



OPEN ACCESS

EDITED BY

Mauricio César De Marzi,
National University of Luján, Argentina

REVIEWED BY

Alexandr Ceasovschi,
Grigore T. Popa University of Medicine and
Pharmacy, Romania
Magdalena Beata Skarżyńska,
Medical University of Warsaw, Poland

*CORRESPONDENCE

Xiaoyun Zhao

✉ zxydoctor@163.com

RECEIVED 05 March 2024

ACCEPTED 10 May 2024

PUBLISHED 23 May 2024

CITATION

Chen Z, Suo Y, Du X and Zhao X (2024)
Genetically predicted N-
methylhydroxyproline levels mediate the
association between naive CD8+ T cells and
allergic rhinitis: a mediation Mendelian
randomization study.
Front. Immunol. 15:1396246.
doi: 10.3389/fimmu.2024.1396246

COPYRIGHT

© 2024 Chen, Suo, Du and Zhao. This is an
open-access article distributed under the terms
of the [Creative Commons Attribution License
\(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use, distribution or reproduction
in other forums is permitted, provided the
original author(s) and the copyright owner(s)
are credited and that the original publication
in this journal is cited, in accordance with
accepted academic practice. No use,
distribution or reproduction is permitted
which does not comply with these terms.

Genetically predicted N-methylhydroxyproline levels mediate the association between naive CD8+ T cells and allergic rhinitis: a mediation Mendelian randomization study

Zhengjie Chen^{1,2}, Ying Suo^{1,2}, Xintao Du^{1,2}
and Xiaoyun Zhao^{1,2,3,4*}

¹Clinical School of Thoracic, Tianjin Medical University, Tianjin, China, ²Department of Respiratory and Critical Care Medicine, Tianjin Chest Hospital, Tianjin, China, ³Department of Respiratory and Critical Care Medicine, Chest Hospital of Tianjin University, Tianjin, China, ⁴DeepinBreath Union Laboratory, Tianjin Chest Hospital, Tianjin, China

Background: Allergic rhinitis (AR), a prevalent chronic inflammatory condition triggered by immunoglobulin E (IgE), involves pivotal roles of immune and metabolic factors in its onset and progression. However, the intricacies and uncertainties in clinical research render current investigations into their interplay somewhat inadequate.

Objective: To elucidate the causal relationships between immune cells, metabolites, and AR, we conducted a mediation Mendelian randomization (MR) analysis.

Methods: Leveraging comprehensive publicly accessible summary-level data from genome-wide association studies (GWAS), this study employed the two-sample MR research method to investigate causal relationships among 731 immune cell phenotypes, 1400 metabolite levels, and AR. Additionally, employing the mediation MR approach, the study analyzed potential mediated effect of metabolites in the relationships between immune cells and AR. Various sensitivity analysis methods were systematically employed to ensure the robustness of the results.

Results: Following false discovery rate (FDR) correction, we identified three immune cell phenotypes as protective factors for AR: Naive CD8br %CD8br (odds ratio (OR): 0.978, 95% CI = 0.966–0.990, $P = 4.5 \times 10^{-4}$), CD3 on CD39+ activated Treg (OR: 0.947, 95% CI = 0.923–0.972, $P = 3 \times 10^{-5}$), HVEM on CD45RA-CD4+ (OR: 0.967, 95% CI = 0.948–0.986, $P = 4 \times 10^{-5}$). Additionally, three metabolite levels were identified as risk factors for AR: N-methylhydroxyproline levels (OR: 1.219, 95% CI = 1.104–1.346, $P = 9 \times 10^{-5}$), N-acetylneuraminic acid levels (OR: 1.133, 95% CI = 1.061–1.211, $P = 1.7 \times 10^{-4}$), 1-stearoyl-2-arachidonoyl-glycerol (18:0/20:4) levels (OR: 1.058, 95% CI = 1.029–1.087, $P = 5 \times 10^{-5}$). Mediation MR analysis indicated a causal relationship between Naive CD8br %CD8br and N-methylhydroxyproline levels, acting as a protective factor (OR: 0.971, 95% CI = 0.950–0.992, $P = 8.31 \times 10^{-3}$). The mediated effect was -0.00574, accounting for

26.1% of the total effect, with a direct effect of -0.01626 . Naive CD8+ T cells exert a protective effect on AR by reducing N-methylhydroxyproline levels.

Conclusion: Our study, delving into genetic information, has substantiated the intricate connection between immune cell phenotypes and metabolite levels with AR. This reveals a potential pathway to prevent the onset of AR, providing guiding directions for future clinical investigations.

KEYWORDS

allergic rhinitis, immunity, metabolites, causal inference, Mendelian randomization, mediation

1 Introduction

Allergic rhinitis (AR), a prevalent chronic inflammatory condition triggered by immunoglobulin E (IgE) in response to inhaled allergens (1), manifests widely across age groups, with estimated prevalence rates in European populations ranging from 17% to 28.5% (2, 3). Moreover, its incidence has steadily increased in recent years (4–6), imposing substantial health and psychological burdens due to its seasonal and recurrent nature.

Long-term exposure to allergens may compromise the airway epithelial cells' barrier function, inducing inflammatory responses in the airways and triggering the onset of AR (7). Elevated levels of group 2 innate lymphoid cells (ILC2s) have been observed in AR patients (8), while myeloid dendritic cells (mDCs) are capable of secreting IL-33 to activate ILC2s via the IL-33/ST2 pathway (9). Additionally, IL-25, IL-33, and thymic stromal lymphopoietin (TSLP) can individually or collectively stimulate ILC2s to produce IL-4, IL-5, and IL-13, initiating immune responses that may manifest as inflammation and allergic symptoms in nasal mucosal tissues, such as congestion and rhinorrhea (10). Moreover, memory-type pathogenic Th2 cells, detected in the peripheral blood of symptomatic AR patients, exhibit heightened IL-5 and IL-9 levels compared to conventional Th2 cells (11), potentially correlating with clinical symptoms (12). Furthermore, a circulating memory B cell subset expressing high levels of CD23 correlates with allergen-specific IgE levels and symptom severity in AR patients (13). Dendritic cells, as primary antigen-presenting cells, uptake allergens and present them to CD4+ T cells, provoking allergen-induced inflammation (14, 15). Thus, allergic rhinitis, characterized as a chronic hypersensitivity disorder, involves diverse immune cells and cytokine-mediated responses in its progression (16). Elucidating these possible pathways is crucial for advancing AR treatment.

With the continuous advancement of high-throughput sequencing technology, there has been a more profound exploration of immune cell functions. Immune cells undergo intricate genetic regulatory processes, and a deeper understanding of these mechanisms holds the potential to unveil the mysteries of

the immune system. Identifying and targeting specific points can enhance drug design and development, offering novel perspectives for the treatment and prevention of AR (17).

Allergen Immunotherapy (AIT) induces allergen tolerance by disrupting immune pathogenic mechanisms of allergic reactions and is currently considered an effective treatment for AR (18). Retrospective cohort studies have demonstrated that AIT can reduce the incidence of asthma attacks and pneumonia in AR patients (19). Over the years, various treatment forms have been developed for different patients, including Subcutaneous Immunotherapy (SCIT), Sublingual Immunotherapy (SLIT), and Intralymphatic Immunotherapy (ILIT) (20). However, owing to the complexity and uncertainty of clinical studies, our understanding of the interaction between the immune system and AR remains somewhat limited. Convincing evidence regarding the regulatory effects of AIT on immune cells in the human body is lacking due to variations in race, region, and individual factors. This may be attributed to insufficient sample sizes in real-world studies, divergent study designs, and challenges in eliminating confounding factors. Given that AIT is an individualized treatment approach based on the clinical and immunological characteristics of patients, identifying and validating biomarkers as treatment targets for AR has become a crucial direction in current AIT research (21).

With the disclosure of phenotype-related genetic information, a novel scientific research method has emerged—Mendelian randomization (MR). In recent years, MR has gained popularity as a research approach that employs genetic information loci strongly correlated with exposure factors as instrumental variables (IVs) to infer causal relationships between exposure factors and study outcomes. The method is grounded in Mendel's laws of inheritance, where alleles are randomly allocated to offspring during meiosis, largely mitigating the influence of confounding factors and reverse causation (22). Immune cells play a pivotal role in the onset and progression of AR. However, current research has only addressed a fraction of them. Due to the intricate biological mechanisms of the immune system, the precise roles of many immune cells remain incompletely understood. Hence, we

conducted a comprehensive causal association analysis between immune cell phenotypes and AR through MR, leveraging extensive genome-wide association studies (GWAS) summary-level data of immune cell phenotypes. The goal is to reveal the intricate relationship between immune cell phenotypes and AR.

Employing a two-sample MR approach, we conducted a study investigating the causal relationships involving 731 immune cell phenotypes and AR. Additionally, recognizing the involvement of metabolite levels in the onset and progression of AR, we further evaluated the causal associations between 1400 blood metabolite levels and AR. As part of the MR research, mediation MR can assess the effects of intermediate factors between exposure and outcome, offering insights into whether exposure factors exert their effects through these intermediaries (23). Consequently, we also scrutinized the mediated effect of blood metabolite levels in the relationship between immune cell phenotypes and AR. It is imperative to emphasize that for immune cell phenotypes with a causal relationship to AR, there should be no reverse causation. Accordingly, for positive results, we conducted an MR analysis of AR and immune cell phenotypes to eliminate reverse causation.

2 Materials and methods

2.1 MR assumptions

It is essential to clarify that MR relies on three fundamental assumptions: (1) The assumption of association, suggesting a robust correlation between the selected genetic variations serving as IVs and the exposure factors. (2) The assumption of independence, stating that genetic variations are unrelated to confounding factors. (3) The assumption of exclusivity, proposing that genetic variations can solely influence the outcome through exposure (24). The study adheres to the STROBE-MR statement (25). Furthermore, the GWAS summary-level data used in this research are publicly accessible, and the ethics committees of each institutional review board granted written informed consent from all participants in individual studies. Since this study involves a secondary analysis of previously published data, ethical approval is considered not required. Significantly, there was no observed overlap, as samples of AR, immune cells, and metabolites originated from distinct consortia.

2.2 GWAS data sources for AR

Summary-level data regarding AR were extracted from a comprehensive European population-wide GWAS conducted within the FinnGen project in Finland (26). We specifically selected the most recent R10 version summary data on AR, which encompasses 12,240 cases and 392,069 controls. The diagnosis of AR relied on the International Classification of Diseases codes, specifically ICD-10 (J30.1-J30.4) and ICD-9 (477). Additional details can be accessed at (https://www.finnngen.fi/en/access_results).

2.3 Immune cells GWAS data sources

A genetic variation analysis was conducted on 731 immune cell phenotypes within a cohort of 3,757 Sardinian individuals. This analysis covered 118 absolute cell counts (AC), 389 median fluorescence intensities (MFI) representing surface antigen levels, 32 morphological features (MP), and 192 relative cell counts (RC) (17). The GWAS data for these phenotypes can be accessed on the IEU Open GWAS project (<https://gwas.mrcieu.ac.uk>) through identifiers (ebi-a-GCST0001391 to ebi-a-GCST90002121).

2.4 Metabolites GWAS data sources

In another investigation with 8,299 participants from the Canadian Longitudinal Study on Aging (CLSA) cohort, researchers performed a GWAS on 1,091 blood metabolites and 309 metabolite ratios (27). This study yields vital insights into elucidating the genetic architecture of metabolites. The summary data for 1,400 metabolite levels are available in the GWAS Catalog (<https://www.ebi.ac.uk/gwas>) under identifiers (GCST90199621 to GCST90201020).

2.5 Selection of IVs

In MR analysis, we employed single nucleotide polymorphisms (SNPs) strongly correlated with the exposure as IVs. Recent studies revealed a limited number of SNPs strongly associated with immune cell phenotypes and blood metabolite levels using a genome-wide significance threshold of $P < 5 \times 10^{-8}$. Therefore, drawing on prior research, we adopted a less stringent threshold ($P < 1 \times 10^{-5}$) for SNPs selection (28, 29). Linkage disequilibrium was eliminated within a genetic distance of $\text{kb} = 10,000$ (correlation coefficient $r^2 < 0.001$) to ensure SNP independence. In reverse MR analysis, SNPs in AR summary data were chosen with a threshold of $P < 5 \times 10^{-8}$, and the same method ($\text{kb} = 10,000$, $r^2 < 0.001$) was applied to eliminate linkage disequilibrium (30). Palindrome SNPs, having identical alleles on the forward and reverse strands, were excluded due to the ambiguity in determining the effect allele strand. Finally, for each SNP meeting the specified criteria, R^2 and F-statistic were calculated, excluding SNPs with F-statistic < 10 as weak IVs, indicating a weak correlation between genetic variation and exposure (31).

2.6 Statistical analysis

Data analyses were conducted using MRPRESSO (version 1.0) and TwoSampleMR (version 0.5.6) packages in R software (version 4.3.1). Employing five MR analysis methods, with the Inverse Variance Weighted (IVW) method as the primary one, we assessed the causal relationship between exposure and outcome. To ensure robustness, we conducted sensitivity analyses using methods with different assumptions about horizontal pleiotropy, such as MR-Egger and Mendelian Randomization Pleiotropy

Residual Sum and Outlier (MR-PRESSO). MR-Egger analysis evaluated instrumental variable pleiotropy, with a non-zero intercept indicating bias in IVW estimates (32). MR-PRESSO identified horizontal pleiotropy through a global test and, if necessary, corrected potential pleiotropy by removing outliers (33). Additionally, we implemented the “leave-one-out” method to exclude abnormal SNPs, preventing undue influence of individual SNPs on the causal relationship between exposure and outcome. Furthermore, heterogeneity assessment used the Cochran Q test, and a P-value below 0.05 indicated present heterogeneity. In such cases, we employed the IVW random-effects model to estimate causal effects. In the absence of heterogeneity, the IVW fixed-effects model was considered the result of the IVW method. Moreover, considering multiple MR analyses between different exposures and a single outcome, we controlled for potential false-positive results due to multiple hypothesis testing using the false discovery rate (FDR) (34). Finally, we visually represented the results through forest plots, funnel plots, scatter plots, and “leave-one-out” plots.

3 Results

3.1 Exploration of the causal effect of immune cell phenotypes on AR

For the three GWAS summary-level datasets mentioned above, we selected IVs based on established significance threshold levels and included them in this study. Their F-statistics were all above 10, indicating that weak instrument bias is unlikely to be significant (Supplementary Tables 1–3, Sheet 1). Employing the IVW method as the primary MR analysis, we conducted a two-sample MR analysis investigating the relationship between immune cell phenotypes and AR, accompanied by tests for heterogeneity and pleiotropy. Post multiple testing correction using the FDR method, we identified two immune cell phenotypes demonstrating a causal relationship with AR at a significance level of 0.05, both exerting protective effects: CD3 on CD39+ activated Treg (Treg panel) and HVEM on CD45RA- CD4+ (Maturation stages of T cell). The odds ratio (OR) of CD3 on CD39+

activated Treg against AR, calculated by the IVW method, was 0.947 (95%CI = 0.923–0.972, $P = 3 \times 10^{-5}$, $P_{FDR} = 0.02278$), and the OR of HVEM on CD45RA- CD4+ against AR, calculated by IVW, was 0.967 (95%CI = 0.948–0.986, $P = 4 \times 10^{-5}$, $P_{FDR} = 0.01463$). Additionally, following insights from previous research, a $P_{FDR} < 0.2$ was considered suggestive correlation (35). At a significance level of 0.2, we identified a third immune cell phenotype with a causal relationship with AR, also displaying a protective effect: Naive CD8br %CD8br (Maturation stages of T cell panel). The OR of Naive CD8br %CD8br against AR, calculated by IVW, was 0.978 (95%CI = 0.966–0.990, $P = 4.5 \times 10^{-4}$, $P_{FDR} = 0.10862$). Results from the other four MR analysis methods for these three immune cell phenotypes and AR mirrored those of the IVW method, with OR values less than 1. MR-Egger and MR-PRESSO indicated no horizontal pleiotropy (P-values > 0.05, Supplementary Table 1, Sheet 1), confirming the reliability of the analysis results. Forest plots representing three immune cell phenotypes as exposure and AR as an outcome are illustrated in Figure 1. Scatter plots in Figures 2A–C, and Supplementary Figures 1–3 A, B, C offer comprehensive insights into funnel plots, ‘leave-one-out’ plots, and individual forest plots. Additionally, the results of five MR analyses for the above three immune cell phenotypes and AR are provided in the Supplementary Table 1, Sheet 2.

3.2 Exploration of the causal effect of AR on immune cell phenotypes

Additionally, to meet the requirements of mediation MR studies, we further treated AR as the exposure and the three aforementioned immune cell phenotypes as outcomes, conducting a reverse MR analysis with IVW as the primary method. We did not observe any reverse causal relationships between the three immune cell phenotypes and AR (P-values all greater than 0.05). Details of the heterogeneity and pleiotropy tests are provided in the Supplementary Table 2, Sheet 1, the results of five MR analyses for AR and three immune cell phenotypes are provided in the Supplementary Table 2, Sheet 2, and the forest plots are illustrated in Figure 3.

exposure	outcome	method	Pval	P_FDR	OR(95% CI)
Naive CD8br %CD8br	Allergic rhinitis	MR Egger	0.07287		0.985 (0.970 – 1.001)
		Weighted median	0.02026		0.979 (0.961 – 0.997)
		Inverse variance weighted	0.00045	0.10862	0.978 (0.966 – 0.990)
		Simple mode	0.09962		0.970 (0.935 – 1.005)
		Weighted mode	0.02691		0.983 (0.969 – 0.997)
CD3 on CD39+ activated Treg	Allergic rhinitis	MR Egger	0.00140		0.923 (0.883 – 0.964)
		Weighted median	0.00079		0.937 (0.903 – 0.973)
		Inverse variance weighted	0.00003	0.02278	0.947 (0.923 – 0.972)
		Simple mode	0.02299		0.932 (0.881 – 0.987)
		Weighted mode	0.00178		0.937 (0.904 – 0.972)
HVEM on CD45RA- CD4+	Allergic rhinitis	MR Egger	0.03259		0.965 (0.937 – 0.994)
		Weighted median	0.02176		0.974 (0.953 – 0.996)
		Inverse variance weighted	0.00004	0.01463	0.967 (0.948 – 0.986)
		Simple mode	0.61295		0.984 (0.927 – 1.045)
		Weighted mode	0.04574		0.974 (0.951 – 0.998)

FIGURE 1 Forest plots show the causal effect of three immune cell phenotypes on AR. OR, odds ratio; CI, confidence interval.

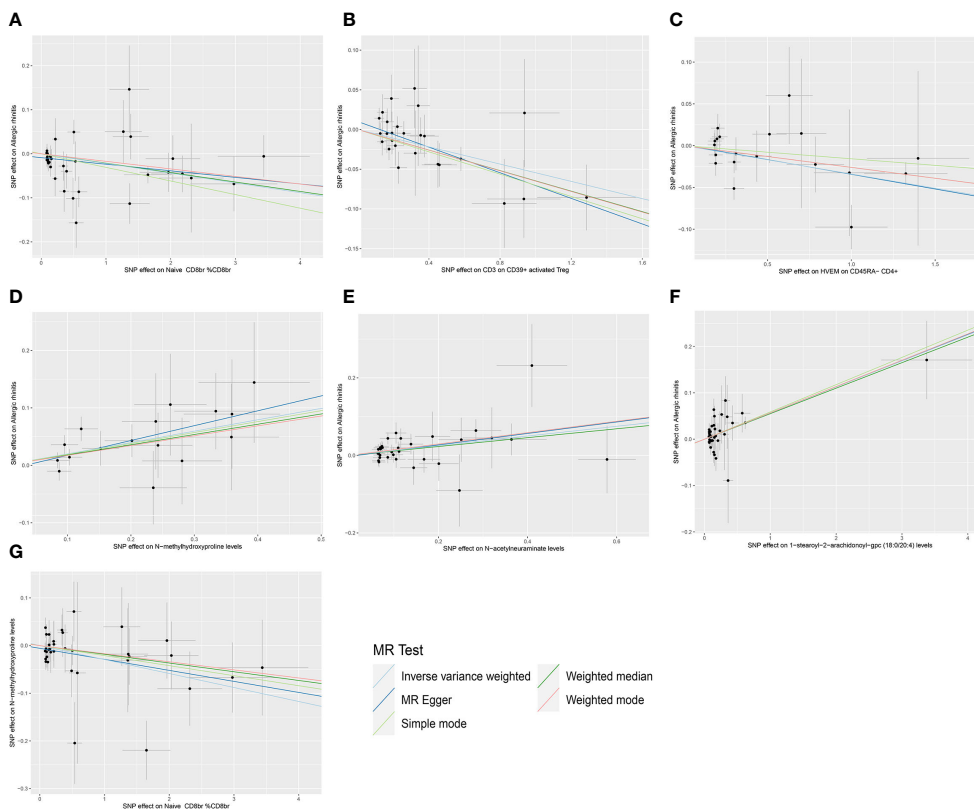


FIGURE 2 Scatter plots of MR analysis. (A) The causal effect of Naive CD8br % CD8br on AR. (B) The causal effect of CD3 on CD39+ activated Treg on AR. (C) The causal effect of HVEM on CD45RA- CD4+ on AR. (D) The causal effect of N-methylhydroxyproline levels on AR. (E) The causal effect of N-acetylneuraminate levels on AR. (F) The causal effect of 1-stearoyl-2-arachidonoyl-gpc (18:0/20:4) levels on AR. (G) The causal effect of Naive CD8br % CD8br on N-methylhydroxyproline levels.

3.3 Exploration of the causal effect of metabolite levels on AR

Subsequently, a two-sample MR analysis was conducted with metabolite levels as the exposure and AR as the outcome, utilizing IVW as the primary analytical method. Despite no significant

associations at the 0.05 level after FDR correction for multiple testing, we identified three metabolite levels with causal relationships with AR at the 0.2 level, all acting as risk factors: N-methylhydroxyproline levels, N-acetylneuraminate levels, and 1-stearoyl-2-arachidonoyl-gpc (18:0/20:4) levels. The OR of N-methylhydroxyproline levels against AR, calculated by the IVW

exposure	outcome	method	Pval	OR(95% CI)
Allergic rhinitis	Naive CD8br %CD8br	MR Egger	0.87530	0.950 (0.511 – 1.765)
		Weighted median	0.43532	1.078 (0.892 – 1.303)
		Inverse variance weighted	0.36674	1.067 (0.927 – 1.227)
		Simple mode	0.16549	1.245 (0.940 – 1.649)
		Weighted mode	0.43246	1.097 (0.880 – 1.368)
Allergic rhinitis	CD3 on CD39+ activated Treg	MR Egger	0.91669	0.952 (0.391 – 2.317)
		Weighted median	0.61710	1.076 (0.808 – 1.433)
		Inverse variance weighted	0.63096	1.055 (0.847 – 1.315)
		Simple mode	0.83412	1.040 (0.733 – 1.475)
Allergic rhinitis	HVEM on CD45RA- CD4+	MR Egger	0.51346	0.631 (0.170 – 2.340)
		Weighted median	0.21655	0.786 (0.537 – 1.151)
		Inverse variance weighted	0.39550	0.879 (0.654 – 1.183)
		Simple mode	0.44497	0.813 (0.491 – 1.347)
		Weighted mode	0.31973	0.794 (0.518 – 1.216)

FIGURE 3 Forest plots show the causal effect of AR on three immune cell phenotypes. OR, odds ratio; CI, confidence interval.

method, was 1.219 (95% CI = 1.104–1.346, $P = 9 \times 10^{-5}$, $P_{FDR} = 0.06427$). The OR of N-acetylneuraminate levels against AR, calculated by the IVW method, was 1.133 (95% CI = 1.061–1.211, $P = 1.7 \times 10^{-4}$, $P_{FDR} = 0.08076$). The OR of 1-stearoyl-2-arachidonoyl-gpc (18:0/20:4) levels against AR, calculated by the IVW method, was 1.058 (95% CI = 1.029–1.087, $P = 5 \times 10^{-5}$, $P_{FDR} = 0.07172$). Results from the other four methods in the MR analysis between these three metabolite levels and AR were consistent with the IVW method, with OR values all greater than 1. MR-Egger and MR-PRESSO indicated no horizontal pleiotropy (P-values all greater than 0.05, [Supplementary Table 3, Sheet 1](#)), confirming the reliability of the analysis results. Forest plots for these three metabolite levels as exposure and AR as the outcome are presented in [Figure 4](#), scatter plots in [Figures 2D–F](#), and [Supplementary Figures 1–3D–F](#) offer comprehensive insights into funnel plots, ‘leave-one-out’ plots, and individual forest plots. Furthermore, the detailed results for the five MR analyses between above three metabolite levels and AR can be found in the [Supplementary Table 3, Sheet 2](#).

3.4 Exploration of the causal effect of immune cell phenotypes on metabolite levels

Following the identification of immune cell phenotypes and metabolite levels causally associated with AR, we conducted two-sample MR analyses between them. Since no reverse causal relationships were found in the second step of the MR analysis, the three immune cell phenotypes from the initial step met the criteria for mediation MR analysis. Due to the relatively small number of tests (only three tests between multiple exposures and a single outcome), we refrained from performing FDR correction in this segment of the study, considering a P-value less than 0.05 as statistically significant. Utilizing IVW as the primary analytical method, we established a causal relationship between the Naive CD8br %CD8br phenotype and N-methylhydroxyproline levels, indicating a protective effect. The OR of Naive CD8br %CD8br

method, was 0.971 (95% CI = 0.950–0.992, $P = 8.31 \times 10^{-3}$). OR values from all five MR analysis methods were consistently greater than 1, and both MR-Egger and MR-PRESSO indicated no horizontal pleiotropy (P-values all greater than 0.05, [Supplementary Table 4, Sheet 1](#)), affirming the robustness of the results. For the remaining eight MR analyses, given that the IVW method is the primary MR analysis method, we refrain from considering causal relationships between them, as their IVW method P-values were all greater than 0.05. The forest plots for these three immune cell phenotypes as exposure and three metabolite levels as the outcome are displayed in [Figure 5](#), while [Figure 2G](#) illustrates the scatter plot for Naive CD8br %CD8br and N-methylhydroxyproline levels. [Supplementary Figures 1–3G](#) offer comprehensive insights into funnel plots, ‘leave-one-out’ plots, and individual forest plots. Moreover, the detailed results for the five MR analyses between three metabolite levels and three metabolite levels can be found in the [Supplementary Table 4, Sheet 2](#).

3.5 Genetically predicted N-methylhydroxyproline levels mediate the association between Naive CD8+ T cells and AR

Finally, the findings of our mediation MR analysis were summarized, and an intermediary factor was identified. [Figure 6](#) presents the summarized forest plot, while [Figure 7](#) depicts the schematic diagram of the mediation MR analysis. Notably, Naive CD8br %CD8br demonstrated a protective role in relation to N-methylhydroxyproline levels ($\beta = -0.029$, $P = 8.31 \times 10^{-3}$), with N-methylhydroxyproline levels identified as a risk factor for AR ($\beta = 0.198$, $P = 9 \times 10^{-5}$). Furthermore, Naive CD8+ %CD8+ emerged as a protective factor against AR ($\beta = -0.022$, $P = 4.5 \times 10^{-4}$). The mediated effect was -0.00574, accounting for 26.1% of the total effect, while the direct effect was -0.01626. Consequently, Naive CD8+ T cells contribute to the protection against AR by mitigating N-methylhydroxyproline levels.

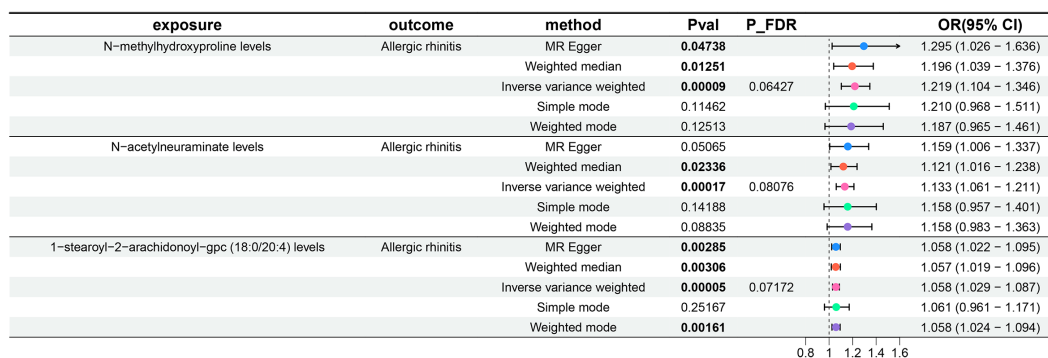


FIGURE 4 Forest plots show the causal effect of three metabolite levels on AR. OR, odds ratio; CI, confidence interval.

against N-methylhydroxyproline levels, calculated by the IVW

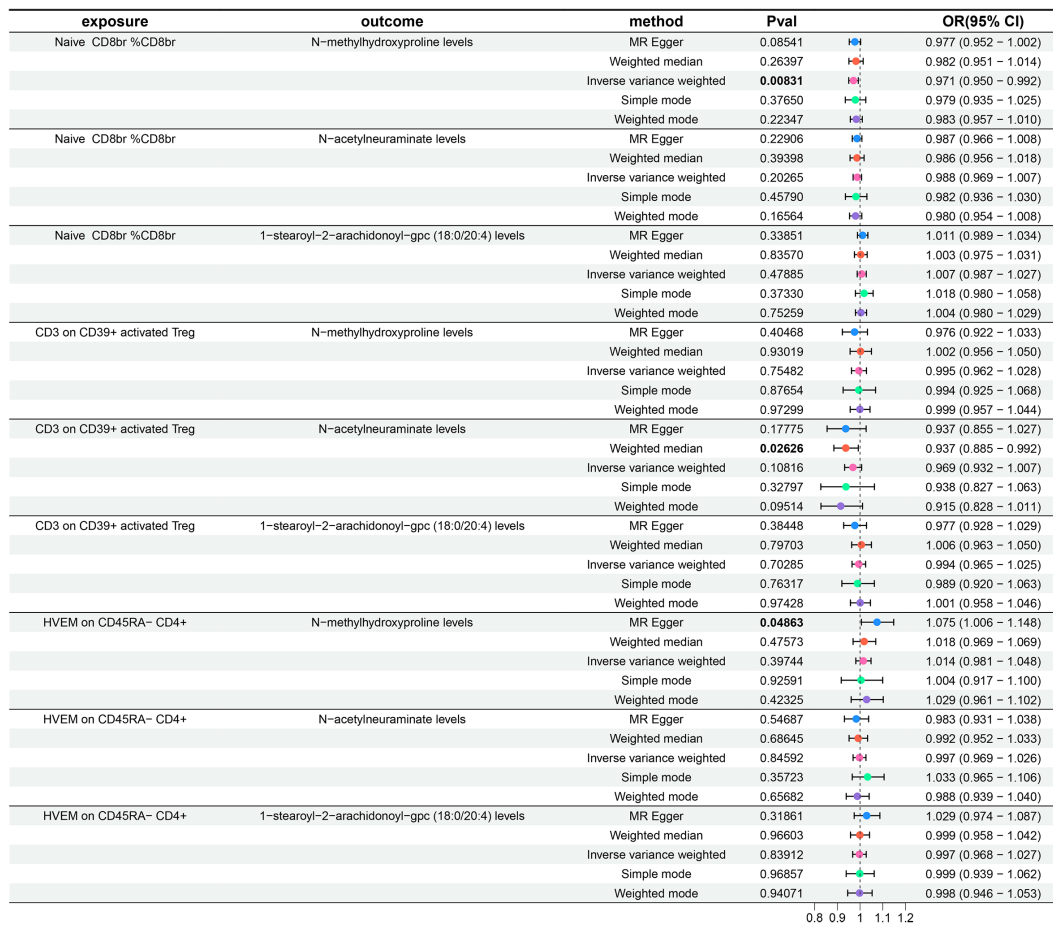


FIGURE 5 Forest plots show the causal effect of three immune cell phenotypes on three metabolite levels. OR, odds ratio; CI, confidence interval.

4 Discussion

Utilizing publicly available GWAS summary-level data, we systematically explored the causal relationships between 731 immune cell phenotypes and AR. Furthermore, acknowledging

the potential mediating influence of metabolites, we expanded our investigation to encompass the causal associations between 1400 metabolite levels and AR.

Previous studies have conducted MR analyses to investigate the causal relationship between eight immune-related diseases and AR.

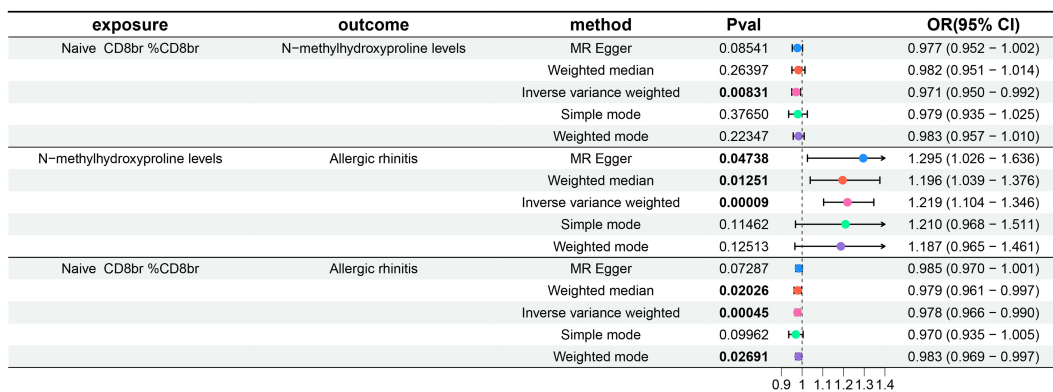
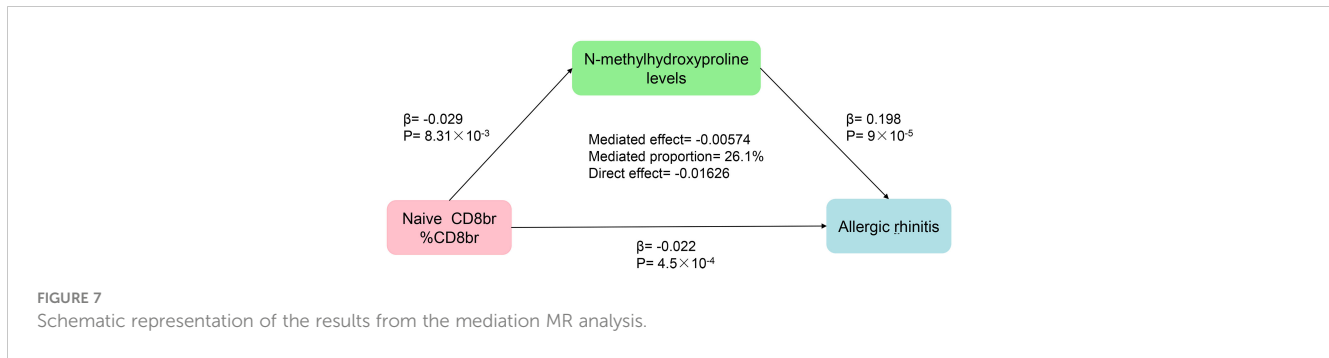


FIGURE 6 Forest plot of mediation MR analysis. OR, odds ratio; CI, confidence interval.



It indicates that atopic dermatitis (AD), asthma, and Crohn's disease (CD) elevate the risk of AR, whereas Graves' disease (GD) and systemic lupus erythematosus (SLE) potentially mitigate this risk (36). Another MR investigation scrutinized 486 circulating metabolites and 55 targeted urinary metabolites concerning their association with atopic dermatitis, allergic rhinitis, and asthma. Meta-analysis unveiled several circulating and urinary metabolites potentially linked to the onset and advancement of allergic diseases (37). Undoubtedly, the findings of these studies carry substantial implications for formulating preventive and therapeutic interventions for allergic rhinitis. Building upon prior research, this study employed GWAS summary-level data comprising 731 immune cell phenotypes and 1400 metabolite levels. Initially, it conducted separate MR analyses to identify immune cell phenotypes and metabolite levels potentially causally linked to allergic rhinitis. Subsequently, via mediation MR analysis, it investigated and uncovered a potential pathway, thus holding significant implications for guiding clinical practice.

To our knowledge, this study is the pioneering use of mediation MR analysis to investigate the causal links among immune phenotypes, metabolites, and AR. Following multiple testing correction of P-values using the FDR method, our findings indicate a total of three immune cell phenotypes acting as protective factors for AR, alongside three metabolite levels identified as risk factors for the condition. Notably, Naive CD8+ T Cells were discerned to exert a protective effect on AR by mitigating the levels of N-methylhydroxyproline. The STROBE-MR checklist pertaining to this study is available in [Supplementary Table 5](#).

The human immune system comprises innate and adaptive components. Unlike the nonspecific actions of the innate immune system, the adaptive immune system involves T lymphocytes and B lymphocytes, acts with specificity, and induces immune memory, enabling rapid responses in subsequent encounters. T lymphocytes, vital in cellular immune responses, consist of two main subsets: CD4-positive (CD4+) T cells, known as helper T cells (Th), and CD8-positive (CD8+) T cells, known as cytotoxic T cells (CTL) (38). These cells play crucial roles in pathogen defense. Naive CD8+ T cells, stimulated by antigens, differentiate into effector and memory T cells, participating in respective immune processes (39). Furthermore, the existence of Naive CD8+ T cells empowers the organism to fend off novel, undetected infections and ailments (40). Our investigation identified a heightened ratio of Naive CD8+ T cells to the overall CD8+ T cell count, potentially diminishing the

susceptibility to AR. Naive CD8+ T cells constitute the reservoir of CD8+ T cells, having recently matured in the thymus without encountering antigens yet, and can promptly react, producing effector and memory T cells for immunological assaults upon exposure to the relevant allergen. Certain research has indicated that, under chronic stimulation or specific pathological conditions, such as chronic infections or cancer, CD8+ T cells may experience immune exhaustion, characterized by a gradual loss of function and activity in response to sustained stimuli (41). Regulatory CD8+ T cells have inhibitory effects during AR occurrences, impacting inflammatory responses (42–44). Though not primary participants in allergic reactions, CD8+ T cells may exhibit exhaustion in response to allergens. Thus, the reservoir of Naive CD8+ T cells might have a significant regulatory function in the immune response of AR patients. However, this represents a speculative hypothesis concerning the potential causes of chronic recurrent episodes in AR patients, and the precise mechanisms necessitate additional validation through clinical research.

The CD3 on CD39+ activated Treg refers to a distinct subset within CD4 regulatory T cells expressing both CD3 and CD39 on its surface. Our investigation demonstrates that an elevation in the CD3 on CD39+ activated Treg correlates with a decreased risk of AR. Treg, categorized within the CD4+ T cell subset, play a pivotal role in the immune response to AR. Study suggests that diverse Toll-like receptors play crucial immunomodulatory roles in the pathogenesis of AR (45, 46). Notably, TICAM-1, functioning as a vital adaptor protein in Toll-like receptor signaling domains, participates in the signal transduction of AR. Treg can exert significant immunomodulatory effects on AR through the TICAM-1 pathway (47). The findings of this study highlight that a specific cell subset within Treg may confer a protective effect against AR, holding substantial implications for guiding AIT strategies.

HVEM on CD45RA- CD4+ T cells denote a distinct subset of CD4+ T cells expressing HVEM on the cell surface while lacking CD45RA, belonging to the mature stage of T lymphocytes. Herpes virus entry mediator (HVEM) belongs to the tumor necrosis factor receptor superfamily (TNFRSF) and plays a pivotal role in immune response regulation, contributing to the preservation of mucosal immune homeostasis (48, 49). Several studies have underscored HVEM's role in maintaining T cell homeostasis within the intestinal epithelium and safeguarding against invasion by enteropathogenic bacteria (50, 51). However, current clinical research on the involvement of HVEM in AR remains limited. CD45RA, a

member of the CD45 antigen family, is a glycoprotein located on the surface of lymphocytes (52). Under specific antigenic stimulation, the expression of CD45RA on the surface of T cells is constrained, commonly regarded as an indicator of naive T cells (53). Consequently, HVEM on CD45RA⁻ CD4⁺ T cells signifies mature CD4⁺ T cells expressing HVEM. Our study discloses that a subset of mature CD4⁺ T cells may serve as a protective factor in AR. This finding is anticipated to offer valuable guidance for future AIT.

N-methylhydroxyproline is a derivative of proline. Extensive research has demonstrated that proline metabolism plays a crucial role in diverse biological processes, including cell signal transduction, stress response, and energy generation. Additionally, it may contribute to the pathogenic mechanisms of microbial invaders. Consequently, inhibiting the proline metabolism process emerges as a potential therapeutic strategy against the intrusion of specific pathogens into the body (54). The involvement of proline metabolism in the immune response triggered by allergens remains unclear. This investigation brings to light that N-methylhydroxyproline may pose a risk factor for AR, suggesting a promising avenue for future exploration in AR research.

N-acetylneuraminic acid, also known as sialic acid, represents a neuraminic acid embellished with an acetyl group, typically positioned at the terminal end of polysaccharide chains (55). Current research underscores that diverse pathogenic bacteria can exploit sialic acid for camouflage, eluding detection by the host's innate immune system, thus gaining entry to enact their pathogenic pathways. Moreover, sialic acid can function as a nutritional source for select pathogenic bacteria (56, 57). This study posits that N-acetylneuraminic acid may constitute a potential risk factor for AR. An elevation in N-acetylneuraminic acid metabolite levels could, to some extent, create a conducive environment for the infiltration of pathogenic bacteria. When certain pathogenic microorganisms act as allergens, they might instigate an immune response in the body, culminating in the onset of AR. Presently, there exists a research gap concerning the correlation between N-acetylneuraminic acid and AR, necessitating further clinical investigations to validate these findings.

1-stearoyl-2-arachidonoyl-gpc is a lipid molecule derived from the amalgamation of stearic acid and arachidonic acid, constituting a glycerophospholipid. This investigation postulates that 1-stearoyl-2-arachidonoyl-GPC may play a role in the onset of AR. Regrettably, there is presently a dearth of clinical research concerning this metabolite, necessitating subsequent exploration in the future.

Through mediation MR analysis, we have discerned that N-methylhydroxyproline levels mediate the correlation between Naive CD8⁺ T cells and AR. The protective impact of Naive CD8⁺ T cells on AR manifests through a reduction in the N-methylhydroxyproline levels. It is conjectured that Naive CD8⁺ T cells, following exposure to the corresponding allergen, may influence N-methylhydroxyproline through a specific pathway, thereby mitigating the risk of AR occurrence.

This study conducted MR analysis utilizing publicly available comprehensive GWAS summary-level data, incorporating a large sample size and employing diverse testing methods to mitigate

potential biases, thus demonstrating considerable statistical robustness. However, the study does have several limitations. Firstly, the absence of individual information on AR patients precludes further subgroup analysis. Secondly, the primary focus of this study is to investigate the potential mediated effect of metabolites between immune cell phenotypes and AR. Consequently, the impact of AR on immune cell phenotypes was not comprehensively analyzed. Thirdly, MR studies hinge on genetic variations for causal inference; therefore, to authenticate our research findings, additional clinical studies remain imperative. Furthermore, our study may serve as a guiding framework for identifying and validating biomarkers as therapeutic targets for AR in clinical practice, introducing new perspectives to future AIT, and fostering clinical advancements.

5 Conclusion

Our MR analysis explored the causal relationships between several immune phenotypes and metabolite levels with AR, shedding light on several factors potentially implicated in the occurrence of AR. Our study has alleviated the influence of confounding factors in clinical research, bolstering the reliability of the study results. Furthermore, employing mediation MR analysis, we unveiled a potential pathway to prevent the onset of AR, providing novel research directions for AIT.

In conclusion, this study represents a pioneering exploration into the genetic perspective of AR pathogenesis, providing fresh insights for future treatment and prevention strategies. Building upon this research, it holds promise for innovative clinical interventions for AR patients. Nonetheless, further investigations are warranted to validate these findings and extend their applicability to broader populations.

Data availability statement

The original contributions presented in the study are included in the article/[Supplementary Material](#). Further inquiries can be directed to the corresponding author.

Ethics statement

Ethical approval was not required for the study involving humans in accordance with the local legislation and institutional requirements. Written informed consent to participate in this study was not required from the participants or the participants' legal guardians/next of kin in accordance with the national legislation and the institutional requirements.

Author contributions

ZC: Methodology, Project administration, Software, Writing – original draft. YS: Data curation, Visualization, Writing – original

draft. XD: Validation, Writing – original draft. XZ: Funding acquisition, Supervision, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This work was funded by grants from National Clinical Key Specialty Construction Project, Tianjin Key Medical Discipline (Specialty) Construction Project (TJYXZDXK-049A).

Acknowledgments

The authors express sincere gratitude to the researchers and participants involved in the original GWAS and the FinnGen biobank for their efforts in collecting and managing extensive data resources. Special appreciation is extended to those who actively contributed to this study.

References

- Greiner AN, Hellings PW, Rotiroli G, Scadding GK. Allergic rhinitis. *Lancet (London England)*. (2011) 378:2112–22. doi: 10.1016/S0140-6736(11)60130-X
- Bauchau V, Durham SR. Prevalence and rate of diagnosis of allergic rhinitis in Europe. *Eur Respir J*. (2004) 24:758–64. doi: 10.1183/09031936.04.00013904
- Ozdoganoglu T, Songu M. The burden of allergic rhinitis and asthma. *Ther Adv Respir Disease*. (2012) 6:11–23. doi: 10.1177/1753465811431975
- Leth-Møller KB, Skaaby T, Linneberg A. Allergic rhinitis and allergic sensitisation are still increasing among Danish adults. *Allergy*. (2020) 75:660–8. doi: 10.1111/all.14046
- Wang XD, Zheng M, Lou HF, Wang CS, Zhang Y, Bo MY, et al. An increased prevalence of self-reported allergic rhinitis in major Chinese cities from 2005 to 2011. *Allergy*. (2016) 71:1170–80. doi: 10.1111/all.12874
- Mims JW. Epidemiology of allergic rhinitis. *Int Forum Allergy Rhinology*. (2014) 4 Suppl 2:S18–20. doi: 10.1002/alf.21385
- Gauvreau GM, El-Gammal AI, O'Byrne PM. Allergen-induced airway responses. *Eur Respir J*. (2015) 46:819–31. doi: 10.1183/13993003.00536-2015
- Zhong H, Fan XL, Yu QN, Qin ZL, Chen D, Xu R, et al. Increased innate type 2 immune response in house dust mite-allergic patients with allergic rhinitis. *Clin Immunol*. (2017) 183:293–9. doi: 10.1016/j.clim.2017.09.008
- Peng YQ, Qin ZL, Fang SB, Xu ZB, Zhang HY, Chen D, et al. Effects of myeloid and plasmacytoid dendritic cells on ILC2s in patients with allergic rhinitis. *J Allergy Clin Immunol*. (2020) 145:855–67.e8. doi: 10.1016/j.jaci.2019.11.029
- Camelo A, Rosignoli G, Ohne Y, Stewart RA, Overed-Sayer C, Sleeman MA, et al. IL-33, IL-25, and TSLP induce a distinct phenotypic and activation profile in human type 2 innate lymphoid cells. *Blood Adv*. (2017) 1:577–89. doi: 10.1182/bloodadvances.2016002352
- Wambre E, Bajzik V, DeLong JH, O'Brien K, Nguyen QA, Speake C, et al. A phenotypically and functionally distinct human T(H)2 cell subpopulation is associated with allergic disorders. *Sci Transl Med*. (2017) 9:eaa9171. doi: 10.1126/scitranslmed.aam9171
- Iinuma T, Okamoto Y, Morimoto Y, Arai T, Sakurai T, Yonekura S, et al. Pathogenicity of memory Th2 cells is linked to stage of allergic rhinitis. *Allergy*. (2018) 73:479–89. doi: 10.1111/all.13295
- Yao Y, Wang N, Chen CL, Pan L, Wang ZC, Yunis J, et al. CD23 expression on switched memory B cells bridges T-B cell interaction in allergic rhinitis. *Allergy*. (2020) 75:2599–612. doi: 10.1111/all.14288
- Poposki JA, Peterson S, Welch K, Schleimer RP, Hulse KE, Peters AT, et al. Elevated presence of myeloid dendritic cells in nasal polyps of patients with chronic rhinosinusitis. *Clin Exp Allergy: J Br Soc Allergy Clin Immunol*. (2015) 45:384–93. doi: 10.1111/cea.12471

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fimmu.2024.1396246/full#supplementary-material>

- He YQ, Qiao YL, Xu S, Jiao WE, Yang R, Kong YG, et al. Allergen induces CD11c (+) dendritic cell autophagy to aggravate allergic rhinitis through promoting immune imbalance. *Int Immunopharmacology*. (2022) 106:108611. doi: 10.1016/j.intimp.2022.108611
- Galli SJ, Tsai M, Piliponsky AM. The development of allergic inflammation. *Nature*. (2008) 454:445–54. doi: 10.1038/nature07204
- Orrù V, Steri M, Sidore C, Marongiu M, Serra V, Olla S, et al. Complex genetic signatures in immune cells underlie autoimmunity and inform therapy. *Nat Genet*. (2020) 52:1036–45. doi: 10.1038/s41588-020-0684-4
- Pfaar O, Bousquet J, Durham SR, Kleine-Tebbe J, Larché M, Roberts G, et al. One hundred and ten years of Allergen Immunotherapy: A journey from empiric observation to evidence. *Allergy*. (2022) 77:454–68. doi: 10.1111/all.15023
- Fritzscheing B, Contoli M, Porsbjerg C, Buchs S, Larsen JR, Elliott L, et al. Long-term real-world effectiveness of allergy immunotherapy in patients with allergic rhinitis and asthma: Results from the REACT study, a retrospective cohort study. *Lancet Regional Health Europe*. (2022) 13:100275. doi: 10.1016/j.lanep.2021.100275
- Pfaar O, Agache I, de Blay F, Bonini S, Chaker AM, Durham SR, et al. Perspectives in allergen immunotherapy: 2019 and beyond. *Allergy*. (2019) 74 Suppl 108:3–25. doi: 10.1111/all.14077
- Licari A, Castagnoli R, Brambilla I, Tosca MA, De Filippo M, Marseglia G, et al. Biomarkers of immunotherapy response in patients with allergic rhinitis. *Expert Rev Clin Immunol*. (2018) 14:657–63. doi: 10.1080/1744666X.2018.1504679
- Smith GD, Ebrahim S. 'Mendelian randomization': can genetic epidemiology contribute to understanding environmental determinants of disease? *Int J Epidemiol*. (2003) 32:1–22. doi: 10.1093/ije/dyg070
- Sanderson E. Multivariable mendelian randomization and mediation. *Cold Spring Harbor Perspect Med*. (2021) 11:a038984. doi: 10.1101/cshperspect.a038984
- Emdin CA, Khera AV, Kathiresan S. Mendelian randomization. *Jama*. (2017) 318:1925–6. doi: 10.1001/jama.2017.17219
- Skrivankova VW, Richmond RC, Woolf BAR, Yarmolinsky J, Davies NM, Swanson SA, et al. Strengthening the reporting of observational studies in epidemiology using mendelian randomization. *Jama*. (2021) 326:1614. doi: 10.1136/bmj.n2233
- Kurki MI, Karjalainen J, Palta P, Sipilä TP, Kristiansson K, Donner KM, et al. FinnGen provides genetic insights from a well-phenotyped isolated population. *Nature*. (2023) 613:508–18. doi: 10.1038/s41586-022-05473-8
- Chen Y, Lu T, Pettersson-Kymmer U, Stewart ID, Butler-Laporte G, Nakanishi T, et al. Genomic atlas of the plasma metabolome prioritizes metabolites implicated in human diseases. *Nat Genet*. (2023) 55:44–53. doi: 10.1038/s41588-022-01270-1

28. Fei Y, Yu H, Wu Y, Gong S. The causal relationship between immune cells and ankylosing spondylitis: a bidirectional Mendelian randomization study. *Arthritis Res Ther.* (2024) 26:24. doi: 10.1186/s13075-024-03266-0
29. Yu XH, Yang YQ, Cao RR, Bo L, Lei SF. The causal role of gut microbiota in development of osteoarthritis. *Osteoarthritis cartilage.* (2021) 29:1741–50. doi: 10.1016/j.joca.2021.08.003
30. Burgess S, Butterworth A, Thompson SG. Mendelian randomization analysis with multiple genetic variants using summarized data. *Genet Epidemiol.* (2013) 37:658–65. doi: 10.1002/gepi.21758
31. Papadimitriou N, Dimou N, Tsilidis KK, Banbury B, Martin RM, Lewis SJ, et al. Physical activity and risks of breast and colorectal cancer: a Mendelian randomisation analysis. *Nat Commun.* (2020) 11:597. doi: 10.1038/s41467-020-14389-8
32. Bowden J, Davey Smith G, Burgess S. Mendelian randomization with invalid instruments: effect estimation and bias detection through Egger regression. *Int J Epidemiol.* (2015) 44:512–25. doi: 10.1093/ije/dyv080
33. Verbanck M, Chen CY, Neale B, Do R. Detection of widespread horizontal pleiotropy in causal relationships inferred from Mendelian randomization between complex traits and diseases. *Nat Genet.* (2018) 50:693–8. doi: 10.1038/s41588-018-0099-7
34. Benjamini Y, Drai D, Elmer G, Kafkafi N, Golani I. Controlling the false discovery rate in behavior genetics research. *Behav Brain Res.* (2001) 125:279–84. doi: 10.1016/s0166-4328(01)00297-2
35. Ma Z, Zhao M, Zhao H, Qu N. Causal role of immune cells in generalized anxiety disorder: Mendelian randomization study. *Front Immunol.* (2023) 14:1338083. doi: 10.3389/fimmu.2023.1338083
36. Zhao J, Zhang M, Li Z. Association between immune-related disease and allergic rhinitis: A two-sample mendelian randomization study. *Am J Rhinology Allergy.* (2023) 38:31–7. doi: 10.1177/19458924231207131
37. Tu J, Wen J, Luo Q, Li X, Wang D, Ye J. Causal relationships of metabolites with allergic diseases: a trans-ethnic Mendelian randomization study. *Respir Res.* (2024) 25:94. doi: 10.1186/s12931-024-02720-6
38. Medzhitov R. Recognition of microorganisms and activation of the immune response. *Nature.* (2007) 449:819–26. doi: 10.1038/nature06246
39. Kaech SM, Wherry EJ. Heterogeneity and cell-fate decisions in effector and memory CD8+ T cell differentiation during viral infection. *Immunity.* (2007) 27:393–405. doi: 10.1016/j.immuni.2007.08.007
40. Saxena A, Dagur PK, Biancotto A. Multiparametric flow cytometry analysis of naïve, memory, and effector T cells. *Methods Mol Biol (Clifton NJ).* (2019) 2032:129–40. doi: 10.1007/978-1-4939-9650-6_8
41. Kurachi M. CD8(+) T cell exhaustion. *Semin immunopathology.* (2019) 41:327–37. doi: 10.1007/s00281-019-00744-5
42. Lin L, Dai F, Wei J, Chen Z. CD8(+) Tregs ameliorate inflammatory reactions in a murine model of allergic rhinitis. *Allergy asthma Clin immunology: Off J Can Soc Allergy Clin Immunol.* (2021) 17:74. doi: 10.1186/s13223-021-00577-8
43. Lin L, Dai F, Wei J, Chen Z. Influences of CD8(+) tregs on peripheral blood mononuclear cells from allergic rhinitis patients. *Laryngoscope.* (2021) 131:E316–e23. doi: 10.1002/lary.28759
44. Qiu S, Duan X, Geng X, Xie J, Gao H. Antigen-specific activities of CD8+ T cells in the nasal mucosa of patients with nasal allergy. *Asian Pacific J Allergy Immunol.* (2012) 30:107–13.
45. Golshiri-Isfahani A, Amizadeh M, Arababadi MK. The roles of toll like receptor 3, 7 and 8 in allergic rhinitis pathogenesis. *Allergologia immunopathologia.* (2018) 46:503–7. doi: 10.1016/j.aller.2017.09.026
46. Radman M, Golshiri A, Shamsizadeh A, Zainodini N, Bagheri V, Arababadi MK, et al. Toll-like receptor 4 plays significant roles during allergic rhinitis. *Allergologia immunopathologia.* (2015) 43:416–20. doi: 10.1016/j.aller.2014.04.006
47. Zhang LX, Liu T. [Treg influences the pathogenesis of allergic rhinitis through TICAM-1 pathway]. *Lin chuang er bi yan hou tou jing wai ke za zhi = J Clin otorhinolaryngology head Neck surgery.* (2018) 32:1763–6. doi: 10.13201/j.issn.1001-1781.2018.22.020
48. Shui JW, Kronenberg M. HVEM: An unusual TNF receptor family member important for mucosal innate immune responses to microbes. *Gut Microbes.* (2013) 4:146–51. doi: 10.4161/gmic.23443
49. Shui JW, Kronenberg M. HVEM is a TNF receptor with multiple regulatory roles in the mucosal immune system. *Immune network.* (2014) 14:67–72. doi: 10.4110/in.2014.14.2.67
50. Seo GY, Takahashi D, Wang Q, Mikulski Z, Chen A, Chou TF, et al. Epithelial HVEM maintains intraepithelial T cell survival and contributes to host protection. *Sci Immunol.* (2022) 7:eabm6931. doi: 10.1126/sciimmunol.abm6931
51. Seo GY, Shui JW, Takahashi D, Song C, Wang Q, Kim K, et al. LIGHT-HVEM signaling in innate lymphoid cell subsets protects against enteric bacterial infection. *Cell Host Microbe.* (2018) 24:249–60.e4. doi: 10.1016/j.chom.2018.07.008
52. Henson SM, Riddell NE, Akbar AN. Properties of end-stage human T cells defined by CD45RA re-expression. *Curr Opin Immunol.* (2012) 24:476–81. doi: 10.1016/j.coi.2012.04.001
53. Hermiston ML, Xu Z, Weiss A. CD45: a critical regulator of signaling thresholds in immune cells. *Annu Rev Immunol.* (2003) 21:107–37. doi: 10.1146/annurev.immunol.21.120601.140946
54. Christgen SL, Becker DF. Role of proline in pathogen and host interactions. *Antioxidants Redox Signaling.* (2019) 30:683–709. doi: 10.1089/ars.2017.7335
55. Angata T, Varki A. Chemical diversity in the sialic acids and related alpha-keto acids: an evolutionary perspective. *Chem Rev.* (2002) 102:439–69. doi: 10.1021/cr000407m
56. Severi E, Hood DW, Thomas GH. Sialic acid utilization by bacterial pathogens. *Microbiol (Reading England).* (2007) 153:2817–22. doi: 10.1099/mic.0.2007/009480-0
57. Vimr ER, Kalivoda KA, Deszo EL, Steenbergen SM. Diversity of microbial sialic acid metabolism. *Microbiol Mol Biol reviews: MMBR.* (2004) 68:132–53. doi: 10.1128/MMBR.68.1.132-153.2004