Check for updates

OPEN ACCESS

EDITED BY Yuumi Nakamura, Osaka University, Japan

<mark>REVIEWED BY</mark> Tomoko Sumitomo, Tokushima University, Japan

*CORRESPONDENCE Chie Miyabe Chie.miyabe@marianna-u.ac.jp

RECEIVED 25 December 2023 ACCEPTED 05 February 2024 PUBLISHED 23 February 2024

CITATION

Ohashi A, Murayama MA, Miyabe Y, Yudoh K and Miyabe C (2024) Streptococcal infection and autoimmune diseases. *Front. Immunol.* 15:1361123. doi: 10.3389/fimmu.2024.1361123

COPYRIGHT

© 2024 Ohashi, Murayama, Miyabe, Yudoh and Miyabe. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Streptococcal infection and autoimmune diseases

Ayaka Ohashi¹, Masanori A. Murayama², Yoshishige Miyabe¹, Kazuo Yudoh³ and Chie Miyabe^{3*}

¹Department of Immunology and Parasitology, St. Marianna University School of Medicine, Kawasaki, Japan, ²Department of Animal Models for Human Diseases, Institute of Biomedical Science, Kansai Medical University, Osaka, Japan, ³Department of Frontier Medicine, Institute of Medical Science, St. Marianna University School of Medicine, Kawasaki, Japan

Excessive activation of immune cells by environmental factors, such as infection or individual genetic risk, causes various autoimmune diseases. *Streptococcus* species are gram-positive bacteria that colonize the nasopharynx, respiratory tract, gastrointestinal tract, genitourinary tract, and skin. Group A *Streptococcus* (GAS) species cause various symptoms, ranging from mild infections, such as tonsillitis and pharyngitis, to serious infections, such as necrotizing fasciitis and streptococcal toxic shock syndrome. The contribution of GAS infections to several autoimmune diseases, including acute rheumatic fever, vasculitis, and neuropsychiatric disorders, has been studied. In this review, we focus on the association between streptococcal infections and autoimmune diseases, and discuss current research on the mechanisms underlying the initiation and progression of autoimmune diseases.

KEYWORDS

Streptococcus, infection, autoimmune diseases, nephritis-associated plasmin receptor (NAPlr), molecular mimicry

1 Introduction

Streptococci are anaerobic, gram-positive organisms that constitute a heterogeneous group of bacteria that include more than 100 species (1). Streptococci colonize mucosal tissues in the nasopharynx, respiratory tract, gastrointestinal tract, genitourinary tract, and skin (2). Some of these bacteria are harmless to humans; however, some pathogenic species can cause severe infections (3). Streptococcal infections were first reported in 4th century BC, when Hippocrates described the disease erysipelas; however, major advancements were made through microscopic observation and the subsequent description of their forms by Anton van Leeuwenhoek (4). *Streptococcus* was first isolated from the uteri and blood of women with puerperal fever by Louis Pasteur in 1879 and was shown to be the etiological microbe responsible for a condition that caused the highest mortality rates among women and newborns (4). Streptococci were classified based on the Lancefield scheme, which groups streptococcal strains according to the carbohydrate composition of cell wall

antigens, such as polysaccharides (groups A, B, C, E, F, and G), teichoic acids (groups D and N), and lipoteichoic acid (group H) (2).

Streptococcus pyogenes, also known as Group A Streptococcus (GAS), causes mild infections, such as respiratory tract infections, including tonsillitis and pharyngitis, and rarely causes serious infections such as necrotizing fasciitis and streptococcal toxic shock syndrome (STSS), which shows similar symptoms to toxic shock syndrome (TSS) caused by *Staphylococcus aureus* (5). GAS is also known to colonize the skin, causing superficial infections such as pyoderma and impetigo (5, 6). The prevalence of GAS among healthy children with no signs or symptoms of pharyngitis is 12% (7). GAS is considered the most clinically important Streptococcal group due to the high number of cases and clinically important postinfectious sequelae, such as post-streptococcal glomerulonephritis and acute rheumatic fever (8).

Autoimmune diseases are heterogeneous disorders characterized by autoreactive immune responses that result in immune-mediated organ damage (9). Autoimmune diseases are heterogeneous, with a wide range of clinical presentations and courses. Autoimmune diseases can affect virtually every organ in the body, including the heart, blood vessels, skin, joints, kidneys, and central nervous system (10, 11). The association between autoimmune diseases and microbes has been widely reported (12–14). For example, immunoglobulin (Ig)A vasculitis (IgAV) is often triggered by a prior infection, such as an upper respiratory tract infection caused by various microorganisms, including *Streptococcus*, *S. aureus*, and varicella-zoster virus (13). Although *Streptococcus* is associated with a variety of autoimmune rheumatic

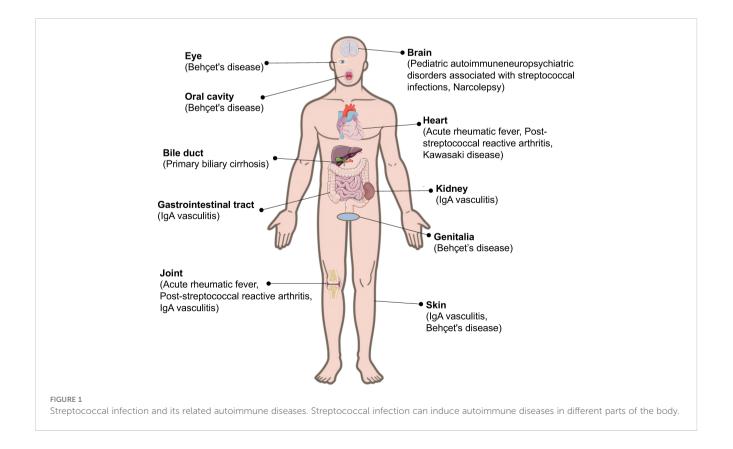
diseases (Figure 1), it has not received as much attention as other bacteria, such as *S. aureus*.

In this review, we provide an overview of the association between streptococcal infections and autoimmune diseases, thereby contributing to our understanding of the pathogenesis of autoimmune diseases.

2 Arthritis

2.1 Acute rheumatic fever

Acute rheumatic fever (ARF) is an autoimmune response to GAS infection. ARF may develop approximately two-four weeks after pharyngeal infection with GAS (15), and is characterized by inflammation of the joints (35-66%) and heart (50-70%), typically manifesting as polyarthritis and valvular regurgitation (16, 17). These symptoms are followed by chorea (10-30%), subcutaneous nodules (0-10%), and erythema marginatum (<6%), which are less common but highly specific manifestations of ARF (17). In 1944, the major and minor manifestations of ARF were defined in the Jones criteria to facilitate accurate ARF diagnoses (18). In most cases, the manifestations of acute rheumatic fever resolve without sequelae. However, valvular lesions can persist in some cases, leading to chronic rheumatic heart disease (RHD). Although ARF itself is not usually life-threatening, cardiac involvement can result in heart failure, stroke, or death (16). It is reported that ARF develops in 0.3-3% of people with untreated or ineffectively treated GAS infections (typically children between the ages of 5



and 14 years) (19). It is also estimated that 33 million cases of RHD are responsible for over 275,000 deaths each year (15). Although the pathogenesis of ARF remains unclear, it is considered to be the result of an autoimmune response to pharyngeal infection with GAS in genetically predisposed individuals, which is mediated by molecular mimicry (16). The cell wall of GAS is composed of NacetylBD-glucosamine linked to a polymeric rhamnose backbone (20). GAS contains M, T, and R surface proteins, as well as lipoteichoic acid, which are involved in bacterial adherence to epithelial cells (20). The M protein is the most important antigenic structure in bacteria and shares structural homology with alpha helical coiled-coil human proteins, such as cardiac myosin, tropomyosin, keratin, laminin, vimentin, and valvular proteins. Mice immunized with streptococcal cell wall and membrane components produce monoclonal antibodies that recognize the human antigens of cardiac structures (21). M proteins can be divided into groups with a preponderance of various symptoms (22). The accumulated data indicate that the biological characteristics of GAS, which has an affinity for the throat, are related to the number of M protein types 1, 3, 5, 6, 14, 18, 19, 24, and 29, the particular emm gene patterns, and share epitopes with human heart tissue (23). Although a potential role of Superantigen (SAg) in the development of RHD has previously been studied, experiments designed to test this question remain scarce and are conflicting (15).

2.2 Post-streptococcal reactive arthritis

Post-streptococcal reactive arthritis (PSRA) is another type of arthritis associated with streptococcal infections in patients who do not fulfill the Jones criteria for the diagnosis of ARF (24). After acute streptococcal pharyngitis, prolonged symptoms of arthritis and multisystem manifestations that do not fulfill the Jones criteria for the diagnosis of ARF indicate PSRA (25). In ARF, arthritis usually develops 10-28 days after GAS-mediated pharyngitis, while in PSRA, arthritis appears after a shorter "incubation" period; approximately 7-10 days after infection (26). The presentation could be similar to that of acute septic arthritis with a sudden onset of fever and severe joint pain (27). Patterns of joint involvement may be monoarticular, polyarticular, migratory, additive, or chronic. The most commonly involved joints are the large joints, such as the knees and ankles, rather than the small joints of the hands and wrists (25). Arthritis is usually nondestructive and self-limiting; however, symptoms may last for months. The response of patients with PSRA to aspirin has been found to be less effective when compared to those with ARF. Whether PSRA is distinct from ARF has not yet been fully addressed. However, since there are substantial clinical, immunological, and genetic differences between PSRA and ARF, PSRA is considered to be a distinct entity (24, 26). There is limited literature on the pathogenesis of PSRA and the hypothesis has not been fully validated yet, but cross reactivity between virulent

molecules of GAS and articular proteins such as keratin, vimentin, and laminin has been extrapolated (28).

3 Vascular disease

3.1 IgA vasculitis

IgAV, previously referred to as Henoch-Schönlein purpura, is a type of vasculitis characterized by IgA-dominant immune deposits that affect small vessels, including the skin, kidney, gastrointestinal tract, and joints (29). Although the disease mechanism has not yet been elucidated, previous studies have identified various infections as a potential trigger for this disease (30). Several bacterial and viral species are associated with the pathogenesis of IgAV. Representative causative pathogens include Streptococcus, S. aureus, Helicobacter pylori, varicella-zoster virus, hepatitis virus, parvovirus, human immunodeficiency virus, cytomegalovirus, and Clostridium difficile (31). Among them, the most frequently isolated microbe is β -hemolytic Streptococcus (32). Nephritis-associated plasmin receptor (NAPlr) has been isolated from GAS as a nephritogenic protein in post-streptococcal acute glomerulonephritis (PSAGN) and was shown to be identical to streptococcal glyceraldehyde-3phosphate dehydrogenase (GAPDH) (33). NAPlr deposition has been detected in the glomeruli of early phase PSAGN, and its distribution is identical to that of plasmin activity, suggesting that NAPlr causes glomerular damage by trapping plasmin and maintaining plasmin activity (33). NAPlr immunoreactivity has been observed in the glomeruli of patients with IgAV with nephritis (34). Recently, we demonstrated that NAPlr was positive in 75.0% of skin tissues derived from patients with IgAV (14). NAPlr immunoreactivity was detected around small dermal vessels, and its distribution was similar to the area where plasmin activity was observed, suggesting that NAPIr-positive perivascular deposits may trigger neutrophil infiltration in IgAV (14). Similar to several other vascular disorders, anti-endothelial cell antibodies (AECA) have been detected in patients with IgAV (35). In IgAV, AECA are exclusively the IgA isotype and directed toward \u03b32-glycoprotein I on the endothelial cell surface (36). IgA AECA may bind to the endothelial cells and enhance IL-8 production via mitogenactivated protein kinase/extracellular regulated kinase pathway and ICAM-1 expression, which is responsible for the perivascular neutrophil infiltration and leukocytoclastic vasculitis in IgAV (37).

3.2 Kawasaki disease

Kawasaki disease (KD) is the most common form of systemic vasculitis affecting children and infants (38). KD initially presents with high fever, mucocutaneous inflammation, and cervical lymphadenopathy, followed by inflammation of the coronary arteries and other cardiovascular structures (39). KD is the leading cause of acquired heart disease in children in developed

countries (40). Scarlet fever caused by GAS presents with symptoms similar to those of KD, including exanthema, enanthema, and cervical lymphadenopathy. However, purulent tonsillitis and the absence of conjunctivitis in scarlet fever can help differentiate between the two diseases (38). Infectious agents are suspected to trigger KD due to the clinical symptoms and seasonal peaks (41). Several microorganisms, such as Streptococci, S. aureus, Propionibacterium acnes, and Yersinia pseudotuberculosis, have been reported to cause KD; however, their causal relationships are yet to be confirmed (42). Metagenomic approaches have revealed the presence of Streptococcus spp. in the lymph nodes and guts of patients with KD (43, 44). SAgs are the powerful T cell stimulators that simultaneously activate the major histocompatibility complex class II (MHC II) molecules and specific V β segments of T cell receptors, leading to the activation of various immune cells (45). In Kawasaki disease, S. aureus strains expressing various SAg have been isolated, in particular the toxic shock syndrome toxin-1 (TSST-1), and analysis of the VB repertoire on the T cell receptor of circulating T cells indicated T cell expansion compatible with SAdriven T cell proliferation (46). The SAg genes of GAS, such as SPE-A, SPE-G, and SPE-J, were also detected more frequently in the stools of children with KD than in those without KD, suggesting that GAS is harbored in the upper airways or gastrointestinal tracts of patients with KD (47). The SAgs of GAS also function as highly potent activators of T lymphocytes and are recognized as causative molecules of the STSS (48). Of interest, some of the streptococcal and staphylococcal SAgs have amino acid sequence homologies, indicating that they have all evolved from a single primordial superantigen (49, 50). For example, TSST-1 of S. aureus and SMEZ of GAS have similar amino acid sequences, and cause TSS and STSS, respectively (50). Although less is known about the relationship between SAgs and autoimmune diseases including KD, it is possible that the variation of the structure of SAgs could result in differences in the affinity toward MHC II or the T cell receptors, which may determine the distinct clinical manifestations as a consequence.

4 Neuroimmune diseases

4.1 Pediatric autoimmune neuropsychiatric disorders associated with streptococcal infections(PANDAS)

Pediatric autoimmune neuropsychiatric disorders associated with streptococcal infections (PANDAS) are acute, sudden onset, obsessive-compulsive disorders accompanied by a previous streptococcal infection (51). PANDAS is characterized by impairment in the basal ganglia, which are responsible for switching between motor and mental behaviors (52). The clinical course of PANDAS is characterized by relapsing-remitting symptoms with significant psychiatric comorbidities accompanying the exacerbations, such as emotional lability,

separation anxiety, nighttime fears and bedtime rituals, cognitive deficits, oppositional behaviors, and motoric hyperactivity (53). A major virulence factor of GAS, namely M protein types 5, 6, and 19, has been detected in the human brain using immunofluorescence staining (54). It has been hypothesized that M proteins trigger the production of antineuronal antibodies, such as antibodies against both D1 and D2 receptors, tubulin and lysoganglioside (55-58). These antineuronal autoantibodies are elevated in the serum and cerebrospinal fluid of patients with PANDAS. Antineuronal antibodies are considered to bind to the receptors on the surface of neuronal cells and trigger the signaling cascade of calcium calmodulin-dependent protein kinase II, tyrosine hydroxylase, which may lead to excess dopamine and the manifestations of PANDAS (59). Studies on mice infected intranasally with GAS have demonstrated the importance of GAS infections and its role in opening the blood-brain barrier, which must be broken to allow IgG to penetrate the brain (21).

4.2 Narcolepsy

Narcolepsy is a sleep disorder characterized by excessive daytime sleepiness, cataplexy, sleep paralysis, hallucinations, and fragmented nocturnal sleep. It usually occurs during adolescence or young adulthood (60). Narcolepsy is strongly associated with the presence of the HLA-DQB1n0602 allele, a variant of the HLA-DQB1 gene. An inciting event, such as infection or vaccination, triggers the destruction of hypocretin-secreting neurons in the hypothalamus, leading to symptoms of narcolepsy (61). A high prevalence of streptococcal throat infections prior to onset has been reported, particularly in prepubertal and peripubertal children (62). It has been hypothesized that Streptococcus has a tropism for hypocretinsecreting neutrons, and its infections can increase the risk of narcolepsy through the activation of general immunity or increased hemato-encephalic barrier permeability to T cells (63). However, despite extensive research, auto-antibodies directed toward hypocretin-secreting neurons have not been proved (64).

5 Other autoimmune/ inflammatory diseases

5.1 Primary biliary cirrhosis

Primary biliary cirrhosis (PBC) is a chronic liver disease that predominantly affects middle-aged women and is characterized by the destruction of small intrahepatic bile ducts, portal inflammation, and progressive scarring (65). The presence of elevated antimitochondrial antibody titers facilitates its recognition and the early detection of associated autoimmune diseases such as Hashimoto's thyroiditis (66). Several bacterial strains have been proposed to cause PBC via molecular mimicry (67). A number of microorganisms have been identified in the tissue

or body fluids of patients with PBC, including Escherichia coli, retroviruses, Epstein-Barr virus, Streptococcus intermedius, S. aureus, and P. acnes (66, 67). Patients with PBC possess higher IgM and IgA class anti-lipoteichoic acid serum titers, which are major components of the gram-positive streptococcal cell wall that mediate the attachment of bacteria to host tissues (68). IgM class S. intermedius titers are significantly higher in the serum of patients with PBC, and bacterial histone-like DNA-binding proteins may be involved in the of PBC (69). Metagenomic analysis of gut microbes in patients with PBC revealed that Streptococcus sp. increased nearly five-fold in mean relative abundance, and may serve as a biomarker to distinguish between patients with PBC and healthy individuals (70). The most widely studied mechanism explaining these observations is the molecular mimicry between bacterial antigens and human epitopes. Another hypothesis is that when bacteria enter the mucosa, bacterial lipoic acid residues may be modified by xenobiotics to form immunoreactive products (67). As most xenobiotics are metabolized in the liver, the potential for liverspecific protein alterations would increase.

5.2 Behçet's disease

Behçet's Disease (BD) is a systemic inflammatory syndrome characterized by relapsing episodes of oral aphthous ulcers, genital ulcers, skin lesions, ocular lesions, and other manifestations including vascular, gastrointestinal, or neurological involvement (71). Histocompatibility leukocyte antigen (*HLA*)-*B**51 is the strongest genetic risk factor of BD (72). Infectious agents (e.g., *Streptococcus sanguinis*, herpes simplex virus, and mycobacteria) have long been proposed as triggering factors for BD pathogenesis (72). An old study reported an abnormal abundance of *S. sanguinis* in the ulcers of patients with BD (73). Another study demonstrated that patients with BD exhibited a positive skin prick specific to intraoral Streptococci (74). Antigens from *S. sanguinis*

are suspected to have high homology with human heat-shock proteins (HSPs) and cross-reactivity leads to an immune response (75). HSPs, which are mainly expressed in mitochondria, are conserved in microorganisms and mammals, and serve as antigens for T cell activation (73). Antibodies against *S. sanguinis* and GAS have been detected more frequently in patients with BD than in controls (76). Hyperactivity against microorganisms and increased serum levels of alarmins in patients with genetic risk factors are considered to cause constitutive NF-kB hyperactivation and neutrophil activation, which are prominent features of BD (72).

6 Future directions/conclusions

The ability of leukocytes to enter tissues in response to immune stimuli is a central feature of host defense (13). However, the excessive activation of leukocytes aggravates inflammation, leading to various autoimmune diseases. GAS can cause diverse skin and respiratory tract symptoms, and occasionally triggers autoimmune sequelae that persist after the infection resolves due to specific features, such as the direct deposition of streptococcal antigens, molecular mimicry, SAgs, modification of bacterial antigens by xenobiotics, or tropism for a specific organ (Table 1). The mechanism by which Streptococci induce persistent autoimmunity has been partially elucidated; however, some questions remain. The common streptococcal antigens that induce autoimmunity and the precise mechanisms by which these antigens promote immune cell attack in specific organs have not been identified. It is also difficult to verify the cause-and-effect relationship between microbes and diseases due to the continuous interaction between these two factors and the lack of appropriate animal models. Further studies in human and animal models are necessary to verify the role of Streptococci in autoimmune diseases, and to identify novel therapeutic and preventive strategies.

TABLE 1 Summarized findings of the association of Streptococcal infection with autoimmune diseases.

Disease	Antigens (Human/Streptococcus)	Immune cells	Mechanism	References
Acute rheumatic fever	cardiac myosin, tropomyosin, keratin, laminin, vimentin	T cells, B cells	molecular mimicry	(21, 23)
IgA vasculitis	NAPlr β2-glycoprotein I	Neutrophils T cells, B cells	deposition of the bacterial antigen molecular mimicry	(14, 34) (35–37)
Kawasaki disease	SPE-A, SPE-G, SPE-J	T cells	superantigens	(46, 47)
PANDAS	dopamine D1 and D2 receptors lysogangliosid, tubulin	T cells, B cells	molecular mimicry	(53–56)
Narcolepsy	hypocretin-secreting neurons?	T cells, B cells?	molecular mimicry?	(61, 62)
Primary biliary cirrhosis	mitochondrial antigens, histone	T cells, B cells	molecular mimicry xenobiotics	(64–67) (65)
Behçet's disease	human heat-shock proteins	T cells, B cells	molecular mimicry	(71, 73)

NAPIr, nephritis-associated plasmin receptor; PANDAS, Pediatric autoimmune neuropsychiatric disorders associated with streptococcal infections.

05

Author contributions

AO: Visualization, Writing – original draft, Writing – review & editing. MM: Writing – review & editing. YM: Supervision, Writing – review & editing. CM: Writing – original draft, Writing – review & editing, Conceptualization.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This work was supported in part by Takeda Medical Research Foundation Research Grant, The Mochida Memorial Foundation for Medical and Pharmaceutical Research, and JSPS KAKENHI (Grant Number 23K07883) to CM.

References

1. Andam CP, Hanage WP. Mechanisms of genome evolution of Streptococcus. Infect Genet Evol. (2015) 33:334-42. doi: 10.1016/j.meegid.2014.11.007.

2. Nobbs AH, Lamont RJ, Jenkinson HF. Streptococcus adherence and colonization. *Microbiol Mol Biol Rev.* (2009) 73:407–50. doi: 10.1128/MMBR.00014-09.

3. Syed S, Viazmina L, Mager R, Meri S, Haapasalo K. Streptococci and the complement system: interplay during infection, inflammation and autoimmunity. *FEBS Lett.* (2020) 594:2570–85. doi: 10.1002/1873-3468.13872.

 Ferretti J, Köhler W. History of streptococcal research. In: Ferretti JJ, Stevens DL, Fischetti VA, editors. Streptococcus pyogenes: Basic Biology to Clinical Manifestations. University of Oklahoma Health Sciences Center © The University of Oklahoma Health Sciences Center., Oklahoma City (OK (2016).

5. Walker MJ, Barnett TC, McArthur JD, Cole JN, Gillen CM, Henningham A, et al. Disease manifestations and pathogenic mechanisms of Group A Streptococcus. *Clin Microbiol Rev.* (2014) 27:264–301. doi: 10.1128/CMR.00101-13.

6. Brouwer S, Rivera-Hernandez T, Curren BF, Harbison-Price N, De Oliveira DMP, Jespersen MG, et al. Pathogenesis, epidemiology and control of Group A Streptococcus infection. *Nat Rev Microbiol.* (2023) 21:431–47. doi: 10.1038/s41579-023-00865-7.

7. Shaikh N, Leonard E, Martin JM. Prevalence of streptococcal pharyngitis and streptococcal carriage in children: a meta-analysis. *Pediatrics*. (2010) 126:e557-64. doi: 10.1542/peds.2009-2648.

8. Cunningham MW. Pathogenesis of group A streptococcal infections. Clin Microbiol Rev. (2000) 13:470-511. doi: 10.1128/CMR.13.3.470.

9. Konig MF. The microbiome in autoimmune rheumatic disease. Best Pract Res Clin Rheumatol. (2020) 34:101473. doi: 10.1016/j.berh.2019.101473.

10. Fairweather D, Rose NR. Women and autoimmune diseases. *Emerg Infect Dis.* (2004) 10:2005–11. doi: 10.3201/eid1011.040367.

11. Miyabe Y, Miyabe C, Iwai Y, Luster AD. Targeting the chemokine system in rheumatoid arthritis and vasculitis. *Jma J.* (2020) 3:182–92. doi: 10.31662/jmaj.2020-0019.

12. Miyabe C, Miyabe Y, Miyata R, Ishiguro N. Pathogens in vasculitis: is it really idiopathic? Jma J. (2021) 4:216-24. doi: 10.31662/jmaj.2021-0021.

13. Miyata R, Miyabe C, Oki H, Motooka D, Nakamura S, Miyabe Y, et al. Alteration of microbial composition in the skin and blood in vasculitis. *Sci Rep.* (2023) 13:15317. doi: 10.1038/s41598-023-42307-7.

14. Miyabe C, Oda T, Miyata R, Miyabe Y, Ishiguro N. Nephritis-associated plasmin receptor in the cutaneous vessels in IgA vasculitis. *J Dermatol.* (2023) 50:102–3. doi: 10.1111/1346-8138.16574.

15. Hurst JR, Kasper KJ, Sule AN, McCormick JK. Streptococcal pharyngitis and rheumatic heart disease: the superantigen hypothesis revisited. *Infect Genet Evol.* (2018) 61:160–75. doi: 10.1016/j.meegid.2018.03.006.

16. Karthikeyan G, Guilherme L. Acute rheumatic fever. Lancet. (2018) 392:161-74. doi: 10.1016/S0140-6736(18)30999-1.

17. Gewitz MH, Baltimore RS, Tani LY, Sable CA, Shulman ST, Carapetis J, et al. Revision of the Jones Criteria for the diagnosis of acute rheumatic fever in the era of Doppler echocardiography: a scientific statement from the American Heart Association. *Circulation.* (2015) 131:1806–18. doi: 10.1161/CIR.00000000000205.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The author(s) declared that they were an editorial board member of Frontiers, at the time of submission. This had no impact on the peer review process and the final decision.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

18. Auala T, Zavale BG, Mbakwem A, Mocumbi AO. Acute rheumatic fever and rheumatic heart disease: highlighting the role of group A streptococcus in the global burden of cardiovascular disease. *Pathogens*. (2022) 11. doi: 10.3390/pathogens11050496.

19. Dooley LM, Ahmad TB, Pandey M, Good MF, Kotiw M. Rheumatic heart disease: A review of the current status of global research activity. *Autoimmun Rev.* (2021) 20:102740. doi: 10.1016/j.autrev.2020.102740.

20. Girschick HJ, Guilherme L, Inman RD, Latsch K, Rihl M, Sherer Y, et al. Bacterial triggers and autoimmune rheumatic diseases. *Clin Exp Rheumatol.* (2008) 26:S12–7.

21. Cunningham MW. Molecular mimicry, autoimmunity, and infection: the crossreactive antigens of group A streptococci and their sequelae. *Microbiol Spectr.* (2019) 7. doi: 10.1128/microbiolspec.GPP3-0045-2018.

22. Jansen TL, Janssen M, van Riel PL. Grand rounds in rheumatology: acute rheumatic fever or post-streptococcal reactive arthritis: a clinical problem revisited. *Br J Rheumatol.* (1998) 37:335–40. doi: 10.1093/rheumatology/37.3.335.

23. Martin DR. Rheumatogenic and nephritogenic group A streptococci. Myth or reality? An opening lecture. Adv Exp Med Biol. (1997) 418:21–7.

24. Barash J, Mashiach E, Navon-Elkan P, Berkun Y, Harel L, Tauber T, et al. Differentiation of post-streptococcal reactive arthritis from acute rheumatic fever. *J Pediatr.* (2008) 153:696–9. doi: 10.1016/j.jpeds.2008.05.044.

25. Li EK. Rheumatic disorders associated with streptococcal infections. *Baillieres Best Pract Res Clin Rheumatol.* (2000) 14:559–78. doi: 10.1053/berh.2000.0093.

26. Uziel Y, Perl L, Barash J, Hashkes PJ. Post-streptococcal reactive arthritis in children: a distinct entity from acute rheumatic fever. *Pediatr Rheumatol Online J.* (2011) 9:32. doi: 10.1186/1546-0096-9-32.

27. Padhan P, Danda D. Clinical spectrum of post-streptococcal reactive arthritis (PSRA) revisited: Juvenile versus adult-onset disease. *Int J Rheum Dis.* (2019) 22:750–1. doi: 10.1111/1756-185X.13524.

28. Ahmed S, Padhan P, Misra R, Danda D. Update on post-streptococcal reactive arthritis: narrative review of a forgotten disease. *Curr Rheumatol Rep.* (2021) 23:19. doi: 10.1007/s11926-021-00982-3.

29. Jennette JC, Falk RJ, Bacon PA, Basu N, Cid MC, Ferrario F, et al. 2012 revised international chapel hill consensus conference nomenclature of vasculitides. *Arthritis Rheum*. (2013) 65:1–11. doi: 10.1002/art.37715.

30. Brogan P, Eleftheriou D. Vasculitis update: pathogenesis and biomarkers. *Pediatr Nephrol.* (2018) 33:187-98. doi: 10.1007/s00467-017-3597-4.

31. Sugino H, Sawada Y, Nakamura M. IgA vasculitis: etiology, treatment, biomarkers and epigenetic changes. *Int J Mol Sci.* (2021) 22. doi: 10.3390/ijms22147538.

32. Duquesnoy B. Henoch-Schönlein purpura. Baillieres Clin Rheumatol. (1991) 5:253-61. doi: 10.1016/S0950-3579(05)80282-1.

33. Oda T, Yoshizawa N, Yamakami K, Sakurai Y, Takechi H, Yamamoto K, et al. The role of nephritis-associated plasmin receptor (NAPlr) in glomerulonephritis associated with streptococcal infection. *J BioMed Biotechnol.* (2012), 2012 417675. doi: 10.1155/2012/417675.

34. Masuda M, Nakanishi K, Yoshizawa N, Iijima K, Yoshikawa N. Group A streptococcal antigen in the glomeruli of children with Henoch-Schönlein nephritis. *Am J Kidney Dis.* (2003) 41:366–70. doi: 10.1053/ajkd.2003.50045.

35. Legendre P, Régent A, Thiebault M, Mouthon L. Anti-endothelial cell antibodies in vasculitis: A systematic review. *Autoimmun Rev.* (2017) 16:146–53. doi: 10.1016/ j.autrev.2016.12.012.

36. Yang YH, Chang CJ, Chuang YH, Hsu HY, Yu HH, Lee JH, et al. Identification and characterization of IgA antibodies against β2-glycoprotein I in childhood Henoch-Schönlein purpura. *Br J Dermatol.* (2012) 167:874–81. doi: 10.1111/bjd.2012.167.issue-4.

37. Guilpain P, Mouthon L. Antiendothelial cells autoantibodies in vasculitisassociated systemic diseases. *Clin Rev Allergy Immunol.* (2008) 35:59–65. doi: 10.1007/s12016-007-8069-3.

38. Hedrich CM, Schnabel A, Hospach T. Kawasaki disease. Front Pediatr. (2018) 6:198. doi: 10.3389/fped.2018.00198.

39. Newburger JW, Takahashi M, Burns JC. Kawasaki disease. J Am Coll Cardiol. (2016) 67:1738–49. doi: 10.1016/j.jacc.2015.12.073.

40. Taubert KA, Rowley AH, Shulman ST. Seven-year national survey of Kawasaki disease and acute rheumatic fever. *Pediatr Infect Dis J.* (1994) 13:704–8. doi: 10.1097/0000654-199408000-00005.

41. Miyabe C, Miyabe Y, Bricio-Moreno L, Lian J, Rahimi RA, Miura NN, et al. Dectin-2-induced CCL2 production in tissue-resident macrophages ignites cardiac arteritis. *J Clin Invest*. (2019) 129:3610–24. doi: 10.1172/JCI123778.

42. Hara T, Yamamura K, Sakai Y. The up-to-date pathophysiology of Kawasaki disease. Clin Transl Immunol. (2021) 10:e1284. doi: 10.1002/cti2.1284.

43. Kinumaki A, Sekizuka T, Hamada H, Kato K, Yamashita A, Kuroda M. Characterization of the gut microbiota of Kawasaki disease patients by metagenomic analysis. *Front Microbiol.* (2015) 6:824. doi: 10.3389/fmicb.2015.00824.

44. Katano H, Sato S, Sekizuka T, Kinumaki A, Fukumoto H, Sato Y, et al. Pathogenic characterization of a cervical lymph node derived from a patient with Kawasaki disease. *Int J Clin Exp Pathol.* (2012) 5:814–23.

45. Esposito S, Polinori I, Rigante D. The gut microbiota-host partnership as a potential driver of Kawasaki syndrome. *Front Pediatr.* (2019) 7:124. doi: 10.3389/fped.2019.00124.

46. Meissner HC, Leung DY. Superantigens, conventional antigens and the etiology of Kawasaki syndrome. *Pediatr Infect Dis J.* (2000) 19:91–4. doi: 10.1097/00006454-200002000-00001.

47. Suenaga T, Suzuki H, Shibuta S, Takeuchi T, Yoshikawa N. Detection of multiple superantigen genes in stools of patients with Kawasaki disease. *J Pediatr.* (2009) 155:266–70. doi: 10.1016/j.jpeds.2009.03.013.

48. McCormick JK, Yarwood JM, Schlievert PM. Toxic shock syndrome and bacterial superantigens: an update. *Annu Rev Microbiol.* (2001) 55:77–104. doi: 10.1146/annurev.micro.55.1.77.

49. Schlievert PM, Bohach GA, Ohlendorf DH, Stauffacher CV, Leung DY, Murray DL, et al. Molecular structure of staphylococcus and streptococcus superantigens. *J Clin Immunol.* (1995) 15:4s–10s. doi: 10.1007/BF01540887.

50. Proft T, Fraser JD. Bacterial superantigens. *Clin Exp Immunol.* (2003) 133:299–306. doi: 10.1046/j.1365-2249.2003.02203.x.

51. Prato A, Gulisano M, Scerbo M, Barone R, Vicario CM, Rizzo R. Diagnostic approach to pediatric autoimmune neuropsychiatric disorders associated with streptococcal infections (PANDAS): A narrative review of literature data. *Front Pediatr.* (2021) 9:746639. doi: 10.3389/fped.2021.746639.

52. Baj J, Sitarz E, Forma A, Wróblewska K, Karakuła-Juchnowicz H. Alterations in the nervous system and gut microbiota after β -hemolytic streptococcus group A infection-characteristics and diagnostic criteria of PANDAS recognition. *Int J Mol Sci.* (2020) 21. doi: 10.3390/ijms21041476.

53. Swedo SE, Leonard HL, Garvey M, Mittleman B, Allen AJ, Perlmutter S, et al. Pediatric autoimmune neuropsychiatric disorders associated with streptococcal infections: clinical description of the first 50 cases. *Am J Psychiatry.* (1998) 155:264–71. doi: 10.1176/ajp.155.2.264.

54. Bronze MS, Dale JB. Epitopes of streptococcal M proteins that evoke antibodies that cross-react with human brain. *J Immunol.* (1993) 151:2820–8. doi: 10.4049/jimmunol.151.5.2820.

55. Kirvan CA, Cox CJ, Swedo SE, Cunningham MW. Tubulin is a neuronal target of autoantibodies in Sydenham's chorea. *J Immunol.* (2007) 178:7412–21. doi: 10.4049/ jimmunol.178.11.7412.

56. Brimberg L, Benhar I, Mascaro-Blanco A, Alvarez K, Lotan D, Winter C, et al. Behavioral, pharmacological, and immunological abnormalities after streptococcal exposure: a novel rat model of Sydenham chorea and related neuropsychiatric disorders. *Neuropsychopharmacology*. (2012) 37:2076–87. doi: 10.1038/npp.2012.56.

57. Ben-Pazi H, Stoner JA, Cunningham MW. Dopamine receptor autoantibodies correlate with symptoms in Sydenham's chorea. *PloS One.* (2013) 8:e73516. doi: 10.1371/journal.pone.0073516.

58. Cox CJ, Zuccolo AJ, Edwards EV, Mascaro-Blanco A, Alvarez K, Stoner J, et al. Antineuronal antibodies in a heterogeneous group of youth and young adults with tics and obsessive-compulsive disorder. *J Child Adolesc Psychopharmacol.* (2015) 25:76–85. doi: 10.1089/cap.2014.0048.

59. Orefici G, Cardona F, Cox CJ, Cunningham MW. Pediatric autoimmune neuropsychiatric disorders associated with streptococcal infections (PANDAS). In: Ferretti JJ, Stevens DL, Fischetti VA, editors. *Streptococcus pyogenes: Basic Biology to Clinical Manifestations*. University of Oklahoma Health Sciences Center © The University of Oklahoma Health Sciences Center, Oklahoma City (OK (2016).

60. Rocca FL, Pizza F, Ricci E, Plazzi G. Narcolepsy during childhood: an update. *Neuropediatrics*. (2015) 46:181–98. doi: 10.1055/s-00000041.

61. Viorritto EN, Kureshi SA, Owens JA. Narcolepsy in the pediatric population. Curr Neurol Neurosci Rep. (2012) 12:175-81. doi: 10.1007/s11910-011-0246-3.

62. Aran A, Lin L, Nevsimalova S, Plazzi G, Hong SC, Weiner K, et al. Elevated antistreptococcal antibodies in patients with recent narcolepsy onset. *Sleep*. (2009) 32:979– 83. doi: 10.1093/sleep/32.8.979.

63. Lopes DA, Coelho FM, Pradella-Hallinan M, de Araújo Melo MH, Tufik S. Infancy narcolepsy: Streptococcus infection as a causal factor. *Sleep Sci.* (2015) 8:49–52. doi: 10.1016/j.slsci.2015.02.002.

64. Kornum BR, Faraco J, Mignot E. Narcolepsy with hypocretin/orexin deficiency, infections and autoimmunity of the brain. *Curr Opin Neurobiol.* (2011) 21:897–903. doi: 10.1016/j.conb.2011.09.003.

65. Kaplan MM. Primary biliary cirrhosis. N Engl J Med. (1996) 335:1570-80. doi: 10.1056/NEJM199611213352107.

66. Quigley EM. Primary biliary cirrhosis and the microbiome. Semin Liver Dis. (2016) 36:349–53. doi: 10.1055/s-0036-1594006.

67. Baio P, Brucato A, Buskila D, Gershwin ME, Giacomazzi D, Lopez LR, et al. Autoimmune diseases and infections: controversial issues. *Clin Exp Rheumatol.* (2008) 26:S74–80.

68. Haruta I, Hashimoto E, Kato Y, Kikuchi K, Kato H, Yagi J, et al. Lipoteichoic acid may affect the pathogenesis of bile duct damage in primary biliary cirrhosis. *Autoimmunity*. (2006) 39:129–35. doi: 10.1080/08916930600623841.

69. Haruta I, Kikuchi K, Hashimoto E, Kato H, Hirota K, Kobayashi M, et al. A possible role of histone-like DNA-binding protein of Streptococcus intermedius in the pathogenesis of bile duct damage in primary biliary cirrhosis. *Clin Immunol.* (2008) 127:245–51. doi: 10.1016/j.clim.2008.01.010.

70. Lv LX, Fang DQ, Shi D, Chen DY, Yan R, Zhu YX, et al. Alterations and correlations of the gut microbiome, metabolism and immunity in patients with primary biliary cirrhosis. *Environ Microbiol.* (2016) 18:2272–86. doi: 10.1111/1462-2920.13401.

71. Sakane T, Takeno M, Suzuki N, Inaba G. Behçet's disease. N Engl J Med. (1999) 341:1284–91. doi: 10.1056/NEJM199910213411707.

72. Perazzio SF, Andrade LEC, de Souza AWS. Understanding Behçet's disease in the context of innate immunity activation. *Front Immunol.* (2020) 11:586558. doi: 10.3389/fimmu.2020.586558.

73. Joubert M, André M, Barnich N, Billard E. Microbiome and Behçet's disease: a systematic review. *Clin Exp Rheumatol.* (2023) 41:2093-104. doi: 10.55563/ clinexprheumatol/zbt4gx.

74. Togashi A, Saito S, Kaneko F, Nakamura K, Oyama N. Skin prick test with selfsaliva in patients with oral aphthoses: a diagnostic pathergy for Behcet's disease and recurrent aphthosis. *Inflammation Allergy Drug Targets*. (2011) 10:164–70. doi: 10.2174/187152811795564109.

75. Leccese P, Alpsoy E. Behçet's disease: an overview of etiopathogenesis. Front Immunol. (2019) 10:1067. doi: 10.3389/fimmu.2019.01067.

76. Yokota K, Hayashi S, Fujii N, Yoshikawa K, Kotake S, Isogai E, et al. Antibody response to oral streptococci in Behçet's disease. *Microbiol Immunol.* (1992) 36:815–22. doi: 10.1111/j.1348-0421.1992.tb02083.x.