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Exposing the latent phenotype of Gulf War Illness: examination of the mechanistic mediators of cognitive dysfunction

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Though it has been over 30 years since the 1990–1991 Gulf War (GW), the pathophysiology of Gulf War Illness (GWI), the complex, progressive illness affecting approximately 30% of GW Veterans, has not been fully characterized. While the symptomology of GWI is broad, many symptoms can be attributed to immune and endocrine dysfunction as these critical responses appear to be dysregulated in many GWI patients. Since such dysregulation emerges in response to immune threats or stressful situations, it is unsurprising that clinical studies suggest that GWI may present with a latent phenotype. This is most often observed in studies that include an exercise challenge during which many GWI patients experience an exacerbation of symptoms. Unfortunately, very few preclinical studies include such physiological stressors when assessing their experimental models of GWI, which creates variable results that hinder the elucidation of the mechanisms mediating GWI. Thus, the purpose of this review is to highlight the clinical and preclinical findings that investigate the inflammatory component of GWI and support the concept that GWI may be characterized as having a latent phenotype. We will mainly focus on studies assessing the progressive cognitive impairments associated with GWI and emphasize the need for physiological stressors in future work to create a more unified hypothesis that can identify potential therapeutics for this patient population.

KEYWORDS

acetylcholinesterase, pyridostigmine bromide, acetylcholine, hippocampus, lipopolysaccharide, stress

1 Introduction

Nearly one third of soldiers that served in the 1990–1991 Gulf War (GW) continue to suffer from an unexplained, multi-symptom illness (1). While the potential for a “Gulf War Illness” (GWI) was initially debated by the Departments of Defense (DoD) and Veterans Affairs (VA), it was ultimately recognized as a unique combination of symptoms solely

affecting GW Veterans in 1997 (2, 3). This recognition came after multiple case studies and congressional hearings reported significantly higher rates of unexplained health issues and poorer overall health in GW Veterans compared to other cohorts of Veterans from this era (4, 5). Such symptoms included cardiovascular abnormalities such as tachycardia and hypertension, gastrointestinal (GI) disturbances including diarrhea and irritable bowel syndrome (IBS), and respiratory impairments including emphysema and chronic bronchitis (1, 6). Many GW Veterans also reported neurological disturbances including chronic headaches, sleep disturbances, persistent depression, and memory and attentional impairments (1, 7).

As the symptomology of GWI is quite varied, the state of Kansas created the Kansas Persian Gulf War Veterans Health Initiative Program to establish explicit criteria for GWI diagnoses (8). This work identified six main categories of GWI symptomology: fatigue/sleep, somatic pain, neurologic/cognitive/mood, GI symptoms, respiratory symptoms, and skin symptoms (8). Now known as the Kansas definition, GWI is diagnosed if a patient reports symptoms in at least three of these six categories (8). The Center for Disease Control established a broader definition of GWI that only includes mood/cognition, fatigue, and musculoskeletal categories (9). GWI is diagnosed using these criteria if a patient reports one chronic symptom in two of the three categories (9). Though different, both the Kansas and CDC definitions have been recommended for diagnostic use by the DoD and VA.

Utilizing these standardized definitions, it is estimated that GWI affects approximately 200,000–250,000 of the 700,000 troops deployed to the GW (1). While GWI has been reported in both men and women, the majority of cases are seen in men as over 95% of GW Veterans are male (6). GWI is not only complex in its symptomology, but also in its progression as 10-year and 20-year follow-up surveys reported that many GWI symptoms had worsened in affected Veterans (7, 10, 11). This progression includes declines in neurological measures, including cognitive function and neuropsychiatric disorders (7, 12, 13), which coincides with volume loss in the cerebellum and brainstem of some patients (14, 15). For additional information on the initiation and progression of GWI, readers are referred to Gulf War and Health Volumes 1–11 from the Institute of Medicine Committee on Gulf War and Health. Adding to its complexity, many symptoms of GWI, including cognitive deficits, are exacerbated when patients experience stressors and many GW Veterans complain of severe post-exertional malaise (16). Such reports suggest that GWI may present with a latent phenotype, as the full symptomology may only appear after a stressor is presented. Unfortunately, the number of preclinical and clinical studies examining these stimulus-dependent symptoms are limited. Thus, the purpose of this review is to illustrate the latent phenotype of GWI and to highlight the importance of including physiological stressors in future GWI studies in order to fully elucidate the mechanisms driving this complex pathology, as well as to identify potential therapeutics.

2 GWI and inflammation

While this complex disease remains poorly understood, immune dysfunction is now considered a hallmark feature of

GWI as numerous clinical studies have found a persistent pro-inflammatory state in GWI patients. Early studies reported that relative to controls, GWI patients exhibited elevated numbers of plasma T cells and B cells, while the number of natural killer cells was reduced approximately 2–8 years after their GW service (17, 18). Additionally, studies found higher plasma levels of pro-inflammatory markers such as interferon-gamma (IFN- γ), interleukin (IL)-2, and tumor necrosis factor alpha (TNF- α) in GWI patients at similar post-deployment time points (17, 19). This immune profile has been validated by more recent studies that also found elevations in these cytokines, as well as IL-6 and C-reactive protein (CRP) in samples collected between 2010 and 2013 (20, 21). Along with higher levels of pro-inflammatory cytokines, another study found a down regulation of the anti-inflammatory cytokines IL-4 and IL-13 in the serum of GWI patients almost 25 years after their GW service (22). Chronic inflammation resulting from sustained elevations of pro-inflammatory cytokines in plasma has been implicated in various diseases such as IBS, rheumatoid arthritis, and psoriasis, all of which have been reported in patients with GWI (6, 7, 12, 23–25). Recently, similarities between GWI and post-acute COVID-19 syndrome (PACS) or “long COVID” symptomology have been drawn, suggesting that this chronic inflammation may be due to an unidentified, persistent antigen (26). Thus, while the mechanistic basis of this immune dysfunction has not been fully elucidated, it is likely that the aberrant immune signaling is contributing to a broad range of pathological settings, including GWI.

Beyond its peripheral effects, systemic inflammation is also known to produce neuroinflammation through humoral, cellular, and neural pathways (27). This includes the passage of circulating immune cells and cytokines into the central nervous system (CNS) through various transport mechanisms (28). Additionally, vagal afferent fibers innervating the periphery can detect the presence of pro-inflammatory signals and relay this information to the CNS (29). Collectively, this detection of peripheral inflammation triggers an inflammatory response within the CNS that can compromise the overall health and function of neurons when such pathways are chronically activated. As such, some researchers have hypothesized that chronic, peripheral inflammation may be a mechanistic mediator in the cognitive decline observed in GWI patients through the initiation of chronic neuroinflammation. For example, the cytokine high-mobility group box 1 (HMGB1) is elevated in plasma samples of GWI patients (30) and intraperitoneally (i.p.) administered HMGB1 has been shown to activate microglia and produce memory impairments and depressive-like behavior in mice (31–34). Similar conclusions were drawn in a separate study that found that adding serum of GWI patients to neural cultures caused a reduction in cell growth and an increase in apoptosis which disrupted neural network communication (35). These investigators suggested that such responses occurred because the serum of GWI patients contains neuropathogenic factors as these impairments were ameliorated by the addition of serum from healthy GW Veterans (35). This neuroinflammatory component of GWI is further supported by recent findings that visualized greater amounts of activated microglia and astrocytes in the brains of GWI patients relative to healthy GW Veterans and civilians with *in vivo* imaging (36). Taken

together, these studies identify neuroinflammation as a putative causative factor in the neurological complications in GWI patients, but additional studies are needed to identify specific CNS biomarkers and fully elucidate the mechanism(s) responsible, including the contribution of peripheral immune dysfunction.

Along with immune dysfunction, dysregulated endocrine signaling has also been implicated in GWI (6, 7, 16). These impairments have been associated with abnormal hypothalamic-pituitary-adrenal (HPA) axis activity, which regulates the stress response by producing cortisol through the release of corticotropin releasing factor (CRF) and adrenocorticotrophin hormone (ACTH) (37). HPA axis dysfunction in GWI subjects was observed following a dexamethasone suppression test, which assesses the feedback activity of the HPA axis by mimicking cortisol release which should suppress ACTH and in turn, the release of cortisol (38). Specifically, approximately 15 years after the GW, GWI patients exhibited a greater suppression of cortisol levels than controls, which may be indicative of impaired pituitary or adrenal responses (39). Golier and colleagues expanded upon these findings and reported that GW Veterans exhibit significantly lower levels of plasma ACTH and a higher cortisol:ACTH ratio, which they attributed to an enhanced sensitivity to cortisol's feedback effects and reduced HPA axis activity (40). Importantly, cortisol and other glucocorticoids released by the HPA axis perform critical anti-inflammatory actions and the attenuation of this axis in GW Veterans may be exacerbating the observed immune dysfunction (41, 42).

3 Potential causes and mechanisms of GWI

In view of these clinical observations, peripheral and/or central inflammation is proposed to be a mechanistic mediator of GWI pathology. As such, it is important to identify potential exposures during deployment that could be responsible for these inflammatory responses in GWI Veterans. Soldiers were exposed to several hazards in the Gulf including the use of pesticides and insecticides, airborne particulates from dust and sand, depleted uranium in military vehicles, smoke from oil well fires, and severe stress from combat (43). Additionally, soldiers were administered various vaccines and pyridostigmine bromide (PB), a pharmacological prophylaxis against potential nerve gas attacks (44, 45). While the list of harmful exposures during the GW is extensive, it is interesting that multiple exposures, namely PB, nerve gases, pesticides/insecticides, and stress, all greatly impact the cholinergic system, a key regulator of cognition and anti-inflammatory responses in both the periphery and CNS.

3.1 Pyridostigmine bromide and nerve gases

PB is the dimethylcarbamate ester of 3-hydroxy-1-methylpyridinium bromide that was originally used for the treatment of myasthenia gravis

(MG) as it acts an acetylcholinesterase (AChE) inhibitor (46, 47). PB blocks the enzymatic activity of AChE by binding to the serine residue in the enzyme active site, rendering it inactive (48). Being a carbamate compound, PB is considered a reversible AChE inhibitor as the binding of the AChE-carbamate complex that inhibits AChE action is quite unstable and will spontaneously split from the enzyme by hydrolysis (49). Importantly, PB's inhibitory actions are confined to the periphery due to its quaternary ammonium group that limits lipid solubility and prevents passage through the blood-brain barrier (BBB) (50). While PB is well known for its therapeutic properties, many other AChE inhibitors are better known for their deleterious actions. Organophosphate (OP) AChE inhibitors, such as some nerve gases, are much more toxic than PB as they permanently inhibit the enzyme by non-reversible phosphorylation (49). Such irreversible AChE inhibitors produce a dangerous accumulation of acetylcholine (ACh) in the synaptic cleft that compromises normal muscle control, which becomes lethal if the respiratory muscles are affected and asphyxiation occurs (51). OP AChE inhibitors have been used as chemical warfare agents as they can be vaporized or aerosolized to attack large groups through their inhalation and absorption into the skin (48).

Saddam Hussein was thought to possess large stockpiles of these OP AChE inhibitors during the GW, namely the G-series nerve agents, sarin and soman (52). PB was identified as a potential prophylactic against irreversible OP AChEs due to its ability to compete for AChE binding (53). Preclinical studies before the GW showed that carbamate AChEs, such as PB, significantly improved survival rates after nerve agent exposure when used as a pretreatment (44, 45, 53). Thus, PB was used as an investigational drug in the GW, with each soldier receiving 21-tablet blister packs and instructed to take one 30 mg tablet every 8 hours to ensure continuous protection (46, 54). This prescribed dose suppressed plasma AChE activity by approximately 50% (55), which replicated the dose tested in preclinical models (45). Many soldiers reported immediate side effects to PB, with most involving the GI system, but such symptoms were not considered incapacitating (56). The rationale for this prophylactic approach had practical implications, especially in view of reports from the DoD estimating that perhaps 100,000 personnel were possibly exposed to low levels of sarin nerve agent after the Khamisiyah munitions depot demolition (57). More recent studies have correlated higher rates of mild cognitive impairment (MCI) and smaller hippocampal volumes in GW Veterans that recall hearing chemical alarm sounds more frequently, which was attributed to greater nerve agent exposure, but the prophylactic use of PB during these incidents is not clear (58). Indeed, while the DoD reports that approximately 250,000 military personnel took PB during the GW, its self-administration led to a wide variety of dosing regimens (59). Despite its varied use, PB exposure has been consistently correlated with GWI presentation as many GWI symptoms can be attributed to cholinergic toxicity (43, 60–62). Currently, the use of PB for MG is not attributed to any long-term side effects beyond mild GI disturbances (63), but adverse effects have not been widely studied in individuals without MG, or when combined with other ACh-altering exposures.

3.2 Pesticides/insecticides

Another form of prophylaxis during the GW was the liberal use of pesticides and insecticides as a means to control vector-borne diseases. Due to this aggressive approach, the DoD has estimated at least 41,000 servicemembers may have been overexposed to such chemicals, including chlorpyrifos, permethrin, and *N,N*-diethyl-meta-toluamide (DEET). Importantly, like PB, these chemicals are known to alter AChE activity (50, 64, 65). Specifically, chlorpyrifos is an irreversible, OP AChE inhibitor similar to nerve gases that acts as an extremely potent pesticide (66). Chlorpyrifos and other OP AChE inhibitors were used during the GW as fumigating agents for campgrounds (67). Permethrin is a synthetic pyrethroid and insecticide that causes respiratory paralysis in arthropods by blocking sodium channels (65). This chemical was applied to soldiers' uniforms and skin through aerosol sprays (68). While it is not classified as an AChE inhibitor, permethrin has been shown to inhibit AChE in multiple brain regions when administered to rats intragastrically (65). Lastly, DEET is a common insect repellent used in the GW that works through similar mechanisms as permethrin and is thought to have weak AChE activity (69). As with PB, the use of such pesticides/insecticides during the GW varied greatly among soldiers, but their neurotoxic effects have been proposed as a causative factor of GWI.

3.3 War-related stress

Though many of the aforementioned GW exposures have been linked to GWI, the incidence of GWI is more prevalent in ground forces that faced combat compared to forces that remained in relief areas (8). This disparity led researchers to hypothesize that the physical and psychological stress of combat may also contribute to GWI presentation (70–76). As in most military conflicts, GW soldiers experienced several psychological stressors, including the lack of social contact with family, threat of attack, and fear of a friend or self being killed (77). Importantly, though such stressors are known to produce post-traumatic stress disorder (PTSD) in Veteran populations, studies have concluded that the symptomology of GW Veterans is distinct from PTSD (78). While PTSD has been shown to exacerbate GWI symptoms (78), adjusting for PTSD diagnoses revealed worse overall health outcomes in GW Veterans relative to non-GW Veterans from this era, independent of PTSD prevalence (79).

As described above, the stress response is primarily mediated by the HPA axis and chronic stress, such as the prolonged stress of deployment, can dysregulate the feedback mechanisms of the HPA axis and lead to systemic inflammation, and both are seen in GWI patients (19, 37). Along with peripheral inflammation, chronic stress can affect the central nervous system (CNS) by disrupting the brain regions that regulate the stress response. This includes the hippocampus, which is an essential integration center for learning and memory in the mammalian brain (80). From a morphological perspective, chronic stress reduces neurogenesis of dentate gyrus granule neurons and induces dendritic atrophy of pyramidal

neurons within this region, which can produce memory and attentional deficits like those observed in GWI (81, 82). Similar effects of chronic stress on neuronal structure and function are also observed in the prefrontal cortex (PFC) (83), a region that mediates higher-order cognitive functions such as attention and decision making (84). While the stress response within the CNS involves numerous neurotransmitters, ACh is a key mediator as cholinergic neurotransmission increases in both the hippocampus and PFC in response to a stressor (85). This increased efflux is thought to be adaptive for acute stressors as it can promote synaptic plasticity, but more severe or chronic stress can elicit excessive ACh efflux in these brain regions that produces maladaptive stress responses and impairs cognitive function (86). Thus, the synergistic effects of AChE inhibitors and war-related stress on cholinergic neurotransmission may be especially relevant to the cognitive deficits observed in GWI.

3.4 Acetylcholine in GWI

3.4.1 Role of acetylcholine in cognition

A large portion of the cholinergic neurons in the CNS are found in the basal forebrain (BF) nuclei, a heterogeneous set of structures located in the rostroventral forebrain (87). These structures include the medial septum and the diagonal band of Broca which send cholinergic projections to the hippocampus (88). Clinical studies have shown the importance of ACh neurotransmission in the hippocampus by using cholinergic antagonists to impair performance in memory tasks (89, 90). In addition to hippocampal projections, the BF also contains the nucleus basalis and substantia innominata, which house cholinergic projections that innervate the PFC (88). It is well established that BF cholinergic neurons are involved in PFC-dependent cognitive processes, as lesioning BF projections in rodents results in significant impairments in attentional tasks (91–95). Besides cholinergic neurons, ACh receptors are also found on other neurons in the PFC and hippocampus including glutamatergic, GABAergic, noradrenergic, and dopaminergic neurons (85). Thus, ACh is also considered a neuromodulator as it can mediate the release of these other neurotransmitters and regulate neuronal excitability, synaptic plasticity, and coordinate the firing of groups of neurons to adapt to environmental stimuli (96–99).

3.4.2 The cholinergic anti-inflammatory pathway

Among its modulatory actions, cholinergic neurotransmission has also been recognized as a key regulator of inflammation through the cholinergic anti-inflammatory pathway (100). The peripheral cholinergic anti-inflammatory pathway was first identified by Kevin Tracey's group who discovered the presence of $\alpha 7$ nicotinic acetylcholine receptors (nAChRs) on macrophages. (101). *In vitro* studies found that activation of $\alpha 7$ nAChRs by ACh reduced the expression of pro-inflammatory cytokines, specifically IL-1 β , IL-6, IL-18, and TNF- α in macrophages exposed to the endotoxin lipopolysaccharide (LPS) (101). These *in vitro* studies were later confirmed *in vivo* when $\alpha 7$ nAChR deficient mice exhibited suppressed anti-inflammatory responses to an LPS challenge

compared to wild type mice (102). Later studies identified $\alpha 7$ nAChRs on other immune cells including monocytes (103, 104), T-cells (105, 106), and B-cells (107).

Peripherally, ACh's anti-inflammatory actions are mediated by the vagus nerve (100). Vagal afferent fibers innervate a multitude of peripheral organs and alert the CNS about any homeostatic disturbances through their projections to the nucleus tractus solitarius (NTS) in the brainstem (29). Vagal afferents can detect such disruptions as they contain receptors for endotoxins and the pro-inflammatory cytokines they produce (108). Once this information is relayed to the NTS, it is propagated to other brain regions involved in processing visceral information, such as the hypothalamus, amygdala, cortex, and locus coeruleus (109–111). To restore homeostasis, vagal efferent fibers residing in the dorsal motor nucleus of the vagus within the brainstem trigger the release of norepinephrine (NE) from the splenic nerve in the celiac plexus (112). NE then triggers the release of ACh from lymphocytes through $\beta 2$ -adrenergic receptors and ACh attenuates the release of pro-inflammatory mediators by binding to $\alpha 7$ nAChRs on immune cells (112). This anti-inflammatory vagal reflex is best illustrated in vagotomized mice experiencing endotoxemia after high dose LPS administration while sham animals do not (101). Importantly, the immunosuppressive actions of the vagus nerve are incredibly efficient and occur much faster than other immune responses (113).

As $\alpha 7$ nAChRs are also expressed in the CNS, more recent studies have investigated if a central cholinergic anti-inflammatory pathway exists. Evidence for such a pathway was first supported by studies that identified $\alpha 7$ nAChRs on microglia and astrocytes, key CNS immune cells (114, 115). *In vitro* studies have used ACh and nicotine, a nAChR agonist, in microglial cultures to show that activation of $\alpha 7$ nAChRs suppresses LPS-induced increases in TNF- α and this response is blocked by the $\alpha 7$ nAChR antagonist α -bungarotoxin (115). The immunosuppressive role of microglial $\alpha 7$ nAChRs has also been observed *in vivo* as intracerebroventricular administration of the $\alpha 7$ nAChR agonist PNU282987 attenuated LPS-induced microglial activation and sickness behavior in mice (116). Astrocytic $\alpha 7$ nAChRs elicit similar effects as nicotine also dampens the pro-inflammatory response of cultured astrocytes activated by IL-1 β (117). As neuroinflammation has been implicated in a variety of neurodegenerative diseases including Alzheimer's disease, Parkinson's disease, and multiple sclerosis, disruptions in the cholinergic anti-inflammatory pathway may be a mechanistic mediator of these diseases (118, 119). Indeed, many preclinical studies have investigated $\alpha 7$ nAChR agonists as potential therapeutics for neurodegenerative diseases driven by neuroinflammation (120–122). The therapeutic effects of $\alpha 7$ nAChR agonists have also been investigated in chronic inflammatory diseases in the periphery including rheumatoid arthritis, ulcerative colitis, and diabetes (123–125). This area of research may be especially relevant for GWI studies as GW Veterans experience similar inflammation-mediated symptoms and immune dysregulation (19, 126). Furthermore, many GWI symptoms involve organ systems that are highly innervated by the vagus nerve (i.e., heart, lungs, GI tract) (7, 127). Unfortunately, little is known about the potential dysfunction of the peripheral and/or

central cholinergic anti-inflammatory network in GWI, including how these deficits may contribute to its latent phenotype.

3.5 Latent phenotype of GWI

Perhaps the best illustration of the latent phenotype of GWI emerges when patients are subjected to an exercise challenge. It is well established that fatigue and post-exertional malaise are common symptoms of GWI (16), but exercise has also been shown to exacerbate other GWI symptoms including musculoskeletal pain and cognitive impairments (128–130). Various imaging studies have been performed to understand this latent phenotype of GWI and why exercise can worsen these symptoms. One group reported that while performing a working memory task after bicycling, the dorsal midbrain and cerebella dermis became deactivated in GWI patients, but not healthy controls (129). Additionally, this bicycling challenge separated GWI patients into two phenotypes, with one group exhibiting orthostatic tachycardia and brainstem atrophy, and the other group experiencing severe pain and exhibiting cortical atrophy (131). Another study that analyzed cerebrospinal fluid after bicycling found elevations in glutamate in a subset of GWI patients, which may be indicative of exercise-induced excitotoxicity (132).

It has been suggested that such negative responses to exercise may be due to sensitized immune and endocrine responses as GWI patients subjected to a bicycle test also showed abnormal immune responses. Specifically, exercise induced a heightened pro-inflammatory state in GWI patients relative to healthy controls as the inflammatory activities of nuclear factor kappa B (NF- κ B) and IL-6 were increased. Other studies from this group report that exercise reduced the expression of genes associated with the activity of natural killer cells in GWI patients (133). Interestingly, many of these genes are also involved in glucocorticoid receptor signaling pathways and salivary cortisol was significantly lower in GWI patients after exercise (133). These reductions provide further evidence that diminished HPA axis activity may be a key feature of GWI and its latent phenotype as most symptoms are exacerbated following an exercise challenge. It is important to note that these findings occurred when an aerobic, cycling challenge was used, but lower intensity exercises, such as yoga, have been shown to decrease GWI symptom severity, especially chronic pain (134, 135). Thus, the pathophysiology of GWI may be due to sensitized responses to certain stressors, but the mechanisms responsible for these aberrant immune and endocrine responses have not been fully elucidated.

4 Lessons from GWI studies in rodents: focus on inflammation

As noted above, clinical studies suggest that while no specific neurotoxic chemical exposure explains the illnesses observed in GW Veterans (2), administration of AChE inhibitors in combination with the stress of deployment likely contributes to the initiation and progression of GWI. In view of these observations, many preclinical

studies have investigated the neurological effects of PB administration alone and in combination with pesticides and/or stress. [For a recent comprehensive review of GWI models, please see (66)]. Interestingly, though the treatment paradigms vary, inflammation is a common feature among many GWI animal models, which is consistent with clinical findings in GWI patients. For example, plasma levels of pro-inflammatory cytokines, including IL-1 β , TNF- α , and IFN- γ , were significantly elevated in a rat model of GWI approximately 6 weeks after exposure to GW chemicals (PB + permethrin + DEET) and restraint stress (RS) (136). These animals also exhibited muscle atrophy, which recapitulates the musculoskeletal disturbances seen in GWI patients (136). Another group investigated the long-term effects of their GWI model (PB + permethrin + DEET + RS) and saw similar increases in plasma IL-1 β and TNF- α levels, as well as IL-1 α and fibroblast growth factor 2 (FGF2) levels more than 6 months after treatment (137). The same group also observed increased expression of oxidative stress markers in the plasma of these rats and reproduced these findings in a second study (137, 138).

Given the established relationship between neuroinflammation and cognitive decline, many studies have assessed neuroinflammation in rodent models of GWI, especially at delayed timepoints, as a potential cause of the progressive cognitive deficits observed in patients. Similar to plasma cytokines, elevations in pro-inflammatory cytokines (IL-1 β , TNF- α , IFN- γ), were reported in the brains of a GWI mouse model (PB + permethrin) almost 1 year after the cessation of treatment (139). More region-specific studies found elevations in pro-inflammatory mediators in the hippocampus (137), and cortex (140) of a GWI rat model 6–10 months after GW chemical exposure (PB + permethrin + DEET + RS). As astrocytes and microglia are the immune cells within the CNS that drive the release of pro-inflammatory signals, many studies report an upregulation of markers for astrocyte and microglia activation in the hippocampus and cortex of GWI rodents (139, 141–143). Furthermore, some GWI rodent models exhibit decreases in neurogenesis in the dentate gyrus, as well as reduced hippocampal volume approximately 4 months after GW chemical exposure (PB + permethrin + DEET + RS) (141, 143). A more recent study reported that GWI mice (PB + chlorpyrifos + DEET) also exhibit dendritic atrophy and decreases in spine density in dentate gyrus granule neurons (144). Taken together, these studies suggest that GW chemicals produce long-lasting inflammation within the CNS which likely involves reactive astrocytes and microglia producing pro-inflammatory cytokines, thereby leading to deficits in neuronal structure. Given that these changes are occurring in the cortex and hippocampus, such findings identify a role for neuroinflammation in the progressive cognitive decline observed in many GW Veterans suffering from GWI.

While these studies suggest a causal relationship between peripheral and central inflammation as a mechanistic mediator of GWI, other laboratories have reported contrasting findings. For example, Zakirova and colleagues did not observe any significant differences in brain cytokine levels (IL-1 β , TNF- α and IFN- γ) in their mouse model of GWI (PB + permethrin) (145). Additionally, plasma levels of these cytokines were reduced, further adding to the

uncertainty of whether an inflammatory phenotype is a key feature of GWI rodents (145). Similar findings were reported by our group which saw a reduction in plasma levels of the pro-inflammatory cytokines IL-1 α , IL-2, IL-6, IL-12 and IFN- γ in rats that received PB treatment 3 months prior (72). Our follow-up studies also support Zakirova's findings as we did not observe any difference in hippocampal or PFC cytokine levels in saline-treated rats previously exposed to PB (75). However, we found that a prior history of PB treatment exacerbates the cytokine response in the plasma (TNF- α , cytokine-induced neutrophil chemoattractant 3; CINC-3), hippocampus (IL-1 β , IL-12, granulocyte-macrophage colony stimulating factor; GM-CSF) and PFC (IL-12, GM-CSF) when these animals are challenged with a peripheral injection of LPS 3 months after treatment cessation (75). These results suggest that instead of a persistent inflammatory phenotype, GWI may be better characterized as a dysregulation of pro-inflammatory responses and anti-inflammatory feedback mechanisms. Indeed, we also observed that the LPS challenge potentiates ACh efflux in the hippocampus of PB-treated rats 3 months after exposure, which should result in a suppression of pro-inflammatory cytokines, not an exaggeration (75). This potentiation was not specific to an immune challenge, as PB-treated rats also exhibited greater ACh efflux after an immobilization stress challenge relative to vehicle-treated controls, at this delayed timepoint (76). Importantly, these elevations in ACh: 1) were not due to basal differences in ACh levels or alterations in AChE activity within the hippocampus; and 2) only emerged when a physiological stressor was presented months after treatment cessation (75, 76). Such findings mirror the sensitized immune and stress responses observed in GWI patients, further suggesting that dysregulation of the central cholinergic anti-inflammatory pathway is a causative factor in the pathogenesis of GWI. Such results are also consistent with the concept that a latent phenotype is a characteristic feature of GWI. Unfortunately, there are limited studies including such stressors in GWI research, making it difficult to fully elucidate the latent phenotype of GWI and how inflammation may be contributing to the clinical symptoms.

5 Lessons from GWI studies in rodents: focus on cognitive studies

Another area of GWI research that could greatly benefit from the inclusion of stressors and thereby reveal a latent phenotype is the behavioral studies aiming to reproduce the cognitive deficits observed in GW Veterans. Though many laboratories have assessed learning and memory in rodents exposed to GW-related chemicals and/or stress, the results from these studies are quite varied; see Table 1 for outcomes and treatment paradigms. For example, Crawford and coworkers reported that PB + permethrin-treated mice exhibit deficits in the Barnes maze compared to control mice approximately 3 months post treatment (150) and also 15–16 months post treatment (151). However, other behavioral measures yielded more equivocal findings. For example, a study assessing working memory in a rat model of GWI with a delayed alternation task found that stress alone impairs performance, but the combination of PB and stress did not show synergistic effects on working memory (70). Additionally, several groups have utilized

novel object recognition (NOR) and novel place recognition testing in GWI models and saw significant impairments in both tasks in a rat model of GWI more than 3 months after treatment cessation (140, 149, 152, 153). Conversely, another group did not observe any deficits in NOR in their rat model of GWI at this 3 month timepoint (141). Interestingly, we observed both results when we assessed NOR performance in PB-treated rats approximately 3 months after treatment cessation, as impairments were only seen in the presence of a physiological stressor (76). Specifically, 3 months after

treatment cessation, PB-treated rats that were administered saline (i.p.) before familiar object presentation did not exhibit any deficits in NOR performance 24-hours later (76). However, when these rats were challenged with LPS (i.p.) the following week, PB-treated rats displayed significant impairments in 24-hour memory, while the performance of the vehicle-treated controls was not affected by the LPS challenge (76). Thus, including an immune challenge in our behavioral studies allowed us to unmask hippocampal-dependent memory deficits in our GWI model. It is interesting to speculate

TABLE 1 Effects of different GWI treatment paradigms on learning and memory measures in rodents.

GWl paradigm	Testing period(s) relative to Rx	Sex/Species	Outcomes	Reference
PB +/- FS stress	During treatment	Male SD rats	Transient decreases in alternation performance in FS group; no individual or additive effects with PB.	(70)
PB +/- PCA for 12 days	Post Rx days 15, 16, 17, 18, 19, 52, 113, 119	Male Wistar rats	Time to escape water maze was increased in PB + stress rats at days 16 through 119.	(146)
PB + PER for 10 days	Post Rx days 51, 86, 115	Male CD1 mice	Using the MWM , escape latency was reduced in GWI mice during training. In the probe trials, escape latency was reduced in GWI mice at d51 and d 86, but increased at d115.	(147)
PB + PER + DEET + RS for 28 days	Post Rx days 20–28	Male and female C57BL6 mice	In the MWM , differences in spatial learning were not observed in GWI Rx mice compared to control mice.	(148)
PB + PER + DEET +/- RRS for 28 days	3 months post Rx	SD rats	GWI Rx alone or in combination with RRS impaired learning during MWM training trials and impaired spatial memory in the 24-hour probe trial. In the NOR , GWI Rx + RRS rats exhibited reduced exploration times with the novel object but also exhibited decreases in total exploration time; therefore, DI was unaffected.	(141)
PB + PER + DEET + RRS for 28 days	3 months post Rx	SD rats	Compared to control rats, GWI rats exhibited decreased place recognition in the OLT and decreased object recognition in the NOR . ITI in these tests was one-hour.	(149)
PB + PER for 10 days	Post Rx days 18, 56, 77, 106	Male C57BL6 mice	In the BM , long term memory deficits were exclusively observed 106 days post GWI Rx.	(150)
PB + PER for 10 days	15- and 16-months post Rx	Male C57BL6 mice	In the BM , path lengths in the training sessions were greater in GWI mice compared to controls. In the 24-probe trial, path lengths were greater in GWI mice 15-months and 16-months post Rx.	(151)
DFP for 5 days	3 months post Rx	Male SD rats	In the OLT with a one-hour ITI, DFP mice exhibited reductions in DI compared to control rats.	(152)
DFP for 5 days	6 months post Rx	Male SD rats	In the NOR , DFP mice exhibited decreased DI following a one-hour ITI compared to control rats.	(153)
PB +/-RRS	10 days and 3 months post Rx	Male SD rats	In CFC , acquisition of freezing was unchanged but context-induced freezing was reduced in PB + RRS rats 10 days post-Rx. Three-months, post Rx, PB or PB + RRS rats did not exhibit differences in context freezing.	(72)
PB + PER + DEET + RRS for 28 days	3 months post Rx	Male and female B6.Cg-Tg EGFP mice	In the MWM , spatial learning and memory was similar in GWI mice and control mice.	(154)
PB +/-RRS	10 days post Rx	Male SD rats	Cue-conditioned freezing was reduced in PB-treated rats compared to control rats	(73)
PB + PER + DEET + RRS for 28 days	10 months post Rx	Male SD rats	GWI rats exhibited decreased object recognition in the NOR and decreased place recognition in the OLT .	(140)
PB +/-RRS	10 days and 3 months post Rx	Male SD rats	NOR performance was not affected 10 days post Rx. Three months post Rx, NOR performance is impaired in PB + RRS rats 24-hours following LPS administration. In the MWM , training performance was unaffected but PB-treated rats exhibited spatial learning deficits in the 24-hour probe trial.	(76)

BM, Barnes maze; CFC, contextual fear conditioning; DEET, N,N,-diethyl-meta-toluamide; DFP, diisopropyl flurophosphate; DI, discrimination index; FS, foot-shock; ITI, inter-trial interval; LPS, lipopolysaccharide; MWM, Morris water maze; NOR, novel object recognition; OLT, object location test; PB, pyridostigmine bromide; PCA, pole-climbing avoidance stress; PER, permethrin; RRS, repeated restraint stress; SD, Sprague-Dawley.

that the addition of an immune challenge would have resulted in more consistent behavioral outcomes across these other studies.

While cognitive performance varied across GWI models and laboratories in the tests described above, more consistent findings have been reported when hippocampal-dependent spatial learning and memory was assessed with water maze tests. For example, while some studies did not identify deficits in the Morris water maze (MWM) in a mouse model of GWI (154), a rat model of GWI exhibited deficits in water maze performance as early as 16 days after treatment cessation and these impairments remained at days 52, 113 and 199 (146). Other studies using a mouse model of GWI reported that MWM performance declines over time, as impairments were not seen at days 20–30 post-treatment (148), but emerged by day 115 (147). While we only assessed MWM performance 3 months after treatment cessation, we observed similar deficits in PB-treated rats, specifically during the 24-hour probe trial (76). Similarly, the group that did not observe deficits in NOR in their rat model of GWI 3 months post exposure reported significant impairments in MWM performance at this timepoint (141). Importantly, relative to the other behavioral paradigms, the MWM is innately stressful due to the swimming component (160), which further supports the idea that stressors must be included in GWI studies in order for cognitive impairments to fully emerge.

The stimulus-dependent cognitive impairments that are emerging in GWI rodent models provide further evidence of a latent phenotype of GWI and are consistent with clinical studies in which an exercise challenge is used to exacerbate cognitive deficits in GWI patients (133, 161, 162). Our studies also implicate the cholinergic system as a mechanistic mediator of GWI and suggest that deficits in the central cholinergic anti-inflammatory network may be responsible for the dysregulated immune and stress responses observed in GWI patients (Figure 1). In support of this concept, we have reported that hippocampal ACh efflux is potentiated in PB-treated rats when challenged with physiological stressors months later. These enhancements in cholinergic transmission may have cognitive consequences as sustained elevations in ACh levels are known to impair memory consolidation (75, 76, 163). Furthermore, a history of PB treatment appears to dysregulate ACh's anti-inflammatory actions in the hippocampus as the LPS-induced elevation in ACh efflux is accompanied by greater levels of pro-inflammatory hippocampal cytokines, which also contribute to cognitive deficits (75, 76, 164–166). While we have only assessed the effects of a single physiological stressor in PB-treated rats (i.e., LPS treatment or immobilization stress), GW Veterans continue to face daily life stressors and occasional infections. Thus, the repeated activation of this sensitized cholinergic system within the CNS is likely exacerbated over time and may explain the progressive immune dysregulation, chronic inflammation, and cognitive deficits observed in GWI patients.

The potential sensitization of the central cholinergic system in GWI patients is especially interesting as GW Veterans continue to age. Much like GWI, aging is associated with a “primed” immune system, both in the periphery and CNS, which causes heightened pro-inflammatory responses to stressors (167–169). Additionally, it is well established that aged brains exhibit reduced cholinergic transmission, and the loss of BF cholinergic neurons projecting to brain regions mediating cognition is a hallmark feature of age-

related cognitive decline (ARCD) and Alzheimer's disease (AD) (170). Similar deficits have been observed in GWI studies as 12% of GW Veterans (median age of 48 years) exhibit reduced hippocampal volumes and MCI (58). This rate of MCI is significantly greater than the general population, as the prevalence of MCI in individuals aged 60–64 and 65–69 is 6.7% and 8.4%, respectively (171). These findings suggest that GWI is causing accelerated brain aging, and our rodent studies suggest that this acceleration is due to lasting impairments in the cholinergic anti-inflammatory network. While our rat model of GWI exhibits the same immune sensitivity observed in aging studies, more longitudinal studies are needed to determine if repeated stressors produce similar reductions in hippocampal volume and a loss of cholinergic transmission.

6 Potential therapeutics for GWI

Though the mechanisms driving GWI pathology remain undefined, the evidence of immune dysregulation in both preclinical and clinical studies highlight the potential therapeutic benefit of anti-inflammatory interventions for this patient population (Table 2). One anti-inflammatory compound that has shown promise in preclinical GWI studies is Lacto-N-fucopentaose III (LNFPIII), an immunomodulatory glycan found in human breast milk (175). Treating a mouse model of GWI with LNFPIII either 7 months or 11 months post-exposure decreased hippocampal levels of IL-6 and increased hippocampal levels of brain-derived neurotrophic factor (BDNF) and nerve growth factor (NGF) (172). Additionally, the same group of investigators found that LNFPIII ameliorates the deficits in hippocampal synaptic plasticity and transmission observed in their mouse model (172, 176). Another family of compounds that have shown anti-inflammatory effects in GWI studies are activators of nuclear factor [erythroid-derived 2]-like 2 (Nrf2), which is a transcription factor that regulates antioxidant responses. This includes melatonin, which normalized Nrf2 levels in a rat model of GWI as well as improved recognition memory (155), and *tert*-butylhydroquinone (tBHQ), which ameliorated reductions in dendritic complexity of dentate granule cells observed in a mouse model of GWI (144). Additionally, various GWI studies have assessed the therapeutic benefit of the antioxidant curcumin. Curcumin has been shown to normalize a number of genes related to mitochondrial respiration in a rat model of GWI, which was associated with reduced hippocampal inflammation, enhanced neurogenesis, and improved performance on cognitive assessments (143). Similar findings were reported in a small clinical study that saw reduced GWI symptom severity in curcumin-treated patients relative to placebo-treated controls (156). Clinical studies have also been performed with the antioxidant L-Carnosine (β -alanine-L-histidine), and GWI patients showed significant improvements in cognitive function after 12 weeks of treatment (157).

Beyond anti-inflammatory compounds, GWI researchers have also assessed the therapeutic potential of vagal nerve stimulation (VNS) as it can increase the vagus nerve's anti-inflammatory actions. Performing VNS in a mouse model of GWI over 30 weeks after the initial GW chemical exposure reduced the

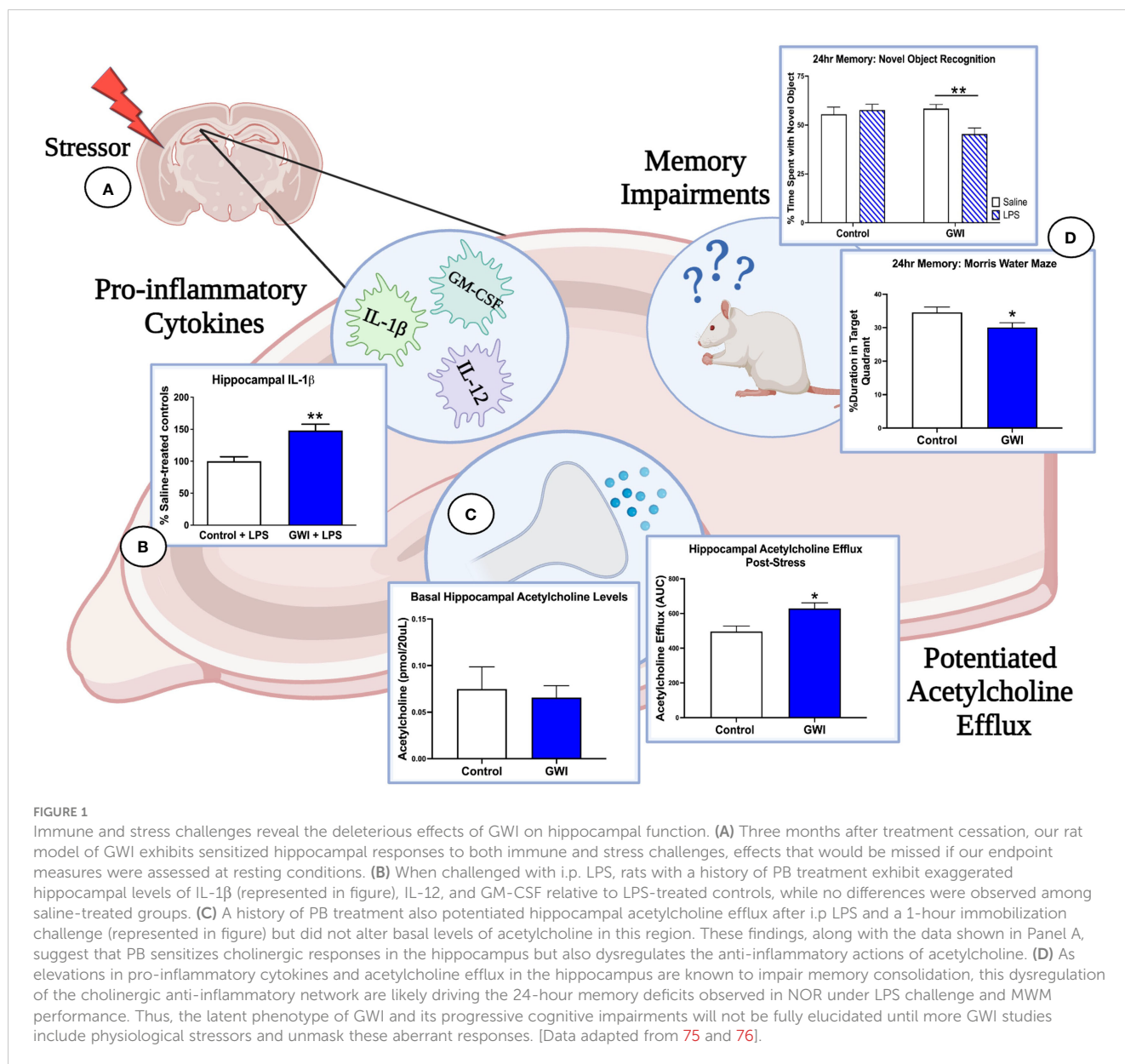


FIGURE 1

Immune and stress challenges reveal the deleterious effects of GWI on hippocampal function. (A) Three months after treatment cessation, our rat model of GWI exhibits sensitized hippocampal responses to both immune and stress challenges, effects that would be missed if our endpoint measures were assessed at resting conditions. (B) When challenged with i.p. LPS, rats with a history of PB treatment exhibit exaggerated hippocampal levels of IL-1 β (represented in figure), IL-12, and GM-CSF relative to LPS-treated controls, while no differences were observed among saline-treated groups. (C) A history of PB treatment also potentiated hippocampal acetylcholine efflux after i.p. LPS and a 1-hour immobilization challenge (represented in figure) but did not alter basal levels of acetylcholine in this region. These findings, along with the data shown in Panel A, suggest that PB sensitizes cholinergic responses in the hippocampus but also dysregulates the anti-inflammatory actions of acetylcholine. (D) As elevations in pro-inflammatory cytokines and acetylcholine efflux in the hippocampus are known to impair memory consolidation, this dysregulation of the cholinergic anti-inflammatory network are likely driving the 24-hour memory deficits observed in NOR under LPS challenge and MWM performance. Thus, the latent phenotype of GWI and its progressive cognitive impairments will not be fully elucidated until more GWI studies include physiological stressors and unmask these aberrant responses. [Data adapted from 75 and 76].

intensity of the astrocytic marker, glial fibrillary acidic protein (GFAP), in the hippocampus and improved cognitive performance in the object location test (158). VNS also ameliorated the reduced hippocampal neurogenesis observed in this GWI model and moderately improved performance in the pattern separation task (159). While this intervention may be promising for the cognitive deficits associated with GWI, VNS may not provide relief for GWI Veterans who experience chronic pain as Veterans with GWI that underwent VNS for 10 weeks did not see a reduction in widespread pain or migraine severity or frequency (177). Though some studies have shown positive effects of VNS on disorders associated with chronic pain (178, 179) and migraines (180), other studies suggest that VNS can lower pain thresholds in rodents and humans (181, 182). Thus, future clinical studies are needed to assess the potential benefits of VNS for GWI-related symptoms, especially cognitive impairments, as well as possible adverse effects in this patient population.

Along with VNS, there are also pharmacological interventions that can restore the cholinergic anti-inflammatory response that may be diminished in GWI patients and potentially alleviate hippocampal neuroinflammation and cognitive impairments. Positive allosteric modulators of $\alpha 7$ nAChRs ($\alpha 7$ PAMs) are emerging as potential therapeutics for a variety of pathologies, ranging from ulcerative colitis to AD, for their anti-inflammatory and pro-cognitive actions (183–185). Unlike agonists that have the potential to elicit receptor desensitization, $\alpha 7$ PAMs are an attractive option as they only augment natural cholinergic signaling (186). Additionally, the allosteric binding site creates higher binding specificity than current pharmacological treatments (187). PNU-120596 is an $\alpha 7$ PAM that has been shown to cross the BBB and inhibit LPS-induced neuroinflammation in rodent studies (188, 189). Furthermore, PNU-120596 has been shown to enhance cognition and prevent the cognitive deficits associated with LPS administration, making it an excellent candidate for future GWI studies (189–193). Collectively,

TABLE 2 Therapeutic interventions targeting inflammation in GWI.

Intervention	Rationale	Species Tested	Outcomes	Reference
Lacto-N-fucopentaose III	Immunomodulatory glycan with anti-inflammatory properties	Male C57BL6 mice	Reduced hippocampal IL-6 levels. Increased hippocampal BDNF and NGF levels. Ameliorated hippocampal synaptic plasticity.	(172)
Melatonin	Nrf2 activator	Male SD rats	Normalized Nrf2 levels in hippocampus. Reduced astrocyte hypertrophy in hippocampus. Reduced number of activated microglia in hippocampus. Improved performance in NOR and OLT	(155)
<i>tert</i> -butylhydroquinone	Nrf2 activator	Male C57BL6 mice	Increases dendritic complexity of dentate gyrate cells.	(144)
Curcumin	Antioxidant	Male SD rats	Normalized genes associated with mitochondrial respiration. Reduced astrocyte hypertrophy in hippocampus. Reduced number of activated microglia in hippocampus. Enhanced hippocampal neurogenesis. Improved performance in OLT and NOR	(143)
		Human	Reduced symptom severity (self-reported).	(156)
L-Carnosine	Antioxidant	Human	Improved cognitive function.	(157)
Low Glutamate Diet	Reduce excitotoxicity by eliminating dietary amino acids	Human	Reduced serum levels of IL-1 β .	(173)
			Reduced symptom severity (self-reported).	(174)
Vagal Nerve Stimulation	Stimulate the vagal-mediated cholinergic anti-inflammatory pathway	Male CD1 mice	Reduced GFAP intensity in hippocampus. Improved performance in OLT.	(158)
			Increased hippocampal neurogenesis Moderately improved performance in PST	(159)

BDNF, brain derived neurotrophic factor; GFAP, glial fibrillary acidic protein; IL, interleukin; NGF, nerve growth factor; NOR, novel object recognition; Nrf2, nuclear factor [erythroid-derived 2]-like 2; OLT, object location test; PST, pattern separation task; SD, Sprague Dawley.

these intervention studies further support the hypothesis that peripheral and CNS inflammation, likely resulting from dysregulation of the cholinergic anti-inflammatory network, is a mechanistic mediator of GWI. More importantly, these studies suggest that restoration of the activity of the cholinergic anti-inflammatory network represents a provocative treatment strategy to ameliorate the progressive cognitive deficits observed in Veterans with GWI.

7 Conclusions

In sum, the studies highlighted throughout this review provide strong evidence for a dysregulation of immune and endocrine signaling in GWI, which likely contributes to the progressive cognitive impairments observed in patients. While some studies implicate chronic neuroinflammation as a chief mediator of the cognitive deficits seen in GWI, other studies suggest that such deficits are due to disruptions in central cholinergic signaling that only emerge when a stressor is presented. One major debate within GWI research has been the effect of AChE inhibitors on these cognitive deficits as they should not cross the BBB and affect central cholinergic activity. Our studies support this idea as we do not observe any differences in basal levels of ACh or AChE activity in the hippocampus of our rat model of GWI. However, our studies suggest that PB sensitizes the cholinergic system to physiological stressors, including

immune and stress challenges, and these aberrant responses are impairing memory consolidation. Furthermore, the repeated activation of this sensitized cholinergic system over the lifespan likely dysregulates ACh signaling, including its important anti-inflammatory actions, potentiating peripheral and central inflammation and accelerating cognitive decline. These findings are highly consistent with clinical data that report an exacerbation of GWI symptoms, especially cognitive function, when GWI patients are performing an exercise challenge. While this is an exciting area of research, the field needs future studies to include such stressors in preclinical models, as well as alternative stressors in clinical assessments to fully understand the latent phenotype of GWI. These additions will provide better opportunities to determine if peripheral inflammation is driving cognitive deficits or if the central cholinergic system becomes dysregulated through an alternative mechanism. Most importantly, such studies will build upon this strong foundation of GWI research and help identify potential therapeutics that will slow or stop this debilitating disease.

Author contributions

HB: Conceptualization, Formal analysis, Funding acquisition, Writing – original draft, Writing – review & editing. LR: Conceptualization, Formal analysis, Funding acquisition, Writing – original draft, Writing – review & editing.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

References

- Auxéméry Y. [The Gulf War Syndrome twenty years on]. *Encephale*. (2013) 39:332–8. doi: 10.1016/j.encep.2012.11.003. (Le syndrome de la guerre du Golfe vingt ans après.)
- Kerr KJ. Gulf War illness: an overview of events, most prevalent health outcomes, exposures, and clues as to pathogenesis. *Rev Environ Health*. (2015) 30:273–86. doi: 10.1515/revheh-2015-0032
- Britannica E. Persian Gulf War (2023). Available at: <https://www.britannica.com/event/Persian-Gulf-War>.
- Program. *Evaluation of the US Department of Defense Persian Gulf Comprehensive Clinical Evaluation Program*. Washington, DC: National Academy Press (1996).
- Kang HK, Mahan CM, Lee KY, Magee CA, Murphy FM. Illnesses among United States veterans of the Gulf War: a population-based survey of 30,000 veterans. *J Occup Environ Med*. (2000) 42:491–501. doi: 10.1097/00043764-200005000-00006
- Nettleman M. Gulf War Illness: challenges persist. *Trans Am Clin Climatol Assoc*. (2015) 126:237–47.
- Blanchard MS, Eisen SA, Alpern R, Karlinsky J, Toomey R, Reda DJ, et al. Chronic multisymptom illness complex in Gulf War I veterans 10 years later. *Am J Epidemiol*. (2005) 163:66–75. doi: 10.1093/aje/kwj008
- Steele L. Prevalence and patterns of Gulf War illness in Kansas veterans: association of symptoms with characteristics of person, place, and time of military service. *Am J Epidemiol*. (2000) 152:992–1002. doi: 10.1093/aje/152.10.992
- Fukuda K, Nisenbaum R, Stewart G, Thompson WW, Robin L, Washko RM, et al. Chronic multisymptom illness affecting Air Force veterans of the Gulf War. *Jama*. (1998) 280:981–8. doi: 10.1001/jama.280.11.981
- Kang HK, Li B, Mahan CM, Eisen SA, Engel CC. Health of US veterans of 1991 Gulf War: a follow-up survey in 10 years. *J Occup Environ Med*. (2009) 51(4):401–10. doi: 10.1097/JOM.0b013e3181a2feeb
- Mawson AR, Croft AM. Gulf war illness: unifying hypothesis for a continuing health problem. *Int J Environ Res Public Health*. (2019) 16:111. doi: 10.3390/ijerph16010111
- Li B, Mahan CM, Kang HK, Eisen SA, Engel CC. Longitudinal health study of US 1991 Gulf War veterans: changes in health status at 10-year follow-up. *Am J Epidemiol*. (2011) 174:761–8. doi: 10.1093/aje/kwr154
- Chao LL. Olfactory and cognitive decrements in 1991 Gulf War veterans with gulf war illness/chronic multisymptom illness. *Environ Health*. (2024) 23:14. doi: 10.1186/s12940-024-01058-2
- Christova P, James LM, Engdahl BE, Lewis SM, Carpenter AF, Georgopoulos AP. Subcortical brain atrophy in Gulf War Illness. *Exp Brain Res*. (2017) 235:2777–86. doi: 10.1007/s00221-017-5010-8
- Zhang Y, Avery T, Vakhtin AA, Mathersul DC, Tranvinh E, Wintermark M, et al. Brainstem atrophy in Gulf War Illness. *Neurotoxicology*. (2020) 78:71–9. doi: 10.1016/j.neuro.2020.02.006
- Lindheimer JB, Stegner AJ, Wylie GR, Klein-Adams JC, Almassi NE, Ninneman JV, et al. Post-exertional malaise in veterans with Gulf War Illness. *Int J Psychophysiol*. (2020) 147:202–12. doi: 10.1016/j.ijpsycho.2019.11.008

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- Zhang Q, Zhou XD, Denny T, Ottenweller JE, Lange G, LaManca JJ, et al. Changes in immune parameters seen in Gulf War veterans but not in civilians with chronic fatigue syndrome. *Clin Diagn Lab Immunol*. (1999) 6:6–13. doi: 10.1128/cdli.6.1.6-13.1999
- Vojdani A, Thrasher JD. Cellular and humoral immune abnormalities in Gulf War veterans. *Environ Health Perspect*. (2004) 112:840–6. doi: 10.1289/ehp.6881
- Skowera A, Hotopf M, Sawicka E, Varela-Calvino R, Unwin C, Nikolaou V, et al. Cellular immune activation in Gulf War veterans. *J Clin Immunol*. (2004) 24:66–73. doi: 10.1023/B:JOCI.0000018065.64685.82
- Johnson GJ, Leis LA, Slater BC, Bach RR. Elevated platelet count, C-reactive protein and thromboxane analog-induced platelet aggregation in patients with Gulf War veterans' illnesses: evidence of a chronic inflammatory state? *Blood Coagul Fibrinolysis*. (2013) 24:736–41. doi: 10.1097/MBC.0b013e328362627f
- Butterick TA, Trembley JH, Hocum Stone LL, Muller CJ, Rudquist RR, Bach RR. Gulf War Illness-associated increases in blood levels of interleukin 6 and C-reactive protein: biomarker evidence of inflammation. *BMC Res Notes*. (2019) 12:1–5. doi: 10.1186/s13104-019-4855-2
- Khaiboullina SF, DeMeirleir KL, Rawat S, Berk GS, Gaynor-Berk RS, Mijatovic T, et al. Cytokine expression provides clues to the pathophysiology of Gulf War illness and myalgic encephalomyelitis. *Cytokine*. (2015) 72:1–8. doi: 10.1016/j.cyto.2014.11.019
- Bardoel BW, Elaine F, Sollberger G, Zychlinsky A. The balancing act of neutrophils. *Cell Host Microbe*. (2014) 15:526–36. doi: 10.1016/j.chom.2014.04.011
- Jang DI, Lee AH, Shin HY, Song HR, Park JH, Kang TB, et al. The role of tumor necrosis factor alpha (TNF- α) in autoimmune disease and current TNF- α Inhibitors in therapeutics. *Int J Mol Sci*. (2021) 22. doi: 10.3390/ijms22052719
- Herrero-Cervera A, Soehnlein O, Kenne E. Neutrophils in chronic inflammatory diseases. *Cell Mol Immunol*. (2022) 19:177–91. doi: 10.1038/s41423-021-00832-3
- James LM, Georgopoulos AP. At the root of 3 "Long" Diseases: persistent antigens inflicting chronic damage on the brain and other organs in Gulf War Illness, long-COVID-19, and chronic fatigue syndrome. *Neurosci Insights*. (2022) 17:26331055221114817. doi: 10.1177/26331055221114817
- Andersson U, Tracey KJ. Neural reflexes in inflammation and immunity. *J Exp Med*. (2012) 209:1057–68. doi: 10.1084/jem.20120571
- Banks WA. Blood-brain barrier transport of cytokines: a mechanism for neuropathology. *Curr Pharm design*. (2005) 11:973–84. doi: 10.2174/1381612053381684
- Tracey KJ. The inflammatory reflex. *Nature*. (2002) 420:853–9. doi: 10.1038/nature01321
- Garza-Lombó C, Thang M, Greve HJ, Mumaw CL, Messenger EJ, Ahmed C, et al. Circulating HMGB1 is elevated in veterans with Gulf War Illness and triggers the persistent pro-inflammatory microglia phenotype in male C57Bl/6j mice. *Transl Psychiatry*. (2021) 11:390. doi: 10.1038/s41398-021-01517-1
- Gao H-M, Zhou H, Zhang F, Wilson BC, Kam W, Hong J-S. HMGB1 acts on microglia Mac1 to mediate chronic neuroinflammation that drives progressive neurodegeneration. *J Neurosci*. (2011) 31:1081–92. doi: 10.1523/JNEUROSCI.3732-10.2011

32. Vacas S, Degos V, Tracey KJ, Maze M. High-mobility group box 1 protein initiates postoperative cognitive decline by engaging bone marrow-derived macrophages. *Anesthesiology*. (2014) 120:1160–7. doi: 10.1097/ALN.0000000000000045
33. Wu T-Y, Liu L, Zhang W, Zhang Y, Liu Y-Z, Shen X-L, et al. High-mobility group box-1 was released actively and involved in LPS induced depressive-like behavior. *J Psychiatr Res*. (2015) 64:99–106. doi: 10.1016/j.jpsychires.2015.02.016
34. Lian Y-J, Gong H, Wu T-Y, Su W-J, Zhang Y, Yang Y-Y, et al. Ds-HMGB1 and fr-HMGB induce depressive behavior through neuroinflammation in contrast to nonoxid-HMGB1. *Brain Behavior Immun*. (2017) 59:322–32. doi: 10.1016/j.bbi.2016.09.017
35. Georgopoulos AP, Tsilibary EC, Souto EP, James LM, Engdahl BE, Georgopoulos A. Adverse effects of Gulf War Illness (GWI) serum on neural cultures and their prevention by healthy serum. *J Neurol Neuromedicine*. (2018) 3:19–27. doi: 10.29245/2572.942x/2018/2.1177
36. Alshelh Z, Albrecht DS, Bergan C, Akeju O, Clauw DJ, Conboy L, et al. *In-vivo* imaging of neuroinflammation in veterans with Gulf War illness. *Brain Behav Immun*. (2020) 87:498–507. doi: 10.1016/j.bbi.2020.01.020
37. Papadimitriou A, Priftis KN. Regulation of the hypothalamic-pituitary-adrenal axis. *Neuroimmunomodulation*. (2009) 16:265–71. doi: 10.1159/000216184
38. Oei TI. The dexamethasone suppression test as a variable in clinical diagnosis and research: a review. *J R Soc Med*. (1988) 81:31–3. doi: 10.1177/014107688808100114
39. Golier JA, Schmeidler J, Legge J, Yehuda R. Enhanced cortisol suppression to dexamethasone associated with Gulf War deployment. *Psychoneuroendocrinology*. (2006) 31:1181–9. doi: 10.1016/j.psyneuen.2006.08.005
40. Golier JA, Schmeidler J, Legge J, Yehuda R. Twenty-four hour plasma cortisol and adrenocorticotropic hormone in Gulf War veterans: relationships to posttraumatic stress disorder and health symptoms. *Biol Psychiatry*. (2007) 62:1175–8. doi: 10.1016/j.biopsych.2007.04.027
41. Liberman AC, Budziński ML, Sokn C, Gobbi RP, Steininger A, Arzt E. Regulatory and mechanistic actions of glucocorticoids on T and inflammatory cells. *Front Endocrinol*. (2018) 9:235. doi: 10.3389/fendo.2018.00235
42. Tapp ZM, Godbout JP, Kokiko-Cochran ON. A tilted axis: maladaptive inflammation and HPA axis dysfunction contribute to consequences of TBI. *Front Neurol*. (2019) 10:345. doi: 10.3389/fneur.2019.00345
43. White RF, Steele L, O'Callaghan JP, Sullivan K, Binns JH, Golomb BA, et al. Recent research on Gulf War illness and other health problems in veterans of the 1991 Gulf War: Effects of toxicant exposures during deployment. *Cortex*. (2016) 74:449–75. doi: 10.1016/j.cortex.2015.08.022
44. Gordon J, Leadbeater L, Maidment MP. The protection of animals against organophosphate poisoning by pretreatment with a carbamate. *Toxicol Appl Pharmacol*. (1978) 43:207–16. doi: 10.1016/S0041-008X(78)80045-3
45. von Bredow JD, Adams NL, Groff WA, Vick JA. Effectiveness of oral pyridostigmine pretreatment and cholinergic-oxime therapy against soman intoxication in nonhuman primates. *Fundam Appl Toxicol*. (1991) 17:761–70. doi: 10.1093/toxsci/17.4.761
46. Sox HC, Liverman CT, Fulco CE. *Gulf War and health: Volume 1: Depleted uranium, sarin, pyridostigmine bromide, and vaccines* Vol. 1. National Academies Press (2000).
47. Keesey JC. A history of treatments for myasthenia gravis. *Semin Neurol*. (2004). doi: 10.1055/s-2004-829584
48. Pohanka M. Cholinesterases, a target of pharmacology and toxicology. *BioMed Pap Med Fac Univ Palacky Olomouc Czech Repub*. (2011) 155:219–29. doi: 10.5507/bp.2011.036
49. Colović MB, Krstić DZ, Lazarević-Pašti TD, Bondžić AM, Vasić VM. Acetylcholinesterase inhibitors: pharmacology and toxicology. *Curr Neuropharmacol*. (2013) 11:315–35. doi: 10.2174/1570159x11311030006
50. Golomb BA. Acetylcholinesterase inhibitors and Gulf War illnesses. *Proc Natl Acad Sci U.S.A.* (2008) 105:4295–300. doi: 10.1073/pnas.0711986105
51. Sidell FR, Wilde H. Medical aspects of chemical and biological warfare. *JAMA-Journal Am Med Association-International Edition*. (1998) 280:1197. doi: 10.1001/jama.280.13.1199
52. Mukherjee S, Gupta RD. Organophosphorus nerve agents: types, toxicity, and treatments. *J Toxicol*. (2020) 2020:3007984. doi: 10.1155/2020/3007984
53. Berry WK, Davies DR. The use of carbamates and atropine in the protection of animals against poisoning by 1,2,2-trimethylpropyl methylphosphonofluoridate. *Biochem Pharmacol*. (1970) 19:927–34. doi: 10.1016/0006-2952(70)90256-X
54. Rettig RA. *Military use of drugs not yet approved by the FDA for CW/BW defense: Lessons from the Gulf War* Vol. 9. United States: Minnesota Historical Society (1999). doi: 10.7249/MR1018.9
55. Marino MT, Schuster BG, Brueckner RP, Lin E, Kaminski A, Lasseter KC. Population pharmacokinetics and pharmacodynamics of pyridostigmine bromide for prophylaxis against nerve agents in humans. *J Clin Pharmacol*. (1998) 38:227–35. doi: 10.1002/j.1552-4604.1998.tb04420.x
56. Keeler JR, Hurst CG, Dunn MA. Pyridostigmine used as a nerve agent pretreatment under wartime conditions. *Jama*. (1991) 266:693–5. doi: 10.1001/jama.266.5.693
57. Chao LL, Rothlind JC, Cardenas VA, Meyerhoff DJ, Weiner MW. Effects of low-level exposure to sarin and cyclosarin during the 1991 Gulf War on brain function and brain structure in US veterans. *Neurotoxicology*. (2010) 31:493–501. doi: 10.1016/j.jneuro.2010.05.006
58. Chao LL. The prevalence of mild cognitive impairment in a convenience sample of 202 gulf war veterans. *Int J Environ Res Public Health*. (2020) 17. doi: 10.3390/ijerph17191758
59. Institute of Medicine (US) Committee on the Evaluation of the Department of Defense Comprehensive Clinical Evaluation Program. *Adequacy of the Comprehensive Clinical Evaluation Program: A Focused Assessment*. National Academies Press (US). (1997).
60. Haley RW, Kurt TL, Hom J. Is there a Gulf war syndrome?: searching for syndromes by factor analysis of symptoms. *Jama*. (1997) 277:215–22. doi: 10.1001/jama.277.3.215
61. Pope C, Karanth S, Liu J. Pharmacology and toxicology of cholinesterase inhibitors: uses and misuses of a common mechanism of action. *Environ Toxicol Pharmacol*. (2005) 19:433–46. doi: 10.1016/j.etap.2004.12.048
62. Steele L, Sastre A, Gerkovich MM, Cook MR. Complex factors in the etiology of Gulf War illness: wartime exposures and risk factors in veteran subgroups. *Environ Health Perspect*. (2012) 120:112–8. doi: 10.1289/ehp.1003399
63. Remijn-Nelissen L, Verschuuren J, Tannemaat MR. The effectiveness and side effects of pyridostigmine in the treatment of myasthenia gravis: a cross-sectional study. *Neuromuscul Disord*. (2022) 32:790–9. doi: 10.1016/j.nmd.2022.09.002
64. Eells JT, Bandettini PA, Holman PA, Propp JM. Pyrethroid insecticide-induced alterations in mammalian synaptic membrane potential. *J Pharmacol Exp Ther*. (1992) 262:1173–81.
65. Rao GV, Jagannatha Rao KS. Modulation in acetylcholinesterase of rat brain by pyrethroids *in vivo* and an *in vitro* kinetic study. *J Neurochemistry*. (1995) 65:2259–66. doi: 10.1046/j.1471-4159.1995.65052259.x
66. Ribeiro ACR, Deshpande LS. A review of pre-clinical models for Gulf War Illness. *Pharmacol Ther*. (2021) 228:107936. doi: 10.1016/j.pharmthera.2021.107936
67. Cecchine G, Golomb BA, Hilborne LH, Spector DM, Anthony C. A review of the scientific literature as it pertains to gulf war illnesses. In: *Pesticides*, vol. 8. Santa Monica, CA: RAND Corporation (2000).
68. Binns JH, Barlow C, Bloom FE, Clauw DJ, Golomb BA, Graves JC, et al. Gulf War illness and the health of Gulf War veterans. In: *Scientific Findings and Recommendations*. US Government Printing Office, Washington DC (2008).
69. Swale DR, Sun B, Tong F, Bloomquist JR. Neurotoxicity and mode of action of N, N-diethyl-meta-toluamide (DEET). *PLoS One*. (2014) 9:e103713. doi: 10.1371/journal.pone.0103713
70. Kant GJ, Bauman RA, Feaster SR, Anderson SM, Saviolakis GA, Garcia GE. The combined effects of pyridostigmine and chronic stress on brain cortical and blood acetylcholinesterase, corticosterone, prolactin and alternation performance in rats. *Pharmacol Biochem Behav*. (2001) 70:209–18. doi: 10.1016/S0091-3057(01)00596-2
71. Amourette C, Lamproglou I, Barbier L, Fauquette W, Zoppe A, Viret R, et al. Gulf War illness: Effects of repeated stress and pyridostigmine treatment on blood-brain barrier permeability and cholinesterase activity in rat brain. *Behav Brain Res*. (2009) 203:207–14. doi: 10.1016/j.bbr.2009.05.002
72. Macht VA, Woodruff JL, Grillo CA, Wood CS, Wilson MA, Reagan LP. Pathophysiology in a model of Gulf War Illness: Contributions of pyridostigmine bromide and stress. *Psychoneuroendocrinology*. (2018) 96:195–202. doi: 10.1016/j.psyneuen.2018.07.015
73. Macht VA, Woodruff JL, Maissy ES, Grillo CA, Wilson MA, Fadel JR, et al. Pyridostigmine bromide and stress interact to impact immune function, cholinergic neurochemistry and behavior in a rat model of Gulf War Illness. *Brain Behav Immun*. (2019) 80:384–93. doi: 10.1016/j.bbi.2019.04.015
74. Macht VA, Woodruff JL, Burzynski HE, Grillo CA, Reagan LP, Fadel JR. Interactions between pyridostigmine bromide and stress on glutamatergic neurochemistry: Insights from a rat model of Gulf War Illness. *Neurobiol Stress*. (2020) 12:100210. doi: 10.1016/j.yjnstr.2019.100210
75. Burzynski HE, Macht VA, Woodruff JL, Crawford JN, Erichsen JM, Piroli GG, et al. Pyridostigmine bromide elicits progressive and chronic impairments in the cholinergic anti-inflammatory pathway in the prefrontal cortex and hippocampus of male rats. *Neurobiol Stress*. (2022) 18:100446. doi: 10.1016/j.yjnstr.2022.100446
76. Burzynski HE, Ayala KE, Frick MA, Dufala HA, Woodruff JL, Macht VA, et al. Delayed cognitive impairments in a rat model of Gulf War Illness are stimulus-dependent. *Brain Behav Immun*. (2023) 113:248–58. doi: 10.1016/j.bbi.2023.07.003
77. Gifford RK, Ursano RJ, Stuart JA, Engel CC. Stress and stressors of the early phases of the Persian Gulf War. *Philos Trans R Soc B: Biol Sci*. (2006) 361:585–91. doi: 10.1098/rstb.2006.1818
78. Jeffrey M, Collado F, Kibler J, DeLucia C, Messer S, Klimas N, et al. Post-traumatic stress impact on health outcomes in Gulf War Illness. *BMC Psychol*. (2021) 9:57. doi: 10.1186/s40359-021-00561-2
79. Proctor SP, Heeren T, White RF, Wolfe J, Borgos MS, Davis JD, et al. Health status of Persian Gulf War veterans: self-reported symptoms, environmental exposures and the effect of stress. *Int J Epidemiol*. (1998) 27:1000–10. doi: 10.1093/ije/27.6.1000
80. McEwen BS. Plasticity of the hippocampus: adaptation to chronic stress and allostatic load. *Ann N Y Acad Sci*. (2001) 933:265–77. doi: 10.1111/j.1749-6632.2001.tb05830.x

81. Gould E, Tanapat P, McEwen BS, Flugge G, Fuchs E. Proliferation of granule cell precursors in the dentate gyrus of adult monkeys is diminished by stress. *Proc Natl Acad Sci*. (1998) 95:3168–71. doi: 10.1073/pnas.95.6.3168
82. Magariños AM, McEwen BS. Chronic stress alters synaptic terminal structure in hippocampus. *Proc Natl Acad Sci USA*. (1997) 94:14002–8.
83. Radley JJ, Rocher AB, Janssen WG, Hof PR, McEwen BS, Morrison JH. Reversibility of apical dendritic retraction in the rat medial prefrontal cortex following repeated stress. *Exp Neurol*. (2005) 196:199–203. doi: 10.1016/j.jepneurol.2005.07.008
84. Arnsten AF. Stress signalling pathways that impair prefrontal cortex structure and function. *Nat Rev Neurosci*. (2009) 10:410–22. doi: 10.1038/nrn2648
85. Picciotto MR, Higley MJ, Mineur YS. Acetylcholine as a neuromodulator: cholinergic signaling shapes nervous system function and behavior. *Neuron*. (2012) 76:116–29. doi: 10.1016/j.neuron.2012.08.036
86. Mineur YS, Mose TN, Vanopdenbosch L, Etherington IM, Ogbjesi C, Islam A, et al. Hippocampal acetylcholine modulates stress-related behaviors independent of specific cholinergic inputs. *Mol Psychiatry*. (2022) 27:1829–38. doi: 10.1038/s41380-021-01404-7
87. Mesulam M-M, Mufson EJ, Levey AI, Wainer BH. Cholinergic innervation of cortex by the basal forebrain: Cytochemistry and cortical connections of the septal area, diagonal band nuclei, nucleus basalis (Substantia innominata), and hypothalamus in the rhesus monkey. *J Comp Neurol*. (1983) 214:170–97. doi: 10.1002/cne.902140206
88. Woolf NJ. Cholinergic systems in mammalian brain and spinal cord. *Prog Neurobiol*. (1991) 37:475–524. doi: 10.1016/0301-0082(91)90006-M
89. Atri A, Sherman S, Norman KA, Kirchoff BA, Nicolas MM, Greicius MD, et al. Blockade of central cholinergic receptors impairs new learning and increases proactive interference in a word paired-associate memory task. *Behav Neurosci*. (2004) 118:223–36. doi: 10.1037/0735-7044.118.1.223
90. Green A, Ellis KA, Ellis J, Bartholomew CF, Ilic S, Croft RJ, et al. Muscarinic and nicotinic receptor modulation of object and spatial n-back working memory in humans. *Pharmacol Biochem Behav*. (2005) 81:575–84. doi: 10.1016/j.pbb.2005.04.010
91. McGaughy J, Kaiser T, Sarter M. Behavioral vigilance following infusions of 192 IgG-saporin into the basal forebrain: selectivity of the behavioral impairment and relation to cortical AChE-positive fiber density. *Behav Neurosci*. (1996) 110:247–65. doi: 10.1037//0735-7044.110.2.247
92. Turchi J, Sarter M. Cortical acetylcholine and processing capacity: effects of cortical cholinergic deafferentation on crossmodal divided attention in rats. *Cogn Brain Res*. (1997) 6:147–58. doi: 10.1016/S0926-6410(97)00027-X
93. McGaughy J, Sarter M. Sustained attention performance in rats with intracortical infusions of 192 IgG-saporin-induced cortical cholinergic deafferentation: effects of physostigmine and FG 7142. *Behav Neurosci*. (1998) 112:1519–25. doi: 10.1037//0735-7044.112.6.1519
94. Dalley JW, Theobald DE, Bouger P, Chudasama Y, Cardinal RN, Robbins TW. Cortical cholinergic function and deficits in visual attentional performance in rats following 192 IgG-saporin-induced lesions of the medial prefrontal cortex. *Cereb Cortex*. (2004) 14:922–32. doi: 10.1093/cercor/bhh052
95. Newman LA, McGaughy J. Cholinergic deafferentation of prefrontal cortex increases sensitivity to cross-modal distractors during a sustained attention task. *J Neurosci*. (2008) 28:2642–50. doi: 10.1523/jneurosci.5112-07.2008
96. Wonnacott S. Presynaptic nicotinic ACh receptors. *Trends Neurosci*. (1997) 20:92–8. doi: 10.1016/S0166-2236(96)10073-4
97. Rice ME, Cragg SJ. Nicotine amplifies reward-related dopamine signals in striatum. *Nat Neurosci*. (2004) 7:583–4. doi: 10.1038/nn1244
98. Zhang H, Sulzer D. Frequency-dependent modulation of dopamine release by nicotine. *Nat Neurosci*. (2004) 7:581–2. doi: 10.1038/nn1243
99. Kawai H, Lazar R, Metherate R. Nicotinic control of axon excitability regulates thalamocortical transmission. *Nat Neurosci*. (2007) 10:1168–75. doi: 10.1038/nn1956
100. Pavlov VA, Wang H, Czura CJ, Friedman SG, Tracey KJ. The cholinergic anti-inflammatory pathway: a missing link in neuroimmunomodulation. *Mol Med*. (2003) 9:125–34. doi: 10.1007/BF03402177
101. Borovikova LV, Ivanova S, Zhang M, Yang H, Botchkina GI, Watkins LR, et al. Vagus nerve stimulation attenuates the systemic inflammatory response to endotoxin. *Nature*. (2000) 405:458–62. doi: 10.1038/35013070
102. Wang H, Yu M, Ochani M, Amella CA, Tanovic M, Susarla S, et al. Nicotinic acetylcholine receptor alpha7 subunit is an essential regulator of inflammation. *Nature*. (2003) 421:384–8. doi: 10.1038/nature01339
103. Hamano R, Takahashi HK, Iwagaki H, Yoshino T, Nishibori M, Tanaka N. Stimulation of $\alpha 7$ nicotinic acetylcholine receptor inhibits CD14 and the toll-like receptor 4 expression in human monocytes. *Shock*. (2006) 26:358–64. doi: 10.1097/01.shk.0000228168.86845.60
104. Yoshikawa H, Kurokawa M, Ozaki N, Nara K, Atou K, Takada E, et al. Nicotine inhibits the production of proinflammatory mediators in human monocytes by suppression of I- κ B phosphorylation and nuclear factor- κ B transcriptional activity through nicotinic acetylcholine receptor $\alpha 7$. *Clin Exp Immunol*. (2006) 146:116–23. doi: 10.1111/j.1365-2249.2006.03169.x
105. Razani-Boroujerdi S, Boyd RT, Dávila-García MI, Nandi JS, Mishra NC, Singh SP, et al. T cells express alpha7-nicotinic acetylcholine receptor subunits that require a functional TCR and leukocyte-specific protein tyrosine kinase for nicotine-induced Ca²⁺ response. *J Immunol*. (2007) 179:2889–98. doi: 10.4049/jimmunol.179.5.2889
106. De Rosa MJ, Dionisio L, Agriello E, Bouzat C, Esandi Mdel C. Alpha 7 nicotinic acetylcholine receptor modulates lymphocyte activation. *Life Sci*. (2009) 85:444–9. doi: 10.1016/j.lfs.2009.07.010
107. Skok MV, Kalashnik EN, Koval LN, Tsetlin VI, Utkin YN, Chingex J-P, et al. Functional nicotinic acetylcholine receptors are expressed in B lymphocyte-derived cell lines. *Mol Pharmacol*. (2003) 64:885–9. doi: 10.1124/mol.64.4.885
108. Goehler LE, Gaykema RP, Hansen MK, Anderson K, Maier SF, Watkins LR. Vagal immune-to-brain communication: a visceral chemosensory pathway. *Autonomic Neurosci*. (2000) 85:49–59. doi: 10.1016/S1566-0702(00)00219-8
109. Berthoud HR, Neuhuber WL. Functional and chemical anatomy of the afferent vagal system. *Auton Neurosci*. (2000) 85:1–17. doi: 10.1016/s1566-0702(00)00215-0
110. Grill HJ, Hayes MR. Hindbrain neurons as an essential hub in the neuroanatomically distributed control of energy balance. *Cell Metab*. (2012) 16:296–309. doi: 10.1016/j.cmet.2012.06.015
111. Hachem LD, Wong SM, Ibrahim GM. The vagus afferent network: emerging role in translational connectomics. *Neurosurg Focus*. (2018) 45:E2. doi: 10.3171/2018.6.Focus18216
112. Rosas-Ballina M, Olofsson PS, Ochani M, Valdés-Ferrer SI, Levine YA, Reardon C, et al. Acetylcholine-synthesizing T cells relay neural signals in a vagus nerve circuit. *Science*. (2011) 334:98–101. doi: 10.1126/science.1209985
113. Czura CJ, Friedman SG, Tracey KJ. Neural inhibition of inflammation: the cholinergic anti-inflammatory pathway. *J Endotoxin Res*. (2003) 9:409–13. doi: 10.1179/096805103225002755
114. Sharma G, Vijayaraghavan S. Nicotinic cholinergic signaling in hippocampal astrocytes involves calcium-induced calcium release from intracellular stores. *Proc Natl Acad Sci*. (2001) 98:4148–53. doi: 10.1073/pnas.071540198
115. Shytle RD, Mori T, Townsend K, Vendrame M, Sun N, Zeng J, et al. Cholinergic modulation of microglial activation by $\alpha 7$ nicotinic receptors. *J Neurochemistry*. (2004) 89:337–43. doi: 10.1046/j.1471-4159.2004.02347.x
116. Navarro E, Norden DM, Trojanowski PJ, Godbout JP, López MG. Central activation of Alpha7 nicotinic signaling attenuates LPS-induced neuroinflammation and sickness behavior in adult but not in aged animals. *Molecules*. (2021) 26. doi: 10.3390/molecules26082107
117. Revathikummar P, Bergqvist F, Gopalakrishnan S, Korotkova M, Jakobsson PJ, Lampa J, et al. Immunomodulatory effects of nicotine on interleukin 1 β activated human astrocytes and the role of cyclooxygenase 2 in the underlying mechanism. *J Neuroinflamm*. (2016) 13:256. doi: 10.1186/s12974-016-0725-1
118. Dumas JA, Newhouse PA. The cholinergic hypothesis of cognitive aging revisited again: cholinergic functional compensation. *Pharmacol Biochem Behav*. (2011) 99:254–61. doi: 10.1016/j.pbb.2011.02.022
119. Gamage R, Wagon I, Rossetti I, Childs R, Niedermayer G, Chesworth R, et al. Cholinergic modulation of glial function during aging and chronic neuroinflammation. *Front Cell Neurosci*. (2020) 14:577912. doi: 10.3389/fncel.2020.577912
120. Reale M, Di Bari M, Di Nicola M, D'Angelo C, De Angelis F, Velluto L, et al. Nicotinic receptor activation negatively modulates pro-inflammatory cytokine production in multiple sclerosis patients. *Int Immunopharmacol*. (2015) 29:152–7. doi: 10.1016/j.intimp.2015.06.034
121. Takata K, Amamiya T, Mizoguchi H, Kawanishi S, Kuroda E, Kitamura R, et al. Alpha7 nicotinic acetylcholine receptor-specific agonist DMXB A (GTS-21) attenuates A β accumulation through suppression of neuronal γ -secretase activity and promotion of microglial amyloid- β phagocytosis and ameliorates cognitive impairment in a mouse model of Alzheimer's disease. *Neurobiol Aging*. (2018) 62:197–209. doi: 10.1016/j.neurobiolaging.2017.10.021
122. Hua Y, Yang B, Chen Q, Zhang J, Hu J, Fan Y. Activation of $\alpha 7$ nicotinic acetylcholine receptor protects against 1-methyl-4-phenylpyridinium-induced astroglial apoptosis. *Front Cell Neurosci*. (2019) 13:507. doi: 10.3389/fncel.2019.00507
123. Marrero MB, Lucas R, Salet C, Hauser TA, Mazurov A, Lippello PM, et al. An $\alpha 7$ nicotinic acetylcholine receptor-selective agonist reduces weight gain and metabolic changes in a mouse model of diabetes. *J Pharmacol Exp Ther*. (2010) 332:173–80. doi: 10.1124/jpet.109.154633
124. Zhu X, Dai S, Xia B, Gong J, Ma B. Activation of the alpha 7 nicotinic acetylcholine receptor mitigates osteoarthritis progression by inhibiting NF- κ B/NLRP3 inflammasome activation and enhancing autophagy. *PLoS One*. (2021) 16:e0256507. doi: 10.1371/journal.pone.0256507
125. Pu W, Su Z, Wazir J, Zhao C, Wei L, Wang R, et al. Protective effect of $\alpha 7$ nicotinic acetylcholine receptor activation on experimental colitis and its mechanism. *Mol Med*. (2022) 28:104. doi: 10.1186/s10020-022-00532-2
126. Johnson GJ, Slater BC, Leis LA, Rector TS, Bach RR. Blood biomarkers of chronic inflammation in Gulf War Illness. *PLoS One*. (2016) 11:e0157855. doi: 10.1371/journal.pone.0157855
127. Farmer AD, Albu-Soda A, Aziz Q. Vagus nerve stimulation in clinical practice. *Br J Hosp Med (Lond)*. (2016) 77:645–51. doi: 10.12968/hmed.2016.77.11.645
128. Cook DB, Stegner AJ, Ellingson LD. Exercise alters pain sensitivity in Gulf War veterans with chronic musculoskeletal pain. *J Pain*. (2010) 11:764–72. doi: 10.1016/j.jpain.2009.11.010

129. Washington SD, Rayhan RU, Garner R, Provenzano D, Zajur K, Addiego FM, et al. Exercise alters cerebellar and cortical activity related to working memory in phenotypes of Gulf War Illness. *Brain Commun.* (2020) 2:fcz039. doi: 10.1093/braincomms/fcz039
130. Boruch AE, Lindheimer JB, Ninneman JV, Wylie GR, Alexander T, Klein-Adams JC, et al. Exercise-induced changes in gene expression do not mediate post exertional malaise in Gulf War illness. *Brain Behav Immun Health.* (2023) 29:100612. doi: 10.1016/j.bbih.2023.100612
131. Rayhan RU, Stevens BW, Raksit MP, Ripple JA, Timbol CR, Adewuyi O, et al. Exercise challenge in Gulf War Illness reveals two subgroups with altered brain structure and function. *PLoS One.* (2013) 8:e63903. doi: 10.1371/journal.pone.0063903
132. Baraniuk JN, Kern G, Narayan V, Cheema A. Exercise modifies glutamate and other metabolic biomarkers in cerebrospinal fluid from Gulf War Illness and Myalgic encephalomyelitis / Chronic Fatigue Syndrome. *PLoS One.* (2021) 16:e0244116. doi: 10.1371/journal.pone.0244116
133. Whistler T, Fletcher MA, Lonergan W, Zeng X-R, Lin J-M, LaPerriere A, et al. Impaired immune function in Gulf War illness. *BMC Med Genomics.* (2009) 2:1–13. doi: 10.1186/1755-8794-2-12
134. Bayley PJ, Schulz-Heik RJ, Cho R, Mathersul D, Collery L, Shankar K, et al. Yoga is effective in treating symptoms of Gulf War illness: A randomized clinical trial. *J Psychiatr Res.* (2021) 143:563–71. doi: 10.1016/j.jpsychires.2020.11.024
135. Allende S, Mathersul DC, Schulz-Heik JR, Avery TJ, Mahoney L, Bayley PJ. Yoga is effective for treating chronic pain in veterans with Gulf War Illness at long-term follow-up. *BMC Complement Med Ther.* (2023) 23:319. doi: 10.1186/s12906-023-04145-y
136. Ramirez-Sanchez I, Navarrete-Yañez V, Garate-Carrillo A, Lara-Hernandez M, Espinosa-Raya J, Moreno-Ulloa A, et al. Restorative potential of (-)-epicatechin in a rat model of Gulf War illness muscle atrophy and fatigue. *Sci Rep.* (2021) 11:21861. doi: 10.1038/s41598-021-01093-w
137. Shetty GA, Hattiangady B, Upadhy D, Bates A, Attaluri S, Shuai B, et al. Chronic oxidative stress, mitochondrial dysfunction, Nrf2 activation and inflammation in the hippocampus accompany heightened systemic inflammation and oxidative stress in an animal model of gulf war illness. *Front Mol Neurosci.* (2017) 10:182. doi: 10.3389/fnmol.2017.00182
138. Shetty AK, Attaluri S, Kodali M, Shuai B, Shetty GA, Upadhy D, et al. Monosodium lumnol reinstates redox homeostasis, improves cognition, mood and neurogenesis, and alleviates neuro-and systemic inflammation in a model of Gulf War Illness. *Redox Biol.* (2020) 28:101389. doi: 10.1016/j.redox.2019.101389
139. Joshi U, Evans JE, Joseph R, Emmerich T, Saltiel N, Lungmus C, et al. Oleoylethanolamide treatment reduces neurobehavioral deficits and brain pathology in a mouse model of Gulf War Illness. *Sci Rep.* (2018) 8:12921. doi: 10.1038/s41598-018-31242-7
140. Madhu LN, Attaluri S, Kodali M, Shuai B, Upadhy R, Gitai D, et al. Neuroinflammation in Gulf War Illness is linked with HMGB1 and complement activation, which can be discerned from brain-derived extracellular vesicles in the blood. *Brain Behav Immun.* (2019) 81:430–43. doi: 10.1016/j.bbi.2019.06.040
141. Parihar VK, Hattiangady B, Shuai B, Shetty AK. Mood and memory deficits in a model of Gulf War illness are linked with reduced neurogenesis, partial neuron loss, and mild inflammation in the hippocampus. *Neuropsychopharmacology.* (2013) 38:2348–62. doi: 10.1038/npp.2013.158
142. Zakirova Z, Tweed M, Crynen G, Reed J, Abdullah L, Nissanka N, et al. Gulf War agent exposure causes impairment of long-term memory formation and neuropathological changes in a mouse model of Gulf War Illness. *PLoS One.* (2015) 10:e0119579. doi: 10.1371/journal.pone.0119579
143. Kodali M, Hattiangady B, Shetty GA, Bates A, Shuai B, Shetty AK. Curcumin treatment leads to better cognitive and mood function in a model of Gulf War Illness with enhanced neurogenesis, and alleviation of inflammation and mitochondrial dysfunction in the hippocampus. *Brain Behav Immun.* (2018) 69:499–514. doi: 10.1016/j.bbi.2018.01.009
144. Murray KE, Ratliff WA, Delic V, Citron BA. Gulf War toxicant-induced reductions in dendritic arbors and spine densities of dentate granule cells are improved by treatment with a Nrf2 activator. *Brain Res.* (2024) 1823:148682. doi: 10.1016/j.brainres.2023.148682
145. Zakirova Z, Reed J, Crynen G, Horne L, Hassan S, Mathura V, et al. Complementary proteomic approaches reveal mitochondrial dysfunction, immune and inflammatory dysregulation in a mouse model of Gulf War Illness. *Proteomics Clin Appl.* (2017) 11. doi: 10.1002/prca.201600190
146. Lamproglou I, Barbier L, Diserbo M, Fauvelle F, Fauquette W, Amourette C. Repeated stress in combination with pyridostigmine: Part I: Long-term behavioural consequences. *Behav Brain Res.* (2009) 197:301–10. doi: 10.1016/j.bbr.2008.08.031
147. Abdullah L, Crynen G, Reed J, Bishop A, Phillips J, Ferguson S, et al. Proteomic CNS profile of delayed cognitive impairment in mice exposed to Gulf War agents. *Neuromolecular Med.* (2011) 13:275–88. doi: 10.1007/s12017-011-8160-z
148. Abdullah L, Evans JE, Bishop A, Reed JM, Crynen G, Phillips J, et al. Lipidomic profiling of phosphocholine-containing brain lipids in mice with sensorimotor deficits and anxiety-like features after exposure to Gulf War agents. *Neuromolecular Med.* (2012) 14:349–61. doi: 10.1007/s12017-012-8192-z
149. Hattiangady B, Mishra V, Kodali M, Shuai B, Rao X, Shetty AK. Object location and object recognition memory impairments, motivation deficits and depression in a model of Gulf War illness. *Front Behav Neurosci.* (2014) 8:78. doi: 10.3389/fnbeh.2014.00078
150. Zakirova Z, Crynen G, Hassan S, Abdullah L, Horne L, Mathura V, et al. A chronic longitudinal characterization of neurobehavioral and neuropathological cognitive impairment in a mouse model of gulf war agent exposure. *Front Integr Neurosci.* (2015) 9:71. doi: 10.3389/fnint.2015.00071
151. Abdullah L, Evans JE, Joshi U, Crynen G, Reed J, Mouzon B, et al. Translational potential of long-term decreases in mitochondrial lipids in a mouse model of Gulf War Illness. *Toxicology.* (2016) 372:22–33. doi: 10.1016/j.tox.2016.10.012
152. Phillips KF, Deshpande LS. Repeated low-dose organophosphate DFP exposure leads to the development of depression and cognitive impairment in a rat model of Gulf War Illness. *Neurotoxicology.* (2016) 52:127–33. doi: 10.1016/j.neuro.2015.11.014
153. Phillips KF, Deshpande LS. Chronic neurological morbidities and elevated hippocampal calcium levels in a DFP-based rat model of Gulf War Illness. *Mil Med.* (2018) 183:552–5. doi: 10.1093/milmed/usx148
154. Carreras I, Aytan N, Mellott T, Choi JK, Lehar M, Crabtree L, et al. Anxiety, neuroinflammation, cholinergic and GABAergic abnormalities are early markers of Gulf War illness in a mouse model of the disease. *Brain Res.* (2018) 1681:34–43. doi: 10.1016/j.brainres.2017.12.030
155. Madhu LN, Kodali M, Attaluri S, Shuai B, Melissari L, Rao X, et al. Melatonin improves brain function in a model of chronic Gulf War Illness with modulation of oxidative stress, NLRP3 inflammasomes, and BDNF-ERK-CREB pathway in the hippocampus. *Redox Biol.* (2021) 43:101973. doi: 10.1016/j.redox.2021.101973
156. Donovan EK, Kekes-Szabo S, Lin JC, Massey RL, Cobb JD, Hodgins KS, et al. A Placebo-Controlled, Pseudo-Randomized, Crossover Trial of Botanical Agents for Gulf War Illness: Curcumin (Curcuma longa), Boswellia (Boswellia serrata), and French Maritime Pine Bark (Pinus pinaster). *Int J Environ Res Public Health.* (2021) 18. doi: 10.3390/ijerph18052468
157. Baraniuk JN, El-Amin S, Corey R, Rayhan R, Timbol C. Carnosine treatment for gulf war illness: a randomized controlled trial. *Glob J Health Sci.* (2013) 5:69–81. doi: 10.5539/gjhs.v5n3p69
158. Venkatasamy L, Nizamutdinov D, Jenkins J, Shapiro LA. Vagus nerve stimulation ameliorates cognitive impairment and increased hippocampal astrocytes in a mouse model of gulf war illness. *Neurosci Insights.* (2021) 16:26331055211018456. doi: 10.1177/26331055211018456
159. Iannucci J, Nizamutdinov D, Shapiro LA. Neurogenesis and chronic neurobehavioral outcomes are partially improved by vagus nerve stimulation in a mouse model of Gulf War illness. *Neurotoxicology.* (2022) 90:205–15. doi: 10.1016/j.neuro.2022.04.001
160. Harrison FE, Hosseini AH, McDonald MP. Endogenous anxiety and stress responses in water maze and Barnes maze spatial memory tasks. *Behav Brain Res.* (2009) 198:247–51. doi: 10.1016/j.bbr.2008.10.015
161. Broderick G, Kreitz A, Fuite J, Fletcher MA, Vernon SD, Klimas N. A pilot study of immune network remodeling under challenge in Gulf War Illness. *Brain Behav Immun.* (2011) 25:302–13. doi: 10.1016/j.bbi.2010.10.011
162. Broderick G, Ben-Hamo R, Vashishtha S, Efroni S, Nathanson L, Barnes Z, et al. Altered immune pathway activity under exercise challenge in Gulf War Illness: an exploratory analysis. *Brain Behav Immun.* (2013) 28:159–69. doi: 10.1016/j.bbi.2012.11.007
163. Rogers JL, Kesner RP. Cholinergic modulation of the hippocampus during encoding and retrieval. *Neurobiol Learn Mem.* (2003) 80:332–42. doi: 10.1016/s1074-7427(03)00063-7
164. Bellinger FP, Madamba S, Siggins GR. Interleukin 1 beta inhibits synaptic strength and long-term potentiation in the rat CA1 hippocampus. *Brain Res.* (1993) 628:227–34. doi: 10.1016/0006-8993(93)90959-Q
165. Gately MK, Renzetti LM, Magram J, Stern AS, Adorini L, Gubler U, et al. The interleukin-12/interleukin-12-receptor system: role in normal and pathologic immune responses. *Annu Rev Immunol.* (1998) 16:495–521. doi: 10.1146/annurev.immunol.16.1.495
166. Barrientos RM, Frank MG, Hein AM, Higgins EA, Watkins LR, Rudy JW, et al. Time course of hippocampal IL-1 β and memory consolidation impairments in aging rats following peripheral infection. *Brain Behavior Immun.* (2009) 23:46–54. doi: 10.1016/j.bbi.2008.07.002
167. Godbout JP, Chen J, Abraham J, Richwine AF, Berg BM, Kelley KW, et al. Exaggerated neuroinflammation and sickness behavior in aged mice following activation of the peripheral innate immune system. *FASEB J.* (2005) 19:1329–31. doi: 10.1096/fj.05-3776fj
168. Norden DM, Muccigrosso MM, Godbout JP. Microglial priming and enhanced reactivity to secondary insult in aging, and traumatic CNS injury, and neurodegenerative disease. *Neuropharmacology.* (2015) 96:29–41. doi: 10.1016/j.neuropharm.2014.10.028
169. Macht VA, Reagan LP. Chronic stress from adolescence to aging in the prefrontal cortex: A neuroimmune perspective. *Front Neuroendocrinol.* (2018) 49:31–42. doi: 10.1016/j.yfrne.2017.12.001
170. Chang Q, Gold PE. Age-related changes in memory and in acetylcholine functions in the hippocampus in the Ts65Dn mouse, a model of Down syndrome. *Neurobiol Learn Mem.* (2008) 89:167–77. doi: 10.1016/j.nlm.2007.05.007
171. Petersen RC, Lopez O, Armstrong MJ, Getchius TSD, Ganguli M, Gloss D, et al. Practice guideline update summary: Mild cognitive impairment. *Neurology.* (2018) 90:126–35. doi: 10.1212/WNL.0000000000004826

172. Brown KA, Preston CJ, Carpenter JM, Ludwig HD, Norberg T, Harn DA, et al. *Lacto-N-fucopentaose-III (LNFPIII) ameliorates acute aberrations in hippocampal synaptic transmission in a Gulf War Illness animal model. Brain Res.* (2021) 1766:147513. doi: 10.1016/j.brainres.2021.147513
173. Holton KF, Ramachandra SS, Murray SL, Baron M, Baraniuk JN. Effect of the low glutamate diet on inflammatory cytokines in veterans with Gulf War Illness (GWI): A pilot study. *Life Sci.* (2021) 280:119637. doi: 10.1016/j.lfs.2021.119637
174. Holton KF, Kirkland AE, Baron M, Ramachandra SS, Langan MT, Brandley ET, et al. The Low Glutamate Diet Effectively Improves Pain and Other Symptoms of Gulf War Illness. *Nutrients.* (2020) 12(9):2593. doi: 10.3390/nu12092593
175. Carpenter JM, Gordon HE, Ludwig HD, Wagner JJ, Harn DA, Norberg T, et al. Neurochemical and neuroinflammatory perturbations in two Gulf War Illness models: Modulation by the immunotherapeutic LNFPIII. *Neurotoxicology.* (2020) 77:40–50. doi: 10.1016/j.neuro.2019.12.012
176. Carpenter JM, Brown KA, Diaz AN, Dockman RL, Benbow RA, Harn DA, et al. Delayed treatment with the immunotherapeutic LNFPIII ameliorates multiple neurological deficits in a pesticide-nerve agent prophylactic mouse model of Gulf War Illness. *Neurotoxicol Teratol.* (2021) 87:107012. doi: 10.1016/j.ntt.2021.107012
177. Natelson BH, Stegner AJ, Lange G, Khan S, Bate M, Sotolongo A, et al. Vagal nerve stimulation as a possible non-invasive treatment for chronic widespread pain in Gulf Veterans with Gulf War Illness. *Life Sci.* (2021) 282:119805. doi: 10.1016/j.lfs.2021.119805
178. Lange G, Janal MN, Maniker A, FitzGibbons J, Fobler M, Cook D, et al. Safety and efficacy of vagus nerve stimulation in fibromyalgia: A phase I/II proof of concept trial. *Pain Med.* (2011) 12:1406–13. doi: 10.1111/j.1526-4637.2011.01203.x
179. Oshinsky ML, Murphy AL, Hekierski H Jr., Cooper M, Simon BJ. Noninvasive vagus nerve stimulation as treatment for trigeminal allodynia. *Pain.* (2014) 155(5):1037–42. doi: 10.1016/j.pain.2014.02.009
180. Straube A, Ellrich J, Eren O, Blum B, Ruscheweyh R. Treatment of chronic migraine with transcutaneous stimulation of the auricular branch of the vagal nerve (auricular t-VNS): a randomized, monocentric clinical trial. *J Headache Pain.* (2015) 16:63. doi: 10.1186/s10194-015-0543-3
181. Ness TJ, Fillingim RB, Randich A, Backensto EM, Faught E. Low intensity vagal nerve stimulation lowers human thermal pain thresholds. *Pain.* (2000) 86:81–5. doi: 10.1016/S0304-3959(00)00237-2
182. Nizamutdinov D, Mukherjee S, Deng C, Stauss HM, Shapiro IA. Gulf War agents pyridostigmine bromide and permethrin cause hypersensitive nociception that is restored after vagus nerve stimulation. *Neurotoxicology.* (2018) 69:93–6. doi: 10.1016/j.neuro.2018.09.007
183. Yang T, Xiao T, Sun Q, Wang K. The current agonists and positive allosteric modulators of $\alpha 7$ nAChR for CNS indications in clinical trials. *Acta Pharm Sin B.* (2017) 7:611–22. doi: 10.1016/j.apsb.2017.09.001
184. Tregellas JR, Wylie KP. Alpha7 nicotinic receptors as therapeutic targets in Schizophrenia. *Nicotine Tob Res.* (2019) 21:349–56. doi: 10.1093/ntr/nty034
185. Rueda Ruzafa L, Cedillo JL, Hone AJ. Nicotinic acetylcholine receptor involvement in inflammatory bowel disease and interactions with gut microbiota. *Int J Environ Res Public Health.* (2021) 18. doi: 10.3390/ijerph18031189
186. Abdel-Magid A. Allosteric modulators: an emerging concept in drug discovery. *ACS medicinal Chem Lett.* (2015) 6:104–7. doi: 10.1021/ml5005365
187. Williams DK, Wang J, Papke RL. Investigation of the molecular mechanism of the $\alpha 7$ nicotinic acetylcholine receptor positive allosteric modulator PNU-120596 provides evidence for two distinct desensitized states. *Mol Pharmacol.* (2011) 80:1013–32. doi: 10.1124/mol.111.074302
188. Christensen DZ, Mikkelsen JD, Hansen HH, Thomsen MS. Repeated administration of alpha7 nicotinic acetylcholine receptor (nAChR) agonists, but not positive allosteric modulators, increases alpha7 nAChR levels in the brain. *J Neurochem.* (2010) 114:1205–16. doi: 10.1111/j.1471-4159.2010.06845.x
189. Alzarea S, Rahman S. Effects of alpha-7 nicotinic allosteric modulator PNU 120596 on depressive-like behavior after lipopolysaccharide administration in mice. *Prog Neuropsychopharmacol Biol Psychiatry.* (2018) 86:218–28. doi: 10.1016/j.pnpbp.2018.05.018
190. Callahan PM, Hutchings EJ, Kille NJ, Chapman JM, Terry AV Jr. Positive allosteric modulator of $\alpha 7$ nicotinic-acetylcholine receptors, PNU-120596 augments the effects of donepezil on learning and memory in aged rodents and non-human primates. *Neuropharmacology.* (2013) 67:201–12. doi: 10.1016/j.neuropharm.2012.10.019
191. Pandya AA, Yakel JL. Effects of neuronal nicotinic acetylcholine receptor allosteric modulators in animal behavior studies. *Biochem Pharmacol.* (2013) 86:1054–62. doi: 10.1016/j.bcp.2013.05.018
192. Phenis D, Vunck SA, Valentini V, Arias H, Schwarcz R, Bruno JP. Activation of alpha7 nicotinic and NMDA receptors is necessary for performance in a working memory task. *Psychopharmacol (Berl).* (2020) 237:1723–35. doi: 10.1007/s00213-020-05495-y
193. Potasiewicz A, Krawczyk M, Gzielo K, Popik P, Nikiforuk A. Positive allosteric modulators of alpha 7 nicotinic acetylcholine receptors enhance procognitive effects of conventional anti-Alzheimer drugs in scopolamine-treated rats. *Behav Brain Res.* (2020) 385:112547. doi: 10.1016/j.bbr.2020.112547