Check for updates

OPEN ACCESS

EDITED BY Kay-Dietrich Wagner, University of Nice Sophia Antipolis, France

REVIEWED BY Santi M. Mandal, Indian Institute of Technology Kharagpur, India Chengye Che, The Affiliated Hospital of Qingdao University, China

*CORRESPONDENCE Wensong Zhang Zhangws@jlu.edu.cn

RECEIVED 25 June 2024 ACCEPTED 06 December 2024 PUBLISHED 23 December 2024

CITATION

Zhou H, Zhang H, Bi M and Zhang W (2024) The role of PPAR in fungal keratitis. *Front. Immunol.* 15:1454463. doi: 10.3389/fimmu.2024.1454463

COPYRIGHT

© 2024 Zhou, Zhang, Bi and Zhang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

The role of PPAR in fungal keratitis

Hongyan Zhou¹, Hong Zhang¹, Miaomiao Bi¹ and Wensong Zhang²*

¹Department of Ophthalmology, The China-Japan Union Hospital of Jilin University, Changchun, Jilin, China, ²Department of Ophthalmology, The Second Hospital of Jilin University, Changchun, Jilin, China

The treatment of fungal keratitis(FK) remains challenging due to delayed fungal detection and the limited effectiveness of antifungal drugs. Fungal infection can activate both innate and adaptive immune responses in the cornea. Fungi stimulate the production of oxidative stress-related biomarkers and mediate the infiltration of neutrophils, macrophages, and T cells. These cells can induce infiltration of cytokines, chemokines, and matrix metalloproteinases (MMPs), leading to corneal tissue damage and even corneal perforation. The signaling pathway regulates the expression of inflammatory cytokines in fungal keratitis. Immune inflammatory damage is the main mechanism of FK, and oxidative stress damage is also involved in this infection process. Peroxisome proliferatoractivated receptor (PPAR) is a member of the nuclear hormone receptor superfamily, with different subtypes of PPAR a, PPAR β/δ , and PPAR γ . PPARs play important roles in the antioxidant response, anti-inflammatory, lipid metabolism, neuroprotection, and immune regulation processes. PPAR γ can promote macrophage polarization and reduce oxidative stress damage by regulating ROS production. PPAR has made some progress in the treatment of eye diseases: PPARa agonists can inhibit diabetes keratopathy and corneal neuropathy. PPARa agonists inhibit early immature angiogenesis in corneal alkali burns and have potential therapeutic effects on inflammatory corneal angiogenesis. PPARs can control the progression of dry eye disease and improve the condition of meibomian gland dysfunction. Based on this, we explored the potential roles of PPARs in the treatment of FK.

KEYWORDS

PPAR, fungal keratitis, infection and inflammation, cytokines, oxidative stress, immunology

1 Introduction

The treatment of fungal infections is an important part of global public health. Fungi is the most important environmental biomarker. Its biodiversity is very important and highly sensitive to environmental changes. Fungal resistance can lead to uncontrolled infections that can be life-threatening (1, 2). Microbial keratitis includes bacteria, viruses, fungi, and protozoa. The

incidence of FK is increasing year by year. FK accounts for 40% to 50% of all cases of microbial keratitis. FK can cause corneal damage and endophthalmitis, which can eventually lead to vision loss. The main causative agents of FK are Fusarium and Aspergillus (3, 4). Trauma, weakened immune function, ocular surface disease, and wearing contact lenses are the main causes of FK. The lag of fungal test results is challenging (5). Fungal resistance and corneal transplant rejection are currently challenges in the treatment of aggressive FK (6).The difficulties of drug therapy with FK lie in the limited permeability of antifungal drugs, low bioavailability, poor pharmacokinetic properties and corneal toxicity. The lack of donors for corneal transplantation is a dilemma for fungal treatment (7).

Exploring new adjuvant treatment options in clinical practice is a top priority. PPAR can be a good option. PPARs are the members of nuclear hormone receptor superfamilies with different subtypes PPAR α , PPAR β/δ , and PPAR γ . PPARs can regulate energy homeostasis, and oxidative stress, and determine cell fate (8). Impaired peroxidase function can lead to different diseases, which are associated with severe clinical symptoms. Oxidative stress damage is also involved in the pathogenesis of FK (9). Pro-inflammatory cytokines, oxidative stress damage, macrophage reactive oxygen species, and mitochondrial reactive oxygen species levels during FK pathogenesis are our therapeutic targets (10). PPAR has definite anti-inflammatory and antioxidant effects. One of the key functions of peroxisomes is the oxidation of fatty acids, which can be mobilized and transferred to peroxisomes and mitochondria, thus efficiently exchanging metabolites (11, 12). Excessive inflammation and immune damage during FK can cause irreversible damage to corneal tissue. PPARa is a transcription factor that can modulate inflammation. PPARa functional modulators inhibit inflammation by blocking peripheral immune cells and the Toll-like receptor-4(TLR-4)/the nuclear factor-kB(NF-kB)pathway (13). Numerous studies suggest that PPAR γ agonists may be effective therapeutic agents for inflammation-related diseases (14). PPAR activators inhibit the expression of inflammatory response genes such as IL-2, IL-6, IL-8, TNFα, and MMP by inhibiting the NF-κB signal transducer, the activator of the transcription(STAT) and Activator Protein -1(AP-1) signaling pathways (15).

Finding appropriate reagents to fight inflammation, modulate an overactive immune response, and resist oxidative stress damage is essential to prevent infection-related corneal damage. Peroxisome could be the ideal target. In this review, we emphasize corneal fungal infections that require progress to better express treatment of FK related to PPARs.

2 Immunity to fungi in the cornea

Fungi infection activated the corneal innate and adaptive immune response (16).

2.1 Innate immunity to FK

Innate immunity is the host's first line of defense against microbial infection. Fungal infections in immunocompromised patients can cause severe tissue damage. The corneal cells specifically recognizes patterns associated with molecular patterns (PAMPs) on the fungal surface through pattern recognition receivers (PRRs). PRRs play a clinical role in innate immunity, mediating the infiltration of neutrophils, macrophages, and T cells, and ultimately the destruction of fungi. The infiltration of these cells can lead to infiltration of cascades of amplified inflammatory and chemokines, which can eventually lead to corneal tissue damage and even corneal perforation. The innate immune function relies on the recognition of PAMPs, and damage-associated molecular patterns (DAMPs) of pathogen by PRRs (17, 18). Ficolin-A (FCN-A) is a class of soluble PRRs that play an important role in innate immunity. Downregulation of FCN-A reduces the inflammatory response and decreases the expression of tumor necrosis factor(TNF-a), p-p38, p-JNK, and mitogen-activated protein kinase(MAPK)in corneal infected mice, and promotes macrophage polarization (19). The proportion of dendritic cells (DCs) in the early stage of FK generally decreases with fungal infection, while the proportion of macrophages, monocytes, and neutrophils increases dramatically in the early stage and then gradually decreases as inflammation resolves. Activation of adaptive immune cells is also observed in the later stages of infection. Activation of early acquired immune cells favors fungal clearance, which gradually decreases in the later stages of inflammation to reduce the damage to corneal tissue by the corresponding cytokines (20). β -glucan on A. fumigatus germinate conidia which activates Dectin-1 on corneal macrophages produce interleukin-1(IL-1β), and CXC ligand-1 (CXCL₁), recruiting neutrophils from the corneal stroma through IL-1R₁/myeloid differentiation primary response gene 88 (MYD88) dependent activation and killing fungi through the TLR4 dependent pathway (21). Fumigatus-infected corneal DCs could drive naive CD4+ T-cell proliferation and promote the production of Th17 cytokines and IL-22. Activated DC produces thymic stromal lymphopoietin (TSLP) leading to a Th17-type inflammatory response via the Janus kinase-signal transducer and activator of transcription pathway(JAK/STAT)signaling pathway (22). Fumigatus-infected mouse cornea induces macrophage infiltration through a TLR-4-mediated signaling pathway. and produce proinflammatory factors IL-1 β , tumour necrosis factor- α (TNF- α), and IL-6 in the corneal epithelium and stroma. The antifungal and antiinflammatory reagents should be found in the therapy of FK (23).T cells follicular helper, monocytes, macrophages, and mast cells might play a key role in Fusarium keratitis using analysis of immune infiltration (24). Inhibition of macrophage phagocytosis and killing function can mediate the protective immune effect of Aspergillus fumigatus (25, 26). In the process of FK infection, finding effective macrophage pathogen clearance while reducing unnecessary inflammatory reactions to alleviate tissue damage is the immune regulatory target that scholars are dedicated to searching for (27).

2.2 Adaptive immunity to FK

2.2.1 Cellular immunity

Immunity through the mucosal surface stimulates IgA antibodies in tears which can prevent the development of keratitis

(28). T helper cells(Th17)and regulatory T (Treg)cells share a common precursor cell (naive CD4 T cells), Th17 originated from neutrophils, plays a key role in the severity of autoimmune and inflammatory infection, while Treg cells inhibit Immune inflammatory damage and maintain immune homeostasis.Th17/ Treg cells balance signaling pathways were triggered by T cell receptors, costimulatory receptors, and cytokines, as well as various metabolic pathways and gut microbiota (29). CD4+T cell-driven inflammation leads to irreversible damage to the cornea. and can lead to corneal nerve damage (30, 31). The regulation of Th17/ Treg differentiation will determine the outcome of fungal keratitis. This will be a potential site (32, 33).

2.2.2 Humoral immunity

Local and systemic immunity inhibits pathogen responses in some keratitis, such as infectious bovine keratoconjunctivitis (34). Anti-amoebic antigens were detected in tear and ocular lysates, and different levels of Immunoglobulin M(IGM) and Immunoglobulin A(IGA) antibodies were detected during the course of the disease (35). The keratitis vaccine has not been clinically promoted. Treatment of corneas infected with Pseudomonas aeruginosa was effective with immune serum from animals immunized with Pseudomonas aeruginosa membrane vesicles, and the results were effective. Topical treatment with immune serum reduces corneal clinical scores (36).

3 Corneal inflammations in FK

The treatment of FK needs to be based on preventing spore adhesion and disrupting the formation of fungal biofilms, reducing the expression of inflammatory factors in corneal epithelial cells, and inducing reactive oxygen species (ROS) levels in corneal epithelial cells (24). Many cytokines and chemokines are involved in the progression of FK. NF-KB pathway, phosphatidylinositol-3 kinase (PI3K)/protein kinase B(AKT) pathway like PI3K and AKT, and MAPK pathway like p38 regulate the expression of inflammatory cytokines. The fungus induces an increase in TLR4/ MyD88 expression in corneal tissue. It can be used as an effective way of action against fungal infection (37, 38). In infectious keratitis, corneal stromal MMPs, especially MMP-9 and MMP-2, increase activity, inducing corneal collagen degradation and ulcer formation (39). Macrophage inhibitory protein 2 (MIP-2) and intercellular adhesion molecule 1 (ICAM-1) attract neutrophils to the inflammatory site in the cornea (40, 41). Neutrophils are the earliest cells in the body's immune response and are an important part of fighting pathogens (42). The corneal inflammatory response to fungal infection is caused by the accumulation of inflammatory cells. Neutrophils trigger the expression of cytokines and chemokines. These cytokines and chemokines deposit immune cells, and excessive immune responses can cause immune damage, corneal opacity, and vision loss. Therefore, antiinflammatory is an important direction of FK treatment (43-46). Downregulated the p-p65/p65 and p-IkB/IkB protein ratios, attenuate the inflammatory response and fungal burden in FK (47). DC-derived TSLP promotes the Th17-type inflammatory response through the JAK/STAT signaling pathway. Corneal damage in Aspergillus fumigatus-infected keratitis can be reversed by inhibition of the JAK/STAT signaling pathway with specific inhibitors (22).

4 Oxidative stress damage in FK

The fungus stimulates the production of oxidative stress-related markers (ROS, HNE, NO, MDA mitochondrial DNA 8-OHdG, and aconitase-2) in corneal cells. Fungal infections also increase HMOX1 and COX2 expressions and inhibit the levels of antioxidant enzymes, superoxide dismutase-1 (SOD1), glutathione peroxidase-1 (GPx1), and peroxide resin-4 (PRDX4) (48). Upregulation of HO-1 leads to the conversion of heme to carbon monoxide, which acts as an inhibitor of the NF- κ B pathway, leading to decreased expression of pro-inflammatory cytokines. Inflammation is the main culprit in exacerbating fungal infections (49, 50). Activation of the Nrf2/(HO)-1 pathway reduces the innate immune response mediated by oxidative stress.Nrf2 plays an important role in promoting corneal wound healing (51). Inhibition of oxidative stress damage is an effective way to alleviate corneal damage in fungal infections (52) (Figure 1).

5 The potential role of PPARs in FK

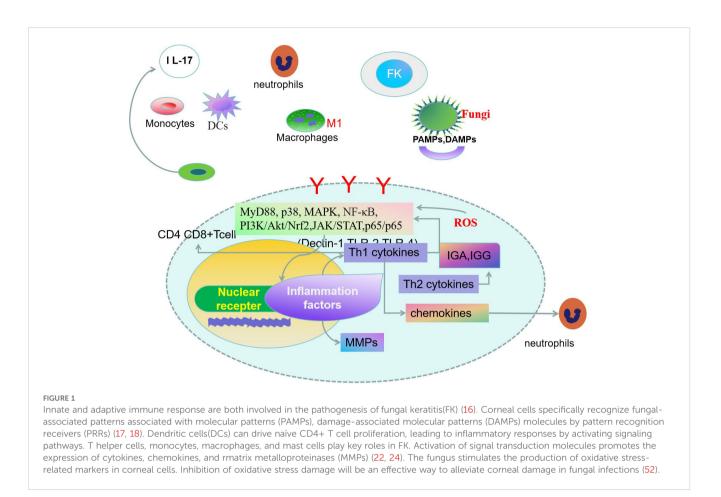
The anti-inflammatory and antioxidant effects of PPAR agonists in various inflammatory diseases have been confirmed, but their roles in FK is worth exploring.

5.1 The role of peroxisome in liver disease

PPAR enhances the production of high-density lipids and inhibits the proliferation of vascular smooth muscle cells. PPARα prevents cardiac complications through adenosine-activated protein kinase, TGF-β activated kinase, and NF-κB pathway (53). PPARα/γ agonists can reduce cardiovascular risk by improving the serum atherosclerotic lipoprotein profile of patients with nonalcoholic fatty liver disease (54). The dual agonist of PPAR a,δ, efebranol, can significantly improve the relevant biochemical indicators of cholestasis and is expected to treat primary biliary cholangitis caused by liver fibrosis (55, 56).

5.2 The role of peroxisome in nerve protection

PPARγ agonists are being studied for the treatment of neurodegenerative diseases including Alzheimer's and Parkinson's disease (57, 58). The biology of PPARγ inhibits transmembrane



spanning transporters or alpha-amino-3-hydroxy-5-methyl-4isoxazolepropionic acid(AMPA) receptors thus alleviating brain swelling and changes around tumors (57–59). PPAR γ coactivator 1 α plays an important role in maintaining mitochondrial function, protecting peripheral nerves, and alleviating chemotherapy-induced peripheral neuropathy (60). PPARa is selectively expressed in certain brain regions and neuronal glial cells and regulates antioxidant responses, neurotransmission, neuroinflammation, neurogenesis, and glial cell proliferation differentiation. PPAR agonists perform a protective role in neurodegenerative diseases and neuropsychiatric disorders (61).

5.3 The role of peroxisome in inflammatory diseases

PPAR is a ligand-dependent transcription factor belonging to the nuclear receptor family. The basic function of PPAR α is to regulate the oxidation of fatty acids. PPAR β/δ is involved in lipid metabolism, wound healing, embryonic development, and inflammation (62–64). PPAR agonists have recently demonstrated promising short-term biochemical responses in patients with primary cholangitis (65). PPARy performs anti-inflammatory and anti-fibrotic effects in different disease models (64). PPARy agonists can be used to alleviate allergic inflammation and inhibit proinflammatory gene expression programs (66). Recombinant Human Fibroblast Growth Factor 21(rhFGF21)Regulates the secretion of pro- and anti-inflammatory cytokines to improve neurological deficits in behavioral testing. It attenuates macrophage polarization to the M1 phenotype and peripheral immune cell accumulation, inhibits NF-KB, and upregulates PPARγ to inhibit pro-inflammatory cytokine expression (67). In the treatment of immunoglobulin A nephropathy rat model, the addition of Yi-Shen-Hua-Shi granules modulates immune and inflammatory damage by the PPAR γ /NF- κ B pathway (68). PPAR- γ is a transcription factor with anti-inflammatory capacity. Nuclear receptors are important bridges between lipid signaling molecules and transcription responses. Macrophages downregulate PPARy and its target genes under inflammatory conditions, thereby decreasing lipid stress in these cells. PPARy is required during the resolution phase of the inflammatory response, and the absence of PPARy is associated with a sustained immune response. Signaling pathways (p38, NFKB, ERK1/2) activated by TLR2 and TL₄ were inhibited by PPAR γ in various situations (69–72).

5.4 Existing research results of peroxisome in eye diseases

Both PPAR α and PPAR γ could be detected in the cornea, conjunctiva, meibomian gland, and lacrimal gland. PPAR γ is expressed at higher levels than PPAR α in all tissues (73).

5.4.1 Dry eye

PPAR α , β/δ , γ can be expressed in the conjunctiva and lacrimal glands. Decreased tear secretion, shortened BUT, and increased corneal staining were shown in Dry eye rats and diabetic rats. The expression of PPAR γ decreased, while the expression of TNF- α , IL-1 β , IL-6, MyD88, and TGF- β increased in both dry-eye rats and diabetic rats (74). Excessive keratinization of the meibomian gland epithelium leads to obstruction of the meibomian opening, meibomian stasis, and cystic dilation of the ducts, resulting in secondary disuse acinar atrophy and glandular shedding (75). PPARy is responsible for the formation of ductal lumens and the differentiation of epithelial cells into meibocyte phenotype during meibomian gland development. The decrease in PPARy receptor expression is the key role of meibomian gland atrophy and degeneration (76). PPAR ligands have neuroprotective and antiinflammatory effects (77), making them a promising drug choice for the treatment of dry eye syndrome.

5.4.2 Corneal alkali burn

In the rat corneal alkali burn model, PPAR α , PPAR γ agonists, and their combinations of ophthalmic fluid were administered respectively in the corneal alkali burn model. PPAR α , PPAR γ , and PPAR α , γ agonists inhibit inflammatory cells, neovascularization, and fibrotic changes. PPAR γ agonists facilitate M_2 macrophages and promote wound healing, The combination of PPAR α with γ agonists may be a new therapy strategy for corneal alkali burns (78, 79). The increased expression level of PPAR γ mRNA can serve as one of the factors for evaluating the prognosis of corneal alkali burn (80).

PPARγ can reduce the expression of downstream inflammatory molecules by regulating signaling molecules. The enhanced expression of PPARγ protein in spinal microglia can inhibit the activity of the NF - κ B pathway, increase the expression of M2 polarization-related genes and significantly reduce the level of inflammatory factors (81). Oxidative stress and inflammatory reaction play an important role in the development of diabetes. Curcumin has been shown to have anti-inflammatory effects in combating changes induced by hyperglycemia by regulating signaling pathways such as PPAR - γ and NF - κ B, as well as high expression of IL-1 β , IL-6, and chemokines such as MCP-1 (82). By regulating the polarization of M2 macrophages through the PPARγ/NF κ B axis and IL-6/IL-6R signaling pathway, the proliferation, migration, and glucose metabolism of lung adenocarcinoma cells are regulated (83).

5.4.3 corneal epithelial injury

In a model of delayed corneal epithelial injury, PPAR agonists or vehicle ophthalmic fluids are locally instilled into the rat cornea, and the corneal epithelial healing process is assessed by fluorescein staining. The area of corneal epithelial defect in the PPAR agonist ophthalmic fluid group was significantly smaller than that in the vehicle group. PPAR γ administration can inhibit the expression of inflammatory cytokines and NF- κ B thus promoting corneal epithelial healing (84). The expression of PPAR γ and PPAR β/δ is reduced in acute epidermal barrier injury, promoting the progression of the disease (85). PPAR β/δ agonists promote the infiltration of M₂ macrophages and vascular endothelial cells and promote the expression of vascular endothelial growth factor A mRNA. Thus, PPAR β/δ ligands promote corneal neovascularization as well as wound healing processes (86).

5.4.4 Diabetic nerve damage

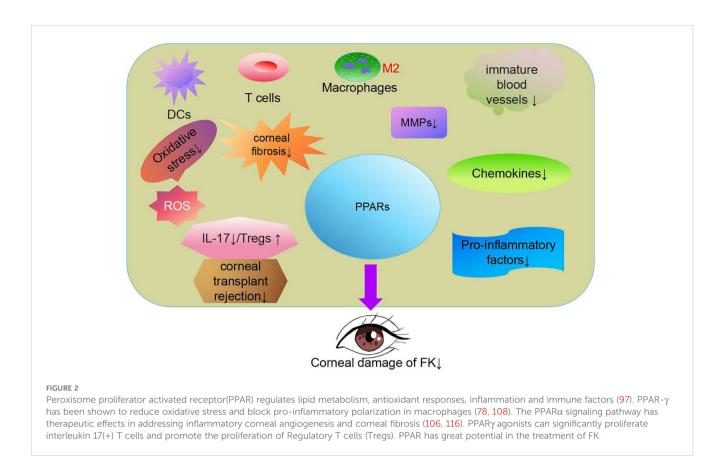
The strong nerve innervation of the cornea ensures its maintenance of the ocular surface homeostasis. Corneal diabetes neuropathy changes the shape and density of the intranasal plexus and nerves. The length and the fiber density of corneal nerve fiber, corneal nerve branch density, twist coefficient, and bead frequency are quantitative analysis parameters of nerve fibers in corneal diabetes neuropathy (87). PPARa agonists stimulate corneal nerve regeneration, reduce nerve edema, significantly improve corneal nerve fiber density and corneal nerve fiber width, and upregulate the neurotrophic signaling pathway (88). The protective effect of PPARa on diabetes keratopathy and corneal neuropathy is achieved by restoring the level of neurotrophic factors in the cornea. The density of corneal nerve fibers in PPAR α -/- mice gradually decreased. The corneal sensitivity of PPAR α -/- mice continues to decrease. PPARa agonist has therapeutic potential in the treatment of diabetes keratopathy (89). Fenofibrate(PPAR alpha agonist)can delay the development of retinopathy and reduce macular edema, reducing the demand for laser therapy (90, 91). Fenofibrate has good effects on lipid control, inflammation, angiogenesis, and cell apoptosis. These factors are considered important in the development of DR. Fenofibrate has the function of protecting the retina (92). Fenofibrate is the treatment choice for DR. More understanding of biomarkers related to inflammation and angiogenesis in diabetes retinopathy will help to develop treatment methods for advanced retinopathy (93).

5.5 The possible mechanism of PPARs involving in FK

PPARs play multiple roles in the β - oxidation of fatty acids and the metabolism of arachidonic acid metabolites. PPAR is also a key regulatory factor in controlling inflammation and has great therapeutic potential in inflammatory diseases (14). Given this, based on our analysis of the aforementioned literature, we attempt to demonstrate the therapeutic potential of PPAR in FK.

5.5.1 Immune regulation of PPARs

Immune damage plays a crucial role in the pathogenesis of FK. In the early stage of inflammation, the innate immune response plays a protective role, while in the late stage of inflammation,



immune damage leads to the formation of corneal ulcers and severe decline in vision. PPAR γ regulates macrophage polarization, DC function, and T-cell activation (64). PPAR α is an important target through which Melatonin regulates autophagy to alleviate the hepatocyte lipid accumulation of Cadmium (94).

PPARα is an important player in innate immune regulation. To distinguish between self and foreign molecules and cells, the pattern recognition receptor(PRR) binds specific molecule PAMPs from certain groups of common pathogens of viral, bacterial, or fungal origin. In the event of an invasion, phagocytosis is triggered and the production of cytotoxic compounds helps to destroy the engulfed particles. PPARα acts as a transcription factor to exert a strong influence on intracellular signal transduction events, fighting a disruptive cytokine storm (95, 96). Peroxisomes are key regulators that regulate immunity, paving the way for potential therapeutic intervention in FK by its immunomodulatory function.

5.5.2 Inhibition effect of PPARs on oxidative stress damage

Oxidative stress promotes the formation of liver fibrosis. Inhibition of ROS-mediated effects and modulation of major antioxidant responses has emerged as therapeutic targets for the prevention of liver injury. ROS is involved in signal transduction, which regulates inflammation and immune factors (97). PPAR γ has been shown to reduce oxidative stress and block pro-inflammatory polarization of macrophages. PPAR γ regulates ROS production by inhibiting Inducible nitric oxide synthase (iNOS) or enhancing the activity of endothelial nitric oxide synthase(eNOS) (98, 99).

Upregulation of antioxidant enzymes as described above can alleviate the condition of FK, and peroxisome can participate in the treatment process of FK through its potential to reduce oxidative stress damage (100). The relative research outcome offers the potential targets for therapeutic strategies to FK.

5.5.3 Inhibition effect of PPARs on inflammations

During the course of FK disease, the retinoid X receptor (RXR) recognizes PAMP and activates a large number of upregulated cytokines/chemokines through signal transduction molecules. Fungal antigens activate CD4+T, activate Th₁, Th₂, and Th₁₇, and regulate T cell production. The production of related cytokines determines the final outcome of fungal inflammation (101). Chemokines recruit neutrophils, regulate vascularization, play a key role in inflammatory responses, and are widely involved in a variety of complex physiological processes (102, 103). Chemokines are involved in FK pathogenesis mentioned above. CC chemokine receptor 1 (CCR1) is a member of the chemokine family and its receptor family, playing a role in the autoimmune response. Inhibiting the MAPK signaling pathway of CCR1 and PPAR - Y reduces inflammatory response and can downregulate the expression of iNOS, IL-1 B, COX-2, and MMP13. Chemokine inhibitors reduce the inflammatory response through the PPARy pathway (104). PPAR agonists have recently been demonstrated promising short-term biochemical responses in patients with primary cholangitis. PPAR-y plays a central and important role in the inhibition of inflammation (65). Peroxisome can be exploited to develop new anti-inflammatory drugs to treat FK.

5.5.4 Inhibition effect of PPARs on corneal neovascularization

Infection is an important factor in promoting corneal neovascularization, which can cause corneal opacity and lead to decreased vision. And often leads to chronic inflammatory circulation (105). The balance between angiogenic factors and antiangiogenic factors is an important cause of corneal neovascularization. Anti-Vascular endothelial growth factor(anti-VEGF) drugs are effective in the treatment of neovascular eye diseases such as age-related macular degeneration, diabetes retinopathy, macular edema, neovascular glaucoma, and other neovascular diseases. Keratitis can cause pathological corneal neovascularization, and the inflammatory genes have a time-dependent effect on capillary regression and corneal transparency recovery. When endogenous capillaries disappear, corneal capillaries become thinner, accompanied by the disappearance of inflammatory cells. Anti-VEGF drugs have great potential in treating corneal neovascularization (106). Endogenous capillary regression is characterized by progressive thinning and remodeling of neovascularization in the rat cornea and in vivo inflammatory cell regression. VEGF ligand receptor signaling is inhibited during vascular regression and PPARa/RXRa is activated in the later stages, which may be responsible for inhibiting proinflammatory genes and immature blood vessels, and helping to prune and remodel capillaries. The administration of PPAR α , a γ agonist, promotes corneal wound healing and provides a new therapeutic strategy (78, 107).

5.5.5 Inhibition effect of PPARs on corneal fibrosis and scar formation

After the corneal injury, transforming growth factor beta-1,2 (TGF β -1,2) is activated and enters the corneal stroma, promoting the transition of keratinocytes near the injury site to myofibroblasts, excreting a large amount of disordered extracellular matrix, forming corneal scarring, and causing vision loss (108–111). PPAR γ can inhibit inflammation, fibrosis, and cell differentiation activated by TGF β (112). Impaired PPAR α signaling can affect FXR activation by inducing the TGF β signaling pathway, P53 signaling pathway, and PI3K-AKT-mTOR signaling pathway, reducing the liver's ability to inhibit inflammation, and ultimately leading to apoptosis and fibrosis (113–115). PPAR plays an important role in inflammation, angiogenesis, and fibrosis formation, and these pathological processes are also important causes of FK (116). PPAR agonists were considered promising therapeutic targets to reduce corneal neovascularization, fibrosis, and inflammation (117).

5.5.6 Inhibition effect of PPARs on corneal transplant rejection in FK

In cases of FK, corneal transplantation is an effective measure for restoring vision, but fungal recurrence and corneal transplant rejection are important reasons for surgical failure. In cases of corneal vascularization or severe infection, the failure rate of corneal transplantation can reach as high as 35% to 70% in patients undergoing corneal transplantation. The rejection reaction of corneal transplantation leads to irreversible damage to the corneal endothelium and corneal opacity (118). The loss of donor corneal endothelial cells is progressive after corneal transplant, which can lead to late-stage graft failure (119). T cell activation is an important cause of corneal transplant rejection. Activated T cells proliferate to form CD4+T helper cells, causing a delayed hypersensitive immune response against the same antigen on the transplant, resulting in damage to corneal tissue (120, 121). After T cells recognize foreign antigens, antigen-presenting cells (APCs) trigger innate and adaptive immunity. CD4+T cells differentiate into Th₁, Th₂, Treg, and Th₁₇ cells, causing tissue inflammation and immune damage (122, 123). PPARy agonists can significantly inhibit the proliferation of IL-17 (+) T cells and promote the proliferation of Tregs in a heart transplant model, which can help protect heart allografts. Given the anti-inflammatory and antiimmunity effects of PPARy agonists, the agonists were used to treat acute and chronic allograft rejection reactions (124-126) (Figure 2).

6 Conclusion

FK is an infectious disease that seriously affects eye health. Due to the widespread use of antibiotics and steroid drugs, dysbiosis of the microbiota leads to a continuous increase in fungal infections. At present, there is a lack of effective antifungal drugs, and fungal resistance remains an urgent problem to be solved. PPAR is a member of the nuclear receptor family, widely expressed in the cornea, and has made research progress in various eye diseases. Excessive immune inflammatory damage in FK cases resulted in severe damage to corneal tissue. The occurrence of corneal transplant rejection is an important reason for the failure of FK vision restoration. Corneal scarring in the late stage of PK is also an important treatment direction. The anti-inflammatory, immunomodulatory, immunosuppressive, antioxidant, and anti-fibrotic functions of PPAR are a promising choice for future FK treatment. This will contribute to better treatment outcomes for patients with FK.

Author contributions

HYZ: Data curation, Formal analysis, Resources, Writing – original draft. HZ: Investigation, Project administration, Writing – original draft. MB: Formal analysis, Project administration, Validation, Writing – original draft. WZ: Data curation, Formal analysis, Investigation, Supervision, Validation, Visualization, Writing – original draft, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This work was supported by the Jilin Province Department of Finance (the Health Research Talent Project of Jilin Province, grant number2023SCZ86)

and Higher Education Association of Jilin Province (grant number: JGJX2023D13).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

References

1. Sashika D, Warnasuriya DU, Manamgoda DS, Biles C. Fungi as environmental bioindicators. *Sci Total Environ.* (2023) :892164583. doi: 10.1016/j.scitotenv.2023.164583

2. Lionakis MS, Drummond RA, Hohl TM. Immune responses to human fungal pathogens and therapeutic prospects. *Nat Rev Immunol.* (2023) 23:433-52. doi: 10.1038/s41577-022-00826-w

3. Zhou K, Wu J, Chen J, Zhou Ye, Chen X, Wu Q, et al. Schaftoside ameliorates oxygen glucose deprivation-induced inflammation associated with the TLR4/Myd88/ Drp1-related mitochondrial fission in BV2 microglia cells. *J Pharmacol Sci.* (2019) 139:15–22. doi: 10.1016/j.jphs.2018.10.012

4. Kim PS, Shin JH, Jo DS, Shin DW, Choi DH, Kim WJ, et al. Anti-melanogenic activity of schaftoside in Rhizoma Arisaematis by increasing autophagy in B16F1 cells. *Biochem Biophys Res Commun.* (2018) 503:309. doi: 10.1016/j.bbrc.2018.06.021

5. Castano G, Elnahry AG, Mada PK. Fungal keratitis. Treasure Island (FL: StatPearls Publishing (2024).

6. Abbondante S, Leal SM, Clark HL, Ratitong B, Sun Y, Ma L-J, et al. Immunity to pathogenic fungi in the eye. *Semin Immunol.* (2023) 67:101753. doi: 10.1016/j.smim.2023.101753

7. Veerdonk FLvde, Gresnigt MS, Romani L, Netea MG, Latgé J-P. Aspergillus fumigatus morphology and dynamic host interactions. *Nat Rev Microbiol.* (2017) 15:661–74. doi: 10.1038/nrmicro.2017.90

8. Zhang Y, Xiao B, Liu Y, Wu S, Xiang Q, Xiao Y, et al. Roles of PPAR activation in cancer therapeutic resistance: Implications for combination therapy and drug development. *Eur J Pharmacol.* (2024) 964:176304. doi: 10.1016/j.ejphar.2023.176304

9. Fan Y, Li C, Peng X, Jiang N, Hu L, Gu L, et al. Perillaldehyde ameliorates aspergillus fumigatus keratitis by activating the nrf2/HO-1 signaling pathway and inhibiting dectin-1-mediated inflammation. *Invest Ophthalmol Vis Sci.* (2020) 61:51. doi: 10.1167/iovs.61.6.51

10. Wang Z, Lin J, Wang Q, Fu Y, Gu L, Tian X, et al. Guiqiu Zhao Rosmarinic acid alleviates fungal keratitis caused by Aspergillus fumigatus by inducing macrophage autophagy. *Exp Eye Res.* (2024) 244:109944. doi: 10.1016/j.exer.2024.109944

11. Dixon SJ, Olzmann JA. The cell biology of ferroptosis. Nat Rev Mol Cell Biol. (2024) 25:424-42. doi: 10.1038/s41580-024-00703-5

12. Pinna G. Role of PPAR-allopregnanolone signaling in behavioral and inflammatory gut-brain axis communications. *Biol Psychiatry*. (2023) 94:609–18. doi: 10.1016/j.biopsych.2023.04.025

13. Wanders RJA, Vaz FrédéricM, Waterham HR, Ferdinandusse S. Fatty acid oxidation in peroxisomes: enzymology, metabolic crosstalk with other organelles and peroxisomal disorders. *Adv Exp Med Biol.* (2020) 1299:55–70. doi: 10.1007/978-3-030-60204-8_5

14. Lee –T, Yang C-C, Yang C-M. Harnessing peroxisome proliferator-activated receptor γ agonists to induce Heme Oxygenase-1: a promising approach for pulmonary inflammatory disorders. *Cell Commun Signal.* (2024) 22:125. doi: 10.1186/s12964-024-01501-4

15. Chinetti G, Fruchart JC, Staels B. Peroxisome proliferator-activated receptors (PPARs): nuclear receptors at the crossroads between lipid metabolism and inflammation. *Inflammation Res.* (2000) 49:497–505. doi: 10.1007/s000110050622

16. Karthikeyan RS, Jr SML, Prajna NV, Dharmalingam K, Geiser DM, Pearlman E, et al. Expression of innate and adaptive immune mediators in human corneal tissue infected with Aspergillus or fusarium. J Infect Dis. (2011) 204:942–50. doi: 10.1093/infdis/jir426

17. Xue Y, Tuipulotu DE, Tan WH, Kay C, Man SiM. Emerging activators and regulators of inflammasomes and pyroptosis. *Trends Immunol.* (2019) 40:1035–52. doi: 10.1016/j.it.2019.09.005

18. Liston A, Masters SL. Homeostasis-altering molecular processes as mechanisms of inflammasome activation. *Nat Rev Immunol.* (2017) 17:208–14. doi: 10.1038/nri.2016.151

19. Zhu G, Zhao G, Lin J, Li C, Wang Q, Xu Q, et al. FCN-A mediates the inflammatory response and the macrophage polarization in Aspergillus fumigatus keratitis of mice by activating the MAPK signaling pathway. *Int Immunopharmacol.* (2020) 83:106473. doi: 10.1016/j.intimp.2020.106473

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

20. Xu X, Wei. Y, Pang J, Wei Z, Wang L, Chen Q, et al. Time-course transcriptomic analysis reveals the crucial roles of PANoptosis in fungal keratitis. *Invest Ophthalmol Vis Sci.* (2023) 64:6. doi: 10.1167/iovs.64.3.6

21. Sixto M Leal, Cowden S, Hsia Y-C, Ghannoum MA, Momany M, Pearlman E. Distinct roles for Dectin-1 and TLR4 in the pathogenesis of Aspergillus fumigatus keratitis. *PloS Pathog.* (2010) 6:e1000976. doi: 10.1371/journal.ppat.1000976

22. Han F, Guo H, Wang L, Zhang Y, Sun L, Dai C, et al. TSLP produced by aspergillus fumigatus-stimulated DCs promotes a th17 response through the JAK/ STAT signaling pathway in fungal keratitis. *Invest Ophthalmol Vis Sci.* (2020) 61:24. doi: 10.1167/iovs.61.14.24

23. Luan J, Zhu. Y, Lin J, Zhang Y, Xu Q, Zhan Lu, et al. Quercetin protects against Aspergillus fumigatus keratitis by reducing fungal load and inhibiting TLR-4 induced inflammatory response. *Cytokine*. (2023) 171:156356. doi: 10.1016/j.cyto.2023.156356

24. Teng X, Xiong X, Sha X, Lei Y, Diao Y, Liu J, et al. Identification of hub genes and pathways of ferroptosis in Fusarium keratitis by bioinformatics methods. *Front Cell Infect Microbiol.* (2023) 13:1103471.eCollection 2023. doi: 10.3389/fcimb.2023.1103471.eCollection2023

25. Feng Z, Gu L, Lin J, Wang Q, Yu B, Yao X, et al. Formononetin protects against Aspergillus fumigatus Keratitis: Targeting inflammation and fungal load. *Int Immunopharmacol.* (2024) 132:112046. doi: 10.1016/j.intimp.2024.112046

26. Yu F, Jiang W, Zhang Li, Jiang N. IDO regulates macrophage functions by inhibiting the CCL2/CCR2 signaling pathway in fungal keratitis. *Cornea*. (2023) 42:1005–15. doi: 10.1097/ICO.00000000003309

27. Ghosh S, Khathun AHH, Athulya GS, Vignesh P, Mathan L, Mudaraddi N, et al. Host cell-type and pathogen-specific immunomodulatory functions of macrophage migration inhibitory factor (MIF) in infectious keratitis. *Exp Eye Res.* (2023) 236:109669. doi: 10.1016/j.exer.2023.109669

28. Borecka A, Bielawska-DrÓzd A, Skotarczak BogumiŁa, Adamska MaŁgorzata, CieŚlik P, Antos-Bielska MaŁgorzata, et al. Acanthamoeba - pathogen and vector of highly pathogenic bacteria strains to healthy and immunocompromised individuals. *Cent Eur J Immunol.* (2020) 45:228–32. doi: 10.5114/ceji.2020.97667

29. Lee GR. The balance of th17 versus treg cells in autoimmunity. Int J Mol Sci. (2018) 19:730. doi: 10.3390/ijms19030730.\

30. Park SJ, Riccio RE, Kopp SJ, Ifergan I, Miller SD, Longnecker R. herpesvirus entry mediator binding partners mediate immunopathogenesis of ocular herpes simplex virus 1 infection. *mBio.* (2020) 11:e00790-20. doi: 10.1128/mBio.00790-20

31. Royer DJ, Echegaray-Mendez J, Lin L, Gmyrek GB, Mathew R, Saban DR, et al. Complement and CD4+ T cells drive context-specific corneal sensory neuropathy. . *Elife.* (2019) 8:e48378. doi: 10.7554/eLife.48378

32. Ji Q, Liu J, Dong Y, Wang L, Dong K, Setiz B, et al. Exosomes derived from thymic stromal lymphopoietin-treated dendritic cells regulate T helper 17/regulatory T cell differentiation via miR-21/Smad7 axis. *Exp Cell Res.* (2021) 398:112393. doi: 10.1016/j.yexcr.2020.112393

33. Yan J-B, Luo. M, Chen Z-Y, He. B-H. The function and role of the th17/treg cell balance in inflammatory bowel disease. *J Immunol Res.* (2020) 2020:8813558. doi: 10.1155/2020/8813558.eCollection2020

34. Maier G, Connor AMO, Sheedy D. The evidence base for prevention of infectious bovine keratoconjunctivitis through vaccination. *Vet Clin North Am Food Anim Pract.* (2021) 37:341–53. doi: 10.1016/j.cvfa.2021.03.009

35. Kim M-J, Ham A-J, Park A-Y, Sohn H-J, Shin H-J, Quan F-S, et al. Detection of acanthamoeba from acanthamoeba keratitis mouse model using acanthamoeba-specific antibodies. *Microorganisms*. (2022) 10:1711. doi: 10.3390/microorganisms10091711

36. Ito S, Nakamura J, Fukuta M, Ura T, Teshigawara T, Fukushima J, et al. Prophylactic and therapeutic vaccine against Pseudomonas aeruginosa keratitis using bacterial membrane vesicles. *Vaccine*. (2021) 39:3152–60. doi: 10.1016/ jvaccine.2021.04.035

37. Chen X, Lu D, Liu W, Xie J, Lu Z, Yang H, et al. Therapeutic effect of Atractylenolide I on Aspergillus fumigatus keratitis by affecting MyD88/ NF- κB

pathway and IL-1 β , IL-10 expression. Cytokine. (2023) 162:156112. doi: 10.1016/ j.cyto.2022.156112

38. Diao W, Yin M, Qi Y, Fu Y, Gu L, Lin J, et al. Resveratrol has neuroprotective effects and plays an anti-inflammatory role through Dectin-1/p38 pathway in Aspergillus fumigatus keratitis. *Cytokine*. (2024) 179:156626. doi: 10.1016/j.cyto.2024.156626

39. García-López C, Rodríguez-Calvo-de-Mora M, Borroni D, Sánchez-González José-María, Romano V, Rocha-de-Lossada C. The role of matrix metalloproteinases in infectious corneal ulcers. *Surv Ophthalmol.* (2023) 68:929–39. doi: 10.1016/j.survophthal.2023.06.007

40. He S, Zhang. H, Liu S, Liu H, Chen G, Xie Y, et al. [amp]]gamma; δ T cells regulate the expression of cytokines but not the manifestation of fungal keratitis. *Exp Eye Res.* (2015) 135:93–101. doi: 10.1016/j.exer.2015.03.022

41. Kimura K, Orita T, Nomi N, Fujitsu Y, Nishida T, Sonoda K-H. Identification of common secreted factors in human corneal fibroblasts exposed to LPS, poly(I:C), or zymosan. *Exp Eye Res.* (2012) 96:157–62. doi: 10.1016/j.exer.2011.10.015

42. Zhao K, Hu F, Zhang Z, Yin X, Wang He, Li M. 0.01% Hypochlorous acid treats aspergillus fumigatus keratitis in rats by reducing fungal load and inhibiting the inflammatory response. *Transl Vis Sci Technol.* (2023) 12:3. doi: 10.1167/tvst.12.8.3

43. Brault M, Oberst A. Controlled detonation: evolution of necroptosis in pathogen defense. *Immunol Cell Biol.* (2017) 95:131–6. doi: 10.1038/icb.2016.117

44. Underhill DM, Pearlman E. Immune interactions with pathogenic and commensal fungi: A two-way street. *Immunity.* (2015) 43:845–58. doi: 10.1016/j.immuni.2015.10.023

45. Zhan Lu, Peng X, Lin J, Zhang Y, Gao H, Zhu Y, et al. Honokiol reduces fungal load, toll-like receptor-2, and inflammatory cytokines in aspergillus fumigatus keratitis. *Invest Ophthalmol Vis Sci.* (2020) 61:48. doi: 10.1167/iovs.61.4.48

46. Margalit A, Kavanagh K. The innate immune response to Aspergillus fumigatus at the alveolar surface. *FEMS Microbiol Rev.* (2015) 39:670–87. doi: 10.1093/femsre/fuv018

47. Tang H, Huang L, Hu J. Inhibition of the m6A Methyltransferase METTL3 Attenuates the Inflammatory Response in Fusarium solani-Induced Keratitis via the NF- κ B Signaling Pathway. *Invest Ophthalmol Vis Sci.* (2022) 63:2. doi: 10.1167/iovs.63.11.2

48. Hua X, Chi W, Su L, Li J, Zhang Z, Yuan X. ROS-induced Oxidative Injury involved in Pathogenesis of Fungal Keratitis via p38 MAPK Activation. *Sci Rep.* (2017) 7:10421. doi: 10.1038/s41598-017-09636-w

49. Tsuji G, Takahara M, Uchi H, Matsuda T, Chiba T, Takeuchi S, et al. Identification of ketoconazole as an AhR-Nrf2 activator in cultured human keratinocytes: the basis of its anti-inflammatory effect. *J Invest Dermatol.* (2012) 132:59-68. doi: 10.1038/jid.2011.194

50. Yu H, Qiu J, Ma L-J, Hu Y-J, Li P, Wan J-B. Phytochemical and phytopharmacological review of Perilla frutescens L. (Labiatae), a traditional ediblemedicinal herb in China. *Food Chem Toxicol.* (2017) 108:375–91. doi: 10.1016/ j.fct.2016.11.023

51. Hayashi R, Himori N, Taguchi K, Ishikawa Y, Uesugi K, Ito M, et al. The role of the Nrf2-mediated defense system in corneal epithelial wound healing. *Free Radic Biol Med.* (2013) 61:333–42. doi: 10.1016/j.freeradbiomed.2013.04.008

52. Yu B, Wang Q, Zhang L, Lin J, Feng Z, Wang Z, et al. Ebselen improves fungal keratitis through exerting anti-inflammation, anti-oxidative stress, and antifungal effects. *Redox Biol.* (2024) 73:103206. doi: 10.1016/j.redox.2024.103206

53. Sharma S, Sharma D, Dhobi M, Wang D, Tewari D. An insight to treat cardiovascular diseases through phytochemicals targeting PPAR- α . Mol Cell Biochem. (2024) 479:707–32. doi: 10.1007/s11010-023-04755-7

54. Siddiqui MS, Parmar D, Sheikh F, Sarin SK, Cisneros L, Gawrieh S, et al. Saroglitazar, a dual PPAR α / γ Agonist, improves atherogenic dyslipidemia in patients with non-cirrhotic nonalcoholic fatty liver disease: A pooled analysis. *Clin Gastroenterol Hepatol.* (2023) 21:2597–605. doi: 10.1016/j.cgh.2023.01.018

55. Kowdley KV, Bowlus CL, Levy C, Akarca US, Alvares-da-Silva MR, Andreone P, et al. Efficacy and safety of elafibranor in primary biliary cholangitis. *N Engl J Med.* (2024) 390:795–805. doi: 10.1056/NEJMoa2306185

56. Hirschfield GM, Shiffman ML, Gulamhusein A, Kowdley KV, Vierling JM, Levy C, et al. Seladelpar efficacy and safety at 3 months in patients with primary biliary cholangitis: ENHANCE, a phase 3, randomized, placebo-controlled study. *Hepatology*. (2023) 78:397–415. doi: 10.1097/HEP.000000000000395

57. Nowell J, Blunt E, Gupta D, Edison P. Antidiabetic agents as a novel treatment for Alzheimer's and Parkinson's disease. *Ageing Res Rev.* (2023) 89:101979. doi: 10.1016/j.arr.2023.101979

58. Esposito E, Impellizzeri D, Mazzon E, Paterniti I, Cuzzocrea S. Neuroprotective activities of palmitoylethanolamide in an animal model of Parkinson's disease. *PloS One.* (2012) 7:e41880. doi: 10.1371/journal.pone.0041880

59. Yakubov E, Schmid S, Hammer A, Chen D, Dahlmanns JK, Mitrovic I. Ferroptosis and PPAR-gamma in the limelight of brain tumors and edema. *Front Oncol.* (2023) 13:1176038. doi: 10.3389/fonc.2023.1176038

60. Zhai M, Hu H, Zheng Yi, Wu B, Sun W. PGC1 α : an emerging therapeutic target for chemotherapy-induced peripheral neuropathy. *Ther Adv Neurol Disord.* (2023) 16:17562864231163361. doi: 10.1177/17562864231163361

61. Fidaleo M, Fanelli F, Ceru MP, Moreno S. Neuroprotective properties of peroxisome proliferator-activated receptor alpha (PPAR α) and its lipid ligands. *Curr Med Chem*. (2014) 21:2803–21. doi: 10.2174/0929867321666140303143455

62. Kersten S, Stienstra R. The role and regulation of the peroxisome proliferator activated receptor alpha in human liver. *Biochimie*. (2017) 136:75–84. doi: 10.1016/j.biochi.2016.12.019

63. Chinetti-Gbaguidi G, Staels B. PPAR β in macrophages and atherosclerosis. Biochimie. (2017) 136:59–64. doi: 10.1016/j.biochi.2016.12.008

64. Liu Yu, Wang J, Luo S, Zhan Yi, Lu Q. The roles of PPARγ and its agonists in autoimmune diseases: A comprehensive review. J Autoimmun. (2020) 113:102510. doi: 10.1016/j.jaut.2020.102510

65. Tanaka A, Corpechot C. PPAR agonists in PBC: Where do we go from here? Or how to choose between the new and the old. *Clin Res Hepatol Gastroenterol.* (2024) 48:102358. doi: 10.1016/j.clinre.2024.102358

66. Stark JM, Coquet JM, Tibbitt CA. The role of PPAR- γ in allergic disease. Curr Allergy Asthma Rep. (2021) 21:45. doi: 10.1007/s11882-021-01022-x

67. Wang D, Liu F, Zhu L, Lin P, Han F, Wang X, et al. FGF21 alleviates neuroinflammation following ischemic stroke by modulating the temporal and spatial dynamics of microglia/macrophages. *J Neuroinflammation*. (2020) 17:257. doi: 10.1186/s12974-020-01921-2

68. Xu R, Zhang J, Hu X, Xu P, Huang S, Cui S, et al. Yi-shen-hua-shi granules modulate immune and inflammatory damage via the ALG3/PPAR γ /NF- κ B pathway in the treatment of immunoglobulin a nephropathy. *J Ethnopharmacol.* (2024) 319:117204. doi: 10.1016/j.jep.2023.117204

69. Nagy ZS, Czimmerer Z, Szanto A, Nagy L. Pro-inflammatory cytokines negatively regulate PPAR γ mediated gene expression in both human and murine macrophages via multiple mechanisms. *Immunobiology*. (2013) 218:1336–44. doi: 10.1016/j.imbio.2013.06.011

70. Heming M, Gran. S, Jauch S-L, Fischer-Riepe L, Russo A, Klotz L, et al. Peroxisome proliferator-activated receptor- γ Modulates the response of macrophages to lipopolysaccharide and glucocorticoids. *Front Immunol.* (2018) 9:893. doi: 10.3389/fimmu.2018.00893

71. Croasdell A, Duffney PF, Kim N, Lacy SH, Sime PJ, Phipps RP. PPARγ and the innate immune system mediate the resolution of inflammation. *PPAR Res.* (2015) 2015:549691. doi: 10.1155/2015/549691

72. Alatshan A, Benkő S. Nuclear receptors as multiple regulators of NLRP3 inflammasome function. *Front Immunol.* (2021) 12:630569. doi: 10.3389/fimmu.2021.630569

73. Mu P-Y, Chu C, Yu Di, Shao Y, Zhao S-Z. PPARγ: the dominant regulator among PPARs in dry eye lacrimal gland and diabetic lacrimal gland. *Int J Ophthalmol.* (2020) 13:860–9. doi: 10.18240/ijo.2020.06.02

74. Zhang Su, Gu H, Hu N. Role of peroxisome proliferator-activated receptor γ in ocular diseases. J Ophthalmol. (2015) 2015:275435. doi: 10.1155/2015/275435

75. Knop E, Knop N, Millar T, Obata H, Sullivan DA. The international workshop on meibomian gland dysfunction: report of the subcommittee on anatomy, physiology, and pathophysiology of the meibomian gland. *Invest Ophthalmol Vis Sci.* (2011) 52:1938–78. doi: 10.1167/iovs.10-6997c

76. Hwang HoS, Parfitt GJ, Brown DJ, Jester JV. Meibocyte differentiation and renewal: Insights into novel mechanisms of meibomian gland dysfunction (MGD). *Exp Eye Res.* (2017) 163:37–45. doi: 10.1016/j.exer.2017.02.008

77. Escandon P, Vasini B, Whelchel AE, Nicholas SE, Matlock HG, Ma J-X. The role of peroxisome proliferator-activated receptors in healthy and diseased eyes. *Exp Eye Res.* (2021) 208:108617. doi: 10.1016/j.exer.2021.108617

78. Nakano Y, Arima T, Tobita Y, Uchiyama M, Shimizu A, Takahashi H. Combination of peroxisome proliferator-activated receptor (PPAR) alpha and gamma agonists prevents corneal inflammation and neovascularization in a rat alkali burn model. *Int J Mol Sci.* (2020) 21:5093. doi: 10.3390/ijms21145093

79. Aiello F, Afflitto GG, Li J-PO, Martucci A, Cesareo M, Nucci C. CannabinEYEds: the endocannabinoid system as a regulator of the ocular surface nociception, inflammatory response, neovascularization and wound healing. *J Clin Med.* (2020) 9:4036. doi: 10.3390/jcm9124036

80. Peng W-Y, Wang F, Yang S-J, Sun Q-Y, Zhou H-S, Li X, et al. Therapeutic effects of a novel venom abstract (ZK002) solution in an alkali-burned corneal wound-healing model. *Mol Vis.* (2023) 29:317–28.

81. Li Bo, Mei X-F. Naringin may promote functional recovery following spinal cord injury by modulating microglial polarization through the PPAR-γ/NF-κB signaling pathway. *Brain Res.* (2023) 1821:148563. doi: 10.1016/j.brainres.2023.148563

82. Zamanian MY, Alsaab HO, Golmohammadi M, Yumashev A, Jabba AM, Abid MK, et al. NF-κB pathway as a molecular target for curcumin in diabetes mellitus treatment: Focusing on oxidative stress and inflammation. *Cell Biochem Funct.* (2024) 42:e4030. doi: 10.1002/cbf.4030

 Lin G, Lin L, Chen X, Chen L, Yang J, Chen Y. PPAR-γ/NF-kB/AQP3 axis in M2 macrophage orchestrates lung adenocarcinoma progression by upregulating IL-6. *Cell Death Dis.* (2024) 15:532. doi: 10.1038/s41419-024-06919-9

84. Tobita Y, Arima T, Nakano Y, Uchiyama M, Shimizu A, Takahashi H. Effects of selective peroxisome proliferator activated receptor agonists on corneal epithelial wound healing. *Pharm (Basel)*. (2021) 14:88. doi: 10.3390/ph14020088

85. Blunder S, Krimbacher T, Moosbrugger-Martinz V, Gruber R, Schmuth M, Dubrac S. Keratinocyte-derived IL-1 β induces PPARG downregulation and PPARD upregulation in human reconstructed epidermis following barrier impairment. *Exp Dermatol.* (2021) 30:1298–308. doi: 10.1111/exd.14323

86. Tobita Y, Arima T, Nakano Y, Uchiyama M, Shimizu A, Takahashi H. Peroxisome proliferator-activated receptor beta/delta agonist suppresses inflammation and promotes neovascularization. *Int J Mol Sci.* (2020) 21:5296. doi: 10.3390/ijms21155296

87. Roszkowska AM, Licitra C, Tumminello G, Postorino EI, Colonna MR, Aragona P. Corneal nerves in diabetes-The role of the *in vivo* corneal confocal microscopy of the subbasal nerve plexus in the assessment of peripheral small fiber neuropathy. *Surv Ophthalmol.* (2021) 66:493–513. doi: 10.1016/j.survophthal.2020.09.003

88. Teo CHY, Lin MT-Y, Lee IXY, Koh S-K, Zhou L, Goh DS, et al. Oral peroxisome proliferator-activated receptor- α Agonist enhances corneal nerve regeneration in patients with type 2 diabetes. *Diabetes*. (2023) 72:932–46. doi: 10.2337/db22-0611

89. Greg Matlock H, Qiu F, Malechka V, Zhou K, Cheng R, Benyajati S, et al. Pathogenic role of PPARα Downregulation in corneal nerve degeneration and impaired corneal sensitivity in diabetes. *Diabetes*. (2020) 69:1279–91. doi: 10.2337/db19-0898

90. Chou Y, Ma J, Su X, Zhong Y. Emerging insights into the relationship between hyperlipidemia and the risk of diabetic retinopathy. *Lipids Health Dis.* (2020) 19:241. doi: 10.1186/s12944-020-01415-3

91. Keech AC, Mitchell PA, Summanen, O'Day J, Davis TME, Moffitt MS. Effect of fenofibrate on the need for laser treatment for diabetic retinopathy (FIELD study): a randomised controlled trial. *Lancet.* (2007) 370:1687–97. doi: 10.1016/S0140-6736(07) 61607-9

92. Noonan JE, Jenkins AJ, Ma J-X, Keech AC, Wang JJ, Lamoureux EL. An update on the molecular actions of fenofibrate and its clinical effects on diabetic retinopathy and other microvascular end points in patients with diabetes. *Diabetes*. (2013) 62:3968– 75. doi: 10.2337/db13-0800

93. Jenkins AJ, Joglekar MV, Hardikar AA, Keech AC, O'Neal DN, Januszewski AS. Biomarkers in diabetic retinopathy. *Rev Diabetes Stud.* (2015) 12:159–95. doi: 10.1900/RDS.2015.12.159

94. Sun J, Bian Y, Ma Y, Ali W, Wang T, Yuan Y, et al. Melatonin alleviates cadmium-induced nonalcoholic fatty liver disease in ducks by alleviating autophagic flow arrest via PPAR- α and reducing oxidative stress. *Poult Sci.* (2023) 102:102835. doi: 10.1016/j.psj.2023.102835

95. Grabacka M, Pierzchalska Małgorzata, Płonka PrzemysławM, Pierzchalski P. The role of PPAR alpha in the modulation of innate immunity. *Int J Mol Sci.* (2021) 22:10545. doi: 10.3390/ijms221910545

96. Yang De, Han Z, Oppenheim JJ. Alarmins and immunity. *Immunol Rev.* (2017) 280:41-56. doi: 10.1111/imr.12577

97. Blas-García A, Apostolova N, Blas-García A. Novel therapeutic approaches to liver fibrosis based on targeting oxidative stress. *Antioxidants (Basel)*. (2023) 12:1567. doi: 10.3390/antiox12081567

98. Luo W, Xu Q, Wang Qi, Wu H, Hua J. Effect of modulation of PPAR- γ activity on Kupffer cells M1/M2 polarization in the development of non-alcoholic fatty liver disease. *Sci Rep.* (2017) 7:44612. doi: 10.1038/srep44612

99. Chen H, Tan H, Wan J, Zeng Y, Wang J, Wang H, et al. PPAR-gamma signaling in nonalcoholic fatty liver disease: Pathogenesis and therapeutic targets. *Pharmacol Ther.* (2023) 245:108391. doi: 10.1016/j.pharmthera.2023.108391

100. Fang X, Cao J, Tao Z, Yang Z, Dai Y, Zhao L. Hydroxytyrosol attenuates ethanol-induced liver injury by ameliorating steatosis, oxidative stress and hepatic inflammation by interfering STAT3/iNOS pathway. *Redox Rep.* (2023) 28:2187564. doi: 10.1080/13510002.2023.2187564

101. Peng L, Zhong J, Xiao Y, Wang B, Li S, Deng Y. Therapeutic effects of an anti-IL-6 antibody in fungal keratitis: Macrophage inhibition and T cell subset regulation. *Int Immunopharmacol.* (2020) 85:106649. doi: 10.1016/j.intimp.2020.106649

102. Legler DF, Thelen M. Chemokines: chemistry, biochemistry and biological function. *Chimia (Aarau).* (2016) 70:856–9. doi: 10.2533/chimia.2016.856

103. Mughees M, Kaushal JB, Sharma G, Wajid S, Batra SK, Siddiqui JA. Chemokines and cytokines: Axis and allies in prostate cancer pathogenesis. *Semin Cancer Biol.* (2022) 86:497–512. doi: 10.1016/j.semcancer.2022.02.017

104. Xu H, Chen S, Meng C, He Yi, Huang X-J, You H-B. Inhibition of CC chemokine receptor 1 ameliorates osteoarthritis in mouse by activating PPAR- γ . Mol Med. (2024) 30:74. doi: 10.1186/s10020-024-00823-w

105. Nicholas MP, Mysore N. Corneal neovascularization. Exp Eye Res. (2021) 202:108363. doi: 10.1016/j.exer.2020.108363

106. Chang J-H, Garg NK, Lunde E, Han K-Y, Jain S, Azar DT. Corneal neovascularization: an anti-VEGF therapy review. *Surv Ophthalmol.* (2012) 57:415-29. doi: 10.1016/j.survophthal.2012.01.007

107. Mukwaya A, Lennikov A, Xeroudaki M, Mirabelli P, Lachota M, Jensen L. Time-dependent LXR/RXR pathway modulation characterizes capillary remodeling in inflammatory corneal neovascularization. *Angiogenesis*. (2018) 21:395–413. doi: 10.1007/s10456-018-9604-y

108. Wilson SE. Topical losartan: practical guidance for clinical trials in the prevention and treatment of corneal scarring fibrosis and other eye diseases and disorders. J Ocul Pharmacol Ther. (2023) 39:191–206. doi: 10.1089/jop.2022.0174

109. Wilson SE. TGF beta -1, -2 and -3 in the modulation of fibrosis in the cornea and other organs. *Exp Eye Res.* (2021) 207:108594. doi: 10.1016/j.exer.2021.108594

110. Chaurasia SS, Kaur H, Medeiros FWde, Smith SD, Wilson SE. Dynamics of the expression of intermediate filaments vimentin and desmin during myofibroblast differentiation after corneal injury. *Exp Eye Res.* (2009) 89:133–9. doi: 10.1016/ j.exer.2009.02.022

111. Kökény Gábor, Calvier L, Hansmann G. PPARγ and TGFβ-major regulators of metabolism, inflammation, and fibrosis in the lungs and kidneys. *Int J Mol Sci.* (2021) 22:10431. doi: 10.3390/ijms221910431

112. Ma Y, Lu Li, Tan K, Li Z, Guo T, Wu Y, et al. Reduced peroxisome proliferatoractivated receptor- α and bile acid nuclear receptor NR1H4/FXR may affect the hepatic immune microenvironment of biliary atresia. *Front Immunol.* (2022) 13:875593. doi: 10.3389/fimmu.2022.875593

113. Chen L, Li L, Chen J, Li L, Zheng Z, Ren J, et al. Oleoylethanolamide, an endogenous PPAR- α ligand, attenuates liver fibrosis targeting hepatic stellate cells. Oncotarget. (2015) 6:42530–40. doi: 10.18632/oncotarget.6466

114. Comella F, Lama A, Pirozzi C, Annunziata C, Piegari G, Sodano F, et al. Oleoylethanolamide attenuates acute-to-chronic kidney injury: *in vivo* and *in vitro* evidence of PPAR- α involvement. *BioMed Pharmacother*. (2024) 171:116094. doi: 10.1016/j.biopha.2023.116094

115. Barbier O, Pineda Torra I, Duguay Y, Blanquart C, Fruchart J-C, Glineur C, et al. Pleiotropic actions of peroxisome proliferator-activated receptors in lipid metabolism and atherosclerosis. *Arterioscler Thromb Vasc Biol.* (2002) 22:717–26. doi: 10.1161/01.atv.0000015598.86369.04

116. Wang X, Tang L, Zhang Z, Li W, Chen Y. Keratocytes promote corneal neovascularization through VEGFr3 induced by PPAR α -inhibition. *Exp Eye Res.* (2020) 193:107982. doi: 10.1016/j.exer.2020.107982

117. Santana-Garrido Álvaro, Reyes-Goya C, Arroyo-Barrios A, André H, Vázquez CM, Mate A. Hypertension secondary to nitric oxide depletion produces oxidative imbalance and inflammatory/fibrotic outcomes in the cornea of C57BL/6 mice. *J Physiol Biochem.* (2022) 78:915–32. doi: 10.1007/s13105-022-00916-2

118. Gurnani B, Czyz CN, Mahabadi N, Havens SJ. Corneal graft rejection. In: *StatPearls*. StatPearls Publishing, Treasure Island (FL (2024).

119. Liu S, Wong YL, Walkden A. Current perspectives on corneal transplantation. *Clin Ophthalmol.* (2022) 16:631–46. doi: 10.2147/OPTH.S289359

120. Niederkorn JY. Immune mechanisms of corneal allograft rejection. Curr Eye Res. (2007) 32:1005–16. doi: 10.1080/02713680701767884

121. Moshirfar M, Basharat NF, Seitz TS, Ply BK, Ronquillo YC, Hoopes PC. Corneal transplant rejections in patients receiving immune checkpoint inhibitors. J Clin Med. (2022) 11:5647. doi: 10.3390/jcm11195647

122. Lan Xu, Zhang J, Ren S, Wang H, Shao Bo, Qin Y, et al. Oxymatrine combined with rapamycin to attenuate acute cardiac allograft rejection. *Heliyon*. (2024) 10: e29448. doi: 10.1016/j.heliyon.2024.e29448

123. Sousa CRe. Dendritic cells in a mature age. *Nat Rev Immunol.* (2006) 6:476–83. doi: 10.1038/nri1845

124. Piccotti JR, Li K, Chan SY, Eichwald EJ, Bishop DK. Cytokine regulation of chronic cardiac allograft rejection: evidence against a role for Th1 in the disease process. *Transplantation*. (1999) 67:1548–55. doi: 10.1097/00007890-199906270-00008

125. Ye P, Li J, Wang S, Xie A, Sun W, Xia J. Eicosapentaenoic acid disrupts the balance between Tregs and IL-17+ T cells through PPAR γ nuclear receptor activation and protects cardiac allografts. J Surg Res. (2012) 173:161–70. doi: 10.1016/j.jss.2010.08.052

126. Huang X, Ren L, Ye P, Cheng C, Wu J, Wang S, et al. Peroxisome proliferatoractivated receptor γ deficiency in T cells accelerates chronic rejection by influencing the differentiation of CD4+ T cells and alternatively activated macrophages. *PloS One.* (2014) 9:e112953. doi: 10.1371/journal.pone.0112953