Check for updates

OPEN ACCESS

EDITED BY Sajjad Ahmad, Abasyn University, Pakistan

REVIEWED BY Irfan Manzoor, Government College University, Pakistan Anam Naz, University of Lahore, Pakistan

*CORRESPONDENCE Alaa Karkashan 🕅 askarkashan@uj.edu.sa

RECEIVED 04 March 2024 ACCEPTED 22 April 2024 PUBLISHED 10 May 2024

CITATION

Karkashan A (2024) Immunoinformatics assisted profiling of West Nile virus proteome to determine immunodominant epitopes for the development of next-generation multipeptide vaccine. *Front. Immunol.* 15:1395870. doi: 10.3389/fimmu.2024.1395870

COPYRIGHT

© 2024 Karkashan. This is an open-access article distributed under the terms of the **Creative Commons Attribution License (CC BY)**. The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Immunoinformatics assisted profiling of West Nile virus proteome to determine immunodominant epitopes for the development of nextgeneration multi-peptide vaccine

Alaa Karkashan*

Department of Biological Sciences, College of Sciences, University of Jeddah, Jeddah, Saudi Arabia

Emerging infectious diseases represent a significant threat to global health, with West Nile virus (WNV) being a prominent example due to its potential to cause severe neurological disorders alongside mild feverish conditions. Particularly prevalent in the continental United States, WNV has emerged as a global concern, with outbreaks indicating the urgent need for effective prophylactic measures. The current problem is that the absence of a commercial vaccine against WNV highlights a critical gap in preventive strategies against WNV. This study aims to address this gap by proposing a novel, multivalent vaccine designed using immunoinformatics approaches to elicit comprehensive humoral and cellular immune responses against WNV. The objective of the study is to provide a theoretical framework for experimental scientists to formulate of vaccine against WNV and tackle the current problem by generating an immune response inside the host. The research employs reverse vaccinology and subtractive proteomics methodologies to identify NP_041724.2 polyprotein and YP_009164950.1 truncated flavivirus polyprotein NS1 as the prime antigens. The selection process for epitopes focused on B and T-cell reactivity, antigenicity, water solubility, and non-allergenic properties, prioritizing candidates with the potential for broad immunogenicity and safety. The designed vaccine construct integrates these epitopes, connected via GPGPG linkers, and supplemented with an adjuvant with the help of another linker EAAAK, to enhance immunogenicity. Preliminary computational analyses suggest that the proposed vaccine could achieve near-universal coverage, effectively targeting approximately 99.74% of the global population, with perfect coverage in specific regions such as Sweden and Finland. Molecular docking and immune simulation studies further validate the potential efficacy of the vaccine, indicating strong binding affinity with toll-like receptor 3 (TLR-3) and promising immune response profiles, including significant antibody-mediated and cellular responses. These findings present the vaccine construct as a viable candidate for further development and testing. While the theoretical and computational results are promising, advancing from in-silico predictions to a tangible vaccine requires comprehensive laboratory validation. This next step is essential to confirm the vaccine's efficacy and safety in eliciting an immune response against WNV. Through this study, we propose a novel approach to vaccine development against WNV and contribute to the broader field of

immunoinformatics, showcasing the potential to accelerate the design of effective vaccines against emerging viral threats. The journey from hypothesis to practical solution embodies the interdisciplinary collaboration essential for modern infectious disease management and prevention strategies.

KEYWORDS

computer-assisted vaccine designing, reverse vaccinology, immunoinformatics, molecular docking, MD simulation

Introduction

The West Nile virus WNV is the causative agent of vector-borne infectious illnesses primarily observed within the United States. The primary mode of transmission is predominantly by the bite of a mosquito that has been infected. The transmission of the WNV occurs when an infected mosquito transfers the disease-causing agent to an intermediate host, typically other animals. Subsequently, these infective animals can then transmit the virus to human individuals (1). Instances of WNV infections manifest during the period of mosquito activity, commencing from the summer months and extending through the autumn season. Currently, there is a lack of commercially accessible vaccines or medications for the treatment of WNV infection in human populations. Auspiciously, a majority of individuals afflicted with WNV infection do not experience symptoms of illness. Approximately 20% of persons who are infected experience the manifestation of a fever and accompanying symptoms. Moreover, it has been shown that approximately 1 out of every 150 individuals who are infected with the disease acquire a severe and potentially lethal condition (2). Furthermore, it is worth noting that a majority of individuals within various communities, approximately 80%, do not exhibit any signs of infection. However, in extreme instances, multiple symptoms may manifest, such as high-grade fever, neck stiffness, tremors, muscle weakness, eyesight loss, paralysis, and numbness (3). The process of recuperation following a severe illness might span from several weeks to many months. In certain cases, where the virus impacts the central nervous system, it may result in enduring impairments (4). Given its significant pathogenicity and the existence of effective medicine and immunization, it is imperative to implement concerted measures to address the infection (5).

Antibiotics are generally ineffective in treating viral infections due to the significant diversity and high mutation rate of viral genomes. Nonetheless, alternative treatments, including hydration and pain relievers, can offer relief (6). Vaccination offers an alternative strategy for the permanent eradication of infections. The risk of infection can be reduced through the use of appropriate insect repellent and by adopting protective measures such as wearing long-sleeved shirts and pants to decrease the chances of mosquito bites (7). The virus has been detected in several bird species, notably jays and crows, which have shown symptoms and mortality linked to the infection. Furthermore, reports suggest that birds can become infected with the virus within a week, and there is evidence of viral spread within bird populations. The genetic structure of the West Nile Virus (WNV) comprises a singlestranded RNA molecule with a positive-sense orientation (8). The primary role of the genome is twofold: firstly, to serve as the sole viral mRNA, and secondly, to act as the template for the production of the complementary minus-strand RNA (9).

The production of vaccines is a highly significant subject in the contemporary period, as it pertains to a diverse array of transmissible and infectious diseases. Additionally, vaccines can be employed as a preventive measure against certain diseases, including autoimmune conditions. Historically, the utilization of pasture vaccinology for vaccine development was characterized by its cost-effectiveness, lengthy duration, and inherent risks, occasionally resulting in the inability to successfully cultivate and propagate the desired study specimens inside laboratory settings (10).

Vaccination aims to confer protective immunity to the host organism against pathogens by eliciting both innate and acquired immune responses, which are subsequently activated upon encountering the pathogens (11). The genomes of various bacterial and viral pathogens have been made accessible in biological databases through the utilization of bioinformatics technologies. Reverse vaccinology utilizes the available databases by leveraging the sequencing data for various objectives (12). Bioinformatics, immunoinformatics, and reverse vaccinology have emerged as fundamental and indispensable approaches in the field of vaccine development (13). In the aforementioned analysis, the researcher can collect and analyze sequential data to build vaccines against various dangerous bacterial species (14).

The utilization of computer-aided techniques in vaccine development shows promise in establishing a theoretical framework for the creation of multi-epitope vaccine constructs that may effectively target various pathogenic species. This strategy involves identifying potential vaccine candidates within the entire proteome (15). Multi-epitope-based vaccinations mostly comprise several antigenic epitopes rather than solitary epitopes, so eliciting a robust and secure immune response within the host organism (16). Typically, the identification of anticipated epitopes relies on experimental data obtained on a case-by-case basis. The hepatitis C virus served as a prime illustration in the pursuit of developing a very effective recombinant vaccine. The development of epitope-based vaccine design has been ongoing for the past thirty years. In comparison to entire pathogen vaccinations, multi-epitope vaccines have lower inherent immunogenicity. However, the construction of epitope-based vaccines offers a safe means of inducing immunogenicity and effectively protecting the host (17).

The objective of the present study was to develop a multiepitope vaccination construct targeting specific viral infections across their whole proteome. Furthermore, the study attempted to evaluate the docking contact between the suggested vaccine construct and various receptors on immune cells. Furthermore, molecular dynamic modelling was conducted to analyze the motion of the docked complexes, since this is of utmost importance for antigenic presentation, processing, and the induction of an appropriate immune response in the host against specific pathogens. The discovery has the potential to mitigate the challenges encountered in traditional vaccinology through the utilization of pan-genome-based vaccinology methods. This strategy can yield substantial cost and time savings in the vaccine formulation process.

Research methodology

Figure 1 Schematic representation of the study conducted for the multi-epitope vaccine construct.

Proteins selection and prioritization of possible vaccine candidates (epitopes)

The investigation was initiated by conducting a comprehensive extraction of the proteome of WNV from the NCBI database, which was subsequently followed by the prioritization of potential vaccine



FIGURE 1

Methodology flow diagram, followed by the designing of a multiepitopes vaccine candidate against WNV by applying several Immunoinformatics and biophysical approaches. targets (18). During the phase of vaccine target prioritization, the examination of physiochemical properties was conducted. This analysis involved examining many factors such as molecular weight, theoretical isoelectric point (pI), instability index, aliphatic index, and GRAVY. The ProtParam web server (19) was utilized for this purpose. A variety of computational tools were used to evaluate antigenicity, allergenicity, and toxicity using the following computational tools: VaxiJen 2.0 (20), Allerton (21) and ToxinPred (22). To identify potential vaccine candidates that are likely to exhibit antigenicity, lack allergenicity, and possess non-toxic properties, a rigorous selection process is necessary to mitigate the risk of triggering autoimmune responses. A BLASTp analysis was conducted to identify and eradicate duplicate proteins to reduce the risk of triggering auto-immune responses (23).

Epitope mapping and selection phase

The multi-epitope-based vaccine construct is composed of a sequence of short peptides that overlap with each other. This design allows for an effective enhancement of the immune response due to the antigenic and immunogenic properties of the epitopes. The study utilized the NP_041724.2 polyprotein and YP_009164950.1 truncated flavivirus polyprotein NS1 prime proteins as antigenic and physio-chemically stable proteins to predict and map epitopes. B cell epitopes were predicted and selected using the "IEDB" online web server (24). In addition, the epitopes of B-cells were employed for the prediction of T-cell epitopes in the major histocompatibility complex (MHC-I and MHC-II). The predicted epitopes were ranked based on the lowest percentile rank (15). The epitopes that were predicted were further evaluated for their antigenicity, allergenicity, water solubility, and toxicity using VaxiJen 2.0, AllerTop 2.0, and the Solubility Peptide Calculator available at https://pepcalc.com/peptidesolubility-calculator.php. Afterwards, we proceeded to choose epitopes that were deemed to be potentially antigenic, non-allergic, non-toxic and had good water solubility (25).

Vaccine construction 3-dimensional structure prediction and processing phase

The utilization of multi-epitope-based vaccines represents a highly promising strategy within the field of reverse vaccinology (26). The field of vaccinology research (RV) encompasses contemporary approaches that aim to identify potential vaccine candidates within the core proteome of pathogens, applying various bioinformatics and immunoinformatics technologies. The multi-epitope vaccine design primarily consists of diverse antigenic epitopes that are non-allergenic, highly soluble in water, and non-toxic (27). During the construction phase of the multi-epitope-based vaccine (MEVC), the shortlisted epitopes were connected using GPGPG linkers. Furthermore, to improve the effectiveness of the vaccine construct, the designed vaccine construct was linked with the cholera toxin-B subunit adjuvant, which is known to induce immune efficacy (28).

Choice of linker in the vaccine construct

The utilization of GPGPG linkers in our vaccine design is a deliberate choice, guided by their well-documented efficacy in maintaining structural stability and epitope presentation. GPGPG linkers are short, flexible sequences that provide sufficient space between epitopes, thereby reducing structural hindrance and potential steric clashes. This spatial separation is crucial for the independent folding and processing of each epitope, ensuring that each component of the vaccine is effectively recognized by the immune system (29). The flexibility offered by GPGPG linkers also facilitates the physical separation of functional domains within the vaccine construct, minimizing interference in their respective immune elicitation roles. This is particularly important in multiepitope vaccines, where the goal is to present multiple antigens to the immune system in a manner that mimics natural infection closely (16). Alternative linkers, such as EAAAK could theoretically be used in vaccine constructs and may influence the vaccine's efficacy differently. We used an EAAAK linker to join an adjuvant with a vaccine construct The specifically used linkers have many advantages for example, they are known for their rigid helical structure, which could enhance the structural stability of the construct but might limit the flexibility and independent orientation of linked epitopes. On the other hand, EKK linkers, owing to their positive charge, could potentially improve the solubility of the vaccine construct but may also affect the overall immunogenicity due to their shorter length and less pronounced structural separation capabilities (30). Ultimately, the choice of GPGPG was predicated on achieving an optimal balance between structural flexibility, solubility, and the effective presentation of epitopes to the immune system. This careful consideration ensures that our vaccine construct can elicit a broad and robust immune response, a critical factor in the successful development of a vaccine against West Nile Virus (31).

Following the development of the multi-epitope vaccine construct, the vaccine construct underwent subsequent processing for the examination of its physiochemical properties. This analysis was conducted using the ProtParam tool (19), which can be accessed at https://web.expasy.org/protparam/. In addition, the vaccine design underwent examination for antigenicity, allergenicity, toxicity, and water solubility utilizing VaxiJen 2.0, AllerTop 2.0, ToxinPred, ProtParam, and INNOVAGEN web tools (32). The three-dimensional structure of the vaccine construct was created utilizing the "Scratch protein predictor online tool" (28). The vaccine structure underwent additional refinement using the GalaxyRefine web-based tool, the refined structure of the vaccine construct was then selected for further processing (33).

Disulfide engineering

Following the refining of disulfide linkages, engineering was conducted using the web-based program Design 2.0. This resulted in the creation of a mutant structure, denoted as MEVC, whereby all enzymatic sensitive residues were substituted with cysteine amino acid residues (34).

Population coverage analysis

The examination of population coverage was conducted using the Immune Epitope Database (IEDB) online database (24). This database was utilized to analyze the selected epitopes for the vaccine construct in terms of their coverage across global populations and individual countries (35).

C-IMMSIM immune simulation analysis

The analysis of immune simulation was conducted using in silico methods, specifically through the utilization of the C-IMMSIM internet web server (36). In the context of C-IMMSIM immunological simulation analysis, distinct immune responses of B and T-cells were found towards the vaccine design (37).

Molecular docking analysis of vaccine construct

Molecular docking analysis is a computational-based analysis that is mainly used for binding interaction analysis between ligand and receptor molecules (Vaccine and immune cells receptors (38, 39), In the molecular docking analysis phase immune cell receptor molecule, was selected as a receptor for the design of vaccine construct, and pdb structure of the Toll-like receptor-3 (TLR-3) was retrieved from protein data bank (pdb) (40) and prepare for docking analysis cluspro 2.0 web tool was utilized (38), the target receptor TLR-3 plays a vital role in the immunogenicity in the pathogenesis of viral pathogenic species (42). Both the vaccine and receptor PDB files were uploaded to the cluspro 2.0 web server and submitted for docking analysis. The "cluspro 2.0 webserver" generated top-10 docked complexes of vaccine molecule with target receptor (TLR-3).

Selection of host receptor - TLR3

In the design of our proposed multi-epitope vaccine against West Nile Virus (WNV), we specifically targeted Toll-like receptor 3 (TLR3) due to its critical role in the innate immune system's response to viral infections. TLR3, a pattern recognition receptor, identifies double-stranded RNA (dsRNA), a molecular pattern associated with viral replication, including that of flaviviruses such as WNV. The activation of TLR3 initiates a cascade of antiviral defenses, notably the induction of type I interferon responses, which are paramount in controlling viral load and spread early in the infection. Moreover, TLR3's involvement in the maturation and activation of dendritic cells underscores its importance in bridging innate and adaptive immunity, thereby enhancing the antigen presentation process and the subsequent activation of a targeted immune response (43).

Normal mode simulation analysis

To explore the dynamic motion of the docked complex normal mode simulation analysis was done using the iMOD normal mode analysis server. In the mode simulation analysis, the stability and three-dimensional structure of the construct with the docked structure were analyzed. For the normal mode simulation analysis the pdb file of the docked complex was used having the lowest binding energy score (15, 44).

Molecular dynamic analysis

Molecular dynamic simulation analysis was applied for the dynamic behavior analysis of docked molecules in the specific period of 100 nanoseconds (ns). The MD simulation analysis was done by using AMBER 21 packages (45). For simulation analysis top-1 docked compound was selected which was generated by cluspro 2.0, the selection was based on the lowest binding energy score. The simulation was done mainly in three phases, system preparation, pre-processing, and trajectory analysis phase. In the trajectories analysis phase mainly three trajectories "root mean square deviation (RMSD)", "root mean square fluctuation (RMSF)" and radius of gyration (RoG) were created. In root mean square deviation analysis time-dependent deviation was analyzed while in root mean square fluctuation residues dependent fluctuation of the residues of the protein was analyzed. Moreover, the docked complex compactness and relaxation were assessed through the radius of gyration analysis as the compactness is crucial for docked complexes for appropriate stability (46).

Results

Subtractive proteomic analysis

In subtractive proteomic analysis, we extracted two proteins of WNV using accession numbers>NP_041724.2 and >YP_009164950. The proteins were subjected to physiochemical properties analysis and both the proteins were predicted with stable physiochemical properties as tabulated in Table 1.

The physiochemical stable proteins were considered for further subtractive proteomics analysis, in antigenicity analysis both proteins were predicted as probable antigenic with 0.5407 and 0.6015, >NP_041724.2, >YP_009164950 respectively, in water solubility and allergenicity analysis both the proteins were predicted good water soluble and non-allergic. In homology analysis, the selected proteins were predicted as non-similar to human and human microbiota analysis. Finally, the selected epitopes were selected for the epitopes prediction and prioritization phase.

Epitope prediction and prioritization analysis phase

In the epitopes prediction and prioritization phase B cell epitopes were predicted from the selected proteins in this phase both B and T cells (MHC-I and MHC-II) epitopes were predicted the predicted epitopes are mentioned in the following, Table 2 mentions B and B cells derived T-cells Epitopes. The B cells peptide was then used for the T-cells (MHC-I and MHC-II) Prediction, based on the lowest percentile ranks the epitopes were prioritized and selected for further analysis as described in the following Table 3.

Epitopes screening phase

The predicted epitopes underwent further screening, including MHC prediction, antigenicity, allergenicity, toxicity, and water solubility analysis. Only the probable antigenic, well-binding to DRB*0101, water-soluble, and non-toxic epitopes were chosen for the design of the epitope-based vaccine construct. The shortlisted epitopes are presented in Table 4.

Epitopes vaccine construction MEVC and evaluation

In the epitope-based vaccine construction phase, the final shortlisted epitopes were utilized to create a multi-epitope vaccine construct. This construct primarily comprised four selected epitopes with varying lengths, connected by GPGPG and EAAAK linkers, along with an adjuvant. The adjuvant was linked via the EAAA linker. Following the assembly of the linear epitope-based vaccine construct, it underwent evaluation for physicochemical properties analysis.

TABLE 1 Physiochemical properties analysis of the selected proteins.

	Proteins accession number	Physiochemical properties Analysis							
		Number of amino acids	Molecular weight (kDa)	Theoretical pl	Instability index	Aliphatic index	GRAVY		
1	>NP_041724.2	3430	38.010982	8.63	34.02	86.14	-0.15		
2	>YP_009164950	1191	13.02591	8.66	36.55	80.4	-0.162		

TABLE 2 B-cell epitopes, predicted from shortlisted proteins.

S.NO	Selected Proteins	B-cells Peptides	
1	>NP_041724.2 polyprotein	orotein NRRSTKQKKRGGTA	
		TRAACPTMGEAHNEKRAD	
		PTTVESHGKIGATQA	
		IDGPETEECPTANRAW	
		AGPRSNHNRRPGYKTQNQGPWDEGRV	
		SCEHRGPAARTTTESGK	
		DITWESDAEITGSSER	
		GKNVKNVQTKPGVFKTPEGEIGA	
		LDYPTGTSGSPIVDKNG	
		ATPPGTSDPFPESNAPISDMQTEIPDRAWNT	
		KSYETEYPKCKNDD	
		SAITAASAAQRRGRIGRNPSQVGDEYCYGGHTNEDDSNFA	
		FDGPRTNTILEDNNE	
		GWQAEAMRSAQRRT	
		ITEVDRSAAKHARREGNITGGHPVSRGT	
		RVQEVKGYTKGGPGHEEPQLV	
		KKTWKGPQFEEDVNLGSGTRAV	
		SSTWHQDANHPYRTWNYH	
		KVDTKAPEPPEGV	
		SAITAASAAQRRGRIGRNPSQVGDEYCYGGHTNEDDSNF	
		ITEVDRSAAKHARREGNITGGHPVSRGT	
		RVQEVKGYTKGGPGHEEPQLV	
		KKTWKGPQFEEDVNLGSGTRAV	
		SSTWHQDANHPYRTWNYH	
		QNQWKNAREAVEDPK	
2	>YP_009164950.1 truncated flavivirus	NRRSTKQKKRGGTA	
		PTTVESHGKIGATQA	
		EVTVDCEPRSGIDT	
		AGPRSNHNRRPGYKTQNQGPWDEGRV	
		SCEHRGPAARTTTESGK	
		RVLGHPGGPSQEVDGQDQHSSYHACTPSPSVW	
		NRRSTKQKKRGGTA	

The analysis indicated that the designed vaccine construct consists of 180 amino acid sequences, with a molecular weight of 19.81774 kilodaltons (KDa), a Theoretical pI of 9.91, and an Aliphatic index of 66.28. The instability evaluation yielded a value of 40.78, suggesting that the vaccine construct is classified as stable. Subsequently, the designed sequence of the vaccine construct was utilized for three-dimensional structure prediction. Figure 2A illustrates a 3D representation of the multi-epitope vaccine construct (MEVC). Furthermore the tertiary of the vaccine construct was validated through Ramachandran analysis, in the Ramachandran Plot statistics we observed that 128 (83.1%) amino acid residues are present in the most favored regions, while 21 (13.6%) amino acid residues are predicted in the additional allowed regions, moreover, 4 (2.6%) number of amino acids residues were observed in the Generously allowed regions and 1 (0.6%) several amino acids residues were observed in the Disallowed regions. Next

TABLE 3 MHC (I & II) predicted epitopes from B-cell epitopes with the lowest percentile rank.

MHC-I	Lowest percentile rank MHC-II		Lowest percentile rank	
NRRSTKQKK	2 NRRSTKQKKRGGT		49	
TMGEAHNEK	EAHNEK 0.67 RAACPTMGEAHNEK		25	
HGKIGATQA	2.1	VESHGKIGATQA	3.4	
ETEECPTANR	0.09	IDGPETEECPTANRA	54	
NQGPWDEGR	3	YKTQNQGPWDEGRV	40	
SCEHRGPAAR	3.3	SCEHRGPAARTTT	9.6	
ESDAEITGSS	3.7	WESDAEITGSSE	2.4	
KNVKNVQTK	0.48	GKNVKNVQTKPGV	13	
GTSGSPIVDK	0.05	PTGTSGSPIVDK	11	
ESNAPISDM	0.24	FPESNAPISDMQTE	13	
KSYETEYPK	0.02	KSYETEYPKCKN	8.9	
TAASAAQRR	0.05	SAITAASAAQRRG	1.8	
DGPRTNTIL	1.2	FDGPRTNTILEDN	8.5	
EAMRSAQRR	0.07	GWQAEAMRSAQRR	18	
EAMRSAQRR	0.06	EGNITGGHPVSR	3.3	
RVQEVKGYTK	0.07	RVQEVKGYTKGG	7.4	
GPQFEEDVNL	0.57	GPQFEEDVNLGSG	3.9	
TWHQDANHPY	0.83	SSTWHQDANHPY	0.81	
KAPEPPEGV	0.32	VDTKAPEPPEGV	42	
TAASAAQRR	0.05	SAITAASAAQRRGR	2.1	
NITGGHPVSR	0.25	EGNITGGHPVSR	3.3	
RVQEVKGYTK	0.07	RVQEVKGYTKGGP	20	
GPQFEEDVNL	0.57	GPQFEEDVNLGS	3.3	
TWHQDANHPY	0.83	SSTWHQDANHPY	0.81	
NQWKNAREA	1.3	QNQWKNAREAVEDPK	16	
NRRSTKQKK	2.4	NRRSTKQKKRGG	24	
HGKIGATQA	2.1	TVESHGKIGATQA	4.2	
EVTVDCEPR	0.06	EVTVDCEPRSGI	9.5	
RPGYKTQNQ	1.9	RPGYKTQNQGPWD	34	
SCEHRGPAAR	3.3	SCEHRGPAARTTT	9.6	
HACTPSPSVW	0.03	HSSYHACTPSPSVW	15	
NRRSTKQKK	2	NRRSTKQKKRGGT	49	
TMGEAHNEK	0.67	RAACPTMGEAHNEK	25	
HGKIGATQA	2.1	VESHGKIGATQA	3.4	
EVTVDCEPR	0.06	IDGPETEECPTANRA	54	
RPGYKTQNQ	1.9	YKTQNQGPWDEGRV	40	
SCEHRGPAAR	3.3	SCEHRGPAARTTT	9.6	
HACTPSPSVW	0.03	WESDAEITGSSE	2.4	

Shortlisted epitopes	Calculated DRB*0101 Allele binding score	Antigenicity	Allergenicity	Toxicity	Water solubility
NRRSTKQKK	30.06	0.2751			Good water-
KSYETEYPK	37	0.0578	Non-	Non- toxic peptides	
TAASAAQRR	22	1.1437	allergens peptides		soluble peptides
NRRSTKQKK	30	0.2751			

TABLE 4 Characteristics of selected epitopes for designing of epitopes-based vaccine construct.

154 (100.0%) Non-glycine and non-proline residues were also observed. Further 14, 10 glycine and proline residues respectively observed and 2 End-residues (excl. Gly and Pro), the above statistic was based on a total number of 180 amino acid residues of the vaccine construct and it represents that the vaccine construct structure is stable, as the Ramachandran analysis graph is mentioned in Figure 2B.

Furthermore, in disulfide engineering the original structure is mutated and replaced enzymatic sensitive amino acid residues with cysteine amino acid (Leu 4-Asn 35, His 20- Asn 25, Ala 17-Lue 29, Cys 30- Asn 35, Gln 37-Lue 41, Arg50-Gly 75, Tyr 97-Cys 150, c \Cys 107- Ala 127, Pro 151-Thr 157) pairs of residues were selected to be mutated. The original and mutated structures of the designed vaccine construct are presented in following Figures 3A, B.

Next Secondary structure was generated, and the secondary structure predicted 180 residues, 1 chain 9 (47%) helices and 0 beta sheets, Figure 4 presented the description of the secondary structure of the vaccine construct.

Cloning of MEVC and codon optimization

The sequences were first converted to DNA sequences. "ATGATCAAACTGAAATTTGGCGTCTTCTTCACCGT CCTGCTGTCTTCTGCTTACGCTCACGGTACCCCGC AGAACATCACCGACCTGTGCGCTGAATACCTTACG CTCACGGTACCCCGCAGAACATCACCGACCTGTGC GCTGAATACCTCTCTGGCTGGTAAACGTGAAATGG CTATCATCACCTTCAAAAACGGTGCTATCTTCCAGG TTGAAGTTCCGGGTTCTCAGCACATCGACTCTCAGA AAAAAGCTATCGAACGTATGAAAGACACCCTGCGTA TCGCTTACCTGACCGAAGCTAAAGTTGAAAAACTGTG CGTTTGGAACAACAAAACCCCGCACGCTATCGCTGCTA TCTCTATGGCTAACGAAGCTGCTGCTAAAAACCGTCGT TCTACCAAACAGAAAAAGGTCCGGGTCCGGGTAAATCT TACGAAACCGAATACCCGAAAGGTCCGGGTCCGGGTAC CGCTGCTTCTGCTGCTCAGCGTCGTGGTCCGGGTCCGGG TAACCGTCGTTCTACCAAACAGAAAAAA" with (0.9607476511968707) codon adaption value "CAI-Value of the improved sequence" and (49.44) GC-Content of the improved sequence, The improved DNA sequence then inserted in pET-28a vector as pet28a vector map is presented in the following Figure 5.

Interaction analysis

For interaction analysis and the binding ability of MEVC to immune cell receptors molecular docking was performed, the docking analysis was performed through the online cluspro 2.0 web tool, and the tool generated the Top 10 docked complexes. Table 5 mentions docked complexes with the lowest binding energy score. The Top 1 docked complex was then subjected to molecular dynamic simulation analysis. The intermolecular docked confirmation is represented in Figure 6.



(A) Multi-epitopes based vaccine construct against West Nile Virus, consisting upon selected epitopes, GPGPG and EAAAK linkers and Cholera Toxin B subunit adjuvant. (B) Ramachandran Plot of the tertiary structure of vaccine construct.

08



Normal mode molecular dynamic simulation

The IModS webserver does the precarious analysis of the docked structure by adjusting the force field of the docked complex concerning different interval times. The findings represent less deformation at each residue level. The Eigenvalues value of the complex is 1.96817e - 06. Next, the Lowest RMSD and highest co-related regions in the heat maps predicted that there are better interactions of each residue. The following Figure 7 explains the outcome trajectories of the normal mode simulation analysis. In the following trajectories, the structure assessment of the docked complexes was examined in Figure 7A the mobility and structure strength of the docked complexes have been analyzed, and predicted that there is proper binding strength between the vaccine and receptors. In the structural variances analysis of the docked complexes, we observed that the structures of the docked complexes are compact and no abrupt variances occur during simulation time the variances plot is mentioned in Figure 7B

while the covariance plot is presented in Figure 7C. The elastic network related to the atom is mentioned in Figure 7D. Furthermore deformability of the docked structure was also analyzed and we observed that there is no deformation occurring in the docked complexes, the deformability plot is mentioned in Figure 7E. The beta factor plot of the NMA is presented in Figure 7F and the graph presents that the docked complexes are stable and compact. Moreover, the eigenvalues plot is presented in Figure.

Molecular dynamic simulation analysis

In molecular dynamic simulation analysis, the dynamic movement of the docked complexes was analyzed, and the trajectories of MD simulation were analyzed on the base of kinetics and total net binding energy (RMSD and radius of gyration). In MD simulation RMSD was analyzed as RMSD is the measurement of the average distance between the backbone atoms of the protein. RMSD plots predicted the highest deviation at 80 ns



Frontiers in Immunology



but at the end time of the simulation, the plot became stable and showed stability as the RSMD graph is presented in Figure 8 top plot. Next radius of gyration analysis was performed for competence and relaxation of the docked complexes at 70 nanoseconds the RoG plot showed fluctuation however at the end of simulation time we observed stability in the RoG plot which predicted that the vaccine and immune receptor are compact as the RoG graph is presented in Figure 8 bottom plot.

C-immune simulation analysis

In the c-immune simulation analysis, we observed the immuneinducing ability of the vaccine construct, the server predicted that the MEVC could produce a strong immune response in the host body against the designed vaccine construct. The immune system response was detected in the form of humoral and cellular immunity Figure 9A represents antibody response toward the designed vaccine in the form of IgM and IgG followed by IgG1 and IgG2 as overall the immune response is presented by different colors peaks. Next cellular immune was also observed in the form of interferon and several other types of cytokines and tumor growth factor beta and tumor necrosis factor a (TNFa) as the cellular immune response as presented in Figure 9B.

Population coverage analysis

In the population coverage analysis, we observed that the selected epitopes can cover the worldwide population (Class combined "MHC-I and MHC-II) as we predicted that World, North America, Central America, West Indies, South America, Central Africa, South Africa, East Africa, Southwest Asia,

Southeast Asia, South Asia, Oceania, 99.74%, 99.89%,53.80%, 99.69%, 95.15%, 94.79%, 95.27%, 97.08%,95.79%, 97.66%, 98.70%, 97.88% respectively while the other countries population coverage are presented in Figure 10. However different other countries and their population coverage are tabulated in Supplementary Table 1 (S1).

Discussion

A multitude of WNV cases have been reported throughout the United States, predominantly observed during the summer season. It is noteworthy that the annual incidence of WNV exhibits global variability, primarily attributed to periodic outbreaks (47). The challenge of addressing WNV lies in the lack of a suitable commercial vaccine, as the availability of a vaccine would provide a valuable and secure means of eradicating infectious diseases on a global scale. The process of vaccine development necessitates a substantial allocation of labor and financial resources (38). The application of reverse vaccinology, bioinformatics, and immunoinformatics has significantly alleviated this burden. Presently, researchers employ reverse vaccinology pipelines to discern numerous antigenic vaccine targets against various bacterial and viral pathogens by analyzing the proteins of said pathogens (48). Reverse vaccinology mainly uses the expressed genomic sequences of the pathogens for the identification of vaccine candidates, as in pasture base vaccine development vaccines are created by using pathogenic organisms which can cause harmful effects during vaccine formulation, but in the case of reverse vaccinology the experimental scientist the purified and possible vaccine candidate for vaccine formulation (49).

The absence of a vaccine targeting the aforementioned virus can be attributed to the relatively low rates of mortality and morbidity associated with it. However, it is imperative to acknowledge that our

TABLE 5	Top- 10) generated	docked	complexes	of	vaccine	with	TLR-	-3)	•
---------	---------	-------------	--------	-----------	----	---------	------	------	-----	---

Cluster	Members	Representative	Weighted Score		
0	0 91 Center		-924.4		
		Lowest Energy	-938.2		
1	89	Center	-791.5		
		Lowest Energy	-960.3		
2	60	Center	-915.0		
		Lowest Energy	-952.1		
3	49	Center	-878.4		
		Lowest Energy	-878.4		
4	45	Center	-983.5		
		Lowest Energy	-983.5		
5		Center	-859.1		
		Lowest Energy	-859.1		
6	35	Center	-750.9		
		Lowest Energy	-910.5		
7	28	Center	-827.8		
		Lowest Energy	-891.4		
8	27	Center	-789.4		
		Lowest Energy	-839.9		
9	23	Center	-846.7		
		Lowest Energy	-929.3		
10	20	Center	-789.2		
		Lowest Energy	-834.6		

global community remains persistently vulnerable to numerous potential epidemics and endemics. Notable instances include the emergence of Bolivian Hemorrhagic fever and the coronavirus infection in 2019 (50). The investigation does not necessitate exclusive attention on the outer membrane complete proteins of the pathogens. It is worth noting that through the utilization of the immunoinformatics approach, researchers have successfully identified potential multi-antigenic vaccine targets that exhibit a high degree of antigenicity. These findings hold significant promise for the advancement of vaccine development (51). The potential vaccine should be probable antigenic, physiochemical stable, good water soluble, non-allergic, nontoxic and good water-soluble. The purpose of the said characteristics is to target only probable antigenic epitopes, physiochemical stable, good water soluble non-allergic and nontoxic respectively, the only antigenic epitopes can elicit an immune response so only antigenic epitopes were considered for further filtering, also to avoid allergic and toxic reaction toward vaccine candidates. the allergic and toxic epitopes were removed (28). In epitopes prediction specifically two antigenic proteins >NP_041724.2 polyprotein and >YP_009164950.1 truncated flavivirus were used for epitope prediction using the IEDB online web server, and the epitopes were selected for further filters based on the lowest percentile score (24).

The viral poly proteins mainly play a vital role in the budding of the virus by binding to the host cell membrane and gathering the viral genomic material (DNA or RNA) into the nucleocapsid that produces the core of a virion particle. The WNV polyprotein is the first virus protein that is produced by the infected cells, but the main role of the target protein plays an important role in the virus assembly formation, and it can also block the process of host cell apoptosis during the viral replication process. Next, the second target protein of the WNV mainly plays a vital role in the process of WNV propagation, with attachment, entrance, replication, assembly formation and release of complete viral particles. So we target these proteins specifically to predict the epitopes and reduce the viral propagation by generation of immune responses (52).

The selected epitopes were further filtered through various subtractive immunoinformatics filters, like antigenicity, allergenicity, water solubility and toxicity, to activate proper immune responses and avoid allergic and toxic reactions toward the vaccine candidate. The probable antigenic epitopes could evoke proper immune reaction inside the host against specific pathogens, next allergic response with





the vaccine is one of the main concerns, to avoid the allergic response, all the allergic epitopes were discarded using AllerTOP v. 2.0 online web server and only the non-allergic epitopes were processed further. Followed by allergenicity prevention from the toxic immune response is also vital to prevent the host from a toxic reaction. Utilizing all the immunoinformatics approaches probable antigenic and good vaccine target epitopes were selected for a multi-epitope vaccine designed against target pathogens (23, 25).

Multi-epitope vaccine construct is mainly composed of multiantigenic peptides rather than whole proteins and can be considered a promising strategy against tumors and viral infections. The epitopes-based vaccine was designed by joining the selected



FIGURE 8

Root mean square deviation (RMSD) and radius of gyration (RoG) plots of docked complexes (Vaccine and Toll-like receptors 3), top one represents RMSD while the bottom one represents RoG.



epitopes through GPGPG linkers and joined with the Cholera Toxin B subunit adjuvant. The GPGPG linkers were used to avoid flexibility in the vaccine construct while the adjuvant was used for inducing immunodeficiency of the vaccine to construct. After designing the sequences of the vaccine construct, and being subjected to Scratch Protein Predictor for structure prediction the vaccine construct, the 3D structure was then refined through another software GalaxyRefine and all the present loops from the structure to the structural stability and competence. Next, the structure minimization of the vaccine was done before docking analysis to optimize the active sites of the target receptors (53).

The crucial aspect in the induction of an appropriate immune response against pathogens lies in the intricate interplay between vaccine construction and immune cell receptors. The interaction analysis involved conducting a Molecular docking analysis to examine the vaccine's binding affinity with the TLR-3 molecule. The docking analysis was conducted utilizing the ClusPro 2.0 web tool and predicted the interaction of vaccine construct with the target immune cell receptor, ClusPro 2.0 is a widely recognized webserver for molecular docking analysis. The tool was employed to generate a set of Top-10 models based on their respective binding energy scores, thereby providing valuable insights into the docking process. The assessment of the vaccine construct's stability in conjunction with the



receptor molecule is of utmost importance. To fulfil this objective, we conducted an analysis utilizing normal mode simulation and molecular dynamic simulation techniques were employed. Additionally, we examined the deformability of the structure throughout the simulation time and predicted that the vaccine construct and immune cell receptor have proper binding affinity in a dynamic environment. Afterwards, C-immune simulation analysis of the vaccine construct has the potential to elicit an immune response against WNV through the activation of both B and T-cells, thereby enhancing the immune system's ability to combat the virus (54). Researchers can utilize the proposed vaccine construct during the development of experimental vaccines targeting specific pathogens. The inclusion of experimental validation will significantly contribute to the theoretical foundation of the vaccine.

Conclusion

The organism that is responsible for the transmission of the WNV is primarily mosquitoes. This virus induces a range of clinical manifestations, such as elevated and fluctuating body temperature, Cephalalgia, cervical rigidity, altered mental state, profound confusion, unconsciousness, involuntary muscle contractions, convulsions, muscular debility, visual impairment, sensory numbness, and motor paralysis. These symptoms have been observed within the United States. To propose a potentially effective strategy for mitigating the presence of disease-causing surface-associated proteins, a theoretical foundation for a vaccine construct was devised utilizing an advanced bioinformatics methodology known as next-generation sequencing. The vaccine construct that was designed consisted of various epitopes, adjuvants, and linkers. An analysis of its physiochemical properties, structure, antigenicity, and allergenicity was conducted. The designed vaccine construct, which was deemed potentially antigenic, non-toxic, and non-allergic, was then utilized for further analysis. Utilizing biophysical methodologies, including docking and simulation analysis, we investigated the binding mode of the vaccine with TLR-3. Our analysis revealed promising results, suggesting that the vaccine construct possesses the capability to bind with the desired target receptors. Moreover, our analysis of the C-immune system revealed a significant induction of immune response against the targeted pathogens upon administration of the vaccine construct. Additionally, the designed vaccine demonstrated the ability to stimulate the production of various interferons. Nevertheless, it is highly advisable to conduct an experimental study to further substantiate the *insilico* findings.

Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material. Further inquiries can be directed to the corresponding author.

Author contributions

AK: Writing - original draft.

Funding

The author(s) declare that no financial support was received for the research, authorship, and/or publication of this article.

References

 Colpitts TM, Conway MJ, Montgomery RR, Fikrig E. West Nile Virus: biology, transmission, and human infection. *Clin Microbiol Rev.* (2012) 25:635–48. doi: 10.1128/ CMR.00045-12

2. Elizondo-Quiroga D, Elizondo-Quiroga A. West Nile virus and its theories, a big puzzle in Mexico and Latin America. *J Glob Infect Dis.* (2013) 5:168. doi: 10.4103/0974-777X.122014

3. Venkat H, Krow-Lucal E, Hennessey M, Jones J, Adams L, Fischer M, et al. Notes from the field: concurrent outbreaks of St. Louis encephalitis virus and West Nile virus disease—Arizona, 2015. *MMWR Morb Mortal Wkly Rep.* (2015) 64:1349–50. 10.15585/mmwr.mm6448a5

4. Davis LE, DeBiasi R, Goade DE, Haaland KY, Harrington JA, Harnar JB, et al. West Nile virus neuroinvasive disease. Ann Neurol Off J Am Neurol Assoc Child Neurol Soc. (2006) 60:286-300. doi: 10.1002/ana.20959

 Coffin CS, Fung SK, Alvarez F, Cooper CL, Doucette KE, Fournier C, et al. Management of hepatitis B virus infection: 2018 guidelines from the Canadian Association for the Study of Liver Disease and Association of Medical Microbiology and Infectious Disease Canada. *Can Liver J.* (2018) 1:156–217. doi: 10.3138/ canlivj.2018-0008

6. Luong T, Salabarria A-C, Roach DR. Phage therapy in the resistance era: where do we stand and where are we going? *Clin Ther.* (2020) 42:1659–80. doi: 10.1016/j.clinthera.2020.07.014

7. Klein SRM, Foster AO, Feagins DA, Rowell JT, Erovenko IV. Optimal voluntary and mandatory insect repellent usage and emigration strategies to control the chikungunya outbreak on Reunion Island. *PeerJ.* (2020) 8:e10151. doi: 10.7717/peerj.10151

 McLean RG. West Nile virus in North American birds. Ornithol Monogr. (2006) 60:44–64. doi: 10.2307/40166827

9. Rampersad S, Tennant P. Replication and expression strategies of viruses. *Viruses*. (2018), 55. doi: 10.1016/B978-0-12-811257-1.00003-6

10. Zhang H, Chen J. Current status and future directions of cancer immunotherapy. *J Cancer.* (2018) 9:1773. doi: 10.7150/jca.24577

11. Kobayashi M, Vekemans J, Baker CJ, Ratner AJ, Le Doare K, Schrag SJ. Group B Streptococcus vaccine development: present status and future considerations, with emphasis on perspectives for low and middle-income countries. *F1000Research*. (2016) 5. doi: 10.12688/f1000research

12. Bambini S, Rappuoli R. The use of genomics in microbial vaccine development. Drug Discovery Today. (2009) 14:252-60. doi: 10.1016/j.drudis.2008.12.007

13. Ullah A, Ullah Khan S, Haq MU, Ahmad S, Irfan M, Asif M, et al. Computational study to investigate Proteus mirabilis proteomes for multi-epitope vaccine construct design. *J Biomol Struct Dyn.* (2022) 41:1–12. doi: 10.1080/07391102.2022.2153920

Conflict of interest

The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fimmu.2024. 1395870/full#supplementary-material

14. Nawaz M, Ullah A, Al-Harbi AI, Haq MU, Hameed AR, Ahmad S, et al. Genome-Based Multi-Antigenic Epitopes Vaccine Construct Designing against Staphylococcus hominis Using Reverse Vaccinology and Biophysical Approaches. *Vaccines.* (2022) 10:1729. doi: 10.3390/vaccines10101729

15. Yousaf M, Ismail S, Ullah A, Bibi S. Immuno-informatics profiling of monkeypox virus cell surface binding protein for designing a next-generation multi-valent peptidebased vaccine. *Front Immunol.* (2022) 13. doi: 10.3389/fimmu.2022.1035924

16. Rida T, Ahmad S, Ullah A, Ismail S, Tahir ul Qamar M, Afsheen Z, et al. Pan-genome analysis of oral bacterial pathogens to predict a potential novel multi-epitopes vaccine candidate. *Int J Environ Res Public Health*. (2022) 19:8408. doi: 10.3390/ijerph19148408

17. Khan S, Irfan M, Hameed AR, Ullah A, Abideen SA, Ahmad S, et al. Vaccinomics to design a multi-epitope-based vaccine against monkeypox virus using surface-associated proteins. *J Biomol Struct Dyn.* (2022) 41:1–10. doi: 10.1080/07391102.2022.2158942

18. Pruitt KD, Tatusova T, Maglott DR. NCBI reference sequences (RefSeq): a curated non-redundant sequence database of genomes, transcripts and proteins. *Nucleic Acids Res.* (2007) 35:D61–5. doi: 10.1093/nar/gkl842

19. Chason, Eric, Tong Su, and Zhaoxia Rao. "Computational tool for analyzing stress in thin films." Surface and Coatings Technology (2023) 74:130099

20. Areeshi MY. Prediction of Epitope Based Vaccine Candidates against Macaca fascicularis PV Type 2 Virus Using In-silico Approaches. *Egypt Acad J Biol Sci C Physiol Mol Biol.* (2018) 10:49–57. doi: 10.21608/eajbsc.2018.13655

21. Dimitrov I, Bangov I, Flower DR, Doytchinova I. AllerTOP v. 2—a server for in silico prediction of allergens. J Mol Model. (2014) 20:2278. doi: 10.1007/s00894-014-2278-5

22. Sharma N, Naorem LD, Jain S, Raghava GPS. ToxinPred2: an improved method for predicting toxicity of proteins. *Brief Bioinform*. (2022). 23(5):1–12. doi: 10.1093/bib/bbac174

23. Malik M, Khan S, Ullah A, Hassan M, Ahmad S, Al-Harbi AI, et al. Proteomewide Screening of Potential Vaccine Targets against Brucella melitensis. *Vaccines*. (2023) 11:263. doi: 10.3390/vaccines11020263

24. Vita R, Overton JA, Greenbaum JA, Ponomarenko J, Clark JD, Cantrell JR, et al. The immune epitope database (IEDB) 3.0. *Nucleic Acids Res.* (2015) 43:D405–12. doi: 10.1093/nar/gku938

25. Mushtaq M, Khan S, Hassan M, Al-Harbi AI, Hameed AR, Khan K, et al. Computational Design of a Chimeric Vaccine against Plesiomonas shigelloides Using Pan-Genome and Reverse Vaccinology. *Vaccines*. (2022) 10:1886. doi: 10.3390/vaccines10111886

26. Albekairi TH, Alshammari A, Alharbi M, Alshammary AF, Tahir ul Qamar M, Anwar T, et al. Design of a Multi-Epitope Vaccine against Tropheryma whipplei Using

Immunoinformatics and Molecular Dynamics Simulation Techniques. Vaccines. (2022) 10:691. doi: 10.3390/vaccines10050691

27. Gul S, Ahmad S, Ullah A, Ismail S, Khurram M, Tahir ul Qamar M, et al. Designing a Recombinant Vaccine against Providencia rettgeri Using Immunoinformatics Approach. *Vaccines*. (2022) 10:189. doi: 10.3390/vaccines10020189

28. Alharbi M, Alshammari A, Alasmari AF, Alharbi SM, Tahir ul Qamar M, Ullah A, et al. Designing of a Recombinant Multi-Epitopes Based Vaccine against Enterococcus mundtii Using Bioinformatics and Immunoinformatics Approaches. *Int J Environ Res Public Health.* (2022) 19:3729. doi: 10.3390/ijerph19063729

29. Ud-Din M, Albutti A, Ullah A, Ismail S, Ahmad S, Naz A, et al. Vaccinomics to design a multi-epitopes vaccine for Acinetobacter baumannii. *Int J Environ Res Public Health*. (2022) 19:5568. doi: 10.3390/ijerph19095568

30. Ullah A, Ahmad S, Ismail S, Afsheen Z, Khurram M, Tahir ul Qamar M, et al. Towards A Novel Multi-Epitopes Chimeric Vaccine for Simulating Strong Immune Responses and Protection against Morganella morganii. *Int J Environ Res Public Health*. (2021) 18:10961. doi: 10.3390/ijerph182010961

31. Anwar T, Ismail S, Parvaiz F, Abbasi SW, A. Al-Abbasi F, M. Alghamdi A, et al. Computational design of experimentally validated multi-epitopes vaccine against hepatitis E virus: An immunological approach. *PloS One.* (2023) 18:e0294663. doi: 10.1371/journal.pone.0294663

32. Ismail S, Ahmad S, Azam SS. Vaccinomics to design a novel single chimeric subunit vaccine for broad-spectrum immunological applications targeting nosocomial Enterobacteriaceae pathogens. *Eur J Pharm Sci.* (2020) 146. doi: 10.1016/j.ejps.2020.105258

33. Ko J, Park H, Heo L, Seok C. GalaxyWEB server for protein structure prediction and refinement. *Nucleic Acids Res.* (2012) 40:W294–7. doi: 10.1093/nar/gks493

34. Abbasi BA, Saraf D, Sharma T, Sinha R, Singh S, Sood S, et al. Identification of vaccine targets & design of vaccine against SARS-CoV-2 coronavirus using computational and deep learning-based approaches. *PeerJ.* (2022) 10:e13380. 10.7717/peerj.13380

35. Alshammari A, Alharbi M, Alghamdi A, Alharbi SA, Ashfaq UA, Tahir ul Qamar M, et al. Computer-Aided Multi-Epitope Vaccine Design against Enterobacter xiangfangensis. *Int J Environ Res Public Health*. (2022) 19:7723. doi: 10.3390/ijerph19137723

36. Meem FS. An in-silico approach to design a vaccine against beta lactamase enzyme of mycobacterium tuberculosis (Doctoral dissertation, Brac University). (2022).

37. Irfan M, Khan S, Hameed AR, Al-Harbi AI, Abideen SA, Ismail S, et al. Computational Based Designing of a Multi-Epitopes Vaccine against Burkholderia mallei. *Vaccines*. (2022) 10:1580. doi: 10.3390/vaccines10101580

38. Kozakov D, Hall DR, Xia B, Porter KA, Padhorny D, Yueh C, et al. The ClusPro web server for protein–protein docking. *Nat Protoc.* (2017) 12:255–78. doi: 10.1038/nprot.2016.169

39. Alshammari A, Alasmari AF, Alharbi M, Ali N, Muhseen ZT, Ashfaq UA, et al. Novel Chimeric Vaccine Candidate Development against Leptotrichia buccalis. *Int J Environ Res Public Health.* (2022) 19:10742. doi: 10.3390/ijerph191710742

40. Burley SK, Bhikadiya C, Bi C, Bittrich S, Chao H, Chen L, et al. RCSB Protein Data Bank (RCSB. org): delivery of experimentally-determined PDB structures

alongside one million computed structure models of proteins from artificial intelligence/machine learning. *Nucleic Acids Res.* (2023) 51:D488–508. 0.1093/nar/gkac1077

41. Pettersen EF, Goddard TD, Huang CC, Couch GS, Greenblatt DM, Meng EC, et al. UCSF Chimera—a visualization system for exploratory research and analysis. *J Comput Chem.* (2004) 25:1605–12. doi: 10.1002/jcc.20084

42. Yujuan C, Junhong LIN, Yao Z, Xianping MA, Huashan YI. Toll-like receptor 3 (TLR3) regulation mechanisms and roles in antiviral innate immune responses. *J Zhejiang Univ Sci B*. (2021) 22:609. doi: 10.1631/jzus.B2000808

43. Daffis S, Samuel MA, Suthar MS, Gale M, Diamond MS. Toll-like receptor 3 has a protective role against West Nile virus infection. *J Virol.* (2008) 82:10349–58. doi: 10.1128/JVI.00935-08

44. Elshafei SO, Mahmoud NA, Almofti YA. Immunoinformatics, molecular docking and dynamics simulation approaches unveil a multi-epitope-based potent peptide vaccine candidate against the avian leukosis virus. *Sci Rep.* (2024) 14:2870. doi: 10.1038/s41598-024-53048-6

45. Case DA, Aktulga HM, Belfon K, Ben-Shalom I, Brozell SR, Cerutti DS, et al. *Amber 2021*. San Francisco: University of California (2021).

46. Suleman M, Balouch AR, Randhawa AW, Khan T, Muddassir M, Ullah A, et al. Characterization of proteome-wide antigenic epitopes to design proteins specific and proteome-wide ensemble vaccines against heartland virus using structural vaccinology and immune simulation approaches. *Microb Pathog.* (2022) 168:105592. doi: 10.1016/j.micpath.2022.105592

47. Shocket MS, Verwillow AB, Numazu MG, Slamani H, Cohen JM, El Moustaid F, et al. Transmission of West Nile and five other temperate mosquito-borne viruses peaks at temperatures between 23 C and 26 C. *Elife.* (2020) 9:e58511. doi: 10.7554/eLife.58511

48. Ulbert S. West Nile virus vaccines-current situation and future directions. *Hum Vaccin Immunother.* (2019) 15:2337-42. doi: 10.1080/21645515.2019.1621149

49. Delany I, Rappuoli R, Seib KL. Vaccines, reverse vaccinology, and bacterial pathogenesis. *Cold Spring Harb Perspect Med.* (2013) 3:a012476. doi: 10.1101/ cshperspect.a012476

50. Nayir T, Nazlican E, Şahin M, Kara F, Meşe EA. Effects of immunization program on morbidity and mortality rates of vaccine-preventable diseases in Turkey. *Turkish J Med Sci.* (2020) 50:1909–15. 10.3906/sag-2008-177

51. Habib A, Liang Y, Xu X, Zhu N, Xie J. Immunoinformatic Identification of Multiple Epitopes of gp120 Protein of HIV-1 to Enhance the Immune Response against HIV-1 Infection. *Int J Mol Sci.* (2024) 25:2432. doi: 10.3390/ijms25042432

52. Villanueva RA, Rouillé Y, Dubuisson J. Interactions between virus proteins and host cell membranes during the viral life cycle. *Int Rev Cytol.* (2005) 245:171–244. doi: 10.1016/S0074-7696(05)45006-8

53. Zhang L. Multi-epitope vaccines: a promising strategy against tumours and viral infections. *Cell Mol Immunol.* (2018) 15:182–4. doi: 10.1038/cmi.2017.92

54. Albaqami FF, Altharawi A, Althurwi HN, Alharthy KM, Tahir ul Qamar M, Muhseen ZT, et al. Development of a novel vaccine candidates against cardiobacterium valvarum through reverse vaccinology and computational approaches. *BioMed Res Int.* (2023) 2023. doi: 10.1155/2023/6325568

15