#### Check for updates

### OPEN ACCESS

EDITED BY Wenhong Deng, Renmin Hospital of Wuhan University, China

REVIEWED BY De Luo, Essen University Hospital, Germany

\*CORRESPONDENCE Xinrui Gao 🖾 gaoxr1899@163.com Hao Chi 🖾 Chihao7511@gmail.com

<sup>†</sup>These authors have contributed equally to this work

RECEIVED 28 December 2023 ACCEPTED 29 January 2024 PUBLISHED 13 February 2024

#### CITATION

He R, Huang S, Lu J, Su L, Gao X and Chi H (2024) Unveiling the immune symphony: decoding colorectal cancer metastasis through immune interactions. *Front. Immunol.* 15:1362709. doi: 10.3389/fimmu.2024.1362709

#### COPYRIGHT

© 2024 He, Huang, Lu, Su, Gao and Chi. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# Unveiling the immune symphony: decoding colorectal cancer metastasis through immune interactions

Ru He<sup>1†</sup>, Shangke Huang<sup>2†</sup>, Jiaan Lu<sup>1†</sup>, Lanqian Su<sup>1</sup>, Xinrui Gao<sup>3\*</sup> and Hao Chi<sup>1\*</sup>

<sup>1</sup>Clinical Medical College, Southwest Medical University, Luzhou, China, <sup>2</sup>Department of Oncology, The Affiliated Hospital of Southwest Medical University, Luzhou, China, <sup>3</sup>Department of Oncology, Yongchuan Hospital of Traditional Chinese Medicine, Chongqing, China

Colorectal cancer (CRC), known for its high metastatic potential, remains a leading cause of cancer-related death. This review emphasizes the critical role of immune responses in CRC metastasis, focusing on the interaction between immune cells and tumor microenvironment. We explore how immune cells, through cytokines, chemokines, and growth factors, contribute to the CRC metastasis cascade, underlining the tumor microenvironment's role in shaping immune responses. The review addresses CRC's immune evasion tactics, especially the upregulation of checkpoint inhibitors like PD-1 and CTLA-4, highlighting their potential as therapeutic targets. We also examine advanced immunotherapies, including checkpoint inhibitors and immune cell transplantation, to modify immune responses and enhance treatment outcomes in CRC metastasis. Overall, our analysis offers insights into the interplay between immune molecules and the tumor environment, crucial for developing new treatments to control CRC metastasis and improve patient prognosis, with a specific focus on overcoming immune evasion, a key aspect of this special issue.

#### KEYWORDS

metastatic colorectal cancer, immunological characteristic, immune escape mechanisms, tumor microenvironment, TME, PD-L1, immunization checkpoints

# 1 Background

Colorectal cancer (CRC), a major global health concern, recorded 1.93 million new cases and approximately 916,000 deaths in 2020, making it the second deadliest cancer worldwide (1, 2). A significant proportion of early-stage CRC patients (25%-50%) progress to metastasis, mainly affecting the liver, lungs, peritoneum, and distant lymph nodes, with a five-year survival rate for metastatic CRC at about 14% (3). The interaction between the immune system and tumor growth, involving phases of elimination, equilibrium, and

escape, is crucial in CRC progression (4). Despite progress in targeted and immunotherapies, which offer advantages over systemic chemotherapy, there are still limitations in current treatments for metastatic CRC (3): Patients with microsatellite stable tumors (5), lack of biomarkers to predict the response to immunotherapy and so on. This review focuses on the roles of immune cells and molecules in CRC metastasis, particularly exploring the mechanisms of immune tolerance and evasion. We aim to evaluate the potential and limitations of these components in clinical applications, highlighting the need for novel therapeutic strategies to overcome immune evasion in metastatic CRC, and to foste r future research in this vital area.

# 2 The role of immune cells in mediating the metastasis cascade of CRC

In the progression of colorectal cancer (CRC), the immune system is fundamentally instrumental (6). The tumor microenvironment (TME), encompassing immune cells, fibroblasts, endothelial cells, and matrix proteins, significantly influences tumor growth and metastatic spread, impacting processes like seeding, proliferation, and evasion of immune response. Key players within the TME include tumor-associated macrophages (TAMs), CD4+ T (helper T) cells, dendritic cells (DCs), regulatory T cells (Tregs), and tumor-associated neutrophils (TANs), each playing a vital role in CRC's metastatic journey.

### 2.1 Tumor-associated macrophages

Traditionally, TAMs are divided into "M1" and "M2" types, which are associated with pro-inflammatory, immune-activating, anti-tumor properties, and immune-suppressive, tumor-promoting properties, respectively (7). M1 type produces pro-inflammatory and immune-activating cytokines like IL-1 $\beta$ , IL-6, and TNF- $\alpha$ . While generally seen as anti-tumor, their effectiveness depends on the environment. For instance, in colitis models, M1 type TAMs can promote chronic inflammation, increasing the risk of CRC. Studies have shown that M1 type TAMs can promote tumor invasion and metastasis by secreting factors like MMP-9. Conversely, M2 type TAMs accelerate tumor progression and invasion by secreting immune-suppressive factors like IL-10 and TGF-B (Figure 1, Table 1). Understanding the dual roles of TAMs opens avenues for targeted therapies. Modulating the balance between M1 and M2 phenotypes may offer a strategy to harness the anti-tumor potential of TAMs, potentially influencing treatment outcomes.

### 2.2 Helper T

Cells CD4+ T cells, particularly the TH17 subset, have been linked to the hepatic metastasis of colorectal cancer (CRC), also

known as CRLM. Research has shown that an increase in these cells after surgery correlates with a worsened prognosis (8) (Figure 1, Table 1). Strategies aimed at modulating these cells may hold promise in improving post-surgery outcomes.

### 2.3 Dendritic cells

As crucial antigen-presenting cells, DCs play a pivotal role in the immune response. They capture and present external antigens to T cells. Within CRC, it's often myeloid DCs that predominate. These cells are integral in cell-mediated immunity and in nudging naïve CD4+ T cells towards a TH1 phenotype. They are also noted for their presence at the tumor's invasive edge and their link to lymph node invasion (9, 10) (Figure 1, Table 1). Strategies that maintain the balance of DCs(Lymophoid DCs and myeloid DCs) and T cells(TH1 and TH2) could be explored as a potential adjunct to existing treatments, aiming to improve outcomes, particularly in lymph node-involved cases.

### 2.4 Regulatory T cells

In the tumor microenvironment (TME), Tregs are key players in immune suppression. Their presence is often tied to heightened metastasis and poorer outcomes in various cancers (11). In CRC, the role of Tregs in the TME is multifaceted, with indications of their dual functionality (12–14) (Table 1). Understanding the nuanced role of Tregs in CRC provides insights for targeted interventions. Modulating Treg activity within the TME may be a viable strategy to disrupt immune evasion mechanisms and improve treatment responses.

### 2.5 Tumor-associated neutrophils

The link between TANs and patient prognosis is a growing research focus. These cells are implicated in promoting tumor invasion, partly through mechanisms like angiogenesis regulation (Figure 1) and increasing tumor cell resistance to VEGF inhibitors. They also contribute to metastasis by accelerating the breakdown of the basement membrane (15, 16) (Table 1). Strategies aimed at regulating TAN functions may enhance the efficacy of existing therapies, especially those targeting angiogenesis.

# 2.6 Other cells in the tumor microenvironment

Beyond immune cells, the TME comprises stromal cells, the extracellular matrix (ECM), cancer-associated fibroblasts (CAFs), and endothelial cells (ECs), all playing roles in metastasis (17). The ECM, made up of structural proteins such as collagen and proteoglycans, undergoes remodeling by TME cells, affecting both its structure and function (18). It serves not just as a scaffold for



tumor cells but also in roles like intercellular adhesion and paracrine signaling, contributing to tumor growth, immune evasion, and metastasis (19–23). CAFs are crucial for ECM upkeep, fibrosis, angiogenesis, immune suppression, invasion, and chemoresistance. The vascular system, composed of ECs, peripheral cells, smooth muscle cells, and progenitor cells, plays a significant role in CRC invasion and therapy targeting (24). The liver is often the primary metastasis site in CRC (25), with interactions between tumor and liver cells, liver sinusoidal endothelial cells (LSECs), and others fostering CAF formation and enhancing cellular stemness and epithelial-mesenchymal transition via exosomes (26, 27). Comprehensive approaches targeting various components of the TME, including ECM, CAFs, and vascular elements, may be crucial. Strategies aimed at disrupting these interactions could potentially hinder metastasis and improve the efficacy of CRC treatments.

# 3 Immune-related molecules in colorectal cancer

Colorectal cancer (CRC) is significantly influenced by the TME, where tumor cells interact intricately with various stromal cells. This interaction is facilitated by a range of soluble elements like cytokines, growth factors, chemokines, and components of the extracellular matrix. In CRC's pathogenesis, cytokines are particularly crucial. Chronic inflammation, for example, often leads to CRC, exacerbated by the dysregulated expression of cytokines.

CRC exhibits a rise in both pro-inflammatory and antiinflammatory cytokines, which are deeply intertwined with the progression and potential outcomes of the disease. The proinflammatory cytokine, IL-17, for instance, not only intensifies

Category	Role/Function in CRC Metastasis	Key Points/Findings	References
Tumor-Associated Macrophages (TAMs)	Mediate tumor growth and metastasis	M1 type: pro-inflammatory, can promote/inhibit tumor; M2 type: immune- suppressive, promote tumor progression	(7)
CD4+ T Cells	Influence hepatic metastasis	TH17 subset linked to worsened prognosis after surgery	(8)
Dendritic Cells (DCs)	Antigen presentation, immune response initiation	Myeloid DCs predominant in CRC; link to lymph node invasion	(9, 10)
Regulatory T Cells (Tregs)	Immune suppression within TME	Associated with heightened metastasis and poorer outcomes	(11–14)
Tumor-Associated Neutrophils (TANs)	Promote tumor invasion and metastasis	Involved in angiogenesis regulation, resistance to VEGF inhibitors	(15, 16)
Cytokines (e.g., IL-17, TNF- α, IL-6)	Modulate tumor growth and immune response	Both pro-inflammatory and anti-inflammatory cytokines play roles in CRC pathogenesis	(28-32)
Cytokines (IL-17A, IL-17F, and IL-22)	Display anti-tumor effects in CRC	Aiding immune cell recruitment, tissue repair, and reducing inflammation and angiogenesis-promoting factors (IL-17A has a dual role in tumor development)	(36)
Immune Evasion Mechanisms (PD-1, PD-L1, CTLA-4)	Inhibit immune cell activation and response	High PD-L1 levels in metastases; TGF- $\beta$ impairs DCs; CTLA-4 inhibits T cell activation	(44, 46–49)
Emerging Immunotherapies	Target immune checkpoints, enhance anti-tumor immunity	PD-1 inhibitors (e.g., nivolumab) show promise, especially in MSI-H/dMMR CRC cases	(57–59)

TABLE 1 The different aspects of immune response-related molecules in CRC metastasis.

tumor-caused inflammation but also assists cancer cells in evading immune detection (28) (Table 1). The IL-23/IL-17 pathway's role in fostering colorectal tumor development is also notable (29). Further, high levels of IL-23, its receptor, and IL-17A correlate with unfavorable prognosis and swift metastatic progression in CRC cases (30).

Cytokines like TNF- $\alpha$ , IL-1 $\beta$ , and IL-6 are pivotal in the intestinal response to inflammation, fostering intricate cell interactions within the intestinal milieu. These interactions involve cells such as intestinal epithelial cells, paneth cells, macrophages, and goblet cells, all contributing to the chronic inflammation seen in inflammatory bowel diseases like Crohn's disease and ulcerative colitis. Both TNF- $\alpha$  and IL-6 are implicated in CRC progression, activating oncogenic transcription factors like NF- $\kappa$ B and STAT3 (31, 32).

Moreover, cytokines IL-11, IL-17A, and IL-22 show heightened protein-level expression in CRC, playing a role in the disease's development in both humans and mice (33–35). IL-17A, IL-17F, and IL-22 display anti-tumor effects in CRC, aiding immune cell recruitment, tissue repair, and reducing inflammation and angiogenesis-promoting factors (36). Interestingly, IL-17A has a dual role in tumor development, whereas IL-17F exhibits tumorsuppressing properties (36) (Table 1).

In addition, the CXCL12/CXCR4 axis activation is linked with the spread of colorectal cancer and peritoneal macrophage involvement (37, 38). The TGF- $\beta$  signaling pathway is also believed to play a crucial role in CRC by governing epithelialmesenchymal transition (EMT) (39). The intricate interplay of abnormally expressed molecules like cytokines, chemokines, growth factors, and enzymes remodeling the matrix contributes to CRC pathogenesis, ultimately affecting disease outcomes (40–43).

# 4 Immune evasion mechanisms of CRC cells

# 4.1 Cancer cells suppress immune cells and immune molecules

To evade immune detection, cancer cells deploy various strategies to inhibit immune cells and molecules. On the surfaces of activated T cells, B cells, and natural killer cells, PD-1, a transmembrane protein, functions as an immune checkpoint receptor. Its main role is to modulate immune responses, diminishing inflammatory activity, thereby serving as a crucial immune checkpoint. Its ligand, PD-L1, another transmembrane protein, is found on tumor cells, macrophages, and other immune cells. These proteins are part of the immune checkpoint protein family. The interaction between PD-L1 and PD-1 suppresses the activation, growth, and tumor-fighting capabilities of CD8+ T cells, aiding in the tumor's immune evasion (44). Research indicates that PD-L1 levels are higher in lung (45) and liver metastases (46) of colorectal cancer (CRC) than in primary tumors. Cancer cells also produce TGF-B, which impairs dendritic cells (DCs) (47). CTLA-4, a receptor on T cells, inhibits T cell activation and growth by interacting with B7 molecules (CD80 and CD86) on antigen-presenting cells. In CRC, increased regulatory T cells (Tregs) in the tumor microenvironment may upregulate CTLA-4, which dampens immune responses through high CTLA-4 expression. This upregulation, along with changes in cytokines and chemokines, can promote CTLA-4 expression. Enhanced CTLA-4 expression further impedes anti-tumor immune responses by competing with CD28 for B7 molecules, thereby reducing T cell activation and proliferation (48) (Figure 1). This creates an immunosuppressive milieu, aiding tumor immune evasion and

10.3389/fimmu.2024.1362709

survival. Additionally, cancer cells limit cytotoxic T cells (CTL) activity through various mechanisms. For instance, TNF-a in CT26 CRC cells heightens PD-L1 expression in stromal cells, which curbs CD8+ T cell Granzyme secretion (49) (Table 1). PD-L1 on tumor and stromal cells binds to PD-1 on T cells, further repressing T cell activation. In colorectal cancer, PD-L1 levels are lower in CRC cells but higher in TAMs, which also hinder the immune response through PD-1 expression and support CRC cell immune evasion by releasing CCL5. Thus, TAMs are key targets in cancer immunotherapy. Advanced stages of CRC, characterized by high PD-L1 expression, are linked with more aggressive disease and poorer survival rates (50). Targeting the PD-1/PD-L1 pathway has become a focal point in immunotherapy. Other cells in the TME also suppress CTL function by releasing TGF- $\beta$ , which lowers the expression of critical lytic enzymes like perforin, Granzyme, and Fas ligand (51). Furthermore, the CCL5/CCR5 signaling pathway recruits regulatory T cells, effectively suppressing CTLs in CRC mouse models (52). Studies by Wei et al.,2017 demonstrate that high PD-L1 expression in colorectal liver metastasis tumors (CRLMs) and primary tumors correlates with an abundance of CD4+ T cells and CTLs (53). Similarly, Katz et al.,2009 observed that high density of CTLs was positively correlated with a 10-year survival rate after CRLM resection, and in multivariate analysis, a high CD8+/low CD4+ phenotype was significantly associated with better long-term survival (54).

# 4.2 Immune evasion mechanisms of CRC cells involving exosomes

Research indicated (55)that small extracellular vesicles (sEVs) are secreted by CRC cells. These sEVs, when absorbed by macrophages, enhance M2 polarization and PD-L1 expression. This process escalates the population of PD-L1+CD206+ macrophages and diminishes T cell activity within the TME CRC. Subsequent studies have identified miR-21-5p and miR-200a within sEVs as crucial in macrophage regulation. By influencing the PTEN/AKT and SCOS1/ STAT1 pathways, these miRNAs foster M2 polarization and PD-L1 expression in macrophages. This leads to a decrease in CD8+ T cell activity and promotes tumor progression (Figure 1). Hence, focusing on sEV-miRNAs specific to CRC and PD-L1 in TAMs could emerge as an innovative approach to treat CRC and boost the effectiveness of anti-PD-L1 treatments.

## 5 Emerging field of immunotherapy

In the TME, various immune cells such as tumor-infiltrating lymphocytes, TAMs, and TANs play a role. Immunotherapy offers new opportunities for the treatment of solid tumors, especially therapies targeting immune checkpoints. Although there has been progress in immunotherapy for colorectal cancer, there are still obstacles. In clinical trials, immunotherapeutic drugs for metastatic colorectal cancer generally show moderate effects, especially in patients with microsatellite stable tumors (5). Challenges include the lack of biomarkers to predict the response to immunotherapy, the relationship between tumor mutation burden and treatment response, the association of immune cells in the TME with treatment outcomes, and the expression of suppressive immune nucleic acid toxic molecules (56). Significant results have been achieved in the treatment of colorectal cancer patients with microsatellite instability (MSI-H)/DNA mismatch repair deficiencies (dMMR) using PD-1 inhibitors such as nivolumab and other immune checkpoint inhibitors (57). Tumor-infiltrating lymphocytes, particularly CD8+ T cells, play an important role in anti-tumor activity. However, tumor cells evade immune attack by expressing inhibitory immune checkpoint receptors such as PD-1, CTLA-4, and LAG-3, as well as regulatory T cell immune suppressive molecules like TGF- $\beta$  and IL-10. In colorectal cancer, the expression of PD-1 and CTLA-4 on T cells and regulatory T cells is associated with a poor prognosis. Therefore, immune checkpoint molecules are potential targets for immunotherapy in colorectal cancer. Blocking immune checkpoints opens a new era in cancer treatment. Targeting immune checkpoints in the tumor microenvironment of colorectal cancer is a novel cancer treatment method that changes the function of immune cells. The response of patients is related to pro-tumoral and anti-tumoral immune cells in the TME (such as TILs, TAMs, and TANs) (58, 59) (Table 1). Anti-PD-1, anti-PD-L1, and anti-CTLA-4 are promising immune checkpoint blockers in colorectal cancer. Furthermore, the emergence of new immune checkpoints like TIM-3 and LAG-3, which can inhibit T or NK cell activity (60), is noteworthy. Combining immune checkpoint blockers with other treatments has shown positive results and may become part of successful colorectal cancer treatment. Immune checkpoint blockers have made some progress in cancer treatment, but still face challenges such as treatment resistance and immune toxicity. Therefore, combining them with other treatment strategies like immune adjuvants and tumor vaccines may enhance their effects. In addition, there are numerous potential targets for immunotherapy in colorectal cancer (CRC), such as the IL-33/ST2 axis (61-63), MDSCs (64), and KRAS (65-67). Immune checkpoint blockers in tumor immunotherapy have broad application prospects and provide a new direction for treatment.

Currently, various immunotherapy strategies are under research, including non-specific immunotherapy, specific immunotherapy, and cell therapy. One or more of these strategies are expected to be used in the clinical treatment of colorectal cancer in the future. Developing effective immunotherapy strategies is crucial for addressing colorectal cancer. Future research will focus on immune escape mechanisms, the immune function of lymphocytes, and the impact of individual genetic polymorphisms on treatment response.

### 6 Discussion

The development and metastasis of colorectal cancer (CRC) are closely related to the interactions between immune cells and molecules in the tumor microenvironment (TME). The different immune cells mentioned in this review, such as Tumor-Associated Macrophages (TAMs), CD4+ T cells, Dendritic Cells (DCs), Regulatory T cells (Tregs), and Tumor-Associated Neutrophils (TANs), each play specific roles and mechanisms in the metastasis of CRC. For instance, the dual nature of M1 and M2 macrophages in TAMs in promoting and inhibiting tumor progression highlights the complexity of the immune system in cancer development. Moreover, the role of CD4+ T cells in liver metastasis of CRC and the critical role of DCs in antigen presentation further emphasize the diversity and importance of immune cells in the TME.

The immune evasion mechanisms of colorectal cancer cells are a key area in cancer research. CRC cells evade the immune system's surveillance and attack through various mechanisms, such as the secretion of sEVs, upregulation of PD-L1 expression, and the action of CTLA-4. These findings not only reveal the complexity of tumor immune evasion but also provide potential targets for developing new immunotherapy strategies.

The development of immunotherapy has brought new hope for the treatment of colorectal cancer. The application of immune checkpoint inhibitors, especially in patients with high microsatellite instability CRC, has shown significant therapeutic effects. However, these treatments have limited effects in patients with microsatellite stable tumors, highlighting the need for more personalized and targeted treatment approaches. Combining immune checkpoint inhibitors with other treatment methods, such as targeted therapy and chemotherapy, may improve treatment effectiveness and reduce drug resistance.

Future research should focus on a deeper understanding of immune escape mechanisms, improving the function of immune cells, and exploring the impact of individual genetic polymorphisms on the response to immunotherapy. Additionally, new therapeutic strategies, such as specific immunotherapies and cell therapies, are expected to play a significant role in the treatment of colorectal cancer. Integrating these aspects of research can not only deepen our understanding of the CRC immune microenvironment but also pave the way for developing more effective treatment strategies.

In summary, the treatment of colorectal cancer is a multifaceted and multilevel challenge that requires the integration of knowledge and technology from different fields. By gaining a deeper understanding of the complex relationship between the immune system and CRC, we can move towards developing more effective and precise treatment methods.

### Author contributions

RH: Conceptualization, Data curation, Writing – original draft. SH: Writing – original draft. JL: Conceptualization, Data curation, Writing – original draft. LS: Writing – original draft. XG: Conceptualization, Writing – original draft, Writing – review & editing. HC: Conceptualization, Supervision, Writing – original draft, Writing – review & editing.

# Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This study was supported by the Student Innovation and Entrepreneurship Program of Southwest Medical University (No: 202310632090).

# Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

# References

1. Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* (2021) 71:209–49. doi: 10.3322/caac.21660

2. Chi H, Huang J, Yan Y, Jiang C, Zhang S, Chen H, et al. Unraveling the role of disulfidptosis-related LncRNAs in colon cancer: a prognostic indicator for immunotherapy response, chemotherapy sensitivity, and insights into cell death mechanisms. *Front Mol Biosci* (2023) 10:1254232. doi: 10.3389/fmolb.2023.1254232

3. Shin AE, Giancotti FG, Rustgi AK. Metastatic colorectal cancer: mechanisms and emerging therapeutics. *Trends Pharmacol Sci* (2023) 44:222-36. doi: 10.1016/j.tips.2023.01.003

4. Dunn GP, Old LJ, Schreiber RD. The three Es of cancer immunoediting. Annu Rev Immunol (2004) 22:329–60. doi: 10.1146/annurev.immunol.22.012703.104803

5. Ghiringhelli F, Fumet JD. Is there a place for immunotherapy for metastatic microsatellite stable colorectal cancer? *Front Immunol* (2019) 10:1816. doi: 10.3389/fimmu.2019.01816

6. Hanus M, Parada-Venegas D, Landskron G, Wielandt AM, Hurtado C, Alvarez K, et al. Microbiota, and microbial metabolites: the unresolved triad in colorectal cancer microenvironment. *Front Immunol* (2021) 12:612826. doi: 10.3389/fimmu.2021.612826

7. Mantovani A, Sica A. Macrophages, innate immunity and cancer: balance, tolerance, and diversity. *Curr Opin Immunol* (2010) 22:231–7. doi: 10.1016/j.coi.2010.01.009

8. Kroemer M, Turco C, Spehner L, Viot J, Idirène I, Bouard A, et al. Investigation of the prognostic value of CD4 T cell subsets expanded from tumor-infiltrating lymphocytes of colorectal cancer liver metastases. *J Immunother Cancer* (2020) 8: e001478. doi: 10.1136/jitc-2020-001478

9. Liu YJ, Kanzler H, Soumelis V, Gilliet M. Dendritic cell lineage, plasticity and cross-regulation. *Nat Immunol* (2001) 2:585–9. doi: 10.1038/89726

10. Dadabayev AR, Sandel MH, Menon AG, Morreau H, Melief CJ, Offringa R, et al. Dendritic cells in colorectal cancer correlate with other tumor-infiltrating immune cells. *Cancer Immunol Immunother* (2004) 53:978–86. doi: 10.1007/s00262-004-0548-2

11. Kampan NC, Madondo MT, McNally OM, Stephens AN, Quinn MA, Plebanski M. Interleukin 6 present in inflammatory ascites from advanced epithelial ovarian cancer patients promotes tumor necrosis factor receptor 2-expressing regulatory T cells. *Front Immunol* (2017) 8:1482. doi: 10.3389/fimmu.2017.01482

12. Olguín JE, Medina-Andrade I, Rodríguez T, Rodríguez-Sosa M, Terrazas LI. Relevance of regulatory T cells during colorectal cancer development. *Cancers (Basel)* (2020) 12:1888. doi: 10.3390/cancers12071888

13. Aristin Revilla S, Kranenburg O, Coffer PJ. Colorectal cancer-infiltrating regulatory T cells: functional heterogeneity, metabolic adaptation, and therapeutic targeting. *Front Immunol* (2022) 13:903564. doi: 10.3389/fimmu.2022.903564

14. Whiteside TL. What are regulatory T cells (Treg) regulating in cancer and why? Semin Cancer Biol (2012) 22:327–34. doi: 10.1016/j.semcancer.2012.03.004

15. Mizuno R, Kawada K, Itatani Y, Ogawa R, Kiyasu Y, Sakai Y. The role of tumorassociated neutrophils in colorectal cancer. *Int J Mol Sci 20* (2019) 20:529. doi: 10.3390/ ijms20030529

16. Piccard H, Muschel RJ, Opdenakker G. On the dual roles and polarized phenotypes of neutrophils in tumor development and progression. *Crit Rev Oncol Hematol* (2012) 82:296–309. doi: 10.1016/j.critrevonc.2011.06.004

17. Quail DF, Joyce JA. Microenvironmental regulation of tumor progression and metastasis. *Nat Med* (2013) 19:1423–37. doi: 10.1038/nm.3394

18. Ricciardelli C, Rodgers RJ. Extracellular matrix of ovarian tumors. Semin Reprod Med (2006) 24:270–82. doi: 10.1055/s-2006-948556

19. Liotta LA. Adhere, degrade, and move: the three-step model of invasion. *Cancer Res* (2016) 76:3115–7. doi: 10.1158/0008-5472.CAN-16-1297

20. Huang J, Zhang L, Wan D, Zhou L, Zheng S, Lin S, et al. Extracellular matrix and its therapeutic potential for cancer treatment. *Signal Transduct Target Ther* (2021) 6:153. doi: 10.1038/s41392-021-00544-0

21. Naba A, Clauser KR, Lamar JM, Carr SA, Hynes RO. Extracellular matrix signatures of human mammary carcinoma identify novel metastasis promoters. *Elife* (2014) 3:e01308. doi: 10.7554/eLife.01308

22. Eble JA, Niland S. The extracellular matrix in tumor progression and metastasis. *Clin Exp Metastasis* (2019) 36:171–98. doi: 10.1007/s10585-019-09966-1

23. Kaushik N, Kim S, Suh Y, Lee SJ. Proinvasive extracellular matrix remodeling for tumor progression. Arch Pharm Res (2019) 42:40–7. doi: 10.1007/s12272-018-1097-0

24. Carmeliet P, Jain RK. Molecular mechanisms and clinical applications of angiogenesis. *Nature* (2011) 473:298-307. doi: 10.1038/nature10144

25. Li J, Yuan Y, Yang F, Wang Y, Zhu X, Wang Z, et al. Expert consensus on multidisciplinary therapy of colorectal cancer with lung metastases (2019 edition). J Hematol Oncol (2019) 12:16. doi: 10.1186/s13045-019-0702-0

26. Sagmeister S, Eisenbauer M, Pirker C, Mohr T, Holzmann K, Zwickl H, et al. New cellular tools reveal complex epithelial-mesenchymal interactions in hepatocarcinogenesis. *Br J Cancer* (2008) 99:151–9. doi: 10.1038/sj.bjc.6604440

27. Kim H, Lee S, Shin E, Seong KM, Jin YW, Youn H, et al. The emerging roles of exosomes as EMT regulators in cancer. *Cells* (2020) 9:861. doi: 10.3390/cells9040861

28. Fabre J, Giustiniani J, Garbar C, Antonicelli F, Merrouche Y, Bensussan A, et al. Targeting the tumor microenvironment: the protumor effects of IL-17 related to cancer type. *Int J Mol Sci* (2016) 17:1433. doi: 10.3390/ijms17091433

29. Grivennikov SI, Wang K, Mucida D, Stewart CA, Schnabl B, Jauch D, et al. Adenoma-linked barrier defects and microbial products drive IL-23/IL-17-mediated tumour growth. *Nature* (2012) 491:254–8. doi: 10.1038/nature11465

30. Wang K, Karin M. The IL-23 to IL-17 cascade inflammation-related cancers. *Clin Exp Rheumatol* (2015) 33:S87–90.

31. Chung SS, Wu Y, Okobi Q, Adekoya D, Atefi M, Clarke O, et al. Proinflammatory cytokines IL-6 and TNF- $\alpha$  Increased telomerase activity through NF- $\kappa$ B/STAT1/STAT3 activation, and withaferin A inhibited the signaling in colorectal cancer cells. *Mediators Inflammation* (2017) 2017:5958429. doi: 10.1155/2017/5958429

32. De Simone V, Franzè E, Ronchetti G, Colantoni A, Fantini MC, Di Fusco D, et al. Th17-type cytokines, IL-6 and TNF- $\alpha$  synergistically activate STAT3 and NF-kB to promote colorectal cancer cell growth. *Oncogene* (2015) 34:3493–503. doi: 10.1038/onc.2014.286

33. Nishina T, Deguchi Y, Ohshima D, Takeda W, Ohtsuka M, Shichino S, et al. Interleukin-11-expressing fibroblasts have a unique gene signature correlated with poor prognosis of colorectal cancer. *Nat Commun* (2021) 12:2281. doi: 10.1038/s41467-021-22450-3

34. Xie Z, Qu Y, Leng Y, Sun W, Ma S, Wei J, et al. Human colon carcinogenesis is associated with increased interleukin-17-driven inflammatory responses. *Drug Des Devel Ther* (2015) 9:1679–89. doi: 10.2147/DDDT.S79431

35. Khare V, Paul G, Movadat O, Frick A, Jambrich M, Krnjic A, et al. IL10R2 overexpression promotes IL22/STAT3 signaling in colorectal carcinogenesis. *Cancer Immunol Res* (2015) 3:1227–35. doi: 10.1158/2326-6066.CIR-15-0031

36. De Simone V, Pallone F, Monteleone G, Stolfi C. Role of T(H)17 cytokines in the control of colorectal cancer. *Oncoimmunology* (2013) 2:e26617. doi: 10.4161/onci.26617

37. Yu X, Shi W, Zhang Y, Wang X, Sun S, Song Z, et al. CXCL12/CXCR4 axis induced miR-125b promotes invasion and confers 5-fluorouracil resistance through enhancing autophagy in colorectal cancer. *Sci Rep* (2017) 7:42226. doi: 10.1038/ srep42226

38. Chatterjee M, von Ungern-Sternberg SN, Seizer P, Schlegel F, Büttcher M, Sindhu NA, et al. Platelet-derived CXCL12 regulates monocyte function, survival, differentiation into macrophages and foam cells through differential involvement of CXCR4-CXCR7. *Cell Death Dis* (2015) 6:e1989. doi: 10.1038/cddis.2015.233

39. Li S, Zhang J, Qian S, Wu X, Sun L, Ling T, et al. S100A8 promotes epithelial-mesenchymal transition and metastasis under TGF- $\beta$ /USF2 axis in colorectal cancer. *Cancer Commun (Lond)* (2021) 41:154–70. doi: 10.1002/cac2.12130

40. Liang J, Chen M, Hughes D, Chumanevich AA, Altilia S, Kaza V, et al. CDK8 selectively promotes the growth of colon cancer metastases in the liver by regulating gene expression of TIMP3 and matrix metalloproteinases. *Cancer Res* (2018) 78:6594–606. doi: 10.1158/0008-5472.CAN-18-1583

41. Duarte S, Baber J, Fujii T, Coito AJ. Matrix metalloproteinases in liver injury, repair and fibrosis. *Matrix Biol* (2015) 44-46:147–56. doi: 10.1016/j.matbio.2015.01.004

42. Zeng X, Zhou J, Xiong Z, Sun H, Yang W, Mok MTS, et al. Cell cycle-related kinase reprograms the liver immune microenvironment to promote cancer metastasis. *Cell Mol Immunol* (2021) 18:1005–15. doi: 10.1038/s41423-020-00534-2

43. Bao S, Ouyang G, Bai X, Huang Z, Ma C, Liu M, et al. Periostin potently promotes metastatic growth of colon cancer by augmenting cell survival via the Akt/ PKB pathway. *Cancer Cell* (2004) 5:329–39. doi: 10.1016/S1535-6108(04)00081-9

44. Ratajczak K, Grel H, Olejnik P, Jakiela S, Stobiecka M. Current progress, strategy, and prospects of PD-1/PDL-1 immune checkpoint biosensing platforms for cancer diagnostics, therapy monitoring, and drug screening. *Biosens Bioelectron* (2023) 240:115644. doi: 10.1016/j.bios.2023.115644

45. Schweiger T, Berghoff AS, Glogner C, Glueck O, Rajky O, Traxler D, et al. Tumor-infiltrating lymphocyte subsets and tertiary lymphoid structures in pulmonary metastases from colorectal cancer. *Clin Exp Metastasis* (2016) 33:727–39. doi: 10.1007/ s10585-016-9813-y

46. Wang HB, Yao H, Li CS, Liang LX, Zhang Y, Chen YX, et al. Rise of PD-L1 expression during metastasis of colorectal cancer: Implications for immunotherapy. *J Dig Dis* (2017) 18:574–81. doi: 10.1111/1751-2980.12538

47. Gabrilovich DI, Ostrand-Rosenberg S, Bronte V. Coordinated regulation of myeloid cells by tumours. *Nat Rev Immunol* (2012) 12:253–68. doi: 10.1038/nri3175

48. Schwartz JC, Zhang X, Fedorov AA, Nathenson SG, Almo SC. Structural basis for co-stimulation by the human CTLA-4/B7-2 complex. *Nature* (2001) 410:604–8. doi: 10.1038/35069112

49. O'Malley G, Treacy O, Lynch K, Naicker SD, Leonard NA, Lohan P, et al. Stromal cell PD-L1 inhibits CD8(+) T-cell antitumor immune responses and promotes colon cancer. *Cancer Immunol Res* (2018) 6:1426–41. doi: 10.1158/2326-6066.CIR-17-0443

50. Topalian SL, Drake CG, Pardoll DM. Targeting the PD-1/B7-H1(PD-L1) pathway to activate anti-tumor immunity. *Curr Opin Immunol* (2012) 24:207–12. doi: 10.1016/j.coi.2011.12.009

51. Mariathasan S, Turley SJ, Nickles D, Castiglioni A, Yuen K, Wang Y, et al. TGF $\beta$  attenuates tumour response to PD-L1 blockade by contributing to exclusion of T cells. *Nature* (2018) 554:544–8. doi: 10.1038/nature25501

52. Tan MC, Goedegebuure PS, Belt BA, Flaherty B, Sankpal N, Gillanders WE, et al. Disruption of CCR5-dependent homing of regulatory T cells inhibits tumor growth in a murine model of pancreatic cancer. *J Immunol* (2009) 182:1746–55. doi: 10.4049/ jimmunol.182.3.1746

53. Wei SC, Levine JH, Cogdill AP, Zhao Y, Anang NAS, Andrews MC, et al. Distinct cellular mechanisms underlie anti-CTLA-4 and anti-PD-1 checkpoint blockade. *Cell* (2017) 170:1120–1133.e17. doi: 10.1016/j.cell.2017.07.024

54. Katz SC, Pillarisetty V, Bamboat ZM, Shia J, Hedvat C, Gonen M, et al. T cell infiltrate predicts long-term survival following resection of colorectal cancer liver metastases. *Ann Surg Oncol* (2009) 16:2524–30. doi: 10.1245/s10434-009-0585-3

55. Yin Y, Liu B, Cao Y, Yao S, Liu Y, Jin G, et al. Colorectal cancer-derived small extracellular vesicles promote tumor immune evasion by upregulating PD-L1 expression in tumor-associated macrophages. *Adv Sci (Weinh)* (2022) 9:2102620. doi: 10.1002/advs.202102620

56. Chiba S, Baghdadi M, Akiba H, Yoshiyama H, Kinoshita I, Dosaka-Akita H, et al. Tumor-infiltrating DCs suppress nucleic acid-mediated innate immune responses through interactions between the receptor TIM-3 and the alarmin HMGB1. *Nat Immunol* (2012) 13:832–42. doi: 10.1038/ni.2376

57. Kasi PM, Budde G, Krainock M, Aushev VN, Koyen Malashevich A, Malhotra M, et al. (ctDNA) serial analysis during progression on PD-1 blockade and later CTLA-4 rescue in patients with mismatch repair deficient metastatic colorectal cancer. *J Immunother Cancer* (2022) 10:e003312. doi: 10.1136/jitc-2021-003312

58. Bonavita E, Galdiero MR, Jaillon S, Mantovani A. Phagocytes as corrupted policemen in cancer-related inflammation. *Adv Cancer Res* (2015) 128:141–71. doi: 10.1016/bs.acr.2015.04.013

59. Saleh R, Elkord E. Acquired resistance to cancer immunotherapy: Role of tumormediated immunosuppression. *Semin Cancer Biol* (2020) 65:13–27. doi: 10.1016/ j.semcancer.2019.07.017

60. Yang M, Du W, Yi L, Wu S, He C, Zhai W, et al. Checkpoint molecules coordinately restrain hyperactivated effector T cells in the tumor microenvironment. *Oncoimmunology* (2020) 9:1708064. doi: 10.1080/2162402X.2019.1708064

61. Cui G, Qi H, Gundersen MD, Yang H, Christiansen I, Sørbye SW, et al. Dynamics of the IL-33/ST2 network in the progression of human colorectal adenoma to sporadic colorectal cancer. *Cancer Immunol Immunother* (2015) 64:181–90. doi: 10.1007/s00262-014-1624-x

62. Maywald RL, Doerner SK, Pastorelli L, De Salvo C, Benton SM, Dawson EP, et al. IL-33 activates tumor stroma to promote intestinal polyposis. *Proc Natl Acad Sci U.S.A.* (2015) 112:E2487–96. doi: 10.1073/pnas.1422445112

63. Akimoto M, Maruyama R, Takamaru H, Ochiya T, Takenaga K. Soluble IL-33 receptor sST2 inhibits colorectal cancer Malignant growth by modifying the tumour microenvironment. *Nat Commun* (2016) 7:13589. doi: 10.1038/ncomms13589

64. Groth C, Hu X, Weber R, Fleming V, Altevogt P, Utikal J, et al. Immunosuppression mediated by myeloid-derived suppressor cells (MDSCs) during tumour progression. Br J Cancer (2019) 120:16–25. doi: 10.1038/s41416-018-0333-1 65. Dias Carvalho P, Guimarães CF, Cardoso AP, Mendonça S, Costa M, Oliveira MJ, et al. KRAS oncogenic signaling extends beyond cancer cells to orchestrate the microenvironment. *Cancer Res* (2018) 78:7–14. doi: 10.1158/0008-5472.CAN-17-2084

66. Ji H, Houghton AM, Mariani TJ, Perera S, Kim CB, Padera R, et al. K-ras activation generates an inflammatory response in lung tumors. *Oncogene* (2006) 25:2105–12. doi: 10.1038/sj.onc.1209237

67. Swidnicka-Siergiejko AK, Gomez-Chou SB, Cruz-Monserrate Z, Deng D, Liu Y, Huang H, et al. Chronic inflammation initiates multiple forms of K-Ras-independent mouse pancreatic cancer in the absence of TP53. *Oncogene* (2017) 36:3149–58. doi: 10.1038/onc.2016.461