; see Figure S5 for all models for other taxa and seamounts).

Distribution Predictions for Coral Reef VME Habitat

The ensemble-abundance models for coral reefs predicted that this VME habitat was sparsely distributed on all seamounts (**Figure 8**). Where it was predicted to occur at the summit-slope break and ridges on the seamount flanks, the patch size of this habitat was typically small with few relatively large patches (range from 625 to 42,500 m²), and did not total more than 0.6 km² or 0.09% of the modeled area of any single seamount (**Table 6**, **Figure 9**).

DISCUSSION

The results of our study contribute to the growing number of examples that demonstrate the potential for using high-resolution habitat suitability models for the conservation and management of VMEs in the deep sea. Our models predict that suitable habitat for the stony coral VME indicator species *S. variabilis* is found at distinct breaks in slope and on elevated ridge-like features on seamounts. The general pattern of distribution predicted by our models conforms to expectations from previous observations of the distribution of stony corals at seamounts, canyons and continental slopes, and the topographic-related factors that are thought to control their distribution at these locations (e.g., Frederiksen et al., 1992; Thiem et al., 2006; Reveillaud et al., 2008). The predicted small-scale distribution patterns for *S. variabilis*, and the topographic variables identified as being important in these models (Aspect, Slope, SlopeSD, Curvature), are similar to those resulting from previous habitat suitability models for similar coral species found elsewhere (e.g., Dolan et al., 2008; Howell et al., 2011; Rengstorf et al., 2012).

Habitat suitability models for crinoids and brisingids, taxa that are considered to be indicators of VME habitat (Parker et al.,

TABLE 5 | Variable contributions to each type of habitat suitability model for each VME indicator taxon.

	RAC	Curvature	Aspect	Ruggedness	SlopeSD	Depth	Slope	Backscatter	Seamount
PRESENCE-ABSENCE MODELS									
<i>S. variabilis</i>									
BRT	45.3	3.7	18.1	4.1	7.6	5.2	8.3	3.1	4.5
GAM	55.5	6.8	1.5	5.0	11.7	8.5	7.5	1.6	2.1
RF	26.0	9.3	8.8	9.8	11.5	11.6	9.3	10.3	3.5
Brisingida									
BRT	42.6	8.4	16.1	8.4	8.8	6.4	5.1	2.3	2.1
GAM	39.6	15.9	0.5	16.2	14.7	5.7	6.2	0.7	0.5
RF	24.7	10.3	9.9	11.3	12.5	10.1	9.2	9.7	2.3
Crinoidea									
BRT	40.2	6.6	12.8	6.9	5.0	6.8	10.4	2.2	9.1
GAM	38.9	10.7	0.2	8.3	12.4	9.2	16.7	3.5	0.0
RF	23.4	8.8	9.1	9.7	9.7	10.6	12.6	8.3	7.7
ABUNDANCE MODELS									
<i>S. variabilis</i>									
BRT	26.2	17.3	19.1	7.6	6.1	6.0	10.9	5.1	1.8
GAM	48.5	7.8	0.6	9.2	11.5	8.9	9.4	1.2	2.9
RF	28.0	9.7	7.4	10.8	10.4	8.5	10.7	12.0	2.6
Brisingida									
BRT	32.0	16.4	15.6	11.8	9.6	7.1	5.1	2.1	0.4
GAM	47.7	16.7	0.1	16.6	11.2	3.1	3.9	0.3	0.3
RF	20.1	13.6	11.3	11.7	12.8	8.8	9.3	10.4	1.9
Crinoidea									
BRT	23.4	9.1	28.0	4.8	5.4	8.4	7.8	5.3	7.8
GAM	52.4	8.7	0.3	8.0	9.3	7.9	11.0	2.4	0.1
RF	25.4	9.6	11.4	9.7	8.7	10.2	10.4	9.3	5.4

Values are the percentage influence of each variable in each model. See text for an explanation of how these values were calculated. BRT, Boosted Regression Tree; GAM, General Additive Model; RF, Random Forest.

2009), were correlated with those for the stony coral *S. variabilis*. However, the correlations were not strong, particularly for brisingids. As such, in a habitat suitability modeling context, these taxa are not likely to be useful model proxies for the presence of coral habitat. Many observations suggest an association between the occurrence of crinoids and brisingids and the stony coral, and the contribution of their presence to the “VME indicator score” used by observers on fishing vessels operating the “move-on rule” (Parker et al., 2009) appears to be supported to some extent by the present study. However, despite their co-occurrence with stony corals in areas of high relief and hard substrate, there is also a growing body of data showing that both of these taxa can occur on bare rock surfaces, and that Brisingids can occur on soft sediment seabeds.

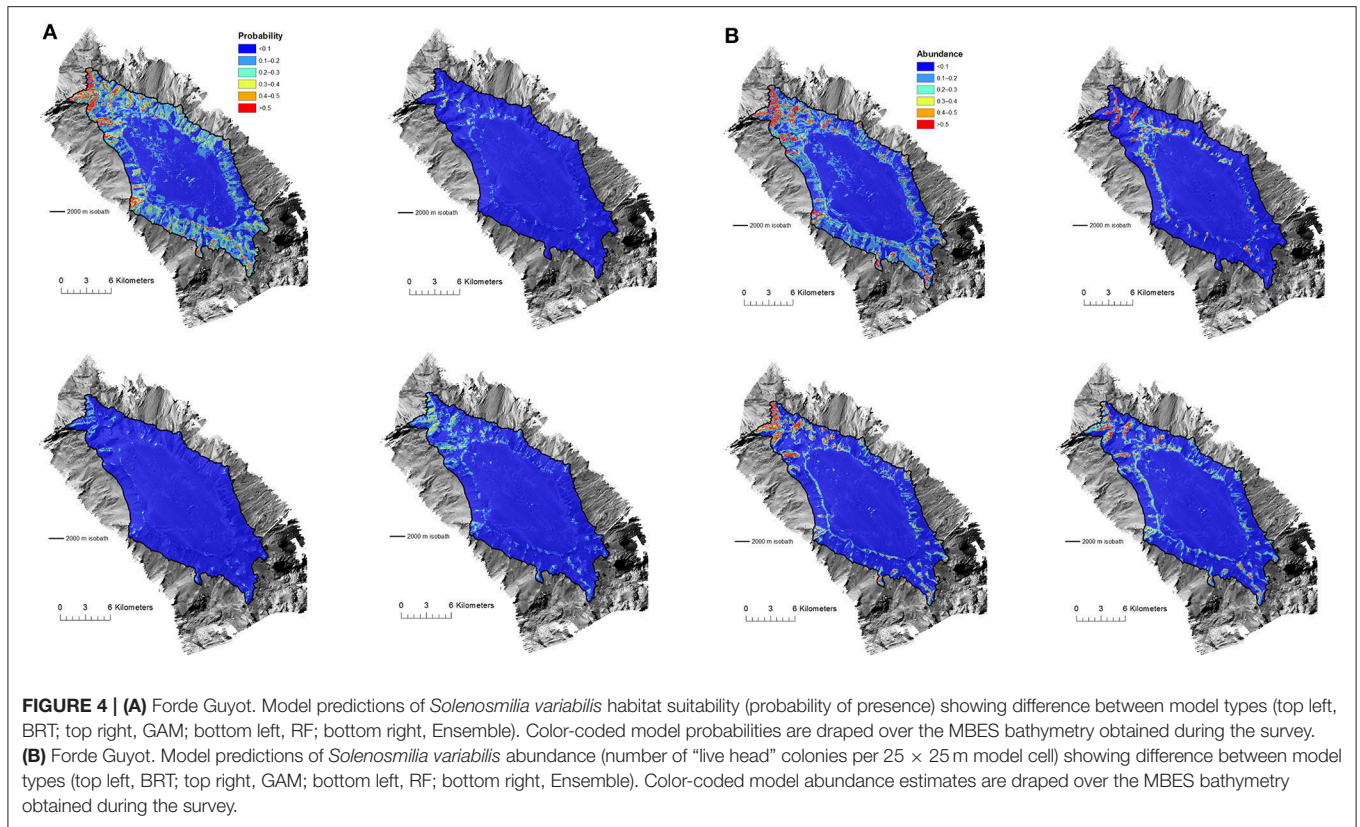
Below we discuss the results of our study with particular reference to improving high-resolution habitat suitability modeling, identifying and mapping VME habitat, and implications for spatial management in the SPRFMO Convention Area. For discussion about the limitations of the model types and general modeling methodology used in this study, please see Anderson et al. (2016b).

High-Resolution Habitat Suitability Mapping

The efficacy of habitat suitability modeling, using seafloor camera imagery and bathymetric and backscatter data obtained

from multibeam surveys to make high-resolution predictive distribution maps for benthic species of conservation or management significance in the deep sea was first demonstrated off SW Ireland by Dolan et al. (2008). Since then similar models for VME indicator taxa have been made and developed for the same area and elsewhere in the NE Atlantic Ocean (Guinan et al., 2009; Howell et al., 2011; Rengstorf et al., 2012, 2013, 2014; Tong et al., 2013; Robert et al., 2016). However, none of these studies produced abundance-based models for VME indicator taxa, nor used measures of abundance to model directly the distribution of VME habitat.

Howell et al. (2011) modeled the distribution of the presence of the stony coral *Lophelia pertusa* and coral reef habitat, but the latter was based on data from the subjective identification of reefs by eye. Nonetheless, this study demonstrated the importance of distinguishing between models for VME indicator taxa and VME habitat, and the authors suggested that “mapping efforts should focus on the habitat rather than the species at fine (100 m) scales” (Howell et al., 2011). Robert et al. (2016) used quantitative density data in their high resolution models but did so indirectly by modeling the distribution of benthic assemblages (including a *L. pertusa*-dominated assemblage) that were first identified objectively using multivariate analyses based on abundance data for individual taxa. Their models also used the occurrence of coral and coral rubble substrata (classes derived from backscatter data) as predictor variables, and thus appear to include a degree



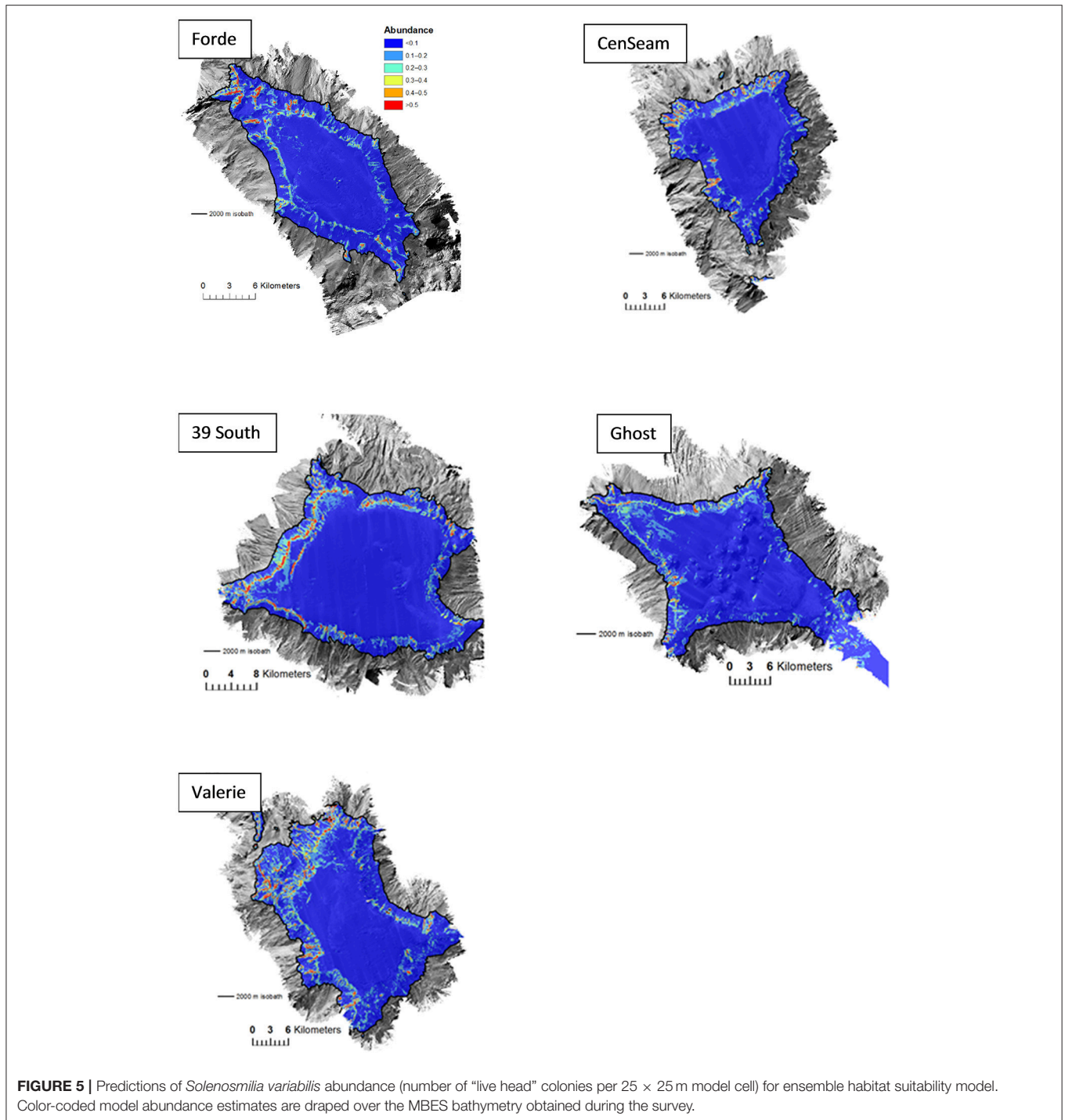
of circularity (at least for predicting the occurrence of the *L. pertusa*-dominated assemblage). Rengstorf et al. (2014) used species occurrence proportion data for *L. pertusa* (i.e., number of presence records compared to the number of absence records in a grid cell) as a measure of coral prevalence. While they did not use abundance data in their models, they nonetheless found that models that included the semi-quantitative proportional data performed better than models based on presence-absence data, had lower coefficients of uncertainty, and were considered more reliable (Rengstorf et al., 2014).

Rengstorf et al. (2014) advocated the use of proportion data because of the time-consuming nature of obtaining true abundance data for benthic taxa from seabed imagery. However, where abundance data are available, they can significantly improve model performance and provide a more comprehensive understanding of spatial patterns, as clearly demonstrated by the present study.

Mapping VME Habitat

FAO (2009) notes that “merely detecting the presence of an element itself (i.e., a VME indicator taxa) is not sufficient to identify a VME.” Thus, as well as model improvements for VME indicator taxa, abundance data also offer the prospect of making distribution models for VME habitat based on a density threshold for habitat-forming taxa, rather than using data based solely on subjectively identifying VME habitat, or data for benthic assemblages that act as a proxy for VME habitat. However, VME habitat is not always or consistently defined by

the density of the habitat-forming taxa. For example, definitions of coral reef habitat are not provided by FAO (2009) in their initial list of VMEs, and the longer description of coral reef habitats provided in support of the OSPAR List of Threatened and/or Declining Species and Habitats does not indicate any density or coverage thresholds (Hall-Spencer and Stehfest, 2009). In the few studies where deep-sea coral reef habitats are identified based on a particular density or coverage in a VME mapping context, values differ depending on the type of coral habitat. For example, Vertino et al. (2010) defined 20–40% coverage of dead and/or live coral colonies per video frame (~2.4 m²) for “Coral framework and hardground” habitat compared with >60% coverage for “Coral framework” habitat. These threshold-based identifications of VME habitat often include both dead and live corals, which reflects the typical composition of coral reef habitats. The amount of dead coral can be extensive, as found in the present study, by Thresher et al. (2011) off Tasmania, and by Vertino et al. (2010) in the Mediterranean where they noted live colonies rarely cover more than 30% of the seafloor. In the present study we constructed predictive maps of VME habitat based on the definition for coral reefs and thickets (as “sensitive environments”) provided by the regulations associated with New Zealand’s EEZ Environmental Effects Act. However, this threshold-based definition is essentially a subjective regional translation of the “you know it when you see it” descriptions provided by the likes of OSPAR, rather than a density of corals determined directly from study of the structural or functional attributes that distinguish coral reefs as VMEs.



While our attempt to predict the distribution of VME habitat is currently defensible because no consensus exists about what density of corals is necessary to generate the structural and functional attributes of a VME, the lack of a robust density threshold-based definition of VME habitat potentially limits the acceptance of these maps for conservation and fisheries management purposes (see below). Nonetheless, the results of the present study are in line with the developments envisaged for habitat suitability modeling in order that predictive distribution

maps can be better used for the spatial management of VMEs (Ross and Howell, 2013; Reiss et al., 2014; Rengstorf et al., 2014; Vierod et al., 2014; Robert et al., 2016). These improvements include our use of an abundance-based multi-model ensemble approach to produce high-resolution predictive distribution maps, and presentation of spatially explicit measures of model uncertainty for VME indicator taxa and VME habitat across a group of seamounts that are targeted by a specific fishery.

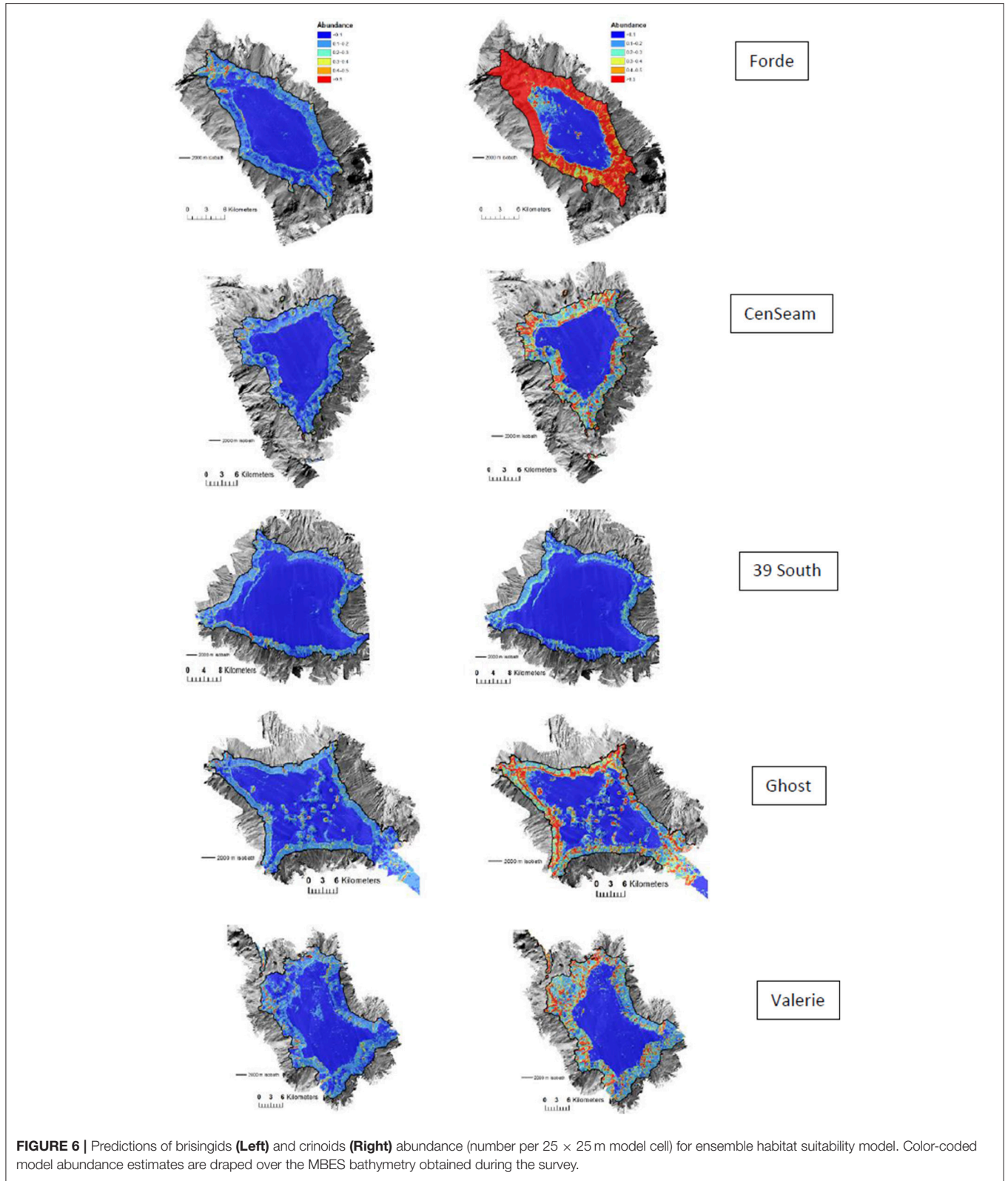


FIGURE 6 | Predictions of brisingids (**Left**) and crinoids (**Right**) abundance (number per 25 × 25 m model cell) for ensemble habitat suitability model. Color-coded model abundance estimates are draped over the MBES bathymetry obtained during the survey.

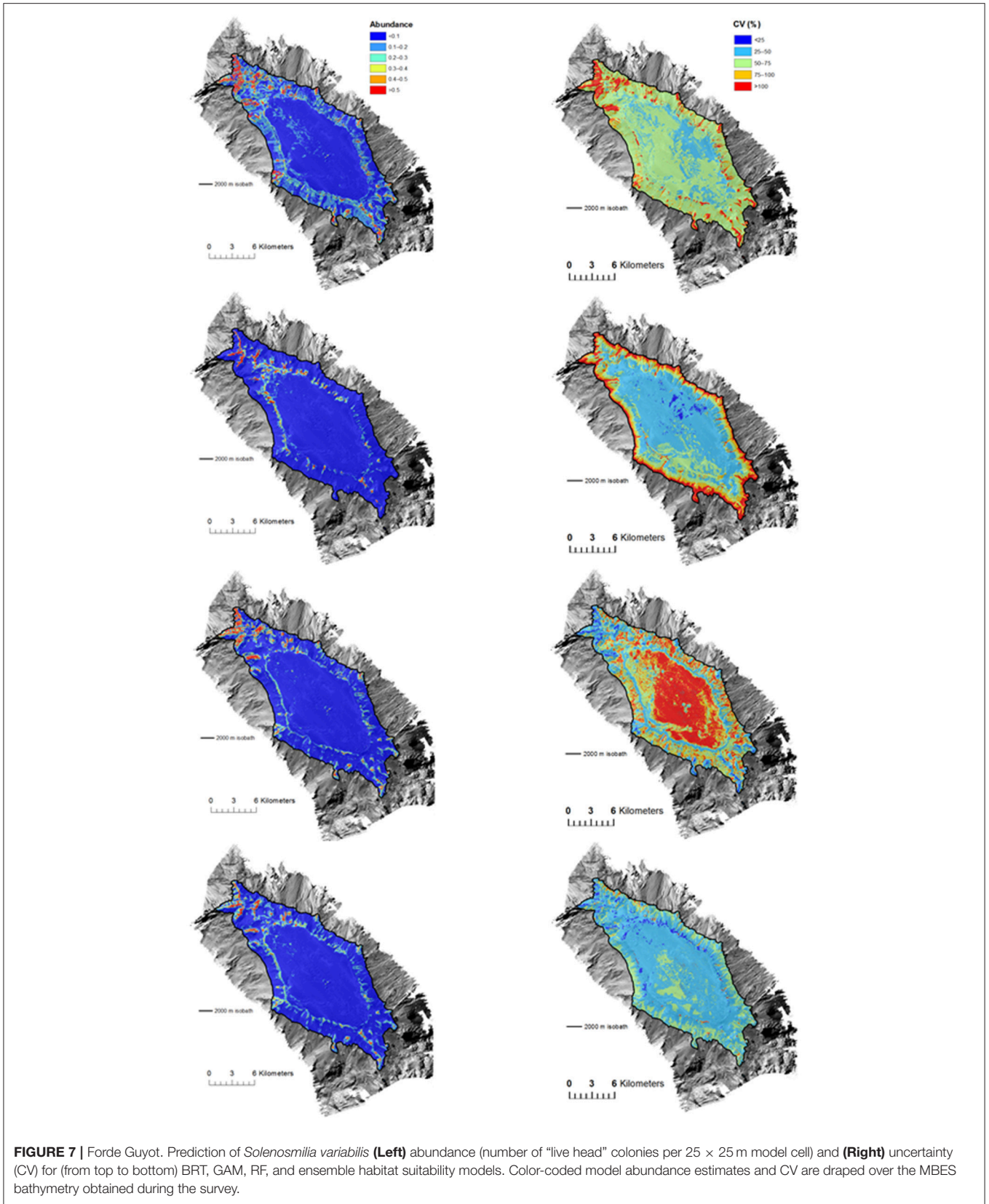


FIGURE 7 | Forde Guyot. Prediction of *Solenosmilia variabilis* (**Left**) abundance (number of “live head” colonies per 25 × 25 m model cell) and (**Right**) uncertainty (CV) for (from top to bottom) BRT, GAM, RF, and ensemble habitat suitability models. Color-coded model abundance estimates and CV are draped over the MBES bathymetry obtained during the survey.

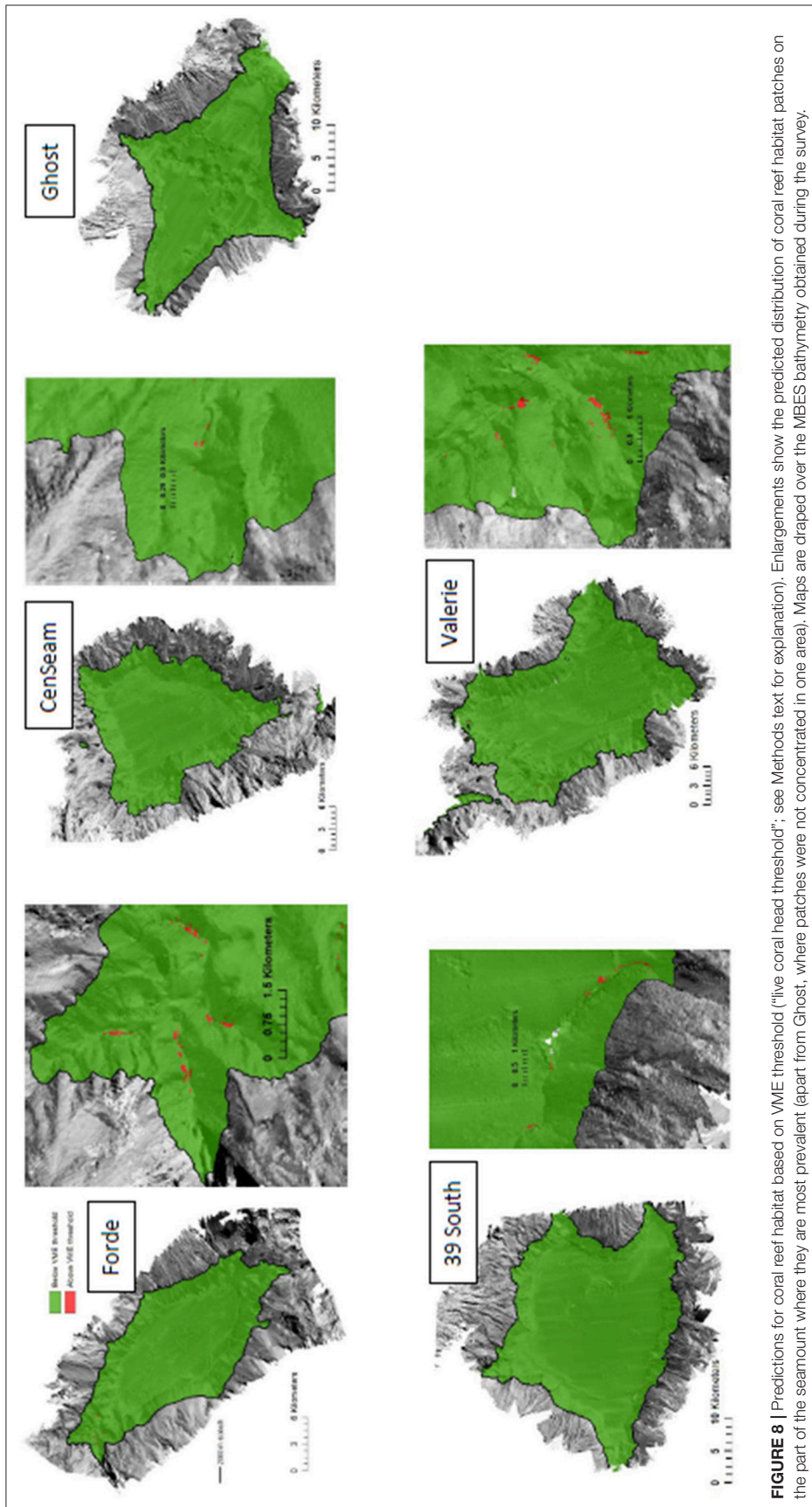


FIGURE 8 | Predictions for coral reef habitat based on VME threshold (“live coral head threshold”; see Methods text for explanation). Enlargements show the predicted distribution of coral reef habitat patches on the part of the seamount where they are most prevalent (apart from Ghost, where patches were not concentrated in one area). Maps are draped over the MBES bathymetry obtained during the survey.

TABLE 6 | Total seamount area within the modeled depth range (265–2,000 m), estimated area and proportion of VME coral reef habitat, and range of VME habitat patch size.

Seamount	Total area (km ²)	Area of VME coral reef habitat (km ²)	Proportion of VME coral reef habitat area (%)	Patch size range (m ²)
Forde	253	0.202	0.080	625–30,000
CenSeam	348	0.032	0.009	625–3,750
39 South	725	0.176	0.024	625–22,500
Ghost	568	0.008	0.001	625–1,250
Valerie	678	0.578	0.085	625–42,500

On each seamount the smallest estimated patch is by default a single 25 × 25 m model cell (625 m²).

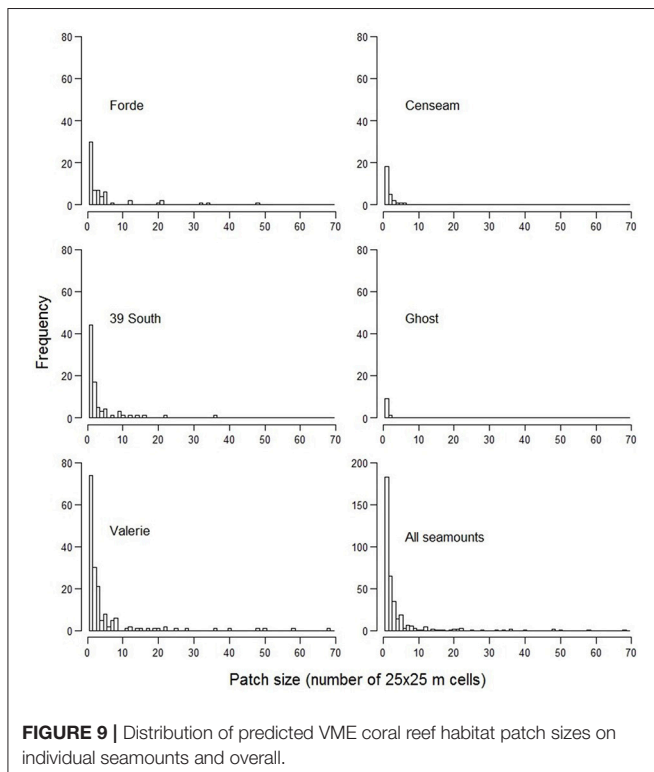


FIGURE 9 | Distribution of predicted VME coral reef habitat patch sizes on individual seamounts and overall.

Implications for Spatial Management in the SPRFMO Convention Area

New Zealand deepwater fishing stakeholders have proposed that, because their bottom trawling occurs exclusively in relation to seamounts in the SPRFMO Convention Area, and that commercially viable fish concentrations vary within and between seamounts, as do presumably the abundance of benthic species, spatial management measures should be based on individual seamounts (<https://www.sprfmo.int/assets/Meetings/Meetings-2013-plus/SC-Meetings/2nd-SC-Meeting-2014/Papers/SC-02-INF-04-Management-by-Sea-floor-Feature-of-Deepwater-Fisheries-in-the-South-Pacific-Ocean-b.pdf>). That is, open and closed areas should be defined on an individual seamount basis, with a sufficient fraction of each seamount closed to ensure adequate protection of VMEs. Fishing stakeholders have argued

for this approach because they believe that spatial management based on large, regular shaped areas (e.g., areas defined by 20-min of latitude and longitude) precludes them from fishing areas within closed areas that are not suitable for VMEs, and that it is not necessary to impose large closure areas that might extend over most or all of a seamount. Furthermore, with the accurate deployment of fishing gear using modern navigational methods, it is possible to effectively avoid relatively small closed areas where it is known that VMEs do occur on a seamount. The fishing stakeholders accept that in order for this strategy to be implemented it is necessary to map the distribution of VMEs at high-resolution on each seamount within the broader fishing area. As we have demonstrated through the present study, habitat suitability modeling using multibeam data and seafloor images can achieve this mapping goal.

The ensemble habitat suitability models for *S. variabilis* predicted that concentrations of relatively high abundances of this VME indicator taxon typically occur in the north/northwestern areas of the seamounts, and sometimes in southern areas. However, relatively high abundances of the coral (>5 live heads 25 m⁻²) were predicted to occur around the entire seamount at the summit-slope break, mostly on ridge-like features that extend down slope from the flat top of the seamount summit (860–2,000 m—maximum modeled depth). This result suggests that only the flat topped and soft sediment summits are relatively large and discrete areas of consistently low habitat suitability for *S. variabilis*, and thus where bottom fishing could occur with a low likelihood of encountering this VME indicator taxon. However, these areas are not generally favored by fishers. Fishing patterns revealed by logbook data and commercial fishers plotter lines differ between seamounts, but indicate that fishers tend to target either small localized hill features on the summits (e.g., Ghost, 39 South), or the upper slopes and flanks of seamounts (e.g., Valerie, JCM) where over 90% of tows occur away from the summit plateau on the flanks to depths of 1,200 m. Our habitat suitability models predict that areas of relatively high abundance of *S. variabilis* are likely to be encountered by bottom trawling the seamount flanks. Based on the results of the abundance models for this VME indicator taxon, and current knowledge of the areas of interest to fishers, designing spatial management measures on an individual seamount level would be problematic.

The abundance models were also used to identify and predict coral reef habitat. This VME was predicted to occur as relatively small patches in only a few relatively isolated areas on seamounts. This limited distribution and abundance of VME habitat on seamounts conforms to the experience of the fishers, and thus supports one of their arguments for reducing the size of closed areas on seamounts, and adopting an individual seamount-based spatial management approach for the Louisville Seamount Chain. However, this conclusion is predicated on the veracity of the threshold-based approach by which VME habitat was identified in the present study. If predictive models are to be used for designing spatial management at the within-seamount scale, research is required to establish coverage and/or density estimates that relate to the specific structural and functional attributes that define a VME. For example, Cathelot et al. (2015)

recently demonstrated that an area of 1,767 m² dominated by live corals with a density of visible live polyps of ~33 per 100 cm² can make a significant contribution to local ecosystem function in the deep sea. Some of the predicted coral reef habitat patches in the present study were greater than the size examined by Cathelot et al. (2015) (0, 18, 27, 38, and 41% of patches on the study seamounts were predicted to occupy ≥3 cells i.e., ≥1,875 m²) but most were smaller. So while it is probable that some of the coral reef habitat on the Louisville Seamounts Chain is providing important ecosystem function, the significance of the more numerous smaller patches of coral habitat is uncertain. The necessary patch size or polyp/live head density to represent significant sites of benthic respiration and organic carbon cycling, or significant habitat to support biodiversity or juvenile fish is unknown. Thus, we recommend that further research is needed to investigate and develop the way in which VME habitats are identified and included in habitat suitability modeling approaches (regardless of the resolution or spatial scale of the models) for the purpose of informing conservation and management. Such work should include multi-modeling approaches that incorporate the functional traits of species to improve the ecological interpretation of results, and support a broader ecosystem approach to management.

AUTHOR CONTRIBUTIONS

AR and MC conceived of the study. AR, MC, and DB designed the field survey. MC, DB, OA, and AP collected data from the field survey. DB, AM, and AP analyzed image and multibeam data from the survey. OA and SG undertook the modeling. AR, OA, SG, MC, and DB interpreted all data. AR led the

writing, and drafted text for the manuscript. OA, SG, MC, DB, and AP contributed draft text to the manuscript, and all authors revised and approved the final version of the manuscript.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fmars.2017.00335/full#supplementary-material>

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Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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