Check for updates

OPEN ACCESS

EDITED BY Efe Sezgin, Izmir Institute of Technology, Türkiye

REVIEWED BY

Ebru Menekse, Ankara City Hospital, Türkiye Marko Samardžija, University of Zagreb, Croatia Marwa H. Niazy, Cairo University, Egypt

*CORRESPONDENCE Jing Luo 1318497642@qq.com Jiqiao Yang jqyangscu@163.com

RECEIVED 17 September 2023 ACCEPTED 22 February 2024 PUBLISHED 11 March 2024

CITATION

Wang X, He X, Liu J, Zhang H, Wan H, Luo J and Yang J (2024) Immune pathogenesis of idiopathic granulomatous mastitis: from etiology toward therapeutic approaches. *Front. Immunol.* 15:1295759. doi: 10.3389/fimmu.2024.1295759

COPYRIGHT

© 2024 Wang, He, Liu, Zhang, Wan, Luo and Yang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Immune pathogenesis of idiopathic granulomatous mastitis: from etiology toward therapeutic approaches

Xiaoli Wang¹, Xiujing He^{2,3}, Junzhi Liu⁴, Haiyan Zhang⁵, Hangyu Wan⁵, Jing Luo⁵* and Jiqiao Yang^{1,2}*

¹Breast Center, Department of General Surgery, West China Hospital, Sichuan University, Chengdu, China, ²Clinical Research Center for Breast, State Key Laboratory of Biotherapy, West China Hospital, Sichuan University, Chengdu, China, ³Laboratory of Tumor Targeted and Immune Therapy, State Key Laboratory of Biotherapy, West China Hospital, Sichuan University, Chengdu, China, ⁴West China School of Medicine/West China Hospital, Sichuan University, Chengdu, China, ⁵Department of Breast Surgery, Sichuan Provincial Maternity and Child Health Care Hospital, Chengdu, China

Idiopathic granulomatous mastitis (IGM) is a noncancerous, chronic inflammatory disorder of breast with unknown causes, posing significant challenges to the quality of life due to its high refractoriness and local aggressiveness. The typical symptoms of this disease involve skin redness, a firm and tender breast mass and mastalgia; others may include swelling, fistula, abscess (often without fever), nipple retraction, and peau d'orange appearance. IGM often mimics breast abscesses or malignancies, particularly inflammatory breast cancer, and is characterized by absent standardized treatment options, inconsistent patient response and unknown mechanism. Definite diagnosis of this disease relies on core needle biopsy and histopathological examination. The prevailing etiological theory suggests that IGM is an autoimmune disease, as some patients respond well to steroid treatment. Additionally, the presence of concurrent erythema nodosum or other autoimmune conditions supports the autoimmune nature of the disease. Based on current knowledge, this review aims to elucidate the autoimmune-favored features of IGM and explore its potential etiologies. Furthermore, we discuss the immune-mediated pathogenesis of IGM using existing research and propose immunotherapeutic strategies for managing this condition.

KEYWORDS

idiopathic granulomatous mastitis, autoimmune, autoimmunity, etiology, pathogenesis

Introduction

Idiopathic granulomatous mastitis (IGM), first described in 1972 (1), is a nonspecific, refractory inflammatory breast disease mostly affecting young parous women (2, 3). It is

characterized by the sudden onset of breast lumps, abscesses, sinus tracts, and ulcers (2). IGM was once considered an orphan disease; however, in recent years, the incidence has been calculated to compromise 1.8% of benign breast diseases (4) and 24% of inflammatory breast diseases (5). Interestingly, the prevalence of IGM varies regionally or racially, with higher rates observed in Middle Eastern, Mediterranean, Asian, and Hispanic populations compared to Western Caucasian populations (6–9).

IGM is not cancerous, but can be highly recalcitrant and locally aggressive, significantly impacting patients' quality of life and potentially leaving permanent cosmetic deformities (10). As a mysterious entity to date, IGM is still difficult to diagnose and treat. The etiology remains largely unclear and clinicians have an inconsistent understanding of the disease, resulting in a lack of a gold standard for IGM management. While a portion of patients respond to corticosteroids, antibiotics, doxycycline, and methotrexate (2), relapse rates have been reported as high as 5-78% (5, 11). Surgery is not a common treatment, but some patients underwent multiple surgeries and, in extreme cases, mastectomies due to refractory pain, abscesses, fistulas, sinuses, and healing complications (12–15).

Most commonly, IGM is characterized by a firm, tender breast mass, skin redness, and mastalgia (3). Other symptoms include swelling, fistula, abscess (often without fever (16)), nipple retraction, and peau d'orange appearance (17). The extramammary manifestations includes erythema nodosum and regional lymphadenopathy, present in a quarter of patients (18). Ultrasound, as an imaging modality, is commonly used for diagnosis of IGM, indicating irregular hypoechoic masses, multiple hypoechoic masses, parenchymal heterogeneity, or heterogeneous echogenic areas on ultrasonographic examination (19). The common mammographic features are focal or diffuse asymmetric density (20), while magnetic resonance imaging is not routinely utilized in the evaluation of IGM (9).

Clinically and radiologically, IGM often mimics breast abscess or malignancy, especially inflammatory breast cancer (14), therefore definitive diagnosis relies on core needle biopsy and histopathologic examination. IGM is histologically characterized by lobular-centered epithelioid noncaseating granulomas, with occasional sterile microabscesses. Epithelioid histiocytes are predominantly neutrophils and eosinophils. Langhans cells, lymphocytes, and plasma cells generated by the eosinophilic cellsdominated inflammatory reactions are also observed in the lobular center of the breast parenchyma. Depending on the disease severity, the inflammatory responses may extend and obliterate the lobularcentric pattern. It should be noted that the diagnosis of IGM is not clear until inflammation caused by other specific causes has been excluded, including inflammatory breast malignancies, tuberculosis, infective mastitis caused by known pathogens, sarcoidosis, Wegener's granulomatosis, granulomatosis with polyangiitis, diabetic mastopathy, foreign body reaction, and IgG4-related sclerosing mastitis (3, 10, 21).

Currently, the most widely supported etiological theory suggests that IGM is an autoimmune disease (22). The fact that some patients responded well to steroid treatment and the observation of concomitant erythema nodosum (23) or other autoimmune conditions further supported the autoimmune nature of the disease. Nonetheless, previous studies have failed to find an association between antinuclear antibody (ANA) levels and the risk, medical response, or recurrence of IGM (24). Benson and colleagues proposed a tentative pathogenic pathway for IGM: 1) the ductal epithelial lining damage that is possibly secondary to the retention of ductal secretions; 2) efflux of ductal contents from the lumen into the surrounding lobular connective tissue; 3) development of local inflammation in response to extravasated ductal contents; 4) migration of lymphocytes and macrophages into periductal zones; and 5) local granulomatous response with the formation of noncaseating granulomas (Figure 1) (25). Other possible etiologies include hyperprolactinemia, hormonal imbalance, microbiological agents, and genetic factors (13, 26). In this manuscript, we provide critical insights into the current understanding involving the pathogenetic features of IGM, the complex natures and potential therapeutic strategies of a condition that is confounding the medical community.

Autoimmune-favored features of IGM

Erythema nodosum and arthritis

Erythema nodosum (EN), first described in 1987, is a rare but typical systemic manifestation of IGM (27). EN is blood vesselassociated hypodermal septal inflammatory panniculitis characterized by reddish, tender nodules, typically of bilateral subcutaneous pretibial regions and occasionally seen on arms (28). The reported frequency of EN among IGM patients ranges from 6.6% to 58.1% (28-31). The results of a retrospective, comparative parallelarm study involving 43 patients with IGM coexisting with EN and 43 with IGM only suggested that the coexistence with EN presented higher fistula distribution and recurrence rates, indicating a worse prognosis for IGM patients (28, 32). In addition, IGM patients with EN often have more extensive lesions or bilateral breasts involved, suggesting a more severe disease (29). Inflammatory arthritis/ arthralgia is another extramammary manifestation of IGM with a lower incidence than EN (33). In clinical observations, regression of arthritis in IGM patients is usually synchronized with the remission of local (mammary) symptoms (34). Despite a low occurrence, the simultaneous involvement of EN and arthritis in IGM patients has been reported previously (35). Therefore, EN and arthritis might have autoimmune mechanisms similar to those of IGM in disease development.

EN can be triggered by various antigenic stimuli (34) and is associated with a range of pathological conditions, such as drug reactions, infection, tuberculosis, and malignancies (36). Similar to IGM, EN also has an unelucidated pathogenesis. It has been hypothesized that increased immune complexes, cytokines, and chemokine levels in the blood might lead to EN in severe IGM cases by triggering septal panniculitis induced by neutrophilic recruitment and activation in pretibial subcutaneous tissue (29). It is widely believed that systematic involvement, such as EN and arthritis, supports an autoimmune component in etiology (34, 37).

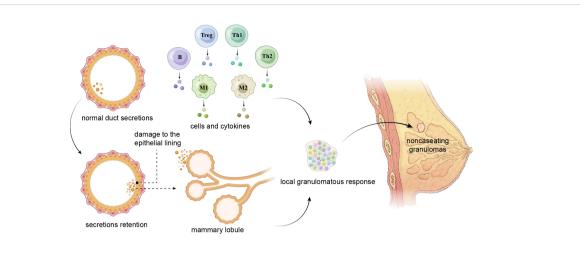


FIGURE 1

A tentative pathogenic pathway for IGM proposed by Benson and colleagues. The damage of the epithelial lining caused by retention of ductal secretions leads to the leakage of content from the lumen into the surrounding lobular connective tissue, and the extravasation initiates local inflammatory reactions in which lymphocytes and macrophages are involved. These cells migrate to the periductal area, produce cytokines, and trigger a local granulomatous response that pathologically presents with noncaseating granulomas. Th, helper T cell; Treg, regulatory T cell; M, macrophage; B, B lymphocyte.

Coexistence with other autoimmune diseases

Apart from systematic symptoms, IGM has also been reported to coexist with autoimmune diseases, such as systemic lupus erythematosus (SLE) (38) and Sjögren's syndrome (39, 40), simultaneously within an individual. This simultaneous presence of two or more autoimmune diseases in a patient is defined as polyautoimmunity and is considered a common phenomenon in rheumatic and autoimmune diseases (41). As reported, 33% of patients suffering from SLE and 1/3 of patients with Sjögren's syndrome have developed another autoimmune disease (42, 43). The polyautoimmunity among IGM patients indicates the importance of shared etiopathogenesis with other autoimmune diseases.

Seasonal fluctuation of disease onset

Although there is no solid statistical evidence, seasonal fluctuations are noticed in the onset of IGM (32, 44). In a retrospective study of 44 patients treated for IGM, an increased incidence of IGM was observed during the warm seasons from 1994 to 2018 (44). A similar pattern was noticed in a single-institutional retrospective analysis of 134 IGM patients, with a slightly higher incidence observed in summer and late spring (32). However, despite the observation of the above trends in the distribution of onset times, the above two studies were unable to draw statistical conclusions. The small sample size may account for this, especially when divided into twelve months.

Seasonality has been well recognized in autoimmune disorders (45). For example, it has been suggested that the onset of autoimmune encephalitis was dominant in summer-autumn (46). Moreover, rheumatoid arthritis (RA) activity in the upper and lower extremities peaks in spring (47). The loss of tolerance to self-

antigens contributes to the development of autoimmune diseases (48). Environmental factors with seasonal fluctuation patterns, such as temperature and humidity, vitamin D levels that varied with ultraviolet radiation exposure, and melatonin secretion were indicated to influence disease onset and progression (45). Vitamin D levels are associated with disease activity in multiple sclerosis (MS), SLE, psoriasis, and RA. Melatonin exerts anti-inflammatory properties by affecting different arms of the immune system including but not limited to the inhibition of inflammatory cytokines (49). Melatonin levels, which reach their minimum in spring, may aggravate MS and SLE (45). The incidence increases in spring and early summer in children with primary immune thrombocytopenia (ITP) (50). A similar pattern has been observed in acute adult ITP, which may be attributed to seasonal variations in certain areas of infection that lead to proinflammatory cytokine release and human antigen mimicry (32). In addition, seasonal variations have also been reported in the production of hormones (e.g. growth hormone and insulin-like growth factor) and self-reactive autoantibodies (e.g. anti-Jo-1 autoantibodies, anti-SRP autoantibodies, anti-proteinase 3 antibodies, and autoantibodies to islet cell antigens or pancreatic tyrosine phosphatase) (51). In IGM patients, although the current evidence of seasonal fluctuations and hidden mechanisms is far from solid, the trend observed adds to the suspicion of its nature as an autoimmune disease.

Triggering events and precipitating factors of IGM

Pregnancy and parturition

IGM is typically found in childbearing-age women with a history of pregnancy and/or lactation within 5 years (52–55). In a

multicenter retrospective analysis including 22 breast centers with a total of 720 IGM patients, the overall incidence rate of the first recurrence was 17% (122/720), and the re-recurrence after treatment was 3% (22/720), with a statistically significant association between recurrence and the history of pregnancy (p=0.0008) and of breastfeeding (p=0.03), respectively (5). Therefore, the history of pregnancy and/or lactation (rather than the moment of pregnancy and breastfeeding) is seen as a risk factor for IGM recurrence.

The relevance between history of pregnancy and/or lactation and autoimmune diseases has been discussed from time to time. Both local and systemic factors accounted for the pathogenic mechanisms of IGM. Benson and his colleagues proposed a tentative pathogenic pathway for IGM centered on the mammary ducts. Possibly secondary to the retained ductal secretions, damage to the ductal epithelial lining induced the outflow of contents from the lumen into the surrounding lobular connective tissue which in turn developed a local inflammation in response to extravasated contents. Macrophages and lymphocytes involved in the above mentioned local inflammation migrate to the periductal zones and induce a local granulomatous response characterized by the formation of noncaseating granulomas (25). Additionally, damage to the ductal epithelium may occur as a result of milk stasis in the absence of breastfeeding (33). Systemic factors are prominent among the effects of pregnancy on many autoimmune diseases (25). RA often began several months after delivery, the disease activity flared in 46.7% of patients postpartum but interestingly improved in 60% during pregnancy, as reported in a meta-analysis (56). Another study also observed an increased risk of new-onset RA during the first 3 months postpartum, especially in primigravida for whom this risk increased to almost 11-fold (57). Up to 30% of young women with Graves' disease had a history of pregnancy in the 12 months prior to the onset (58). It is indicated that pregnancy is a significant risk factor for susceptible women and postpartum autoimmune diseases are not rare. There is a great chance that the onset and progression of autoimmune and/or inflammatory diseases are regulated by T cell cytokine-mediated responses not only during the gestational and the postpartum periods but also decades after pregnancy, with progesterone and estrogens inducing a T helper 2 (Th2)-dominated response which further prompts the worsening of Th2-type autoimmune diseases, and impressing the Th1/Th17-dominated response which facilitates the improvement of Th1/Th17-type autoimmune diseases (59). Therefore, the pathogenic mechanism and the exact correlation between IGM and a recent history of pregnancy and/or lactation need to be further illuminated.

Hormonal disorders

Hormonal imbalance has a notable impact on the pathogenesis of IGM (60–62). Diagnosed with hyperprolactinemia secondary to a prolactinoma, IGM patients in two case reports improved after treatment with prolactin inhibitors (63, 64). In nonpregnant IGM women with schizophrenia or bipolar disorder, hyperprolactinemia was secondary to risperidone therapy (65–67); however, in some cases, prolactin (PRL) levels may be normal (68).

The relationship between PRL and immune system has been elucidated to a great extent in the last decade, shedding new lights on the field of immunoendocrinology. Hyperprolactinemia has been described in the active phase of some non-organ-specific autoimmune diseases (e.g. SLE, RA) and organ-specific autoimmune diseases (e.g. celiac disease, type 1 diabetes mellitus, Addison's disease, and autoimmune thyroid disease) (69). A study recruiting 102 patients with hyperprolactinemia showed a possible autoimmune condition in which levels of interleukin-4 (IL-4) and thyroid peroxidase antibodies (TPOAb) are higher (70). PRL increased the Th1 lymphocytes-induced synthesis of interferon gamma (IFN- γ) and IL-2, and activated Th2 lymphocytes-induced production of autoantibody, therefore of worthy interest was the association between hyperprolactinemia and levels of cytokines and specific/nonspecific antibodies in autoimmune diseases (69). An animal experiment concluded that persistently elevated levels of serum prolactin interfered with B cell tolerance induction by impairing B cell receptor (BCR)-mediated clonal deletion, deregulating receptor editing, and lowering the threshold for anergic B cell activation, thereby promoting autoreactivity (71). Different from the sex hormones that experience a cliff-like drop at delivery and a slow recovery postpartum, prolactin increases slightly during pregnancy and rises exponentially at delivery and during lactation (72). Future studies may confirm the specific association between IGM and hormonal disorders and unravel the pathogenic mechanism.

Trauma and duct epithelial damage

Trauma and ductal perforation have been reported as triggering factors for IGM. The proposed mechanism of IGM involves the following steps: ductal epithelial damage, leakage of luminal secretions into surrounding lobular connective tissue, local inflammation, migration of macrophages and lymphocytes into the region, and local granulomatous inflammatory response (17). Secretion extravasation, as a result of ductal damage, causes migration of macrophages and leukocytes into the tissue, leading to local inflammation (33). In contrast, another tentative pathogenesis of IGM advocates that an insult to the ductal epithelial cells, perhaps by directing against antigens within lactations, triggers the formation of local granulomatous (36). Specifically, ductal perforation elicits an inflammatory response, and the subsequent immune presentation of mammary antigens induces autoimmunity, contributing to reactive T cell-mediated inflammation and centrilobular granuloma formation (9). There is a strong possibility that skin lesions may be a consequence of epitope spreading following an immunological event activated within the breast tissue (36). In addition to the lactation process, trauma may be responsible for ductal epithelial damage. Other trauma-related conditions suspected of causing epithelial damage have been reported, such as one case of underlying IGM was

exacerbated by ductoscopy. A 41-year-old woman in this case report went to the outpatient clinic with a 2-year history of rightsided pathologic nipple discharge, excluding remarkable medical history and drug history. She developed pain and a 4-5 cm palpable mass in the lower outer quadrant four days after ductoscopy. The patient initially responded well to antibiotics but ten days later presented with abscesses that were finally incised and drained by surgery. A few days after a week of slow recovery, a new red palpable mass developed in the upper outer quadrant of the breast, and the histopathology of an ultrasound-guided biopsy showed granulomatous mastitis (73). Furthermore, trauma and ductal perforation are emerging as risk factors for other systemic immune disorders. Twenty nine patients with traumatic brain injury completed a follow-up of 3 years and the result suggested significantly worse pituitary dysfunction in antipituitary antibody (APA)-positive patients than in APA-negative patients (46.2% vs 12.5%, P=0.04), with a corresponding OR of 2.25 (95% CI 1.1-4.6) (74). A detailed, responsible, and practical investigation of the trauma mechanism may be a high priority for further research in some patients with IGM.

Smoking

A recognition is led to that smoking is possibly one of the risk factors for IGM, but no definite conclusions have been drawn thus far. The reported proportions of IGM patients who smoke were 0%-77.8% (55, 75–78). A 11-year retrospective study of 77 patients with IGM found two groups (high flare *vs* low flare) were largely comparable except for smoking as a risk factor. Specifically, smokers added to 10 times greater odds of having a high IGM flare than non-smokers (P = 0.031) (12). A multicenter and retrospective analysis of 720 patients with IGM also found a significant association between smoking and IGM recurrence (p=0.01) (5). Therefore, investigators consider smoking as one of the risk factors for IGM recurrence.

The association between smoking and autoimmunity has long been proposed (79, 80). Epidemiologic studies have stated that tobacco smoking is associated with the development, disease course, symptom severity, and outcome of autoimmune diseases, such as RA, systemic sclerosis (SSc) and SLE (81). Smoking modulates immune system through multiple and complex mechanisms, including the immunosuppression, induction of apoptosis, induction of the inflammatory response, alteration of cytokine balance and the synthesis of anti-DNA antibody generated by DNA damage (81). It promotes autoimmune diseases by releasing intracellular proteins from reactive oxygen species (ROS)-activated or injured cells, altering antigen presentation in cigarette smokeimpaired cells, modifying regulatory B- and T-cell functions, dysregulating DNA demethylation and upregulating immune genes (12, 82-84). The existence of susceptible genes further expands the understanding about the role of smoking in autoimmunity (81). However, a clearer relationship between IGM and smoking-caused pathogenic mechanism still remains to be elucidated.

Microorganisms

IGM presents as an inflammatory mass in the peripheral tissues of the breast or simultaneous peripheral (rarely central) inflammation at multiple sites with abscesses and/or inflammation and ulceration of the skin (55, 85–87). Partly for this reason, infections or microorganisms are naturally considered as a potential risk factor for IGM. A fine needle aspiration culture of the abscess in a female patient with IGM revealed *Enterococcus* avium (88). Tissue culture from a repeat biopsy in a 28-year-old woman grew *Mycobacterium mucogenicum*, a rare cause of skin and soft tissue infections (89). As a possible infectious etiology, *Corynebacterium* species (90), especially *Corynebacterium kroppenstedtii* (91, 92), have been identified in the progression of IGM. Microorganisms detected by culture in previous studies are cluttered, random, and lack regularity, making it difficult to guide clinical practice.

Infections provide necessary antigens by mimicking or altering self-antigens, leading to an increase of overall immune reactivity level or an antigen "spillage" (93, 94). In addition, infections serve as inflammatory context that favors the activation of innate immune responses (95). Importantly, the Epstein–Barr virus has been implicated as an inciting agent for several autoimmune diseases, including MS and SLE (96, 97).

In the apparent absence of infection, microorganisms also influence the initiation and progression of an autoimmune responses. Populations of commensal microorganisms ("microbiota") profoundly affect the induction of diseases such as inflammatory bowel disease on humans (98). Changes in the composition of the microbiota are common in autoimmune diseases, giving rise to a condition termed dysbiosis (99, 100). For example, Enterococcus gallinarum, found in the liver of SLE patients with cross-reactivity, developed an autoimmune response to the Ro antigen (101).

Others

The primary function of Alpha-1-antitrypsin (AAT), a glycoprotein synthesized by hepatic cells, is to prevent the destructive effects of proteases secreted by activated neutrophils. Schelfout et al (102) demonstrated AAT deficiency in a 37-year-old female patient with IGM. According to current studies, we authors define no immunological events, but hypothesize that AAT deficiency could be one of the etiologic factors; therefore, further investigations are needed to support this argument.

Immune pathogenesis of IGM

Cellular dysregulation

Hypothesis that autoimmunity plays a role in IGM has long been considered, and the dominance of T lymphocytes found in some studies partially forms the basis for this theory (Figure 2). A study

10.3389/fimmu.2024.1295759

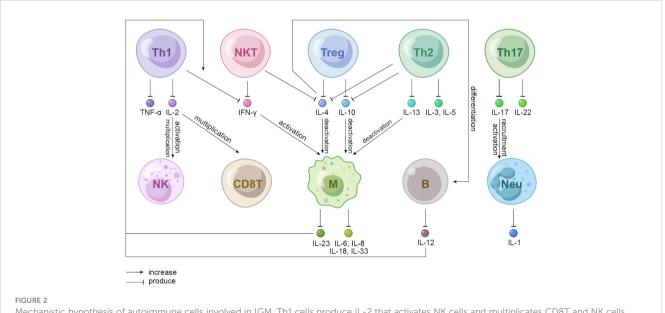
recruiting 51 pathologically proven IGM patients (26 in activity; 25 in remission) and 28 healthy volunteers as controls pointed that lymphocyte subsets in the peripheral blood determined their respective impacts on etiopathogenesis. To be more specific, the percentage of Th cells was lower in patients with IGM than controls regardless of disease severity, perhaps because the recruitment of Th cells to granulomatous breast tissue contributes to the low T cell counts in the peripheral blood of patients. However, the absolute count of cytotoxic T lymphocytes increased as Th cells decreased, presenting with higher percentage and greater absolute count both of natural killer (NK) cells and natural killer T (NKT) cells in IGM patients than controls (8). NKT cells suppressed granuloma formation by modulating the production of IFN-y and IL-10 according to research performed on mouse livers (103). That is, observed changes in T cells, NK cells and NKT cells imply systemic immune dysregulation in patients with IGM which is recognized as a localized form of granulomatous disorder. In a large cohort study of granulomatous mastitis in which microbiologic studies were negative, IGM showed significantly more plasma cells than tuberculous mastitis in terms of histologic features. Current understanding possibly accounts for the autoimmune aetiology, which would draw in humoral responses and thus more plasma cells (104). A recent study suggested that CD68- and CD163-positive M2-type macrophages were privileged in the course of plasma cell mastitis (PCM) and granulomatous lobular mastitis (GLM). CD 57-positive NK cells even provided a practical basis for clinical staging of GLM. The Neutrophil-to-lymphocyte ratio (NLR), one of the preoperative inflammatory parameters, was significantly associated with IGM recurrence, highlighting that a high NLR predicted poor patient outcomes (105, 106).

IL family/cytokine

Cytokines regulate the growth, differentiation, maturation, and response of cells (Table 1). In general, levels of IL-4, IL-5, and IL-13 increased in allergic diseases; levels of IL-17 and IL-23 increased in autoimmune diseases; and levels of IL2 and IFN- γ increased in sarcoidosis and granulomatous diseases (112). The levels of IL-33 (107), IL-6 (108), IL-8, IL-10, IL-17 (109), IL-22, and IL-23 (110) were all significantly higher in IGM patients than healthy controls. IL-6 level was even significantly higher in patients with severe disease than those with mild and moderate disease, indicating that IL-6 is a biomarker for assessing disease severity in IGM (108). However, a recent study showed that values of IL-1 β , IL-10, and IL-18 were significantly lower in the acute IGM group than the control group (111).

HLA

A study covering 48 patients with IGM and 50 healthy controls reported that the frequency of human leukocyte antigens (HLA)-A*10, HLA-A*2403, HLA-B*18 and HLA-DR*17 was significantly



Mechanistic hypothesis of autoimmune cells involved in IGM. Th1 cells produce IL-2 that activates NK cells and multiplicates CD8T and NK cells. IFN- γ produced by NKT and Th1 cells activates macrophages that produces a variety of interleukins among which IL-23 increases Th1-produced IFN- γ . IL-4 (produced by NKT, Treg, and Th2 cells), IL-10 (produced by Treg and Th2 cells) and IL-13 (produced by Th2 cells) all deactivate macrophages; IL-4 also induces the differentiation of B cells. Th17 cells produce IL-22 that promotes proliferation, remodeling, and repair of tissues and organs to maintain innate host defense mechanism and IL-17 that participates in the activation and recruitment of neutrophils. Th, helper T cell; Treg, regulatory T cell; NKT, natural killer T cell; NK, natural killer cell; CD8T, cluster differentiation 8 positive T cell, also known as CTL, cytotoxic T lymphocyte; M, macrophage; B, B lymphocyte; Neu, neutrophil.

TABLE 1 Reported cytokines associated with IGM.

IL family	Type of research	Sample size	Method	Results (statistically significant)	Meaning	Reason
IL-33 [(107)]	Case- control study	25 IGM patients 32 BC patients 30 HCs	Human IL-33 (Platinum ELISA kit, BMS2048/ BMS2048TEN, eBioscience, Vienna, Austria)	IGM>BC>HCs	Distinguishing diagnoses of IGM and BC (along with the radiological and pathological findings).	The induction of T- helper type 2 immune response
IL-6 [(108)]	Case- control study	11 IGM patients -mild (n = 1) -moderate (n = 4)- severe (n = 6) 7 HCs	ProcartaPlex magnetic bead- based multiplex assay (Cat. No. EPX180-12165-901; eBioscience, Vienna, Austria)	1. IGM >HCs 2. IGM: severe>moderate 3. positive correlation with time to resolution	Biomarkers for disease severity and time to resolution in IGM patients.	The stimulation of IL-1 and Toll-like receptors
IL-8 IL-10 IL-17 [(109)]	Case- control study	47 IGM patients -active (n=21) -remission (n=26) 30 HCs	Human enzyme-linked immunosorbent assay kits (Elabscience [®] , Houston, USA).	IL-8、IL-10、IL- 17: IGM>HCs IL-10: active IGM>HCs	Indicating the presence of serious immune dysregulation in IGM patients.	IL-8 mRNA is upregulated by the deficiency of NADPH oxidase in CGD
IL-22 IL-23 [(110)]	Case- control study	26 IGM patients 15 HCs	Elabscience Human IL-22 ELISA Kit, cat. no. E-EL- H0107; and Elabscience Human IL-23 ELISA Kit, cat. no. E-EL-H0106	IGM >HCs	Supporting the etiopathogenesis of IGM in favor of the autoinflammatory thesis.	Activated CD4+ cells initiate inflammation by releasing inflammatory cytokines
IL-1β TNF-α IL-10 IL-18 [(111)]	Case– control study	32 IGM patients -active (n=19) -remission (n=13) 18 HCs	Flow cytometry (FACSLyric TM ,BD, USA)	IGM <hcs active IGM<hcs< td=""><td>Cytokines, which play roles in known autoimmune and Granulomatous reactions, especially cytokines related to Th1 and Th17, might not play a role in the etiopathogenesis in GM patients.</td><td>-Inflammasome Th1 may not play roles in IGM etiopathogenesis。 -The cytokine measurement method was bead-based immunoassay on Flow Cytometry instead of ELISA.</td></hcs<></hcs 	Cytokines, which play roles in known autoimmune and Granulomatous reactions, especially cytokines related to Th1 and Th17, might not play a role in the etiopathogenesis in GM patients.	-Inflammasome Th1 may not play roles in IGM etiopathogenesis。 -The cytokine measurement method was bead-based immunoassay on Flow Cytometry instead of ELISA.

BC, breast cancer; HCs, healthy controls; ELISA, enzyme-linked immunosorbent assay; NADPH, nicotinamide adenine dinucleotide phosphate; CGD, chronic granulomatous disease.

higher in the patient group than the control group (p=0.012, p=0.012, p=0.001 and p=0.005, respectively), which may help explain the etiopathogenesis. However, further data based on large samples and geographically diverse regions are needed (113).

Others

The detected high level of soluble triggering receptor expressed on myeloid cells-1 (sTREM-1) is dedicated to IGM according to a case-control study. In particular, blockade of TREM may be a promising treatment option for resistant or multiple recurrent patients with IGM (114). In contrast to breast cancer, some preoperative parameters (C-reactive protein, albumin, fibrinogen, fibrinogen/albumin, NLR, etc.) have considerable potential to be early sensitive biomarkers of IGM, providing a useful guide for differential diagnosis of two diseases (115).

Immunotherapeutic approaches in IGM

The watch-and-wait approach

Often, no specific treatment is required for IGM, especially for patients with small (<5 cm), unilateral lesions without sinus formation or abscess. IGM is usually a self-limiting inflammatory disease and complete resolution may take 5 to 20 months (116, 117). Observation, known as the watch-and-wait procedure or expectant treatment, seems one of the most appropriate treatment approaches for patients who do no desire medical intervention and are concerned about surgical complications.

In a study of 120 women with IGM, 112 patients recovered spontaneously without surgical or medical treatment (117). A metaanalysis of 10 retrospective studies from five countries unmasked comparable recurrence rates between patients managed with

10.3389/fimmu.2024.1295759

conservative (observation and medicine) and surgical treatment approaches (OR 1.25, 95% CI 0.51 – 3.03) (118). However, a recent meta-analysis of 71 studies including 4735 patients demonstrated a significant difference in relapse rates with expectant management compared with other treatments (p = 0.023) (119). Clinically, treatment plans should be tailored according to symptom severity and patient preference.

Conventional immunosuppressive agents

Steroids

Steroids, also known as corticosteroids or glucocorticoids, are effective and commonly used conservative therapies for IGM patients. Steroids and methotrexate (MTX), with or without surgery, are regarded as the treatment for IGM patients (61). Painful unilateral small (< 5 cm) lesions with minimal discharge or ulceration can be treated with prednisone at a dose of 0.5 mg/kg per day, while those with multiple, bilateral lesions (\geq 5 cm) or significant skin ulcers, secretions/fistulas can be treated with prednisone at a dose of 0.5-1 mg/kg per day or 10-15 mg oral MTX weekly accompanied by daily folic acid supplementation (120, 121). A prospective cohort study of 49 women with biopsy-proven granulomatous mastitis was conducted from 2006 to 2010. Fifty-nine percent of patients initially receiving antibiotics reached incomplete resolution and 90% were prescribed oral steroids. Of those who received steroids, 80% had complete resolution with a median time of 159 days (interquartile range, IQR 120-241 days) (120). A meta-analysis grouped 106 patients into surgical management, oral steroids, and oral steroids plus surgical management groups and found that oral steroids may be a widely acceptable option for patients with concerns about surgical scarring (121).

High-dose prednisolone has a high success rate and a low recurrence rate with a reduction of the need for surgery in treatment of IGM (122). For localized disease, observation, steroids and excision can be applied (123). Under certain circumstances, pregnancy precluded the use of most drugs, but local corticosteroids was effective in terms of treatment response and duration, need for surgery, reduced recurrence and side effects (124). It is generally believed that noninvasive treatments should be initially performed due to the self-limiting nature of IGM. A greater number of valuable studies are needed to determine the appropriate dose of topical steroids and to estimate the efficacy of this therapy (125). The route of administration also influences the efficacy and safety of steroids. A prospective single-arm study reported a novel treatment for patients with nonlactating mastitis (NLM) (27 patients with IGM). After local anesthesia, an appropriate amount of irrigation solution was pumped into the ducts, followed by a daily breast massage lasting for two weeks. During this study, oral antibiotics and fine-needle aspiration for breast abscess were allowed; however, oral intake of corticosteroids and surgical drainage were not allowed. This study found that the 1-year clinical complete response (cCR) rate of NLM patients was more than 90% with insignificant adverse events (126, 127). Based on the available evidence, well-designed randomized controlled trials are encouraged to compare the efficacy and safety of ductal lavage with traditional strategies (surgery, steroids, observation alone, etc.).

Steroids are remarkably useful in acute phase of inflammatory diseases and autoimmune diseases by inhibiting a wide range of immune responses (128). Two decades ago, a basic experiment illustrated that the stability of messenger RNA (mRNA) encoding interleukins, tumor necrosis factor (TNF), and granulocytemacrophage colony-stimulating factor diminished for the presence of glucocorticoids (129). In the low to moderate dose range, glucocorticoids have variable effects on T lymphocyte subsets. Following glucocorticoid administration, total T cells were slightly reduced in the circulation, while immature CD4+ cells were more affected than mature CD4+ cells, Th17+ cells, and CD8+ cells. All above mentioned subsets transiently increased after hydrocortisone administration (130, 131). The percentage of circulating regulatory T cells increased in patients with lupus or sarcoidosis treated with intravenous methylprednisolone or prednisone (130-132). High dose of glucocorticoids causes a rapid depletion of most circulating T cells due to a combination of effects (e.g. enhanced circulatory emigration (133); inhibition of IL-2 signaling (134); induction of apoptosis) (135-137).

Immune modulators

Immunosuppressive drugs are essential in the management of IGM. For patients who failed treatment of antibiotics, steroids and surgery, 94% of them improved and 75% achieved remission after 15 months of treatment with MTX (138). Complete drug-free remission was obtained in 45 (88.2%) of 51 patients treated with immunosuppressive drugs. MTX is preferred as an effective corticosteroid-tapering agent in initial treatment, and azathioprine (AZA) acts as an alternative in patients intolerant to MTX, both of which regulate the inflammatory process and prevent further complications (123, 139-141). Although the treatment algorithm after diagnosis of IGM is still controversial, steroid treatment should be administered first in widespread disease and then MTX or surgery should be considered depending on the response to steroids (123). If IGM patients do not respond to steroids alone, low-dose MTX and prednisone should be considered as an alternative to surgery (142). Meanwhile, the combination of steroids and methotrexate is an effective and reliable option to ensure long-term remission (143). Colchicine and hydroxychloroquine were used in six and four patients with IGM, respectively. Of the six patients treated with colchicine, four did not relapse; and only one patient treated with hydroxychloroquine as a monotherapy healed (11). In 2017, a case of IGM that was refractory to MTX and intralesional and systemic steroids but responded well to mycophenolate mofetil 1500mg twice daily (144), and a recent prospective study also showed the promising results of mycophenolate mofetil in the treatment management of IGM (145).

Rationale of antibodies and other immune approaches

Several TNF inhibitors were approved by the Food and Drug Administration (FDA). To reduce side effects and costs, most

patients with mild or moderate disease are treated with MTX before adding or switching to a TNF inhibitor. These agents may be used alone or in combination with other medications (e.g. prednisone, MTX, hydroxychloroquine). A 32-year-old female patient in a pilot report of IGM was successfully treated with etanercept in a combination with MTX; therefore, the improvement achieved with TNF inhibitors in this case offered a different therapeutic option for patients with a similar condition (146). A premenopausal Hispanic woman with refractory IGM had an excellent response to adalimumab despite aggressive steroid and MTX therapy (147). The latest case with overlapping clinical features of IGM, hidradenitis suppurativa, and pyoderma gangrenosum was successfully treated with adalimumab and eventually achieved complete resolution of all lesions without relapse even after discontinuation of biologic therapy (148).

Biologic agents that deplete B cells (e.g., rituximab) and inhibit factors which activate B cells (e.g., belimumab) are both used to treat a variety of rheumatic diseases. In addition to antibody production, B cells present antigens to T cells, activate T cells, and promote the production of proinflammatory cytokines (149). Rituximab, for example, eliminates CD20-positive B cells, induces complement-mediated cytotoxicity, and stimulates apoptosis but has little or nonspecific effects on autoantibody titers; thus, the combination of the above effects may explain disease activity (150).

Several pathways mediating receptor signal transduction have been targeted by orally administered chemical compounds (e.g. small-molecule kinase inhibitors) (151, 152). A pan-janus kinase (JAK) inhibitor was approved in Japan and Korea for the treatment of rheumatoid arthritis patients with an inadequate response to disease-modifying antirheumatic drugs (153, 154); while another selective JAK-1 inhibitor, approved by the European Medicines Agency, showed rapid and significant improvements in disease activity compared with placebo (155, 156). A systematic literature review and meta-analysis of IGM demonstrated the superiority of TNF-blocking agents in combination with MTX (157). A recombinant IL-1 receptor antagonist is significantly less potent than TNF inhibitors in most patients with RA (158, 159). Moreover, the regimen of a IL-1receptor antagonist plus a TNF inhibitor or other biologic agents has not been recommended due to an increased incidence of serious adverse events, including serious infections (160, 161). IL-6 inhibitors have been successfully used in the treatment of RA and other rheumatic diseases (162).

Characteristics that cannot be explained by immune pathogenesis/unsolved questions and unmet medical needs

ANA

Clinical research has not demonstrated the clinical utility of many autoantibodies (e.g. ANA) in IGM (163). Another study did

not unravel the autoimmune property by determining the levels of ANA and extractable nuclear antigen (ENA) in patients with IGM (24). Altintoprak and colleagues also evaluated the levels of ANA and ENA in 26 parous patients with IGM, although no indicator for ANA or ENA levels favoring the preeminent impact of autoimmunity on the pathogenesis was found (62).

Conclusion

IGM is a chronic benign inflammatory breast disease of unknown etiology, but of autoimmune-favorable features such as erythema nodosum and arthritis, coexistence with other autoimmune diseases and seasonal fluctuations in disease onset. Triggering events and precipitating factors are also discussed in this systemic review (including pregnancy and parturition, hormonal disorders, trauma and ductal perforation, smoking, microorganisms, and others). Cellular dysregulation, IL family/ cytokines, HLA and others might participate in the immune pathogenesis of IGM. As conventional immunosuppressive agents, steroids and immunomodulators (MTX, AZA, colchicine and hydroxychloroquine) play an important role in treatment. Emerging novel immunological approaches are potential alternatives with a range of biologic agents targeting B cells, kinase inhibitors, and biologic cytokine inhibitors. However, the findings of most clinical studies on autoantibodies do not reach clinical utility and cannot be explained by immune pathogenesis. In the future, more research is expected to focus on the etiology, immunologic mechanism and treatment of IGM.

Search strategy

PubMed, Scopus, Embase and Cochrane databases were systematically searched for related articles and reviews. The predefined search terms are "IGM," "idiopathic granulomatous mastitis," "GLM," "granulomatous lobular mastitis," "NPM," "non-puerperal mastitis," "NLM," "nonlactational mastitis,". Synonyms were combined by the Boolean operator (OR) and paratactic terms were combined by the Boolean operator (AND) when searching relevant content for the corresponding section.

Author contributions

JY: Conceptualization, Funding acquisition, Methodology, Writing – original draft, Writing – review & editing. XW: Data curation, Formal analysis, Validation, Visualization, Writing – original draft, Writing – review & editing. XH: Conceptualization, Supervision, Writing – review & editing. JZL: Data curation, Formal analysis, Investigation, Visualization, Writing – review & editing. HZ: Conceptualization, Formal analysis, Investigation, Supervision, Validation, Writing – review & editing. HW: Conceptualization, Investigation, Methodology, Supervision, Validation, Writing – review & editing. JL: Conceptualization, Investigation, Methodology, Supervision, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This work was supported by Natural Science Foundation of Sichuan (NO. 2022NSFSC1347).

References

1. Kessler E, Wolloch Y. Granulomatous mastitis: a lesion clinically simulating carcinoma. Am J Clin Pathol. (1972) 58:642-6. doi: 10.1093/ajcp/58.6.642

2. Steuer AB, Stern MJ, Cobos G, Castilla C, Joseph KA, Pomeranz MK, et al. Clinical characteristics and medical management of idiopathic granulomatous mastitis. *JAMA Dermatol.* (2020) 156:460–4. doi: 10.1001/jamadermatol.2019.4516

3. Alsaleh N. Assertive clinical practice in managing patients with idiopathic granulomatous mastitis: Review of literature. *Ann Med Surg.* (2021) 70:102792. doi: 10.1016/j.amsu.2021.102792

4. Li J. Diagnosis and treatment of 75 patients with idiopathic lobular granulomatous mastitis. J Invest Surg. (2019) 32:414-20. doi: 10.1080/08941939.2018.1424270

5. Uysal E, Soran A, Sezgin EGranulomatous Mastitis Study G. Factors related to recurrence of idiopathic granulomatous mastitis: what do we learn from a multicentre study? *ANZ J Surg.* (2018) 88:635–9. doi: 10.1111/ans.14115

6. Hashmi D, Al Samaraee A, Marks B, Fasih T. Idiopathic granulomatous mastitis: a diagnostic dilemma. Br J Hosp Med. (2020) 81:1–4. doi: 10.12968/hmed.2019.0287

7. Wolfrum A, Kümmel S, Theuerkauf I, Pelz E, Reinisch M. Granulomatous mastitis: a therapeutic and diagnostic challenge. *Breast Care.* (2018) 13:413-8. doi: 10.1159/000495146

8. Emsen A, Köksal H, Uçaryilmaz H, Kadoglou N, Artac H. The alteration of lymphocyte subsets in idiopathic granulomatous mastitis. *Turk J Med Sci.* (2021) 51:1905–11. doi: 10.3906/sag-2012-192

9. Godazandeh G, Shojaee L, Alizadeh-Navaei R, Hessami A. Corticosteroids in idiopathic granulomatous mastitis: a systematic review and meta-analysis. *Surg Today.* (2021) 51:1897-905. doi: 10.1007/s00595-021-02234-4

10. Chan CW. The treatment conundrum that is idiopathic granulomatous mastitis. Ann Acad Med Singap. (2021) 50:596–7. doi: 10.47102/annals-acadmedsg.

11. Neel A, Hello M, Cottereau A, Graveleau J, De Faucal P, Costedoat-Chalumeau N, et al. Long-term outcome in idiopathic granulomatous mastitis: a western multicentre study. *QJM*. (2013) 106:433–41. doi: 10.1093/qjmed/hct040

12. Tang ELS, Ho CSB, Chan PMY, Chen JJC, Goh MH, Tan EY. The therapeutic dilemma of idiopathic granulomatous mastitis. *Ann Acad Med Singap*. (2021) 50:598–605. doi: 10.47102/annals-acadmedsg.

13. Zhang X, Li Y, Zhou Y, Liu D, Chen L, Niu K, et al. A systematic surgical approach for the treatment of idiopathic granulomatous mastitis: a case series. *Gland Surg.* (2020) 9:261–70. doi: 10.21037/gs

14. Azizi A, Prasath V, Canner J, Gharib M, Sadat Fattahi A, Naser Forghani M, et al. Idiopathic granulomatous mastitis: Management and predictors of recurrence in 474 patients. *Breast J.* (2020) 26:1358–62. doi: 10.1111/tbj.13822

15. Wang J, Zhang Y, Lu X, Xi C, Yu K, Gao R, et al. Idiopathic granulomatous mastitis with skin rupture: a retrospective cohort study of 200 patients who underwent surgical and nonsurgical treatment. *J Invest Surg.* (2021) 34:810–5. doi: 10.1080/08941939.2019.1696905

16. Tekin L, Elibol FD. Is there any relationship between granulomatous mastitis and seasons? An analysis of seasonal frequency, clinical, and radiologic findings. *Eur J Breast Health.* (2020) 16:235. doi: 10.5152/ejbh.

17. Bayrak BY, Cam I, Eruyar AT, Utkan NZ. Clinicopathological evaluation of idiopathic granulomatous mastitis patients: A retrospective analysis from a tertiary care hospital in Turkey. *Ann Diagn Pathol.* (2021) 55:151812. doi: 10.1016/j.anndiagpath.2021.151812

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

18. Chalmers R, McClellan P, Silva V, Shutt N, Restini C. Red flags for the differential diagnosis of granulomatous mastitis: a case report. J Med Case Rep. (2020) 14:1–7. doi: 10.1186/s13256-020-02563-x

19. Toprak N, Toktas O, Ince S, Gunduz A, Yokus A, Akdeniz H, et al. Does ARFI elastography complement B-mode ultrasonography in the radiological diagnosis of idiopathic granulomatous mastitis and invasive ductal carcinoma? *Acta Radiol.* (2022) 63:28–34. doi: 10.1177/0284185120983568

20. Dursun M, Yilmaz S, Yahyayev A, Salmaslioglu A, Yavuz E, Igci A, et al. Multimodality imaging features of idiopathic granulomatous mastitis: outcome of 12 years of experience. *Radiol Med.* (2012) 117:529–38. doi: 10.1007/s11547-011-0733-2

21. Çetinkaya G, Kozan R, Emral AC, Tezel E. Granulomatous mastitis, watch and wait is a good option. Ir J Med Sci. (2021) 190:1117-22. doi: 10.1007/s11845-020-02406-0

22. Deng J, Yu L, Yang Y, Feng X-J, Sun J, Liu J, et al. Steroids administered after vacuum-assisted biopsy in the management of idiopathic granulomatous mastitis. *J Clin Pathol.* (2017) 70:827–31. doi: 10.1136/jclinpath-2016-204287

23. Gümüş M, Akkurt ZM, Gümüş H. Is erythema nodosum coexisting with lesions of the breast a suggestive sign for idiopathic granulomatous mastitis? *Turk J Surg.* (2018) 34:71. doi: 10.5152/turkjsurg.

24. Altintoprak F, Karakece E, Kivilcim T, Dikicier E, Cakmak G, Celebi F, et al. Idiopathic granulomatous mastitis: an autoimmune disease? *ScientificWorldJournal*. (2013) 2013:148727. doi: 10.1155/2013/148727

25. Benson J, Dumitru D. Idiopathic granulomatous mastitis: presentation, investigation and management. *Future Oncol (London England).* (2016) 12:1381–94. doi: 10.2217/fon-2015-0038

26. Kıvılcım T, Altıntoprak F, Memiş B, Ferhatoğlu M, Kartal A, Dikicier E, et al. Role of bacteriological agents in idiopathic granulomatous mastitis: real or not? *Eur J Breast Health*. (2018) 15:32–6. doi: 10.5152/ejbh.2018.4249

27. Adams D, Hubscher S, Scott D. Granulomatous mastitis-a rare cause of erythema nodosum. *Postgrad Med J.* (1987) 63:581-2. doi: 10.1136/pgmj.63.741.581

28. Velidedeoğlu M, Papila Kundaktepe B, Mete B, Uğurlu S. Idiopathic granulomatous mastitis associated with erythema nodosum may indicate a worse prognosis. *Int J Rheum Dis.* (2021) 24:1370–7. doi: 10.1111/1756-185X.14218

29. Çetin K, Sıkar HE, Güllüoğlu BM. Idiopathic granulomatous mastitis with erythema nodosum: is it a variant of clinical presentation indicating treatment resistance? A retrospective cohort study. *Breast J.* (2020) 26:1645–51. doi: 10.1111/tbj.13944

30. Otto TS, Argobi Y, Lerwill MJ, Smith GP, Fedeles F. A retrospective study of idiopathic granulomatous mastitis diagnosis and clinical presentation. J Am Acad Dermatol. (2022) 86:467–9. doi: 10.1016/j.jaad.2021.09.055

31. Li S-B, Xiong Y, Han X-R, Liu Z-Y, Lv X-L, Ning P. Pregnancy associated granulomatous mastitis: clinical characteristics, management, and outcome. *Breastfeed Med.* (2021) 16:759–64. doi: 10.1089/bfm.2021.0023

32. Koksal H. What are the new findings with regard to the mysterious disease idiopathic granulomatous mastitis? *Surg Today.* (2021) 51:1158-68. doi: 10.1007/s00595-020-02204-2

33. Ringsted S, Friedman M. A rheumatologic approach to granulomatous mastitis: A case series and review of the literature. *Int J Rheum Dis.* (2021) 24:526–32. doi: 10.1111/1756-185X.14065

34. Şener Bahçe Z, Aktaş H. Patients with idiopathic granulomatous mastitis accompanied by erythema nodosum. *Int J Clin Pract.* (2021) 75:e13928. doi: 10.1111/ ijcp.13928

35. Akin M, Karabacak H, Esendağli G, Yavuz A, Gültekin S, Dikmen K, et al. Coexistence of idiopathic granulomatous mastitis and erythemanodosum: successful treatment with corticosteroids. *Turk J Med Sci.* (2017) 47:1590–2. doi: 10.3906/sag-1611-100

36. Koufakis T, Gabranis I. Erythema nodosum leads to the diagnosis of pulmonary tuberculosis. Pan Afr Med J. (2014) 18:291. doi: 10.11604/pamj.2014.18.291.5170

37. Pandey TS, Mackinnon JC, Bressler L, Millar A, Marcus EE, Ganschow PS. Idiopathic granulomatous mastitis—a prospective study of 49 women and treatment outcomes with steroid therapy. *Breast J.* (2014) 20:258–66. doi: 10.1111/tbj.2014.20.issue-3

38. Zhang L, Shi T, Yang Y, Zhang F. An SLE patient with prolactinoma and recurrent granulomatous mastitis successfully treated with hydroxychloroquine and bromocriptine. *Lupus*. (2014) 23:417–20. doi: 10.1177/0961203313520059

39. Letourneux C, Diemunsch P, Korganow A-S, Akladios CY, Bellocq J-P, Mathelin C. First report of granulomatous mastitis associated with Sjögren's syndrome. *World J Surg Oncol.* (2013) 11:1–5. doi: 10.1186/1477-7819-11-268

40. Yazigi G, Trieu BH, Landis M, Parikh JG, Mangal M. Granulomatous mastitis: A rare case with Sjogren's syndrome and complications. *Cureus.* (2019) 11:e5359. doi: 10.7759/cureus.5359

41. Matusiewicz A, Stróżyńska-Byrska J, Olesińska M. Polyautoimmunity in rheumatological conditions. *Int J Rheum Dis.* (2019) 22:386–91. doi: 10.1111/1756-185X.13454

42. Chambers S, Charman S, Rahman A, Isenberg D. Development of additional autoimmune diseases in a multiethnic cohort of patients with systemic lupus erythematosus with reference to damage and mortality. *Ann Rheum Dis.* (2007) 66:1173–7. doi: 10.1136/ard.2006.062984

43. Lazarus M, Isenberg D. Development of additional autoimmune diseases in a population of patients with primary Sjögren's syndrome. *Ann Rheum Dis.* (2005) 64:1062–4. doi: 10.1136/ard.2004.029066

44. Yaghan RJ, Ayoub NM, Hamouri S, Al-Mohtaseb A, Gharaibeh M, Yaghan L, et al. The role of establishing a multidisciplinary team for idiopathic granulomatous mastitis in improving patient outcomes and spreading awareness about recent disease trends. *Int J Breast Canc.* (2020) 2020:5243958. doi: 10.1155/2020/5243958

45. Watad A, Azrielant S, Bragazzi NL, Sharif K, David P, Katz I, et al. Seasonality and autoimmune diseases: the contribution of the four seasons to the mosaic of autoimmunity. *J Autoimmun.* (2017) 82:13–30. doi: 10.1016/j.jaut.2017.06.001

46. Lai Q-L, Cai M-T, Zheng Y, Zhang T-Y, Fang G-L, Shen C-H, et al. Seasonal variation in autoimmune encephalitis: A multi-center retrospective study. *J Neuroimmunol.* (2021) 359:577673. doi: 10.1016/j.jneuroim.2021.577673

47. Mori H, Sawada T, Nishiyama S, Shimada K, Tahara K, Hayashi H, et al. Influence of seasonal changes on disease activity and distribution of affected joints in rheumatoid arthritis. *BMC Musculoskelet Disord*. (2019) 20:1–8. doi: 10.1186/s12891-019-2418-2

48. Valesini G, Gerardi MC, Iannuccelli C, Pacucci VA, Pendolino M, Shoenfeld Y. Citrullination and autoimmunity. *Autoimmun Rev.* (2015) 14:490–7. doi: 10.1016/j.autrev.2015.01.013

49. Galbo H, Kall L. Circadian variations in clinical symptoms and concentrations of inflammatory cytokines, melatonin, and cortisol in polymyalgia rheumatica before and during prednisolone treatment: a controlled, observational, clinical experimental study. *Arthritis Res Ther.* (2016) 18:1–10. doi: 10.1186/s13075-016-1072-4

50. Kühne T, Imbach P, Bolton-Maggs PH, Berchtold W, Blanchette V, Buchanan GR, et al. Newly diagnosed idiopathic thrombocytopenic purpura in childhood: an observational study. *Lancet.* (2001) 358:2122–5. doi: 10.1016/S0140-6736(01)07219-1

51. Schlesinger N, Schlesinger M. Seasonal variation of rheumatic diseases. *Discovery Med.* (2009) 5:64–9.

52. Velidedeoglu M, Umman V, Kilic F, Celik V, Gazioglu E, Hatipoglu E, et al. Idiopathic granulomatous mastitis: introducing a diagnostic algorithm based on 5 years of follow-up of 152 cases from Turkey and a review of the literature. *Surg Today*. (2022) 52:668–80. doi: 10.1007/s00595-021-02367-6

53. Imoto S, Kitaya T, Kodama T, Hasebe T, Mukai K. Idiopathic granulomatous mastitis: case report and review of the literature. *Japan J Clin Oncol*. (1997) 27:274–7. doi: 10.1093/jjco/27.4.274

54. Gautier N, Lalonde L, Tran-Thanh D, El Khoury M, David J, Labelle M, et al. Chronic granulomatous mastitis: Imaging, pathology and management. *Eur J Radiol.* (2013) 82:e165–175. doi: 10.1016/j.ejrad.2012.11.010

55. Al-Khaffaf B, Knox F, Bundred N. Idiopathic granulomatous mastitis: a 25-year experience. J Am Coll Surgeons. (2008) 206:269-73. doi: 10.1016/j.jamcollsurg.2007.07.041

56. Jethwa H, Lam S, Smith C, Giles I. Does rheumatoid arthritis really improve during pregnancy? A systematic review and metaanalysis. *J Rheumatol.* (2019) 46:245–50. doi: 10.3899/jrheum.180226

57. Silman A, Kay A, Brennan P. Timing of pregnancy in relation to the onset of rheumatoid arthritis. *Arthritis Rheumatol.* (1992) 35:152-5. doi: 10.1002/art.1780350205

58. Jansson R, Dahlberg PA, Winsa B, Meirik O, Säfwenberg J, Karlsson A. The postpartum period constitutes an important risk for the development of clinical Graves' disease in young women. *Acta Endocrinol.* (1987) 116:321–5. doi: 10.1530/acta.0.1160321

59. Piccinni MP, Lombardelli L, Logiodice F, Kullolli O, Parronchi P, Romagnani S. How pregnancy can affect autoimmune diseases progression? *Clin Mol Allergy*. (2016) 14:11. doi: 10.1186/s12948-016-0048-x

60. Sheybani F, Naderi H, Gharib M, Sarvghad M, Mirfeizi Z. Idiopathic granulomatous mastitis: Long-discussed but yet-to-be-known. *Autoimmunity*. (2016) 49:236–9. doi: 10.3109/08916934.2016.1138221

61. Sheybani F, Sarvghad M, Naderi H, Gharib M. Treatment for and clinical characteristics of granulomatous mastitis. *Obstet Gynecol.* (2015) 125:801-7. doi: 10.1097/AOG.00000000000734

62. Altintoprak F, Kivilcim T, Ozkan OV. Aetiology of idiopathic granulomatous mastitis. World J Clin Cases. (2014) 2:852-8. doi: 10.12998/wjcc.v2.i12.852

63. Agrawal A, Pabolu S. A rare case of idiopathic granulomatous mastitis in a nulliparous woman with hyperprolactinemia. *Cureus.* (2019) 11:e4680. doi: 10.7759/ cureus.4680

64. Destek S, Gul VO, Ahioglu S, Serin KR. Pituitary adenoma and hyperprolactinemia accompanied by idiopathic granulomatous mastitis. *Case Rep Endocrinol.* (2017) 2017:3974291. doi: 10.1155/2017/3974291

65. Holla SN, Vittalrao A, Kamath A, Kamalkishore MK, Ommurugan B. Risperidone induced granulomatous mastitis secondary to hyperprolactinemia in a non-pregnant woman-A rare case report in a bipolar disorder. *J OF Clin AND Diagn Res.* (2017) 11:FD01–3. doi: 10.7860/JCDR/2017/20733.9278

66. Lin CH, Hsu CW, Tsao TY, Chou J. Idiopathic granulomatous mastitis associated with risperidone-induced hyperprolactinemia. *Diagn Pathol.* (2012) 7:2. doi: 10.1186/1746-1596-7-2

67. Li J, McGregor H. Idiopathic granulomatous mastitis associated with hyperprolactinemia: A nonoperative approach. *Breast J.* (2017) 23:742–4. doi: 10.1111/tbj.12914

68. Cai R, Zhao J, Qiao Z, Li Y. Idiopathic granulomatous mastitis with normal prolactin level caused by risperidone. *Asian J Surg.* (2021) 44:763–4. doi: 10.1016/ j.asjsur.2021.02.026

69. Bellis AD, Bizzarro A, Pivonello R, Lombardi G, Bellastella A. Prolactin and autoimmunity. *Pituitary*. (2005) 8:25–30. doi: 10.1007/s11102-005-5082-5

70. Kalsi AK, Halder A, Jain M, Chaturvedi PK, Mathew M, Sharma JB. Association of raised levels of IL-4 and anti-TPO with hyperprolactinemia. *Am J Reprod Immunol.* (2019) 81:e13085. doi: 10.1111/ajj.13085

71. Saha S, Gonzalez J, Rosenfeld G, Keiser H, Peeva E. Prolactin alters the mechanisms of B cell tolerance induction. *Arthritis Rheumatol.* (2009) 60:1743–52. doi: 10.1002/art.24500

72. Borba V, Zandman-Goddard G, Shoenfeld Y. Exacerbations of autoimmune diseases during pregnancy and postpartum. *Best Pract Res Clin Endocrinol Metab.* (2019) 33:101321. doi: 10.1016/j.beem.2019.101321

73. Makineli S, van Diest PJ, Fernandez MA, Witkamp AJ. Idiopathic granulomatous mastitis after ductoscopy: A case report. *Int J Surg Case Rep.* (2021) 88:106540. doi: 10.1016/j.ijscr.2021.106540

74. Tanriverdi F, De Bellis A, Bizzarro A, Sinisi A, Bellastella G, Pane E, et al. Antipituitary antibodies after traumatic brain injury: is head trauma-induced pituitary dysfunction associated with autoimmunity? *Eur J Endocrinol.* (2008) 159:7–13. doi: 10.1530/EJE-08-0050

75. Özel L, Ünal A, Ünal E, Kara M, Erdoğdu E, Krand O, et al. Granulomatous mastitis: is it an autoimmune disease? Diagnostic and therapeutic dilemmas. *Surg Today.* (2012) 42:729–33. doi: 10.1007/s00595-011-0046-z

76. Oran EŞ, Gürdal SÖ, Yankol Y, Öznur M, Calay Z, Tunacı M, et al. Management of idiopathic granulomatous mastitis diagnosed by core biopsy: a retrospective multicenter study. *Breast J.* (2013) 19:411–8. doi: 10.1111/tbj.2013.19.issue-4

77. Baslaim MM, Khayat HA, Al-Amoudi SA. Idiopathic granulomatous mastitis: a heterogeneous disease with variable clinical presentation. *World J Surg.* (2007) 31:1677–81. doi: 10.1007/s00268-007-9116-1

78. Asoglu O, Ozmen V, Karanlik H, Tunaci M, Cabioglu N, Igci A, et al. Feasibility of surgical management in patients with granulomatous mastitis. *Breast J.* (2005) 11:108–14. doi: 10.1111/j.1075-122X.2005.21576.x

79. Shoenfeld Y, Tincani A. Autoantibodies-the smoke and the fire. *Autoimmunity*. (2005) 38:1-2. doi: 10.1080/14756360400021882

80. Perricone C, Versini M, Ben-Ami D, Gertel S, Watad A, Segel MJ, et al. Smoke and autoimmunity: The fire behind the disease. *Autoimmun Rev.* (2016) 15:354–74. doi: 10.1016/j.autrev.2016.01.001

81. Harel-Meir M, Sherer Y, Shoenfeld Y. Tobacco smoking and autoimmune rheumatic diseases. Nat Clin Pract Rheumatol. (2007) 3:707–15. doi: 10.1038/ncprheum0655

82. Klareskog L, Stolt P, Lundberg K, Källberg H, Bengtsson C, Grunewald J, et al. A new model for an etiology of rheumatoid arthritis: smoking may trigger HLA-DR (shared epitope)-restricted immune reactions to autoantigens modified by citrullination. *Arthritis Rheumatol.* (2006) 54:38–46. doi: 10.1002/art.21575

83. Lee S-H, Goswami S, Grudo A, Song L-Z, Bandi V, Goodnight-White S, et al. Antielastin autoimmunity in tobacco smoking–induced emphysema. *Nat Med.* (2007) 13:567–9. doi: 10.1038/nm1583

84. Strickland FM, Li Y, Johnson K, Sun Z, Richardson BC. CD4+ T cells epigenetically modified by oxidative stress cause lupus-like autoimmunity in mice. *Autoimmunity*. (2015) 62:75–80. doi: 10.1016/j.jaut.2015.06.004

85. Wilson JP, Massoli N, Marshall J, Foss RM, Copeland EM, Grobmyer SR. Idiopathic granulomatous mastitis: in search of a therapeutic paradigm. *Am Surg.* (2007) 73:798. doi: 10.1177/000313480707300813

86. Maffini F, Baldini F, Bassi F, Luini A, Viale G. Systemic therapy as a first choice treatment for idiopathic granulomatous mastitis. *J Cutan Pathol.* (2009) 36:689–91. doi: 10.1111/j.1600-0560.2008.01102.x

87. Tuli R, O'Hara BJ, Hines J, Rosenberg AL. Idiopathic granulomatous mastitis masquerading as carcinoma of the breast: a case report and review of the literature. *Int Semin Surg Oncol.* (2007) 4:21. doi: 10.1186/1477-7800-4-21

88. Albogami M, Alsaedy A, Matrood RA, Alotaibi Y, Al Ahmadi BA. Case of a female patient presenting with idiopathic granulomatous mastitis with superimposed enterococcus avium infection. *Cureus*. (2022) 14:e29997. doi: 10.7759/cureus.29997

89. Krueger K, Guggina LM. Mycobacterium mucogenicum skin and soft tissue infection of the breast mimicking idiopathic granulomatous mastitis. *BMJ Case Rep.* (2019) 12:e231900. doi: 10.1136/bcr-2019-231900

90. Stary CM, Lee YS, Balfour J. Idiopathic granulomatous mastitis associated with corynebacterium sp. Infection. *Hawaii Med J.* (2011) 70:99–101.

91. Taylor G, Paviour S, Musaad S, Jones W, Holland D. A clinicopathological review of 34 cases of inflammatory breast disease showing an association between corynebacteria infection and granulomatous mastitis. *Pathology*. (2003) 35:109–19. doi: 10.1080/0031302031000082197

92. Lu C, Marcucci V, Kibbe E, Patel R. Granulomatous mastitis secondary to Corynebacterium requiring surgical intervention: a complicated diagnosis. *J Surg Case Rep.* (2023) 2023:rjad211. doi: 10.1093/jscr/rjad211

93. Mackay IR, Leskovsek NV, Rose NR. Cell damage and autoimmunity: a critical appraisal. J Autoimmun. (2008) 30:5–11. doi: 10.1016/j.jaut.2007.11.009

94. Rose NR. Negative selection, epitope mimicry and autoimmunity. *Curr Opin Immunol.* (2017) 49:51–5. doi: 10.1016/j.coi.2017.08.014

95. Fairweather D, Frisancho-Kiss S, Rose NR. Viruses as adjuvants for autoimmunity: evidence from Coxsackievirus-induced myocarditis. *Rev Med Virol.* (2005) 15:17–27. doi: 10.1002/rmv.445

96. James JA, Harley JB, Scofield RH. Epstein-Barr virus and systemic lupus erythematosus. *Curr Opin Rheumatol.* (2006) 18:462-7. doi: 10.1097/01.bor.0000240355.37927.94

97. Ascherio A, Munger KL. EBV and autoimmunity. *Curr Top Microbiol Immunol.* (2015) 390:365–85. doi: 10.1007/978-3-319-22822-8_15

 Elson CO, Alexander KL. Host-microbiota interactions in the intestine. *Dig Dis.* (2015) 33:131–6. doi: 10.1159/000369534

99. Scher JU, Sczesnak A, Longman RS, Segata N, Ubeda C, Bielski C, et al. Expansion of intestinal Prevotella copri correlates with enhanced susceptibility to arthritis. *eLife*. (2013) 2:e01202. doi: 10.7554/eLife.01202

100. Azzouz D, Omarbekova A, Heguy A, Schwudke D, Gisch N, Rovin B, et al. Lupus nephritis is linked to disease-activity associated expansions and immunity to a gut commensal. *Ann Rheum Dis.* (2019) 78:947–56. doi: 10.1136/annrheumdis-2018-214856

101. Manfredo Vieira S, Hiltensperger M, Kumar V, Zegarra-Ruiz D, Dehner C, Khan N, et al. Translocation of a gut pathobiont drives autoimmunity in mice and humans. *Science*. (2018) 359:1156–61. doi: 10.1126/science.aar7201

102. Schelfout K, Tjalma WA, Cooremans ID, Coeman DC, Colpaert CG, Buytaert PM. Observations of an idiopathic granulomatous mastitis. *Eur J Obstet Gynecol Reprod Biol.* (2001) 97:260–2. doi: 10.1016/S0301-2115(00)00546-7

103. Kobayashi T, Kawamura H, Kanda Y, Matsumoto H, Saito S, Takeda K, et al. Natural killer T cells suppress zymosan A-mediated granuloma formation in the liver by modulating interferon- γ and interleukin-10. *Immunology*. (2012) 136:86–95. doi: 10.1111/j.1365-2567.2012.03562.x

104. Lacambra M, Thai TA, Lam CC, Yu AM, Pham HT, Tran PV, et al. Granulomatous mastitis: the histological differentials. *J Clin Pathol.* (2011) 64:405–11. doi: 10.1136/jcp.2011.089565

105. Cetinkaya OA, Celik SU, Terzioglu SG, Eroglu A. The predictive value of the neutrophil-to-lymphocyte and platelet-to-lymphocyte ratio in patients with recurrent idiopathic granulomatous mastitis. *Eur J Breast Health*. (2020) 16:61–5. doi: 10.5152/ejbh.

106. Li Q, Wan J, Feng Z, Shi J, Wei W. Predictive significance of the preoperative neutrophil-lymphocyte ratio for recurrence in idiopathic granulomatous mastitis patients. *Am Surg.* (2023) 89:5577–83. doi: 10.1177/00031348231161793

107. Yigitbasi M, Guntas G, Atak T, Sonmez C, Yalman H, Uzun H. The role of interleukin-33 as an inflammatory marker in differential diagnosis of idiopathic granulomatous mastitis and breast cancer. J Invest Surg: Off J Acad Surg Res. (2017) 30:272–6. doi: 10.1080/08941939.2016.1240270

108. Huang YM, Lo C, Cheng CF, Lu CH, Hsieh SC, Li KJ. Serum C-reactive protein and interleukin-6 levels as biomarkers for disease severity and clinical outcomes in patients with idiopathic granulomatous mastitis. J Clin Med. (2021) 10:2077. doi: 10.3390/jcm10102077

109. Koksal H, Vatansev H, Artac H, Kadoglou N. The clinical value of interleukins-8, -10, and -17 in idiopathic granulomatous mastitis. *Clin Rheumatol.* (2020) 39:1671– 7. doi: 10.1007/s10067-020-04925-8

110. Saydam M, Yilmaz KB, Sahin M, Yanik H, Akinci M, Yilmaz I, et al. New findings on autoimmune etiology of idiopathic granulomatous mastitis: serum IL-17, IL-22 and IL-23 levels of patients. *J Invest Surg.* (2021) 34:993–7. doi: 10.1080/08941939.2020.1725190

111. Cakir C, Nayci AE, Ferlengez E, Guler M, Idiz UO. Cytokines the etiology of idiopathic granulomatous mastitis. *J Coll Physicians Surg Pak.* (2022) 32:869–73. doi: 10.29271/jcpsp

112. Zhou F, Yu L, Ma Z, Yu Z. Granulomatous lobular mastitis. *Chronic Dis Trans Med.* (2016) 2:17–21. doi: 10.1016/j.cdtm.2016.02.004

113. Koksal H. Human leukocyte antigens class I and II in patients with idiopathic granulomatous mastitis. Am J Surg. (2019) 218:605-8. doi: 10.1016/j.amjsurg.2019.01.038

114. Ates D, Doner HC, Kurban S, Koksal H. The effect of soluble TREM-1 in idiopathic granulomatous mastitis. *Immunol Invest.* (2022) 51:839–50. doi: 10.1080/08820139.2021.1879846

115. Velidedeoglu M, Kundaktepe BP, Aksan H, Uzun H. Preoperative fibrinogen and hematological indexes in the differential diagnosis of idiopathic granulomatous mastitis and breast cancer. *Medicina (Kaunas).* (2021) 57:698. doi: 10.21203/rs.3.rs-460509/v1

116. Bouton ME, Jayaram L, O'Neill PJ, Hsu CH, Komenaka IK. Management of idiopathic granulomatous mastitis with observation. *Am J Surg.* (2015) 210:258–62. doi: 10.1016/j.amjsurg.2014.08.044

117. Davis J, Cocco D, Matz S, Hsu C, Brown M, Lee J, et al. Re-evaluating if observation continues to be the best management of idiopathic granulomatous mastitis. *Surgery*. (2019) 166:1176–80. doi: 10.1016/j.surg.2019.06.030

118. Zhou F, Liu L, Liu L, Yu L, Wang F, Xiang Y, et al. Comparison of conservative versus surgical treatment protocols in treating idiopathic granulomatous mastitis: A meta-analysis. *Breast Care (Basel)*. (2020) 15:415–20. doi: 10.1159/000503602

119. Fattahi AS, Amini G, Sajedi F, Mehrad-Majd H. Factors affecting recurrence of idiopathic granulomatous mastitis: A systematic review. *Breast J.* (2023) 2023:9947797. doi: 10.1155/2023/9947797

120. Pandey TS, Mackinnon JC, Bressler L, Millar A, Marcus EE, Ganschow PS. Idiopathic granulomatous mastitis-a prospective study of 49 women and treatment outcomes with steroid therapy. *Breast J.* (2014) 20:258–66. doi: 10.1111/tbj.2014.20.issue-3

121. Lei X, Chen K, Zhu L, Song E, Su F, Li S. Treatments for idiopathic granulomatous mastitis: systematic review and meta-analysis. *Breastfeed Med.* (2017) 12:415–21. doi: 10.1089/bfm.2017.0030

122. Montazer M, Dadashzadeh M, Toomatari S. 95 comparison of the outcome of low dose and high-dose corticosteroid in the treatment of idiopathic granulomatous mastitis. *Asian Pacific J Cancer Prev.* (2020) 21:993–6. doi: 10.31557/APJCP.2020.21.4.993

123. Akbulut S, Yilmaz D, Bakir S. Methotrexate in the management of idiopathic granulomatous mastitis: review of 108 published cases and report of four cases. *Breast J.* (2011) 17:661–8. doi: 10.1111/tbj.2011.17.issue-6

124. Toktas O, Toprak N. Treatment results of intralesional steroid injection and topical steroid administration in pregnant women with idiopathic granulomatous mastitis. *Eur J Breast Health*. (2021) 17:283–7. doi: 10.4274/ejbh

125. Gunduz Y, Altintoprak F, Tatli Ayhan L, Kivilcim T, Celebi F. Effect of topical steroid treatment on idiopathic granulomatous mastitis: clinical and radiologic evaluation. *Breast J.* (2014) 20:586–91. doi: 10.1111/tbj.2014.20.issue-6

126. Hu T, Li S, Huang H, Huang H, Tan L, Chen Y, et al. Multicentre, randomised, open-label, non-inferiority trial comparing the effectiveness and safety of ductal lavage versus oral corticosteroids for idiopathic granulomatous mastitis: a study protocol. *BMJ Open.* (2020) 10:e036643. doi: 10.1136/bmjopen-2019-036643

127. Chen K, Zhu L, Hu T, Tan C, Zhang J, Zeng M, et al. Ductal lavage for patients with nonlactational mastitis: A single-arm, proof-of-concept trial. *J Surg Res.* (2019) 235:440–6. doi: 10.1016/j.jss.2018.10.023

128. Chatham W, Kimberly R. Treatment of lupus with corticosteroids. Lupus. (2001) 10:140-7. doi: 10.1191/096120301675075008

129. Tobler A, Meier R, Seitz M, Dewald B, Baggiolini M, Fey M. Glucocorticoids downregulate gene expression of GM-CSF, NAP-1/IL-8, and IL-6, but not of M-CSF in human fibroblasts. *Blood*. (1992) 79:45–51. doi: 10.1182/blood.V79.1.45.45

130. Haynes B, Fauci A. The differential effect of in *vivo* hydrocortisone on the kinetics of subpopulations of human peripheral blood thymus-derived lymphocytes. *J Clin Invest.* (1978) 61:703–7. doi: 10.1172/JCI108982

131. Mathian A, Jouenne R, Chader D, Cohen-Aubart F, Haroche J, Fadlallah J, et al. Regulatory T cell responses to high-dose methylprednisolone in active systemic lupus erythematosus. *PloS One*. (2015) 10:e0143689. doi: 10.1371/journal.pone.0143689

132. Huang H, Lu Z, Jiang C, Liu J, Wang Y, Xu Z. Imbalance between Th17 and regulatory T-Cells in sarcoidosis. *Int J Mol Sci.* (2013) 14:21463–73. doi: 10.3390/ ijms141121463

133. Fauci A, Dale D, Balow J. Glucocorticosteroid therapy: mechanisms of action and clinical considerations. *Ann Internal Med.* (1976) 84:304–15. doi: 10.7326/0003-4819-84-3-304

134. Paliogianni F, Ahuja S, Balow J, Balow J, Boumpas D. Novel mechanism for inhibition of human T cells by glucocorticoids. Glucocorticoids inhibit signal transduction through IL-2 receptor. *J Immunol (Baltimore Md: 1950).* (1993) 151:4081–9. doi: 10.4049/jimmunol.151.8.4081

135. Webster J, Huber R, Hanson R, Collier P, Haws T, Mills J, et al. Dexamethasone and tumor necrosis factor-alpha act together to induce the cellular inhibitor of apoptosis-2 gene and prevent apoptosis in a variety of cell types. *Endocrinology*. (2002) 143:3866–74. doi: 10.1210/en.2002-220188

136. McKinley L, Alcorn J, Peterson A, Dupont R, Kapadia S, Logar A, et al. TH17 cells mediate steroid-resistant airway inflammation and airway hyperresponsiveness in mice. *J Immunol (Baltimore Md: 1950).* (2008) 181:4089–97. doi: 10.4049/jimmunol.181.6.4089

137. Ramesh R, Kozhaya L, McKevitt K, Djuretic I, Carlson T, Quintero M, et al. Pro-inflammatory human Th17 cells selectively express P-glycoprotein and are refractory to glucocorticoids. *J Exp Med.* (2014) 211:89–104. doi: 10.1084/jem.20130301

138. Postolova A, Troxell ML, Wapnir IL, Genovese MC. Methotrexate in the treatment of idiopathic granulomatous mastitis. *J Rheumatol.* (2020) 47:924–7. doi: 10.3899/jrheum.181205

139. Akbulut S, Arikanoglu Z, Senol A, Sogutcu N, Basbug M, Yeniaras E, et al. Is methotrexate an acceptable treatment in the management of idiopathic granulomatous mastitis? *Arch Gynecol Obstet*. (2011) 284:1189–95. doi: 10.1007/s00404-010-1825-2

140. Tekgoz E, Colak S, Cinar M, Yilmaz S. Treatment of idiopathic granulomatous mastitis and factors related with disease recurrence. *Turk J Med Sci.* (2020) 50:1380–6. doi: 10.3906/sag-2003-93

141. Konan A, Kalyoncu U, Dogan I, Kiliç Y, Karakoç D, Akdogan A, et al. Combined long-term steroid and immunosuppressive treatment regimen in granulomatous mastitis. *Breast Care (Basel)*. (2012) 7:297–301. doi: 10.1159/000341388

142. Kafadar MT, Bahadir MV, Girgin S. Low-dose methotrexate use in idiopathic granulomatous mastitis: an alternative treatment method. *Breast Care (Basel).* (2021) 16:402–7. doi: 10.1159/000513879

143. Kehribar DY, Duran TI, Polat AK, Ozgen M. Effectiveness of methotrexate in idiopathic granulomatous mastitis treatment. *Am J Med Sci.* (2020) 360:560–5. doi: 10.1016/j.amjms.2020.05.029

144. Di Xia F, Ly A, Smith GP. Mycophenolate mofetil as a successful therapy for idiopathic granulomatous mastitis. *Dermatol Online J.* (2017) 23:13030/qt51g1k0zm. doi: 10.5070/D3237035736

145. Shahin AA, Khallaf E, Salaheldin LA, Soliman SAM, Rezk YS, Niazy MH. Idiopathic granulomatous mastitis in seventy seven-female patients: Management, follow up of an overlooked immune-mediated disease, and review of literature. *Egyptian Rheumatologist.* (2023) 45:183–9. doi: 10.1016/j.ejr.2023.03.003

146. Wang ST, Lin JC, Li CF, Lee YH. A successful case of etanercept used for idiopathic granulomatous mastitis. *Breast J.* (2019) 25:343-5. doi: 10.1111/tbj.2019.25.issue-2

147. Cadena-Semanate RE, Estrella-Tapia LF, Contreras-Yametti FI, Contreras-Yametti JE, Salazar-Molina RD. Adalimumab in a patient with refractory idiopathic granulomatous mastitis: A case report. *Breast J.* (2021) 27:99–102. doi: 10.1111/tbj.14050

148. Breznik V, Marko PB. A case of overlapping clinical features of idiopathic granulomatous mastitis, hidradenitis suppurativa, and pyoderma gangrenosum

successfully treated with adalimumab. Case Rep Dermatol. (2022) 14:98-106. doi: 10.1159/000523801

149. Engel P, Gómez-Puerta J, Ramos-Casals M, Lozano F, Bosch X. Therapeutic targeting of B cells for rheumatic autoimmune diseases. *Pharmacol Rev.* (2011) 63:127–56. doi: 10.1124/pr.109.002006

150. Pateinakis P, Pyrpasopoulou A. CD20+ B cell depletion in systemic autoimmune diseases: common mechanism of inhibition or disease-specific effect on humoral immunity? *BioMed Res Int.* (2014) 2014:973609. doi: 10.1155/2014/973609

151. Bonilla-Hernán M, Miranda-Carús M, Martin-Mola E. New drugs beyond biologics in rheumatoid arthritis: the kinase inhibitors. *Rheumatol (Oxford England)*. (2011) 50:1542–50. doi: 10.1093/rheumatology/ker192

152. Kerschbaumer A, Smolen J, Nash P, Doerner T, Dougados M, Fleischmann R, et al. Points to consider for the treatment of immune-mediated inflammatory diseases with Janus kinase inhibitors: a systematic literature research. *RMD Open.* (2020) 6: e001374. doi: 10.1136/rmdopen-2020-001374

153. Takeuchi T, Tanaka Y, Tanaka S, Kawakami A, Iwasaki M, Katayama K, et al. Efficacy and safety of peficitinib (ASP015K) in patients with rheumatoid arthritis and an inadequate response to methotrexate: results of a phase III randomised, doubleblind, placebo-controlled trial (RAJ4) in Japan. *Ann rheumatic Dis.* (2019) 78:1305–19. doi: 10.1136/annrheumdis-2019-215164

154. Nakayamada S, Kubo S, Iwata S, Tanaka Y. Recent progress in JAK inhibitors for the treatment of rheumatoid arthritis. *BioDrugs: Clin Immunotherapeut biopharmaceuticals Gene Ther.* (2016) 30:407–19. doi: 10.1007/s40259-016-0190-5

155. Westhovens R, Rigby W, van der Heijde D, Ching D, Stohl W, Kay J, et al. Filgotinib in combination with methotrexate or as monotherapy versus methotrexate monotherapy in patients with active rheumatoid arthritis and limited or no prior exposure to methotrexate: the phase 3, randomised controlled FINCH 3 trial. *Ann rheumatic Dis.* (2021) 80:727–38. doi: 10.1136/annrheumdis-2020-219213

156. Genovese M, Kalunian K, Gottenberg J, Mozaffarian N, Bartok B, Matzkies F, et al. Effect of filgotinib vs placebo on clinical response in patients with moderate to severe rheumatoid arthritis refractory to disease-modifying antirheumatic drug therapy: the FINCH 2 randomized clinical trial *JAMA*. (2019) 322:315–25. doi: 10.1001/jama.2019.9055

157. Aaltonen K, Virkki L, Malmivaara A, Konttinen Y, Nordström D, Blom M. Systematic review and meta-analysis of the efficacy and safety of existing TNF blocking agents in treatment of rheumatoid arthritis. *PloS One*. (2012) 7:e30275. doi: 10.1371/journal.pone.0030275

158. Donahue K, Gartlehner G, Jonas D, Lux L, Thieda P, Jonas B, et al. Systematic review: comparative effectiveness and harms of disease-modifying medications for rheumatoid arthritis. *Ann Internal Med.* (2008) 148:124–34. doi: 10.7326/0003-4819-148-2-200801150-00192

159. Mertens M, Singh J. Anakinra for rheumatoid arthritis. Cochrane Database Systemat Rev. (2009) 1):CD005121. doi: 10.1002/14651858

160. Weisman M. What are the risks of biologic therapy in rheumatoid arthritis? An update on safety. *J Rheumatol Supplement.* (2002) 65:33–8.

161. Genovese M, Cohen S, Moreland L, Lium D, Robbins S, Newmark R, et al. Combination therapy with etanercept and anakinra in the treatment of patients with rheumatoid arthritis who have been treated unsuccessfully with methotrexate. *Arthritis rheumatism.* (2004) 50:1412–9. doi: 10.1002/art.20221

162. Akira S, Taga T, Kishimoto T. Interleukin-6 in biology and medicine. Adv Immunol. (1993) 54:1-78. doi: 10.1016/S0065-2776(08)60532-5

163. Koksal H. The clinical utility of autoantibodies in patients with idiopathic granulomatous mastitis. *J Invest Surg.* (2022) 35:325-9. doi: 10.1080/08941939.2020.1861666