



OPEN ACCESS

EDITED BY

Vinay Kumar,
The Pennsylvania State University,
United States

REVIEWED BY

Shravan Girada,
University of California, San Diego,
United States
Dharmendra Bhatt,
National Institutes of Health (NIH),
United States

*CORRESPONDENCE

Guangrui Huang
✉ hgr@ucm.edu.cn
Chen Li
✉ casio1981@163.com

[†]These authors have contributed
equally to this work and share
first authorship

RECEIVED 08 September 2024

ACCEPTED 07 November 2024

PUBLISHED 26 November 2024

CITATION

Geng S, Hu B, Guan Y, Jiang Y, Shu Z, Li C
and Huang G (2024) Advances of the
multifaceted functions of PSTPIP2 in
inflammatory diseases.
Front. Immunol. 15:1492878.
doi: 10.3389/fimmu.2024.1492878

COPYRIGHT

© 2024 Geng, Hu, Guan, Jiang, Shu, Li and
Huang. This is an open-access article
distributed under the terms of the [Creative
Commons Attribution License \(CC BY\)](#). The
use, distribution or reproduction in other
forums is permitted, provided the original
author(s) and the copyright owner(s) are
credited and that the original publication in
this journal is cited, in accordance with
accepted academic practice. No use,
distribution or reproduction is permitted
which does not comply with these terms.

Advances of the multifaceted functions of PSTPIP2 in inflammatory diseases

Shaohui Geng ^{1†}, Bohan Hu ^{2†}, Yiwei Guan ^{2†}, Yijin Jiang ²,
Zixuan Shu ², Chen Li ^{3*} and Guangrui Huang ^{1*}

¹School of Life Sciences, Beijing University of Chinese Medicine, Beijing, China, ²School of Chinese Materia Medica, Beijing University of Chinese Medicine, Beijing, China, ³Department of Rheumatology, Fangshan Hospital, Beijing University of Chinese Medicine, Beijing, China

The complex interaction between the immune system and autoinflammatory disorders highlights the centrality of autoimmune mechanisms in the pathogenesis of autoinflammatory diseases. With the exploration of PSTPIP2, it has been discovered to play an inhibitory role in immune diseases, suggesting its potential utility in the research and treatment of rheumatic diseases. This review outlines the mechanisms of PSTPIP2 in chronic multifocal osteomyelitis (CMO), rheumatoid arthritis (RA), synovitis-acne-pustulosis-hyperostosis-osteitis (SAPHO) syndrome, liver diseases, renal diseases, pressure ulcer sepsis and diabetic obesity. The mechanisms include inhibiting the IL-1 β inflammatory responses, NF- κ B, ERK phosphorylation etc., promoting Er β , and modulating the polarization of macrophage to prevent the inflammatory diseases. This review summarized current findings and offered perspectives on future research directions, laying a foundation for applying of PSTPIP2 in inflammatory diseases.

KEYWORDS

PSTPIP2, inflammatory diseases, osteomyelitis, inflammation, SAPHO syndrome

1 Introduction

As research into immune inhibitory proteins progresses, proline-serine-threonine phosphatase-interacting protein 2 (PSTPIP2), a member of the Fes/CIP4 homology-Bin/Amphiphysin/Rvs (F-BAR) domain family, has been identified as an adaptor protein residing on the cell membrane (1–3). Serving as an immune suppressor, PSTPIP2 inhibits inflammation and mitigates the damage inflicted by the immune system on the body (4–6). Its efficacy is anchored in its modulation of multiple signaling cascades, notably by inhibiting the functional activity of IL-1 β , modulating the chemokine CXCL2 within neutrophil granules, and suppressing the production of reactive oxygen species (ROS) via the neutrophil NOX2 NADPH oxidase. Furthermore, PSTPIP2 demonstrates its role as an immune suppressor by effectively dampening inflammation and alleviating immune-induced tissue damage through these multifaceted mechanisms. It accomplishes this by inhibiting the activity of IL-1 β , fine-tuning CXCL2 within neutrophil granules, and suppressing the

generation of reactive oxygen species (ROS) by neutrophil NOX2 NADPH oxidase. Additionally, PSTPIP2 exerts inhibitory effects on osteoclastogenesis. By modulating these pathways, PSTPIP2 dampens immune responses, thereby mitigating the harm inflicted on the body (7).

The damage wrought by immune responses on the human body poses a formidable clinical hurdle. This condition, characterized by abnormal immune activation, often manifests as fever, joint swelling, pain, and even deformation, along with skin redness, rash, and itching. These impairments frequently pose significant therapeutic challenges (8–12). Currently, there are no specific drugs available, and most therapies aim to alleviate these symptoms by suppressing the overall immune system of the body, which can be highly detrimental to the patient (13–16). Consequently, research into these immune-mediated disorders is of paramount importance. PSTPIP2, as a relevant protein, underscores its significance value in immune disease research by suppressing immune responses and mitigating the damage caused by the immune system to the body (17–19). While researchers are actively delving into the role of PSTPIP2 in various immune disorders, the precise mechanisms underlying its function in some of these diseases, such as its inhibition of IL-1 β , remain incompletely understood. A deeper exploration of these mechanisms is imperative for advancing our comprehension of PSTPIP2's potential therapeutic applications.

This comprehensive review offers an extensive perspective on the advancements in understanding the mechanisms of PSTPIP2 in context of autoinflammatory diseases, as vividly depicted in **Figure 1**. We systematically outline the role of PSTPIP2 in CMO, RA, SAPHO syndrome, liver diseases, renal diseases, pressure ulcer sepsis and diabetic obesity. We hope to provide insights and guidance for future research directions and drug design related to PSTPIP2 in inflammatory diseases.

2 Osteomyelitis

Osteomyelitis, characterized by infection and bone destruction, presents with pain in the affected area, along with fever, weight loss, localized redness and swelling, and tenderness on palpation (20–23). This chapter delves into the pivotal role of PSTPIP2 protein in chronic multifocal osteomyelitis (CMO). The pathogenetic role of PSTPIP2 in osteomyelitis is shown in **Figure 2**. PSTPIP2, a crucial mediator in autoinflammatory diseases, can initiate or exacerbate the symptoms of osteomyelitis when absent or mutated. Recent research have unveiled that PSTPIP2 collaborates with the suppression of pro-inflammatory factors like IL-1 β , the regulation of megakaryocyte and neutrophil functions, and the interaction with proteins such as protein tyrosine phosphatases containing a PEST domain (PEST-PTPs) to collectively inhibit inflammatory responses. Additionally, the emerging role of PSTPIP2 in modulating the gut microbiome in chronic multifocal osteomyelitis presents a novel perspective for disease treatment.

The CMO represents a distinctive subtype of osteomyelitis primarily characterized by bone pain and pyrexia. Due to the unclear pathogenesis, the treatment rate remains low (24–27). Scholars had demonstrated that the absence of the PSTPIP2

protein in animals leads to the development of autoinflammatory manifestations, suggesting a role for PSTPIP2 in controlling the onset of autoinflammatory diseases. These discoveries provide novel insights into the pathogenesis of CMO and other related autoinflammatory disorders, setting the stage for future research endeavors in this field. Ferguson, PJ et al. (28) utilized backcross strategies to localize the CMO gene and identified the presence of the PSTPIP2 gene within the region. They discovered a single base pair mutation, suggesting that mutations in PSTPIP2 may serve as a genetic determinant contributing to the autoimmune inflammatory phenotype observed in CMO mice. Similarly, Chen, TC (29) et al. induced PSTPIP2 mutations in C57BL/6 J mice through N-ethyl-N-nitrosourea mutagenesis, and these mice exhibited inflammatory responses in areas such as claws. Furthermore, Chitu, V et al. (30) found that asymptomatic PSTPIP2 (CMO) mice had an increased number of macrophage precursors in their spleens. The lack of PSTPIP2 facilitated the proliferation of macrophage progenitors and augmented the responsiveness of mature macrophages to activating stimuli. This combination effect predisposes the organism to excessive and sustained inflammatory responses, ultimately resulting in autoimmune inflammatory diseases.

2.1 IL-1 β -mediated pathway

The inhibitory effect of PSTPIP2 on IL-1 β also holds considerable significance in CMO. Drobek, A et al. (31) demonstrated that the C-terminal key tyrosine residue region of PSTPIP2 is crucial for its inhibition of IL-1 β processing in neutrophils, through its binding to the inhibitory enzymes CSK and SHIP1. Furthermore, the inhibitory capacity of SHIP1 enhances this process. Similarly, Gurung, P et al. (32) revealed the combined role of IL-1 β in driving the disease progression in PSTPIP2(CMO) mice. The number of IL-1 receptors (IL-1Ra) in PSTPIP2-deficient mice also impacts disease severity. Cassel SL et al. (33) demonstrated that the absence of IL-1RI in CMO mice significantly shortened the disease onset time and reduced the degree of bone lesions, indicating that controlling the number of IL-1RI could be a potential therapeutic direction for CMO caused by PSTPIP2 deficiency. In addition, Lukens, JR et al. (34, 35) identified PSTPIP2 as a negative regulator of caspase-1-mediated autonomous IL-1 β production. This finding underscores the mechanism by which PSTPIP2 exerts its anti-inflammatory effects by modulating IL-1 β levels, thereby preventing the unchecked activation of inflammatory responses that could lead to autoimmune or autoinflammatory conditions.

2.2 Neutrophil-mediated pathway

Kralova, J et al. (10) discovered that in addition to IL-1 β , PSTPIP2 negatively regulates the pathway of neutrophil NOX2 NADPH oxidase, which generates reactive oxygen species (ROS). PSTPIP2(CMO) neutrophils exhibit extremely high superoxide production in response to various stimuli, implicating dysregulated NADPH oxidase activity as a pivotal mediator of autoimmune inflammatory bone damage in PSTPIP2(CMO) mice. Furthermore,

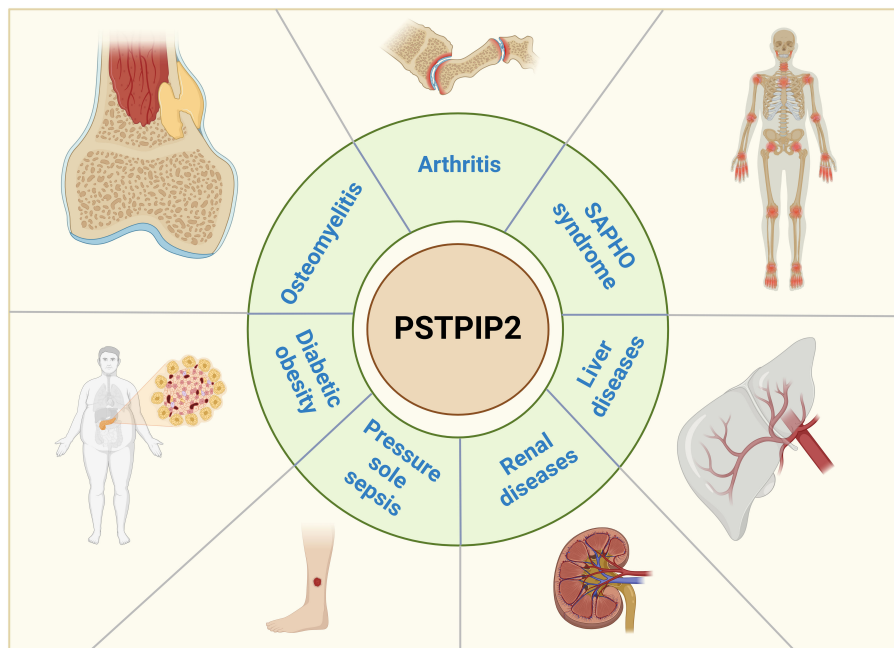


FIGURE 1 Research progress on the mechanism of action of PSTPIP2 in inflammatory diseases.

PSTPIP2 inhibits the exaggerated neutrophil responses to various stimuli induced by ROS produced by neutrophil NOX2 NADPH oxidase, thereby suppressing inflammation. Pavliuchenko, N et al. (36) used mouse strains with disrupted PEST or SHIP1 binding sites in PSTPIP2 to demonstrate that when PEST-PTPs cannot bind to PSTPIP2, it leads to dysregulation of the chemokine CXCL2 in neutrophils, causing symptomatic disease.

2.3 Gut microbiota-mediated pathway

Lukens, JR et al. (35) investigated the characteristics of the gut microbiota in PSTPIP2(CMO) mice and elucidated that diet-related changes in the gut microbiota composition play a pivotal role in regulating caspase-1 and caspase-8-mediated IL-1 β maturation, which subsequently impacts the development of osteomyelitis in

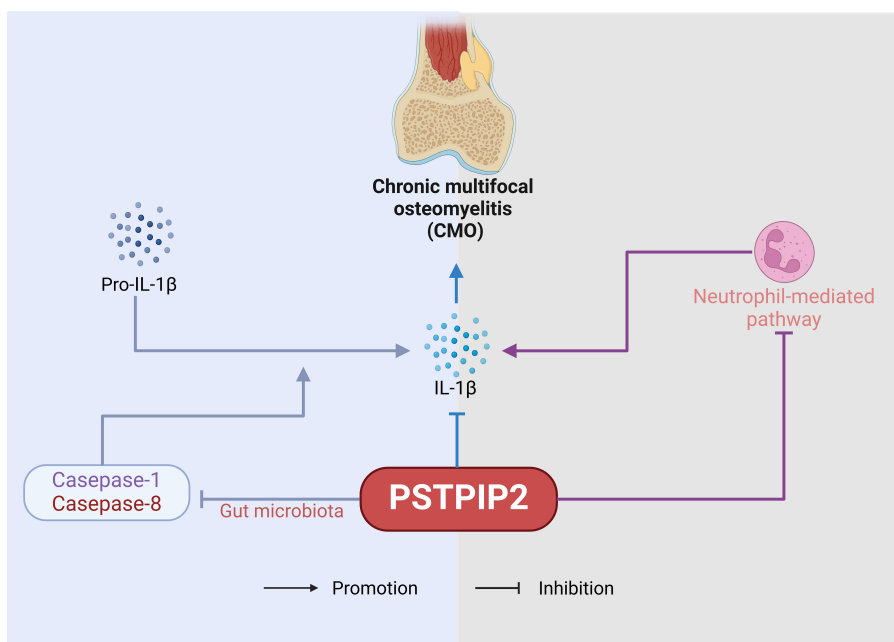


FIGURE 2 The pathogenetic role of PSTPIP2 in osteomyelitis.

these animals. Their findings hint at the promising potential of dietary interventions aimed at modulating the gut microbiota as a therapeutic strategy for the treatment of chronic multifocal osteomyelitis (CMO) arising from PSTPIP2 deficiency. However, the specific relationship between PSTPIP2 protein and the gut microbiota remains to be further investigated.

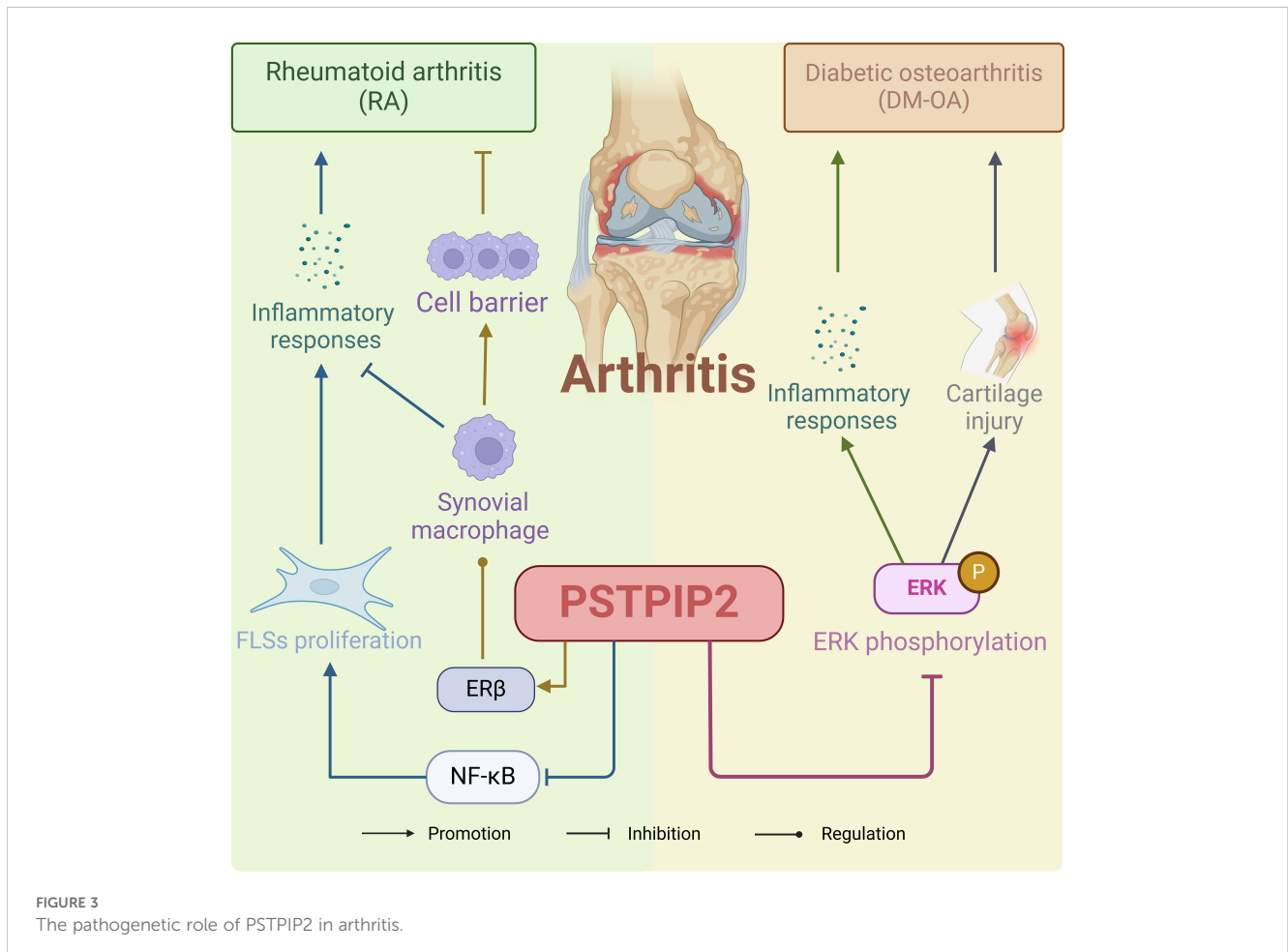
3 Arthritis

Arthritis represents a diverse group of inflammatory diseases affecting human joints and their surrounding tissues, manifesting as bone hyperplasia, and ligament tissue alterations, among other symptoms (37–40). Emerging research has shown that PSTPIP2 can inhibit osteoclast development and thereby prevent the onset of arthritis. The pathogenic role of PSTPIP2 in arthritis is shown in Figure 3. Chitu, V et al. (41) delved into the mechanism underlying PSTPIP2's regulation of osteoclast development by examining cmo models unable to express PSTPIP2 and Lupu models with PSTPIP2 dysfunction. They found that PSTPIP2 acts as a negative feedback regulator of CSF-1R signaling, inhibiting TRAP expression, and osteoclast precursor fusion, thus suppressing inflammation and osteoclastogenesis, where PSTPIP2 tyrosine phosphorylation and a functional F-BAR domain played an important role. Similarly, Tsujita, K et al. (42) discovered the complex interplay between

PSTPIP2 and other F-BAR domain proteins in regulating cellular processes that impact inflammation and tissue remodeling. Sztacho, M (43) discovered that PSTPIP2 plays a role in regulating podosome assembly within the podosome/sealed dynamics monitoring mechanism.

3.1 Rheumatoid arthritis

Rheumatoid arthritis (RA) is a chronic autoimmune inflammatory disease (44–48), characterized by erosive arthritis with synovitis as its underlying pathological mechanism. Initial symptoms include morning stiffness, swelling, and pain in the joints, which can progressively evolve into joint deformity (49–52). Fibroblast-like synoviocytes (FLS) play a crucial role in its pathogenesis (53–56). Yao Y (3) conducted a study utilizing an arthritis animal model to investigate the function of PSTPIP2 in FLS and demonstrated that PSTPIP2 exerts inhibitory effects on FLS proliferation and inflammatory responses. Moreover, the expression mechanism of PSTPIP2 is closely related to the NF- κ B signaling pathway. Grosse, J et al. (57) found that PSTPIP2 has anti-inflammatory effects in macrophages. Yao, Y (58) explored the molecular mechanism of PSTPIP2's anti-bone erosion effects by overexpressing PSTPIP2 protein *in vivo* experiments. They discovered that PSTPIP2 regulates synovial macrophage polarization and dynamics through Estrogen



Receptor Beta, forming an immune barrier (F4/80(+)/PSTPIP2(hi) cell-enriched zone) at the joint, thereby controlling rheumatoid arthritis bone erosion. This suggests that locally regulating PSTPIP2 expression in the joint microenvironment may be a potential strategy for controlling rheumatoid arthritis bone erosion.

3.2 Diabetic osteoarthritis

Diabetic osteoarthritis (DM-OA) often affects individuals with inadequate glycemic control despite prolonged oral hypoglycemic medication and stands as a prominent cause of disability (59–62). Its primary symptoms include joint pain, stiffness, and swelling (63–65). PSTPIP2 has a mitigating effect on DM-OA and bone damage. Li, M et al. (66) investigated the potential pathways of PSTPIP2 influencing DM-OA progression by overexpressing PSTPIP2 through intra-articular injection of lentiviral vectors. They found that PSTPIP2 overexpression alleviates synovial inflammation and bone damage in DM-OA by inhibiting ERK phosphorylation. In addition, Liu, L et al. (67) also discovered PSTPIP2 overexpression caused enhanced activation of Src family kinases and subsequently reduced ERK phosphorylation, and verified that PSTPIP2 upregulation repressed megakaryocyte development in primary mouse bone marrow cells.

4 SAPHO syndrome

SAPHO syndrome is an uncommon condition primarily affecting bones and skin, with its primary diagnostic feature being chronic multifocal osteitis (68–71). The protein PSTPIP2 plays crucial roles in macrophage activation, neutrophil migration, and osteoclast differentiation (72). Liao HJ et al. (73) generated PSTPIP2 knockout (Pstpip2^{-/-}) mice and observed that all Pstpip2^{-/-} mice developed an inflammatory disease resembling SAPHO syndrome. Notably, inflamed tissues exhibited significant elevations in chemokines attracting neutrophils and IL-1 β , hinting at a potential role for PSTPIP2 in innate immunity and autoinflammatory bone diseases, possibly linked to the pathogenesis of human SAPHO syndrome. Marzano, AV et al. (74) summarized that in SAPHO syndrome, the activation of PSTPIP2 inflammasomes is thought to contribute to the induction of innate immune system dysfunction. However, there are also different points of view, Hurtado-Nedelec M (75) analyzed the PSTPIP2 gene in patients with SAPHO syndrome. Compared to controls, no specific or more frequent rare variations in this gene were observed in SAPHO patients, indicating no correlation between PSTPIP2 variations and SAPHO syndrome. Therefore, further research is imperative to unravel and elucidate the underlying mechanistic connections between PSTPIP2 and SAPHO syndrome.

5 Liver system diseases

Liver system diseases, encompassing a range of disorders that impair liver function, such as liver injury and hepatitis, represent a complex spectrum of conditions. Liver injury is usually caused by

external forces or viral infections (76, 77), whereas alcoholic liver injury (ALI) is caused by chronic excessive alcohol consumption and manifests as fatigue, anorexia, bloating, and diarrhea (78). Figure 4 illustrates the pathogenic role of PSTPIP2 in liver system diseases. This chapter emphasizes the multifaceted nature of PSTPIP2 in liver disease, suggesting its potential application in therapeutic strategies and the need for further research.

5.1 Alcoholic liver injury

Liver injury, a relatively common condition, affects the liver and is characterized by varying degrees of hepatocyte damage induced by external forces or viral infections (79–82). Alcoholic liver injury (ALI), specifically, arises from prolonged and excessive alcohol consumption, manifesting with symptoms such as fatigue, anorexia, abdominal distension, and diarrhea (83–86). Yin, NN et al. (87) utilized an ethanol (EtOH)-fed mouse model and an EtOH-induced AML-12 cell model to demonstrate that PSTPIP2 regulates hepatocyte apoptosis in ALI through the signal transducer and activator of transcription 3 (STAT3) pathway. Xu, JJ et al. (88) found that in ALI, ethanol induces aberrant methylation of PSTPIP2 and elevates the expression of proteins such as DNMT3a. Furthermore, the silencing of DNMT3a significantly restored ethanol-induced low PSTPIP2 expression and inhibited ethanol-induced inflammation.

5.2 Liver fibrosis and hepatitis

Hepatitis is a diverse range of conditions caused by bacteria, viruses, parasites, alcohol, drugs, chemicals, and autoimmune factors, leading to impaired liver function and abnormal liver function indicators (79, 89–91). PSTPIP2 has been implicated in alleviating hepatic fibrosis and inflammation. For instance, Yang, Y et al. (92) investigated the function of PSTPIP2 in hepatic fibrosis through adeno-associated virus (AAV9)-mediated PSTPIP2 overexpression, investigating the molecular mechanisms underlying PSTPIP2-regulated hepatic fibrosis. They discovered that increased PSTPIP2 expression alleviates hepatic fibrosis and inflammation in mice by modulating macrophage polarization. However, contrasting research suggests that PSTPIP2 may promote the progression of hepatitis C. Chao, TC et al. (93) employed a lentiviral-based RNA interference (RNAi) screening approach to identify PSTPIP2 as a potential cellular factor involved in HCV replication. They further demonstrated the importance of PSTPIP2's membrane-deforming ability in HCV replication, proposing that PSTPIP2 facilitates membrane alterations and participates in the formation of membrane webs, which are crucial for HCV replication complexes. Therefore, a more detailed investigation into the mechanisms of PSTPIP2 in different types of hepatitis is warranted.

6 Renal system diseases

The kidney is a highly susceptible genitourinary organ prone to damage from trauma, spontaneous rupture, iatrogenic injuries, and

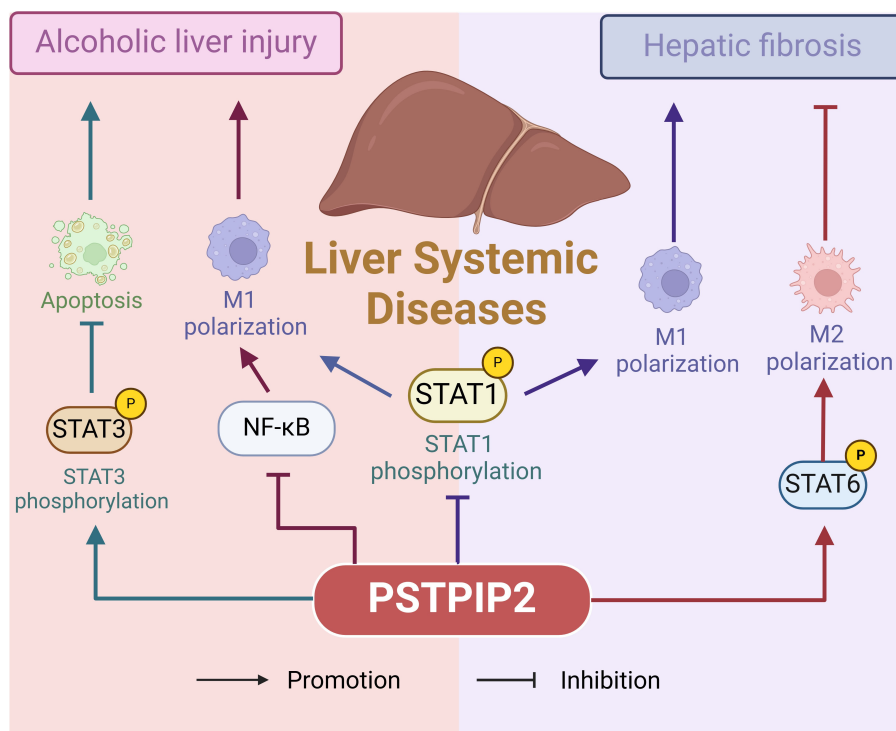


FIGURE 4
The pathogenic role of PSTPIP2 in liver system diseases.

other factors, resulting in symptoms such as hematuria, pain, and shock (94–96). The previous research has shown a close association between PSTPIP2 and inflammatory diseases, with histone deacetylases potentially mediating the expression of PSTPIP2. The pathogenic role of PSTPIP2 in renal systemic diseases is shown in Figure 5. In the context of renal injury, Zhu, H et al. (97) conducted a study to determine the specific role of PSTPIP2 in cisplatin-induced acute kidney injury (AKI), discovering that cisplatin might silence PSTPIP2 through histone acetylation. Xu, CT et al. (98) experimentally found that histone deacetylase (HDAC)-mediated PSTPIP2 silencing may contribute to the development of aristolochic acid nephropathy (AAN). Du, CL et al. (99) demonstrated that neutrophils and neutrophil extracellular traps (NETs) play crucial roles in AAN, and proposed that therapeutic targets targeting PSTPIP2/nuclear factor (NF)- κ B/IL-19/IL-20R β could offer novel strategies for reducing aristolochic acid I-mediated acute kidney injury and apoptosis.

7 Other conditions

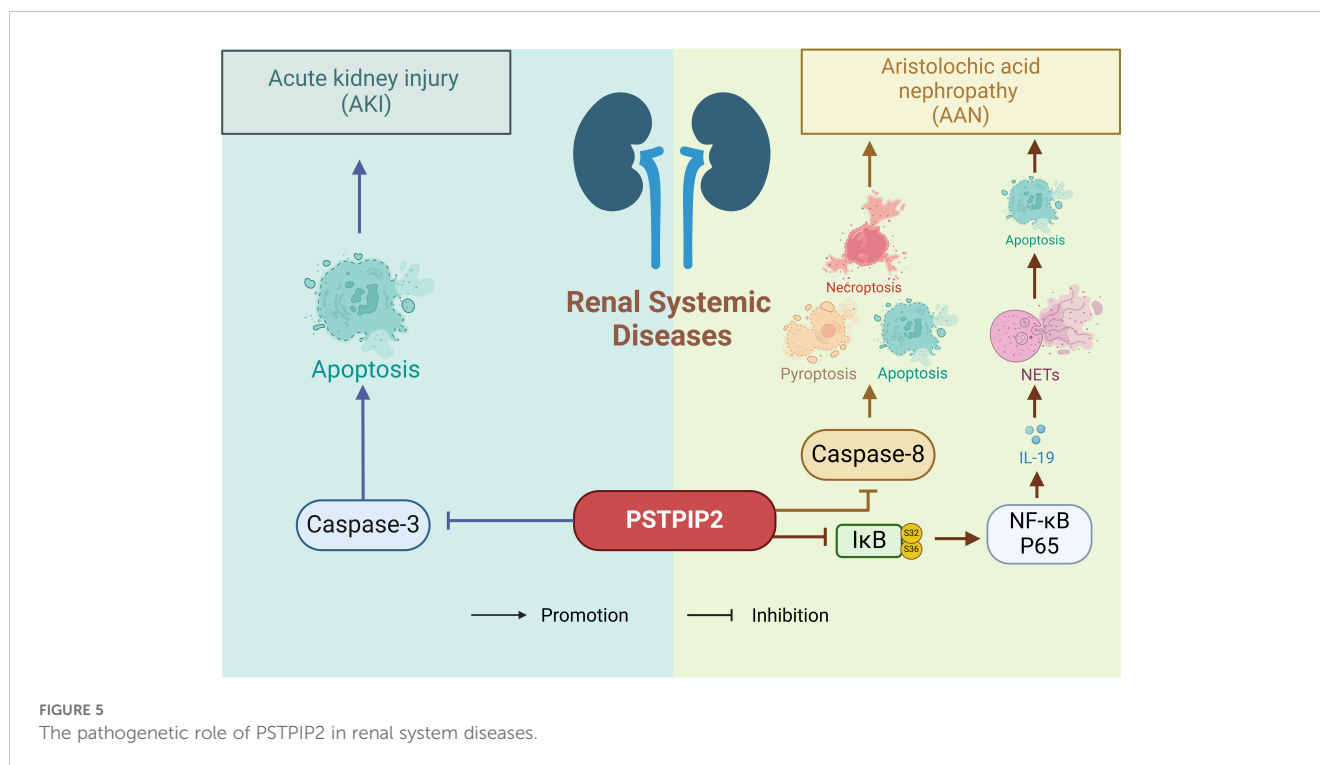
7.1 Pressure sore sepsis

Pressure sore sepsis is a chronic condition characterized by deep skin breakdown accompanied by pus and bleeding (100–102). PSTPIP2 exhibits anti-inflammatory effects in pressure-sore sepsis. Wang, XX et al. (103) compared the expression levels of

PSTPIP2 in peripheral blood samples from 20 patients suffering from sepsis secondary to pressure ulcers and 10 healthy controls. They found that patients with sepsis due to pressure ulcers had lower levels of PSTPIP2 in their peripheral blood. Further, lipopolysaccharide (LPS)-induced THP-1 cells expressed lesser amounts of PSTPIP2 compared to untreated control cells. Additionally, the transfection of PSTPIP2 resulted in reduced levels of IL-6, IL-1 β , and TNF- α , while also inhibiting the activation of the NF- κ B signaling pathway. These findings collectively suggest that PSTPIP2 is associated with the severity of pressure sore sepsis and exerts anti-inflammatory effects, implying potential anti-inflammatory roles of PSTPIP2 in other skin-related inflammatory conditions that remain to be explored.

7.2 Diabetic obesity

Diabetic obesity refers to a condition where individuals have both diabetes and obesity, often leading to increased health risks and complications such as cardiovascular diseases, hypertension, and metabolic disorders (104–106). In the realm of diabetic obesity, PSTPIP2 also alleviates obesity-related tissue inflammation in diabetic mice. Xu, J et al. (107) established a diabetic mouse model through a high-fat diet (HFD) and discovered that PSTPIP2 promotes M2 macrophage polarization via activation of PPAR γ , thereby mitigating obesity-related adipose tissue inflammation and insulin resistance in diabetic mice. These



findings hint at the potential of PSTPIP2 as a therapeutic target for diabetes, presenting a novel therapeutic trajectory for managing this condition.

8 Outlook

This review comprehensively examines the mechanisms of PSTPIP2 in diverse diseases, with a particular emphasis on its advancements in osteomyelitis, arthritis, SAPHO syndrome, hepatic diseases, renal diseases, and other inflammatory conditions. As a pivotal immunomodulatory protein, PSTPIP2 exhibits broad-spectrum immunosuppressive effects through multiple pathways, including inhibition of IL-1 β , modulation of neutrophil and macrophage activity, and regulation of gut microbiota.

While considerable progress has been made in elucidating the mechanisms of PSTPIP2 in various diseases, numerous challenges persist and avenues for future research remain unexplored. Firstly, the intricate mechanisms of PSTPIP2 in diverse diseases remain incompletely understood, particularly its dual and sometimes paradoxical roles in different liver disease manifestations. For instance, while PSTPIP2 exhibits protective effects against acute liver injury (ALI) and fibrosis, it paradoxically facilitates viral replication in hepatitis C. A profound understanding of these mechanisms is paramount for the development of targeted therapeutic strategies that can harness the immunomodulatory potential of PSTPIP2 without exacerbating undesirable effects. Secondly, the intricate relationship between PSTPIP2 and the gut microbiota merits thorough investigation. The gut microbiota serves as a crucial modulator of immune responses, and its intricate

interplay with PSTPIP2 may offer novel insights into potential dietary interventions for disorders related to PSTPIP2 deficiency. By exploring this interaction, there is potential to uncover dietary interventions or probiotics that could be employed to address PSTPIP2 deficiency-related disorders, thereby providing patients with an alternative or adjunctive therapeutic approach. Moreover, the development of PSTPIP2-targeted therapeutics remains a crucial research direction. Currently, there are no commercially available drugs specifically targeting PSTPIP2, despite its promising therapeutic potential across a range of immune-mediated diseases. Progress in this area will necessitate overcoming substantial scientific and technical challenges, including the identification of specific binding sites on PSTPIP2 and the development of molecules capable of effectively modulating its activity without eliciting adverse effects. Additionally, future studies should explore the potential synergies between PSTPIP2-targeted therapies and current treatment modalities for immune-mediated diseases. Specifically, the integration of PSTPIP2 modulators with conventional immunosuppressants could potentially augment treatment efficacy and mitigate the risk of adverse events.

In conclusion, by conducting in-depth investigations into the mechanisms underlying PSTPIP2 and addressing the associated challenges, we can pave the way for innovative therapeutic strategies for a diverse array of immune-mediated diseases. These endeavors will not only elevate treatment efficacy and enhance patients' quality of life but also contribute significantly to a deeper understanding of the intricate interplay between the immune system and various diseases. This holistic approach is imperative for advancing the field of immunomodulatory therapies and improving clinical outcomes for patients with immune-mediated disorders.

Author contributions

SG: Writing – original draft. BH: Writing – original draft. YG: Writing – original draft. YJ: Writing – original draft. ZS: Writing – original draft. GH: Writing – review & editing. CL: Writing – review & editing, Methodology, Conceptualization, Validation, Funding acquisition, Visualization.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This work was supported by grants from the National Natural Science Foundation of China (82074246, 82374272); The Fundamental Research Funds for the Central Universities (2023-JYB-XJSJ-013); Beijing University of Traditional Chinese Medicine College Student Innovation and Entrepreneurship Training Program Project (202410026006, X202410026077); Beijing University Student Innovation and Entrepreneurship Training Inter school Cooperation Program (202498132).

References

1. Yao Y, Cai XY, Yu HX, Xu QQ, Li XF, Yang Y, et al. PSTPIP2 attenuates joint damage and suppresses inflammation in adjuvant-induced arthritis. *Eur J Pharmacol.* (2019) 859:172558. doi: 10.1016/j.ejphar.2019.172558
2. Phillips FC, Gurung P, Kanneganti TD. Microbiota and caspase-1/caspase-8 regulate IL-1 β -mediated bone disease. *Gut Microbes.* (2016) 7:334–41. doi: 10.1080/19490976.2016.1182289
3. Yao Y, Yu HX, Liu YR, Xu QQ, Li XF, Meng XM, et al. PSTPIP2 inhibits the inflammatory response and proliferation of fibroblast-like synoviocytes. *Front Pharmacol.* (2018) 9. doi: 10.3389/fphar.2018.01432
4. Su RJ, Sun M, Wang W, Zhang JL, Zhang L, Zhen JL, et al. A novel immunosuppressor, (5R)-5-hydroxytryptolide, alleviates movement disorder and neuroinflammation in a 6-OHDA hemiparkinsonian rat model. *Aging Dis.* (2017) 8:31–43. doi: 10.14336/AD.2016.0929
5. Albiñana V, Sanz-Rodríguez F, Recio-Poveda L, Bernabéu C, Botella LM. Immunosuppressor FK506 increases endoglin and activin receptor-like kinase 1 expression and modulates transforming growth factor- β 1 signaling in endothelial cells. *Mol Pharmacol.* (2011) 79:833–43. doi: 10.1124/mol.110.067447
6. Liu XL, Liu TL, Xiao YF. Protective effect of immunosuppressor FTY720 against CVB3 induced-viral myocarditis. *Eur Heart J.* (2017) 38:234–. doi: 10.1093/eurheartj/ehx502.P1110
7. Biggoggero M, Gabbriellini L, Meroni PL. Type I interferon therapy and its role in autoimmunity. *Autoimmunity.* (2010) 43:248–54. doi: 10.3109/08916930903510971
8. Krainer J, Siebenhandl S, Weinhäusel A. Systemic autoinflammatory diseases. *J Autoimmun.* (2020) 109:102421. doi: 10.1016/j.jaut.2020.102421
9. Loayza M, Carter K, Pang Y, Bhatt AL. Altered fetal microglia phenotypes are associated with abnormal neurogenesis following maternal immune activation. *J Invest Med.* (2020) 68(2):584–85. doi: 10.1136/jim-2020-SRM.374
10. Kralova J, Drobek A, Prochazka J, Spoutil F, Fabisik M, Glatzova D, et al. Dysregulated NADPH oxidase promotes bone damage in murine model of autoinflammatory osteomyelitis. *J Immunol.* (2020) 204:1607–20. doi: 10.4049/jimmunol.1900953
11. Zhu CS, Sun B, Nie AZ, Zhou Z. The tRNA-associated dysregulation in immune responses and immune diseases. *Acta Physiol.* (2020) 228:e13391. doi: 10.1111/apha.v228.2
12. de Jesus AA, Goldbach-Mansky R. Genetically defined autoinflammatory diseases. *Oral Dis.* (2016) 22:591–604. doi: 10.1111/odi.2016.22.issue-7
13. Blank M, Shoenfeld Y. B cell targeted therapy in autoimmunity. *J Autoimmun.* (2007) 28:62–8. doi: 10.1016/j.jaut.2007.02.001
14. Yasmeen F, Pirzada RH, Ahmad B, Choi B, Choi S. Understanding autoimmunity: mechanisms, predisposing factors, and cytokine therapies. *Int J Mol Sci.* (2024) 25:7666. doi: 10.3390/ijms25147666

Acknowledgments

We would like to express our sincere gratitude to [BioRender.com](https://www.biorender.com) and Figdraw for providing the platform to draw the scheme. Thanks to ERNIE Bot (yiyian. baidu. com, V3.3.0) for the helps in language polishing.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

15. Hosseinalizadeh H, Rabiee F, Eghbalifard N, Rajabi H, Kliionsky DJ, Rezaee A. Regulating the regulatory T cells as cell therapies in autoimmunity and cancer. *Front Med-Lausanne.* (2023) 10. doi: 10.3389/fmed.2023.1244298
16. Wang LF, Wang FS, Gershwin ME. Human autoimmune diseases: a comprehensive update. *J Intern Med.* (2015) 278:369–95. doi: 10.1111/joim.2015.278.issue-4
17. Monzani F, Caraccio N, Dardano A, Ferrannini E. Thyroid autoimmunity and dysfunction associated with type I interferon therapy. *Clin Exp Med.* (2004) 3:199–210. doi: 10.1007/s10238-004-0026-3
18. Krysiak R, Szkrobka W, Okopien B. The effect of ezetimibe/statin combination and high-dose statin therapy on thyroid autoimmunity in women with hashimoto's thyroiditis and cardiovascular disease: A pilot study. *Exp Clin Endocr Diab.* (2016) 124(9):577–81. doi: 10.1055/s-0042-113872
19. Shoenfeld Y. The future of autoimmunity. *Clin Rev Allerg Immun.* (2012) 42:113–20. doi: 10.1007/s12016-011-8297-4
20. Schmitt SK. Osteomyelitis. *Infect Dis Clin N Am.* (2017) 31:325. doi: 10.1016/j.idc.2017.01.010
21. Wang XK, Zhang MR, Zhu TT, Wei QH, Liu GY, Ding JX. Flourishing antibacterial strategies for osteomyelitis therapy. *Adv Sci.* (2023) 10:2206154. doi: 10.1002/advs.202206154
22. Kavanagh N, Ryan EJ, Widaa A, Sexton G, Fennell J, O'Rourke S, et al. Staphylococcal osteomyelitis: disease progression, treatment challenges, and future directions. *Clin Microbiol Rev.* (2018) 31:e00084–17. doi: 10.1128/CMR.00084-17
23. Acierio S, Angrisani F, Marino A, Caporali RF, Cimaz R, Giani T. Canakinumab treatment in a young girl with refractory chronic recurrent multifocal osteomyelitis associated with pyoderma gangrenosum. *Int J Rheum Dis.* (2022) 25(11):1333–8. doi: 10.1111/1756-185X.14425
24. El-Shanti HI, Ferguson PJ. Chronic recurrent multifocal osteomyelitis - A concise review and genetic update. *Clin Orthop Relat R.* (2007) 462:11–9. doi: 10.1097/BLO.0b013e3180986d73
25. Stojkic I, Rubin E, Lee JY, Lee ASE, Ogbu EA. An unusual presentation of chronic recurrent multifocal osteomyelitis involving bones of the skull. *Int J Rheum Dis.* (2023) 26(8):1579–81. doi: 10.1111/1756-185X.14629
26. Yang CJ, Yoshida T, Sugimoto S, Teranishi M, Sone M. Facial nerve palsy associated with chronic recurrent multifocal osteomyelitis. *Pediatr Int.* (2021) 63:1125–7. doi: 10.1111/ped.14563
27. Ssi-Yan-Kai G, Brun N, Hamida K, Marson V, Castelain CS, Marson F. Chronic recurrent multifocal osteomyelitis: A case report in adult. *J Radiol.* (2011) 92:1131–4. doi: 10.1016/j.jradio.2011.05.016
28. Ferguson PJ, Bing XY, Vasef MA, Ochoa LA, Mahgoub A, Waldschmidt TJ, et al. A missense mutation in pstpip2 is associated with the murine autoinflammatory

- disorder chronic multifocal osteomyelitis. *Bone*. (2006) 38:41–7. doi: 10.1016/j.bone.2005.07.009
29. Chen TC, Wu JJS, Chang WP, Hsu PN, Hsieh ST, Shyu BC. Spontaneous inflammatory pain model from a mouse line with Ni-ethyl-N-nitrosourea mutagenesis. *J BioMed Sci*. (2012) 19(1):55. doi:10.1186/1423-0127-19-55
30. Chitu V, Ferguson PJ, de Bruijn R, Schlueter AJ, Ochoa LA, Waldschmidt TJ, et al. Primed innate immunity leads to autoinflammatory disease in PSTPIP2-deficient mice. *Blood*. (2009) 114:2497–505. doi: 10.1182/blood-2009-02-204925
31. Drobek A, Kralova J, Skopcová T, Kucova M, Novák P, Angelisová P, et al. PSTPIP2, a protein associated with autoinflammatory disease, interacts with inhibitory enzymes SHIP1 and csk. *J Immunol*. (2015) 195:3416–26. doi: 10.4049/jimmunol.1401494
32. Gurung P, Burton A, Kanneganti TD. NLRP3 inflammasome plays a redundant role with caspase 8 to promote IL-1 β -mediated osteomyelitis. *P Natl Acad Sci USA*. (2016) 113:4452–7. doi: 10.1073/pnas.1601636113
33. Cassel SL, Janczy JR, Bing XY, Wilson SP, Olivier AK, Otero JE, et al. Inflammasome-independent IL-1 β mediates autoinflammatory disease in Pstpip2-deficient mice. *P Natl Acad Sci USA*. (2014) 111:1072–7. doi: 10.1073/pnas.1318685111
34. Lukens JR, Gross JM, Calabrese C, Iwakura Y, Lamkanfi M, Vogel P, et al. Critical role for inflammasome-independent IL-1 β production in osteomyelitis. *P Natl Acad Sci USA*. (2014) 111:1066–71. doi: 10.1073/pnas.1318688111
35. Lukens JR, Gurung P, Vogel P, Johnson GR, Carter RA, McGoldrick DJ, et al. Dietary modulation of the microbiome affects autoinflammatory disease. *Nature*. (2014) 516:246. doi: 10.1038/nature13788
36. Pavliuchenko N, Duric I, Kralova J, Fabisik M, Spoutil F, Prochazka J, et al. Molecular interactions of adaptor protein PSTPIP2 control neutrophil-mediated responses leading to autoinflammation. *Front Immunol*. (2022) 13. doi: 10.3389/fimmu.2022.1035226
37. Hao Y, Xu Z, Xueping L, Shijie F. The effect of Duhuo Parasitic Decoction on knee joint function, pain, and serum inflammatory factors in patients with knee osteoarthritis. *Chin Arch Traditional Chin Med*, 1–8. Available online at: <https://link.cnki.net/urlid/21.1546.r.20240828.1623.019>.
38. Barbour KE, Helmick CG, Boring M, Brady TJ. Vital signs: prevalence of doctor-diagnosed arthritis and arthritis-attributable activity limitation - United States, 2013–2015. *MMWR-Morbidity Mortal W*. (2017) 66:246–53. doi: 10.15585/mmwr.mm6609e1
39. Hootman JM, Helmick CG, Barbour KE, Theis KA, Boring MA. Updated projected prevalence of self-reported doctor-diagnosed arthritis and arthritis-attributable activity limitation among US adults, 2015–2040. *Arthritis Rheumatol*. (2016) 68:1582–7. doi: 10.1002/art.39692
40. Tajik N, Frech M, Schulz O, Schäler F, Lucas S, Azizov V, et al. Targeting zonulin and intestinal epithelial barrier function to prevent onset of arthritis. *Nat Commun*. (2020) 11(1):1995. doi: 10.1038/s41467-020-15831-7
41. Chitu V, Nacu V, Charles JF, Henne WM, McMahon HT, Nandi S, et al. PSTPIP2 deficiency in mice causes osteopenia and increased differentiation of multipotent myeloid precursors into osteoclasts. *Blood*. (2012) 120:3126–35. doi: 10.1182/blood-2012-04-425595
42. Tsujita K, Kondo A, Kurisu S, Hasegawa J, Itoh T, Takenawa T. Antagonistic regulation of F-BAR protein assemblies controls actin polymerization during podosome formation. *J Cell Sci*. (2013) 126:2267–78. doi: 10.1242/jcs.122515
43. Sztacho M, Segelitz S, Sanchez-Fernandez MA, Czupalla C, Niehage C, Hoflack B. BAR proteins PSTPIP1/2 regulate podosome dynamics and the resorption activity of osteoclasts. *PLoS One*. (2016) 11(10):e0164829. doi: 10.1371/journal.pone.0164829
44. Verheul MK, Fearon U, Trouw LA, Veale DJ. Biomarkers for rheumatoid and psoriatic arthritis. *Clin Immunol*. (2015) 161:2–10. doi: 10.1016/j.clim.2015.04.005
45. Wang Y, Chen SJ, Du KZ, Liang CX, Wang SQ, Boadi EO, et al. Traditional herbal medicine: Therapeutic potential in rheumatoid arthritis. *J Ethnopharmacol*. (2021) 279:114368. doi: 10.1016/j.jep.2021.114368
46. England BR, Thiele GM, Anderson DR, Mikuls TR. Increased cardiovascular risk in rheumatoid arthritis: mechanisms and implications. *Bmj-Brit Med J*. (2018) 361: k1036. doi: 10.1136/bmj.k1036
47. Burmester GR, Pope JE. Novel treatment strategies in rheumatoid arthritis. *Lancet*. (2017) 389:2338–48. doi: 10.1016/S0140-6736(17)31491-5
48. Lu H, Lu X, Xie QH, Wan HL, Sun YY. TTC4 inhibits NLRP3 inflammation in rheumatoid arthritis by HSP70. *Int J Rheum Dis*. (2023) 26(9):1751–9. doi: 10.1111/1756-185X.14818
49. Tureson C, Matteson EL. Management of extra-articular disease manifestations in rheumatoid arthritis. *Curr Opin Rheumatol*. (2004) 16:206–11. doi: 10.1097/00002281-200405000-00007
50. Jang S, Kwon EJ, Lee JJ. Rheumatoid arthritis: pathogenic roles of diverse immune cells. *Int J Mol Sci*. (2022) 23(2):905. doi: 10.3390/ijms23020905
51. Bullock J, Rizvi SA, Saleh AM, Ahmed SS, Do DP, Ansari RA, et al. Rheumatoid arthritis: A brief overview of the treatment. *Med Prin Pract*. (2018) 27:501–7. doi: 10.1159/000493930
52. Li X, Tong XN, Yeung WWY, Kuan P, Yum SHH, Chui CSL, et al. Two-dose COVID-19 vaccination and possible arthritis flare among patients with rheumatoid arthritis in Hong Kong. *Ann Rheum Dis*. (2022) 81:564–8. doi: 10.1136/annrheumdis-2021-221571
53. Liang QC, Tian YJ, Liu ZJ, Yu DJ, Guo HB, Sun FL. Co-culture of bone marrow mesenchymal stem cells and fibroblast-like synoviocytes (RA-FLS) alleviates rheumatoid arthritis cell apoptosis by inhibiting inflammatory response. *J Biomater Tissue Engineering*. (2023) 13:294–300. doi: 10.1166/jbt.2023.3254
54. Wu ZW, Ma D, Yang HL, Gao JF, Zhang GL, Xu K, et al. Fibroblast-like synoviocytes in rheumatoid arthritis: Surface markers and phenotypes. *Int Immunopharmacol*. (2021) 93:107392. doi: 10.1016/j.intimp.2021.107392
55. Nygaard G, Firestein GS. Restoring synovial homeostasis in rheumatoid arthritis by targeting fibroblast-like synoviocytes. *Nat Rev Rheumatol*. (2020) 16:316–33. doi: 10.1038/s41584-020-0413-5
56. Lin J, Liu BX, Zou HM. Inhibition of miR-5571-3p attenuates proliferation and invasion in rheumatoid arthritis fibroblast-like synoviocytes by upregulating OAZ3. *Indian J Pharm Sci*. (2020) 82:119. doi: 10.36468/pharmaceutical-sciences.spl.110
57. Grosse J, Chitu V, Marquardt A, Hanke P, Schmittwolf C, Zeitlmann L, et al. Mutation of mouse Myp/Pstpip2 causes a macrophage autoinflammatory disease. *Blood*. (2006) 107:3350–8. doi: 10.1182/blood-2005-09-3556
58. Yao Y, Cai XY, Zhang M, Zhang X, Ren FJ, Zheng Y, et al. PSTPIP2 regulates synovial macrophages polarization and dynamics via ER β in the joint microenvironment. *Arthritis Res Ther*. (2022) 24(1):247. doi: 10.1186/s13075-022-02939-y
59. Matsunaga M, Lim E, Davis J, Chen JJ. Dietary quality associated with self-reported diabetes, osteoarthritis, and rheumatoid arthritis among younger and older US adults: A cross-sectional study using NHANES 2011–2016. *Nutrients*. (2021) 13(2):545. doi: 10.3390/nu13020545
60. Mohajer B, Moradi K, Dolatshahi M, Guermazi A, Najafzadeh N, Roemer F, et al. Changes in thigh muscle quality and their association with knee osteoarthritis progression in diabetic patients: A longitudinal study on osteoarthritis initiative data. *Osteoarthr Cartilage*. (2022) 30:S43–S4. doi: 10.1016/j.joca.2022.02.046
61. Chan DC, Chiu CY, Lan KC, Weng TI, Yang RS, Liu SH. Transplantation of human skeletal muscle-derived progenitor cells ameliorates knee osteoarthritis in streptozotocin-induced diabetic mice. *J Orthop Res*. (2017) 35:1886–93. doi: 10.1002/jor.23503
62. Uebelhart D, Branco J, Bruyere O, Chapurlat RD, Cooper C, Dennison EM, et al. Management of osteoarthritis in diabetic patients. *Osteoporosis Int*. (2019) 30(2): S192–S192.
63. Wang HJ, Giambini H, Chen JW, Wang QS, Hou HG, Luo SM, et al. Diabetes mellitus accelerates the progression of osteoarthritis in streptozotocin-induced diabetic mice by deteriorating bone microarchitecture, bone mineral composition, and bone strength of subchondral bone. *Ann Transl Med*. (2021) 9(9):768. doi: 10.21037/atm-20-6797
64. Arce NR, Muruges D, Hum N, Sebastian A, Jbeily E, Christiansen B, et al. Diabetes promotes mild osteoarthritis in the streptozotocin-induced diabetic mouse model. *FASEB J*. (2021) 35(1). doi:10.1096/fasebj.2021.35.S1.02490
65. Korpar I, Frangez I. Preservation surgery of septic osteoarthritis and osteomyelitis in the diabetic foot using S53P4 bioactive glass-A case series. *Appl Sci-Basel*. (2023) 13(5):3310. doi: 10.3390/app13053310
66. Li M, Xiao YB, Wang XT, Zhuang JP, Zhou CL. Proline-serine-threonine phosphatase-interacting protein 2 alleviates diabetes mellitus-osteoarthritis in rats through attenuating synovial inflammation and cartilage injury. *Orthop Surg*. (2021) 13:1398–407. doi: 10.1111/os.13000
67. Liu L, Wen Q, Gong R, Gilles L, Stankiewicz MJ, Li W, et al. dysregulation contributes to aberrant terminal differentiation in GATA-1-deficient megakaryocytes by activating LYN. *Cell Death Dis*. (2014) 5:e988. doi: 10.1038/cddis.2013.512
68. Van den Eynde M, Lecluyse K, Chioccoli C, Brouckaert M, Caussin E, Lammens P. Crohn's disease and the SAPHO syndrome during treatment with infliximab: a case report and review of literature. *Gastroen Clin Biol*. (2007) 31(6-7):607–10. doi: 10.1016/S0399-8320(07)89438-3
69. Nissen MJ, Syrogiannopoulou A, Gabay C. Coexistence of seropositive rheumatoid arthritis and SAPHO syndrome. *Joint Bone Spine*. (2013) 80:674–6. doi: 10.1016/j.jbspin.2013.03.020
70. Jiang HX, Chen NG, Li C. SAPHO syndrome can cause masseter muscle involvement. *Int J Rheum Dis*. (2023) 26(2):398–400. doi: 10.1111/1756-185X.14513
71. Cao FJ, Hou XJ, Kang TL, Shi XJ, Ma WG, Zhang Y, et al. SAPHO syndrome complicated with relapsing polychondritis: A case report. *Int J Rheum Dis*. (2023) 26(10):2060–3. doi: 10.1111/1756-185X.14693
72. Misiak-Galazka M, Jeziorkowska R, Sikorska-Siudek K, Wolska H. SAPHO syndrome: case report. *Postep Derm Alergol*. (2012) 29(2):136–8.
73. Liao HJ, Chyuan IT, Wu CS, Lin SW, Chen KH, Tsai HF, et al. Increased neutrophil infiltration, IL-1 production and a SAPHO syndrome-like phenotype in PSTPIP2-deficient mice. *Rheumatology*. (2015) 54:1317–26. doi: 10.1093/rheumatology/keu481
74. Marzano AV, Borghi A, Meroni PL, Cugno M. Pyoderma gangrenosum and its syndromic forms: evidence for a link with autoinflammation. *Brit J Dermatol*. (2016) 175:882–91. doi: 10.1111/bjd.2016.175.issue-5
75. Hurtado-Nedelec M, Chollet-Martin S, Chapeton D, Hugot JP, Hayem G, Gérard B. Genetic susceptibility factors in a cohort of 38 patients with SAPHO syndrome: A study of PSTPIP2, NOD2, and LPIN2 genes. *J Rheumatol*. (2010) 37:401–9. doi: 10.3899/jrheum.090456

76. Yan MZ, Huo YZ, Yin ST, Hu HB. Mechanisms of acetaminophen-induced liver injury and its implications for therapeutic interventions. *Redox Biol.* (2018) 17:274–83. doi: 10.1016/j.redox.2018.04.019
77. Huang TW, Zhang K, Wang JQ, He KH, Zhou XT, Nie SP. Quercetin alleviates acrylamide-induced liver injury by inhibiting autophagy-dependent ferroptosis. *J Agr Food Chem.* (2023) 71:7427–39. doi: 10.1021/acs.jafc.3c01378
78. Li HZ, Shi JL, Zhao L, Guan JQ, Liu F, Huo GC, et al. KLDS1.0344 and KLDS1.0901 mixture prevents chronic alcoholic liver injury in mice by protecting the intestinal barrier and regulating gut microbiota and liver-related pathways. *J Agr Food Chem.* (2021) 69:183–97. doi: 10.1021/acs.jafc.0c06346
79. Arefhosseini S, Aghajani T, Tutunchi H, Ebrahimi-Mameghani M. Association of systemic inflammatory indices with anthropometric measures, metabolic factors, and liver function in non-alcoholic fatty liver disease. *Sci Rep-Uk.* (2024) 14(1):12829. doi: 10.1038/s41598-024-63381-5
80. Duarte S, Saber J, Fujii T, Coito AJ. Matrix metalloproteinases in liver injury, repair and fibrosis. *Matrix Biol.* (2015) 44-46:147–56. doi: 10.1016/j.matbio.2015.01.004
81. McGill MR. The past and present of serum aminotransferases and the future of liver injury biomarkers. *Excli J.* (2016) 15:817–28. doi: 10.17179/excli2016-800
82. Huang H, Tohme S, Al-Khafaji AB, Tai S, Loughran P, Chen L, et al. Damage-associated molecular pattern-activated neutrophil extracellular trap exacerbates sterile inflammatory liver injury. *Hepatology.* (2015) 62:600–14. doi: 10.1002/hep.27841
83. Jung JC, Lee YH, Kim SH, Kim KJ, Kim KM, Oh S, et al. Hepatoprotective effect of licorice, the root of *Glycyrrhiza uralensis* Fischer, in alcohol-induced fatty liver disease. *BMC Complem Altern M.* (2016) 16:19. doi:10.1186/s12906-016-0997-0
84. Yang YM, Cho YE, Hwang S. Crosstalk between oxidative stress and inflammatory liver injury in the pathogenesis of alcoholic liver disease. *Int J Mol Sci.* (2022) 23(2):774. doi:10.3390/ijms23020774
85. Louvet A, Mathurin P. Alcoholic liver disease: mechanisms of injury and targeted treatment. *Nat Rev Gastro Hepat.* (2015) 12:231–42. doi: 10.1038/nrgastro.2015.35
86. Wrzosek L, Ciocan D, Hugot C, Spatz M, Dupoux M, Houron C, et al. Microbiota tryptophan metabolism induces aryl hydrocarbon receptor activation and improves alcohol-induced liver injury. *Gut.* (2021) 70:1299–308. doi: 10.1136/gutjnl-2020-321565
87. Yin NN, Chen X, Sun YY, Yang L, Zhang YF, Niu XN, et al. PSTPIP2 protects against alcoholic liver injury and invokes STAT3-mediated suppression of apoptosis. *Biochem Pharmacol.* (2024) 225:116334. doi: 10.1016/j.bcp.2024.116334
88. Xu JJ, Zhu L, Li HD, Du XS, Li JJ, Yin NN, et al. DNMT3a-mediated methylation of PSTPIP2 enhances inflammation in alcohol-induced liver injury via regulating STAT1 and NF- κ B pathway. *Pharmacol Res.* (2022) 177:106125. doi: 10.1016/j.phrs.2022.106125
89. Rivero-Juarez A, Frias M, Perez AB, Pineda JA, Reina G, Fuentes-Lopez A, et al. C infection as an emerging cause of acute hepatitis in Spain: First report in Europe. *J Hepatol.* (2022) 77:326–31. doi: 10.1016/j.jhep.2022.01.028
90. Heymann F, Hamesch K, Weiskirchen R, Tacke F. The concanavalin A model of acute hepatitis in mice. *Lab Anim-Uk.* (2015) 49:12–20. doi: 10.1177/0023677215572841
91. Mack CL, Adams D, Assis DN, Kerker N, Manns MP, Mayo MJ, et al. Diagnosis and management of autoimmune hepatitis in adults and children: 2019 practice guidance and guidelines from the american association for the study of liver diseases. *Hepatology.* (2020) 72:671–722. doi: 10.1002/hep.31065
92. Yang Y, Wu XQ, Li WX, Huang HM, Li HD, Pan XY, et al. PSTPIP2 connects DNA methylation to macrophage polarization in CCL4-induced mouse model of hepatic fibrosis. *Oncogene.* (2018) 37:6119–35. doi: 10.1038/s41388-018-0383-0
93. Chao TC, Su WC, Huang JY, Chen YC, Jeng KS, Wang HD, et al. Proline-serine-threonine phosphatase-interacting protein 2 (PSTPIP2), a host membrane-deforming protein, is critical for membranous web formation in hepatitis C virus replication. *J Virol.* (2012) 86:1739–49. doi: 10.1128/JVI.06001-11
94. Armstrong LB, Mooney DP. Pediatric renal injury: which injury grades warrant close follow-up. *Pediatr Surg Int.* (2018) 34:1183–7. doi: 10.1007/s00383-018-4355-9
95. ter Maaten JM, Damman K, Verhaar MC, Paulus WJ, Duncker DJ, Cheng C, et al. Connecting heart failure with preserved ejection fraction and renal dysfunction: the role of endothelial dysfunction and inflammation. *Eur J Heart Fail.* (2016) 18:588–98. doi: 10.1002/ejhf.2016.18.issue-6
96. van Swelm RPL, Wetzels JFM, Swinkels DW. The multifaceted role of iron in renal health and disease. *Nat Rev Nephrol.* (2020) 16:77–98. doi: 10.1038/s41581-019-0197-5
97. Zhu H, Jiang WJ, Zhao HZ, He CS, Tang XH, Xu SB, et al. PSTPIP2 inhibits cisplatin-induced acute kidney injury by suppressing apoptosis of renal tubular epithelial cells. *Cell Death Dis.* (2020) 11(12):1057. doi: 10.1038/s41419-020-03267-2
98. Xu CT, Wang Q, Du CL, Chen L, Zhou ZN, Zhang ZM, et al. Histone deacetylase-mediated silencing of PSTPIP2 expression contributes to aristolochic acid nephropathy-induced PANoptosis. *Brit J Pharmacol.* (2024) 181:1452–73. doi: 10.1111/bph.v181.9
99. Du CL, Xu CT, Jia PC, Cai N, Zhang ZM, Meng WN, et al. PSTPIP2 ameliorates aristolochic acid nephropathy by suppressing interleukin-19-mediated neutrophil extracellular trap formation. *Elife.* (2024) 13:e89740. doi: 10.7554/eLife.89740.sa2
100. Ma Q, Gao XL, Xiao Y, Ke XS. Multivariate analysis and nursing intervention strategies for intraoperative pressure ulcers in patients undergoing craniocerebral microsurgery. *Asian J Surg.* (2024) 47:233–6. doi: 10.1016/j.asjsur.2023.08.025
101. Lemyze M, Mallat J, Vangrunderbeek N, Granier M. Under-the-nose versus over-the-nose face mask to prevent facial pressure sores during face mask-delivered noninvasive ventilation for acute hypercapnic respiratory failure: A randomized controlled trial. *Crit Care Med.* (2023) 51:1177–84. doi: 10.1097/CCM.0000000000005902
102. Dong DX, Lv XL, Jiang QS, Zhang JJ, Gu ZY, Yu WM, et al. Multifunctional electrospun polycaprolactone/chitosan/hEGF/lidocaine nanofibers for the treatment of 2 stage pressure ulcers. *Int J Biol Macromol.* (2024) 256(2):128533. doi: 10.1016/j.jbiomac.2023.128533
103. Wang XX, Wang YL, Luo JJ, Wang LP, Guo LY, Zhu XX. PSTPIP2 is associated with disease severity in patients with pressure ulcer sepsis and has anti-inflammatory effects. *Allergol Immunopath.* (2023) 51:23–8. doi: 10.15586/aei.v51i5.939
104. Chen JH, Li YT, Niu ZM, He ZP, Xie YJ, Hernandez J, et al. Investigating the causal association of generalized and abdominal obesity with microvascular complications in patients with type 2 diabetes: A community-based prospective study. *Diabetes Obes Metab.* (2024) 26:2796–810. doi: 10.1111/dom.15598
105. Ahmad KA, Prakash AS, Chandra A, Jha RK. Randomized controlled trial assessing PBI-4547's impact on glycemic control, diabetic nephropathy, and obesity in type 2 diabetes mellitus patients. *Diabetes.* (2024) 73(1):1134–P. doi: 10.2337/db24-1134-P
106. Chen ZY, Zhong XJ, Lin RY, Liu SL, Cao H, Chen HJ, et al. Type 2 diabetes: is obesity for diabetic retinopathy good or bad? A cross-sectional study. *Nutr Metab.* (2024) 21(1):68. doi: 10.1186/s12986-024-00842-8
107. Xu J, Lv HY. PSTPIP2 alleviates obesity associated adipose tissue inflammation and insulin resistance in diabetes mice through promoting M2 macrophage polarization via activation of PPAR γ . *J Diabetes Complicat.* (2023) 37(6):108479. doi: 10.1016/j.jdiacomp.2023.108479

Glossary

SAPHO	synovitis-acne-pustulosis-hyperostosis-osteitis	CMO	chronic multifocal osteomyelitis
RA	rheumatoid arthritis	FLS	fibroblast-like synoviocytes
ROS	reactive oxygen species	DM-OA	Diabetic osteoarthritis
PSTPIP2	proline-serine-threonine phosphatase-interacting protein 2	ALI	Alcoholic liver injury
F-BAR	Fes/CIP4 homology-Bin/Amphiphysin/Rvs	STAT3	signal transducer and activator of transcription 3
CAO	chronic aseptic osteomyelitis	AAV9	adeno-associated virus
ENU	N-ethyl-N-nitrosourea	RNAi	RNA interference
SNP	single nucleotide polymorphism	AKI	acute kidney injury
CRMO	chronic recurrent multifocal osteomyelitis	HDAC	histone deacetylase
MCP-1	monocyte chemoattractant protein-1	AAN	aristolochic acid nephropathy
CSK	C-terminal Src kinase	NETs	neutrophil extracellular traps
ERK	extracellular signal-regulated kinase	LPS	lipopolysaccharide
PEST-PTPs	protein tyrosine phosphatases containing a PEST domain	HFD	high-fat diet
SHIP1	Src homology 2 domain-containing inositol-5'-phosphatase 1	IL-1Ra	IL-1 receptors
CSK	C-terminal Src kinase		