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Harnessing microbial antigens as cancer antigens: a promising avenue for cancer immunotherapy

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Immunotherapy has revolutionized cancer treatment by leveraging the immune system's innate capabilities to combat malignancies. Despite the promise of tumor antigens in stimulating anti-tumor immune responses, their clinical utility is hampered by limitations in eliciting robust and durable immune reactions, exacerbated by tumor heterogeneity and immune evasion mechanisms. Recent insights into the immunogenic properties of host homologous microbial antigens have sparked interest in their potential for augmenting anti-tumor immunity while minimizing off-target effects. This review explores the therapeutic potential of microbial antigens and their challenges in clinical translation. We further explore the intricate relationship between microorganisms and tumor development, elucidating the concept of molecular mimicry and its implications for immune recognition of tumor-associated antigens. Finally, we discuss methodologies for identifying and characterizing microbial antigen peptides, highlighting their immunogenicity and prospects for therapeutic application.

KEYWORDS

microbial antigen peptide, tumor antigen, microorganism, immunotherapy, molecular mimicry

Highlights

- Microbial peptides mimicking tumor antigens.
- Elevated immunogenicity of host-homologous microbial antigens surpassing tumor-associated counterparts.
- Bacterial antigenic peptides as promising candidates for tumor vaccines.

• Exploring host-homologous microbial antigens through innovative strategies.

1 Introduction

Immunotherapy has ushered in a new era in cancer treatment, offering tailored strategies to harness the immune system's intrinsic capabilities in combating malignancies (1, 2). Central to the success of immunotherapy are tumor antigens, which serve as crucial targets for immune recognition and subsequent elimination of cancerous cells while sparing normal tissues from collateral damage (3). However, despite the promise of cancer antigens in immunotherapeutic interventions, their clinical efficacy remains limited due to various challenges, including the constrained repertoire of antigens capable of eliciting robust and enduring anti-tumor immune responses (4). Moreover, the intricate landscape of tumor heterogeneity, characterized by variations in antigen expression profiles among patients and within individual tumors, poses a formidable barrier to the development of effective immunotherapeutic approaches (5). Furthermore, cancer cells employ a repertoire of immune evasion mechanisms, such as antigen downregulation and alteration of antigen processing pathways, further hampering immune recognition and compromising therapeutic outcomes (6). Consequently, there is a critical need to identify and validate novel tumor antigens through comprehensive profiling of tumor cells and the surrounding microenvironment, with the aim of uncovering immunogenic targets accessible to immune effectors. Recent advancements have highlighted the unique immunogenic properties of host homologous microbial antigens, which offer the potential to elicit robust immune responses against tumors while minimizing off-target effects on normal tissues (7). For instance, Wells et al. proposed the concept of pathogenic peptide homology as a strategy for evaluating neoantigen immunogenicity, suggesting that comparisons with exogenous microbial peptides could facilitate the identification of neoantigens for cancer immunotherapy (8, 9). In this review, we aim to explore the therapeutic potential of homologous microbial antigen peptides in tumor immunotherapy. We commence by providing an overview of tumor antigens and their characteristics, followed by an examination of the influence of microorganisms on tumor development and treatment. Subsequently, we summary the concept of molecular mimicry, elucidating the structural similarities between microbial antigens and tumor-associated antigens. Finally, we present a comprehensive overview of methods for obtaining microbial antigen peptides, discussing their immunogenicity and potential applications in cancer immunotherapy.

2 Tumor specific antigen and tumor associated antigen

Tumor antigens encompass two primary categories: tumor-specific antigens (TSAs), also known as neoantigens, and tumor-associated

antigens (TAAs). TSAs originate from nonsynonymous mutations or other genetic alterations, leading to the production of mutant peptides exclusively expressed in tumor cells (10). In contrast, noncanonical neoepitopes arise from various mechanisms such as alternative splicing, post-translational modifications, RNA editing, and aberrant mRNA translation (9). These tumor neoantigens, as immunogenic nonautoantigens, demonstrate specificity in their expression by tumor cells, thereby eliciting highly targeted immune responses against the malignancy (9, 11). Consequently, neoantigens exhibit two defining characteristics: exclusive presence on tumor cells and the capacity to evade central immune tolerance mechanisms, effectively stimulating T cell responses. For example, a recently identified melanoma epitope has been recognized by CD4⁺ T cells, with a neoantigen vaccine predominantly activating this subset of immune cells (12, 13). Crucial to targeted tumor therapy leveraging neoantigens is the identification of those possessing robust immunogenicity capable of eliciting potent T cell responses (9). Notably, several neoantigens share sequence homology with peptides derived from microbial sources, with accumulating evidence indicating their heightened immunogenic potential compared to pathogenic antigen homologs and nonhomologous neoantigens (14, 15).

The personalized nature of neoantigens presents a significant hurdle for their widespread application, as they are uniquely tailored to individual patients and thus impractical for large-scale use (13, 16, 17). Furthermore, the dynamic mutation landscape of tumors can lead to the continuous generation of novel neoantigens, allowing tumors to evade immune surveillance (10). In contrast, TAAs exhibit broader relevance across various malignancies, as they are self-antigens overexpressed in tumor cells while also shared with normal tissues. Consequently, TAAs represent promising targets for the development of cancer vaccines on a larger scale (10). For instance, the melanomaassociated antigen (MAGE) gene family-encoded antigens MZ2-E and MZ2-D have been identified as elicitors of anti-tumor immune responses mediated by cytotoxic T cells since the 1990s (18, 19). However, the ubiquitous expression of TAAs in normal tissues poses a challenge, potentially inducing immune tolerance and impeding effective anti-tumor immune responses (20, 21). Moreover, TAAspecific T cells may inadvertently target corresponding normal tissues, precipitating autoimmune reactions (22, 23). In light of these challenges, Tagliamonte et al. proposed that exploiting the homology between TAAs and microbial antigens may offer a promising strategy to enhance the efficacy of cancer immunotherapy (10).

3 How intratumoral microorganisms shape the immune landscape within tumors

3.1 Reciprocity between intratumoral microorganisms and tumor cells

The term "microbiome in tumors" encompasses the diverse microbial communities residing within tumor tissues or their

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immediate surroundings, comprising bacteria, fungi, viruses, and other microorganisms (24). These entities can inhabit tumor cells directly or parasitize tissues, blood, or adjacent body fluids. Emerging research indicates that various types of tumors harbor distinct microbial compositions, suggesting a close association between microbial diversity and tumor types (25, 26). In the context of breast cancer, for instance, the presence of specific microorganisms within breast cancer tissues has been implicated in facilitating tumor metastasis, highlighting the functional significance of the tumor microbiome (27). Moreover, the spatial organization of microflora within tumors reflects a non-random distribution pattern, occupying discrete niches that modulate immune responses and epithelial cell functions. Interactions between these microorganisms and tumor cells can promote cancer progression through various mechanisms, including direct stimulation of cell proliferation, suppression of immune responses, and modulation of the tumor microenvironment (28).

The influence mechanisms of tumor-associated microorganisms on tumor development have garnered considerable attention. On one hand, these microorganisms may induce DNA damage, manipulate host gene expression, or interfere with signaling pathways, thereby promoting tumorigenesis (29, 30). Alternatively, they can contribute to tumor progression by inducing immunosuppressive states, fostering pro-inflammatory responses, and activating carcinogenic pathways (31). Furthermore, microbial metabolites may interfere with chemotherapy efficacy, leading to drug resistance (32). Conversely, the presence of microorganisms within tumors may induce inhibitory effects on tumor progression. Studies suggest that the diversity of tumor-associated microorganisms correlates with the survival outcomes of pancreatic cancer patients, with interplay observed between the microbial compositions of pancreatic tumors and the intestinal microbiota (33). Notably, certain microbial compounds produced within tumors may enhance anti-tumor immunity, though their precise origins remain unclear (34). Moreover, the tumor microbiome can influence the efficacy of immunotherapy by modulating immune responses and altering tumor immunogenicity (31). In addition, bacterial influences on tumor outcomes extend to transcriptional pathways that directly impact inflammation, tumor growth, and immune cell infiltration, underscoring the multifaceted nature of microbial contributions to cancer pathogenesis (28, 33).

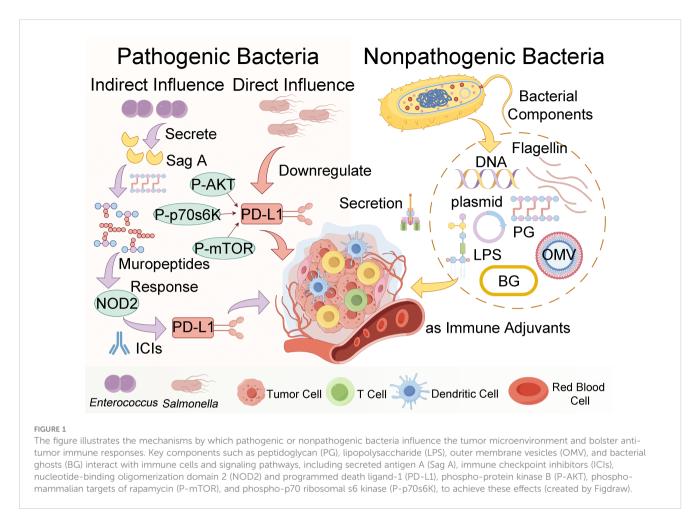
3.2 Unveiling the mechanisms: microbialmediated immune regulation

Immunotherapy for tumors encompasses a spectrum of modalities, including molecular targeted therapy utilizing monoclonal antibodies, immune checkpoint inhibitor (ICI) therapy, adoptive cellular immunotherapy such as chimeric antigen receptor T cell (CAR-T) therapy, cytokine therapy involving interleukin-2 (IL-2) and interferon-gamma (IFN- γ), and tumor vaccines (35, 36). Among these approaches, ICI therapy stands out as a mature and extensively studied modality. Its primary mechanism involves the upregulation of

T cell-mediated immune killing by targeting co-suppressor molecules, notably programmed death protein-1/programmed death ligand-1 (PD-1/PD-L1), thereby bolstering endogenous host immunity and counteracting tumor immune evasion (37, 38). Despite the clinical approval of several ICIs for cancer treatment, challenges persist, including low response rates and the emergence of drug resistance in many cases (6, 39-43). The intricate interplay between intestinal microorganisms and host immunity can indirectly influence cancer patients' responses to ICIs (Figure 1) (44, 45). For instance, active Enterococcus secreted antigen A (Sag A), may enhance host immunity by interacting with nucleotide-binding oligomerization domain 2 (NOD2), potentially augmenting the anti-tumor effects of anti-PD-L1 therapy (46). In the realm of anti-tumor immunity, microbial communities exert regulatory control over metabolites through the stimulation of pathogen-related molecular patterns, thereby contributing to the formation of an adaptive immune repertoire by inducing cross-reactive T cell responses (44, 47-49).

Indeed, bacteria serve as natural immune antigens capable of activating immune cells to mount anti-tumor responses (Figure 1). Studies have demonstrated the potential of *Salmonella* in inducing direct tumor cell death and modulating the tumor microenvironment via immune-mediated mechanisms, thereby promoting tumor suppression (50, 51). For instance, *Salmonella* has been shown to downregulate the expression of programmed death-ligand 1 (PD-L1) in tumors, thus altering tumor immune tolerance (52). Moreover, recent research by Gao et al. has unveiled the therapeutic potential of a mixture of four Clostridium strains, designated CC4, in enhancing programmed cell death protein 1 (PD-1) therapy. This innovative approach holds promise as an independent immunotherapeutic strategy for melanoma and colorectal cancer (53).

Bacteria also possess the potential to modulate immune cells as therapeutic agents, as depicted in Figure 1. Non-pathogenic bacteria can interact with phagocytes and regulate various immune cells within the tumor microenvironment, including macrophages, dendritic cells, and neutrophils (54, 55). The early induction of monocyte-derived dendritic cells (DCs) serves as a hallmark of successful activation of the tumor immune response (56). Bacterial factors are frequently utilized as immune adjuvants in DC-based anti-tumor treatments, exemplified by the use of 50S ribosomal protein from Mycobacterium tuberculosis and w-Mycobacterium acid mutase TBCM, as well as FcyR secreted by Staphylococcus aureus (56, 57). NK cells, belonging to the innate lymphocyte (ILC) family, exhibit versatile immune functions within tumors (58). Bacteria such as Salmonella and Mycobacteria have been shown to impede the progression of numerous cancers in an NK celldependent manner (59, 60). This effect is mediated by the stimulation of NK cells to produce IFN-y, which orchestrates the accumulation, activation, and cytotoxicity of NK cells, thereby restraining tumor metastasis (61). A large number of preclinical evidence highlights the reliance of bacteria on T cells to instigate anti-tumor adaptive immune responses (47). For instance, a vaccine targeting Helicobacter pylori activates CD3⁺ T cell immune responses and suppresses the growth of germinal center (GC) cells. Consequently, the T cell immunity induced by bacteria cross-reacts with cancer cell major histocompatibility complex class I (MHC-I) restriction antigens, underscoring the role



of microbial-specific T cells in bolstering anti-tumor immune responses (62, 63).

4 Use of microbial peptides as cancer neoantigens for immunotherapy

4.1 Molecular mimicry of microbial peptides to human antigens

The concept of molecular mimicry, also known as antigen mimicry, was first proposed in 1964 to elucidate the phenomenon wherein infectious pathogens display structural similarities to antigens expressed by human cells, enabling them to evade host immune responses and establish more invasive infections (64). Molecular mimicry not only facilitates pathogen immune evasion but can also precipitate autoimmune diseases. For instance, the epitope SVYRYYGL (SVY) expressed in *Bifidobacterium brevis* bears resemblance to the epitope SIYRYYGL (SIY) found in tumors, enabling SVY-specific T cells to recognize and suppress tumor growth (65). Similarly, the antigenic epitope tail tape measure protein 1 (TMP1) within the genome of the bacteriophage *Enterococcus hirae* shares similarities with the tumor antigen proteasome β subunit 4 (PSMB4). This molecular mimicry can activate CD8⁺ T cells and enhance the efficacy of PD-1 blocking therapy (62). Numerous prior studies, as highlighted by Ting et al., have established a link between various autoimmune diseases and molecular mimicry involving antigens derived from microorganisms (66, 67). Given the vast proteomic diversity of the intestinal microbiome, it is conceivable that additional microbial antigens exhibiting high homology with cancer antigens may be identified (68).

4.2 Identification of microbial antigens as cancer neoantigens for cancer therapy

Zitvogel et al. proposed two hypotheses to elucidate the microbial mechanisms underlying tumor immune surveillance (44). Alongside the non-antigen pathway, which involves microorganisms modulating T cell anti-tumor activity through metabolites derived from pathogen-associated molecular patterns (PAMPs), an antigen pathway exists (69). In this antigen pathway, microbial antigens bearing high similarity to tumor antigens activate specific anti-tumor T cells, thereby influencing the immune system and inducing T cell cross-reactivity (44, 70).

Microbial antigens (Ags) encompass surface or internal molecules of microorganisms capable of eliciting immune responses. These substances include various bacterial components such as flagellin, lipopolysaccharides (LPS), cyclic dinucleotides, and bacterial metabolites including toxins, monophosphate lipids, and

transforming growth factors (55). Of particular interest is the principle of microbial resistance, which underscores the recognition of tumor-associated bacterial peptides sharing homology with human peptides. Research by Kalaora et al. demonstrated that polypeptides produced by bacteria infiltrating tumor cells can be displayed on the surface of tumor cells, subsequently recognized by the human immune system (63). Through the utilization of 16S rRNA gene sequencing and human leukocyte antigen (HLA) peptidomics, they identified a peptide library derived from intracellular bacteria, revealing 248 HLA-I peptides and 35 HLA-II peptides from 41 bacterial species across 17 melanoma metastases from 9 patients. These bacterial polypeptides constitute a novel class of tumor antigens previously unrecognized (63). Compared to human HLA peptides, bacterial peptides exhibit noticeably higher hydrophobicity, rendering them more conducive to antigen presentation and T cell recognition (63, 71, 72). In addition, clustering analysis of HLA-I peptides indicates that bacterial peptides typically range between 8 and 13 amino acids in length, exhibiting relatively low complexity. This structural simplicity may facilitate immune recognition and subsequent presentation to immune cells for eliciting immune responses (63). It is noteworthy that the selection of bacterial species for immunotherapy should favor those with known negative impacts on host responses and immune system function (63), rather than "protective" bacteria (73, 74).

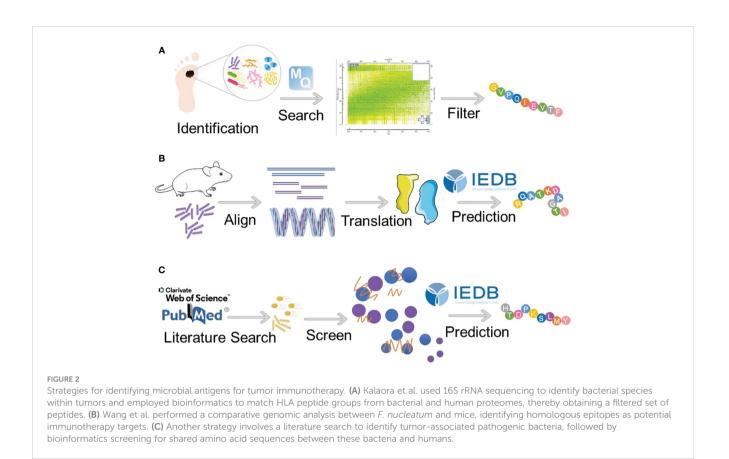
Therefore, we emphasize that the presence of cancer cells and bacteria within the tumor microenvironment may serve as a novel source of epitopes for cancer immunotherapy. The advantages of utilizing these bacterial peptides are manifold: Firstly, neoantigens bearing homology to microbial peptides exhibit heightened immunogenicity compared to non-homologous antigens (9); Secondly, from a host perspective, bacterial peptides are foreign to the host organism, potentially eliciting a more robust immune response (75); Furthermore, microbial molecular mimicry can impede tumor growth by eliciting T cell cross-reactivity (9); Lastly, antigens sharing homology with pathogenic bacteria can be efficiently expressed within bacterial vectors, offering significant convenience for the development of tumor vaccines (76).

4.3 Methods and tools for predicting microbial antigen peptides

In the paper by Naghavian et al., the authors investigated the potential activation of microbial peptides on tumor-infiltrating lymphocytes in glioblastoma, with the objective of elucidating whether microbial antigens could augment immunoreactivity against glioblastomas (77). Their experimental approach focused on assessing immune peptides that bind to HLA class II bacterial peptides in glioblastoma patients. Meanwhile, Kalaora et al. employed 16S rRNA sequencing to identify bacterial species within corresponding tumors. Subsequently, they conducted an analysis of the proteome of the identified bacterial species alongside the human proteome to extract relevant HLA peptide groups, thereby deriving qualified filter peptides (refer to Figure 2A). Notably, a key step in their analysis involved comparing the HLA peptide repertoire with the proteome of the corresponding bacteria (63). Furthermore, in the study conducted by Wang et al., a comparative analysis between the genome of *Fusobacterium nucleatum* (*F. nucleatum*) and the mouse genome unveiled the presence of neoantigens originating from bacteria (78). These neoantigens were observed to lack sequence similarity with mice, as well as homologous epitopes sharing similarity between bacteria and mice (see Figure 2B).

In this review, we present a method akin to those employed by Kalaora et al. and Wang et al. for the identification of humanized bacterial antigen peptides (63, 78). Our approach begins by compiling a comprehensive list of previously reported pathogenic bacteria known to be associated with tumors. Subsequently, utilizing sophisticated bioinformatics tools, we conduct sequence analyses to identify amino acid sequences within these bacteria that exhibit a high degree of similarity to those present in the human proteome. Following sequence identification, the identified amino acid sequences are subjected to rigorous computational analysis within the Immune Epitope Database (IEDB). Leveraging advanced algorithms such as NetMHCPan, we predict the potential for T cell major MHC-I antigen presentation. The final step of our method involves the meticulous selection of bacterial antigen peptides based on their robust predictive scores and strong affinity for MHC binding. This rigorous selection process ensures the prioritization of peptides with the highest likelihood of eliciting potent T cell responses against tumor cells (Figure 2C).

Researchers have developed numerous readily accessible software or tools similar to NetMHCpan for predicting antigenic peptides currently (Table 1) (79-88). For instance, MHCpred, based on machine learning algorithms, can rapidly and accurately identify potential MHC-binding peptides by predicting the affinity between peptides and MHC molecules (81). While software such as MaxQuant is not specifically designed for antigen peptide prediction, it aids in the identification of potential antigenic peptides through the analysis of proteomic data (87). MHC molecules consist of two main variants, MHC-I and MHC-II. MHC-I predominantly binds peptides derived from intracellular proteins, while MHC-II mainly interacts with peptides originating from extracellular proteins. Both MHC systems function by presenting non-self protein-derived peptides to T cells, thereby regulating responses against foreign entities (89). The binding between MHC and antigenic peptides represents the most selective step in the antigen presentation pathway, highlighting the significance of predicting peptide-MHC interactions for anticipating the specificity of T cell immune responses (80). In addition, MHC-I molecules bind peptides typically ranging from 8 to 10 amino acid residues, with the ability to accommodate longer peptide ligands (90-92). MHC-II molecules bind peptides of slightly larger average size, usually in the range of 10 to 15 amino acid residues, but can also interact with much longer amino acid sequences (93). In experimental practice, we can adjust and choose appropriate parameters within these software or tools based on the characteristics of MHC molecules to predict antigenic peptides effectively.



4.4 Immunogenicity of microbial peptides

Immunogenicity, as defined, encompasses the capacity of a molecule or substance to provoke an immune response, reflecting the intensity or magnitude of that response (94). When the innate immune system proves insufficient in fully eradicating a pathogen infection, the adaptive immune system, constituting the second line of defense, comes into play. In scenarios where specific antibodies have been generated from prior natural infections or vaccinations, the adaptive immune system is primed to mount a direct response upon encountering corresponding pathogens (95). An effective adaptive immune response hinges upon the precise pairing of an epitope with complementary T lymphocyte receptors (TcRs) (95). Importantly, immunogenicity and immunological memory are intimately linked. Immunological memory, a hallmark of adaptive immunity, can be engendered within the central or peripheral immune systems and stands as a primary outcome of successful specific responses (96). A robust immune response is typified by the sustained production of effector lymphocytes. Notably, the immunogenicity of an antigen inversely correlates with the quantity of antigen required to trigger an immune response. Consequently, heightened immunogenicity translates to reduced antigen demand for eliciting a response, fostering stronger peripheral memory and bolstering affinity in epitope or target interactions. In essence, the magnitude of immunogenicity profoundly influences the potency and duration of the immune response (95).

Indeed, the immunogenicity of antigens is governed by a multitude of factors, encompassing protein expression, antigen processing and presentation, peptide-MHC binding affinity, and the stability of resultant complexes, among others (97). Strong immune responses elicited by antigenic peptides underscore the efficient binding and presentation of these peptides by MHC molecules to active TcRs. In addition, the capacity of numerous MHC molecules to bind specific peptides suggests a relatively weak MHC restriction, permitting the presentation of these peptides to T cells by various MHC molecule types (95, 98). At the core of peptide immunogenicity lies antigen presentation, comprising the delivery of antigens to MHC molecules, loading of peptides onto MHC molecules, and subsequent display of MHC molecules at the cell surface (see Figure 3) (95, 98). The process is analogous to HLA molecules replacing the "housekeeping" peptide with a short amino acid sequence (8-20 aa) from the external antigen, aligning and complementing the antigen-binding groove on the HLA molecule (99). These amino acid sequences are securely anchored within the HLA binding groove through physicochemical interactions, facilitating engagement with corresponding TcRs on T lymphocytes. Subsequently, HLA molecules complexed with peptides are transported to the surface of antigen-presenting cells (APCs) via antigen presentation (95, 98). Moreover, the establishment of an optimal cellular milieu is imperative for fostering and sustaining the responsiveness of antigen-specific T cells and B cells. This involves epitope recognition and the interaction of specific TcRs with antigenic determinants.

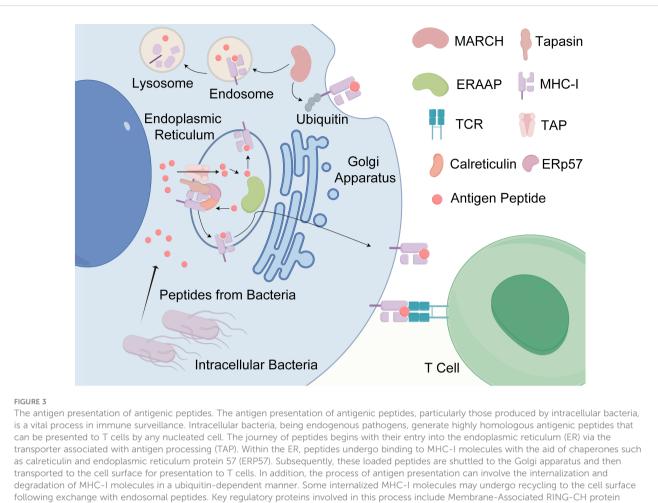
TABLE 1 The software or tools to predict antigenic peptides.

Software	Principle	Function	Туре	Reference
IEDB	Containing information on immune epitopes, the molecular targets of adaptive immune responses	Including tools for predicting peptide immunogenicity, such as MHC I/II binding prediction and T-cell epitope prediction	Database	(79)
NetMHCpan	Using artificial neural networks (ANN) and binding affinity and mass spectrometry eluted ligand data for training Predicting the binding of p sequences of MHC		Online Software	(80)
NetCTLpan	Combining predictions of proteasomal cleavage, transporter associated with antigen processing (TAP) transport efficiency, and MHC class I binding affinity to generate an MHC class I pathway likelihood score	Predicting for all MHC class I molecules with known protein sequence and allow predictions for 8-, 9-, 10-, and 11-mer peptides	Online Software	(81)
SYFPEITHI	Providing extensive MHC I and MHC II binding data	Predicting peptide binding	Database	(82)
MixMHCpred	Describing preferred peptide sequences established for each allele by a mixture model based on position weight matrices (pwms), and train on mass spectrometry elution data alone	Scoring different peptides and prioritizing the most likely hla-i ligands. As it is trained on naturally presented Peptides	Software	(83)
MHCflurry	Training the antigen processing model combining new models for MHC class I binding and antigen processing	predicting the comprehensive MHC class I presentation	Software	(84)
ProPred	Providing extensive MHC binding data	Predicting the ability of MHC class I and MHC class II molecules to bind peptides	Online Software	(85)
RANKPEP	Providing a position specific scoring matrix (PSSM) or profiling derived from a set of peptides known to bind to a specific MHC molecule	Predicting whether other peptides might bind and anticipating possible T-cell epitopes within a protein	Online Software	(86)
MaxQuant	For mass spectrometry (MS)-based analysis	Mainly using for proteomics research, but can also being used to help identify potential antigenic peptides in specific cases	Software	(87)
DeepNeo	Training models using deep learning algorithms combined with large-scale genomic and epigenomic data from tumor samples	Predicting the immunogenic neoantigen	Software	(88)

Furthermore, the balance between cytokines and other immune mediators considerably influences the outcomes of T cell activation and differentiation processes. Consequently, this intricate interplay can either amplify or attenuate the inherent immunogenicity of antigenic peptides (95, 100).

4.5 Validation of microbial peptide immunogenicity

To validate the activity of predicted microbial peptides *in vivo*, it is imperative to substantiate their immunogenicity through a series of rigorous laboratory experiments (8). Presently, *in vitro* validation procedures for immunogenicity encompass HLA binding assays and immunological analyses aimed at determining whether the selected peptides can be recognized by existing T cells. A crucial tool in this validation process is the Class I peptide binding assay, a classic competitive assay method. This assay relies on the inhibitory effect exerted by a specific radiolabeled peptide segment with high affinity, which competes for binding to purified MHC molecules harboring specific allele genes. Through this method, the quantitative measurement of the peptide's binding capacity to MHC Class I molecules is achievable (101). In addition, the MHC Class I multimer binding assay plays a crucial role in elucidating whether particular peptide segments are capable of eliciting recognition by T cells via MHC Class I molecules. This assay is crucial for assessing the immunogenicity of peptide segments and their potential to stimulate immune responses (102-104). Moreover, the nanoparticle (NP) pull-down assay serves as a valuable technique in investigating protein interactions, particularly pertinent in the realms of immunology and cancer research. This assay facilitates the exploration of interactions between peptide-MHC I complexes and T cell receptors, providing insights into the immunological responses induced by the peptides (105). The selection of appropriate methods for immunogenicity validation hinges upon the specific objectives and requirements of the study at hand. For instance, in the study conducted by Wang et al., the researchers opted to employ an ELISpot assay coupled with T cell stimulation utilizing candidate peptides presented by dendritic cells. This methodological approach enabled them to gauge the specific T cell response triggered by the treatment regimen, thereby confirming the immunogenic activity of the peptides within the context of their study's objectives (78). By employing these rigorous validation techniques, researchers can



(MARCH) and ER-associated aminopeptidases (ERAP) (created by Figdraw).

ascertain the immunogenicity of predicted microbial peptides, laying a solid foundation for their potential application *in vivo* and furthering our understanding of their therapeutic efficacy in cancer treatment.

4.6 Application of microbial antigen

Neoantigens, akin to bacterial antigenic peptides (Table 2) (49, 62, 63, 65, 77, 78, 106–111), emerge as promising candidates for tumor vaccines, capable of inciting tumor-specific immune responses. In mice, cross-reactivity between symbiotic bacteria and tumor antigens has been demonstrated to initiate anti-tumor responses (65). Specifically, $CD8^+$ T cells have shown proficiency in recognizing peptides derived from *E. hirae* bacteriophage and tumor antigens through molecular mimicry cross-recognition (62). An approved therapeutic approach for bladder cancer involves the direct infusion of *Mycobacterium bovis* strain extract into the bladder, stimulating a helper T cell 1 (TH1) immune response from CD4⁺ T cells and imparting enduring protection in murine models (112, 113). Furthermore, Naghavian et al.'s findings illustrate that T cell clones TCC88 can directly target neoantigens in glioblastoma, exhibiting robust responses not only towards various glioblastoma-derived peptides but also towards targets originating from a diverse array of bacteria and intestinal microflora. These T cell clones elicit cross-reactive responses against tumor targets in numerous tumor-infiltrating lymphocytes (TIL) and peripheral blood memory T cells (77). Moreover, Wang et al.'s study suggests that the depletion of bacteria within tumors exposes bacterial epitopes, thereby precipitating immune responses aimed at eradicating tumor cells. In addition, the presence of homologous epitopes shared by bacteria and the host remarkably contributes to anti-tumor immunity. In a model of carcinogenic *F. nucleatum* infection, T cells exhibited responsiveness to epitopes shared by both *F. nucleatum* and the host, underscoring the potential of microbial epitope exposure in augmenting cancer treatment efficacy (78).

5 Conclusion and prospect

Tumor neoantigens have long been investigated as potential targets for cancer therapy, yet challenges persist regarding their applicability on a large scale and their efficacy against immune escape mechanisms induced by tumor mutations. Conversely, tumor-associated antigens, while prevalent across various

TABLE 2 The potential bacterial antigenic peptides for cancer therapy.

Disease Type	Host Species	Bacterial Species	Homologous Bacte- rial Peptides	Reference
Melanoma	Homo sapiens	Bacillus polymyxa Escherichia coli	GAGIGVLTA QAGIGILLA	(106)
Melanoma	Homo sapiens	Chlamydia trachomatis	MLSGIGIFFI	(107)
Melanoma	Homo sapiens	Mycoplasma penetrans	YIFAACLLLI	(108)
Pancreatic cancer	Homo sapiens	Francisella tularensis	KLLPEGYWV	(48)
Non-small-cell lung cancer	Homo sapiens	bacteriophage Enterococcus hirae	TSLARFANI KLAKFASVV	(62)
Melanoma	Mus musculus Homo sapiens	Bifidobacterium breve Fusobacterium nucleatum Staphylococcus aureus Staphylococcus capitis	SVYRYYGL(SVY) ITELNSPVL, ITNTGAVTV, SLTDKISII, SVVVDELFEV, VLTDTYLTL EGNIDFITL, LSDLGKSIY AIGVAASILY, LEGTVLDTL, TPIVAVNAL, VTSGVTAAY	(65)
Colon cancer	Mus musculus	Ruminococcaceae Bacteroidales/Prevotella/ Muribaculacee Bacteria_unclassified Duncaniella/ Bacteroides/Bacteroidales	RLAGFFPRL LGPWRSGGVL SLPGSWRSL YIALFDGFI	(109)
Type 1 diabetes	Homo sapiens	Parabacteroides distasonis 33B Bacteroides sp.CAG:144 Ruminococcus gnavus CAG:126 Coprococcus eutactus Clostridiales bacterium VE202–14 Corynebacterium genitalium Lactobacillus vaginalis Lymphocystis disease virus 1 Cyprinid herpesvirus 1 Grouper iridovirus Burkholderia multivorans Bradyrhizobium japonicum SEMIA 5079 Streptomyces griseus Tetrasphaera japonica T1-X7 Saccharomonospora halophila Brevundimonas sp.BAL3 Metarhizium robertsif	RILVELLYLVCSEYL LDFKEALYLGCGDRT DPRRSALYLFCGKRC NHDKEALYIYCDETE VRAGYALFLVCDEEK FVHEDALHLVCGERI LOSMEIPYLVCGERE AHLVAALORVCGNRG SHPNVFIALVCGERG GELIDALTEHCGDRG LHLARALYEMCGEFP VSGKHALYLYCGERG RDRVEALRLVCGEAM HWLVEIAYLVCGDRR TAHGVAEYLVCGERR WVGFETLYLYCGERI DHWDEAGFLVCGERG	(110)
Autoimmune diseases	Homo sapiens	Streptomyces Mobaraensis	PSRMKAVIYSKHF NESAPAASSAGP APAASSAGPSFRAP REVASVMNRALE LCTAGFMPSAGEAAA AADNGAGEETKSY	(111)
Colorectal cancer	Mus musculus	Fusobacterium nucleatum	GVPQIEVTF, RQATKDAGTI, LADDNFSTIV, LADDNFSTI, RGVPQIEVTF	(78)
Glioblastoma	Homo sapiens	Pectobacterium carotovorum Pectobacterium carotovorum Pectobacterium carotovorum Chlorobium phaeobacteroides	TPILVDGKDVMPEVN TPILVDGKDVMPE NTPILVDGKDVMPE ALPVIETQAGDVSAYIP	(77)

(Continued)

TABLE 2 Continued

Disease Type	Host Species	Bacterial Species	Homologous Bacte- rial Peptides	Reference
		Geobacter lovleyi Chlorobium phaeobacteroides Listeria welshimeri Chlamydia pneumoniae Chlamydia pneumoniae Geobacter lovleyi Chlamydia pneumoniae Pectobacterium carotovorum Buchnera aphidicola Geobacter lovleyi Streptomyces coelicolor Geobacter lovleyi Moorella thermoacetica	LNVLRIINEPTAAA LPVIETQAGDVSAYIP PAPATTFAHLD LQNIIPASTGAAK LQNIIPASTGAAKA NIIPASTGAAKA NVLRIINEPTAAA QNIIPASTGAAKA SNTPILVDGKDVMPE FRVPTANV GLNVLRIINEPTAAA DPAPATTFAHLDATTVLSR LNVLRIINEPTAA NPVDILTYVAWKISG	

malignancies, often elicit immune tolerance due to their expression in normal cells. Both neoantigens and tumor-associated antigens exhibit limited immunogenicity, highlighting the need for alternative approaches in cancer immunotherapy. Microbial antigen peptides, distinguished by their exogenous nature, possess robust immunogenicity, offering promising avenues for cancer treatment. Our review extensively explores the intricate interplay between microorganisms and tumorigenesis, shedding light on the immune mechanisms orchestrated by microbial antigens within the host immune system. By elucidating these mechanisms, we underscore the key role of microorganisms in tumor therapy and underscore the potential of microbial antigens in shaping the landscape of tumor immunotherapy. Studies have demonstrated that bacterial peptides with homologous epitopes to host antigens can induce potent immune responses, offering potential for antitumor therapy (63, 78).

Hence, the variation in intratumoral microorganisms dictates the presence of microbial antigens in a tumor-specific manner analogous to TAAs. For example, Clostridium difficile (C. difficile) is an important pathogenic bacterium that leads to colorectal cancer. Therefore, it is necessary to focus on C. difficile to explore microbial antigens for the prevention and treatment of colorectal cancer. Current research indicates that there are hundreds to thousands of different microbes present in tumors, but apart from C. difficile in colorectal cancer and human papilloma virus that can induce uterine cancer, there is still a lack of consensus on the major microbiota in other tumors. With the further study of microbiota in tumors, some specific bacteria are gradually being discovered. For example, in non-small cell lung cancer (NSCLC) tissues, the presence of specific taxa such as Prevotella, Streptococcus and Veillonella can lead to Phosphoinositide 3-kinase (PI3K) and protein kinase B(AKT) signaling activation (114). Pernigoni et al. found that specific Ruminococcus isolates were enriched in patients with castration-resistant prostate cancer (CRPC) and that these bacteria were able to synthesize dehydroepiandrosterone (DHEA) from pregnenolone, a precursor of testosterone (114). Developing bacterial antigens tailored to different tumor subtypes based on specific bacteria present in various tumors will be an important direction for identifying and characterizing bacterial antigens in the near future. Therefore, strategic integration of microbial antigen peptides into cancer therapy holds promise for enhancing immune responses against tumors.

We propose several avenues for leveraging microbial antigens in cancer therapy. Firstly, incorporation of microbial antigens into cancer vaccines or immunotherapeutic regimens can exploit their inherent immunogenicity to prime the immune system against cancer cells. In addition, the phenomenon of molecular mimicry between microbial and cancer antigens presents opportunities for developing personalized cancer immunotherapies based on individual microbial exposure histories. Moreover, therapeutic cancer vaccines incorporating microbial antigens offer potential for targeted cancer cell recognition and elimination, particularly when combined with tumor-specific antigens and advanced vaccine delivery systems. In practical clinical applications, we recommend using bacterial peptides identified by aligning the human reference genome or proteome with intratumoral bacteria for a more broadspectrum cancer vaccine. These peptides can be encapsulated within diverse carriers, including liposomes, hydrogels, and engineered bacteria, and administered via routes like intramuscular and intravenous injections, as well as oral delivery. The process releases bacterial antigens, stimulating the generation of CD8⁺ T cells with anti-tumor capabilities in the body. For a more personalized treatment, identifying the bacterial species within the patient's tumor through metagenomics and aligning them with the patient's genome or proteome is necessary to select personalized bacterial homologous peptides. Furthermore, combination therapy approaches integrating microbial antigens with immune checkpoint inhibitors, chemotherapy, or other modalities hold promise for synergistically enhancing anti-tumor immune responses. By capitalizing on the complementary mechanisms of action of different agents, such combinatorial strategies offer potential for overcoming tumor immune evasion mechanisms and improving treatment outcomes.

Although microbial peptides have been recognized for their potential effects in tumor immune therapy, there are several challenges in clinical application. Of particular importance is the mutagenicity of tumors, which may hinder the long-term persistence of memory T cells stimulated by microbial antigen peptides, relegating them to a supporting role rather than a primary therapeutic modality. At second, the abundance of intratumoral microorganisms makes the screening for anti-tumor bacterial antigens even more challenging. Although computerassisted screening has significantly narrowed down the range of peptides, the diversity of microorganisms means that each type of tumor will produce a vast array of microbial peptides. How to achieve high-throughput screening of tumor antigens will be a major challenge in developing anti-tumor microbial antigens. Therefore, in practical applications, it is crucial to have a sensible screening process, which may involve more peptidomics approaches. Thirdly, the initial screening of tumor antigens relies more on animal experiments, and the significant differences between animals and humans pose a risk of peptide failure in human bodies. Furthermore, despite some research findings from in vitro and in vivo animal experiments, there is still a lack of sufficient clinical studies, especially, to fully demonstrate the effectiveness of microbial antigens. Last but not least, if our prediction involves aligning the human reference genome with the homology of intratumor bacteria, it might lead to immune cells activated by microbial antigen peptides attacking normal body cells instead of specifically targeting tumor cells.

In conclusion, microbial antigens represent a promising frontier in cancer therapy, offering unique advantages in terms of immunogenicity, cross-reactivity, and versatility. Moving forward, concerted efforts are warranted to further elucidate the mechanisms underlying the anti-tumor immune response elicited by microbial antigens and optimize their integration into clinical practice. By harnessing the power of microbial antigens, we can advance towards more effective and personalized cancer treatments, thereby transforming the landscape of cancer therapy.

Author contributions

TZ: Project administration, Software, Writing – original draft. XLZ: Data curation, Formal Analysis, Methodology, Validation, Writing – original draft. JC: Data curation, Formal analysis, Writing

References

1. Mellman I, Coukos G, Dranoff G. Cancer immunotherapy comes of age. *Nature*. (2011) 480:480–9. doi: 10.1038/nature10673

2. Renner K, Singer K, Koehl GE, Geissler EK, Peter K, Siska PJ, et al. Metabolic hallmarks of tumor and immune cells in the tumor microenvironment. *Front Immunol.* (2017) 8:248. doi: 10.3389/fimmu.2017.00248

3. Schumacher TN, Schreiber RD. Neoantigens in cancer immunotherapy. Sci (New York NY). (2015) 348:69–74. doi: 10.1126/science.aaa4971

 Sahin U, Türeci Ö. Personalized vaccines for cancer immunotherapy. Sci (New York NY). (2018) 359:1355–60. doi: 10.1126/science.aar7112

5. McGranahan N, Swanton C. Clonal heterogeneity and tumor evolution: past, present, and the future. Cell. (2017) 168:613-28. doi: 10.1016/j.cell.2017.01.018

6. Ribas A, Wolchok JD. Cancer immunotherapy using checkpoint blockade. Sci (New York NY). (2018) 359:1350-5. doi: 10.1126/science.aar4060

7. Gopalakrishnan V, Helmink BA, Spencer CN, Reuben A, Wargo JA. The influence of the gut microbiome on cancer, immunity, and cancer immunotherapy. *Cancer Cell.* (2018) 33:570–80. doi: 10.1016/j.ccell.2018.03.015

8. Wells DK, van Buuren MM, Dang KK, Hubbard-Lucey VM, Sheehan KCF, Campbell KM, et al. Key parameters of tumor epitope immunogenicity revealed

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through a consortium approach improve neoantigen prediction. *Cell.* (2020) 183:818–834.e813. doi: 10.1016/j.cell.2020.09.015

9. Tian J, Ma J. The value of microbes in cancer neoantigen immunotherapy. *Pharmaceutics*. (2023) 15:2138. doi: 10.3390/pharmaceutics15082138

10. Tagliamonte M, Cavalluzzo B, Mauriello A, Ragone C, Buonaguro FM, Tornesello ML, et al. Molecular mimicry and cancer vaccine development. *Mol cancer.* (2023) 22:75. doi: 10.1186/s12943-023-01776-0

11. Schumacher TN, Scheper W, Kvistborg P. Cancer neoantigens. Annu Rev Immunol. (2019) 37:173–200. doi: 10.1146/annurev-immunol-042617-053402

12. Kreiter S, Vormehr M, van de Roemer N, Diken M, Löwer M, Diekmann J, et al. Mutant MHC class II epitopes drive therapeutic immune responses to cancer. *Nature*. (2015) 520:692–6. doi: 10.1038/nature14426

13. Ott PA, Hu Z, Keskin DB, Shukla SA, Sun J, Bozym DJ, et al. An immunogenic personal neoantigen vaccine for patients with melanoma. *Nature*. (2017) 547:217–21. doi: 10.1038/nature22991

14. Leng Q, Tarbe M, Long Q, Wang F. Pre-existing heterologous T-cell immunity and neoantigen immunogenicity. *Clin Trans Immunol.* (2020) 9:e01111. doi: 10.1002/cti2.1111

15. Łuksza M, Riaz N, Makarov V, Balachandran VP, Hellmann MD, Solovyov A, et al. A neoantigen fitness model predicts tumour response to checkpoint blockade immunotherapy. *Nature*. (2017) 551:517–20. doi: 10.1038/nature24473

16. Strønen E, Toebes M, Kelderman S, van Buuren MM, Yang W, van Rooij N, et al. Targeting of cancer neoantigens with donor-derived T cell receptor repertoires. *Sci* (*New York NY*). (2016) 352:1337–41. doi: 10.1126/science.aaf2288

17. Sahin U, Derhovanessian E, Miller M, Kloke BP, Simon P, Löwer M, et al. Personalized RNA mutanome vaccines mobilize poly-specific therapeutic immunity against cancer. *Nature*. (2017) 547:222-6. doi: 10.1038/nature23003

18. van der Bruggen P, Traversari C, Chomez P, Lurquin C, De Plaen E, Van den Eynde B, et al. A gene encoding an antigen recognized by cytolytic T lymphocytes on a human melanoma. *Sci (New York NY).* (1991) 254:1643–7. doi: 10.1126/science.1840703

19. Gaugler B, Van den Eynde B, van der Bruggen P, Romero P, Gaforio JJ, De Plaen E, et al. Human gene MAGE-3 codes for an antigen recognized on a melanoma by autologous cytolytic T lymphocytes. *J Exp Med.* (1994) 179:921–30. doi: 10.1084/ jem.179.3.921

20. Buonaguro L, Tagliamonte M. Selecting target antigens for cancer vaccine development. Vaccines (Basel). (2020) 8:615. doi: 10.3390/vaccines8040615

21. Theobald M, Biggs J, Hernández J, Lustgarten J, Labadie C, Sherman LA. Tolerance to p53 by A2.1-restricted cytotoxic T lymphocytes. J Exp Med. (1997) 185:833–41. doi: 10.1084/jem.185.5.833

22. Buonaguro L, Petrizzo A, Tornesello ML, Buonaguro FM. Translating tumor antigens into cancer vaccines. *Clin Vaccine immunol: CVI.* (2011) 18:23-34. doi: 10.1128/CVI.00286-10

23. Dudley ME, Wunderlich JR, Robbins PF, Yang JC, Hwu P, Schwartzentruber DJ, et al. Cancer regression and autoimmunity in patients after clonal repopulation with antitumor lymphocytes. *Sci (New York NY).* (2002) 298:850–4. doi: 10.1126/science.1076514

24. Chen Y, Liu B, Wei Y, Kuang DM. Influence of gut and intratumoral microbiota on the immune microenvironment and anti-cancer therapy. *Pharmacol Res.* (2021) 174:105966. doi: 10.1016/j.phrs.2021.105966

25. Nejman D, Livyatan I, Fuks G, Gavert N, Zwang Y, Geller LT, et al. The human tumor microbiome is composed of tumor type-specific intracellular bacteria. *Sci (New York NY).* (2020) 368:973–80. doi: 10.1126/science.aay9189

26. Wang M, Yu F, Li P. Intratumor microbiota in cancer pathogenesis and immunity: from mechanisms of action to therapeutic opportunities. *Front Immunol.* (2023) 14:1269054. doi: 10.3389/fimmu.2023.1269054

27. Fu A, Yao B, Dong T, Chen Y, Yao J, Liu Y, et al. Tumor-resident intracellular microbiota promotes metastatic colonization in breast cancer. *Cell.* (2022) 185:1356–1372.e1326. doi: 10.1016/j.cell.2022.02.027

28. Galeano Niño JL, Wu H, LaCourse KD, Kempchinsky AG, Baryiames A, Barber B, et al. Effect of the intratumoral microbiota on spatial and cellular heterogeneity in cancer. *Nature*. (2022) 611:810–7. doi: 10.1038/s41586-022-05435-0

29. Xie Y, Xie F, Zhou X, Zhang L, Yang B, Huang J, et al. Microbiota in tumors: from understanding to application. *Adv Sci (Weinheim Baden-Wurttemberg Germany).* (2022) 9:e2200470. doi: 10.1002/advs.202200470

30. Knippel RJ, Drewes JL, Sears CL. The cancer microbiome: recent highlights and knowledge gaps. *Cancer discov.* (2021) 11:2378–95. doi: 10.1158/2159-8290.CD-21-0324

31. Pushalkar S, Hundeyin M, Daley D, Zambirinis CP, Kurz E, Mishra A, et al. The pancreatic cancer microbiome promotes oncogenesis by induction of innate and adaptive immune suppression. *Cancer discov.* (2018) 8:403–16. doi: 10.1158/2159-8290.CD-17-1134

32. Geller LT, Barzily-Rokni M, Danino T, Jonas OH, Shental N, Nejman D, et al. Potential role of intratumor bacteria in mediating tumor resistance to the chemotherapeutic drug gemcitabine. *Sci (New York NY).* (2017) 357:1156-60. doi: 10.1126/science.aah5043

33. Riquelme E, Zhang Y, Zhang L, Montiel M, Zoltan M, Dong W, et al. Tumor microbiome diversity and composition influence pancreatic cancer outcomes. *Cell.* (2019) 178:795–806.e712. doi: 10.1016/j.cell.2019.07.008

34. Zitvogel L, Daillère R, Roberti MP, Routy B, Kroemer G. Anticancer effects of the microbiome and its products. *Nat Rev Microbiol.* (2017) 15:465–78. doi: 10.1038/nrmicro.2017.44

35. O'Donnell JS, Teng MWL, Smyth MJ. Cancer immunoediting and resistance to T cell-based immunotherapy. *Nat Rev Clin Oncol.* (2019) 16:151–67. doi: 10.1038/ s41571-018-0142-8

36. Zhang Y, Zhang Z. The history and advances in cancer immunotherapy: understanding the characteristics of tumor-infiltrating immune cells and their therapeutic implications. *Cell Mol Immunol.* (2020) 17:807–21. doi: 10.1038/s41423-020-0488-6

37. Wei SC, Duffy CR, Allison JP. Fundamental mechanisms of immune checkpoint blockade therapy. *Cancer discov.* (2018) 8:1069–86. doi: 10.1158/2159-8290.CD-18-0367

38. Pesce S, Greppi M, Grossi F, Del Zotto G, Moretta L, Sivori S, et al. PD/1-PD-ls checkpoint: insight on the potential role of NK cells. *Front Immunol.* (2019) 10:1242. doi: 10.3389/fimmu.2019.01242

39. SChadendorf D, Hodi FS, Robert C, Weber JS, Margolin K, Hamid O, et al. Pooled analysis of long-term survival data from phase II and phase III trials of ipilimumab in unresectable or metastatic melanoma. *J Clin Oncol.* (2015) 33:1889– 94. doi: 10.1200/JCO.2014.56.2736

40. Anagnostou V, Smith KN, Forde PM, Niknafs N, Bhattacharya R, White J, et al. Evolution of neoantigen landscape during immune checkpoint blockade in non-small cell lung cancer. *Cancer discov.* (2017) 7:264–76. doi: 10.1158/2159-8290.CD-16-0828

41. Magen A, Hamon P, Fiaschi N, Soong BY, Park MD, Mattiuz R, et al. Intratumoral dendritic cell-CD4(+) T helper cell niches enable CD8(+) T cell differentiation following PD-1 blockade in hepatocellular carcinoma. *Nat Med.* (2023) 29:1389–99. doi: 10.1038/s41591-023-02345-0

42. Kennedy LB, Salama AKS. A review of cancer immunotherapy toxicity. CA: Cancer J Clin. (2020) 70:86–104. doi: 10.3322/caac.21596

43. Yang K, Halima A, Chan TA. Antigen presentation in cancer - mechanisms and clinical implications for immunotherapy. *Nat Rev Clin Oncol.* (2023) 20:604–23. doi: 10.1038/s41571-023-00789-4

44. Zitvogel L, Ayyoub M, Routy B, Kroemer G. Microbiome and anticancer immunosurveillance. Cell. (2016) 165:276-87. doi: 10.1016/j.cell.2016.03.001

45. Zhang L, Jiang L, Yu L, Li Q, Tian X, He J, et al. Inhibition of UBA6 by inosine augments tumour immunogenicity and responses. *Nat Commun.* (2022) 13:5413. doi: 10.1038/s41467-022-33116-z

46. Griffin ME, Espinosa J, Becker JL, Luo JD, Carroll TS, Jha JK, et al. Enterococcus peptidoglycan remodeling promotes checkpoint inhibitor cancer immunotherapy. *Sci* (*New York NY*). (2021) 373:1040–6. doi: 10.1126/science.abc9113

47. Zitvogel L, Kroemer G. Cross-reactivity between microbial and tumor antigens. *Curr Opin Immunol.* (2022) 75:102171. doi: 10.1016/j.coi.2022.102171

48. Tanoue T, Morita S, Plichta DR, Skelly AN, Suda W, Sugiura Y, et al. A defined commensal consortium elicits CD8 T cells and anti-cancer immunity. *Nature*. (2019) 565:600–5. doi: 10.1038/s41586-019-0878-z

49. Balachandran VP, Luksza M, Zhao JN, Makarov V, Moral JA, Remark R, et al. Identification of unique neoantigen qualities in long-term survivors of pancreatic cancer. *Nature*. (2017) 551:512–6. doi: 10.1038/nature24462

50. Al-Saafeen BH, Fernandez-Cabezudo MJ, Al-Ramadi BK. Integration of salmonella into combination cancer therapy. *Cancers*. (2021) 13:3228. doi: 10.3390/ cancers13133228

51. Guo Y, Chen Y, Liu X, Min JJ, Tan W, Zheng JH. Targeted cancer immunotherapy with genetically engineered oncolytic Salmonella typhimurium. *Cancer lett.* (2020) 469:102-10. doi: 10.1016/j.canlet.2019.10.033

52. Chen MC, Pangilinan CR, Lee CH. Salmonella breaks tumor immune tolerance by downregulating tumor programmed death-ligand 1 expression. *Cancers.* (2019) 12:57. doi: 10.3390/cancers12010057

53. Gao G, Ma T, Zhang T, Jin H, Li Y, Kwok LY, et al. Adjunctive Probiotic Lactobacillus rhamnosus Probio-M9 Administration Enhances the Effect of Anti-PD-1 Antitumor Therapy via Restoring Antibiotic-Disrupted Gut Microbiota. *Front Immunol.* (2021) 12:772532. doi: 10.3389/fimmu.2021.772532

54. Jenkins RW, Barbie DA, Flaherty KT. Mechanisms of resistance to immune checkpoint inhibitors. *Br J cancer.* (2018) 118:9–16. doi: 10.1038/bjc.2017.434

55. Zhou M, Tang Y, Xu W, Hao X, Li Y, Huang S, et al. Bacteria-based immunotherapy for cancer: a systematic review of preclinical studies. *Front Immunol.* (2023) 14:1140463. doi: 10.3389/fimmu.2023.1140463

56. Kuhn S, Hyde EJ, Yang J, Rich FJ, Harper JL, Kirman JR, et al. Increased numbers of monocyte-derived dendritic cells during successful tumor immunotherapy with immune-activating agents. *J Immunol (Baltimore Md: 1950)*. (2013) 191:1984–92. doi: 10.4049/jimmunol.1301135

57. Jeong H, Lee SY, Seo H, Kim DH, Lee D, Kim BJ. Potential of Mycobacterium tuberculosis chorismate mutase (Rv1885c) as a novel TLR4-mediated adjuvant for dendritic cell-based cancer immunotherapy. *Oncoimmunology.* (2022) 11:2023340. doi: 10.1080/2162402X.2021.2023340

58. Coria LM, Ibañez AE, Tkach M, Sabbione F, Bruno L, Carabajal MV, et al. A brucella spp. Protease inhibitor limits antigen lysosomal proteolysis, increases cross-presentation, and enhances CD8+ T cell responses. *J Immunol (Baltimore Md: 1950).* (2016) 196:4014–29. doi: 10.4049/jimmunol.1501188

59. Portevin D, Young D. Natural killer cell cytokine response to M. bovis BCG Is associated with inhibited proliferation, increased apoptosis and ultimate depletion of NKp44 (+)CD56(bright) cells. *PloS One.* (2013) 8:e68864. doi: 10.1371/journal.pone.0068864

60. Esteso G, Aguiló N, Julián E, Ashiru O, Ho MM, Martín C, et al. Natural killer anti-tumor activity can be achieved by *in vitro* incubation with heat-killed BCG. *Front Immunol.* (2021) 12:622995. doi: 10.3389/fimmu.2021.622995

61. Lin Q, Rong L, Jia X, Li R, Yu B, Hu J, et al. IFN-γ-dependent NK cell activation is essential to metastasis suppression by engineered Salmonella. *Nat Commun.* (2021) 12:2537. doi: 10.1038/s41467-021-22755-3

62. Fluckiger A, Daillère R, Sassi M, Sixt BS, Liu P, Loos F, et al. Cross-reactivity between tumor MHC class I-restricted antigens and an enterococcal bacteriophage. *Sci* (*New York NY*). (2020) 369:936–42. doi: 10.1126/science.aax0701

63. Kalaora S, Nagler A, Nejman D, Alon M, Barbolin C, Barnea E, et al. Identification of bacteria-derived HLA-bound peptides in melanoma. *Nature.* (2021) 592:138–43. doi: 10.1038/s41586-021-03368-8

64. Damian RT. Molecular mimicry: antigen sharing by parasite and host and its consequences. Am Nat. (1964) 98:129-149. doi: 10.1086/282313

65. Bessell CA, Isser A, Havel JJ, Lee S, Bell DR, Hickey JW, et al. Commensal bacteria stimulate antitumor responses via T cell cross-reactivity. *JCI Insight*. (2020) 5: e135597. doi: 10.1172/jci.insight.135597

66. Gil-Cruz C, Perez-Shibayama C, De Martin A, Ronchi F, van der Borght K, Niederer R, et al. Microbiota-derived peptide mimics drive lethal inflammatory cardiomyopathy. *Sci (New York NY).* (2019) 366:881–6. doi: 10.1126/science.aav3487

67. Greiling TM, Dehner C, Chen X, Hughes K, Iñiguez AJ, Boccitto M, et al. Commensal orthologs of the human autoantigen Ro60 as triggers of autoimmunity in lupus. *Sci Trans Med.* (2018) 10:eaan2306. doi: 10.1126/scitranslmed.aan2306

68. Ting NL, Lau HC, Yu J. Cancer pharmacomicrobiomics: targeting microbiota to optimise cancer therapy outcomes. *Gut.* (2022) 71:1412–25. doi: 10.1136/gutjnl-2021-326264

69. Bai L, Li W, Zheng W, Xu D, Chen N, Cui J. Promising targets based on pattern recognition receptors for cancer immunotherapy. *Pharmacol Res.* (2020) 159:105017. doi: 10.1016/j.phrs.2020.105017

70. Hayase E, Jenq RR. Role of the intestinal microbiome and microbial-derived metabolites in immune checkpoint blockade immunotherapy of cancer. *Genome Med.* (2021) 13:107. doi: 10.1186/s13073-021-00923-w

71. Vigneron N, Abi Habib J, Van den Eynde BJ. Learning from the proteasome how to fine-tune cancer immunotherapy. *Trends cancer*. (2017) 3:726–41. doi: 10.1016/j.trecan.2017.07.007

72. Chowell D, Krishna S, Becker PD, Cocita C, Shu J, Tan X, et al. TCR contact residue hydrophobicity is a hallmark of immunogenic CD8+ T cell epitopes. *Proc Natl Acad Sci United States America*. (2015) 112:E1754–1762. doi: 10.1073/pnas.1500973112

73. Gopalakrishnan V, Spencer CN, Nezi L, Reuben A, Andrews MC, Karpinets TV, et al. Gut microbiome modulates response to anti-PD-1 immunotherapy in melanoma patients. *Sci (New York NY).* (2018) 359:97–103. doi: 10.1126/science.aan4236

74. Matson V, Fessler J, Bao R, Chongsuwat T, Zha Y, Alegre ML, et al. The commensal microbiome is associated with anti-PD-1 efficacy in metastatic melanoma patients. *Sci (New York NY).* (2018) 359:104–8. doi: 10.1126/science.aao3290

75. Iwasaki A, Medzhitov R. Control of adaptive immunity by the innate immune system. *Nat Immunol.* (2015) 16:343–53. doi: 10.1038/ni.3123

76. Schreiber RD, Old LJ, Smyth MJ. Cancer immunoediting: integrating immunity's roles in cancer suppression and promotion. *Sci (New York NY).* (2011) 331:1565–70. doi: 10.1126/science.1203486

77. Naghavian R, Faigle W, Oldrati P, Wang J, Toussaint NC, Qiu Y, et al. Microbial peptides activate tumour-infiltrating lymphocytes in glioblastoma. *Nature*. (2023) 617:807–17. doi: 10.1038/s41586-023-06081-w

78. Wang M, Rousseau B, Qiu K, Huang G, Zhang Y, Su H, et al. Killing tumorassociated bacteria with a liposomal antibiotic generates neoantigens that induce antitumor immune responses. *Nat Biotechnol.* (2023). doi: 10.1038/s41587-023-01957-8

79. Vita R, Overton JA, Greenbaum JA, Ponomarenko J, Clark JD, Cantrell JR, et al. The immune epitope database (IEDB) 3.0. *Nucleic Acids Res.* (2015) 43:D405–412. doi: 10.1093/nar/gku938

80. Reynisson B, Alvarez B, Paul S, Peters B, Nielsen M. NetMHCpan-4.1 and NetMHCIIpan-4.0: improved predictions of MHC antigen presentation by concurrent motif deconvolution and integration of MS MHC eluted ligand data. *Nucleic Acids Res.* (2020) 48:W449–w454. doi: 10.1093/nar/gkaa379

81. Guan P, Hattotuwagama CK, Doytchinova IA, Flower DR. MHCPred 2.0: an updated quantitative T-cell epitope prediction server. *Appl Bioinf.* (2006) 5:55–61. doi: 10.2165/00822942-200605010-00008

82. Schuler MM, Nastke MD, Stevanovikć S. SYFPEITHI: database for searching and T-cell epitope prediction. *Methods Mol Biol.* (2007) 409:75–93. doi: 10.1007/978-1-60327-118-9_5

83. Gfeller D, Schmidt J, Croce G, Guillaume P, Bobisse S, Genolet R, et al. Improved predictions of antigen presentation and TCR recognition with MixMHCpred2.2 and PRIME2.0 reveal potent SARS-CoV-2 CD8(+) T-cell epitopes. *Cell sys.* (2023) 14:72–83.e75. doi: 10.1016/j.cels.2022.12.002

84. O'Donnell TJ, Rubinsteyn A, Laserson U. MHCflurry 2.0: improved pan-allele prediction of MHC class I-presented peptides by incorporating antigen processing. *Cell sys.* (2020) 11:42–48.e47. doi: 10.1016/j.cels.2020.06.010

85. Singh H, Raghava GP. ProPred: prediction of HLA-DR binding sites. *Bioinf* (Oxford England). (2001) 17:1236-7. doi: 10.1093/bioinformatics/17.12.1236

86. Reche PA, Glutting J-P, Reinherz EL. Prediction of MHC class I binding peptides using profile motifs. *Hum Immunol.* (2002) 63:701–9. doi: 10.1016/S0198-8859(02) 00432-9

87. Tyanova S, Temu T, Cox J. The MaxQuant computational platform for mass spectrometry-based shotgun proteomics. *Nat Protoc.* (2016) 11:2301–19. doi: 10.1038/nprot.2016.136

88. Kim JY, Bang H, Noh SJ, Choi JK. DeepNeo: a webserver for predicting immunogenic neoantigens. *Nucleic Acids Res.* (2023) 51:W134-w140. doi: 10.1093/nar/gkad275

89. Blum JS, Wearsch PA, Cresswell P. Pathways of antigen processing. Annu Rev Immunol. (2013) 31:443–73. doi: 10.1146/annurev-immunol-032712-095910 90. Matsumura M, Fremont DH, Peterson PA, Wilson IA. Emerging principles for the recognition of peptide antigens by MHC class I molecules. *Sci (New York NY)*. (1992) 257:927–34. doi: 10.1126/science.1323878

91. Bouvier M, Wiley DC. Importance of peptide amino and carboxyl termini to the stability of MHC class I molecules. *Sci (New York NY)*. (1994) 265:398–402. doi: 10.1126/science.8023162

92. Zacharias M, Springer S. Conformational flexibility of the MHC class I alpha1alpha2 domain in peptide bound and free states: a molecular dynamics simulation study. *Biophys J.* (2004) 87:2203–14. doi: 10.1529/biophysj.104.044743

93. Chicz RM, Urban RG, Lane WS, Gorga JC, Stern LJ, Vignali DA, et al. Predominant naturally processed peptides bound to HLA-DR1 are derived from MHC-related molecules and are heterogeneous in size. *Nature.* (1992) 358:764–8. doi: 10.1038/358764a0

94. Gulati P. Janeway's Immunobiology, 7th Edition, Vol. 37. Murphy K, Travers P, Walport M editors. New York: Garland Science (2009), pp. 134–4. doi: 10.1002/ bmb.20272.

95. Mahanty S, Prigent A, Garraud O. Immunogenicity of infectious pathogens and vaccine antigens. *BMC Immunol.* (2015) 16:31. doi: 10.1186/s12865-015-0095-y

96. Shlomchik MJ, Weisel F. Germinal center selection and the development of memory B and plasma cells. *Immunol Rev.* (2012) 247:52–63. doi: 10.1111/j.1600-065X.2012.01124.x

97. Assarsson E, Sidney J, Oseroff C, Pasquetto V, Bui HH, Frahm N, et al. A quantitative analysis of the variables affecting the repertoire of T cell specificities recognized after vaccinia virus infection. *J Immunol (Baltimore Md: 1950).* (2007) 178:7890–901. doi: 10.4049/jimmunol.178.12.7890

98. Pishesha N, Harmand TJ, Ploegh HL. A guide to antigen processing and presentation. *Nat Rev Immunol.* (2022) 22:751–64. doi: 10.1038/s41577-022-00707-2

99. Gras S, Burrows SR, Turner SJ, Sewell AK, McCluskey J, Rossjohn J. A structural voyage toward an understanding of the MHC-I-restricted immune response: lessons learned and much to be learned. *Immunol Rev.* (2012) 250:61–81. doi: 10.1111/j.1600-065X.2012.01159.x

100. Malik B, Rath G, Goyal AK. Are the anatomical sites for vaccine administration selected judiciously? *Int immunopharmacol.* (2014) 19:17–26. doi: 10.1016/j.intimp.2013.12.023

101. Sidney J, Southwood S, Moore C, Oseroff C, Pinilla C, Grey HM, et al. Measurement of MHC/peptide interactions by gel filtration or monoclonal antibody capture. *Curr Protoc Immunol.* (2013) 18:13. doi: 10.1002/0471142735.im1803s100

102. Andersen RS, Kvistborg P, Frøsig TM, Pedersen NW, Lyngaa R, Bakker AH, et al. Parallel detection of antigen-specific T cell responses by combinatorial encoding of MHC multimers. *Nat Protoc.* (2012) 7:891–902. doi: 10.1038/nprot.2012.037

103. Toebes M, Coccoris M, Bins A, Rodenko B, Gomez R, Nieuwkoop NJ, et al. Design and use of conditional MHC class I ligands. *Nat Med.* (2006) 12:246–51. doi: 10.1038/nm1360

104. Kvistborg P, Shu CJ, Heemskerk B, Fankhauser M, Thrue CA, Toebes M, et al. TIL therapy broadens the tumor-reactive CD8(+) T cell compartment in melanoma patients. *Oncoimmunology*. (2012) 1:409–18. doi: 10.4161/onci.18851

105. Peng S, Zaretsky JM, Ng AHC, Chour W, Bethune MT, Choi J, et al. Sensitive detection and analysis of neoantigen-specific T cell populations from tumors and blood. *Cell Rep.* (2019) 28:2728–2738.e2727. doi: 10.1016/j.celrep.2019.07.106

106. Loftus DJ, Castelli C, Clay TM, Squarcina P, Marincola FM, Nishimura MI, et al. Identification of epitope mimics recognized by CTL reactive to the melanoma/melanocytederived peptide MART-1(27-35). J Exp Med. (1996) 184:647–57. doi: 10.1084/jem.184.2.647

107. Dutoit V, Rubio-Godoy V, Pittet MJ, Zippelius A, Dietrich P-Y, Legal FA, et al. Degeneracy of antigen recognition as the molecular basis for the high frequency of naive A2/Melan-a peptide multimer(+) CD8(+) T cells in humans. *J Exp Med.* (2002) 196:207–16. doi: 10.1084/jem.20020242

108. Vujanovic L, Mandic M, Olson WC, Kirkwood JM, Storkus WJ. A mycoplasma peptide elicits heteroclitic CD4+ T cell responses against tumor antigen MAGE-A6. *Clin Cancer Res.* (2007) 13:6796–806. doi: 10.1158/1078-0432.CCR-07-1909

109. Tomasi M, Dalsass M, Beghini F, Zanella I, Caproni E, Fantappie L, et al. Commensal bifidobacterium strains enhance the efficacy of neo-epitope based cancer vaccines. *Vaccines (Basel)*. (2021) 9:1356. doi: 10.3390/vaccines9111356

110. Girdhar K, Huang Q, Chow IT, Vatanen T, Brady C, Raisingani A, et al. A gut microbial peptide and molecular mimicry in the pathogenesis of type 1 diabetes. *Proc Natl Acad Sci U. S. A.* (2022) 119:e2120028119. doi: 10.1073/pnas.2120028119

111. Lerner A, Benzvi C, Vojdani A. Cross-reactivity and sequence similarity between microbial transglutaminase and human tissue antigens. *Sci Rep.* (2023) 13:17526. doi: 10.1038/s41598-023-44452-5

112. Antonelli AC, Binyamin A, Hohl TM, Glickman MS, Redelman-Sidi G. Bacterial immunotherapy for cancer induces CD4-dependent tumor-specific immunity through tumor-intrinsic interferon- γ signaling. *Proc Natl Acad Sci United States America*. (2020) 117:18627–37. doi: 10.1073/pnas.2004421117

113. Kates M, Nirschl T, Sopko NA, Matsui H, Kochel CM, Reis LO, et al. Intravesical BCG induces CD4(+) T-cell expansion in an immune competent model of bladder cancer. *Cancer Immunol Res.* (2017) 5:594–603. doi: 10.1158/2326-6066.CIR-16-0267

114. El Tekle G, Garrett WS. Bacteria in cancer initiation, promotion and progression. Nat Rev Cancer. (2023) 23:600–18. doi: 10.1038/s41568-023-00594-2

Glossary

Co	ntin	ued
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NK cells	natural killer cells
TSAs	tumor-specific antigens
TAAs	tumor-associated antigens
MAGE	melanoma-associated antigen
ICI	immune checkpoint inhibitor
CAR-T	
IL-2	chimeric antigen receptor T cell interleukin-2
IFN-γ	interferon-gamma
PD-1/PD-L1	programmed death protein-1/programmed death ligand-1
Sag A	secreted antigen A
NOD2	nucleotide-binding oligomerization domain 2
DCs	dendritic cells
ILC	innate lymphocyte
GC cells	germinal center cells
MHC	histocompatibility complex class
PG	peptidoglycan
LPS	lipopolysaccharide
OMV	outer membrane vesicles
BG	bacterial ghosts
P-AKT	phospho-protein kinase B
P-mTOR	phospho-mammalian targets of rapamycin
P-p70s6K	phospho-p70 ribosomal s6 kinase
TMP1	tail tape measure protein 1
PSMB4	proteasome β subunit 4
PAMPs	pathogen-associated molecular patterns
Ags	Microbial antigens
HLA	human leukocyte antigen
IEDB	Immune Epitope Database
TcRs	T lymphocyte receptors
ТАР	transporter associated with antigen processing
ERP57	endoplasmic reticulum protein 57
MARCH	Membrane-Associated RING-CH protein
ERAAP	ER-associated aminopeptidases
APCs	antigen-presenting cells
NP	nanoparticle
TH1	helper T cell 1
TIL	tumor-infiltrating lymphocytes
ANN	artificial neural networks

PWMs	position weight matrices
PSSM	position specific scoring matrix
MS	mass spectrometry
NSCLC	non-small cell lung cancer
PI3K	Phosphoinositide 3-kinase
AKT	protein kinase B
CRPC	castration-resistant prostate cancer
DHEA	dehydroepiandrosterone

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