Check for updates

OPEN ACCESS

EDITED BY Lynn Xiaoling Qiang, Northwell Health, United States

REVIEWED BY Hailong Song, University of Pennsylvania, United States Ivana Kawikova, National Institute of Mental Health, Czechia

*CORRESPONDENCE Jianguo Xu ⊠ xujg@scu.edu.cn Min He ⊠ hemin19910306@wchscu.cn

RECEIVED 03 October 2024 ACCEPTED 16 December 2024 PUBLISHED 09 January 2025

CITATION

Wang R, Xu J and He M (2025) Blood leukocyte-based clusters in patients with traumatic brain injury. *Front. Immunol.* 15:1504668. doi: 10.3389/fimmu.2024.1504668

COPYRIGHT

© 2025 Wang, Xu and He. This is an openaccess article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Blood leukocyte-based clusters in patients with traumatic brain injury

Ruoran Wang¹, Jianguo Xu^{1*} and Min He^{2*}

¹Department of Neurosurgery, West China Hospital, Sichuan University, Chengdu, Sichuan, China, ²Department of Critical Care Medicine, West China Hospital, Sichuan University, Chengdu, Sichuan, China

Background: Leukocytes play an important role in inflammatory response after a traumatic brain injury (TBI). We designed this study to identify TBI phenotypes by clustering blood levels of various leukocytes.

Methods: TBI patients from the Medical Information Mart for Intensive Care-III (MIMIC-III) database were included. Blood levels of neutrophils, lymphocytes, monocytes, basophils, and eosinophils were collected by analyzing the first blood sample within 24 h since admission. Overall, TBI patients were divided into clusters following the K-means clustering method using blood levels of five types of leukocytes. The correlation between identified clusters and mortality was tested by univariate and multivariate logistic regression analyses. The Kaplan–Meier method was used to verify the survival difference between identified TBI clusters.

Results: A total of 172 (cluster 1), 791 (cluster 2), and 636 (cluster 3) TBI patients were divided into three clusters with the following percentages, 10.8%, 49.5%, and 39.8%, respectively. Cluster 1 had the lowest Glasgow Coma Scale (GCS) and the highest Injury Severity Score (ISS) while cluster 2 had the highest GCS and the lowest ISS. The mortality rates of the three clusters were 25.6%, 13.3%, and 18.1%, respectively. The multivariate logistic regression indicated that cluster 1 had a higher mortality risk (OR = 2.211, p = 0.003) than cluster 2, while cluster 3 did not show a significantly higher mortality risk than cluster 2 (OR = 1.285, p = 0.163). Kapan–Meier analysis showed that cluster 1 had shorter survival than cluster 2 and cluster 3.

Conclusion: Three TBI phenotypes with different inflammatory statuses and mortality rates were identified based on blood levels of leukocytes. This classification is helpful for physicians to evaluate the prognosis of TBI patients.

KEYWORDS

traumatic brain injury, leukocytes, clusters, phenotypes, K-means

01

1 Introduction

Occurring widely with an estimated incidence of 69 million each year globally, traumatic brain injury (TBI) brings huge burdens to families of patients and to social economy (1). The mortality of TBI patients is high, especially severe TBI, with the mortality rate ranging from 23.0%-38.8% (2, 3). Accurate and convenient risk stratification of admitted TBI patients in the early phase is helpful for clinicians in making personalized treatments. The conventional tool of risk stratification for TBI is the Glasgow Coma Scale (GCS), which is evaluated based on clinical symptoms. However, single GCS could not comprehensively reflect the severity and progression of TBI patients due to the complex pathophysiological process of TBI and would be influenced by intubation and status of sedation. Identifying TBI phenotypes with different risks of clinical outcomes using laboratory biomarkers may make up for the insufficiency of GCS and guide clinicians in making specific treatment options based on pathophysiological processes.

Previous studies have explored phenotypes of some critically ill patients with acute kidney injury, sepsis, or acute respiratory distress syndrome using clustering methods (4–7). Composed of neutrophils, lymphocytes, monocytes, basophils, and eosinophils, leukocytes are recruited to the injured brain tissue and activated to participate in neuroinflammation after TBI (8–13). Previous studies have used the value of a single type of leukocytes or the neutrophil-to-lymphocyte ratio to reflect the inflammatory status and predict the prognosis of TBI (14–19). There is no study yet using blood values of all types of leukocytes to evaluate the prognosis of TBI. We designed this study to identify TBI phenotypes based on blood values of leukocytes using the K-means clustering method.

2 Materials and methods

2.1 Patients

The data for this study were extracted from the Medical Information Mart for Intensive Care-III (MIMIC-III) database produced by the Beth Israel Deaconess Medical Center (BIDMC). This database was approved by the institutional review boards of the Massachusetts Institute of Technology (MIT) and BIDMC. All patients included in this database were de-identified and anonymized for privacy protection. Written informed consent was waived due to the nature of the database study. The study design was approved by the review board of West China Hospital (2021-1598).

The MIMIC-III is a free, public database of collected clinical records of critically ill patients who received treatments in intensive care units of the BIDMC (Boston, MA) between 2001 and 2012. The MIMIC-III database was designed and produced by MIT (Cambridge, MA) and received ethical approval from the institutional review boards of MIT and BIDMC, respectively. The diagnosis of TBI was identified according to ICD-9 codes: 80000-80199; 80300-80499; and 8500-85419. Overall, from 2,680 TBI patients, some were excluded due to the following standards: (1)

lacked records of leukocytes on the first day (n = 1,018); (2) lacked records of GCS on admission (n = 25); and (3) lacked records of vital signs on admission (n = 38) (Figure 1). A total of 1,599 TBI patients were finally included in the study.

2.2 Data collection

Baseline characteristics including age, gender, blood pressure, SpO₂, and comorbidities (diabetes, hypertension, hyperlipidemia, coronary heart disease, and cancer) were recorded. Disease severity was evaluated by including Glasgow Coma Scale (GCS), Injury Severity Score (ISS), and Sequential Organ Failure Assessment (SOFA). The values of blood biochemical and blood routine examination were obtained by analyzing the first blood sample within 24 h since admission. The number of leukocytes including neutrophils, lymphocytes, monocytes, basophils, and eosinophils was part of the blood routine examination.

2.3 Statistical analysis

Overall, TBI patients were divided into clusters by the unsupervised K-means clustering method using values of five kinds of leukocytes (neutrophils, lymphocytes, monocytes, basophils, and eosinophils). The K-means clustering is an iterative algorithm that involves pre-dividing data into K groups, randomly selecting K objects as the initial clustering centers, and then calculating the distance between each object and each seed clustering center. Each object is assigned to the nearest cluster center. The cluster centers and the objects assigned to them represent a cluster. The optimal number of clusters was determined according to the gap statistic criterion. The differences in baseline characteristics and clinical outcomes among clusters were analyzed by Kruskal-Wallis test and Pearson's χ^2 test. Additionally, inflammatory markers including neutrophil-to-lymphocyte ratio (NLR), monocyte-to-lymphocyte ratio (MLR), platelet-to-lymphocyte ratio (PLR), neutrophil-to-



monocyte ratio (NMR), and systemic inflammation index (SII = platelet \times neutrophil/lymphocyte) were compared between clusters. The correlation between TBI clusters and mortality was confirmed by univariate and multivariate logistic regression analyses. The Kaplan–Meier method was used to verify the survival difference among identified TBI clusters.

A two-sided p value < 0.05 was considered statistically significant. The R software (version 3.6.1; R Foundation) was used for statistical analyses.

3 Results

3.1 Comparison of characteristics among identified TBI clusters

The optimal number of TBI clusters determined by the gap statistic criterion was 3 (Figure 2A). Two components of the cluster plots explained the 61.9% variability of the point (Figure 2B). A total of 172, 791, and 636 patients were divided into cluster 1, cluster 2, and cluster 3, respectively, with percentages of 10.8%, 49.5%, and 39.8% (Table 1). Age (p < 0.001), incidence of comorbidities including diabetes (p = 0.008), hypertension (p < 0.001), hyperlipidemia (p = 0.004), coronary heart disease (p = 0.019), and cancer (p < 0.001) showed different distributions among the three clusters. Cluster 1 had the lowest GCS and the highest ISS while cluster 2 had the highest GCS and the lowest ISS. The SOFA score did not differ among the three clusters (p = 0.256). Cluster 1 had the highest value of WBC, neutrophil, lymphocyte, monocyte, platelet, RBC, hemoglobin, and glucose (Figure 3). Cluster 2 had the lowest value of WBC, neutrophil, monocyte, platelet, RBC, hemoglobin, and glucose. Regarding inflammatory markers, cluster 1 showed a higher inflammatory status reflected by higher levels of NLR, MLR, PLR, NMR, and SII. In contrast, cluster 2 showed a lower inflammatory status indicated by lower levels of NLR, MLR, PLR, NMR, and SII. Similarly, cluster 1 had a higher incidence of mechanical ventilation, 30-day mortality, and longer length of ICU stay and length of hospital stay (Figure 4). Cluster 2 had a lower incidence of mechanical ventilation, 30-day mortality, and shorter length of ICU stay and length of hospital stay.

3.2 Relationship between identified clusters and outcomes of TBI

The mortality rates of the three clusters were 25.6%, 13.3%, and 18.1%, respectively. Univariate regression showed that cluster 1 (OR = 2.246, p < 0.001) and cluster 3 (OR = 1.442, p = 0.003) had higher mortality risks than cluster 2 (Table 2). Additionally, age (p < 0.001), diabetes (p < 0.001), hypertension (p = 0.216), cancer (p = 0.005), diastolic blood pressure (p = 0.038), GCS (p < 0.001), ISS (p < 0.001), SOFA (p < 0.001), SAH (p = 0.040), platelet (p = 0.016), RBC (p < 0.001), hemoglobin (p < 0.001), glucose (p < 0.001), and prothrombin time (p = 0.006) were significantly associated with TBI mortality. After adjusting the confounding effects of those factors, multivariate logistic regression indicated that cluster 1 had a higher mortality risk (OR = 2.211, p = 0.003) than cluster 2 while cluster 3 did not show a significantly higher mortality risk than cluster 2 (OR = 1.285, p = 0.163). Kapan-Meier analysis found that cluster 1 (p < 0.001) had shorter survival than cluster 2 (p < 0.001) and cluster 3 (p = 0.024; Figure 5). Cluster 3 had shorter survival than cluster 2 (p = 0.009).

4 Discussion

We found that the three TBI clusters had different inflammatory statuses through the clustering of peripheral leukocytes. Cluster 1 with a higher inflammatory status had a worse prognosis than cluster 2 with a lower inflammatory status. TBI could induce an increase in leukocytes soon after an initial injury. Neutrophils would be released from the bone marrow by stimulating cortisol and catecholamines after TBI (20). The activated microglia after TBI would then activate the endothelial cell and recruit the peripheral neutrophil to infiltrate the injured area by releasing inflammatory chemokines and cytokines (8, 9). Then, activated T cells and monocytes/macrophages would be recruited to the injured brain area and participate in the response of the adaptive immune system for the injured brain tissue (10–13). These cells would interactively activate and promote a secondary brain injury by aggravating neuroinflammation and immune response (21).

The relatively lower level of lymphocytes in the cluster and the higher level of lymphocytes in cluster 2 indicated that the peripheral immunosuppressive status might be related with a higher



TABLE 1 Baseline characteristics of included TBI patients.

Variables	Overall patients (n=1599)	Cluster 1 Cluster 2 (n=172, 10.8%) (n=791, 49.5%)		Cluster 3 (n=636, 39.8%)	р			
Age (years)	67.6 (47.2-82.3)	49.1 (28.0-73.9)	71.6 (54.0-83.2)	66.7 (43.4-82.2)	< 0.001			
Male gender (n, %)	959 (60.0 %)	108 (62.8%)	475 (60.1%)	376 (59.1%)	0.683			
Comorbidities								
Diabetes (n, %)	271 (16.9%)	18 (10.5%)	154 (19.5%)	99 (15.6%)	0.008			
Hypertension (n, %)	642 (40.2%)	45 (26.2%)	354 (44.8%)	243 (38.2%)	< 0.001			
Hyperlipidemia (n, %)	234 (14.6%)	16 (9.3%)	138 (17.4%)	80 (12.6%)	0.004			
Coronary heart disease (n, %)	238 (14.9%)	14 (8.1%)	131 (16.6%)	93 (14.6%)	0.019			
Cancer (n, %)	201 (12.6%)	12 (7.0 %)	133 (16.8%)	56 (8.8%) <0				
Systolic blood pressure (mmHg)	133 (117-148)	126 (112-144)	135 (120-150)	132 (116-147)	< 0.001			
Diastolic blood pressure (mmHg)	67 (56-78)	(56-78) 63 (56-75) 67 (57-79)		67 (55-78) 0				
SpO ₂ (%)	99 (97-100)	100 (98-100)	98 (96-100)	99 (97-100)	< 0.001			
GCS	13 (7-15)	13 (7-15) 7 (5-12) 14 (9-15)		10 (6-15) <0.0				
ISS	16 (16-22)	22 (16-29)	16 (16-17)	16 (16-25)	< 0.001			
SOFA	3 (1-4)	3 (1-5)	3 (1-4)	3 (1-4)	0.256			
Intracranial injury types								
EDH (n, %)	322 (20.1%)	54 (31.4%)	119 (15.0%)	149 (23.4%)	< 0.001			
SDH (n, %)	909 (56.8%)	86 (50.0%)	481 (60.8%)	342 (53.8%)	0.005			
SAH (n, %)	589 (36.8%)	78 (45.3%)	259 (32.7%)	252 (39.6%)	0.001			
Laboratory tests								
WBC (10^9/L)	11.30 (8.20-15.50)	23.30 (20.70-26.80)	8.20 (6.60-9.70)	14 (12.40-16.20)	< 0.001			
Neutrophil (10^9/L)	9.04 (6.05-12.72)	19.40 (17.80-22.40)	5.97 (4.53-7.53)	11.78 (10.50-13.72)	< 0.001			
Lymphocyte (10^9/L)	1.26 (0.85-1.91)	1.64 (1.03-2.72)	1.31 (0.85-1.88)	1.19 (0.82-1.81)	< 0.001			
Monocyte (10^9/L)	0.44 (0.32-0.64)	0.84 (0.62-1.31	0.37 (0.28-0.49)	0.50 (0.36-0.71)	< 0.001			
Basophil (10^9/L)	0.03 (0.01-0.05)	5) 0.04 (0-0.07) 0.03 (0.01-0.05)		0.03 (0.01-0.05)	0.387			
Eosinophil (10^9/L)	0.07 (0.02-0.16)	0.05 (0-0.19) 0.08 (0.03-0.17)		0.05 (0.02-0.13)	< 0.001			
Platelet (10^9/L)	230 (184-286)	287 (228-344) 212 (161-267)		237 (198-292)	< 0.001			
RBC (10^9/L)	4.12 (3.66-4.57)	4.38 (3.89-4.77) 4.05 (3.59-4.45)		4.17 (3.71-4.62)	< 0.001			
Hemoglobin (g/dL)	12.7 (11.2-14.1)	13.3 (11.9-14.7) 12.5 (11.1-1		12.9 (11.4-14.2)	< 0.001			
Glucose (mg/dL)	131 (109-163)	152 (125-187)	118 (102-148)	142 (118-174)	< 0.001			
Prothrombin time (s)	13.1 (12.3-14.4)	13.4 (12.6-14.5)	13.2 (12.3-14.6)	13.0 (12.3-14.2)	0.128			
Inflammatory markers								
NLR	6.92 (4.01-11.74)	12.50 (7.50-19.40)	4.42 (2.70-7.05)	9.84 (6.54-14.22)	< 0.001			
MLR	0.35 (0.23-0.55)	0.50 (0.36-0.77)	0.28 (0.19-0.42)	0.41 (0.29-0.60)	< 0.001			
PLR	174.89 (114.25-267.03)	174.95 (105.02-293.63)	156.78 (103.09-234.09)	202.55 (128.62-296.06)	< 0.001			
NMR	18.66 (13.21-27.42)	23.184 (15.60-31.52)	15.46 (11.00-21.30)	23.162 (16.60-31.82)	< 0.001			
SII	1597.50 (821.59-2856.91)	3521.05 (2157.55-5700.14)	887.66 (527.63-1537.54)	2400.64 (1497.13-3552.40)	< 0.001			
Platelet transfusion during the first day (n, %)	152 (9.5%)	10 (5.8%)	95 (12.0%)	47 (7.4%)	0.003			

(Continued)

TABLE 1 Continued

Variables	Overall patients (n=1599)	Cluster 1 (n=172, 10.8%)	Cluster 2 (n=791, 49.5%)	Cluster 3 (n=636, 39.8%)	р
Inflammatory markers					
RBC transfusion during the first day (n, %)	105 (6.6%)	13 (7.6%)	50 (6.3%)	42 (6.6%)	0.837
Mechanical ventilation (n, %)	688 (43.0%)	107 (62.2%)	257 (32.5%)	324 (50.9%)	< 0.001
30-day mortality (n, %)	264 (16.5%)	44 (25.6%)	105 (13.3%)	115 (18.1%)	< 0.001
Length of ICU stay (days)	2.4 (1.2-6.0)	4.0 (2.0 -9.8)	2.1 (1.1-4.6)	2.7 (1.3-7.2)	< 0.001
Length of hospital stay (days)	6.9 (3.8-14.4)	9.0 (4.5-16.8)	6.5 (3.7-12.0)	7.7 (3.6-16.1)	0.003

SpO₂, pulse oxygen saturation; GCS, Glasgow Coma Scale; ISS, Injury Severity Score; SOFA, sequential organ failure assessment; EDH, epidural hematoma; SDH, subdural hematoma; SAH, subarachnoid hemorrhage; WBC, white blood cell; RBC, red blood cell; NLR, neutrophil to lymphocyte ratio; MLR, monocyte to lymphocyte ratio; PLR, platelet to lymphocyte ratio; NMR, neutrophil to monocyte ratio; SII, systemic immune inflammation index = platelet × neutrophil/lymphocyte.



FIGURE 3

(A) Blood neutrophil level in clusters. (B) Blood lymphocyte level in clusters. (C). Blood monocyte level in clusters. (D) Blood basophil level in clusters. (E) Blood eosinophil level in clusters. (F) Neutrophil-to-lymphocyte ratio (NLR) level in clusters. (G) Monocyte-to-lymphocyte (MLR) level in clusters. (H) Platelet-to-lymphocyte (PLR) level in clusters. (I) Neutrophil-to-monocyte (NMR) level in clusters. (J) Systemic inflammation index (SII) level in clusters. *** means p value<0.001



TABLE 2 Univariate and multivariate logistic regression analysis of risk factors for mortality in TBI patients.

	U	Unadjusted analysis			Adjusted analysis		
Variables	OR	95% CI	P value	OR	95% CI	P value	
Age	1.027	1.020-1.035	< 0.001	1.046	1.036-1.057	<0.001	
Male gender	0.840	0.643-1.097	0.200				
Diabetes	1.790	1.304-2.458	<0.001	1.002	0.993-1.011	0.628	
Hypertension	1.183	0.906-1.545	0.216				
Hyperlipidemia	1.205	0.842-1.725	0.307				
Coronary heart disease	1.137	0.793-1.631	0.483				
Cancer	1.661	1.162-2.374	0.005	1.288	0.859-1.931	0.220	
Systolic blood pressure	0.999	0.993-1.004	0.610				
Diastolic blood pressure	0.992	0.984-1.000	0.038	1.569	1.028-2.397	0.037	
SpO ₂ (%)	0.998	0.968-1.029	0.900				
GCS	0.857	0.831-0.883	<0.001	0.833	0.801-0.865	<0.001	
ISS	1.032	1.017-1.047	<0.001	1.033	1.013-1.054	0.001	
SOFA	1.273	1.210-1.338	<0.001	1.162	1.093-1.234	< 0.001	
EDH	1.112	0.805-1.535	0.519				
SDH	0.999	0.765-1.304	0.991				
SAH	1.325	1.013-1.734	0.040	1.527	1.114-2.095	0.009	
Leukocytes clusters			< 0.001			0.012	
Cluster 2	1.000	Reference		1.000	Reference		
Cluster 1	2.246	1.506-3.348	<0.001	2.211	1.310-3.733	0.003	
Cluster 3	1.442	1.081-1.923	0.003	1.285	0.903-1.827	0.163	
Platelet	0.998	0.997-1.000	0.016	1.855	1.162-2.960	0.010	
RBC	0.659	0.547-0.793	< 0.001	0.782	0.669-0.915	0.002	
Hemoglobin	0.837	0.788-0.890	< 0.001	0.999	0.997-1.001	0.260	
Glucose	1.009	1.006-1.011	< 0.001	1.003	1.000-1.005	0.035	
Prothrombin time	1.022	1.006-1.037	0.006	1.015	0.999-1.031	0.062	

OR, odds ratio; CI, confidence interval; SpO₂, pulse oxygen saturation; GCS, Glasgow Coma Scale; ISS, Injury Severity Score; SOFA, sequential organ failure assessment; EDH, epidural hematoma; SDH, subdural hematoma; SAH, subarachnoid hemorrhage; RBC, red blood cell.

inflammatory status and worse outcome. Actually, previous studies found that TBI could cause the acute increase of cortisol levels both in serum and cerebrospinal fluid (22, 23). The increased plasma cortisol in the early phase after TBI would prevent the lymphocyte egress from the secondary lymphoid tissues, which may be an endogenous protective response to inhibit the excessive infiltration of T cells in the injured brain area, subsequently amplifying neuroinflammation (24). However, the peripheral immunosuppressive status is certainly associated with a higher risk of infection among extracranial organs.

Many previous studies used the value of a single type of leukocytes or neutrophil-to-lymphocyte ratio to evaluate the risk of poor prognosis of TBI (14–19), while not all types of leukocytes were included and analyzed to evaluate the prognosis risk, which was a limitation of these studies. We performed this study to identify inflammatory clusters of TBI patients with different mortality risks by comprehensively clustering multiple types of leukocytes, namely, neutrophils, lymphocytes, monocytes, basophils, and eosinophils. Three inflammatory clusters of TBI were identified and were consistent with their GCS. This classification is beneficial for physicians to stratify risks and make personalized treatment schedules. It may also be helpful to identify TBI subgroups benefiting from specific treatments targeted at inflammation.

Several limitations were unavoidable in this study. Firstly, a number of patients were excluded from this study mainly due to lack of records on leukocytes on the first day, causing a selection bias. The classification we identified should be further verified in other medical centers with more generalized TBI patients and a prospective design collecting levels of leukocytes at a fixed time period, as early as possible, after admission. Secondly, specific subgroups of lymphocytes were not analyzed including NK cells, B cells, cytotoxic T cells, and regulatory T cells due to lack of records



from the MIMIC-III database. Thirdly, common inflammation markers such as C-reactive protein, interleukin-1, interlukin-6, TNF- α , and interferon- γ were not analyzed among the three clusters due to lack of records from MIMIC-III. Future studies could be performed to measure these markers among our identified clusters and evaluate the consistency between these markers and clusters with different inflammatory statuses. Fourthly, functional outcomes and recovery statuses were not recorded in MIMIC-III, so we were unable to analyze the relationship between the discovered three clusters and these outcomes. Future studies could be designed to verify the importance of the three clusters on functional outcomes and recovery statuses. Finally, agents with immunomodulatory effects or cytotoxicity were not particularly extracted and analyzed, which may limit the reliability of our classifications.

5 Conclusion

Three TBI clusters with different inflammatory statuses and prognoses were identified based on levels of blood leukocytes. This classification is beneficial for physicians in evaluating the prognosis and making personalized treatments for TBI patients.

Data availability statement

Publicly available datasets were analyzed in this study. This data can be found here: https://physionet.org/content/mimiciii/1.4/.

Ethics statement

The studies involving humans were approved by Ethics Committee of the West China hospital (2021-1598). The studies were conducted in accordance with the local legislation and institutional requirements. The participants provided their written informed consent to participate in this study.

Author contributions

RW: Conceptualization, Data curation, Formal Analysis, Writing – original draft. JX: Funding acquisition, Supervision, Validation, Writing – review & editing. MH: Funding acquisition, Supervision, Validation, Writing – review & editing, Project administration.

Funding

The author(s) declare that financial support was received for the research, authorship, and/or publication of this article. This study was funded by the Sichuan Science and Technology Program (24QYCX0411, 2024YFHZ0070), the 1•3•5 project for disciplines of excellence—Clinical Research Incubation Project, West China Hospital, Sichuan University (2020HXFH036), and the National Natural Science Foundation of China (82173175).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Generative AI statement

The author(s) declare that no Generative AI was used in the creation of this manuscript.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated

References

1. Dewan MC, Rattani A, Gupta S, Baticulon RE, Hung YC, Punchak M, et al. Estimating the global incidence of traumatic brain injury. *J Neurosurg.* (2018) 2018:1–18. doi: 10.3171/2017.10.JNS17352

2. Roozenbeek B, Chiu YL, Lingsma HF, Gerber LM, Steyerberg EW, Ghajar J, et al. Predicting 14-day mortality after severe traumatic brain injury: application of the IMPACT models in the brain trauma foundation TBI-trac[®] New York State database. *J Neurotrauma*. (2012) 29:1306–12. doi: 10.1089/neu.2011.1988

3. Dawes AJ, Sacks GD, Cryer HG, Gruen JP, Preston C, Gorospe D, et al. Intracranial pressure monitoring and inpatient mortality in severe traumatic brain injury: A propensity score-matched analysis. *J Trauma acute Care Surg.* (2015) 78:492– 501; discussion 501-492. doi: 10.1097/TA.000000000000559

4. Boussina A, Wardi G, Shashikumar SP, Malhotra A, Zheng K, Nemati S. Representation learning and spectral clustering for the development and external validation of dynamic sepsis phenotypes: observational cohort study. *J Med Internet Res.* (2023) 25:e45614. doi: 10.2196/45614

5. Lai CF, Liu JH, Tseng LJ, Tsao CH, Chou NK, Lin SL, et al. Unsupervised clustering identifies sub-phenotypes and reveals novel outcome predictors in patients with dialysis-requiring sepsis-associated acute kidney injury. *Ann Med.* (2023) 55:2197290. doi: 10.1080/07853890.2023.2197290

6. Sinha P, Kerchberger VE, Willmore A, Chambers J, Zhuo H, Abbott J, et al. Identifying molecular phenotypes in sepsis: an analysis of two prospective observational cohorts and secondary analysis of two randomised controlled trials. *Lancet Respir Med.* (2023) 11(11):965-74. doi: 10.1016/S2213-2600(23)00237-0

7. Chen H, Yu Q, Xie J, Liu S, Pan C, Liu L, et al. Longitudinal phenotypes in patients with acute respiratory distress syndrome: a multi-database study. *Crit Care (London England).* (2022) 26:340. doi: 10.1186/s13054-022-04211-w

8. Zhou H, Lapointe BM, Clark SR, Zbytnuik L, Kubes P. A requirement for microglial TLR4 in leukocyte recruitment into brain in response to lipopolysaccharide. *J Immunol (Baltimore Md: 1950).* (2006) 177:8103-10. doi: 10.4049/jimmunol.177.11.8103

 Clark RS, Schiding JK, Kaczorowski SL, Marion DW, Kochanek PM. Neutrophil accumulation after traumatic brain injury in rats: comparison of weight drop and controlled cortical impact models. *J Neurotrauma*. (1994) 11:499–506. doi: 10.1089/ neu.1994.11.499

10. Holmin S, Mathiesen T. Biphasic edema development after experimental brain contusion in rat. *Neurosci Lett.* (1995) 194:97–100. doi: 10.1016/0304-3940(95)11737-H

11. Beschorner R, Nguyen TD, Gözalan F, Pedal I, Mattern R, Schluesener HJ, et al. CD14 expression by activated parenchymal microglia/macrophages and infiltrating monocytes following human traumatic brain injury. *Acta neuropathologica*. (2002) 103:541–9. doi: 10.1007/s00401-001-0503-7

12. Ziebell JM, Morganti-Kossmann MC. Involvement of pro- and antiinflammatory cytokines and chemokines in the pathophysiology of traumatic brain organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

injury. Neurotherapeutics: J Am Soc Exp Neurother. (2010) 7:22-30. doi: 10.1016/ j.nurt.2009.10.016

13. Jassam YN, Izzy S, Whalen M, McGavern DB, El Khoury J. Neuroimmunology of traumatic brain injury: time for a paradigm shift. *Neuron*. (2017) 95:1246–65. doi: 10.1016/j.neuron.2017.07.010

14. Mukherjee S, Sivakumar G, Goodden JR, Tyagi AK, Chumas PD. Prognostic value of leukocytosis in pediatric traumatic brain injury. *J Neurosurg Pediatr.* (2020) 27:335–45. doi: 10.3171/2020.7.PEDS19627

15. Li Z, Wu X, Wu X, Yu J, Yuan Q, Du Z, et al. Admission circulating monocytes level is an independent predictor of outcome in traumatic brain injury. *Brain injury*. (2018) 32:515–22. doi: 10.1080/02699052.2018.1429023

16. Matias LF, Pimentel MD, Medeiros MF, Rocha FR, Gambetta MV, Lopes SC. Predictive value of neutrophil-to-lymphocyte ratio and neutrophil-to-monocyte ratio in severe traumatic brain injury: a retrospective cohort. *J neurosurgical Sci.* (2023) 68 (5):604–11. doi: 10.23736/S0390-5616.23.05877-0

17. Alexiou GA, Lianos GD, Zika J, Alexiou ES, Voulgaris S. Neutrophil-tolymphocyte ratio to predict prognosis in traumatic brain injury. *Am J Emergency Med.* (2022) 56:341. doi: 10.1016/j.ajem.2021.09.017

18. Chen L, Xia S, Zuo Y, Lin Y, Qiu X, Chen Q, et al. Systemic immune inflammation index and peripheral blood carbon dioxide concentration at admission predict poor prognosis in patients with severe traumatic brain injury. *Front Immunol.* (2022) 13:1034916. doi: 10.3389/fimmu.2022.1034916

19. Ge X, Zhu L, Li M, Li W, Chen F, Li Y, et al. A novel blood inflammatory indicator for predicting deterioration risk of mild traumatic brain injury. *Front Aging Neurosci.* (2022) 14:878484. doi: 10.3389/fnagi.2022.878484

20. Hazeldine J, Naumann DN, Toman E, Davies D, Bishop JRB, Su Z, et al. Prehospital immune responses and development of multiple organ dysfunction syndrome following traumatic injury: A prospective cohort study. *PloS Med.* (2017) 14:e1002338. doi: 10.1371/journal.pmed.1002338

21. Simon DW, McGeachy MJ, Bayır H, Clark RS, Loane DJ, Kochanek PM. The farreaching scope of neuroinflammation after traumatic brain injury. *Nat Rev Neurol.* (2017) 13:171–91. doi: 10.1038/nrneurol.2017.13

22. Wagner AK, McCullough EH, Niyonkuru C, Ozawa H, Loucks TL, Dobos JA, et al. Acute serum hormone levels: characterization and prognosis after severe traumatic brain injury. *J Neurotrauma*. (2011) 28:871–88. doi: 10.1089/neu.2010.1586

23. Santarsieri M, Niyonkuru C, McCullough EH, Dobos JA, Dixon CE, Berga SL, et al. Cerebrospinal fluid cortisol and progesterone profiles and outcomes prognostication after severe traumatic brain injury. *J Neurotrauma*. (2014) 31:699–712. doi: 10.1089/neu.2013.3177

24. Dong T, Zhi L, Bhayana B, Wu MX. Cortisol-induced immune suppression by a blockade of lymphocyte egress in traumatic brain injury. *J Neuroinflamm*. (2016) 13:197. doi: 10.1186/s12974-016-0663-y