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Stem-like CD8⁺ T cells in cancer

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Stem-like CD8⁺ T cells (T_{SL}) are a subset of immune cells with superior persistence and antitumor immunity. They are TCF1⁺ PD-1⁺ and important for the expansion of tumor specific CD8⁺ T cells in response to checkpoint blockade immunotherapy. In acute infections, naïve CD8⁺ T cells differentiate into effector and memory CD8⁺ T cells; in cancer and chronic infections, persistent antigen stimulation can lead to T cell exhaustion. Recent studies have highlighted the dichotomy between late dysfunctional (or exhausted) T cells (T_{LD}) that are TCF1⁻ PD-1⁺ and self-renewing TCF1⁺ PD-1⁺ T_{SL} from which they derive. TCF1⁺ T_{SL} cells are considered to have stem cell-like properties akin to memory T cell populations and can give rise to cytotoxic effector and transitory T cell phenotypes (T_{TE}) which mediate tumor control. In this review, we will discuss recent advances made in research on the formation and expansion of T_{SL}, as well as distinct niches required for their differentiation and maintenance in the setting of cancer. We will also discuss potential strategies to generate these cells, with clinical implications for stemness enhancement in vaccine design, immune checkpoint blockade (ICB), and adoptive T cell therapies.

KEYWORDS

stem-like CD8 T cells (T_{SL}), chronic viral infection, cancer models, immune, tertiary lymphoid structure (TLS), tumor microenvironment (TME)

1 Introduction

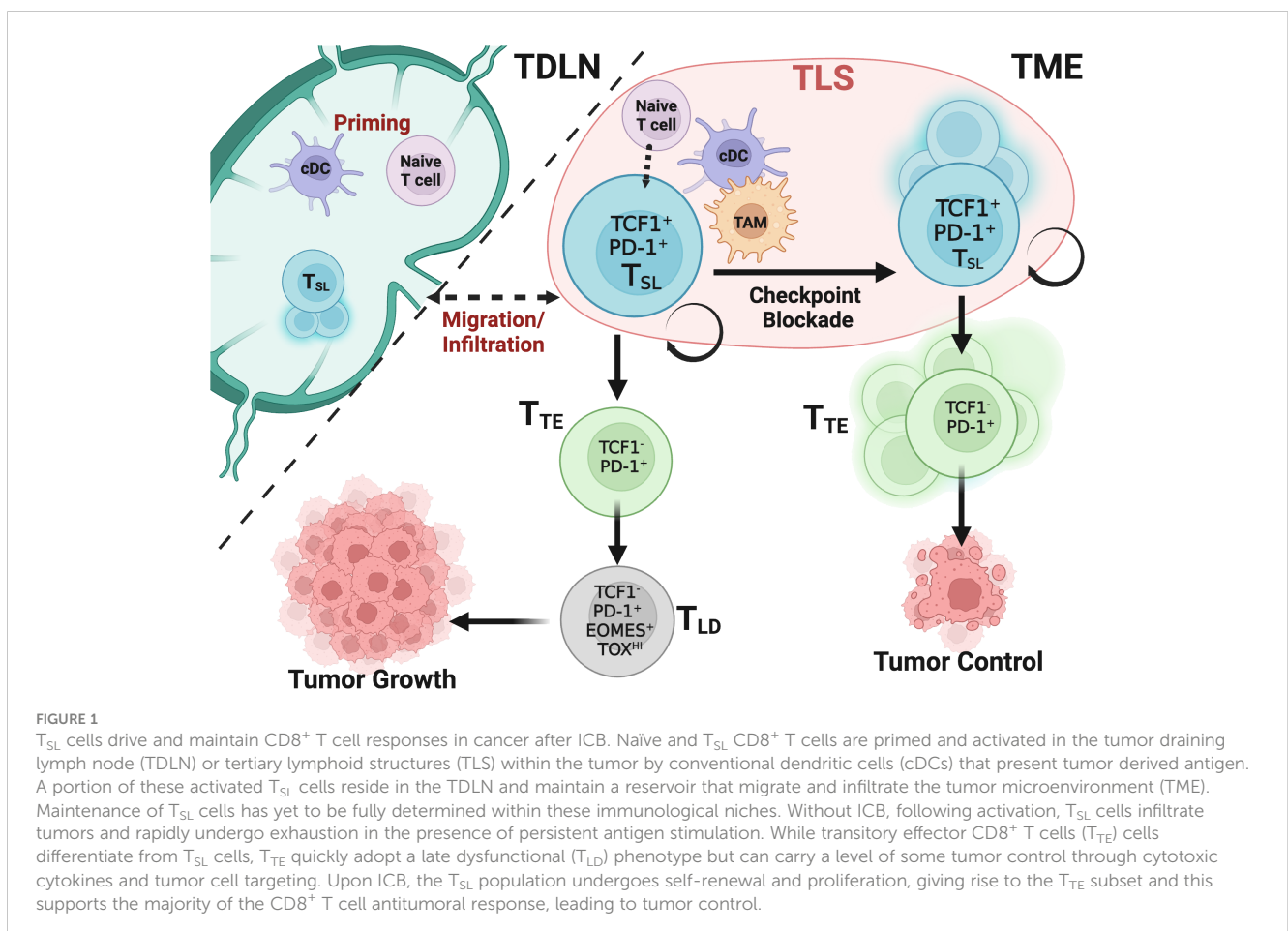
Immune checkpoint blockade (ICB) therapy has generated impressive success in recent years as 15~30% of cancer patients treated with ICB experience durable remissions (1). It has been proposed that ICB can reverse exhausted or late dysfunctional CD8⁺ T cells (T_{LD}) to an effector-like state. However, recent studies have shown T_{LD} cells have a terminally differentiated phenotype and may not be readily rescued. Rather, proliferative bursts of a relatively undifferentiated population of “stem-like” T cells (T_{SL}) occur after ICB, which has been correlated with clinical benefit. These T_{SL} are identified by their expression of transcription factor T cell factor-1 (TCF1), along with intermediate expression of inhibitory receptor, programmed cell death protein-1 (PD-1). TCF1⁺ PD-1⁺ T_{SL} cells have the ability to expand, self-renew, and differentiate into transitory effector-like T cells (T_{TE}) and T_{LD} cells. T_{SL} cells have been identified to play a vital role in sustaining the CD8⁺ T cell response in both chronic infection and cancer. Their presence is associated with

positive clinical outcomes of checkpoint immunotherapies in patients with melanoma, colorectal, and non-small cell lung cancer (NSCLC) (2–4). Here we will review the latest developments regarding T_{SL} population formation and expansion, along with the specific niches required for their maintenance and differentiation in the context of cancer. We will also explore potential approaches to produce T_{SL} cells and discuss the therapeutic implications of enhancing stemness in adoptive T cell therapies, ICB, and vaccine design.

2 Formation, expansion, and hallmarks of stem-like $CD8^+$ T cells

Stem-like $CD8^+$ T cells have emerged as key players in response to ICB, as a subset of cells that retain stemness, have memory potential, and a high proliferative capacity. Targeting the PD-1:PD-L1 pathway with ICB treatment drives the expansion of these cells. This was first observed in chronic infection models (5–8) and subsequently in mouse and human cancers (2–4, 9, 10). As shown in Figure 1, the proliferation burst encompasses not only expansion of T_{SL} 's cells' downstream T_{TE} progeny, but also self-renewal of the T_{SL} population. T_{SL} self-propagate an epigenetically distinct, stable pool of T_{SL} cells that persists during active disease. This population is armed for subsequent proliferative bursts and fuels a downstream

differentiated effector population in an antigen-dependent manner. T_{SL} cells survive and persist following antigen withdrawal, similar to conventional memory cells. Additionally, they can mount a recall response and continue to produce terminally differentiated progeny (11, 12). Although this subset is more proliferative than other differentiated exhausted subsets, compared to conventional memory cells, T_{SL} have reduced proliferative capacity and cytokine function (13). T_{SL} cells do share many markers with memory and naïve T cells (Figure 2; Table 1). Markers such as CD62L and CD27 are more commonly expressed on naïve and memory populations, while CCR7 and CD28 are often expressed by both naïve and T_{SL} cells. They are also induced/maintained by some similar transcription factors (TFs) including TCF1, BCL6, FOXO1, STAT3, JUN, MYB, BACH2, EOMES, TOX and ID3 (5–7, 14, 15). However, while T_{SL} cells share many memory and stem-like features, they are committed to the exhaustion lineage, and transfer an exhausted phenotype to their progeny (16). While ICB treatment results in the expansion or proliferative bursts of this stem-like population, these cells and their effector progeny show distinct epigenetic features and metabolic state of exhausted T cells (17–19). Studies have observed that although commitment toward the T cell exhaustion phenotype begins as early as 5 days, it requires time for the epigenetic imprint to stabilize where it cannot be overcome by ICB (16, 20–22). The TF nuclear factor of activated T cells (NFAT) plays a pivotal role in effector and exhaustion responses of $CD8^+$ T cells and induces the effector



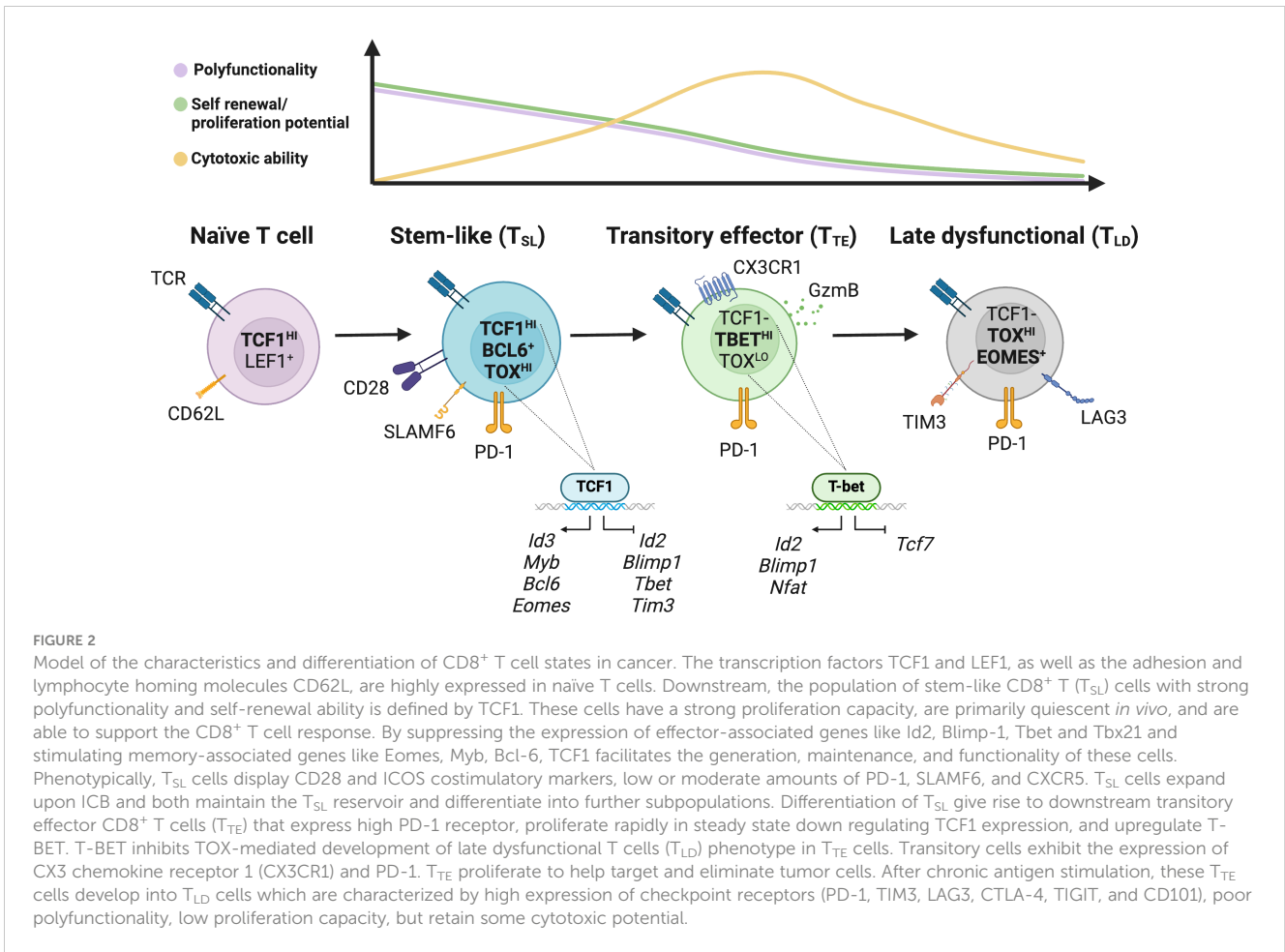


TABLE 1 Summary of the transcription factors, biomarkers, and key features that define CD8⁺ T cell subsets in cancer.

Murine	Human	Both	Naïve T cell	T Stem-like (T _{SL})	Transitory Effector (T _{TE})	Late Dysfunctional (T _{LD})
Blue	Red	Black				
Transcription Factors			TCF1, LEF1	TCF1, LEF1, EOMES, TOX, MYB, FOXO1, JUN, STAT3 <i>ID3, BCL6, BACH2, EGR2</i>	TBET, BLIMP1, BATF, IRF4, ID2, NFAT, RUNX3, NR4A	EOMES, TOX, BATF, NFAT
Biomarkers			CD62L CCR7 CD28 CD27 <i>CD45RA</i> <i>CD45</i>	TCF1 PD-1 LY108/SLAMF6 CXCR5 CD28 ICOS CCR7 CD69 <i>CD45RO</i>	PD-1 GZMB TBET LAG3 CX3CR1 <i>CD45RO</i>	PD-1 TIM3 LAG3 TIGIT CD101 CTLA4 CX3CR1 <i>CD45RO</i>
Key Features			Immature cell Circulate in lymph and blood Feeds downstream subsets	Self-renewal Expands/proliferate after ICB Persistent population pool Feeds downstream effector subsets	Effector/cytotoxic killing to control tumor growth	Increased expression of inhibitory receptors Limited killing capacity and proliferation

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program with its associate TF activator protein 1 (AP-1) and its subunits JUN/FOS (23). In the absence of AP-1, NFAT induces a program of negative feedback leading to T cell exhaustion. Downstream targets of NFAT: TOX, NR4A1, NR4A2 are critical in enforcing T cell exhaustion in T_{SL} cells (24–27). Absence of TOX results in the loss of the T_{SL} population and loss over time in their effector progeny in chronic infection and tumor models (5, 7, 25, 26). Likewise, a recent study reported double deletion of NR4A1/NR4A2 in $CD8^+$ tumor-infiltrating lymphocytes (TILs) resulted in murine tumor eradication after transfer as well as expansion of T_{SL} population with increased chromatin accessibility of several stem-like/memory-related genes (28). T_{SL} cells, however, do not express other co-inhibitory, exhausted T cell markers (TIM3, TIGIT, CTLA4) but do express low to intermediate levels of PD-1, not as a marker of exhaustion but rather activation (29). PD-1 has been shown to help preserve the co-expressing PD-1⁺ TCF1⁺ T_{SL} population by attenuating TCR and co-stimulatory CD28, and by repressing downstream effector differentiation (22, 30, 31). T_{SL} also express other markers such as inducible T cell costimulator (ICOS) molecule, CD28, CXCR5, SLAMF6 (also known as LY108), which denote a population of cells that have experienced antigen and require lymphoid homing (3–6, 14, 32). In chronic viral infection, T_{SL} infiltrate B cell follicles correlating with CXCR5 expression on T_{SL} whereas in tumors, SLAMF6 is highly expressed and positively correlates with TCF1 levels (2, 4, 9, 33).

Another critical feature of T_{SL} cells is the uniform expression of TCF1, encoded by the *Tcf7* gene, which is essential for the formation and function of this population (3, 5–7, 10, 14). Originally identified as a TF essential for thymocyte development, both TCF1 and its homologue LEF1, are now known to promote memory T cell differentiation and inhibit effector differentiation (34, 35). Open chromatin sites in T_{SL} cells are highly enriched in the TCF/LEF motif, similar to naïve T cells, and overlap frequently with TCF1 binding peaks, suggesting direct regulation by TCF1 (20, 35, 36). Studies in chronic infection and tumor models have shown that loss of TCF1 in $CD8^+$ T cells limits their maintenance, function, and overall response to ICB, but does not diminish their overall function (3, 7, 10, 37). Additionally, a preclinical tumor study showing ectopic expression of TCF1 skews TILs to adopt a T_{SL} phenotype while enhancing their polyfunctionality and further suppressing inhibitory receptors and modulating the transcriptome to further suppress TFs like BLIMP1, RUNX3, and TOX to improve viral and tumor control (38). A recent study disputes that tumor immunogenicity dictates reliance on TCF1 for ICB efficacy (39). However, antitumor responses in poorly immunogenic tumors can be improved by optimizing T cell priming through either vaccination or enhancing antigen presentation on tumors (39). Additionally, frequency of *Tcf7*-expressing $CD8^+$ T cells in melanoma can correlate to positive response to ICB, whereas in advanced clear cell renal carcinoma patients, it failed to predict any clinical outcomes (40, 41). How TCF1 directly aids in forming and expanding this crucial stem-like population within its environment is still debated.

Together, the key features that define the formation and expansion of T_{SL} cells encompass multiple regulatory pathways. Many of the features of T_{SL} are similar to other well defined T cell

subsets, therefore it is crucial to establish how regulatory mechanisms operate uniquely in the T_{SL} population in a variety of environments. We have described how T_{SL} cells self-renew while maintaining an exhausted lineage; next we will delve into how this subset continues to feed into the pool of $CD8^+$ T cells and help sustain responses to ICB.

3 Differentiation and maintenance of stem-like $CD8^+$ T cells

Studies from chronic viral infection and tumor models have characterized two populations of epigenetically and spatially distinct populations of $CD8^+$ T cells: TCF1⁺ PD-1⁺ TIM3⁻ $CD8^+$ T_{SL} and their progeny, TCF1⁻ PD-1⁺ $CD8^+$ T transitory effector-like $CD8^+$ T cells (T_{TE}) (3–7, 9, 10). The T_{TE} cells become terminally differentiated, late dysfunctional TCF1^{low/-} PD-1⁺ TIM3⁺ T cells (T_{LD}) that carry distinct transcriptional and epigenetic programs that differ from those seen in traditional memory and effector populations, both in cancer and chronic viral infection (Figures 1, 2; Table 1) (8, 19, 22). It has been shown that T_{SL} drive the proliferative response after immunotherapy and are often associated with clinical benefit, while T_{LD} populations have limited survival and re-expansion potential (3–7, 10, 42). T_{SL} cells and their progeny are committed to an exhausted phenotype, however a unique feature of the T_{SL} population being its ability to be stimulated to expand by ICB, whereas T_{LD} cells cannot be reinvigorated (5, 7, 8, 16, 43). On the other hand, the majority of the tumor specific population exhibits a T_{LD} phenotype, which may indicate a continuous immune response that requires a precursor population generating and infiltrating from external locations (37, 44–49).

The generation and maintenance of T_{SL} cells may be significantly impacted by varying environmental cues. In chronic infection, most T_{SL} cells are located within B cell follicles and the T cell zone of the spleen while their progeny exist within the red pulp taking up residency rather than migration (6, 14, 50). Contrastingly in tumors, T_{SL} cells migrate between perivascular niches or tertiary lymphoid structures (TLS) within the tumor and reservoirs in the tumor-draining lymph node (TDLN) (Figure 1) (3, 9, 32, 51–61). Blocking migration using sphingosine 1-phosphate receptor 1 (S1P1)-agonist FTY720 in multiple preclinical tumor models prevented tumor regression and challenged the understanding that anti-PD-1 immunotherapy primarily targets intratumoral T cells. This also suggests that T_{SL} migration to TDLN may even be required for T_{SL} cell maintenance (51, 52, 55, 60, 61). These specific tissue niches likely have two purposes for maintaining T_{SL} cells: to sequester away this population from inflammatory cues that quickly drive differentiation into exhausted phenotypes and to provide close, tightly regulated contact with antigen-presenting cells (APCs) such as dendritic cells (DCs) (60, 62). Recent preclinical research also implies that molecularly distinct lymph-node resident $CD8^+$ memory-like and T_{SL} cells are sole mediators of ICB (61, 63). Two additional recent studies in non-small cell lung cancer (NSCLC) and head and neck squamous cell carcinoma (HNSCC)

respectively, also support the idea that T_{SL} cells respond to immunotherapy within the lymph nodes (64, 65). Clusters of T_{SL} populations and APCs are also linked to significant T cell infiltration in human malignancies, whereas their absence may lead to immune evasion (9, 34). While T_{SL} cells are clustered with APCs and even $CD4^+$ T cells within TLSs creating a supportive network to promote effective differentiation into T_{TE} subsets, T_{LD} cells are more scattered throughout the tumor parenchyma where they can readily engage with target cells (57, 66–68). Evidence also suggests that immunotherapy responses in sarcoma, melanoma and renal cell carcinoma are favorably linked with TLSs containing B cells and T_{SL} cells (33, 69, 70). In tumors that possess obstacles preventing the infiltration of T cells, such as solid tumors, immune cell niches can persist and harbor concentrated populations of T_{SL} cells that are aggregated with APCs (71). Cells in such niches were able to rapidly regenerate the immune response in patients with brain metastases and these immune niches were prognostic for local disease control (71). Thus, it is likely the interactions of T_{SL} cells with DCs and B cells within these niches are influential in the maintenance and function of T_{SL} and are required for durable $CD8^+$ T cell responses. As previously mentioned, epigenetic analysis of T_{SL} in chronic infection compared to T_{LD} revealed unique open chromatin sites, and T_{SL} subsets show increased accessibility to XCL1 which is involved in the interactions between DCs and T cells (3, 4, 36, 72). XCL1 expressed on T cells promotes the recruitment of $XCR1^+$ conventional type 1 DCs (cDC1) which have superior antigen processing and cross-presentation capabilities (73). Several groups have highlighted the necessity of cDC1s in sustaining T_{SL} cells and inducing the proliferative burst after ICB within the TLS as well as in maintaining the TDLN T_{SL} reservoir in preclinical tumor and chronic infection models, and patient samples (60, 62, 74). Additionally, the B7/CD28 pathway, expressed on DCs and T cells respectively, may have a role in structuring how these interactions sustain the immune response as T_{SL} have high CD28 expression that is necessary for the proliferative burst after ICB (75, 76). By blocking B7 costimulatory molecule on APCs or deletion of CD28 on T cells, effective responses to PD-1/PD-L1 ICB were diminished (76).

Another environmental cue being investigated is the CXCR3 pathway as a significant axis of immunotherapy response that regulates the infiltration and spatial positioning of T cells near APCs expressing the ligands CXCL9/10/11 within the murine and human tumor microenvironment (TME) (54, 77, 78). As multiple myeloid populations within the TME express the ligand CXCL9, including both DCs and tumor-associated macrophages (TAMs), and these chemokines are broadly induced in response to treatment, it remains another avenue to investigate in the maintenance of T_{SL} within TLSs (79–82). In the TME, macrophages are more abundant and express higher levels of CXCL9 than compared to DCs and may play a more prominent role in the TME compared to DCs in the TDLN (81).

Differentiation of T_{SL} into their cytolytic progeny T_{TE} cells has proved vital to the efficacy of ICB. The maintenance of this population via the TDLN reservoir or in TLSs within the tumor additionally have gained recognition in contributing to improved clinical outcomes. Many of the networks and signaling pathways

involved in these environments will likely aid in determining future successes of therapeutics.

4 Therapeutic potential of stem-like $CD8^+$ T cells in cancer

4.1 Immune checkpoint blockade

ICB therapy against inhibitory receptors PD-1 and CTLA4 of TILs has shown success in mounting a T cell response against tumors in many cancer types. Efficacy is highest in tumors with more mutational burden and typically higher TIL infiltration suggesting leveraging an already present immune response. Prior to the role of T_{SL} , it was thought that T_{LD} being “rescued” from their late dysfunctional phenotype to a less exhausted, more effector phenotype was the primary mechanism of ICB (83). Some clinical studies have shown an abundance of cells with a T_{TE} or T_{LD} phenotype rather than T_{SL} cells can provide a better predictor for response to ICB (84–88). While $TCF1^+$ expression by TILs in human melanoma coincides with clinical benefit of ICB, $TCF1$ is produced also by bystander TILs which are less relevant for antitumor response. High frequencies of $TCF1^+$ $PD-1^+$ T_{SL} thus may be an unreliable biomarker as a portion of these cells are not tumor-specific (40, 89). Likely, the ratio of T_{SL} to more differentiated TILs may represent a more suitable biomarker for outcome prediction as T_{SL} frequencies are comparable to those observed in responders versus non-responders (4, 40). In chronic infection and tumor models, T_{SL} have been shown to be critical in amplifying the response to ICB by self-renewal, expansion, and differentiation into T_{TE} , supplying the pool of cytotoxic cells and mediating disease control (90). Given their crucial role in ICB, it is imperative to effectively control T_{SL} cells. Continuous driving of differentiation by immune checkpoints can negatively impact maintenance of T_{SL} cells and ultimately result in loss of the ability to expand and differentiate, driving patients toward a refractory state (22, 91, 92). Bi-specific antibody therapy has shown promising outcomes in patients with hematologic malignancies, although in cancers more resistant to ICB and favorable outcomes are limited. One drug construct uses an anti-PD-1 molecule as a targeting moiety fused to a stimulatory IL-2 variant (IL-2v) to deliver IL-2 to $PD-1^+$ T cells in the TME. Combining with anti-PD-L1 treatment resulted in murine tumor regression, enhanced infiltration of the T_{SL} population, and reprogramming of TAMs (93). It is important to note, prolonged exposure of T cells to bispecifics through continuous infusion can also cause cells to adopt the T_{LD} phenotype and therefore must be carefully evaluated (93, 94). Other therapeutic strategies taken to clinical trial include inhibiting cell division, T cell receptor (TCR) signaling, or epigenetic pathways to hinder T_{SL} differentiation (18, 19, 95–97). Additionally, depleting or altering T cell signaling pathways in T_{SL} cells have shown to promote stem-like phenotype retention, allowing these cells to persist in harsh environments that would otherwise push these populations towards T_{LD} phenotype, and instead still produce effective T_{TE}

progeny (98–100). Clinical data has also shown that ICB therapy induced expansion of pre-treatment T_{SL} cells present in patients who were responders compared to non-responders which had more pre-treatment T_{LD} phenotypes, experienced therapy resistance (10, 40, 48, 49, 59, 90).

Quantity or presence of T_{SL} alone may be insufficient as a marker of response, because as previously mentioned, APC-dense niches or TLSs tolerant for T_{SL} self-renewal or expansion, may additionally be required for effective responses. Clinical observations have revealed that tumors with such regions correlate with favorable therapeutic responses (51, 59, 71, 87, 88). Additionally in other preclinical studies it has been observed that blocking T cell egress from TDLN, surgically removing the TDLN, or disrupting the migration of T cells from the TME diminishes ICB response (51–54, 61). Further new studies from patient samples of NSCLC, HNSCC, and melanoma also indicate T_{SL} cells respond to ICB directly in the TDLN, displaying local clonal expansion and subsequent migration of these new clones to the TME (44, 64, 65, 83, 90). Therefore, targeting the establishment and cultivation of these regions within the TME or TDLN, to enhance T_{SL} maintenance and differentiation, could further increase efficacy (101, 102).

4.2 Adoptive cellular therapy

This therapy encompasses two main approaches: ex vivo expansion of TILs or genetic modification of peripheral blood mononuclear cells (PBMC)-derived T cells for tumor specific subsets and subsequent reintroduction into the patient. Ex vivo manufacturing and expansion strategies to induce T_{SL} cells include introducing IL-7, IL-15, and IL-21 to promote expression of associated genes like *TCF7*, *Eomes*, and *Bcl6* (103–107), or promoting Notch signaling upstream of *TCF1* (108, 109). Suppressing genes associated with late dysfunctional or exhaustive phenotypes such as *Tbet*, *BATF*, *EOMES* pharmacologically ex vivo can maintain stem-like genes (*TCF1/LEF1*) and retains T_{SL} cell polyfunctionality (110, 111). Numerous studies of both preclinical models and patients of ACT observe that less differentiated, memory and stem-like cells elicit more of an effective antitumoral response (112–118). Genetically engineering T cells using retroviral transduction to incorporate a tumor reactive TCR or a chimeric antigen receptor (CAR-T) has become standard of care for many hematologic malignancies (119–125). Increased populations of terminally exhausted CD8⁺ CAR-T cells present in pre-treatment product correlate with worse outcomes however, presence of more naïve and memory-like CAR-T phenotypes are correlated with increased response rates (126–128). Although extensive clinical research into T_{SL} phenotypes in CAR-T products has yet to be conducted, one recent study identified that PD-1⁺ TCF1⁺ stem-like CAR-T and PD-1⁺ TIM3⁺ effector-like CAR-T correlated with improved clinical outcomes (129). This study highlights the importance of PD-1 expression on CAR-T cells post-infusion as a marker of activation rather than exhaustion for optimal activation as well as the potential for optimizing stem-like phenotypes in CAR-T subsets to potentially improve clinical outcomes.

Study of the epigenetic landscape of T_{SL}, T_{TE} and T_{LD} subsets has revealed several targets for controlling the differentiation and antitumor response and are now in preclinical CAR-T models (130–133). Exploration of the chromatin accessibility of CAR-T cells at the single cell level, both *in vitro* and *in vivo*, identified two distinct subsets (133). The subsets consisted of intermediate exhausted CAR-T cells enriched for TFs of T_{SL} cells (*JUN/FOS*) and another with enriched motifs of *BATF* and *IRF4* resembling terminally exhausted or the T_{LD} CAR-T subset. CAR-T cells with knockdown of *BATF*, *IRF4* or *NR4A* expression had enhanced effector function, inhibited exhaustion and prolonged CAR-T cell persistence *in vivo* (133, 134). A dual knockout of genes *PRDM1* (encoding *BLIMP1* TF) and *NR4A* in preclinical murine models, skewed CAR-T cell phenotypes toward T_{SL} subsets and away from T_{LD}, improving antitumor responses and not achieved by single knockouts (132).

Additionally, several preclinical CAR-T models targeting overexpression of TFs specific for T_{SL} such as *c-Jun* and *FOXO1*, promote stem-like phenotypes, enhanced expansion potential, persistence and therapeutic efficacy *in vivo* (130, 131). Other factors such as hub transcription factors, like *FOXP1* and *KLF2* that have high numbers of enhancers that are positioned in the center of gene regulatory networks, can serve as checkpoints that control lineage-defining TFs between stem-like and effector CAR-T, and the decision between effector and late dysfunctional CAR-T cells, respectively (135). While harnessing the power of T_{SL} cell phenotype in CAR-T therapy by targeting key transcriptional regulators may lead to further successful trials, investigating the relationships of other immune cells or combination therapy in altering other environmental cues could be crucial to their advancement.

Pre-existing TLS or APC-dense niches may also be required for generating stem-like CAR-T phenotypes and catering to the cultivation of these environments may also increase their persistence (63, 136, 137). Utilizing stem-like CD8⁺ T cells and their respective molecular determinants as biomarkers of response to CAR-T may also prove beneficial within a clinical setting.

Cancer immunotherapy such as ICB and CAR-T rely on T cell infiltration. The accumulated evidence above shows that combining multiple therapeutic agents is crucial for cancer immunotherapy and targeting stem-like CD8⁺ T cells requires more than one approach.

4.3 Cancer vaccination

Studies in the forefront of cancer vaccination are seeking to harness the self-renewal, long lasting durability, and sustainability of the T_{SL} subset by targeting common tumor antigens or patient specific neoantigens (neoAg) (138–140). Recent advances in genomic sequencing have led to personalized cancer vaccines targeting neoAg. Early studies show feasibility in mice and clinical trials, but neoAg targeted CD8⁺ T cell responses have been limited (139–145). Coupling self-assembling nanoparticle vaccine platform technology, exploiting its ability to quickly drain via lymphatics to DCs and enhance antigen presentation to CD8s, the SNP-7/8a intravenous vaccination generated more T_{SL} cells that

are receptive to ICB in a therapeutic murine model (146). Additionally, adenovirus (Ad)-vectored vaccines encoding tumor neoAg combined with ICB have been shown to eradicate large tumors and increases in T_{SL} cells in the TDLN and T_{TE} cells within the TME in mice and have translated into similar results within the clinic (147). Further, studies harnessing not only T_{SL} cells but also other tumor targeting progenitors, like stem-like natural killer (NK) cells are gaining interest. Introduced at the contraction phase after immunization with an artificial adjuvant vector cell (aAVC), an IL-2/anti-IL-2 monoclonal antibody complex (IL-2Cx) combination activated stem-like subsets that correlated with therapeutic responses, and induced long-term memory CD8⁺ T cells that conferred protection against tumor rechallenge in a leukemic model (148). While tumor vaccine trial successes have been mixed, expanding the population of tumor specific T_{SL} cells is likely the key consideration for the future of favorable tumor vaccine outcomes.

5 Conclusions and outlook

The role of stem-like T cells has been underscored in recent studies, highlighting their potential to improve the antitumor effect of immunotherapies. However, to fully exploit this potential, a complete understanding of how T_{SL} cells form, maintain, and function is necessary. Recent advances in deciphering this subsets' key characteristics and hallmarks have led even further to questions that require investigation. The most vital questions and potential targets will likely center around T_{SL} and APC interactions within their relevant niches in a variety of models. The targeting and harnessing of T_{SL} cells will require multiple points of application.

In conclusion, while significant strides have been made in understanding the role and potential of T_{SL} cells in cancer therapy, there is still much work to be done. Future research should focus on elucidating the regulatory circuits that control these cells, understanding the APC interactions with intratumoral T_{SL} cells and within niches, and developing methods for T_{SL} cell generation. These efforts will be crucial in harnessing T_{SL} cells for therapeutic interventions and enhancing immunotherapy against cancer. The exploration of combination therapies and strategies to

maintain the “stemness” of T cells represent promising avenues for future research and could revolutionize cancer treatment.

Author contributions

CS: Conceptualization, Writing – original draft. ND: Writing – review & editing. XH: Conceptualization, Supervision, Writing – review & editing. YY: Conceptualization, Funding acquisition, Supervision, Writing – review & editing.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

- Sharma P, Allison JP. Dissecting the mechanisms of immune checkpoint therapy. *Nat Rev Immunol*. (2020) 20:75–6. doi: 10.1038/s41577-020-0275-8
- Brummelman J, Mazza EMC, Alvisi G, Colombo FS, Grilli A, Mikulak J, et al. High-dimensional single cell analysis identifies stem-like cytotoxic CD8(+) T cells infiltrating human tumors. *J Exp Med*. (2018) 215:2520–35. doi: 10.1084/jem.20180684
- Siddiqui I, Schaeuble K, Chennupati V, Fuertes Marraco SA, Calderon-Copete S, Pais Ferreira D, et al. Intratumoral Tcf1(+)/PD-1(+)/CD8(+) T cells with stem-like properties promote tumor control in response to vaccination and checkpoint blockade immunotherapy. *Immunity*. (2019) 50:195–211 e110. doi: 10.1016/j.immuni.2018.12.021
- Miller BC, Sen DR, Al Abosy R, Bi K, Virkud YV, LaFleur MW, et al. Subsets of exhausted CD8(+) T cells differentially mediate tumor control and respond to checkpoint blockade. *Nat Immunol*. (2019) 20:326–36. doi: 10.1038/s41590-019-0312-6
- Im SJ, Hashimoto M, Gerner MY, Lee J, Kissick HT, Burger MC, et al. Defining CD8+ T cells that provide the proliferative burst after PD-1 therapy. *Nature*. (2016) 537:417–21. doi: 10.1038/nature19330
- He R, Hou S, Liu C, Zhang A, Bai Q, Han M, et al. Follicular CXCR5- expressing CD8(+) T cells curtail chronic viral infection. *Nature*. (2016) 537:412–28. doi: 10.1038/nature19317
- Utzschneider DT, Chormoy M, Chennupati V, Pousse L, Ferreira DP, Calderon-Copete S, et al. T cell factor 1-expressing memory-like CD8(+) T cells sustain the immune response to chronic viral infections. *Immunity*. (2016) 45:415–27. doi: 10.1016/j.immuni.2016.07.021
- Sen DR, Kaminski J, Barnitz RA, Kurachi M, Gerdemann U, Yates KB, et al. The epigenetic landscape of T cell exhaustion. *Science*. (2016) 354:1165–9. doi: 10.1126/science.aae0491
- Jansen CS, Prokhnevska N, Master VA, Sanda MG, Carlisle JW, Bilen MA, et al. An intra-tumoral niche maintains and differentiates stem-like CD8 T cells. *Nature*. (2019) 576:465–70. doi: 10.1038/s41586-019-1836-5
- Kurtulus S, Madi A, Escobar G, Klapholz M, Nyman J, Christian E, et al. Checkpoint blockade immunotherapy induces dynamic changes in PD-1(-)/CD8(+)

- tumor-infiltrating T cells. *Immunity*. (2019) 50:181–194 e186. doi: 10.1016/j.immuni.2018.11.014
11. Wieland D, Kemming J, Schuch A, Emmerich F, Knolle P, Neumann-Haefelin C, et al. TCF1(+) hepatitis C virus-specific CD8(+) T cells are maintained after cessation of chronic antigen stimulation. *Nat Commun*. (2017) 8:15050. doi: 10.1038/ncomms15050
 12. Tonnerre P, Wolski D, Subudhi S, Aljabban J, Hoogveen RC, Damasio M, et al. Differentiation of exhausted CD8(+) T cells after termination of chronic antigen stimulation stops short of achieving functional T cell memory. *Nat Immunol*. (2021) 22:1030–41. doi: 10.1038/s41590-021-00982-6
 13. Beltra JC, Manne S, Abdel-Hakeem MS, Kurachi M, Giles JR, Chen Z, et al. Developmental relationships of four exhausted CD8(+) T cell subsets reveals underlying transcriptional and epigenetic landscape control mechanisms. *Immunity*. (2020) 52:825–841 e828. doi: 10.1016/j.immuni.2020.04.014
 14. Leong YA, Chen Y, Ong HS, Wu D, Man K, Deleage C, et al. CXCR5(+) follicular cytotoxic T cells control viral infection in B cell follicles. *Nat Immunol*. (2016) 17:1187–96. doi: 10.1038/ni.3543
 15. Petrovas C, Ferrando-Martinez S, Gerner MY, Casazza JP, Pegu A, Deleage C, et al. Follicular CD8 T cells accumulate in HIV infection and can kill infected cells *in vitro* via bispecific antibodies. *Sci Transl Med*. (2017) 9. doi: 10.1126/scitranslmed.aag2285
 16. Pauken KE, Sammons MA, Odorizzi PM, Manne S, Godec J, Khan O, et al. Epigenetic stability of exhausted T cells limits durability of reinvigoration by PD-1 blockade. *Science*. (2016) 354:1160–5. doi: 10.1126/science.aaf2807
 17. Guo Y, Xie YQ, Gao M, Zhao Y, Franco F, Wenes M, et al. Metabolic reprogramming of terminally exhausted CD8(+) T cells by IL-10 enhances anti-tumor immunity. *Nat Immunol*. (2021) 22:746–56. doi: 10.1038/s41590-021-00940-2
 18. Gabriel SS, Tsui C, Chisanga D, Weber F, Llano-Leon M, Gubser PM, et al. Transforming growth factor-beta-regulated mTOR activity preserves cellular metabolism to maintain long-term T cell responses in chronic infection. *Immunity*. (2021) 54:1698–1714 e1695. doi: 10.1016/j.immuni.2021.06.007
 19. Ghoneim HE, Fan Y, Moustaki A, Abdelsamed HA, Dash P, Dogra P, et al. *De novo* epigenetic programs inhibit PD-1 blockade-mediated T cell rejuvenation. *Cell*. (2017) 170:142–157 e119. doi: 10.1016/j.cell.2017.06.007
 20. Philip M, Fairchild L, Sun L, Horste EL, Camara S, Shakiba M, et al. Chromatin states define tumor-specific T cell dysfunction and reprogramming. *Nature*. (2017) 545:452–6. doi: 10.1038/nature22367
 21. Utzschneider DT, Gabriel SS, Chisanga D, Gloury R, Gubser PM, Vasanthakumar A, et al. Early precursor T cells establish and propagate T cell exhaustion in chronic infection. *Nat Immunol*. (2020) 21:1256–66. doi: 10.1038/s41590-020-0760-z
 22. Chen Z, Ji Z, Ngio SF, Manne S, Cai Z, Huang AC, et al. TCF-1-centered transcriptional network drives an effector versus exhausted CD8 T cell-fate decision. *Immunity*. (2019) 51:840–855 e845. doi: 10.1016/j.immuni.2019.09.013
 23. Martinez GJ, Pereira RM, Aijo T, Kim EY, Marangoni F, Pipkin ME, et al. The transcription factor NFAT promotes exhaustion of activated CD8(+) T cells. *Immunity*. (2015) 42:265–78. doi: 10.1016/j.immuni.2015.01.006
 24. Alfei F, Kanev K, Hofmann M, Wu M, Ghoneim HE, Roelli P, et al. TOX reinforces the phenotype and longevity of exhausted T cells in chronic viral infection. *Nature*. (2019) 571:265–9. doi: 10.1038/s41586-019-1326-9
 25. Khan O, Giles JR, McDonald S, Manne S, Ngio SF, Patel KP, et al. TOX transcriptionally and epigenetically programs CD8(+) T cell exhaustion. *Nature*. (2019) 571:211–8. doi: 10.1038/s41586-019-1325-x
 26. Yao C, Sun HW, Lacey NE, Ji Y, Moseman EA, Shih HY, et al. Single-cell RNA-seq reveals TOX as a key regulator of CD8(+) T cell persistence in chronic infection. *Nat Immunol*. (2019) 20:890–901. doi: 10.1038/s41590-019-0403-4
 27. Seo H, Chen J, Gonzalez-Avalos E, Samaniego-Castruita D, Das A, Wang YH, et al. TOX and TOX2 transcription factors cooperate with NR4A transcription factors to impose CD8(+) T cell exhaustion. *Proc Natl Acad Sci USA*. (2019) 116:12410–5. doi: 10.1073/pnas.1905675116
 28. Srirat T, Hayakawa T, Mise-Omata S, Nakagawara K, Ando M, Shichino S, et al. NR4a1/2 deletion promotes accumulation of TCF1(+) stem-like precursors of exhausted CD8(+) T cells in the tumor microenvironment. *Cell Rep*. (2024) 43:113898. doi: 10.1016/j.celrep.2024.113898
 29. Sharpe AH, Pauken KE. The diverse functions of the PD1 inhibitory pathway. *Nat Rev Immunol*. (2018) 18:153–67. doi: 10.1038/nri.2017.108
 30. Burger ML, Cruz AM, Crossland GE, Gaglia G, Ritch CC, Blatt SE, et al. Antigen dominance hierarchies shape TCF1(+) progenitor CD8 T cell phenotypes in tumors. *Cell*. (2021) 184:4996–5014 e4926. doi: 10.1016/j.cell.2021.08.020
 31. Shakiba M, Zumbo P, Espinosa-Carrasco G, Menocal L, Dunder F, Carson SE, et al. TCR signal strength defines distinct mechanisms of T cell dysfunction and cancer evasion. *J Exp Med*. (2022) 219(2). doi: 10.1084/jem.20201966
 32. Connolly KA, Kuchroo M, Venkat A, Khatun A, Wang J, William I, et al. A reservoir of stem-like CD8(+) T cells in the tumor-draining lymph node preserves the ongoing antitumor immune response. *Sci Immunol*. (2021) 6:eabg7836. doi: 10.1126/sciimmunol.abg7836
 33. Cabrita R, Lauss M, Sanna A, Donia M, Skaarup Larsen M, Mitra S, et al. Tertiary lymphoid structures improve immunotherapy and survival in melanoma. *Nature*. (2020) 577:561–5. doi: 10.1038/s41586-019-1914-8
 34. Escobar G, Mangani D, Anderson AC. T cell factor 1: A master regulator of the T cell response in disease. *Sci Immunol*. (2020) 5(53). doi: 10.1126/sciimmunol.abb9726
 35. Zhao X, Shan Q, Xue HH. TCF1 in T cell immunity: a broadened frontier. *Nat Rev Immunol*. (2022) 22:147–57. doi: 10.1038/s41577-021-00563-6
 36. Jadhav RR, Im SJ, Hu B, Hashimoto M, Li P, Lin JX, et al. Epigenetic signature of PD-1+ TCF1+ CD8 T cells that act as resource cells during chronic viral infection and respond to PD-1 blockade. *Proc Natl Acad Sci USA*. (2019) 116:14113–8. doi: 10.1073/pnas.1903520116
 37. Zehn D, Thimme R, Lugli E, de Almeida GP, Oxenius A. ‘Stem-like’ precursors are the fount to sustain persistent CD8(+) T cell responses. *Nat Immunol*. (2022) 23:836–47. doi: 10.1038/s41590-022-01219-w
 38. Shan Q, Hu S, Chen X, Danahy DB, Badovinac VP, Zang C, et al. Ectopic Tcf1 expression instills a stem-like program in exhausted CD8(+) T cells to enhance viral and tumor immunity. *Cell Mol Immunol*. (2021) 18:1262–77. doi: 10.1038/s41423-020-0436-5
 39. Escobar G, Tooley K, Oliveras JP, Huang L, Cheng H, Bookstaver ML, et al. Tumor immunogenicity dictates reliance on TCF1 in CD8(+) T cells for response to immunotherapy. *Cancer Cell*. (2023) 41:1662–1679 e1667. doi: 10.1016/j.ccell.2023.08.001
 40. Sade-Feldman M, Yizhak K, Bjorgaard SL, Ray JP, de Boer CG, Jenkins RW, et al. Defining T cell states associated with response to checkpoint immunotherapy in melanoma. *Cell*. (2018) 175:998–1013 e1020. doi: 10.1016/j.cell.2018.10.038
 41. Fical M, Jegede OA, Sant’Angelo M, Hou Y, Flaifel A, Pignon JC, et al. Expression of T-cell exhaustion molecules and human endogenous retroviruses as predictive biomarkers for response to nivolumab in metastatic clear cell renal cell carcinoma. *Clin Cancer Res*. (2021) 27:1371–80. doi: 10.1158/1078-0432.CCR-20-3084
 42. Blackburn SD, Shin H, Freeman GJ, Wherry EJ. Selective expansion of a subset of exhausted CD8 T cells by alphaPD-L1 blockade. *Proc Natl Acad Sci USA*. (2008) 105:15016–21. doi: 10.1073/pnas.0801497105
 43. Philip M, Schietinger A. CD8(+) T cell differentiation and dysfunction in cancer. *Nat Rev Immunol*. (2022) 22:209–23. doi: 10.1038/s41577-021-00574-3
 44. Yost KE, Chang HY, Satpathy AT. Recruiting T cells in cancer immunotherapy. *Science*. (2021) 372:130–1. doi: 10.1126/science.abd1329
 45. Duhon T, Duhon R, Montler R, Moses J, Moudgil T, de Miranda NF, et al. Co-expression of CD39 and CD103 identifies tumor-reactive CD8 T cells in human solid tumors. *Nat Commun*. (2018) 9:2724. doi: 10.1038/s41467-018-05072-0
 46. Li H, van der Leun AM, Yofe I, Lubling Y, Gelbard-Solodkin D, van Akkooi ACJ, et al. Dysfunctional CD8 T cells form a proliferative, dynamically regulated compartment within human melanoma. *Cell*. (2019) 176:775–789 e718. doi: 10.1016/j.cell.2018.11.043
 47. Simoni Y, Becht E, Fehlings M, Loh CY, Koo SL, Teng KWW, et al. Bystander CD8(+) T cells are abundant and phenotypically distinct in human tumor infiltrates. *Nature*. (2018) 557:575–9. doi: 10.1038/s41586-018-0130-2
 48. Oliveira G, Stromhaug K, Klaeger S, Kula T, Frederick DT, Le PM, et al. Phenotype, specificity and avidity of antitumor CD8(+) T cells in melanoma. *Nature*. (2021) 596:119–25. doi: 10.1038/s41586-021-03704-y
 49. Caushi JX, Zhang J, Ji Z, Vaghshias A, Zhang B, Hsiue EH, et al. Author Correction: Transcriptional programs of neoantigen-specific TIL in anti-PD-1-treated lung cancers. *Nature*. (2021) 598:E1. doi: 10.1038/s41586-021-03893-6
 50. Im SJ, Konieczny BT, Hudson WH, Masopust D, Ahmed R. PD-1+ stemlike CD8 T cells are resident in lymphoid tissues during persistent LCMV infection. *Proc Natl Acad Sci USA*. (2020) 117:4292–9. doi: 10.1073/pnas.1917298117
 51. Dammeyer F, van Gulijk M, Mulder EE, Lukkes M, Klaase L, van den Bosch T, et al. The PD-1/PD-L1-checkpoint restrains T cell immunity in tumor-draining lymph nodes. *Cancer Cell*. (2020) 38:685–700 e688. doi: 10.1016/j.ccell.2020.09.001
 52. Spitzer MH, Carmi Y, Reticker-Flynn NE, Kwek SS, Madhiredy D, Martins MM, et al. Systemic immunity is required for effective cancer immunotherapy. *Cell*. (2017) 168:487–502 e415. doi: 10.1016/j.cell.2016.12.022
 53. Fransen MF, Schoonderwoerd M, Knopf P, Camps MG, Hawinkels LJ, Kneilling M, et al. Tumor-draining lymph nodes are pivotal in PD-1/PD-L1 checkpoint therapy. *JCI Insight*. (2018) 3(23). doi: 10.1172/jci.insight.124507
 54. Chow MT, Ozga AJ, Servis RL, Frederick DT, Lo JA, Fisher DE, et al. Intratumoral activity of the CXCR3 chemokine system is required for the efficacy of anti-PD-1 therapy. *Immunity*. (2019) 50:1498–1512 e1495. doi: 10.1016/j.immuni.2019.04.010
 55. Li Z, Tuong ZK, Dean I, Willis C, Gaspal F, Fiancette R, et al. *In vivo* labeling reveals continuous trafficking of TCF1+ T cells between tumor and lymphoid tissue. *J Exp Med*. (2022) 219(6). doi: 10.1084/jem.20210749
 56. Prokhnevskina N, Cardenas MA, Valanparambil RM, Sobierajska E, Barwick BG, Jansen C, et al. CD8(+) T cell activation in cancer comprises an initial activation phase in lymph nodes followed by effector differentiation within the tumor. *Immunity*. (2023) 56:107–124 e105. doi: 10.1016/j.immuni.2022.12.002
 57. Hua Y, Vella G, Rambow F, Allen E, Antoranz Martinez A, Duhamel M, et al. Cancer immunotherapies transition endothelial cells into HEVs that generate TCF1(+) T lymphocyte niches through a feed-forward loop. *Cancer Cell*. (2022) 40:1600–1618 e1610. doi: 10.1016/j.ccell.2022.11.002
 58. Hoch T, Schulz D, Eling N, Gomez JM, Levesque MP, Bodenmiller B. Multiplexed imaging mass cytometry of the chemokine milieu in melanoma characterizes features of the response to immunotherapy. *Sci Immunol*. (2022) 7:eabk1692. doi: 10.1126/sciimmunol.abki692

59. Magen A, Hamon P, Fiaschi N, Soong BY, Park MD, Mattiuz R, et al. Intratumoral dendritic cell-CD4(+) T helper cell niches enable CD8(+) T cell differentiation following PD-1 blockade in hepatocellular carcinoma. *Nat Med.* (2023) 29:1389–99. doi: 10.1038/s41591-023-02345-0
60. Schenkel JM, Herbst RH, Canner D, Li A, Hillman M, Shanahan SL, et al. Conventional type I dendritic cells maintain a reservoir of proliferative tumor-antigen specific TCF-1(+) CD8(+) T cells in tumor-draining lymph nodes. *Immunity.* (2021) 54:2338–2353 e2336. doi: 10.1016/j.immuni.2021.08.026
61. Huang Q, Wu X, Wang Z, Chen X, Wang L, Lu Y, et al. The primordial differentiation of tumor-specific memory CD8(+) T cells as bona fide responders to PD-1/PD-L1 blockade in draining lymph nodes. *Cell.* (2022) 185:4049–66.e4025. doi: 10.1016/j.cell.2022.09.020
62. Dahling S, Mansilla AM, Knopper K, Grafen A, Utzschneider DT, Ugur M, et al. Type I conventional dendritic cells maintain and guide the differentiation of precursors of exhausted T cells in distinct cellular niches. *Immunity.* (2022) 55:656–670 e658. doi: 10.1016/j.immuni.2022.03.006
63. Gebhardt T, Park SL, Parish IA. Stem-like exhausted and memory CD8(+) T cells in cancer. *Nat Rev Cancer.* (2023) 23:780–98. doi: 10.1038/s41568-023-00615-0
64. Pai JA, Hellmann MD, Sauter JL, Mattar M, Rizvi H, Woo HJ, et al. Lineage tracing reveals clonal progenitors and long-term persistence of tumor-specific T cells during immune checkpoint blockade. *Cancer Cell.* (2023) 41:776–790 e777. doi: 10.1016/j.ccell.2023.03.009
65. Rahim MK, Okholm TLH, Jones KB, McCarthy EE, Liu CC, Yee JL, et al. Dynamic CD8(+) T cell responses to cancer immunotherapy in human regional lymph nodes are disrupted in metastatic lymph nodes. *Cell.* (2023) 186:1127–1143 e1118. doi: 10.1016/j.cell.2023.02.021
66. Zander R, Schauder D, Xin G, Nguyen C, Wu X, Zajac A, et al. CD4(+) T cell help is required for the formation of a cytolytic CD8(+) T cell subset that protects against chronic infection and cancer. *Immunity.* (2019) 51:1028–1042 e1024. doi: 10.1016/j.immuni.2019.10.009
67. Cui C, Wang J, Fagerberg E, Chen PM, Connolly KA, Damo M, et al. Neoantigen-driven B cell and CD4 T follicular helper cell collaboration promotes anti-tumor CD8 T cell responses. *Cell.* (2021) 184:6101–6118 e6113. doi: 10.1016/j.cell.2021.11.007
68. Im SJ, Obeng RC, Nasti TH, McManus D, Kamphorst AO, Gunisetty S, et al. Characteristics and anatomic location of PD-1(+)TCF1(+) stem-like CD8 T cells in chronic viral infection and cancer. *Proc Natl Acad Sci USA.* (2023) 120:e2221985120. doi: 10.1073/pnas.2221985120
69. Helmink BA, Reddy SM, Gao J, Zhang S, Basar R, Thakur R, et al. B cells and tertiary lymphoid structures promote immunotherapy response. *Nature.* (2020) 577:549–55. doi: 10.1038/s41586-019-1922-8
70. Petitprez F, de Reynies A, Keung EZ, Chen TW, Sun CM, Calderaro J, et al. B cells are associated with survival and immunotherapy response in sarcoma. *Nature.* (2020) 577:556–60. doi: 10.1038/s41586-019-1906-8
71. Jansen CS, Prabhu RS, Pagadala MS, Chappa P, Goyal S, Zhou C, et al. Immune niches in brain metastases contain TCF1+ stem-like T cells, are associated with disease control and are modulated by preoperative SRS. *Res Sq.* (2023) 23:rs.3.rs-2722744. doi: 10.12103/rs.3.rs-2722744/v1
72. Carmona SJ, Siddiqui I, Bilous M, Held W, Gfeller D. Deciphering the transcriptomic landscape of tumor-infiltrating CD8 lymphocytes in B16 melanoma tumors with single-cell RNA-Seq. *Oncimmunology.* (2020) 9:1737369. doi: 10.1080/2162402X.2020.1737369
73. Broz ML, Binnewies M, Boldajipour B, Nelson AE, Pollack JL, Erle DJ, et al. Dissecting the tumor myeloid compartment reveals rare activating antigen-presenting cells critical for T cell immunity. *Cancer Cell.* (2014) 26:638–52. doi: 10.1016/j.ccell.2014.09.007
74. Meiser P, Knolle MA, Hirschberger A, de Almeida GP, Bayerl F, Lacher S, et al. A distinct stimulatory cDC1 subpopulation amplifies CD8(+) T cell responses in tumors for protective anti-cancer immunity. *Cancer Cell.* (2023) 41:1498–1515 e1410. doi: 10.1016/j.ccell.2023.06.008
75. Hui E, Cheung J, Zhu J, Su X, Taylor MJ, Wallweber HA, et al. T cell costimulatory receptor CD28 is a primary target for PD-1-mediated inhibition. *Science.* (2017) 355:1428–33. doi: 10.1126/science.aaf1292
76. Kamphorst AO, Wieland A, Nasti T, Yang S, Zhang R, Barber DL, et al. Rescue of exhausted CD8 T cells by PD-1-targeted therapies is CD28-dependent. *Science.* (2017) 355:1423–7. doi: 10.1126/science.aaf0683
77. Dangaj D, Bruand M, Grimm AJ, Ronet C, Barras D, Duttgupta PA, et al. Cooperation between constitutive and inducible chemokines enables T cell engraftment and immune attack in solid tumors. *Cancer Cell.* (2019) 35:885–900 e810. doi: 10.1016/j.ccell.2019.05.004
78. House IG, Savas P, Lai J, Chen AXY, Oliver AJ, Teo ZL, et al. Macrophage-derived CXCL9 and CXCL10 are required for antitumor immune responses following immune checkpoint blockade. *Clin Cancer Res.* (2020) 26:487–504. doi: 10.1158/1078-0432.CCR-19-1868
79. Petty AJ, Li A, Wang X, Dai R, Heyman B, Hsu D, et al. Hedgehog signaling promotes tumor-associated macrophage polarization to suppress intratumoral CD8+ T cell recruitment. *J Clin Invest.* (2019) 129:5151–62. doi: 10.1172/JCI128644
80. Rashidian M, LaFleur MW, Verschoor VL, Dongre A, Zhang Y, Nguyen TH, et al. Immuno-PET identifies the myeloid compartment as a key contributor to the outcome of the antitumor response under PD-1 blockade. *Proc Natl Acad Sci USA.* (2019) 116:16971–80. doi: 10.1073/pnas.1905005116
81. Qu Y, Wen J, Thomas G, Yang W, Prior W, He W, et al. Baseline frequency of inflammatory Cxcl9-expressing tumor-associated macrophages predicts response to avelumab treatment. *Cell Rep.* (2020) 32:107873. doi: 10.1016/j.celrep.2020.107873
82. Marcovecchio PM, Thomas G, Salek-Ardakani S. CXCL9-expressing tumor-associated macrophages: new players in the fight against cancer. *J Immunother Cancer.* (2021) 9. doi: 10.1136/jitc-2020-002045
83. Huang AC, Postow MA, Orlowski RJ, Mick R, Bengsch B, Manne S, et al. T-cell invigoration to tumor burden ratio associated with anti-PD-1 response. *Nature.* (2017) 545:60–5. doi: 10.1038/nature22079
84. Thommen DS, Koelzer VH, Herzog P, Roller A, Trefny M, Dimeloe S, et al. A transcriptionally and functionally distinct PD-1(+) CD8(+) T cell pool with predictive potential in non-small-cell lung cancer treated with PD-1 blockade. *Nat Med.* (2018) 24:994–1004. doi: 10.1038/s41591-018-0057-z
85. Clarke J, Panwar B, Madrigal A, Singh D, Gujar R, Wood O, et al. Single-cell transcriptomic analysis of tissue-resident memory T cells in human lung cancer. *J Exp Med.* (2019) 216:2128–49. doi: 10.1084/jem.20190249
86. Zhang Y, Chen H, Mo H, Hu X, Gao R, Zhao Y, et al. Single-cell analyses reveal key immune cell subsets associated with response to PD-L1 blockade in triple-negative breast cancer. *Cancer Cell.* (2021) 39:1578–1593 e1578. doi: 10.1016/j.ccell.2021.09.010
87. Li K, Tandurella JA, Gai J, Zhu Q, Lim SJ, Thomas DL 2nd, et al. Multi-omic analyses of changes in the tumor microenvironment of pancreatic adenocarcinoma following neoadjuvant treatment with anti-PD-1 therapy. *Cancer Cell.* (2022) 40:1374–1391 e1377. doi: 10.1016/j.ccell.2022.10.001
88. Bassez A, Vos H, Van Dyck L, Floris G, Arijis I, Desmedt C, et al. A single-cell map of intratumoral changes during anti-PD1 treatment of patients with breast cancer. *Nat Med.* (2021) 27:820–32. doi: 10.1038/s41591-021-01323-8
89. Held W, Siddiqui I, Schaeuble K, Speiser DE. Intratumoral CD8(+) T cells with stem cell-like properties: Implications for cancer immunotherapy. *Sci Transl Med.* (2019) 11(515). doi: 10.1126/scitranslmed.aay6863
90. Liu B, Hu X, Feng K, Gao R, Xue Z, Zhang S, et al. Temporal single-cell tracing reveals clonal revival and expansion of precursor exhausted T cells during anti-PD-1 therapy in lung cancer. *Nat Cancer.* (2022) 3:108–21. doi: 10.1038/s43018-021-00292-8
91. Odorizzi PM, Pauken KE, Paley MA, Sharpe A, Wherry EJ. Genetic absence of PD-1 promotes accumulation of terminally differentiated exhausted CD8+ T cells. *J Exp Med.* (2015) 212:1125–37. doi: 10.1084/jem.20142237
92. Tsui C, Kretschmer L, Rapelius S, Gabriel SS, Chisanga D, Knopper K, et al. MYB orchestrates T cell exhaustion and response to checkpoint inhibition. *Nature.* (2022) 609:354–60. doi: 10.1038/s41586-022-05105-1
93. Tichet M, Wullschlegler S, Chryplewicz A, Fournier N, Marcone R, Kauzlaric A, et al. Bispecific PD1-IL2v and anti-PD-L1 break tumor immunity resistance by enhancing stem-like tumor-reactive CD8(+) T cells and reprogramming macrophages. *Immunity.* (2023) 56:162–179 e166. doi: 10.1016/j.immuni.2022.12.006
94. Philipp N, Kazerani M, Nicholls A, Vick B, Wulf J, Straub T, et al. T-cell exhaustion induced by continuous bispecific molecule exposure is ameliorated by treatment-free intervals. *Blood.* (2022) 140:1104–18. doi: 10.1182/blood.2022015956
95. Lelliott EJ, Kong IY, Zethoven M, Ramsbottom KM, Martelotto LG, Meyran D, et al. CDK4/6 inhibition promotes antitumor immunity through the induction of T-cell memory. *Cancer Discovery.* (2021) 11:2582–601. doi: 10.1158/2159-8290.CD-20-1554
96. Ebert PJR, Cheung J, Yang Y, McNamara E, Hong R, Moskalenko M, et al. MAP kinase inhibition promotes T cell and anti-tumor activity in combination with PD-L1 checkpoint blockade. *Immunity.* (2016) 44:609–21. doi: 10.1016/j.immuni.2016.01.024
97. Verma V, Jafarzadeh N, Boi S, Kundu S, Jiang Z, Fan Y, et al. MEK inhibition reprograms CD8(+) T lymphocytes into memory stem cells with potent antitumor effects. *Nat Immunol.* (2021) 22:53–66. doi: 10.1038/s41590-020-00818-9
98. LaFleur MW, Nguyen TH, Cox MA, Miller BC, Yates KB, Gillis JE, et al. PTPN2 regulates the generation of exhausted CD8(+) T cell subpopulations and restrains tumor immunity. *Nat Immunol.* (2019) 20:1335–47. doi: 10.1038/s41590-019-0480-4
99. Pelly VS, Moeini A, Roelofsen LM, Bonavita E, Bell CR, Hutton C, et al. Anti-inflammatory drugs remodel the tumor immune environment to enhance immune checkpoint blockade efficacy. *Cancer Discovery.* (2021) 11:2602–19. doi: 10.1158/2159-8290.CD-20-1815
100. Liu C, Somasundaram A, Manne S, Gocher AM, Szymczak-Workman AL, Vignali KM, et al. Neuropilin-1 is a T cell memory checkpoint limiting long-term antitumor immunity. *Nat Immunol.* (2020) 21:1010–21. doi: 10.1038/s41590-020-0733-2
101. Francis DM, Manspeaker MP, Schudel A, Sestito LF, O'Melia MJ, Kissick HT, et al. Blockade of immune checkpoints in lymph nodes through locoregional delivery augments cancer immunotherapy. *Sci Transl Med.* (2020) 12(563). doi: 10.1126/scitranslmed.aay3575
102. Shen Y, Connolly E, Aiello M, Zhou C, Chappa P, Song H, et al. Radiation and anti-PD-L1 synergize by stimulating a stem-like T cell population in the tumor-draining lymph node. *Res Sq.* (2024) 6:rs.3.rs-3921977. doi: 10.1158/1538-7445.AM2024-LB084
103. Cieri N, Camisa B, Cocchiarella F, Forcato M, Oliveira G, Provasi E, et al. IL-7 and IL-15 instruct the generation of human memory stem T cells from naive precursors. *Blood.* (2013) 121:573–84. doi: 10.1182/blood-2012-05-431718

104. Sabatino M, Hu J, Sommariva M, Gautam S, Fellowes V, Hocker JD, et al. Generation of clinical-grade CD19-specific CAR-modified CD8+ memory stem cells for the treatment of human B-cell Malignancies. *Blood*. (2016) 128:519–28. doi: 10.1182/blood-2015-11-683847
105. Zanon V, Pilipow K, Scamardella E, De Paoli F, De Simone G, Price DA, et al. Curtailed T-cell activation curbs effector differentiation and generates CD8(+) T cells with a naturally-occurring memory stem cell phenotype. *Eur J Immunol*. (2017) 47:1468–76. doi: 10.1002/eji.201646732
106. Lee J, Lee K, Bae H, Lee K, Lee S, Ma J, et al. IL-15 promotes self-renewal of progenitor exhausted CD8 T cells during persistent antigenic stimulation. *Front Immunol*. (2023) 14:1117092. doi: 10.3389/fimmu.2023.1117092
107. Romine KA, MacPherson K, Cho HJ, Kosaka Y, Flynn PA, Byrd KH, et al. BET inhibitors rescue anti-PD1 resistance by enhancing TCF7 accessibility in leukemia-derived terminally exhausted CD8(+) T cells. *Leukemia*. (2023) 37:580–92. doi: 10.1038/s41375-023-01808-0
108. Kondo T, Morita R, Okuzono Y, Nakatsukasa H, Sekiya T, Chikuma S, et al. Notch-mediated conversion of activated T cells into stem cell memory-like T cells for adoptive immunotherapy. *Nat Commun*. (2017) 8:15338. doi: 10.1038/ncomms15338
109. Ando M, Kondo T, Tomisato W, Ito M, Shichino S, Srirat T, et al. Rejuvenating effector/exhausted CAR T cells to stem cell memory-like CAR T cells by resting them in the presence of CXCL12 and the NOTCH ligand. *Cancer Res Commun*. (2021) 1:41–55. doi: 10.1158/2767-9764.CRC-21-0034
110. Kagoya Y, Nakatsugawa M, Yamashita Y, Ochi T, Guo T, Anczurowski M, et al. BET bromodomain inhibition enhances T cell persistence and function in adoptive immunotherapy models. *J Clin Invest*. (2016) 126:3479–94. doi: 10.1172/JCI86437
111. Mousset CM, Hobo W, Ji Y, Fredrix H, De Giorgi V, Allison RD, et al. Ex vivo AKT-inhibition facilitates generation of polyfunctional stem cell memory-like CD8(+) T cells for adoptive immunotherapy. *Oncimmunology*. (2018) 7:e1488565. doi: 10.1080/2162402X.2018.1488565
112. Gattinoni L, Klebanoff CA, Palmer DC, Wrzesinski C, Kerstann K, Yu Z, et al. Acquisition of full effector function *in vitro* paradoxically impairs the *in vivo* antitumor efficacy of adoptively transferred CD8+ T cells. *J Clin Invest*. (2005) 115:1616–26. doi: 10.1172/JCI24480
113. Gattinoni L, Zhong XS, Palmer DC, Ji Y, Hinrichs CS, Yu Z, et al. Wnt signaling arrests effector T cell differentiation and generates CD8+ memory stem cells. *Nat Med*. (2009) 15:808–13. doi: 10.1038/nm.1982
114. Gattinoni L, Lugli E, Ji Y, Pos Z, Paulos CM, Quigley MF, et al. A human memory T cell subset with stem cell-like properties. *Nat Med*. (2011) 17:1290–7. doi: 10.1038/nm.2446
115. Chapuis AG, Ragnarsson GB, Nguyen HN, Chaney CN, Pufnock JS, Schmitt TM, et al. Transferred WT1-reactive CD8+ T cells can mediate antileukemic activity and persist in post-transplant patients. *Sci Transl Med*. (2013) 5:174ra127. doi: 10.1126/scitranslmed.3004916
116. Xu Y, Zhang M, Ramos CA, Duret A, Liu E, Dakhova O, et al. Closely related T-memory stem cells correlate with *in vivo* expansion of CAR-CD19-T cells and are preserved by IL-7 and IL-15. *Blood*. (2014) 123:3750–9. doi: 10.1182/blood-2014-01-552174
117. Fraietta JA, Lacey SF, Orlando EJ, Pruteanu-Malinici I, Gohil M, Lundh S, et al. Determinants of response and resistance to CD19 chimeric antigen receptor (CAR) T cell therapy of chronic lymphocytic leukemia. *Nat Med*. (2018) 24:563–71. doi: 10.1038/s41591-018-0010-1
118. Pilipow K, Scamardella E, Puccio S, Gautam S, De Paoli F, Mazza EM, et al. Antioxidant metabolism regulates CD8+ T memory stem cell formation and antitumor immunity. *JCI Insight*. (2018) 3(18). doi: 10.1172/jci.insight.122299
119. Neelapu SS, Locke FL, Bartlett NL, Lekakis LJ, Miklos DB, Jacobson CA, et al. Axicabtagene ciloleucel CART-cell therapy in refractory large B-cell lymphoma. *N Engl J Med*. (2017) 377:2531–44. doi: 10.1056/NEJMoa1707447
120. Schuster SJ, Bishop MR, Tam CS, Waller EK, Borchmann P, McGuirk JP, et al. Tisagenlecleucel in adult relapsed or refractory diffuse large B-cell lymphoma. *N Engl J Med*. (2019) 380:45–56. doi: 10.1056/NEJMoa1804980
121. Abramson JS, Palomba ML, Gordon LI, Lunning MA, Wang M, Arnason J, et al. Lisocabtagene maraleucel for patients with relapsed or refractory large B-cell lymphomas (TRANSCEND NHL 001): a multicenter seamless design study. *Lancet*. (2020) 396:839–52. doi: 10.1016/S0140-6736(20)31366-0
122. Locke FL, Miklos DB, Jacobson CA, Perales MA, Kersten MJ, Oluwole OO, et al. Axicabtagene ciloleucel as second-Line therapy for large B-Cell lymphoma. *New Engl J Med*. (2022) 386:640–54. doi: 10.1056/NEJMoa2116133
123. Kamdar M, Solomon SR, Arnason J, Johnston PB, Glass B, Bachanova V, et al. Lisocabtagene maraleucel versus standard of care with salvage chemotherapy followed by autologous stem cell transplantation as second-line treatment in patients with relapsed or refractory large B-cell lymphoma (TRANSFORM): results from an interim analysis of an open-label, randomized, phase 3 trial. *Lancet*. (2022) 399:2294–308. doi: 10.1016/S0140-6736(22)00662-6
124. Jacobson CA, Chavez JC, Sehgal AR, William BM, Munoz J, Salles G, et al. Axicabtagene ciloleucel in relapsed or refractory indolent non-Hodgkin lymphoma (ZUMA-5): a single-arm, multicenter, phase 2 trial. *Lancet Oncol*. (2022) 23:91–103. doi: 10.1016/S1470-2045(21)00591-X
125. Fowler NH, Dickinson M, Dreyling M, Martinez-Lopez J, Kolstad A, Butler J, et al. Tisagenlecleucel in adult relapsed or refractory follicular lymphoma: the phase 2 ELARA trial. *Nat Med*. (2022) 28:325–32. doi: 10.1038/s41591-021-01622-0
126. Deng Q, Han G, Puebla-Osorio N, Ma MCJ, Strati P, Chasen B, et al. Characteristics of anti-CD19 CAR T cell infusion products associated with efficacy and toxicity in patients with large B cell lymphomas. *Nat Med*. (2020) 26:1878–87. doi: 10.1038/s41591-020-1061-7
127. Locke FL, Rossi JM, Neelapu SS, Jacobson CA, Miklos DB, Ghobadi A, et al. Tumor burden, inflammation, and product attributes determine outcomes of axicabtagene ciloleucel in large B-cell lymphoma. *Blood Adv*. (2020) 4:4898–911. doi: 10.1182/bloodadvances.2020002394
128. Monfrini C, Stella F, Aragona V, Magni M, Ljevar S, Vella C, et al. Phenotypic composition of commercial anti-CD19 CAR T cells affects *in vivo* expansion and disease response in patients with large B-cell lymphoma. *Clin Cancer Res*. (2022) 28:3378–86. doi: 10.1158/1078-0432.CCR-22-0164
129. Denlinger N, Song NJ, Zhang X, Jeon H, Peterson C, Wang Y, et al. Postinfusion PD-1+ CD8+ CAR T cells identify patients responsive to CD19 CAR T-cell therapy in non-Hodgkin lymphoma. *Blood Adv*. (2024) 8:3140–53. doi: 10.1182/bloodadvances.2023012073
130. Lynn RC, Weber EW, Sotillo E, Gennert D, Xu P, Good Z, et al. c-Jun overexpression in CAR T cells induces exhaustion resistance. *Nature*. (2019) 576:293–300. doi: 10.1038/s41586-019-1805-z
131. Chan JD, Scheffler CM, Munoz I, Sek K, Lee JN, Huang YK, et al. FOXP1 enhances CAR T cell stemness, metabolic fitness and efficacy. *Nature*. (2024) 629:201–10. doi: 10.1038/s41586-024-07242-1
132. Jung IY, Narayan V, McDonald S, Rech AJ, Bartoszek R, Hong G, et al. BLIMP1 and NR4A3 transcription factors reciprocally regulate antitumor CAR T cell stemness and exhaustion. *Sci Transl Med*. (2022) 14:eabn7336. doi: 10.1126/scitranslmed.abn7336
133. Jiang P, Zhang Z, Hu Y, Liang Z, Han Y, Li X, et al. Single-cell ATAC-seq maps the comprehensive and dynamic chromatin accessibility landscape of CAR-T cell dysfunction. *Leukemia*. (2022) 36:2656–68. doi: 10.1038/s41375-022-01676-0
134. Chen J, Lopez-Moyado IF, Seo H, Lio CJ, Hempleman LJ, Sekiya T, et al. NR4A transcription factors limit CAR T cell function in solid tumors. *Nature*. (2019) 567:530–4. doi: 10.1038/s41586-019-0985-x
135. Zhu Z, Lou G, Teng XL, Wang H, Luo Y, Shi W, et al. FOXP1 and KLF2 reciprocally regulate checkpoints of stem-like to effector transition in CAR T cells. *Nat Immunol*. (2024) 25:117–28. doi: 10.1038/s41590-023-01685-w
136. Scholler N, Perbost R, Locke FL, Jain MD, Turcan S, Danan C, et al. Tumor immune contexture is a determinant of anti-CD19 CAR T cell efficacy in large B cell lymphoma. *Nat Med*. (2022) 28:1872–82. doi: 10.1038/s41591-022-01916-x
137. Reinhard K, Rengstl B, Oehm P, Michel K, Billmeier A, Hayduk N, et al. An RNA vaccine drives expansion and efficacy of claudin-CAR-T cells against solid tumors. *Science*. (2020) 367:446–53. doi: 10.1126/science.aay5967
138. Mizukoshi E, Nakagawa H, Tamai T, Kitahara M, Fushimi K, Nio K, et al. long-term surviving cancer patients harbor self-renewing tumor-specific CD8(+) T cells. *Nat Commun*. (2022) 13:3123. doi: 10.1038/s41467-022-30861-z
139. Sahin U, Derhovanessian E, Miller M, Kloke BP, Simon P, Lower M, et al. Personalized RNA mutanome vaccines mobilize poly-specific therapeutic immunity against cancer. *Nature*. (2017) 547:222–6. doi: 10.1038/nature23003
140. Ott PA, Hu Z, Keskin DB, Shukla SA, Sun J, Bozym DJ, et al. An immunogenic personal neoantigen vaccine for patients with melanoma. *Nature*. (2017) 547:217–21. doi: 10.1038/nature22991
141. Kreiter S, Vormehr M, van de Roemer N, Diken M, Lower M, Diekmann J, et al. Mutant MHC class II epitopes drive therapeutic immune responses to cancer. *Nature*. (2015) 520:692–6. doi: 10.1038/nature14426
142. Yadav M, Jhunjhunwala S, Phung QT, Lupardus P, Tanguay J, Bumbaca S, et al. Predicting immunogenic tumor mutations by combining mass spectrometry and exome sequencing. *Nature*. (2014) 515:572–6. doi: 10.1038/nature14001
143. Li B, Jing P, Zheng G, Pi C, Zhang L, Yin Z, et al. Neo-intline: integrated pipeline enables neoantigen design through the in-silico presentation of T-cell epitope. *Signal Transduct Target Ther*. (2023) 8:397. doi: 10.1038/s41392-023-01644-9
144. Hilf N, Kuttruff-Coqui S, Frenzel K, Bukur V, Stevanovic S, Gouttefangeas C, et al. Actively personalized vaccination trial for newly diagnosed glioblastoma. *Nature*. (2019) 565:240–5. doi: 10.1038/s41586-018-0810-y
145. Yu YJ, Shan N, Li LY, Zhu YS, Lin LM, Mao CC, et al. Preliminary clinical study of personalized neoantigen vaccine therapy for microsatellite stability (MSS)-advanced colorectal cancer. *Cancer Immunol Immunother*. (2023) 72:2045–56. doi: 10.1007/s00262-023-03386-7
146. Baharom F, Ramirez-Valdez RA, Tobin KKS, Yamane H, Dutertre CA, Khalilnezhad A, et al. Intravenous nanoparticle vaccination generates stem-like TCF1(+) neoantigen-specific CD8(+) T cells. *Nat Immunol*. (2021) 22:41–52. doi: 10.1038/s41590-020-00810-3
147. D'Alise AM, Brasu N, De Intinis C, Leoni G, Russo V, Langone F, et al. Adenoviral-based vaccine promotes neoantigen-specific CD8(+) T cell stemness and tumor rejection. *Sci Transl Med*. (2022) 14:eabo7604. doi: 10.1126/scitranslmed.abo7604
148. Shimizu K, Ueda S, Kawamura M, Aoshima H, Satoh M, Nakabayashi J, et al. Combination of cancer vaccine with CD122-biased IL-2/anti-IL-2 Ab complex shapes the stem-like effector NK and CD8(+) T cells against tumor. *J Immunother Cancer*. (2023) 11(7). doi: 10.1136/jitc-2022-006409