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RECEIVED 06 August 2024
ACCEPTED 14 November 2024
PUBLISHED 02 December 2024

CITATION
Guo D, Yan J, Yang Z, Chen M, Zhong W,
Yuan X and Yu S (2024) The immune
regulatory role of exosomal miRNAs and their
clinical application potential in heart failure.
Front. Immunol. 15:1476865.
doi: 10.3389/fimmu.2024.1476865

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The immune regulatory role of exosomal miRNAs and their clinical application potential in heart failure

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Heart failure (HF) is a complex and debilitating condition characterized by the heart's inability to pump blood effectively, leading to significant morbidity and mortality. The abnormality of immune response is a key factor in the progression of HF, contributing to adverse cardiac remodeling and dysfunction. Exosomal microRNAs (miRNAs) play a pivotal role in regulating gene expression and cellular function, which are integral to the crosstalk between cardiac and immune cells, influencing immune cell functions, such as macrophage polarization, T cell activity, and cytokine production, thereby modulating various pathological processes of HF, such as inflammation, fibrosis, and cardiac dysfunction. This review emphasizes the immune-regulatory role of exosomal miRNAs in HF and highlights their clinical potential as diagnostic biomarkers and therapeutic agents.

KEYWORDS

heart failure, exosome, miRNA, immune modulation, macrophage polarization

1 Introduction

Heart failure (HF) is a multifactorial syndrome characterized by the heart's inability to pump sufficient blood to meet the body's metabolic demands, remaining a leading cause of morbidity and mortality worldwide (1). The global prevalence of heart failure is estimated to be approximately 64 million people, with a prevalence rate of 1-2% in the general adult population (2). The morbidity and mortality rates of HF vary across different age groups, with a marked increase in the incidence and mortality as age advances. Among middle-aged adults, the incidence rate increases sharply, with estimates ranging from 4.7% to 13.3% (3). Compared with young adults, mortality rates are higher in middle-aged adults, older adults, and the elderly, with a 1-year mortality rate of 17.1%, 24.7%, and 39.9%, respectively (4). The pathophysiology of HF is complex and involves neurohormonal

dysregulation, myocardial remodeling, and immunological abnormalities (5). The hyperactivity of renin-angiotensin-aldosterone system causes increased fluid retention and worsening cardiac function (6). Structural changes in the myocardium, such as fibrosis and hypertrophy, lead to ventricular stiffening and impaired relaxation (7). Among these factors, dysfunction of immune system plays a crucial role in the progression of HF, contributing to adverse cardiac remodeling, fibrosis, and ventricular dysfunction (8). With the progression of HF to advanced stage, patients fail to respond adequately to standard pharmacological treatments, often requiring additional interventions such as diuretics and inotropes, which can have limited long-term efficacy and potential adverse effects (9). Understanding the regulatory mechanisms governing immune modulation in HF is vital for developing novel therapeutic strategies.

Exosomes are small extracellular vesicles (EVs) ranging from 30 to 150 nm in diameter that are responsible for cell-to-cell communication (10). Exosomes are released by almost all cell types and carry diverse cargoes, including proteins, lipids, and nucleic acids, which reflect the physiological state of their cell of origin (11). Exosomal microRNAs (miRNAs), short non-coding RNA molecules that post-transcriptionally regulate gene expression by binding to target messenger RNAs (mRNAs), have emerged as critical regulators of gene expression and cellular function (12). During HF progression, exosomal miRNAs play crucial roles in various biological processes, including inflammation, apoptosis, and cellular remodeling, participating in cardiac hypertrophy and fibrosis (11). Moreover, their stability in circulation enhances the utility as reliable indicators of various pathological states of HF (13). Thus, exosomal miRNAs may serve as promising markers and therapeutic targets in HF. Notably, exosomal miRNAs are pivotal in orchestrating the complex interplay between cardiac and immune cells by modulating various immune cell functions, including macrophage polarization, dendritic cell (DC) maturation, T cell response, and cytokine production, thereby modulating the inflammatory response (14–16). Numerous studies have identified specific exosomal miRNA signatures associated with HF, suggesting their potential as non-invasive diagnostic biomarkers (17, 18). Furthermore, the therapeutic application of stem cell-derived exosomal miRNAs is an area of active investigation, as they have shown potential in preclinical models to reduce inflammation, mitigate fibrosis, and promote cardiac repair (19, 20).

This review concisely summarizes the biogenesis, processing, and functions of exosomes and exosomal miRNAs, as well as focuses on their immune regulatory roles of exosomal miRNAs in HF. It discusses the interplay between exosomal miRNAs and various immune cells, and their impact on the inflammatory milieu in HF. Additionally, it explores the clinical application potential of exosomal miRNAs, highlighting their potential as diagnostic biomarkers and therapeutic agents.

2 Biogenesis and biological functions of exosomes

The biogenesis of exosomes involves several molecular mechanisms that ensure proper formation, cargo loading, and release of EVs. This process begins with the formation of early endosomes, which mature into multivesicular bodies (MVBs). These MVBs contain

intraluminal vesicles (ILVs) that are eventually released as exosomes upon fusion with the plasma membrane (21). The endosomal sorting complex required for transport (ESCRT) machinery, a multi-protein complex involved in the biogenesis of MVB, facilitates the sorting of ubiquitinated proteins into ILVs (22). Several specific molecules, such as tetraspanins and lipid rafts, contribute to the selective loading of cargo into exosomes, including specific proteins, lipids, nucleic acids, and other molecules, which ensure that exosomes carry distinct molecular signatures that dictate their functional roles (23). The transport and fusion of MVB are required for exosome release. The transport of MVBs to the plasma membrane is mediated by the cytoskeletal elements and motor proteins. Rab GTPases, particularly Rab35, are crucial regulators of MVB trafficking and fusion with the plasma membrane, facilitating exosome release (24). Moreover, fusion of MVBs with the plasma membrane is a tightly controlled process that involves soluble N-ethylmaleimide-sensitive factor attachment protein receptor (SNARE) proteins and other fusion machinery components, ensuring the timely release of exosomes into the extracellular space (25).

Exosomes exert pivotal biological functions in intercellular communication by transporting a diverse array of bioactive molecules between cells. Exosomes facilitate the transfer of metabolic and genetic information, thereby influencing the behavior and function of recipient cells. Their roles are particularly significant in immune modulation, inflammation, and cancer progression. For instance, exosomes can modulate immune responses by transporting antigens and other immune-related molecules, thereby influencing immune cell activation and response (26). They can also carry pro-inflammatory molecules that exacerbate inflammatory responses and are involved in the pathogenesis of inflammatory diseases (27). Additionally, exosomes are involved in cancer dynamics. They can promote tumor growth and metastasis by transferring oncogenic proteins and RNAs, facilitating processes such as epithelial-mesenchymal transition, and enhancing drug resistance (28). Thus, exosome-mediated communication by delivering its molecular cargo to recipient cells, plays a vital role in various physiological and pathological processes. While the cargo composition of exosomes is diverse and context-dependent, it is essential to consider the heterogeneity within exosome populations. Different subpopulations of exosomes can carry distinct sets of molecules that may exert various biological functions.

3 An overview of exosomal miRNAs

MiRNAs are small, non-coding RNA molecules that play critical roles in the regulation of gene expression. They have garnered significant attention owing to their involvement in various physiological and pathological processes, including cardiovascular diseases (29). Exosomes have emerged as crucial vehicles for intercellular communication and transport a variety of bioactive molecules, including miRNAs (30). Exosomal miRNAs have the potential to influence recipient cell function and behavior, making them important players in cell-to-cell communication and disease pathogenesis.

3.1 Biogenesis of miRNAs

The biogenesis of miRNAs is a complex, multi-step process involving both canonical and non-canonical pathways, which are tightly regulated to ensure proper gene expression and cellular function (Figure 1). Initially, miRNAs are transcribed as primary miRNAs (pri-miRNAs) by RNA polymerase II, which are then processed in the nucleus by the microprocessor complex, consisting of the endoribonuclease Drosha and its cofactor DGCR8, resulting in precursor miRNAs (pre-miRNAs) (31). These pre-miRNAs are subsequently exported to the cytoplasm by Exportin-5, where they undergo further cleavage by another endoribonuclease, Dicer, to form miRNA duplexes (32). The miRNA duplexes are then loaded onto Argonaute (AGO) proteins to form the RNA-induced silencing complex (RISC), where one strand of the duplex is retained as mature miRNA, while the other strand is typically degraded (33). This mature miRNA within the RISC complex can then bind to complementary sequences in the 3' untranslated regions (UTRs) of target mRNAs, leading to mRNA degradation or translational repression, thereby post-transcriptionally regulating gene expression (34).

In addition to the canonical pathway, non-canonical pathways also exist, in which miRNAs can be processed independently of Drosha, DGCR8, or Dicer. For instance, mirtrons are a class of miRNAs that bypass Drosha processing and are spliced directly from introns (35). The regulation of miRNA biogenesis is further modulated by various RNA-binding proteins (RBPs) and other

regulatory factors that can enhance or inhibit different stages of miRNA processing. These factors include protein phosphorylation events that link miRNA biogenesis to cellular signaling pathways, and epigenetic modifications such as DNA methylation and histone modifications that influence the transcription and processing of miRNAs (36, 37). Additionally, the stability and turnover of miRNAs are controlled by mechanisms such as RNA methylation and uridylation, which can affect the half-life of miRNAs and their ability to regulate their target mRNAs (38). The biogenesis of miRNAs is not only crucial for normal cellular processes but also has significant implications in disease states. Understanding the detailed mechanisms of miRNA biogenesis, including both canonical and non-canonical pathways, and the regulatory networks involved, provides valuable insights into their roles in HF.

3.2 Exosomal miRNA sorting and transport

The sorting of miRNAs into exosomes is a highly selective and regulated process (Figure 1). Several main mechanisms have been proposed for the sorting of miRNAs into exosomes. The first is the involvement of specific RBPs such as heterogeneous nuclear ribonucleoproteins (hnRNPs) and Ago2, which recognize and bind to specific sequence motifs in miRNAs, facilitating their sorting into exosomes (39, 40). Furthermore, membranous proteins involved in EV biogenesis such as caveolin-1 and neutral sphingomyelinase 2, also play

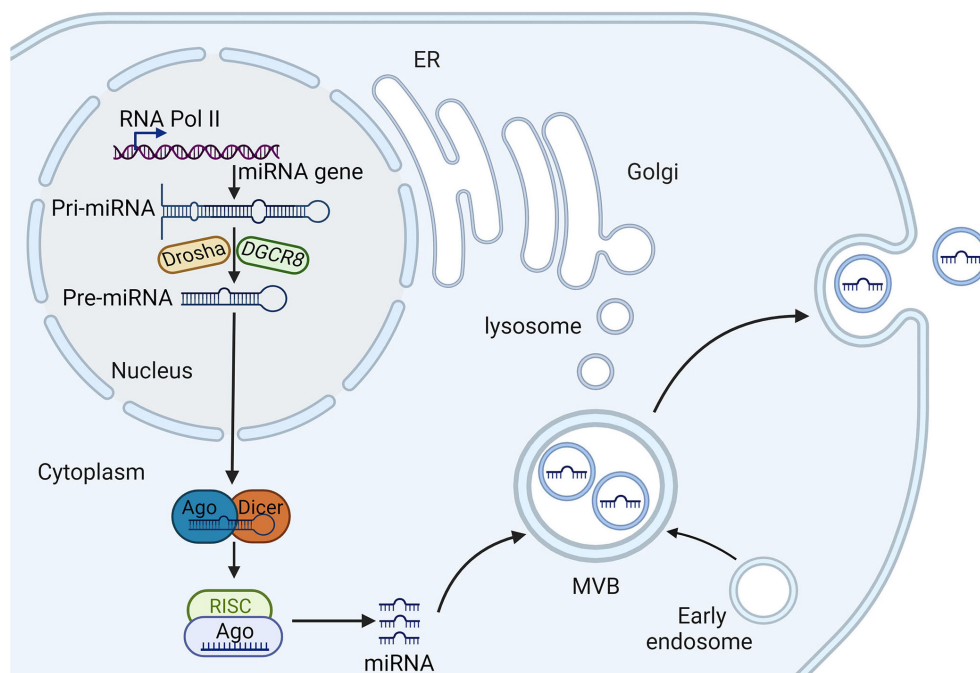


FIGURE 1

The biogenesis and sorting of exosomal miRNA. RNA polymerase II transcribes miRNA genes, generating pri-miRNA, which is processed in the nucleus by Drosha and DGCR8, producing pre-miRNA. Pre-miRNA is then transported to the cytoplasm and is further processed in the cytoplasm by Dicer, generating mature miRNA, which is incorporated into the target gene. Early endosomes are generated by endocytosis of parent cell and then undergo the second invagination of the plasma membrane, thus forming, which are subsequently either degraded or released from the cell as exosomes. Several proteins are responsible for miRNA entry into the exosomes, such as heterogeneous nuclear ribonucleoproteins and neutral sphingomyelinase 2. miRNA, microRNA; AGO, argonaute; pre-miRNAs, precursor miRNAs; pri-miRNAs, primary miRNAs; RISCs, RNA-induced silencing complexes; MVBs, multivesicular bodies.

crucial roles in the selective sorting of miRNAs into EVs (41, 42). Besides, the selective loading of miRNAs into exosomes is mediated by specific sequence motifs. For example, the EXOmotif, a short sequence present in some miRNAs, has been identified as a determinant for miRNA sorting into exosomes (43). Components of the ESCRT complex, such as Alix and TSG101, have been implicated in the incorporation of specific miRNAs into exosomes (44).

Once exosomes are formed and loaded with miRNAs, they are released into the extracellular space through the fusion of MVB with the plasma membrane (45). Exosomal miRNAs are protected from degradation by RNases in the extracellular environment owing to their encapsulation within the lipid bilayer of exosomes. This protection enhances the stability of miRNAs, allowing them to be transported over long distances and to remain functional upon delivery to recipient cells (46). Exosomal miRNAs can then be taken up by recipient cells through various mechanisms, including endocytosis, phagocytosis, and direct fusion with the plasma membrane (47). The specific mechanism of uptake depends on the cell type and the surrounding microenvironment.

3.3 Exosomal miRNA functions

Exosomal miRNAs can regulate gene expression in recipient cells by binding to complementary sequences in target mRNAs, leading to mRNA degradation or translational repression (48). This regulatory function allows exosomal miRNAs to influence various cellular processes, including proliferation, differentiation, apoptosis, and immune responses (49). Besides, exosomal miRNAs play a ligand-like role in recipient cells, in which they function as direct agonists of specific receptor families by interacting with proteins, thereby affecting cellular processes and disease progression (50). Therefore, exosomal miRNAs play a critical role in intercellular communication by transferring genetic information between cells, thereby modulating the behavior of recipient cells, contributing to pathological processes of various diseases. Understanding the biogenesis, sorting, transport, and function of exosomal miRNAs is essential for elucidating their roles in HF.

While the general biological processes for exosomal miRNA biogenesis and function are understood, the identification of sorting signals that determine which miRNAs are packaged into exosomes is still incomplete. Also, the precise mechanisms by which exosomal miRNAs mediate intercellular communication and influence recipient cell behavior are not fully understood.

4 Immunomodulatory roles of exosomal miRNAs from different cell types

Exosomal miRNAs play significant immunomodulatory roles in HF by influencing various pathological processes, such as myocardial hypertrophy, injury, infarction, and ventricular remodeling. Various cell-derived exosomal miRNAs are involved in modulating immune responses, inflammation, and tissue repair

by regulating the expression of genes related to various cellular processes and signaling pathways in HF (Figure 2).

4.1 Cardiac cells

Exosomal miRNAs from cardiac cells play a crucial role in modulating immune responses during HF progression. These miRNAs are involved in mediating intercellular communication and influencing immune cell behavior. For example, miR-146a-5p, when encapsulated in exosomes, is capable of activating various cardiac cell types, including cardiac fibroblasts and cardiomyocytes. This activation contributes to myocardial inflammation and cardiomyocyte dysfunction through the engagement of Toll-like receptor (TLR) 7 in nucleic acid sensing. Consequently, this process leads to the activation of immune cells, such as CD45⁺ leukocytes, monocytes, and neutrophils, as well as the subsequent production of proinflammatory cytokines, including C-X-C motif ligand (CXCL) 2, interleukin (IL)-6, and tumor necrosis factor (TNF)- α , which exacerbates cardiac inflammation (14). Indeed, compared to healthy individuals, the level of exosomal miR-146a-5p in patients with myocardial infarction is decreased. Further mechanistic evaluations revealed that exosomal miR-146a-5p derived from cardiomyocytes can modulate macrophage polarization, promoting M1 macrophage polarization and inhibiting M2 polarization, which in turn induces an inflammatory reaction; whereas, miR-146a-5p can downregulate the expression of TNF receptor-associated factor 6 to exert an anti-inflammatory effect (51). Similarly, when cardiomyocytes are exposed to hypoxic conditions, exosomal miRNAs, including miR-10a, miR-10b, miR-143, and miR-423, are elevated to induce IL-6 expression in macrophages, thus aggravating cardiac inflammation (52). Exosomes from hypertrophic cardiomyocytes also play a role in immune modulation by triggering the secretion of inflammatory cytokines IL-6 and IL-8 in macrophages. Notably, miR-155 has been identified as a crucial factor in initiating the inflammatory response, as it activates key signaling pathways such as extracellular signal-regulated kinase (ERK), c-Jun N-terminal kinase (JNK), and p38 (53). Consequently, these findings suggest that exosomal miRNAs influence macrophage phenotype and behavior to modulate immune responses, contributing to the repair process of infarcted myocardium.

Exosomal miRNAs from cardiosphere-derived cells (CDCs) are involved in modulating immune responses in HF. It has been shown that CDCs-derived exosomal miRNAs carry clusters of immune-related and cardiac-related molecular biomarkers, suggesting their significant roles in immune modulation and cardiac repair (54). In rat and pig models of myocardial infarction, exosomal transfer of miR-181b from CDCs into macrophages can modify their polarization state and confer cardioprotective efficacy by reducing the expression of protein kinase C δ (55). Following ischemic myocardial injury, exosomal miR-26a secreted by CDCs polarizes macrophages toward the phenotype with enhanced phagocytic capacity, which enhances macrophage efferocytosis and cardioprotection by sustaining the expression of MerTK and complement factor C1q, a phagocytosis facilitator (56). Likewise, exosomes from CDCs attenuate

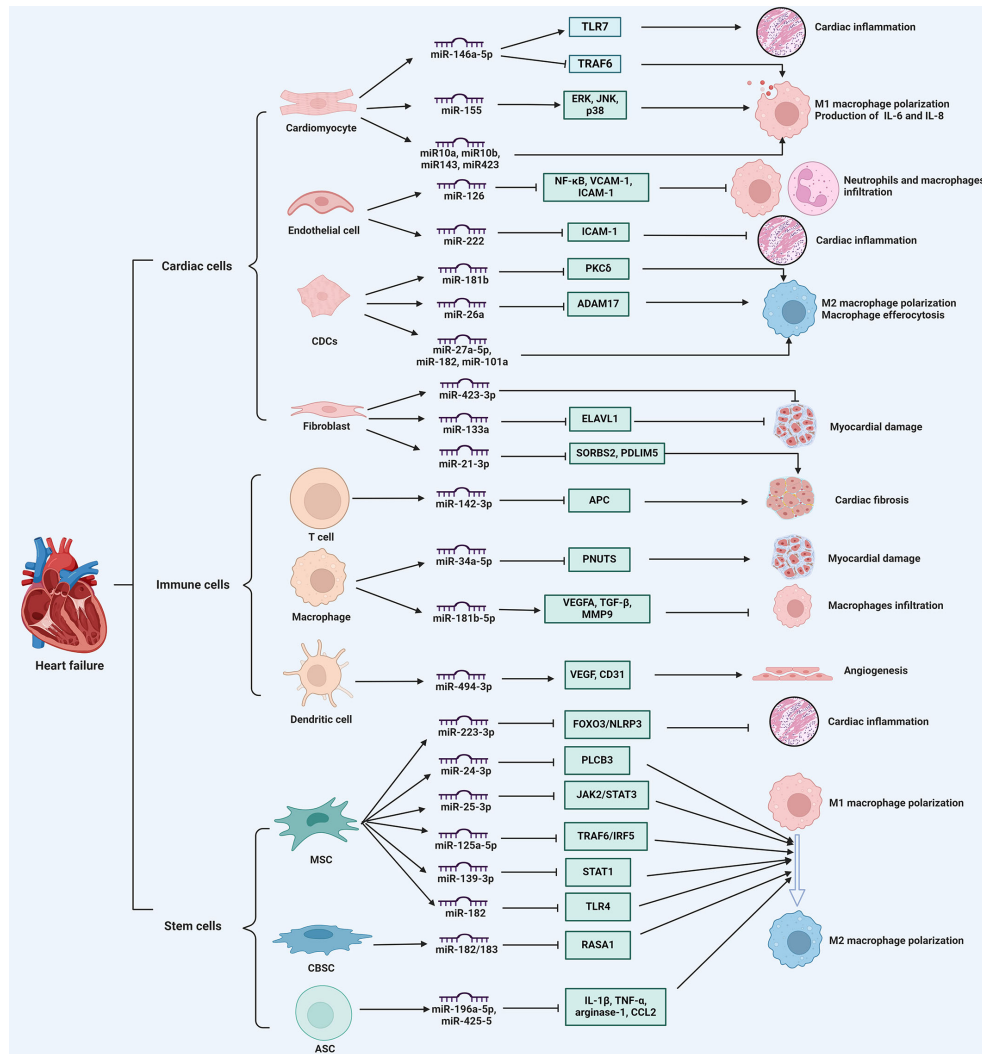


FIGURE 2

The immune regulatory role of exosomal miRNAs in heart failure. exosomal miRNAs derived from various cell types, including cardiac cells, immune cells, and stem cells, regulate immune cell infiltration, cytokine secretion, macrophage phenotype switch, thereby influencing myocardial inflammation, cardiomyocyte apoptosis, angiogenesis, and cardiac fibrosis, ultimately participating in the progression of heart failure. ADAM17, a disintegrin and metalloproteinase 17; Ang II, angiotensin II; APC, adenomatous polyposis coli; ASCs, adipose-derived stem cells; CBSCs, cortical bone stem cells; CDCs, cardiosphere-derived cells; ELAVL1, drosophila like RNA-binding protein 1; ERK, extracellular signal-regulated kinase; IL, interleukin; JNK, c-Jun N-terminal kinase; MSCs, mesenchymal stem cells; NFAM1, NFAT activating molecule 1; PDLIM5, PDZ and LIM domain 5; PKCδ, protein kinase C δ; PNUMS, serine/threonine-protein phosphatase 1 regulatory subunit 10; RASA1, ras p21 protein activator 1; SORBS2, sorbin and SH3 domain-containing protein 2; STAT1, signal transducer and activator of transcription 1; TLR4, toll-like receptor 4; TRAF6, TNF receptor-associated factor 6. ⊥ indicates an inhibitory effect and → indicates a promoting effect.

macrophage infiltration and myocardial infarct size, and improve cardiac function by transferring miR-27a-5p, miR-182, and miR-101a to macrophages and regulating M2 macrophage polarization after myocardial ischemia/reperfusion (I/R) injury (57). Overall, exosomal miRNAs from CDCs play multifaceted roles in modulating immune responses, exhibiting their potential therapeutic applications in the management and treatment of HF.

Additionally, exosomal miRNAs derived from endothelial cells modulate immune cell functions and affect various pathological processes of HF. It is reported that miR-222 from endothelial microparticles reduces the expression of intercellular adhesion

molecule (ICAM)-1 in recipient endothelial cells, which diminishes monocyte adhesion and promotes anti-inflammatory effects, protecting against cardiac dysfunction from myocardial infarction (58). Moreover, in septic mice, elevated miR-126 levels in endothelial cells mitigate severe cardiomyopathy and cardiac dysfunction by reducing the expression of adhesion molecules vascular cell adhesion molecule 1 and ICAM-1, thus further decreasing the accumulation of immune cells in the myocardium and inflammatory cytokine secretion (59). Hence, endothelial cell-derived exosomal miRNAs play an immunomodulatory role in influencing immune cell behaviors and cardiac inflammation during HF progression.

Fibroblast-derived exosomal miRNAs also regulate cardiac remodeling and fibrosis in HF. For instance, miR-21-3p, a passenger strand miRNA enriched in fibroblast-derived exosomes, induces cardiomyocyte hypertrophy by targeting proteins such as sorbin and SH3 domain-containing protein 2 and PDZ and LIM domain 5, suggesting its involvement in pathological cardiac remodeling (60). During the acute phase of ischemia-reperfusion injury, miR-423-3p expression is enhanced in fibroblast-derived exosomes, which increases cell viability and reduces apoptosis in cardiomyocytes under hypoxia/reperfusion condition (61). Further studies revealed that fibroblast-derived exosomal miR-133a protects cardiomyocytes against myocardial ischemia/reperfusion-induced injury through suppression of pyroptosis by suppressing the expression of drosophila like RNA-binding protein 1 (62). Hence, fibroblast-derived exosomal miRNAs predominantly exhibit cardioprotective effects, their dysregulation can contribute to pathological remodeling and cardiac fibrosis in HF.

4.2 Immune cells

Immune cells, particularly macrophages and DCs, have been implicated in the progression of HF through their involvement in inflammation and tissue remodeling (63, 64). In addition to resident immune cells, peripheral immune cells are recruited to participate in inflammatory processes and pathological remodeling in response to cardiac injury (65). In chronic HF patients, overexpressed exosomal miR-126 in plasma is associated with increased levels of IL-10, ICAM-1 and TNF- α in activated peripheral blood mononuclear cells (PBMCs) (66). In fact, circulating exosomes from experimental autoimmune myocarditis selectively load abundant miR-142, which causes CD4⁺ T cell immunometabolic dysfunction and exacerbates cardiac injury via repressing the methyl-CpG binding domain protein 2 and suppressor of cytokine signaling 1 (15). Also, in an acute myocardial infarction mouse model, exosomal miR-342-3p is suppressed to increase the expression of NFAT activating molecule 1, which is linked to DC maturation and cardiac inflammation (16). Besides, circulating exosomal miRNAs derived immune cells are overexpressed in HF patients, indicating the activation of peripheral immune system. These exosomal miRNAs, particularly those secreted by monocytes, macrophages, DCs, and T cells, are correlated with disease severity during HF progression (67). Thus, circulating exosomal miRNAs derived from immune cells can regulate immune responses and influence disease outcomes.

Macrophages, derived from tissue-resident cells or circulating monocytes, undergo significant phenotypic changes following cardiac injury, playing crucial roles in the inflammatory response and tissue repair in HF (68). The polarization of macrophages into pro-inflammatory (M1) and anti-inflammatory (M2) types is regulated by exosomal miRNAs, which can either exacerbate or mitigate inflammation and subsequent cardiac damage (69). For instance, during I/R-induced myocardial injury, exosomal miR-155-5p activates the intracellular JAK/STAT1 pathway in

macrophages, promoting M1 macrophage proliferation and exacerbating cardiac inflammation (70). Also, in I/R-induced pig models of myocardial infarction, exosomal miR-181b-5p from M2 macrophages suppresses the number of CCR2⁺ macrophages in favor of the polarization of macrophages toward the M2 phenotype, which facilitates the migration and pro-angiogenic vascular endothelial growth factor (VEGF) expression by endothelial cells, leading to improved angiogenesis and cardioprotective effect (71). Similarly, the upregulated levels of exosomal miR-34a-5p in programmed cell death 1 inhibitor-treated macrophages can mediate cardiomyocyte injury and cardiac senescence through inhibiting the expression of serine/threonine-protein phosphatase 1 regulatory subunit 10 (72). These findings imply that exosomal miRNAs derived from macrophages mediate inflammatory responses and myocardial damage, which are key factors in adverse cardiac remodeling and HF progression. In addition, as key modulators of immune responses and inflammation, DCs are antigen-presenting cells that contribute to systemic immune surveillance and regulate the inflammatory response during the development of cardiac fibrosis in HF (73). It has been demonstrated that exosomal miR-494 3p derived from DCs enhances the expression of VEGF in the infarcted myocardium of myocardial infarction (MI) model mice, which facilitates tube formation by cardiac microvascular endothelial cells, thereby promoting angiogenesis post-MI (74). Moreover, studies have shown that CD4⁺ T cells in HF patients exhibit increased activation and clonal expansion, which is associated with elevated levels of pro-inflammatory cytokines such as IL-6 and TNF- α , further contributing to adverse cardiac remodeling and sustained inflammation (75). Exosomal miRNAs from CD4⁺ T cells play a crucial role in the development of HF. For example, exosomes derived from activated CD4⁺ T cells have been shown to evoke pro-fibrotic effects in cardiac fibroblasts, thereby aggravating cardiac fibrosis and dysfunction post-infarction. This is primarily mediated through miR-142-3p, which is enriched in exosomes from CD4⁺ T cells and targets the adenomatous polyposis coli gene, a negative regulator of the WNT signaling pathway, thus promoting cardiac fibroblast activation and fibrosis (76). Overall, the evidence strongly supports the crucial role of exosomal miRNAs from immune cells, including macrophages, DCs, and CD4⁺ T cells in the development and progression of HF, making them a focal point for future research and clinical applications in this disease.

4.3 Stem cells

Stem cells, particularly mesenchymal stem cells (MSCs), have emerged as a promising therapeutic option for improving cardiac function in patients with HF. They have been shown to promote angiogenesis and the formation of new blood vessels, which improves blood supply to the ischemic heart tissue and supports myocardial repair (19). Of importance, stem cells are involved in the paracrine effect, where they secrete various growth factors, cytokines, and exosomes that modulate the local cardiac environment, reduce

inflammation, and inhibit apoptosis of existing cardiomyocytes (77). Exosomes secreted from MSCs can induce the differentiation of pro-inflammatory macrophages towards a pro-resolving phenotype, reduce generation of pro-inflammatory cytokines, and facilitate the resolution of inflammation within the myocardial infarct area (78). By entrapping MSC-derived exosomes onto the surface of the elastomeric scaffolds, the number of M2 macrophages is enhanced to improve myocardial regeneration during the inflammatory and wound healing responses (79). Thus, exosomes derived from stem cells play a significant role in modulating immune responses, which is crucial for the therapeutic management of HF.

Specifically, exosomal miRNAs from MSCs have shown promising cardioprotective effects by modulating macrophage polarization. For instance, exosomes derived from MSCs are confirmed to enhance M2 macrophage polarization and further reduce inflammation and infarct size in a mouse model of myocardial I/R, primarily through the action of miR-182, which targets the toll-like receptor 4 pathway (80). Similarly, exosomes from nicorandil-pretreated MSCs have been shown to promote cardiac repair post-myocardial infarction by upregulating miR-125a-5p, which inhibits the TRAF6/IRF5 signaling pathway, thereby enhancing M2 polarization (81). Additionally, exosomes from human umbilical cord MSCs are verified to improve cardiac function by increasing M2 macrophage polarization through the downregulation of phospholipase C beta 3 and inactivation of the nuclear factor kappa B (NF- κ B) pathway, mediated by miR-24-3p (82). These findings suggest exosomal miRNAs from MSCs can promote M2 polarization and reduce cardiac inflammation, further aiding heart repair in HF. Furthermore, exosomes from MSCs repress M1 polarization by miRNA-mediated cellular signaling and stress responses in HF. It is reported that bone marrow MSC-derived exosomal miR-25-3p confers cardioprotective effects in I/R-induced MI mice and downregulates proinflammatory cytokines in LPS-stimulated macrophages by inactivating the JAK2/STAT3 signaling pathway and subsequent inhibiting M1-like macrophage polarization (83). Consistent with this result, exosomal miR-139-3p from MSCs reduces the amount of M1 macrophages and facilitates cardiac repair in acute MI through suppressing downstream signal transducer and activator of transcription 1 (84). Of interest, in addition to macrophages, MSC-derived exosomal miRNAs like miR-223-3p can also be transferred into cardiomyocytes to inhibit LPS-induced cardiomyocyte inflammation and pyroptosis, which helps alleviate immune cell infiltration, inflammatory cytokine secretion, and cardiac dysfunction (85). Therefore, exosomal miRNAs derived from MSCs exert immunomodulatory effects via affecting the macrophage polarization and ameliorating cardiac inflammation in HF.

Moreover, other stem cells, like cortical bone and adipose stem cells, is verified to secrete exosomal miRNAs to exert cardioprotective effects in HF. In the myocardium post-MI model mice, cortical bone stem cell-derived exosomes which are enriched in miR-182/183, promote the polarization of macrophages and T cells towards a pro-reparative phenotype, which enhances oxidative phosphorylation rate and reduces reactive oxygen species (ROS) via the Ras p21 protein activator 1 axis, thereby improving structural remodeling and

cardiac function (86). Besides, exosome-derived miR-196a-5p and miR-425-5p from adipose stem cells halt ischemia-induced mitochondrial dysfunction and ROS production in cardiomyocytes, and promote angiogenesis by polarizing macrophages toward the anti-inflammatory M2 immunophenotype (87).

In conclusion, exosomal miRNAs derived from various cell types, including cardiac cells, immune cells, and stem cells, represent a versatile and potent class of molecules with potential to modulate immune responses and influencing HF outcomes (Figure 2). These exosomal miRNAs are involved in macrophage polarization, DC maturation, and T cell response, thereby modulating inflammatory reactions and cardiac remodeling by acting on various molecular targets and signaling pathways, highlighting their potential as multifaceted therapeutic agents. Identification of exosomal miRNA signatures as biomarkers for HF diagnosis and prognosis is a promising approach. Integrating exosomal miRNA profiling into clinical practice could enhance the precision of HF management. Exosomal miRNAs derived from various cell types, such as cardiomyocytes, fibroblasts, endothelial cells, and immune cells, exert immunomodulatory effects and further affect cardiac structures and functions in HF. However, the functional diversity of exosomal miRNAs in regulation of immune responses in HF requires further investigation. Besides, temporal changes in exosomal miRNA profiles during the progression of HF and their correlation with disease stages and outcomes are not thoroughly explored.

5 Clinical application potential of exosomal miRNAs in HF

5.1 Diagnostic markers

Exosomal miRNAs are stable within the circulatory system due to their encapsulation in EVs, which protect them from degradation, making them reliable candidates for diagnostic purposes (17). The analysis of exosomal miRNAs in plasma or serum holds significant promise as a non-invasive biomarker for the early diagnosis of HF (18). A comprehensive characterization of miRNAs in plasma exosomes from dilated cardiomyopathy (DCM) patients with chronic HF identified 92 differentially expressed miRNAs correlated with several enriched pathways, suggesting their potential roles in the pathogenesis and clinical management of DCM (13). Research on patients with acute HF owing to DCM revealed exosomal-derived miR-92b-5p is upregulated, highlighting its potential as a non-invasive diagnostic biomarker (88). Similarly, it is reported that levels of miR129-5p isolated from plasma microvesicles are inversely related to the degree of clinical HF in patients with univentricular heart disease, indicating that miR129-5p is a sensitive and specific biomarker for this disease independent of ventricular morphology or palliation stage (89). Besides, the diagnostic potential of exosomal miRNAs extends beyond HF to other cardiovascular complications, such as HF with preserved ejection fraction in diabetic patients, where miR-30d-5p and miR-126a-5p are found to be correlated with reduced cardiac output and exosomal expression, supporting their release from the heart and

association with diabetic HF with preserved ejection fraction (90). Likewise, in HF patients with reduced ejection fraction, elevated circulating exosomal miR-92b-5p is positively correlated with left atrial diameter, left ventricular diastolic diameters, and systolic diameters, acting as a promising diagnostic biomarker for this disease (91). The identification of exosomal miRNAs like miR-7856-5p in HF patients with depressive symptoms further underscores its diagnostic value, showing high sensitivity and specificity for severe depressive symptoms among HF patients (92). Moreover, dysregulated exosomal miRNAs reflect pathological processes during HF progression. For example, it has been shown that plasma exosomal miR-425 and miR-744, which serve as negative regulators of cardiac fibrosis by suppressing TGF- β 1 expression, possess the potential to be a biomarker to predict cardiac fibrosis and HF (93). Consistently, circulating exosomal miR-146a is upregulated in response to systemic inflammation, implying its potential role in reflecting the inflammatory state of HF (94). As for central inflammation in chronic HF, circulating exosomal let-7g-5p and let-7i-5p are confirmed to reduce the neuroinflammation, which is associated with sympathetic hyperactivity in chronic HF (95). Of interest, three p53-responsive exosomal miRNAs (miR-194, miR-34a, and miR-192), which function as circulating regulators of HF development via the p53 pathway, are elevated by the early convalescent stage of acute MI, underscoring their diagnostic value, with a developed three-circulating exosomal miRNA model predicting the risk of development of ischemic HF after acute MI (96).

Taken together, dysregulated exosomal miRNAs indicate their potential as diagnostic factors for HF (Table 1). However, there are still challenges in identifying optimal exosomal miRNA candidates for early diagnosis and screening in patients. A promising strategy is to apply next-generation sequencing to winnow the vital exosomal miRNAs in these patients with different disease conditions and stages. Thus, further clarification of exosomal miRNA-related markers could provide a new angle for prognostic indicators of HF.

TABLE 1 Exosomal miRNAs act as potential diagnostic markers in HF.

Exosomal miRNAs	Expression	Clinical relevance	Ref.
miR-92b-5p	Up	DCM-related acute HF	(88)
miR129-5p	Down	HF with univentricular heart disease	(89)
miR-30d-5p, miR-126a-5p	Down	Diabetic HF with preserved ejection fraction	(90)
miR-92b-5p	Up	HF with reduced ejection fraction	(91)
miR-7856-5p	Up	HF with depressive symptoms	(92)
miR-425, miR-744	Down	HF with cardiac fibrosis	(93)
miR-146a	Up	HF with systematic inflammation	(94)
let-7g-5p, let-7i-5p	Down	HF with neuroinflammation	(95)
miR-194, miR-34a, miR-192	Up	Ischemic HF with acute MI	(96)

5.2 Therapeutic targets

Exosomal miRNA-based therapy holds significant potential in reversing cardiac remodeling and improving cardiac function in HF patients. For instance, in an MI rat model, exosomes loaded with miR-126 and miR-146a mimics promote the expression of vascular endothelial growth factor and tube formation, which helps to decrease infarct size and facilitate angiogenesis, as well as reduces the production of collagen fibers and cardiac fibrosis (97). By delivering exosomal miRNA-21 into cardiomyocytes and endothelial cells, cell apoptosis and cardiac function impairment are inhibited in a preclinical MI animal model (98). Thus, exosomal miRNA-based therapy could revolutionize HF treatment by reversing cardiac remodeling and improving cardiac function.

Specifically, MSC-derived exosomes have shown promise in cardiac repair and regeneration owing to their ability to carry specific miRNAs that provide anti-apoptotic, anti-inflammatory, anti-fibrotic, and angiogenic effects within the infarcted heart (99). For example, miR-129-5p loaded into MSC-derived exosomes has been demonstrated to ameliorate oxidative stress, cell apoptosis, inflammation, and fibrosis in cardiomyocytes in mice with HF by inhibiting the TRAF3/NF- κ B signaling pathway, thereby alleviating ventricular dysfunction and improving cardiac function (100). Following myocardial I/R injury, MSC-derived exosomes act as tools for the specific delivery of miR-181a, which exert immune-suppressing effect and mitigate cardiac inflammation (101). Moreover, exosomal miRNAs derived from MSCs also alleviate MI-mediated cardiac dysfunction, thus delaying HF progression. It is reported that miR-200b-3p overexpressing MSC-derived exosomes enhance cardioprotective abilities by reducing

TABLE 2 Exosomal miRNAs derived from MSCs act as potential therapeutic targets in HF.

Exosomal miRNAs	Downstream targets	Outcomes	Ref.
miR-129-5p	TRAF3/NF- κ B	Inhibit oxidative stress, cell apoptosis, inflammation, and fibrosis in cardiomyocytes	(100)
miR-181a	Unknown	Exert immune-suppressing effect and mitigate cardiac inflammation	(101)
miR-200b-3p	BCL2L1	Reduce cardiomyocyte apoptosis, cardiac inflammation, and myocardial fibrosis	(102)
miR-302d-3p	BCL6/MD2/NF- κ B	Attenuate inflammation and cardiac remodeling and promote cardiac function recovery	(103)
miR-223	P53/S100A9	Relieve myocardial fibrosis and induce the angiogenesis	(104)
miR-223	SEMA3A, STAT3	Represses inflammation and cardiomyocyte death	(105)
miR-141	PTEN/ β -catenin	Ameliorate myocardial injury	(106)
miR-29a-3p	HMGB1/TLR4/AKT	Inhibit inflammatory response and improve cardiac function	(107)

cardiomyocyte apoptosis, cardiac inflammation, and myocardial fibrosis by suppressing BCL2L11 in MI mice (102). Similarly, exosomes from MSCs containing miR-302d-3p have been shown to impede inflammation and cardiac remodeling and promote cardiac function recovery in MI-treated mice by diminishing hypoxic cardiomyocyte apoptosis via the BCL6/MD2/NF- κ B axis (103). Also, human umbilical cord MSC-derived exosomes loaded with miR-223 can relieve myocardial fibrosis and inflammation infiltration, and induce angiogenesis through the P53/S100A9 axis, thereby promoting myocardial repair in MI rats (104). The therapeutic potential of MSC-derived exosomes extends beyond MI to sepsis-induced myocardial injury during HF progression. For instance, in CLP-induced septic mice, exosomal miR-223 derived from MSCs is transferred to cardiomyocytes, which represses inflammation and cell death via inhibiting the expression of SEMA3A and STAT3, alleviating sepsis-induced cardiac injury (105). In the same preclinical setting, bone marrow MSC-derived exosomal miR-141 is found to bind to ameliorate myocardial injury in septic mice through regulating the PTEN/ β -catenin axis (106). Intriguingly, in severe acute pancreatitis-induced rats, miR-29a-3p can be transferred into cardiomyocytes by MSC-derived exosomes, which inhibits inflammatory responses and improves cardiac function to attenuate myocardial injury via inactivating the HMGB1/TLR4/AKT signaling pathway (107).

Collectively, exosomal miRNAs postpone the pathological processes of HF, including mitigating cell apoptosis, oxidative stress, and inflammatory response, thereby improving cardiac function (Table 2). Thus, exosomal miRNA-based therapies may be beneficial for HF treatment by transferring protective miRNAs into myocardial injury areas. Accumulating evidence supports the use of MSC-derived exosomal miRNAs as a novel and effective therapeutic approach to improve cardiac function in patients with HF, offering a new avenue for treatment that leverages the natural regenerative capabilities of stem cells and their exosomal cargo. Despite these promising results, challenges such as understanding the interaction mechanisms with host cells, distribution within the body, metabolism, and long-term safety need to be addressed to optimize the clinical application of exosomal miRNA-based therapies.

6 Conclusion and future directions

This review emphasizes the multifaceted roles of exosomal miRNAs derived from cardiac cells, immune cells, and stem cells in modulating immune responses, including macrophage polarization, DC maturation, and T cell response during HF progression. Owing to their stability in bodily fluids and their specific expression profiles in HF patients, exosomal miRNAs have shown promise as non-invasive biomarkers for early diagnosis of HF. The therapeutic potential of exosomal miRNAs lies in their ability to alleviate pathological processes of HF, such as apoptosis, inflammatory responses, fibrosis, cardiac remodeling and cardiac dysfunction at the molecular level. Stem cell-derived exosomes loaded with specific miRNAs could offer targeted therapies, reducing adverse immune reactions and promoting cardiac repair. Despite the potential, several challenges need addressing, including the efficient delivery of exosomal miRNAs, ensuring their

stability and activity *in vivo*, and overcoming the potential for off-target effects. Future research should focus on a deeper understanding of the molecular mechanisms underlying the immunomodulatory functions of exosomal miRNAs. This includes identifying specific miRNA targets, understanding their interactions with immune cell receptors, and elucidating the signaling pathways involved. Besides, the development of standardized protocols for the isolation and characterization of exosomes and their miRNA cargo is crucial, which ensures reproducibility and comparability of results across different studies, facilitating the translation of research findings into clinical practice. Furthermore, extensive clinical studies are needed to validate the diagnostic potential of exosomal miRNAs. Large-scale, multi-center trials will help establish their clinical utility and reliability as biomarkers for HF. Importantly, the safety and efficacy of exosomal miRNA-based therapies must be thoroughly evaluated, including assessing potential off-target effects, immune reactions, and long-term outcomes.

Author contributions

DG: Writing – original draft, Writing – review & editing. JY: Methodology, Supervision, Writing – review & editing. ZY: Methodology, Supervision, Writing – review & editing. MC: Methodology, Supervision, Writing – review & editing. WZ: Methodology, Supervision, Writing – review & editing. XY: Methodology, Supervision, Writing – review & editing. SY: Funding acquisition, Methodology, Project administration, Supervision, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This work was supported by the Scientific Research Program of Chinese Medicine in Heilongjiang Province (No. ZHY2023-174) and the Scientific Research Program of Chinese Medicine in Heilongjiang Province (No. ZHY2020-148).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

- Redfield MM, Borlaug BA. Heart failure with preserved ejection fraction: A review. *JAMA*. (2023) 329:827–38. doi: 10.1001/jama.2023.2020
- Shahim B, Kapelios CJ, Savarese G, Lund LH. Global public health burden of heart failure: an updated review. *Card Fail Rev*. (2023) 9:e11. doi: 10.15420/cfr.2023.05
- Groenewegen A, Rutten FH, Mosterd A, Hoes AW. Epidemiology of heart failure. *Eur J Heart Fail*. (2020) 22:1342–56. doi: 10.1002/ehf.1858
- Teixeira A, Parenica J, Park JJ, Ishihara S, AlHabib KF, Laribi S, et al. Clinical presentation and outcome by age categories in acute heart failure: results from an international observational cohort. *Eur J Heart Fail*. (2015) 17:1114–23. doi: 10.1002/ehf.330
- Lam CSP, Docherty KF, Ho JE, McMurray JVV, Myhre PL, Omland T. Recent successes in heart failure treatment. *Nat Med*. (2023) 29:2424–37. doi: 10.1038/s41591-023-02567-2
- Maryam, Varghese TP, T B. Unraveling the complex pathophysiology of heart failure: insights into the role of renin-angiotensin-aldosterone system (RAAS) and sympathetic nervous system (SNS). *Curr Probl Cardiol*. (2024) 49:102411. doi: 10.1016/j.cpcardiol.2024.102411
- Barriault A, Iftikhar U, Stone JA. Cardiac rehabilitation in heart failure with reduced ejection fraction: pathophysiology, benefits and precautions. *Can J Cardiol*. (2024) 19:S0828–282X(24)01032–8. doi: 10.1016/j.cjca.2024.10.014
- Markousis-Mavrogenis G, Baumhove L, Al-Mubarak AA, Aboumsallem JP, Bomer N, Voors AA, et al. Immunomodulation and immunopharmacology in heart failure. *Nat Rev Cardiol*. (2024) 21:119–49. doi: 10.1038/s41569-023-00919-6
- Butta C, Roberto M, Tuttolomondo A, Petrantoni R, Miceli G, Zappia L, et al. Old and new drugs for treatment of advanced heart failure. *Curr Pharm Des*. (2020) 26:1571–83. doi: 10.2174/1381612826666191226165402
- Tang X, Leng M, Tang W, Cai Z, Yang L, Wang L, et al. The roles of exosome-derived microRNAs in cardiac fibrosis. *Molecules*. (2024) 29:1199. doi: 10.3390/molecules29061199
- Tian C, Ziegler JN, Zucker IH. Extracellular vesicle microRNAs in heart failure: pathophysiological mediators and therapeutic targets. *Cells*. (2023) 12:2145. doi: 10.3390/cells12172145
- Zheng D, Huo M, Li B, Wang W, Piao H, Wang Y, et al. The role of exosomes and exosomal microRNA in cardiovascular disease. *Front Cell Dev Biol*. (2020) 8:616161. doi: 10.3389/fcell.2020.616161
- Zhang L, Zhang G, Lu Y, Gao J, Qin Z, Xu S, et al. Differential expression profiles of plasma exosomal microRNAs in dilated cardiomyopathy with chronic heart failure. *J Cell Mol Med*. (2023) 27:1988–2003. doi: 10.1111/jcmm.17789
- Shimada BK, Yang Y, Zhu J, Wang S, Suen A, Kronstadt SM, et al. Extracellular miR-146a-5p induces cardiac innate immune response and cardiomyocyte dysfunction. *Immunohorizons*. (2020) 4:561–72. doi: 10.4049/immunohorizons.2000075
- Sun P, Wang N, Zhao P, Wang C, Li H, Chen Q, et al. Circulating Exosomes Control CD4(+) T Cell Immunometabolic Functions via the Transfer of miR-142 as a Novel Mediator in Myocarditis. *Mol Ther*. (2020) 28:2605–20. doi: 10.1016/j.jymthe.2020.08.015
- Zhu J, Chen Z, Peng X, Zheng Z, Le A, Guo J, et al. Extracellular Vesicle-Derived circITGB1 Regulates Dendritic Cell Maturation and Cardiac Inflammation via miR-342-3p/NFAM1. *Oxid Med Cell Longev*. (2022) 2022:8392313. doi: 10.1155/2022/8392313
- Zhou R, Wang L, Zhao G, Chen D, Song X, Momtazi-Borojeni AA, et al. Circulating exosomal microRNAs as emerging non-invasive clinical biomarkers in heart failure: Mega bio-roles of a nano bio-particle. *IUBMB Life*. (2020) 72:2546–62. doi: 10.1002/iub.2396
- Oh JG, Lee P, Gordon RE, Sahoo S, Kho C, Jeong D. Analysis of extracellular vesicle miRNA profiles in heart failure. *J Cell Mol Med*. (2020) 24:7214–27. doi: 10.1111/jcmm.15251
- Yahyazadeh R, Baradaran Rahimi V, Askari VR. Stem cell and exosome therapies for regenerating damaged myocardium in heart failure. *Life Sci*. (2024) 351:122858. doi: 10.1016/j.lfs.2024.122858
- Farahzadi R, Fathi E, Valipour B, Ghaffary S. Stem cells-derived exosomes as cardiac regenerative agents. *Int J Cardiol Heart Vasc*. (2024) 52:101399. doi: 10.1016/j.ijcha.2024.101399
- Han QF, Li WJ, Hu KS, Gao J, Zhai WL, Yang JH, et al. Exosome biogenesis: machinery, regulation, and therapeutic implications in cancer. *Mol Cancer*. (2022) 21:207. doi: 10.1186/s12943-022-01671-0
- Hessvik NP, Llorente A. Current knowledge on exosome biogenesis and release. *Cell Mol Life Sci*. (2018) 75:193–208. doi: 10.1007/s00018-017-2595-9
- Lee YJ, Shin KJ, Chae YC. Regulation of cargo selection in exosome biogenesis and its biomedical applications in cancer. *Exp Mol Med*. (2024) 56:877–89. doi: 10.1038/s12276-024-01209-y
- Zhang Y, Liu Y, Liu H, Tang WH. Exosomes: biogenesis, biologic function and clinical potential. *Cell Biosci*. (2019) 9:19. doi: 10.1186/s13578-019-0282-2
- Krylova SV, Feng D. The machinery of exosomes: biogenesis, release, and uptake. *Int J Mol Sci*. (2023) 24:36674857. doi: 10.3390/ijms24021337
- Kang J, Hua P, Wu X, Wang B. Exosomes: efficient macrophage-related immunomodulators in chronic lung diseases. *Front Cell Dev Biol*. (2024) 12:1271684. doi: 10.3389/fcell.2024.1271684
- Zhang R, Li M, Li H, Ran X, Jin F, Tan Q, et al. Immune cell-derived exosomes in inflammatory disease and inflammatory tumor microenvironment: A review. *J Inflammation Res*. (2024) 17:301–12. doi: 10.2147/JIR.S421649
- Zhao J, Li X, Liu L, Zhu Z, He C. Exosomes in lung cancer metastasis, diagnosis, and immunologically relevant advances. *Front Immunol*. (2023) 14:1326667. doi: 10.3389/fimmu.2023.1326667
- Schoettler FI, Fatehi Hassanabad A, Jadli AS, Patel VB, Fedak PWM. Exploring the role of pericardial miRNAs and exosomes in modulating cardiac fibrosis. *Cardiovasc Pathol*. (2024) 73:107671. doi: 10.1016/j.carpath.2024.107671
- Long M, Cheng M. Small extracellular vesicles associated miRNA in myocardial fibrosis. *Biochem Biophys Res Commun*. (2024) 727:150336. doi: 10.1016/j.bbrc.2024.150336
- Treiber T, Treiber N, Meister G. Regulation of microRNA biogenesis and its crosstalk with other cellular pathways. *Nat Rev Mol Cell Biol*. (2019) 20:5–20. doi: 10.1038/s41580-018-0059-1
- Wu K, He J, Pu W, Peng Y. The role of exportin-5 in microRNA biogenesis and cancer. *Genomics Proteomics Bioinf*. (2018) 16:120–6. doi: 10.1016/j.gpb.2017.09.004
- Hynes C, Kakumani PK. Regulatory role of RNA-binding proteins in microRNA biogenesis. *Front Mol Biosci*. (2024) 11:1374843. doi: 10.3389/fmolb.2024.1374843
- Rani V, Sengar RS. Biogenesis and mechanisms of microRNA-mediated gene regulation. *Biotechnol Bioeng*. (2022) 119:685–92. doi: 10.1002/bit.28029
- Komatsu S, Kitai H, Suzuki HI. Network regulation of microRNA biogenesis and target interaction. *Cells*. (2023) 12:306. doi: 10.3390/cells12020306
- Salim U, Kumar A, Kulshreshtha R, Vivekanandan P. Biogenesis, characterization, and functions of mirtrons. *Wiley Interdiscip Rev RNA*. (2022) 13:e1680. doi: 10.1002/wrna.1680
- Liu Z, Wang M, Cheng A, Ou X, Mao S, Yang Q, et al. Gene regulation in animal miRNA biogenesis. *Epigenomics*. (2022) 14(19):1197–212. doi: 10.2217/epi-2022-0214
- Michlewski G, Cáceres JF. Post-transcriptional control of miRNA biogenesis. *RNA*. (2019) 25:1–16. doi: 10.1261/rna.068692.118
- Villarroya-Beltri C, Gutiérrez-Vázquez C, Sánchez-Cabo F, Pérez-Hernández D, Vázquez J, Martín-Cofreces N, et al. Sumoylated hnRNP2B1 controls the sorting of miRNAs into exosomes through binding to specific motifs. *Nat Commun*. (2013) 4:2980. doi: 10.1038/ncomms3980
- Frank F, Sonenberg N, Nagar B. Structural basis for 5'-nucleotide base-specific recognition of guide RNA by human AGO2. *Nature*. (2010) 465:818–22. doi: 10.1038/nature09039
- Groot M, Lee H. Sorting mechanisms for microRNAs into extracellular vesicles and their associated diseases. *Cells*. (2020) 9:1044. doi: 10.3390/cells9041044
- Kosaka N, Iguchi H, Hagiwara K, Yoshioka Y, Takeshita F, Ochiya T. Neutral sphingomyelinase 2 (nSMase2)-dependent exosomal transfer of angiogenic microRNAs regulate cancer cell metastasis. *J Biol Chem*. (2013) 288:10849–59. doi: 10.1074/jbc.M112.446831
- Ramanathan S, Shenoda BB, Lin Z, Alexander GM, Huppert A, Sacan A, et al. Inflammation potentiates miR-939 expression and packaging into small extracellular vesicles. *J Extracell Vesicles*. (2019) 8:1650595. doi: 10.1080/20013078.2019.1650595
- Gibbings DJ, Ciaudo C, Erhardt M, Voynet O. Multivesicular bodies associate with components of miRNA effector complexes and modulate miRNA activity. *Nat Cell Biol*. (2009) 11:1143–9. doi: 10.1038/ncb1929
- Montecalvo A, Larregina AT, Shufesky WJ, Stolz DB, Sullivan ML, Karlsson JM, et al. Mechanism of transfer of functional microRNAs between mouse dendritic cells via exosomes. *Blood*. (2012) 119:756–66. doi: 10.1182/blood-2011-02-338004
- O'Brien J, Hayder H, Zayed Y, Peng C. Overview of microRNA biogenesis, mechanisms of actions, and circulation. *Front Endocrinol (Lausanne)*. (2018) 9:402. doi: 10.3389/fendo.2018.00402
- Zeng H, Guo S, Ren X, Wu Z, Liu S, Yao X. Current strategies for exosome cargo loading and targeting delivery. *Cells*. (2023) 12:1416. doi: 10.3390/cells12101416
- Robson A. Exosome-derived microRNAs improve cardiac function. *Nat Rev Cardiol*. (2021) 18:150–1. doi: 10.1038/s41569-020-00498-w
- Isaac R, Reis FCG, Ying W, Olefsky JM. Exosomes as mediators of intercellular crosstalk in metabolism. *Cell Metab*. (2021) 33:1744–62. doi: 10.1016/j.cmet.2021.08.006
- Fabbri M, Paone A, Calore F, Galli R, Gaudio E, Santhanam R, et al. MicroRNAs bind to Toll-like receptors to induce prometastatic inflammatory response. *Proc Natl Acad Sci U.S.A.* (2012) 109:E2110–6. doi: 10.1073/pnas.1209414109
- Chen C, Cai S, Wu M, Wang R, Liu M, Cao G, et al. Role of cardiomyocyte-derived exosomal microRNA-146a-5p in macrophage polarization and activation. *Dis Markers*. (2022) 2022:2948578. doi: 10.1155/2022/2948578

52. Siegel PM, Schmich J, Barinov G, Bojti I, Vecednik C, Simanjuntak NR, et al. Cardiomycocyte microvesicles: proinflammatory mediators after myocardial ischemia? *J Thromb Thrombolysis*. (2020) 50:533–42. doi: 10.1007/s11239-020-02156-x
53. Yu H, Qin L, Peng Y, Bai W, Wang Z. Exosomes Derived From Hypertrophic Cardiomyocytes Induce Inflammation in Macrophages via miR-155 Mediated MAPK Pathway. *Front Immunol*. (2020) 11:606045. doi: 10.3389/fimmu.2020.606045
54. López E, Marinario F, de Pedro M, Sánchez-Margallo FM, Gómez-Serrano M, Ponath V, et al. The immunomodulatory signature of extracellular vesicles from cardiosphere-derived cells: A proteomic and miRNA profiling. *Front Cell Dev Biol*. (2020) 8:321. doi: 10.3389/fcell.2020.00321
55. de Couto G, Gallet R, Cambier L, Jaghatspanyan E, Makkar N, Dawkins JF, et al. Exosomal microRNA transfer into macrophages mediates cellular postconditioning. *Circulation*. (2017) 136:200–14. doi: 10.1161/circulationaha.116.024590
56. de Couto G, Jaghatspanyan E, DeBerge M, Liu W, Luther K, Wang Y, et al. Mechanism of enhanced merTK-dependent macrophage efferocytosis by extracellular vesicles. *Arterioscler Thromb Vasc Biol*. (2019) 39:2082–96. doi: 10.1161/atvbaha.119.313115
57. Ma Y, Su M, Qian W, Xuan Y, Chen T, Zhou R, et al. CXCR4-overexpressed exosomes from cardiosphere-derived cells attenuate myocardial ischemia/reperfusion injury by transferring miRNA to macrophages and regulating macrophage polarization. *Cell Mol Biol (Noisy-le-grand)*. (2023) 69:98–103. doi: 10.14715/cmb/2023.69.12.16
58. Jansen F, Yang X, Baumann K, Przybilla D, Schmitz T, Flender A, et al. Endothelial microparticles reduce ICAM-1 expression in a microRNA-222-dependent mechanism. *J Cell Mol Med*. (2015) 19:2202–14. doi: 10.1111/jcmm.12607
59. Zhang X, Wang X, Fan M, Tu F, Yang K, Ha T, et al. Endothelial HSPA12B Exerts Protection Against Sepsis-Induced Severe Cardiomyopathy via Suppression of Adhesion Molecule Expression by miR-126. *Front Immunol*. (2020) 11:566. doi: 10.3389/fimmu.2020.00566
60. Bang C, Batkai S, Dargwal S, Gupta SK, Foinquinos A, Holzmann A, et al. Cardiac fibroblast-derived microRNA passenger strand-enriched exosomes mediate cardiomyocyte hypertrophy. *J Clin Invest*. (2014) 124:2136–46. doi: 10.1172/JCI70577
61. Luo H, Li X, Li T, Zhao L, He J, Zha L, et al. microRNA-423-3p exosomes derived from cardiac fibroblasts mediates the cardioprotective effects of ischaemic post-conditioning. *Cardiovasc Res*. (2019) 115:1189–204. doi: 10.1093/cvr/cvy231
62. Liu N, Xie L, Xiao P, Chen X, Kong W, Lou Q, et al. Cardiac fibroblasts secrete exosome microRNA to suppress cardiomyocyte pyroptosis in myocardial ischemia/reperfusion injury. *Mol Cell Biochem*. (2022) 477:1249–60. doi: 10.1007/s11010-021-04343-7
63. Xie L, Chen J, Wang Y, Jin C, Xie Y, Ma H, et al. Emerging roles of macrophages in heart failure and associated treatment approaches. *Ther Adv Chronic Dis*. (2023) 14:20406223231168755. doi: 10.1177/20406223231168755
64. Saleh D, Jones RTL, Schroth SL, Thorp EB, Feinstein MJ. Emerging roles for dendritic cells in heart failure. *Biomolecules*. (2023) 13:1535. doi: 10.3390/biom13101535
65. Abplanalp WT, John D, Cremer S, Assmus B, Dorsheimer L, Hoffmann J, et al. Single-cell RNA-sequencing reveals profound changes in circulating immune cells in patients with heart failure. *Cardiovasc Res*. (2021) 117:484–94. doi: 10.1093/cvr/cvaa101
66. Natrus L, Labudzynski D, Muzychenko P, Chernovol P, Klys Y. Plasma-derived exosomes implement miR-126-associated regulation of cytokines secretion in PBMCs of CHF patients *in vitro*. *Acta BioMed*. (2022) 93:e2022066. doi: 10.23750/abm.v93i3.12449
67. Vilella-Figuerola A, Padró T, Roig E, Mirabet S, Badimon L. New factors in heart failure pathophysiology: Immunity cells release of extracellular vesicles. *Front Cardiovasc Med*. (2022) 9:939625. doi: 10.3389/fcvm.2022.939625
68. Wang K, Sun X, Sun Y, Jiao B, Yao J, Hu Y, et al. Transcriptional regulation of macrophages in heart failure. *Front Cardiovasc Med*. (2023) 10:1148041. doi: 10.3389/fcvm.2023.1148041
69. Wang H, Ye X, Spanos M, Wang H, Yang Z, Li G, et al. Exosomal non-coding RNA mediates macrophage polarization: roles in cardiovascular diseases. *Biol (Basel)*. (2023) 12:745. doi: 10.3390/biology12050745
70. Ge X, Meng Q, Wei L, Liu J, Li M, Liang X, et al. Myocardial ischemia-reperfusion induced cardiac extracellular vesicles harbour proinflammatory features and aggravate heart injury. *J Extracell Vesicles*. (2021) 10:e12072. doi: 10.1002/jev2.12072
71. Li L, Cao J, Li S, Cui T, Ni J, Zhang H, et al. M2 macrophage-derived sEV regulate pro-inflammatory CCR2(+) macrophage subpopulations to favor post-AMI cardiac repair. *Adv Sci (Weinh)*. (2023) 10:e2202964. doi: 10.1002/adv.202202964
72. Xia W, Chen H, Chen D, Ye Y, Xie C, Hou M. PD-1 inhibitor inducing exosomal miR-34a-5p expression mediates the cross talk between cardiomyocyte and macrophage in immune checkpoint inhibitor-related cardiac dysfunction. *J Immunother Cancer*. (2020) 8:e001293. doi: 10.1136/jitc-2020-001293
73. Nagai T, Honda S, Sugano Y, Matsuyama TA, Ohta-Ogo K, Asaumi Y, et al. Decreased myocardial dendritic cells is associated with impaired reparative fibrosis and development of cardiac rupture after myocardial infarction in humans. *J Am Heart Assoc*. (2014) 3:e000839. doi: 10.1161/jaha.114.000839
74. Liu H, Zhang Y, Yuan J, Gao W, Zhong X, Yao K, et al. Dendritic cell-derived exosomal miR-494-3p promotes angiogenesis following myocardial infarction. *Int J Mol Med*. (2021) 47:315–25. doi: 10.3892/ijmm.2020.4776
75. Liu M, Xia N, Zha L, Yang H, Gu M, Hao Z, et al. Increased expression of protein tyrosine phosphatase nonreceptor type 22 alters early T-cell receptor signaling and differentiation of CD4(+) T cells in chronic heart failure. *FASEB J*. (2024) 38:e23386. doi: 10.1096/fj.202300663R
76. Cai L, Chao G, Li W, Zhu J, Li F, Qi B, et al. Activated CD4(+) T cells-derived exosomal miR-142-3p boosts post-ischemic ventricular remodeling by activating myofibroblast. *Aging (Albany NY)*. (2020) 12:7380–96. doi: 10.18632/aging.103084
77. Rheault-Henry M, White I, Grover D, Atoui R. Stem cell therapy for heart failure: Medical breakthrough, or dead end? *World J Stem Cells*. (2021) 13:236–59. doi: 10.4252/wjsc.v13.i4.236
78. Amaro-Prellezo E, Gómez-Ferrer M, Hakobyan L, Ontoria-Oviedo I, Peiró-Molina E, Tarazona S, et al. Extracellular vesicles from dental pulp mesenchymal stem cells modulate macrophage phenotype during acute and chronic cardiac inflammation in athymic nude rats with myocardial infarction. *Inflammation Regen*. (2024) 44:25. doi: 10.1186/s41232-024-00340-7
79. Chachques JC, Gardin C, Lila N, Ferroni L, Migonney V, Falentin-Daudre C, et al. Elastomeric cardiowrap scaffolds functionalized with mesenchymal stem cells-derived exosomes induce a positive modulation in the inflammatory and wound healing response of mesenchymal stem cell and macrophage. *Biomedicine*. (2021) 9:824. doi: 10.3390/biomedicine9070824
80. Zhao J, Li X, Hu J, Chen F, Qiao S, Sun X, et al. Mesenchymal stromal cell-derived exosomes attenuate myocardial ischaemia-reperfusion injury through miR-182-regulated macrophage polarization. *Cardiovasc Res*. (2019) 115:1205–16. doi: 10.1093/cvr/cvz040
81. Gong ZT, Xiong YY, Ning Y, Tang RJ, Xu JY, Jiang WY, et al. Nicorandil-pretreated Mesenchymal Stem Cell-Derived Exosomes Facilitate Cardiac Repair After Myocardial Infarction via Promoting Macrophage M2 Polarization by Targeting miR-125a-5p/TRAF6/IRF5 Signaling Pathway. *Int J Nanomedicine*. (2024) 19:2005–24. doi: 10.2147/ijn.s441307
82. Zhu F, Chen Y, Li J, Yang Z, Lin Y, Jiang B, et al. Human Umbilical Cord Mesenchymal Stem Cell-Derived Exosomes Attenuate Myocardial Infarction Injury via miR-24-3p-Promoted M2 Macrophage Polarization. *Adv Biol (Weinh)*. (2022) 6:e2200074. doi: 10.1002/adbi.202200074
83. Du J, Dong Y, Song J, Shui H, Xiao C, Hu Y, et al. BMSC-derived exosome-mediated miR-25-3p delivery protects against myocardial ischemia/reperfusion injury by constraining M1-like macrophage polarization. *Mol Med Rep*. (2024) 30:142. doi: 10.3892/mmr.2024.13266
84. Ning Y, Huang P, Chen G, Xiong Y, Gong Z, Wu C, et al. Atorvastatin-pretreated mesenchymal stem cell-derived extracellular vesicles promote cardiac repair after myocardial infarction via shifting macrophage polarization by targeting microRNA-139-3p/Stat1 pathway. *BMC Med*. (2023) 21:96. doi: 10.1186/s12916-023-02778-x
85. Pan L, Yan B, Zhang J, Zhao P, Jing Y, Yu J, et al. Mesenchymal stem cells-derived extracellular vesicles-shuttled microRNA-223-3p suppress lipopolysaccharide-induced cardiac inflammation, pyroptosis, and dysfunction. *Int Immunopharmacol*. (2022) 110:108910. doi: 10.1016/j.intimp.2022.108910
86. Yang Y, Johnson J, Troupes CD, Feldsott EA, Kraus L, Megill E, et al. miR-182/183-Rasa1 axis induced macrophage polarization and redox regulation promotes repair after ischemic cardiac injury. *Redox Biol*. (2023) 67:102909. doi: 10.1016/j.redox.2023.102909
87. de Almeida Oliveira NC, Neri EA, Silva CM, Valadão IC, Fonseca-Alaniz MH, Zogbi C, et al. Multicellular regulation of miR-196a-5p and miR-425-5 from adipose stem cell-derived exosomes and cardiac repair. *Clin Sci (Lond)*. (2022) 136:1281–301. doi: 10.1042/cs20220216
88. Wu T, Chen Y, Du Y, Tao J, Zhou Z, Yang Z. Serum exosomal miR-92b-5p as a potential biomarker for acute heart failure caused by dilated cardiomyopathy. *Cell Physiol Biochem*. (2018) 46:1939–50. doi: 10.1159/000489383
89. Ramachandran S, Lowenthal A, Ritner C, Lowenthal S, Bernstein HS. Plasma microvesicle analysis identifies microRNA 129-5p as a biomarker of heart failure in univentricular heart disease. *PLoS One*. (2017) 12:e0183624. doi: 10.1371/journal.pone.0183624
90. Huang JP, Chang CC, Kuo CY, Huang KJ, Sokal EM, Chen KH, et al. Exosomal microRNAs miR-30d-5p and miR-126a-5p Are Associated with Heart Failure with Preserved Ejection Fraction in STZ-Induced Type 1 Diabetic Rats. *Int J Mol Sci*. (2022) 23:7514. doi: 10.3390/ijms23147514
91. Wu T, Chen Y, Du Y, Tao J, Li W, Zhou Z, et al. Circulating exosomal miR-92b-5p is a promising diagnostic biomarker of heart failure with reduced ejection fraction patients hospitalized for acute heart failure. *J Thorac Dis*. (2018) 10:6211–20. doi: 10.21037/jtd.2018.10.52
92. Wang R, Huang K, Feng Y, Duan J, Ying H, Shi Q, et al. Exo-miR-144-3p as a promising diagnostic biomarker for depressive symptoms in heart failure. *Neurobiol Dis*. (2024) 192:106415. doi: 10.1016/j.nbd.2024.106415
93. Wang L, Liu J, Xu B, Liu YL, Liu Z. Reduced exosome miR-425 and miR-744 in the plasma represents the progression of fibrosis and heart failure. *Kaohsiung J Med Sci*. (2018) 34:626–33. doi: 10.1016/j.kjms.2018.05.008

94. Beg F, Wang R, Saeed Z, Devaraj S, Masoor K, Nakshatri H. Inflammation-associated microRNA changes in circulating exosomes of heart failure patients. *BMC Res Notes*. (2017) 10:751. doi: 10.1186/s13104-017-3090-y
95. Xiao YC, Wang W, Gao Y, Li WY, Tan X, Wang YK, et al. The peripheral circulating exosomal microRNAs related to central inflammation in chronic heart failure. *J Cardiovasc Transl Res*. (2022) 15:500–13. doi: 10.1007/s12265-022-10266-5
96. Matsumoto S, Sakata Y, Suna S, Nakatani D, Usami M, Hara M, et al. Circulating p53-responsive microRNAs are predictive indicators of heart failure after acute myocardial infarction. *Circ Res*. (2013) 113:322–6. doi: 10.1161/circresaha.113.301209
97. Shafei S, Khanmohammadi M, Ghanbari H, Nooshabadi VT, Tafti SHA, Rabbani S, et al. Effectiveness of exosome mediated miR-126 and miR-146a delivery on cardiac tissue regeneration. *Cell Tissue Res*. (2022) 390:71–92. doi: 10.1007/s00441-022-03663-4
98. Song Y, Zhang C, Zhang J, Jiao Z, Dong N, Wang G, et al. Localized injection of miRNA-21-enriched extracellular vesicles effectively restores cardiac function after myocardial infarction. *Theranostics*. (2019) 9:2346–60. doi: 10.7150/thno.29945
99. Nasser MI, Masood M, Adlat S, Gang D, Zhu S, Li G, et al. Mesenchymal stem cell-derived exosome microRNA as therapy for cardiac ischemic injury. *BioMed Pharmacother*. (2021) 143:112118. doi: 10.1016/j.biopha.2021.112118
100. Yan F, Cui W, Chen Z. Mesenchymal stem cell-derived exosome-loaded microRNA-129-5p inhibits TRAF3 expression to alleviate apoptosis and oxidative stress in heart failure. *Cardiovasc Toxicol*. (2022) 22:631–45. doi: 10.1007/s12012-022-09743-9
101. Wei Z, Qiao S, Zhao J, Liu Y, Li Q, Wei Z, et al. miRNA-181a over-expression in mesenchymal stem cell-derived exosomes influenced inflammatory response after myocardial ischemia-reperfusion injury. *Life Sci*. (2019) 232:116632. doi: 10.1016/j.lfs.2019.116632
102. Wan J, Lin S, Yu Z, Song Z, Lin X, Xu R, et al. Protective effects of microRNA-200b-3p encapsulated by mesenchymal stem cells-secreted extracellular vesicles in myocardial infarction via regulating BCL2L1. *J Am Heart Assoc*. (2022) 11:e024330. doi: 10.1161/jaha.121.024330
103. Liu Y, Guan R, Yan J, Zhu Y, Sun S, Qu Y. Mesenchymal stem cell-derived extracellular vesicle-shuttled microRNA-302d-3p represses inflammation and cardiac remodeling following acute myocardial infarction. *J Cardiovasc Transl Res*. (2022) 15:754–71. doi: 10.1007/s12265-021-10200-1
104. Yang M, Liao M, Liu R, Zhang Q, Zhang S, He Y, et al. Human umbilical cord mesenchymal stem cell-derived extracellular vesicles loaded with miR-223 ameliorate myocardial infarction through P53/S100A9 axis. *Genomics*. (2022) 114:110319. doi: 10.1016/j.ygeno.2022.110319
105. Wang X, Gu H, Qin D, Yang L, Huang W, Essandoh K, et al. Exosomal miR-223 contributes to mesenchymal stem cell-elicited cardioprotection in polymicrobial sepsis. *Sci Rep*. (2015) 5:13721. doi: 10.1038/srep13721
106. Pei Y, Xie S, Li J, Jia B. Bone marrow-mesenchymal stem cell-derived exosomal microRNA-141 targets PTEN and activates β -catenin to alleviate myocardial injury in septic mice. *Immunopharmacol Immunotoxicol*. (2021) 43:584–93. doi: 10.1080/08923973.2021.1955920
107. Ren S, Pan L, Yang L, Niu Z, Wang L, Feng H, et al. miR-29a-3p transferred by mesenchymal stem cells-derived extracellular vesicles protects against myocardial injury after severe acute pancreatitis. *Life Sci*. (2021) 272:119189. doi: 10.1016/j.lfs.2021.119189

Glossary

AGO	argonaute	CDCs	cardiosphere-derived cells
CXCL	C-X-C motif ligand	DCM	dilated cardiomyopathy
DC	dendritic cell	ESCRT	endosomal sorting complex required for transport
ERK	extracellular signal-regulated kinase	EVs	extracellular vesicles
HF	heart failure	hnRNPs	heterogeneous nuclear ribonucleoproteins
ICAM	intercellular adhesion molecule	IL	interleukin
I/R	ischemia/reperfusion	JNK	c-Jun N-terminal kinase
mRNAs	messenger RNAs	MI	myocardial infarction
miRNAs	microRNAs	MSCs	mesenchymal stem cells
NF- κ B	nuclear factor kappa B	PBMCs	peripheral blood mononuclear cells
pre-miRNAs	precursor miRNAs	pri-miRNAs	primary miRNAs
RBPs	RNA-binding proteins	RISC	RNA-induced silencing complex
ROS	reactive oxygen species	TNF	tumor necrosis factor
TLR	toll-like receptor	UTRs	untranslated regions.
VEGF	vascular endothelial growth factor		