Check for updates

OPEN ACCESS

EDITED BY Li-Shu Wang, Medical College of Wisconsin, United States

REVIEWED BY Johnna Francis Varghese,

Brigham and Women's Hospital and Harvard Medical School, United States Denis Gris, Université de Sherbrooke, Canada Junji Xing, Houston Methodist Research Institute, United States

*CORRESPONDENCE Tünde Fekete fekete.tunde@med.unideb.hu

RECEIVED 14 March 2024 ACCEPTED 20 June 2024 PUBLISHED 28 June 2024

CITATION

Pázmándi K, Szöllősi AG and Fekete T (2024) The "root" causes behind the antiinflammatory actions of ginger compounds in immune cells. *Front. Immunol.* 15:1400956. doi: 10.3389/fimmu.2024.1400956

COPYRIGHT

© 2024 Pázmándi, Szöllősi and Fekete. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

The "root" causes behind the anti-inflammatory actions of ginger compounds in immune cells

Kitti Pázmándi, Attila Gábor Szöllősi and Tünde Fekete*

Department of Immunology, Faculty of Medicine, University of Debrecen, Debrecen, Hungary

Ginger (Zingiber officinale) is one of the most well-known spices and medicinal plants worldwide that has been used since ancient times to treat a plethora of diseases including cold, gastrointestinal complaints, nausea, and migraine. Beyond that, a growing body of literature demonstrates that ginger exhibits anti-inflammatory, antioxidant, anti-cancer and neuroprotective actions as well. The beneficial effects of ginger can be attributed to the biologically active compounds of its rhizome such as gingerols, shogaols, zingerone and paradols. Among these compounds, gingerols are the most abundant in fresh roots, and shogaols are the major phenolic compounds of dried ginger. Over the last two decades numerous in vitro and in vivo studies demonstrated that the major ginger phenolics are able to influence the function of various immune cells including macrophages, neutrophils, dendritic cells and T cells. Although the mechanism of action of these compounds is not fully elucidated yet, some studies provide a mechanistic insight into their anti-inflammatory effects by showing that ginger constituents are able to target multiple signaling pathways. In the first part of this review, we summarized the current literature about the immunomodulatory actions of the major ginger compounds, and in the second part, we focused on the possible molecular mechanisms that may underlie their anti-inflammatory effects.

KEYWORDS

ginger, gingerol, shogaol, immune cell, cytokine, signaling

1 Introduction

The rhizome of ginger (*Zingiber officinale*) has been used as a dietary food supplement across China and Southeast Asia since ancient times. Ginger was first documented as an herbal medicine around 3000 BC in China (1). It was primarily recommended as a remedy for cold, fever, leprosy and tetanus (1). Other medicinal uses of ginger include the treatment of nausea, upset stomach and it's use as a digestive aid (2). The U.S. Food and Drug Administration (FDA) classified ginger root as a safe herbal supplement that can be used in

complementary and alternative medicine preparations (3). Due to its proven beneficial effects, in 2012 the European Medicines Agency listed the ginger rhizome as an approved treatment modality in the prevention of nausea and vomiting associated with motion sickness (4). In the last few years a myriad of studies indicated that ginger possesses various biological activities such as anti-inflammatory, antioxidant, anti-microbial, anti-cancer and neuroprotective effects. It has also been revealed that the pharmacological benefits of ginger can be credited to the bioactive compounds of its rhizome.

To date, more than 400 chemical compounds such as lipids, terpenes, carbohydrates and phenolic compounds have been identified in the ginger rhizome (5, 6). The phenolic compounds or phenolics, which are usually referred to as the nonvolatile components of ginger, are mainly responsible for its pharmacological activities and consist of gingerols, shogaols, paradols and zingerone (7). Gingerols are the main pungent components of fresh ginger that can be differentiated based on their unbranched alkyl side chain length. Among them, 6gingerol is the most abundant constituent that is followed by 8-gingerol and 10-gingerol (8). Upon drying or heating of ginger root the thermally labile gingerols undergo dehydration reactions to form the corresponding shogaols, which are twice as pungent as gingerols (9). Although shogaols are scarcely found in fresh ginger root, 6-shogaol is the predominant bioactive compound in the dried rhizome. At high temperatures or by microbial metabolism shogaols might be partly transformed to paradols, which represent a minor but important bioactive constituent of ginger (10). Cooking or drying can also convert gingerol into zingerone through a retro-aldol reaction (11). In addition, a number of other gingerol derivatives such as gingerdion, gingerdiol, dehydrogingerdione and other minor components have been isolated from ginger rhizome (12). A recent study using synthetic strategy to prepare 6-gingerol derivatives indicates that several of those show promising anti-platelet and antioxidant activities; however, their biological effects needs to be further elucidated (13). The major bioactive compounds of ginger and the conversion of 6-gingerol into 6-shogaol, paradol and zingerone are shown in Figure 1.

In the last two decades, especially since the outbreak of coronavirus disease 2019 (Covid-19) pandemic, herbal plants have gained special interest as an alternative or complementary medicine due to their beneficial medicinal properties and minimal adverse effects (14, 15). Ginger is one of the most researched medicinal plants due to its long-known health benefits and high bioactive agent content. In the last few years, a number of studies have proven the pharmacological potential of ginger-derived phytochemicals; however, the cellular and molecular mechanisms behind their activities are not fully revealed yet. It was shown that ginger phytochemicals might exert their anti-inflammatory effects by modulating the function of various immune cells such as macrophages, neutrophils, T cells and dendritic cells (DC) (5). It was also demonstrated that ginger constituents have the potential to regulate key signaling pathways such as the nuclear factor-kappa B (NF-KB), mitogen activated protein kinase (MAPK) and phosphatidylinositol-3-kinase (PI3K)/Akt/mammalian target of rapamycin (mTOR) signaling cascades in these cell types. Some studies also provided a deeper insight into the mechanism of action of ginger constituents by showing that those are able to target several signaling molecules such as the AMP-activated protein kinase (AMPK), nuclear factor erythroid 2-related factor 2 (NRF2), heme oxygenase-1 (HO-1), and peroxisome proliferator-activated receptor gamma (PPARγ).

Although ginger rhizome is abundant in bioactive compounds, the in vitro and in vivo studies mainly focus on gingerols and shogaols, since those possess the most significant pharmacological effects. Some reports also suggest that shogaols exhibit more potent biological activities then gingerols (16). In particular, the loss of the hydroxyl group in their sidechains might increase their lipophilicity and thus shogaols show a superior bioavailability compared to gingerols (9). Reviews published in the last few years focused on the general biological effects (5), anti-cancer (17, 18) or neuroprotective (19) activities of ginger constituents. Some reports also addressed the anti-inflammatory actions of ginger extract, or one or the other components in general (20, 21). Nevertheless, the current literature lacks a comprehensive review on the immunomodulatory effects of ginger phytochemicals at the cellular and molecular level. Most importantly, an update is needed to discuss the scientific merit of the most recently published research articles. Therefore, in this recent review we aimed to collate all the available evidence on the effects of the major ginger constituents on different immune cell types and to highlight the latest advances in our understanding of their mechanisms of anti-inflammatory actions.

2 Effects of ginger compounds on the cells of the immune system

Ginger has been used for centuries for its anti-emetic effect. Recent clinical trials demonstrated that ginger supplementation can be used as an adjuvant therapy for managing and preventing chemotherapy-induced nausea and vomiting as well (22, 23). Additionally, some clinical studies also suggested that ginger can effectively ameliorate chronic pain in conditions such as arthritis (24, 25). Interestingly, the pain-relieving properties of ginger constituents are similar to that exerted by the non-steroidal anti-



FIGURE 1

Conversion of 6-gingerol into 6-shogaol, paradol and zingerone. Upon heating or drying 6-gingerol is mostly converted to 6-shogaol through dehydration and to a lesser extent to zingerone through a retro-aldol reaction. Moreover, 6-shogaol is transformed to 6paradol by microbial metabolism. inflammatory drugs such as ibuprofen (26, 27). Research data also indicates that the pain reducing effect of ginger is linked to its high anti-inflammatory capacity [reviewed in (28)]. Indeed, a number of *in vitro* and *in vivo* studies demonstrated that the bioactive compounds of ginger are able to dampen the inflammatory responses of various immune cells. Most of these studies focused on the innate immune machinery; however, there are a few reports available on the effects of the major ginger compounds on T cells as well. This chapter summarizes those studies that investigated the *in vitro* effects of ginger phenolics on different immune cell types.

2.1 Effects of ginger phenolics on macrophages

Macrophages serve as the first line of defense against invading pathogens and represent a highly plastic cell population with divergent phenotypes and functions. Generally, macrophages play an essential role in clearing out microbial pathogens by producing antimicrobial molecules such as reactive oxygen species (ROS) and nitric oxide (NO). In response to pathogenic stimuli macrophages also secrete various types of cytokines and chemokines and thus contribute to the initiation of inflammatory responses. Owing to their high functional plasticity, macrophages are also involved in the resolution of inflammation and restoration of homeostasis (29). Nevertheless, abnormal activation and polarization of macrophages has been suggested to contribute to the pathogenesis of different autoimmune diseases (30).

The very first study investigating the effects of ginger components on immune cells was published in 2003. Ippoushi et al. demonstrated that 6-gingerol decreased the production of NO and the protein levels of inducible NO synthase (iNOS) in lipopolysaccharide (LPS)stimulated J774.1 murine macrophages (31). 6-gingerol also protected against DNA and protein damage by suppressing the peroxynitrite-induced single strand breaks in supercoiled plasmid DNA as well as the formation of nitrotyrosine in these cells (31). In mouse peritoneal macrophages, 6-gingerol decreased the LPStriggered production of the inflammatory cytokines interleukin 1 beta (IL-1 β), tumor necrosis factor (TNF), IL-12 and the chemokine CCL5 (RANTES), whereas it had no effect on the upregulation of major histocompatibility complex (MHC) class II, and the costimulatory molecules CD80 (B7.1) and CD86 (B7.2) (32). The authors also gave some mechanistic insight by demonstrating that 6-gingerol was able to suppress the LPS-induced activation of NF-κB (32). In the human monocytic U937 cell line, 6-, 8-, 10-gingerols efficiently reduced the mRNA level of cyclooxygenase-2 (COX-2), a key enzyme responsible for prostaglandin E2 (PGE₂) production, and thus were more potent inhibitors of PGE₂ production compared to 6shogaol (33). On the contrary, 6-shogaol more effectively decreased the level of PGE₂, COX-2 and iNOS than 6-gingerol in LPSstimulated murine RAW 264.7 cells (34). The authors further demonstrated that 6-shogaol blocked the LPS-mediated phosphorylation and degradation of inhibitor kBa (IKBa) that suppressed the subsequent phosphorylation, nuclear translocation and transcriptional activity of NF-KB p65 in RAW 264.7 cells. Furthermore, 6-shogaol reduced the activation of PI3K and Akt and interfered with the MAPK signaling pathway by attenuating the phosphorylation of extracellular-regulated kinase 1/2 (ERK1/2) but not that of p38 in LPS-stimulated RAW 264.7 macrophages (34). Similarly, another study demonstrated that 6-gingerol inhibited iNOS and TNF production through the suppression of I κ B α phosphorylation and NF- κ B activation (35). It was also shown that 6-gingerol inhibits the LPS-triggered intracellular Ca²⁺ mobilization and ROS generation, most likely by blocking the cytosol-tomembrane translocation of protein kinase C- α (PKC- α) (35).

In LPS-stimulated primary microglia cells, 6-shogaol significantly decreased NO levels by reducing iNOS activity, suppressed PGE₂ production by downregulating COX-2 expression, and reduced IL-1 β and TNF secretion by inhibiting IkB α phosphorylation and degradation, and thereby NF-kB activation (36). Nevertheless, in contrast to the results obtained by RAW264.7 macrophages (30), 6-shogaol inhibited the LPSstimulated activation of p38 and JNK but not that of ERK1/2 in primary microglia cells (32). A few years later similar results were published showing that 6-shogaol suppressed LPS-induced IL-1β, IL-6, TNF and PGE₂ production by inhibiting the phosphorylation and nuclear translocation of NF-KB in BV2 cells (37). As a new concept the authors suggested that 6-shogaol blocks the LPSinduced inflammatory mediator production through activating PPARy, which is a known inhibitor of NF- κ B activation (33). Similar to 6-shogaol, 6-gingerol also effectively inhibited the LPSstimulated expression of iNOS and production of NO, IL-1β and IL-6 in primary mouse microglia (38). Furthermore, 6-gingerol was able to suppress the phosphorylation of Akt, mTOR and signal transducer and activator of transcription 3 (STAT3), the latter of which is a crucial signaling intermediate for TLR4-induced inflammatory responses in macrophages (39).

In primary mouse calvarial osteoblasts, 6-shogaol also inhibited the IL-1-induced expression of PGE_2 by suppressing the enzymatic activity of COX-2 and PGE synthase that resulted in decreased receptor activator of NF- κ B (RANKL) production and thus reduction of osteoclast differentiation (40).

Finally, two reports investigated the effect of gingerols and shogaols on the nucleotide-binding oligomerization domain-like receptor family pyrin domain containing 3 (NLRP3) inflammasome-mediated responses. The first study compared the inhibitory effects of 6-, 8-, 10-gingerols and 6-, 8-, 10- shogaols in human THP-1 macrophages stimulated with LPS and adenosine 5'-triphosphate (ATP) (41). It was demonstrated that 10-gingerol and all the shogaols effectively reduced the secretion of TNF and IL-1 β as well as the protein levels of NLRP3 and caspase-1. In general, shogaols show a higher anti-inflammatory capacity than the corresponding gingerols that could be attributed to the α , β -unsaturated carbonyl group in the structure of shogaols. Among the investigated phytochemicals 6-shogaol was the most potent inhibitor of NLRP3-mediated inflammasome activation. Interestingly, the results also indicate that the increase in alkyl side chain length weakens the anti-inflammatory potential of shogaols, while enhancing that of gingerols (41). The other study was conducted on RAW 264.7 cells and mouse bone marrow-derived macrophages (BM-DMs), in which 6-gingerol greatly reduced the release of caspase-1 p20 as well as the production of IL-1 β and IL-18 in response to ATP and LPS (42). 6gingerol significantly suppressed the phosphorylation of p38, JNK and ERK1/2 in RAW 264.7 cells, while it inhibited only ERK1/2 activation in BM-DMs. Treatment of RAW 264.7 cells with an ERK agonist reversed the inhibitory effects of 6-gingerol on caspase-1 p20 release suggesting that 6-gingerol exerts its effect through blocking MAPK activation (42).

Recent *in vivo* studies further suggest that 6-gingerol might inhibit NLPR3 inflammasome activation by reducing oxidative stress. In the lung tissues of LPS-subjected rats, 6-gingerol repressed the levels of oxidative stress markers and the expression of NLRP3 inflammasome (43). 6-gingerol also decreased the arsenic-trioxide induced production of ROS and the expression of NLRP3 inflammasome components in the liver tissues of mice (44). These current findings indicate that 6-gingerol might exert its anti-inflammatory effects by inhibiting the ROS-NLRP3 inflammasome pathway.

Table 1 summarizes the effects of the major ginger compounds on macrophages, whereas Figure 2A shows the signaling pathways,

TABLE 1 Effects of the major ginger phenolics on macrophages.

which are affected by the regulatory actions of major ginger phenolics in these cells.

2.2 Effects of the major ginger phenolics on neutrophil granulocytes

As phagocytic cells of the innate immune system, neutrophil granulocytes take an important part in clearing infectious agents and cellular debris from the human body. In addition to their phagocytic capacity, activated neutrophils extrude neutrophil extracellular traps (NETs), composed of DNA, histones, and antimicrobial enzymes, such as neutrophil elastase (NE) and myeloperoxidase (MPO), in which pathogens are immobilized and exposed to a lethal dose of effector proteins (45). A large body of evidence also indicates that through NET release

Macrophages							
Compounds	Cell types	Challenges	Observed effects	Potential mechanisms	Ref.		
6-gingerol	J774.1 murine macrophage cell line	LPS	 reduced NO, iNOS levels protection against DNA and protein damage 	-	(31)		
6-gingerol	mouse peritoneal macrophage	LPS	• decreased IL-1β, IL-12, TNF, CCL5	• suppressed NF-κB activity	(32)		
6-, 8-, 10-gingerols and 6-shogaol	human monocytic U937 cell line	LPS	 reduced PGE₂ production gingerols reduced COX- 2 expression 	-	(33)		
6-shogaol	RAW 264.7 murine macrophages cell line	LPS	 reduced PGE₂ production reduced COX-2, iNOS levels 	 blocked ΙκΒα phosphorylation and degradation suppressed NF-κB activity inhibited PI3K/Akt and ERK1/2 activation 	(34)		
6-gingerol	RAW 264.7 murine macrophages cell line	LPS	 inhibited iNOS and TNF expression reduced intracellular Ca²⁺ mobilization decreased ROS generation 	- suppressed IKB phosphorylation, NF-KB nuclear translocation and PKC- α translocation	(35)		
6-shogaol	primary microglia	LPS	 reduced IL-1β, TNF and PGE₂ production downregulated COX- 2 expression 	 blocked ΙκΒα phosphorylation and nuclear translocation decreased NF-κB activity inhibited p38 and JNK activation 	(36)		
6-shogaol	BV2 murine microglial cell line	LPS	\bullet suppressed IL-1 β , IL-6, TNF and PGE ₂ production	 reduced NF-κB phosphorylation and nuclear translocation increased PPARγ activity 	(37)		
6-gingerol	primary mouse microglia cell	LPS	 reduced NO, iNOS levels reduced IL-1β and IL- production 	• reduced Akt, mTOR and STAT3 phosphorylation	(38)		
6-shogaol	primary mouse calvarial osteoblast	IL-1α	 reduced PGE₂ expression decreased RANKL production and osteoclast differentiation 	- suppressed COX2 and \mbox{PGE}_2 synthase activity	(40)		
6-, 8-, 10- gingerols/shogaols	THP-1 human monocytic cell line	LPS+ATP	 reduced TNF and IL-1β secretion reduced NLRP3 and caspase- 1 levels 	-	(41)		
6-gingerol	RAW 264.7 macrophage cell line and mouse BM-DM	LPS+ATP	 reduced IL-1β and IL-18 secretion reduced caspase-1 p20 release 	• suppressed MAPK activation	(42)		



neutrophils are implicated in the pathogenesis of autoimmune diseases such as lupus (46).

It was first published in 2010 that 6-,8-,10-gingerols and 6shogaol are able to effectively inhibit ROS generation by human neutrophils in response to formylmethionine-leucyl-phenylalanine (fMLP), a strong inducer of neutrophil activation (16). Among the tested ginger phenolics, 6-shogaol showed the highest potential to suppress fMLP-induced ROS release, while 6-gingerol had the weakest inhibitory capacity (16). More than 10 years later, a study from 2021 also investigated the anti-inflammatory properties of gingerols in neutrophils. Ali et al. demonstrated that 6-, 8-, and 10gingerols suppressed NET-associated MPO levels, thereby netosis of human neutrophils in response to LPS, phorbol 12-myristate 13acetate (PMA) and various lupus-relevant stimuli as well (47). All three gingerols suppressed the formation of hydrogen-peroxide (H_2O_2) in neutrophils indicating that gingerols attenuate netosis by inhibiting the generation of ROS. Further, 6-gingerol also decreased the activity of phosphodiesterase 4 (PDE4), which by regulating cAMP levels plays a prominent anti-inflammatory effect in basically all cells involved in inflammation (48). Concomitantly, 6-gingerol increased the intracellular levels of cAMP and thus that of cAMP-dependent protein kinase A (PKA), which exerts anti-inflammatory activity by promoting the production of anti-inflammatory cytokines and by reducing the transcriptional activity of NF- κ B (48). Mechanistically, the authors proposed that the antineutrophil effects of 6-gingerol depend, at least partially, on its ability to inhibit PDE4 activity (47).

A year later it was published that zingerone, a less-studied component of ginger, also displays antineutrophil potential (49). In PMA-stimulated mouse bone marrow-derived neutrophils, zingerone significantly suppressed ROS production and formation of NET, as indicated by the decreased level of DNA-associated MPO. Interestingly, the bactericidal activity of neutrophils was not affected by zingerone treatment either *in vitro* or *in vivo*. In particular, zingerone did not alter the phagocytic capacity of neutrophils *in vitro* and had no effect on bacteria dissemination *in vivo*. Nevertheless, zingerone treatment significantly increased the levels of the nuclear NRF2 and HO-1 in PMA-stimulated neutrophils. Further it was shown that a specific inhibitor of NRF2 could efficiently reverse the protective effect of zingerone in septic mice (49).

In contrast with the previous findings, a recent study reported that 6-gingerol increased fMLP-stimulated CXCL8 chemokine secretion and ROS production in primary human neutrophils (50). Furthermore, 6-gingerol also increased the expression of neutrophil surface markers such as CD11b and CD66, as well as the expression of formyl peptide receptor 1 (FPR1), which may lead to increased responsiveness to its ligand, fMLP. In this particular study, the authors used a very low concentration of 6-gingerol (50 nM), which is generally a hundred- and thousand-fold lower compared to the doses used in previous publications on neutrophils and other immune cell types. The report also suggested that 6-gingerol applied in a low dose facilitates neutrophil functions through binding to the transient receptor potential cation channel subfamily V member 1 (TRPV1), the potential of which as a mechanism of action we discuss in chapter 4.4 in detail.

The effects of the major ginger phenolics on neutrophil granulocytes are included in Table 2, whereas their antineutrophil actions are shown in Figure 2B.

2.3 Effects of 6-gingerol and 6-shogaol on DCs

DCs represent a heterogeneous population, which acting as the most potent professional antigen presenting cells (APCs) bridge the innate and adaptive immune systems. Expressing a wide repertoire of innate receptors, DCs can recognize a broad range of pathogen-, and danger-associated molecular patterns (PAMPs and DAMPs), and then migrate to secondary lymphoid organs to present antigens to T cells. Activated DCs upregulate costimulatory molecules and produce polarizing cytokines to drive T cell differentiation, and initiate different types of antigen-specific adaptive immune responses. In the steady-state, in the absence of activation signals, antigen presentation by DCs leads to T cell unresponsiveness and tolerance (51). In addition to their well-known functions of controlling innate and adaptive immunity, accumulating evidence indicates that DCs are implicated in the pathogenesis and pathomechanism of various autoimmune disorders (52).

Although DCs play a central role in the coordination of immune responses, only two studies investigated how gingerderived phytochemicals might affect their functionality. Han et al. studied the effects of 6-gingerol on mouse BM-DCs, and found that 6-gingerol was able to remarkably reduce the production of TNF, IL-1 β , IL-6 and IL-23 as well as the expression of CD80, CD86 and MHC II in response to LPS exposure (53). 6-gingerol treated BM-DCs also had an inferior capacity to prime T helper 17 (Th17) cell polarization upon co-culture with CD4+ naïve T cells. Further, it was demonstrated that 6-gingerol prevented the LPS-induced phosphorylation of NF- κ B, JNK and ERK1/2 suggesting that 6-gingerol suppressed the activation of BM-DCs by interfering with the NF- κ B and MAPK signaling cascades (53).

Our research group has recently reported that 6-gingerol and 6shogaol are also able to modulate the phenotypical and functional properties of human DCs (54). The studies so far have investigated the effect of 6-gingerol or 6-shogaol on the TLR4-mediated response of immune cells. In our experiments, monocyte-derived DCs (moDCs) were first treated with 6-gingerol and 6-shogaol then were exposed to various TLR ligands including agonists of TLR4, TLR2/1 and TLR7/8. We found that 6-gingerol and 6-shogaol could significantly decrease the TLR-triggered expression of CD40, CD83, CD86 and HLA-DQ in moDCs. Furthermore, 6-gingerol and 6shogaol significantly reduced the secretion of TNF, IL-6 and IL-10 by TLR-stimulated moDCs. Similarly, both compounds significantly reduced the Escherichia coli-triggered secretion of inflammatory cytokines, and thus their capacity to promote a Th1 phenotype in naïve CD4+ T cells (54). Investigating the mechanism underlying the actions of these compounds, we found that both 6gingerol and 6-shogaol could significantly decrease the TLRinduced nuclear translocation of NF-KB p65, while not affecting IkB α degradation. Both compounds were also able to reduce the

TABLE 2	Effects of	the major	ginger	phenolics	on neutrophil	granulocytes.
---------	------------	-----------	--------	-----------	---------------	---------------

Neutrophil granulocytes								
Compounds	Cell types	Challenges	Observed effects	Potential mechanisms	Ref.			
6-,8-,10-gingerols and 6-shogaol	human neutrophil	fMLP	• suppressed ROS release	-	(16)			
6-, 8-, and 10-gingerol	human neutrophil	LPS, PMA	suppressed netosisreduced ROS generation	decreased PDE4 activityincreased cAMP levels and PKA	(47)			
zingerone	mouse BM- derived neutrophil	РМА	suppressed ROS productionreduced netosis	• activated the NRF2 signaling pathway	(49)			
6-gingerol	human neutrophil	fMLP	 increased ROS and CXCL8 production increased CD11b, CD66, FPR1 expression 	binding to TRPV1	(50)			

TLR-triggered activation of the MAPK pathway. Further, we demonstrated that 6-gingerol decreased mTOR complex 2 (mTORC2)-mediated Akt phosphorylation, whereas it had no effect on mTORC1-mediated p7086K phosphorylation in TLR-stimulated moDCs. In contrast, 6-shogaol could greatly suppress the TLR-mediated activation of both mTOR complexes. Further, we found that 6-shogaol but not 6-gingerol could significantly increase the activity of AMPK, the nuclear translocation of NRF2 and the expression of HO-1. Altogether, our data imply that both ginger-derived compounds attenuate human DC functionality via suppressing the NF- κ B, MAPK and mTOR signaling pathways. Further, we demonstrated that compared to 6-gingerol, 6-shogaol possesses a stronger inhibitory activity on moDCs that might be attributed to its ability to activate AMPK and the NRF2/HO-1 system (54).

The effects of 6-gingerol and 6-shogaol on DCs are summarized in Table 3, and illustrated in Figure 2C.

2.4 Effects of gingerols on T cells

As specialized cells of the adaptive immune system, T cells play a central role in directing immune responses against pathogens, allergens, or tumors. While CD4+ T helper cells modulate immune responses by facilitating the activity of other immune cells, regulatory T cells (Treg) contribute to the maintenance of immune homeostasis and CD8+ cytotoxic T cells are essential to the elimination of cancer cells and virally infected cells (55). Nevertheless, it must be noted that specific populations of T cells such as Th17 cells might contribute to the pathogenesis of various chronic inflammatory disorders (56).

The first study regarding the effects of gingerols on the activation and effector function of T cells was published in 2015. Bernard et al. found that 6-, 8- and 10-gingerols inhibited the proliferation of mouse spleen-derived CD3+ T cell in response to anti-CD2/CD3/CD28-coated beads (Dynabeads) or syngeneic DCs (57). 8- and 10-gingerols also significantly decreased the expression of the activation markers CD25 and CD69. All three gingerols reduced the Dynabead-stimulated production of interferon-gamma (IFN- γ), but did not affect IL-4 synthesis. Moreover, 8- and 10-gingerols decreased IL-2 secretion as well. In general, the data

suggest that 8- and 10-gingerols are more potent inhibitors of T cell activation and proliferation than 6-gingerol (57).

In accordance with the above, another study demonstrated that 6-gingerol is able to suppress the production of IFN- γ and IL-4 by mouse spleen-derived CD4+ T cells cultured under Th1 or Th2 polarizing conditions, respectively (58). 6-gingerol also significantly suppressed the proliferation of pan T cells isolated from ovalbumin (OVA)-sensitized mice. In addition, 6-gingerol abrogated the staphylococcal enterotoxin B (SEB)-induced proliferation of unprimed T cells and the proliferation of human Jurkat T cells in response to anti-CD3/CD28 and IL-2. Pre-treatment with 6gingerol also inhibited the activation of p38, ERK1/2 and JNK, as well as the nuclear translocation of NF- κ B and c-fos transcription factors in Jurkat cells stimulated with PMA and ionomycin. Thus, the results suggest that 6-gingerol suppresses T cell activation and proliferation through inhibiting NF- κ B and activating protein 1 (AP-1) activation (58).

The effects of 6-gingerol on CD8+ T cells were studied only in the context of cancer. Two publications reported that 6-gingerol is able to increase the number of tumor-infiltrating CD8+ T cell in mice that might play an important role in the anti-tumor effect of ginger (59, 60). The anti-cancer activity of ginger compounds is not in the scope of the recent review; however, it has been discussed in detail elsewhere (17, 18). The effects of gingerols on CD4+ T cells are summarized in Table 4, and illustrated in Figure 2D.

In conclusion, the above data indicate that the bioactive compounds of ginger can efficiently modulate the functionality of different immune cell types, mostly via interfering with the activity of various signaling pathways in them. Nevertheless, opposing data were also published regarding the effects of different ginger phenolics that might be attributed to the varying cell types, culture conditions and stimuli applied in these studies. Furthermore, the effect of ginger phenolics on other immunce cell types such as NK cells, basophils, and eosinophils have not been explored yet. Furthermore, in lymphomatic cell lines with B cell origin only the anti-cancer potential of these compounds were investigated (61, 62).

Altogether, the pleiotropic effects of ginger phenolics suggest that their mechanism of action likely involves multiple regulatory mechanisms, which is discussed in chapter 4 in details.

TABLE 3	Effects	of	6-gingerol	and	6-shogaol	on	DCs.
---------	---------	----	------------	-----	-----------	----	------

DCs							
Compounds	Cell types	Challenges	Observed effects	Potential mechanisms	Ref.		
6-gingerol	mouse BM-DC	LPS	 reduced TNF, IL-1β, IL-6 and IL-23 secretion reduced CD80, CD86 and MHC II expression reduced Th17 priming capacity 	 inhibited NF-κB, JNK and ERK1/ 2 activation 	(53)		
6-gingerol, 6-shogaol	human moDC	various TLR ligands	 decreased costimulatory molecule and MHC II expression reduced TNF, IL-6 and IL-10 production decreased capacity to prime Th1 T cells 	 inhibited NF-κB nuclear translocation reduced mTOR and MAPK activity 6-shogaol increased AMPK and NRF2 activity 	(54)		

TABLE 4 Effects of gingerols on T cells.

T cells									
Compounds	Cell types	Challenges	Observed effects	Potential mechanisms	Ref.				
6-, 8- and 10-gingerols	mouse spleen-derived CD3+ T cell	Dynabead, syngeneic DCs	 decreased proliferation reduced expression of CD25 and CD69 reduced IFNγ and IL-2 	-	(57)				
6-gingerol	mouse spleen-derived CD4+ T cell	ovalbumin, Th1 or Th2 polarizing conditions, SEB	 decreased proliferation reduced IFNγ and IL- 4 production 	-	(58)				
6-gingerol	Jurkat human T cell line	PMA, ionomycin anti-CD3/CD28 and IL-2	reduced proliferationdecreased T cell activation	 inhibited MAPK activation suppressed NF-κB and AP-1 activation 	(58)				

3 Protective effects of ginger phenolics in animal models of autoinflammatory and autoimmune diseases

Abnormal activation of the immune system can lead to the generation of autoinflammatory or autoimmune disorders. The pathogenesis of autoimmune diseases is characterized by loss of tolerance against self-tissues, appearance of autoreactive T and B cells, and production of autoantibodies. On the contrary, autoinflammatory diseases are mainly caused by altered innate immunity. They are also characterized by the activation of inflammasomes, and lack of self-reactive antibodies and T cells (63). Although there is a wide range of treatment options to manage symptoms, currently there is no cure for these disorders. As we introduced above several reports suggest that gingerols and shogaols might also alleviate disease symptoms due to their strong antioxidant and anti-inflammatory activities. These assumptions were later also supported by data from animal models of inflammation such as sepsis (42, 64, 65) or neuroinflammation (36, 38). Here we summarize the most recent in vivo animal data on the efficacy of gingerols and shogaols in the treatment of autoinflammatory and autoimmune diseases.

Ulcerative colitis (UC), one of the main forms of inflammatory bowel disease, is characterized by chronic recurrent inflammation of the large intestine. The etiology and pathogenesis of these diseases is not yet fully understood, although it appears to involve both autoinflammatory and autoimmune traits (66). A study investigated the therapeutic efficacy of intraperitoneally-injected 6-, 8-, and 10-gingerols in a dextran sulfate sodium (DSS)induced rat colitis model (67). Results show that all three gingerols attenuated DSS-induced symptoms of colitis and accelerated the healing of mucosal damage. Gingerols elevated the activity of the anti-inflammatory enzyme superoxide dismutase (SOD), while reduced the activity of MPO, a marker of neutrophil infiltration, in the colon tissue. All three gingerols also reduced the DSS-induced serum levels of the pro-inflammatory cytokines TNF and IL-1 β (67).

A subsequent study further explored the mechanism behind the anti-inflammatory effects of 6-gingerol in the DSS-induced mouse

colitis model (68). Orally-administered 6-gingerol significantly decreased the DSS-induced weight loss of mice. It was demonstrated that 6-gingerol decreased IL-17 levels, while it increased IL-10 levels both in the serum and bowel tissues of DSS-treated mice. Moreover, 6-gingerol inhibited the DSS-induced phosphorylation of I κ B and the phosphorylation and nuclear translocation of p65 in the bowel tissue (68).

Another study investigated the efficacy of orally-administered 6-shogaol-loaded nanoparticles in the mouse model of DSSinduced colitis (69). Similar to gingerols, 6-shogaol also alleviated colitis symptoms and accelerated wound repair. In addition, 6shogaol significantly decreased the levels of TNF, IL-6, IL-1 β and iNOS, while it increased the expression of the anti-inflammatory genes HO-1 and NRF2 in colon tissues of DSS-treated mice (69).

A recent study compared the anti-colitis efficacy of 6-, 8-, 10gingerols and shogaols in the DSS-induced colitis mouse model (70). All components were able to prevent DSS-induced weight loss, colon length reduction and IL-1 β , IL-6 and IFN- γ serum levels in DSS-treated mice. Among the ginger compounds, 8-, and 10shogaols, which were also able to significantly decrease DSSinduced serum levels of TNF, showed the greatest efficacy compared to the other compounds. 6-, 8-, and 10-gingerols and 6-, 8-, and 10-shogaols also downregulated iNOS and COX-2 protein levels and significantly reduced phosphorylation of NF-KB in colonic tissue. Interestingly, 10-shogaol was found to be the most potent in its ability to block iNOS and COX-2 expression and NFκB activity. It was also the most effective in increasing the expression of tight junction proteins, thus the intestinal integrity of DSS-treated mice. The study concluded that 8-, and 10-shogaols are highly efficient in their ability to suppress colitis symptoms and inflammation, thus may serve as better candidates for the treatment of colitis than the corresponding gingerols.

Multiple sclerosis (MS) is a T cell-mediated autoimmune disease of the central nervous system that is characterized by immune-mediated demyelination in the spinal cord and cerebral cortex (71). The neuroprotective effects of ginger phenolics were extensively studied in animal models of neurotoxicity and brain damage [reviewed in (19)]; however, so far only two studies investigated the immunomodulatory activity of gingerols and shogaols in experimental autoimmune encephalomyelitis (EAE), the mouse model of MS. The first study showed that 6-gingerol

decreased the inflammatory infiltration and demyelination in the white matter of spinal cord of EAE mice (53). 6-gingerol-treated mice had lower numbers of inflammatory cells including DCs and T cells in the spleen. Further, splenocytes from 6-gingerol treated mice produced significantly lower levels of IL-17 and GM-CSF indicating that 6-gingerol inhibited Th17 cell polarization in vivo (53). In addition, 6-gingerol lowered the percentage of leukocytes such as CD11c+MHC II+ DCs, CD45+CD11b+ monocytes and Th17 cells in the central nervous system (CNS) of EAE mice. This suggests that 6-gingerol inhibits inflammatory cell infiltration in the CNS. The next study investigated whether 6-shogaol or its metabolite, 6-paradol could ameliorate EAE symptoms and inflammation (72). Administration of 6-shogaol or 6-paradol significantly reduced the clinical signs of the disease. Histological analysis showed that both components decreased demyelination, cell accumulation and TNF expression in the spinal cord of EAE mice. In addition, 6-shogaol and 6-paradol markedly reduced astrogliosis and microglial activation of EAE mice. These findings suggest that the neuroprotective effects of 6-gingerol and 6-paradol in EAE might be associated to the dampened inflammatory responses in the CNS (72).

The protective role of 6-gingerol was also investigated in the context of systemic autoimmune diseases such as systemic lupus erythematosus (SLE) and rheumatoid arthritis (RA). SLE affects almost every organ in the body and is characterized by the appearance of anti-nuclear autoantibodies and circulating immune complexes. In a lupus mouse model 6-gingerol administration significantly reduced serum levels of anti-dsDNA, cell-free DNA and also MPO-DNA complexes, the latter of which serves as a marker of NET formation (47). 6-gingerol also greatly reduced the serum levels of the pro-inflammatory cytokines TNF and IFN-y. In line with that, 6-gingerol suppressed netosis and thus large-vein thrombosis in a mouse model of antiphospholipid syndrome (APS) as well. Pharmacokinetic studies further revealed that 6-gingerol accumulated in neutrophils, even while its plasma level was dropping (47). These results are in line with previous reports showing that ginger phenolics are converted into glucuronide conjugates and thus are rapidly cleared from the plasma. In tissues, these conjugated forms are then reconverted into their free active form by specific enzymes such as β glucuronidase (73, 74).

RA is also a systemic chronic immune-mediated disorder, which is characterized by synovial inflammation, joint damage, loss of function and deformities. IL-17 is one of the key cytokines in promoting inflammation and thus cartilage damage in RA (75). Therefore, in a recent study, 315 natural extracts were tested in their ability to inhibit IL-17-induced IL-6 production by synovial cells (76). Among these extracts, dried ginger and in particular its specific component 8-shogaol showed the highest inhibitory activity against IL-17-mediated IL-6 secretion by synovial cells and macrophages. Thereafter, the anti-arthritic potency of 8shogaol was investigated in adjuvant induced arthritis (AIA), a rat model of RA (76). Treatment with 8-shogaol reduced paw thickness and weight loss of AIA rats and improved their walking performance as well. Furthermore, 8-shogaol treatment decreased bone erosion and cellular infiltration into the joints of AIA rats. Upon 8-shogaol treatment the levels of TNF, IL-6 and IL-8 were markedly downregulated both in the serum and synovial tissues of AIA rats. These data suggest that 8-shogaol has the potential to ameliorate disease severity and inflammation in RA (76).

The above data indicate that the bioactive compounds of ginger exert a general anti-inflammatory effect by efficiently decreasing immune reactions in a number of autoimmune and inflammatory disease models, thus it is plausible that ginger-derived phytochemicals might serve as supplementary agents in the therapy of these diseases. The effects of the major ginger phenolics in animal models of autoimmune and autoinflammatory diseases are included in Table 5.

4 Proposed mechanism of action of gingerols and shogaols

As we have introduced above ginger constituents are able to interfere with common inflammatory signaling pathways including the NF- κ B, MAPK and PI3K/Akt/mTOR cascades in different immune cell types; however, the previously introduced *in vitro* studies did not, or rarely revealed the mechanism behind the antiinflammatory actions of ginger phenolics. Nevertheless, mounting evidence, mostly from *in vivo* studies and experiments with nonimmune cell types, indicates that ginger constituents might affect the inflammatory signaling pathways by targeting different upstream regulatory molecules such as AMPK, NRF2, or PPAR γ . Recent publications also suggest that these compounds might exert their actions through binding to the TRPV1 ion channel. In this chapter we summarize our current knowledge about the possible mechanisms of immunomodulation by different ginger phenolics.

4.1 The mechanism of action of ginger phenolics through AMPK activation

AMPK is a central regulator of diverse physiological and metabolic processes, which can be dysregulated in pathological conditions, such as cancer, obesity or chronic inflammatory diseases (77). Generally, AMPK blocks mTORC1 activation by phosphorylating two of its regulatory proteins. On the one hand AMPK phosphorylates and thereby increases the activity of Tuberous Sclerosis Complex 2 (TSC2), an upstream inhibitor of mTORC1. On the other hand, it phosphorylates Raptor, a scaffold protein of mTORC1 that also leads to mTORC1 inactivation and consequently to the suppression of anabolic processes and inflammation (78). Accumulating evidence suggests a negative correlation between AMPK activity and inflammation (77). Chronic inflammation is associated with reduced AMPK activity and thereby aberrant mTOR activity. Thus, targeting AMPK might serve as a potential therapeutic strategy for treating inflammatoryrelated diseases (79). Many phytochemicals such as resveratrol, quercetin, and curcumin act as natural activators of AMPK (80). Recent studies also suggest that ginger phytochemicals might also exert their anti-inflammatory effects on immune cells via suppressing mTOR and increasing AMPK activity.

Compounds	Models	Administration routes	Observed effects	Potential mechanisms	Ref.
6-, 8-, and 10-gingerols	DSS-induced rat colitis model	intraperitoneal	 attenuated DSS-induced symptoms of colitis accelerated mucosal damage healing decreased neutrophil infiltration in the colon reduced TNF and IL-1β serum levels 	increased SOD activitydecreased MPO activity	(67)
6-gingerol	DSS-induced mouse colitis model	oral	 decreased DSS-induced weight loss decreased IL-17 levels and increased IL-10 levels both in serum and bowel tissue 	 inhibited IKB phosphorylation suppressed NF-KB nuclear translocation in the bowel tissue 	(68)
6-shogaol	DSS-induced mouse colitis model	oral	 alleviated colitis symptoms accelerated wound repair decreased the levels of TNF, IL-6, IL-1β and iNOS 	• increased HO-1 and NRF2 expression in the colon tissues	(69)
6-, 8-, 10-gingerols and 6-, 8-, 10-shogaols	DSS-induced mouse colitis model	oral	 prevent DSS-induced weight loss, colon length reduction reduced IL-1β, IL-6 and IFN-γ serum levels downregulated iNOS and COX-2 levels 	• reduced NF-κB phosphorylation	(70)
6-gingerol	EAE mice	intraperitoneal	 decreased inflammatory cell infiltration and demyelination in spinal cord and CNS lowered number of inflammatory cells in the spleen inhibited Th17 polarization 	-	(53)
6-shogaol, 6-paradol	EAE mice	oral	 decreased demyelination, cell accumulation and TNF expression in spinal cord reduced astrogliosis and microglial activation 	-	(72)
6-gingerol	lupus and APS mouse model	intraperitoneal	 reduced serum level of anti-dsDNA, cell-free DNA, MPO-DNA complexes, TNF, IFN-γ suppressed netosis inhibited thrombosis in APS 	-	(47)
8-shogaol	AIA rats	intraperitoneal	 reduced paw thickness and weight loss improved walking decreased bone erosion and cellular infiltration to the joints reduced TNF, IL-6 and IL-8 levels in serum and synovial tissues 	-	(76)

TABLE 5 In vivo effects of ginger phenolics in animal models of autoinflammatory and autoimmune diseases.

Initially, studies reported that 6-shogaol inhibit inflammation by suppressing the phosphorylation of key mTOR pathway components. First it was demonstrated that 6-shogaol inhibited the LPS-induced phosphorylation and activation of NF- κ B by interfering with the PI3K/Akt and MAPK signaling pathways in murine macrophages (34). Thereafter, it was shown that 6-shogaol inhibited TNF-induced disassembly of tight junctions via inhibition of NF- κ B and PI3K/Akt in the human colonic epithelial cell line HT-29/B6 (81). Another study further demonstrated that 6gingerol suppressed LPS-induced microglia activation by downregulating the Akt/mTOR pathway activity (38).

Later it was also demonstrated that 6-gingerol indirectly suppress mTOR activity by promoting the activation of AMPK. First it was reported that ginger extract restored the high fat diet (HFD)-induced downregulation of AMPK activity in the liver of rats (82). Another study presented that 6-gingerol attenuates hepatic steatosis, inflammation and oxidative stress in HFD-fed mice via activating AMPK (83). In particular, intragastrical administration of 6-gingerol decreased the concentration of TNF, IL-6 and ROS, while it increased the phosphorylation level of liver kinase B1 (LKB1) and AMPK in the liver of HFD-fed mice. LKB1 is one of the master upstream kinases of AMPK, which forms a trimeric complex with two other proteins and then phosphorylates and activates AMPK. The authors suggest that 6-gingerol enhanced the stability of the LKB1 complex that consequently increased LKB1/AMPK pathway activity in the liver of mice. These results were also strengthened by *in vitro* experiments using palmitic acid-treated HepG2 cells, in which 6-gingerol reduced lipid accumulation and oxidative stress by increasing LKB1 complex stability and thus LKB1/AMPK pathway activity (83).

Recent observations further suggest that the anti-diabetic and anti-cancer efficacy of ginger phenolics are also associated with their ability to induce AMPK activation. It was reported that 6-shogaol and 6-paradol promote glucose utilization in mouse adipocytes that could be attributed to the increased activity of AMPK (84). Furthermore, 6-gingerol was found to suppress oral cancer cell growth by inducing the activation of AMPK and blocking mTOR pathway activity (85), while 10-gingerol was shown to inhibit the proliferation and migration of vascular smooth muscle cells by AMPK activation (86). The latter study also demonstrated a stable binding between 10-gingerol and AMPK by molecular docking studies and surface plasmon resonance imaging analysis. This was the first study showing a direct interaction between 10-gingerol and AMPK that proposed 10-gingerol as a natural agonist of AMPK. Nevertheless, there are no available data on the binding affinity of other ginger phytochemicals to AMPK.

Although, some reports demonstrated that gingerols and shogaols decrease PI3K/Akt/mTOR signaling pathway activity in immune cells, the regulatory role of AMPK activity was not investigated in these studies. We have recently reported that 6shogaol was able to inhibit mTORC1 signaling through upregulating AMPK activity in moDCs, while 6-gingerol was not able to do so (54). Nevertheless, further studies are needed to reveal whether the anti-inflammatory activities of gingerols and shogaols could be generally linked to increased AMPK activity.

Figure 3A shows how the activation of AMPK by the major ginger phenolics might affect the mTOR and thus the NF-KB signaling pathways.

4.2 Anti-inflammatory actions of ginger phenolics through NRF2 regulation

NRF2 is a transcription factor that plays a pivotal role in the regulation of antioxidant and anti-inflammatory responses (87). Besides inducing the expression of an array of enzymes involved in antioxidant defense and detoxification, it is able to negatively regulate NF-KB activity by multiple mechanisms. For instance, by inducing antioxidant enzymes NRF2 decreases ROS levels that consequently inhibit oxidative stress-mediated NF-KB activation. NRF2 is also able to prevent IkBa proteasomal degradation and thus the nuclear translocation of NF-KB. In addition, a series of in vitro and in vivo studies indicated that NRF2-mediated HO-1 expression substantially contributes to the anti-inflammatory activity of NRF2. As an important antioxidant enzyme HO-1 catalyzes the degradation of toxic free heme that leads to the



Schematic representation of the possible mechanism of action of ginger phenolics in immune cells (A) Ginger phenolics suppress mTORC1 functionality by increasing AMPK activity. Generally, mTORC1 supports protein synthesis and thus cell growth and proliferation by direct phosphorylation of p70S6 kinase (p70S6K) and eukaryotic initiation factor 4E-binding protein 1 (4E-BP1). In immune cells mTORC1 is also involved in NF-kB activation and its target gene expression. (B) Ginger-derived bioactive compounds might also exert their anti-inflammatory effects by activating NRF2, which induces the expression of an array of antioxidant and cytoprotective genes. NRF2 also inhibits NF-KB activity by preventing IκBα proteasomal degradation and thus blocks the nuclear translocation of NF-κB. (C) Ginger phenolics might also exert their anti-inflammatory effects by activating PPARy. By interacting with p65 NF-xB, PPARy can induce its ubiquitination and proteolytic degradation. Stimulation of PPAR-y limits NF-kB dependent gene transcription through transrepression as well. Moreover, PPARy supports the expression of inhibitory proteins such as IxBa via transactivation. (D) Ginger constituents can directly interact with TRPV1 channels and might act as either TRPV1 agonists or antagonists. Generally, TRPV1 mediated Ca²⁺ influx facilitates the activation of different transcription factors that might be facilitated or inhibited by gingerderived bioactive compounds. NFAT: nuclear factor of activated T cells, PPRE: PPAR response elements.

release of anti-inflammatory products, such as carbon monoxide and biliverdin (88). Beyond its enzymatic functions, HO-1 exerts non-canonical, signaling functions, through which HO-1 is able to control essential cellular signaling processes such as gene expression, protein translation, and DNA repair mechanisms (89).

In the last few years many studies proposed that ginger-derived bioactive compounds exert their anti-inflammatory effects by activating the NRF2/HO-1 axis. A study suggest that this is especially true for shogaols, which in contrast to gingerols, bear an α,β -unsaturated carbonyl group in their side chain (90). Molecular modeling revealed that 6-shogaol is able to increase HO-1 levels through a Michael reaction between its α , β unsaturated carbonyl group and Kelch-like ECH-associated protein 1 (Keap1), which acts as a negative regulator of NRF2. This interaction initiates conformational changes in Keap1 that allows the release of NRF2 from Keap1 leading to its nuclear translocation. In the nucleus NRF2 binds to the antioxidant response element (ARE) and induces the expression of various antioxidant and anti-inflammatory genes. In that particular study, 6-shogaol was found to upregulate the protein levels of HO-1 and suppress thrombin-induced NO release in rat microglia, while 6gingerol was not able to do so (90). 6-shogaol also increased the nuclear import of NRF2 in BV2 microglia and significantly enhanced the HO-1 mRNA level in primary-cultured microglia (90). Similarly, we have recently demonstrated that 6-shogaol increases the expression and nuclear translocation of NRF2 and enhances the protein levels of HO-1, while 6-gingerol does not affect that in human moDCs (54). In an in vivo mouse ICH model, 6shogaol increased striatal HO-1 protein levels and rescued neuron loss (90). Another study demonstrated that 6-shogaol protects against renal ischemia-reperfusion (I/R) injury as well. 6-shogaol pretreatment significantly decreased the expression of various proinflammatory cytokines and chemokines in mice subjected to renal I/R. Mechanistically, 6-shogaol reduced kidney inflammation by attenuating NF-KB activation and inducing HO-1 expression (91). In the same year it was published that 6-shogaol prevented UVBinduced inflammation and oxidative stress through modulating NRF2 signaling in human epidermal keratinocytes (HaCaT cells) (92). 6-shogaol significantly decreased the UVB-triggered expression of IL-6, TNF and IL-10, while reduced the phosphorylation of ERK, JNK and p38 MAPKs in human epidermal keratinocytes. The authors further demonstrated that 6-shogaol prevented UVB-induced depletion of NRF2 and elevated HO-1 protein levels in HaCaT cells. In another in vitro model, 6shogaol treatment increased the expression of HO-1 in LPSstimulated human umbilical vein endothelial cells (HUVECs) as well (93). Similar to 6-shogaol, 6-gingerol was also able to activate the NRF2/Keap1 signaling pathway. It was demonstrated that 6gingerol significantly reduced the expression of Keap1 and increased that of NRF2 in the nuclear fraction of buccal pouch tissues of 7,12-dimethylbenz(a)anthracened (DMBA)-treated hamsters (94). By doing so, 6-gingerol prevented buccal pouch carcinogenesis through inhibiting the expression of inflammatory and cell proliferation markers such as IL-6, TNF- α , IL-1 β , iNOS, COX-2 and cyclin D, while inducing pro-apoptotic markers such as Bax in DMBA-induced hamsters. These findings were further strengthened by another study showing that 6-gingerol also alleviates sepsis-induced liver injury through activating the NRF2 pathway (65). Pre-treatment with 6-gingerol attenuated cecal ligation and puncture (CLP)-induced hepatic inflammation and injury by increasing the protein levels of NRF2 and HO-1 in liver homogenates of C57BL/6 mice. These results were further confirmed by an in vitro model. 6-gingerol suppressed ATPinduced pyroptosis, IL-1B and caspase-1 secretion and ROS production by preventing the downregulation of NRF2 and HO-1 protein levels in LPS-primed RAW 264.7 cells (65). Furthermore, it was shown that 6-gingerol decreased cardiac injury via the NRF2/ HO-1 pathway in both mouse and cell models of diabetic cardiomyopathy (DCM) (95). In the mouse heart 6-gingerol inhibited the expression of ferroptosis-related proteins, while enhanced the expression of anti-ferroptosis-related proteins. In addition, 6-gingerol treatment decreased the levels of IL-1β, IL-6, and TNF- α in serum and heart tissues of diabetic mice. In H9c2 cardiac cells, 6-gingerol also inhibited ferroptosis and inflammation evoked by palmitic acid and high glucose administration. Most importantly, 6-gingerol enhanced the NRF2 and HO-1 protein levels both in the mouse and cell models of DCM. Hence, the authors suggested that 6-gingerol might protect against DCM by inhibiting ferroptosis and inflammation via activating the NRF2/ HO-1 pathway (95). A novel study suggests that 6-gingerol attenuates sepsis-induced acute lung injury by suppressing NLRP3 inflammasome activation via NRF2 activation (43). In LPS-induced rats, 6-gingerol repressed the expression of various oxidative stress markers, inflammatory cytokines and NLRP3 inflammasome components in the lung tissues and inhibited the infiltration of inflammatory cells into the lungs. In addition, 6gingerol prevented the LPS-mediated downregulation of NRF2 and HO-1 levels in the lung tissues of rats. Intraperitoneal injection of ML385, an NRF2 inhibitor, reversed the protective effect of 6gingerol against LPS-induced oxidative stress and inflammation suggesting that 6-gingerol exerts its anti-inflammatory effects through activating the NRF2/HO-1 axis (43). Similar to 6gingerol, zingerone, a less-studied natural compound of ginger, also reduced organ injury through activating the NRF2 signaling pathway in a CLP-induced sepsis model (49). In particular, administration of zingerone alleviated ROS accumulation and systematic inflammation in septic mice and inhibited NET formation both in vivo and in vitro. The results further suggest that zingerone attenuates NET formation and inflammation via NRF2-dependent ROS inhibition (49).

As the above data indicate many studies have demonstrated that the major bioactive compounds of ginger are able to activate the NRF2/HO-1 axis that seems to greatly contribute to their antiinflammatory effects. Lately, emerging evidence indicate a cooperation between AMPK and NRF2 signaling as well. In particular, AMPK might function as a positive upstream regulator of the NRF2/HO-1 system and thus can lead to the transactivation of specific target genes (96). Interestingly, the AMPK-mediated enhancement of the NRF2/HO-1 response does not seem to depend on mTOR inhibition (97). In our studies, we found that 6-shogaol is able to increase the activity of both AMPK and NRF2 in human DCs (54). Nevertheless, it needs to be further elucidated whether these components contribute separately to the ant-inflammatory potential of 6-shogaol or AMPK acts as an upstream regulator of NRF2.

It is also plausible that certain phytochemicals might control NRF2 activity through the E3 ubiquitin ligase tripartite motifcontaining 29 (TRIM29), also known as ataxia telangiectasia group D complementing (ATDC). Recent evidences indicate that TRIM29 plays a versatile role in orchestrating inflammatory responses. In alveolar macrophages, TRIM29 negatively regulates the LPS-induced pro-inflammatory cytokine production by inducing the ubiquitination and degradation of NF-KB essential modulator (NEMO) (98). In mice, TRIM29 controls proinflammatory cytokine production by acting as a central regulator of protein kinase RNA-like endoplasmic reticulum kinase (PERK)driven endoplasmic reticulum (ER) stress response (99). Mechanistic studies have also shown that TRIM29 is also able to bind to and sequester KEAP1, thereby preventing NRF2 from proteasomal degradation in pancreatic cancer cells (100). To the best of our knowledge, it has never been investigated whether plantderived compounds are able to control TRIM29; however, as the previous reports suggest TRIM29 acts as a multifunctional protein, and thus might regulate immune responses by various mechanisms. In the future, it could also be explored whether phytochemicals control NRF2 activity through TRIM29 that might help to better understand the mechanism behind their anti-inflammatory actions.

Figure 3B represents how ginger phenolics might exert their anti-inflammatory effects via activating the NRF2 signaling pathway.

4.3 Mode of action of ginger-derived compounds through PPARy regulation

PPARy is a member of the nuclear receptor superfamily that is widely expressed in immune and endothelial cells (101). It is able to regulate various biological functions, such as inflammatory responses and lipid metabolism. Most importantly, PPAR γ can suppress inflammation by promoting the inactivation of NF- κ B through direct or indirect mechanisms. PPAR γ can cause ubiquitination and proteolytic degradation of p65 NF- κ B, and it can also promote the expression of inhibitory proteins such as I κ B α or HO-1. Many dietary phytochemicals target PPAR γ (102), however, only a few studies investigated whether 6-gingerol and 6-shogaol act as PPAR γ agonists.

In rats subjected to ventilator-induced lung injury, GW9662, a selective PPAR γ inhibitor abolished the protective effect of 6-gingerol (103). In particular, the 6-gingerol-mediated decrease in proinflammatory cytokine release, neutrophil accumulation and oxidative stress in lung tissues of rats challenged with mechanical ventilation was partially reversed by the inhibition of PPAR γ (103). 6-shogaol was also suggested to exert its anti-inflammatory effects in LPS-activated BV2 microglia by activating PPAR γ (37). In these cells 6-shogaol significantly reduced LPS-induced NF- κ B activation, pro-inflammatory cytokine and PGE₂ release that could be reversed by GW9662 treatment. In contrast, another study presented that GW9662 could not reverse the inhibitory effect of 6-shogaol on TNF-induced disassembly of tight junctions in human colonic HT-29/B6 cells (81). Although data on the relation of ginger phytochemicals to PPAR γ activity is scarce and contradictory, we cannot exclude the possibility that the anti-inflammatory effects of gingerols and shogaols are at least partially dependent on PPAR γ activity.

Figure 3C shows that 6-gingerol and 6-shogaol might inhibit the NF-κB pathway through activating PPARγ.

4.4 The mechanism of action of gingerderived phytochemicals through targeting TRPV1

TRPV1 is primarily expressed in sensory neurons and plays an essential role in heat sensation and nociception. Accumulating data suggests that TRPV1 is also expressed in various mammalian immune cells, especially in macrophages, neutrophils, DCs and T cells, where it modulates various functions such as cytokine release, migration or phagocytic activity (104). Nevertheless, exploring the role of TRPV1 on immune cells is hindered by the sometimes-contradictory results gained from human and animal cells. Specifically, TRPV1 activation promotes the maturation and activation of murine DCs (105), while inhibits the differentiation, maturation, phagocytosis and cytokine secretion of human moDCs (106).

The major pungent compounds of ginger share certain structural characteristics with capsaicin, the main pungent constituent found in chili pepper, and the prototypic agonist of TRPV1 (107). In particular, gingerol, shogaol and zingerone contain the same vanillyl head group as capsaicin; therefore, these ginger constituents can directly activate TRPV1 channels, though with a much lower potency than capsaicin (108). Specifically, a study showed that while capsaicin shows strong potency on TRPV1, 6-shogaol and 6-gingerol have presented only moderate potency on the channel (109). It was further demonstrated that 6-shogaol, 6-gingerol, and zingerone bind to the same ligand-binding pocket in TRPV1 channels as capsaicin; however, the distinct structural features in their tails cause large differences in their potency (110). 6-shogaol is the most similar to capsaicin regarding its molecular structure. The aliphatic tail of 6shogaol similar in length to that of capsaicin, and 6-shogaol has the same functional groups, which are able to form hydrogen bonds with the TRPV1 channels. capsaicin. The major difference between 6shogaol and capsaicin is that the C=C bond is located at the tip of the tail in capsaicin, whereas it is at base of the tail in 6-shogaol. Although, shifting the C=C bond limits the rotational freedom of the tail of 6-shogaol, 6-shogaol represents the strongest agonist of TRPV1 among the ginger compounds (110). In comparison to 6shogaol, 6-gingerol is slightly weaker in its potency to induce TRPV1 activity. This might be due to the presence of a hydroxyl group instead of a double bond in the tail of 6-gingerol that reduces its lengths. Finally, zingerone is a weak agonist for TRPV1 channels that might be explained by the shortness of its aliphatic tail. Altogether, the study suggests that the length of the aliphatic tail, the presence or absence of the hydroxyl group, and the position of the C=C bond in the tail all might influence the ability of ginger phenolics to bind and activate TRPV1 (110).

In a rat model of I/R injury 6-gingerol was found to inhibit NLRP3-mediated inflammation and neuronal apoptosis, while it

upregulated autophagy (111). The mechanistic findings indicate that 6-gingerol exerts anti-apoptotic and anti-inflammatory effects during cerebral I/R injury via dissociating TRPV1 from Fas-associated factor 1 (FAF1), which is able to modulate the sensitivity and thus the activity of TRPV1 to various stimuli.

The ligand-induced activation of TRPV1 by ginger compounds in non-neuronal cell types is poorly characterized. So far, only one study investigated whether the ligand-induced activation of TRPV1 by 6-gingerol is able to affect the functionality of neutrophils (50). The analyses showed that 6-gingerol increased intracellular Ca²⁺ concentrations and the expression of common surface markers such as CD11b and CD66b in neutrophils. Moreover, 6-gingerol enhanced fMLP-stimulated CXCL8 secretion and ROS production that could be reversed by pharmacological inhibition of TRPV1. Here the authors applied 6-gingerol at a low dose of 50 nM, which is a dietary relevant concentration that could be reached in blood plasma after consumption of 1 liter of ginger tea (50). In contrast, 50 µM of geranylacetone, a natural sesquiterpenoid, acting as a TRPV1 agonist inhibited fMLP-induced migration of neutrophils and CXCL8-induced intracellular Ca²⁺ mobilization (112). These observations suggest that lower concentrations of TRPV1 ligands might induce immune cell functions, while higher TRPV1 ligand concentrations in the µM range rather inhibit immune responses. Therefore, further studies are needed to reveal the exact mode of action of ginger phenolics on TRPV1.

Figure 3D shows that acting through TRPV1, various ginger phenolics might affect the activity of various signaling pathways.

5 Concluding remarks and future perspectives

In this review, our goal was to compile the most relevant findings on the immunomodulatory actions of the major ginger phenolics. We mainly focused on the anti-inflammatory effects of these compounds on different immune cells and presented a variety of studies, which demonstrated the possible mode of their antiinflammatory actions. Though, comparative studies are sparse, most of the findings indicate that 6-shogaol exhibit a stronger anti-inflammatory and antioxidant capacity than 6-gingerol that might be attributed to the presence of the α,β -unsaturated ketone moiety in its skeleton. Although the exact mechanism of action of ginger-derived bioactive compounds are not yet fully elucidated, data show that the application of these compounds results in the successful inhibition of common signaling pathways such as the NF-KB and PI3K/Akt/mTOR signaling cascades that consequently leads to the suppression of inflammatory responses by immune cells. Interestingly, contradictory results were obtained regarding the action of gingerols and shogaols on the activity of MAPK signaling components; however, the reason for the varying results has not been revealed yet. Research data summarized above further suggest that ginger phytochemicals exert their pleiotropic effects via targeting multiple regulatory molecules including AMPK, PPARy, NRF2 and also TRPV1. In addition, recent in silico molecular docking and molecular dynamics stimulation studies revealed that 6-gingerol has a binding affinity towards COX-1/2 and 5- lipoxygenase, thus it has the potential to directly target various enzymes associated with inflammation (113).

Ginger has been used traditionally as an herbal medicine for the treatment of many maladies, and due to their potent immunomodulatory capacity several reports proposed gingerderived bioactive compounds as candidates for the management and prevention of autoimmune diseases. Nevertheless, clinical studies are lacking, probably due to the poor water solubility and adsorption of ginger phenolics. Among the organic solvents, ethanol has been proposed as an optimal solvent since it is able to retain the bioactivity of ginger compounds (114, 115). Pharmacokinetic studies also revealed that once ginger is consumed gingerols and shogaols are absorbed quickly and are majorly present in the form of water soluble glucuronide conjugates in the human plasma (74, 116). Although the level of ginger phenolics and their metabolites is low and drops quickly in the serum, experiments with mice demonstrated that gingerols and shogaols may reach higher concentrations in tissues in comparison to the serum levels due to the accumulation of free and conjugated forms of ginger phenolics in various organs (73).

Nevertheless, further studies are needed to increase the solubility, stability, half-life time and thus the therapeutic efficiency of ginger phenolics. Recently, more and more projects are focusing on the development of novel formulation techniques and drug delivery systems to increase the efficacy and to reduce the toxicity of the bioactive compounds of ginger (104, 117–120). Current studies indicate that nanoformulation, ensuring a sustained release of ginger constituents, might provide an effective delivery system. Possible nanotechnological approaches include carriers, such as nanovesicles, exosome-like nanoparticles, nanostructure lipid carrier, and emulsions (20). Optimization of nano-drug delivery systems for the targeted delivery and controlled release of ginger phenolics is still in its infancy; however, it is feasible that in the future ginger supplements might provide an alternative or most probably a complementary therapeutic approach to treat various inflammatory and autoimmune disorders.

Author contributions

KP: Writing – original draft, Writing – review & editing. AS: Writing – original draft, Writing – review & editing. TF: Writing – original draft, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This work was supported by the National Research, Development and Innovation Office (NKFIH PD 135193 to TF, FK 142782 to KP, and FK 134993 to AGS).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated

References

1. Valnet J. The Practice of Aromatherapy: A Classic Compendium of Plant Medicines and Their Healing Properties. Rochester, Vt: Healing Arts Press (1990).

2. Nikkhah Bodagh M, Maleki I, Hekmatdoost A. Ginger in gastrointestinal disorders: A systematic review of clinical trials. *Food Sci Nutr.* (2019) 7:96–108. doi: 10.1002/fsn3.807

3. Safety Assessment of Zingiber officinale (Ginger) . Available online at: https://cir-safety.org/sites/default/files/ginger032021slr.pdf.

4. Assessment report on Zingiber officinale Roscoe, rhizoma . Available online at: https://www.ema.europa.eu/en/documents/herbal-report/final-assessment-report-zingiber-officinale-roscoe-rhizoma_en.pdf.

5. Mao QQ, Xu XY, Cao SY, Gan RY, Corke H, Beta T, et al. Bioactive compounds and bioactivities of ginger (Zingiber officinale roscoe). *Foods*. (2019) 8. doi: 10.3390/ foods8060185

6. Unuofin JO, Masuku NP, Paimo OK, Lebelo SL. Ginger from farmyard to town: nutritional and pharmacological applications. *Front Pharmacol.* (2021) 12:779352. doi: 10.3389/fphar.2021.779352

7. Li LL, Cui Y, Guo XH, Ma K, Tian P, Feng J, et al. Pharmacokinetics and tissue distribution of gingerols and shogaols from ginger (Zingiber officinale rosc.) in rats by UPLC(-)Q-exactive(-)HRMS. *Molecules*. (2019) 24. doi: 10.3390/molecules24030512

8. Qiao Q, Du Q. Preparation of the monomers of gingerols and 6-shogaol by flash high speed counter-current chromatography. *J Chromatogr A*. (2011) 1218:6187–90. doi: 10.1016/j.chroma.2010.12.062

9. Bhattarai S, Tran VH, Duke CC. The stability of gingerol and shogaol in aqueous solutions. J Pharm Sci. (2001) 90:1658-64. doi: 10.1002/jps.1116

10. You H, Ireland B, Moeszinger M, Zhang H, Snow L, Krepich S, et al. Determination of bioactive nonvolatile ginger constituents in dietary supplements by a rapid and economic HPLC method: Analytical method development and single-laboratory validation. *Talanta.* (2019) 194:795–802. doi: 10.1016/j.talanta.2018.10.075

11. Utama-Ang N, Sida S, Wanachantararak P, Kawee-Ai A. Development of edible Thai rice film fortified with ginger extract using microwave-assisted extraction for oral antimicrobial properties. *Sci Rep.* (2021) 11:14870. doi: 10.1038/s41598-021-94430-y

12. Chrubasik S, Pittler MH, Roufogalis BD. Zingiberis rhizoma: a comprehensive review on the ginger effect and efficacy profiles. *Phytomedicine*. (2005) 12:684–701. doi: 10.1016/j.phymed.2004.07.009

13. Ahmed SHH, Gonda T, Agbadua OG, Girst G, Berkecz R, Kusz N, et al. Preparation and evaluation of 6-gingerol derivatives as novel antioxidants and antiplatelet agents. *Antioxidants (Basel)*. (2023) 12. doi: 10.3390/antiox12030744

14. Salmeron-Manzano E, Garrido-Cardenas JA, Manzano-Agugliaro F. Worldwide research trends on medicinal plants. *Int J Environ Res Public Health.* (2020) 17. doi: 10.3390/ijerph17103376

15. Mathai RV, Jindal MK, Mitra JC, Sar SK. COVID-19 and medicinal plants: A critical perspective. *Forensic Sci Int Anim Environ*. (2022) 2:100043. doi: 10.1016/j.fsiae.2022.100043

16. Dugasani S, Pichika MR, Nadarajah VD, Balijepalli MK, Tandra S, Korlakunta JN. Comparative antioxidant and anti-inflammatory effects of [6]-gingerol, [8]-gingerol, [10]-gingerol and [6]-shogaol. *J Ethnopharmacol.* (2010) 127:515–20. doi: 10.1016/j.jep.2009.10.004

17. de Lima RMT, Dos Reis AC, de Menezes APM, Santos JVO, Filho J, Ferreira JRO, et al. Protective and therapeutic potential of ginger (Zingiber officinale) extract and [6]-gingerol in cancer: A comprehensive review. *Phytother Res.* (2018) 32:1885–907. doi: 10.1002/ptr.6134

18. Ma RH, Ni ZJ, Zhu YY, Thakur K, Zhang F, Zhang YY, et al. A recent update on the multifaceted health benefits associated with ginger and its bioactive components. *Food Funct*. (2021) 12:519–42. doi: 10.1039/D0FO02834G

19. Arcusa R, Villano D, Marhuenda J, Cano M, Cerda B, Zafrilla P. Potential role of ginger (Zingiber officinale roscoe) in the prevention of neurodegenerative diseases. *Front Nutr.* (2022) 9:809621. doi: 10.3389/fnut.2022.809621

20. Yucel C, Karatoprak GS, Acikara OB, Akkol EK, Barak TH, Sobarzo-Sanchez E, et al. Immunomodulatory and anti-inflammatory therapeutic potential of gingerols and their nanoformulations. *Front Pharmacol.* (2022) 13:902551. doi: 10.3389/fphar.2022.902551

21. Bischoff-Kont I, Furst R. Benefits of ginger and its constituent 6-shogaol in inhibiting inflammatory processes. *Pharm (Basel)*. (2021) 14. doi: 10.3390/ph14060571

organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

22. Crichton M, Marshall S, Isenring E, Lohning A, McCarthy AL, Molassiotis A, et al. Effect of a standardized ginger root powder regimen on chemotherapy-induced nausea and vomiting: A multicenter, double-blind, placebo-controlled randomized trial. *J Acad Nutr Diet.* (2024) 124:313–30.e6. doi: 10.1016/j.jand.2023.09.003

23. Choi J, Lee J, Kim K, Choi HK, Lee SA, Lee HJ. Effects of ginger intake on chemotherapy-induced nausea and vomiting: A systematic review of randomized clinical trials. *Nutrients.* (2022) 14. doi: 10.3390/nu14234982

24. Aryaeian N, Shahram F, Mahmoudi M, Tavakoli H, Yousefi B, Arablou T, et al. The effect of ginger supplementation on some immunity and inflammation intermediate genes expression in patients with active Rheumatoid Arthritis. *Gene.* (2019) 698:179–85. doi: 10.1016/j.gene.2019.01.048

25. Mozaffari-Khosravi H, Naderi Z, Dehghan A, Nadjarzadeh A, Fallah Huseini H. Effect of ginger supplementation on proinflammatory cytokines in older patients with osteoarthritis: outcomes of a randomized controlled clinical trial. *J Nutr Gerontol Geriatr.* (2016) 35:209–18. doi: 10.1080/21551197.2016.1206762

26. Menon P, Perayil J, Fenol A, Rajan Peter M, Lakshmi P, Suresh R. Effectiveness of ginger on pain following periodontal surgery - A randomized cross-over clinical trial. *J Ayurveda Integr Med.* (2021) 12:65–9. doi: 10.1016/j.jaim.2020.05.003

27. Rayati F, Hajmanouchehri F, Najafi E. Comparison of anti-inflammatory and analgesic effects of Ginger powder and Ibuprofen in postsurgical pain model: A randomized, double-blind, case-control clinical trial. *Dent Res J (Isfahan).* (2017) 14:1–7. doi: 10.4103/1735-3327.201135

28. Rondanelli M, Fossari F, Vecchio V, Gasparri C, Peroni G, Spadaccini D, et al. Clinical trials on pain lowering effect of ginger: A narrative review. *Phytother Res.* (2020) 34:2843–56. doi: 10.1002/ptr.6730

29. Austermann J, Roth J, Barczyk-Kahlert K. The good and the bad: monocytes' and macrophages' Diverse functions in inflammation. *Cells.* (2022) 11. doi: 10.3390/ cells11121979

30. Yang S, Zhao M, Jia S. Macrophage: Key player in the pathogenesis of autoimmune diseases. *Front Immunol.* (2023) 14:1080310. doi: 10.3389/fimmu.2023.1080310

31. Ippoushi K, Azuma K, Ito H, Horie H, Higashio H. [6]-Gingerol inhibits nitric oxide synthesis in activated J774.1 mouse macrophages and prevents peroxynitrite-induced oxidation and nitration reactions. *Life Sci.* (2003) 73:3427–37. doi: 10.1016/j.lfs.2003.06.022

32. Tripathi S, Maier KG, Bruch D, Kittur DS. Effect of 6-gingerol on proinflammatory cytokine production and costimulatory molecule expression in murine peritoneal macrophages. *J Surg Res.* (2007) 138:209–13. doi: 10.1016/j.jss.2006.07.051

33. Lantz RC, Chen GJ, Sarihan M, Solyom AM, Jolad SD, Timmermann BN. The effect of extracts from ginger rhizome on inflammatory mediator production. *Phytomedicine*. (2007) 14:123–8. doi: 10.1016/j.phymed.2006.03.003

34. Pan MH, Hsieh MC, Hsu PC, Ho SY, Lai CS, Wu H, et al. 6-Shogaol suppressed lipopolysaccharide-induced up-expression of iNOS and COX-2 in murine macrophages. *Mol Nutr Food Res.* (2008) 52:1467–77. doi: 10.1002/mnfr.200700515

35. Lee TY, Lee KC, Chen SY, Chang HH. 6-Gingerol inhibits ROS and iNOS through the suppression of PKC-alpha and NF-kappaB pathways in lipopolysaccharide-stimulated mouse macrophages. *Biochem Biophys Res Commun.* (2009) 382:134–9. doi: 10.1016/j.bbrc.2009.02.160

36. Ha SK, Moon E, Ju MS, Kim DH, Ryu JH, Oh MS, et al. 6-Shogaol, a ginger product, modulates neuroinflammation: a new approach to neuroprotection. *Neuropharmacology*. (2012) 63:211–23. doi: 10.1016/j.neuropharm.2012.03.016

37. Han Q, Yuan Q, Meng X, Huo J, Bao Y, Xie G. 6-Shogaol attenuates LPS-induced inflammation in BV2 microglia cells by activating PPAR-gamma. *Oncotarget.* (2017) 8:42001–6. doi: 10.18632/oncotarget.v8i26

38. Liu Y, Deng S, Zhang Z, Gu Y, Xia S, Bao X, et al. 6-Gingerol attenuates microglia-mediated neuroinflammation and ischemic brain injuries through AktmTOR-STAT3 signaling pathway. *Eur J Pharmacol.* (2020) 883:173294. doi: 10.1016/ j.ejphar.2020.173294

39. Balic JJ, Albargy H, Luu K, Kirby FJ, Jayasekara WSN, Mansell F, et al. STAT3 serine phosphorylation is required for TLR4 metabolic reprogramming and IL-1beta expression. *Nat Commun.* (2020) 11:3816. doi: 10.1038/s41467-020-17669-5

40. Hwang YH, Kim T, Kim R, Ha H. The natural product 6-gingerol inhibits inflammation-associated osteoclast differentiation via reduction of prostaglandin E(2) levels. *Int J Mol Sci.* (2018) 19. doi: 10.3390/ijms19072068

41. Ho SC, Chang YH. Comparison of inhibitory capacities of 6-, 8- and 10gingerols/shogaols on the canonical NLRP3 inflammasome-mediated IL-1beta secretion. *Molecules*. (2018) 23. doi: 10.3390/molecules23020466

42. Zhang FL, Zhou BW, Yan ZZ, Zhao J, Zhao BC, Liu WF, et al. 6-Gingerol attenuates macrophages pyroptosis via the inhibition of MAPK signaling pathways and predicts a good prognosis in sepsis. *Cytokine*. (2020) 125:154854. doi: 10.1016/j.cyto.2019.154854

43. Pan Q, Liu P, Wan M. 6-Gingerol attenuates sepsis-induced acute lung injury by suppressing NLRP3 inflammasome through NRF2 activation. *Folia Histochem Cytobiol.* (2023) 61:68–80. doi: 10.5603/FHC.a2023.0002

44. Wu Y, Sun X, Li H, Chu X, Xue Y, Qi J, et al. 6-Gingerol attenuates arsenic trioxide-induced liver injury by inhibiting pyroptosis and ROS-NLRP3 inflammatory signaling pathway: Based on network pharmacology analysis and experiment verification. *J Funct Foods.* (2023) 105:105551. doi: 10.1016/j.jff.2023.105551

45. Masucci MT, Minopoli M, Del Vecchio S, Carriero MV. The emerging role of neutrophil extracellular traps (NETs) in tumor progression and metastasis. *Front Immunol.* (2020) 11:1749. doi: 10.3389/fimmu.2020.01749

46. Lee KH, Kronbichler A, Park DD, Park Y, Moon H, Kim H, et al. Neutrophil extracellular traps (NETs) in autoimmune diseases: A comprehensive review. *Autoimmun Rev.* (2017) 16:1160–73. doi: 10.1016/j.autrev.2017.09.012

47. Ali RA, Gandhi AA, Dai L, Weiner J, Estes SK, Yalavarthi S, et al. Antineutrophil properties of natural gingerols in models of lupus. *JCI Insight.* (2021) 6. doi: 10.1172/ jci.insight.138385

48. Tavares LP, Negreiros-Lima GL, Lima KM, PMR ES, Pinho V, Teixeira MM, et al. Blame the signaling: Role of cAMP for the resolution of inflammation. *Pharmacol Res.* (2020) 159:105030. doi: 10.1016/j.phrs.2020.105030

49. Zhu Y, Wang D, Luo J, Jie J, Liu H, Peng L, et al. Zingerone Inhibits the Neutrophil Extracellular Trap Formation and Protects against Sepsis via NRF2-Mediated ROS Inhibition. *Oxid Med Cell Longev.* (2022) 2022:3990607. doi: 10.1155/2022/3990607

50. Andersen G, Kahlenberg K, Krautwurst D, Somoza V. [6]-gingerol facilitates CXCL8 secretion and ROS production in primary human neutrophils by targeting the TRPV1 channel. *Mol Nutr Food Res.* (2023) 67:e2200434. doi: 10.1002/mnfr.202200434

51. Ganguly D, Haak S, Sisirak V, Reizis B. The role of dendritic cells in autoimmunity. *Nat Rev Immunol.* (2013) 13:566–77. doi: 10.1038/nri3477

52. Coutant F, Miossec P. Altered dendritic cell functions in autoimmune diseases: distinct and overlapping profiles. *Nat Rev Rheumatol.* (2016) 12:703–15. doi: 10.1038/ nrrheum.2016.147

53. Han JJ, Li X, Ye ZQ, Lu XY, Yang T, Tian J, et al. Treatment with 6-gingerol regulates dendritic cell activity and ameliorates the severity of experimental autoimmune encephalomyelitis. *Mol Nutr Food Res.* (2019) 63:e1801356. doi: 10.1002/mnfr.201801356

54. Pazmandi K, Agics B, Szollosi AG, Bacsi A, Fekete T. Ginger-derived bioactive compounds attenuate the Toll-like receptor mediated responses of human dendritic cells. *Eur J Pharmacol.* (2024) 967:176399. doi: 10.1016/j.ejphar.2024.176399

55. Kumar BV, Connors TJ, Farber DL. Human T cell development, localization, and function throughout life. *Immunity*. (2018) 48:202-13. doi: 10.1016/j.immuni.2018.01.007

56. Sun L, Su Y, Jiao A, Wang X, Zhang B. T cells in health and disease. Signal Transduct Target Ther. (2023) 8:235. doi: 10.1038/s41392-023-01471-y

57. Bernard M, Furlong SJ, Power Coombs MR, Hoskin DW. Differential inhibition of T lymphocyte proliferation and cytokine synthesis by [6]-gingerol, [8]-gingerol, and [10]-gingerol. *Phytother Res.* (2015) 29:1707–13. doi: 10.1002/ptr.5414

58. Kawamoto Y, Ueno Y, Nakahashi E, Obayashi M, Sugihara K, Qiao S, et al. Prevention of allergic rhinitis by ginger and the molecular basis of immunosuppression by 6-gingerol through T cell inactivation. *J Nutr Biochem.* (2016) 27:112–22. doi: 10.1016/j.jnutbio.2015.08.025

59. Deng X, Chen D, Sun X, Dong J, Huang J. Effects of ginger extract and its major component 6-gingerol on anti-tumor property through mitochondrial biogenesis in CD8(+) T cells. *J Food Sci.* (2022) 87:3307–17. doi: 10.1111/1750-3841.16228

60. Ju SA, Park SM, Lee YS, Bae JH, Yu R, An WG, et al. Administration of 6gingerol greatly enhances the number of tumor-infiltrating lymphocytes in murine tumors. *Int J Cancer.* (2012) 130:2618–28. doi: 10.1002/ijc.26316

61. Chatupheeraphat C, Nantasenamat C, Deesrisak K, Roytrakul S, Anurathapan U, Tanyong D. Bioinformatics and experimental studies of anti-leukemic activity from 6-gingerol demonstrate its role in p53 mediated apoptosis pathway. *EXCLI J.* (2020) 19:582–95. doi: 10.17179/excli2019-2008

62. Najafi Dorcheh S, Rahgozar S, Talei D. 6-Shogaol induces apoptosis in acute lymphoblastic leukaemia cells by targeting p53 signalling pathway and generation of reactive oxygen species. *J Cell Mol Med.* (2021) 25:6148–60. doi: 10.1111/jcmm.16528

63. Havnaer A, Han G. Autoinflammatory disorders: A review and update on pathogenesis and treatment. *Am J Clin Dermatol.* (2019) 20:539-64. doi: 10.1007/s40257-019-00440-y

64. Rodrigues FAP, Santos A, de Medeiros P, Prata MMG, Santos TCS, da Silva JA, et al. Gingerol suppresses sepsis-induced acute kidney injury by modulating methylsulfonylmethane and dimethylamine production. *Sci Rep.* (2018) 8:12154. doi: 10.1038/s41598-018-30522-6

65. Hong MK, Hu LL, Zhang YX, Xu YL, Liu XY, He PK, et al. 6-Gingerol ameliorates sepsis-induced liver injury through the NRF2 pathway. *Int Immunopharmacol.* (2020) 80:106196. doi: 10.1016/j.intimp.2020.106196

66. Padoan A, Musso G, Contran N, Basso D. Inflammation, autoinflammation and autoimmunity in inflammatory bowel diseases. *Curr Issues Mol Biol.* (2023) 45:5534–57. doi: 10.3390/cimb45070350

67. Zhang F, Ma N, Gao YF, Sun LL, Zhang JG. Therapeutic effects of 6-gingerol, 8-gingerol, and 10-gingerol on dextran sulfate sodium-induced acute ulcerative colitis in rats. *Phytother Res.* (2017) 31:1427–32. doi: 10.1002/ptr.5871

68. Sheng Y, Wu T, Dai Y, Xu L, Zhong Y, Xue Y, et al. 6-gingerol alleviates inflammatory injury in DSS-induced ulcerative colitis mice by regulating NF-kappaB signaling. *Ann Palliat Med.* (2020) 9:1944–52. doi: 10.21037/apm

69. Zhang M, Xu C, Liu D, Han MK, Wang L, Merlin D. Oral delivery of nanoparticles loaded with ginger active compound, 6-shogaol, attenuates ulcerative colitis and promotes wound healing in a murine model of ulcerative colitis. *J Crohns Colitis.* (2018) 12:217–29. doi: 10.1093/ecco-jcc/jjx115

70. Kim HR, Noh EM, Kim SY. Anti-inflammatory effect and signaling mechanism of 8-shogaol and 10-shogaol in a dextran sodium sulfate-induced colitis mouse model. *Heliyon*. (2023) 9:e12778. doi: 10.1016/j.heliyon.2022.e12778

71. Stampanoni Bassi M, Iezzi E, Centonze D. Multiple sclerosis: Inflammation, autoimmunity and plasticity. *Handb Clin Neurol.* (2022) 184:457–70. doi: 10.1016/B978-0-12-819410-2.00024-2

72. Sapkota A, Park SJ, Choi JW. Neuroprotective effects of 6-shogaol and its metabolite, 6-paradol, in a mouse model of multiple sclerosis. *Biomol Ther (Seoul)*. (2019) 27:152–9. doi: 10.4062/biomolther.2018.089

73. Mukkavilli R, Yang C, Tanwar RS, Saxena R, Gundala SR, Zhang Y, et al. Pharmacokinetic-pharmacodynamic correlations in the development of ginger extract as an anticancer agent. *Sci Rep.* (2018) 8:3056. doi: 10.1038/s41598-018-21125-2

74. Zick SM, Djuric Z, Ruffin MT, Litzinger AJ, Normolle DP, Alrawi S, et al. Pharmacokinetics of 6-gingerol, 8-gingerol, 10-gingerol, and 6-shogaol and conjugate metabolites in healthy human subjects. *Cancer Epidemiol Biomarkers Prev.* (2008) 17:1930–6. doi: 10.1158/1055-9965.EPI-07-2934

75. Miossec P. Local and systemic effects of IL-17 in joint inflammation: a historical perspective from discovery to targeting. *Cell Mol Immunol.* (2021) 18:860–5. doi: 10.1038/s41423-021-00644-5

76. Jo S, Samarpita S, Lee JS, Lee YJ, Son JE, Jeong M, et al. 8-Shogaol inhibits rheumatoid arthritis through targeting TAK1. *Pharmacol Res.* (2022) 178:106176. doi: 10.1016/j.phrs.2022.106176

77. Jeon SM. Regulation and function of AMPK in physiology and diseases. *Exp Mol Med.* (2016) 48:e245. doi: 10.1038/emm.2016.81

78. Cork GK, Thompson J, Slawson C. Real talk: the inter-play between the mTOR, AMPK, and hexosamine biosynthetic pathways in cell signaling. *Front Endocrinol (Lausanne).* (2018) 9:522. doi: 10.3389/fendo.2018.00522

79. Suto T, Karonitsch T. The immunobiology of mTOR in autoimmunity. J Autoimmun. (2020) 110:102373. doi: 10.1016/j.jaut.2019.102373

80. Keerthana CK, Rayginia TP, Shifana SC, Anto NP, Kalimuthu K, Isakov N, et al. The role of AMPK in cancer metabolism and its impact on the immunomodulation of the tumor microenvironment. *Front Immunol.* (2023) 14:1114582. doi: 10.3389/fimmu.2023.1114582

81. Luettig J, Rosenthal R, Lee IM, Krug SM, Schulzke JD. The ginger component 6shogaol prevents TNF-alpha-induced barrier loss via inhibition of PI3K/Akt and NFkappaB signaling. *Mol Nutr Food Res.* (2016) 60:2576–86. doi: 10.1002/mnfr.201600274

82. Hashem RM, Rashed LA, Hassanin KMA, Hetta MH, Ahmed AO. Effect of 6gingerol on AMPK- NF-kappaB axis in high fat diet fed rats. *BioMed Pharmacother*. (2017) 88:293–301. doi: 10.1016/j.biopha.2017.01.035

83. Liu Y, Li D, Wang S, Peng Z, Tan Q, He Q, et al. 6-gingerol ameliorates hepatic steatosis, inflammation and oxidative stress in high-fat diet-fed mice through activating LKB1/AMPK signaling. *Int J Mol Sci.* (2023) 24. doi: 10.3390/ijms24076285

84. Wei CK, Tsai YH, Korinek M, Hung PH, El-Shazly M, Cheng YB, et al. 6-paradol and 6-shogaol, the pungent compounds of ginger, promote glucose utilization in adipocytes and myotubes, and 6-paradol reduces blood glucose in high-fat diet-fed mice. *Int J Mol Sci.* (2017) 18. doi: 10.3390/ijms18010168

85. Zhang H, Kim E, Yi J, Hai H, Kim H, Park S, et al. [6]-gingerol suppresses oral cancer cell growth by inducing the activation of AMPK and suppressing the AKT/ mTOR signaling pathway. *In Vivo.* (2021) 35:3193–201. doi: 10.21873/invivo.12614

86. Deng B, Jiang XL, Xu YC, Chen S, Cai M, Deng SH, et al. 10-Gingerol, a natural AMPK agonist, suppresses neointimal hyperplasia and inhibits vascular smooth muscle cell proliferation. *Food Funct.* (2022) 13:3234–46. doi: 10.1039/D1FO03610F

87. Saha S, Buttari B, Panieri E, Profumo E, Saso L. An overview of NRF2 signaling pathway and its role in inflammation. *Molecules*. (2020) 25. doi: 10.3390/molecules25225474

88. Campbell NK, Fitzgerald HK, Dunne A. Regulation of inflammation by the antioxidant haem oxygenase 1. *Nat Rev Immunol.* (2021) 21:411–25. doi: 10.1038/s41577-020-00491-x

89. Dennery PA. Signaling function of heme oxygenase proteins. *Antioxid Redox Signal.* (2014) 20:1743–53. doi: 10.1089/ars.2013.5674

90. Ohnishi M, Ohshita M, Tamaki H, Marutani Y, Nakayama Y, Akagi M, et al. Shogaol but not gingerol has a neuroprotective effect on hemorrhagic brain injury: Contribution of the alpha, beta-unsaturated carbonyl to heme oxygenase-1 expression. *Eur J Pharmacol.* (2019) 842:33–9. doi: 10.1016/j.ejphar.2018.10.029

91. Han SJ, Kim M, D'Agati VD, Lee HT. 6-Shogaol protects against ischemic acute kidney injury by modulating NF-kappaB and heme oxygenase-1 pathways. *Am J Physiol Renal Physiol.* (2019) 317:F743–F56. doi: 10.1152/ajprenal.00182.2019

92. Chen F, Tang Y, Sun Y, Veeraraghavan VP, Mohan SK, Cui C. 6-shogaol, a active constiuents of ginger prevents UVB radiation mediated inflammation and oxidative stress through modulating NRF2 signaling in human epidermal keratinocytes (HaCaT cells). *J Photochem Photobiol B.* (2019) 197:111518. doi: 10.1016/j.jphotobiol.2019.111518

93. Bischoff-Kont I, Primke T, Niebergall LS, Zech T, Furst R. Ginger constituent 6shogaol inhibits inflammation- and angiogenesis-related cell functions in primary human endothelial cells. *Front Pharmacol.* (2022) 13:844767. doi: 10.3389/fphar.2022.844767

94. Sun Y, Ren J, Wang F. [6]-Gingerol impedes 7,12-dimethylbenz(a)anthraceneinduced inflammation and cell proliferation-associated hamster buccal pouch carcinogenesis through modulating NRF2 signaling events. *J Biochem Mol Toxicol.* (2021) 35:e22689. doi: 10.1002/jbt.22689

95. Wu S, Zhu J, Wu G, Hu Z, Ying P, Bao Z, et al. 6-gingerol alleviates ferroptosis and inflammation of diabetic cardiomyopathy via the NRF2/HO-1 pathway. *Oxid Med Cell Longev*. (2022) 2022:3027514. doi: 10.1155/2022/3027514

96. Matzinger M, Fischhuber K, Poloske D, Mechtler K, Heiss EH. AMPK leads to phosphorylation of the transcription factor NRF2, tuning transactivation of selected target genes. *Redox Biol.* (2020) 29:101393. doi: 10.1016/j.redox.2019.101393

97. Zimmermann K, Baldinger J, Mayerhofer B, Atanasov AG, Dirsch VM, Heiss EH. Activated AMPK boosts the NRF2/HO-1 signaling axis-A role for the unfolded protein response. *Free Radic Biol Med.* (2015) 88:417–26. doi: 10.1016/j.freeradbiomed.2015.03.030

98. Xing J, Weng L, Yuan B, Wang Z, Jia L, Jin R, et al. Identification of a role for TRIM29 in the control of innate immunity in the respiratory tract. *Nat Immunol.* (2016) 17:1373–80. doi: 10.1038/ni.3580

99. Wang J, Lu W, Zhang J, Du Y, Fang M, Zhang A, et al. Loss of TRIM29 mitigates viral myocarditis by attenuating PERK-driven ER stress response in male mice. *Nat Commun.* (2024) 15:3481. doi: 10.1038/s41467-024-44745-x

100. Purohit V, Wang L, Yang H, Li J, Ney GM, Gumkowski ER, et al. ATDC binds to KEAP1 to drive NRF2-mediated tumorigenesis and chemoresistance in pancreatic cancer. *Genes Dev.* (2021) 35:218–33. doi: 10.1101/gad.344184.120

101. Korbecki J, Bobinski R, Dutka M. Self-regulation of the inflammatory response by peroxisome proliferator-activated receptors. *Inflammation Res.* (2019) 68:443–58. doi: 10.1007/s00011-019-01231-1

102. Venkataraman B, Ojha S, Belur PD, Bhongade B, Raj V, Collin PD, et al. Phytochemical drug candidates for the modulation of peroxisome proliferatoractivated receptor gamma in inflammatory bowel diseases. *Phytother Res.* (2020) 34:1530–49. doi: 10.1002/ptr.6625

103. Hong W, Zhi FX, Kun TH, Hua FJ, Huan Ling L, Fang F, et al. 6-Gingerol attenuates ventilator-induced lung injury via anti-inflammation and antioxidative stress by modulating the PPARgamma/NF-kappaBsignalling pathway in rats. *Int Immunopharmacol.* (2021) 92:107367. doi: 10.1016/j.intimp.2021.107367

104. Erin N, Szallasi A. Carcinogenesis and metastasis: focus on TRPV1-positive neurons and immune cells. *Biomolecules*. (2023) 13. doi: 10.3390/biom13060983

105. Basu S, Srivastava P. Immunological role of neuronal receptor vanilloid receptor 1 expressed on dendritic cells. *Proc Natl Acad Sci U.S.A.* (2005) 102:5120–5. doi: 10.1073/pnas.0407780102

106. Toth BI, Benko S, Szollosi AG, Kovacs L, Rajnavolgyi E, Biro T. Transient receptor potential vanilloid-1 signaling inhibits differentiation and activation of human dendritic cells. *FEBS Lett.* (2009) 583:1619–24. doi: 10.1016/j.febslet.2009.04.031

107. Szallasi A, Blumberg PM. Resiniferatoxin and its analogs provide novel insights into the pharmacology of the vanilloid (capsaicin) receptor. Life Sci. (1990) 47:1399–408. doi: 10.1016/0024-3205(90)90518-V

108. Dedov VN, Tran VH, Duke CC, Connor M, Christie MJ, Mandadi S, et al. Gingerols: a novel class of vanilloid receptor (VR1) agonists. *Br J Pharmacol.* (2002) 137:793–8. doi: 10.1038/sj.bjp.0704925

109. Ohbuchi K, Mori Y, Ogawa K, Warabi E, Yamamoto M, Hirokawa T. Detailed analysis of the binding mode of vanilloids to transient receptor potential vanilloid type I (TRPV1) by a mutational and computational study. *PloS One.* (2016) 11:e0162543. doi: 10.1371/journal.pone.0162543

110. Yin Y, Dong Y, Vu S, Yang F, Yarov-Yarovoy V, Tian Y, et al. Structural mechanisms underlying activation of TRPV1 channels by pungent compounds in gingers. *Br J Pharmacol.* (2019) 176:3364–77. doi: 10.1111/bph.14766

111. Luo J, Chen J, Yang C, Tan J, Zhao J, Jiang N, et al. 6-Gingerol protects against cerebral ischemia/reperfusion injury by inhibiting NLRP3 inflammasome and apoptosis via TRPV1/FAF1 complex dissociation-mediated autophagy. *Int Immunopharmacol.* (2021) 100:108146. doi: 10.1016/j.intimp.2021.108146

112. Schepetkin IA, Kushnarenko SV, Ozek G, Kirpotina LN, Sinharoy P, Utegenova GA, et al. Modulation of human neutrophil responses by the essential oils from ferula akitschkensis and their constituents. *J Agric Food Chem.* (2016) 64:7156–70. doi: 10.1021/acs.jafc.6b03205

113. Rudrapal M, Eltayeb WA, Rakshit G, El-Arabey AA, Khan J, Aldosari SM, et al. Dual synergistic inhibition of COX and LOX by potential chemicals from Indian daily spices investigated through detailed computational studies. *Sci Rep.* (2023) 13:8656. doi: 10.1038/s41598-023-35161-0

114. Mukherjee S, Mandal N, Dey A, Mondal B. An approach towards optimization of the extraction of polyphenolic antioxidants from ginger (Zingiber officinale). *J Food Sci Technol.* (2014) 51:3301–8. doi: 10.1007/s13197-012-0848-z

115. Tohma H, Gülçin I, Bursal E, Gören AC, Alwasel SH, Köksal E. Antioxidant activity and phenolic compounds of ginger (Rosc.) determined by HPLC-MS/MS. J Food Meas Charact. (2017) 11:556–66. doi: 10.1007/s11694-016-9423-z

116. Yu Y, Zick S, Li X, Zou P, Wright B, Sun D. Examination of the pharmacokinetics of active ingredients of ginger in humans. *AAPS J.* (2011) 13:417–26. doi: 10.1208/s12248-011-9286-5

117. Mu J, Zhuang X, Wang Q, Jiang H, Deng ZB, Wang B, et al. Interspecies communication between plant and mouse gut host cells through edible plant derived exosome-like nanoparticles. *Mol Nutr Food Res.* (2014) 58:1561–73. doi: 10.1002/mnfr.201300729

118. Nair HB, Sung B, Yadav VR, Kannappan R, Chaturvedi MM, Aggarwal BB. Delivery of antiinflammatory nutraceuticals by nanoparticles for the prevention and treatment of cancer. *Biochem Pharmacol.* (2010) 80:1833–43. doi: 10.1016/j.bcp.2010.07.021

119. Sumithaa C, Manjunathan T, Mazuryk O, Peters S, Pillai RS, Brindell M, et al. Nanoencapsulation of ru(p-cymene) complex bearing ginger-based natural product into liposomal nanoformulation to improve its cellular uptake and antiproliferative activity. ACS Appl Bio Mater. (2022) 5:3241–56. doi: 10.1021/acsabm.2c00231

120. Amorndoljai P, Taneepanichskul S, Niempoog S, Nimmannit U. A comparative of ginger extract in nanostructure lipid carrier (NLC) and 1% Diclofenac gel for treatment of knee osteoarthritis (OA). *J Med Assoc Thai*. (2017) 100:447–56.