



OPEN ACCESS

EDITED BY

Elmostafa Bahraoui,
INSERM U1043 Centre de Physiopathologie
de Toulouse Purpan, France

REVIEWED BY

Simon H. Bridge,
Northumbria University, United Kingdom
Daniel Sepúlveda-Crespo,
Carlos III Health Institute (ISCIII), Spain

*CORRESPONDENCE

Anna C. Hearps
✉anna.hearps@burnet.edu.au

[†]These authors have contributed equally to
this work

RECEIVED 08 December 2023

ACCEPTED 25 January 2024

PUBLISHED 14 February 2024

CITATION

Hearps AC, Vootukuru N,
Ebrahimnezhaddarzi S, Harney BL, Boo I,
Nguyen L, Pavlyshyn D, Dietze PM,
Drummer HE, Thompson AJ, Jaworowski A,
Hellard ME, Sacks-Davis R and Doyle JS
(2024) Injecting drug use and hepatitis C virus
infection independently increase biomarkers
of inflammatory disease risk which are
incompletely restored by curative
direct-acting antiviral therapy.
Front. Immunol. 15:1352440.
doi: 10.3389/fimmu.2024.1352440

COPYRIGHT

© 2024 Hearps, Vootukuru,
Ebrahimnezhaddarzi, Harney, Boo, Nguyen,
Pavlyshyn, Dietze, Drummer, Thompson,
Jaworowski, Hellard, Sacks-Davis and Doyle.
This is an open-access article distributed under
the terms of the [Creative Commons Attribution
License \(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use, distribution or
reproduction in other forums is permitted,
provided the original author(s) and the
copyright owner(s) are credited and that the
original publication in this journal is cited, in
accordance with accepted academic
practice. No use, distribution or reproduction
is permitted which does not comply with
these terms.

Injecting drug use and hepatitis C virus infection independently increase biomarkers of inflammatory disease risk which are incompletely restored by curative direct-acting antiviral therapy

Anna C. Hearps^{1,2*†}, Nikil Vootukuru^{1,3†},
Salimeh Ebrahimnezhaddarzi¹, Brendan L. Harney^{1,2,4},
Irene Boo¹, Long Nguyen¹, Damian Pavlyshyn¹,
Paul M. Dietze^{1,5}, Heidi E. Drummer^{1,6,7},
Alexander J. Thompson⁸, Anthony Jaworowski^{1,2},
Margaret E. Hellard^{1,2,4,6,9}, Rachel Sacks-Davis^{1,4,9}
and Joseph S. Doyle^{1,2}

¹Disease Elimination Program, Burnet Institute, Melbourne, VIC, Australia, ²Department of Infectious Diseases, The Alfred and Monash University, Melbourne, VIC, Australia, ³Department of Gastroenterology, Eastern Health and Monash University, Melbourne, VIC, Australia, ⁴School of Public Health and Preventive Medicine, Monash University, Melbourne, VIC, Australia, ⁵National Drug Research Institute, Curtin University, Melbourne, VIC, Australia, ⁶The Peter Doherty Institute for Infection and Immunity, University of Melbourne, Melbourne, VIC, Australia, ⁷Department of Microbiology, Monash University, Melbourne, VIC, Australia, ⁸Department of Gastroenterology, St Vincent's Hospital and the University of Melbourne, Melbourne, VIC, Australia, ⁹Melbourne School of Population and Global Health, The University of Melbourne, Melbourne, VIC, Australia

Background: Hepatitis C virus (HCV) infections are more prevalent in people who inject drugs (PWID) who often experience additional health risks. HCV induces inflammation and immune alterations that contribute to hepatic and non-hepatic morbidities. It remains unclear whether curative direct acting antiviral (DAA) therapy completely reverses immune alterations in PWID.

Methods: Plasma biomarkers of immune activation associated with chronic disease risk were measured in HCV-seronegative (n=24) and HCV RNA+ (n=32) PWID at baseline and longitudinally after DAA therapy. Adjusted generalised estimating equations were used to assess longitudinal changes in biomarker levels. Comparisons between community controls (n=29) and HCV-seronegative PWID were made using adjusted multiple regression modelling.

Results: HCV-seronegative PWID exhibited significantly increased levels of inflammatory biomarkers including soluble (s) TNF-RII, IL-6, sCD14 and sCD163 and the diabetes index HbA1c as compared to community controls. CXCL10, sTNF-RII, vascular cell adhesion molecule-1 and lipopolysaccharide binding protein (LBP) were additionally elevated in PWID with viremic HCV infection as compared to HCV- PWID. Whilst curative DAA therapy reversed

some biomarkers, others including LBP and sTNF-RII remained elevated 48 weeks after HCV cure.

Conclusion: Elevated levels of inflammatory and chronic disease biomarkers in PWID suggest an increased risk of chronic morbidities such as diabetes and cardiovascular disease. HCV infection in PWID poses an additional disease burden, amplified by the incomplete reversal of immune dysfunction following DAA therapy. These findings highlight the need for heightened clinical surveillance of PWID for chronic inflammatory diseases, particularly those with a history of HCV infection.

KEYWORDS

people who inject drugs, hepatitis C virus, inflammation, direct-acting antivirals, biomarkers

Background

Chronic hepatitis C virus (HCV) infection is a major public health issue (1), with approximately 58.6 million people living with chronic HCV and an estimated 1.5 million new infections each year (1, 2). HCV primarily infects liver hepatocytes causing inflammation and liver damage leading to morbidities including liver cirrhosis and hepatocellular carcinoma (3). HCV also causes extrahepatic manifestations including increased risk of non-hodgkins lymphoma, kidney and rheumatic disease, type 2 diabetes and cardiovascular disease (4, 5). These extrahepatic morbidities are likely mediated at least in part by systemic activation of circulating immune cells and inflammation in response to viral particles and antigens during viremia, as well as cellular damage resulting from virus infection. Similar to HCV, chronic HIV infection is also associated with increased risk of multiple inflammatory diseases (6) and this risk persists despite viral suppression with antiretroviral therapy, implying some impacts of chronic viral disease are not fully reversed upon suppression of viral replication.

HCV can now be cured with direct acting antiviral (DAA) therapy, which provides substantial reductions in both hepatic and non-hepatic sequelae of HCV (5). However, people treated for HCV with interferon-free DAA therapy continue to experience substantially higher rates of all-cause mortality after HCV cure, ranging from 3-14 times higher than the general population depending on the extent of liver disease at the time of treatment (7). It is thus unclear whether DAA therapy completely reverses HCV-related pathologies that may contribute to increased risk of both hepatic and extrahepatic morbidities. Understanding systemic HCV-associated changes which persist post-cure are highly relevant for the ongoing clinical management of individuals with a history of HCV infection.

Inflammation is a key pathogenic mechanism of liver damage during HCV infection and may also drive extrahepatic comorbidities such as cardiovascular disease which are more prevalent in people with

HCV (reviewed in (5)). Chronic HCV infection is associated with systemic immune activation and elevated levels of interferons, inflammatory biomarkers including IL-6 and TNF/TNF receptors, chemokines such as CXCL10 and markers of myeloid cell activation including soluble (s) markers sCD163 and sCD14 (8-13). Conflicting data exist regarding the impact of HCV elimination on reversal of immune activation; whilst some biomarkers including CXCL10, IL-10 and sCD163 are consistently reduced following HCV elimination (8, 9, 13-18), others including IL-6, TNF and sCD14 are unaffected or inconsistently altered by DAA therapy (9, 13-16). Importantly, numerous studies suggest that DAA therapy does not restore levels of many parameters including sCD163 and CXCL10 to those found in comparable, HCV-seronegative populations (8, 13, 14, 17-19). Chronic inflammation is associated with increased risk of a number of chronic morbidities and mortality in both the general population and in the setting of HIV and HCV infection (20-23). Therefore, understanding whether elements of HCV-associated inflammation persist post DAA therapy is relevant for the long-term clinical management of individuals after HCV cure.

In most high-income countries, the HCV epidemic is concentrated in populations of people who inject drugs (PWID) who represent up to 80% of new HCV infections in many countries (24). PWID experience additional health risks associated with injecting drug use (25-27) and behavioural factors such as tobacco smoking, alcohol use and diet, which increase chronic inflammation (28, 29), may further contribute to poor health. The additional burden of HCV in PWID may therefore potentiate inflammatory diseases and/or compromise the restorative potential of DAA-mediated HCV cure on immune function. However, to date there is limited information regarding the discrete effects of HCV and substance use-related risks on inflammatory pathways in PWID populations who are disproportionately affected by HCV.

In this study we aimed to delineate the independent effects of both HCV infection and injecting drug use on soluble inflammatory

and immune activation biomarkers among people with HCV who inject drugs. Furthermore, we investigated whether these biomarkers normalised to levels comparable to HCV-uninfected PWID after curative DAA therapy.

Materials and methods

Study design

Cohorts and participants

This retrospective cohort study included a total of 85 participants in three distinct groups (1); participants with no history of injecting drug use or HCV infection (general community controls; “CC”, n=29); (2) participants with a history of injecting drug use but not HCV infection (“PWID/HCV-”, n=24) and (3) participants with a history of injecting drug use and current HCV infection (“PWID/HCV+”, n=32). Participants in the PWID groups were selected from three prospective cohort studies (Networks 2, SuperMIX and TAP), which all included PWID recruited from Melbourne, Australia. These studies involved regular study visits which included structured interviews and blood collection (30–32). PWID participants were primarily Australian-born with approximately 80% reporting recent heroin use, with a minority of participants reporting meth/amphetamine or pharmaceutical opiate use (30–32).

Inclusion/exclusion criteria

A history of ever using injecting drugs was an inclusion criterion for both PWID cohorts. PWID/HCV- participants were defined as those with a negative HCV antibody test and were included if they were comparable in age (within 5 years) and sex to those in the PWID/HCV+ group. Inclusion criteria for PWID/HCV+ participants were being HCV antibody and RNA positive at baseline, completion of 12 weeks of DAA therapy with sofosbuvir/velpatasvir, achieving sustained viral response after 12 weeks of DAA therapy, maintaining undetectable HCV RNA at all post-DAA timepoints up to and including 48 weeks and the availability of stored samples at the required timepoints. CC were volunteers from the general community with no self-reported history of acute or chronic disease (including HCV infection) or injecting drug use (33, 34). CC were sex-matched and of a comparable age (within 5 years) to PWID groups. The study was approved by the Alfred Hospital Research Ethics Committee (approval #40/18).

Analysis timepoints

Samples from PWID/HCV+ participants were analysed at baseline (prior to initiating DAA therapy; “pre-treatment”), after 12 weeks of DAA when HCV viremia was undetectable (end of treatment; “EOT”) and 48 weeks after DAA initiation (“48wk post-DAA”). Due to sample availability, the EOT timepoint analysis included samples taken at 8 (n=2), 12 (n=29) or 28 (n=1) weeks after DAA commencement whilst the 48wk post-DAA timepoint included samples taken at 48 (n=29), 60 (n=2) or 68 (n=1) weeks post DAA commencement. Pre-HCV infection samples were

available for some individuals in the PWID/HCV+ group which were also included in the analysis. HCV genotypes were not determined here, however a previous analysis of PWID populations from which these samples were obtained reported a predominance of genotype 1 (45%) and 3 (42%, primarily 3a) (30), consistent with other reports regarding HCV genotype prevalence in Australia (35, 36). Participants in the CC and PWID/HCV+ groups were analysed at a single timepoint.

Data collection, blood sampling and biomarker analysis

For both PWID groups, socio-demographical, behavioural (including drug injecting frequency) and clinical (HCV antibody, HCV RNA, platelets, white cell count, alanine/aspartate transaminase [ALT/AST], bilirubin, albumin, protein) data were obtained from the Networks 2, TAP and SuperMIX databases. Available data for the CC group was limited to age, sex and self-reported medical history.

Cryopreserved plasma samples for each group were used to measure plasma levels of immune biomarkers relevant to chronic disease and/or known to be altered by HCV including markers of (1) immune activation (CXCL10), (2) endothelial activation (soluble vascular cell adhesion molecule-1 [VCAM]), (3) inflammation (soluble TNF-receptor II [sTNF-RII], IL-6, high sensitivity C-reactive protein [hsCRP]), (4) coagulation (d-dimer) and (5) myeloid cell activation (sCD14, sCD163 and lipopolysaccharide binding protein [LBP]) using customised human magnetic Luminex assays (R&D Systems, Minneapolis, MN), high sensitivity Quantikine ELISA (for IL-6; R&D Systems) or quantitative immunoturbidity assay (for CRP; ARCHITECT, Abbott Laboratories, Abbott Park, IL). Levels of glycated haemoglobin (HbA1c; indicative of diabetes risk) were measured by ELISA (Aviva Systems Biology, San Diego, CA).

Data analysis

All statistical analyses were performed using the R programming environment, version 4.1. Differences between baseline characteristics of the study groups were assessed using Chi-squared and Fisher exact tests for categorical variables and Wilcoxon rank sum and Wilcoxon rank sum exact tests for continuous variables. All inflammatory markers were log-transformed to account for non-normal distribution. Multivariable linear regression models were used to evaluate differences in the log values of each inflammatory and immune activation marker between CC and PWID/HCV- control groups. Models were adjusted for age and sex given the potential effects of these variable on the parameters measured. History of injecting drug use was assessed as a binary exposure variable. Generalised estimating equation models with an autoregressive structure were used to evaluate differences between the log values of each inflammatory marker among PWID/HCV- and PWID/HCV+ by group and DAA-therapy related timepoint (PWID/HCV- group vs

PWID/HCV+ group at pre-treatment, end of treatment and 48 weeks post treatment timepoints). Models were adjusted for age (continuous variable), sex (binary variable) and injection frequency (daily injecting drug use as a binary variable; participants with missing data were excluded from these analyses). Statistical significance was assessed at $p < 0.05$ using two-sided p-values.

Results

Patient characteristics

Demographic and clinical information for the cohorts utilised in this study are described in Tables 1, 2. Basic demographic information was available for CC participants whilst more detailed information was available for PWID groups. Groups were well matched for age and sex and there were no reported cases of HIV or HBV co-infection. At first included timepoint, all participants in the PWID/HCV- and PWID/HCV+ groups reported a history of injecting drug use; 75% and 100% of HCV- and HCV+ PWID participants reported injecting drug use in the past month and 43% and 42% reported daily injecting drug use over past month (defined as injecting ≥ 7 times per week), respectively. PWID/HCV+ were more likely to be unemployed but also more likely to be in stable housing as compared to PWID/HCV- ($p = 0.032$ for both in unadjusted analyses, Table 2). Liver function biomarkers (ALT and AST) were elevated in PWID/HCV+ ($p \leq 0.001$ for both) whilst plasma albumin was slightly lower ($p = 0.015$, Table 2).

People with a history of injecting drug use exhibit elevated levels of a range of biomarkers of inflammation and chronic disease risk

A preliminary analysis was conducted comparing participants in the CC and PWID/HCV- groups to identify any differences in

TABLE 1 Baseline cohort description of community controls and HCV-people who inject drugs (PWID).

Characteristic	PWID (HCV-)	Community Controls	p-value ²
n	24	29	
Sex			0.650
Female	8 (33%)	8 (28%)	
Male	16 (67%)	21 (72%)	
Age	37 (32, 41) ¹	40 (33, 47) ¹	0.386
Unknown	3	0	
Injecting Drugs Monthly	18 (75%)	0 (0%)	<0.001

¹Median (IQR); n (%).

²Pearson's Chi-squared test; Wilcoxon rank sum test; Fisher's exact test.

TABLE 2 Baseline demographics of people who inject drugs (PWID) cohorts.

Characteristic	PWID/HCV- ¹	PWID/HCV+ ¹	p-value ²
n	24	32	
Sex			0.495
Female	8 (33%)	8 (25%)	
Male	16 (67%)	24 (75%)	
Age	37 (32, 41)	39 (34, 47)	0.239
Employment Status			0.032
Employed	7 (29%)	2 (6%)	
Unemployed	17 (71%)	29 (94%)	
Unknown	0	1	
Accommodation Status ³			0.032
Stable	17 (71%)	29 (94%)	
Unstable	7 (29%)	2 (6%)	
Unknown	0	1	
Alcohol Use			0.123
Never	7 (32%)	12 (46%)	
Less than 4 times a week	8 (36%)	12 (46%)	
4 or more times a week	7 (32%)	2 (8%)	
Unknown	2	6	
Age Commenced Injecting	16.5 (15.8, 21.2)	18.0 (15.0, 21.0)	0.858
Unknown	0	1	
Injecting drug use – past month	18 (75%)	25 (100%)	0.010
Unknown	0	7	
Injecting Drugs Daily	9 (43%)	10 (42%)	0.936
Unknown	3	8	
ALT (U/L)	19 (15, 28)	50 (38, 78)	<0.001
Unknown	0	1	
AST (U/L)	24 (21, 36)	43 (35, 62)	0.001
Unknown	2	1	
White cell count (x10 ⁹ /L)	7.00 (5.32, 8.38)	7.40 (6.20, 8.50)	0.757
Unknown	18	1	
Platelets (x10 ⁹ /L)	253 (233, 344)	243 (184, 308)	0.343
Unknown	18	1	
Total Bilirubin (umol/L)	6.0 (5.0, 8.8)	7.0 (4.0, 12.0)	0.435
Unknown	2	1	
Albumin (g/L)	41.0 (39.0, 43.0)	38.0 (37.0, 40.0)	0.015

(Continued)

TABLE 2 Continued

Characteristic	PWID/HCV ⁻¹	PWID/HCV ⁺¹	p-value ²
Unknown	2	1	
Total Protein (g/L)	74.0 (69.0, 77.0)	74.0 (71.5, 77.0)	0.949
Unknown	2	1	

¹Median (IQR); n (%).

²Wilcoxon rank sum exact test; Pearson's Chi-squared test; Fisher's exact test; Wilcoxon rank sum test. P values were derived from unadjusted analyses.

³Stable accommodation included owner occupied, rental and rent free. All other forms of accommodation including boarding, boarding house, squat, homeless/street were defined as unstable.

ALT, alanine transaminase; AST, aspartate transaminase.

Bold values indicate those which are statistically significant (p<0.05).

biomarker levels (Figure 1). Multivariable linear regression analysis indicated that a lifetime history of injecting drug use was associated with significantly higher levels of multiple inflammatory biomarkers including sTNF-RII and IL-6 (adjusted p<0.001 for both) and the myeloid cell activation markers sCD163 and sCD14 (p=0.048 and 0.003 respectively; Table 3, where a negative beta coefficient indicates lower biomarker levels in CC). Plasma levels of glycated haemoglobin (HbA1c), which indicates glycaemic control over the past 3 months and is thus a relevant biomarker for diabetes, were also significantly elevated in PWID/HCV⁻ as compared to the CC group (p<0.001). Elevated levels of the coagulation marker d-dimer and the acute phase inflammatory factor hsCRP in PWID/HCV⁻ did not reach statistical significance (p=0.058 and 0.051 respectively). Age and sex were not significantly related to levels of the abovementioned biomarkers in this analysis.

To investigate whether recent injecting drug use contributed to the abovementioned differences, regression analyses adjusting for injecting drug use within the past month was performed (also adjusting for age and sex). These analyses indicated individuals who had injected drugs within the previous month exhibited significantly higher levels of sTNF-RII, d-dimer, HbA1c (p<0.01 for all) and sCD14 (p=0.031; Supplementary Table 1). These data indicate that this cohort of PWID seronegative for HCV exhibit elevated levels of a range of soluble biomarkers including those associated with increased risk of chronic diseases, and that recent injecting drug use may contribute to elevated levels of some of these factors.

Viremic HCV infection is associated with elevated levels of biomarkers indicative of immune and endothelial activation

Given the abovementioned differences in biomarker levels in PWID/HCV⁻ as compared to those who did not report having injected drugs, we assessed the specific effect of viremic HCV infection on immune activation biomarkers exclusively in PWID populations. For these analyses, we compared 24 PWID/HCV⁻ (median age [IQR] 37 [32, 41], 33% female, Table 2) and 32 PWID with viremic HCV infection at baseline (PWID/HCV⁺; median age 39 [34, 47]; 25% female) with a median [IQR] viral load of 202,229 [62,237 - 1,987,626] copies/mL at enrolment. Generalised estimating equation models were used adjusting for age, sex and daily injecting drug use (Table 4, Figure 2). Viremic HCV infection was associated with elevated levels of the immune

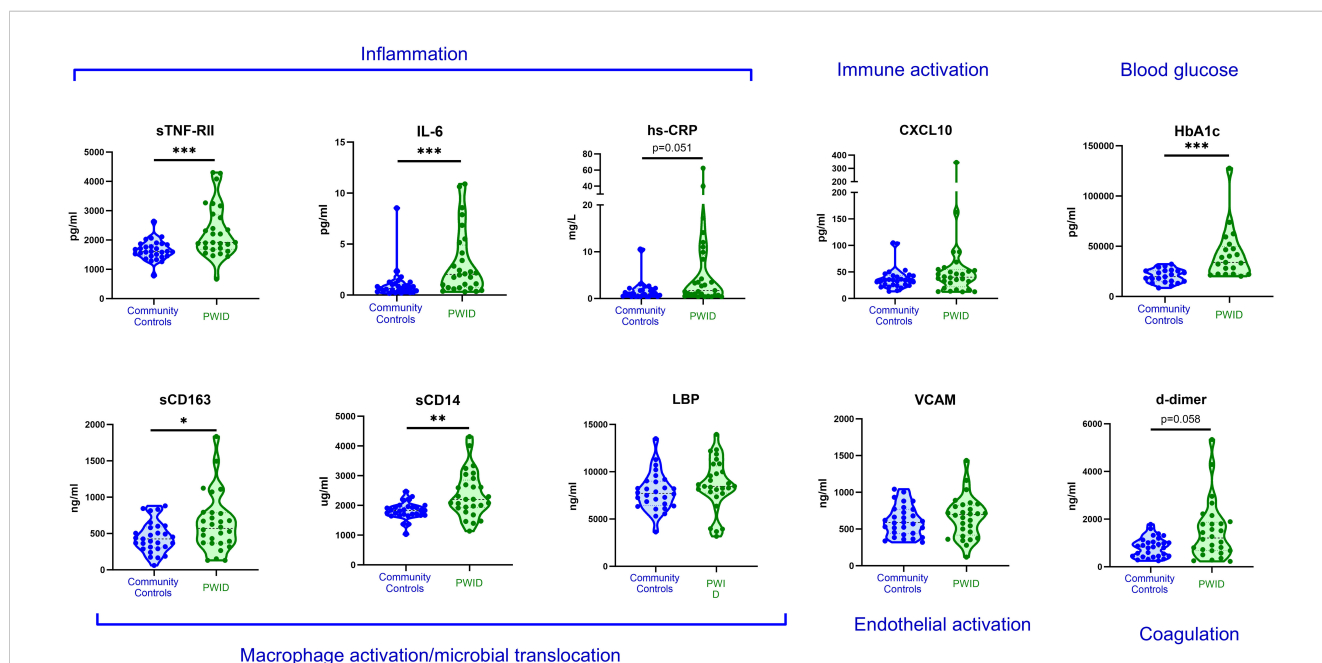


FIGURE 1

Plasma biomarkers levels in community controls and PWID. Plasma biomarkers of inflammation and chronic disease risk were measured in HCV-community controls and PWID of a comparable age and sex. Annotations in blue text indicate association of each biomarker with relevant biological process. *, ** and *** indicate p value <0.05, 0.01 and 0.001 respectively as determined by multivariable linear regression analysis adjusted for age, sex and recent injecting drug use as described in Table 3.

TABLE 3 Multivariable Linear Regression analysis of inflammatory biomarkers in community controls with reference to PWID.

	Sex: Male		Age (10/years)		History of Injecting Drugs	
	Beta (95% CI) ¹	p-value	Beta (95% CI) ¹	p-value	Beta (95% CI) ¹	p-value
Log CXCL10	0.14 (-0.23 to 0.51)	0.437	-0.10 (-0.32 to 0.12)	0.365	-0.12 (-0.47 to 0.22)	0.472
Log sTNF-RII	0.02 (-0.15 to 0.19)	0.836	0.04 (-0.06 to 0.14)	0.431	-0.31 (-0.47 to -0.15)	<0.001
Log VCAM	0.10 (-0.17 to 0.37)	0.457	0.01 (-0.14 to 0.17)	0.866	-0.04 (-0.28 to 0.21)	0.774
Log sCD163	0.03 (-0.34 to 0.40)	0.860	-0.03 (-0.24 to 0.19)	0.812	-0.35 (-0.69 to 0.00)	0.048
Log D-Dimer	-0.36 (-0.75 to 0.03)	0.073	0.22 (-0.01 to 0.45)	0.064	-0.35 (-0.72 to 0.01)	0.058
Log sCD14	-0.09 (-0.25 to 0.07)	0.249	0.03 (-0.07 to 0.12)	0.551	-0.23 (-0.37 to -0.08)	0.003
Log LBP	-0.15 (-0.35 to 0.04)	0.121	0.01 (-0.10 to 0.13)	0.810	-0.02 (-0.20 to 0.16)	0.829
Log IL-6	-0.03 (-0.62 to 0.56)	0.924	0.02 (-0.33 to 0.37)	0.901	-0.98 (-1.5 to -0.43)	<0.001
Log hsCRP	-0.26 (-1.0 to 0.50)	0.491	0.19 (-0.25 to 0.64)	0.386	-0.71 (-1.4 to 0.00)	0.051
Log HbA1c	-0.07 (-0.40 to 0.27)	0.685	-0.01 (-0.20 to 0.17)	0.905	-0.64 (-0.93 to -0.34)	<0.001

¹CI, Confidence Interval.

s, soluble; TNF-RII, tumor necrosis factor receptor II; VCAM, vascular cellular adhesion molecule; LBP, lipopolysaccharide binding protein; IL-6, interleukin-6; hsCRP, high sensitivity C-reactive protein; HbA1c, glycated haemoglobin.

Bold values indicate those which are statistically significant (p<0.05).

activation marker CXCL10 (p=0.007), the inflammatory marker sTNF-RII (p=0.002), the endothelial activation marker VCAM (p=0.014) and also the acute phase protein LBP (p=0.005), which binds to bacterial lipopolysaccharide (LPS; Table 4, Figures 2, 3). Levels of other biomarkers including IL-6, sCD163, sCD14 and d-dimer which were elevated in PWID/HCV- as compared with CC, were not significantly different in viremic PWID/HCV+ as compared with PWID/HCV- (Table 4). Increasing age was associated with higher levels of the blood coagulation biomarker d-dimer but lower levels of HbA1c (p ≤ 0.01 for both, Table 4). Sex

did not significantly alter any parameter, whilst the association between daily injection frequency and higher d-dimer levels approached statistical significance (p=0.057, data not shown), consistent with observations made comparing PWID/HCV- to CC (Table 3). Taken together, these analyses indicate that in addition to increased inflammation identified above to be associated with a history of injecting drug use, PWID with viremic HCV infection exhibit significantly elevated levels of additional biomarkers of inflammation as compared to PWID without HCV.

TABLE 4 Generalised estimating equation analysis of inflammatory biomarkers in PWID/HCV+ pre and post DAA therapy compared to HCV-uninfected PWID.

	Timepoint (Ref: PWID/HCV-)						Age (10/years)	
	Pre-treatment		EOT		48wk post-DAA		Beta (95% CI) ¹	p-value
	Beta (95% CI) ¹	p-value	Beta (95% CI) ¹	p-value	Beta (95% CI) ¹	p-value		
Log CXCL10	0.50 (0.14 to 0.87)	0.007	-0.26 (-0.59 to 0.08)	0.134	-0.16 (-0.65 to 0.32)	0.508	-0.21 (-0.44 to 0.03)	0.083
Log sTNF-RII	0.31 (0.11 to 0.52)	0.002	0.09 (-0.10 to 0.28)	0.337	0.25 (0.05 to 0.45)	0.014	-0.01 (-0.14 to 0.12)	0.866
Log VCAM	0.42 (0.08 to 0.76)	0.014	0.11 (-0.23 to 0.45)	0.530	0.16 (-0.19 to 0.51)	0.367	0.10 (-0.13 to 0.33)	0.392
Log sCD163	0.18 (-0.19 to 0.55)	0.333	-0.15 (-0.48 to 0.17)	0.348	-0.30 (-0.66 to 0.05)	0.095	0.07 (-0.13 to 0.28)	0.489
Log d-dimer	0.13 (-0.22 to 0.48)	0.460	0.11 (-0.22 to 0.43)	0.524	0.07 (-0.28 to 0.41)	0.707	0.23 (0.07 to 0.39)	0.005
Log sCD14	0.07 (-0.08 to 0.22)	0.331	0.15 (0.01 to 0.29)	0.040	0.07 (-0.10 to 0.25)	0.411	0.00 (-0.08 to 0.08)	0.996
Log LBP	0.19 (0.06 to 0.33)	0.005	0.20 (0.06 to 0.34)	0.006	0.18 (0.02 to 0.35)	0.030	-0.02 (-0.12 to 0.09)	0.759
Log IL-6	-0.23 (-0.85 to 0.39)	0.472	0.00 (-0.54 to 0.53)	0.987	0.03 (-0.55 to 0.61)	0.922	-0.17 (-0.50 to 0.16)	0.314
Log hsCRP	0.41 (-0.46 to 1.3)	0.355	NT		0.47 (-0.43 to 1.4)	0.305	-0.18 (-0.64 to 0.27)	0.430
Log HbA1c	NT		NT		-0.04 (-0.30 to 0.22)	0.759	-0.17 (-0.30 to -0.04)	0.010

¹CI, Confidence Interval; NT, Not tested.

NB Sex and daily injecting drug use were included as covariates in the analysis but did not significantly affect the parameters shown (not shown).

Bold values indicate those which are statistically significant (p<0.05).

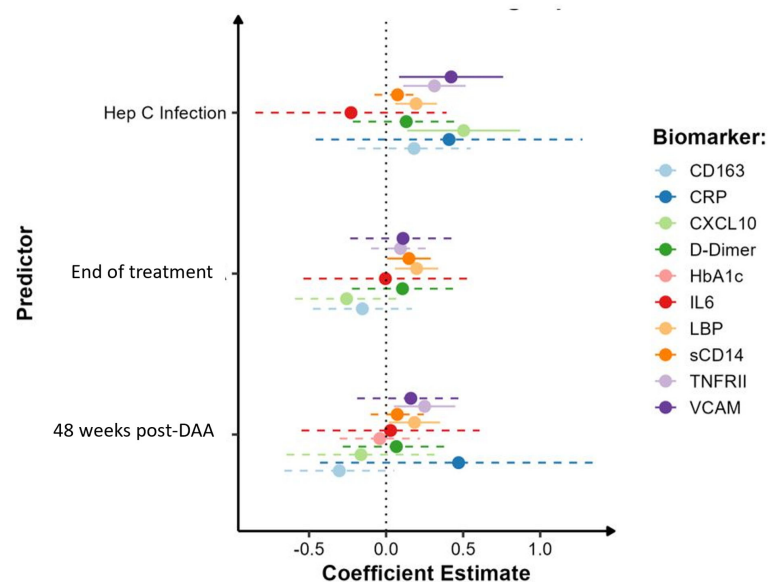


FIGURE 2

Impact of HCV infection and DAA therapy on biomarker levels in PWID. Model estimates of inflammatory biomarkers by HCV infection (pre-treatment timepoint), at end of DAA treatment and 48 weeks post-DAA therapy in HCV+ PWID with reference to HCV-uninfected PWID. Analysis was adjusted for sex, age and daily injecting drug use. Solid lines indicate parameters which were significantly different to HCV-uninfected PWID.

Impact of DAA therapy on immune biomarkers in PWID with HCV

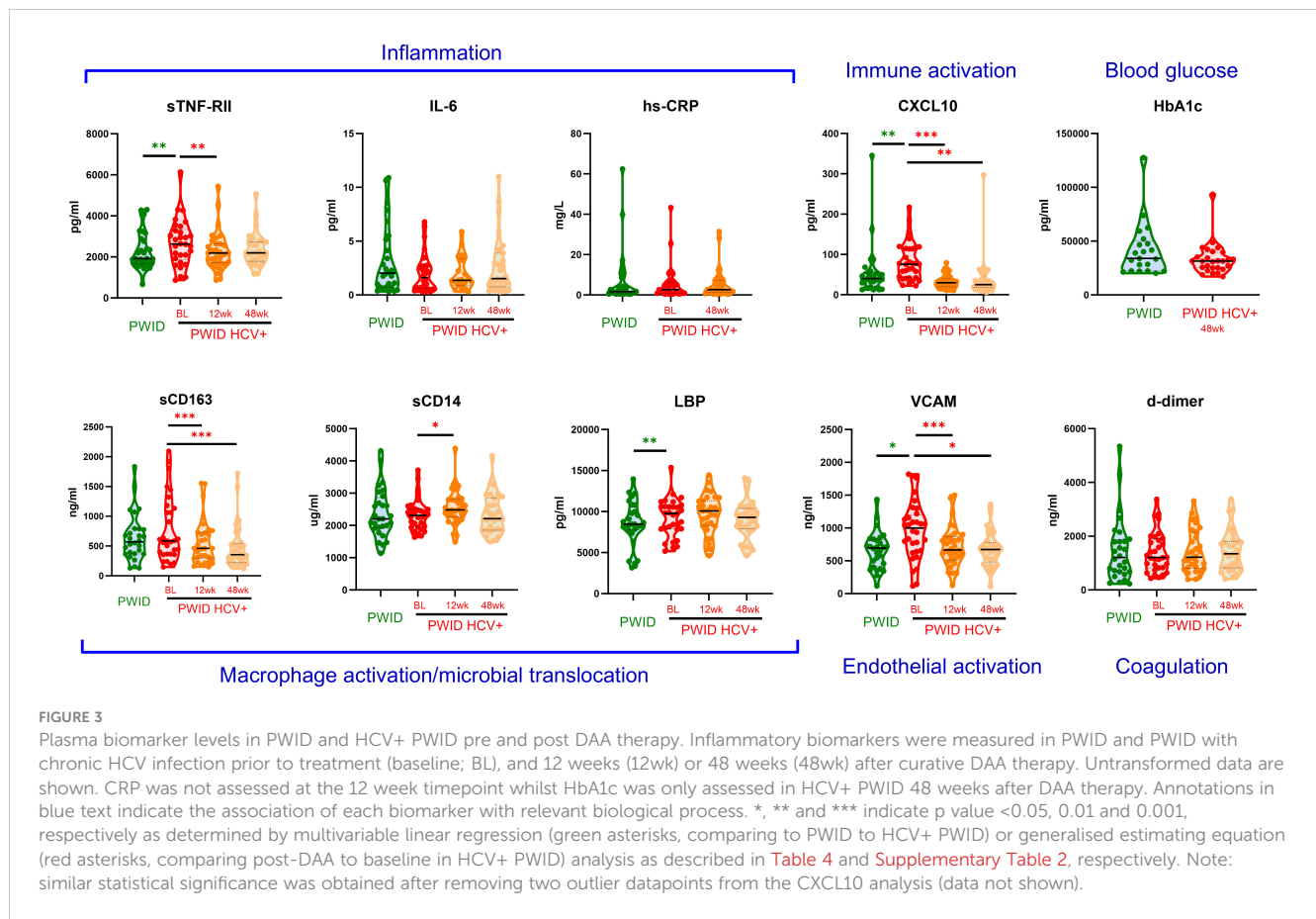
All participants in the PWID/HCV+ cohort initiated DAA therapy with sofosbuvir/velpatasvir and achieved sustained virological response at the end of treatment (EOT). HCV RNA was not detected in any participant between the EOT and 48wk timepoints (data not shown). Analysis of changes in immune biomarkers over time after DAA therapy indicated that levels of CXCL10 and VCAM, which were both elevated in PWID/HCV+ prior to treatment, were significantly reduced by DAA therapy ($p < 0.05$ for all timepoints as compared to pre-treatment values, [Supplementary Table 2](#) and [Figure 3](#)). Comparison of post-DAA biomarker levels in PWID/HCV+ with PWID/HCV- participants revealed similar levels of CXCL10 and VCAM at EOT and 48 wk timepoints, implying a complete reversal of HCV-associated effects by DAA therapy ([Table 4](#), [Figure 2](#)). sTNF-R11 levels were reduced in PWID/HCV+ participants at the EOT but not 48wk post-DAA timepoints ($p = 0.002$ and 0.453 , respectively, compared with pre-treatment; [Supplementary Table 2](#)), and 48wk timepoint levels remained significantly elevated as compared to the PWID/HCV- participants ($p = 0.014$, [Table 4](#)). In contrast, elevated levels of the endotoxin-binding factor LBP in PWID/HCV+ participants were not significantly altered by DAA therapy ($p \geq 0.85$, [Supplementary Table 2](#)) and remained higher than PWID/HCV- individuals at both post-treatment timepoints ($p = 0.006$ and 0.030 at EOT and 48 weeks respectively, [Table 4](#)).

Regarding myeloid cell activation markers, DAA therapy was associated with a decrease in sCD163 ($p < 0.001$ for EOT and 48wks, [Supplementary Table 2](#)) despite levels not being significantly different to PWID/HCV- at baseline, whilst sCD14 was

significantly elevated in PWID/HCV+ at EOT ($p = 0.039$ compared to pre-treatment, [Supplementary Table 2](#)) and was significantly higher than PWID/HCV- ($p = 0.040$, [Table 4](#)). Other immune biomarkers measured including hsCRP, IL-6, HbA1c and d-dimer which were not different between PWID/HCV- and PWID/HCV+ prior to treatment were also not altered following DAA therapy. These analyses were adjusted for daily injecting drug use, which was a missing parameter in some PWID participants ([Table 2](#)). Thus modelling analyses were repeated without adjustment for this parameter to allow all participants to be included, which yielded very similar results (not shown). Taken together, these analyses suggest that DAA-mediated HCV cure reverses some elements of immune and endothelial activation induced by HCV viremia, but certain biomarkers of inflammation (i.e. sTNF-R11) and LPS signalling/regulation (i.e. LBP) are not fully reversed whilst others (i.e. sCD14) are actually increased by DAA therapy and remain elevated in PWID/HCV+ despite DAA-mediated HCV elimination.

Discussion

Curative DAA therapy for HCV prevents substantial morbidity and mortality, but it is unclear whether DAA therapy leads to a complete reversal of HCV-related immune alterations and thus related pathologies. We identified elevated levels of numerous soluble biomarkers associated with chronic disease risk in PWID, compared to those with no history of injecting drug use, including inflammatory and monocyte activation biomarkers. PWID with chronic HCV exhibited elevated levels of additional parameters including biomarkers of endothelial activation and only some of these parameters were normalised by curative DAA therapy. These



data suggest PWID may be at increased risk and thus should be closely monitored for the development of inflammatory morbidities. Furthermore, PWID with a history of HCV infection may possess additional and unique immunological alterations which persist after HCV cure and may potentiate the development of chronic diseases.

The specific biomarkers identified to be elevated in PWID with no history of HCV included the inflammatory biomarkers sTNF-RII and IL-6 and markers of myeloid cell activation (sCD14 and sCD163). These findings are consistent with previous observations in people who use drugs (37–40) and suggest a persistent state of inflammation and myeloid cell activation. Elevated levels of these biomarkers are associated with increased risk and severity of cardiovascular disease and mortality in both the general population and in individuals with underlying chronic infections such as HIV (21–23, 41, 42), however their relationship with chronic disease risk and mortality in PWID with and without HCV is not known and warrants further investigation.

Elevated levels of these inflammatory markers in PWID may be driven by a number of behavioural and sociodemographic factors including, but not limited to, diet, tobacco and alcohol use (32% of PWID in the present study reported using alcohol ≥ 4 times/week). Tobacco use and moderate to heavy alcohol consumption are associated with heightened inflammation (28) and these risks are prevalent in PWID populations (43). Similarly, there is a well-established link between diet and systemic inflammation (29, 44)

and substance use is known to affect diet and compromise nutrition, which is potentiated by factors including unstable housing (45). There are also mechanisms directly related to injecting drug use itself such as inflammatory and immunomodulatory effects of the drugs injected including methamphetamine, opioids and accompanying filler agents (46–50). Injury and infection related to injection site and injection practices can result from contaminated injecting equipment and/or the introduction of pathogens, foreign bodies and skin contaminants during the process of injecting (25, 27, 51). Over 60% of PWID report a history of skin and soft tissue infections with subcutaneous/intradermal injecting practices and vascular damage associated with injecting contributing to this risk (26). Injection frequency and duration correlate positively with markers of immune activation including d-dimer and sCD14 (39), whilst methamphetamine users with HIV who injected the drug exhibited elevated levels of numerous inflammatory biomarkers (including IL-6, sTNF-RI and sCD163) compared to those who did not inject (52). Together, these findings support a direct contribution of drug injection to systemic inflammation in PWID irrespective of HCV status, and more research is needed to characterise these risks and determine their long-term health implications.

PWID in the present study also exhibited substantially elevated levels of HbA1c, which is a well validated indicator of type 2 diabetes risk, diabetic complications and cardiovascular disease

(53). These metabolic alterations are likely associated with heightened inflammation but may also be influenced by lifestyle-related differences such as diet which is often altered in PWID (45). These data are consistent with the well documented association of injecting drug use with poor health outcomes and suggest inflammatory and cardiometabolic alterations may predispose them to increased risk of chronic inflammatory diseases such as cardiovascular disease and diabetes. Future studies are warranted to better characterise diabetes prevalence in PWID and identify contributing risk factors.

Analysis of the impact of active HCV infection in PWID revealed a further elevation of sTNF-RII levels as compared to never-infected PWID, as well as significant increases in plasma levels of CXCL10, VCAM and LBP in PWID/HCV+, consistent with previous findings (8, 9, 12, 17, 54). We did not detect an increase in inflammatory factors previously associated with chronic HCV such as sCD163, sCD14 or IL-6 in our cohort of PWID with HCV, potentially due to the pre-existing elevation of these parameters observed in all PWID. However, we did observe a significant reduction in sCD163 levels following DAA initiation, implying viremic HCV was contributing to heightened sCD163 levels and by association heightened macrophage activation prior to DAA initiation.

DAA therapy was associated with normalisation of certain activation markers (e.g., CXCL10, VCAM) to levels comparable to uninfected PWID, suggesting these parameters are closely linked with HCV viremia and are thus largely reversed following HCV cure. Although DAA therapy initially reduced sTNF-RII, levels at 48 weeks post treatment were higher in PWID treated for HCV as compared to never-infected PWID. In contrast, LBP levels were not significantly reduced by DAA therapy and remained elevated as compared to HCV-uninfected PWID up to 48 weeks post HCV cure. LBP is primarily produced by hepatocytes and is involved in the inflammatory response to microbial products such as bacterial LPS. Elevated LBP observed here in PWID with current or past HCV infection may indicate impaired hepatocyte/liver function due to persistent tissue damage or may be due to enduring damage to the gut or other mucosal barriers, resulting in heightened translocation of bacterial products. The latter is consistent with elevated plasma LPS levels in untreated HCV (55) and is analogous to the compromised gut barrier integrity and heighten microbial translocation observed in chronic treated HIV infection, which is known to contribute to persistent immune activation and inflammatory comorbidities (56). Elevated LBP and LPS levels are associated with dietary glucose and diabetes (57), so the observed elevation in HbA1c and LBP in the present cohort may be indicative of metabolic alterations. Little is known regarding the clinical implications of elevated LBP levels in people with HCV, however in HIV/HCV co-infection it is associated with more advanced liver fibrosis (58) and in HIV, elevated LPS and LBP are linked with increased risk of cardiovascular disease and dementia (59, 60), supporting the clinical relevance of these findings. Follow-up studies are required to identify the relationship between persistently elevated levels of inflammatory factors identified here and

clinical outcomes in individuals after HCV cure, especially in PWID.

Interestingly, we also observed a significant increase in sCD14 levels in PWID with HCV after 12 weeks of DAA therapy consistent with previous finding (13), although this was normalised after 48 weeks. This may be related to transient immune reconstitution induced by viral suppression and resultant increase in myeloid cell activity, however this remains to be explored. Consistent with the well-established relationship between inflammation and impaired liver function, all biomarker analysed here (except HbA1c) were significantly associated by multivariable regression analysis with indices of liver function including AST, ALT and the AST to platelet ratio index (APRI) score (data not shown), the latter being a surrogate marker of liver disease in HCV (61). People treated for HCV with DAA therapy are at heightened risk of mortality related not only to liver disease but also extrahepatic cancers and circulatory diseases, and the presence of comorbidities further increases mortality risk (7). The findings presented here of a persistent inflammatory state in individuals post HCV cure may therefore be relevant to these long term health outcomes and it would be of interest to determine whether these biomarkers were associated with morbidity and mortality post HCV cure.

The limitations of this study include the modest sample size due to the selection criteria and sample availability at the required timepoints and a lack of data pertaining to lifestyle factors such as body mass index, level of physical activity, as well as medical history and current medications. A longer duration of HCV viremia may heighten the immune changes measured here due to more extensive immune activation/exhaustion and a greater accumulation of cellular and tissue damage; this parameter could not be controlled for since time of infection is difficult to determine. Injection practices were not assessed in this analysis but would be useful to include in future studies. Finally, we had limited demographic data available in our never infected community control participants which precluded formal investigation of behavioural factors which may contribute to the observed biomarker differences. Given their influence on the parameters measured here, future studies are warranted to specifically assess the contribution of behavioural factors to chronic inflammatory disease risk.

Conclusions

These data add to a growing body of evidence that curative DAA therapy may not completely reverse immunological changes included by HCV infection, and biomarkers relevant to the development of chronic morbidities may remain elevated in some individuals. This study also highlights the particular vulnerability of PWID with HCV to these effects given the overlapping impact of injecting drug use and related lifestyle factors on immunological changes relevant to the development of chronic inflammatory diseases. These findings suggest ongoing clinical management of PWID with a history of HCV infection should include heightened vigilance for chronic inflammatory diseases even after curative HCV treatment.

Data availability statement

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

Ethics statement

Written informed consent to participate in this study was obtained from all participants. This work was approved by the Alfred Hospital Research and Ethics Committee (Melbourne, Australia).

Author contributions

AH: Conceptualization, Formal analysis, Funding acquisition, Investigation, Resources, Writing – original draft. NV: Formal analysis, Writing – original draft. SE: Investigation, Writing – review & editing. BH: Data curation, Project administration, Validation, Writing – review & editing. IB: Resources, Writing – review & editing. LN: Data curation, Project administration, Writing – review & editing. DP: Formal analysis, Writing – review & editing. PD: Resources, Writing – review & editing. AD: Resources, Writing – review & editing. AT: Resources, Writing – review & editing. AJ: Conceptualization, Supervision, Writing – review & editing. MH: Resources, Writing – review & editing. RS-D: Data curation, Formal analysis, Supervision, Writing – original draft. JD: Conceptualization, Funding acquisition, Resources, Supervision, Writing – original draft.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This

References

1. World Health Organisation. Updated recommendations on treatment of adolescents and children with chronic HCV infection: policy brief. Geneva: World Health Organization (2022). Available at: <https://iris.who.int/bitstream/handle/10665/357085/9789240052710-eng.pdf?sequence=1>.
2. Polaris Observatory HCV Collaborators. Global change in hepatitis C virus prevalence and cascade of care between 2015 and 2020: a modelling study. *Lancet Gastroenterol Hepatol* (2022) 7(5):396–415. doi: 10.1016/S2468-1253(21)00472-6
3. Manns MP, Maasoumy B. Breakthroughs in hepatitis C research: from discovery to cure. *Nat Rev Gastroenterol Hepatol* (2022) 19(8):533–50. doi: 10.1038/s41575-022-00608-8
4. Negro F, Forton D, Craxi A, Sulkowski MS, Feld JJ, Manns MP. Extrahepatic morbidity and mortality of chronic hepatitis C. *Gastroenterology*. (2015) 149(6):1345–60. doi: 10.1053/j.gastro.2015.08.035
5. Mazzaro C, Quartuccio L, Adinolfi LE, Roccatello D, Pozzato G, Nevola R, et al. A review on extrahepatic manifestations of chronic hepatitis C virus infection and the impact of direct-acting antiviral therapy. *Viruses* (2021) 13(11):2249. doi: 10.3390/v13112249
6. Hearps AC, Martin GE, Rajasurir R, Crowe SM. Inflammatory co-morbidities in HIV+ individuals: learning lessons from healthy ageing. *Curr HIV/AIDS Rep* (2014) 11(1):20–34. doi: 10.1007/s11904-013-0190-8
7. Hamill V, Wong S, Benselin J, Krajen M, Hayes PC, Mutimer D, et al. Mortality rates among patients successfully treated for hepatitis C in the era of interferon-free

research as funded in part by an internal grant received from the Burnet Institute, Melbourne Australia.

Acknowledgments

We would like to thank the study participants for their generous commitment to this study. We gratefully acknowledge the contribution to this work of the Victorian Operational Infrastructure Support Program received by the Burnet Institute.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fimmu.2024.1352440/full#supplementary-material>

- antivirals: population based cohort study. *BMJ*. (2023) 382:e074001. doi: 10.1136/bmj-2022-074001
8. Mascia C, Vita S, Zuccalà P, Marocco R, Tieghi T, Savinelli S, et al. Changes in inflammatory biomarkers in HCV-infected patients undergoing direct acting antiviral-containing regimens with or without interferon. *PLoS One* (2017) 12(6):e0179400. doi: 10.1371/journal.pone.0179400
9. Saraiva GN, do Rosário NF, Medeiros T, Leite PEC, Lacerda GS, de Andrade TG, et al. Restoring inflammatory mediator balance after sofosbuvir-induced viral clearance in patients with chronic hepatitis C. *Mediators Inflamm* (2018) 2018:8578051. doi: 10.1155/2018/8578051
10. Zeremski M, Petrovic LM, Talal AH. The role of chemokines as inflammatory mediators in chronic hepatitis C virus infection. *J Viral Hepat* (2007) 14(10):675–87. doi: 10.1111/j.1365-2893.2006.00838.x
11. Shin EC, Sung PS, Park SH. Immune responses and immunopathology in acute and chronic viral hepatitis. *Nat Rev Immunol* (2016) 16(8):509–23. doi: 10.1038/nri.2016.69
12. Babiker A, Hassan M, Muhammed S, Taylor G, Poonia B, Shah A, et al. Inflammatory and cardiovascular diseases biomarkers in chronic hepatitis C virus infection: A review. *Clin Cardiol* (2020) 43(3):222–34. doi: 10.1002/clc.23299
13. Kostadinova L, Shive CL, Zebrowski E, Fuller B, Rife K, Hirsch A, et al. Soluble markers of immune activation differentially normalize and selectively associate with

improvement in AST, ALT, albumin, and transient elastography during IFN-free HCV therapy. *Pathog Immun* (2018) 3(1):149–63. doi: 10.20411/pai.v3i1.242

14. Anthony DD, Sulkowski MS, Smeaton LM, Damjanovska S, Shive CL, Kowal CM, et al. Hepatitis C virus (HCV) direct-acting antiviral therapy in persons with human immunodeficiency virus-HCV genotype 1 coinfection resulting in high rate of sustained virologic response and variable in normalization of soluble markers of immune activation. *J Infect Dis* (2020) 222(8):1334–44. doi: 10.1093/infdis/jiaa254

15. Villani R, Facciorusso A, Bellanti F, Tamborra R, Piscazzi A, Landriscina M, et al. DAAs Rapidly Reduce Inflammation but Increase Serum VEGF Level: A Rationale for Tumor Risk during Anti-HCV Treatment. *PLoS One* (2016) 11(12):e0167934. doi: 10.1371/journal.pone.0167934

16. Radmanić L, Bodulić K, Šimić P, Vince A, Lepej S. The effect of treatment-induced viral eradication on cytokine and growth factor expression in chronic hepatitis C. *Viruses* (2022) 14(8):1613. doi: 10.3390/v14081613

17. Rosário NF, Saraiva GDN, Medeiros T, Gandini M, Lacerda G, Leite PEC, et al. Successful DAA therapy for chronic hepatitis C reduces HLA-DR on monocytes and circulating immune mediators: A long-term follow-up study. *Immunol Lett* (2020) 228:15–23. doi: 10.1016/j.imlet.2020.09.002

18. Gorin JB, Malone DFG, Strunz B, Carlsson T, Aleman S, Bjorkstrom NK, et al. Plasma FABP4 is associated with liver disease recovery during treatment-induced clearance of chronic HCV infection. *Sci Rep* (2020) 10(1):2081. doi: 10.1038/s41598-020-58768-z

19. Hengst J, Falk CS, Schlaphoff V, Deterding K, Manns MP, Cornberg M, et al. Direct-acting antiviral-induced hepatitis C virus clearance does not completely restore the altered cytokine and chemokine milieu in patients with chronic hepatitis C. *J Infect Dis* (2016) 214(12):1965–74. doi: 10.1093/infdis/jiw457

20. Koenig W, Löwel H, Baumert J, Meisinger C. C-reactive protein modulates risk prediction based on the Framingham Score: implications for future risk assessment: results from a large cohort study in southern Germany. *Circulation* (2004) 109(11):1349–53. doi: 10.1161/01.CIR.0000120707.98922.E3

21. Ridker PM, Rifai N, Stampfer MJ, Hennekens CH. Plasma concentration of interleukin-6 and the risk of future myocardial infarction among apparently healthy men. *Circulation* (2000) 101(15):1767–72. doi: 10.1161/01.CIR.101.15.1767

22. Schnabel RB, Yin X, Larson MG, Yamamoto JF, Fontes JD, Kathiresan S, et al. Multiple inflammatory biomarkers in relation to cardiovascular events and mortality in the community. *Arterioscler Thromb Vasc Biol* (2013) 33(7):1728–33. doi: 10.1161/ATVBAHA.112.301174

23. Vos AG, Idris NS, Barth RE, Klipstein-Grobusch K, Grobbee DE. Pro-inflammatory markers in relation to cardiovascular disease in HIV infection. A systematic review. *PLoS One* (2016) 11(1):e0147484. doi: 10.1371/journal.pone.0147484

24. Nelson PK, Mathers BM, Cowie B, Hagan H, Des Jarlais D, Horyniak D, et al. Global epidemiology of hepatitis B and hepatitis C in people who inject drugs: results of systematic reviews. *Lancet* (2011) 378(9791):571–83. doi: 10.1016/S0140-6736(11)61097-0

25. Smith ME, Robinowitz N, Chaulk P, Johnson KE. High rates of abscesses and chronic wounds in community-recruited injection drug users and associated risk factors. *J Addict Med* (2015) 9(2):87–93. doi: 10.1097/ADM.0000000000000993

26. Allaw F, Zakhour J, Kanj SS. Community-acquired skin and soft-tissue infections in people who inject drugs. *Curr Opin Infect Dis* (2023) 36(2):67–73. doi: 10.1097/QCO.0000000000000902

27. Dwyer R, Topp L, Maher L, Power R, Hellard M, Walsh N, et al. Prevalences and correlates of non-viral injecting-related injuries and diseases in a convenience sample of Australian injecting drug users. *Drug Alcohol Depend* (2009) 100(1–2):9–16. doi: 10.1016/j.drugalcdep.2008.08.016

28. O'Connor MF, Irwin MR. Links between behavioral factors and inflammation. *Clin Pharmacol Ther* (2010) 87(4):479–82. doi: 10.1038/clpt.2009.255

29. Smidowicz A, Regula J. Effect of nutritional status and dietary patterns on human serum C-reactive protein and interleukin-6 concentrations. *Adv Nutr* (2015) 6(6):738–47. doi: 10.3945/an.115.009415

30. Sacks-Davis R, Daraganova G, Aitken C, Higgs P, Tracy L, Bowden S, et al. Hepatitis C virus phylogenetic clustering is associated with the social-injecting network in a cohort of people who inject drugs. *PLoS One* (2012) 7(10):e47335. doi: 10.1371/journal.pone.0047335

31. Horyniak D, Higgs P, Jenkinson R, Degenhardt L, Stooze M, Kerr T, et al. Establishing the Melbourne Injecting Drug User Cohort Study (MIX): rationale, methods, and baseline and twelve-month follow-up results. *Harm Reduct J* (2013) 10:11. doi: 10.1186/1477-7517-10-11

32. Goutzamanis S, Spelman T, Harney B, Dietze P, Stooze M, Higgs P, et al. Patient-reported outcomes of the Treatment and Prevention Study: A real-world community-based trial of direct-acting antivirals for hepatitis C among people who inject drugs. *J Viral Hepat* (2021) 28(7):1068–77. doi: 10.1111/jvh.13516

33. Hearps AC, Martin GE, Angelovich TA, Cheng WJ, Maisa A, Landay AL, et al. Aging is associated with chronic innate immune activation and dysregulation of monocyte phenotype and function. *Aging Cell* (2012) 11(5):867–75. doi: 10.1111/j.1474-9726.2012.00851.x

34. Martin GE, Gouillou M, Hearps AC, Angelovich TA, Cheng AC, Lynch F, et al. Age-associated changes in monocyte and innate immune activation markers occur more rapidly in HIV infected women. *PLoS One* (2013) 8(1):e55279. doi: 10.1371/journal.pone.0055279

35. Walker MR, Li H, Teutsch S, Betz-Stablein B, Luciani F, Lloyd AR, et al. Incident hepatitis C virus genotype distribution and multiple infection in Australian prisons. *J Clin Microbiol* (2016) 54(7):1855–61. doi: 10.1128/JCM.00287-16

36. White PA, Zhai X, Carter I, Zhao Y, Rawlinson WD. Simplified hepatitis C virus genotyping by heteroduplex mobility analysis. *J Clin Microbiol* (2000) 38(2):477–82. doi: 10.1128/JCM.38.2.477-482.2000

37. Ownby RL, Kumar AM, Benny Fernandez J, Moleon-Borodowsky I, Gonzalez L, Eisdorfer S, et al. Tumor necrosis factor-alpha levels in HIV-1 seropositive injecting drug users. *J Neuroimmune Pharmacol* (2009) 4(3):350–8. doi: 10.1007/s11481-009-9150-x

38. Mehandru S, Deren S, Kang SY, Banfield A, Garg A, Garmon D, et al. Behavioural, mucosal and systemic immune parameters in HIV-infected and uninfected injection drug users. *J Addict Res Ther* (2015) 6(4):1–8. doi: 10.4172/2155-6105.1000257

39. Deren S, Cleland CM, Lee H, Mehandru S, Markowitz M. Brief report: the relationship between injection drug use risk behaviors and markers of immune activation. *J Acquir Immune Defic Syndr* (2017) 75(1):e8–e12. doi: 10.1097/QAI.0000000000001270

40. Markowski M, Deren S, Cleland C, La Mar M, Silva E, Batista P, et al. Chronic hepatitis C virus infection and the proinflammatory effects of injection drug use. *J Infect Dis* (2016) 214(9):1376–82. doi: 10.1093/infdis/jiw373

41. Reiner AP, Lange EM, Jenny NS, Chaves PH, Ellis J, Li J, et al. Soluble CD14: genome-wide association analysis and relationship to cardiovascular risk and mortality in older adults. *Arterioscler Thromb Vasc Biol* (2013) 33(1):158–64. doi: 10.1161/ATVBAHA.112.300421

42. Jaworowski A, Hearps AC, Angelovich TA, Hoy JF. How monocytes contribute to increased risk of atherosclerosis in virologically-suppressed HIV-positive individuals receiving combination antiretroviral therapy. *Front Immunol* (2019) 10:1378. doi: 10.3389/fimmu.2019.01378

43. Irvin R, Chander G, Falade-Nwulia O, Astemborski J, Starbird L, Kirk GD, et al. Overlapping epidemics of alcohol and illicit drug use among HCV-infected persons who inject drugs. *Addict Behav* (2019) 96:56–61. doi: 10.1016/j.addbeh.2019.04.023

44. Itsiopoulos C, Mayr HL, Thomas CJ. The anti-inflammatory effects of a Mediterranean diet: a review. *Curr Opin Clin Nutr Metab Care* (2022) 25(6):415–22. doi: 10.1097/MCO.0000000000000872

45. Mahboub N, Rizk R, Karavetian M, de Vries N. Nutritional status and eating habits of people who use drugs and/or are undergoing treatment for recovery: a narrative review. *Nutr Rev* (2021) 79(6):627–35. doi: 10.1093/nutrit/nuaa095

46. Chan YY, Yang SN, Lin JC, Chang JL, Lin JG, Lo WY. Inflammatory response in heroin addicts undergoing methadone maintenance treatment. *Psychiatry Res* (2015) 226(1):230–4. doi: 10.1016/j.psychres.2014.12.053

47. Azzoni L, Giron LB, Vadrevu S, Zhao L, Lalley-Chareczko L, Hiserodt E, et al. Methadone use is associated with increased levels of sCD14, immune activation, and inflammation during suppressed HIV infection. *J Leukoc Biol* (2022) 112(4):733–44. doi: 10.1002/JLB.4A1221-678RR

48. Papageorgiou M, Raza A, Fraser S, Nurgali K, Apostolopoulos V. Methamphetamine and its immune-modulating effects. *Maturitas* (2019) 121:13–21. doi: 10.1016/j.maturitas.2018.12.003

49. Al-Hashimi M, Scott SW, Thompson JP, Lambert DG. Opioids and immune modulation: more questions than answers. *Br J Anaesth* (2013) 111(1):80–8. doi: 10.1093/bja/aet153

50. Nakajima A, Yamada K, Nagai T, Uchiyama T, Miyamoto Y, Mamiya T, et al. Role of tumor necrosis factor-alpha in methamphetamine-induced drug dependence and neurotoxicity. *J Neurosci* (2004) 24(9):2212–25. doi: 10.1523/JNEUROSCI.4847-03.2004

51. Phillips KT, Stein MD. Risk practices associated with bacterial infections among injection drug users in Denver, Colorado. *Am J Drug Alcohol Abuse* (2010) 36(2):92–7. doi: 10.3109/00952991003592311

52. Miller M, Lee JY, Fulcher JA, Roach ME, Dilworth SE, Chahine A, et al. Getting to the point: Methamphetamine injection is associated with biomarkers relevant to HIV pathogenesis. *Drug Alcohol Depend* (2020) 213:108133. doi: 10.1016/j.drugalcdep.2020.108133

53. Sherwani SI, Khan HA, Ekhzaimy A, Masood A, Sakharkar MK. Significance of hbA1c test in diagnosis and prognosis of diabetic patients. *biomark Insights* (2016) 11:95–104. doi: 10.4137/BMLS38440

54. Nien HC, Hsu SJ, Su TH, Yang PJ, Sheu JC, Wang JT, et al. High serum lipopolysaccharide-binding protein level in chronic hepatitis C viral infection is reduced by anti-viral treatments. *PLoS One* (2017) 12(1):e0170028. doi: 10.1371/journal.pone.0170028

55. Hu S, Ghabril M, Amet T, Hu N, Byrd D, Yang K, et al. HIV-1 coinfection profoundly alters intrahepatic chemokine but not inflammatory cytokine profiles in HCV-infected subjects. *PLoS One* (2014) 9(2):e86964. doi: 10.1371/journal.pone.0086964

56. Marquez M, Fernandez Gutierrez del Alamo C, Giron-Gonzalez JA. Gut epithelial barrier dysfunction in human immunodeficiency virus-hepatitis C virus coinfecting patients: Influence on innate and acquired immunity. *World J Gastroenterol* (2016) 22(4):1433–48. doi: 10.3748/wjg.v22.i4.1433

57. Gomes JMG, Costa JA, Alfnas RCG. Metabolic endotoxemia and diabetes mellitus: A systematic review. *Metabolism*. (2017) 68:133–44. doi: 10.1016/j.metabol.2016.12.009
58. He J, Shi R, Duan S, Ye R, Yang Y, Wang J, et al. Microbial translocation is associated with advanced liver fibrosis among people with HIV. *HIV Med* (2022) 23(9):947–58. doi: 10.1111/hiv.13279
59. Ancuta P, Kamat A, Kunstman KJ, Kim EY, Autissier P, Wurcel A, et al. Microbial translocation is associated with increased monocyte activation and dementia in AIDS patients. *PLoS One* (2008) 3(6):e2516. doi: 10.1371/journal.pone.0002516
60. Trevillyan JM, Moser C, Currier JS, Sallam T. Immune biomarkers in the prediction of future myocardial infarctions in people with human immunodeficiency virus. *Clin Infect Dis* (2020) 70(8):1764–7. doi: 10.1093/cid/ciz765
61. Lin ZH, Xin YN, Dong QJ, Wang Q, Jiang XJ, Zhan SH, et al. Performance of the aspartate aminotransferase-to-platelet ratio index for the staging of hepatitis C-related fibrosis: an updated meta-analysis. *Hepatology*. (2011) 53(3):726–36. doi: 10.1002/hep.24105