Check for updates

OPEN ACCESS

EDITED BY Haoyu Liu, Yangzhou University, China

REVIEWED BY Anders Johansson, Umeå University, Sweden Hongfei Li, Zhejiang Ocean University, China

*CORRESPONDENCE Xingwen Xie X q13309329949@163.com

RECEIVED 17 August 2024 ACCEPTED 09 September 2024 PUBLISHED 26 September 2024

CITATION

Qi P, Chen X, Tian J, Zhong K, Qi Z, Li M and Xie X (2024) The gut homeostasis-immune system axis: novel insights into rheumatoid arthritis pathogenesis and treatment. *Front. Immunol.* 15:1482214. doi: 10.3389/fimmu.2024.1482214

COPYRIGHT

© 2024 Qi, Chen, Tian, Zhong, Qi, Li and Xie. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

The gut homeostasis-immune system axis: novel insights into rheumatoid arthritis pathogenesis and treatment

Peng Qi¹, Xin Chen², Jiexiang Tian², Kexin Zhong¹, Zhonghua Qi¹, Menghan Li¹ and Xingwen Xie^{1,2*}

¹Gansu University of Traditional Chinese Medicine, Lanzhou, China, ²Affiliated Hospital of Gansu University of Traditional Chinese Medicine, Lanzhou, China

Rheumatoid arthritis is a widely prevalent autoimmune bone disease that imposes a significant burden on global healthcare systems due to its increasing incidence. In recent years, attention has focused on the interaction between gut homeostasis and the immune system, particularly in relation to bone health. Dysbiosis, which refers to an imbalance in the composition and function of the gut microbiota, has been shown to drive immune dysregulation through mechanisms such as the release of pro-inflammatory metabolites, increased gut permeability, and impaired regulatory T cell function. These factors collectively contribute to immune system imbalance, promoting the onset and progression of Rheumatoid arthritis. Dysbiosis induces both local and systemic inflammatory responses, activating key pro-inflammatory cytokines such as tumor necrosis factor-alpha, Interleukin-6, and Interleukin-17, which exacerbate joint inflammation and damage. Investigating the complex interactions between gut homeostasis and immune regulation in the context of Rheumatoid arthritis pathogenesis holds promise for identifying new therapeutic targets, revealing novel mechanisms of disease progression, and offering innovative strategies for clinical treatment.

KEYWORDS

gut homeostasis, immune system, rheumatoid arthritis, pathogenesis, novel therapeutic approaches

1 Introduction

Rheumatoid arthritis (RA) is a multifaceted, chronic autoimmune disorder characterized by persistent inflammation across multiple joints, leading to joint damage and functional impairment. This condition is typified by the production of autoantibodies, chronic synovitis, and sustained inflammation (1). Affecting approximately 1% of the global population, RA exhibits a higher prevalence among females (2). Hallmark manifestations include widespread joint swelling, tenderness, and systemic inflammation

driven by autoantibodies (2). The inflammatory pathways in RA are marked by alterations in the T helper 1 cell profile, which result in an imbalance between anti-inflammatory and pro-inflammatory cytokines (2). Autoantibodies linked to RA, such as rheumatoid factor, anti-citrullinated protein antibodies, and anti-carbamylated antibodies, can be detected in serum long before the clinical manifestation of the disease (3, 4). Although joint involvement is a hallmark of RA. Affected joint spaces exhibit the release of cytokines such as Interleukin-1, Interleukin-6, and tumor necrosis factor-alpha, accompanied by reduced levels of Interleukin-11, Interleukin-13, and Interleukin-10 (IL-10) (5). However, the pathogenesis of RA may involve extra-articular sites, including the lungs, periodontal tissues, gut microbiota, or citrullination processes in adipose tissue. This may also explain why some anticitrullinated protein antibody-positive patients with joint pain present with normal synovial tissue (6).

In recent years, microbial factors have garnered significant attention for their association with the pathogenesis of RA. Patients with RA exhibit notable dysbiosis of the oral microbiome, which can be partially restored through RA treatment, with the extent of recovery closely correlated with the patient's therapeutic response (7). Epidemiological studies reveal that periodontitis, a chronic infectious oral disease, is highly prevalent among RA patients and is strongly linked to the disease. Porphyromonas gingivalis, a key pathogen in periodontitis, has been implicated in the development of RA due to its ability to synthesize bacterial peptidylarginine deiminase (7). This enzyme induces citrullination of α - and β -fibrin chains within the synovium, generating autoantigens that promote the production of anti-citrullinated protein antibodies (7). These anti-citrullinated protein antibodies form immune complexes with citrullinated proteins, which bind to fragment crystallizable region and complement component 5a receptors on immune and inflammatory cells, triggering a complex cascade of immune responses and the release of inflammatory mediators, ultimately resulting in synovial inflammation and the onset of RA (8).

The gut microbiota plays a pivotal role in modulating host immune responses and is intricately linked to the development of several autoimmune conditions (2). The complex interplay between gut health and immune system homeostasis may be a crucial determinant in the etiology and progression of RA. Regulation of gut and immune system homeostasis is vital for overall health. Gut homeostasis involves the composition, function, and stability of the gut microbiome, as well as the integrity of both physical and immune barriers (9). The gut microbiota, often considered an invisible organ, plays a crucial role in maintaining intestinal barrier function, nutrient absorption, and overall immune and metabolic balance (10, 11). One of its primary metabolic functions is aiding the digestion of indigestible food residues, such as polysaccharides, oligosaccharides, and unabsorbed sugars and alcohols (12). The intestinal barrier consists of multiple layers: an outer mucus layer, a symbiotic gut microbiota, defense proteins like antimicrobial peptides and secretory immunoglobulin A; a middle layer of intestinal epithelial cells; and an inner layer of innate and adaptive immune cells (13, 14). The mucosal immune system helps protect the body by activating and regulating effector cells. When this immune system is disrupted, pathogenic flora can escape the intestinal tract (15). Evidence suggests that gut microbiome dysbiosis is a critical environmental factor in RA pathogenesis and may provoke abnormal immune responses (16, 17). Gut microbiota dysbiosis increases intestinal mucosal permeability, elevating endotoxin levels in the bloodstream and disrupting the balance of downstream metabolites that regulate systemic inflammation. These disruptions can initiate chronic systemic inflammatory responses, potentially influencing the onset and progression of osteoarthritis (18). Although the role of the gut-immune homeostatic axis in RA has been explored, its intricate regulatory network is not yet fully understood. Further research into how the gut microbiota and its metabolites affect RA is valuable for understanding RA pathogenesis and developing novel treatment strategies.

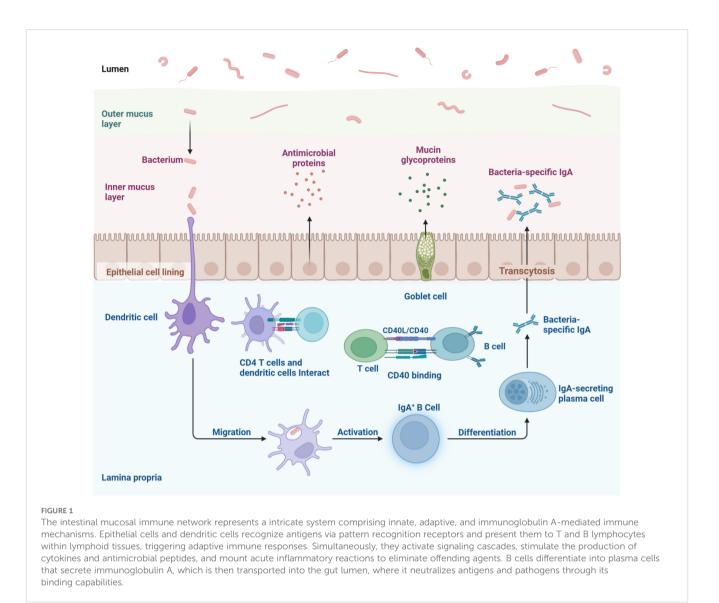
2 Interactions between gut homeostasis and the immune system

Gut microbes are essential for sustaining immune system balance. Changes in the gut microbiota and their metabolites can trigger inflammatory disorders related to the immune system, while a compromised intestinal barrier increases permeability and stimulates immune responses (19). The gastrointestinal tract houses the majority of the body's immune cells and constantly interacts with the gut microbiota, influencing their function and characteristics. The gut microbiota's delicate equilibrium in fostering either commensal or symbiotic relationships enables continuous bidirectional communication with the host immune system (20). System is shown in Figure 1.

2.1 Interactions between the gut microbiome and the immune system

The interactions between gut microbes and the host immune system are complex and context-dependent. The critical role of the gut microbiome in immune system development and response has been well established (21, 22). The microbiota is essential for regulating immune cell activities and inflammatory cytokines, thereby normalizing immune responses (23). Various gut microbes modulate immune responses differently. For example, segmented filamentous bacteria (SFB) promote the development of intestinal helper T cells 17 (Th17), and the SFB-dependent Th17 response inhibits bacterial translocation in mice with constitutively activated myosin light chain kinase (24). SFB enhances autoimmune responses in a context-dependent manner while strengthening intestinal barrier integrity. *Clostridium*, on the other hand, induces regulatory T cells (Treg) (25, 26).

Abbreviations: RA, Rheumatoid arthritis; SFB, Segmented filamentous bacteria; Th17, Helper T cells 17; Treg, Regulatory T cells; IL-10, Interleukin-10; LPS, Lipopolysaccharide; LGS, leaky gut syndrome; PAD, peptidyl-arginine deiminase; TJ, tight junction.



Additionally, distinct populations of Th17 cells exist in the gut, with their development largely influenced by different microorganisms. SFB-induced Th17 cells maintain homeostasis, whereas Th17 cells induced by Citrobacter are pro-inflammatory (27). Different bacterial species trigger distinct systemic immune responses. Probiotics like Lactobacillus and Bifidobacterium promote immune regulation by increasing regulatory T cell numbers, thereby maintaining immune tolerance and suppressing inflammatory responses. In contrast, harmful bacteria, such as those in the Enterobacteriaceae family, activate the Nuclear Factor kappa-light-chain-enhancer of Activated B cells signaling pathway and promote the release of pro-inflammatory cytokines, leading to systemic inflammation. Clinical studies have shown that elevated levels of Prevotella and Veillonella are closely associated with the development of autoimmune diseases. These bacteria exacerbate conditions like rheumatoid arthritis and inflammatory bowel disease by disrupting the gut barrier and facilitating the translocation of pathogens and their metabolites (28). Furthermore, during an inflammatory response, the intestinal microbiota can become disturbed, leading to an increase in harmful bacteria and a decrease in beneficial ones, which may exacerbate inflammation. For instance, the expansion of Pseudomonas aeruginosa causes intestinal inflammation and disrupts epithelial barrier integrity, allowing bacterial antigens to enter systemic circulation and activate immune responses at distant sites (29). Dysbiosis can result in the overactivation of T-helper 1 cells and Th17 cells, leading to excessive production of proinflammatory cytokines such as tumor necrosis factor, interleukin-6, and interleukin-17. This triggers systemic inflammation and immune dysregulation. Such abnormal cellular immune responses are pivotal in the pathogenesis and progression of autoimmune diseases like rheumatoid arthritis (19).

2.2 Gut microbial metabolite interactions with the immune system

The gut microbiota generates a variety of metabolites, including SCFAs, amines, polyamines, vitamins, and other small molecules. These metabolites are transported through the bloodstream to

tissues and organs throughout the body, where they influence immune responses and regulate inflammatory processes. Metabolites derived from gut microbiota are crucial in modulating the development and function of both adaptive and innate immune cells. IL-10 produced by effector T cells is a key selfregulatory mechanism that maintains immune balance (30). Shortchain fatty acids enhance IL-10 production in T-helper 1 Cells cells through a GPR43-dependent pathway and inhibit histone deacetylase histone deacetylase activation during T helper 1 and Th17 differentiation (22). Interleukin-22, a member of the IL-10 family, is vital for preventing intestinal inflammation, with CD4+ T cells being a major source of Interleukin-22 during chronic inflammation. Short-chain fatty acids stimulate Interleukin-22 production in CD4+ T cells via the G protein-coupled receptor 41 pathway and reduce histone deacetylase activity (22). Butyratetreated dendritic cells support Treg differentiation and inhibit Thelper 1 Cells differentiation by increasing the expression of immunosuppressive enzymes such as indoleamine 2,3dioxygenase 1 and aldehyde dehydrogenase 1 family member A2, through an SLC5A8-dependent mechanism (31). Microbial tryptophan metabolites, including indole and its derivatives, interact with aryl hydrocarbon receptors and influence B cell development, differentiation, cytokine production, and regulation through aryl hydrocarbon receptors signaling (32-34). Lipopolysaccharide (LPS), a component of Gram-negative bacteria in the gut, These metabolites are able to enter the bloodstream, initiates a cell-factor cascade that contributes to T cell-mediated inflammatory processes (35). Additionally, bile acids and their metabolites modulate immune responses by regulating signaling pathways and balancing Th17 and Treg cells (36).

2.3 Interactions between the gut barrier and the immune system

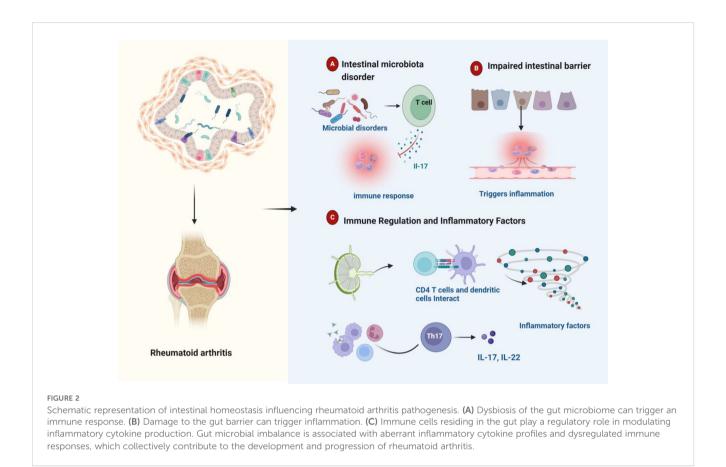
The gut barrier, which includes intestinal epithelial cells, tight junction proteins, and mucus layers (37). The integrity of the gut barrier is vital for preventing pathogen invasion and maintaining immune homeostasis. When this barrier is compromised, undigested food particles, pathogens, and toxins can penetrate into the bloodstream, triggering systemic immune responses and promoting chronic inflammation (38). Dysfunction of the gut barrier has been associated with various immune-mediated diseases, including rheumatoid arthritis and inflammatory bowel disease (39). Gutresident immune cells, such as dendritic cells, goblet cells, and T cells, are essential for regulating both local and systemic immune responses by detecting and responding to gut microbes and their metabolites (40). Disruption of the gut barrier can cause apoptosis of intestinal epithelial cells and create a pro-inflammatory environment, which includes the activation of autoreactive Th17 cells and other helper T cells (19). Pro-inflammatory cytokines like tumor necrosis factor-alpha and interferon-gamma can damage tight junctions (41-43), whereas immunosuppressive cytokines such as interleukin-10 and transforming growth factor-beta help to preserve their integrity (44). A weakened mucosal immune defense reduces Immunoglobulin A secretion or causes Immunoglobulin A dysfunction, while dysbiosis activates B cells, leading to abnormal levels of pro-inflammatory immunoglobulins like Immunoglobulin G. This can elevate rheumatoid arthritisspecific antibodies, further promoting systemic autoimmune responses (29, 45).

3 Impact of the gut homeostasisimmune system axis on rheumatoid arthritis

RA is complex and multifactorial (46, 47). The human body is home to over 100 trillion microbes, predominantly residing in the gut (48). Advances in bacterial DNA sequencing technology have elucidated the relationship between gut bacteria and RA, underscoring the significant role of the gut microbiota in the disease (29, 47). Research has shown notable differences in the gut microbiome between RA patients and healthy individuals, and these differences can affect RA manifestations (49, 50). The effect of intestinal homeostasis on rheumatoid arthritis is shown in Figure 2. Pro-inflammatory gut pathogens reshape the immune landscape by overactivating the innate immune system, which subsequently drives abnormal activation of the adaptive immune system, inducing RA. The gut microbiota composition also modulates sex hormone levels, influencing RA risk. Hormonal deficiencies increase intestinal permeability, elevating Th17 cells, nuclear factor kappa-light-chain-enhancer of activated B cells, Interleukin-17, and tumor necrosis factor-alevels in peripheral blood, which promotes bone resorption. Gut homeostasis imbalance is closely linked to RA pathogenesis through several mechanisms, including immune modulation by microbial metabolites and local inflammation initiated by bacterial components in synovial tissue (51).

3.1 Gut microbiome dysbiosis regulates inflammatory responses and influences rheumatoid arthritis

In healthy individuals, the gut microbiota is characterized by high diversity and stability, creating a dynamically balanced ecosystem crucial for maintaining gut homeostasis and immune balance (46). Increasing research focuses on the interaction of the gut microbiome in RA patients. The gut microbiota of RA patients undergoes significant alterations, characterized by a decrease in beneficial bacteria and an increase in pathogenic species. This dysbiosis is closely associated with immune system imbalance, driving inflammatory responses and exacerbating RA severity. Studies have demonstrated that in RA patients, significant upregulation is observed in bacterial genera such as Klebsiella, *Enterobacteriaceae*, *Eggerthella*, and *Flavobacteriaceae*, while genera like *Clostridium*, *Blautia*, and *Enterococcus* are downregulated (28). The proportion of Firmicutes is notably reduced in the RA gut, whereas Bacteroidetes levels are relatively



increased (28). Firmicutes members, including lactobacilli and bifidobacteria, exert anti-inflammatory effects and modulate immune responses. Conversely, some Bacteroidetes produce toxins and metabolites that exacerbate inflammation and RA. In treatment-naive, new-onset RA patients, Prevotella is significantly enriched and shows gene-cluster rearrangement; its 27-kDa protein stimulates a T helper 1 response in 42% of these patients (52, 53). Lactobacillus casei has been shown to significantly reduce the expression of interferon-gamma, tumor necrosis factor-alpha, and interleukin-1 beta, thereby preventing joint damage (54). Analysis of stool metabolites in RA patients reveals higher levels of glycerophospholipids, benzene and its derivatives, and cholesterol, while sphingolipids and tryptophan downstream metabolites are found at lower levels (28). In a collagen-induced arthritis mouse model, etanercept significantly decreased the abundance of Escherichia/Shigella while increasing the relative proportions of Tannerella, Lactobacillus, and Clostridium XIVa. Furthermore, certain natural compounds, such as clematichinenoside, have shown promise in improving RA-related gut dysbiosis and have been proven effective in reducing arthritis symptoms (55).

3.2 Impaired intestinal barrier and autoimmune activation

The intestinal barrier, consisting of epithelial cells, tight junction proteins, and mucus layers, primarily functions to prevent pathogens and harmful substances from entering the bloodstream (56, 57). This multilayered barrier is crucial for maintaining gut homeostasis and systemic immune responses. The integrity of the gut barrier is essential for maintaining health. LPS is widely used as a biomarker to assess gut barrier dysfunction, directly indicating the translocation of endotoxins into the bloodstream. Other markers, such as D-lactate and L-lactate, produced by gut bacteria, indirectly reflect changes in gut permeability (58). Barrier proteins, such as zonulin and adhesion molecules, are crucial for maintaining tight junctions, and their circulating levels reflect the functional status of the barrier. When the gut mucosal barrier is impaired, the disruption of tight junctions increases permeability, allowing undigested food particles, toxins, and pathogens to cross the compromised barrier and enter the bloodstream. Impairment of the intestinal barrier leads to tight junction breakdown, increased intestinal permeability, and the translocation of undigested food particles, toxins, and pathogens into the bloodstream, a condition known as leaky gut syndrome (LGS) (59). When these substances enter the bloodstream, they are identified by the immune system as "non-self," triggering an immune response that produces autoantibodies and inflammatory mediators. LGS initiates inflammatory responses in both the gut and peripheral tissues (60, 61) and is associated with autoimmune disorders such as type 1 diabetes mellitus, multiple sclerosis, rheumatoid arthritis, and celiac disease (62-64). LGS can induce the release of pro-inflammatory cytokines, including tumor necrosis factor-alpha, Interleukin-1 β , and Interleukin-6, by activating dendritic and goblet cells. These cytokines can elicit local inflammation and affect the immune system systemically, leading

to widespread inflammation and perpetuation (38). Additionally, LGS can enhance autoantigen exposure, stimulate autoimmune responses, and produce autoantibodies, thereby triggering and exacerbating RA.

3.3 Immune regulation and inflammatory mediators

The innate immune cells present in the gut-associated lymphoid tissue serve as the primary line of defense against foreign substances in the gastrointestinal tract. Dysbiosis of the gut microbiota can lead to inappropriate activation of these innate immune cells, resulting in an increased production of pro-inflammatory cytokines such as interleukin-12, interleukin-23, and type I interferon, coupled with a concomitant reduction in anti-inflammatory cytokines like transforming growth factor-\u03b3 and interleukin-10 produced by gutresident leukocytes (39). In the context of autoimmunity, adaptive lymphocytes, particularly T and B cells, play a pivotal role, with their aberrant activation contributing to the development of RA. Proinflammatory antigens originating from the gut can trigger maladaptive responses in the immune system by altering the immune milieu through excessive activation of innate immunity (19). Microbial antigens presented by dendritic cells and goblet cells can activate CD4+ T cells, leading to the differentiation of inflammatory T cell subsets. Notably, Th17 cells, a pro-inflammatory subset of CD4+ T cells, are characterized by their production of interleukin-17 (65). Conversely, CD4+ T cells can also differentiate into Tregs, which aid in suppressing Th17 responses (66, 67). An increased Th17/Treg ratio has been strongly implicated in RA, with this balance being closely regulated by the gut microbiota and its metabolites (39, 68). Furthermore, microbial antigens can stimulate follicular helper T cells, promoting the activation of B lymphocytes and subsequent production of pathogenic autoantibodies that may contribute to the pathogenesis of RA (39). Gut microbiota dysbiosis can trigger the migration of autoreactive cells to the joints, leading to local joint inflammation (69). These autoreactive cells activate macrophages, promoting the production of inflammatory cytokines. Additionally, cytokines such as tumor necrosis factor-a, Interleukin-6, and Interleukin-1 induce fibroblasts to secrete matrix metalloproteinases and receptor activator of nuclear factor KB ligand, further mediating bone and cartilage destruction and driving the progression of RA (70). These intricate interactions underscore the significant role of the gut microbiota in modulating systemic inflammatory responses and suggest that gut dysbiosis, inflammatory cytokines, and immune responses are interconnected factors influencing the development of RA (40).

4 Role of the gut-immune system axis in the pathogenesis of rheumatoid arthritis

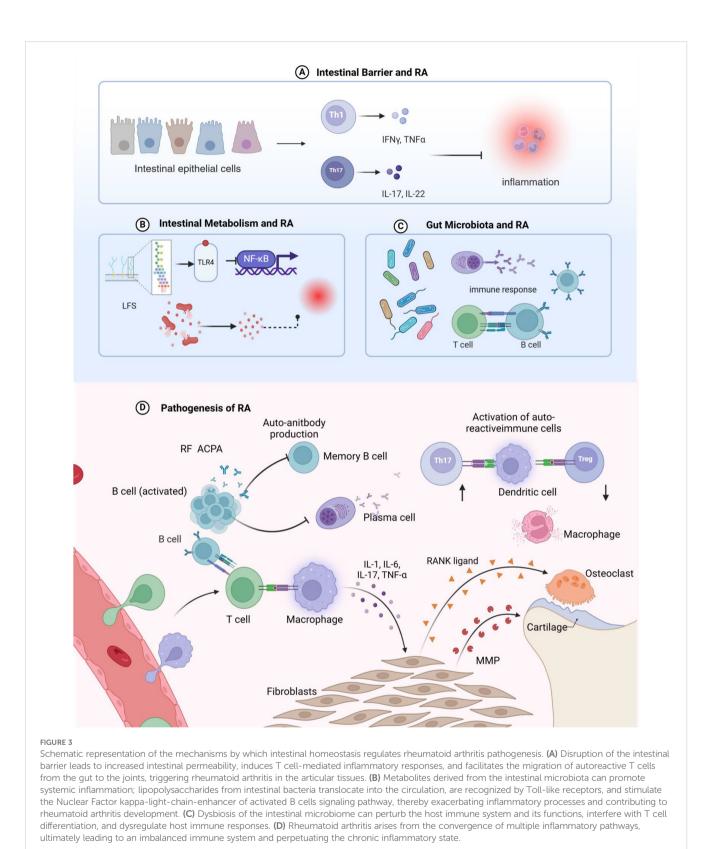
RA may be intricately linked to how disruptions in gut homeostasis influence immune function through metabolites produced by the gut microbiota (28, 71). Immune T and B cells in the intestinal mucosa have specific phenotypes and functions that are modulated by the microbiota (72). For instance, bacterial peptidoglycan components found in the synovial tissue of RA patients may contribute to inflammation within the joint microenvironment (53, 73). Recent research indicates that changes in gut microbiota composition in RA patients are a crucial factor in initiating abnormal systemic immune responses (38, 74, 75). The gut-immune system axis in the pathogenesis of rheumatoid arthritis shown in Figure 3.

4.1 Gut microbiota - immune system - rheumatoid arthritis

The mechanisms underlying the association between gut microbiome disturbances and RA have been extensively studied. Imbalance in the intestinal microbiota can impact the host immune system and its functions. The gut microbiota can activate antigenpresenting cells, such as dendritic cells, which alters cytokine production and antigen-presenting processes, interferes with T cell differentiation and function, and modulates the host immune response (76). Gut microbes interact with pattern recognition receptors, key innate immune receptors that detect pathogen-associated molecular patterns, including Toll-like receptors (50). Additionally, gut microbes can promote peptide citrullination through the enzymatic action of peptidyl-arginine deiminase (PAD). The human intestinal epithelium is a primary source of citrullinated peptides, and PAD activity is present in the human gut, with some gut microbes encoding functional microbial PADs (77). Consequently, the intestine can act as a source of citrullinated peptides and other mucosal surface antigens. Citrullination of peptides mediated by Porphyromonas gingivalisexpressed PAD is a significant factor in linking periodontitis with increased susceptibility to RA (50, 78). For instance, Prevotelladerived Pc-p27 can induce a Th1-mediated immune response by binding to human leukocyte antigen DR molecules in RA patients (2). This association is further supported by immunoglobulin A antibody responses to Pc-p27 in patients with acute and chronic RA, which are linked to the production of Th17 cell factors and anticitrullinated protein antibodies (53). Moreover, gut microbes influence the host immune system by regulating T cell differentiation and disrupting the balance between Th17 and Treg cells. In a mouse model of RA, specific changes in the gut microbiota may enhance the pathogenic role of Th17 cells while diminishing the inhibitory effects of Tregs thereby promoting Th17-mediated mucosal inflammation (50).

4.2 Gut metabolites - immune system - rheumatoid arthritis

When Gram-negative bacteria overgrow, they produce excessive amounts of LPS. Dysfunctional gut microbiota can disrupt gut barrier function, allowing pro-inflammatory substances like LPS to enter the circulatory system through the compromised barrier, thereby triggering systemic inflammation (79). Elevated LPS is recognized by Toll-like receptor 4, which activates the Nuclear Factor kappa-



light-chain-enhancer of activated B cells signaling pathway and initiates inflammation (80). Additionally, LPS can activate the complement alternative pathway, which plays a critical role in the development of arthritis, potentially contributing to its progression (81). *Bacteroides* fragilis stimulates T helper 1 responses during early

colonization by producing polysaccharide A. It may also contribute to RA pathogenesis by increasing intestinal permeability, reducing tight junction (TJ) protein expression, and affecting epithelial production of interleukin-17A (50, 82). Research shows that SCFAs, such as butyrate, can cross the gut-blood barrier and enter systemic

circulation, regulating the function of distant joints and immune cells. In RA patients, SCFA production is reduced, leading to impaired Treg cell function. Lower butyrate levels in the bloodstream are associated with increased inflammatory markers, such as C-reactive protein and interleukin-6, which enhance pro-inflammatory responses and accelerate RA progression. This underscores the direct influence of gut metabolite balance on inflammation. Additionally, fecal analyses reveal a significant reduction in butyrate-producing microbiota in RA patients, further supporting the link between gut microbial metabolism and RA progression (83, 84). Thus, imbalances in gut-derived metabolites directly affect the onset and development of RA through multiple pathways.

4.3 Intestinal epithelial barrier - immune system - rheumatoid arthritis

Destruction of the intestinal barrier leads to leakage of intestinal contents, creation of a pro-inflammatory environment, and the activation and infiltration of autoreactive Th17 and other effector T cells (38). Our data indicate that gut barrier dysfunction occurs prior to the clinical onset of arthritis in mouse models (85). Human studies also reveal elevated serum markers associated with impaired gut barrier function before RA onset, which correlates with a higher risk of developing RA later (85). Disruption of the gut barrier allows bacterial components to translocate across the intestinal wall and enter the bloodstream, triggering a robust immune response and inducing the release of pro-inflammatory cytokines, such as tumor necrosis factor- α and interleukin-6 (86). These pro-inflammatory mediators are key drivers of RA pathogenesis, promoting synovial inflammation and tissue destruction.

The TJ between intestinal epithelial cells is crucial for maintaining intestinal barrier integrity. Zonulin, a regulatory protein secreted by intestinal epithelial cells in response to dietary or microbial stimuli, modulates TJ function and intestinal epithelial permeability (87). Zonulin affects the expression of TJ proteins such as Zonula Occludens-1, occludin, and claudin-1 (88). In mouse models, elevated zonulin levels lead to the degradation of essential TJ proteins like Zonula Occludens-1 and an increase in claudin-2 and claudin-15. This combination disrupts TJ integrity, causing significant barrier dysfunction and increased intestinal permeability (89). This disruption not only initiates T-cell-mediated intestinal mucosal inflammation but also facilitates the migration of autoreactive T cells, including T-helper 1 Cells and Th17 cells, from the gut to the joints, thereby triggering RA.

5 A novel perspective on rheumatoid arthritis treatment targeting the gutimmune axis

Changes in gut microbiota composition and diversity can affect the onset and progression of RA by influencing the immune system (85). Interventions targeting the gut microbiome in RA patients aim to reshape microbiota composition and diversity, thereby alleviating autoimmune inflammatory responses. Recent studies in rat models have explored the efficacy and safety of strategies like probiotic supplementation and gut barrier stabilization in the management of RA (90).

5.1 Development of biomarkers and diagnostic tools - prevention before disease onset

The development of biomarkers and diagnostic tools is essential for the clinical study of RA. Gut microbiota composition and serum cytokine levels are key biomarkers for early diagnosis, monitoring disease activity, and evaluating treatment responses. The gut microbiota in RA patients differs significantly from that in healthy individuals, and these differences could serve as potential markers for early diagnosis (91). Notably, a significant increase in the proportion of Bacteroidetes and a decrease in Firmicutes are characteristic changes in RA (92). Early diagnosis of RA is critical for slowing joint damage. In early-stage RA patients, Prevotella abundance increases, while Bacteroides decreases (52). Transplanting gut microbiota from early RA patients into germfree sakuragi mouse model mice induces severe arthritis (93). During active RA, Haemophilus decreases, whereas Lactobacillus salivarius increases (94). Furthermore, the relative abundance of Collinsella and Akkermansia is higher in patients with active RA compared to those in remission (95). These findings suggest that gut microbiota alterations significantly influence RA severity, and shifts in microbial composition could serve as important biomarkers for RA diagnosis. Additionally, variations in serum levels of proand anti-inflammatory cytokines can reflect disease activity and treatment efficacy. For instance, elevated levels of tumor necrosis factor-alpha and Interleukin-6 are often associated with increased disease activity, while reduced levels of Interleukin-10 indicate weakened anti-inflammatory mechanisms (19, 96). Comprehensive analysis of gut microbiota and serum cytokine changes can lead to the development of novel diagnostic tools based on the gut-immune axis, supporting early diagnosis and personalized treatment of RA. Recent advancements, such as high-throughput sequencing and metabolomics, enable precise analysis of gut microbiota composition and its metabolites, offering in-depth information on biomarkers. Integrating multiomics data with machine learning algorithms can enhance diagnostic accuracy and sensitivity. The adoption of these advanced technologies will advance diagnostic tools for RA and strengthen clinical practice.

5.2 Treatment of rheumatoid arthritis targeting intestinal homeostasis

RA is a chronic, multisystem autoimmune disease characterized by inflammation in peripheral joints. The primary goals of RA treatment are to alleviate pain and swelling and to control disease progression. Methotrexate is commonly used as the first-line treatment for RA; however, individual sensitivity and tolerance to methotrexate can vary widely. Additionally, methotrexate may accumulate in the kidneys, liver, and pleural and peritoneal effusions, with significant variability in its clearance rate. The human gut microbiota and its enzymatic products influence the bioavailability, clinical efficacy, and toxicity of many drugs through both direct and indirect mechanisms. Moreover, various drugs and active compounds achieve therapeutic effects by normalizing gut microbiota composition, thereby regulating immune cell function (19). An emerging therapeutic approach involves targeting intestinal homeostasis to manage RA, and preliminary clinical studies have shown promising results. Regulating the intestinal microbiota and improving the balance of intestinal homeostasis and immune function offer potential therapeutic benefits for RA.

5.2.1 Microbiome modulators

Probiotics directly modulate the immune system by downregulating Toll-like Receptor expression, thereby reducing inflammation (55). They also regulate antigen-presenting cells. Kwon et al. further demonstrated that probiotics induce a Treg immune response in experimental RA models, significantly promoting the differentiation of T cells into forkhead box P3-expressing Tregs, which play a critical role in regulating and suppressing the inflammatory cascade (97). Additionally, probiotics produce gut metabolites, such as short-chain fatty acids, which have antibacterial and anti-inflammatory properties and can alleviate some symptoms of rheumatoid arthritis (55). Probiotic strains such as Lactobacillus and Bifidobacterium are known to produce beneficial short-chain fatty acids and contribute to maintaining intestinal mucosal homeostasis (98). Hatakka et al. evaluated the impact of Lactobacillus rhamnosus supplementation in patients with stable RA who were not receiving disease-modifying antirheumatic drugs. Although no significant differences were observed in inflammatory markers or clinical disease status compared to the placebo group, patients receiving the probiotic supplementation reported subjective improvements in health (99). In a study conducted RA patients underwent an eight-week probiotic regimen comprising Lactobacillus acidophilus, Lactobacillus casei, and Bifidobacterium bifidum. Compared to the control group, this intervention group exhibited enhanced disease activity scores 28 and reduced levels of high-sensitivity C-reactive protein, suggesting a favorable impact on clinical disease markers (100, 101). Furthermore, a compound probiotic capsule containing Lactobacillus casei LC-11, Lactobacillus acidophilus LA-14, Lactococcus lactis LL-23, Bifidobacterium lactis BL-04, and Bifidobacterium bifidum BB-06 also demonstrated therapeutic potential in patients with active RA (102).

Commensal bacteria play a pivotal role in regulating epithelial barrier function. While dysregulated LPS induction can trImmunoglobulin Ger inflammatory responses, microbial byproducts such as butyrate and short-chain fatty acids contribute to maintaining TJ integrity (38). Given the involvement of commensal microbiota in the development of LPS-related autoimmune diseases, interventions targeting the microbiota are emerging as novel strategies for preventing or treating such conditions (103, 104). Secondary bile acids, notably lithocholic acid, have exhibited anti-inflammatory effects in collagen-induced arthritis models, indicating their potential therapeutic role in RA (105). Additionally, propionate derived from *Bacteroides* fragilis may offer an alternative or complementary approach to current RA therapies (106). The dietary indole derivative indole-3-carbinol has been

demonstrated to mitigate adjuvant-induced arthritis (107). Sinomenine, a compound that enhances specific gut microbiota-derived indole tryptophan metabolites, activates the aryl hydrocarbon receptor aryl hydrocarbon receptors, and modulates the Nuclear Factor kappa-lightchain-enhancer of activated B cells and mitogen-activated protein kinase pathways, can alleviate RA-associated inflammation and immune responses (108).

5.2.2 Maintenance of intestinal barrier integrity

The zonulin family of peptides is a key regulator of intestinal tight junctions, highly expressed in autoimmune-prone mice and humans, and can predict the transition from autoimmunity to inflammatory arthritis. Elevated serum zonulin levels are strongly associated with gut barrier leakage, malnutrition, and inflammation. Studies suggest that restoring gut barrier function in the pre-arthritis phase using butyrate or type 1 cannabinoid receptor agonists can effectively inhibit the progression of arthritis (85). Treatments aimed at improving intestinal barrier function can help alleviate arthritis symptoms. For instance, microbial metabolites like butyrate are known to regulate epithelial tight junction protein expression effectively and restore intestinal barrier function, highlighting their role in mitigating the impact of microbial dysregulation on barrier integrity (109). Butyrate treatment has been shown to not only restore epithelial barrier function and gastrointestinal permeability to fluorescein Isothiocyanate-dextran but also significantly reduce the incidence of arthritis (109). In preclinical arthritis models, specific inhibition of the zonulin peptide by the antagonist larazotide acetate decreased the development of arthritis by nearly 50% and reduced intestinal permeability, thus preventing immune cell migration from the gut to the joints (85). Therapeutic interventions targeting zonulin can restore tight junction structure, improve barrier function, and partially protect joints from inflammatory damage (110). Early intervention in autoimmune diseases such as rheumatoid arthritis can improve long-term outcomes and increase remission rates. Thus, targeting zonulin and intestinal barrier function early in the disease provides a novel therapeutic strategy for modulating autoimmune development (79). The expression of hypoxia-inducible factor-1ain intestinal epithelial cells is crucial for maintaining barrier integrity. By inhibiting necroptosis in intestinal epithelial cells, hypoxia-inducible factor-10thelps preserve the intestinal epithelial barrier and is identified as a key regulatory factor in RA treatment (111). There is a potential mechanistic link between disrupted intestinal barrier function and arthritic autoimmune reactions (69, 85, 112, 113). Therefore, reinforcing or restoring intestinal barrier integrity is considered a promising strategy for RA treatment.

6 Conclusions

RA is a complex autoimmune disease involving multiple factors, with the gut homeostasis-immune system axis being particularly crucial. The bidirectional regulation between gut homeostasis and the immune system forms a regulatory network that significantly affects RA. RA is closely associated with gut dysbiosis, characterized by an increase in harmful bacteria and a decrease in beneficial bacteria. Gut homeostasis interacts with the immune system to influence RA progression. The gut microbiota and its metabolites regulate immune cells through multiple pathways, affecting immune responses. Gut microbes participate in antigen activation and regulate tight junction proteins to modulate gut mucosal permeability, thereby influencing RA development. Excessive production of metabolites like LPS can damage the gut barrier, promoting inflammation and accelerating RA progression. SCFAs and beneficial bacteria play essential roles in maintaining gut homeostasis and exert anti-inflammatory effects, but their levels are reduced in RA patients. Gut microbiota composition analysis has become a valuable tool for predicting RA susceptibility and controlling disease progression. By integrating gut microbiota data with serum cytokine profiles, new diagnostic approaches based on the gut-immune axis can be developed, offering crucial support for early diagnosis and personalized treatment of RA. Probiotics can induce Treg immune responses, promoting the differentiation of T cells into forkhead box P3-expressing Tregs. Moreover, SCFAs and other metabolites possess antibacterial and anti-inflammatory properties, helping to alleviate RA symptoms.

Therapeutic strategies targeting the gut microbiome have shown potential in modulating the immune system, offering new avenues for RA treatment. However, several limitations remain: the exact mechanisms by which specific microbiota or their metabolites influence RA pathogenesis through immune modulation are not fully understood; the molecular processes linking dysbiosis or intestinal barrier disruption to systemic inflammation and RA need further elucidation. The impact of intestinal homeostasis on arthritis and its underlying mechanisms requires more detailed investigation. The clinical efficacy and safety of RA treatments that focus on regulating intestinal homeostasis need to be thoroughly evaluated and validated. Future research should include additional clinical studies to confirm the effects and mechanisms of gut microbiota regulation in RA prevention and treatment. Investigating the action mechanisms of specific probiotics or metabolites, developing novel microbiome-targeted therapies, and integrating traditional drug treatments with lifestyle interventions could enhance the comprehensive management of RA. Further exploration of drug metabolism in RA and the interplay between gut homeostasis and the immune system is also needed. Advancing our understanding of strategies to regulate intestinal homeostasis may offer new insights and therapeutic targets, improving patient outcomes and quality of life.

References

1. Scott DL, Wolfe F, Huizinga TWJ. Rheumatoid arthritis. *Lancet Lond Engl.* (2010) 376:1094–108. doi: 10.1016/S0140-6736(10)60826-4

2. Attur M, Scher JU, Abramson SB, Attur M. Role of intestinal dysbiosis and nutrition in rheumatoid arthritis. *Cells.* (2022) 11:2436. doi: 10.3390/cells11152436

3. Evangelatos G, Fragoulis GE, Koulouri V, Lambrou GI. MicroRNAs in rheumatoid arthritis: From pathogenesis to clinical impact. *Autoimmun Rev.* (2019) 18:102391. doi: 10.1016/j.autrev.2019.102391

4. Fu L, Jin L, Yan L, Shi J, Wang H, Zhou B, et al. Comprehensive review of genetic association studies and meta-analysis on miRNA polymorphisms and rheumatoid arthritis and systemic lupus erythematosus susceptibility. *Hum Immunol.* (2016) 77:1–6. doi: 10.1016/j.humimm.2014.09.002

Author contributions

PQ: Investigation, Writing – original draft, Writing – review & editing. XC: Writing – review & editing. JT: Writing – review & editing. KZ: Writing – review & editing. ZQ: Writing – review & editing. ML: Writing – review & editing. XX: Funding acquisition, Methodology, Writing – original draft, Writing – review & editing.

Funding

The author(s) declare that financial support was received for the research, authorship, and/or publication of this article. This research was supported by the National Nature Fund Regional Project (82160911, 81860864). Central University Basic Scientific Research Business Expenses Project (31920210041). The special project of science and technology development under the guidance of the central government (YDZX2020620002356).

Acknowledgments

The authors acknowledge the use of Biorender to create schematic representations in Figures 1–3. The agreement numbers associated with this use are DI276B2BXU, EQ276B2VN6, QM279W085F.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.Abbreviation

5. Peroumal D, Abimannan T, Tagirasa R, Parida JR, Singh SK, Padhan P, et al. Inherent low Erk and p38 activity reduce Fas Ligand expression and degranulation in T helper 17 cells leading to activation induced cell death resistance. *Oncotarget.* (2016) 7:54339–59. doi: 10.18632/oncotarget.10913

6. Petrovská N, Prajzlerová K, Vencovský J, Šenolt L, Filková M. The pre-clinical phase of rheumatoid arthritis: From risk factors to prevention of arthritis. *Autoimmun Rev.* (2021) 20:102797. doi: 10.1016/j.autrev.2021.102797

7. Araújo VMA, Melo IM, Lima V. Relationship between periodontitis and rheumatoid arthritis: review of the literature. *Mediators Inflamm*. (2015) 2015:259074. doi: 10.1155/2015/259074

8. Konig MF, Abusleme L, Reinholdt J, Palmer RJ, Teles RP, Sampson K, et al. Aggregatibacter actinomycetemcomitans-induced hypercitrullination links periodontal

infection to autoimmunity in rheumatoid arthritis. Sci Transl Med. (2016) 8:369ra176. doi: 10.1126/scitranslmed.aaj1921

 Qi P, Lv J, Yan X, Bai L, Zhang L. Microfluidics: insights into intestinal microorganisms. Microorganisms. (2023) 11:1134. doi: 10.3390/microorganisms11051134

10. Qi P, Lv J, Bai LH, Yan XD, Zhang L. Effects of hypoxemia by acute high-altitude exposure on human intestinal flora and metabolism. *Microorganisms*. (2023) 11:2284. doi: 10.3390/microorganisms11092284

11. Allam-Ndoul B, Castonguay-Paradis S, Veilleux A. Gut microbiota and intestinal trans-epithelial permeability. *Int J Mol Sci.* (2020) 21:6402. doi: 10.3390/ ijms21176402

12. Kim B, Choi HN, Yim JE. Effect of diet on the gut microbiota associated with obesity. J Obes Metab Syndr. (2019) 28:216–24. doi: 10.7570/jomes.2019.28.4.216

13. Di Tommaso N, Gasbarrini A, Ponziani FR. Intestinal barrier in human health and disease. *Int J Environ Res Public Health*. (2021) 18:12836. doi: 10.3390/ ijerph182312836

14. Camilleri M, Madsen K, Spiller R, Greenwood-Van Meerveld B, Verne GN. Intestinal barrier function in health and gastrointestinal disease. *Neurogastroenterol Motil.* (2012) 24:503–12. doi: 10.1111/j.1365-2982.2012.01921.x

15. Gomaa EZ. Human gut microbiota/microbiome in health and diseases: a review. Antonie Van Leeuwenhoek. (2020) 113:2019–40. doi: 10.1007/s10482-020-01474-7

16. Kau AL, Ahern PP, Griffin NW, Goodman AL, Gordon JI. Human nutrition, the gut microbiome and the immune system. *Nature*. (2011) 474:327–36. doi: 10.1038/ nature10213

17. Taneja V. Arthritis susceptibility and the gut microbiome. FEBS Lett. (2014) 588:4244–9. doi: 10.1016/j.febslet.2014.05.034

18. Wei J, Zhang Y, Hunter D, Zeng C, Lei G. The gut microbiome-joint axis in osteoarthritis. *Sci Bull.* (2023) 68:759–62. doi: 10.1016/j.scib.2023.03.024

19. Zhao T, Wei Y, Zhu Y, Xie Z, Hai Q, Li Z, et al. Gut microbiota and rheumatoid arthritis: From pathogenesis to novel therapeutic opportunities. *Front Immunol.* (2022) 13:1007165. doi: 10.3389/fimmu.2022.1007165

20. Shin C, Kim YK. Autoimmunity in microbiome-mediated diseases and novel therapeutic approaches. *Curr Opin Pharmacol.* (2019) 49:34–42. doi: 10.1016/j.coph.2019.04.018

21. Noverr MC, Huffnagle GB. Does the microbiota regulate immune responses outside the gut? *Trends Microbiol.* (2004) 12:562–8. doi: 0.1016/j.tim.2004.10.008

22. Yang W, Cong Y. Gut microbiota-derived metabolites in the regulation of host immune responses and immune-related inflammatory diseases. *Cell Mol Immunol.* (2021) 18:866–77. doi: 10.1038/s41423-021-00661-4

23. Macpherson AJ, Harris NL. Interactions between commensal intestinal bacteria and the immune system. *Nat Rev Immunol.* (2004) 4:478-85. doi: 10.1038/nri1373

24. Edelblum KL, Sharon G, Singh G, Odenwald MA, Sailer A, Cao S, et al. The microbiome activates CD4 T-cell-mediated immunity to compensate for increased intestinal permeability. *Cell Mol Gastroenterol Hepatol.* (2017) 4:285–97. doi: 10.1016/j.jcmgh.2017.06.001

25. Ivanov II, Atarashi K, Manel N, Brodie EL, Shima T, Karaoz U, et al. Induction of intestinal Th17 cells by segmented filamentous bacteria. *Cell.* (2009) 139:485–98. doi: 10.1016/j.cell.2009.09.033

26. Atarashi K, Tanoue T, Shima T, Imaoka A, Kuwahara T, Momose Y, et al. Induction of colonic regulatory T cells by indigenous Clostridium species. *Science*. (2011) 331:337-41. doi: 10.1126/science.1198469

27. Omenetti S, Bussi C, Metidji A, Iseppon A, Lee S, Tolaini M, et al. The intestine harbors functionally distinct homeostatic tissue-resident and inflammatory th17 cells. *Immunity.* (2019) 51:77–89.e6. doi: 10.1016/j.immuni.2019.05.004

28. Yu D, Du J, Pu X, Zheng L, Chen S, Wang N, et al. The gut microbiome and metabolites are altered and interrelated in patients with rheumatoid arthritis. *Front Cell Infect Microbiol.* (2022) 11:763507. doi: 10.3389/fcimb.2021.763507

29. Tsetseri MN, Silman AJ, Keene DJ, Dakin SG. The role of the microbiome in rheumatoid arthritis: a review. *Rheumatol Adv Pract.* (2023) 7:rkad034. doi: 10.1093/rap/rkad034

30. Kalliolias GD, Papavassiliou AG. Stabilizing the integrity of intestinal barrier to control arthritis. *Arthritis Res Ther.* (2024) 26:135. doi: 10.1186/s13075-024-03378-7

31. Rosser EC, Piper CJM, Matei DE, Blair PA, Rendeiro AF, Orford M, et al. Microbiota-derived metabolites suppress arthritis by amplifying aryl-hydrocarbon receptor activation in regulatory B cells. *Cell Metab.* (2020) 31:837–851.e10. doi: 10.1016/j.cmet.2020.03.003

32. Li J, Bhattacharya S, Zhou J, Phadnis-Moghe AS, Crawford RB, Kaminski NE. Aryl hydrocarbon receptor activation suppresses EBF1 and PAX5 and impairs human B lymphopoiesis. *J Immunol Baltim Md.* (1950) 199:3504–15. doi: 10.4049/jimmunol.1700289

33. Vaidyanathan B, Chaudhry A, Yewdell WT, Angeletti D, Yen WF, Wheatley AK, et al. The aryl hydrocarbon receptor controls cell-fate decisions in B cells. *J Exp Med.* (2017) 214:197–208. doi: 10.1084/jem.20160789

34. Piper CJM, Rosser EC, Oleinika K, Nistala K, Krausgruber T, Rendeiro AF, et al. Aryl hydrocarbon receptor contributes to the transcriptional program of IL-10-producing regulatory B cells. *Cell Rep.* (2019) 29:1878–1892.e7. doi: 10.1016/j.celrep.2019.10.018

35. Lin L, Zhang K, Xiong Q, Zhang J, Cai B, Huang Z, et al. Gut microbiota in preclinical rheumatoid arthritis: From pathogenesis to preventing progression. *J Autoimmun.* (2023) 141:103001. doi: 10.1016/j.jaut.2023.103001

36. Xu X, Wang M, Wang Z, Chen Q, Chen X, Xu Y, et al. The bridge of the gutjoint axis: Gut microbial metabolites in rheumatoid arthritis. *Front Immunol.* (2022) 13:1007610. doi: 10.3389/fimmu.2022.1007610

37. Untersmayr E, Brandt A, Koidl L, Bergheim I. The intestinal barrier dysfunction as driving factor of inflammaging. *Nutrients*. (2022) 14:949. doi: 10.3390/nu14050949

38. Kinashi Y, Hase K. Partners in leaky gut syndrome: intestinal dysbiosis and autoimmunity. *Front Immunol.* (2021) 12:673708. doi: 10.3389/fimmu.2021.673708

39. Zhang X, di Chen B, Zhao LD, Li H. The gut microbiota: emerging evidence in autoimmune diseases. *Trends Mol Med.* (2020) 26:862–73. doi: 10.1016/j.molmed.2020.04.001

40. Li Y, Zhang SX, Yin XF, Zhang MX, Qiao J, Xin XH, et al. The gut microbiota and its relevance to peripheral lymphocyte subpopulations and cytokines in patients with rheumatoid arthritis. *J Immunol Res.* (2021) 2021:6665563. doi: 10.1155/2021/6665563

41. Ye D, Ma I, Ma TY. Molecular mechanism of tumor necrosis factor-alpha modulation of intestinal epithelial tight junction barrier. *Am J Physiol Gastrointest Liver Physiol.* (2006) 290:G496–504. doi: 10.1152/ajpgi.00318.2005

42. Mankertz J, Amasheh M, Krug SM, Fromm A, Amasheh S, Hillenbrand B, et al. TNFalpha up-regulates claudin-2 expression in epithelial HT-29/B6 cells via phosphatidylinositol-3-kinase signaling. *Cell Tissue Res.* (2009) 336:67–77. doi: 10.1007/s00441-009-0751-8

43. Bruewer M, Utech M, Ivanov AI, Hopkins AM, Parkos CA, Nusrat A. Interferon-gamma induces internalization of epithelial tight junction proteins via a macropinocytosis-like process. *FASEB J Off Publ Fed Am Soc Exp Biol.* (2005) 19:923–33. doi: 10.1096/fj.04-3260com

44. Howe KL, Reardon C, Wang A, Nazli A, McKay DM. Transforming growth factor-beta regulation of epithelial tight junction proteins enhances barrier function and blocks enterohemorrhagic Escherichia coli O157:H7-induced increased permeability. *Am J Pathol.* (2005) 167:1587–97. doi: 10.1016/S0002-9440(10)61243-6

45. Rescigno M. The intestinal epithelial barrier in the control of homeostasis and immunity. *Trends Immunol.* (2011) 6):256–64. doi: 10.1016/j.it.2011.04.003

46. Dong Y, Yao J, Deng Q, Li X, He Y, Ren X, et al. Relationship between gut microbiota and rheumatoid arthritis: A bibliometric analysis. *Front Immunol.* (2023) 14:1131933. doi: 10.3389/fimmu.2023.1131933

47. Catrina AI, Deane KD, Scher JU. Gene, environment, microbiome and mucosal immune tolerance in rheumatoid arthritis. *Rheumatol Oxf Engl.* (2016) 55:391–402. doi: 10.1093/rheumatology/keu469

48. Qin J, Li R, Raes J, Arumugam M, Burgdorf KS, Manichanh C, et al. A human gut microbial gene catalogue established by metagenomic sequencing. *Nature*. (2010) 464:59–65. doi: 10.1038/nature08821

49. Brusca SB, Abramson SB, Scher JU. Microbiome and mucosal inflammation as extra-articular triggers for rheumatoid arthritis and autoimmunity. *Curr Opin Rheumatol.* (2014) 26:101–7. doi: 10.1097/BOR.000000000000008

 Horta-Baas G, Romero-Figueroa MDS, Montiel-Jarquín AJ, Pizano-Zárate ML, García-Mena J, Ramírez-Durán N. Intestinal dysbiosis and rheumatoid arthritis: A link between gut microbiota and the pathogenesis of rheumatoid arthritis. *J Immunol Res.* (2017) 2017:4835189. doi: 10.1155/2017/4835189

51. Pacifici R. Bone remodeling and the microbiome. *Cold Spring Harb Perspect Med.* (2018) 8:a031203. doi: 10.1101/cshperspect.a031203

52. Scher JU, Sczesnak A, Longman RS, Segata N, Ubeda C, Bielski C, et al. Expansion of intestinal Prevotella copri correlates with enhanced susceptibility to arthritis. *eLife*. (2013) 2:e01202. doi: 10.7554/eLife.01202

53. Pianta A, Arvikar S, Strle K, Drouin EE, Wang Q, Costello CE, et al. Evidence of the immune relevance of prevotella copri, a gut microbe, in patients with rheumatoid arthritis. *Arthritis Rheumatol Hoboken NJ.* (2017) 69:964–75. doi: 10.1002/art.40003

54. Parada-Turska J, Rzeski W, Zgrajka W, Majdan M, Kandefer-Szerszeń M, Turski W. Kynurenic acid, an endogenous constituent of rheumatoid arthritis synovial fluid, inhibits proliferation of synoviocytes *in vitro*. *Rheumatol Int*. (2006) 26:422–6. doi: 10.1007/s00296-005-0057-4

55. Opoku YK, Asare KK, Ghartey-Quansah G, Afrifa J, Bentsi-Enchill F, Ofori EG, et al. Intestinal microbiome-rheumatoid arthritis crosstalk: The therapeutic role of probiotics. *Front Microbiol.* (2022) 13:996031. doi: 10.3389/fmicb.2022.996031

56. Shi N, Li N, Duan X, Niu H. Interaction between the gut microbiome and mucosal immune system. *Mil Med Res.* (2017) 4:14. doi: 10.1186/s40779-017-0122-9

57. König J, Wells J, Cani PD, García-Ródenas CL, MacDonald T, Mercenier A, et al. Human intestinal barrier function in health and disease. *Clin Transl Gastroenterol.* (2016) 7:e196. doi: 10.1038/ctg.2016.54

58. Zhang Y, Li X, Luo Z, Ma L, Zhu S, Wang Z, et al. ECM1 is an essential factor for the determination of M1 macrophage polarization in IBD in response to LPS stimulation. *Proc Natl Acad Sci U.S.A.* (2020) 117(6):3083–92. https://pubmed.ncbi. nlm.nih.gov/31980528/.

59. Ahmad R, Sorrell MF, Batra SK, Dhawan P, Singh AB. Gut permeability and mucosal inflammation: bad, good or context dependent. *Mucosal Immunol.* (2017) 10:307–17. doi: 10.1038/mi.2016.128

60. Fasano A, Shea-Donohue T. Mechanisms of disease: the role of intestinal barrier function in the pathogenesis of gastrointestinal autoimmune diseases. *Nat Clin Pract Gastroenterol Hepatol.* (2005) 2:416–22. doi: 10.1038/ncpgasthep0259

61. Mu Q, Kirby J, Reilly CM, Luo XM. Leaky gut as a danger signal for autoimmune diseases. *Front Immunol.* (2017) 8:598. doi: 10.3389/fimmu.2017.00598

62. Johansson MEV, Hansson GC. Immunological aspects of intestinal mucus and mucins. Nat Rev Immunol. (2016) 16:639–49. doi: 10.1038/nri.2016.88

63. Bevins CL, Salzman NH. Paneth cells, antimicrobial peptides and maintenance of intestinal homeostasis. *Nat Rev Microbiol.* (2011) 9:356–68. doi: 10.1038/nrmicro2546

64. Johansen FE, Pekna M, Norderhaug IN, Haneberg B, Hietala MA, Krajci P, et al. Absence of epithelial immunoglobulin A transport, with increased mucosal leakiness, in polymeric immunoglobulin receptor/secretory component-deficient mice. *J Exp Med.* (1999) 190:915–22. doi: 10.1084/jem.190.7.915

65. Wang Y, Yin Y, Chen X, Zhao Y, Wu Y, Li Y, et al. Induction of intestinal th17 cells by flagellins from segmented filamentous bacteria. *Front Immunol.* (2019) 10:2750. doi: 10.3389/fimmu.2019.02750

66. Lina C, Conghua W, Nan L, Ping Z. Combined treatment of etanercept and MTX reverses Th1/Th2, Th17/Treg imbalance in patients with rheumatoid arthritis. *J Clin Immunol.* (2011) 31:596–605. doi: 10.1007/s10875-011-9542-6

67. Chen X, Oppenheim JJ. Th17 cells and Tregs: unlikely allies. *J Leukoc Biol*. (2014) 95:723–31. doi: 10.1189/jlb.1213633

68. Cheng H, Guan X, Chen D, Ma W. The th17/treg cell balance: A gut microbiotamodulated story. *Microorganisms*. (2019) 7:583. doi: 10.3390/microorganisms7120583

69. Brandl C, Bucci L, Schett G, Zaiss MM. Crossing the barriers: Revisiting the gut feeling in rheumatoid arthritis. *Eur J Immunol.* (2021) 51:798–810. doi: 10.1002/eji.202048876

70. Pacifici R. T cells, osteoblasts, and osteocytes: interacting lineages key for the bone anabolic and catabolic activities of parathyroid hormone. *Ann N Y Acad Sci.* (2016) 1364:11–24. doi: 10.1111/nyas.2016.1364.issue-1

71. He J, Chu Y, Li J, Meng Q, Liu Y, Jin J, et al. Intestinal butyrate-metabolizing species contribute to autoantibody production and bone erosion in rheumatoid arthritis. *Sci Adv.* (2022) 8:eabm1511. doi: 10.1126/sciadv.abm1511

72. Honda K, Littman DR. The microbiota in adaptive immune homeostasis and disease. *Nature*. (2016) 535:75–84. doi: 10.1038/nature18848

73. Schrijver IA, Melief MJ, Tak PP, Hazenberg MP, Laman JD. Antigen-presenting cells containing bacterial peptidoglycan in synovial tissues of rheumatoid arthritis patients coexpress costimulatory molecules and cytokines. *Arthritis Rheumatol.* (2000) 43:2160–8. doi: 10.1002/1529-0131(200010)43:10<2160::AID-ANR3>3.0.CO;2-T

74. Chen B, Sun L, Zhang X. Integration of microbiome and epigenome to decipher the pathogenesis of autoimmune diseases. *J Autoimmun*. (2017) 83:31–42. doi: 10.1016/j.jaut.2017.03.009

75. Zhang X, Zhang D, Jia H, Feng Q, Wang D, Liang D, et al. The oral and gut microbiomes are perturbed in rheumatoid arthritis and partly normalized after treatment. *Nat Med.* (2015) 21:895–905. doi: 10.1038/nm.3914

76. Guerreiro CS, Calado Â, Sousa J, Fonseca JE. Diet, microbiota, and gut permeability-the unknown triad in rheumatoid arthritis. *Front Med.* (2018) 5:349. doi: 10.3389/fmed.2018.00349

77. Lerner A, Matthias T. Rheumatoid arthritis-celiac disease relationship: joints get that gut feeling. Autoimmun Rev. (2015) 14:1038–47. doi: 10.1016/j.autrev.2015.07.007

78. Lerner A, Aminov R, Matthias T. Dysbiosis may trigger autoimmune diseases via inappropriate post-translational modification of host proteins. *Front Microbiol.* (2016) 7:84. doi: 10.3389/fmicb.2016.00084

79. Peng Y, Huang Y, Li H, Li C, Wu Y, Chen ZS, et al. Huangqin Qingre Chubi Capsule inhibits rheumatoid arthritis by regulating intestinal flora and improving intestinal barrier. *Front Pharmacol.* (2024) 15:1422245. doi: 10.3389/fphar.2024.1422245

80. Zhao Z, Ning J, Bao XQ, Shang M, Ma J, Li G, et al. Fecal microbiota transplantation protects rotenone-induced Parkinson's disease mice via suppressing inflammation mediated by the lipopolysaccharide-TLR4 signaling pathway through the microbiota-gut-brain axis. *Microbiome*. (2021) 9:226. doi: 10.1186/s40168-021-01107-9

81. Murayama MA, Kakuta S, Inoue A, Umeda N, Yonezawa T, Maruhashi T, et al. CTRP6 is an endogenous complement regulator that can effectively treat induced arthritis. *Nat Commun.* (2015) 6:8483. doi: 10.1038/ncomms9483

82. Chen J, Wright K, Davis JM, Jeraldo P, Marietta EV, Murray J, et al. An expansion of rare lineage intestinal microbes characterizes rheumatoid arthritis. *Genome Med.* (2016) 8:43. doi: 10.1186/s13073-016-0299-7

83. Kim D, Yoo SA, Kim WU. Gut microbiota in autoimmunity: potential for clinical applications. *Arch Pharm Res.* (2016) 39:1565–76. doi: 10.1007/s12272-016-0796-7

84. Zhong D, Wu C, Zeng X, Wang Q. The role of gut microbiota in the pathogenesis of rheumatic diseases. *Clin Rheumatol.* (2018) 37:25–34. doi: 10.1007/s10067-017-3821-4

85. Tajik N, Frech M, Schulz O, Schälter F, Lucas S, Azizov V, et al. Targeting zonulin and intestinal epithelial barrier function to prevent onset of arthritis. *Nat Commun.* (2020) 11:1995. doi: 10.1038/s41467-020-15831-7

86. Neunlist M, Toumi F, Oreschkova T, Denis M, Leborgne J, Laboisse CL, et al. Human ENS regulates the intestinal epithelial barrier permeability and a tight junctionassociated protein ZO-1 via VIPergic pathways. *Am J Physiol Gastrointest Liver Physiol.* (2003) 285(5):G1028–36. https://pubmed.ncbi.nlm.nih.gov/12881224/.

87. Campisi L, Barbet G, Ding Y, Esplugues E, Flavell RA, Blander JM. Apoptosis in response to microbial infection induces autoreactive TH17 cells. *Nat Immunol.* (2016) 17:1084–92. doi: 10.1038/ni.3512

88. Romero-Figueroa MDS, Ramírez-Durán N, Montiel-Jarquín AJ, Horta-Baas G. Gut-joint axis: Gut dysbiosis can contribute to the onset of rheumatoid arthritis via multiple pathways. *Front Cell Infect Microbiol.* (2023) 13:1092118. doi: 10.3389/fcimb.2023.1092118

89. Van Itallie CM, Fanning AS, Bridges A, Anderson JM. ZO-1 stabilizes the tight junction solute barrier through coupling to the perijunctional cytoskeleton. *Mol Biol Cell*. (2009) 20:3930–40. doi: 10.1091/mbc.e09-04-0320

90. Pan H, Guo R, Ju Y, Wang Q, Zhu J, Xie Y, et al. A single bacterium restores the microbiome dysbiosis to protect bones from destruction in a rat model of rheumatoid arthritis. *Microbiome*. (2019) 7:107. doi: 10.1186/s40168-019-0719-1

91. Hanchi H, Mottawea W, Sebei K, Hammami R. The genus enterococcus: between probiotic potential and safety concerns-an update. *Front Microbiol.* (2018) 9:1791. doi: 10.3389/fmicb.2018.01791

92. Lee JY, Mannaa M, Kim Y, Kim J, Kim GT, Seo YS. Comparative analysis of fecal microbiota composition between rheumatoid arthritis and osteoarthritis patients. *Genes.* (2019) 10:748. doi: 10.3390/genes10100748

93. Maeda Y, Kurakawa T, Umemoto E, Motooka D, Ito Y, Gotoh K, et al. Dysbiosis contributes to arthritis development via activation of autoreactive T cells in the intestine. *Arthritis Rheumatol Hoboken NJ*. (2016) 68:2646–61. doi: 10.1002/art.v68.11

94. Liu X, Zou Q, Zeng B, Fang Y, Wei H. Analysis of fecal Lactobacillus community structure in patients with early rheumatoid arthritis. *Curr Microbiol.* (2013) 67:170–6. doi: 10.1007/s00284-013-0338-1

95. Chiang HI, Li JR, Liu CC, Liu PY, Chen HH, Chen YM, et al. An association of gut microbiota with different phenotypes in chinese patients with rheumatoid arthritis. *J Clin Med.* (2019) 8:1770. doi: 10.3390/jcm8111770

96. Feng J, Zhao F, Sun J, Lin B, Zhao L, Liu Y, et al. Alterations in the gut microbiota and metabolite profiles of thyroid carcinoma patients. *Int J Cancer.* (2019) 144:2728–45. doi: 10.1002/ijc.v144.11

97. Kwon HK, Lee CG, So JS, Chae CS, Hwang JS, Sahoo A, et al. Generation of regulatory dendritic cells and CD4+Foxp3+ T cells by probiotics administration suppresses immune disorders. *Proc Natl Acad Sci U S A.* (2010) 107:2159–64. doi: 10.1073/pnas.0904055107

98. Bungau SG, Behl T, Singh A, Sehgal A, Singh S, Chigurupati S, et al. Targeting probiotics in rheumatoid arthritis. *Nutrients*. (2021) 13:3376. doi: 10.3390/nu13103376

99. Hatakka K, Martio J, Korpela M, Herranen M, Poussa T, Laasanen T, et al. Effects of probiotic therapy on the activity and activation of mild rheumatoid arthritisa pilot study. *Scand J Rheumatol.* (2003) 32:211–5. doi: 10.1080/03009740310003695

100. Vaghef-Mehrabany E, Alipour B, Homayouni-Rad A, Sharif SK, Asghari-Jafarabadi M, Zavvari S. Probiotic supplementation improves inflammatory status in patients with rheumatoid arthritis. *Nutr Burbank Los Angel Cty Calif.* (2014) 30:430–5. doi: 10.1016/j.nut.2013.09.007

101. Alipour B, Homayouni-Rad A, Vaghef-Mehrabany E, Sharif SK, Vaghef-Mehrabany L, Asghari-Jafarabadi M, et al. Effects of Lactobacillus casei supplementation on disease activity and inflammatory cytokines in rheumatoid arthritis patients: a randomized double-blind clinical trial. *Int J Rheum Dis.* (2014) 17:519–27. doi: 10.1111/apl.2014.17.issue-5

102. Cannarella LAT, Mari NL, Alcântara CC, Iryioda TMV, Costa NT, Oliveira SR, et al. Mixture of probiotics reduces inflammatory biomarkers and improves the oxidative/nitrosative profile in people with rheumatoid arthritis. *Nutr Burbank Los Angel Cty Calif.* (2021) 89:111282. doi: 10.1016/j.nut.2021.111282

103. de Oliveira GLV, Leite AZ, Higuchi BS, Gonzaga MI, Mariano VS. Intestinal dysbiosis and probiotic applications in autoimmune diseases. *Immunology.* (2017) 152:1-12. doi: 10.1111/imm.2017.152.issue-1

104. Marietta E, Mangalam AK, Taneja V, Murray JA. Intestinal Dysbiosis in, and Enteral Bacterial Therapies for, Systemic Autoimmune Diseases. *Front Immunol.* (2020) 11:573079. doi: 10.3389/fimmu.2020.573079

105. Li ZY, Zhou JJ, Luo CL, Zhang LM. Activation of TGR5 alleviates inflammation in rheumatoid arthritis peripheral blood mononuclear cells and in mice with collagen II –induced arthritis. *Mol Med Rep.* (2019) 20:4540–50. doi: 10.3892/mmr.2019.10711

106. Chen H, Fu X, Wu X, Zhao J, Qiu F, Wang Z, et al. Gut microbial metabolite targets HDAC3-FOXK1-interferon axis in fibroblast-like synoviocytes to ameliorate rheumatoid arthritis. *Bone Res.* (2024) 12:31. doi: 10.1038/s41413-024-00336-6

 Hasan H, Ismail H, El-Orfali Y, Khawaja G. Therapeutic benefits of Indole-3-Carbinol in adjuvant-induced arthritis and its protective effect against methotrexate induced-hepatic toxicity. BMC Complement Altern Med. (2018) 18:337. doi: 10.1186/ s12906-018-2408-1

108. Jiang ZM, Zeng SL, Huang TQ, Lin Y, Wang FF, Gao XJ, et al. Sinomenine ameliorates rheumatoid arthritis by modulating tryptophan metabolism and activating aryl hydrocarbon receptor via gut microbiota regulation. *Sci Bull.* (2023) 68:1540–55. doi: 10.1016/j.scib.2023.06.027

109. Plöger S, Stumpff F, Penner GB, Schulzke JD, Gäbel G, Martens H, et al. Microbial butyrate and its role for barrier function in the gastrointestinal tract. *Ann N Y Acad Sci.* (2012) 1258:52–9. doi: 10.1111/j.1749-6632.2012.06553.x

110. Deane KD, El-Gabalawy H. Pathogenesis and prevention of rheumatic disease: focus on preclinical RA and SLE. *Nat Rev Rheumatol.* (2014) 10:212–28. doi: 10.1038/nrrheum.2014.6

111. Lyu P, Wen J, Zhang W, Liu N, Stolzer I, Gießl A, et al. Expression of HIF1 α in intestinal epithelium restricts arthritis inflammation by inhibiting

RIPK3-induced cell death machinery. Ann Rheum Dis. (2024) 83:984-97. doi: 10.1136/ard-2023-224491

112. Matei DE, Menon M, Alber DG, Smith AM, Nedjat-Shokouhi B, Fasano A, et al. Intestinal barrier dysfunction plays an integral role in arthritis pathology and can be targeted to ameliorate disease. *Med N Y N.* (2021) 2:864–883.e9. doi: 10.1016/j.medj.2021.04.013

113. Zaiss MM, Joyce Wu HJ, Mauro D, Schett G, Ciccia F. The gut-joint axis in rheumatoid arthritis. Nat Rev Rheumatol. (2021) 17:224-37. doi: 10.1038/s41584-021-00585-3