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*CORRESPONDENCE Ruixiao Song M18796972431@163.com

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Research progress of SIRTs activator resveratrol and its derivatives in autoimmune diseases

Xiaolong Yu^{1,2,3}, Mingkai Chen², Jiabiao Wu² and Ruixiao Song^{2,3*}

¹Jiangsu Key Laboratory of Immunity and Metabolism, Xuzhou Medical University, Xuzhou, Jiangsu, China, ²Wujin Hospital Affiliated with Jiangsu University, Changzhou, Jiangsu, China, ³The Wujin Clinical College of Xuzhou Medical University, Changzhou, Jiangsu, China

Autoimmune diseases (AID) have emerged as prominent contributors to disability and mortality worldwide, characterized by intricate pathogenic mechanisms involving genetic, environmental, and autoimmune factors. In response to this challenge, a growing body of research in recent years has delved into genetic modifications, yielding valuable insights into AID prevention and treatment. Sirtuins (SIRTs) constitute a class of NAD-dependent histone deacetylases that orchestrate deacetylation processes, wielding significant regulatory influence over cellular metabolism, oxidative stress, immune response, apoptosis, and aging through epigenetic modifications. Resveratrol, the pioneering activator of the SIRTs family, and its derivatives have captured global scholarly interest. In the context of AID, these compounds hold promise for therapeutic intervention by modulating the SIRTs pathway, impacting immune cell functionality, suppressing the release of inflammatory mediators, and mitigating tissue damage. This review endeavors to explore the potential of resveratrol and its derivatives in AID treatment, elucidating their mechanisms of action and providing a comprehensive analysis of current research advancements and obstacles. Through a thorough examination of existing literature, our objective is to advocate for the utilization of resveratrol and its derivatives in AID treatment while offering crucial insights for the formulation of innovative therapeutic approaches.

KEYWORDS

resveratrol, derivative, SIRTs, sirtuins, autoimmune diseases, SIRTs activators

1 Introduction

Autoimmune diseases (AID) represent a formidable threat to human health, stemming from the immune system's erroneous targeting of the body's own tissues. Recent years have witnessed a steady rise in AID incidence, underscoring its status as a critical global public health concern. AID presents with a spectrum of clinical phenotypes due to the production of specific immune cells and autoantibodies during inflammatory immune responses. These assaults on tissues or organs vary widely in their severity (1). Depending on the extent of tissue damage, AID is broadly classified into two principal categories: systemic autoimmune diseases, such as systemic lupus erythematosus (SLE), rheumatoid arthritis (RA), systemic sclerosis (SSc), among others; and organspecific autoimmune diseases, exemplified by Graves disease (GD), type 1 diabetes mellitus (T1DM), inflammatory bowel disease (IBD), pulmonary fibrosis (PF), and others. The clinical ramifications of AID are profound; for instance, SLE can afflict multiple organs and bodily systems, with severe cases precipitating lupus nephritis, lupus cerebritis, and other crises, posing life-threatening risks (2). RA is characterized by joint degradation, resulting in significant rates of disability (3). Severe manifestations of GD can precipitate cardiovascular irregularities and thyroid crises (4). Individuals with T1DM confront heightened risks of complications such as cardiovascular ailments, retinopathy, and neuropathy (5). Overall, AID is marked by a complex etiology, diverse clinical presentations, and myriad sequelae, exerting considerable impact on individuals' quality of life and productivity, and imposing substantial economic and psychological burdens on society and families. Presently, the treatment landscape for autoimmune diseases predominantly relies on immunosuppressive agents and biologics; however, these modalities often entail notable side effects, efficacy limitations, considerable interindividual variability, and an inability to effectuate cures. Consequently, the pursuit of precise and efficacious treatment modalities has emerged as a paramount objective in AID research.

2 Pathophysiological characteristics of autoimmune diseases

In normal circumstances, the immune system can recognize and respond to external threats, including pathogens and damaged tissues, thereby maintaining the body's health. However, when the immune system receives abnormal stimuli or its regulation is imbalanced, it can lead to excessive activation, resulting in inappropriate immune responses. It is noteworthy that this abnormal activation can manifest in various forms, such as the overactivation of immune cells, attacks on self-antigens, inflammatory reactions, and more. Additionally, genetic and environmental factors play crucial roles in the abnormal activation of the immune system. The primary mechanisms of immune system abnormal activation include several aspects.

Firstly, the immune system incorrectly identifies its own tissues or molecules as foreign pathogens, which is related to changes in the expression pattern of self-antigens or immune tolerance disorders. Immune tolerance disorder refers to the disruption of immune balance through central and peripheral immune tolerance disorders and abnormal presentation of self-antigens, causing the immune system to mistakenly perceive its own tissues as external threats, resulting in damage to self-tissues (6). Secondly, the abnormal function of immune regulatory cells (such as regulatory T cells) fails to effectively suppress excessive immune responses, leading to the occurrence of autoimmune diseases. Abnormalities in the number and function of regulatory T (Treg) cells, the inhibition of Treg cells by effector T cells, and the increase in abnormal T cell activity can all lead to abnormal activation of the autoimmune system (7). Among them, the balance between Treg cells and T helper 17 (Th17) cells is crucial, as it can restrain the malignant cycle of autoimmunity and block the pathways leading to autoimmune diseases. For instance, the impairment of Treg cells' stability and the abnormal proliferation of Th17 cells can lead to the activation of other immune cells, thereby driving acute autoimmune responses (8). Research has demonstrated that inhibiting the activity of IL-2 inducible T-cell kinase (ITK) with specific inhibitors can regulate the translocation of Foxo1 and effectively modulate the balance between Th17 and Treg cells (9). This pathway inhibits the phosphorylation process of phospholipase C-gamma 1 (PLC-y1), mitigating autoimmune reactions and diminishing the production of antigen-specific antibodies. Furthermore, evidence suggests that in the presence of antigenpresenting cells, CD4+ lymphocytes are prompted to activate RORgt via the JAK2/STAT3 pathway, leading to their differentiation into Th17 cells (10). Building on this, research conducted by Wang et al. indicates that obstructing the leptin pathway not only prevents the differentiation of Th17 cells but also enhances the transcription of FOXP3, fostering the conversion of CD4+ lymphocytes into Treg cells (11). Consequently, inhibiting the leptin pathway reinstates the equilibrium between Treg and Th17 cells, thereby reducing cell death attributed to autoimmune inflammation. Thirdly, when the immune system is abnormally activated, inflammatory mediators are excessively released, triggering immune inflammatory reactions. These inflammatory mediators, while regulating and transmitting immune signals, can also cause tissue inflammation and damage. Tumor necrosis factoralpha (TNF- α) is involved in the inflammatory processes of various autoimmune diseases such as RA and Crohn's disease (12). Interleukin-1 (IL-1) can cause fever and promote the production of other inflammatory mediators, and is associated with SLE (13). Transforming growth factor-beta (TGF-B) plays a crucial role in controlling immune cell activity and maintaining immune tolerance, but in some cases, it may also promote disease development (14).Fourthly, in Zhernakova's 2009 whole-genome study, some single nucleotide polymorphisms (SNPs) were found to share and have unique pathological pathways in the development of autoimmune diseases (15). Among them, the most prominent findings were within the MHC locus, which carries multiple signaling pathways from classical pathways of MHCI/II genes to non-classical pathways of MHCIII genes. Within the MHCIII region, multiple genes are involved, including genes encoding complement factor 4 (C4A) and tumor necrosis factor (TNF) (16). In SLE, the HLA-DR3 allele is associated with the production of anti-DNA

antibodies (17), while in Sjogren's syndrome, HLA-DRB1 is associated with the production of Ro antibodies (18). Therefore, the strong association between autoimmune diseases, specific autoantibodies, and specific MHC alleles plays an important role in the occurrence of autoimmune diseases. Fifthly, environmental factors such as infections, drugs, radiation, chemicals, etc., may trigger AID by affecting the function of the immune system or inducing the expression of self-antigens. Smoking is an important risk factor for diseases such as SLE, GD, idiopathic inflammatory myopathy, and IBD (19), and it can promote disease development through various pathways. Cigarette smoke activates innate immune responses and contains Toll-like receptor (TLR) stimulating compounds, directly activating TLR4 signaling and triggering the pro-inflammatory pathway of TLR4 agonists (20, 21). Signals are transmitted through molecules with genetic polymorphisms associated with systemic immunity, providing a direct connection and mechanistic basis for gene-environment interactions. Therefore, specific risk factors will increase the disease risk of individuals with specific genetic backgrounds. Of course, chemicals and drugs such as thiazides, hydralazine, calcium channel blockers, proton pump inhibitors, and interferon-alpha can also induce AID through various mechanisms, including inhibition of central or peripheral tolerance, changes in gene transcription in T and B cells, abnormal cytokine or cytokine receptor function, chromatin structure modification, and antigen modification (22).

3 The important role of SIRTs in autoimmune diseases

In 1979, Klar discovered, through mapping studies of the brewing yeast genome, that Sirtuins are mammalian homologs of the yeast silent information regulator 2 (Sir2), thus ushering in the era of research on the SIRTs family (23). Subsequent research indicated that Sirtuins are a class III histone deacetylase dependent on NAD+, present in both higher vertebrates and unicellular eukaryotes within the kingdom Animalia (24). In 1999, Frye et al. identified five human sirtuins, namely SIRT1, SIRT2, SIRT3, SIRT4, and SIRT5, using the amino acid sequence of Sir2 from brewing yeast as a probe (25). Later, the Frye team used human SIRT4 as a probe for similar identification and ultimately discovered two new human sirtuins (SIRT6 and SIRT7) (26). Thus, all seven members of the SIRTs family have been discovered, gradually entering people's field of vision.

In recent years, research on genetic epigenetic modifications has continued to deepen. Zentner's study revealed that histones can undergo various covalent translational modifications, including acetylation, methylation, phosphorylation, ubiquitination, and more (27). Subsequently, proteins are recruited to specific modified histones, serving as docking sites, to regulate all chromatin templating processes, such as replication, transcription, and DNA repair. This gene modification creates an open and accessible chromatin environment at all epigenetic markers on the genome, regulating the transcription and repair of DNA (28). However, in this process of genetic modification, enzymes catalyzing the reactions are crucial. The SIRTs family, as NADdependent histone deacetylases, can deacetylate lysine residues on histones, regulating the activity of many transcription factors (29). This leads to a silencing effect on numerous target proteins in epigenetics, such as Forkhead box O class (FoxOs), p53, nuclear factor-kB (NFkB), nuclear factor E2-related factor 2 (Nrf2), hypoxia-inducible factor-1 α (HIF-1 α), AMP-activated protein kinase (AMPK), b-catenin, peroxisome proliferator-activated receptor gamma coactivator 1-alpha (PCG-1a), proliferatoractivated receptor gamma (PPARg), and others (30). Through these regulatory mechanisms, SIRTs participate in physiological processes such as inflammation, oxidative stress, mitochondrial function, immune response, and cell differentiation. Particularly, by modulating various cellular signaling pathways, SIRTs play a crucial role in cellular metabolism and DNA repair, making them a promising therapeutic target. Their role in immune regulation includes modulating both innate and adaptive immunity.

3.1 The mechanism of SIRTs regulating innate immunity

Innate immunity represents the body's initial line of defense against pathogens, characterized by its nonspecific, rapid, and broad response. The innate immune system comprises a range of intrinsic immune cells and molecules present from birth, including natural killer (NK) cells, macrophages, monocytes, dendritic cells, neutrophils, eosinophils, and basophils, among others. Unlike adaptive immunity, innate immunity operates independently of prior pathogen exposure and does not confer immune memory against specific pathogens. It is noteworthy that the SIRTs family can modulate the function of innate immune cells at multiple levels. Generally, decreased sirtuin levels can precipitate various inflammatory states in the body, including autoimmunity, cellular senescence, and degenerative changes in tissues and organs.

3.1.1 SIRTs and macrophages

Macrophages arise from early embryonic erythro-myeloid progenitors or from adult infiltrating monocytes. Upon antigen stimulation, macrophages are activated and polarized into proinflammatory or anti-inflammatory phenotypes, namely classical activation (M1) macrophages and alternative activation (M2) macrophages. M1 macrophages execute cytotoxic and tissuedamaging pro-inflammatory functions, while M2 macrophages play a crucial role in resolving inflammation and tissue repair (31). Within the SIRTs family, SIRT1, SIRT2, SIRT6, and SIRT7 participate in multiple specific stimuli and downstream signaling events of macrophage polarization, with significant implications for balancing macrophage polarization. Several studies have shown that deletion of the SIRT1, SIRT2, and SIRT6 genes in macrophages results in a significant increase in acetylation levels of the p65 subunit of NF-kB, leading to increased expression of NF-kB target genes IL-6, TNF- α , and IL-1 β , inducing immune inflammation (32, 33). In other words, SIRT1, SIRT2, and SIRT6 inhibit inflammation in macrophages by deacetylating NF-kB (34). Rothgiesser et al.

found that SIRT2 interacts with p65 in mouse embryonic fibroblasts, leading to deacetylation of p65 subunit of NF-kB at Lys 310, thereby increasing the expression of p65 acetylationdependent target gene subsets, regulating the expression of specific NF-kB-dependent genes, and controlling a large number of target genes involved in immune and inflammatory responses, cell proliferation, differentiation, and apoptosis (35). Santos-Barriopedro et al. found that SIRT6 can bind to the histone methyltransferase Suv39h1 specific for H3K9me3 and induce monoubiquitination at the conserved cysteine residue in the PRE-SET domain of Suv39h1 (36). Upon activation of the NF-κB signaling pathway, SIRT6 upregulates the activity of the NF-KB inhibitor (IkBa) through cysteine monoubiquitination and chromatin eviction by Suv39h1, thus achieving inhibition of the NF-KB pathway and attenuating the aggravation of the immuneinflammatory response by SIRT6. In addition, the expression of SIRT7 decreases in an age-dependent manner in leukocytes of healthy individuals, and Phorbol-12-myristate-13-acetate(PMA)mediated monocyte-to-macrophage differentiation in the monocytic THP-1 cell line increases SIRT7 expression, while SIRT7 overexpression also increases differentiation of nonstimulated THP-1 cells (37).

3.1.2 SIRTs and NK cells

NK cells are cytotoxic lymphocytes that play a crucial role in innate immunity against viral infections and tumors. NK cells secrete perforins and granzymes, inducing apoptosis in target cells through the expression of cell death ligands on their surface. Additionally, NK cells secrete various pro-inflammatory cytokines, including TNF- α and interferon-beta (IFN- β), which play important roles in maintaining and amplifying immune responses through interactions with macrophages and dendritic cells (38). Studies have indicated a specific increase in the expression of SIRT2 in liver NK cells induced in mice with liver cancer, suggesting that SIRT2 promotes the activity of liver NK cells in response to hepatocellular carcinoma (HCC). Furthermore, overexpression of SIRT2 enhances the secretion of proinflammatory cytokines and cytotoxic granules by NK cells, thereby exhibiting increased anti-tumor activity (39). Additionally, SIRT2 activity is associated with increased phosphorylation of extracellular signal-regulated kinases 1/2 (Erk1/2) and p38 mitogen-activated protein kinase (MAPK), which are two important signaling pathways for NK cell activity (40). Therefore, SIRT2 holds potential value in enhancing the antitumor effects mediated by liver NK cells.

3.1.3 SIRTs and dendritic cells

Dendritic cells are antigen-presenting cells that, under steadystate conditions, exhibit high phagocytic activity and continuously present self-antigens to restrain T cell reactivity. Upon infection, dendritic cells mature, leading to an increased expression of costimulatory receptors, including CD80, CD86, and MHC-II molecules (41). When dendritic cells show weakened responses to pathogens but enhanced responses to self-antigens, it results in an increased expression of pro-inflammatory cytokines, leading to the breakdown of immune tolerance and inflammatory responses.

Research indicates that the expression of SIRT1 in dendritic cells increases when toll-like receptors (TLRs) are stimulated. Conversely, the absence of SIRT1 leads to changes in T cell polarization (42). In 2012, Alvarez et al. demonstrated that SIRT1 promotes the production of the signature cytokines IL-12 and TGF- β 1 in a hypoxia-inducible factor α (HIF1 α)-dependent manner, thereby regulating the differentiation of Th1 cells and Treg cells, participating in the process of immune tolerance regulation (43). Subsequently, Yang et al. found that in dendritic cells, SIRT1 interacts with interferon regulatory factor 1 (IRF1), a transcription factor associated with the expression of interleukin 27 (IL-27), and produces a deacetylation effect. This effect reduces the binding of IRF1 to the gene promoter of il-27p28, silencing it and decreasing the production of IL-27, promoting the differentiation of Th17 cells, which are a pro-inflammatory subset of T cells (44). In summary, SIRT1 is a major regulatory factor for cytokine production in dendritic cells and holds significant importance for the subsequent generation of T cell subsets. Apart from SIRT1, SIRT6 also participates in the differentiation and maturation of dendritic cells. Studies confirm that inhibiting SIRT6 significantly hinders the differentiation of monocytes into dendritic cells, resulting in immature dendritic cells with significantly reduced expression of CD86, CD80, and MHC-II molecules. Additionally, these dendritic cells exhibit increased phagocytic ability and a further decrease in the ability to stimulate lymphocyte proliferation (45, 46). At the same time, there is an increased proportion of cells producing TNF- α and IL-6, indicating that SIRT6 regulates the production of cytokines in these cells.

3.2 Mechanisms of SIRTs regulation in adaptive immunity

Adaptive immunity constitutes the highly specific and memorydriven immune response of an organism against specific pathogens. In contrast to innate immunity, which broadly recognizes nonspecific antigens present in pathogens and damaged host cells, the adaptive immune system possesses high antigen recognition specificity, immunological memory, and adaptability to diverse pathogens. Primarily composed of T cells and B cells, the adaptive immune system features B cell receptors (BCRs) or T cell receptors (TCRs) expressed on their cell membranes. While tolerating self-antigens, these cells have the capability to recognize and mount highly specific immune responses against particular antigens.

3.2.1 SIRTs and T lymphocytes

T lymphocytes comprise helper CD4+ T cells and cytotoxic CD8+ T cells, activated by TCR-specific antigens during cell-cell contact. Fagnoni et al. (47) propose an association between SIRT1 and the accumulation of CD8+CD28- T cells. Effros et al. (48) discovered that under conditions of CD28 co-stimulation

deficiency, CD8+CD28- T cells exhibit heightened cytotoxicity, express pro-inflammatory cytokines, and display characteristics indicative of replicative senescence. During aging, SIRT1 undergoes autophagy-mediated degradation in various organs, including the spleen and thymus. Jeng and colleagues observed a notable decrease in SIRT1 levels in human CD8+ memory T cells, particularly CD8 +CD28- T cells, without alterations in their gene expression. SIRT1, is a deacetylase for many transcription factors, including FOXO1, and regulates several cellular processes, such as proliferation, differentiation, and apoptosis. SIRT1 activation triggers apoptosis by directly inhibiting FOXO1 expression through deacetylation. Consequently, the loss of SIRT1 amplifies proteasomal degradation of its target FOXO1, thereby diminishing the cytotoxicity of memory T cells by enhancing their glycolytic capacity (49). Studies indicate a substantial decrease in SIRT2 levels in the spleens of aged rats, with an even more precipitous decline in SIRT2 levels in the thymus. Moreover, Sidler et al. (50) found that among the seven sirtuin genes encoded by the mammalian genome, only the expression level of SIRT2 decreases with age. This may be linked to the relationship between SIRT2 and Histone H4K16, a target of SIRT2 deacetylation in aging yeast (51). Reduced DNA methylation and H3K9me3 with age suggest the loss of heterochromatin as aging progresses. Additionally, SIRT6 plays a role in immune aging and inflammatory responses by regulating T cell inflammatory responses. A study of SIRT6 in aging demonstrated that targeted deletion of SIRT6 in T cells or the myeloid lineage recapitulated the inflammatory and fibrotic phenotype in the liver, suggesting autonomous regulation of inflammation by SIRT6 in immune cells (52). Importantly, SIRT1 levels are significantly upregulated in patients with GD, promoting the deacetylation of FOXP3 and mediating the simultaneous loss of IL-17+ T cells and Treg cells in Graves' patients (53). Overexpression of SIRT1 in patients with IBD induces concurrent inhibition of Th17 cell differentiation and Treg cell differentiation, exacerbating colitis development. In conclusion, SIRTs actively participate in T cell differentiation processes, influencing immune responses and contributing to the occurrence of autoimmune diseases.

3.2.2 SIRTs and B lymphocytes

B lymphocytes constitute the cornerstone of adaptive immunity. Mature B cells predominantly reside in the spleen and lymph nodes, displaying antigen-specific immunoglobulins on their cell membranes. Upon infection, antigen-specific B cell clones are activated, leading to their differentiation into plasma cells that secrete antibodies or memory B cells.

The intrinsic relationship between SIRTs and B lymphocytes is currently under exploration. Nonetheless, it is established that SIRTs are linked to immunoglobulin class switch recombination (CSR), B cell homeostasis, and autoimmune suppression. Research indicates a significant role for SIRT7 in non-homologous end joining (NHEJ) repair. Regarding NHEJ, as one of the main DNA double-strand break repair modes in cells. NHEJ is different from homologous recombination in that it does not require homologous DNA as a repair template and can repair broken DNA at all stages of the cell cycle (54). In addition, NHEJ is involved in the recombination of diversified antibodies and T-cell receptors. Reduced expression of SIRT7 during aging may partly contribute to the diminished clonal diversity of newborn B cells, associated with immunosenescence (55). In a study by Gan in 2020, it was discovered that the activation of immunoglobulin CSR is mediated by activation-induced cytidine deaminase, and its expression in mature B cells is subject to epigenetic regulation. Enhanced activity of SIRT1 can lead to the deacetylation of H3K9Ac/H3K14Ac in the promoter of activation-induced cytidine deaminase, resulting in reduced expression and impaired CSR (56). Moreover, SIRT1 is crucial for B cell antigen presentation, as the loss of SIRT1 in B cells leads to decreased levels of MHC-II molecules expressed, thereby reducing their cross-presentation with CD4+ T cells (57). In summary, the downregulation of SIRT1 with age may compromise B cell immunity and contribute to immunosenescence.

Notably, during immunosenescence, inflammation and AID are commonly observed. In mature B cells of SLE patients, significantly lower levels of SIRT1 have been observed, and SIRT1 levels are negatively correlated with the frequency of CD19+ B cells in these patients (58). Activation of SIRT1 has a protective effect on RA patients, partly due to a reduction in the production of autoantibodies by B cells (59). Serum levels of SIRT1 in asthma patients are positively correlated with IgE levels (60), while anti-SIRT1 autoantibodies are more abundant in inflammatory diseases such as ankylosing spondylitis (61). In summary, SIRT1, by inhibiting the autoimmune response of B cells and reducing the cascade of immune inflammation, plays a crucial role in preventing severe invasion of various systems and organs in the body, maintaining the stability of the immune environment, and ensuring the normal functioning of the immune system.

4 The development history of resveratrol and its derivatives

Resveratrol (RES) is a natural phytotoxin found in many plants, produced in response to ultraviolet radiation, damage, fungal, or bacterial infections (62). It is present in various natural foods, including grapes, blueberries, and mulberries, with significant variations in concentration among different foods (63). The chemical structure of RES consists of two aromatic rings connected by a methylene bridge, with both trans and cis isomers naturally occurring. The primary medicinal effects of RES stem from the trans-resveratrol, making the structure-activity relationship crucial in determining RES bioactivity (64). However, due to its rapid absorption and metabolism in the human body, RES exhibits low plasma concentrations and tissue distribution (65). Consequently, there has been widespread interest in finding methods to enhance the bioavailability of RES.

RES is a small molecule with a molecular weight of 228.247 g/mol and various functional groups, including phenolic hydroxyl, aromatic rings, and double bonds (66). These functional groups provide opportunities to modify RES into active derivatives with diversified therapeutic effects. Therefore, designing and synthesizing novel resveratrol derivatives to enhance the therapeutic effects of RES has become a hotspot of interest among pharmacologists. Over the past few decades, research on various natural and synthetic resveratrol derivatives, particularly methoxylation, hydroxylation, and halogenation derivatives, has received special attention. Here, we provide a comprehensive summary of the chemical structures, mechanisms of action, and clinical effects of methoxylated, hydroxylated, and halogenated derivatives of RES (Table 1).

In 2003, David Sinclair and colleagues utilized the innovative "FluordeLys" (FdL) method and discovered the first-generation SIRT1 activator – RES (93), also the first SIRTs activator discovered in humans. As a pioneer of SIRTs activators, RES has rich clinical applications and promising therapeutic prospects.

Combretastatin A-4, a cis-configured methylated resveratrol derivative, was first isolated from Combretum caffrum in 1995. However, it gained prominence in 2003 due to its phosphate salt, which selectively disrupts the microtubule protein cytoskeleton of endothelial cells (94), exhibiting potent anti-tumor and anti-vascular effects, leading to the development of the market drug ombrabulin (95).

In 2004, Rimando and colleagues found that Pterostilbene (96), structurally highly similar to RES, also possesses potent anti-tumor, anti-inflammatory, and antioxidant activities (97), serving as a naturally occurring dimethylated resveratrol derivative from blueberries. The dimethoxy structure of Pterostilbene enhances its lipophilicity and cellular membrane permeability, with better metabolic stability than RES, displaying superior pharmacokinetic properties (98). Research indicates that Pterostilbene can more effectively prevent azoxymethane (AOM)-induced colon cancer by activating the antioxidant signaling pathway mediated by NF-E2-related factor 2 (Nrf2) compared to RES (99).

Piceid, a non-glycosylated resveratrol derivative, is the most abundant form of RES in nature (100). It is primarily found in alcohol-free grape juice and is also a major extract component of the traditional herbal medicine Polygonum cuspidatum, used to treat heart diseases, including atherosclerosis and myocarditis (101). Jeong et al. discovered in 2010 that Piceid inhibits tyrosinase in melanocytes, thus blocking melanin production without adversely affecting cell viability. Additionally, Piceid inhibits tyrosinase, tyrosinase-related protein 1 (TRP1), tyrosinase-related protein 2 (TRP2) and microphthalmia-associated transcription factor (MITF), leading to reduced melanin synthesis (102). Hence, Piceid may be a potential candidate for a skin whitening agent. Recent studies show that the team led by Kobayashi et al. isolated a resveratrol glucosyltransferase gene from three grapevine species and introduced it into kiwi plants via Agrobacterium-mediated gene transfer, producing the glycosylated form of RES - piceid (103).

In 2011, Hsieh et al. observed in prostate cancer cells LNCaP that resveratrol derivatives triacetyl-resveratrol and trimethoxy-resveratrol are effective anti-CaP agents (104). Due to their evasion of further enzymatic glucuronidation or sulfation, they exhibit efficacy comparable to RES in chemoprevention and treatment. Furthermore, triacetyl-resveratrol preferentially targets the inhibition of CaP proliferation through the p53 signaling pathway, while trimethoxy-resveratrol acts by controlling the cell cycle and inducing apoptosis, achieving a dual anti-CaP activity (105). As the most extensively studied and structurally simplest resveratrol derivatives to date, tri-methylation and tri-acetylation

resveratrol derivatives play indispensable roles in regulating the growth and gene expression of prostate cancer cells LNCaP (106).

Pallidol is one of the most common resveratrol oligomers, relatively easy to obtain. It effectively scavenges reactive oxygen species (ROS) and activates the Keap1-Nrf2 pathway, upregulating the expression of antioxidant enzymes (107). In 2015, the team led by Matsuura B developed an efficient and scalable oxidative dimerization method for the tert-butylated resveratrol derivative, resulting in the unique and stable quinone methylation product pallidol, the most effective synthesis method of pallidol to date (108).

Duan et al. designed and synthesized a series of resveratrol derivatives in 2016, proven to be effective lysine-specific demethylase 1 (LSD1) inhibitors (109). Among them, the compounds (Z)-3-((E)-2bromo-4,5-dihydroxystyryl)-N'-hydroxybenzimidamide (4e) and (Z)-4-((E)-2-bromo-4,5-dihydroxystyryl)-N'-hydroxybenzimidamide (4m) were identified as the most effective LSD1 inhibitors, dose-dependently inducing an increase in dimethylation of Lys4 of histone H3 in MGC-803 cells, while significantly increasing the mRNA levels of CD86 (an alternative cellular biomarker of LSD1 activity) in MGC-803 cells (110), demonstrating potent intracellular inhibition of LSD1 with potential anticancer activity. Furthermore, inhibiting LSD1 can enhance the binding of histone H3 lysine 4 dimethylation (H3K4me2) to its promoter sequence, upregulating the expression of transferrin receptor (TFRC) and acyl-CoA synthetase long-chain family member 4 (ACSL4). Subsequently, this promotes the accumulation of intracellular iron and the synthesis of unsaturated fatty acids, leading to ferroptosis. Ferroptosis, to some extent, can inhibit tumor growth and metastasis. Therefore, these resveratrol derivatives may become a novel option for cancer therapy.

In 2020, Yang et al. found that Piceatannol (111), a hydroxylated resveratrol derivative, exhibits biological functions similar to RES (112). In MDA-MB-231 breast cancer cells, it inhibits cancer cell invasion, migration, adhesion, and reduces the activity of matrix metalloproteinase-9 (MMP-9), thereby suppressing breast cancer cell proliferation, invasion, and metastasis (111, 113). Meanwhile, Piceatannol reduces drug metabolism rates, increases its bioavailability, and surpasses RES in terms of drug potency by participating in multiple steps such as apoptosis, cell proliferation, and clonogenicity (114).

In 2022, Innets's research found that the methoxy derivative of RES, 4,4-(ethane-1,2-diyl) bis(2-methoxyphenol) (RD2), exhibits high affinity between the ATP binding site and the allosteric site of the Akt molecule (115). In other words, RD2 is a potential inhibitor of Akt. RD2 blocks the PI3K/Akt/mTOR pathway (116), leading to the activation of apoptosis, reduction in cell proliferation rate, and inhibition of cancer cell metastasis, making it a potential drug for the treatment of non-small cell lung cancer (117).

In 2023, Fragopoulou's research showed that the methoxy derivatives of RES have better antiplatelet effects, with the most effective derivative being the 4'-methoxy derivative, exhibiting approximately 2.5 orders of magnitude of antiplatelet activity against thrombin receptor activating peptide (TRAP)-induced platelet aggregation, indicating its potential as an antiplatelet agent comparable to protease-activated receptor-1 (PAR1) inhibitor vorapaxar (118).

Classification	Name	Chemical structure	Differences with resveratrol	Mechanism of action	Clinical effect	References
Methoxylated resveratrol derivatives	Pterostilbene		More lipophilic than resveratrol.	 Inhibits various signaling pathways. Suppresses ethanol-induced DNA oxidative damage. 	 Inhibits oxidative stress. 2. Reduces the chemosensitivity of pancreatic cancer, melanoma, leukemia, breast cancer, lung cancer, and gastric cancer. Promotes apoptosis. 4. Anti-invasive, anti-metastatic, and anti-inflammatory. Treats hypertension and diabetes. 	(67–70)
	Trimethoxystilbene		Stronger by 30-100 times in inhibiting endothelial cell proliferation and morphological transformation compared to resveratrol.	1. Downregulates the activity of the PI3K/AKT signaling pathway. 2. Acts as an anti-angiogenic compound to block neovascular formation. 3. As a vascular-targeting agent, it can cause rupture of immature blood vessels. 4. Promotes bundling of microtubules and formation of spot-like structures of DCLK1-microtubule complexes.	1. Used in chemotherapy for breast cancer, lung cancer, and liver cancer. 2. Vascular-targeted therapy. 3. Inhibits HCV.	(71-73)
	Tetramethoxy stilbene	, , , , , , , , , , , , , , , , , , ,	Possesses higher biological potency and bioavailability than resveratrol.	 Induces the release of a large amount of cytochrome C into the cytoplasm by increasing mitochondrial permeability at the nuclear periphery. Exhibits anti-angiogenic activity by inhibiting the phosphorylation of multiple downstream signaling components of VEGFR2. 	1. Inhibits the growth of various cancers, including colon cancer, prostate cancer, ovarian cancer, and liver cancer. 2. Inhibits cytochrome P450. 3. Regulates hypertension and cardiac fibrosis.	(74–77)
	Pentamethoxystilbene		Far exceeds resveratrol in inhibiting cell growth.	Induces cell cycle G1 phase arrest and regulates G1 phase proteins.	1. Inhibits the growth of breast cancer and colon cancer. 2. Inhibits cytochrome P450.	(78, 79)
Hydroxylated resveratrol derivatives	Dihydroxystilbenes		Exhibits stronger biological effects than resveratrol.	1. Inhibits cancer progression by blocking G1 phase cell transition and tumor growth. 2. Reduces the secretion of endothelin-1 and mRNA levels of endothelin-1 in human endothelial cells. 3. Protects against hemin-induced lipid peroxidation and ROS production.	1. Used in chemotherapy for lung cancer and breast cancer. 2. Manages vascular abnormalities. 3. Inhibits oxidative stress reactions.	(80, 81)

(Continued)

TABLE 1 Continued

Classification	Name	Chemical structure	Differences with resveratrol	Mechanism of action	Clinical effect	References
	Tetrahydroxystilbene		Stronger anti-oxidant and anti-cancer activity than resveratrol.	1. Inhibits oxidation of LDL-C in plasma, platelet aggregation, and inflammation. 2. Antiproliferative, cytotoxic, and inhibitory effects on cancer cell growth. 3. Inhibits mutations in the leucine-rich repeat kinase-2 and uptake of 5-hydroxytryptamine.	 Regulates atherosclerosis, hypertension, and myocardial ischemia. Reduces the chemosensitivity of liver cancer, leukemia, and cervical cancer. 3. Inhibits cell oxidation. 4. Antibacterial. Effective against Parkinson's and Alzheimer's disease. 	(82-84)
	Hexahydroxystilbene	$ \begin{array}{c} & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ $	Most effective radical scavenger among resveratrol derivatives, with higher anti-HIV-1 activity than resveratrol.	1. Causes imbalance of deoxyribonucleotide triphosphate pools within cells. 2. Inhibits virus attachment and reverse transcription.	1. Inhibits the growth of various malignant tumors, including breast cancer, colon cancer, leukemia, melanoma, and glioma. 2. Inhibits HIV-1. 3. Selective COX-2 inhibitor. 4. Inhibits oxidative stress.	(85–87)
	(E)-2,6-dibromo-4-(3,5- dibromostyryl)phenol		Higher bioavailability than resveratrol.	Stabilizes the natural tetramer of transthyretin and modifies the quaternary structure of monomeric transthyretin in solution.	Cardioprotective effects.	(88)
Halogenated resveratrol derivatives	(E)-3,5-di-fluoro- 4'-acetoxystilbene		Stronger anti-tumor activity than resveratrol.	Inhibits upregulation of ABC superfamily cell transport proteins.	Resists cell proliferation.	(89)
	3,4,5-trimethoxy-4'-brom- cis-stilbene	of of of other states of the s	Stronger anti-cancer effects than resveratrol.	Inhibits G2/M phase tumor cell growth.	Inhibits lung cancer cell growth.	(90)

(Continued)

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TABLE 1 Continued

Classification	Name	Chemical structure	Differences with resveratrol	Mechanism of action	Clinical effect	References
	4'-Bromo-Resveratrol		More effective in inhibiting SIRT3 than resveratrol.	Effectively inhibits SIRT1 and SIRT3 by extending the bromophenyl moiety of the active site.	Therapeutic effects on aging, transcription, apoptosis, and inflammation-related diseases.	(91)
	2-bromo-resveratrol	HO FOH BIT COH	MIC values against Candida albicans are 3 times lower than resveratrol.	_	1. Inhibits microbial growth. 2. Inhibits cell proliferation.	(92)
	2-chloro-resveratrol	HO CI OH	MIC values against Candida albicans are 30 times lower than resveratrol.	_	1. Inhibits microbial growth. 2. Inhibits cell proliferation.	(92)

Here, we have retrospectively reviewed the discovery timeline and development process of resveratrol and its derivatives, providing a brief overview of their specific efficacy in clinical applications (Figure 1).

5 Resveratrol and its derivatives in the research progress of autoimmune diseases

5.1 Systemic autoimmune diseases

5.1.1 Rheumatoid arthritis (RA)

RA is a systemic autoimmune disease characterized by erosive joint damage and systemic inflammation, affecting multiple systems (119). It is among the most prevalent systemic autoimmune diseases, with increasing prevalence and disability rates annually, particularly among middle-aged and elderly women aged 30–60, in regions such as North America, Europe, and Asia (120). The hazards of RA primarily involve joint damage and deformity, alongside systemic inflammation and elevated cardiovascular risk, significantly impacting patients' quality of life and imposing a substantial global economic burden (121–123).

Oxidative stress is widely recognized as a major contributor to inflammation and arthritis. An increase in chemical reactions within the body, damage to antioxidant defense systems, and an imbalance between oxidants and antioxidants can lead to a sharp rise in ROS, culminating in the accumulation of reactive oxygen molecules and defects in DNA, RNA, and proteins (124). Studies have demonstrated that RES effectively scavenges radicals such as hydroxyl and superoxide, thereby inhibiting lipid peroxidation and DNA damage induced by excessive ROS production (125). Consequently, RES exhibits remarkable anti-inflammatory and antioxidant effects in RA. During oxidative stress, nuclear factor erythroid 2-related factor 2 (Nrf2), a transcription factor with antioxidant and anti-inflammatory properties, counteracts mitochondrial decay by interacting with antioxidant response elements (AREs) (126). Building upon this theoretical foundation, Li et al. revealed that RES can prevent calcium deposition and mitochondrial decay by activating the SIRT1/Nrf2 pathway, thereby inhibiting the phosphorylation and acetylation of NF-κB p65 and suppressing synovial hyperplasia and joint inflammation (127).

Given SIRT1's capability to deacetylate various transcription factors, including NF- κ B, it can modulate downstream cellular pathways associated with oxidative stress (128). Consequently, activating SIRT1-mediated NF- κ B deacetylation to inhibit oxidative stress and inflammation has emerged as a potential therapeutic strategy for RA. In 2012, Rathore identified that NF- κ B p65 directly targets the miR-29a-3p and miR-23a-3p promoters, downregulating the transcription levels of miR-29a-3p and miR-23a-3p, thus inducing immune inflammatory responses and oxidative stress in the body, leading to joint synovial damage (129, 130). Subsequently, Wang's research in 2019 demonstrated that RES can inhibit the acetylation and phosphorylation of NF- κ B p65, increase the transcriptional activity of miR-29a-3p and miR-



23a-3p promoters, block fibroblast-like synoviocyte (FLS) proliferation, and mitigate oxidative stress induction mediated by miR-29a-3p and miR-23a-3p promoters, thereby alleviating ROS-induced damage to joints and surrounding tissues (131). In summary, RES regulates the activation of the SIRT1/NF-κB/miR-29a-3p/Keap1 pathway and the SIRT1/NF-κB/miR-23a-3p/cul3 pathway, subsequently activating the Nrf2-ARE signaling pathway, effectively inhibiting oxidative stress resulting from ROS accumulation and mitigating joint inflammation, positioning it as a promising candidate drug for RA prevention and treatment.

Thalhamer et al.'s research highlights the pivotal role of MAPK signaling pathways in RA, particularly emphasizing the significance of p38 MAPK (132, 133). Essentially, ROS trigger inflammation by activating the MAPK signaling pathway, which subsequently induces the production of pro-inflammatory cytokines, perpetuating a detrimental cycle of immune-driven inflammation resulting in severe bone erosion (134). In 2018, Yang et al. demonstrated that the functional inhibition of p38 MAPK by RES underpins its protective effects against RA progression (135). Specifically, RES can activate SIRT3, thereby restoring enzymatic activities of isocitrate dehydrogenase 2 (IDH2), superoxide dismutase 2 (SOD2), and glutathione peroxidase (GSH-Px) in endothelial cells, and promoting the deacetylation of SOD2 and forkhead box O3A (FoxO3A) (136, 137). Through this mechanism, RES diminishes ROS accumulation, inflammation, and angiogenesis in synovial tissue, inhibits p38 MAPK activation, and consequently exerts anti-inflammatory, anti-angiogenic, cellinhibitory, and pro-apoptotic effects, exhibiting preventive potential against RA both in vitro and in vivo. Therefore, targeting p38 MAPK represents a promising approach for RA treatment, and RES emerges as a promising candidate for clinical RA therapy.

Khojah and colleagues conducted a randomized controlled clinical trial to investigate the efficacy of RES therapy in treating RA, wherein they found that RES effectively reduces the levels of TNF- α , IL-1 β , IL-6, monocyte chemoattractant protein 1, and soluble receptor activator of NF-KB ligand in both serum and joint tissues (138). Subsequent studies indicate that RES achieves this reduction by upregulating SIRT1 expression levels, resulting in a significant decrease in NF-KB expression and thereby mitigating inflammation and bone degradation (139). Furthermore, RES demonstrates regulatory effects on inflammatory arthritis in rodents by selectively inhibiting cellular and humoral responses associated with RA development (59). Additionally, RES induces apoptosis in RA patients' fibroblast-like synoviocytes (FLS) by activating caspase-8, leading to substantial apoptotic cell death through both mitochondrial signaling pathway convergence and caspase-dependent pathways (140). Consequently, the emergence of RES presents a promising, efficacious, and safe therapeutic option for RA treatment (141).

5.1.2 Systemic lupus erythematosus (SLE)

SLE patients produce autoantibodies such as antinuclear antibodies (ANA) that attack their own tissues and organs, releasing large amounts of inflammatory mediators such as TNF- α and IL-6, thereby triggering immune and inflammatory responses (142). SLE is a globally distributed systemic autoimmune disease

involving genetic susceptibility genes and environmental factors such as infections, drugs, and ultraviolet light in its pathogenesis (143). Currently, SLE is widely recognized worldwide due to its complex clinical manifestations and severe complications. Its hazards mainly include lupus nephritis, cardiovascular diseases, central nervous system damage, risks of infection and bleeding, affecting female fertility, and increasing pregnancy risks. Severe cases may lead to systemic multi-organ damage and lupus crisis (144–148).

Research indicates a close relationship between CD4+ T cells and B cells in SLE (149). In humoral immunity, helper T cells interact with B lymphocytes, stimulating their proliferation and differentiation, leading to the production of autoantibodies against nuclear components, thereby attacking self-tissues and organs (150), ultimately triggering systemic autoimmune diseases. However, the SIRT1 activator, RES, inhibits activator protein-1 (AP-1) transcriptional activity by up-regulating SIRT1 expression levels, further inhibiting T cell activation and maintaining peripheral T cell tolerance (151). Simultaneously, RES can also induce the deacetylation of Fas, Bcl-2, Bax, and p53 by activating SIRT1 and SIRT3, and trigger Caspase 9-mediated cell apoptosis (152, 153). This mechanism achieves an inhibitory effect on CD4+ T cells and B cells, leading to a reduction in the production of antinuclear antibodies and the suppression of SLE immune inflammation-mediated damage to systemic tissues (154).

In 2010, Dorrie and colleagues discovered that antigenpresenting cells (APCs) present antigens to initial T cells to initiate immune responses during the recognition phase of the immune response, and some APCs present antigens to differentiated T cells during the effector phase to trigger antigen elimination. RES can influence the maturation of dendritic cells and their antigen-presenting ability in vitro (155). Additionally, RES can inhibit COX expression by suppressing NF-KB activation and inhibit TNF- α -induced inflammation response in fibroblasts by exciting SIRT1 (156), thereby reducing the deposition of immune complexes in the kidneys (157). Therefore, inflammation inhibition mediated by RES may also be beneficial in reducing the occurrence of lupus nephritis. In their investigation of lupus nephritis, Miyake et al. delineated the pivotal roles of Th1 and Th17 cells in the pathogenesis of diffuse proliferative lupus nephritis, while emphasizing the significance of Th2 cell cytokines in membranous lupus nephritis (158). Subsequently, Wang et al. utilized RES as a therapeutic intervention and elucidated its dosedependent reduction in the Th1/Th2 cell ratio, thereby ameliorating the deleterious effects associated with diffuse proliferative lupus nephritis (159). Moreover, this study yielded a groundbreaking discovery, demonstrating that RES effectively mitigates proteinuria, diminishes IgG and IgM deposition in renal tissues, and attenuates renal histopathological lesions. These findings collectively suggest that RES holds promise in halting the progression of lupus nephritis.

Neuropsychiatric lupus erythematosus (NPSLE) is mainly characterized by brain vasculitis mediated by vascular endothelial growth factor (VEGF) (160). The polymorphism of genes encoding VEGF is closely related to the increased incidence of neuropsychiatric lupus (161). In 2018, Kalinowska-Lyszczarz et al. treated lupus-prone atherosclerotic mice with RES and found a

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significant decrease in VEGF expression, indicating that RES can reduce the occurrence of NPSLE inflammation, promote neuronal survival, and improve memory (162). The latest research indicates that any trends observed in RES-treated NPSLE mice are A2A receptor-dependent (163). Moreover, in microglia, RES counteracts the negative effects of A2A receptors by exciting SIRT1, reducing the expression of fractalkine (CX3CL1), inhibiting inflammatory responses, and improving cognitive function (164). In conclusion, the anti-inflammatory properties of RES can exert strong protective effects in NPSLE.

5.1.3 Systemic sclerosis (SSc)

SSc is a chronic autoimmune connective tissue disease characterized by fibrosis and vascular abnormalities of the skin and internal organs. The disease usually severely affects the skin, oesophagus, lungs, heart, kidneys, and other organ systems, and manifests itself in a diverse range of clinical symptoms and modes of disease progression (165). The onset of SSc is geographically and ethnically diverse. The incidence of SSc is notably higher in regions such as Europe, North America, and Australia, while relatively low rates are observed in Asia, Africa, and Latin America. Additionally, the prevalence of SSc is lower among individuals of African descent and Asians, whereas those of European descent and Indian Americans exhibit higher rates. Typically, SSc affects individuals in the young to middle-aged demographic, spanning ages 20 to 50. Factors contributing to its onset include infections, occupational exposures, substance use, and lifestyle choices (166). The risks associated with SSc are multifaceted. Firstly, there's the potential for damage to both the skin and internal organs. SSc induces fibrosis and dysfunction in these areas, where skin sclerosis can significantly impact appearance and quality of life, while organ involvement may result in compromised function, such as pulmonary fibrosis, cardiac issues, and renal impairment. Secondly, vascular complications are common. SSc often presents with both microangiopathy and macrovascular lesions, potentially leading to symptoms like Raynaud's phenomenon and hypertension. Thirdly, there's immune system dysregulation to consider. SSc is characterized by autoimmune processes, wherein aberrant immune activation can spur the production of autoantibodies and inflammatory cytokines, precipitating tissue inflammation and damage. Lastly, joint and muscle involvement can occur, manifesting as arthritis, joint swelling, and muscle weakness, which can significantly hinder daily activities and motor function (167).

Research suggests that the decline in SIRTs levels and activity contributes to the pathogenesis of SSc (168). Conversely, activating SIRTs can induce anti-fibrotic effects, offering therapeutic benefits for SSc patients. Specifically, when SIRT1 is activated, it inhibits the TGF-b/SMAD signaling pathway, reducing collagen release by fibroblasts and blocking tissue fibrosis (169). Notably, Zhu and colleagues discovered that RES inhibits inflammatory factor expression by activating the SIRT1/mTOR signaling pathway. By downregulating mTOR, it diminishes collagen levels, alleviating skin inflammation and fibrosis in SSc (170). Additionally, RES enhances SIRT3 expression, increases intracellular TGF-b deacetylation, hinders intracellular TGF-b signaling and fibrotic responses, and mitigates the activated phenotype of fibroblasts in SSc (171). This leads to a substantial reduction in mitochondrial and cytoplasmic ROS accumulation in fibroblasts, lessening oxidative stress damage (172). In conclusion, RES improves fibrosis by activating SIRT1 and SIRT3, thereby inhibiting TGFb-induced signal transduction. Moreover, RES activates SIRT1 to obstruct the mTOR pathway, restraining collagen fiber accumulation. Consequently, RES emerges as a potential pharmaceutical intervention for treating SSc.

5.2 Organ-specific autoimmune diseases

5.2.1 Graves disease (GD)

GD arises from the immune system erroneously attacking thyroid tissue, leading to hyperactivity of the thyroid gland and excessive secretion of thyroid hormones, causing an elevated metabolic rate. Approximately one-third of patients may also develop eye conditions, such as protruding eyes and eyelid swelling, known as Graves' ophthalmopathy (GO) (173). GD, a prevalent autoimmune disease globally, predominantly affects women aged 20–40, with genetics and environmental factors closely linked to its onset. Particularly, environmental factors such as smoking, excessive iodine intake, and ionizing radiation play significant roles (174). Hazards associated with GD encompass thyroid dysfunction, thyroid storm, eye complications, an increased risk of cardiovascular diseases, and osteoporosis (175–178).

Kim et al. illustrated that RES inhibits adipogenesis in orbital fibroblasts associated with GO (179). Acting as a broad-spectrum SIRTs activator, RES diminishes the rate of ROS generation triggered by oxidative stress and lowers heme oxygenase-1 (HO-1) levels through the activation of SIRT1/3. This concurrent reduction extends to superoxide dismutase Cu/Zn-SOD (SOD1), catalase, and thioredoxin (Trx), thereby reducing adipocyte numbers and lipid droplet accumulation. Furthermore, RES induces adipocyte apoptosis within fibroblasts (180, 181). Consequently, RES holds promise in mitigating orbital lipid deposition, ameliorating ocular complications of GO, and emerging as a significant therapeutic modality for managing GD and its ocular sequelae. Nonetheless, the research in this domain remains limited, necessitating further investigation into RES's therapeutic effects on GO via the SIRT1 signalingdependent pathway.

5.2.2 Type 1 diabetes mellitus (T1DM)

T1DM is a chronic metabolic disorder characterized by insufficient or complete lack of insulin production, leading to elevated blood sugar levels. This disease is typically caused by autoimmune destruction of pancreatic β -cells, rendering insulin unable to be effectively secreted (182). T1DM is more common in children, adolescents, or young adults and is associated with factors such as family history, viral infections, early-life environment, and dietary factors. It is worth noting that T1DM is associated with other autoimmune diseases (such as RA, autoimmune thyroid diseases, etc.), suggesting the presence of common genetic or immune factors (183). Its hazards mainly include several aspects: Firstly, cardiovascular diseases. Hyperglycemia increases the risk of cardiovascular diseases, including myocardial infarction, stroke, atherosclerosis, etc., which are among the main causes of death in diabetic patients (184). Secondly, retinal lesions. High blood sugar damages retinal blood vessels, leading to retinal lesions and potential blindness (185). Thirdly, renal damage. High blood sugar harms the kidneys, leading to diabetic nephropathy, which may require dialysis or kidney transplantation in severe cases (186). Fourthly, neuropathy. Hyperglycemia damages the nervous system, causing peripheral neuropathy (such as sensory abnormalities, pain) and autonomic neuropathy (such as gastrointestinal dysfunction, cardiovascular dysfunction) (187). Fifthly, foot complications. Neuropathy and vascular changes increase the risk of foot infections and ulcers, potentially leading to amputation (188).

In 2015, Côté et al.'s study showed that compared with saline, RES significantly increased the levels of NAD+ and its ratio to NADH in the duodenal mucosa (189). Subsequently, by detecting the acetylation level of liver kinase B1 (Lkb1) in cells after RES treatment, it was further confirmed that RES significantly increased SIRT1 activity (190-192). Specifically, RES activates AMPK, thereby activating SIRT1 to achieve its biological effects (192). The activation of the Glp1r-Pka dependent signaling in the duodenum is downstream of the metformin-AMPK sensory pathway (193), so the duodenum naturally becomes a site of action for RES. Therefore, RES reduces hepatic glucose production (HGP) by activating the duodenal AMPK-Sirt1→Glp1r-Pka dependent neural pathway, exerting a hypoglycemic effect similar to metformin (194). This not only lays the foundation for the development of intestine-specific targeted therapy but also collaboratively reduces HGP and blood sugar in diabetic and obese patients, becoming a new targeted treatment for diabetes.

RES is not only one of the new therapeutic drugs for lowering blood sugar but also has broad prospects in the prevention and treatment of diabetes-related complications. In 2016, Park et al. showed that RES can increase the expression of adiponectin receptors 1/2 (AdipoR 1/2) and activate the AMPK-SIRT1-PGC- 1α axis, thereby alleviating endothelial dysfunction caused by lipotoxicity, oxidative stress, and apoptosis, and preventing diabetic nephropathy (195, 196). At the same time, RES can also prevent diabetes-induced nephritis and mesangial cell proliferation by inhibiting the Akt/NF κ B pathway (197). Subsequent studies have found that RES reduces ROS production and lowers blood sugar by activating the AMPK pathway and inhibiting the expression of NADPH oxidase 4 (NOX 4) in renal tubular epithelial cells (198). In summary, RES reduces renal fibrosis and restores renal function by regulating the AMPK/SIRT1/NOX 4/ROS signaling pathway (199), achieving prevention and control of diabetic nephropathy, and reducing the immune-inflammatory damage of the kidneys caused by diabetes.

Negi et al. found that long-term high blood sugar levels can cause oxidative stress in the body, leading to the activation of the transcription factor NF- κ B in peripheral neurons (200). However, NF- κ B can mediate the release of a large number of proinflammatory cytokines such as iNOS, TNF- α , IL-6, and COX-2, thereby driving nerve damage mediated by neuroinflammation and causing peripheral neuropathy (201). In 2020, Huang et al. found that RES can reduce the inflammatory mediators in diabetic neuropathy (202). Specifically, RES upregulates the expression of SIRT1 and downregulates the expression of NF- κ B, thereby reducing the activity of pro-inflammatory factors (such as TNF- α , IL-6, and iNOS) associated with neurodegeneration (203). Furthermore, RES can also ameliorate sensory motor disturbances associated with diabetic neuropathy by inhibiting the activity of TNF- α and nicotinamide adenine dinucleotide phosphate oxidase through activating SIRT1 (204, 205). Therefore, RES reduces the release of pro-inflammatory factors in diabetic patients, prevents oxidative stress from attacking neuronal cells, protects nerve function, and alleviates neuropathy.

In vitro studies have shown that RES reduces the expression of VEGF in human retinal endothelial cells and inhibits high glucoseinduced cell proliferation (206). It can also inhibit the migration of retinal endothelial cells by activating the PI3K/Akt pathway mediated by matrix-derived factors (207). Therefore, RES has potential value in preventing diabetic retinal lesions.

5.2.3 Inflammatory bowel disease (IBD)

IBD is a group of chronic intestinal disorders, mainly including Crohn's disease (CD) and ulcerative colitis (UC). The hallmark of these diseases is chronic inflammation of the intestinal mucosa, often accompanied by ulcer formation (208). The incidence of IBD varies in different regions, with higher rates in North America, Europe, and Australia, and relatively lower rates in Asia, Africa, and Latin America. IBD is more common in adolescents and young adults aged 20–40 and is associated with genetic factors, lifestyle, dietary habits, and infections. Particularly, smoking is closely associated with an increased risk of IBD (209). Its hazards mainly include intestinal inflammation and ulcers, malnutrition, intestinal obstruction, intestinal fistulas, and an increased risk of colorectal cancer (210–212).

Many in vitro and animal studies have shown that RES can reduce the severity of intestinal inflammation in IBD. However, the beneficial effects of RES on treating IBD are attributed to multiple mechanisms, which ultimately lead to the inhibition of the inflammatory cascade response (213). Ren et al. demonstrated that RES primarily exerts its inhibitory effect on NF-KB by activating SIRT1, thus mitigating the severity of IBD (214). Specifically, RES suppresses the transcriptional activity of p65 and the ubiquitination of NF-KB essential modulator (NEMO) in a dose-dependent approach, thereby inhibiting NF-KB activation mediated by the NF-KB kinase. Singh et al. found that RES not only downregulates TH1 responses to reduce the secretion of inflammatory factors such as IL-1 β , IL-6, and TNF- α in colitic mice but also reduces the percentage of CXCR3+ T cells. More importantly, RES significantly induces the expression of immunosuppressive CD11b+ Gr-1+ myeloid-derived suppressor cells (MDSCs) in the colon, which helps suppress local effector T cell responses (215). Furthermore, RES markedly diminishes the populations of inflammatory CD4+ and CD8+ T cells, B cells, NK cells, and myeloid-derived suppressor cells (MDSCs), underscoring its efficacy in reversing the advancement of chronic colitis (216).

In a 2012 study on UC, it was proposed that IL-17, IL-10, and transforming growth factor $\beta 1$ (TGF- $\beta 1$) participate in the

development of TH17 through two important pathways: mTOR-HIF-1 β -TH17 and IL-6-STAT3-HIF-1 β -TH17. RES precisely regulates the balance between Treg and TH17 cells by activating the SIRT1/mTOR/ HIF-1 β /TH17 signaling pathway, leading to cell cycle arrest in intestinal smooth muscle cells, increased apoptosis, and decreased collagen synthesis (217). Subsequently, in a study on CD, RES effectively reduced the expression of insulin-like growth factor 1 (IGF-1) and procollagen mRNA by activating SIRT1, thereby significantly decreasing the activity of inflammatory cytokines (IL-1 β , IL-6, and TNF- α) and the fibrosis-promoting factor (TGF- β 1) (218). This study elucidated that RES not only mitigates the inflammatory response but also impedes the fibrotic process of the cecal wall.

Furthermore, Arslan et al. found that RES can enhance the activity of superoxide dismutase (SOD), catalase (CAT), and glutathione (GSH) *in vivo* (219). The study report by Serra showed that RES is more effective than 5-aminosalicylic acid and can activate two pathways, nuclear factor erythroid 2-related factor 2 (Nrf2) (220), and PPAR- γ in human intestinal cells (221, 222). It is worth noting that these two pathways are currently considered as new therapeutic targets for IBD.

In summary, RES can inhibit ROS in the intestines and increase the activity of antioxidant enzymes through multiple pathways, thereby suppressing oxidative stress to maintain the balance between oxidants and antioxidants in the body, to some extent reducing the activity of IBD and improving the quality of life of patients with IBD.

5.2.4 Pulmonary fibrosis (PF)

PF is a chronic progressive lung disease and a component of connective tissue disorders. Its hallmark is the excessive proliferation and deposition of fibrous tissue in the lung tissue, leading to irreversible lung damage. This fibrosis can damage the alveoli and lung interstitium, making the lungs stiff and losing elasticity, thereby affecting respiratory function (223). PF has a relatively low incidence but is increasing annually, occurring more frequently in adult males over 60 years of age, and is associated with prolonged exposure to harmful chemicals (such as silica dust, asbestos, silicates, etc.) and inhalation of toxic gases (such as nitrogen oxides, iron oxide dust, etc.) (224). Its hazards mainly include the following aspects: firstly, impaired respiratory function. PF leads to excessive fibrous tissue proliferation in the alveoli and lung interstitium, thereby affecting lung function. Impaired lung function can cause symptoms such as dyspnea, shortness of breath, and in severe cases, may even require respiratory assistance from a ventilator, severely affecting the patient's quality of life (225). Secondly, cardiovascular complications. PF can lead to the occurrence of cardiovascular complications such as pulmonary arterial hypertension and right heart failure, which can be lifethreatening in severe cases (226). Thirdly, increased risk of cancer. Long-term chronic inflammation and tissue damage in the lungs may increase the risk of developing lung cancer (227).

In 2016, Li et al. discovered that the SIRT1 agonist RES reduces systemic oxidative stress by activating the Nrf2-related antioxidant defense mechanism, thus exerting a protective effect on PF (228). Importantly, Nrf2 can induce various downstream signaling targets, including NAD(P)H quinone dehydrogenase 1 (HO-1/NQO1), NADPH oxidase 4 (NOX4), and GSH. These signaling targets participate in the formation of multiple signaling pathways, such as the SIRT1-Nrf2-HO-1 pathway, thereby achieving anti-fibrotic effects both in vivo and in vitro through mechanisms such as inhibiting immune-inflammatory responses and oxidative stress, which significantly attenuates PF (229). The latest research indicates that RES can alleviate PF by inhibiting TGF-β-activated kinase 1 (TAK1) (230). TAK1 is a kinase involved in TGF-β activation, participating in fibroblast proliferation, collagen deposition, and scar formation (231). Zhang et al. demonstrated that moderate concentrations of RES can activate SIRT1, leading to enhanced expression of AMPK, reducing the activity of inflammatory cytokines such as IL-1ß and macrophage inflammatory protein- 1α (mip- 1α), thereby preventing nitric oxide (NO) release, inhibiting iNOS expression, and suppressing NF-KB nuclear translocation (232). In studies on human alveolar epithelial cells (AEC2), RES upregulates the expression of SIRT1, thereby improving mitochondrial membrane potential, reducing ROS levels, decreasing cell apoptosis, ultimately reducing high oxygeninduced cell damage (233). Additionally, the activation of the SIRT1/p53 signaling pathway by RES can also delay the aging of AEC2 and participate in the treatment process of PF through its antioxidant and anti-inflammatory responses (234). Therefore, the SIRT1 and Nrf2 pathways induced by RES are likely to become potential therapeutic targets for PF.

6 The challenges faced in the clinical application of resveratrol and its derivatives

In recent years, there has been a surge in attention toward resveratrol and its derivatives as activators of the SIRTs family, following the discovery of their roles in metabolic regulation, cell apoptosis, cancer prevention, and other cellular processes. These compounds are recognized for their potential clinical benefits, including antioxidative, anti-inflammatory, anti-cancer, and cardiovascular protective properties. However, despite their broad range of potential applications, the clinical development of RES is hindered by its remarkably low bioavailability and susceptibility to oxidative degradation in biological formulations, thereby impeding the attainment of doses conducive to human health (106). Consequently, there is a critical need to explore various types of resveratrol derivatives to broaden and enhance the therapeutic potential of compounds resembling resveratrol, representing an urgent research imperative.

6.1 The severe inadequacy in bioavailability

The most pressing issue currently regarding the clinical application of RES is its bioavailability. RES's bioavailability in the human body is affected by gradually increasing doses and repeated administration, with an oral absorption rate of approximately 75%,

primarily through epithelial diffusion. However, extensive metabolism predominantly occurs in the intestines and liver, resulting in an oral bioavailability far below 1%. Even with gradual dose increases and repeated administration, this situation remains largely unchanged (65). In mammals, RES is chiefly metabolized in the intestines and liver, primarily converting to glucuronides and sulfates (235, 236). Nonetheless, its metabolism is exceedingly rapid, with plasma concentrations of RES diminishing rapidly within 30-60 minutes post-administration, posing challenges for systemic medication (237). A comparative study by Ndiave investigated three RES administration models-drinking water, oral feeding, and sustained-release pellets implanted near tumors (238). Surprisingly, RES exhibited no effect on melanoma in the body (239), with plasma RES levels remaining very low and rapidly converting to its metabolites such as resveratrol glucoside and resveratrol. Similarly, RES application in various areas such as lung cancer, intestinal tumors, and leukemia has shown markedly low bioavailability (240, 241). Efforts to address this issue are underway. For instance, a study by Niles and others in 2003 demonstrated that leveraging RES's characteristics via epithelial diffusion for skin cancer treatment not only tackled its rapid metabolism but also significantly enhanced its bioavailability (242). However, enhancing RES's bioavailability remains a critical concern in the medical community.

6.2 The poor biological stability

RES, a bioactive polyphenol, is widely present in various foods. Despite its strong beneficial effects on human health, such as antioxidant, anti-aging, cardioprotective, neuroprotective, and chemopreventive properties (243), its use as a drug is still greatly limited by its poor bioavailability, and the tendency for autooxidation and photosensitivity further restricts the development of its pharmaceutical formulations (244). The instability of RES is attributed to its deprotonation in alkaline media and subsequent auto-oxidation, degradation, or polymerization processes. RES exists in cis and trans isomeric forms, with the trans isomer being more biologically active and widely used as SIRTs activators (245). However, the isomerization of trans-resveratrol is influenced by various factors such as exposure time and wavelength, physical state of the molecule, temperature, and pH (64). Research by Goldberg et al. suggests that trans-resveratrol is unstable under light and heat; when exposed to ultraviolet radiation for several hours, transresveratrol can convert to cis-resveratrol, significantly losing its biological stability (246). Additionally, Robinson et al. investigated the stability of both resveratrol isomers in aqueous solutions and the kinetic effects of this process (247). The results indicate that under light protection, trans-resveratrol can remain stable for at least 42 hours in neutral buffer and at least 28 days in acidic medium. Therefore, the urgent need to address the issues of RES stability in terms of photo and water stability is crucial to maintain its beneficial effects, prevent the formation of oxidation or photooxidation products, and avoid harm from its metabolites to human health. Recent evidence suggests that the limitations of RES stability and bioavailability can be circumvented by developing new formulations or synthesizing new prodrugs (248). Studies by Dhakar et al. found that RES formulated into nanosponge encapsulated systems achieved higher water solubility and drug loading than single drugs. Furthermore, nanosponges dissolve drugs in a controlled manner, significantly prolonging the release duration of RES. Moreover, the antioxidant activity of RES is further enhanced in the presence of nanosponges. Most importantly, nanosponges exhibit biocompatibility and do not cause significant cytotoxicity to the organism. Therefore, future research should focus on the development of new products to make RES more soluble and delay its metabolism.

6.3 The dosage range and safety assessment

In addition to low bioavailability, determining the appropriate dosage range of RES and assessing the long-term safety of its use are also important challenges. In 2010, Brown et al. reported toxicity data from a dose-escalation study describing the intake of RES by 44 healthy volunteers (249). No serious adverse events were detected through clinical, biochemical, or hematological parameters, with the most common toxicity being gastrointestinal reactions. According to the Common Terminology Criteria for Adverse Events (CTCAE) from the National Cancer Institute, approximately 90% of reported events were classified as mild. However, most adverse reactions occurred in populations taking the two highest doses or individuals ingesting more than 1 gram of RES per day. Therefore, for future clinical development of RES, intake for one month at doses as high as 1 gram per day is safe and well tolerated (250). It is important to note that the concentration of RES varies in different tissues and organs. For example, there is a high concentration of RES in colonic tissue, far exceeding the concentrations required for in vitro activity, but the concentration in other tissues may be much lower than the appropriate in vitro concentration (251). Therefore, the clinical efficacy of RES likely depends to a large extent on whether its metabolites have significant activity or can regenerate RES (252). However, some studies have found different results, such as low concentrations of RES exhibiting biological activity and displaying a biphasic dose-response relationship (253). Further research is needed to explore the relationship between RES dosage and safety.

6.4 Adverse reactions

6.4.1 Cell toxicity

Bolton et al. found that the metabolites of RES have cytoprotective effects but can also induce cytotoxicity or immunotoxicity (254). During the metabolism of RES, cytochrome P450 generates quinone compounds through hydroxylation reactions, producing the metabolite piceatannol. Piceatannol possesses anti-inflammatory and antioxidant properties. It not only enhances the expression of the antioxidant enzyme heme oxygenase-1 (HO-1) in human breast epithelial cells by inducing Nrf2 (255), but also inhibits the downregulation of the anti-apoptotic B-cell lymphoma 2 protein (Bcl-2), thereby

suppressing oxidative stress and apoptosis induced by hydrogen peroxide and peroxynitrite (256). The quinone metabolites of RES are also associated with toxic effects, involving oxidative stress and alkylation mechanisms (257). For example, piceatannol can induce the inhibition of P450 oxidase or alkylation of certain proteins such as Keap1, Nrf2, I kappa B kinase (IKK), where NF-KB can lead to severe hepatorenal toxicity (258). Moreover, quinone compounds can deplete GSH, affect the function of nicotinamide adenine dinucleotide phosphate oxidase (NOX), and ultimately lead to oxidative stress reactions, damaging normal organisms (254). Studies have shown that tyrosinase, a key enzyme involved in melanin biosynthesis, participates in RES metabolism. RES can serve as a substrate for tyrosinase, generating active quinone compounds (259). The generated quinone compounds can decay to form oligomers, which as pro-oxidants, can trigger cytotoxicity in melanocytes, ultimately leading to the development of skin tumors (260).

6.4.2 DNA damage

Research has shown that increasing intake of RES can better clear ROS, thus, RES has a cellular protective effect (259). However, under certain conditions, antioxidants may also act as pro-oxidants, leading to accelerated lipid peroxidation or induced DNA damage (261). In fact, whether RES exhibits pro-oxidant or antioxidant activity depends on RES concentration, form, processing conditions, and its redox status (262, 263). When acting as a prooxidant molecule in vitro, RES can cause DNA damage, reduce multiple DNA repair pathways, and activate cytotoxicity and apoptosis pathways (264). Zuo et al. found that RES exhibits preferential cytotoxicity in malignant tumor cells, leading to higher electron transfer between RES and copper ions (265). Therefore, RES and copper-induced DNA damage may be one of the cytotoxic mechanisms of RES against cancer cells (266). It is noteworthy that the pro-oxidative action of RES has pro-apoptotic function in different types of cancer cells, and its ability to induce DNA breaks has potential therapeutic value in combating cancer cells (267). However, there are also reports that RES triggers p53dependent apoptosis by activating topoisomerase II, thereby inducing DNA damage in colon cancer cells (268). In U2OS and A549 cancer cells treated with RES, the proportion of DNA doublestrand breaks significantly increased. This phenomenon may also be mediated by RES-induced pro-oxidative effects and regulation of the CXCR2-p53 pathway (269).

6.4.3 Induction of oxidative stress

Giordo et al. found that RES severely impacts cellular redox state (270). In this regard, low doses of RES have various beneficial effects, such as protecting cells and tissues from the effects of neurodegeneration, cardiovascular diseases, cancer, diabetes, and obesity-related diseases, and prolonging organism lifespan (271). However, there is substantial evidence indicating that RES exhibits a biphasic concentration-dependent effect. That is, both *in vivo* and *in vitro*, RES acts as an antioxidant at low doses and a pro-oxidant at high doses (272). The pro-oxidative effect of RES is typically associated with the downregulation of phosphorylated pkb/Akt, cell damage, and apoptosis. Meanwhile, ROS-mediated mitochondrial damage produced by cytochrome P450 enzyme cyp2c9 in high doses of RES-induced oxidative damage seems to be involved as well (273). Heo et al. proposed that RES can induce apoptosis and cell cycle arrest in malignant melanoma cells (274). Additionally, Kim found that RES induces caspase-dependent cell death in ovarian cancer cells through a ROS-dependent mechanism. It is worth mentioning that in their 2022 study, Elbaz et al. found that resveratrol also triggers the downregulation of miR-144 by upregulating the expression level of Nrf2.Consequently, resveratrol showed hepatorenal antioxidant, anti-inflammatory, and antiapoptotic activities as manifested by improvement in the antioxidant markers along with a decline in NF- κ B, TNF- α , and caspase-3 expressions. In summary, this study demonstrated that resveratrol has potential therapeutic effects in alleviating liver and kidney damage induced by nonsteroidal anti-inflammatory drugs (275).

6.4.4 Drug interactions

Currently, an increasing number of studies have found indirect interactions between RES and other drugs, leading to decreased activity or overexpression of major cellular systems involved in drug metabolism-drug transporters and CYP450 enzyme activity (276). Previous studies have shown that RES can blunt the function and expression of drug transporters, thereby increasing the anti-proliferative activity of various drugs and reducing their bioavailability (277). For example, RES inhibits drug transport proteins such as P-glycoprotein, multidrug resistance-associated protein 2 (MRP2), and organic anion transport proteins (OAT1/ OAT3), reducing the clearance of methotrexate in the kidneys and increasing the risk of liver toxicity (278, 279). Additionally, RES can enhance the anticoagulant activity of warfarin, increasing the risk of bleeding (280). Interestingly, combination therapy with RES has also been reported to attenuate the effects of other drugs. For instance, RES can diminish the effects of human immunodeficiency virus (HIV) protease inhibitors (281) and interact with inhibitors of 3hydroxy-3-methylglutaryl-coenzyme A reductase (HMG-CoA reductase) (282), immunosuppressants (283), antiarrhythmic drugs (284), antihistamines (285), and calcium channel agonists (286), thereby reducing the bioactivity and utilization of drugs and affecting drug efficacy.

6.5 The turning point between basic research and clinical practice

In addition, the majority of research on RES is currently still in the basic research stage, and transitioning it into clinical practice promptly poses a significant challenge. The lag in clinical application is due to RES being a natural compound with many clinically relevant targets, which have different dose-response curves, tissue distributions, and modifiers. Effectively addressing these issues is currently a challenge. New paradigms and methods need to be developed, including better molecular modeling to predict interactions; large-scale screening for toxicity or other common effects; and affinity-based methods to identify drug interactions, among others (287).

7 Conclusion and outlook

This study systematically explores the mechanism and potential applications of resveratrol and its derivatives in the treatment of AID. Through literature review, we found that RES, as a SIRTs activator, has significant therapeutic potential in regulating immune cell function and suppressing the release of inflammatory factors, providing new insights for the treatment of AID.

Despite some encouraging progress, several challenges persist and must be addressed. These challenges encompass the bioavailability, stability, and safety of resveratrol and its derivatives. Future research endeavors should focus on, but not be restricted to, the following key areas: firstly, delving deeper into assessing the efficacy and safety of resveratrol and its derivatives across various types of AID. Secondly, enhancing the bioavailability, stability, and specificity of resveratrol and its derivatives through advancements in synthesis, drug design, and targeted delivery systems. Thirdly, formulating personalized treatment plans tailored to individual genetic, phenotypic, and lifestyle factors to achieve the precision medicine goal, considering the potential variation in the impact of resveratrol and its derivatives among individuals. Lastly, exploring the molecular mechanisms of the SIRTs pathway in immune regulation and devising more selective and specific SIRTs modulators to mitigate adverse reactions and side effects.

In conclusion, resveratrol, as a potential AID treatment, holds expansive application prospects; however, further research and clinical validation are imperative. Collaborative efforts within the medical community are essential to propel the advancement of this field, offering patients more efficacious and safer treatment alternatives.

Author contributions

XY: Data curation, Funding acquisition, Investigation, Methodology, Writing – original draft. MC: Investigation, Resources, Visualization, Writing – review & editing. JW: Investigation, Resources, Validation, Writing – review & editing. **References**

1. Karagianni P, Tzioufas AG. Epigenetic perspectives on systemic autoimmune disease. J Autoimmun. (2019) 104:102315. doi: 10.1016/j.jaut.2019.102315

 Scott DL, Wolfe F, Huizinga TW. Rheumatoid arthritis. Lancet. (2010) 376:1094– 108. doi: 10.1016/S0140-6736(10)60826-4

4. Bartalena L. Diagnosis and management of Graves disease: a global overview. *Nat Rev Endocrinol.* (2013) 9:724–34. doi: 10.1038/nrendo.2013.193

 Burn P. Type 1 diabetes. Nat Rev Drug Discov. (2010) 9:187–8. doi: 10.1038/nrd3097
 Long A, Kleiner A, Looney RJ. Immune dysregulation. J Allergy Clin Immunol. (2023) 151:70–80. doi: 10.1016/j.jaci.2022.11.001 RS: Conceptualization, Funding acquisition, Resources, Validation, Writing – review & editing.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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7. Cheru N, Hafler DA, Sumida TS. Regulatory T cells in peripheral tissue tolerance and diseases. *Front Immunol.* (2023) 14:1154575. doi: 10.3389/fimmu.2023.1154575

8. Zhan Q, Zhang J, Lin Y, Chen W, Fan X, Zhang D. Pathogenesis and treatment of Sjogren's syndrome: Review and update. *Front Immunol.* (2023) 14:1127417. doi: 10.3389/fimmu.2023.1127417

9. Chen Y, Liang R, Shi X, Shen R, Liu L, Liu Y, et al. Targeting kinase ITK treats autoimmune arthritis via orchestrating T cell differentiation and function. *BioMed Pharmacother*. (2023) 169:115886. doi: 10.1016/j.biopha.2023.115886

^{2.} Kiriakidou M, Ching CL. Systemic lupus erythematosus. Ann Intern Med. (2020) 172:ITC81–96. doi: 10.7326/AITC202006020

^{10.} Huang Y, Ba X, Wang H, Shen P, Han L, Lin W, et al. Triptolide alleviates collagen-induced arthritis in mice by modulating Treg/Th17 imbalance through the JAK/PTEN-STAT3 pathway. *Basic Clin Pharmacol Toxicol.* (2023) 133:43–58. doi: 10.1111/bcpt.13880

11. Wang W, Zhang BT, Jiang QL, Zhao HQ, Xu Q, Zeng Y, et al. Leptin receptor antagonist attenuates experimental autoimmune thyroiditis in mice by regulating Treg/Th17 cell differentiation. *Front Endocrinol (Lausanne)*. (2022) 13:1042511. doi: 10.3389/fendo.2022.1042511

12. Han Z, Li C, Han S, Han Y, Qiu J, Shi Y, et al. Meta-analysis: polymorphisms in TNF-alpha gene promoter and Crohn's disease. *Aliment Pharmacol Ther.* (2010) 32:159–70. doi: 10.1111/j.1365-2036.2010.04340.x

13. Ziaee V, Tahghighi F, Moradinejad MH, Harsini S, Mahmoudi M, Rezaei A, et al. Interleukin-6, interleukin-1 gene cluster and interleukin-1 receptor polymorphisms in Iranian patients with juvenile systemic lupus erythematosus. *Eur Cytokine Netw.* (2014) 25:35–40. doi: 10.1684/ecn.2014.0352

14. Monteiro M, Almeida CF, Caridade M, Ribot JC, Duarte J, Agua-Doce A, et al. Identification of regulatory Foxp3+ invariant NKT cells induced by TGF-beta. *J Immunol.* (2010) 185:2157–63. doi: 10.4049/jimmunol.1000359

15. Zhernakova A, van Diemen CC, Wijmenga C. Detecting shared pathogenesis from the shared genetics of immune-related diseases. *Nat Rev Genet.* (2009) 10:43–55. doi: 10.1038/nrg2489

16. Scofield RH. Genetics of systemic lupus erythematosus and Sjogren's syndrome. Curr Opin Rheumatol. (2009) 21:448-53. doi: 10.1097/BOR.0b013e32832f0861

17. Taylor KE, Chung SA, Graham RR, Ortmann WA, Lee AT, Langefeld CD, et al. Risk alleles for systemic lupus erythematosus in a large case-control collection and associations with clinical subphenotypes. *PloS Genet.* (2011) 7:e1001311. doi: 10.1371/ journal.pgen.1001311

18. Gottenberg JE, Busson M, Loiseau P, Cohen-Solal J, Lepage V, Charron D, et al. In primary Sjogren's syndrome, HLA class II is associated exclusively with autoantibody production and spreading of the autoimmune response. *Arthritis Rheumatol.* (2003) 48:2240–5. doi: 10.1002/art.11103

 Gustafsson JT, Gunnarsson I, Kallberg H, Pettersson S, Zickert A, Vikerfors A, et al. Cigarette smoking, antiphospholipid antibodies and vascular events in Systemic Lupus Erythematosus. *Ann Rheum Dis.* (2015) 74:1537–43. doi: 10.1136/annrheumdis-2013-205159

20. Nadigel J, Préfontaine D, Baglole CJ, Maltais F, Bourbeau J, Eidelman DH, et al. Cigarette smoke increases TLR4 and TLR9 expression and induces cytokine production from CD8(+) T cells in chronic obstructive pulmonary disease. *Respir Res.* (2011) 12:149. doi: 10.1186/1465–9921-12–149

21. Taucher E, Mykoliuk I, Lindenmann J, Smolle-Juettner FM. Implications of the immune landscape in COPD and lung cancer: smoking versus other causes. *Front Immunol.* (2022) 13:846605. doi: 10.3389/fimmu.2022.846605

22. Chang C, Gershwin ME. Drugs and autoimmunity-a contemporary review and mechanistic approach. J Autoimmun. (2010) 34:J266–75. doi: 10.1016/j.jaut.2009.11.012

23. Klar AJ, Fogel S, Macleod K. MAR1-a regulator of the HMa and HMalpha loci in SACCHAROMYCES CEREVISIAE. *Genetics*. (1979) 93:37–50. doi: 10.1093/genetics/ 93.1.37

24. Finkel T, Deng CX, Mostoslavsky R. Recent progress in the biology and physiology of sirtuins. *Nature*. (2009) 460:587–91. doi: 10.1038/nature08197

25. Frye RA. Characterization of five human cDNAs with homology to the yeast SIR2 gene: Sir2-like proteins (sirtuins) metabolize NAD and may have protein ADP-ribosyltransferase activity. *Biochem Biophys Res Commun.* (1999) 260:273–9. doi: 10.1006/bbrc.1999.0897

26. Frye RA. Phylogenetic classification of prokaryotic and eukaryotic Sir2-like proteins. *Biochem Biophys Res Commun.* (2000) 273:793-8. doi: 10.1006/bbrc.2000.3000

27. Zentner GE, Henikoff S. Regulation of nucleosome dynamics by histone modifications. *Nat Struct Mol Biol.* (2013) 20:259–66. doi: 10.1038/nsmb.2470

28. Schep R, Brinkman EK, Leemans C, Vergara X, van der Weide RH, Morris B, et al. Impact of chromatin context on Cas9-induced DNA double-strand break repair pathway balance. *Mol Cell*. (2021) 81:2216–2230.e10. doi: 10.1016/j.molcel.2021.03.032

29. Houtkooper RH, Pirinen E, Auwerx J. Sirtuins as regulators of metabolism and healthspan. *Nat Rev Mol Cell Biol.* (2012) 13:225–38. doi: 10.1038/nrm3293

30. Shen P, Deng X, Chen Z, Ba X, Qin K, Huang Y, et al. SIRT1: A potential therapeutic target in autoimmune diseases. *Front Immunol.* (2021) 12:779177. doi: 10.3389/fimmu.2021.779177

31. Chaintreuil P, Kerreneur E, Bourgoin M, Savy C, Favreau C, Robert G, et al. The generation, activation, and polarization of monocyte-derived macrophages in human Malignancies. *Front Immunol.* (2023) 14:1178337. doi: 10.3389/fimmu.2023.1178337

32. Schug TT, Xu Q, Gao H, Peres-da-Silva A, Draper DW, Fessler MB, et al. Myeloid deletion of SIRT1 induces inflammatory signaling in response to environmental stress. *Mol Cell Biol.* (2010) 30:4712–21. doi: 10.1128/MCB.00657–10

33. Lo Sasso G, Menzies KJ, Mottis A, Piersigilli A, Perino A, Yamamoto H, et al. SIRT2 deficiency modulates macrophage polarization and susceptibility to experimental colitis. *PloS One.* (2014) 9:e103573. doi: 10.1371/journal.pone.0103573

34. Zou Y, Zhang J, Xu J, Fu L, Xu Y, Wang X, et al. SIRT6 inhibition delays peripheral nerve recovery by suppressing migration, phagocytosis and M2-polarization of macrophages. *Cell Biosci.* (2021) 11:210. doi: 10.1186/s13578-021-00725-y

35. Rothgiesser KM, Erener S, Waibel S, Lüscher B and Hottiger MO. SIRT2 regulates NF- κ B-dependent gene expression through deacetylation of p65 Lys310. J Cell Sci. (2010) 123:4251–8. doi: 10.1242/jcs.073783

36. Santos-Barriopedro I, Bosch-Presegue L, Marazuela-Duque A, de la Torre C, Colomer C, Vazquez BN, et al. SIRT6-dependent cysteine monoubiquitination in the PRE-SET domain of Suv39h1 regulates the NF-kappaB pathway. *Nat Commun.* (2018) 9:101. doi: 10.1038/s41467-017-02586-x

37. Kaiser A, Schmidt M, Huber O, Frietsch JJ, Scholl S, Heidel FH, et al. SIRT7: an influence factor in healthy aging and the development of age-dependent myeloid stemcell disorders. *Leukemia*. (2020) 34:2206–16. doi: 10.1038/s41375–020-0803–3

38. Paolini R, Bernardini G, Molfetta R, Santoni A. NK cells and interferons. Cytokine Growth Factor Rev. (2015) 26:113–20. doi: 10.1016/j.cytogfr.2014.11.003

39. Zhang M, Acklin S, Gillenwater J, Du W, Patra M, Yu H, et al. SIRT2 promotes murine melanoma progression through natural killer cell inhibition. *Sci Rep.* (2021) 11:12988. doi: 10.1038/s41598-021-92445-z

40. Chen M, Xu M, Zhu C, Wang H, Zhao Q, Zhou F. Sirtuin2 enhances the tumoricidal function of liver natural killer cells in a mouse hepatocellular carcinoma model. *Cancer Immunol Immunother*. (2019) 68:961–71. doi: 10.1007/s00262–019-02337–5

41. Bak SP, Barnkob MS, Bai A, Higham EM, Wittrup KD, Chen J. Differential requirement for CD70 and CD80/CD86 in dendritic cell-mediated activation of tumor-tolerized CD8 T cells. *J Immunol.* (2012) 189:1708–16. doi: 10.4049/jimmunol.1201271

42. Liu G, Bi Y, Xue L, Zhang Y, Yang H, Chen X, et al. Dendritic cell SIRT1-HIF1alpha axis programs the differentiation of CD4+ T cells through IL-12 and TGFbeta1. *Proc Natl Acad Sci U S A*. (2015) 112:E957–65. doi: 10.1073/pnas.1420419112

43. Alvarez Y, Rodriguez M, Municio C, Hugo E, Alonso S, Ibarrola N, et al. Sirtuin 1 is a key regulator of the interleukin-12 p70/interleukin-23 balance in human dendritic cells. *J Biol Chem.* (2012) 287:35689–701. doi: 10.1074/jbc.M112.391839

44. Yang H, Lee SM, Gao B, Zhang J, Fang D. Histone deacetylase sirtuin 1 deacetylates IRF1 protein and programs dendritic cells to control Th17 protein differentiation during autoimmune inflammation. *J Biol Chem.* (2013) 288:37256–66. doi: 10.1074/jbc.M113.527531

45. Lasigliè D, Boero S, Bauer I, Morando S, Damonte P, Cea M, et al. Sirt6 regulates dendritic cell differentiation, maturation, and function. *Aging (Albany NY).* (2016) 8:34–49. doi: 10.18632/aging.100870

46. Li Y, Jin J, Wang Y. SIRT6 widely regulates aging, immunity, and cancer. Front Oncol. (2022) 12:861334. doi: 10.3389/fonc.2022.861334

47. Fagnoni FF, Vescovini R, Mazzola M, Bologna G, Nigro E, Lavagetto G, et al. Expansion of cytotoxic CD8+ CD28- T cells in healthy ageing people, including centenarians. *Immunology*. (1996) 88:501–7. doi: 10.1046/j.1365-2567.1996.d01-689.x

48. Effros RB, Dagarag M, Spaulding C, Man J. The role of CD8+ T-cell replicative senescence in human aging. *Immunol Rev.* (2005) 205:147–57. doi: 10.1111/j.0105-2896.2005.00259.x

49. Anwar HM, Hamad SR, Salem GEM, Soliman RHM, Elbaz EM. Inflammatory Modulation of miR-155 Inhibits Doxorubicin-Induced Testicular Dysfunction via SIRT1/FOXO1 Pathway: Insight into the Role of Acacetin and Bacillus cereus Protease. *Appl Biochem Biotechnol.* (2022) 194:5196–219. doi: 10.1007/s12010-022-03992-8

50. Sidler C, Woycicki R, Ilnytskyy Y, Metz G, Kovalchuk I, Kovalchuk O. Immunosenescence is associated with altered gene expression and epigenetic regulation in primary and secondary immune organs. *Front Genet.* (2013) 4:211. doi: 10.3389/fgene.2013.00211

51. Dang W, Steffen KK, Perry R, Dorsey JA, Johnson FB, Shilatifard A, et al. Histone H4 lysine 16 acetylation regulates cellular lifespan. *Nature*. (2009) 459:802–7. doi: 10.1038/nature08085

52. Xiao C, Wang RH, Lahusen TJ, Park O, Bertola A, Maruyama T, et al. Progression of chronic liver inflammation and fibrosis driven by activation of c-JUN signaling in Sirt6 mutant mice. *J Biol Chem.* (2012) 287:41903–13. doi: 10.1074/jbc.M112.415182

53. Zhang D, Qiu X, Li J, Zheng S, Li L, Zhao H. MiR-23a-3p-regulated abnormal acetylation of FOXP3 induces regulatory T cell function defect in Graves' disease. *Biol Chem.* (2019) 400:639–50. doi: 10.1515/hsz-2018–0343

54. Chen S, Lee L, Naila T, Fishbain S, Wang A, Tomkinson AE, et al. Structural basis of long-range to short-range synaptic transition in NHEJ. *Nature*. (2021) 593:294–8. doi: 10.1038/s41586-021-03458-7

55. Vazquez BN, Thackray JK, Simonet NG, Kane-Goldsmith N, Martinez-Redondo P, Nguyen T, et al. SIRT7 promotes genome integrity and modulates non-homologous end joining DNA repair. *EMBO J.* (2016) 35:1488–503. doi: 10.15252/embj.201593499

56. Gan H, Shen T, Chupp DP, Taylor JR, Sanchez HN, Li X, et al. B cell Sirt1 deacetylates histone and non-histone proteins for epigenetic modulation of AID expression and the antibody response. *Sci Adv.* (2020) 6:eaay2793. doi: 10.1126/sciadv.aay2793

57. Wang Y, Bi Y, Chen X, Li C, Li Y, Zhang Z, et al. Histone deacetylase SIRT1 negatively regulates the differentiation of interleukin-9-producing CD4(+) T cells. *Immunity.* (2016) 44:1337–49. doi: 10.1016/j.immuni.2016.05.009

58. Qiu Y, Zhou X, Liu Y, Tan S, Li Y. The role of sirtuin-1 in immune response and systemic lupus erythematosus. *Front Immunol.* (2021) 12:632383. doi: 10.3389/fmmu.2021.632383

59. Xuzhu G, Komai-Koma M, Leung BP, Howe HS, McSharry C, McInnes IB, et al. Resveratrol modulates murine collagen-induced arthritis by inhibiting Th17 and B-cell function. *Ann Rheum Dis.* (2012) 71:129–35. doi: 10.1136/ard.2011.149831 60. Wang Y, Li D, Ma G, Li W, Wu J, Lai T, et al. Increases in peripheral SIRT1: a new biological characteristic of asthma. *Respirology*. (2015) 20:1066–72. doi: 10.1111/ resp.12558

61. Hu Q, Sun Y, Li Y, Shi H, Teng J, Liu H, et al. Anti-SIRT1 autoantibody is elevated in ankylosing spondylitis: a potential disease biomarker. *BMC Immunol.* (2018) 19:38. doi: 10.1186/s12865-018-0280-x

62. Nakata R, Takahashi S, Inoue H. Recent advances in the study on resveratrol. *Biol Pharm Bull.* (2012) 35:273–9. doi: 10.1248/bpb.35.273

63. Burns J, Yokota T, Ashihara H, Lean ME, Crozier A. Plant foods and herbal sources of resveratrol. J Agric Food Chem. (2002) 50:3337–40. doi: 10.1021/jf0112973

64. Francioso A, Mastromarino P, Masci A, d'Erme M, Mosca L. Chemistry, stability and bioavailability of resveratrol. *Med Chem.* (2014) 10:237–45. doi: 10.2174/15734064113096660053

65. Walle T. Bioavailability of resveratrol. Ann N Y Acad Sci. (2011) 1215:9–15. doi: 10.1111/j.1749-6632.2010.05842.x

66. Yang T, Wang L, Zhu M, Zhang L, Yan L. Properties and molecular mechanisms of resveratrol: a review. *Pharmazie*. (2015) 70:501–6.

67. McCormack D, McFadden D. A review of pterostilbene antioxidant activity and disease modification. *Oxid Med Cell Longev.* (2013) 2013:575482. doi: 10.1155/2013/575482

68. Guo Y, Zhang L, Li F, Hu CP, Zhang Z. Restoration of sirt1 function by pterostilbene attenuates hypoxia-reoxygenation injury in cardiomyocytes. *Eur J Pharmacol.* (2016) 776:26–33. doi: 10.1016/j.ejphar.2016.02.052

69. Rimando AM, Nagmani R, Feller DR, Yokoyama W. Pterostilbene, a new agonist for the peroxisome proliferator-activated receptor alpha-isoform, lowers plasma lipoproteins and cholesterol in hypercholesterolemic hamsters. *J Agric Food Chem.* (2005) 53:3403–7. doi: 10.1021/jf0580364

70. Zhang F, Shi JS, Zhou H, Wilson B, Hong JS, Gao HM. Resveratrol protects dopamine neurons against lipopolysaccharide-induced neurotoxicity through its antiinflammatory actions. *Mol Pharmacol.* (2010) 78:466–77. doi: 10.1124/mol.110.064535

71. Yang YT, Weng CJ, Ho CT, Yen GC. Resveratrol analog-3,5,4'-trimethoxytrans-stilbene inhibits invasion of human lung adenocarcinoma cells by suppressing the MAPK pathway and decreasing matrix metalloproteinase-2 expression. *Mol Nutr Food Res.* (2009) 53:407–16. doi: 10.1002/mnfr.200800123

72. Gao G, Wang X, Qin X, Jiang X, Xiang D, Xie L, et al. Effects of trimethoxystilbene on proliferation and apoptosis of pulmonary artery smooth muscle cells. *Cell Biochem Biophys.* (2012) 64:101–6. doi: 10.1007/s12013–012-9377-7

73. Nguyen CB, Kotturi H, Waris G, Mohammed A, Chandrakesan P, May R, et al. (Z)-3,5,4'-trimethoxystilbene limits hepatitis C and cancer pathophysiology by blocking microtubule dynamics and cell-cycle progression. *Cancer Res.* (2016) 76:4887–96. doi: 10.1158/0008–5472.CAN-15-2722

74. Androutsopoulos VP, Fragiadaki I, Spandidos DA, Tosca A. The resveratrol analogue, 3,4,5,4'-trans-tetramethoxystilbene, inhibits the growth of A375 melanoma cells through multiple anticancer modes of action. *Int J Oncol.* (2016) 49:1305–14. doi: 10.3892/ijo.2016.3635

75. Cichocki M, Baer-Dubowska W, Wierzchowski M, Murias M, Jodynis-Liebert J. 3,4,5,4'-trans-tetramethoxystilbene (DMU-212) modulates the activation of NF-kappaB, AP-1, and STAT3 transcription factors in rat liver carcinogenesis induced by initiation-promotion regimen. *Mol Cell Biochem.* (2014) 391:27–35. doi: 10.1007/s11010-014-1983-9

76. Crofton JT, Share L, Shade RE, Lee-Kwon WJ, Manning M, Sawyer WH. The importance of vasopressin in the development and maintenance of DOC-salt hypertension in the rat. *Hypertension*. (1979) 1:31–8. doi: 10.1161/01.hyp.11.31

77. Malik KU, Jennings BL, Yaghini FA, Sahan-Firat S, Song CY, Estes AM, et al. Contribution of cytochrome P450 1B1 to hypertension and associated pathophysiology: a novel target for antihypertensive agents. *Prostaglandins Other Lipid Mediat.* (2012) 98:69–74. doi: 10.1016/j.prostaglandins.2011.12.003

78. Pan MH, Lin CL, Tsai JH, Ho CT, Chen WJ. 3,5,3',4',5'-pentamethoxystilbene (MR-5), a synthetically methoxylated analogue of resveratrol, inhibits growth and induces G1 cell cycle arrest of human breast carcinoma MCF-7 cells. *J Agric Food Chem.* (2010) 58:226–34. doi: 10.1021/jf903067g

79. Lee SK, Kim Y, Kim MY, Chun YJ, Kim S. Potent inhibition of recombinant human cytochrome p-450 1A1 by pentamethoxystilbene. *J Toxicol Environ Health A*. (2004) 67:1987–2000. doi: 10.1080/15287390490514642

80. Kimura Y, Sumiyoshi M, Baba K. Antitumor and antimetastatic activity of synthetic hydroxystilbenes through inhibition of lymphangiogenesis and M2 macrophage differentiation of tumor-associated macrophages. *Anticancer Res.* (2016) 36:137–48.

81. Balan KV, Wang Y, Chen SW, Chen JC, Zheng LF, Yang L, et al. Proteasomeindependent down-regulation of estrogen receptor-alpha (ERalpha) in breast cancer cells treated with 4,4'-dihydroxy-trans-stilbene. *Biochem Pharmacol.* (2006) 72:573–81. doi: 10.1016/j.bcp.2006.05.023

82. Tang YL, Chan SW. A review of the pharmacological effects of piceatannol on cardiovascular diseases. *Phytother Res.* (2014) 28:1581–8. doi: 10.1002/ptr.5185

83. Duarte N, Kayser O, Abreu P, Ferreira MJ. Antileishmanial activity of piceatannol isolated from Euphorbia lagascae seeds. *Phytother Res.* (2008) 22:455–7. doi: 10.1002/ptr.2334

84. Bastianetto S, Dumont Y, Han Y, Quirion R. Comparative neuroprotective properties of stilbene and catechin analogs: action via a plasma membrane receptor site? *CNS Neurosci Ther.* (2009) 15:76–83. doi: 10.1111/j.1755-5949.2008.00074.x

85. Mikula-Pietrasik J, Sosinska P, Wierzchowski M, Piwocka K, Ksiazek K. Synthetic resveratrol analogue, 3,3',4,4',5,5'-hexahydroxy-trans-stilbene, accelerates senescence in peritoneal mesothelium and promotes senescence-dependent growth of gastrointestinal cancers. *Int J Mol Sci.* (2013) 14:22483–98. doi: 10.3390/ijms141122483

86. Paulitschke V, Schicher N, Szekeres T, Jager W, Elbling L, Riemer AB, et al. 3,3',4,4',5,5'-hexahydroxystilbene impairs melanoma progression in a metastatic mouse model. J Invest Dermatol. (2010) 130:1668–79. doi: 10.1038/jid.2009.376

87. Singh G, Pai RS. Recent advances of resveratrol in nanostructured based delivery systems and in the management of HIV/AIDS. *J Control Release*. (2014) 194:178–88. doi: 10.1016/j.jconrel.2014.09.002

88. Bourgault S, Choi S, Buxbaum JN, Kelly JW, Price JL, Reixach N. Mechanisms of transthyretin cardiomyocyte toxicity inhibition by resveratrol analogs. *Biochem Biophys Res Commun.* (2011) 410:707–13. doi: 10.1016/j.bbrc.2011.04.133

89. Moran BW, Anderson FP, Devery A, Cloonan S, Butler WE, Varughese S, et al. Synthesis, structural characterisation and biological evaluation of fluorinated analogues of resveratrol. *Bioorg Med Chem.* (2009) 17:4510–22. doi: 10.1016/j.bmc.2009.05.007

90. Lee EJ, Min HY, Joo Park H, Chung HJ, Kim S, Nam Han Y, et al. G2/M cell cycle arrest and induction of apoptosis by a stilbenoid, 3,4,5-trimethoxy-4'-bromo-cis-stilbene, in human lung cancer cells. *Life Sci.* (2004) 75:2829–39. doi: 10.1016/ j.lfs.2004.07.002

91. Nguyen GT, Gertz M, Steegborn C. Crystal structures of Sirt3 complexes with 4'bromo-resveratrol reveal binding sites and inhibition mechanism. *Chem Biol.* (2013) 20:1375–85. doi: 10.1016/j.chembiol.2013.09.019

92. Nagaradja E, Bentabed-Ababsa G, Scalabrini M, Chevallier F, Philippot S, Fontanay S, et al. Deprotometalation-iodolysis and computed CH acidity of 1,2,3and 1,2,4-triazoles. Application to the synthesis of resveratrol analogues. *Bioorg Med Chem.* (2015) 23:6355–63. doi: 10.1016/j.bmc.2015.08.031

93. Dai H, Sinclair DA, Ellis JL, Steegborn C. Sirtuin activators and inhibitors: Promises, achievements, and challenges. *Pharmacol Ther.* (2018) 188:140-54. doi: 10.1016/j.pharmthera.2018.03.004

94. Karatoprak GS, Kupeli Akkol E, Genc Y, Bardakci H, Yucel C, Sobarzo-Sanchez E. Combretastatins: an overview of structure, probable mechanisms of action and potential applications. *Molecules*. (2020) 25:2560. doi: 10.3390/molecules25112560

95. Cirla A, Mann J. Combretastatins: from natural products to drug discovery. Nat Prod Rep. (2003) 20:558–64. doi: 10.1039/b306797c

96. Rimando AM, Kalt W, Magee JB, Dewey J, Ballington JR. Resveratrol, pterostilbene, and piceatannol in vaccinium berries. *J Agric Food Chem.* (2004) 52:4713–9. doi: 10.1021/jf040095e

97. Remsberg CM, Yanez JA, Ohgami Y, Vega-Villa KR, Rimando AM, Davies NM. Pharmacometrics of pterostilbene: preclinical pharmacokinetics and metabolism, anticancer, antiinflammatory, antioxidant and analgesic activity. *Phytother Res.* (2008) 22:169–79. doi: 10.1002/ptr.2277

98. Lin HS, Yue BD, Ho PC. Determination of pterostilbene in rat plasma by a simple HPLC-UV method and its application in pre-clinical pharmacokinetic study. *BioMed Chromatogr.* (2009) 23:1308–15. doi: 10.1002/bmc.1254

99. Chiou YS, Tsai ML, Nagabhushanam K, Wang YJ, Wu CH, Ho CT, et al. Pterostilbene is more potent than resveratrol in preventing azoxymethane (AOM)induced colon tumorigenesis via activation of the NF-E2-related factor 2 (Nrf2)mediated antioxidant signaling pathway. *J Agric Food Chem.* (2011) 59:2725–33. doi: 10.1021/jf2000103

100. Chen Y, Zhang DQ, Liao Z, Wang B, Gong S, Wang C, et al. Anti-oxidant polydatin (piceid) protects against substantia nigral motor degeneration in multiple rodent models of Parkinson's disease. *Mol Neurodegener*. (2015) 10:4. doi: 10.1186/1750-1326-10-4

101. Romero-Pérez AI, Ibern-Gómez M, Lamuela-Raventós RM, de la Torre-Boronat MC. Piceid, the major resveratrol derivative in grape juices. *Chem.* (1999) 47:1533–6. doi: 10.1021/jf981024g

102. Jeong ET, Jin MH, Kim MS, Chang YH, Park SG. Inhibition of melanogenesis by piceid isolated from Polygonum cuspidatum. *Arch Pharm Res.* (2010) 33:1331–8. doi: 10.1007/s12272-010-0906-x

103. Kobayashi S, Ding CK, Nakamura Y, Nakajima I, Matsumoto R. Kiwifruits (Actinidia deliciosa) transformed with a Vitis stilbene synthase gene produce piceid (resveratrol-glucoside). *Plant Cell Rep.* (2000) 19:904–10. doi: 10.1007/s002990000203

104. Hsieh TC, Huang YC, Wu JM. Control of prostate cell growth, DNA damage and repair and gene expression by resveratrol analogues, in vitro. *Carcinogenesis*. (2011) 32:93–101. doi: 10.1093/carcin/bgq230

105. Singh BR, Singh BN, Khan W, Singh HB, Naqvi AH. ROS-mediated apoptotic cell death in prostate cancer LNCaP cells induced by biosurfactant stabilized CdS quantum dots. *Biomaterials*. (2012) 33:5753–67. doi: 10.1016/j.biomaterials.2012.04.045

106. Li C, Xu X, Tao Z, Sun C, Pan Y. Resveratrol derivatives: an updated patent review (2012-2015). *Expert Opin Ther Pat.* (2016) 26:1189-200. doi: 10.1080/ 13543776.2016.1215435 107. Li C, Xu X, Tao Z, Wang XJ, Pan Y. Resveratrol dimers, nutritional components in grape wine, are selective ROS scavengers and weak Nrf2 activators. *Food Chem.* (2015) 173:218–23. doi: 10.1016/j.foodchem.2014.09.165

108. Matsuura BS, Keylor MH, Li B, Lin Y, Allison S, Pratt DA, et al. A scalable biomimetic synthesis of resveratrol dimers and systematic evaluation of their antioxidant activities. *Angew Chem Int Ed Engl.* (2015) 54:3754–7. doi: 10.1002/anie.201409773

109. Duan YC, Guan YY, Zhai XY, Ding LN, Qin WP, Shen DD, et al. Discovery of resveratrol derivatives as novel LSD1 inhibitors: Design, synthesis and their biological evaluation. *Eur J Med Chem.* (2017) 126:246–58. doi: 10.1016/j.ejmech.2016.11.035

110. Abdulla A, Zhao X, Yang F. Natural polyphenols inhibit lysine-specific demethylase-1 in vitro. J Biochem Pharmacol Res. (2013) 1:56-63.

111. Yang MF, Yao X, Chen LM, Gu JY, Yang ZH, Chen HF, et al. Synthesis and biological evaluation of resveratrol derivatives with anti-breast cancer activity. *Arch Pharm (Weinheim)*. (2020) 353:e2000044. doi: 10.1002/ardp.202000044

112. Piotrowska H, Kucinska M, Murias M. Biological activity of piceatannol: leaving the shadow of resveratrol. *Mutat Res.* (2012) 750:60–82. doi: 10.1016/j.mrrev.2011.11.001

113. Ko HS, Lee HJ, Kim SH, Lee EO. Piceatannol suppresses breast cancer cell invasion through the inhibition of MMP-9: involvement of PI3K/AKT and NF-kappaB pathways. J Agric Food Chem. (2012) 60:4083–9. doi: 10.1021/jf205171g

114. Song NR, Hwang MK, Heo YS, Lee KW, Lee HJ. Piceatannol suppresses the metastatic potential of MCF10A human breast epithelial cells harboring mutated H-ras by inhibiting MMP-2 expression. *Int J Mol Med.* (2013) 32:775–84. doi: 10.3892/ ijmm.2013.1449

115. Innets B, Thongsom S, Petsri K, Racha S, Yokoya M, Moriue S, et al. Akt/mTOR targeting activity of resveratrol derivatives in non-small lung cancer. *Molecules*. (2022) 27:8268. doi: 10.3390/molecules27238268

116. Iksen, Pothongsrisit S, Pongrakhananon V. Targeting the PI3K/AKT/mTOR signaling pathway in lung cancer: an update regarding potential drugs and natural products. *Molecules*. (2021) 26:4100. doi: 10.3390/molecules26134100

117. Chen K, Shang Z, Dai AL, Dai PL. Novel PI3K/Akt/mTOR pathway inhibitors plus radiotherapy: Strategy for non-small cell lung cancer with mutant RAS gene. *Life Sci.* (2020) 255:117816. doi: 10.1016/j.lfs.2020.117816

118. Fragopoulou E, Gkotsi K, Petsini F, Gioti K, Kalampaliki AD, Lambrinidis G, et al. Synthesis and biological evaluation of resveratrol methoxy derivatives. *Molecules*. (2023) 28:5547. doi: 10.3390/molecules28145547

119. Padyukov L. Genetics of rheumatoid arthritis. Semin Immunopathol. (2022) 44:47-62. doi: 10.1007/s00281-022-00912-0

120. Myasoedova E, Davis JM 3rd, Crowson CS, Gabriel SE. Epidemiology of rheumatoid arthritis: rheumatoid arthritis and mortality. *Curr Rheumatol Rep.* (2010) 12:379-85. doi: 10.1007/s11926-010-0117-y

121. He X, Liu J, Liang C, Badshah SA, Zheng K, Dang L, et al. Osteoblastic PLEKHO1 contributes to joint inflammation in rheumatoid arthritis. *EBioMedicine*. (2019) 41:538–55. doi: 10.1016/j.ebiom.2019.02.009

122. Prete M, Racanelli V, Digiglio L, Vacca A, Dammacco F, Perosa F. Extraarticular manifestations of rheumatoid arthritis: An update. *Autoimmun Rev.* (2011) 11:123–31. doi: 10.1016/j.autrev.2011.09.001

123. Figus FA, Piga M, Azzolin I, McConnell R, Iagnocco A. Rheumatoid arthritis: Extra-articular manifestations and comorbidities. *Autoimmun Rev.* (2021) 20:102776. doi: 10.1016/j.autrev.2021.102776

124. Maiese K. The impact of aging and oxidative stress in metabolic and nervous system disorders: programmed cell death and molecular signal transduction crosstalk. *Front Immunol.* (2023) 14:1273570. doi: 10.3389/fimmu.2023.1273570

125. Tian J, Chen JW, Gao JS, Li L, Xie X. Resveratrol inhibits TNF-alpha-induced IL-1beta, MMP-3 production in human rheumatoid arthritis fibroblast-like synoviocytes via modulation of P13kinase/Akt pathway. *Rheumatol Int.* (2013) 33:1829–35. doi: 10.1007/s00296-012-2657-0

126. Strom J, Xu B, Tian X, Chen QM. Nrf2 protects mitochondrial decay by oxidative stress. FASEB J. (2016) 30:66-80. doi: 10.1096/fj.14-268904

127. Li G, Xia Z, Liu Y, Meng F, Wu X, Fang Y, et al. SIRT1 inhibits rheumatoid arthritis fibroblast-like synoviocyte aggressiveness and inflammatory response via suppressing NF-kappaB pathway. *Biosci Rep.* (2018) 38:BSR20180541. doi: 10.1042/BSR20180541

128. Katto J, Engel N, Abbas W, Herbein G, Mahlknecht U. Transcription factor NF κ B regulates the expression of the histone deacetylase SIRT1. *Clin Epigenetics*. (2013) 5:11. doi: 10.1186/1868-7083-5-11

129. Rathore MG, Saumet A, Rossi JF, de Bettignies C, Tempe D, Lecellier CH, et al. The NF-kappaB member p65 controls glutamine metabolism through miR-23a. *Int J Biochem Cell Biol.* (2012) 44:1448–56. doi: 10.1016/j.biocel.2012.05.011

130. Zhou L, Xu DY, Sha WG, Shen L, Lu GY, Yin X, et al. High glucose induces renal tubular epithelial injury via Sirt1/NF-kappaB/microR-29/Keap1 signal pathway. J Transl Med. (2015) 13:352. doi: 10.1186/s12967-015-0710-y

131. Wang G, Xie X, Yuan L, Qiu J, Duan W, Xu B, et al. Resveratrol ameliorates rheumatoid arthritis via activation of SIRT1-Nrf2 signaling pathway. *Biofactors*. (2020) 46:441–53. doi: 10.1002/biof.1599

132. Thalhamer T, McGrath MA, Harnett MM. MAPKs and their relevance to arthritis and inflammation. *Rheumatol (Oxford)*. (2008) 47:409-14. doi: 10.1093/rheumatology/kem297

133. Schett G, Zwerina J, Firestein G. The p38 mitogen-activated protein kinase (MAPK) pathway in rheumatoid arthritis. *Ann Rheum Dis.* (2008) 67:909–16. doi: 10.1136/ard.2007.074278

134. Mittal M, Siddiqui MR, Tran K, Reddy SP, Malik AB. Reactive oxygen species in inflammation and tissue injury. *Antioxid Redox Signal.* (2014) 20:1126-67. doi: 10.1089/ars.2012.5149

135. Yang G, Chang CC, Yang Y, Yuan L, Xu L, Ho CT, et al. Resveratrol alleviates rheumatoid arthritis via reducing ROS and inflammation, inhibiting MAPK signaling pathways, and suppressing angiogenesis. *J Agric Food Chem.* (2018) 66:12953–60. doi: 10.1021/acs.jafc.8b05047

136. Qiu X, Brown K, Hirschey MD, Verdin E, Chen D. Calorie restriction reduces oxidative stress by SIRT3-mediated SOD2 activation. *Cell Metab.* (2010) 12:662–7. doi: 10.1016/j.cmet.2010.11.015

137. Tseng AHH, Shieh S-S, Wang DL. SIRT3 deacetylates FOXO3 to protect mitochondria against oxidative damage. *Free Radical Biol Med.* (2013) 63:222–34. doi: 10.1016/j.freeradbiomed.2013.05.002

138. Khojah HM, Ahmed S, Abdel-Rahman MS, Elhakeim EH. Resveratrol as an effective adjuvant therapy in the management of rheumatoid arthritis: a clinical study. *Clin Rheumatol.* (2018) 37:2035-42. doi: 10.1007/s10067-018-4080-8

139. Sheng S, Wang X, Liu X, Hu X, Shao Y, Wang G, et al. The role of resveratrol on rheumatoid arthritis: From bench to bedside. *Front Pharmacol.* (2022) 13:829677. doi: 10.3389/fphar.2022.829677

140. Byun HS, Song JK, Kim YR, Piao L, Won M, Park KA, et al. Caspase-8 has an essential role in resveratrol-induced apoptosis of rheumatoid fibroblast-like synoviocytes. *Rheumatol (Oxford)*. (2008) 47:301-8. doi: 10.1093/rheumatology/kem368

141. Long Z, Xiang W, He Q, Xiao W, Wei H, Li H, et al. Efficacy and safety of dietary polyphenols in rheumatoid arthritis: A systematic review and meta-analysis of 47 randomized controlled trials. *Front Immunol.* (2023) 14:1024120. doi: 10.3389/fimmu.2023.1024120

142. Askanase A, Shum K, Mitnick H. Systemic lupus erythematosus: an overview. Soc Work Health Care. (2012) 51:576-86. doi: 10.1080/00981389.2012.683369

143. Barber MRW, Drenkard C, Falasinnu T, Hoi A, Mak A, Kow NY, et al. Global epidemiology of systemic lupus erythematosus. *Nat Rev Rheumatol.* (2021) 17:515–32. doi: 10.1038/s41584–021-00668–1

144. Kronbichler A, Brezina B, Gauckler P, Quintana LF, Jayne DRW. Refractory lupus nephritis: When, why and how to treat. *Autoimmun Rev.* (2019) 18:510–8. doi: 10.1016/j.autrev.2019.03.004

145. Liu Y, Kaplan MJ. Cardiovascular disease in systemic lupus erythematosus: an update. Curr Opin Rheumatol. (2018) 30:441–8. doi: 10.1097/BOR.000000000000528

146. Bougea A, Anagnostou E, Konstantinos G, George P, Triantafyllou N, Kararizou E. A systematic review of peripheral and central nervous system involvement of rheumatoid arthritis, systemic lupus erythematosus, primary sjogren's syndrome, and associated immunological profiles. *Int J Chronic Dis.* (2015) 2015;910352.

147. Apor E, O'Brien J, Stephen M, Castillo JJ. Systemic lupus erythematosus is associated with increased incidence of hematologic Malignancies: a meta-analysis of prospective cohort studies. *Leuk Res.* (2014) 38:1067-71. doi: 10.1016/j.leukres.2014.06.025

148. Bermas BL, Sammaritano LR. Fertility and pregnancy in rheumatoid arthritis and systemic lupus erythematosus. *Fertil Res Pract.* (2015) 1:13. doi: 10.1186/s40738-015-0004-3

149. Li H, Boulougoura A, Endo Y, Tsokos GC. Abnormalities of T cells in systemic lupus erythematosus: new insights in pathogenesis and therapeutic strategies. *J Autoimmun.* (2022) 132:102870. doi: 10.1016/j.jaut.2022.102870

150. Richards HB, Satoh M, Jennette JC, Okano T, Kanwar YS, Reeves WH. Disparate T cell requirements of two subsets of lupus-specific autoantibodies in pristane-treated mice. *Clin Exp Immunol.* (1999) 115:547–53. doi: 10.1046/j.1365-2249.1999.00825.x

151. Zhang J, Lee SM, Shannon S, Gao B, Chen W, Chen A, et al. The type III histone deacetylase Sirt1 is essential for maintenance of T cell tolerance in mice. *J Clin Invest.* (2009) 119:3048–58. doi: 10.1172/JCI38902

152. Ko YC, Chang CL, Chien HF, Wu CH, Lin LI. Resveratrol enhances the expression of death receptor Fas/CD95 and induces differentiation and apoptosis in anaplastic large-cell lymphoma cells. *Cancer Lett.* (2011) 309:46–53. doi: 10.1016/j.canlet.2011.05.014

153. Dorrie J, Gerauer H, Wachter Y, Zunino SJ. Resveratrol induces extensive apoptosis by depolarizing mitochondrial membranes and activating caspase-9 in acute lymphoblastic leukemia cells. *Cancer Res.* (2001) 61:4731–9.

154. Zunino SJ, Storms DH. Resveratrol alters proliferative responses and apoptosis in human activated B lymphocytes in vitro. *J Nutr.* (2009) 139:1603–8. doi: 10.3945/jn.109.105064

155. Svajger U, Obermajer N, Jeras M. Dendritic cells treated with resveratrol during differentiation from monocytes gain substantial tolerogenic properties upon activation. *Immunology.* (2010) 129:525–35. doi: 10.1111/j.1365-2567.2009.03205.x

156. Surh YJ, Chun KS, Cha HH, Han SS, Keum YS, Park KK, et al. Molecular mechanisms underlying chemopreventive activities of anti-inflammatory phytochemicals:

down-regulation of COX-2 and iNOS through suppression of NF-kappa B activation. *Mutat Res.* (2001) 480–481:243–68. doi: 10.1016/S0027-5107(01)00183-X

157. Zhu X, Liu Q, Wang M, Liang M, Yang X, Xu X, et al. Activation of Sirt1 by resveratrol inhibits TNF-alpha induced inflammation in fibroblasts. *PloS One*. (2011) 6: e27081. doi: 10.1371/journal.pone.0027081

158. Miyake K, Akahoshi M, Nakashima H. Th subset balance in lupus nephritis. J BioMed Biotechnol. (2011) 2011:980286. doi: 10.1155/2011/980286

159. Wang ZL, Luo XF, Li MT, Xu D, Zhou S, Chen HZ, et al. Resveratrol possesses protective effects in a pristane-induced lupus mouse model. *PloS One.* (2014) 9: e114792. doi: 10.1371/journal.pone.0114792

160. Taha S, Gamal SM, Nabil M, Naeem N, Labib D, Siam I, et al. Vascular endothelial growth factor G1612A (rs10434) gene polymorphism and neuropsychiatric manifestations in systemic lupus erythematosus patients. *Rev Bras Reumatol Engl Ed.* (2017) 57:149–53. doi: 10.1016/j.rbre.2016.11.007

161. Hoang TTT, Ichinose K, Morimoto S, Furukawa K, Le LHT, Kawakami A. Measurement of anti-suprabasin antibodies, multiple cytokines and chemokines as potential predictive biomarkers for neuropsychiatric systemic lupus erythematosus. *Clin Immunol.* (2022) 237:108980. doi: 10.1016/j.clim.2022.108980

162. Kalinowska-Lyszczarz A, Pawlak MA, Pietrzak A, Pawlak-Bus K, Leszczynski P, Puszczewicz M, et al. Subcortical gray matter atrophy is associated with cognitive deficit in multiple sclerosis but not in systemic lupus erythematosus patients. *Lupus*. (2018) 27:610–20. doi: 10.1177/0961203317735186

163. Kasselman LJ, Renna HA, Voloshyna I, Pinkhasov A, Gomolin IH, Teboul I, et al. Cognitive changes mediated by adenosine receptor blockade in a resveratroltreated atherosclerosis-prone lupus mouse model. *J Tradit Complement Med.* (2022) 12:447–54. doi: 10.1016/j.jtcme.2022.01.006

164. Cope EC, LaMarca EA, Monari PK, Olson LB, Martinez S, Zych AD, et al. Microglia play an active role in obesity-associated cognitive decline. *J Neurosci.* (2018) 38:8889–904. doi: 10.1523/JNEUROSCI.0789–18.2018

165. Denton CP, Khanna D. Systemic sclerosis. Lancet. (2017) 390:1685-99. doi: 10.1016/S0140-6736(17)30933-9

166. Jerjen R, Nikpour M, Krieg T, Denton CP, Saracino AM. Systemic sclerosis in adults. Part I: Clinical features and pathogenesis. *J Am Acad Dermatol.* (2022) 87:937–54. doi: 10.1016/j.jaad.2021.10.065

167. Volkmann ER, Andreasson K, Smith V. Systemic sclerosis. Lancet. (2023) 401:304-18. doi: 10.1016/S0140-6736(22)01692-0

168. Wyman AE, Atamas SP. Sirtuins and accelerated aging in scleroderma. *Curr Rheumatol Rep.* (2018) 20:16. doi: 10.1007/s11926-018-0724-6

169. Wei J, Ghosh AK, Chu H, Fang F, Hinchcliff ME, Wang J, et al. The histone deacetylase sirtuin 1 is reduced in systemic sclerosis and abrogates fibrotic responses by targeting transforming growth factor beta signaling. *Arthritis Rheumatol.* (2015) 67:1323–34. doi: 10.1002/art.39061

170. Zhu X, Chu H, Jiang S, Liu Q, Liu L, Xue Y, et al. Sirt1 ameliorates systemic sclerosis by targeting the mTOR pathway. *J Dermatol Sci.* (2017) 87:149–58. doi: 10.1016/j.jdermsci.2017.04.013

171. Akamata K, Wei J, Bhattacharyya M, Cheresh P, Bonner MY, Arbiser JL, et al. SIRT3 is attenuated in systemic sclerosis skin and lungs, and its pharmacologic activation mitigates organ fibrosis. *Oncotarget.* (2016) 7:69321–36. doi: 10.18632/ oncotarget.12504

172. Shimojima Y, Kishida D, Ichikawa T, Takamatsu R, Nomura S, Sekijima Y. Oxidative stress promotes instability of regulatory T cells in antineutrophil cytoplasmic antibody-associated vasculitis. *Front Immunol.* (2021) 12:789740. doi: 10.3389/fmmu.2021.789740

173. Bartalena L, Tanda ML. Current concepts regarding Graves' orbitopathy. J Intern Med. (2022) 292:692–716. doi: 10.1111/joim.13524

174. Wemeau JL, Klein M, Sadoul JL, Briet C, Velayoudom-Cephise FL. Graves' disease: Introduction, epidemiology, endogenous and environmental pathogenic factors. *Ann Endocrinol (Paris)*. (2018) 79:599–607. doi: 10.1016/j.ando.2018.09.002

175. Chng CL, Lim AY, Tan HC, Kovalik JP, Tham KW, Bee YM, et al. Physiological and metabolic changes during the transition from hyperthyroidism to euthyroidism in graves' Disease. *Thyroid*. (2016) 26:1422–30. doi: 10.1089/thy.2015.0602

176. Panagiotou G, Perros P. Asymmetric graves' Orbitopathy. Front Endocrinol (Lausanne). (2020) 11:611845. doi: 10.3389/fendo.2020.611845

177. Tsymbaliuk I, Unukovych D, Shvets N, Dinets A. Cardiovascular complications secondary to Graves' disease: a prospective study from Ukraine. *PloS One.* (2015) 10: e0122388. doi: 10.1371/journal.pone.0122388

178. Yoshihara A, Yoshimura Noh J, Mukasa K, Watanabe N, Iwaku K, Ohye H, et al. The characteristics of osteoporotic patients in Graves' disease patients newly diagnosed after menopause: a prospective observational study. *Endocr J.* (2016) 63:1113–22. doi: 10.1507/endocrj.EJ16–0261

179. Kim CY, Lee HJ, Chae MK, Byun JW, Lee EJ, Yoon JS. Therapeutic effect of resveratrol on oxidative stress in graves' Orbitopathy orbital fibroblasts. *Invest Ophthalmol Vis Sci.* (2015) 56:6352–61. doi: 10.1167/iovs.15–16870

180. Hondur A, Konuk O, Dincel AS, Bilgihan A, Unal M, Hasanreisoglu B. Oxidative stress and antioxidant activity in orbital fibroadipose tissue in Graves' ophthalmopathy. *Curr Eye Res.* (2008) 33:421–7. doi: 10.1080/02713680802123532

181. Tsai CC, Wu SB, Cheng CY, Kao SC, Kau HC, Lee SM, et al. Increased response to oxidative stress challenge in Graves' ophthalmopathy orbital fibroblasts. *Mol Vis.* (2011) 17:2782–8.

182. DiMeglio LA, Evans-Molina C, Oram RA. Type 1 diabetes. Lancet. (2018) 391:2449-62. doi: 10.1016/S0140-6736(18)31320-5

183. Maahs DM, West NA, Lawrence JM, Mayer-Davis EJ. Epidemiology of type 1 diabetes. *Endocrinol Metab Clin North Am.* (2010) 39:481–97. doi: 10.1016/ j.ecl.2010.05.011

184. Wang Z, Yang T, Fu H. Prevalence of diabetes and hypertension and their interaction effects on cardio-cerebrovascular diseases: a cross-sectional study. *BMC Public Health.* (2021) 21:1224. doi: 10.1186/s12889-021-11122-y

185. Laiginhas R, Madeira C, Lopes M, Neves JS, Barbosa M, Rosas V, et al. Risk factors for prevalent diabetic retinopathy and proliferative diabetic retinopathy in type 1 diabetes. *Endocrine*. (2019) 66:201–9. doi: 10.1007/s12020-019-02047-z

186. Wheelock KM, Cai J, Looker HC, Merchant ML, Nelson RG, Fufaa GD, et al. Plasma bradykinin and early diabetic nephropathy lesions in type 1 diabetes mellitus. *PloS One.* (2017) 12:e0180964. doi: 10.1371/journal.pone.0180964

187. Orlov S, Bril V, Orszag A, Perkins BA. Heart rate variability and sensorimotor polyneuropathy in type 1 diabetes. *Diabetes Care.* (2012) 35:809–16. doi: 10.2337/dc11–1652

188. Migdalis I, Czupryniak L, Lalic N, Leslie RD, Papanas N, Valensi P. The diabetic foot. J Diabetes Res. (2017) 2017:3585617. doi: 10.1155/2017/3585617

189. Côté CD, Rasmussen BA, Duca FA, Zadeh-Tahmasebi M, Baur JA, Daljeet M, et al. Resveratrol activates duodenal Sirt1 to reverse insulin resistance in rats through a neuronal network. *Nat Med.* (2015) 21:498–505. doi: 10.1038/nm.3821

190. Price Nathan L, Gomes Ana P, Ling Alvin JY, Duarte Filipe V, Martin-Montalvo A, North Brian J, et al. SIRT1 is required for AMPK activation and the beneficial effects of resveratrol on mitochondrial function. *Cell Metab.* (2012) 15:675– 90. doi: 10.1016/j.cmet.2012.04.003

191. Park S-J, Ahmad F, Philp A, Baar K, Williams T, Luo H, et al. Resveratrol ameliorates aging-related metabolic phenotypes by inhibiting cAMP phosphodiesterases. *Cell*. (2012) 148:421-33. doi: 10.1016/j.cell.2012.01.017

192. Ciddi V, Dodda D. Therapeutic potential of resveratrol in diabetic complications: *In vitro* and in *vivo* studies. *Pharmacol Rep.* (2014) 66:799-803. doi: 10.1016/j.pharep.2014.04.006

193. Duca FA, Côté CD, Rasmussen BA, Zadeh-Tahmasebi M, Rutter GA, Filippi BM, et al. Metformin activates a duodenal Ampk-dependent pathway to lower hepatic glucose production in rats. *Nat Med.* (2015) 21:506–11. doi: 10.1038/nm.3787

194. Brubaker PL, Lauffer LM, Grieco A, Kim H, Oh AH, Mulherin AJ. Mechanisms underlying metformin-induced secretion of glucagon-like peptide-1 from the intestinal L cell. *Endocrinology*. (2011) 152:4610–9. doi: 10.1210/en.2011–1485

195. Park HS, Lim JH, Kim MY, Kim Y, Hong YA, Choi SR, et al. Resveratrol increases AdipoR1 and AdipoR2 expression in type 2 diabetic nephropathy. *J Transl Med.* (2016) 14:176. doi: 10.1186/s12967–016-0922–9

196. Jin Q, Liu T, Qiao Y, Liu D, Yang L, Mao H, et al. Oxidative stress and inflammation in diabetic nephropathy: role of polyphenols. *Front Immunol.* (2023) 14:1185317. doi: 10.3389/fimmu.2023.1185317

197. Xu F, Wang Y, Cui W, Yuan H, Sun J, Wu M, et al. Resveratrol prevention of diabetic nephropathy is associated with the suppression of renal inflammation and mesangial cell proliferation: possible roles of akt/NF-kappaB pathway. *Int J Endocrinol.* (2014) 2014:289327. doi: 10.1155/2014/289327

198. Chen YJ, Tang ZZ, Du L, Liu Y, Lu Q, Ma TF, et al. A novel compound AB-38b improves diabetes-associated cognitive decline in mice via activation of Nrf2/ARE pathway. *Brain Res Bull.* (2019) 150:160–7. doi: 10.1016/j.brainresbull.2019.05.010

199. He T, Xiong J, Nie L, Yu Y, Guan X, Xu X, et al. Resveratrol inhibits renal interstitial fibrosis in diabetic nephropathy by regulating AMPK/NOX4/ROS pathway. *J Mol Med (Berl)*. (2016) 94:1359–71. doi: 10.1007/s00109-016-1451-y

200. Negi G, Kumar A, Sharma SS. Melatonin modulates neuroinflammation and oxidative stress in experimental diabetic neuropathy: effects on NF-kappaB and Nrf2 cascades. *J Pineal Res.* (2011) 50:124–31. doi: 10.1111/j.1600-079X.2010.00821.x

201. Bahar E, Kim JY, Yoon H. Quercetin Attenuates Manganese-Induced Neuroinflammation by Alleviating Oxidative Stress through Regulation of Apoptosis, iNOS/NF-kappaB and HO-1/Nrf2 Pathways. *Int J Mol Sci.* (2017) 18:1989. doi: 10.3390/ijms18091989

202. Huang DD, Shi G, Jiang Y, Yao C, Zhu C. A review on the potential of Resveratrol in prevention and therapy of diabetes and diabetic complications. *BioMed Pharmacother*. (2020) 125:109767. doi: 10.1016/j.biopha.2019.109767

203. Kumar A, Sharma SS. NF-kappaB inhibitory action of resveratrol: a probable mechanism of neuroprotection in experimental diabetic neuropathy. *Biochem Biophys Res Commun.* (2010) 394:360–5. doi: 10.1016/j.bbrc.2010.03.014

204. Zhang W, Yu H, Lin Q, Liu X, Cheng Y, Deng B. Anti-inflammatory effect of resveratrol attenuates the severity of diabetic neuropathy by activating the Nrf2 pathway. *Aging (Albany NY).* (2021) 13:10659–71. doi: 10.18632/aging.202830

205. Wight RD, Tull CA, Deel MW, Stroope BL, Eubanks AG, Chavis JA, et al. Resveratrol effects on astrocyte function: relevance to neurodegenerative diseases. *Biochem Biophys Res Commun.* (2012) 426:112–5. doi: 10.1016/j.bbrc.2012.08.045

206. Nagineni CN, Raju R, Nagineni KK, Kommineni VK, Cherukuri A, Kutty RK, et al. Resveratrol suppresses expression of VEGF by human retinal pigment epithelial

cells: potential nutraceutical for age-related macular degeneration. *Aging Dis.* (2014) 5:88-100. doi: 10.14366/AD.2014.050088

207. Chan CM, Chang HH, Wang VC, Huang CL, Hung CF. Inhibitory effects of resveratrol on PDGF-BB-induced retinal pigment epithelial cell migration via PDGFRbeta, PI3K/Akt and MAPK pathways. *PloS One.* (2013) 8:e56819. doi: 10.1371/journal.pone.0056819

208. Wehkamp J, Gotz M, Herrlinger K, Steurer W, Stange EF. Inflammatory bowel disease. *Dtsch Arztebl Int*. (2016) 113:72–82. doi: 10.3238/arztebl.2016.0072

209. Mak WY, Zhao M, Ng SC, Burisch J. The epidemiology of inflammatory bowel disease: East meets west. *J Gastroenterol Hepatol.* (2020) 35:380–9. doi: 10.1111/jgh.14872

210. Rogler G. Resolution of inflammation in inflammatory bowel disease. Lancet Gastroenterol Hepatol. (2017) 2:521-30. doi: 10.1016/S2468-1253(17)30031-6

211. Mohan HM, Coffey JC. Surgical treatment of intestinal stricture in inflammatory bowel disease. *J Dig Dis.* (2020) 21:355–9. doi: 10.1111/1751–2980.12880

212. Chang M, Chang L, Chang HM, Chang F. Intestinal and extraintestinal cancers associated with inflammatory bowel disease. *Clin Colorectal Cancer*. (2018) 17:e29–37. doi: 10.1016/j.clcc.2017.06.009

213. Shi Y, Zhou J, Jiang B, Miao M. Resveratrol and inflammatory bowel disease. Ann N Y Acad Sci. (2017) 1403:38–47. doi: 10.1111/nyas.13426

214. Ren Z, Wang L, Cui J, Huoc Z, Xue J, Cui H, et al. Resveratrol inhibits NF-kB signaling through suppression of p65 and IkappaB kinase activities. *Pharmazie.* (2013) 68:689–94.

215. Singh UP, Singh NP, Singh B, Hofseth LJ, Taub DD, Price RL, et al. Role of resveratrol-induced CD11b(+) Gr-1(+) myeloid derived suppressor cells (MDSCs) in the reduction of CXCR3(+) T cells and amelioration of chronic colitis in IL-10(-/-) mice. *Brain Behav Immun.* (2012) 26:72–82. doi: 10.1016/j.bbi.2011.07.236

216. Altamemi I, Murphy EA, Catroppo JF, Zumbrun EE, Zhang J, McClellan JL, et al. Role of microRNAs in resveratrol-mediated mitigation of colitis-associated tumorigenesis in Apc(Min/+) mice. *J Pharmacol Exp Ther.* (2014) 350:99–109. doi: 10.1124/jpet.114.213306

217. Garcia P, Schmiedlin-Ren P, Mathias JS, Tang H, Christman GM, Zimmermann EM. Resveratrol causes cell cycle arrest, decreased collagen synthesis, and apoptosis in rat intestinal smooth muscle cells. *Am J Physiol Gastrointest Liver Physiol.* (2012) 302:G326–35. doi: 10.1152/ajpgi.00083.2011

218. Rahal K, Schmiedlin-Ren P, Adler J, Dhanani M, Sultani V, Rittershaus AC, et al. Resveratrol has antiinflammatory and antifibrotic effects in the peptidoglycanpolysaccharide rat model of Crohn's disease. *Inflammation Bowel Dis.* (2012) 18:613– 23. doi: 10.1002/ibd.21843

219. Arslan A, Ozcicek F, Keskin Cimen F, Altuner D, Yarali O, Kurt N, et al. Protective effect of resveratrol against methotrexate-induced oxidative stress in the small intestinal tissues of rats. *Int J Clin Exp Med.* (2015) 8:10491–500.

220. Serra D, Almeida LM, Dinis TC. Anti-inflammatory protection afforded by cyanidin-3-glucoside and resveratrol in human intestinal cells via Nrf2 and PPAR-gamma: Comparison with 5-aminosalicylic acid. *Chem Biol Interact.* (2016) 260:102–9. doi: 10.1016/j.cbi.2016.11.003

221. Dubuquoy L, Rousseaux C, Thuru X, Peyrin-Biroulet L, Romano O, Chavatte P, et al. PPARgamma as a new therapeutic target in inflammatory bowel diseases. *Gut.* (2006) 55:1341–9. doi: 10.1136/gut.2006.093484

222. Kim J, Cha YN, Surh YJ. A protective role of nuclear factor-erythroid 2-related factor-2 (Nrf2) in inflammatory disorders. *Mutat Res.* (2010) 690:12–23. doi: 10.1016/j.mrfmmm.2009.09.007

223. Yu QY, Tang XX. Irreversibility of pulmonary fibrosis. *Aging Dis.* (2022) 13:73–86. doi: 10.14336/AD.2021.0730

224. Meyer KC. Pulmonary fibrosis, part I: epidemiology, pathogenesis, and diagnosis. *Expert Rev Respir Med.* (2017) 11:343–59. doi: 10.1080/17476348.2017.1312346

225. Cottin V. The impact of emphysema in pulmonary fibrosis. *Eur Respir Rev.* (2013) 22:153–7. doi: 10.1183/09059180.00000813

226. Darwiche T, Collum SD, Bi W, Reynolds JO, Wilson C, Wareing N, et al. Alterations in cardiovascular function in an experimental model of lung fibrosis and pulmonary hypertension. *Exp Physiol.* (2019) 104:568–79. doi: 10.1113/EP087321

227. Horowitz JC, Osterholzer JJ, Marazioti A, Stathopoulos GT. Scar-cinoma": viewing the fibrotic lung mesenchymal cell in the context of cancer biology. *Eur Respir J.* (2016) 47:1842–54. doi: 10.1183/13993003.01201–2015

228. Li S, Zhao G, Chen L, Ding Y, Lian J, Hong G, et al. Resveratrol protects mice from paraquat-induced lung injury: The important role of SIRT1 and NRF2 antioxidant pathways. *Mol Med Rep.* (2016) 13:1833–8. doi: 10.3892/mmr.2015.4710

229. He X, Wang L, Szklarz G, Bi Y, Ma Q. Resveratrol inhibits paraquat-induced oxidative stress and fibrogenic response by activating the nuclear factor erythroid 2-related factor 2 pathway. *J Pharmacol Exp Ther.* (2012) 342:81–90. doi: 10.1124/jpet.112.194142

230. Ramli I, Cheriet T, Posadino AM, Giordo R, Zayed H, Eid AH, et al. Potential therapeutic targets of resveratrol in the prevention and treatment of pulmonary fibrosis. *Front Biosci (Landmark Ed).* (2023) 28:198. doi: 10.31083/j.fbl2809198

231. Li J, Liang C, Zhang ZK, Pan X, Peng S, Lee WS, et al. TAK1 inhibition attenuates both inflammation and fibrosis in experimental pneumoconiosis. *Cell Discov.* (2017) 3:17023. doi: 10.1038/celldisc.2017.23

232. Conte E, Fagone E, Fruciano M, Gili E, Iemmolo M, Vancheri C. Antiinflammatory and antifibrotic effects of resveratrol in the lung. *Histol Histopathol.* (2015) 30:523–9. doi: 10.14670/HH-30.523

233. Yang X, Dong WB, Lei XP, Li QP, Zhang LY, Zhang LP. Resveratrol suppresses hyperoxia-induced nucleocytoplasmic shuttling of SIRT1 and ROS production in PBMC from preterm infants in vitro. *J Matern Fetal Neonatal Med.* (2018) 31:1142–50. doi: 10.1080/14767058.2017.1311310

234. Navarro S, Reddy R, Lee J, Warburton D, Driscoll B. Inhaled resveratrol treatments slow ageing-related degenerative changes in mouse lung. *Thorax.* (2017) 72:451–9. doi: 10.1136/thoraxjnl-2016-208964

235. Burkon A, Somoza V. Quantification of free and protein-bound transresveratrol metabolites and identification of trans-resveratrol-C/O-conjugated diglucuronides - two novel resveratrol metabolites in human plasma. *Mol Nutr Food Res.* (2008) 52:549–57. doi: 10.1002/mnfr.200700290

236. de Santi C, Pietrabissa A, Mosca F, Pacifici GM. Glucuronidation of resveratrol, a natural product present in grape and wine, in the human liver. *Xenobiotica*. (2000) 30:1047–54. doi: 10.1080/00498250010002487

237. Cottart CH, Nivet-Antoine V, Laguillier-Morizot C, Beaudeux JL. Resveratrol bioavailability and toxicity in humans. *Mol Nutr Food Res.* (2010) 54:7–16. doi: 10.1002/mnfr.200900437

238. Ndiaye M, Philippe C, Mukhtar H, Ahmad N. The grape antioxidant resveratrol for skin disorders: promise, prospects, and challenges. *Arch Biochem Biophys.* (2011) 508:164–70. doi: 10.1016/j.abb.2010.12.030

239. Niles RM, Cook CP, Meadows GG, Fu YM, McLaughlin JL, Rankin GO. Resveratrol is rapidly metabolized in athymic (nu/nu) mice and does not inhibit human melanoma xenograft tumor growth. *J Nutr.* (2006) 136:2542–6. doi: 10.1093/jn/136.10.2542

240. Ziegler CC, Rainwater L, Whelan J, McEntee MF. Dietary resveratrol does not affect intestinal tumorigenesis in Apc(Min/+) mice. J Nutr. (2004) 134:5–10. doi: 10.1093/jn/134.1.5

241. Gao X, Xu YX, Divine G, Janakiraman N, Chapman RA, Gautam SC. Disparate in *vitro* and in *vivo* antileukemic effects of resveratrol, a natural polyphenolic compound found in grapes. J Nutr. (2002) 132:2076–81. doi: 10.1093/jn/132.7.2076

242. Niles RM, McFarland M, Weimer MB, Redkar A, Fu YM, Meadows GG. Resveratrol is a potent inducer of apoptosis in human melanoma cells. *Cancer Lett.* (2003) 190:157-63. doi: 10.1016/s0304-3835(02)00676-6

243. Neves AR, Lucio M, Lima JL, Reis S. Resveratrol in medicinal chemistry: a critical review of its pharmacokinetics, drug-delivery, and membrane interactions. *Curr Med Chem.* (2012) 19:1663–81. doi: 10.2174/092986712799945085

244. Amri A, Chaumeil JC, Sfar S, Charrueau C. Administration of resveratrol: What formulation solutions to bioavailability limitations? *J Control Release*. (2012) 158:182–93. doi: 10.1016/j.jconrel.2011.09.083

245. Frau J, Munoz F, Glossman-Mitnik D. A molecular electron density theory study of the chemical reactivity of cis- and trans-resveratrol. *Molecules*. (2016) 21:1650. doi: 10.3390/molecules21121650

246. Goldberg DM, Tsang E, Karumanchiri A, Diamandis E, Soleas G, Ng E. Method to assay the concentrations of phenolic constituents of biological interest in wines. *Anal Chem.* (1996) 68:1688–94. doi: 10.1021/ac951083i

247. Robinson K, Mock C, Liang D. Pre-formulation studies of resveratrol. Drug Dev Ind Pharm. (2015) 41:1464–9. doi: 10.3109/03639045.2014.958753

248. Dhakar NK, Matencio A, Caldera F, Argenziano M, Cavalli R, Dianzani C, et al. Comparative evaluation of solubility, cytotoxicity and photostability studies of resveratrol and oxyresveratrol loaded nanosponges. *Pharmaceutics*. (2019) 11:545. doi: 10.3390/pharmaceutics11100545

249. Brown VA, Patel KR, Viskaduraki M, Crowell JA, Perloff M, Booth TD, et al. Repeat dose study of the cancer chemopreventive agent resveratrol in healthy volunteers: safety, pharmacokinetics, and effect on the insulin-like growth factor axis. *Cancer Res.* (2010) 70:9003–11. doi: 10.1158/0008–5472.CAN-10–2364

250. Almeida L, Vaz-da-Silva M, Falcao A, Soares E, Costa R, Loureiro AI, et al. Pharmacokinetic and safety profile of trans-resveratrol in a rising multiple-dose study in healthy volunteers. *Mol Nutr Food Res.* (2009) 53 Suppl 1:S7–15. doi: 10.1002/mnfr.200800177

251. Hoshino J, Park EJ, Kondratyuk TP, Marler L, Pezzuto JM, van Breemen RB, et al. Selective synthesis and biological evaluation of sulfate-conjugated resveratrol metabolites. *J Med Chem*. (2010) 53:5033–43. doi: 10.1021/jm100274c

252. Patel KR, Scott E, Brown VA, Gescher AJ, Steward WP, Brown K. Clinical trials of resveratrol. Ann N Y Acad Sci. (2011) 1215:161–9. doi: 10.1111/j.1749-6632.2010.05853.x

253. Juhasz B, Mukherjee S, Das DK. Hormetic response of resveratrol against cardioprotection. *Exp Clin Cardiol.* (2010) 15:e134-8.

254. Bolton JL, Dunlap T. Formation and biological targets of quinones: cytotoxic versus cytoprotective effects. *Chem Res Toxicol.* (2017) 30:13–37. doi: 10.1021/acs.chemrestox.6b00256

255. Lee HH, Park SA, Almazari I, Kim EH, Na HK, Surh YJ. Piceatannol induces heme oxygenase-1 expression in human mammary epithelial cells through activation of ARE-driven Nrf2 signaling. *Arch Biochem Biophys.* (2010) 501:142–50. doi: 10.1016/j.abb.2010.06.011

256. Kim HJ, Lee KW, Kim MS, Lee HJ. Piceatannol attenuates hydrogen-peroxideand peroxynitrite-induced apoptosis of PC12 cells by blocking down-regulation of BclXL and activation of JNK. J Nutr Biochem. (2008) 19:459-66. doi: 10.1016/j.jnutbio.2007.06.001

257. Bolton JL, Dunlap TL, Dietz BM. Formation and biological targets of botanical o-quinones. *Food Chem Toxicol.* (2018) 120:700–7. doi: 10.1016/j.fct.2018.07.050

258. Crowell JA, Korytko PJ, Morrissey RL, Booth TD, Levine BS. Resveratrolassociated renal toxicity. *Toxicol Sci.* (2004) 82:614–9. doi: 10.1093/toxsci/kfh263

259. Na JI, Shin JW, Choi HR, Kwon SH, Park KC. Resveratrol as a multifunctional topical hypopigmenting agent. *Int J Mol Sci.* (2019) 20:956. doi: 10.3390/ijms20040956

260. Ito S, Fujiki Y, Matsui N, Ojika M, Wakamatsu K. Tyrosinase-catalyzed oxidation of resveratrol produces a highly reactive ortho-quinone: Implications for melanocyte toxicity. *Pigment Cell Melanoma Res.* (2019) 32:766–76. doi: 10.1111/pcmr.12808

261. Zheng X, Jia B, Tian XT, Song X, Wu ML, Kong QY, et al. Correlation of reactive oxygen species levels with resveratrol sensitivities of anaplastic thyroid cancer cells. *Oxid Med Cell Longev.* (2018) 2018:6235417. doi: 10.1155/2018/6235417

262. Arcanjo NMO, Luna C, Madruga MS, Estevez M. Antioxidant and pro-oxidant actions of resveratrol on human serum albumin in the presence of toxic diabetes metabolites: Glyoxal and methyl-glyoxal. *Biochim Biophys Acta Gen Subj.* (2018) 1862:1938–47. doi: 10.1016/j.bbagen.2018.06.007

263. Martins LA, Coelho BP, Behr G, Pettenuzzo LF, Souza IC, Moreira JC, et al. Resveratrol induces pro-oxidant effects and time-dependent resistance to cytotoxicity in activated hepatic stellate cells. *Cell Biochem Biophys.* (2014) 68:247–57. doi: 10.1007/s12013-013-9703-8

264. de la Lastra CA, Villegas I. Resveratrol as an antioxidant and pro-oxidant agent: mechanisms and clinical implications. *Biochem Soc Trans.* (2007) 35:1156–60. doi: 10.1042/BST0351156

265. Zuo XL, Chen JM, Zhou X, Li XZ, Mei GY. Levels of selenium, zinc, copper, and antioxidant enzyme activity in patients with leukemia. *Biol Trace Elem Res.* (2006) 114:41–53. doi: 10.1385/BTER:114:1:41

266. Hadi SM, Ullah MF, Azmi AS, Ahmad A, Shamim U, Zubair H, et al. Resveratrol mobilizes endogenous copper in human peripheral lymphocytes leading to oxidative DNA breakage: a putative mechanism for chemoprevention of cancer. *Pharm Res.* (2010) 27:979–88. doi: 10.1007/s11095–010-0055–4

267. D'Angelo S, Martino E, Ilisso CP, Bagarolo ML, Porcelli M, Cacciapuoti G. Prooxidant and pro-apoptotic activity of polyphenol extract from Annurca apple and its underlying mechanisms in human breast cancer cells. *Int J Oncol.* (2017) 51:939–48. doi: 10.3892/ijo.2017.4088

268. Demoulin B, Hermant M, Castrogiovanni C, Staudt C, Dumont P. Resveratrol induces DNA damage in colon cancer cells by poisoning topoisomerase II and activates the ATM kinase to trigger p53-dependent apoptosis. *Toxicol Vitro*. (2015) 29:1156–65. doi: 10.1016/j.tiv.2015.04.015

269. Li B, Hou D, Guo H, Zhou H, Zhang S, Xu X, et al. Resveratrol sequentially induces replication and oxidative stresses to drive p53-CXCR2 mediated cellular senescence in cancer cells. *Sci Rep.* (2017) 7:208. doi: 10.1038/s41598-017-00315-4

270. Giordo R, Cossu A, Pasciu V, Hoa PT, Posadino AM, Pintus G. Different redox response elicited by naturally occurring antioxidants in human endothelial cells. *Open Biochem J.* (2013) 7:44–53. doi: 10.2174/1874091X01307010044

271. Baur JA, Pearson KJ, Price NL, Jamieson HA, Lerin C, Kalra A, et al. Resveratrol improves health and survival of mice on a high-calorie diet. *Nature*. (2006) 444:337–42. doi: 10.1038/nature05354

272. Posadino AM, Giordo R, Cossu A, Nasrallah GK, Shaito A, Abou-Saleh H, et al. Flavin oxidase-induced ROS generation modulates PKC biphasic effect of resveratrol on endothelial cell survival. *Biomolecules*. (2019) 9:209. doi: 10.3390/biom9060209 273. Posadino AM, Cossu A, Giordo R, Zinellu A, Sotgia S, Vardeu A, et al. Resveratrol alters human endothelial cells redox state and causes mitochondrialdependent cell death. *Food Chem Toxicol.* (2015) 78:10-6. doi: 10.1016/ jfct.2015.01.017

274. Heo JR, Kim SM, Hwang KA, Kang JH, Choi KC. Resveratrol induced reactive oxygen species and endoplasmic reticulum stress-mediated apoptosis, and cell cycle arrest in the A375SM Malignant melanoma cell line. *Int J Mol Med.* (2018) 42:1427–35. doi: 10.3892/ijmm.2018.3732

275. Elbaz EM, Ahmed KA, Abdelmonem M. Resveratrol mitigates diclofenacinduced hepatorenal toxicity in rats via modulation of miR-144/Nrf2/GSH axis. J Biochem Mol Toxicol. (2022) 36:e23129. doi: 10.1002/jbt.23129

276. Basheer L, Schultz K, Guttman Y, Kerem Z. In silico and in *vitro* inhibition of cytochrome P450 3A by synthetic stilbenoids. *Food Chem.* (2017) 237:895–903. doi: 10.1016/j.foodchem.2017.06.040

277. Wang L, Wang C, Jia Y, Liu Z, Shu X, Liu K. Resveratrol increases antiproliferative activity of bestatin through downregulating P-glycoprotein expression via inhibiting PI3K/akt/mTOR pathway in K562/ADR cells. *J Cell Biochem*. (2016) 117:1233–9. doi: 10.1002/jcb.25407

278. Jia Y, Liu Z, Wang C, Meng Q, Huo X, Liu Q, et al. P-gp, MRP2 and OAT1/ OAT3 mediate the drug-drug interaction between resveratrol and methotrexate. *Toxicol Appl Pharmacol.* (2016) 306:27–35. doi: 10.1016/j.taap.2016.06.030

279. Lee JS, Oh JS, Kim YG, Lee CK, Yoo B, Hong S. Methotrexate-related toxicity in patients with rheumatoid arthritis and renal dysfunction. *Rheumatol Int.* (2020) 40:765–70. doi: 10.1007/s00296-020-04547-y

280. Chiba T, Kimura Y, Suzuki S, Tatefuji T, Umegaki K. Trans-resveratrol enhances the anticoagulant activity of warfarin in a mouse model. J Atheroscler Thromb. (2016) 23:1099–110. doi: 10.5551/jat.31765

281. Symington B, Mapanga RF, Norton GR, Essop MF. Resveratrol co-treatment attenuates the effects of HIV protease inhibitors on rat body weight and enhances cardiac mitochondrial respiration. *PloS One.* (2017) 12:e0170344. doi: 10.1371/journal.pone.0170344

282. Villanueva JA, Sokalska A, Cress AB, Ortega I, Bruner-Tran KL, Osteen KG, et al. Resveratrol potentiates effect of simvastatin on inhibition of mevalonate pathway in human endometrial stromal cells. *J Clin Endocrinol Metab.* (2013) 98:E455–62. doi: 10.1210/jc.2012–3387

283. Klonowska-Szymczyk A, Kulczycka-Siennicka L, Robak T, Smolewski P, Cebula-Obrzut B, Robak E. The impact of agonists and antagonists of TLR3 and TLR9 on concentrations of IL-6, IL10 and sIL-2R in culture supernatants of peripheral blood mononuclear cells derived from patients with systemic lupus erythematosus. *Postepy Hig Med Dosw.* (2017) 71:867–75. doi: 10.5604/01.3001.0010.5266

284. Stephan LS, Almeida ED, Markoski MM, Garavaglia J, Marcadenti A. Red wine, resveratrol and atrial fibrillation. *Nutrients*. (2017) 9:1190. doi: 10.3390/nu9111190

285. Bedada SK, Yellu NR, Neerati P. Effect of resveratrol on the pharmacokinetics of fexofenadine in rats: Involvement of P-glycoprotein inhibition. *Pharmacol Rep.* (2016) 68:338–43. doi: 10.1016/j.pharep.2015.08.018

286. Chai R, Chen Y, Yuan H, Wang X, Guo S, Qi J, et al. Identification of resveratrol, an herbal compound, as an activator of the calcium-activated chloride channel, TMEM16A. *J Membr Biol.* (2017) 250:483–92. doi: 10.1007/s00232–017-9975–9

287. Smoliga JM, Vang O, Baur JA. Challenges of translating basic research into therapeutics: resveratrol as an example. *J Gerontol A Biol Sci Med Sci.* (2012) 67:158–67. doi: 10.1093/gerona/glr062