Check for updates

OPEN ACCESS

EDITED BY DuoYao Cao, Cedars Sinai Medical Center, United States

REVIEWED BY Stephan Immenschuh, Hannover Medical School, Germany Norbert Leitinger, University of Virginia, United States Xizi Zhang, Gilead, United States

*CORRESPONDENCE Antonio Sica antonio.sica@uniupo.it; antonio.sica@humanitasresearch.it

[†]These authors have contributed equally to this work

RECEIVED 15 May 2024 ACCEPTED 01 October 2024 PUBLISHED 24 October 2024

CITATION

Consonni FM, Incerti M, Bertolotti M, Ballerini G, Garlatti V and Sica A (2024) Heme catabolism and heme oxygenase-1expressing myeloid cells in pathophysiology. *Front. Immunol.* 15:1433113. doi: 10.3389/fimmu.2024.1433113

COPYRIGHT

© 2024 Consonni, Incerti, Bertolotti, Ballerini, Garlatti and Sica. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Heme catabolism and heme oxygenase-1-expressing myeloid cells in pathophysiology

Francesca Maria Consonni^{1,2†}, Martina Incerti^{1,2†}, Milena Bertolotti^{3†}, Giulia Ballerini^{1,2†}, Valentina Garlatti^{1,2†} and Antonio Sica^{1,2*}

¹Department of Pharmaceutical Sciences, University of Piemonte Orientale "A. Avogadro", Novara, Italy, ²IRCCS Humanitas Research Hospital, Rozzano, Milan, Italy, ³Navita S.r.l., University of Eastern Piedmont A. Avogadro, Novara, Italy

Although the pathological significance of myeloid cell heterogeneity is still poorly understood, new evidence indicates that distinct macrophage subsets are characterized by specific metabolic programs that influence disease onset and progression. Within this scenario, distinct subsets of macrophages, endowed with high rates of heme catabolism by the stress-responsive enzyme heme oxygenase-1 (HO-1), play critical roles in physiologic and pathological conditions. Of relevance, the substrates of HO-1 activity are the heme groups that derive from cellular catabolism and are converted into carbon monoxide (CO), biliverdin and Fe2+, which together elicit anti-apoptotic, anti-inflammatory activities and control oxidative damage. While high levels of expression of HO-1 enzyme by specialized macrophage populations (erythrophagocytes) guarantee the physiological disposal of senescent red blood cells (i.e. erythrocateresis), the action of HO-1 takes on pathological significance in various diseases, and abnormal CO metabolism has been observed in cancer, hematological diseases, hypertension, heart failure, inflammation, sepsis, neurodegeneration. Modulation of heme catabolism and CO production is therefore a feasible therapeutic opportunity in various diseases. In this review we discuss the role of HO-1 in different pathological contexts (i.e. cancer, infections, cardiovascular, immune-mediated and neurodegenerative diseases) and highlight new therapeutic perspectives on the modulation of the enzymatic activity of HO-1.

KEYWORDS

HO-1, immunosuppression, innate immunity, myeloid cells, immunemetabolism

Introduction

The growing interest on the relevance of myeloid cell specialization in disease (1), is fueling new efforts to therapeutically re-educate their functions (2). Notably, macrophage phenotypic heterogeneity originates during fetal development from embryonic progenitors, which differentiate into self-regenerating subsets of tissue-resident macrophages (TRMs) and become essential for tissue homeostasis and repair (3). A list of abbreviations and acronyms used in the manuscript has been included in Table 1. In adults, blood monocytes are derived primarily from the bone marrow and orchestrate effector and repair functions of defense, also acquiring phenotypic traits of TRMs. In pathology, immunological stresses promote alterations of the myelopoietic output, defined as emergency (4), which lead to the generation of different myeloid populations endowed with specialized functions and distinct metabolic traits (5). This emergency response largely depends on inflammatory signals, which instruct differentiation and maturation of hematopoietic precursors, in a demand-adapted fashion (6). Although such heterogeneity may offer both beneficial and detrimental contributions to therapy, we are not yet able to select the myeloid phenotype with the greatest benefit. New studies are now delineating distinct myeloid subtypes, whose functional specializations appear to take on distinct pathophysiological meanings in the different pathologies, including cancer (7).

The HO-1 enzyme encoded by the Hmox1 gene was first described in 1968 as a key microsomal component, which together with the HO-2 enzyme isoform, encoded by the Hmox2 gene, forms the heme oxygenase (HO) system involved in catalyzing heme oxidation and its related degradation. While Hmox1 can be

induced throughout all tissue types but is most highly expressed within the spleen and liver, Hmox2 is constitutively and ubiquitously expressed, with specific enrichment in the testes and brain (8). Further, in contrast to HO-1, which is mainly involved in iron homeostasis, angiogenesis, mitochondrial function, and regulation of innate and adaptive immunity, regulating tissue responses to damage in pathophysiological states (9), HO-2 appears more as a physiological regulator of cellular functions and is involved in oxygen and redox sensing, neovascularization and neuroprotection (10).

The process of heme synthesis has been extensively reviewed 11-13). While erythrocytes produce around 85% of total heme content in the organism necessary for complete hemoglobinization, the predominant portion of the remaining synthesis occurs in the liver, which is highly enriched in cytochromes p450 (14). The biological activities and metabolism of free heme have been studied extensively, due to its hydrophobic and highly reactive properties, which confer it the ability to induce the formation of hydroxyl radicals, thereby promoting lipid oxidation, protein damage, and cell death (15). While hemoglobin heme undergoes synthesis in erythrocytes and ultimate degradation in the reticuloendothelial system (12), the final products of heme catabolism catalyzes the breakdown of heme into biliverdin, carbon monoxide (CO), and free iron, which display powerful anti-inflammatory and antioxidant potential (Figure 1) (12, 16). As depicted in Figure 1, cytochrome p450 reductase reduces the HO-1/ferrous heme complex, generating ferrous heme which binds and activates molecular oxygen through a protonation reaction, that in turn determines the formation of the reactive intermediate Fe3+-OOH and the formation of hydroxyheme. Next, oxygen activation promotes the conversion of ferric hydroxyheme to



FIGURE 1

The heme oxygenase enzyme reaction and products. Heme is enzymatically degraded into biliverdin (BV), carbon monoxide (CO) and Iron. Biliverdin (BV) is subsequently transformed into bilirubin (BR) by biliverdin reductase (BVR). Iron can be bound by Ferritin, a protein able to store iron. The heme molecule provides a variety of fundamental biological functions as forming of various apo-heme proteins like hemoglobin, nitric oxide synthase and cytochromes. The HO-1-derived product CO could exert anti-inflammatory, anti-proliferation, anti-apoptosis, and vasodilation effects in immune system. BR can perform a crucial role in anti-inflammation, anti-proliferation, anti-apoptosis, anti-oxidation, and free radical scavenger. Moreover, iron-induced ferritin could play a cytoprotective, anti-oxidative and iron storage effect. A list of abbreviations and acronyms used in the manuscript has been included in Table 1.

10.3389/fimmu.2024.1433113

ferrous verdoheme and CO. A final round of oxygen binding and activation of Ferrous verdoheme determines the cleavage of the heme porphyrin ring and the generation of ferric iron biliverdin. Next, NADPH-cytochrome p450 reductase imposes the reduction of ferric biliverdin and the consequent release of free ferrous, as well as of biliverdin that is metabolized to bilirubin by biliverdin reductase (17). This cascade of reactions is thought to occur in respirating organisms whose cell types express and require the regulation of heme. Here, we review the homeostatic and immunoregulatory functions of HO-1, whose induction differentially affect the course of various pathologies.

Role of myeloid heme oxygenase 1 in tumor progression

A vast literature points out that the inducible enzyme HO-1 plays a pivotal role in cellular adaptation to various stress factors, from oxidative insults to hypoxia, thus maintaining redox homeostasis and preventing cell damage through its cytoprotective, antioxidant and pro-surviving properties (18-20). As such, it is not surprising that HO-1 is widely expressed in cancer (i.e. glioblastoma, melanoma, bladder, breast, colorectal, renal cancer, prostate) (21) and that its overexpression correlates with cancer invasiveness, resistance to therapies and poor prognosis in different tumors (21-26). In particular, TCGA pan-cancer analysis shows that Hmox-1 expression is significantly associated with epithelial-mesenchymal transition (EMT) in most tumors and accordingly, HO-1 target genes modulate adhesion, signaling, transport and other key cellular functions in neoplastic cells, promoting tumor cells proliferation and dissemination (27, 28). Indeed, heme and iron metabolism are key determinants of energy metabolism, hence affecting cancer cell functions and immune responses (29), as well as metabolic disorders (30).

In support of this conclusion, genetic polymorphisms of HO-1 promoter have been associated with an increased risk of cancer progression and a high degree of therapy failure (31). Moreover, alteration of the Kelch-like ECH-associated protein 1/nuclear factor erythroid 2-related factor 2 (Keap1/Nrf2) pathway, a major regulator of cytoprotective responses to oxidative that continuously targets Nrf2 for proteasome degradation (32), is also associated with tumor progression (33). Under stress conditions, the conformational change of Keap1 promotes the transcriptional activity of Nrf2 and consequently the expression of antioxidant genes such as heme oxygenase 1 (HO-1) (34). Recently, KEAP1 mutations were connected to adverse outcomes in NSCLC patients undergoing immunotherapy (33). Of relevance, Nrf2 is involved in the regulation of various molecules and enzymatic activities that control drug metabolism (35) as well as in multidrug resistance (36). In agreement, Di Biase et al. reported in the 4T1 breast cancer and B16 melanoma models that fasting mimicking-diet (FMD) caused reduction in HO-1 expression in cancer cells, sensitizing them to doxorubicin and cyclophosphamide, promoting intratumoral accumulation of cytotoxic CD8⁺ T cells and reducing tumor-associated Tregs activation (37).

Despite this evidence mainly poses for the protumoral role of HO-1, controversial anti-tumoral roles of HO-1 has also been reported (i.e. hepatocellular carcinoma, lung mucoepidermoid carcinoma) (38, 39), highlighting the complexity of the HO-1 activity in cancer and suggesting that HO-1 mediated functions are cell type, metabolism and TME dependent (23). These discrepancies also emerge from the specialized role that HO-1 displays during different stages of tumor development. In fact, while HO-1 deficiency in normal cells enhances DNA damage and carcinogenesis, HO-1 overexpression in cancer cells promotes cancer cell proliferation, survival and tumor angiogenesis (40).

HO-1-expressing protumor myeloid cells

The tumor-promoting action of HO-1 is implemented by the biologically active catabolites of heme degradation not only through their action on tumor cells, but also hampering immune cells functionality and favoring pro-metastatic events (41). Borrowing from the physiological role of HO-1-expressing macrophages during the process of erythrophagocytosis (42), HO-1 expression has been frequently reported in malignant cells and tumor associated macrophages (TAMs) (7, 43-46). However, HO-1 can also be expressed by dendritic cells, regulatory T cells and endothelial cells (25, 47, 48). Thus, the tissue-specific contribution of HO-1 in tumor progression and as a therapeutic target still represents a hot undefined topic in cancer research. Within this scattered scenario, the major tumoral source of myeloid-related HO-1 are TAMs (7). These highly plastic cells are able to express distinct transcriptional and functional programs in response to the different cues in the TME, adopting distinct spectra of pro- or antiinflammatory phenotypes (49, 50). Due to the breadth of its tumorpromoting roles and anti-inflammatory activities, HO-1 expression has been associated with biased M2-like TAMs and several recent reports proved its selective up-regulation in specific TAM subsets (7, 22, 44, 45, 51, 52). Reflecting the opposite and dynamic relationship intervening between therapeutic and pathological inflammation towards tumor development (53), the activity of HO-1 appears to be stage-dependent, with preventive function in the early oncogenic phases and of promotion action during tumor progression (54).

In preclinical models of Lewis lung carcinoma (LL2) and pancreatic ductal adenocarcinoma (PDAC), a distinct subset of $HO-1^+$ F4/80^{hi} TAMs with an M2 phenotype co-expressing the fibroblast activation protein alpha (FAP⁺ HO-1⁺ TAMs) and representing 10% of the total TAMs population, has been described to be the major tumor source of HO-1 (51). Conditional ablation of FAP⁺ HO-1⁺ TAMs in an immunogenic ovalbumin (OVA)-expressing LL2 tumor, using diphtheria toxin in a bone marrow chimera of a FAP/diphtheria toxin receptor (DTR) transgenic mouse, or pharmacological inhibition of HO-1 with tin mesoporphyrin (SnMP), decreased LL2 and PDAC tumor growth, confirming the pro-tumoral and immunosuppressive role of TAMsderived HO-1 (51, 55). FAP⁺ TAMs have been described also in human (56) and murine breast tumors (44). By using the preclinical 4T1 model of breast adenocarcinoma, Muliaditan et al. (44) showed that an IL-6dependent protumorigenic FAP⁺ HO-1⁺ TAM population expresses a cytokine profile similar to that which characterizes wound repair. Since inhibition of HO-1 results in a delay in wound closure (57), the action of HO-1 in tumorigenic processes is increasingly consistent with the Dvorak's definition of tumors as "non-healing wounds" (58). Of note, FAP⁺ HO-1⁺ TAMs were predominantly located in the perivascular region of tumors and supported trans-endothelial tumor cells migration and metastatic spread. The study also demonstrated that the heme catabolite carbon monoxide (CO) directly facilitates tumor cells migration and, accordingly, SnMPIX-mediated pharmacological inhibition of HO-1 prevented metastatic spread (44).

In line, a distinct subset of bone marrow-derived M2 polarized F4/80^{hi} HO-1⁺ TAMs was recently described to play a specialized role in forming a pro-metastatic TME, promoting immunosuppression, EMT transition of angiogenesis and inhibition of T cell-mediated antitumor immunity (7). This population accumulates in the blood of cancer bearers (i.e. fibrosarcoma (MN/MCA1) and melanoma (B16/ F10)) and preferentially localizes at the invasive tumor margins under the influence of NF-KB1/CSF1-R/C3a signaling axis, which converges on Nrf2 activation to support HO-1 expression. Importantly, pharmacological inhibition with zinc protoporphyrin IX (ZnPPIX) or myeloid-specific ablation of HO-1 blocked metastasis formation and improved the efficacy of anti-PD-1-mediated immunotherapy (7). Noteworthy, HO-1 expression in peripheral monocyte subsets as well as in tumor lesions discriminates survival among metastatic melanoma patients, suggesting HO-1 myeloid cells as a new prognostic indicator and a novel antimetastatic target (7).

In an aggressive spontaneous mouse model of breast cancer (MMTV-PyMT), TAMs were reported to be the major intratumoral source of HO-1, playing a crucial role in orchestrating immunosuppressive circuits that occur in the TME (22). Consistently, in this experimental setting, specific conditional ablation of HO-1 in the myeloid lineage improved the proportion of cytotoxic CD8⁺ T cells expressing IFNy, granzyme B and TNFa, while pharmacological inhibition of HO-1, using SnMPIX, increased the antitumor activity of 5-fluorouracil (5-FU) in a CD8⁺ T cell-dependent manner (22). Importantly, HO-1 inhibition appeared as a potentially crucial immunotherapy target, hierarchically more important than PD-1. Moreover, treatment of MMTV-PyMT tumors with SnPPIX displayed increase response to chemotherapy compared to anti-PD-1 neutralizing antibody, suggesting that SnPPIX could be used as immune checkpoint therapy (22). The link between HO-1expressing TAMs and cancer progression was further supported in immunogenic OVA-expressing murine thymic lymphoma model (EG/-OVA) (45). In this tumor, HO-1 is upregulated during the differentiation of monocytic Ly6Chi cells into TAMs, becoming a key molecular effector of their immunosuppressive functions. Specific myeloid ablation of HO-1 or its pharmacological inhibition increased the anti-tumor response, improving the antitumor efficacy of a therapeutic anti-tumor OVA vaccine (45). It was also recently described an essential role of CX3CR1+ gut macrophages in resolving inflammation in the intestine, where they protect against colitis-associated cancer by regulating HMOX-1 expression (59).

Noteworthy, HO-1 could directly skew the polarization of macrophages (60); indeed, myeloid-specific ablation of HO-1 in bone marrow-derived macrophages (BM-DMs) treated with either M1- (LPS) or M2- (IL-4) polarizing signals produced an increase in the expression of M1 markers (CXCL10, IL-1b, MCP1), along with the decrease of M2 markers (Arg1 and CD163) (61). Other evidence shows in a murine breast cancer model (4T1) that following phagocytosis of cellular debris from tumor cells treated with chemotherapy (paclitaxel) TAMs upregulate the expression of HO-1, which in turn hinders M1 polarization and attenuates the response to chemotherapy. In contrast, genetic or pharmacological inhibition of HO-1 in TAMs reinstates M1-like polarization by restoring an immunogenic TME during chemotherapy, favoring the recruitment and activation of cytotoxic CD8+ T cells (62, 63). Along this line, Magri et al. reported that pharmacological inhibition of HO-1 in BM-DMs isolated from glioblastoma samples, with the HO-1 inhibitors ZnPPIX and OB-24, significantly reduced cell-tocell- (i.e. PD-L1/PD-1) and soluble-dependent (i.e. IL-10) immunosuppressive mechanisms. Strikingly, HO-1 inhibition also prevented expression of immunosuppressive enzymes involved in amino acid catabolism (i.e. IDO1 and ARG2) (64). HO-1 inhibition by ZnPPIX was reported to repolarize M2-like protumor TAMs to antitumor M1-like macrophages also in the 4T1 breast cancer model (65). The influence of HO-1 on the TME immunoprofile was highlighted by Alaluf et al., who reported that myeloid-specific ablation of HO-1 induced global transcriptional and epigenetic alterations not limited to the conventional M1/M2 polarization state of TAMs, but rather leading to extensive dysregulation of the central molecular signature of the TME (45).

The HO-1/CO pathway in cancer development

CO is a highly toxic gas due to its high affinity for hemoglobin, 250 times greater than oxygen. Due to its strong bond with heme iron, CO inhibits the transport of oxygen in the blood, while interacting with cytochrome c oxygenase and cytochrome p450 it inhibits cellular respiration and promotes tissue death (66). Despite the role of CO gasotransmitter has been the subject of various studies, its role in cancer is still largely unclear and both tumorpromoting and anti-tumor activities have been reported (67). It was demonstrated that CO directly modulates macrophage polarization in vitro, skewing their phenotype toward the anti-inflammatory one (68) and dampening their activation through M1-oriented signaling (i.e. TLR and MAPK) (69), with downregulation of the proinflammatory cytokines TNF α and IL-1 β and increased IL-10 production (20). Consistently, in non-cancer settings, in vitro treatment of human monocyte-derived DCs with CO has been reported to block TLR3- and 4-induced phenotypic maturation and alloreactive T cell proliferation (70).

Accordingly, *in vitro* treatment of macrophages with carbonmonoxide releasing molecules (CORMs) enhances STAT3/STAT6 activation to induce their anti-inflammatory phenotype (7). In contrast, by using the *in vivo* A549 lung carcinoma model, Nemeth et al. showed that exposure to low doses of exogenous CO polarizes macrophages toward a pro-inflammatory M1-like phenotype through ROS-dependent activation of MAPK/Erk1/2-c-myc pathway, which negatively regulates HO-1 expression leading to an anti-tumor effect (71).

HO-1 catabolites can also influence the activation states and phenotype of a variety of cell populations in the TME, enhancing immune evasion and ultimately contributing to tumor progression (41). In this regard, CO inhibits the maturation of dendritic cells (DCs) by maintaining them in a pro-tolerogenic state and supporting HO-1 expression through increased IL-10 production (47, 70). Consistently, HO-1 expression contributes to imprint a pro-tolerogenic signature of DCs (72). The product of heme degradation biliverdin has also been reported to induce IL-10 production in macrophages through a PI3K-Akt dependent pathway (73), while both CO and bilirubin down-regulate expression of MHCII on DCs, restraining their ability to present antigens CD4⁺ T cells (70, 74).

The HO-1/CO axis has also been described to play a critical role in FoxP3-mediated immune suppression (48, 75). In glioma patients, HO-1 mRNA expression has been linked to Foxp3 induction in infiltrating CD4⁺CD25⁺ Treg cells and correlated with glioma progression and grading (48), while in the preclinical model of malignant glioma the expression of HO-1 improved the survival of Tregs in the hypoxic regions of the TME (75). HO-1mediated suppression of T cell proliferation is not only associated with Treg cell expansion, as growing evidence demonstrates the ability of HO-1/CO to directly block the expansion of effector T cell populations (76). CO has been described to suppress the secretion of IL-2, a cytokine required for T-cell entry into the cell cycle (76) and CO-mediated inhibition of CD3-activated T cell proliferation is highly dependent on caspase-3 and -8, which are regulated by $p21^{Cip1}$ (77). HO-1 was also reported to suppress natural killer (NK) activation and their effector functions, through interfering with the expression of activator receptors (NKG2D, NKp46 and NKp30), as well as blocking their ability to secrete IFN γ and TNF α (78). In acute myeloid leukemia (AML), HO-1 has also been described to prevent cytotoxic effects of NK by inhibiting CD48 expression, a ligand of the NK-activating receptor 2B4, through Sirt1-H3K27dependent pathway (79). Numerous studies also highlight an intriguing direct pro-metastatic role of HO-1/CO on tumor cells (28). CO has been reported to increase glioma cell survival and migration (80-82), while HO-1 silencing using small interfering RNAs (siRNAs) causes downregulation of VEGF-induced vimentin and endothelial cell proliferation, resulting in impaired angiogenesis and reduced tumor progression (81). In agreement, myeloid expression of HO-1 facilitates tumor metastasis by promoting the formation of a premetastatic niche and increasing tumor colonization at the metastatic site (82).

The preclinical and clinical observations available today are consistent with the crucial role played by HO-1 in arresting cancer immune recognition and supporting tumor progression and pharmacological targeting of HO-1 and its catabolites is acquiring solid confirmation as promising anticancer therapy.

HO-1 in immune-mediated diseases

HO-1 plays a critical role in the maintenance of immunehomeostasis, as this enzyme elicits a strong impact in both innate and adaptive immune responses. In particular, the expression of heme oxygenase by immune cells and, later on, its products of reaction exert extensive antioxidant and anti-inflammatory properties associated with beneficial outcomes in inflammatory disorders (18). Consistently, a chronic inflammatory condition is observed in *Hmox1*-knockout mice, paralleled to an accumulation of polymorphonuclear cells, increased number of monocytes in the spleen and lymphocyte count in peripheral blood (83).

In accordance with the beneficial role of this cytoprotective enzyme, different polymorphisms in the Hmox1 promoter region determine different levels of HO-1 induction, associated either with protective functions or with increased susceptibility towards autoimmune and inflammatory diseases, such as rheumatoid arthritis (RA) or systemic lupus erythematosus (SLE) (84). Consolidated evidence also supports a key role of HO-1 in controlling intestinal and lung inflammation (85, 86). Supporting this evidence, the presence of a long allele in a (GT)_n microsatellite polymorphism in Hmox-1 gene promoter leading to decreased expression of HO-1, gives rise to a pronounced risk and higher susceptibility to develop autoimmune disease, such systemic lupus erythematosus (87). On the other hand, the presence of a short (S) allele, provoke an increase in the production of HO-1 leading to a protection in developing immune-mediated disorders (18, 88). The up-regulation or the downregulation of this enzyme in immunemediated disorders therefore makes it a possible inflammatory marker for autoimmune diseases (83).

In RA patients a significant increase of HO-1 levels was observed in synovial fluid and peripheral monocytes, as an adaptive mechanism for limiting inflammation and toxicity (89). Furthermore, induction of HO-1 in SLE confers an anti-inflammatory phenotype to monocytes and DCs, while myeloid cells in these patients show downregulated HO-1 levels, suggesting that its deregulation is involved in disease progression (90). According to the HO-1 protective functions, the anti-inflammatory effects of IL-10 in macrophages appear to be mediated via induction of HO-1 (91, 92).

HO-1-derived CO plays a crucial immunomodulatory role, by acting on different immune cells, including dendritic cells, macrophages and regulatory T cells (93). Indeed, HO-1 induction fosters the polarization of macrophages towards an antiinflammatory M2-like profile and promotes a tolerogenic phenotype in DCs (83, 94), both reducing the activation of T cells and favoring the Treg differentiation, resulting in the suppression of autoreactive responses (47). All this reveals the strong impact that HO-1 elicits in the context of immune-mediated diseases, as well as its centrality for the maintenance of immune homeostasis.

HO-1 in inflammatory bowel diseases

Accumulating evidences indicate that induction of HO-1 expression act as an endogenous defensive mechanism to reduce tissue injury in the intestinal tract associated with chronic disease/IBD), LPS, suggesting the HO-1 pa tis (UC) (95). therapeutic target (107). This

inflammatory conditions (i.e. Inflammatory bowel disease/IBD), including Crohn's disease (CD) and Ulcerative Colitis (UC) (95). Chronic intestinal inflammation is promoted by different conditions, including infections and autoimmune diseases, and is characterized by massive immune cell infiltration, edema and alterations of the epithelium structure (96). Of note, HO-1 is shown to be transcriptionally induced in the intestinal tract in response to oxidative stress (97, 98) and pharmacological evidence supports the protective role of HMOX1 in intestinal inflammation. In the model of colitis induction by administration of Dextran Sulfate Sodium (DSS), administration of HMOX1 inductor/ activator CoPP significantly reduced the intestinal histological damage as compared to control animals (99). This protective response was mimicked by administration of the HMOX1 inducer hemin, which also reduced number of Th17 cells and increased number of Treg cells in mesenteric lymph nodes (MLN) and spleen (100).

Of particular note, a strong association between the development of IBD and the immune response to microbial infections has been described, the mechanistic basis of which are still unclear. In this context, it is important to note that HMOX1-like enzymes are expressed in bacteria and CO can therefore directly interact with the heme groups of the bacterial electron transport chain (101). These evidences support an increasingly central role of HO-1 and CO in the cross-talk between the microbiota and the mucosal immune compartment, suggesting HO-1 as a new therapeutic target for inflammatory bowel disease (102).

HO-1 in inflammatory lung diseases

Mounting evidence indicates that in various pulmonary diseases, such as acute respiratory distress syndrome (ARDS) and interstitial lung disease (ILD), an increase in HO-1 expression in alveolar macrophages reflects the activation of an M2 macrophage phenotype polarized against oxidative stress (103, 104). In agreement, serum heme oxygenase (HO)-1 level has been indicated as a potential marker of acute progression of interstitial lung disease (ILD) (105). Preclinical models of acute lung injury (ALI) (i.e., hyperoxia, sepsis, and ventilator-induced lung injury) have highlighted the protective effects of the HO-1/CO system (93). In the lipopolysaccharide (LPS)-induced lung injury mouse model, upregulation of HO-1 by gene transfer limited neutrophil influx and pro-inflammatory response, protecting against ALI. This protective effect was reproduced by treatment with CO (250 p.p.m.), resulting in downregulation of the p38 MAPK-dependent pro-inflammatory cytokine response and upregulation of IL-10 (20). Furthermore, gene transfer of HMOX1 also protected against ALI induced by influenza A virus infection, reducing the influx of inflammatory cells (106). Neutrophil accumulation is a major cause of damage in many lung diseases, including cystic fibrosis (CF). Macrophages from patients with CF have been observed to exhibit a defect in the heme oxygenase-1 (HO-1)/carbon monoxide (CO) pathway. Recent evidence has shown that systemic administration of PP-007, a CO releasing/O2 transfer agent, rescues the PI3K/HO-1 axis in CF macrophages and decreases the hyperinflammatory response to LPS, suggesting the HO-1 pathway as a potential candidate therapeutic target (107). This is consistent with the abundant expression of HO-1 observed in monocytes/macrophages responsible for resolving the inflammatory response, whose dysregulation leads to hyperinflammation in CF, asthma, COPD and fibrotic lung diseases (108, 109). Human respiratory syncytial virus (hRSV) is the leading cause of severe lower respiratory tract infections in children. Upregulation of HO-1 with cobalt metalloporphyrin protoporphyrin IX (CoPP) in hRSV-infected mice significantly reduced disease-related body weight loss and induced an upregulation of IFN- α/β in the lungs, indicating that HO-1 plays an important role in the development of the antiviral type I IFN response in the airways (106). Despite no direct evidence has been collected on the connection between HO-1 and COVID-19 severity, HO-1 expression has been reported in these patients and its serum levels have been proposed as a useful marker for evaluating disease severity (110, 111).

HO-1 in neurodegenerative disorders

The upregulation of HO-1 in the central nervous system (CNS) constitutes a mechanism of cellular adaptation to stress, which crucially mediates the resolution of neuroinflammation through the activation of antioxidant, antiapoptotic and anti-inflammatory properties (112). In agreement, much evidence indicates a cytoprotective effect of HO-1 in the pathogenesis of neurodegenerative diseases, including amyotrophic lateral sclerosis (ALS), including Alzheimer's disease (AD), Huntington's disease (HD) and Parkinson's disease (PD) (113). Accordingly, while Nrf2-dependent activation of HO-1 is related to a protective and beneficial effect in neurons, this pathway drives the transcription of genes involved in iron quenching, chelation and transport (114, 115), as well as in prevention of lipid peroxidation (116, 117), resulting cytoprotective action on cells and tissues.

In contrast, in conditions of excessive stress, the high production of this enzyme, as well as its metabolites by neuronal or immune cells, favors the deposition of mitochondrial iron and the depletion of bioenergy, which favors the development of degenerative diseases such as multiple sclerosis (MS) or Alzheimer's disease (118).

Abnormal pathways of HO-1 induction via unconventional signaling pathways have been investigated. Among these, accumulation of p62, also known as sequestosome 1 (SQSTM-1) and A170 (119), sequesters Keap1 into aggregates, resulting in the inhibition of Keap1-mediated Nrf2 ubiquitination and its subsequent degradation by the proteasome (120). Consistently, in neurodegenerative diseases it was observed an imbalance in the signaling pathway of HO-1 induction, leading to the loss of normal tissues and immune homeostasis, cellular deterioration and progressive development of neurodegeneration (114). In "stressed" astrocytes, excessive HO-1 activity can lead to mitochondrial iron sequestration, macroautophagy, pathological iron deposition, and bioenergy depletion, which contribute to the development of neurodegenerative disease (AD), Parkinson's disease (PD), and Huntington's disease (HD) (121).

HO-1 in multiple sclerosis

Multiple Sclerosis (MS) is a central nervous system demyelinating neurodegenerative immune-mediated disease characterized by the production of inflammatory lesions in the brain, optic nerve and spinal cord (122). This disorder is also marked by a pro and anti-inflammatory cytokine imbalance, which leads on one side to a progressive degeneration of oligodendrocytes and axons, from another to the proliferation of astrocytes (114). The importance of an equilibrium in the expression of HO-1 appears as a pivotal requirement for the maintenance of protective effect and immune homeostasis preventing the progression of the disease. In the early stage of muscular atrophy, the enzyme plays the role of essential regulator of cytoprotective responses (123).

It was reported that the excessive downregulation of Nrf2/HO-1 signaling pathway substantially enhances neuroinflammation and immune dysregulation associated with oligodendrocyte loss, advancing the stage of disease (114). Moreover, a severe reduction of HO-1 expression in peripheral mononuclear cells (PBMCs) from MS patients was demonstrated during the exacerbation of this disorder (124). The Experimental autoimmune encephalomyelitis (EAE) model has been frequently used to better understand the dual and intricate role of HO-1 in inflammation and its contribution to the immune system in MS (123, 125). It was observed that during the acute phase of disease, EAE mice show increased levels of HO-1 expression in microglia, astrocytes and oligodendrocytes (126). Furthermore, the chronic and constant upregulation of HO-1 by oligodendrocytes has been associated with a damage in glial cells, leading to induction of cell death and severe inflammation. Moreover, the plaques deposed in the spinal cord, during the progression of the disease, show a strong mitochondrial iron deposition induced by the secretion of IL-1 and TNF and provoked by the elevated expression of HO-1 (118).

If on one side the chronic over-expression of HO-1 results detrimental, on the other Hmox1-deficient mice showed an aggravated disease, because of the absence of cytoprotective effects elicited by the HO-1\CO axis. Interestingly, the protective features of HO-1, and in particular of CO, in MS has been associated with the inhibition of both CD4⁺ and CD8⁺ T cell activation and MHC II expression by antigen-presenting cells, including DCs, microglia and infiltrating macrophages (126). Induction of HO-1 expression by iron-containing porphyrin hemin and treatment with the CO donor CORM-A1 protects EAE mice from acute disease, showing improvement in clinical score and reduced incidence of disease infiltration by polymorphonuclear cells (123). Although the mechanism of protection of HO-1 in EAE remains to be investigated, the immunomodulation and cytoprotective properties it exerts on the central nervous system likely provide a preventative role in neuroinflammatory diseases.

HO-1 in Alzheimer's disease

Alzheimer's disease (AD) is a neurodegenerative disorder characterized by a set of brain lesions caused by the accumulation of different proteins such as the β -amyloid in fibrillary plaques,

which give rise to a severe state of inflammation and consequent tissue damage (127). In this setting, the maintenance of the expression's balance in HO-1 signaling pathway appears crucial in controlling the disease progression. Accordingly, in neuronal cells this enzyme plays a protective role, mediating the conversion of prooxidant heme into its degradation and antioxidant products biliverdin (BV) and bilirubin (BR), which are crucial in the promotion of an appropriate tissue redox microenvironment (112, 128). It is established that HO-1 is over-expressed in the brain of AD patients, mainly in the hippocampus cerebral cortex, and is co-localized to neurons and neurofibrillary tangles. During acute brain damage this cytoprotective enzyme can be subjected to a protracted up-regulation leading to a detrimental effect for the neuronal tissue (115). The accumulation of mitochondrial iron, enhanced by glial HO-1 activity, leads to toxicity, protein accumulation and neuronal death. Further, the abnormal brain iron mobilization may amplify the oxidative stress and contribute to the progression of AD (129). Supporting the importance of the homeostatic equilibrium in HO-1 expression in AD, recent studies demonstrated that inhibition of HO-1 up-regulation in microglia reduces inflammation in brain lesions (118). Finally, the detrimental upregulation of HO-1 in cerebral AD patients, compared to age-matched non-dementia individuals, makes this enzyme a possible biomarker and therapeutic target for AD (130).

HO-1 in Parkinson's disease

Parkinson disease (PD) is a neurodegenerative disorder characterized by the progressive loss of dopaminergic neurons in substantia nigra, linked to the chronic movement disorder. Also, neuroinflammation and the presence of Lewy bodies, formed by the accumulation of intracellular aggregation of α -synuclein protein, result dangerous for brain loss of function (131, 132). As for other neurodegenerative diseases, also in PD patients the role of HO-1 has been studied for a long time. Of relevance, HO-1 appears overexpressed both in brain and plasma of Parkinson patients and its upregulation by astrocytes promotes the α -synuclein production, with consequent brain impairment and toxicity (115, 118). Lewy bodies contain a huge amount of iron and the overexpression of HO-1, together with its metabolites, contributes to the increase in its concentration. Augmented free iron is also observed in microglia and substantia nigra of PD patients leading to an impairment of cognitive abilities and neuronal iron-induced cell death, with progression of the disease (133). Consistently, it is observed that the upregulation of Nrf2 triggers with cerebral injury and disease progression, making high the expression of HO-1 (134).

HO-1 in cardiovascular diseases

In spite the decline in mortality observed in recent decades, cardiovascular diseases still remain the main cause of death worldwide (135) and represent a urgent unmeet clinical need. Physiological HO-1 expression levels are usually low in most body areas, but increase in response to pathological stimuli, to

10.3389/fimmu.2024.1433113

buffer ongoing inflammation. This key role of HO-1 in tissue homeostasis has been documented by various studies and, remarkably, its cardioprotective effects emerge through its ability to regulate inflammatory processes and mitochondrial functions (30), ultimately mitigating damage to the cardiovascular system (136). Macrophages represent a functionally plastic (137) homeostatic population present in virtually all tissue, where they maintain proper organ functions, in part by recycling iron, regulating heme acquisition and decomposition (138). Macrophages are claimed as central players in the pathogenesis of various cardiovascular diseases (i.e. atherosclerosis, thrombosis and myocardial infarction), through local production of inflammatory cytokines and factors leading to oxidative stress (139). However, their ability to express HO-1 ensures antioxidant defenses and provides cardioprotective and reparative activities (140). As such, pathways of macrophages differentiation and identification of HO-1-expressing subsets may provide new strategies to control tissue injury and/or repair.

HO-1 shapes the anti-inflammatory phenotype of macrophages in CDVs

HO-1 expression by macrophages critically integrates their M2 polarized phenotype, with specialized anti-inflammatory functions (60). Previous studies have demonstrated that HO-1 null macrophages display increasing levels of ROS, enhanced release of proinflammatory mediators and increased levels of oxLDL scavenger receptor A (SR-A) expression (141). Moreover, in response to bacterial products (i.e. lipopolysaccharide) HO-1 is recruited to the cell membrane caveolae through p38 MAPKdependent mechanism, blocking the activated proinflammatory signaling cascade (142). Moreover, stimulation of macrophages with the anti-inflammatory interleukin-10 (IL-10) activates HO-1 expression in a p38 MAPK-dependent manner (92), while the protective activity of IL-10 in response to LPS stimulation is significantly abrogated after the administration of an HO-1 inhibitor. This evidence corroborates the hypothesis that engagement of HO-1 in response to IL-10 is an integral part of the resolution phase of the inflammatory response (143).

The idea of targeting HO-1 in the myeloid cell compartment appears advantageous because the main functions of the heme-HO-1 system have been highlighted in the myeloid-mononuclear system (7, 144, 145) and because cardiac healing after myocardial ischemia depends on the recruitment and local expansion of myeloid cells, particularly macrophages (146). Macrophages play a key role in the physiological processes of the cardiovascular system, such as cardiac contraction (147) and control of blood pressure (148), as well as in pathology, including clearance of necrotic tissue, tissue remodeling and self-renewal (149). Consistently, hemin-induced HO-1 expression provides cardioprotective activity, attenuating ischemic-induced cardiomyocytes senescence and preventing myocardial infarction (150). In agreement, mice deficient for Bach1, a negative regulator of the HO-1-inducing transcription factor Nrf2 (151), have dramatically higher HO-1 expression in the heart and display resistance to ischemiareperfusion-induced myocardial injury (61, 152). The role of HO-1 in alternative M2 activation of macrophages was further confirmed in Bone marrow-derived macrophages (BMDMs) from myeloid-specific *Hmox1*-knockout mice, that showed increased expression of proinflammatory M1-related mediators as CXCL10, IL-1 β , and monocyte chemoattractant protein 1 (MCP-1), along with increased M2-related markers (i.e. Arg1 and CD163) (61). The scavenger receptor CD163, in particular, mediates the internalization and clearance of hemoglobin haptoglobin (Hb-Hp) complexes, thus providing a fundamental step of heme metabolism, and is mostly expressed by macrophages (153).

Of relevance, the chemokine ligand CXCL4/PF4 inhibits CD163 in macrophages, compromising their phagocytic function and exacerbating the progression of atherosclerosis (154). However, alternatively activated, hemoglobin-scavenging CD163⁺ macrophages are present within atherosclerotic lesions and were also proposed to promote angiogenesis, vessel permeability, and leucocyte infiltration, via the CD163/HIF1 α /VEGF-A pathway, thereby exacerbating plaque progression (155). Of note, CD163 exists also as soluble form of CD163 (sCD163), whose levels are induced by Inflammatory factors, including LPS, through activation of tumor necrosis factor- α -converting enzyme (TACE/ADAM) metallopeptidase (156). Importantly, sCD163 levels were recently associated with established CVD risk factors and with carotid intima media thickness (157).

HO-1 in ischemic conditions

The heart is enriched in the number of mitochondria due to its high metabolic demand, making it particularly sensitive to oxidative stress (158). Dysfunctional coronary blood flow following a heart attack usually leads to a hypoxic condition that promotes greater release of free radicals and causes ischemic heart disease, that may activate a persistent inflammatory condition fueling oxidative damage and the development of heart failure (159). In response to oxidative stress, an activated transcription factors signatures (i.e. the nuclear factor erythroid 2-related factor 2/Nrf2, NF-KB, AP-1 and Bach1) leads to HO-1 upregulation (160), likely involved in the defense against pathophysiological stress (161). In particular, Nrf2 binds to the Maf proteins and activates the antioxidant response element (ARE), sustaining and triggering the expression of HO-1 (162). Nrf2 has been appointed as one of the key modulators of related genes involved in the antioxidative process, as glutathione Stransferase (GST), γ -glutamyl cysteine synthetase (γ GCS) and HO-1 (163). During ischemic heart injury, neutrophils are rapidly recruited to the injured myocardium to initiate the inflammatory phase. The transition to resolution is promoted by macrophage efferocytosis of dying neutrophils, characterized by a time and context-dependent phenotypic M2 polarized switch, toward repair and resolution; Treg release of anti-inflammatory signaling also facilitates resolution (164). Importantly, mice with cardiac-restricted HO-1 overexpression resisted ischemia/reperfusion (I/R) injury, with improved contractibility and reduction of infarct size, oxidative damage, inflammatory cell infiltration and apoptosis (150).

Metabolites resulting from heme degradation, bilirubin and biliverdin (165), are reducing species and powerful antioxidants, mostly produced in the spleen following hemoglobin degradation by HO-1 (113). Coherently, low levels of BR correlate to cardiovascular risk, such as ischemic heart disease and hypertension (166).

HO-1 in myocardial infarction

Myocardial infarction is the most acute manifestation of ischemic heart disease and one of the most life-threatening cardiovascular emergences (167). Risk factors also include arterial hypertension, smoking, dyslipidemia, diabetes mellitus, inactivity and early development of atherosclerosis, while elevation of myocardial infraction is found to be one of the cardiac emergency encountered with COVID-19 pandemic (168). ROS are involved in pathological myocardial dysfunction and their elimination through the antioxidant activity of HO-1 can be considered a potential therapeutic approach. Various cell types contribute to the triggering of sterile inflammation during tissue injury and myeloid cells, particularly resident cardiac macrophages of the myocardium, are involved in the regulation of primary innate immune response, stimulated by the activation of PRRs by Damage-associated molecular patterns (DAMPs). This release activates transcriptional factors, such as IRF, NF-KB and AP-1, causing the production of proinflammatory molecular mediators in the damaged heart (169). Suppression of these transcriptional pathways changes the macrophage phenotype favoring an anti-inflammatory M2-like polarization state, expressing pro-angiogenic properties, reducing pathological cardiac remodeling and preserving the ventricular rupture.

During MI, the recruitment of neutrophils and macrophages, enabled by the up-regulation of specific chemotactic signals of the CXC (CXCL2 and CXCL5) and CC (CCL2 and CCL5) chemokine families, guarantees the first line of defense (170, 171). More indepth studies have shown that the healing process occurring after MI involves the early recruitment of inflammatory Ly-6C^{high} and the subsequent accumulation of reparative Ly-6Clow monocytes/ macrophages. Such transition was shown to be controlled by the Nuclear Receptor Subfamily 4 Group A Member 1 (Nr4a1), which crucially modulates both the early Ly-6Chigh monocyte inflammatory and the later Ly-6C^{low} macrophage reparative phases in the infarcted myocardium (172). It has also been observed that the ischemic myocardium accumulates Ly6C^{high} monocytes in greater numbers than their availability in circulation and that their main source is the spleen (173). Of relevance, the post-ischemic influx of Ly6G⁺ neutrophils and Ly6C^{high} monocytes is further exacerbated in Hmox1-deficient hearts, while treatment with recombinant adeno-associated virus (rAAV)-encoding human Hmox-1 reverses such phenotype and attenuates post-ischemic inflammation, in a murine ischemia/ reperfusion model (174). Consistently, HO-1 knock-out mice showed increased tissue dysfunction post-MI, positively correlating with a higher number of circulating Ly6C^{high} monocytes and increased number of proinflammatory macrophages in the damaged cardiac tissue (175).

Strikingly, in neonatal mice, the heart can regenerate fully, without scarring following MI (176). Interestingly, embryonicderived macrophage subsets (MHC-II^{low}CCR2⁻ and MHC- II^{high}CCR2⁻) resident in the adult heart and owning tissue repair properties are lost after adult cardiac injury and are replaced by inflammatory CCR2⁺ monocyte-derived macrophages that interfere with the angiogenic process and the restoration of cardiomyocytes, thus establishing a reduced regenerative potential (177). This evidence appears in line with the observation that proinflammatory monocytes and monocyte-derived macrophages support the reduction of tissue-resident macrophages (176). Of note, after 3 weeks of MI, *Hmox1*-deficient mice express a pool of blood monocytic cells associated with expansion of cardiac macrophages overexpressing the CD11c marker (175), previously shown to release large amounts of inflammatory cytokines (i.e. IL-1 β , TNF- α and IFN γ).

HO-1-expressing macrophages in atherosclerosis

Coronary heart diseases are clinical manifestations of atherosclerosis, a process characterized by the development of atherosclerotic plaques that can lead to obstruction coronary arteries, ischemia of the cardiac tissue and myocardial infarction. Such process is supported by circulation of elevated levels of lowdensity lipoprotein (LDL), undergoing oxidization and accumulating at the level of the intima wall of the vessels. Uptake of oxidized LDL by monocyte-derived macrophages and smooth muscle cells leads to the formation of foam cells which accumulate lipid droplets and crucially contribute to development of atherosclerosis. Infiltrating monocytes are chemoattracted in the inflamed intima, migrating through the vascular wall and generating the primary atherosclerotic lesion, also identified as fatty streak (178).

Activated inflammation at the plaque level releases a cohort of inflammatory mediators that can induce cell death and the development of a central necrotic core, which favors plaque rupture (178). Based on this, emerging therapeutic strategies aim to target and mitigate inflammatory circuits that support plaque development and fragility. Several reports proved the HO-1 expression in atherosclerotic plaques (141), as well as its role in the early stages of atherosclerosis. This is in line with the detrimental role exerted by oxidative stress on the arterial wall in proatherogenic conditions (i.e. diabetes, hypertension or hyperlipidemia) (179). In agreement, it was shown that increased HO-1 activity markedly reduces the chemotactic recruitment of monocytes after exposure to oxidized LDL. It has in fact been verified that the induction of HO-1 by pharmacological or gene transfer approaches may prevent atherogenesis in preclinical hypercholesterolemic murine model (180). In analogy, genetic ablation of HO-1 amplified the development of atherosclerotic lesions and promoted their progression, thus confirming the protective role of HO-1 against atherogenesis (181).

In this scenario, a number of reports suggest that release of heme from intraplaque hemorrhage induces an atheroprotective macrophage phenotype (182–185); indeed, in the complex atherosclerotic plaques' microenvironment, macrophages are simultaneously exposed to a variety of stimuli and, accordingly to

their known functional plasticity, they polarize into multifunctional subsets which collectively cooperate to determine the dynamic evolution of the inflamed plaque, thus influencing atherosclerosis progression (184, 186, 187). In particular, in addition to the continuum of macrophage polarization states described within the M1 and M2 functional extremes (137), plaque-specific macrophage phenotypes, including Mhem and M(Hb) macrophages, related to the presence of hemoglobin and erythrocytes, have been recently identified, which play atheroprotective roles by preventing foam cell formation and lipid accumulation, as they highly express genes associated with reverse cholesterol transport, such as liver X receptor α and β (LXR- α ; LXR- β) and the ATP-binding cassette transporters ABCA1 and ABCG1 (188-190). Due to the increased expression of molecules involved in cholesterol efflux, M(Hb) macrophages are characterized by low levels of lipid accumulation. In particular, M (Hb) macrophages typically express high levels of mannose receptor CD206 and the scavenger receptor for the hemoglobin/haptoglobin (Hb/Hp) complex CD163, participating in the hemoglobin clearance after plaque hemorrhage. These macrophages are described to produce both anti- (IL-10, IL-1Ra) and proinflammatory (VEGF, IL-1β) cytokines, while producing less ROS than other macrophage subtypes and display low iron accumulation because of the upregulation of ferroportin (FPN) (191).

After endocytosis of the Hb/Hp complex and erythrocytes, the released heme group primes intraplaque macrophages polarization toward a Mhem phenotype, with consequent activation of the 5'-AMP-activated protein kinase (AMPK) and the downstream transcription factor 1 (ATF1) (188, 189). Activation of the ATF1/ MAPK signaling pathway therefore leads to the expression of HO-1, LXR- α /LXR- β , and ABCA1/apolipoprotein E (APOE) cascade, preventing foam cell formation. Thus, since iron levels in macrophages may drive cholesterol efflux, targeting macrophages environment manipulating iron levels and/or iron metabolism-related molecules could interfere with the generation of foam cells and the development of atherosclerosis.

Conversely, proatherogenic roles of CD163⁺ macrophages are also reported (155, 192, 193); indeed, in human and mouse atherosclerotic lesions M(Hb) CD163⁺ macrophages are also associated with promotion of angiogenesis, vessel permeability, and leucocyte infiltration through a mechanism involving Hb/Hp/ CD163/HIF1 α -mediated VEGF induction (194). Therefore, the plasticity of these cells makes the scenario more intricate, suggesting that the complete definition of the role of macrophage subpopulations in plaque needs to be further clarified.

HO-1-expressing macrophages in hypertension

Arterial hypertension is associated with an increased risk of cardiovascular disease (CVD) and is responsible for more than 10 million deaths every year (195, 196). While endocrine and renovascular disorders are responsible for secondary hypertension in 5 and 15% of hypertensive patients, primary hypertension refers to patients in which no underlying cause has been found (197).

Despite its clinical relevance, the causes for its occurrence still remain unclear, although the role of angiotensin II (AngII) in its development has been described. In fact, AngII, via its AT₁ receptor, promotes cell growth, inflammation, vasoconstriction, apoptosis, production of extracellular matrix components and the release of ROS (9, 198). Specifically, the increase in the renin-angiotensin system and sympathetic activity contributes to the macrophage mobilization and to its polarization towards the pro-inflammatory phenotype (199). A low degree of inflammation facilitates vascular oxidative stress, leading to the vascular alterations accounting for increased peripheral vascular resistance (200, 201). The regulatory role of heme availability for the synthesis of enzymes such as cyclooxygenase or nitric oxide synthase, both involved in hypertension development, seems to be responsible for many of the beneficial effects of HO-1. Oxidative activity, in fact, activates cyclooxygenases (COX), with the consequent production of prostaglandins and thromboxanes, both contributing to vascular alterations and enhances inflammatory responses (202). HO-1 has been shown to protect against oxidative and inflammatory insults in hypertension, reducing organ damage and blood pressure, not only by its expression at the vascular level, but also by shifting macrophages toward the anti-inflammatory phenotype (203, 204). Moreover, the antioxidant, anti-inflammatory, antiapoptotic, and antiproliferative effects of the end products of HO-1 activity would also provide protective functions (199). In particular, bilirubin (BR) is one of the most powerful plasma scavenger of ROS and RNS, inhibiting lipid peroxidation and peroxynitrite-mediated oxidations, protecting against H2O2 toxicity and increasing NO half-life (205-207). Coherently, an inverse relationship between plasma BR levels and systolic blood pressure was reported (208, 209). Consequently, it was suggested that BR might prevent oxidant-induced microvascular leukocyte adhesion and attenuate endothelial cell dysfunctions (210), thus limiting inflammationdriven hypertension (211, 212). Overall, it appears that pharmacological modulation of oxidative stress and inflammatory macrophages could represent a viable therapy for the control of hypertension. The HO-1 product CO is also a potent modulator of cellular signaling molecules (i.e. p38 MAPK, ERK1/2, JNK, Akt, NF- κ B), which targets mitochondrial activity affecting energy balance and cellular functions (213). Abnormalities of the heme oxygenase/carbon monoxide system have been critically linked to vascular contractility, increased oxidative stress, and imbalanced cellular apoptosis and proliferation in the vascular wall (214). In agreement, Broard et al. have also demonstrated that CO negatively regulates endothelial cell apoptosis, triggering the activation of p38 mitogen-activated protein kinase (MAPK -p38) (215), while pharmacological induction of HO-1 activity decreases blood pressure in spontaneously hypertensive rats (214).

HO-1 in infections

The role of Hmox-1 and its degradation products is known to have strong implications on the ability to control infections and mitigate their pathological consequences (216). Its relevance in infections is further highlighted by the worrying growth of antibiotic-resistant bacterial strains and the continuous appearance of new viruses. In the present section, we discuss emerging evidence on the crucial role of HO-1 in infectious diseases caused by bacteria and viruses.

Bacterial infections

Mycobacterium tuberculosis (Mtb). HO-1 is expressed in lung tissues and is activated by stress signals, including ROS and inflammatory mediators (217, 218). Its contribution in modulating the host response to Mycobacterium tuberculosis (Mtb) infection and controlling disease progression is still highly controversial. It has been previously reported that Mtb infections can induce HO-1 expression in macrophages in vitro and in lung tissue in vivo (219). Its activity in macrophages and lungs is associated with increased heme degradation which promotes CO production. The release of this gaseous molecule, acting as regulatory ligand the heme two-component sensor kinases DosS and DosT, induces the complete Mtb dormancy (Dos) regulon (220, 221), thus promoting the bacilli quiescence and the establishment of a latent infection. A published study by Costa et al. supports the role of HO-1 in Mtb survival and replication in the host. The authors observed that SnPPIX-mediated inhibition of HO-1 significantly reduced lung bacterial load, with comparable to the effects achieved with conventional antibiotic therapy. Furthermore, the combination of SnPPIX and antibiotic therapy promoted faster resolution of Mtb infection (222). More recently, the same group found that SnPPIX treatment causes a reduction in intracellular iron availability in activated macrophages, along with an increase in IFNy-induced NOS2 expression and subsequent NO production, resulting in more effective control of bacterial replication. The authors proposed that HO-1 promotes bacterial survival and proliferation in host cells, through an iron-dependent intracellular mechanism (223). However, contrasting evidence by Chinta et al. indicates that expression of HO-1 in the lungs of patients with Tuberculosis is associated with lower ROS and RNS production. The authors reported in Hmox1-deficient mice that lack of HO-1 expression in the damaged lung regions is positively related to an enhanced ROS and RNS production by neutrophils and macrophages, as well as with increased susceptibility to Mtb infections (224).

It should be noted that in the study proposing HO-1 as a promoter of Mtb infections, HO-1 activity was hindered by pharmacological inhibitors, whereas Chinta et al. conducted their experiments on *Hmox1*-deficient mice. These different approaches used may make it more difficult to interpret the mechanisms underlying MtB infection, since HO-1 deficiency was shown to cause important hematopoietic abnormalities (225). However, the protective action of HO-1 has been reported using another Mycobacterium species (i.e. *Mycobacterium avium*), in which mice genetically deficient in HO-1 were found to be more susceptible to infection. In particular, HO-1 deficiency resulted in an accumulation of heme which exerts a cytotoxic effect in infected macrophages, increasing their necrotic death and the consequent release of proinflammatory cytokines (226). More recently, the importance of the NRF2/HO-1 pathway in determining the outcome of M. avium infection has been demonstrated, thanks to its role in inducing granuloma formation in infected tissue, to counteract bacterial replication (227). Together, these findings might suggest a stage-dependent effect of HO-1, with a cytoprotective role in the initial stages of Mycobacterium infection, which is gradually lost during disease progression, due to massive heme release and subsequent ROS production and oxidative stress (228).

Salmonella thyphimurium

Infection with S. Typhimurium causes severe gastroenteritis, whose associated inflammatory response is essential for this pathogen to colonize the intestinal tract (229). The intestinal inflammation induced by the infection causes an important dysbiosis, associated with disruption of the colonization barrier, with symptoms that typically include weakness, prostration, fever, and diarrhea, which become particularly problematic in immunocompromised individuals (230). Similarly, to Mycobacterium infection, investigation of HO-1 role in infections with Salmonella highlighted its dual role in the outcome of the disease. A great number of studies unravel the protective role of HO-1 in the infections with this pathogen. Particularly, in an in vivo murine model of salmonellosis, HO-1 inhibition with ZnPPIX resulted in an increased apoptosis of liver cells. The mechanistic association of the defective action of HO-1 with a higher susceptibility to S. thyphimurium infection, was validated in vitro, where HO-1 pharmacological inhibition led to a reduced bacterial killing capacity of macrophages (231). Accordingly, treatment with CoPPIX, an inducer of HO-1, in mice infected with S. thyphimurum, may afford protection against the enterocolitis caused by the pathogen (102). In contrast with the above studies, other studies have observed that the ZnPPIX pharmacological inhibition of HO-1 in RAW 264.7 cells, is related to an increased ROS and RNS production that counteract S.thyphimurum survival upon infection, suggesting a detrimental HO-1 effect on disease outcome (232).

Listeria monocytogenes

The Gram-positive bacterium *L. monocytogenes* is borne through contaminated food and infection with this pathogen generally results into mild gastroenteritis. Nonetheless, in elderly or immunocompromised individuals L. monocytogenes infection can lead to severe illnesses, including severe sepsis, meningitis, or encephalitis, that at times may eventually lead to death (233). The literature reports studies both in favor of the protective effect of HO-1 against *L. monocytogenes*, as well as in favor of its infectionpromoting action. That said, in response to poly I:C treatment, mice with myeloid-specific deletion of HO-1 infected with L. monocytogenes showed lower levels of IFN γ in the blood, along with improved survival (234). Indeed, myeloid-specific HO-1 is required for the activation of IFN regulatory factor (IRF) 3 and for the production of IFN- β , with the latter increasing the severity of *L. monocytogenes* infection (235, 236). Wang and colleagues observed that L. monocytogenes infection elicits the Tim-3 receptor signaling pathway in macrophages, which inhibits the NRF2 nuclear translocation causing the sequential decrease of HO-1 expression. This mechanism results in an increased susceptibility to *L. monocytogenes in vivo* and *in vitro*, indicating that HO-1 is instead required to limit the bacterial replication during the infection (237). This puzzling scenario of interpretations on the role of HO-1 clearly indicates that we still need to fully understand how to regulate its activity during the different phases of bacterial growth, into the different infectious contexts.

Viral infections

Recent studies demonstrate that HO-1 has significant antiviral activity against a wide variety of viruses (e.g., influenza virus, respiratory syncytial virus, HIV, hepatitis C virus, hepatitis B virus, enterovirus 71, dengue and Ebola virus) and that this activity may occur through the heme degradation products biliverdin and carbon monoxide, able to respectively inhibit the function of viral proteases and reactive oxygen species production (238). HO-1 indirectly also promotes an antiviral state by inducing activity of IRF3 and the activation of type I IFN antiviral functions (239).

Human Immunodeficiency Virus (HIV). Human immunodeficiency virus (HIV). The human immunodeficiency virus (HIV) is the causative agent of acquired immunodeficiency syndrome (AIDS). The available antiretroviral therapies represent the first line of treatment for AIDS, contributing significantly to reducing the number of deaths (240), but to date it appears unlikely that they will exterminate the virus. In this context, several reports dissected the role of HO-1 in HIV infection, associating its activity with the suppression of the virus replication. HO-1 induction by Hemin treatment significantly suppressed infection and viral replication of both monocytes and T cells inoculated with R5, X4, R5X4 tropic viral strains. Such effect was consistent with the inhibition of Tat-dependent activation of long terminal repeat (LTR) viral promoter. Consistently, the HO-1 inhibitor tin-protoporphyrin-IX (SnPPIX) hampered the macrophage resilience allowing the progression of the infection, indicating that the antiviral protective mechanisms in monocytes are mediated by HO-1 activity (241). Along with this, it was reported that bilirubin hinders the protease activity of HIV, thus affecting virus replication (242). Altogether, the above studies suggest the protective role of HO-1 and heme catabolism against HIV infection (242).



Distinct roles of myeloid HO-1⁺ cells in pathology. Multifaced functions of HO-1⁺ myeloid cells during the onset and progression of several diseases, depending on the pathological contest. MI, myocardial infarction; MS, multiple sclerosis; PD, Parkinson's Disease; AD, Alzheimer Disease; RA, rheumatoid arthritis; Lysteria M, *Lysteria Monocytogenes*; Salmonella T, *Salmonella Thyphimurium*; IBD, Inflammatory Bowel Diseases; LD, Lung Diseases.

Severe acute respiratory syndrome coronavirus 2

SARS-CoV-2 is the causative agent of COVID-19, that can progress to severe pneumonia with acute respiratory distress syndrome (ARDS) (243). The recent studies of the HO-1 role in the onset of the disease and its progression are still obscure. According to few recent studies, treatments with hemin can attenuate cytokine storms in animal model of sepsis, indicating the putative role of HO-1 in the protection against the cytokine storm syndrome observed in COVID-19 patients (244). Moreover, using a renal cell line (Vero76) infected with SARS-CoV-2 and treated with hemin, it was observed a reduced virus replication, indicating the anti-viral role of HO-1 (245). Singh et al. proposed HO-1 as a protective molecule in the early and late stages of SARS-CoV-2 infections. In fact, the induced expression of HO-1, promotes the expression of type I interferon, which coordinates the anti-inflammatory anti-viral host responses and therefore might protect against the damage from exacerbated oxidative stress (246). However, the role of HO-1 in determining the severity of the infection remains partly to be clarified, as recent the stratification of patients into survivors and non-survivors showed a significant increase of blood HO-1 mRNA levels in the later (247).

HO-1 can be released into plasma by leukocytes, macrophages, smooth muscle cells, and endothelial cells activated by oxidative stress or inflammation (248). Grigorov et al. suggested that serum HO-1 concentrations upon hospital admission of COVID-19 patients could be a useful biomarker for clinical management, proposing that the increase of HO-1 in the early stage of the disease could be beneficial, as it would provide protection against oxidative stress and inflammation (249). However, increased HO-1 expression is not necessarily linked to increased heme catabolism and hemolysis, but could rather be a possible consequence of the inflammatory response as, paralleling the upregulation of blood HO-1, was also reported a significant increment of serum ferritin levels (249), an acute phase

TABLE 1 List of abbreviation and acronymous used in the manuscript.

reactant that can induce anti-inflammatory response by reducing the damage caused by free radicals (250, 251).

Conclusions

Accumulating evidence point to the multifaced functions of the inducible HO-1 form as critical determinants of disease development since, depending on the pathological contest, its anti-inflammatory and antioxidant properties may elicit beneficial or detrimental effects. While a general protective role of HO-1 has being highlighted in autoimmune, immune-mediated and cardiovascular diseases, development of different infection and cancers benefit from its enzymatic activity (Figure 2). Several HO-1 agonists and antagonists have been used to investigate its physiological and pathological roles and to develop new potential therapeutic strategies (Table 2). New studies now aim to define the transcriptional mechanisms and differentiation and maturation pathways that regulate HO-1⁺ myeloid populations in pathology. Transcriptional (i.e. Nrf2, Maf, AP-1, p50 NF-kB) (7, 252) and epigenetic events (83), as well as microenvironmental conditions (i.e. hypoxia) (253) finely tune *Hmox-1* gene expression. Above this, new emergency pathways of hematopoiesis display the capacity to promote expansion of circulating monocytes and macrophages endowed with HO-1 activity (7) (Figure 3). Understanding the interconnection between local (microenviroment) and remote (emergency myelopoiesis) mechanisms of HO-1 regulation could improve our ability to therapeutically target its functions in disease, also considering its dynamic and stage-dependent activity observed in distinct pathological contexts. Furthermore, making the biology of HO-1 even more intricate, a body of evidence indicates that the anti-inflammatory role of HO-1 predominates in the early stages of the experimental inflammatory diseases. Indeed, hemin-like compounds or adoptive transfer of hemin-activated macrophages reduced caerulein-induced pancreatitis, serum amylase and lipase,

| Abbreviation | Definition | Abbreviation | Definition |
|--------------|---------------------------------|--------------|-----------------------------------|
| АМРК | 5'-AMP-activated protein kinase | LL2 | Lewis lung carcinoma |
| AD | Alzheimer's disease | LXR | liver X receptor |
| ALS | amyotrophic lateral sclerosis | M(Hb) | hemoglobin-stimulated macrophages |
| AngII | angiotensin II | МАРК | mitogen-activated protein kinase |
| AP-1 | activator protein-1 | MCP-1 | chemoattractant protein 1 |
| APOE | apolipoprotein E | Mhem | macrophages heme |
| Argl | arginase 1 | MI | myocardial infarction |
| ATF1 | transcription factor 1 | MN/MCA1 | fibrosarcoma |
| B16/F10 | melanoma | MS | Multiple Sclerosis |
| BMDMs | Bone marrow-derived macrophages | Mtb | Mycobacterium tuberculosis |

(Continued)

TABLE 1 Continued

| Abbreviation | Definition | Abbreviation | Definition | |
|--------------|---|--------------|---|--|
| BR | bilirubin | NF-κB | nuclear factor kappa-light-chain-enhancer of activated B cells | |
| BV | biliverdin | NK | natural killer | |
| CD163 | hemoglobin/haptoglobin scavenger receptor | NO | nitric oxide | |
| CNS | central nervous system | Nr4a1 | nuclear receptor subfamily 4 group A member 1 | |
| СО | carbon monoxide | Nrf2 | NF-E2-related factor 2 | |
| СОРР | cobalt protoporphyrin IX | OVA | ovalbumin | |
| CORMs | carbon-monoxide releasing molecules | PBMCs | peripheral mononuclear cells | |
| CSF-1 | colony stimulating factor 1 | PD | Parkinson's disease | |
| DAMP | Damage-associated molecular patterns | PD-1 | programmed cell death protein 1 | |
| DCs | dendritic cells | PD-L1 | programmed death-ligand 1 | |
| EAE | experimental autoimmune encephalomyelitis | PDAC | pancreatic ductal adenocarcinoma | |
| EMT | epithelial-mesenchymal transition | PI3K | phosphatidylinositol 3-kinase | |
| FPN | ferroportin | RA | rheumatoid arthritis | |
| GBM | glioblastoma | RNS | reactive nitrogen species | |
| GST | glutathione S-transferase | ROS | reactive oxygen species | |
| Hb-Hp | hemoglobin haptoglobin | SARS-CoV-2 | severe acute respiratory syndrome coronavirus 2 | |
| HD | Huntington's disease | SCD | sickle cell disease | |
| HIF-1α | hypoxia inducible factor | SLE | systemic lupus erythematosus | |
| HIV | Human Immunodeficiency Virus | SnMPIX | tin mesoporphyrin IX | |
| Hmox-1 | heme oxygenase 1 (gene) | SnPPIX | tin protophorphyrin IX | |
| Hmox-2 | heme oxygenase 2 (gene) | TAMs | tumor associated macrophages | |
| HO-1 | heme oxygenase 1 (protein) | TME | tumor microenvironment | |
| HO-2 | heme oxygenase 2 (protein) | TNF | tumor necrosis factor | |
| IFN | interferon | TRMs | tissue-resident macrophages | |
| IL | interleukin | VEGF | vascular endothelial ggrowth factor | |
| IRF | IFN regulatory factor | ZnPPIX | zinc protoporphyrin IX | |
| LDL | low-density lipoprotein | | | |

decreased pancreatic trypsin generation, and protected from lung injury (254), while activation of the NRF2/HO-1 pathway, through administration of Cobalt protoporphyrin IX (CoPP), induced macrophage differentiation toward a Marco^{hi}Hmox1^{hi} antiinflammatory erythrophagocytic phenotype, contributing to an overall decreased inflammatory profile in a murine model of colitis (255). However, the anti-inflammatory action of HO-1inducing compounds observed in these models was evidenced mainly during the onset phase of the inflammatory response, while it was found to be lacking when induced after disease onset.

As discussed above, HO-1 deficiency causes important hematopoietic abnormalities (225) and has an impact on the regulation of innate and adaptive immune responses. Consistently, genetic alteration of HO-1 gene is associated with important hematopoietic and organ dysfunctions. Sickle cell disease (SCD) is an autosomal recessive pathology generated by a genetic point mutation at the level of β -globin locus. The effects generated by this mutation results in an abnormal form of hemoglobin, characterized by red blood cell membrane rigidity and consequently hemolysis (256). Among the pathological complications, SCD bearing individuals shown more susceptibility to precise bacterial infections, including Salmonella, Streptococcus Pneumoniae and Hemophilus Influenzae, resulting in a potent risk factor for invasive bacterial infections, where a splenic dysfunction has been considered the main leading cause (257). Further, tissue iron overload and anemia were previously reported in a human patient and mice lacking HO-1 (225). It was found that resident splenic and liver macrophages were mostly absent in HO-1-



FIGURE 3

Signaling pathways leading to myeloid HO-1 induction in pathology. The scheme shows potential interrelationships between signaling events promoting transcriptional activation of HO-1 in myeloid cells in different pathophysiological conditions, such as tumors, cardiovascular diseases (ischemic conditions, MI, atherosclerosis, and hypertension) neurodegenerative disorders (AD, PD), immune-mediated diseases (SLE, MS and RA) bacterial and viral infections (*Salmonella thyphimurium, Listeria monocytogenes, HIV* and *SARS-CoV-2*). Several noxious stimuli (i.e. ROS, hypoxia, oxidative stress, hemorrhage/heme, free iron) and inflammatory mediators (i.e. IL-6, VEGF, CSF-1, C3a, EGF, HIF-1a) as well as hemoglobin/ haptoglobin through CD163 receptor are known inducers of transcription of HO-1, primarily promoting KEAP-1/Nrf2 nuclear translocation and nuclear export of the transcriptional repressor BACH1. Activation of MAPK/PI3K/AKT, AP-1 and NF-κB by environmental stresses and cytokines has also been implicated in HO-1 activation. HO-1, heme oxygenase-1; MI, myocardial infarction, AD, Alzheimer's disease; PD, Parkinson's disease; SLE, systemic lupus erythematosus; RA, rheumatoid arthritis; MS, multiple sclerosis; CSF-1, colony stimulating factor 1; MAPK, mitogen-activated protein kinase; Nrf2, NF-E2 related factor-2; HIF-1a, hypoxia inducible factor; AP-1, activator protein-1; BACH1, HO-1 transcriptional repressor; ROS, reactive oxygen species; PI3K, phosphatidylinositol 3-kinase. Created by biorender.com.

TABLE 2 HO-1's agonists and antagonists.

| Disease | HO-1 agonist or antagonist | | Mechanism | Impact on disease/therapy |
|-------------------|-------------------------------|--------------------------------------|---|---|
| | LL2 and PDAC | Antagonist (SnMP) (49, 53) | Suppression of immunosuppressive role of TAMs | Decreased tumor growth and progression |
| | 4T1 | Antagonist (SnMPIX) (43, 63) | Abrogation of TAMs pro- angiogenic activity | Reduced metastatic spread |
| Cancer | MN/MCA1;B16/F10 | Antagonist (ZnPPIX) (7) | Reduced immunosuppressive myelopoiesis | Increased activity of anti-PD- 1 immunotherapy |
| | MMTV-PyMT | Antagonist (SnMPIX) (22) | Improved cytotoxicity of CD8 + T cells | Increased antitumor activity of 5-fluorouracil |
| | GBM | Antagonist (ZnPPIX;OB24) (46, 73) | Reduced levels of PD-L1/PD- 1 and IL-10 | Recovered of pro- inflammatory, antitumor TME |
| Neurodegenerative | Multiple Sclerosis | Agonist (122, 125) | ROS damage | Increased levels of astrocytes and oligodendrocytes |
| ⊶≫ disorders | Alzheimer's disease | Agonist (111, 127) | Promotion of correct tissue redox milieu | |

(Continued)

TABLE 2 Continued

| Disease | HO-1 agonist or antagonist | | Mechanism | Impact on disease/therapy |
|----------------------|-------------------------------|----------------------------------|--|--|
| | | | | Reduced severe state of both inflammation and tissue damage |
| | Parkinson's disease | Antagonist (114, 117, 133) | Reduced iron content in Lewy bodies | Mitigation of neuroinflammation, cerebral injury and α-synuclein protein sedimentation |
| | Ischemia | Agonist (112, 160, 165) | ROS damage | Tissue Healing |
| 🎽 🔭 🖒 Cardiovascular | Myocardial infarction | Agonist (174, 175) | Angiogenesi | Cardiac remodeling and preserving the ventricular rupture |
| diseases | Atherosclerosis | Agonist (181–184, 193) | Reduced Foam cells accumulation | Absence of atherosclerotic plaques formation |
| | Hypertension | Agonist (207–213) | Reduction of inflammation- driven hypertension | Decreases blood pressure |
| | Mycobacterium | Antagonist (SnPPIX) (220–222) | Reduced intracellular iron availability in macrophages | Inhibition of bacterial survival and proliferation |
| | Salmonella thyphimurium | Agonist (224) | Reduced necriotic death of macrophages | Cytoprotective role in the initial stages of infection |
| | Listeria monocytogenes | Agonist (CoPPIX) (101) | Improved bacterial killing by macrophages | Decreased susceptibility to bacterial infection |
| Wiral and Bacterial | HIV | Antagonist (ZnPPIX) (230) | Increased ROS and RNS production | Reduced bacterial survival |
| infections | SARS-CoV-2 | Agonist (235) | Promotion of IFN I | Reduced bacterial proliferation |
| | | Agonist (235) (Hemin; CoPPIX) | Inhibition of viral proteases and reactive oxygen species production | Inhibition of Viral replication |
| | | Agonist (244) (Hemin; CoPPIX) | Promotion of IFN I | Repression of exacerbated oxidative stress and promotion of anti- inflammatory response |
| | Lupus erythematosus | Agonist (252) | Polarization of macrophages towards an M2-like profile | Reduced T cell activation and increased Treg differentiation |
| Immune- | Rheumatoid arthritis | Agonist (91) | Limited inflammation and toxicity | Decreased disease progression |
| mediated diseases | IBD | Agonist (99) | Reduction of Th17 cell number; increased Tregs | Reduced inflammation |
| | Lung diseases | Agonist (107–110) | Activation of IFN I response | Resolution of airways inflammation |

Their effects are reported in the different pathological contexts, including Cancer, Neurodegenerative disorders, Cardiovascular diseases, Viral and bacterial infections, and Immune-mediated disorders. The table includes studies and experimental models cited in the manuscript. CoPP, cobalt protoporphyrin IX; ROS, Reactive oxygen species; IFN, interferon; BMDM, bone-marrow derived macrophages; SnMP, tin mesoporphyrin; ZnPPIX, zinc protoporphyrin IX; 4T1, model of breast adenocarcinoma; LL2, Lewis lung carcinoma; PDAC, pancreatic ductal adenocarcinoma; MN/MCA1, fibrosarcoma; B16/F10, melanoma; MMTV-PyMT spontaneous mouse model of breast cancer; GBM, glioblastoma.

deficient conditions and that $HO-1^{-/-}$ macrophages died of exposure to heme released on erythrophagocytosis. The release of heme, following the rupture of $HO-1^{-/-}$ macrophages, caused further tissue inflammation, with initial splenic enlargement progressing to red pulp fibrosis, atrophy, and functional hyposplenism, that recapitulated the asplenia of an HO-1- deficient patient. This genetic evidence further highlights the key

role of HO-1-expressing erythrophagocytes in tissue homeostasis and iron redistribution.

It appears clear that the therapeutic potential of HO-1 in mediating the resolution of inflammation and tissue homeostasis may reside in specific phases of the inflammatory cascade and the regulation of hematopoietic output, thus calling for further study to fully characterize its therapeutic potential in different pathological contexts.

Author contributions

FMC: Funding acquisition, Writing – original draft. MI: Writing – original draft. MB: Writing – review and editing. GB: Writing – original draft. VG: Writing – original draft. AS: Writing – original draft, Supervision, Funding acquisition.

Funding

The author(s) declare that financial support was received for the research, authorship, and/or publication of this article. This work was supported by Associazione Italiana per la Ricerca sul Cancro (AIRC) IG (Nos. 29348); AIRC 5x1000 no. 22757; Ministero Università Ricerca PRIN 2022 project n° 20227YR8AW). Ministero Università Ricerca (PRIN 2022 project n° 2022T3ZALK), Fondazione "Aiace", Bologna, Associazione "Medicine Rocks", Milano and "Augusto per la vita".

References

1. Sica A, Lazzeri M. Specialization determines outcomes in inflammation and cancer. *Nat Immunol.* (2023) 24:1399–401. doi: 10.1038/s41590-023-01592-0

2. Mantovani A, Allavena P, Marchesi F, Garlanda C. Macrophages as tools and targets in cancer therapy. *Nat Rev Drug Discovery*. (2022) 21:799–820. doi: 10.1038/s41573-022-00520-5

3. Lazarov T, Juarez-Carreño S, Cox N, Geissmann F. Physiology and diseases of tissueresident macrophages. *Nature*. (2023) 618:698–707. doi: 10.1038/s41586-023-06002-x

4. Ueha S, Shand FHW, Matsushima K. Myeloid cell population dynamics in healthy and tumor-bearing mice. *Int Immunopharmacol.* (2011) 11:783–8. doi: 10.1016/j.intimp.2011.03.003

5. Strauss L, Sangaletti S, Consonni FM, Szebeni G, Morlacchi S, Totaro MG, et al. RORC1 regulates tumor-promoting "Emergency" Granulo-monocytopoiesis. *Cancer Cell.* (2015) 28:253–69. doi: 10.1016/j.ccell.2015.07.006

6. Caiado F, Pietras EM, Manz MG. Inflammation as a regulator of hematopoietic stem cell function in disease, aging, and clonal selection. J Exp Med. (2021) 218. doi: 10.1084/jem.20201541

7. Consonni FM, Bleve A, Totaro MG, Storto M, Kunderfranco P, Termanini A, et al. Heme catabolism by tumor-associated macrophages controls metastasis formation. *Nat Immunol.* (2021) 22:595–606. doi: 10.1038/s41590-021-00921-5

8. Zhao H, Wong RJ, Nguyen X, Kalish F, Mizobuchi M, Vreman HJ, et al. Expression and regulation of heme oxygenase isozymes in the developing mouse cortex. *Pediatr Res.* (2006) 60:518–23. doi: 10.1203/01.PDR.0000242374.21415.f5

9. Kim YM, Pae HO, Park JE, Lee YC, Woo JM, Kim NH, et al. Heme oxygenase in the regulation of vascular biology: from molecular mechanisms to therapeutic opportunities. *Antioxidants Redox Signaling*. (2011) 14:137–67. doi: 10.1089/ARS.2010.3153

10. Ayer A, Zarjou A, Agarwa A, Stocker R. Heme oxygenases in cardiovascular health and disease. *Physiol Rev.* (2016) 96:1449–508. doi: 10.1152/physrev.00003.2016

11. Ponka P. Tissue-specific regulation of iron metabolism and heme synthesis: Distinct control mechanisms in erythroid cells. *Blood.* (1997) 89:1–25. doi: 10.1182/blood.v89.1.1

12. Ryter SW, Tyrrell RM. The heme synthesis and degradation pathways: Role in oxidant sensitivityHeme oxygenase has both pro- and antioxidant properties. *Free Radical Biol Med.* (2000) 28:289–309. doi: 10.1016/S0891-5849(99)00223-3

13. Ajioka RS, Phillips JD, Kushner JP. Biosynthesis of heme in mammals. *Biochim Biophys Acta*. (2006) 1763:723–36. doi: 10.1016/J.BBAMCR.2006.05.005

14. Balwani M, Desnick RJ. The porphyrias: Advances in diagnosis and treatment. Blood. (2012) 120:4496–504. doi: 10.1182/blood-2012-05-423186

15. Dunaway LS, Loeb SA, Petrillo S, Tolosano E, Isakson BE. Heme metabolism in nonerythroid cells. J Biol Chem. (2024) 300. doi: 10.1016/j.jbc.2024.107132

16. Tenhunen R, Marver HS, Schmid R. The enzymatic conversion of heme to bilirubin by microsomal heme oxygenase. *Proc Natl Acad Sci.* (1968) 61:748–55. doi: 10.1073/pnas.61.2.748

17. Maines MD, Trakshel GM. Purification and characterization of human biliverdin reductase. *Arch Biochem Biophysics*. (1993) 300:320-6. doi: 10.1006/ abbi.1993.1044

Conflict of interest

Author MB was employed by the company Navita S.r.l., while authors AS, FC and VG were consultants of Navita S.r.l.

These authors declare that the research was conducted in full independence.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could beconstrued as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

18. Campbell NK, Fitzgerald HK, Dunne A. Regulation of inflammation by the antioxidant haem oxygenase 1. Nat Rev Immunol. (2021) 21:411–25. doi: 10.1038/s41577-020-00491-x

19. Gozzelino R, Jeney V, Soares MP. Mechanisms of cell protection by heme Oxygenase-1. *Annu Rev Pharmacol Toxicol.* (2010) 50:323–54. doi: 10.1146/annurev.pharmtox.010909.105600

20. Otterbein LE, Bach FH, Alam J, Soares M, Lu HT, Wysk M, et al. Carbon monoxide has anti-inflammatory effects involving the mitogen- activated protein kinase pathway. *Nat Med.* (2000) 6:422–8. doi: 10.1038/74680

21. Ye W, Liu Z, Liu F, Luo C. Heme oxygenase-1 predicts risk stratification and immunotherapy efficacy in lower grade gliomas. *Front Cell Dev Biol.* (2021) 9:760800. doi: 10.3389/fcell.2021.760800

22. Muliaditan T, Opzoomer JW, Caron J, Okesola M, Kosti P, Lall S, et al. Repurposing tin mesoporphyrin as an immune checkpoint inhibitor shows therapeutic efficacy in preclinical models of cancer. *Clin Cancer Res.* (2018) 24 (7):1617–28. doi: 10.1158/1078-0432.CCR-17-2587

23. Nitti M, Ivaldo C, Traverso N, Furfaro AL, Ryter SW. Clinical significance of heme oxygenase 1 in tumor progression. *Antioxidants*. (2021) 10(5). doi: 10.3390/antiox10050789

24. Wang H, Cheng Q, Bao L, Li M, Chang K, Yi X, et al. Cytoprotective role of heme oxygenase-1 in cancer chemoresistance: focus on antioxidant, antiapoptotic, and proautophagy properties. *Antioxidants.* (2023) 12. doi: 10.3390/antiox12061217

25. Nitti M, Piras S, Marinari UM, Moretta L, Pronzato MA, Furfaro AL. HO-1 induction in cancer progression: A matter of cell adaptation. *Antioxidants*. (2017) 6. doi: 10.3390/antiox6020029

26. Khojandi N, Kuehm LM, Piening A, Donlin MJ, Hsueh EC, Schwartz TL, et al. Oxidized lipoproteins promote resistance to cancer immunotherapy independent of patient obesity. *Cancer Immun Res.* (2021) 9(2):214–26. doi: 10.1158/2326-6066.CIR-20-0358

27. Tauber S, Jais A, Jeitler M, Haider S, Husa J, Lindroos J, et al. Transcriptome analysis of human cancer reveals a functional role of Heme Oxygenase-1 in tumor cell adhesion. *Mol Cancer*. (2010) 9. doi: 10.1186/1476-4598-9-200

28. Chen B, Zhang L, Zhou H, Ye W, Luo C, Yang L, et al. HMOX1 promotes lung adenocarcinoma metastasis by affecting macrophages and mitochondrion complexes. *Front Oncol.* (2022) 12:978006. doi: 10.3389/fonc.2022.978006

29. Brown RAM, Richardson KL, Kabir TD, Trinder D, Ganss R, Leedman PJ. Altered iron metabolism and impact in cancer biology, metastasis, and immunology. *Front Oncol.* (2020) 10:476. doi: 10.3389/fonc.2020.00476

30. Jais A, Einwallner E, Sharif O, Gossens K, Lu TTH, Soyal SM, et al. Heme oxygenase-1 drives metaflammation and insulin resistance in mouse and man. *Cell.* (2014) 158:25–40. doi: 10.1016/j.cell.2014.04.043

31. Zhang L, Song FF, Huang YB, Zheng H, Song FJ, Chen KX. Association between the (GT)n polymorphism of the HO-1 gene promoter region and cancer risk: A metaanalysis. *Asian Pacific J Cancer Prev.* (2014) 15:4617–22. doi: 10.7314/ APJCP.2014.15.11.4617 32. Furfaro AL, Traverso N, Domenicotti C, Piras S, Moretta L, Marinari UM, et al. The nrf2/HO-1 axis in cancer cell growth and chemoresistance. Oxid Med Cell Longevity. (2016) 2016. doi: 10.1155/2016/1958174

33. Ricciuti B, Arbour KC, Lin JJ, Vajdi A, Vokes N, Hong L, et al. Diminished efficacy of programmed death-(Ligand)1 inhibition in STK11- and KEAP1-mutant lung adenocarcinoma is affected by KRAS mutation status. *J Thorac Oncol.* (2022) 17:399–410. doi: 10.1016/j.jtho.2021.10.013

34. Tong KI, Katoh Y, Kusunoki H, Itoh K, Tanaka T, Yamamoto M. Keap1 recruits Neh2 through binding to ETGE and DLG motifs: characterization of the two-site molecular recognition model. *Mol Cell Biol.* (2006) 26:2887–900. doi: 10.1128/ MCB.26.8.2887-2900.2006

35. Malhotra D, Portales-Casamar E, Singh A, Srivastava S, Arenillas D, Happel C, et al. Global mapping of binding sites for Nrf2 identifies novel targets in cell survival response through chip-seq profiling and network analysis. *Nucleic Acids Res.* (2010) 38:5718–34. doi: 10.1093/nar/gkq212

36. Hayes JD, Dinkova-Kostova AT. The Nrf2 regulatory network provides an interface between redox and intermediary metabolism. *Trends Biochem Sci.* (2014) 39:199–218. doi: 10.1016/j.tibs.2014.02.002

37. Di Biase S, Lee C, Brandhorst S, Manes B, Buono R, Cheng CW, et al. Fastingmimicking diet reduces HO-1 to promote T cell-mediated tumor cytotoxicity. *Cancer Cell.* (2016) 30:136–46. doi: 10.1016/j.ccell.2016.06.005

38. Zou C, Zou C, Cheng W, Li Q, Han Z, Wang X, et al. Heme oxygenase-1 retards hepatocellular carcinoma progression through the microRNA pathway. *Oncol Rep.* (2016) 36:2715–22. doi: 10.3892/or.2016.5056

39. Tertil M, Golda S, Skrzypek K, Florczyk U, Weglarczyk K, Kotlinowski J, et al. Nrf2-heme oxygenase-1 axis in mucoepidermoid carcinoma of the lung: Antitumoral effects associated with down-regulation of matrix metalloproteinases. *Free Radical Biol Med.* (2015) 89:147–57. doi: 10.1016/j.freeradbiomed.2015.08.004

40. Lin PH, Lan WM, Chau LY. TRC8 suppresses tumorigenesis through targeting heme oxygenase-1 for ubiquitination and degradation. *Oncogene*. (2013) 32:2325–34. doi: 10.1038/onc.2012.244

41. Luu Hoang KN, Anstee JE, Arnold JN. The diverse roles of heme oxygenase-1 in tumor progression. *Front Immunol.* (2021) 9. doi: 10.3389/fimmu.2021.658315

42. Gottlieb Y, Topaz O, Cohen LA, Yakov LD, Haber T, Morgenstern A, et al. Physiologically aged red blood cells undergo erythrophagocytosis *in vivo* but not in *vitro*. *Haematologica*. (2012) 97:994–1002. doi: 10.3324/haematol.2011.057620

43. Mayerhofer M, Florian S, Krauth MT, Aichberger KJ, Bilban M, Marculescu R, et al. Identification of heme oxygenase-1 as a novel BCR/ABL-dependent survival factor in chronic myeloid leukemia. *Cancer Res.* (2004) 64:3148–54. doi: 10.1158/0008-5472.CAN-03-1200

44. Muliaditan T, Caron J, Okesola M, Opzoomer JW, Kosti P, Georgouli M, et al. Macrophages are exploited from an innate wound healing response to facilitate cancer metastasis. *Nat Commun.* (2018) 9. doi: 10.1038/s41467-018-05346-7

45. Alaluf E, Vokaer B, Detavernier A, Azouz A, Splittgerber M, Carrette A, et al. Heme oxygenase-1 orchestrates the immunosuppressive program of tumor-associated macrophages. *JCI Insight*. (2020) 5. doi: 10.1172/jci.insight.133929

46. Was H, Dulak J, Jozkowicz A. Heme oxygenase-1 in tumor biology and therapy. *Curr Drug Targets.* (2012) 11:1551–70. doi: 10.2174/1389450111009011551

47. Chauveau C, Rémy S, Royer PJ, Hill M, Tanguy-Royer S, Hubert FX, et al. Heme oxygenase-1 expression inhibits dendritic cell maturation and proinflammatory function but conserves IL-10 expression. *Blood.* (2005) 106:1694–702. doi: 10.1182/ blood-2005-02-0494

48. El Andaloussi A, Lesniak MS. CD4+CD25+FoxP3+ T-cell infiltration and heme oxygenase-1 expression correlate with tumor grade in human gliomas. *J Neuro-Oncology.* (2007) 83:145–52. doi: 10.1007/s11060-006-9314-y

49. Murray PJ, Allen JE, Biswas SK, Fisher EA, Gilroy DW, Goerdt S, et al. Macrophage activation and polarization: nomenclature and experimental guidelines. *Immunity.* (2014) 41:14–20. doi: 10.1016/j.immuni.2014.06.008

50. Mantovani A, Sozzani S, Locati M, Allavena P, Sica A. Macrophage polarization: Tumor-associated macrophages as a paradigm for polarized M2 mononuclear phagocytes. *Trends Immunol.* (2002) 23:549–55. doi: 10.1016/S1471-4906(02)02302-5

51. Arnold JN, Magiera L, Kraman M, Fearon DT. Tumoral immune suppression by macrophages expressing fibroblast activation protein- α and heme oxygenase-1. *Cancer Immunol Res.* (2014) 2:121–6. doi: 10.1158/2326-6066.CIR-13-0150

52. Nemeth Z, Li M, Csizmadia E, Döme B, Johansson M, Persson JL, et al. Heme oxygenase-1 in macrophages controls prostate cancer progression. *Oncotarget.* (2015) 6:33675–88. doi: 10.18632/oncotarget.5284

53. Kumar S, Chan CJ, Coussens LM. Inflammation and cancer. *Encyclopedia* Immunobiology. (2016) 4:406–15. doi: 10.1016/B978-0-12-374279-7.17002-X

54. Hemmati M, Yousefi B, Bahar A, Eslami M. Importance of heme oxygenase-1 in gastrointestinal cancers: functions, inductions, regulations, and signaling. *J Gastrointestinal Cancer*. (2029) 52(2):454–61. doi: 10.1007/s12029-021-00587-0

55. Akter Z, Salamat N, Yousuf A, Zhang L. The promise of targeting heme and mitochondrial respiration in normalizing tumor microenvironment and potentiating immunotherapy. *Front Oncol.* (2023) 12. doi: 10.3389/fonc.2022.1072739

56. Julia T, Zhang Paul J, Yingtao B, Celine S, Rajrupa M, Stephen TL, et al. Fibroblast activation protein expression by stromal cells and tumor-associated

macrophages in human breast cancer. Hum Pathol. (2013) 44:2549-57. doi: 10.1016/j.humpath.2013.06.016

57. Grochot-Przeczek A, Lach R, Mis J, Skrzypek K, Gozdecka M, Sroczynska P, et al. Heme oxygenase-1 accelerates cutaneous wound healing in mice. *PloS One.* (2009) 4:5803. doi: 10.1371/journal.pone.0005803

58. Flier JS, Underhill LH, Dvorak HF. Tumors: wounds that do not heal. New Engl J Med. (1986) 315:1650–9. doi: 10.1056/nejm198612253152606

59. Marelli G, Erreni M, Anselmo A, Taverniti V, Guglielmetti S, Mantovani A, et al. Heme-oxygenase-1 production by intestinal CX3CR1+ Macrophages helps to resolve inflammation and prevents carcinogenesis. *Cancer Res.* (2017) 77:4472–85. doi: 10.1158/0008-5472.CAN-16-2501

60. Weis N, Weigert A, Von Knethen A, Brü B. Heme oxygenase-1 contributes to an alternative macrophage activation profile induced by apoptotic cell supernatants. *Mol Biol Cell.* (2009) 20:1280–8. doi: 10.1091/mbc.E08

61. Zhang M, Nakamura K, Kageyama S, Lawal AO, Gong KW, Bhetraratana M, et al. Myeloid HO-1 modulates macrophage polarization and protects against ischemiareperfusion injury. *JCI Insight*. (2018) 3. doi: 10.1172/jci.insight.120596

62. Kim SH, Saeidi S, Zhong X, Gwak SY, Muna IA, Park SA, et al. Breast cancer cell debris diminishes therapeutic efficacy through heme oxygenase-1-mediated inactivation of M1-like tumor-associated macrophages. *Neoplasia (United States).* (2020) 22:606–16. doi: 10.1016/j.neo.2020.08.006

63. Kim SH, Kim S-J, Park J, Joe Y, Lee SE, Saeidi S, et al. Reprograming of tumorassociated macrophages in breast tumor-bearing mice under chemotherapy by targeting heme oxygenase-1. *Antioxidants*. (2021) 10:470. doi: 10.3390/antiox10030470

64. Magri S, Musca B, Pinton L, Orecchini E, Belladonna ML, Orabona C, et al. The immunosuppression pathway of tumor-associated macrophages is controlled by heme oxygenase-1 in glioblastoma patients. *International Journal of Cancer*. (2022) 151 (12):2265–77. doi: 10.1002/ijc.34270

65. Deng R, Wang SM, Yin T, Ye TH, Shen GB, Li L, et al. Inhibition of tumor growth and alteration of associated macrophage cell type by an HO-1 inhibitor in breast carcinoma-bearing mice. *Oncol Res.* (2013) 20:473–82. doi: 10.3727/096504013X13715991125684

66. Kinoshita H, Türkan H, Vucinic S, Naqvi S, Bedair R, Rezaee R, et al. Carbon monoxide poisoning. *Toxicol Rep.* (2020) 7:169-73. doi: 10.1016/j.toxrep.2020.01.005

67. Oza PP, Kashfi K. The Triple Crown: NO, CO, and H2S in cancer cell biology. *Pharmacol Ther.* (2023) 249. doi: 10.1016/j.pharmthera.2023.108502

68. Lee TS, Tsai HL, Chau LY. Induction of heme oxygenase-1 expression in murine macrophages is essential for the anti-inflammatory effect of low dose 15-Deoxy-Δ12,14-prostaglandin J2. J Biol Chem. (2003) 278:19325–30. doi: 10.1074/jbc.M300498200

69. Nakahira K, Hong PK, Xue HG, Nakao A, Wang X, Murase N, et al. Carbon monoxide differentially inhibits TLR signaling pathways by regulating ROS-induced trafficking of TLRs to lipid rafts. *J Exp Med.* (2006) 203:2377–89. doi: 10.1084/jem.20060845

70. Rémy S, Blancou P, Tesson L, Tardif V, Brion R, Royer PJ, et al. Carbon monoxide inhibits TLR-induced dendritic cell immunogenicity. *J Immunol.* (2009) 182:1877-84. doi: 10.4049/jimmunol.0802436

71. Nemeth Z, Csizmadia E, Vikstrom L, Li M, Bisht K, Feizi A, et al. Alterations of tumor microenvironment by carbon monoxide impedes lung cancer growth. *Oncotarget* (2016) 7(17):23919–32. Available online at: www.impactjournals.com/ oncotarget.

72. Trojandt S, Bellinghausen I, Reske-Kunz AB, Bros M. Tumor-derived immunomodulators induce overlapping pro-tolerogenic gene expression signatures in human dendritic cells. *Hum Immunol.* (2016) 77:1223–31. doi: 10.1016/j.humimm.2016.08.014

73. Wegiel B, Baty CJ, Gallo D, Csizmadia E, Scott JR, Akhavan A, et al. Cell surface biliverdin reductase mediates biliverdin-induced anti-inflammatory effects via phosphatidylinositol 3-kinase and Akt. *J Biol Chem.* (2009) 284:21369–78. doi: 10.1074/jbc.M109.027433

74. Liu Y, Li P, Lu J, Xiong W, Oger J, Tetzlaff W, et al. Bilirubin possesses powerful immunomodulatory activity and suppresses experimental autoimmune encephalomyelitis. *J Immunol.* (2008) 181:1887–97. doi: 10.4049/jimmunol.181.3.1887

75. Dey M, Chang AL, Wainwright DA, Ahmed AU, Han Y, Balyasnikova IV, et al. Heme oxygenase-1 protects regulatory T cells from hypoxia-induced cellular stress in an experimental mouse brain tumor model. *J Neuroimmunology.* (2014) 266:33–42. doi: 10.1016/j.jneuroim.2013.10.012

76. Pae H-O, Oh G-S, Choi B-M, Chae S-C, Kim Y-M, Chung K-R, et al. Carbon monoxide produced by heme oxygenase-1 suppresses T cell proliferation via inhibition of IL-2 production. *J Immunol.* (2004) 172:4744–51. doi: 10.4049/jimmunol.172.8.4744

77. Song R, Mahidhara RS, Zhou Z, Hoffman RA, Seol D-W, Flavell RA, et al. Carbon monoxide inhibits T lymphocyte proliferation via caspase-dependent pathway. *J Immunol.* (2004) 172:1220–6. doi: 10.4049/jimmunol.172.2.1220

78. Gómez-Lomelí P, Bravo-Cuellar A, Hernández-Flores G, Jave-Suárez LF, Aguilar-Lemarroy A, Lerma-Díaz JM, et al. Increase of IFN- γ and TNF- α production in CD107a + NK-92 cells co-cultured with cervical cancer cell lines pre-treated with the HO-1 inhibitor. *Cancer Cell Int.* (2014) 14. doi: 10.1186/s12935-014-0100-1

79. Zhang T, Fang Q, Liu P, Wang P, Feng C, Wang J. Heme oxygenase 1 overexpression induces immune evasion of acute myeloid leukemia against natural killer cells by inhibiting CD48. *J Trans Med.* (2022) 20. doi: 10.1186/s12967-022-03589-z

80. Castruccio Castracani C, Longhitano L, Distefano A, Di Rosa M, Pittalà V, Lupo G, et al. Heme oxygenase-1 and carbon monoxide regulate growth and progression in glioblastoma cells. *Mol Neurobiol.* (2020) 57:2436–46. doi: 10.1007/s12035-020-01869-7

81. Bauer A, Mylroie H, Thornton CC, Calay D, Birdsey GM, Kiprianos AP, et al. Identification of cyclins A1, E1 and vimentin as downstream targets of heme oxygenase-1 in vascular endothelial growth factor-mediated angiogenesis. *Sci Rep.* (2016) 6. doi: 10.1038/srep29417

82. Lin H-H, Chiang M-T, Chang P-C, Chau L-Y. Myeloid heme oxygenase-1 promotes metastatic tumor colonization in mice. *Cancer Sci.* (2015) 106:299-306. doi: 10.1111/cas.12604

83. Funes SC, Rios M, Fernández-Fierro A, Covián C, Bueno SM, Riedel CA, et al. Naturally derived heme-oxygenase 1 inducers and their therapeutic application to immunemediated diseases. *Front Immunol.* (2020) 11:1467. doi: 10.3389/fimmu.2020.01467

84. Ma LL, Sun L, Wang YX, Sun BH, Li YF, Jin YL. Association between HO-1 gene promoter polymorphisms and diseases (Review). *Mol Med Rep.* (2022) 25. doi: 10.3892/mmr.2021.12545

85. Marelli G, Allavena P. The good and the bad side of heme-oxygenase-1 in the gut. Antioxidants Redox Signaling. (2020) 32:1071–9. doi: 10.1089/ars.2019.7956

86. Konrad FM, Knausberg U, Höne R, Ngamsri KC, Reutershan J. Tissue heme oxygenase-1 exerts anti-inflammatory effects on LPS-induced pulmonary inflammation. *Mucosal Immunol.* (2016) 9:98–111. doi: 10.1038/mi.2015.39

87. Córdova EJ, Martínez-Hernández A, Ramírez-Bello J, Velázquez-Cruz R, Centeno F, Baca V, et al. HMOX1 promoter (GT)n polymorphism is associated with childhood-onset systemic lupus erythematosus but not with juvenile rheumatoid arthritis in a Mexican population. *Clin Exp Rheumatol.* (2012) 30:297–301.

88. Horio T, Morishita E, Mizuno S, Uchino K, Hanamura I, Espinoza JL, et al. Donor heme oxygenase-1 promoter gene polymorphism predicts survival after unrelated bone marrow transplantation for high-risk patients. *Cancers.* (2020) 12. doi: 10.3390/cancers12020424

89. Catrina AI, Svensson CI, Malmström V, Schett G, Klareskog L. Mechanisms leading from systemic autoimmunity to joint-specific disease in rheumatoid arthritis. *Nat Rev Rheumatol.* (2017) 13:79–86. doi: 10.1038/nrrheum.2016.200

90. Funes SC, Ríos M, Gómez-Santander F, Fernández-Fierro A, Altamirano-Lagos MJ, Rivera-Perez D, et al. Tolerogenic dendritic cell transfer ameliorates systemic lupus erythematosus in mice. *Immunology*. (2019) 158:322–39. doi: 10.1111/imm.13119

91. Hull TD, Agarwal A, George JF. The mononuclear phagocyte system in homeostasis and disease: A role for heme oxygenase-1. *Antioxidants Redox Signaling*. (2014) 20:1770-88. doi: 10.1089/ars.2013.5673

92. Lee TS, Chau LY. Heme oxygenase-1 mediates the anti-inflammatory effect of interleukin-10 in mice. Nat Med. (2002) 8:240-6. doi: 10.1038/nm0302-240

93. Ryter SW, Choi AMK. Targeting heme oxygenase-1 and carbon monoxide for therapeutic modulation of inflammation. *Trans Res.* (2016) 167:7–34. doi: 10.1016/j.trsl.2015.06.011

94. Naito Y, Takagi T, Higashimura Y. Heme oxygenase-1 and anti-inflammatory M2 macrophages. *Arch Biochem Biophysics*. (2014) 564:83-8. doi: 10.1016/j.abb.2014.09.005

95. Ramos GP, Papadakis KA. Mechanisms of disease: inflammatory bowel diseases. *Mayo Clinic Proc.* (2019) 94:155–65. doi: 10.1016/j.mayocp.2018.09.013

96. De Souza HSP, Fiocchi C. Immunopathogenesis of IBD: Current state of the art. *Nat Rev Gastroenterol Hepatol.* (2016) 13:13–27. doi: 10.1038/nrgastro.2015.186

97. Takagi T, Naito Y, Mizushima K, Nukigi Y, Okada H, Suzuki T, et al. Increased intestinal expression of heme oxygenase-1 and its localization in patients with ulcerative colitis. J Gastroenterol Hepatol (Australia) J Gastroenterol Hepatol. (2008) 23(Suppl 2):S229–33. doi: 10.1111/j.1440-1746.2008.05443.x

98. Levine AD, Fiocchi C. Immunology of inflammatory bowel disease. Curr Opin Gastroenterol. (2000) 16:306–9. doi: 10.1097/00001574-200007000-00003

99. Sun L, Clavijo PE, Robbins Y, Patel P, Friedman J, Greene S, et al. Inhibiting myeloid-derived suppressor cell trafficking enhances T cell immunotherapy. *JCI Insight*. (2019) 4. doi: 10.1172/jci.insight.126853

100. Zhang L, Zhang Y, Zhong W, Di C, Lin X, Xia Z. Heme oxygenase-1 ameliorates dextran sulfate sodiuminduced acute murine colitis by regulating Th17/ Treg cell balance. J Biol Chem. (2014) 289:26847–58. doi: 10.1074/jbc.M114.590554

101. Rana N, McLean S, Mann BE, Poole RK. Interaction of the carbon monoxidereleasing molecule Ru(CO)3Cl(glycinate) (CORM-3) with Salmonella enterica serovar Typhimurium: *In situ* measurements of carbon monoxide binding by integrating cavity dual-beam spectrophotometry. *Microbiol (United Kingdom)*. (2014) 160:2771–9. doi: 10.1099/mic.0.081042-0

102. Onyiah JC, Sheikh SZ, Maharshak N, Steinbach EC, Russo SM, Kobayashi T, et al. Carbon monoxide and heme oxygenase-1 prevent intestinal inflammation in mice by promoting bacterial clearance. *Gastroenterology*. (2013) 144:789–98. doi: 10.1053/j.gastro.2012.12.025

103. Mumby S, Upton RL, Chen Y, Stanford SJ, Quinlan GJ, Nicholson AG, et al. Lung heme oxygenase-1 is elevated in acute respiratory distress syndrome. *Crit Care Med.* (2004) 32:1130–5. doi: 10.1097/01.CCM.0000124869.86399.F2

104. Fredenburgh LE, Perrella MA, Mitsialis SA. The role of heme oxygenase-1 in pulmonary disease. *Am J Respir Cell Mol Biol.* (2007) 36:158. doi: 10.1165/RCMB.2006-0331TR

105. Tagami Y, Hara Y, Murohashi K, Nagasawa R, Fujii H, Izawa A, et al. Serum heme oxygenase-1 as a prognostic biomarker in patients with acute exacerbation of interstitial lung disease. *Sci Rep.* (2023) 13:1–9. doi: 10.1038/s41598-023-49342-4

106. Hashiba T, Suzuki M, Nagashima Y, Suzuki S, Inoue S, Tsuburai T, et al. Adenovirus-mediated transfer of heme oxygenase-1 cDNA attenuates severe lung injury induced by the influenza virus in mice. *Gene Ther.* (2001) 8:1499–507. doi: 10.1038/SJ.GT.3301540

107. Di Pietro C, Öz HH, Zhang PX, Cheng EC, Martis V, Bonfield TL, et al. Recruitment of monocytes primed to express heme oxygenase-1 ameliorates pathological lung inflammation in cystic fibrosis. *Exp Mol Med.* (2022) 54:639–52. doi: 10.1038/s12276-022-00770-8

108. Bruscia EM, Bonfield TL. Cystic fibrosis lung immunity: the role of the macrophage. J Innate Immun. (2016) 8:550–63. doi: 10.1159/000446825

109. Yamasaki K, Van Eeden SF. Lung macrophage phenotypes and functional responses: Role in the pathogenesis of COPD. *Int J Mol Sci.* (2018) 19. doi: 10.3390/ ijms19020582

110. Chen HY, Tzeng IS, Tsai KW, Wu YK, Cheng CF, Lu KC, et al. Association between heme oxygenase one and sepsis development in patients with moderate-to-critical COVID-19: a single-center, retrospective observational study. *Eur J Med Res.* (2022) 27. doi: 10.1186/s40001-022-00915-5

111. Hara Y, Tsukiji J, Yabe A, Onishi Y, Hirose H, Yamamoto M, et al. Heme oxygenase-1 as an important predictor of the severity of COVID-19. *PloS One.* (2022) 17. doi: 10.1371/journal.pone.0273500

112. Wu YH, Hsieh HL. Roles of heme oxygenase-1 in neuroinflammation and brain disorders. *Antioxidants.* (2022) 11. doi: 10.3390/antiox11050923

113. Consoli V, Sorrenti V, Grosso S, Vanella L. Heme oxygenase-1 signaling and redox homeostasis in physiopathological conditions. *Biomolecules*. (2021) 11. doi: 10.3390/biom11040589

114. Upadhayay S, Mehan S. Targeting Nrf2/HO-1 anti-oxidant signaling pathway in the progression of multiple sclerosis and influences on neurological dysfunctions. *Brain Disord.* (2021) 3. doi: 10.1016/j.dscb.2021.100019

115. Nitti M, Piras S, Brondolo L, Marinari UM, Pronzato MA, Furfaro AL. Heme oxygenase 1 in the nervous system: Does it favor neuronal cell survival or induce neurodegeneration? *Int J Mol Sci.* (2018) 19. doi: 10.3390/ijms19082260

116. Loboda A, Damulewicz M, Pyza E, Jozkowicz A, Dulak J. Role of Nrf2/HO-1 system in development, oxidative stress response and diseases: an evolutionarily conserved mechanism. *Cell Mol Life Sci.* (2016) 73:3221–47. doi: 10.1007/s00018-016-2223-0

117. Barañano DE, Rao M, Ferris CD, Snyder SH. Biliverdin reductase: A major physiologic cytoprotectant. *Proc Natl Acad Sci United States America*. (2002) 99:16093–8. doi: 10.1073/pnas.252626999

118. Liu R, Yang J, Li Y, Xie J, Wang J. Heme oxygenase-1: The roles of both good and evil in neurodegenerative diseases. *J Neurochemistry*. (2023) 167:347–61. doi: 10.1111/jnc.15969

119. Moscat J, Diaz-Meco MT. p62 at the crossroads of autophagy, apoptosis, and cancer. Cell. (2009) 137:1001–4. doi: 10.1016/j.cell.2009.05.023

120. Lau A, Wang X-J, Zhao F, Villeneuve NF, Wu T, Jiang T, et al. A noncanonical mechanism of nrf2 activation by autophagy deficiency: direct interaction between keap1 and p62. *Mol Cell Biol.* (2010) 30:3275–85. doi: 10.1128/mcb.00248-10

121. Ferris CD, Jaffrey SR, Sawa A, Takahashi M, Brady SD, Barrow RK, et al. Haem oxygenase-1 prevents cell death by regulating cellular iron. *Nat Cell Biol.* (1999) 1:152–7. doi: 10.1038/11072

122. Lassmann H, Bradl M. Multiple sclerosis: experimental models and reality. Acta Neuropathologica. (2017) 133:223-44. doi: 10.1007/s00401-016-1631-4

123. Mackern-Oberti J, Riquelme S, Llanos C, Schmidt C, Simon T, Anegon I, et al. Heme oxygenase-1 as a target for the design of gene and pharmaceutical therapies for autoimmune diseases. *Curr Gene Ther.* (2014) 14:218–35. doi: 10.2174/1566523214666140424150308

124. Fagone P, Patti F, Mangano K, Mammana S, Coco M, Touil-Boukoffa C, et al. Heme oxygenase-1 expression in peripheral blood mononuclear cells correlates with disease activity in multiple sclerosis. *J Neuroimmunology*. (2013) 261:82–6. doi: 10.1016/j.jneuroim.2013.04.013

125. Glatigny S, Bettelli E. Experimental autoimmune encephalomyelitis (EAE) as animal models of multiple sclerosis (MS). *Cold Spring Harbor Perspect Med.* (2018) 8:1079–106. doi: 10.1101/cshperspect.a028977

126. Chora ÂA, Fontoura P, Cunha A, Pais TF, Cardoso S, Ho PP, et al. Heme oxygenase-1 and carbon monoxide suppress autoimmune neuroinflammation. *J Clin Invest.* (2007) 117:438–47. doi: 10.1172/JCI28844

127. Metaxas A, Thygesen C, Kempf SJ, Anzalone M, Vaitheeswaran R, Petersen S, et al. Ageing and amyloidosis underlie the molecular and pathological alterations of tau in a mouse model of familial Alzheimer's disease. *Sci Rep.* (2019) 9. doi: 10.1038/ s41598-019-52357-5

128. Hettiarachchi NT, Dallas ML, Al-Owais MM, Griffiths HH, Hooper NM, Scragg JL, et al. Heme oxygenase-1 protects against Alzheimer's amyloid- β 1-42-induced toxicity via carbon monoxide production. *Cell Death Dis.* (2014) 5. doi: 10.1038/cddis.2014.529

129. Fernández-Mendívil C, Luengo E, Trigo-Alonso P, García-Magro N, Negredo P, López MG. Protective role of microglial HO-1 blockade in aging: Implication of iron metabolism. *Redox Biol.* (2021) 38. doi: 10.1016/j.redox.2020.101789

130. Fernández-Mendívil C, Arreola MA, Hohsfield LA, Green KN, Lopez MG. Aging and progression of beta-amyloid pathology in alzheimer's disease correlates with microglial heme-oxygenase-1 overexpression. *Antioxidants*. (2020) 9:1–17. doi: 10.3390/antiox9070644

131. Schipper HM, Song W, Tavitian A, Cressatti M. The sinister face of heme oxygenase-1 in brain aging and disease. *Prog Neurobiol.* (2019) 172:40–70. doi: 10.1016/ j.pneurobio.2018.06.008

132. Sun W, Zheng J, Ma J, Wang Z, Shi X, Li M, et al. Increased plasma heme oxygenase-1 levels in patients with early-stage parkinson's disease. *Front Aging Neurosci.* (2021) 13:621508. doi: 10.3389/fnagi.2021.621508

133. Schipper HM, Song W. A heme oxygenase-1 transducer model of degenerative and developmental brain disorders. *Int J Mol Sci.* (2015) 16:5400–19. doi: 10.3390/ ijms16035400

134. Kang JE, Lim MM, Bateman RJ, Lee JJ, Smyth LP, Cirrito JR, et al. Amyloid- β dynamics are regulated by orexin and the sleep-wake cycle. *Science*. (2009) 326:1005–7. doi: 10.1126/science.1180962

135. Spaan J, Kolyva C, van den Wijngaard J, ter Wee R, van Horssen P, Piek J, et al. Coronary structure and perfusion in health and disease. *Philos Trans R Soc A: Mathematical Phys Eng Sci.* (2008) 366:3137–53. doi: 10.1098/rsta.2008.0075

136. Otterbein LE, Foresti R, Motterlini R. Heme oxygenase-1 and carbon monoxide in the heart: the balancing act between danger signaling and pro-survival. *Circ Res.* (2016) 118:1940–59. doi: 10.1161/CIRCRESAHA.116.306588

137. Sica A, Mantovani A. Macrophage plasticity and polarization: *In vivo* veritas. *J Clin Invest.* (2012) 122:787–95. doi: 10.1172/JCI59643

138. Mosser DM, Hamidzadeh K, Goncalves R. Macrophages and the maintenance of homeostasis. *Cell Mol Immunol.* (2021) 18:579–87. doi: 10.1038/s41423-020-00541-3

139. Barrett TJ. Macrophages in atherosclerosis regression. Arteriosclerosis Thrombosis Vasc Biol. (2020) 40:20-33. doi: 10.1161/ATVBAHA.119.312802

140. Yap J, Cabrera-Fuentes HA, Irei J, Hausenloy DJ, Boisvert WA. Role of macrophages in cardioprotection. Int J Mol Sci. (2019) 20. doi: 10.3390/ijms20102474

141. Orozco LD, Kapturczak MH, Barajas B, Wang X, Weinstein MM, Wong J, et al. Heme oxygenase-1 expression in macrophages plays a beneficial role in atherosclerosis. *Circ Res.* (2007) 100:1703–11. doi: 10.1161/CIRCRESAHA.107.151720

142. Wang XM, Kim HP, Nakahira K, Ryter SW, Choi AMK. The heme oxygenase-1/carbon monoxide pathway suppresses TLR4 signaling by regulating the interaction of TLR4 with caveolin-1. *J Immunol (Baltimore Md: 1950).* (2009) 182:3809–18. doi: 10.4049/JIMMUNOL.0712437

143. Morsi WG, Shaker OG, Ismail EF, Ahmed HH, El-Serafi TI, Maklady FA, et al. HO-1 and VGEF gene expression in human arteries with advanced atherosclerosis. *Clin Biochem.* (2006) 39:1057–62. doi: 10.1016/J.CLINBIOCHEM.2006.08.003

144. Pradhan P, Vijayan V, Gueler F, Immenschuh S. Interplay of heme with macrophages in homeostasis and inflammation. *Int J Mol Sci.* (2020) 21. doi: 10.3390/ ijms21030740

145. Alam MZ, Devalaraja S, Haldar M. The heme connection: Linking erythrocytes and macrophage biology. *Front Immunol.* (2017) 8:33. doi: 10.3389/fimmu.2017.00033

146. Lörchner H, Pöling J, Gajawada P, Hou Y, Polyakova V, Kostin S, et al. Myocardial healing requires Reg3 β -dependent accumulation of macrophages in the ischemic heart. *Nat Med.* (2015) 21:353–62. doi: 10.1038/nm.3816

147. Hulsmans M, Clauss S, Xiao L, Aguirre AD, King KR, Hanley A, et al. Macrophages facilitate electrical conduction in the heart. *Cell.* (2017) 169:510–522.e20. doi: 10.1016/j.cell.2017.03.050

148. MacHnik A, Neuhofer W, Jantsch J, Dahlmann A, Tammela T, MacHura K, et al. Macrophages regulate salt-dependent volume and blood pressure by a vascular endothelial growth factor-C-dependent buffering mechanism. *Nat Med.* (2009) 15:545–52. doi: 10.1038/nm.1960

149. Duncan SE, Gao S, Sarhene M, Coffie JW, Linhua D, Bao X, et al. Macrophage activities in myocardial infarction and heart failure. *Cardiol Res Pract.* (2020) 2020. doi: 10.1155/2020/4375127

150. Shan H, Li T, Zhang L, Yang R, Li Y, Zhang M, et al. Heme oxygenase-1 prevents heart against myocardial infarction by attenuating ischemic injury-induced cardiomyocytes senescence. *EBioMedicine*. (2019) 39:59–68. doi: 10.1016/j.ebiom.2018.11.056

151. Zhang H, Thorwald MA, D'agostino C, Morgan TE, Finch CE, Forman HJ. Inhibiting Bach1 enhanced the activation of Nrf2 signaling and the degradation of HNE in response to oxidative stress. *Alzheimer's dementia: J Alzheimer's Assoc.* (2021) 17: e053235. doi: 10.1002/alz.053235

152. Yano Y, Ozono R, Oishi Y, Kambe M, Yoshizumi M, Ishida T, et al. Genetic ablation of the transcription repressor Bach1 leads to myocardial protection against ischemia/reperfusion in mice. *Genes to Cells.* (2006) 11:791–803. doi: 10.1111/j.1365-2443.2006.00979.x

153. Law SKA, Micklem KJ, Shaw JM, Zhang X -P, Dong Y, Willis AC, et al. A new macrophage differentiation antigen which is a member of the scavenger receptor superfamily. *Eur J Immunol.* (1993) 23:2320–5. doi: 10.1002/eji.1830230940

154. Gleissner CA, Shaked I, Erbel C, Böckler D, Katus HA, Ley K. CXCL4 downregulates the atheroprotective hemoglobin receptor CD163 in human macrophages. *Circ Res.* (2010) 106:203–11. doi: 10.1161/CIRCRESAHA.109.199505

155. Guo L, Akahori H, Harari E, Smith SL, Polavarapu R, Karmali V, et al. CD163+ macrophages promote angiogenesis and vascular permeability accompanied by inflammation in atherosclerosis. *J Clin Invest.* (2018) 128:1106–24. doi: 10.1172/ JCI93025

156. Etzerodt A, Maniecki MB, Møller K, Møller HJ, Moestrup SK. Tumor necrosis factor α-converting enzyme (TACE/ADAM17) mediates ectodomain shedding of the scavenger receptor CD163. *J Leukocyte Biol.* (2010) 88:1201–5. doi: 10.1189/jlb.0410235

157. Durda P, Raffield LM, Lange EM, Olson NC, Jenny NS, Cushman M, et al. Circulating soluble CD163, associations with cardiovascular outcomes and mortality, and identification of genetic variants in older individuals: the cardiovascular health study. J Am Heart Assoc. (2022) 11. doi: 10.1161/JAHA.121.024374

158. Ventura-Clapier R, Garnier A, Veksler V. Transcriptional control of mitochondrial biogenesis: the central role of PGC-1alpha. *Cardiovasc Res.* (2008) 79:208–17. doi: 10.1093/CVR/CVN098

159. Keith M, Geranmayegan A, Sole MJ, Kurian R, Robinson A, Omran AS, et al. Increased oxidative stress in patients with congestive heart failure. *J Am Coll Cardiol.* (1998) 31:1352–6. doi: 10.1016/S0735-1097(98)00101-6

160. Paine A, Eiz-Vesper B, Blasczyk R, Immenschuh S. Signaling to heme oxygenase-1 and its anti-inflammatory therapeutic potential. *Biochem Pharmacol.* (2010) 80:1895–903. doi: 10.1016/j.bcp.2010.07.014

161. Sharma HS, Maulik N, Gho BCG, Das DK, Verdouw PD. Coordinated expression of heme oxygenase-1 and ubiquitin in the porcine heart subjected to ischemia and reperfusion. *Mol Cell Biochem.* (1996) 157:111-6. doi: 10.1007/BF00227888

162. Wakabayashi N, Dinkova-Kostova AT, Holtzclaw WD, Kang M, Kobayashi A, Yamamoto M, et al. Protection against electrophile and oxidant stress by induction of the phase 2 response: fate of cysteines of the Keap1 sensor modified by inducers. *Proc Natl Acad Sci United States America*. (2004) 101:2040–5. doi: 10.1073/ PNAS.0307301101

163. Motohashi H, Shavit JA, Igarashi K, Yamamoto M, Engel JD. The world according to Maf. *Nucleic Acids Res.* (1997) 25:2953–9. doi: 10.1093/NAR/25.15.2953

164. Hausenloy DJ, Yellon DM. Myocardial ischemia-reperfusion injury: A neglected therapeutic target. J Clin Invest. (2013) 123:92–100. doi: 10.1172/JCI62874

165. Stocker R, Yamamoto Y, McDonagh AF, Glazer AN, Ames BN. Bilirubin is an antioxidant of possible physiological importance. *Science*. (1987) 235:1043-6. doi: 10.1126/science.3029864

166. Stojanov M, Stefanovic A, Dzingalasevic G, Ivanisevic J, Miljkovic M, Mandic-Radic S, et al. Total bilirubin in young men and women: Association with risk markers for cardiovascular diseases. *Clin Biochem*. (2013) 46:1516–9. doi: 10.1016/ j.clinbiochem.2013.06.020

167. Gitto M, Novelli L, Cozzi O, Reimers B, Condorelli G, Stefanini GG. Specific characteristics of STEMI in COVID-19 patients and their practical implications. *Kardiologia Polska*. (2022) 80:266–77. doi: 10.33963/KP.a2022.0072

168. Tobin RP, Jordan KR, Robinson WA, Davis D, Borges VF, Gonzalez R, et al. Targeting myeloid-derived suppressor cells using all-trans retinoic acid in melanoma patients treated with Ipilimumab. *Int Immunopharmacol.* (2018) 63:282–91. doi: 10.1016/j.intimp.2018.08.007

169. Topkara VK, Evans S, Zhang W, Epelman S, Staloch L, Barger PM, et al. Therapeutic targeting of innate immunity in the failing heart. *J Mol Cell Cardiol*. (2011) 51:594–9. doi: 10.1016/j.yjmcc.2010.11.003

170. Li W, Hsiao HM, Higashikubo R, Saunders BT, Bharat A, Goldstein DR, et al. Heart-resident CCR2+ macrophages promote neutrophil extravasation through TLR9/ MyD88/CXCL5 signaling. *JCI Insight*. (2016) 1. doi: 10.1172/JCI.INSIGHT.87315

171. Leuschner F, Rauch PJ, Ueno T, Gorbatov R, Marinelli B, Lee WW, et al. Rapid monocyte kinetics in acute myocardial infarction are sustained by extramedullary monocytopoiesis. J Exp Med. (2012) 209:123–37. doi: 10.1084/JEM.20111009

172. Hilgendorf I, Gerhardt LMS, Tan TC, Winter C, Holderried TAW, Chousterman BG, et al. Ly-6Chigh monocytes depend on Nr4a1 to balance both inflammatory and reparative phases in the infarcted myocardium. *Circ Res.* (2014) 114:1611–22. doi: 10.1161/CIRCRESAHA.114.303204

173. Swirski FK, Nahrendorf M, Etzrodt M, Wildgruber M, Cortez-Retamozo V, Panizzi P, et al. Identification of splenic reservoir monocytes and their deployment to inflammatory sites. *Sci (New York NY)*. (2009) 325:612–6. doi: 10.1126/SCIENCE.1175202

174. Hinkel R, Lange P, Petersen B, Gottlieb E, Ng JKM, Finger S, et al. Heme oxygenase-1 gene therapy provides cardioprotection via control of post-ischemic inflammation: an experimental study in a pre-clinical pig model. *J Am Coll Cardiol.* (2015) 66:154–65. doi: 10.1016/J.JACC.2015.04.064

175. Tomczyk M, Kraszewska I, Szade K, Bukowska-Strakova K, Meloni M, Jozkowicz A, et al. Splenic Ly6Chi monocytes contribute to adverse late postischemic left ventricular remodeling in heme oxygenase-1 deficient mice. *Basic Res Cardiol.* (2017) 112. doi: 10.1007/S00395-017-0629-Y

176. Aurora AB, Porrello ER, Tan W, Mahmoud AI, Hill JA, Bassel-Duby R, et al. Macrophages are required for neonatal heart regeneration. *J Clin Invest.* (2014) 124:1382–92. doi: 10.1172/JCI72181

177. Lavine KJ, Epelman S, Uchida K, Weber KJ, Nichols CG, Schilling JD, et al. Distinct macrophage lineages contribute to disparate patterns of cardiac recovery and

remodeling in the neonatal and adult heart. *Proc Natl Acad Sci United States America*. (2014) 111:16029–34. doi: 10.1073/PNAS.1406508111

178. Ross R. The pathogenesis of atherosclerosis: a perspective for the 1990s. *Nature*. (1993) 362:801–9. doi: 10.1038/362801A0

179. Hinds TD, Sodhi K, Meadows C, Fedorova L, Puri N, Kim DH, et al. Increased HO-1 levels ameliorate fatty liver development through a reduction of heme and recruitment of FGF21. *Obes (Silver Spring Md)*. (2014) 22:705–12. doi: 10.1002/OBY.20559

180. Ishikawa K, Sugawara D, Wang XP, Suzuki K, Itabe H, Maruyama Y, et al. Heme oxygenase-1 inhibits atherosclerotic lesion formation in ldl-receptor knockout mice. Circ Res. (2001) 88:506–12. doi: 10.1161/01.RES.88.5.506

181. Yet SF, Layne MD, Liu X, Chen YH, Ith B, Sibinga NES, et al. Absence of heme oxygenase-1 exacerbates atherosclerotic lesion formation and vascular remodeling. *FASEB journal: Off Publ Fed Am Societies Exp Biol.* (2003) 17:1759–61. doi: 10.1096/FJ.03-0187FJE

182. Boyle JJ, Harrington HA, Piper E, Elderfield K, Stark J, Landis RC, et al. Coronary intraplaque hemorrhage evokes a novel atheroprotective macrophage phenotype. *Am J Pathol.* (2009) 174:1097–108. doi: 10.2353/ajpath.2009.080431

183. Yeudall S, Upchurch CM, Leitinger N. The clinical relevance of heme detoxification by the macrophage heme oxygenase system. *Front Immunol.* (2024) 15:1379967. doi: 10.3389/fimmu.2024.1379967

184. Mushenkova NV, Nikiforov NG, Melnichenko AA, Kalmykov V, Shakhpazyan NK, Orekhova VA, et al. Functional phenotypes of intraplaque macrophages and their distinct roles in atherosclerosis development and atheroinflammation. *Biomedicines*. (2022) 10. doi: 10.3390/biomedicines10020452

185. Pradhan P, Vijayan V, Liu B, Martinez-Delgado B, Matamala N, Nikolin C, et al. Distinct metabolic responses to heme in inflammatory human and mouse macrophages – Role of nitric oxide. *Redox Biol.* (2024) 73. doi: 10.1016/j.redox.2024.103191

186. Chen R, Zhang H, Tang B, Luo Y, Yang Y, Zhong X, et al. Macrophages in cardiovascular diseases: molecular mechanisms and therapeutic targets. *Signal Transduction Targeted Ther.* (2024) 9. doi: 10.1038/s41392-024-01840-1

187. Bories GFP, Leitinger N. Macrophage metabolism in atherosclerosis. *FEBS Lett.* (2017) 591:3042–60. doi: 10.1002/1873-3468.12786

188. Boyle JJ, Johns M, Kampfer T, Nguyen AT, Game L, Schaer DJ, et al. Activating transcription factor 1 directs Mhem atheroprotective macrophages through coordinated iron handling and foam cell protection. *Circ Res.* (2012) 110:20–33. doi: 10.1161/CIRCRESAHA.111.247577

189. Li L, Wang L, Li J, Fan Z, Yang L, Zhang Z, et al. Metformin-induced reduction of CD39 and CD73 blocks myeloid-derived suppressor cell activity in patients with ovarian cancer. *Cancer Res.* (2018) 78:1779–91. doi: 10.1158/0008-5472.CAN-17-2460

190. Boyle JJ. Heme and haemoglobin direct macrophage Mhem phenotype and counter foam cell formation in areas of intraplaque haemorrhage. *Curr Opin Lipidology*. (2012) 23:453–61. doi: 10.1097/MOL.0b013e328356b145

191. Finn AV, Nakano M, Polavarapu R, Karmali V, Saeed O, Zhao XQ, et al. Hemoglobin directs macrophage differentiation and prevents foam cell formation in human atherosclerotic plaques. *J Am Coll Cardiol.* (2012) 59:166–77. doi: 10.1016/j.jacc.2011.10.852

192. Yunoki K, Inoue T, Sugioka K, Nakagawa M, Inaba M, Wada S, et al. Association between hemoglobin scavenger receptor and heme oxygenase-1-related anti-inflammatory mediators in human coronary stable and unstable plaques. *Hum Pathol.* (2013) 44:2256–65. doi: 10.1016/j.humpath.2013.05.008

193. Sakamoto A, Kawakami R, Mori M, Guo L, Paek KH, Mosquera JV, et al. CD163+ macrophages restrain vascular calcification, promoting the development of high-risk plaque. *JCI Insight*. (2023) 8. doi: 10.1172/jci.insight.154922

194. Bengtsson E, Hultman K, Edsfeldt A, Persson A, Nitulescu M, Nilsson J, et al. CD163+ macrophages are associated with a vulnerable plaque phenotype in human carotid plaques. *Sci Rep.* (2020) 10. doi: 10.1038/s41598-020-71110-x

195. Oparil S, Acelajado MC, Bakris GL, Berlowitz DR, Cífková R, Dominiczak AF, et al. Hypertension. *Nat Rev Dis Primers*. (2018) 4:1–21. doi: 10.1038/nrdp.2018.14

196. de Freminville JB, Amar L, Azizi M, Mallart-Riancho J. Endocrine causes of hypertension: literature review and practical approach. *Hypertension Res.* (2023) 46:2679–92. doi: 10.1038/s41440-023-01461-1

197. Rimoldi SF, Scherrer U, Messerli FH. Secondary arterial hypertension: When, who, and how to screen? *Eur Heart J.* (2014) 35. doi: 10.1093/eurheartj/eht534

198. Savoia C, Volpe M. Angiotensin receptor modulation and cardiovascular remodeling. JRAAS - J Renin-Angiotensin-Aldosterone System. (2011) 12:381-4. doi: 10.1177/1470320311417750

199. Martínez-Casales M, Hernanz R, Alonso MJ. Vascular and macrophage heme oxygenase-1 in hypertension: A mini-review. *Front Physiol.* (2021) 12:643435. doi: 10.3389/fphys.2021.643435

200. Norlander AE, Madhur MS, Harrison DG. The immunology of hypertension. J Exp Med. (2018) 215:21–33. doi: 10.1084/jem.20171773

201. Caillon A, Paradis P, Schiffrin EL. Role of immune cells in hypertension. Br J Pharmacol. (2019) 176:1818–28. doi: 10.1111/bph.14427

202. Lopes RA, Neves KB, Tostes RC, Montezano AC, Touyz RM. Downregulation of nuclear factor erythroid 2-related factor and associated antioxidant genes contributes

to redox-sensitive vascular dysfunction in hypertension. *Hypertension*. (2015) 66:1240–50. doi: 10.1161/HYPERTENSIONAHA.115.06163

203. Wenzel P, Rossmann H, Müller C, Kossmann S, Oelze M, Schulz A, et al. Heme oxygenase-1 suppresses a pro-inflammatory phenotype in monocytes and determines endothelial function and arterial hypertension in mice and humans. *Eur Heart J.* (2015) 36:3437–46. doi: 10.1093/EURHEARTJ/EHV544

204. Bellner L, Lebovics NB, Rubinstein R, Buchen YD, Sinatra E, Sinatra G, et al. Heme oxygenase-1 upregulation: A novel approach in the treatment of cardiovascular disease. *Antioxidants Redox Signaling.* (2020) 32:1045-60. doi: 10.1089/ars.2019.7970

205. Minetti M, Mallozzi C, Di Stasi AMM, Pietraforte D. Bilirubin is an effective antioxidant of peroxynitrite-mediated protein oxidation in human blood plasma. *Arch Biochem Biophysics.* (1998) 352:165–74. doi: 10.1006/abbi.1998.0584

206. Kwak JY, Takeshige K, Cheung BS, Minakami S. Bilirubin inhibits the activation of superoxide-producing NADPH oxidase in a neutrophil cell-free system. *Biochim Biophys Acta (BBA)/Protein Structure Mol.* (1991) 1076:369–73. doi: 10.1016/0167-4838(91)90478-I

207. Wang WW, Smith DLH, Zucker SD. Bilirubin inhibits iNOS expression and NO production in response to endotoxin in rats. *Hepatology*. (2004) 40:424–33. doi: 10.1002/hep.20334

208. Wang L, Bautista LE. Serum bilirubin and the risk of hypertension. Int J Epidemiol. (2015) 44:142–52. doi: 10.1093/ije/dyu242

209. Chin HJ, Song YR, Kim HS, Park M, Yoon HJ, Na KY, et al. The bilirubin level is negatively correlated with the incidence of hypertension in normotensive Korean population. *J Korean Med Sci.* (2009) 24. doi: 10.3346/jkms.2009.24.S1.S50

210. Kawamura K, Ishikawa K, Wada Y, Kimura S, Matsumoto H, Kohro T, et al. Bilirubin from heme oxygenase-1 attenuates vascular endothelial activation and dysfunction. *Arteriosclerosis thrombosis Vasc Biol.* (2005) 25:155–60. doi: 10.1161/01.ATV.0000148405.18071.6A

211. Hayashi S, Takamiya R, Yamaguchi T, Matsumoto K, Tojo SJ, Tamatani T, et al. Induction of heme oxygenase-1 suppresses venular leukocyte adhesion elicited by oxidative stress: role of bilirubin generated by the enzyme. *Circ Res.* (1999) 85:663–71. doi: 10.1161/01.RES.85.8.663

212. Wölle J, Ferguson E, Keshava C, Devall LJ, Boschelli DH, Newton RS, et al. Inhibition of tumor necrosis factor induced human aortic endothelial cell adhesion molecule gene expression by an alkoxybenzothiophene-2-carboxamide. *Biochem Biophys Res Commun.* (1995) 214:6–10. doi: 10.1006/BBRC.1995.2249

213. Ryter SW, Ma KC, Choi AMK. Carbon monoxide in lung cell physiology and disease. *Am J Physiol - Cell Physiol*. (2018) 314:C211–27. doi: 10.1152/ ajpcell.00022.2017

214. Ndisang JF, Tabien HEN, Wang R. Carbon monoxide and hypertension. J Hypertension. (2004) 22:1057–74. doi: 10.1097/00004872-200406000-00002

215. Brouard S, Otterbein LE, Anrather J, Tobiasch E, Bach FH, Choi AMK, et al. Carbon monoxide generated by heme oxygenase 1 suppresses endothelial cell apoptosis. *J Exp Med.* (2000) 192:1015–25. doi: 10.1084/jem.192.7.1015

216. Singh N, Ahmad Z, Baid N, Kumar A. Host heme oxygenase-1: Friend or foe in tackling pathogens? *IUBMB Life*. (2018) 70:869–80. doi: 10.1002/iub.1868

217. Kassovska-Bratinova S, Yang G, Igarashi K, Dennery PA. Bach1 modulates heme oxygenase-1 expression in the neonatal mouse lung. *Pediatr Res.* (2009) 65:145–9. doi: 10.1203/PDR.0b013e318191eedc

218. Wilks A. Heme oxygenase: Evolution, structure, and mechanism. *Antioxidants Redox Signaling*. (2002) 4:603–14. doi: 10.1089/15230860260220102

219. Chinta KC, Pacl HT, Agarwal A, Steyn AJC. Heme oxygenase-1 as a pharmacological target for host-directed therapy to limit tuberculosis associated immunopathology. *Antioxidants.* (2021) 10:1–18. doi: 10.3390/antiox10020177

220. Kumar A, Deshane JS, Crossman DK, Bolisetty S, Yan BS, Kramnik I, et al. Heme oxygenase-1-derived carbon monoxide induces the Mycobacterium tuberculosis dormancy regulon. *J Biol Chem.* (2008) 283:18032–9. doi: 10.1074/ jbc.M802274200

221. Shiloh MU, Manzanillo P, Cox JS. Mycobacterium tuberculosis Senses Host-Derived Carbon Monoxide during Macrophage Infection. *Cell Host Microbe.* (2008) 3:323–30. doi: 10.1016/j.chom.2008.03.007

222. Costa DL, Namasivayam S, Amaral EP, Arora K, Chao A, Mittereder LR, et al. Pharmacological inhibition of host heme oxygenase-1 suppresses mycobacterium tuberculosis infection *in vivo* by a mechanism dependent on T lymphocytes. *mBio.* (2016) 7:1–18. doi: 10.1128/mBio.01675-16

223. Costa DL, Amaral EP, Namasivayam S, Mittereder LR, Fisher L, Bonfim CC, et al. Heme oxygenase-1 inhibition promotes IFNY- and NOS2-mediated control of Mycobacterium tuberculosis infection. *Mucosal Immunol.* (2021) 14:253–66. doi: 10.1038/s41385-020-00342-x

224. Chinta KC, Rahman MA, Saini V, Glasgow JN, Reddy VP, Lever JM, et al. Microanatomic distribution of myeloid heme oxygenase-1 protects against free radicalmediated immunopathology in human tuberculosis. *Cell Rep.* (2019) 28:3286. doi: 10.1016/j.celrep.2019.08.081

225. Kovtunovych G, Eckhaus MA, Ghosh MC, Ollivierre-Wilson H, Rouault TA. Dysfunction of the heme recycling system in heme oxygenase 1-deficient mice: Effects on macrophage viability and tissue iron distribution. *Blood*. (2010) 116:6054–62. doi: 10.1182/blood-2010-03-272138

226. Silva-Gomes S, Appelberg R, Larsen R, Soares MP, Gomes MS. Heme catabolism by heme oxygenase-1 confers host resistance to Mycobacterium infection. *Infection Immun.* (2013) 81:2536–45. doi: 10.1128/IAI.00251-13

227. Nakajima M, Matsuyama M, Kawaguchi M, Kiwamoto T, Matsuno Y, Morishima Y, et al. Nrf2 regulates granuloma formation and macrophage activation during mycobacterium avium infection via mediating nramp1 and ho-1 expressions. *mBio.* (2021) 12:1–17. doi: 10.1128/mBio.01947-20

228. Reddy VP, Chinta KC, Saini V, Glasgow JN, Hull TD, Traylor A, et al. Ferritin H deficiency in myeloid compartments dysregulates host energy metabolism and increases susceptibility to Mycobacterium tuberculosis infection. *Front Immunol.* (2018) 9:860. doi: 10.3389/fimmu.2018.00860

229. Pinedo LC, Franz E, van den Beld M, Van Goethem N, Mattheus W, Veldman K, et al. Changing epidemiology of Salmonella Enteritidis human infections in the Netherlands and Belgium, 2006 to 2019: a registry-based population study. *Eurosurveillance*. (2022) 27:2101174. doi: 10.2807/1560-7917.ES.2022.27.38.2101174

230. Zelpina E, Noor SM. Non-thypoid salmonella causes food-borne diseases causing : prevention and control. *Indonesian Bull Anim Veterinary Sci.* (2020) 30:221. doi: 10.14334/wartazoa.v30i4.2194

231. Zaki MH, Fujii S, Okamoto T, Islam S, Khan S, Ahmed KA, et al. Cytoprotective function of heme oxygenase 1 induced by a nitrated cyclic nucleotide formed during murine salmonellosis. *J Immunol.* (2009) 182:3746–56. doi: 10.4049/jimmunol.0803363

232. Mitterstiller AM, Haschka D, Dichtl S, Nairz M, Demetz E, Talasz H, et al. Heme oxygenase 1 controls early innate immune response of macrophages to Salmonella Typhimurium infection. *Cell Microbiol.* (2016) 18:1374–89. doi: 10.1111/cmi.12578

233. de Noordhout CM, Devleesschauwer B, Angulo FJ, Verbeke G, Haagsma J, Kirk M, et al. The global burden of listeriosis: a systematic review and meta-analysis. *Lancet Infect Dis.* (2014) 14:1073-82. doi: 10.1016/S1473-3099(14)70870-9

234. Tzima S, Victoratos P, Kranidioti K, Alexiou M, Kollias G. Myeloid heme oxygenase-1 regulates innate immunity and autoimmunity by modulating IFN- β production. J Exp Med. (2009) 206:1167–79. doi: 10.1084/jem.20081582

235. O'Connell RM, Saha SK, Vaidya SA, Bruhn KW, Miranda GA, Zarnegar B, et al. Type I interferon production enhances susceptibility to Listeria monocytogenes infection. J Exp Med. (2004) 200:437-45. doi: 10.1084/jem.20040712

236. Mattei F, Schiavoni G, Belardelli F, Tough DF. IL-15 is expressed by dendritic cells in response to type I IFN, double-stranded RNA, or lipopolysaccharide and promotes dendritic cell activation. *J Immunol.* (2001) 167:1179–87. doi: 10.4049/jimmunol.167.3.1179

237. Wang Z, Sun D, Chen G, Li G, Dou S, Wang R, et al. Tim-3 inhibits macrophage control of Listeria monocytogenes by inhibiting Nrf2. *Sci Rep.* (2017) 7. doi: 10.1038/srep42095

238. Espinoza JA, León MA, Céspedes PF, Gómez RS, Canedo-Marroquín G, Riquelme SA, et al. Heme oxygenase-1 modulates human respiratory syncytial virus replication and lung pathogenesis during infection. *J Immunol.* (2017) 199:212-23. doi: 10.4049/jimmunol.1601414

239. Lehmann E, El-Tantawy WH, Ocker M, Bartenschlager R, Lohmann V, Hashemolhosseini S, et al. The heme oxygenase 1 product biliverdin interferes with hepatitis C virus replication by increasing antiviral interferon response. *Hepatology*. (2010) 51:398–404. doi: 10.1002/hep.23339

240. Collins DR, Gaiha GD, Walker BD. CD8+ T cells in HIV control, cure and prevention. Nat Rev Immunol. (2020) 20:471-82. doi: 10.1038/s41577-020-0274-9

241. Devadas K, Dhawan S. Hemin activation ameliorates HIV-1 infection via heme oxygenase-1 induction. J Immunol. (2006) 176:4252–7. doi: 10.4049/jimmunol.176.7.4252

242. Liu XM, Durante ZE, Peyton KJ, Durante W. Heme oxygenase-1-derived bilirubin counteracts HIV protease inhibitor-mediated endothelial cell dysfunction. *Free Radical Biol Med.* (2016) 94:218–29. doi: 10.1016/j.freeradbiomed.2016.03.003

243. Sica A, Colombo MP, Trama A, Horn L, Garassino MC, Torri V. Immunometabolic status of covid-19 cancer patients. *Physiol Rev.* (2020) 100:1839–50. doi: 10.1152/physrev.00018.2020

244. Rossi M, Piagnerelli M, Van Meerhaeghe A, Zouaoui Boudjeltia K. Heme oxygenase-1 (HO-1) cytoprotective pathway: A potential treatment strategy against coronavirus disease 2019 (COVID-19)-induced cytokine storm syndrome. *Med Hypotheses.* (2020) 144. doi: 10.1016/j.mehy.2020.110242

245. Kim DH, Ahn HS, Go HJ, Kim DY, Kim JH, Lee JB, et al. Hemin as a novel candidate for treating COVID-19 via heme oxygenase-1 induction. *Sci Rep.* (2021) 11. doi: 10.1038/s41598-021-01054-3

246. Singh D, Wasan H, Reeta KH. Heme oxygenase-1 modulation: A potential therapeutic target for COVID-19 and associated complications. *Free Radical Biol Med.* (2020) 161:263–71. doi: 10.1016/j.freeradbiomed.2020.10.016

247. Detsika MG, Nikitopoulou I, Veroutis D, Vassiliou AG, Jahaj E, Tsipilis S, et al. Increase of HO-1 expression in critically ill COVID-19 patients is associated with poor prognosis and outcome. *Antioxidants*. (2022) 11. doi: 10.3390/antiox11071300

248. Kishimoto Y, Sasaki K, Saita E, Niki H, Ohmori R, Kondo K, et al. Plasma heme oxygenase-1 levels and carotid atherosclerosis. *Stroke*. (2018) 49:2230–2. doi: 10.1161/STROKEAHA.118.022256

249. Grigorov I, Pejić S, Todorović A, Drakulić D, Veljković F, Vukajlović JM, et al. Serum high-mobility group box 1 and heme oxygenase-1 as biomarkers in COVID-19 patients at hospital admission. *Int J Mol Sci.* (2023) 24. doi: 10.3390/ijms241713164

250. Kernan KF, Carcillo JA. Hyperferritinemia and inflammation. Int Immunol. (2017) 29:401-9. doi: 10.1093/intimm/dxx031

251. Moreira AC, Mesquita G, Gomes MS. Ferritin: An inflammatory player keeping iron at the core of pathogen-host interactions. *Microorganisms*. (2020) 8. doi: 10.3390/microorganisms8040589

252. Kimura M, Yamamoto T, Zhang J, Itoh K, Kyo M, Kamiya T, et al. Molecular basis distinguishing the DNA binding profile of Nrf2-Maf heterodimer from that of Maf homodimer. J Biol Chem. (2007) 282:33681–90. doi: 10.1074/jbc.M706863200

253. Panchenko MV, Farber HW, Korn JH. Induction of heme oxygenase-1 by hypoxia and free radicals in human dermal fibroblasts. *Am J Physiol - Cell Physiol.* (2000) 278:C92–C101. doi: 10.1152/ajpcell.2000.278.1.c92

254. Nakamichi I, Habtezion A, Zhong B, Contag CH, Butcher EC, Omary MB. Hemin-activated macrophages home to the pancreas and protect from acute pancreatitis via heme oxygenase-1 induction. *J Clin Invest.* (2005) 115:3007–14. doi: 10.1172/JCI24912

255. Paul G, Bataille F, Obermeier F, Bock J, Klebl F, Strauch U, et al. Analysis of intestinal haem-oxygenase-1 (HO-1) in clinical and experimental colitis. *Clin Exp Immunol.* (2005) 140:547–55. doi: 10.1111/j.1365-2249.2005.02775.x

256. Piel FB, Hay SI, Gupta S, Weatherall DJ, Williams TN. Global burden of sickle cell anaemia in children under five, 2010-2050: modelling based on demographics, excess mortality, and interventions. *PloS Med.* (2013) 10. doi: 10.1371/journal.pmed.1001484

257. Williams TN, Uyoga S, Macharia A, Ndila C, McAuley CF, Opi DH, et al. Bacteraemia in Kenyan children with sickle-cell anaemia: a retrospective cohort and case-control study. *Lancet.* (2009) 374:1364–70. doi: 10.1016/S0140-6736(09)61374-X