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\*CORRESPONDENCE Hao Liu Miuhao@nbu.edu.cn

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# The immunomodulatory effects of GLP-1 receptor agonists in neurogenerative diseases and ischemic stroke treatment

Haohui Sun, Yue Hao, Hao Liu\* and Feng Gao

School of Basic Medical Science, School of Medicine, Ningbo University, Ningbo, Zhejiang, China

Glucagon-like peptide-1 (GLP-1) receptor is widely distributed in the digestive system, cardiovascular system, adipose tissue and central nervous system. Numerous GLP-1 receptor-targeting drugs have been investigated in clinical studies for various indications, including type 2 diabetes and obesity (accounts for 70% of the total studies), non-alcoholic steatohepatitis, Alzheimer's disease, and Parkinson's disease. This review presented fundamental information regarding two categories of GLP-1 receptor agonists (GLP-1RAs): peptidebased and small molecule compounds, and elaborated their potential neuroprotective effects by inhibiting neuroinflammation, reducing neuronal apoptosis, and ultimately improving cognitive function in various neurodegenerative diseases. As a new hypoglycemic drug, GLP-1RA has a unique role in reducing the concurrent risk of stroke in T2D patients. Given the infiltration of various peripheral immune cells into brain tissue, particularly in the areas surrounding the infarct lesion, we further investigated the potential immune regulatory mechanisms. GLP-1RA could not only facilitate the M2 polarization of microglia through both direct and indirect pathways, but also modulate the quantity and function of T cell subtypes, including CD4, CD8, and regulatory T cells, resulting into the inhibition of inflammatory responses and the promotion of neuronal regeneration through interleukin-10 secretion. Therefore, we believe that the "Tregs-microglia-neuron/neural precursor cells" axis is instrumental in mediating immune suppression and neuroprotection in the context of ischemic stroke. Given the benefits of rapid diffusion, favorable bloodbrain barrier permeability and versatile administration routes, these small molecule compounds will be one of the important candidates of GLP-1RA. We look forward to the further clinical evidence of small molecule GLP-1RA intervention in ischemic stroke or T2D complicated by ischemic stroke.

#### KEYWORDS

glucagon-like peptide-1 receptor agonists, Treg cells, microglia, IL-10, ischemic stroke

## 1 The neuroprotection of glucagon like peptide-1 receptor agonists in neurodegenerative and cerebrovascular disorders

Glucagon like peptide-1 (GLP-1) is a peptide called enteropancreatin secreted by small intestinal L cells. Physiologically, it can bind to GLP-1 receptor on pancreatic beta cells through endocrine, activate adenylate cyclase, increase intracellular cAMP levels, promote glycolysis and ATP production, and enhance glucose-stimulated insulin secretion. The direct regulation of GLP-1 on insulin expression is significant for type 2 diabetes (T2D) patients with simple glucose tolerance but normal insulin secretion (1). It can avoid adverse reactions and potential risk of pancreatic gland degeneration caused by direct insulin treatment (2). It is worth noting that GLP-1 can easily spread to the central system and exert neuroprotective and neuropathic actions in various neurodegenerative and cerebrovascular disorders (Alzheimer's disease (AD), Parkinson's disease (PD), stroke, etc.) through biological signal transduction (3-5). However, due to the short half-life of endogenous GLP-1 in the body (only about 2 minutes), it will be rapidly degraded by dipeptidyl peptidase-4 (DPP-4) and neutral endopeptidase. Therefore, GLP-1 derivatives with medium to long acting properties have become the main GLP-1RA for clinical applications, such as exendin-4, exenatide, liraglutide, lixisenatide, semaglutide, etc. (6-8). These peptide-based GLP-1RA have been widely studied globally for improving cognitive dysfunction in patients with neurodegenerative diseases (AD and PD) (ID: NCT03456687; NCT01255163; NCT03439943; NCT03659682; NCT01843075; NCT02953665) (9–12).

# 2 Peptide-based GLP-1 derivatives and small molecule compounds of GLP-1RA

There are two main categories of GLP-1 receptor agonists, small molecule compounds and peptide-based GLP-1 derivatives as shown in Figure 1, including exenatide, liraglutide, dulaglutide, etc. Many literatures proved that GLP-1 derivatives performed neuroprotective effects and inhibited neuronal apoptosis (13–15). It was reported that cerebral expression of GLP-1 receptor protein reduced following subarachnoid hemorrhage (SAH), but markedly increased by liraglutide (15). Liraglutide also alleviated SAH-induced early brain injury and inflammation. Exendin-4 not only reduced the infarct size of ischemic stroke, improved the neurologic deficit scores in ischemia/reperfusion-induced rats, but also reduced the potential rtPA-induced hemorrhagic risk through the Wnt/ $\beta$ -catenin signaling pathway (16).

There are also a type of non-peptide small molecules, which are mainly extracted from herbal medicine, such as *Cornus officinalis*, *Gardenia jasminoides*, *Rehmannia glutinosa*, *Lamiophlomis rotata*, and *Strychnos nuzi* (17–20). These small molecule compounds mainly belong to iridoid glycosides, including geniposide,



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Shanzhiside methylester (SM), 8-O-acetyl-SM, morroniside, catalpol, genipin methyl ether and so on (18, 21). These GLP-1RA could be used to treat diabetes and its complicated obesity, fatty liver, hypertension and various cardiovascular diseases (22). In addition, accumulating studies demonstrated that GLP-1RA performed neuroprotective effects in a variety of neurodegenerative diseases (mainly AD, PD and stroke) (23–25).

Even for patients without complications of diabetes, their cognitive functions were still significantly improved (26, 27). The notable therapeutic impact of peptide-based GLP-1 derivatives in clinical neurodegenerative disorders had fostered enthusiasm among researchers to develop small molecule GLP-1RA. It was reported that small molecule GLP-1RA could reduce chronic neuropathic pain and exert neuroprotective effects by improving synaptic plasticity and ameliorating neuronal death (28, 29). GLP-1RA inhibited inflammatory responses in chronic pain through the GLP-1R/PI3K/Akt/ERK-1/2/IL-10 signaling pathway and reduced the release of inflammatory factors such as IL-6, IL-1, tumor necrosis factor alpha (TNF- $\alpha$ ) (30). In addition, Wang Y.X. also found that the analgesic effects of various small molecule GLP-1RA such as lemairamin mainly relied on their promotion of IL-10 secretion and subsequent IL-10R/ $\beta$  endorphin signaling pathways, which effectively alleviated bone cancer pain and neuropathic pain (31-33). In these studies related to analgesia, shanzhizhi methyl ester performed a dose-dependent and persistent (>4 hours) effects against allodynia (17). In rodent models of neurodegenerative diseases, a series of iridoid glycosides represented by geniposide and mononucleoside could effectively resist cellular oxidative damage and counteract neuronal apoptosis (19, 28). In summary, as a promising class of small molecule GLP-1RA, iridoid glycosides represent a potential therapeutic avenue for the management of neurodegenerative disorders.

# 3 The neuroprotection of GLP-1RA in experimental stroke

Many literatures indicated that GLP-1RA were also considered as the latent intervention strategy against ischemic stroke to improve the pathological symptoms and prognosis. The principal neurological function regulated by GLP-1RA could be summarized as follows (Figure 2): (1) GLP-1RA liraglutide and exendin-4 effectively inhibited neuronal apoptosis (15), and ultimately improved early brain injury by suppressing NLRP3 inflammasome and inflammatory reaction in microglia (34-36); (2) Exendin-4 meliorated vascular permeability by inhibiting bloodbrain barrier breakdown and leakage (37), and reduced the hemorrhagic risk of ischemic insults via the Wnt/beta-catenin signaling pathway (16). Besides, exendin-4 also dilated cerebral arterioles, raised cerebral blood flow and finally reversed the cerebral ischemia (38). (3) GLP-1RA (e.g. morroniside, exendin-4) promoted M2 polarization in microglia by upregulating nuclear factor erythroid 2-related factor 2 (28), and further improving synapse growth and neuronal regeneration via the cAMP/PKA pathway (39), and finally reduced cognitive and memory impairment induced by stroke (40). GLP-1RA appeared to exert



neuroprotective effects primarily by regulating microglia, which were the most significant immune cells in the central nervous system (41).

# 4 GLP-1RA reduced the risk of stroke in T2D patients clinically

As a new type of hypoglycemic drug, GLP-RAs are currently mainly used in clinical practice to evaluate the prognosis of various complications in T2D patients, such as stroke, kidney injury, fatty liver, and cardiovascular disease (42, 43). Compared with other new hypoglycemic drugs such as SGLT2 and DPP-4, GLP-1RA have a unique preventive effect in reducing the risk of stroke as shown in Table 1 (43, 45). With the ageing of society, stroke has emerged as the leading cause of mortality and major cause of disability worldwide, greatly increasing the burden of human society (46). According to the degree of severity, stroke-related disability led to premature death with disability-adjusted life-years (DALYs) (47). There was a notable discrepancy in the proportion of deaths resulting from hemorrhagic stroke among different racial and ethnic groups. The proportion of deaths due to hemorrhagic stroke among middle-aged and older people (≥35 years) in the 1990s was 38% for Asians, 32% for Hispanics, 24% for blacks, and 18% for whites (47, 48). The high rates of intracerebral hemorrhage (ICH) in Asian populations have been attributed to the high prevalence of hypertension, smoking and metabolic syndrome (49). Using Caucasians as a reference, the adjusted relative risk (RR[95%CI]) of ICH in Asians was 1.6[1.1-2.3] (P=0.01) (50, 51). The mortality rate increased by 32% from 1990 to 2019 (52). In 2019, there were 2.2 million deaths and 45.9 million DALYs after stroke insults in China (53). By 2050, the burden of stroke in individuals (aged  $\geq 65$  years) will increase significantly: 104.7% in incidence, 218.5% in prevalence, 100.0% in mortality, and 58.9% in DALYs (54).

Many systematic reviews with network meta-analysis summarized the risk of noval hypoglycemic drugs on diabetic complications, including cardiovascular events, nonfatal stroke, renal composite outcomes and fatty liver (42–45). By using random-effects model, the network meta-analysis indicated that the risk profile of DPP-4i was same as placebo in all the outcomes (44). Distinctly, both SGLT-2i and GLP-1RA significantly decreased the risk of cardiovascular events (odds ratios (OR)[95%CI], GLP–1RA: 0.87[0.82-0.93]; SGLT–2i: 0.88[0.82-0.95]), and renal composite outcome (GLP–1RA: 0.86[0.78-0.94); SGLT–2i:0.59[0.52-0.67]) when compared to placebo (Table 1). In addition, only administration of GLP-1RA resulted into lower nonfatal strokes (OR, 0.88[0.77-0.99]) than those receiving placebo. P-rank scores confirmed that GLP-1RA reduced the risk of nonfatal stroke by 80.6%.

The similar viewpoints were also supported by other critical reviews and clinical trials (55–58), and inferred that GLP-1RA could be considered in patients at a high risk of, or with established cardiovascular diseases to improve the prognosis of cardiovascular events (especially nonfatal stroke), whilst SGLT2i could be recommended for patients with heart failure or chronic kidney diseases.

### 5 Immune dysfunction in ischemic stroke and potential immunomodulatory effects of GLP-1RA

# 5.1 The regulatory function of immune cells in the progression of ischemic stroke

The latest research shows that healthy brains do not lack immune cells. Normal cerebrospinal fluid contains about half a million immune cells, with T cells being the main type (about 50% CD4<sup>+</sup> and 20% CD8<sup>+</sup>) (59). These cells interact bidirectionally with resident immune cells microglia and astrocytes, participating in antigen presentation by antigen-presenting cells, activation of T cells, and promotion of lymphatic system circulation in cerebral spinal fluid (60). In the pathological state of ischemic stroke, inflammatory signals released by dead neurons or cytokines and chemical inducers secreted by microglia under stress could drive circulating lymphocytes to recruit to the damaged brain (61, 62). Concurrently, the disruption of the blood-brain barrier results in a notable influx of immune cells into brain tissue. A substantial body of evidence suggests that both peripheral innate immune cells (such as neutrophils, macrophages, and natural killer cells) and adaptive immune cells (like T cells and B cells) are present in the cellular infiltration of the cerebral embolism area. Neutrophils are one of the earliest cells to infiltrate the injured brain. Following cerebral ischemia, the substantial release of various cytokines, chemokines and damage-associated molecular patterns (DAMPs) attributes to the activation and recruitment of neutrophils from the bone marrow, spleen, and peripheral circulation to the site of damage (Table 2). This process entails the adhesion of the peripheral immune cells to endothelial cells, which facilitates their translocation across the blood-brain barrier, ultimately resulting in barrier disruption, cerebral edema, and brain injury, which indirectly promote the

TABLE 1 New hypoglycemic drugs reduced the risk of multiple complications in T2D patients.

Novel hypoglycemic drugs	Cardiovascular events (43, 44)	Nonfatal stroke (43)	Renal events (43, 45)	Fatty liver (42)
GLP-1RA	++	++	++	++
SGLT2i	++		+++	++
DPP-4i				

++ represent significance compared to placebo; +++ represent significance compared to other positive drug (++).

Immunophenotyping of effector cells	Publication date	The infiltrating situation of Immune cell subtypes	
Microglia	2015	After the onset of stroke, microglia are immediately activated. Microglial activity involved in tissue damage peaks within 3 to 5 days. (63)	
Astrocytes	2023	Astrocytes are activated several minutes after stroke. (64)	
CD8 <sup>+</sup> T cells	2021	CD8+ cytotoxic T cells are the first T cell subset to infiltrate the infarct lesion within hours after stroke. (65)	
CD4 <sup>+</sup> T cells	2021	CD4+ T cells appear at approximately 24 hours after ischemia. (65)	
CD3 <sup>+</sup> T cells	2021	The count of CD3+ T cells reached its highest level on the seventh day following pMCAO. (65)	
Neutrophils	2014	After pMCAO (permanent middle cerebral artery occlusion), neutrophils appear in the pia mater and brain parenchyma within 6 and 12 hours, respectively. Neutrophil recruitment peaks within 1 to 3 days and then gradually decreases, reaching a peak between 3 to 5 days after tMCAO. (66)	
Dendritic cells	2010	DCs accumulated in the ischemic hemisphere within 24 hours after MCAO reperfusion, particularly in the infarct border region where T lymphocytes gathered, and this accumulation persisted for at least 7 days. (67)	
Monocytes/macrophage	2012	Macrophages are rarely detected within the first 48 hours, Their level gradually increases, with a peak during the first week after stroke. (68)	

TABLE 2 The infiltrating situation of different subtypes of immune cells after the onset of ischemic stroke.

infiltration of peripheral immune cells (69). The number of infiltrating neutrophils peaks on day 3 after cerebral infarction, persists for at least a week, before gradually declining (70). In contrast, T cells preferentially migrate to the lesion boundary, with a marked increase in a few days after ischemia, reaching a peak within one week and persisting for months (71). CD8<sup>+</sup> cytotoxic T cells are the first T cell subset to infiltrate the infarct lesion within hours after stroke, whereas CD4<sup>+</sup> and natural killer T cells appear at approximately 24 hours after the onset of ischemia (66). In rodent models of stroke, the number of CD3+ T cells reached a peak on day 7 after permanent middle cerebral artery occlusion (pMCAO), followed by a significant proliferative phase (65). Even on day 14 and 28 after pMCAO, the number of CD3<sup>+</sup> T cells significantly increased in the ipsilateral brain, not only in the distal core but also in the corpus callosum (72). In contrast, Treg cells require a considerable period of time, spanning several days, to invade the brain following the onset of ischemia, and maintain in a significant number for more than one month (71). In total, T cell subsets exhibited a long-term activation state in experimental ischemic stroke, which suggests a potential harmful role in the early phase and a protective effect in the later phase (73).

CD4<sup>+</sup> T cells contribute to the exacerbation of cerebral inflammation and the induction of neuronal death (74, 75). The depletion of CD4<sup>+</sup> or CD8<sup>+</sup> T cells in mice subjected to transient MCAO was found to effectively reduce neuronal apoptosis, decrease infarct volume, and promote neurogenesis (76). The removal of CD25<sup>+</sup> T cells (including Treg cells) was found to have a partial inhibitory effect on neurogenesis and hinder the repair of neural function (77). Treg cells primarily prevented secondary brain injury by coordinating lymphocyte and microglia infiltration in ischemic stroke through IL-10 signaling (78, 79). Treg cells could not only interact with microglia (75), but also regulate their polarization from M1 to M2 via IL-10 pathway (28, 80, 81). The aforementioned studies also demonstrated that Treg cells could not only mitigate neurological damage, but also regulate the steady-state of peripheral immune responses, including the correction of immune suppression and the reduction of peripheral inflammation (82). Conversely, M2 microglia promoted nerve regeneration through two pathways: (1) microglia could secrete an insulin-like growth factor (IGF-1), which increased the proliferation of neural precursor cells (NPCs); (2) M2 microglia regulated immunity and the balance between oligodendrogenesis and neurogenesis by secreting IL-4 and IL-10 (83, 84). The abovementioned content on ischemic stroke has constructed a potential concept of "T cell-microglia-neuron/NPCs" axis (60). We can explain the potential mechanism of regulating immunological function and nerve regeneration after ischemic stroke based on this concept.

# 5.2 Regulation of T cell subtypes CD4/CD8 and Treg cells by GLP-1RA

GLP-1R is expressed in various immune cell populations of NOD mice, with a higher proportion of mature  $\mathrm{CD4}^{\scriptscriptstyle +}$  and  $\mathrm{CD8}^{\scriptscriptstyle +}$  T cells (approximately 5%-6%). Following T-cell activation GLP-1R expression increased significantly, particularly in CD8<sup>+</sup> T cells (85). Male Glp1r<sup>-/-</sup> mice could induce a significant increase in peripheral lymphocytes, but significantly reduced the percentage of CD4<sup>+</sup>CD25<sup>+</sup>FOXP3<sup>+</sup> Tregs, and had no effect on cell apoptosis and migration (86). T helper cell 17 (Th17) and regulatory T cells (Treg) are two immune cells with opposing functions. Th17 cells promote inflammatory responses, whereas Treg cells inhibit inflammation and maintain peripheral tolerance. Exenatide could promote the increase of Treg cells in mice, but inhibit the tissue infiltration of Th17 cells, suggesting that exenatide exerts immunomodulatory effects by correcting Th17/Treg imbalance (87). The PI3K/Akt/FOXO1 pathway might be involved in the regulation of Th17/Treg balance by exenatide, while FOXO1 inhibitors could block the regulatory effect of exenatide on Th17/Treg balance. In the NTS model, Glp1r<sup>-/-</sup> mice exhibited immunomodulatory effects, increasing tissue infiltration of neutrophils and T cells. The Th17 marker gene Roryt and the Th1 and Th2 regulatory genes Tbet and Gata3 were also

significantly elevated, accompanied by a significant increase in various systemic inflammation-related genes such as interferon gamma (IFN-y), etc (88). In vitro experiments showed that liraglutide could reduce the proliferation activity of CD4<sup>+</sup>IFN- $\gamma^+$  T cells (Th1) and CD4<sup>+</sup>IL-17a<sup>+</sup> T cells (Th17), but increase the number of CD4<sup>+</sup>IL-4<sup>+</sup> T cells (Th2) and CD4<sup>+</sup>CD25<sup>+</sup> T cells (Treg). And this regulatory effect was ineffective in Glp1r<sup>-/-</sup> mice, indicating that the immunoregulatory and anti-inflammatory effects of liraglutide depended on the GLP-1R signaling pathway (89). Besides, GLP-1RA NLY01 could significantly reduce the proportion of infiltrating white blood cells (CD45<sup>high</sup>) and monocytes (Clec12a<sup>+</sup>) in the central nervous system (CNS), while decreasing the number of effector/ memory T cells (CD4<sup>+</sup>CD44<sup>+</sup>) and inhibiting inflammation in peripheral circulation and CNS (90). In summary, GLP-1RA could reduce the proliferation of effector T cells, promote the generation of Treg cells, and enhance the immunosuppressive function of Treg cells (Figure 3) (91).

Early immune cell infiltration in rodent stroke is dominated by neutrophils and macrophages/microglia, whereas in human ischemic stroke it is different, consisting mainly of neutrophils and T cells (92). In clinical practice, it was also found that immune activation led to a rapid and significant upregulation of GLP-1R expression in naive CD4<sup>+</sup> T cells enriched more than 40-fold from the human body (93). Among different polarization conditions (including Th1, Th2, Th17, and Treg polarization conditions), the expression level of GLP-1R was highest in CD4<sup>+</sup> T cells under Treg polarization conditions, and the proportion of GLP-1R<sup>+</sup> cells could reach 29-34%. These GLP-1R<sup>+</sup> cells were often accompanied by high expression of Foxp3 and CD25 and low expression of IL-7R $\alpha$ (CD127), which were characteristic protein markers of Treg cells. Therefore, this study provided the first evidence of functional GLP-1R expression in human iTreg cells, and suggested that GLP-1R might play a key role in the anti-inflammatory effects attributed to GLP-1RA. In vitro cell therapy also indicated that infusion of Treg



Blocking the recruitment of peripheral effector T cells could significantly reduce neuronal damage and improve clinical prognostic indicators in cerebral ischemia

cells could increase the expression of anti-inflammatory factors (such as IL-10, IL-4, TGF- $\beta$ , IL-2), while reducing the levels of proinflammatory factors (such as IL-17, IFN- $\gamma$ ) (94). We speculated that GLP-1RA might regulate Treg cells with high expression of GLP-1R after activation, and exert anti-inflammatory effects by increasing anti-inflammatory factors such as IL-10.

### 5.3 The "Tregs-Microglia-Neuron/NPCs" axis and GLP-1R/cAMP/PKA/IL-10 mediated immunosuppressive and neuroprotective effects of GLP-1RAs in ischemic stroke

In summary, in the pathological state of ischemic stroke, T cells preferentially migrated to the lesion boundary, and significantly increased after ischemia, peaking in the following weeks and persisting for several months (71). CD8<sup>+</sup> cytotoxic T cells are the first subpopulation of T cells that invade the ischemic brain within a few hours after stroke (66), while Treg cells take many days to infiltrate the brain after the onset of ischemia and remain in significant numbers for more than one month (71). Neuroinflammation after cerebral ischemia is the combined result of the activated resident microglia and infiltrating peripheral leukocytes. Despite the huge difference in their numbers, with only a few thousand lymphocytes invading the brain and more than 50 times higher numbers of resident microglia, microglia could amplify the impact of T cells on the cerebral immune environment, making T cells the largest subtype of white blood cells contributing to secondary inflammatory damage after ischemia (70). In lymphocyte-deficient mice (T and B cells), the number of activated microglia was significantly reduced after ischemic stroke, particularly in the ischemic border zone (95). Furthermore, blocking the recruitment of peripheral effector T cells could significantly reduce neuronal damage and improve clinical prognostic indicators in different cerebral ischemias (Figure 3) (76, 96). Therefore, we speculated that T cells could induce microglial polarization through direct contact, cytokinemediated communication or antigen presentation, thereby amplifying their impact on the cerebral immune milieu.

Further studies showed that different subtypes of T cells had apparently different effects on microglia. Differentiated Th1, Tregs, and IL-10 overexpressing engineered T cells were separately injected into the cerebellar medullary cistern of lymphocyte deficient (Rag1<sup>-/</sup> <sup>-</sup>) mice after 24 hours in MCAO-induced insults. The results showed that Th1 cells polarized microglia towards high expression of INF response related genes (Irf7 and Stat1), which further exacerbated the immune response in the later stages of neurodegenerative diseases. In contrast, single injection of Tregs or IL-10 overexpressing engineered T cells promoted gene expression of chemokines/cytokines in microglia (Ccl2, Ccl7, and Cxcl10), regulated the chemotactic behavior of microglia and neural stem cells, promoted angiogenesis, and was speculated to contribute to the repair of ischemic injury (95). Another study also demonstrated that depletion of Treg cells could increase the area of delayed cerebral infarction and further exacerbate neurofunctional damage (78). The absence of Treg cells enhanced the activation of resident and invading inflammatory cells (including microglia and T cells) after ischemia, which were the main sources of harmful TNF- $\alpha$  and IFN- $\gamma$ , respectively. In addition, IL-10 could counteract the overexpression of cytokines and delayed brain injury caused by Treg cell deficiency. Similarly, IL-10-deficient Treg cells could not ameliorate ischemic injury, indicating that IL-10 signaling plays a crucial role in immune suppression and neuroprotection of Treg cells. The above evidence suggests that crosstalk between microglia and Tregs is a key determinant of neuronal regeneration and the repair of synaptic plasticity after brain injury (97).

On one hand, GLP-1RA could reduce the proliferation of effector T cells, promote the generation and function of Treg cells. Treg cells could not only suppress immune inflammation, but also promote neurogenesis and neural repair (77). Treg cells mainly coordinated the infiltration of lymphocytes and microglia in ischemic brain tissue through IL-10 signaling to prevent secondary brain injury (78, 79). Treg cells not only interacted with microglia (75), but also regulated the polarization transformation of microglia from M1 to M2 type (28, 80, 81). On the other hand, in vitro experiments demonstrated that GLP-1RA morroniside could act directly on primary microglia, and promote the mRNA levels of the M2 microglial cell markers Agr1, C206, IL-4, and IL-10. This meant that GLP-1RA could not only indirectly mediate M2 polarization of microglia by regulating Tregs, but also directly bind to GLP-1R on the membrane surface of microglia, completing M2 polarization through the GLP-1R/cAMP/PKA/IL-10 signaling pathway (28).

Recent research showed that GLP-1RA could reduce plasma TNF- $\alpha$  levels induced by different Toll-like receptor agonists and significantly inhibit neuroinflammation (98). There was no difference in the anti-inflammatory effect of GLP-1RA between  $Glp1r^{\text{Tie2+/+}}$  and  $Glp1r^{\text{Tie2-/-}}$  mice, indicating that this activity was not dependent on the blood system or endothelial cells. In contrast, the anti-inflammatory effect of GLP-1RA was significantly different between  $Glp1r^{\text{Wnt1+/+}}$  and  $Glp1r^{\text{Wnt1-/-}}$ ,  $Glp1r^{\text{Nes+/+}}$  and  $Glp1r^{\text{Nes-/-}}$ . Wnt1 and Nes were used to label neural crest derived cells and neuroendocrine cells, indicating that this activity required the involvement of central nervous system GLP-1R. These studies suggest that GLP-1RA may inhibit TCR signaling in a GLP-1R-dependent manner, thereby reducing systemic and intestinal inflammation induced by CD8<sup>+</sup> T cells and other factors (anti-CD3 induction) (99).

In summary, GLP-1RAs have the potential to inhibit neuroinflammation and promote nerve regeneration through the "Tregs-microglia-neuron/NPCs" axis: (1) GLP-1RAs directly and indirectly inhibit the activation of microglia, promote their M2 polarization, and secrete cytokines such as IL-4, IL-10, TGF -  $\beta$ , IL-2 to exert anti-inflammatory effects (100); (2) GLP-1RAs can also induce Tregs and microglia to secrete IL-10, which further promote microglia to secrete chemokines (Ccl2, Ccl7, and Cxcl10) and IGF-1. After stroke, NPCs have the ability to migrate to the lesion site, and chemokines and IGF-1 can promote NPCs proliferation, migration, and differentiation (101, 102). Depletion of CD25specific antibodies in Treg leads to a decrease in the number of NPCs after experimental stroke (77). Similarly, the increase in the number of Treg cells and the secretion of the key cytokine IL-10 in the lateral ventricle of the ischemic hemisphere are also positively correlated with the increase in NPCs proliferation (103).

### 6 Perspective

Although multiple cytokines (IL-4, IL-10, etc.) together inhibited neuroinflammation in neurological disorders, their functions were significantly different. M2 microglia could be divided into several subtypes such as IL-4-induced M2a and IL-10-induced M2c. The synergistic induction of IL-4 and IL-10 could enhance the expression of M2a-related genes, produce a large amount of CCL24 (eotaxin-2) and promote eosinophil migration (104). In clinical practice focusing on prognostic indicators for ischemic stroke patients found that there was a close negative correlation between IL-10 and NIHSS scores (P=0.0006), but not between IL-4 and NIHSS scores (P=0.088) (105). Comparing the prognosis of ischemic stroke patients between survivors and non survivors, the IL-10 level in the survivors group was significantly higher than that in the non survivors group (P=0.006) (106). Although there is much preclinical evidence as mentioned above, few clinical trials was reported to evaluate the efficacy of GLP-1RAs in the treatment of ischemic stroke or diabetes complicated by ischemic stroke. A letter reported the clinical manifestations of the GLP-1RA exenatide in acute stroke patients for the first time, including treatment regimens and adverse reactions (107). The results showed that 11 patients experienced mild nausea (n=6) and vomiting (n=5), with no serious adverse reactions or deaths. Given the high incidence of vomiting, the author suggested prophylactic administration of antiemetics. The combination of exenatide and antiemetic agents had good safety and tolerability, with no adverse effects on neurological function and prognosis. In addition, we also found that the clinical rationale and protocol design of the TEXAIS (Treatment With Exenatide in Acute Ischemic Stroke) trial (Trial registration: ClinicalTrials.gov/ANZCTR NTA1127 and ACTRN12614001189617) (108, 109). The latest results from the 2023 TEXAIS trial indicated that the use of exenatide (5 µg, twice daily injections for 5 days) did not significantly reduce neurological damage within 7 days in patients with acute ischemic stroke (trial registration numbers: ACTRN12617000409370 and NCT03287076). However, in the exenatide treatment group, the primary outcome rate was 61.2% (n=170), compared to 56.7% in the standard treatment group (n=171), with an adjusted ratio of 1.22 [95% CI, 0.79-1.88] (P=0.38), which meant that the proportion of subjects who achieved the desired effect in exenatide treatment group was higher (110). GLP-1RAs may be an optimal option for ischemic stroke treatment.

It was noted that no clinical trials had yet been completed on AD, PD, or other neurodegenerative disorders. Thus clinical data on GLP-1 agonists in brain-related conditions remain limited. However, in a double-blinded trial recruiting 54 participants with type 2 diabetes were randomized to liraglutide (1.8 mg/day) or placebo for 26 weeks (111). The mRNA expressions of tumor necrosis factor- $\alpha$  (p = 0.004) and interleukin-1 $\beta$  in peripheral blood mononuclear cells were downregulated (p = 0.046) in the liraglutide-treated group. The clinical evidence of GLP-1RAs in anti-inflammatory effect was also proved in semaglutide, exenatide and others (112–115). Chronic neuroinflammation is considered an important factor in cognitive and memory impairment in neurodegenerative diseases. It was

reported that exendin-4 prohibited amyloid- $\beta$ -induced microglial activation, thereby limiting neuroinflammation, reducing the levels of TNF- $\alpha$ , C1q and IL-1 $\alpha$ ; and improving recognition (116). Similar activities of other GLP-1 mimetics were also demonstrated in a rodent PD model (117). Another study also indicated that exendin-4 improved memory impairment by dampening the AMPK/NF- $\kappa$ B pathway, reducing levels of IL-1 $\beta$  and TNF- $\alpha$ , and increasing synaptic protein levels (40). Besides, liraglutide could also improve rat memory and cognition function by reduced neuronal apoptosis, tau phosphorylation, and  $\beta$ -site APP cleaving enzyme 1 levels (118). For a long time, scientists believe that chronic neuroinflammation is crucial in cognitive impairment, and that GLP-1RAs protect synaptic and learning functions from neuroinflammation (119, 120).

A genome-wide association study examining common genetic features of AD had identified neuroinflammatory pathways (mainly associated with TNF) to be a key feature of AD, and a driver of risk for these chronic neurodegenerative impairment (121). Plenty of nonsteroidal anti-inflammatory drugs (NSAIDs) were recruited in many clinical trials for AD treatment (122). NSAIDs primarily reduce inflammation by inhibiting cyclooxygenase enzymes (COX-1 and COX-2), which are responsible for the production of prostaglandins, which are involved in the inflammatory response, including the promotion of pain, fever, and tissue damage. Therefore, the antiinflammatory effect of NSAIDs is mainly applied to peripheral tissues and for short-term inflammation with the limited CNS penetration. Besides, both GLP-1RAs and SGLT2 inhibitors showed the obvious inhibition on neuroinflammation (123, 124). SGLT2 inhibitors could block the reabsorption of glucose in the kidneys, leading to increased glucose excretion and improved glycemic control in type 2 diabetes. The new preclinical evidence also suggested that they might have neuroprotective properties through reducing oxidative stress, modulating inflammatory pathways, and enhancing mitochondrial function (125, 126). However, their effects in humans with brain disorders are still under investigation.

This review was mainly summarized on the basis of research data, and therefore had limitations in many aspects: (1) Although we already knew that GLP-1RA inhibited the activation of microglia and promoted their polarization towards the M2 type, the regulatory mechanism of their induction of polarization in naive CD4<sup>+</sup> T cells was still unknown, in particular their impact on the proportion of Tregs subtypes in T cells at the sites of infarct lesion. (2) The interaction between microglia and T cells may be bidirectional, as microglia express various molecules involved in antigen presentation and T cell regulation, such as MHC II, CD11c, Dectin-1, etc (127). Do GLP-1RA have a regulatory effect on microglial antigen presentation and microglial-T cell balance? (3) Clinical evidence for GLP-1RA in stroke is still lacking. The latest TEXAIS results in 2023 were negative with a certain trend, which might be limited by the small sample size, short treatment duration and lack of medium to longterm therapeutic evaluation. Current research on GLP-1RA in ischemic stroke should be expanded to include large-scale, long-term follow-up clinical trials to further assess its efficacy and safety. Additionally, evaluating the long-term use of GLP-1RA, particularly across different populations, could provide valuable insights into its therapeutic potential and risk profile, which will contribute to developing more effective treatment strategies and improving the prognosis and quality of life for patients with ischemic stroke and diabetes (128, 129).

Finally, this study elucidated the immunosuppressive and neuroregenerative functions of GLP-1RA based on the "Tregsmicroglia-neuron/NPC" axis theory. Given the advantages of small-molecule GLP-1RA, such as rapid diffusion, good BBB permeability and multiple modes of administration, small molecule compounds will be one of the important development trends of GLP-1RA. In the future, we look forward to the clinical research evidence of small-molecule GLP-1RA intervening in ischemic stroke or T2D complicated by ischemic stroke.

### Author contributions

H-HS: Conceptualization, Data curation, Investigation, Methodology, Software, Supervision, Writing – original draft, Writing – review & editing. YH: Data curation, Formal analysis, Methodology, Project administration, Supervision, Validation, Visualization, Writing – original draft. FG: Formal analysis, Funding acquisition, Project administration, Resources, Validation, Visualization, Writing – original draft. HL: Software, Supervision, Validation, Visualization, Writing – original draft, Writing – review & editing.

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### References

1. Drucker DJ. GLP-1 physiology informs the pharmacotherapy of obesity. *Mol Metab.* (2022) 57:101351. doi: 10.1016/j.molmet.2021.101351

2. Jensen VFH, Molck AM, Nowak J, Fels JJ, Lykkesfeldt J, Bogh IB. Prolonged insulin-induced hypoglycaemia reduces ss-cell activity rather than number in pancreatic islets in non-diabetic rats. *Sci Rep.* (2022) 12:14113. doi: 10.1038/s41598-022-18398-z

3. Müller TD, Finan B, Bloom SR, D'Alessio D, Drucker DJ, Flatt PR, et al. Glucagonlike peptide 1 (GLP-1). *Mol Metab.* (2019) 30:72–130. doi: 10.1016/j.molmet.2019.09.010

4. Salcedo I, Tweedie D, Li Y, Greig NH. Neuroprotective and neurotrophic actions of glucagon-like peptide-1: an emerging opportunity to treat neurodegenerative and cerebrovascular disorders. *Br J Pharmacol.* (2012) 166:1586–99. doi: 10.1111/j.1476-5381.2012.01971.x

5. Tramutola A, Arena A, Cini C, Butterfield DA, Barone E. Modulation of GLP-1 signaling as a novel therapeutic approach in the treatment of Alzheimer's disease pathology. *Expert Rev Neurother*. (2017) 17:59-75. doi: 10.1080/14737175.2017.1246183

6. Cai HY, Holscher C, Yue XH, Zhang SX, Wang XH, Qiao F, et al. Lixisenatide rescues spatial memory and synaptic plasticity from amyloid beta protein-induced impairments in rats. *Neuroscience*. (2014) 277:6-13. doi: 10.1016/j.neuroscience.2014.02.022

7. Han WN, Holscher C, Yuan L, Yang W, Wang XH, Wu MN, et al. Liraglutide protects against amyloid-beta protein-induced impairment of spatial learning and memory in rats. *Neurobiol Aging*. (2013) 34:576-88. doi: 10.1016/j.neurobiolaging.2012.04.009

8. Singh NK, Singh A, Varshney M, Agrawal R. A research update on exendin-4 as a novel molecule against parkinson's disease. *Curr Mol Med.* (2023) 23:889–900. doi: 10.2174/1566524023666230529093314

9. Athauda D, Maclagan K, Skene SS, Bajwa-Joseph M, Letchford D, Chowdhury K, et al. Exenatide once weekly versus placebo in Parkinson's disease: a randomised, double-blind, placebo-controlled trial. *Lancet*. (2017) 390:1664–75. doi: 10.1016/S0140-6736(17)31585-4

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### **Conflict of interest**

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10. Egefjord L, Gejl M, Møller A, Brændgaard H, Gottrup H, Antropova O, et al. Effects of liraglutide on neurodegeneration, blood flow and cognition in Alzheimer s disease - protocol for a controlled, randomized double-blinded trial. *Dan Med J.* (2012) 59:A4519.

11. Gejl M, Gjedde A, Egefjord L, Møller A, Hansen SB, Vang K, et al. In alzheimer's disease, 6-month treatment with GLP-1 analog prevents decline of brain glucose metabolism: randomized, placebo-controlled, double-blind clinical trial. *Front Aging Neurosci.* (2016) 8:108. doi: 10.3389/fnagi.2016.00108

12. Mansur RB, Ahmed J, Cha DS, Woldeyohannes HO, Subramaniapillai M, Lovshin J, et al. Liraglutide promotes improvements in objective measures of cognitive dysfunction in individuals with mood disorders: A pilot, open-label study. *J Affect Disord.* (2017) 207:114–20. doi: 10.1016/j.jad.2016.09.056

13. Kuchay MS, Krishan S, Mishra SK, Choudhary NS, Singh MK, Wasir JS, et al. Effect of dulaglutide on liver fat in patients with type 2 diabetes and NAFLD: randomised controlled trial (D-LIFT trial). *Diabetologia*. (2020) 63:2434–45. doi: 10.1007/s00125-020-05265-7

14. Nauck MA, Quast DR, Wefers J, Meier JJ. GLP-1 receptor agonists in the treatment of type 2 diabetes - state-of-the-art. *Mol Metab.* (2021) 46:101102. doi: 10.1016/j.molmet.2020.101102

15. Tu XK, Chen Q, Chen S, Huang B, Ren BG, Shi SS. GLP-1R agonist liraglutide attenuates inflammatory reaction and neuronal apoptosis and reduces early brain injury after subarachnoid hemorrhage in rats. *Inflammation*. (2021) 44:397–406. doi: 10.1007/s10753-020-01344-4

16. Liu C, Sun S, Xie J, Li H, Li T, Wu Q, et al. GLP-1R Agonist Exendin-4 Protects Against Hemorrhagic Transformation Induced by rtPA After Ischemic Stroke via the Wnt/beta-Catenin Signaling Pathway. *Mol Neurobiol.* (2022) 59:3649–64. doi: 10.1007/s12035-022-02811-9

17. Fan H, Li TF, Gong N, Wang YX. Shanzhiside methylester, the principle effective iridoid glycoside from the analgesic herb Lamiophlomis rotata, reduces neuropathic pain by stimulating spinal microglial  $\beta$ -endorphin expression. *Neuropharmacology*. (2016) 101:98–109. doi: 10.1016/j.neuropharm.2015.09.010

 Gong N, Fan H, Ma AN, Xiao Q, Wang YX. Geniposide and its iridoid analogs exhibit antinociception by acting at the spinal GLP-1 receptors. *Neuropharmacology*. (2014) 84:31–45. doi: 10.1016/j.neuropharm.2014.04.007

19. Xu M, Wu HY, Liu H, Gong N, Wang YR, Wang YX. Morroniside, a secoiridoid glycoside from Cornus officinalis, attenuates neuropathic pain by activation of spinal glucagon-like peptide-1 receptors. *Br J Pharmacol.* (2017) 174:580–90. doi: 10.1111/ bph.v174.7

20. Zhu B, Gong N, Fan H, Peng CS, Ding XJ, Jiang Y, et al. Lamiophlomis rotata, an orally available Tibetan herbal painkiller, specifically reduces pain hypersensitivity states through the activation of spinal glucagon-like peptide-1 receptors. *Anesthesiology*. (2014) 121:835–51. doi: 10.1097/ALN.00000000000320

21. Jia Y, Gong N, Li TF, Zhu B, Wang YX. Peptidic exenatide and herbal catalpol mediate neuroprotection via the hippocampal GLP-1 receptor/beta-endorphin pathway. *Pharmacol Res.* (2015) 102:276–85. doi: 10.1016/j.phrs.2015.10.008

22. Mahapatra MK, Karuppasamy M, Sahoo BM. Therapeutic potential of semaglutide, a newer GLP-1 receptor agonist, in abating obesity, non-alcoholic steatohepatitis and neurodegenerative diseases: A narrative review. *Pharm Res.* (2022) 39:1233–48. doi: 10.1007/s11095-022-03302-1

23. Nowell J, Blunt E, Gupta D, Edison P. Antidiabetic agents as a novel treatment for Alzheimer's and Parkinson's disease. *Ageing Res Rev.* (2023) 89:101979. doi: 10.1016/j.arr.2023.101979

24. Rubin R. Could GLP-1 receptor agonists like semaglutide treat addiction, alzheimer disease, and other conditions? JAMA. (2024) 331:1519-21.

25. Victorino DB, Nejm M, Guimaraes-Marques M, Scorza FA, Scorza CA. Repurposing GLP-1 receptor agonists for parkinson's disease: current evidence and future opportunities. *Pharmaceut Med.* (2021) 35:11–9. doi: 10.1007/s40290-020-00374-5

26. Monney M, Jornayvaz FR, Gariani K. GLP-1 receptor agonists effect on cognitive function in patients with and without type 2 diabetes. *Diabetes Metab.* (2023) 49:101470. doi: 10.1016/j.diabet.2023.101470

27. Pelle MC, Zaffina I, Giofre F, Pujia R, Arturi F. Potential role of glucagon-like peptide-1 receptor agonists in the treatment of cognitive decline and dementia in diabetes mellitus. *Int J Mol Sci.* (2023) 24. doi: 10.3390/ijms241411301

28. Liu H, Ou MX, Han QQ. Microglial M2 polarization mediated the neuroprotective effect of morroniside in transient MCAO-induced mice. *Front Pharmacol.* (2021) 12:784329. doi: 10.3389/fphar.2021.784329

29. Tang X, Wu H, Mao X, Li X, Wang Y. The GLP-1 receptor herbal agonist morroniside attenuates neuropathic pain via spinal microglial expression of IL-10 and beta-endorphin. *Biochem Biophys Res Commun.* (2020) 530:494–9. doi: 10.1016/j.bbrc.2020.05.080

30. Ma L, Ju P, Wang W, Wei J, Wang W, Zhao M, et al. Microglial activation of GLP-1R signaling in neuropathic pain promotes gene expression adaption involved in inflammatory responses. *Neural Plast.* (2021) 2021:9923537. doi: 10.1155/2021/9923537

31. Han QQ, Yin M, Wang ZY, Liu H, Ao JP, Wang YX. Cynandione A alleviates neuropathic pain through alpha7-nAChR-dependent IL-10/beta-endorphin signaling complexes. *Front Pharmacol.* (2020) 11:614450.

32. Ma L, Peng S, Wei J, Zhao M, Ahmad KA, Chen J, et al. Spinal microglial betaendorphin signaling mediates IL-10 and exenatide-induced inhibition of synaptic plasticity in neuropathic pain. *CNS Neurosci Ther.* (2021) 27:1157–72. doi: 10.1111/ cns.13694

33. Wang ZY, Han QQ, Deng MY, Zhao MJ, Apryani E, Shoaib RM, et al. Lemairamin, isolated from the Zanthoxylum plants, alleviates pain hypersensitivity via spinal alpha7 nicotinic acetylcholine receptors. *Biochem Biophys Res Commun.* (2020) 525:1087–94. doi: 10.1016/j.bbrc.2020.03.023

34. Huang J, Liu Y, Cheng L, Li J, Zhang T, Zhao G, et al. Glucagon-like peptide-1 cleavage product GLP-1(9-36) reduces neuroinflammation from stroke via the activation of insulin-like growth factor 1 receptor in astrocytes. *Eur J Pharmacol.* (2020) 887:173581. doi: 10.1016/j.ejphar.2020.173581

35. Tu XK, Chen PP, Chen JY, Ding YH, Chen Q, Shi SS. GLP-1R knockdown abrogates the protective effects of liraglutide on ischaemic stroke via inhibition of M2 polarisation and activation of NLRP3 inflammasome by reducing Nrf2 activation. *Neuropharmacology.* (2023) 237:109603. doi: 10.1016/j.neuropharm.2023.109603

36. Zhang H, Liu Y, Cheng L, Ma X, Luo X. Exendin-4 induces a novel extended effect of ischemic tolerance via crosstalk with IGF-1R. *Brain Res Bull.* (2021) 169:145–55. doi: 10.1016/j.brainresbull.2020.11.008

37. Shan Y, Tan S, Lin Y, Liao S, Zhang B, Chen X, et al. The glucagon-like peptide-1 receptor agonist reduces inflammation and blood-brain barrier breakdown in an astrocyte-dependent manner in experimental stroke. *J Neuroinflamm*. (2019) 16:242. doi: 10.1186/s12974-019-1638-6

38. Nizari S, Basalay M, Chapman P, Korte N, Korsak A, Christie IN, et al. Glucagon-like peptide-1 (GLP-1) receptor activation dilates cerebral arterioles, increases cerebral blood flow, and mediates remote (pre)conditioning neuroprotection against ischaemic stroke. *Basic Res Cardiol.* (2021) 116:32. doi: 10.1007/s00395-021-00873-9

39. Yang X, Qiang Q, Li N, Feng P, Wei W, Holscher C. Neuroprotective mechanisms of glucagon-like peptide-1-based therapies in ischemic stroke: an update based on preclinical research. *Front Neurol.* (2022) 13:844697. doi: 10.3389/fneur.2022.844697

40. Zhang LQ, Zhang W, Li T, Yang T, Yuan X, Zhou Y, et al. GLP-1R activation ameliorated novel-object recognition memory dysfunction via regulating hippocampal AMPK/NF-kappaB pathway in neuropathic pain mice. *Neurobiol Learn Mem.* (2021) 182:107463. doi: 10.1016/j.nlm.2021.107463

41. Zhang S. Microglial activation after ischaemic stroke. *Stroke Vasc Neurol.* (2019) 4:71-4. doi: 10.1136/svn-2018-000196

42. Dougherty JA, Guirguis E, Thornby KA. A systematic review of newer antidiabetic agents in the treatment of nonalcoholic fatty liver disease. *Ann Pharmacother*. (2021) 55:65–79. doi: 10.1177/1060028020935105

43. Fei Y, Tsoi MF, Cheung BMY. Cardiovascular outcomes in trials of new antidiabetic drug classes: a network meta-analysis. *Cardiovasc Diabetol.* (2019) 18:112. doi: 10.1186/s12933-019-0916-z

44. Lo CWH, Fei Y, Cheung BMY. Cardiovascular outcomes in trials of new antidiabetic drug classes. *Card Fail Rev.* (2021) 7:e04. doi: 10.15420/cfr

45. Xie Y, Bowe B, Gibson AK, McGill JB, Maddukuri G, Yan Y, et al. Comparative effectiveness of SGLT2 inhibitors, GLP-1 receptor agonists, DPP-4 inhibitors, and sulfonylureas on risk of kidney outcomes: emulation of a target trial using health care databases. *Diabetes Care.* (2020) 43:2859–69. doi: 10.2337/dc20-1890

46. Donnan GA, Fisher M, Macleod M, Davis SM. Stroke. Lancet. (2008) 371:1612– 23. doi: 10.1016/S0140-6736(08)60694-7

47. Grysiewicz RA, Thomas K, Pandey DK. Epidemiology of ischemic and hemorrhagic stroke: incidence, prevalence, mortality, and risk factors. *Neurol Clin.* (2008) 26:871-895, vii. doi: 10.1016/j.ncl.2008.07.003

48. Hori M, Connolly SJ, Zhu J, Liu LS, Lau CP, Pais P, et al. Dabigatran versus warfarin: effects on ischemic and hemorrhagic strokes and bleeding in Asians and non-Asians with atrial fibrillation. *Stroke*. (2013) 44:1891-6. doi: 10.1161/STROKEAHA.113.000990

49. Zhang L, Guo Z, Wu M, Hu X, Xu Y, Zhou Z. Interaction of smoking and metabolic syndrome on cardiovascular risk in a Chinese cohort. *Int J Cardiol.* (2013) 167:250–3. doi: 10.1016/j.ijcard.2011.12.079

50. Klatsky AL, Friedman GD, Sidney S, Kipp H, Kubo A, Armstrong MA. Risk of hemorrhagic stroke in Asian American ethnic groups. *Neuroepidemiology*. (2005) 25:26–31. doi: 10.1159/000085310

51. Venketasubramanian N, Yoon BW, Pandian J, Navarro JC. Stroke epidemiology in south, east, and south-east asia: A review. *J Stroke*. (2017) 19:286–94. doi: 10.5853/ jos.2017.00234

52. Ma Q, Li R, Wang L, Yin P, Wang Y, Yan C, et al. Temporal trend and attributable risk factors of stroke burden in China 1990-2019: an analysis for the global burden of disease study 2019. *Lancet Public Health*. (2021) 6:e897–906. doi: 10.1016/S2468-2667(21)00228-0

53. Zhao Y, Hua X, Ren X, Ouyang M, Chen C, Li Y, et al. Increasing burden of stroke in China: A systematic review and meta-analysis of prevalence, incidence, mortality, and case fatality. *Int J Stroke*. (2023) 18:259–67. doi: 10.1177/17474930221135983

54. Yao M, Ren Y, Jia Y, Xu J, Wang Y, Zou K, et al. Projected burden of stroke in China through 2050. *Chin Med J (Engl)*. (2023) 136:1598–605. doi: 10.1097/CM9.00000000002060

55. Brown E, Heerspink HJL, Cuthbertson DJ, Wilding JPH. SGLT2 inhibitors and GLP-1 receptor agonists: established and emerging indications. *Lancet.* (2021) 398:262–76. doi: 10.1016/S0140-6736(21)00536-5

56. Sposito AC, Berwanger O, de Carvalho LSF, Saraiva JFK. GLP-1RAs in type 2 diabetes: mechanisms that underlie cardiovascular effects and overview of cardiovascular outcome data. *Cardiovasc Diabetol.* (2018) 17:157. doi: 10.1186/s12933-018-0800-2

57. Tsapas A, Avgerinos I, Karagiannis T, Malandris K, Manolopoulos A, Andreadis P, et al. Comparative effectiveness of glucose-lowering drugs for type 2 diabetes: A systematic review and network meta-analysis. *Ann Intern Med.* (2020) 173:278–86. doi: 10.7326/M20-0864

58. Verma S, Bain SC, Honore JB, Fem J, An M, Ep R, et al. Impact of microvascular disease on cardiovascular outcomes in type 2 diabetes: Results from the LEADER and SUSTAIN 6 clinical trials. *Diabetes Obes Metab.* (2020) 22:2193–8. doi: 10.1111/dom.v22.11

59. Meeker RB, Williams K, Killebrew DA, Hudson LC. Cell trafficking through the choroid plexus. *Cell Adh Migr.* (2012) 6:390–6. doi: 10.4161/cam.21054

60. Cramer JV, Benakis C, Liesz A. T cells in the post-ischemic brain: Troopers or paramedics? *J Neuroimmunol.* (2019) 326:33–7.

61. Iadecola C, Anrather J. The immunology of stroke: from mechanisms to translation. *Nat Med.* (2011) 17:796–808. doi: 10.1038/nm.2399

62. Kuric E, Ruscher K. Dynamics of major histocompatibility complex class IIpositive cells in the postischemic brain-influence of levodopa treatment. J Neuroinflamm. (2014) 11:145. doi: 10.1186/s12974-014-0145-z

63. Ritzel RM, Patel AR, Grenier JM, Crapser J, Verma R, Jellison ER, et al. Functional differences between microglia and monocytes after ischemic stroke. J Neuroinflamm. (2015) 12:106. doi: 10.1186/s12974-015-0329-1

64. Pekna M, Siqin S, de Pablo Y, Stokowska A, Torinsson Naluai Å, Pekny M. Astrocyte responses to complement peptide C3a are highly context-dependent. *Neurochem Res.* (2023) 48:1233–41. doi: 10.1007/s11064-022-03743-5

65. Lei TY, Ye YZ, Zhu XQ, Smerin D, Gu LJ, Xiong XX, et al. The immune response of T cells and therapeutic targets related to regulating the levels of T helper cells after ischaemic stroke. *J Neuroinflamm.* (2021) 18:25. doi: 10.1186/s12974-020-02057-z

66. Chu HX, Kim HA, Lee S, Moore JP, Chan CT, Vinh A, et al. Immune cell infiltration in Malignant middle cerebral artery infarction: comparison with transient cerebral ischemia. *J Cereb. Blood Flow Metab.* (2014) 34:450–9. doi: 10.1038/ jcbfm.2013.217

67. Felger JC, Abe T, Kaunzner UW, Gottfried-Blackmore A, Gal-Toth J, McEwen BS, et al. Brain dendritic cells in ischemic stroke: time course, activation state, and origin. *Brain Behav Immun.* (2010) 24:724–37. doi: 10.1016/j.bbi.2009.11.002

68. Hu X, Li P, Guo Y, Wang H, Leak RK, Chen S, et al. Microglia/macrophage polarization dynamics reveal novel mechanism of injury expansion after focal cerebral ischemia. *Stroke*. (2012) 43:3063–70. doi: 10.1161/STROKEAHA.112.659656

69. Jickling GC, Liu D, Ander BP, Stamova B, Zhan X, Sharp FR. Targeting neutrophils in ischemic stroke: translational insights from experimental studies. J Cereb. Blood Flow Metab. (2015) 35:888–901. doi: 10.1038/jcbfm.2015.45

70. Gelderblom M, Leypoldt F, Steinbach K, Behrens D, Choe CU, Siler DA, et al. Temporal and spatial dynamics of cerebral immune cell accumulation in stroke. *Stroke.* (2009) 40:1849–57. doi: 10.1161/STROKEAHA.108.534503

71. Stubbe T, Ebner F, Richter D, Engel O, Klehmet J, Royl G, et al. Regulatory T cells accumulate and proliferate in the ischemic hemisphere for up to 30 days after MCAO. *J Cereb. Blood Flow Metab.* (2013) 33:37–47. doi: 10.1038/jcbfm.2012.128

72. Vindegaard N, Munoz-Briones C, El Ali HH, Kristensen LK, Rasmussen RS, Johansen FF, et al. T-cells and macrophages peak weeks after experimental stroke: Spatial and temporal characteristics. *Neuropathology*. (2017) 37:407–14. doi: 10.1111/ neup.2017.37.issue-5

73. Xie L, Li W, Hersh J, Liu R, Yang SH. Experimental ischemic stroke induces long-term T cell activation in the brain. *J Cereb. Blood Flow Metab.* (2019) 39:2268–76. doi: 10.1177/0271678X18792372

74. Feng Y, He X, Luo S, Chen X, Long S, Liang F, et al. Chronic colitis induces meninges traffic of gut-derived T cells, unbalances M1 and M2 microglia/macrophage and increases ischemic brain injury in mice. *Brain Res.* (2019) 1707:8–17. doi: 10.1016/ j.brainres.2018.11.019

75. Wang S, Zhang H, Xu Y. Crosstalk between microglia and T cells contributes to brain damage and recovery after ischemic stroke. *Neurol Res.* (2016) 38:495–503. doi: 10.1080/01616412.2016.1188473

76. Liesz A, Zhou W, Mracskó É, Karcher S, Bauer H, Schwarting S, et al. Inhibition of lymphocyte trafficking shields the brain against deleterious neuroinflammation after stroke. *Brain.* (2011) 134:704–20. doi: 10.1093/brain/awr008

77. Saino O, Taguchi A, Nakagomi T, Nakano-Doi A, Kashiwamura S, Doe N, et al. Immunodeficiency reduces neural stem/progenitor cell apoptosis and enhances neurogenesis in the cerebral cortex after stroke. *J Neurosci Res.* (2010) 88:2385–97. doi: 10.1002/jnr.v88:11

78. Liesz A, Suri-Payer E, Veltkamp C, Doerr H, Sommer C, Rivest S, et al. Regulatory T cells are key cerebroprotective immunomodulators in acute experimental stroke. *Nat Med.* (2009) 15:192–9. doi: 10.1038/nm.1927

79. Saenz B, Perez HJ, Chavarria A. Regulatory T cells in central nervous system: in health and disease. *Cent Nerv Syst Agents Med Chem.* (2010) 10:326–36. doi: 10.2174/187152410793429674

80. Liu H, Zhang JT, Mou CY, Hao Y, Cui W. Involvement of DAAO overexpression in delayed hippocampal neuronal death. *Cells.* (2022) 11.

81. Liu H, Zhao MJ, Wang ZY, Han QQ, Wu HY, Mao XF, et al. Involvement of damino acid oxidase in cerebral ischaemia induced by transient occlusion of the middle cerebral artery in mice. *Br J Pharmacol.* (2019) 176:3336–49. doi: 10.1111/bph.v176.17

82. Li P, Mao L, Zhou G, Leak RK, Sun BL, Chen J, et al. Adoptive regulatory T-cell therapy preserves systemic immune homeostasis after cerebral ischemia. *Stroke*. (2013) 44:3509–15. doi: 10.1161/STROKEAHA.113.002637

83. Butovsky O, Ziv Y, Schwartz A, Landa G, Talpalar AE, Pluchino S, et al. Microglia activated by IL-4 or IFN-gamma differentially induce neurogenesis and oligodendrogenesis from adult stem/progenitor cells. *Mol Cell Neurosci.* (2006) 31:149–60. doi: 10.1016/j.mcn.2005.10.006

84. Sanchez-Molina P, Almolda B, Gimenez-Llort L, Gonzalez B, Castellano B. Chronic IL-10 overproduction disrupts microglia-neuron dialogue similar to aging, resulting in impaired hippocampal neurogenesis and spatial memory. *Brain Behav Immun.* (2022) 101:231–45. doi: 10.1016/j.bbi.2021.12.026

85. Ben Nasr M, Usuelli V, Dellepiane S, Seelam AJ, Fiorentino TV, D'Addio F, et al. Glucagon-like peptide 1 receptor is a T cell-negative costimulatory molecule. *Cell Metab.* (2024) 36:1302–1319 e1312. doi: 10.1016/j.cmet.2024.05.001

86. Hadjiyanni I, Siminovitch KA, Danska JS, Drucker DJ. Glucagon-like peptide-1 receptor signalling selectively regulates murine lymphocyte proliferation and maintenance of peripheral regulatory T cells. *Diabetologia*. (2010) 53:730–40. doi: 10.1007/s00125-009-1643-x

87. Xu Q, Zhang X, Li T, Shao S. Exenatide regulates Th17/Treg balance via PI3K/ Akt/FoxO1 pathway in db/db mice. *Mol Med.* (2022) 28:144. doi: 10.1186/s10020-022-00574-6

88. Moschovaki Filippidou F, Kirsch AH, Thelen M, Kétszeri M, Artinger K, Aringer I, et al. Glucagon-like peptide-1 receptor agonism improves nephrotoxic serum

nephritis by inhibiting T-cell proliferation. Am J Pathol. (2020) 190:400–11. doi: 10.1016/j.ajpath.2019.10.008

89. Sha S, Liu X, Zhao R, Qing L, He Q, Sun L, et al. Effects of glucagon-like peptide-1 analog liraglutide on the systemic inflammation in high-fat-diet-induced mice. *Endocrine*. (2019) 66:494–502. doi: 10.1007/s12020-019-02081-x

90. Gharagozloo M, Smith MD, Sotirchos ES, Jin J, Meyers K, Taylor M, et al. Therapeutic potential of a novel glucagon-like peptide-1 receptor agonist, NLY01, in experimental autoimmune encephalomyelitis. *Neurotherapeutics*. (2021) 18:1834–48. doi: 10.1007/s13311-021-01088-5

91. da Silva EM, Yariwake VY, Alves RW, de Araujo DR, Andrade-Oliveira V. Crosstalk between incretin hormones, Th17 and Treg cells in inflammatory diseases. *Peptides.* (2022) 155:170834. doi: 10.1016/j.peptides.2022.170834

 Beuker C, Strecker JK, Rawal R, Schmidt-Pogoda A, Ruck T, Wiendl H, et al. Immune cell infiltration into the brain after ischemic stroke in humans compared to mice and rats: a systematic review and meta-analysis. *Transl Stroke Res.* (2021) 12:976– 90. doi: 10.1007/s12975-021-00887-4

93. Rode AKO, Buus TB, Mraz V, Al-Jaberi FAH, Lopez DV, Ford SL, et al. Induced human regulatory T cells express the glucagon-like peptide-1 receptor. *Cells*. (2022) 11. doi: 10.3390/cells11162587

94. Wang G, Yan Y, Xu N, Yin D, Hui Y. Treatment of type 1 diabetes by regulatory T-cell infusion via regulating the expression of inflammatory cytokines. *J Cell Biochem.* (2019) 120:19338–44. doi: 10.1002/jcb.v120.12

95. Benakis C, Simats A, Tritschler S, Heindl S, Besson-Girard S, Llovera G, et al. T cells modulate the microglial response to brain ischemia. *Elife*. (2022) 11. doi: 10.7554/ eLife.82031.sa2

96. Llovera G, Hofmann K, Roth S, Salas-Pérdomo A, Ferrer-Ferrer M, Perego C, et al. Results of a preclinical randomized controlled multicenter trial (pRCT): Anti-CD49d treatment for acute brain ischemia. *Sci Transl Med.* (2015) 7:299ra121. doi: 10.1126/scitranslmed.aaa9853

97. Shi L, Sun Z, Su W, Xu F, Xie D, Zhang Q, et al. Treg cell-derived osteopontin promotes microglia-mediated white matter repair after ischemic stroke. *Immunity*. (2021) 54:1527–1542 e1528. doi: 10.1016/j.immuni.2021.04.022

98. Wong CK, McLean BA, Baggio LL, Koehler JA, Hammoud R, Rittig N, et al. Central glucagon-like peptide 1 receptor activation inhibits Toll-like receptor agonist-induced inflammation. *Cell Metab.* (2024) 36:130–143 e135. doi: 10.1016/j.cmet.2023.11.009

99. Wong CK, Yusta B, Koehler JA, Baggio LL, McLean BA, Matthews D, et al. Divergent roles for the gut intraepithelial lymphocyte GLP-1R in control of metabolism, microbiota, and T cell-induced inflammation. *Cell Metab.* (2022) 34:1514–1531 e1517. doi: 10.1016/j.cmet.2022.08.003

100. Ricci A, Liesz A. A tale of two cells: Regulatory T cell-microglia cross-talk in the ischemic brain. *Sci Transl Med.* (2023) 15:eadj0052. doi: 10.1126/scitranslmed.adj0052

101. Arvidsson A, Collin T, Kirik D, Kokaia Z, Lindvall O. Neuronal replacement from endogenous precursors in the adult brain after stroke. *Nat Med.* (2002) 8:963–70. doi: 10.1038/nm747

102. Gudi V, Skuljec J, Yildiz O, Frichert K, Skripuletz T, Moharregh-Khiabani D, et al. Spatial and temporal profiles of growth factor expression during CNS demyelination reveal the dynamics of repair priming. *PloS One.* (2011) 6:e22623. doi: 10.1371/journal.pone.0022623

103. Wang J, Xie L, Yang C, Ren C, Zhou K, Wang B, et al. Activated regulatory T cell regulates neural stem cell proliferation in the subventricular zone of normal and ischemic mouse brain through interleukin 10. *Front Cell Neurosci.* (2015) 9:361. doi: 10.3389/fncel.2015.00361

104. Makita N, Hizukuri Y, Yamashiro K, Murakawa M, Hayashi Y. IL-10 enhances the phenotype of M2 macrophages induced by IL-4 and confers the ability to increase eosinophil migration. *Int Immunol.* (2015) 27:131–41. doi: 10.1093/intimm/dxu090

105. Sun W, Wang S, Nan S. The prognostic determinant of interleukin-10 in patients with acute ischemic stroke: an analysis from the perspective of disease management. *Dis Markers*. (2021) 2021:6423244. doi: 10.1155/2021/6423244

106. Chernykh ER, Shevela EY, Starostina NM, Morozov SA, Davydova MN, Menyaeva EV, et al. Safety and therapeutic potential of M2 macrophages in stroke treatment. *Cell Transplant*. (2016) 25:1461–71. doi: 10.3727/096368915X690279

107. Daly SC, Chemmanam T, Loh PS, Gilligan A, Dear AE, Simpson RW, et al. Exenatide in acute ischemic stroke. *Int J Stroke.* (2013) 8:E44. doi: 10.1111/ijs.12073

108. McGrath RT, Hocking SL, Priglinger M, Day S, Herkes GK, Krause M, et al. Rationale and design of Short-Term EXenatide therapy in Acute ischaemic Stroke (STEXAS): a randomised, open-label, parallel-group study. *BMJ Open.* (2016) 6: e008203. doi: 10.1136/bmjopen-2015-008203

109. Muller C, Cheung NW, Dewey H, Churilov L, Middleton S, Thijs V, et al. Treatment with exenatide in acute ischemic stroke trial protocol: A prospective, randomized, open label, blinded end-point study of exenatide vs. *standard Care post stroke hyperglycemia Int J Stroke*. (2018) 13:857–62.

110. Bladin CF, Wah Cheung N, Dewey HM, Churilov L, Middleton S, Thijs V, et al. Management of poststroke hyperglycemia: results of the TEXAIS randomized clinical trial. *Stroke*. (2023) 54:2962–71. doi: 10.1161/STROKEAHA.123.044568

111. Zobel EH, Ripa RS, von Scholten BJ, Rotbain Curovic V, Kjaer A, Hansen TW, et al. Effect of liraglutide on expression of inflammatory genes in type 2 diabetes. *Sci Rep.* (2021) 11:18522. doi: 10.1038/s41598-021-97967-0

112. Alharbi SH. Anti-inflammatory role of glucagon-like peptide 1 receptor agonists and its clinical implications. *Ther Adv Endocrinol Metab.* (2024) 15:20420188231222367. doi: 10.1177/20420188231222367

113. Bray JJH, Foster-Davies H, Salem A, Hoole AL, Obaid DR, Halcox JPJ, et al. Glucagon-like peptide-1 receptor agonists improve biomarkers of inflammation and oxidative stress: A systematic review and meta-analysis of randomised controlled trials. *Diabetes Obes Metab.* (2021) 23:1806–22. doi: 10.1111/dom.14399

114. Buldak L, Machnik G, Buldak RJ, Labuzek K, Boldys A, Okopien B. Exenatide and metformin express their anti-inflammatory effects on human monocytes/ macrophages by the attenuation of MAPKs and NFkappaB signaling. *Naunyn Schmiedebergs Arch Pharmacol.* (2016) 389:1103–15.

115. Reppo I, Jakobson M, Volke V. Effects of semaglutide and empagliflozin on inflammatory markers in patients with type 2 diabetes. *Int J Mol Sci.* (2023) 24. doi: 10.3390/ijms24065714

116. Park JS, Kam TI, Lee S, Park H, Oh Y, Kwon SH, et al. Blocking microglial activation of reactive astrocytes is neuroprotective in models of Alzheimer's disease. *Acta Neuropathol Commun.* (2021) 9:78. doi: 10.1186/s40478-021-01180-z

117. Yun SP, Kam TI, Panicker N, Kim S, Oh Y, Park JS, et al. Block of A1 astrocyte conversion by microglia is neuroprotective in models of Parkinson's disease. *Nat Med.* (2018) 24:931–8. doi: 10.1038/s41591-018-0051-5

118. Yu CJ, Ma D, Song LL, Zhai ZN, Tao Y, Zhang Y, et al. The role of GLP-1/GIP receptor agonists in Alzheimer's disease. *Adv Clin Exp Med.* (2020) 29:661–8. doi: 10.17219/acem/121007

119. Chen T, Tian P, Huang Z, Zhao X, Wang H, Xia C, et al. Engineered commensal bacteria prevent systemic inflammation-induced memory impairment and amyloidogenesis via producing GLP-1. *Appl Microbiol Biotechnol.* (2018) 102:7565–75. doi: 10.1007/s00253-018-9155-6

120. Iwai T, Sawabe T, Tanimitsu K, Suzuki M, Sasaki-Hamada S, Oka J. Glucagonlike peptide-1 protects synaptic and learning functions from neuroinflammation in rodents. J Neurosci Res. (2014) 92:446–54. doi: 10.1002/jnr.23335 121. Bellenguez C, Küçükali F, Jansen IE, Kleineidam L, Moreno-Grau S, Amin N, et al. New insights into the genetic etiology of Alzheimer's disease and related dementias. *Nat Genet.* (2022) 54:412–36. doi: 10.1038/s41588-022-01024-z

122. Novoa C, Salazar P, Cisternas P, Gherardelli C, Vera-Salazar R, Zolezzi JM, et al. Inflammation context in Alzheimer's disease, a relationship intricate to define. *Biol Res.* (2022) 55:39. doi: 10.1186/s40659-022-00404-3

123. Chan YH, Chao TF, Chen SW, Lee HF, Li PR, Yeh YH, et al. SGLT2 inhibitors vs GLP-1 receptor agonists and clinical outcomes in patients with diabetes with/ without atrial fibrillation. *J Clin Endocrinol Metab.* (2024) 109:2617–29. doi: 10.1210/ clinem/dgae157

124. Moon JS, Hong JH, Jung YJ, Ferrannini E, Nauck MA, Lim S. SGLT-2 inhibitors and GLP-1 receptor agonists in metabolic dysfunction-associated fatty liver disease. *Trends Endocrinol Metab.* (2022) 33:424-42. doi: 10.1016/j.tem.2022.03.005

125. Mei J, Li Y, Niu L, Liang R, Tang M, Cai Q, et al. SGLT2 inhibitors: a novel therapy for cognitive impairment via multifaceted effects on the nervous system. *Transl Neurodegener*. (2024) 13:41. doi: 10.1186/s40035-024-00431-y

126. Rizzo MR, Di Meo I, Polito R, Auriemma MC, Gambardella A, di Mauro G, et al. Cognitive impairment and type 2 diabetes mellitus: Focus of SGLT2 inhibitors treatment. *Pharmacol Res.* (2022) 176:106062. doi: 10.1016/j.phrs.2022.106062

127. Schetters STT, Gomez-Nicola D, Garcia-Vallejo JJ, Van Kooyk Y. Neuroinflammation: microglia and T cells get ready to tango. *Front Immunol.* (2017) 8:1905. doi: 10.3389/fimmu.2017.01905

128. Goldenberg RM, Cheng AYY, Fitzpatrick T, Gilbert JD, Verma S, Hopyan JJ. Benefits of GLP-1 (Glucagon-like peptide 1) receptor agonists for stroke reduction in type 2 diabetes: A call to action for neurologists. *Stroke*. (2022) 53:1813–22. doi: 10.1161/STROKEAHA.121.038151

129. Malhotra K, Katsanos AH, Lambadiari V, Goyal N, Palaiodimou L, Kosmidou M, et al. GLP-1 receptor agonists in diabetes for stroke prevention: a systematic review and meta-analysis. *J Neurol.* (2020) 267:2117–22. doi: 10.1007/s00415-020-09813-4