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Involvement of neuronal factors in tumor angiogenesis and the shaping of the cancer microenvironment

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Emerging evidence suggests that nerves within the tumor microenvironment play a crucial role in regulating angiogenesis. Neurotransmitters and neuropeptides released by nerves can interact with nearby blood vessels and tumor cells, influencing their behavior and modulating the angiogenic response. Moreover, nerve-derived signals may activate signaling pathways that enhance the production of pro-angiogenic factors within the tumor microenvironment, further supporting blood vessel growth around tumors. The intricate network of communication between neural constituents and the vascular system accentuates the potential of therapeutically targeting neural-mediated pathways as an innovative strategy to modulate tumor angiogenesis and, consequently, neoplastic proliferation. Hereby, we review studies that evaluate the precise molecular interplay and the potential clinical ramifications of manipulating neural elements for the purpose of anti-angiogenic therapeutics within the scope of cancer treatment.

KEYWORDS

angiogenesis, neurovascular crosstalk, growth factors (angiogenesis factors), tumor microenvironment, tumorigenesis

1 Introduction

The tumor microenvironment plays a critical role in the progression of solid malignancies and will alter the response of cancers to therapeutic approaches such as chemotherapy, radiotherapy, or immunotherapy (1-4). The tumor microenvironment encompasses a complex network of multiple cell types, including fibroblasts, vascular endothelial cells, pericytes, adipocytes, neurons, stem cells, and immune cells (5). Interactions between tumor, stromal cells as well as the extracellular matrix alter the phenotypic properties of tumor cells that can facilitate their dissemination. As such, a better understanding of the underlying cellular and molecular mechanisms governing these interactions could lead to the development of novel therapeutic strategies to fight cancer.

Endothelial cells, as part of the tumor vasculature, play an integral role in defining the properties of the tumor microenvironment. Tumor blood vessels, established primarily through angiogenesis, are required to bring oxygen and nutrients to the tumors, as well as to facilitate tumor cell dissemination. However, the tumor vasculature is often highly dysfunctional and poorly perfused, leading to hypoxia, significantly reshaping the tumor microenvironment (6). As such, vascular normalization, which aims to restore the functionality of the vasculature in solid cancers and restore perfusion, could represent an important strategy to modulate the microenvironment within tumors. A better understanding of the defects associated with pathological tumor angiogenesis would be key to identify novel targets to facilitate vascular normalization.

During development, blood vessels and peripheral nerves form highly branched networks that interact and regulate each other's patterning (7). Indeed, alignment of nerves and blood vessels allows the establishment of a physical relationship between these two systems, and they also respond to related families of guidance cues and growth factors during the establishment of their respective patternings (8). In addition to tumor angiogenesis, recent evidence points to the ability of tumors to recruit neurons (neoneurogenesis) and stimulate innervation (axonogenesis) (9). The presence of neuronal cells has been shown to impact the tumor microenvironment through their interactions with tumor cells, immune cells, and endothelial cells. Furthermore, cancer cells can also infiltrate nerves, a process referred to as perineural invasion (10), demonstrating that nerve fibers in tumors can also act as a mechanical support that enables and facilitates cancer cell migration. Despite the close association of nerves and blood vessels during development, the contribution of neurons to tumor angiogenesis remains unclear, although several studies show that neurons within the tumor microenvironment modulate vascular function in tumors (11). This review summarizes the current state of knowledge regarding the interactions between blood vessels and nerves in homeostasis and cancer, including 1) physiological neurogenesis and angiogenesis and evidence of their interdependence, 2) neurogenesis and angiogenesis in the context of cancer and how they are regulated by one another and 3) the function of neurotransmitters/neuropeptides, axon guidance molecules and neurotrophic factors in neuro-tumoral cooperation, and the distinct involvement of sympathetic and sensory nerves in cancer progression.

2 Physiological and pathological angiogenesis

2.1 Developmental angiogenesis

Angiogenesis, the growth of blood vessels from the existing vasculature, occurs in both physiological and pathological contexts to ensure the transport of oxygen, nutrients, and metabolic waste to tissues. The inner wall of blood vessels is made of endothelial cells that form a monolayer, interconnected by junctional molecules such as VE-cadherin and claudins (12). Endothelial cells interact with a basal membrane and are surrounded by mural cells that prevent vascular leakage by limiting hyperpermeability. The contractility of mural cells further contributes to vessel diameter and the regulation of blood pressure (13).

Embryonic vascular development commences with the formation and aggregation of angioblast precursors within the embryo proper, leading to the creation of blood islands in the visceral yolk sac (14). Subsequently, vasculogenesis occurs, in which blood islands come together to establish a primitive network of endothelial tubes, followed by angiogenesis, which orchestrates the expansion and maturation of vessels through intricate molecular mechanisms, encompassing endothelial cell proliferation and remodeling of the initial vascular network (15). Ultimately, the differentiation of these primitive blood vessels into arteries or veins relies on both intrinsic and extrinsic factors. While vessels may exhibit a genetic predisposition for specific phenotypes, external influences such as haemodynamic forces, which encompass shear stress and blood pressure, can also impact the outcome of blood vessel differentiation (16).

During angiogenic sprouting, Vascular Endothelial Growth Factor (VEGF) stimulates endothelial cells, causing them to become pro-migratory and to adopt a tip cell phenotype (17). Their filopodia and motility increase their responsiveness to the extracellular matrix, growth factors and attractive or repulsive signaling molecules to guide their patterning. Therefore, activated tip cells initiate sprouting whereas following endothelial cells, referred to as stalk cells, proliferate to establish the structural and functional integrity of nascent vessels (17). Several bodies of evidence have shown that the nervous system, through the release of growth factor and axon guidance cues, participates in tip cell guidance and stalk cell elongation during the angiogenic process. For example, it has been shown that several axon-regulating molecules belonging to the Slit/Robo, netrin/DCC/Unc5 and neuropilin/plexin/sema families mediate proper vascular patterning (18).

2.2 Tumor angiogenesis

Tumor blood vessels are dilated, disorganized and immature which, combined with a lack of mural cells association, leads to excessive permeability, poor perfusion and increased hypoxia (19). The important secretion of VEGF by tumor cells 1) suppresses PDGFRβ (platelet-derived growth factor receptor beta) signaling in vascular smooth muscle cells via the complexation of PDGFR β and VEGFR2 that assemble into a hybrid receptor, and 2) modulates the activity of GTPases like RhoA with the effect of increasing vascular permeability (20, 21). The junctional integrity of the vessels is compromised following the release of proteolytic enzymes by tumor cells such as Matrix Metalloproteinases (MMPs), elastase and trypsin that that can accelerate VE-cadherin cleavage together with VEGF signaling (22). The release of inflammatory factors such as histamine further contributes to vessel leakages by increasing the permeability of endothelial cells (23). Given that cancer is characterized by uncontrolled cell growth, the surrounding host tissue restricts tumor expansion which induces

mechanical forces of compression that collapse intratumoral vessels, thus suppressing blood flow and lymphatic network functionality by hindering the drainage of excessive interstitial fluid (24). Blood vessel compression limits perfusion, thereby decreasing oxygen and nutrient concentrations, resulting in a hypoxic microenvironment that selects aggressive clones that activate oncogenes and undergo an epithelial-to-mesenchymal transition, which amplifies the metastatic potential of cancer cells (25). Furthermore, reduced perfusion within tumors is well known to limit the therapeutic efficacy of chemotherapy and radiotherapy treatments. Hypoxia is a major driver of tumor angiogenesis. Under normoxic conditions, prolyl hydroxylase domain protein 2 (PHD2) catalyzes the hydrolysis of hypoxia-inducible factors (HIF) transcription factors with oxygen as the co-factor, which target them for ubiquitination by the Von Hippel-Lindau complex and subsequent degradation by the proteasome (26, 27). Under hypoxic conditions, PHD2 becomes inactive, resulting in HIF1 escaping degradation and binding to hypoxia response elements, a DNA motif present in HIF target genes. Increased levels of HIF proteins lead to the expression of genes involved in cellular adaptation against acute and chronic hypoxia such as angiogenesis (28).

3 The nervous system and angiogenesis

It is well-documented that a strong interplay exists between the neuronal and vascular systems. Individually, nerves and blood vessels maintain normal body function, however, the intricate parallel of anatomic and functional growth between the two has also been deemed necessary for enabling normal function, development, and homeostasis of tissues.

At the anatomical level, arteries and nerves often course alongside each other, exhibiting a high degree of organizational similarity (8).

3.1 Neural activity effects on angiogenesis

There is a strong interdependence between blood vessels and neurons, as neurons have a high metabolic demand, which require blood vessels supplying them with nutrients and oxygen and protecting them from pathogens and toxins. In turn, nerves control blood vessel diameter, heart rate, and other hemodynamic attributes thus linking their functional independence with their proximity.

The modality of the strong interplay between nerves and blood vessels varies slightly between the central nervous system (CNS) and the peripheral nervous system (PNS). In the CNS, multiple different neuronal cell types play a role in the differentiation of blood vessels with a conventional pattern of large brain arteries. While in the PNS, the nerves and blood vessels run in parallel to each other due to a phenomenon named "neurovascular congruency" (29).

Studies have documented the crucial link between neural activity and the vascularization of the nervous system. For example, in the visual cortex, inhibition or promotion of neural activity by modulating visual stimuli significantly modulated vascular density (30, 31). Experiments in the mouse primary somatosensory cortex have also demonstrated that somatosensory stimuli are critical in shaping cortical blood vessels (32). These data support the hypothesis that neural activity driven by sensory experiences can shape the development of the vasculature. Accordingly, reduced glutamatergic neurotransmitter release at the thalamocortical synapses in transgenic mice, leading to diminished neuronal activity, also led to decreased vessel density and branch points, suggesting that altered activity along the somatosensory axis results in changes in cortical angiogenesis (33).

Several neurotransmitters have been shown to play a significant role during physiological angiogenesis. Dopamine release has been shown to significantly delay angiogenesis during wound healing, and D2 dopamine receptor antagonists significantly accelerated wound healing in mice by inducing angiogenesis, in part through its upregulation of α 5 β 1 integrin (34). Dopamine was also shown to inhibit vascular development in the retina by restricting VEGFR activation and Notch-Jagged1 signaling (35). GABA and its receptor signaling have also been shown to shape neurovascular interactions and angiogenesis during embryonic brain development. Endothelial cell-specific conditional mouse models of the GABA pathway (Gabrb3ATie2-Cre) resulted in a significant reduction in the vascular density of the embryonic brain which persisted in the adult brain, accompanied by concurrent GABAergic neuronal cell deficits (36). Mice lacking serotonin in the nervous system (Tph2-/-), while not showing obvious angiogenic defects, also displayed altered cardiovascular parameters, including decreased blood pressure, due to diminished sympathetic nerve activity (37). Nicotine, an agonist of the acetylcholine receptor (AChR), was shown to enhance physiological angiogenesis, and improve wound healing or limb ischemia (38, 39). The nicotinic acetylcholine receptor α 3 subtype (α 3-nAChR) was also shown to modulate vascular inflammation by restricting inflammatory cells infiltration and the release of inflammatory cytokines (40). Finally, metabotropic glutamate receptors (mGluR) were also shown to play important roles in the maintenance of vascular integrity. Treatment of human brain endothelial cells with glutamate or selective mGluR group I or III agonists resulted in a time-dependent loss of phosphorylated vasodilator-stimulated phosphoprotein (VASP) and significantly increased endothelial permeability. In vivo, mGluR antagonists significantly decreased blood-brain barrier function (41). Altogether, these studies highlight the multifaceted roles of neurotransmitters in vascular development and function.

3.2 Axon guidance molecules and angiogenesis

At the cellular level, endothelial tip cells and axonal growth cones display striking similarities in both their morphological structures and functions: endothelial tip cells play a crucial role in guiding the development of blood vessels, while growth cones are found at the leading edges of growing axons (42). At the molecular level, endothelial tip cells and axonal growth cones have also been shown to respond to similar families of growth factors and comparable guidance cues (Figure 1) (18).

For example, VEGF, aside from its role in vascular development, has also been shown to participate in the development of the CNS by promoting axonal and dendrite growth *in vivo* and *in vitro* (43–45). In the PNS, VEGF was shown to induce axonal growth in dorsal root ganglia by attracting growth cone (46) and was also shown to promote Schwann cell proliferation and migration through VEGFR2 (47). Several other neural guidance factors and receptors, which were first identified in the patterning of the nervous system, were also found to play a role in the establishment of the vasculature. These include the Semaphorins, Slits, Eph family, and Netrins (18).

Semaphorins/plexins/neuropilins: Semaphorins were initially identified as axonal cone guidance molecules. They have however since been described to have numerous functions in various organs in the body with a variety of different functions. Semaphorins are ligand molecules that may be subclassified into different groups (from classes 1-8) (48). Several of the Semaphorins' effects, in particular their effects on the nervous system and blood vessel formation are mediated by their receptors, the Plexins and Neuropilins (49). Plexins, which were first identified on the surface of axon growth cones of dorsal root ganglia (DRG), cortical, sensory, motor and sympathetic neurons and play a role in signal transduction to steer axon growth away from the source of semaphorin, were also shown to be involved in several processes such as angiogenesis, immunoregulation, and neuronal connectivity (50).

The Semaphorins-Plexin complex plays a crucial role in blood vessel pathfinding. The main classes of semaphorins that have been shown to modulate the patterning of neurons as well as lymphatic and blood vessels are Sema3A, Sema3E, and Sema6D (51–57).

Although they share a similar mechanism of action, the effects of these ligands on blood vessels and neuronal development vary substantially on account of their pattern of expression.

Loss of Plexin D1 function was shown to cause dramatic mispatterning of developing intersegmental vessels in zebrafish, which are no longer restricted to growth near intersomitic boundaries. Conversely, semaphorin overexpression inhibits the growth of intersegmental vessels in a Plexin D1-dependent manner (58). Semaphorins also interact with neuropilins, transmembrane glycoproteins that regulate neurogenesis and angiogenesis by complexing with Plexin receptors/class-3 semaphorin ligands and VEGF receptors. Neuropilin-1 and Neuropilin-2 have been shown to play crucial roles in blood and lymphatic vascular development by regulating VEGF-A and VEGF-C signaling respectively (59–61).

Sema3A, which acts to halt the extension of the axons by collapsing the growing tip or growth cones of axons, has been shown to have a role in the organization of blood vessels alongside peripheral neuronal growth *in vitro*. The fetal skin of Sema3A-deficient mice shows a disorganized peripheral vasculature running parallel to a disorganized PNS, further indicating that the peripheral nerve branching may serve as a template for the development of the developing arterial system (62).

Sema3E is highly expressed in the somites during development, where it acts as a repulsive cue for plexin-D1–expressing endothelial cells of adjacent intersomitic vessels (63). Studies have also demonstrated Sema3E's regulatory effect on VEGF-induced Dll4-Notch signaling. A study conducted on a model of the retinal vasculature of mice concluded that VEGF directly controlled the expression of Plexin-D1, and in turn, the VEGF-induced Delta-like



4-notch signaling pathway was negatively inhibited by the Sema3E-Plexin-D1 complex. This study also found that although Sema3E remains relatively constant throughout the environment of the retina, the expression of Plexin-D1 is controlled by VEGF both temporally and spatially and that there is a role of Sema3E–Plexin-D1 function in modulating angiogenesis via a VEGF-induced feedback mechanism (55).

A knockout model of Sema6D in zebrafish demonstrated a disrupted pattern of intersegmental vessels and primary motor neurons indicating a necessary role of Sema6D in neuronal and endothelial guidance (64). Furthermore, a 2016 study using samples of human gastric cancer tissues has also shown an increased prevalence of Sema6D and Plexin A1 in endothelial vascular cells and an association of Sema6D and plexin-A1 with VEGFR2. The study provided evidence of an *in-vitro* role in angiogenesis and an increased possibility of tumor angiogenesis by VEGFR2 regulation (56). A study published by Toyofuku et al. showed a proangiogenic potential of Sema6D in conjunction with its receptor, Plexin-A1, due to its ability to aid in ventricular endocardial cell migration via the aid of VEGFR2 (65).

Ephrin/Eph: Ephrin ligands are regulators of neuronal and vascular development, which are classified into two groups, Ephrin As and Bs (66). Ephrins and their corresponding receptors, called Eph receptors, are involved in cell communication and guidance through bidirectional signaling (67). The role of ephrins in angiogenesis is complex and involves both stimulatory and inhibitory effects, depending on the context and specific types of ephrins and Eph receptors, which can be subclassified into two classes: EphA (EphA1-EphA8) and EphB (EphB1-Eph4 & EphB6). In the nervous system, ephrins can act as short-range chemoattractants and repellents, and are important in regulating the branching of axons, dendritic morphology, and synapse formation in the developing CNS (68, 69).

In the vascular system, EphrinB2 and EphB4 are expressed in arterial and venous endothelial cells respectively, where they play a crucial role in arteriovenous specification. Targeted inactivation of the genes encoding ephrin-B2 and EphB4 in mice has shown that both are crucial for angiogenic remodeling and embryonic survival (70, 71). Ephrins are involved in the application of hemodynamic factors to the growth of the lumen as well as for the recruitment of mural cells (72). It is also known that EphrinB2 is implicated in VEGFR3 and VEGFR2 internalization processes and therefore participates in the regulation of lymphangiogenesis and angiogenesis (73).

ROBO/Slit: Slits and their receptors, Robo, were initially recognized as factors that attract and repel neurons across the midline aiding in neuronal axon pathfinding, branching, and migration. Indeed, it was demonstrated that specific combinations of Robo receptors in developing axons of the vertebrate spinal cord determine the final destination of these axons after crossing the midline (74). Slits and Robos have also been found to have significant roles in the vascular system. Slit2 and Slit3 are secreted by vascular smooth muscle cells, endothelial cells, and perivascular cells while Robo4, Robo1 and Robo2 have been reported to be expressed in endothelial cells (75).

It was shown that Slit2, acting through Robo1 and Robo2, promotes the migration of endothelial cells by regulating VEGFinduced Rac1 activation and lamellipodia formation (76). Robo4, an endothelial-specific isoform of the Robo family, has also been shown to promote vessel stabilization and prevent permeability through its interaction with Unc5B (77). Other studies have also shown that it can stimulate filopodia formation, cell migration, and angiogenesis suggesting that its biological effects may be context-dependent (78). It was found that the effects of Slit/Robo signaling on angiogenesis were tissue and ligand-dependent. Using Slit2/Slit3 KO mice, it was observed that stromal rather than epithelial Slit inhibit vascular growth by downregulating VEGF via Robo4, and the loss of Slit in the stroma led to an increase in blood vessel vascularity and blood vessel complexity in the mammary gland (79).

SHH: Sonic hedgehog (Shh), which is involved in embryonic morphogenesis, plays a crucial role in the developing nervous system, with multiple functions including the regulation of motor neuron development in the spinal cord, patterning of the ventral spinal cord, guidance of retinal ganglion cell neurons and olfactory sensory neurons, as well as the migration of commissural axons (80). Shh is secreted as a monomer, multimer, or in exovesicles, and exerts its cellular function through its interaction with the transmembrane receptor Patched 1. In doing so, it unleashes Smoothened from its inhibition, which, once released, transduces the Shh signal resulting in the activation of the zinc-finger transcription factors of the Gli family (81).

Shh-induced guidance of commissural axons is a complex process, involving cross-talk with the Wnt signaling pathways. Research findings have indicated that Shh has a function in attracting commissural axons toward the ventral midline during spinal cord development. Notably, Shh exhibits a dual effect on retinal ganglion cell axons: it attracts these axons at lower concentrations and repels them at higher concentrations (82). The importance of Shh in vascular regulation has been made clear across multiple studies which demonstrated its actions on arterial differentiation, vessel branching, and mural recruitment (83). Shh was shown to be a target of PDGF-BB signaling smooth muscle cells, where it is involved in cell migration, and its inhibition in vivo reduced the recruitment of NG2-positive cells to neovessels (83). In one such study, Shh-null mutants displayed an absence of vascular branching in the lungs during development (84). Shh was also shown to indirectly modulate angiogenesis through stromal cells in the perivascular niche, prompting them to express and release pro-angiogenic factors. These factors then operate within the vasculature, instigating the creation of new blood vessels that aid in enhancing the healing processes (85). Noncanonical Shh signaling, which operates independently of the transcriptional changes mediated by Gli transcription factors has also been shown to promote the formation of blood vessels through its effects on the cytoskeleton of endothelial cells by regulating PI3kinase (86), as well as the small monomeric GTPases RhoA and Rac1 (87). These studies highlight the crucial role that the Shh pathway plays in angiogenesis.

Netrins: Netrins are proteins that guide migrating cells and axonal growth cones (88). Netrins have been found to exert their guidance activities via their action on the DCC (Deleted in

Colorectal cancer) and Unc5 receptors (89). Netrins are involved in midline-crossing of commissural axons of the spinal cord through DCC. Netrin-1 has been shown to act both as a chemoattractant for ventrally directed commissural axons, and as chemorepellent for trochlear motor neurons (90, 91). In the PNS, Netrins have been demonstrated to play a role in guiding motor neurons to their target muscles. This was evidenced when mice lacking Netrin exhibited abnormalities in the projection of motor axons (92).

In blood vessels, inhibition of Netrin-1 binding to its receptor DCC suggested that Netrin-1 produced by arterial smooth muscle cells attract DCC-positive sympathetic axons toward the arterial wall (93). A 50% reduction of the expression of expression of Ntn-1 was sufficient to severely impair arterial innervation in small diameter resistance arteries branching off the iliac and femoral artery as well as arteries in the skin and internal organs, such as the esophagus and mesentery. On the other hand, overexpression of netrin-1 in the peritoneal cavity has also been shown to promote the ectopic extension of sympathetic axons away from the arteries, and this process could be blocked by blocking the netrin-1 receptor DCC (93).

Among the receptors for Netrins, the Unc5b receptor has been found to be selectively expressed in arteries and developing capillaries (94). This receptor, which has been reported to have a repulsive effect on endothelial cells, plays a role in controlling the morphogenesis of the vascular system, contributing to both normal developmental angiogenesis and pathological angiogenesis. A recent study also found that it regulates the integrity of the blood brain barrier by modulating Wnt/β-catenin signaling in brain endothelial cells (95). While some reports demonstrate that Netrin-1 negatively regulates capillary branching in the developing vascular system and to reduce endothelial cell migration in vitro (94, 96), other researchers highlight its ability to positively modulate angiogenesis (97, 98). Netrin-1 was shown to promote NO production which then mediates Netrin-1-induced enhancement in endothelial cell growth and migration (99). Netrin-1 also promotes the survival of endothelial cells upon binding to UNC5B, which has been reported to promote apoptosis in the absence of ligand stimulation (100). The multifaceted role of Netrins in angiogenesis may be dependent on their localization as well as the expression of their receptors.

4 The nervous tissue within the tumor microenvironment

Several cellular and soluble components within the microenvironment influence tumor progression by contributing to the formation of new blood and lymphatic vessels (101). Similar to the processes of angiogenesis and lymphangiogenesis, evidence point out to the neoformation of nerve endings inside the tumors, termed neoneurogenesis (102). Nerve endings within tumors have been reported in several cancers such as skin, eye, heart, bladder, prostate, breast, pancreatic, esophageal, melanoma and colon cancers (103–109). Nerves fibers can facilitate tumorigenesis trough their effects on cancer cells directly or by modulating the microenvironment of tumors. It has also been suggested that the

nervous system is functionally involved via the modulation of mediators of tumor progression (102). Increased neurogenesis has been associated with poor patient outcome, enhanced metastatic burden, and higher tumor grade in several models, including colorectal cancer (110), breast cancer (111, 112), pancreatic ductal adenocarcinoma (113), and thyroid cancer (114).The cross-talk between cancer cells and nerve fibers illustrates a synergistic epithelial-neural interaction in which both components secrete factors that favor their rapid growth. In this section, we describe several cellular and molecular processes by which neuronal cells can impact tumor progression and angiogenesis.

4.1 Perineural invasion (PNI)

Nerve fibers in tumors have been described to fulfill a structural purpose, acting as paths that enable the migration of perineural invading cells. This neural remodeling, which involves structural and functional changes in surrounding nerves within the tumor microenvironment results in increased nerve density, abnormal nerve branching, and altered nerve function, which in turn can create a supportive environment for tumor growth and can create a permissive environment for tumor cells to invade nerves and travel to distant sites, contributing to metastasis. Thus, it is likely that the perineural space is enriched in molecules that attract cancer cells and initiate a process called perineural invasion, which was defined as a coverage of at least 33% of the circumference of a nerve by a tumor (115). Research showed that pancreatic and prostate cancer cells are more proliferative and less susceptible to apoptosis when located near a nerve space (116, 117). The underlying mechanism could be the neoformation of axons via PNI based on the evidence that cancer cells secrete neurotrophic factors like NGF and axon guidance molecules like netrin-1 (118, 119). The incidence of PNI thus correlates positively with a poorer clinical outcome. In rectal cancer, the extent of PNI is even used as a prognostic factor (120).

Neoneurogenesis: Besides the ability of cancer cells to invade peripheral nerves, it has been proposed that cancer cells stimulate their own innervations. Therefore, the concept of neoneurogenesis also includes the development of nerve endings (axons) towards the tumor. This is supported by the fact that the shaping of the nervous system depends on a mechanism of extracellular matrix components assisted chemotactic axon guidance (121-123). A coculture system with DRGs from the lumbar spinal cord of mouse embryos at 17 weeks of gestation and cancer cells was successfully used by Ayala and colleagues to 1) estimate neurite (or any projection from the cell body of a neuron) outgrowth, 2) neurite directionality towards cancer cells, 3) the existence of nerve-cancer cell interaction, 4) migration of cancer cells towards and through the nerves (PNI) and 5) the growth rate of cancer cells. They demonstrated that neurites extended from the DRGs to tumors in vitro in human pancreatic and prostate cancer (116, 117, 124). Although the stimulation of neoneurogenesis by tumor-released soluble factors has not yet been demonstrated in human cancers, the mechanism employed by cancer cells to induce neurite outgrowth could probably be similar to the process of nerve recovery after injury: unlike the CNS, the PNS can support long-axon

regeneration after injury. In the adult PNS, Schwann cells or axon insulating cells, secrete growth factors that enable axonal regeneration, such as CNTF (ciliary neurotrophic factor), NGF, BDGF (brain-derived growth factor) and IGF (insulin-like growth factor) (125). Moreover, neurons require the activation of different signaling pathways by cancer cells to promote axonogenesis: GSK3 for microtubule stability and remodeling, PI3K for cell polarity, motility and chemotaxis and Akt for neuronal polarity (126). It has been revealed that growth factors and soluble factors that activate the aforementioned pathways, secreted by Schwann cells during axon regeneration, may also be secreted by cancer cells. Consequently, it is speculated that within the tumor microenvironment, axonogenesis could be under the control of cancer-derived signaling pathways similar to injury-induced axonal regeneration. Furthermore, cancer cells can generate their own extracellular matrix that favors axonal growth (124).

4.2 Neuroinflammation

Tumors can also trigger an inflammatory response in nerves, leading to the release of inflammatory mediators by the latter. These mediators, which include neuropeptides such as substance P and calcitonin gene-related peptide (CGRP), can promote tumor angiogenesis and increase the permeability of blood vessels, facilitating tumor growth and metastasis (127). The release of these neuropeptides and other inflammatory mediators can recruit also immune cells, such as macrophages and neutrophils, to the tumor microenvironment, which, in turn, release additional proinflammatory factors, growth factors, and cytokines that can stimulate tumor cell proliferation and survival. Neuropeptides released during neurogenic inflammation can also influence immune responses within the tumor microenvironment. Substance P and CGRP have been shown to enhance the recruitment and activation of immune cells, including T lymphocytes and dendritic cells (128). This modulation of immune responses can have both proand anti-tumorigenic effects, depending on the specific context.

Neurogenic inflammation mediators can also influence pain perception in cancer patients, as substance P and CGRP, released during neurogenic inflammation, are known modulators of pain sensations (128, 129). Additionally, neurogenic inflammation can impact the response to cancer treatments, such as radiation therapy or chemotherapy, by affecting the sensitivity of tumor cells to these therapies (130).

4.3 Neuronal-derived factors and tumor angiogenesis

Within tumors, nerve cells release several neurotrophic factors that can modulate the proliferation and phenotype of cancer cells, including Nerve Growth Factor (NGF), Neurotrophins (NT), Brain-Derived Neurotrophic Factor (BDNF) or Glial Cell Line-Derived Neurotrophic Factor (GDNF). In addition, axon guidance molecules and their receptors are also known to be related with tumor progression (102). Moreover, the literature shows that several neurotransmitters (epinephrine, norepinephrine, dopamine, serotonine, acetylcholine, glutamate, gammaaminobutyric acid) and neuropeptides (neuropeptide Y, neurohypophyseal, corticotropin-releasing factor, opioid, secretin, somatostatin, tachykinin) are similarly involved in tumor progression, mainly by stimulating the proliferation or migration ability of cancer cells, a key event in metastasis (131). Although by themselves, these factors may not be critical for cancer cell survival, they were shown to facilitate tumor growth in the context of chemotherapy. Given that neurotransmitters and neuropeptides can activate signaling pathways pertaining to cell proliferation and survival, such as the PI3K, MAPK and Akt pathways, their inhibition should constitute a relevant chemo- or targeted therapy sensitization strategy (132). Table 1 lists some of the reported effect of factors released by nerves within tumors and their reported effects on tumorigenesis. The effects of neuronal-derived factors and axon guidance molecules on the tumor microenvironment and angiogenesis are further discussed in the following sub-sections.

4.4 Growth factors

Neurotrophic factors such as NGF and BDNF are known to play roles in tumor angiogenesis in various cancers, including breast cancer and chondrosarcoma. NGF regulates growth, differentiation and survival of peripheral neurons during embryonic development (168). In addition to its well-described action on survival and differentiation of neurons, NGF also acts as a pleiotropic molecule, involved in a wide variety of functions, such as neuropeptide modulation, wound healing and tissue scar. Endothelial cells have been shown to express the NGF receptors TrkA and p75 (169). Stimulation by NGF activates the Ras/ERK and PI3K/Akt signaling cascades in endothelial cells, both of which are involved in controlling cell proliferation and survival and are required for induction of endothelial cell migration (170).

NGF is upregulated in the tumor microenvironment of breast cancer. MDA-MB-231 breast cancer cells secrete NGF, which has been shown to stimulate angiogenesis *in vivo* when injected into immunodeficient mice (171). Moreover, NGF promotes the secretion of VEGF by breast cancer cells, and the angiogenic capacity of NGF is inhibited by *in vivo* administration of anti-VEGF antibodies. In human glioma microvascular endothelial cells, NGF plays a role in tumor angiogenesis by interacting with the α 9 β 1 integrin (172).

Conversely, it was also shown that NGF can also inhibit the growth of HT1080 fibrosarcomas or HepG2 tumors via its effects on innervation and maturation of tumor neovasculature, which regulates blood flow into tumor tissues. Indeed, NGF markedly increased the density of α -smooth muscle actin (α -SMA)-positive cells in the vasculature of HT1080 tumors and significantly reduced blood flow in HepG2 tumors (173). A tumor-suppressive effect of NGF was also observed in prostate tumors by facilitating maturation of tumor blood vessels by migrating smooth muscle cells, and regulating tumor tissue blood flow (174).

TABLE 1 Neurotrophic factors and their effects on tumor progression.

Neurotrophic Factor	Reported Effect	References
Nerve Growth Factor (NGF)	 NGF and its receptor TrkA are overexpressed in various types of cancer, including breast, ovarian, and prostate cancer. Elevated NGF levels are associated with increased tumor growth, angiogenesis, and metastasis. 	(133–140)
Brain-Derived Neurotrophic Factor (BDNF)	 Involved in the progression of several cancers, such as breast, colorectal, and lung cancer. Increased BDNF expression has been associated with tumor growth, invasion, and resistance to chemotherapy. 	(141-145)
Glial Cell Line- Derived Neurotrophic Factor (GDNF)	 Implicated in tumorigenesis; promotes the growth and metastasis of various cancers, including neuroblastoma and pancreatic cancer. GDNF can stimulate angiogenesis and enhance the invasive properties of cancer cells. 	(146-149)
Netrins/Unc5/DCC	 Netrin-1 and Netrin-4 delay the growth of lung, pancreatic and prostate tumors. Netrin-1 promotes the growth of glioma cells by activating NF-κB signaling via UNC5A. Netrin-1 is a positive regulator of malignant tumor metastasis by activating YAP signaling. Netrin-1 induces antiapoptotic effects of acute myeloid leukemia cells through Unc5B. Loss of netrin-1 receptor DCC is associated with a poor prognosis in patients with colorectal tumors, glioblastoma, and breast carcinoma. Netrin-1 stimulates tumor progression in breast, cancer and medulloblastoma 	(96, 97, 150–155)
Semaphorins/ Neuropilins/Plexins	 High expression of Sema3F correlates with poor prognosis and tumor immune infiltration of hepatocellular carcinoma. In breast cancer, Sema3A is a tumor suppressor, downregulated in tumor and negatively correlated with tumor stage. In Lewis lung cancer, Sema3A binding to Nrp1 and PlexinA1/ PlexinA4 coreceptors promotes tumor growth. Sema4C is overexpressed in multiple types of malignant tumors, including breast cancer, esophageal cancer. Sema7A is involved in macrophage-mediated lymphangiogenesis in breast cancer. Sema3A and Sema3B contribute to 	(156–161)

(Continued)

TABLE 1 Continued

Neurotrophic Factor	Reported Effect	References
	immune escape from the anti-tumor effects of CD8+ T cells.	
Slit/ROBO	 Robo1 and Robo4 are significantly upregulated in colorectal carcinoma. Slit2 and Robo1 are associated with an increased metastatic risk and decreased survival in colorectal carcinoma patients. Increased Slit1 expression in prostate tumors Slit/Robo signaling pathway plays a role in enhancing tumor cell migration and promotes tumor metastasis. Slit2/Robo1 signaling upregulates MMP-9 to enhance breast cancer cell invasion. Slit/Robo signaling decreases the proliferative rate and increases the apoptotic rate of oral squamous cells 	(162–167)

BDNF is a neurotrophin essential for neuronal development and survival, synaptic plasticity, and cognitive function (175). BDNF acts on specific subtypes of TrkB-expressing neurons in the CNS and the periphery, and support the survival of existing neurons, as well as the growth and differentiation of new neurons and synapses. Studies have also shown that BDNF promotes angiogenesis by activating TrkB-expressing endothelial cells. The pro-angiogenic effects of BDNF were shown to be mediated via the PI3-kinase-Akt pathway, and modulation of BDNF levels directly correlate with Akt phosphorylation and inhibitors of PI3-kinase abrogate the BDNF responses in endothelial cells (176). BDNF can also indirectly induce angiogenic responses by regulating the expression of VEGF during tissue healing (177). BDNF was also shown to promote angiogenesis through the recruitment of Sca-1 +CD11b+ pro-angiogenic hematopoietic cells which can facilitate neovascularization (178).

BDNF has been shown to contribute to tumor angiogenesis. In chondrosarcoma patients, the expression of BDNF and VEGF proteins is significantly higher and correlated with tumor stage. Knockdown of BDNF leads to decreased VEGF expression and abolishes angiogenesis in in vitro studies and animal models of chondrosarcoma (179). Co-transplantation experiments of BDNFexpressing endothelial cells with transformed mouse liver cells showed that high BDNF-expressing endothelial cells could facilitate tumor angiogenesis and growth of hepatocellular carcinomas whereas knockdown of BDNF by short hairpin RNAs impaired such effects (141). Immunohistochemical staining of human hepatocellular carcinomas revealed upregulation of BDNF and TrkB protein levels in both tumor and endothelial cells. High TrkB expression was associated with shorter overall survival. Interestingly, BDNF was also shown to regulate tumor lymphangiogenesis by modulating the expression of VEGF-C in human chondrosarcoma tissues (180). A positive correlation

between TrkB expression and lymph vessel density was also observed in ovarian cancer (181) and is associated with the expression of VEGF-C and VEGF-D in oral squamous cell carcinoma (182).

4.5 Axon guidance molecules

Axon guidance molecules, including Semaphorins, Ephrins, Slits and their respective receptors Neuropilins, Eph and Robo receptors, have emerged as significant regulators of tumor angiogenesis. These molecules, which regulate axonal pathfinding during neural development, exhibit parallels in their ability to influence blood vessel growth and alignment within tumors (18).

Semaphorins and their receptors, often overexpressed in cancer cells, have been implicated in promoting angiogenic sprouting and guiding blood vessel trajectories. Neuropilin-1, a co-receptor for both Semaphorins and VEGF, stands out as a mediator of angiogenic signaling pathways. Several studies have shown a positive correlation between neuropilin-1 expression and exacerbated angiogenesis and poor prognosis in prostate, colorectal, kidney, lung and breast cancers (183). Neuropilin-1 expression in tumor endothelial cells, through its interaction with VEGF stimulates angiogenesis by enhancing VEGF signaling. It was shown that antibodies blocking VEGF binding to neuropilin-1 enhanced the anti-tumor effect of anti-VEGF antibodies (184). EG00229, an inhibitor of neuropilin-1 has also been shown to possess significant tumor-suppressive effects in gliomas and squamous cell carcinomas (185, 186). Furthermore, Sema3A selective mutants, which can competitively interact with PlexinA4 and prevent neuropilin-1/PlexinA4 interactions have also been shown to accelerate vascular normalization and reduce tissue hypoxia, increase perfusion and inhibit tumor growth in a mouse model of pancreatic cancer (187). Interestingly, in non-small cell lung cancer (NSCLC), there was an observed correlation between Sema4D expression and the development of perfused channels by tumor cells (188). This process, known as vascular mimicry (VM), allows tumors to create matrix-embedded vascular structures containing plasma and blood cells to fulfill the nutrient and metabolic requirements of neoplastic tissues (189, 190). Notably, this occurs without the involvement of pre-existing host endothelial cells. The resulting vascular-like structures do not constitute true blood vessels but rather imitate their function. Xia and colleagues demonstrated that heightened Sema4D expression was associated with an increased presence of VM channels in NSCLC tumors (188). The impact of Sema4D on VM was mediated through the RhoA/ROCK pathway, which governs tumor cell plasticity and migration. Consequently, inhibiting the expression of plexinB1 through RNAi-expressing vectors or blocking the RhoA/ROCK signaling pathway proved effective in reducing VM formation by NSCLC tumor cells (188).

Ephrins and Eph receptors, known for their role in repulsive axonal guidance, have been found to regulate endothelial cell migration and vessel alignment in tumor vasculature. The dual nature of Ephrin-Eph interactions, capable of either promoting or inhibiting angiogenesis depending on context, underscores their complexity in tumor microenvironments. Hypoxia and VEGF have been shown to cause the upregulation of ephrin-A1 in endothelial cells (191). Ephrin-B2 was also found to be uprregulated by VEGF and hypoxia. This was accompanied by increased tumor angiogenesis and stimulation of tumor growth (192). EphA2 forward signaling can also stimulate angiogenesis and it is known as a poor prognostic factor in endometrial cancer (193). Additionally, expression of Eph and ephrins has also been observed within lymphatic vessels, suggesting that it could regulate lymphangiogenesis (194). Therapeutically, it was shown that inhibition of Eph signaling using EphA2 and EphA3 Fc-fusion proteins could inhibit cancer progression and neoangiogenesis (195). Ephrin B1-derived peptides were also shown to suppress gastric cancer dissemination (196).

Netrins have also been implicated in tumor angiogenesis. It was shown that Netrin-1 binding to the Unc5B receptor on tumor endothelial cells limited tumor angiogenesis in models of lung, prostate and pancreatic cancer (96). Netrin-4, which was found to be overexpressed in VEGF-stimulated endothelial cells also negatively regulated angiogenesis by binding to neogenin, leading to the recruitment of Unc5B in PC3 xenograft tumors (150). Netrin-4 was also shown to delay colorectal cancer progression by inhibiting tumor angiogenesis (153). Conversely, netrins were also shown to possess pro-angiogenic activity in cancers. In tumors associated with the CNS, such as retinoblastoma and glioblastoma, netrin-1 did promote neovascularization (197). In patients with medulloblastoma, Netrin-1 also stimulated cell invasiveness and angiogenesis (198).

4.6 Neurotransmitter release

Neurotransmitters in the tumor microenvironment exert significant effects on angiogenesis. These neurotransmitters and stress hormones can either promote or limit neovascularization through several mechanisms such as the regulation of expression of various angiogenic factors such as VEGF or metalloproteases (MMPs) or by modulating VEGF-induced endothelial cell proliferation, migration, and vascular permeability (Figure 2). The intricate interplay between neurotransmitters and angiogenesis suggests potential therapeutic opportunities for targeting the neuronal regulation of blood vessel growth in cancer treatment.

4.7 Dopamine

Dopamine exhibits antiangiogenic properties by blocking VEGFinduced endothelial cell proliferation, migration, and vascular permeability. The antiangiogenic properties of dopamine are mediated by the D2 dopamine receptors (D2R), leading to VEGF receptor 2 endocytosis, thereby preventing VEGF binding, receptor phosphorylation and subsequent signaling steps (199, 200). Studies have shown that dopamine significantly limits tumor neovascularization in several animal models of stomach, breast, and colon cancers (201–203). Depletion of peripheral dopaminergic nerves in mouse tumor models increased neovascularization and endothelial



permeability (204), while treatments with exogenous dopamine inhibited tumor growth and angiogenesis (201). D2R agonists have also been found to abrogate lung tumor progression in syngeneic (Lewis Lung Carcinoma; LLC) and human xenograft (A549) orthotopic murine models and reduce tumor-infiltrating myeloidderived suppressor cells (205). D2R knockout mice also display increased tumor growth due to exacerbated neovascularization (204). A positive correlation between endothelial D2R expression and tumor stage was also detected in lung cancer biopsies (205).

The dopaminergic system also plays a role in preventing vasculogenesis. Depletion of dopamine in the bone marrow leads to increased mobilization of endothelial progenitor cells (EPC) to sarcoma 180 (S180) tumors (206). Conversely, exogenous dopamine decreased VEGF-induced EPC mobilization, partly by reducing MMP-9 expression in the bone marrow.

4.8 Acetylcholine and nicotine

Acetylcholine receptors (AChRs) are expressed in various tissues, including the CNS and PNS, muscle, and non-neural tissues. At the neuromuscular junction, they are the primary receptors responsible for motor nerve-muscle communication, controlling muscle contraction (207). The main AChRs expressed in non-neuronal tissues include homomeric α 7 AChRs as well as several heteromeric AChRs composed of α 3, α 5, β 2, and β 4 subunits, including α 3 β 4-containing AChRs. Endothelial cells have been shown to express all mammalian AChR subunits with the exception of α 2, α 4, γ , and δ (208). AChR signaling, triggered by endogenous agonists like acetylcholine or exogenous chemicals like nicotine, plays a role in promoting tumor angiogenesis. Stimulation of α 7 AChRs can activate integrin $\alpha\nu\beta$ 3 and intracellular ERK MAP kinase signaling, leading to

endothelial cell proliferation (209). This stimulation also induces the expression of various angiogenic factors, such as VEGF, PDGF, FGF, or TGF- β . Nicotine has also been shown to upregulate MMP-2 and -9 in endothelial cells, facilitating angiogenic sprouting (210).

In experimental and animal models of cancer, nicotine promotes angiogenesis and accelerates tumor growth. For instance, in mice with colon cancer tumors (HT-29), nicotine leads to elevated VEGF expression and increased microvascular density (211). Additionally, nicotine induces angiogenesis in lung cancer, where its systemic administration to mice bearing LLC tumors enhances tumor growth by accelerating tumor angiogenesis (38). Nicotine also promotes angiogenesis in gastric tumors through the upregulation of COX2 and VEGFR2 (212). In addition, nicotine also accelerated the growth of syngenic colon CMT93 tumor cells by inducing angiogenesis through the recruitment of bone marrow derived EPCs within the tumor (213). Selective antagonists of α 7 AChRs, such as α -bungarotoxin and methyllycaconitine,have also been shown to inhibit endothelial cell proliferation, thereby limiting the angiogenic response (209) (214),. Isopropyl methylphosphonofluoridate, an irreversible inhibitor of acetylcholinesterase, the enzyme which catalyzes the breakdown of acetylcholine, also increased the number of newly forming blood vessels in L-1 sarcoma. This effect was mediated by the regulation of lymphocyte-induced angiogenesis and the modulation of angiogenic and pro-inflammatory cytokines secretion (215).

4.9 Y-aminobutyric acid (GABA)

Endothelial cells express various subunits of GABA_A receptors, including $\alpha 1$, $\alpha 2$, $\alpha 6$, $\beta 1$, $\beta 2$, $\beta 3$, $\gamma 1$, $\gamma 2$, and $\gamma 3$ subunits (216), where they participate to cerebral angiogenesis and neurogenesis (36). GABA_A signaling induces an inward Cl– current, a transient

increase of calcium in endothelial cells and lead to the recruitment of endothelial nitric oxide synthase (eNOS) to produce NO, contributing to endothelial cell proliferation and angiogenesis (217). However, GABA_A receptor over-expression has also been shown to inhibit VEGF-induced endothelial cell proliferation, migration, and tube formation, as well as VEGFR-2 phosphorylation *in vitro*, and to reduce the phosphorylation of downstream PI3K components, such as PDK1, Akt, mTOR, TSC-2, p70S6K, and 4E-BP1 by directly binding with VEGFR-2 (218).

GABA stimulation has been shown to decrease VEGF levels in serum and tumor xenografts in stress-exposed mice (Panc-1 and BXPC-3) (219). Furthermore, GABA_A receptor overexpression also down-regulated the levels of VEGF and HIF-1 α protein expression in a carcinoma model system (218). Additionally, in a mouse model of cholangiocarcinoma, GABA inhibited VEGF-A/C expression, resulting in decreased cell proliferation and tumor mass (220). GABAARB2, a GABA_A receptor subunit, was detected in the blood vessels of normal thyroid and thyroid tumors but not in thyroid cancer cells. This suggests that GABA signaling could also directly contribute to angiogenesis in thyroid cancer (221).

4.10 Serotonin

Serotonin is produced in the CNS and can be stored in platelets and presynaptic neurons (222). Its receptors (5-HTRs) are located on the cell membrane and are present in various tissues, including the CNS, heart, gastrointestinal tract, blood vessels, and platelets. The thrombotic environment of tumors induces platelet aggregation, leading to serotonin release, hence facilitating tumor angiogenesis. Serotonin has been found to enhance the migration, permeability, and proliferation of HUVEC and human aortic endothelial cells (HAEC) *in vitro* (223–225). This effect is mediated through the 5-HTR1, 5-HTR2, or 5-HTR3 receptors (225, 226). Serotonin also promotes the expansion of CD34positive EPCs *in vitro* (227). Mechanistically, serotonin has been shown to trigger multiple signaling pathways in endothelial cells, including ERK, P70S6K, Src, PI3K, Akt, mTOR, and p38 MAPK (224, 228, 229).

Serotonin has been implicated in tumor angiogenesis. Tryptophan hydroxylase-deficient mice (serotonin-depleted) exhibited decreased microvessel density in MC38 colon cancers, along with higher MMP-12 and angiostatin concentrations in tumors (230). However, VEGF and VEGFR2 levels were similar to those of wild-type (WT) mice. In contrast, in a model of serotonin transporter-deficient mice bearing murine lung carcinoma or melanoma, tumor microvessel density was not affected, suggesting that the effects might be context-dependent (231). In human cancers, the presence of serotonin-positive cells was associated with higher VEGF expression and increased microvessel density in prostate cancer patients (232, 233). Serotonin could also regulate the vascular tone of the arterioles feeding the tumor through various 5-HTRs expressed on vascular smooth muscle cells, thereby regulating blood flow within the tumor (234).

4.11 Glutamate

Glutamate serves as an excitatory neurotransmitter and regulates synaptic activity through its receptors, including metabotropic glutamate receptors (mGluRs). These receptors are primarily expressed in the CNS, where they play a role in mediating neuronal excitability and neurotransmitter release. Studies have also shown that mGluR1, a subtype of mGluRs, is expressed in several human endothelial cell lines. Activation of mGluR1 signaling in endothelial cells has been linked to endothelial cell growth and tube formation (235).

Various cancers, such as melanoma, breast cancer, and gliomas, have been found to express different subtypes of mGluRs, implicating them in disease progression. Inhibition of mGluR1mediated glutamate release resulted in decreased microvessel density in murine breast tumors (4T1) (235). Additionally, using BAY36-7620, a specific inhibitor of mGluR1, led to inhibited tumor growth and prolonged survival in mice with A549 or H1299 nonsmall cell lung tumors (236). The treated tumors showed reduced protein expression of bcl-2, HIF-1a, and VEGF, suggesting potential therapeutic implications. Increased expression of mGluR1 in several human melanoma cell lines was also associated with elevated expression of IL-8 and VEGF (237). This effect was attributed to the activation of the AKT/mTOR/HIF1 signaling pathway. Riluzole, a glutamate release inhibitor, was also shown to reduce microvessel density in breast cancer xenografts, demonstrating the significant role played by mGluR1 in tumor growth (238).

4.12 Neuropeptides

Neurons can secrete neuropeptides, including substance P and CGRP, which have been shown to play a role in modulating angiogenesis. Substance P has been identified as a promoter of angiogenesis. It achieves this by increasing the expression of proangiogenic factors and stimulating the proliferation of endothelial cells (239). Antagonists of the substance P receptor, Neurokinin-1 (NK-1R) promote apoptosis in tumor cells in a concentrationdependent manner, block the migration of cancer cells, prevent metastasis, and inhibit angiogenesis (240).

Neuropeptide Y (NPY) is secreted by tumor cells and exerts its effects through various receptors, particularly the Y2 receptor (Y2R). NPY plays a significant role in cancer development by mediating both proliferation and angiogenesis (241). The main mechanism by which NPY induces angiogenesis involves its direct influence on endothelial cells. NPY stimulates EC proliferation and migration *in vivo*, and it promotes the formation of capillary tubes (242). Moreover, *in vivo* studies have shown that endogenous NPY facilitates the vascularization of ischemic tissues (243). These actions are dependent on the activation of eNOS and, to some extent, on the VEGF pathway. NPY's angiogenic activities are primarily mediated through Y2R, as demonstrated by impaired angiogenesis in Y2R-deficient mice (Y2R -/-) (244). NPY has also been shown to enhance the expression and secretion of VEGF,

contributing to angiogenesis and promoting breast cancer progression (245). The mechanism underlying this effect involves the activation of endothelial cells through eNOS and through the modulation of the VEGF signaling pathway. NPY treatment also caused a significant increase in VEGF expression in 4T1 breast cancer cells, in a Y5R-dependent manner (245).

Administration of a Y2R antagonist (BIIE 0246) decreased tumor angiogenesis and serum VEGF concentration in obese mice with B16F10 melanomas without altering serum VEGFR1 or NO concentrations (246). Treatment with Y2R antagonists also inhibited angiogenesis of HT29 tumors by regulating the activation of the ERK/MAPK signaling pathway in endothelial cells (247). Y5R inhibition also reduced VEGF expression in 4T1 tumor cells (245). It has also been shown that tumor hypoxia can upregulate the expression of Y2R and Y5R in Ewing sarcoma and endothelial cells, which sensitizes them to the proliferative effects of NPY (248). Treatment of both neuroblastoma and Ewing sarcoma xenografts with a Y2R antagonist also resulted in a significant decrease in tumor vascularization (249). However, a Y5R antagonist alone did not inhibit vascularization of neuroblastoma xenografts (250).

CGRP is a neuropeptide widely distributed in the nervous system that exhibits numerous biological activities (251). It is a member of the calcitonin family of peptides and is produced in both peripheral and central neurons. It acts as a potent vasodilator and can function in the transmission of nociception. The CGRP receptors, Calcitonin Receptor-like receptor (CALCRL) and Receptor Activity-Modifying Protein (RAMP1), are ubiquitously expressed, suggesting that the protein may modulate a variety of physiological functions in all major systems. In endothelial cells, CGRP promotes cell proliferation (252) and activates eNOS via the cAMP-PKA pathway (253). In vivo, CGRP increases angiogenesis during placental development, wound healing and tumorigenesis (127, 254, 255). RAMP1 overexpression led to enhanced endothelial cell migration and capillary tube formation (256). Wound healing and wound-induced angiogenesis were also significantly suppressed in RAMP1(-/-) mice, and was associated with reduced expression of VEGF (257).

Using CGRP-knockout mice, it was shown that CGRP facilitates tumor angiogenesis, as tumor growth and tumor-associated angiogenesis in CGRP-knockout mice implanted with LLC tumors were significantly reduced compared with those in wild-type mice (127). The peptide CGRP 8–37, a CGRP receptor antagonist, was also found to block tumor growth and tumor-associated angiogenesis in wild-type mice (127). The expression of VEGF was also lower in the tumor stroma of CGRP-knockout mice compared to WT mice. CGRP precursor expression in the dorsal root ganglion in lung carcinoma-bearing mice were increased compared to those in mice with no tumors. Denervation of sciatic nerves suppressed LLC growth at the sites of denervation compared, suggesting that local expression of CGRP by peripheral nerves could drive tumorigenesis (127).

4.13 Sympathetic nervous system activation

Activation of the sympathetic nervous system, which is responsible for the "fight-or-flight" response, can affect tumor angiogenesis. Sympathetic or adrenergic nerves are closely associated with blood vessels. The release of catecholamines such as noradrenaline by such nerves mediates blood vessel constriction through surrounding smooth muscle cells contraction (258). Adrenergic nerves have also been shown to potentiate tumor angiogenesis by promoting a putative 'angiometabolic switch' in endothelial cells (259). Indeed, it was shown that the infiltration of adrenergic nerves in the microenvironment of prostate tumors resulted in the liberation of noradrenaline by nerve endings that subsequently stimulated the expression of the the ADR β 2 receptor gene in endothelial cells. When noradrenaline binds to ADRB2, a change in endothelial cell metabolism occurs concomitantly with the inhibition of oxidative phosphorylation. As endothelial cells rely on aerobic glycolysis for angiogenesis, ADR β 2 activation promotes tumor angiogenesis which fuels cancer progression. This could be the cause of the observation that beta blockers improve the survival of patients with cancer: beta blockers inhibit sympathetic nerves mediated ADRB2 signaling in endothelial cells which would in turn inhibit tumor angiogenesis. Evidence pointing to this mechanism is found in pancreatic cancer, where the use of beta blockers was correlated with a significant improvement of the survival rate of patients undergoing surgery, compared to control groups (260). β adrenergic signaling also facilitates angiogenesis through a metabolic switch to activate glycolysis in prostate cancer endothelial cells (259).

Stress responses, which can lead to the release of Norepinephrine (NE) and Epinephrine (EPI), also enhance tumor neovascularization in primary ovarian tumors by upregulating the expression of VEGF and MMPs, which could also underlie the effects of β-blockers, which have been found to reverse stressenhanced angiogenesis (261). Furthermore, the activation of β adrenergic in melanoma, colon cancer, ovarian cancer and multiple myeloma has also been shown to lead to the release of IL-6 or IL-8, in addition to VEGF, contributing to the angiogenic response (262). Additionally, β -adrenergic signaling induces VEGF expression in tumor-associated macrophages through cAMP-mediated pathways (263, 264). Mechanistically, NE-induced VEGF expression was shown to be cAMP mediated, which in turn led to the activation of protein kinase A (PKA). β-adrenergic signaling also modulates the expression of MMPs, which degrade extracellular matrix components involved in tumor cell invasion and angiogenesis. Adrenergic stimulation upregulates MMP-2 and MMP-9 through the extracellular signal-related kinase 1/2 pathway in glioblastoma cells (265).

4.14 Sensory nerve-mediated regulation

Sensory nerves have been shown to be present in a several cancers, including cervical cancer, melanoma, gastric cancer, pancreatic cancer as well as basal cell carcinoma, where they can influence tumor angiogenesis through various mechanisms. Sensory nerves that travel alongside blood vessels promote angiogenesis via substance P-mediated signaling during inflammation and through its promotion of VEGF and MMP-9 expression in cancer cells (240, 266, 267). Sensory neuropeptides, CGRP and substance P, are also

involved in the transcriptional upregulation of several angiogenic markers (VEGF, angiopoietin-2, type 4 collagen, MMP-2) in ECs (268, 269). Co-cultures of endothelial cells with sensory neurons also increased the protein level and enzymatic activity of matrix metalloproteinases 2 and 9 (MMP-2/MMP-9) in ECs (269).

It was also shown that sensory neurons could prevent tumor angiogenesis and growth. A recent study showed that depletion of Nav1.8-positive sensory neurons using genetic or pharmacological approaches could enhance melanoma growth and tumor angiogenesis (103). In this study, Prazeres and colleagues demonstrated that ablation of nociceptors in a murine model of B16F10 melanoma enhanced tumor growth and the intra-tumoral blood vessels' area, suggesting that sensory nerves inhibit tumor angiogenesis in this model. Furthermore, using silencing of genetically defined neuronal populations upon binding to smallmolecule designer drugs, a 2021 study showed that inhibition of sensory neurons' activity is sufficient to trigger an increase in melanoma growth and intra-tumoral new blood vessel formation (270). Conversely, chemogenetic stimulation of sensory neurons counteracted melanoma progression, by negatively regulating tumoral growth, angiogenesis and immunosurveillance (270). While the underlying mechanisms by which nociceptors could limit tumor angiogenesis in melanoma remain unclear, it has been speculated that neuropeptides such as substance P, VIP, tachykinins or CGRP could play a role. Furthermore, sensory nerves may tune down sympathetic nerve activity (271), limiting norepinephrine release, which has been shown to strongly induce tumorigenesis and angiogenesis (262, 272, 273).

4.15 Neurovascular coupling

Neurons and blood vessels in the brain are tightly interconnected, and neurovascular coupling refers to the coordination between neuronal activity and local blood flow regulation (29). Changes in neuronal activity can influence the supply of oxygen and nutrients to surrounding tissues, including tumors (274). Disruptions in neurovascular coupling, such as alterations in blood flow regulation, can affect tumor angiogenesis, and perturbations of the neurovascular unit composed of astrocytes, pericytes, and endothelial cells have been described in tumors of the nervous system such as gliomas (274–277).

Recent studies have highlighted the presence of neurovascular uncoupling in association with brain tumors. These disruptions interfere with neurovascular coupling, resulting in excitotoxicity caused by an inadequate vascular response, leading to decreased delivery of oxygen and nutrients, resulting in hypoxia and cell death, initiating a chain of events that ultimately triggers angiogenic growth and the development of poorly formed vasculature (278, 279). Furthermore, it has been suggested that the failure to deliver essential metabolites required for neural firing may accelerate excitotoxicity (279).

Glioma cells have been shown to closely interact with the brain vasculature, migrating along blood vessels and using them as conduits during tumor spread, leading to disruption of structural and functional integrity (279, 280). These interactions further suggest that tumors could impact the coupling between neuronal activity and vascular dynamics during glioma-infiltration and contribute to the disturbed functional and metabolic state of the brain during tumor progression. Indeed, a recent study demonstrated that glioma infiltration led to tumor-localized alterations in neuronal synchrony and neurovascular coupling, interictal events, and seizures accompanied by disrupted hemodynamic responses in mice (274). Changes in vascular perfusion, combined with the marked increase in neuronal activity and metabolic demands, were postulated to lead to tissue hypoxia within the tumor-bearing cortex, which in turn would drive the expression of cytokines and growth factors that stimulate microvascular proliferation.

Apart from neurovascular uncoupling, the growth of tumor cells along the abluminal surface of cerebrovasculature and the disruption of astrocyte function appear to compromise the integrity of the blood-brain barrier tight junctions in gliomas. As a result of gliomas down-regulating tight junction proteins (claudin-1, -3, and -5), the blood-brain barrier becomes more permeable (278, 281). This phenomenon leads to the formation of leaky blood vessels which can accelerate pro-inflammatory responses. Moreover, glioma growth on the abluminal surface of blood vessels can also impair the association of pericytes with the vasculature, contributing to blood-brain barrier breakdown. Consequently, a decrease in pericyte population may not only affect blood-brain barrier permeability, allowing the entry of molecules that are normally filtered out into the brain's extracellular matrix, but also result in neurovascular uncoupling (278).

5 Neurogenesis and tumor progression

A recent study on prostate cancer used lineage tracing of doublecortin-positive neural progenitor cells from the subventricular zone of the brain to examine whether tumor innervation can originate from the CNS (282). It was observed that subventricular zone-derived neural progenitor cells could cross the blood-brain barrier and migrate to the site of the tumor to further populate it. Upon arrival in the tumor microenvironment, these neural progenitor cells mature to form sympathetic nerves that promote tumorigenesis (283). Depletion of these doublecortinpositive neural progenitor cells abolishes tumor initiation and progression. Conversely, transplantation of doublecortin-positive cells from the subventricular zone augments tumor xenograft growth and metastasis in vivo (282). Furthermore, lineage tracing of ex vivo-tagged human gastric and colorectal carcinoma stem cells, which are cells that can both self-renew and differentiate, showed that these cells not only initiated tumorigenesis in xenograft mouse models but also that the tagged human cancer stem cells differentiated into tyrosine hydroxylase-producing sympathetic and vesicular acetylcholine transporter-producing parasympathetic neurons upon engraftment. Inhibition of their differentiation capacity abrogated xenograft tumor growth (284). These data

support the notion that cancer stem cells can mature into heterogenous cell populations, including neuron-like cells, that are found in the tumor microenvironment. Moreover, stem-like progenitors in prostate cancer possess neurogenic gene expression profiles (285).

6 Concluding remarks

This review evaluated the signaling pathways and molecules within the neural systems and examined their potential roles on the development of blood vessels within tumors. The identification of neurons-derived factors within the tumor microenvironment can shed light on molecular processes involved in tumorigenesis. Indeed, forthcoming studies have the potential to enhance existing anti-angiogenic or vascular normalization therapies by delving deeper into the molecular mechanisms underlying angiogenesis and neurogenesis. By studying the impacts of various combinations of neuron-derived factors and guidance cues on endothelial cells and analyzing the factors that modulate the balance between sprouting angiogenesis and quiescence in endothelial cells, we can significantly advance our comprehension of the intricate mechanisms involved in the establishment of the tumor microenvironment and design strategies to improve therapeutic strategies.

Author contributions

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References

1. Hirata E, Sahai E. Tumor microenvironment and differential responses to therapy. *Cold Spring Harb Perspect Med* (2017) 7. doi: 10.1101/cshperspect.a026781

2. Arner EN, Rathmell JC. Metabolic programming and immune suppression in the tumor microenvironment. *Cancer Cell* (2023) 41:421-33. doi: 10.1016/j.ccell.2023.01.009

3. Wu Y, Song Y, Wang R, Wang T. Molecular mechanisms of tumor resistance to radiotherapy. *Mol Cancer* (2023) 22:96. doi: 10.1186/s12943-023-01801-2

4. Tay C, Tanaka A, Sakaguchi S. Tumor-infiltrating regulatory T cells as targets of cancer immunotherapy. *Cancer Cell* (2023) 41:450–65. doi: 10.1016/j.ccell.2023.02.014

5. Xiao Y, Yu D. Tumor microenvironment as a therapeutic target in cancer. *Pharmacol Ther* (2021) 221:107753. doi: 10.1016/j.pharmthera.2020.107753

6. Sobierajska K, Ciszewski WM, Sacewicz-Hofman I, Niewiarowska J. Endothelial cells in the tumor microenvironment. *Adv Exp Med Biol* (2020) 1234:71–86. doi: 10.1007/978-3-030-37184-5_6

7. Takara K, Hayashi-Okada Y, Kidoya H. Neurovascular interactions in the development of the vasculature. *Life (Basel)* (2022) 13. doi: 10.3390/life13010042

8. James JM, Mukouyama Y. Neuronal action on the developing blood vessel pattern. *Semin Cell Dev Biol* (2011) 22:1019–27. doi: 10.1016/j.semcdb.2011.09.010

9. Entschladen F, Palm D, Lang K, Drell TL4, Zaenker KS. Neoneurogenesis: tumors may initiate their own innervation by the release of neurotrophic factors in analogy to lymphangiogenesis and neoangiogenesis. *Med Hypotheses* (2006) 67:33–5. doi: 10.1016/j.mehy.2006.01.015

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10. Yang M-W, Tao L-Y, Jiang Y-S, Yang J-Y, Huo Y-M, Liu D-J, et al. Perineural invasion reprograms the immune microenvironment through cholinergic signaling in pancreatic ductal adenocarcinoma. *Cancer Res* (2020) 80:1991–2003. doi: 10.1158/0008-5472.CAN-19-2689

11. Faulkner S, Jobling P, March B, Jiang CC, Hondermarck H. Tumor neurobiology and the war of nerves in cancer. *Cancer Discovery* (2019) 9:702–10. doi: 10.1158/2159-8290.CD-18-1398

12. Bazzoni G, Dejana E. Endothelial cell-to-cell junctions: molecular organization and role in vascular homeostasis. *Physiol Rev* (2004) 84:869–901. doi: 10.1152/ physrev.00035.2003

13. Armulik A, Genové G, Betsholtz C. Pericytes: developmental, physiological, and pathological perspectives, problems, and promises. *Dev Cell* (2011) 21:193–215. doi: 10.1016/j.devcel.2011.07.001

14. Wilting J, Brand-Saberi B, Kurz H, Christ B. Development of the embryonic vascular system. *Cell Mol Biol Res* (1995) 41:219–32.

15. Adams RH, Alitalo K. Molecular regulation of angiogenesis and lymphangiogenesis. *Nat Rev Mol Cell Biol* (2007) 8:464–78. doi: 10.1038/nrm2183

16. Riha GM, Lin PH, Lumsden AB, Yao Q, Chen C. Roles of hemodynamic forces in vascular cell differentiation. *Ann BioMed Eng* (2005) 33:772–9. doi: 10.1007/s10439-005-3310-9

17. Gerhardt H, Golding M, Fruttiger M, Ruhrberg C, Lundkvist A, Abramsson A, et al. VEGF guides angiogenic sprouting utilizing endothelial tip cell filopodia. *J Cell Biol* (2003) 161:1163–77. doi: 10.1083/jcb.200302047

18. Larrivée B, Freitas C, Suchting S, Brunet I, Eichmann A. Guidance of vascular development: lessons from the nervous system. *Circ Res* (2009) 104:428-41. doi: 10.1161/CIRCRESAHA.108.188144

19. Viallard C, Larrivée B. Tumor angiogenesis and vascular normalization: alternative therapeutic targets. *Angiogenesis* (2017) 20:409–26. doi: 10.1007/s10456-017-9562-9

20. Greenberg JI, Shields DJ, Barillas SG, Acevedo LM, Murphy E, Huang J, et al. A role for VEGF as a negative regulator of pericyte function and vessel maturation. *Nature* (2008) 456:809–13. doi: 10.1038/nature07424

21. Beckers CML, van Hinsbergh VWM, van Nieuw Amerongen GP. Driving Rho GTPase activity in endothelial cells regulates barrier integrity. *Thromb Haemost* (2010) 103:40–55. doi: 10.1160/TH09-06-0403

22. Dejana E, Tournier-Lasserve E, Weinstein BM. The control of vascular integrity by endothelial cell junctions: molecular basis and pathological implications. *Dev Cell* (2009) 16:209–21. doi: 10.1016/j.devcel.2009.01.004

23. Guo M, Breslin JW, Wu MH, Gottardi CJ, Yuan SY. VE-cadherin and betacatenin binding dynamics during histamine-induced endothelial hyperpermeability. *Am J Physiol Cell Physiol* (2008) 294:C977–984. doi: 10.1152/ajpcell.90607.2007

24. Jain RK, Martin JD, Stylianopoulos T. The role of mechanical forces in tumor growth and therapy. *Annu Rev BioMed Eng* (2014) 16:321–46. doi: 10.1146/annurev-bioeng-071813-105259

25. Reymond N, d'Água BB, Ridley AJ. Crossing the endothelial barrier during metastasis. *Nat Rev Cancer* (2013) 13:858–70. doi: 10.1038/nrc3628

26. Giaccia AJ, Simon MC, Johnson R. The biology of hypoxia: the role of oxygen sensing in development, normal function, and disease. *Genes Dev* (2004) 18:2183–94. doi: 10.1101/gad.1243304

27. Mazzone M, Dettori D, de Oliveira RL, Loges S, Schmidt T, Jonckx B, et al. Heterozygous deficiency of PHD2 restores tumor oxygenation and inhibits metastasis via endothelial normalization. *Cell* (2009) 136:839–51. doi: 10.1016/j.cell.2009.01.020

28. Marín-Hernández A, Gallardo-Pérez JC, Ralph SJ, Rodríguez-Enríquez S, Moreno-Sánchez R. HIF-1alpha modulates energy metabolism in cancer cells by inducing over-expression of specific glycolytic isoforms. *Mini Rev Med Chem* (2009) 9:1084-101. doi: 10.2174/138955709788922610

29. Segarra M, Aburto MR, Hefendehl J, Acker-Palmer A. Neurovascular interactions in the nervous system. *Annu Rev Cell Dev Biol* (2019) 35:615–35. doi: 10.1146/annurev-cellbio-100818-125142

30. Argandoña EG, Lafuente JV. Influence of visual experience deprivation on the postnatal development of the microvascular bed in layer IV of the rat visual cortex. *Brain Res* (2000) 855:137–42. doi: 10.1016/S0006-8993(99)02361-6

31. Black JE, Sirevaag AM, Greenough WT. Complex experience promotes capillary formation in young rat visual cortex. *Neurosci Lett* (1987) 83:351–5. doi: 10.1016/0304-3940(87)90113-3

32. Lacoste B, Comin CH, Ben-Zvi A, Kaeser PS, Xu X, Costa da L F, et al. Sensoryrelated neural activity regulates the structure of vascular networks in the cerebral cortex. *Neuron* (2014) 83:1117–30. doi: 10.1016/j.neuron.2014.07.034

33. Narboux-Nême N, Evrard A, Ferezou I, Erzurumlu RS, Kaeser PS, Lainé J, et al. Neurotransmitter release at the thalamocortical synapse instructs barrel formation but not axon patterning in the somatosensory cortex. *J Neurosci* (2012) 32:6183–96. doi: 10.1523/JNEUROSCI.0343-12.2012

34. Shome S, Rana T, Ganguly S, Basu B, Chaki Choudhury S, Sarkar C, et al. Dopamine regulates angiogenesis in normal dermal wound tissues. *PloS One* (2011) 6: e25215. doi: 10.1371/journal.pone.0025215

35. Liang JH, Akhanov V, Ho A, Tawfik M, D'Souza SP, Cameron MA, et al. Dopamine signaling from ganglion cells directs layer-specific angiogenesis in the retina. *Curr Biol* (2023) 33:3821–3834.e5. doi: 10.1016/j.cub.2023.07.040

36. Li S, Kumar T P, Joshee S, Kirschstein T, Subburaju S, Khalili JS, et al. Endothelial cell-derived GABA signaling modulates neuronal migration and postnatal behavior. *Cell Res* (2018) 28:221-48.

37. Alenina N, Kikic D, Todiras M, Mosienko V, Qadri F, Plehm R, et al. Growth retardation and altered autonomic control in mice lacking brain serotonin. *Proc Natl Acad Sci U.S.A.* (2009) 106:10332–7.

38. Heeschen C, Jang JJ, Weis M, Pathak A, Kaji S, Hu RS, et al. Nicotine stimulates angiogenesis and promotes tumor growth and atherosclerosis. *Nat Med* (2001) 7:833–9.

39. Jacobi J, Jang JJ, Sundram U, Dayoub H, Fajardo LF, Cooke JP, et al. Nicotine accelerates angiogenesis and wound healing in genetically diabetic mice. *Am J Pathol* (2002) 161:97–104.

40. Yang C, Li Z, Yan S, He Y, Dai R, Leung GP-H, et al. Role of the nicotinic acetylcholine receptor $\alpha 3$ subtype in vascular inflammation. *Br J Pharmacol* (2016) 173:3235-47.

41. Collard CD, Park KA, Montalto MC, Alapati S, Buras JA, Stahl GL, et al. Neutrophil-derived glutamate regulates vascular endothelial barrier function. *J Biol Chem* (2002) 277:14801-11. doi: 10.1074/jbc.M110557200

42. De Smet F, Segura I, De Bock K, Hohensinner PJ, Carmeliet P. Mechanisms of vessel branching: filopodia on endothelial tip cells lead the way. *Arterioscler Thromb Vasc Biol* (2009) 29:639–49. doi: 10.1161/ATVBAHA.109.185165

43. Licht T, Goshen I, Avital A, Kreisel T, Zubedat S, Eavri R, et al. Reversible modulations of neuronal plasticity by VEGF. *Proc Natl Acad Sci U.S.A.* (2011) 108:5081–6.

44. Jin K, Zhu Y, Sun Y, Mao XO, Xie L, Greenberg DA. Vascular endothelial growth factor (VEGF) stimulates neurogenesis *in vitro* and *in vivo*. *Proc Natl Acad Sci U.S.A.* (2002) 99:11946–50.

45. Licht T, Eavri R, Goshen I, Shlomai Y, Mizrahi A, Keshet E. VEGF is required for dendritogenesis of newly born olfactory bulb interneurons. *Development* (2010) 137:261–71. doi: 10.1242/dev.039636

46. Foehring D, Brand-Saberi B, Theiss C. VEGF-induced growth cone enhancement is diminished by inhibiting tyrosine-residue 1214 of VEGFR-2. *Cells Tissues Organs* (2012) 196:195–205. doi: 10.1159/000334600

47. Schratzberger P, Schratzberger G, Silver M, Curry C, Kearney M, Magner M, et al. Favorable effect of VEGF gene transfer on ischemic peripheral neuropathy. *Nat Med* (2000) 6:405–13. doi: 10.1038/74664

48. Fard D, Tamagnone L. Semaphorins in health and disease. Cytokine Growth Factor Rev (2021) 57:55-63. doi: 10.1016/j.cytogfr.2020.05.006

49. Pellet-Many C, Frankel P, Jia H, Zachary I. Neuropilins: structure, function and role in disease. Biochem J (2008) 411:211–26. doi: 10.1042/BJ20071639

50. Limoni G, Niquille M. Semaphorins and Plexins in central nervous system patterning: the key to it all? *Curr Opin Neurobiol* (2021) 66:224–32. doi: 10.1016/j.conb.2020.12.014

51. Vieira JM, Schwarz Q, Ruhrberg C. Role of the neuropilin ligands VEGF164 and SEMA3A in neuronal and vascular patterning in the mouse. *Novartis Found Symp* (2007) 283:230–5; discussion 235-241. doi: 10.1002/9780470319413.ch18

52. Acevedo LM, Barillas S, Weis SM, Göthert JR, Cheresh DA. Semaphorin 3A suppresses VEGF-mediated angiogenesis yet acts as a vascular permeability factor. *Blood* (2008) 111:2674–80. doi: 10.1182/blood-2007-08-110205

53. Kaneko S, Iwanami A, Nakamura M, Kishino A, Kikuchi K, Shibata S, et al. A selective Sema3A inhibitor enhances regenerative responses and functional recovery of the injured spinal cord. *Nat Med* (2006) 12:1380–9. doi: 10.1038/nm1505

54. Fukushima Y, Okada M, Kataoka H, Hirashima M, Yoshida Y, Mann F, et al. Sema3E-PlexinD1 signaling selectively suppresses disoriented angiogenesis in ischemic retinopathy in mice. *J Clin Invest* (2011) 121:1974–85. doi: 10.1172/JCI44900

55. Kim J, Oh W-J, Gaiano N, Yoshida Y, Gu C. Semaphorin 3E-Plexin-D1 signaling regulates VEGF function in developmental angiogenesis via a feedback mechanism. *Genes Dev* (2011) 25:1399–411. doi: 10.1101/gad.2042011

56. Lu Y, Xu Q, Chen L, Zuo Y, Liu S, Hu Y, et al. Expression of semaphorin 6D and its receptor plexin-A1 in gastric cancer and their association with tumor angiogenesis. *Oncol Lett* (2016) 12:3967–74. doi: 10.3892/ol.2016.5208

57. Bouvrée K, Brunet I, Del Toro R, Gordon E, Prahst C, Cristofaro B, et al. Semaphorin3A, Neuropilin-1, and PlexinA1 are required for lymphatic valve formation. *Circ Res* (2012) 111:437–45. doi: 10.1161/CIRCRESAHA.112.269316

58. Torres-Vázquez J, Gitler AD, Fraser SD, Berk JD, Pham VN, Fishman MC, et al. Semaphorin-plexin signaling guides patterning of the developing vasculature. *Dev Cell* (2004) 7:117–23. doi: 10.1016/j.devcel.2004.06.008

59. Yuan L, Moyon D, Pardanaud L, Bréant C, Karkkainen MJ, Alitalo K, et al. Abnormal lymphatic vessel development in neuropilin 2 mutant mice. *Development* (2002) 129:4797–806. doi: 10.1242/dev.129.20.4797

60. Xu Y, Yuan L, Mak J, Pardanaud L, Caunt M, Kasman I, et al. Neuropilin-2 mediates VEGF-C-induced lymphatic sprouting together with VEGFR3. *J Cell Biol* (2010) 188:115–30. doi: 10.1083/jcb.200903137

61. Kawasaki T, Kitsukawa T, Bekku Y, Matsuda Y, Sanbo M, Yagi T, et al. A requirement for neuropilin-1 in embryonic vessel formation. *Development* (1999) 126:4895–902. doi: 10.1242/dev.126.21.4895

62. Mukouyama Y-S, Gerber H-P, Ferrara N, Gu C, Anderson DJ. Peripheral nervederived VEGF promotes arterial differentiation via neuropilin 1-mediated positive feedback. *Development* (2005) 132:941–52. doi: 10.1242/dev.01675

63. Gu C, Yoshida Y, Livet J, Reimert DV, Mann F, Merte J, et al. Semaphorin 3E and plexin-D1 control vascular pattern independently of neuropilins. *Science* (2005) 307:265–8. doi: 10.1126/science.1105416

64. Sheng J, Xu J, Geng K, Liu D. Sema6D regulates zebrafish vascular patterning and motor neuronal axon growth in spinal cord. *Front Mol Neurosci* (2022) 15:854556. doi: 10.3389/fnmol.2022.854556

65. Toyofuku T, Zhang H, Kumanogoh A, Takegahara N, Yabuki M, Harada K, et al. Guidance of myocardial patterning in cardiac development by Sema6D reverse signalling. *Nat Cell Biol* (2004) 6:1204–11. doi: 10.1038/ncb1193

66. Vreeken D, Zhang H, van Zonneveld AJ, van Gils JM. Ephs and ephrins in adult endothelial biology. *Int J Mol Sci* (2020) 21. doi: 10.3390/ijms21165623

67. Flanagan JG, Vanderhaeghen P. The ephrins and Eph receptors in neural development. Annu Rev Neurosci (1998) 21:309–45. doi: 10.1146/annurev.neuro.21.1.309

68. Ye X, Qiu Y, Gao Y, Wan D, Zhu H. A subtle network mediating axon guidance: intrinsic dynamic structure of growth cone, attractive and repulsive molecular cues, and the intermediate role of signaling pathways. *Neural Plast* (2019) 2019:1719829. doi: 10.1155/2019/1719829

69. Frisén J, Holmberg J, Barbacid M. Ephrins and their Eph receptors: multitalented directors of embryonic development. *EMBO J* (1999) 18:5159–65. doi: 10.1093/emboj/ 18.19.5159

70. Adams RH, Wilkinson GA, Weiss C, Diella F, Gale NW, Deutsch U, et al. Roles of ephrinB ligands and EphB receptors in cardiovascular development: demarcation of

arterial/venous domains, vascular morphogenesis, and sprouting angiogenesis. Genes Dev (1999) 13:295-306. doi: 10.1101/gad.13.3.295

71. Gerety SS, Wang HU, Chen ZF, Anderson DJ. Symmetrical mutant phenotypes of the receptor EphB4 and its specific transmembrane ligand ephrin-B2 in cardiovascular development. *Mol Cell* (1999) 4:403–14. doi: 10.1016/S1097-2765(00) 80342-1

72. Foo SS, Turner CJ, Adams S, Compagni A, Aubyn D, Kogata N, et al. Ephrin-B2 controls cell motility and adhesion during blood-vessel-wall assembly. *Cell* (2006) 124:161–73. doi: 10.1016/j.cell.2005.10.034

73. Wang Y, Nakayama M, Pitulescu ME, Schmidt TS, Bochenek ML, Sakakibara A, et al. Ephrin-B2 controls VEGF-induced angiogenesis and lymphangiogenesis. *Nature* (2010) 465:483–6. doi: 10.1038/nature09002

74. Long H, Sabatier C, Ma L, Plump A, Yuan W, Ornitz DM, et al. Conserved roles for Slit and Robo proteins in midline commissural axon guidance. *Neuron* (2004) 42:213–23. doi: 10.1016/S0896-6273(04)00179-5

75. Tong M, Jun T, Nie Y, Hao J, Fan D. The role of the slit/robo signaling pathway. J Cancer (2019) 10:2694–705. doi: 10.7150/jca.31877

76. Rama N, Dubrac A, Mathivet T, Ní Chárthaigh R-A, Genet G, Cristofaro B, et al. Slit2 signaling through Robo1 and Robo2 is required for retinal neovascularization. *Nat Med* (2015) 21:483–91. doi: 10.1038/nm.3849

77. Koch AW, Mathivet T, Larrivée B, Tong RK, Kowalski J, Pibouin-Fragner L, et al. Robo4 maintains vessel integrity and inhibits angiogenesis by interacting with UNC5B. *Dev Cell* (2011) 20:33–46. doi: 10.1016/j.devcel.2010.12.001

78. Dai C, Gong Q, Cheng Y, Su G. Regulatory mechanisms of Robo4 and their effects on angiogenesis. *Biosci Rep* (2019) 39:BSR20190513. doi: 10.1042/BSR20190513

79. Marlow R, Binnewies M, Sorensen LK, Monica SD, Strickland P, Forsberg EC, et al. Vascular Robo4 restricts proangiogenic VEGF signaling in breast. *Proc Natl Acad Sci U.S.A.* (2010) 107:10520–5.

80. Li X, Li Y, Li S, Li H, Yang C, Lin J. The role of Shh signalling pathway in central nervous system development and related diseases. *Cell Biochem Funct* (2021) 39:180–9. doi: 10.1002/cbf.3582

81. Briscoe J, Thérond PP. The mechanisms of Hedgehog signalling and its roles in development and disease. *Nat Rev Mol Cell Biol* (2013) 14:416–29. doi: 10.1038/ nrm3598

82. Kolpak A, Zhang J, Bao Z-Z. Sonic hedgehog has a dual effect on the growth of retinal ganglion axons depending on its concentration. *J Neurosci* (2005) 25:3432–41. doi: 10.1523/JNEUROSCI.4938-04.2005

83. Yao Q, Renault M-A, Chapouly C, Vandierdonck S, Belloc I, Jaspard-Vinassa B, et al. Sonic hedgehog mediates a novel pathway of PDGF-BB-dependent vessel maturation. *Blood* (2014) 123:2429–37. doi: 10.1182/blood-2013-06-508689

84. Fernandes-Silva H, Correia-Pinto J, Moura RS. Canonical sonic hedgehog signaling in early lung development. J Dev Biol (2017) 5. doi: 10.3390/jdb5010003

85. Pola R, Ling LE, Silver M, Corbley MJ, Kearney M, Blake Pepinsky R, et al. The morphogen Sonic hedgehog is an indirect angiogenic agent upregulating two families of angiogenic growth factors. *Nat Med* (2001) 7:706–11. doi: 10.1038/89083

86. Kanda S, Mochizuki Y, Suematsu T, Miyata Y, Nomata K, Kanetake H. Sonic hedgehog induces capillary morphogenesis by endothelial cells through phosphoinositide 3-kinase. *J Biol Chem* (2003) 278:8244–9. doi: 10.1074/ jbc.M210635200

87. Renault M-A, Roncalli J, Tongers J, Thorne T, Klyachko E, Misener S, et al. Sonic hedgehog induces angiogenesis via Rho kinase-dependent signaling in endothelial cells. J Mol Cell Cardiol (2010) 49:490–8. doi: 10.1016/j.yjmcc.2010.05.003

88. Dickson BJ, Keleman K. Netrins. Curr Biol (2002) 12:R154–155. doi: 10.1016/ S0960-9822(02)00728-5

89. Boyer NP, Gupton SL. Revisiting netrin-1: one who guides (Axons). Front Cell Neurosci (2018) 12:221. doi: 10.3389/fncel.2018.00221

90. Höpker VH, Shewan D, Tessier-Lavigne M, Poo M, Holt C. Growth-cone attraction to netrin-1 is converted to repulsion by laminin-1. *Nature* (1999) 401:69–73. doi: 10.1038/43441

91. Serafini T, Colamarino SA, Leonardo ED, Wang H, Beddington R, Skarnes WC, et al. Netrin-1 is required for commissural axon guidance in the developing vertebrate nervous system. *Cell* (1996) 87:1001–14. doi: 10.1016/S0092-8674(00)81795-X

92. Murray A, Naeem A, Barnes SH, Drescher U, Guthrie S. Slit and Netrin-1 guide cranial motor axon pathfinding via Rho-kinase, myosin light chain kinase and myosin II. *Neural Dev* (2010) 5:16. doi: 10.1186/1749-8104-5-16

93. Brunet I, Gordon E, Han J, Cristofaro B, Broqueres-You D, Liu C, et al. Netrin-1 controls sympathetic arterial innervation. *J Clin Invest* (2014) 124:3230–40. doi: 10.1172/JCI75181

94. Lu X, Le Noble F, Yuan L, Jiang Q, De Lafarge B, Sugiyama D, et al. The netrin receptor UNC5B mediates guidance events controlling morphogenesis of the vascular system. *Nature* (2004) 432:179–86. doi: 10.1038/nature03080

95. Boyé K, Geraldo LH, Furtado J, Pibouin-Fragner L, Poulet M, Kim D, et al. Endothelial Unc5B controls blood-brain barrier integrity. *Nat Commun* (2022) 13:1169. doi: 10.1038/s41467-022-28785-9

96. Larrivée B, Freitas C, Trombe M, Lv X, Delafarge B, Yuan L, et al. Activation of the UNC5B receptor by Netrin-1 inhibits sprouting angiogenesis. *Genes Dev* (2007) 21:2433–47. doi: 10.1101/gad.437807

97. Shimizu A, Nakayama H, Wang P, König C, Akino T, Sandlund J, et al. Netrin-1 promotes glioblastoma cell invasiveness and angiogenesis by multiple pathways including activation of RhoA, cathepsin B, and cAMP-response element-binding protein. *J Biol Chem* (2013) 288:2210–22. doi: 10.1074/jbc.M112.397398

98. Wilson BD, Ii M, Park KW, Suli A, Sorensen LK, Larrieu-Lahargue F, et al. Netrins promote developmental and therapeutic angiogenesis. *Science* (2006) 313:640–4. doi: 10.1126/science.1124704

99. Nguyen A, Cai H. Netrin-1 induces angiogenesis via a DCC-dependent ERK1/2eNOS feed-forward mechanism. *Proc Natl Acad Sci U.S.A.* (2006) 103:6530–5.

100. Castets M, Coissieux M-M, Delloye-Bourgeois C, Bernard L, Delcros J-G, Bernet A, et al. Inhibition of endothelial cell apoptosis by netrin-1 during angiogenesis. *Dev Cell* (2009) 16:614–20. doi: 10.1016/j.devcel.2009.02.006

101. Achen MG, Stacker SA. Tumor lymphangiogenesis and metastatic spread-new players begin to emerge. *Int J Cancer* (2006) 119:1755–60. doi: 10.1002/ijc.21899

102. Mancino M, Ametller E, Gascón P, Almendro V. The neuronal influence on tumor progression. *Biochim Biophys Acta* (2011) 1816:105–18. doi: 10.1016/j.bbcan.2011.04.005

103. Prazeres PHDM, Leonel C, Silva WN, Rocha BGS, Santos GSP, Costa AC, et al. Ablation of sensory nerves favours melanoma progression. *J Cell Mol Med* (2020) 24:9574–89. doi: 10.1111/jcmm.15381

104. Seifert P, Spitznas M. Axons in human choroidal melanoma suggest the participation of nerves in the control of these tumors. *Am J Ophthalmol* (2002) 133:711-3. doi: 10.1016/S0002-9394(02)01329-6

105. Ventura S, Pennefather J, Mitchelson F. Cholinergic innervation and function in the prostate gland. *Pharmacol Ther* (2002) 94:93–112. doi: 10.1016/S0163-7258(02) 00174-2

106. Mitchell BS, Schumacher U, Stauber VV, Kaiserling E. Are breast tumours innervated? Immunohistological investigations using antibodies against the neuronal marker protein gene product 9.5 (PGP 9.5) in benign and Malignant breast lesions. *Eur J Cancer* (1994) 30A:1100–3.

107. Tsang WY, Chan JK. Neural invasion in intraductal carcinoma of the breast. Hum Pathol (1992) 23:202-4. doi: 10.1016/0046-8177(92)90247-Z

108. Kayahara M, Nakagawara H, Kitagawa H, Ohta T. The nature of neural invasion by pancreatic cancer. *Pancreas* (2007) 35:218-23. doi: 10.1097/mpa.0b013e3180619677

109. Lü S-H, Zhou Y, Que H-P, Liu S-J. Peptidergic innervation of human esophageal and cardiac carcinoma. *World J Gastroenterol* (2003) 9:399-403. doi: 10.3748/wjg.v9.i3.399

110. Albo D, Akay CL, Marshall CL, Wilks JA, Verstovsek G, Liu H, et al. Neurogenesis in colorectal cancer is a marker of aggressive tumor behavior and poor outcomes. *Cancer* (2011) 117:4834–45. doi: 10.1002/cncr.26117

111. Zhao Q, Yang Y, Liang X, Du G, Liu L, Lu L, et al. The clinicopathological significance of neurogenesis in breast cancer. *BMC Cancer* (2014) 14:484. doi: 10.1186/1471-2407-14-484

112. Pundavela J, Roselli S, Faulkner S, Attia J, Scott RJ, Thorne RF, et al. Nerve fibers infiltrate the tumor microenvironment and are associated with nerve growth factor production and lymph node invasion in breast cancer. *Mol Oncol* (2015) 9:1626–35. doi: 10.1016/j.molonc.2015.05.001

113. Zhang L, Guo L, Tao M, Fu W, Xiu D. Parasympathetic neurogenesis is strongly associated with tumor budding and correlates with an adverse prognosis in pancreatic ductal adenocarcinoma. *Chin J Cancer Res* (2016) 28:180–6. doi: 10.21147/j.issn.1000-9604.2016.02.05

114. Rowe CW, Dill T, Griffin N, Jobling P, Faulkner S, Paul JW, et al. Innervation of papillary thyroid cancer and its association with extra-thyroidal invasion. *Sci Rep* (2020) 10:1539. doi: 10.1038/s41598-020-58425-5

115. Liebig C, Ayala G, Wilks JA, Berger DH, Albo D. Perineural invasion in cancer: a review of the literature. *Cancer* (2009) 115:3379–91. doi: 10.1002/cncr.24396

116. Ayala GE, Dai H, Ittmann M, Li R, Powell M, Frolov A, et al. Growth and survival mechanisms associated with perineural invasion in prostate cancer. *Cancer Res* (2004) 64:6082–90. doi: 10.1158/0008-5472.CAN-04-0838

117. Dai H, Li R, Wheeler T, Ozen M, Ittmann M, Anderson M, et al. Enhanced survival in perineural invasion of pancreatic cancer: an *in vitro* approach. *Hum Pathol* (2007) 38:299–307. doi: 10.1016/j.humpath.2006.08.002

118. Dollé L, El Yazidi-Belkoura I, Adriaenssens E, Nurcombe V, Hondermarck H. Nerve growth factor overexpression and autocrine loop in breast cancer cells. *Oncogene* (2003) 22:5592–601. doi: 10.1038/sj.onc.1206805

119. Arakawa H. Netrin-1 and its receptors in tumorigenesis. *Nat Rev Cancer* (2004) 4:978–87. doi: 10.1038/nrc1504

120. Ceyhan GO, Liebl F, Maak M, Schuster T, Becker K, Langer R, et al. The severity of neural invasion is a crucial prognostic factor in rectal cancer independent of neoadjuvant radiochemotherapy. *Ann Surg* (2010) 252:797–804. doi: 10.1097/SLA.0b013e3181fcab8d

121. Bixby JL, Harris WA. Molecular mechanisms of axon growth and guidance. Annu Rev Cell Biol (1991) 7:117–59. doi: 10.1146/annurev.cb.07.110191.001001

122. Tomaselli KJ, Reichardt LF, Bixby JL. Distinct molecular interactions mediate neuronal process outgrowth on non-neuronal cell surfaces and extracellular matrices. *J Cell Biol* (1986) 103:2659–72. doi: 10.1083/jcb.103.6.2659

123. Bixby JL, Pratt RS, Lilien J, Reichardt LF. Neurite outgrowth on muscle cell surfaces involves extracellular matrix receptors as well as Ca2+-dependent and -independent cell adhesion molecules. *Proc Natl Acad Sci U.S.A.* (1987) 84:2555–9.

124. Tonge DA, Golding JP, Edbladh M, Kroon M, Ekström PE, Edström A. Effects of extracellular matrix components on axonal outgrowth from peripheral nerves of adult animals *in vitro*. *Exp Neurol* (1997) 146:81–90. doi: 10.1006/exnr.1997.6498

125. Giger RJ, Hollis ER 2nd, Tuszynski MH. Guidance molecules in axon regeneration. *Cold Spring Harb Perspect Biol* (2010) 2:a001867. doi: 10.1101/ cshperspect.a001867

126. Polleux F, Snider W. Initiating and growing an axon. *Cold Spring Harb Perspect Biol* (2010) 2:a001925. doi: 10.1101/cshperspect.a001925

127. Toda M, Suzuki T, Hosono K, Hayashi I, Hashiba S, Onuma Y, et al. Neuronal system-dependent facilitation of tumor angiogenesis and tumor growth by calcitonin gene-related peptide. *Proc Natl Acad Sci U.S.A.* (2008) 105:13550–5.

128. Balood M, Ahmadi M, Eichwald T, Ahmadi A, Majdoubi A, Roversi K, et al. Nociceptor neurons affect cancer immunosurveillance. *Nature* (2022) 611:405–12. doi: 10.1038/s41586-022-05374-w

129. Tu NH, Inoue K, Lewis PK, Khan A, Hwang JH, Chokshi V, et al. Calcitonin related polypeptide alpha mediates oral cancer pain. *Cells* (2023) 12. doi: 10.3390/ cells12131675

130. Russo AF, Hay DL. CGRP physiology, pharmacology, and therapeutic targets: migraine and beyond. *Physiol Rev* (2023) 103:1565-644. doi: 10.1152/ physrev.00059.2021

131. Jiang S-H, Hu L-P, Wang X, Li J, Zhang Z-G. Neurotransmitters: emerging targets in cancer. *Oncogene* (2020) 39:503–15. doi: 10.1038/s41388-019-1006-0

132. Wu Y, Berisha A, Borniger JC. Neuropeptides in cancer: friend and foe? Adv Biol (Weinh) (2022) 6:e2200111. doi: 10.1002/adbi.202200111

133. Molloy NH, Read DE, Gorman AM. Nerve growth factor in cancer cell death and survival. *Cancers (Basel)* (2011) 3:510–30. doi: 10.3390/cancers3010510

134. Demir IE, Tieftrunk E, Schorn S, Friess H, Ceyhan GO. Nerve growth factor & TrkA as novel therapeutic targets in cancer. *Biochim Biophys Acta* (2016) 1866:37–50. doi: 10.1016/j.bbcan.2016.05.003

135. Griffin N, Marsland M, Roselli S, Oldmeadow C, Attia J, Walker MM, et al. The receptor tyrosine kinase trkA is increased and targetable in HER2-positive breast cancer. *Biomolecules* (2020) 10. doi: 10.3390/biom10091329

136. Lagadec C, Meignan S, Adriaenssens E, Foveau B, Vanhecke E, Romon R, et al. TrkA overexpression enhances growth and metastasis of breast cancer cells. *Oncogene* (2009) 28:1960–70. doi: 10.1038/onc.2009.61

137. Descamps S, Lebourhis X, Delehedde M, Boilly B, Hondermarck H. Nerve growth factor is mitogenic for cancerous but not normal human breast epithelial cells. *J Biol Chem* (1998) 273:16659–62. doi: 10.1074/jbc.273.27.16659

138. Com E, Lagadec C, Page A, El Yazidi-Belkoura I, Slomianny C, Spencer A, et al. Nerve growth factor receptor TrkA signaling in breast cancer cells involves Ku70 to prevent apoptosis. *Mol Cell Proteomics* (2007) 6:1842–54. doi: 10.1074/mcp.M700119-MCP200

139. Krygier S, Djakiew D. Neurotrophin receptor p75(NTR) suppresses growth and nerve growth factor-mediated metastasis of human prostate cancer cells. *Int J Cancer* (2002) 98:1–7. doi: 10.1002/ijc.10160

140. Truzzi F, Marconi A, Lotti R, Dallaglio K, French LE, Hempstead BL, et al. Neurotrophins and their receptors stimulate melanoma cell proliferation and migration. J Invest Dermatol (2008) 128:2031–40. doi: 10.1038/jid.2008.21

141. Lam C-T, Yang Z-F, Lau C-K, Tam K-H, Fan S-T, Poon RTP. Brain-derived neurotrophic factor promotes tumorigenesis via induction of neovascularization: implication in hepatocellular carcinoma. *Clin Cancer Res* (2011) 17:3123–33. doi: 10.1158/1078-0432.CCR-10-2802

142. Zhu Y, Zhang C, Zhao D, Li W, Zhao Z, Yao S, et al. BDNF acts as a prognostic factor associated with tumor-infiltrating th2 cells in pancreatic adenocarcinoma. *Dis Markers* (2021) 2021:7842035. doi: 10.1155/2021/7842035

143. Zhang S-Y, Hui L-P, Li C-Y, Gao J, Cui Z-S, Qiu X-S. More expression of BDNF associates with lung squamous cell carcinoma and is critical to the proliferation and invasion of lung cancer cells. *BMC Cancer* (2016) 16:171. doi: 10.1186/s12885-016-2218-0

144. Yang X, Martin TA, Jiang WG. Biological influence of brain-derived neurotrophic factor (BDNF) on colon cancer cells. *Exp Ther Med* (2013) 6:1475–81. doi: 10.3892/etm.2013.1330

145. Meng L, Liu B, Ji R, Jiang X, Yan X, Xin Y. Targeting the BDNF/TrkB pathway for the treatment of tumors. *Oncol Lett* (2019) 17:2031–9.

146. Li Z, Xie J, Fei Y, Gao P, Xie Q, Gao W, et al. GDNF family receptor alpha 2 promotes neuroblastoma cell proliferation by interacting with PTEN. *Biochem Biophys Res Commun* (2019) 510:339–44. doi: 10.1016/j.bbrc.2018.12.169

147. Ito Y, Okada Y, Sato M, Sawai H, Funahashi H, Murase T, et al. Expression of glial cell line-derived neurotrophic factor family members and their receptors in pancreatic cancers. *Surgery* (2005) 138:788–94. doi: 10.1016/j.surg.2005.07.007

148. Chen M, Ba H, Lu C, Dai J, Sun J. Glial cell line-derived neurotrophic factor (GDNF) promotes angiogenesis through the demethylation of the fibromodulin (FMOD) promoter in glioblastoma. *Med Sci Monit* (2018) 24:6137–43. doi: 10.12659/MSM.911669

149. Huang S-M, Chen T-S, Chiu C-M, Chang L-K, Liao K-F, Tan H-M, et al. GDNF increases cell motility in human colon cancer through VEGF-VEGFR1 interaction. *Endocr Relat Cancer* (2014) 21:73–84. doi: 10.1530/ERC-13-0351

150. Lejmi E, Leconte L, Pédron-Mazoyer S, Ropert S, Raoul W, Lavalette S, et al. Netrin-4 inhibits angiogenesis via binding to neogenin and recruitment of Unc5B. *Proc Natl Acad Sci U.S.A.* (2008) 105:12491–6.

151. Chen J-Y, He X-X, Ma C, Wu X-M, Wan X-L, Xing Z-K, et al. Netrin-1 promotes glioma growth by activating NF-κB via UNC5A. *Sci Rep* (2017) 7:5454. doi: 10.1038/s41598-017-05707-0

152. Yin K, Dang S, Cui L, Fan X, Wang L, Xie R, et al. Netrin-1 promotes metastasis of gastric cancer by regulating YAP activity. *Biochem Biophys Res Commun* (2018) 496:76–82. doi: 10.1016/j.bbrc.2017.12.170

153. Eveno C, Broqueres-You D, Feron J-G, Rampanou A, Tijeras-Raballand A, Ropert S, et al. Netrin-4 delays colorectal cancer carcinomatosis by inhibiting tumor angiogenesis. *Am J Pathol* (2011) 178:1861–9. doi: 10.1016/j.ajpath.2010.12.019

154. Zhang K, An X, Zhu Y, Huang L, Yao X, Zeng X, et al. Netrin-1 inducing antiapoptotic effect of acute myeloid leukemia cells in a concentration-dependent manner through the Unc-5 netrin receptor B-focal adhesion kinase axis. *Cancer Biol Ther* (2023) 24:2200705. doi: 10.1080/15384047.2023.2200705

155. Reyes-Mugica M, Rieger-Christ K, Ohgaki H, Ekstrand BC, Helie M, Kleinman G, et al. Loss of DCC expression and glioma progression. *Cancer Res* (1997) 57:382–6.

156. Lou W, Wang W, Chen J, Wang S, Huang Y. ncRNAs-mediated high expression of SEMA3F correlates with poor prognosis and tumor immune infiltration of hepatocellular carcinoma. *Mol Ther Nucleic Acids* (2021) 24:845–55. doi: 10.1016/j.omtn.2021.03.014

157. Wallerius M, Wallmann T, Bartish M, Östling J, Mezheyeuski A, Tobin NP, et al. Guidance molecule SEMA3A restricts tumor growth by differentially regulating the proliferation of tumor-associated macrophages. *Cancer Res* (2016) 76:3166–78. doi: 10.1158/0008-5472.CAN-15-2596

158. Casazza A, Laoui D, Wenes M, Rizzolio S, Bassani N, Mambretti M, et al. Impeding macrophage entry into hypoxic tumor areas by Sema3A/Nrp1 signaling blockade inhibits angiogenesis and restores antitumor immunity. *Cancer Cell* (2013) 24:695–709. doi: 10.1016/j.ccr.2013.11.007

159. Gurrapu S, Pupo E, Franzolin G, Lanzetti L, Tamagnone L. Sema4C/PlexinB2 signaling controls breast cancer cell growth, hormonal dependence and tumorigenic potential. *Cell Death Differ* (2018) 25:1259–75. doi: 10.1038/s41418-018-0097-4

160. Bieniasz-Krzywiec P, Martín-Pérez R, Ehling M, García-Caballero M, Pinioti S, Pretto S, et al. Podoplanin-expressing macrophages promote lymphangiogenesis and lymphoinvasion in breast cancer. *Cell Metab* (2019) 30:917–936.e10. doi: 10.1016/j.cmet.2019.07.015

161. Muratori C, Tamagnone L. Semaphorin signals tweaking the tumor microenvironment. *Adv Cancer Res* (2012) 114:59-85. doi: 10.1016/B978-0-12-386503-8.00003-X

162. Gröne J, Doebler O, Loddenkemper C, Hotz B, Buhr H-J, Bhargava S. Robo1/ Robo4: differential expression of angiogenic markers in colorectal cancer. *Oncol Rep* (2006) 15:1437–43.

163. Zhou W-J, Geng ZH, Chi S, Zhang W, Niu X-F, Lan S-J, et al. Slit-Robo signaling induces Malignant transformation through Hakai-mediated E-cadherin degradation during colorectal epithelial cell carcinogenesis. *Cell Res* (2011) 21:609–26. doi: 10.1038/cr.2011.17

164. Latil A, Chêne L, Cochant-Priollet B, Mangin P, Fournier G, Berthon P, et al. Quantification of expression of netrins, slits and their receptors in human prostate tumors. *Int J Cancer* (2003) 103:306–15. doi: 10.1002/ijc.10821

165. Schmid BC, Rezniczek GA, Fabjani G, Yoneda T, Leodolter S, Zeillinger R. The neuronal guidance cue Slit2 induces targeted migration and may play a role in brain metastasis of breast cancer cells. *Breast Cancer Res Treat* (2007) 106:333–42. doi: 10.1007/s10549-007-9504-0

166. Narayan G, Goparaju C, Arias-Pulido H, Kaufmann AM, Schneider A, Dürst M, et al. Promoter hypermethylation-mediated inactivation of multiple Slit-Robo pathway genes in cervical cancer progression. *Mol Cancer* (2006) 5:16.

167. Dickinson RE, Fegan KS, Ren X, Hillier SG, Duncan WC. Glucocorticoid regulation of SLIT/ROBO tumour suppressor genes in the ovarian surface epithelium and ovarian cancer cells. *PloS One* (2011) 6:e27792.

168. Levi-Montalcini R. The nerve growth factor 35 years later. Science (1987) 237:1154-62.

169. Cantarella G, Lempereur L, Presta M, Ribatti D, Lombardo G, Lazarovici P, et al. Nerve growth factor-endothelial cell interaction leads to angiogenesis *in vitro* and *in vivo*. *FASEB J* (2002) 16:1307–9.

170. Rahbek UL, Dissing S, Thomassen C, Hansen AJ, Tritsaris K. Nerve growth factor activates aorta endothelial cells causing PI3K/Akt- and ERK-dependent migration. *Pflugers Arch* (2005) 450:355–61.

171. Romon R, Adriaenssens E, Lagadec C, Germain E, Hondermarck H, Le Bourhis X. Nerve growth factor promotes breast cancer angiogenesis by activating multiple pathways. *Mol Cancer* (2010) 9:157. doi: 10.1186/1476-4598-9-157

172. Brown MC, Staniszewska I, Lazarovici P, Tuszynski GP, Del Valle L, Marcinkiewicz C. Regulatory effect of nerve growth factor in alpha9beta1 integrindependent progression of glioblastoma. *Neuro Oncol* (2008) 10:968–80. doi: 10.1215/ 15228517-2008-0047 173. Kawasaki H, Goda M, Fukuhara S, Hashikawa-Hobara N, Zamami Y, Takatori S. Nerve growth factor (NGF) has an anti-tumor effects through perivascular innervation of neovessels in HT1080 fibrosarcoma and HepG2 hepatitis tumor in nude mice. *J Pharmacol Sci* (2019) 140:1–7. doi: 10.1016/j.jphs.2019.02.011

174. Goda M, Atagi S, Amitani K, Hobara N, Kitamura Y, Kawasaki H. Nerve growth factor suppresses prostate tumor growth. *J Pharmacol Sci* (2010) 112:463–6. doi: 10.1254/jphs.09354SC

175. Miranda M, Morici JF, Zanoni MB, Bekinschtein P. Brain-derived neurotrophic factor: A key molecule for memory in the healthy and the pathological brain. *Front Cell Neurosci* (2019) 13:363. doi: 10.3389/fncel.2019.00363

176. Kim H, Li Q, Hempstead BL, Madri JA. Paracrine and autocrine functions of brain-derived neurotrophic factor (BDNF) and nerve growth factor (NGF) in brainderived endothelial cells. *J Biol Chem* (2004) 279:33538–46.

177. Zhang Z, Zhang Y, Zhou Z, Shi H, Qiu X, Xiong J, et al. BDNF regulates the expression and secretion of VEGF from osteoblasts via the TrkB/ERK1/2 signaling pathway during fracture healing. *Mol Med Rep* (2017) 15:1362–7.

178. Kermani P, Rafii D, Jin DK, Whitlock P, Schaffer W, Chiang A, et al. Neurotrophins promote revascularization by local recruitment of TrkB+ endothelial cells and systemic mobilization of hematopoietic progenitors. *J Clin Invest* (2005) 115:653–63.

179. Lin C-Y, Hung S-Y, Chen H-T, Tsou H-K, Fong Y-C, Wang S-W, et al. Brainderived neurotrophic factor increases vascular endothelial growth factor expression and enhances angiogenesis in human chondrosarcoma cells. *Biochem Pharmacol* (2014) 91:522–33.

180. Lin C-Y, Hung S-Y, Chen H-T, Tsou H-K, Fong Y-C, Wang S-W, et al. Brainderived neurotrophic factor promotes VEGF-C-dependent lymphangiogenesis by suppressing miR-624-3p in human chondrosarcoma cells. *Cell Death Dis* (2017) 8: e2964.

181. Zheng W, Wang S-W, Chen Y-L, Chou W-Y, Lin T-Y, Chen W-C, et al. Overexpression of tyrosine kinase receptor B promotes metastasis of ovarian serous adenocarcinoma by lymphangiogenesis. *Tumori* (2011) 97:756–61. doi: 10.1177/030089161109700613

182. Sasahira T, Ueda N, Yamamoto K, Bhawal UK, Kurihara M, Kirita T, et al. Trks are novel oncogenes involved in the induction of neovascularization, tumor progression, and nodal metastasis in oral squamous cell carcinoma. *Clin Exp Metastasis* (2013) 30:165–76. doi: 10.1007/s10585-012-9525-x

183. Niland S, Eble JA. Neuropilins in the context of tumor vasculature. *Int J Mol Sci* (2019) 20. doi: 10.3390/ijms20030639

184. Pan Q, Chanthery Y, Liang W-C, Stawicki S, Mak J, Rathore N, et al. Blocking neuropilin-1 function has an additive effect with anti-VEGF to inhibit tumor growth. *Cancer Cell* (2007) 11:53–67. doi: 10.1016/j.ccr.2006.10.018

185. Powell J, Mota F, Steadman D, Soudy C, Miyauchi JT, Crosby S, et al. Small molecule neuropilin-1 antagonists combine antiangiogenic and antitumor activity with immune modulation through reduction of transforming growth factor beta (TGF β) production in regulatory T-cells. *J Med Chem* (2018) 61:4135–54. doi: 10.1021/acs.jmcdchem.8b00210

186. Huang Z, Cheng C, Xiong H, Wang Y, Chen KK, Yang J, et al. NRP1 promotes cell migration and invasion and serves as a therapeutic target in nasopharyngeal carcinoma. *Int J Clin Exp Pathol* (2018) 11:2460–9.

187. Gioelli N, Maione F, Camillo C, Ghitti M, Valdembri D, Morello N, et al. A rationally designed NRP1-independent superagonist SEMA3A mutant is an effective anticancer agent. *Sci Transl Med* (2018) 10:eaah4807. doi: 10.1126/scitranslmed.aah4807

188. Xia Y, Cai X-Y, Fan J-Q, Zhang L-L, Ren J-H, Li Z-Y, et al. The role of sema4D in vasculogenic mimicry formation in non-small cell lung cancer and the underlying mechanisms. *Int J Cancer* (2019) 144:2227–38. doi: 10.1002/ijc.31958

189. Angara K, Borin TF, Arbab AS. Vascular mimicry: A novel neovascularization mechanism driving anti-angiogenic therapy (AAT) resistance in glioblastoma. *Transl Oncol* (2017) 10:650–60. doi: 10.1016/j.tranon.2017.04.007

190. Hendrix MJC, Seftor EA, Seftor REB, Chao J-T, Chien D-S, Chu Y-W. Tumor cell vascular mimicry: Novel targeting opportunity in melanoma. *Pharmacol Ther* (2016) 159:83–92. doi: 10.1016/j.pharmthera.2016.01.006

191. Song Y, Zhao X-P, Song K, Shang Z-J. Ephrin-A1 is up-regulated by hypoxia in cancer cells and promotes angiogenesis of HUVECs through a coordinated cross-talk with eNOS. *PloS One* (2013) 8:e74464. doi: 10.1371/journal.pone.0074464

192. Sasabe E, Tomomura A, Tomita R, Sento S, Kitamura N, Yamamoto T. Ephrin-B2 reverse signaling regulates progression and lymph node metastasis of oral squamous cell carcinoma. *PloS One* (2017) 12:e0188965. doi: 10.1371/journal.pone.0188965

193. Merritt WM, Kamat AA, Hwang J-Y, Bottsford-Miller J, Lu C, Lin YG, et al. Clinical and biological impact of EphA2 overexpression and angiogenesis in endometrial cancer. *Cancer Biol Ther* (2010) 10:1306–14. doi: 10.4161/cbt.10.12.13582

194. Yamashita T, Ohneda K, Nagano M, Miyoshi C, Kaneko N, Miwa Y, et al. Hypoxia-inducible transcription factor-2alpha in endothelial cells regulates tumor neovascularization through activation of ephrin A1. *J Biol Chem* (2008) 283:18926–36. doi: 10.1074/jbc.M709133200

195. Dobrzanski P, Hunter K, Jones-Bolin S, Chang H, Robinson C, Pritchard S, et al. Antiangiogenic and antitumor efficacy of EphA2 receptor antagonist. *Cancer Res* (2004) 64:910–9. doi: 10.1158/0008-5472.CAN-3430-2

196. Tanaka M, Kamata R, Yanagihara K, Sakai R. Suppression of gastric cancer dissemination by ephrin-B1-derived peptide. *Cancer Sci* (2010) 101:87–93. doi: 10.1111/j.1349-7006.2009.01352.x

197. Yang X, Sun H, Tang T, Zhang W, Li Y. Netrin-1 promotes retinoblastomaassociated angiogenesis. Ann Transl Med (2021) 9:1683. doi: 10.21037/atm-21-5560

198. Akino T, Han X, Nakayama H, McNeish B, Zurakowski D, Mammoto A, et al. Netrin-1 promotes medulloblastoma cell invasiveness and angiogenesis, and demonstrates elevated expression in tumor tissue and urine of patients with pediatric medulloblastoma. *Cancer Res* (2014) 74:3716–26. doi: 10.1158/0008-5472.CAN-13-3116

199. Basu S, Nagy JA, Pal S, Vasile E, Eckelhoefer IA, Bliss VS, et al. The neurotransmitter dopamine inhibits angiogenesis induced by vascular permeability factor/vascular endothelial growth factor. *Nat Med* (2001) 7:569–74. doi: 10.1038/87895

200. Sarkar C, Chakroborty D, Mitra RB, Banerjee S, Dasgupta PS, Basu S. Dopamine *in vivo* inhibits VEGF-induced phosphorylation of VEGFR-2, MAPK, and focal adhesion kinase in endothelial cells. *Am J Physiol Heart Circ Physiol* (2004) 287:H1554–1560. doi: 10.1152/ajpheart.00272.2004

201. Sarkar C, Chakroborty D, Chowdhury UR, Dasgupta PS, Basu S. Dopamine increases the efficacy of anticancer drugs in breast and colon cancer preclinical models. *Clin Cancer Res* (2008) 14:2502–10. doi: 10.1158/1078-0432.CCR-07-1778

202. Chakroborty D, Sarkar C, Mitra RB, Banerjee S, Dasgupta PS, Basu S. Depleted dopamine in gastric cancer tissues: dopamine treatment retards growth of gastric cancer by inhibiting angiogenesis. *Clin Cancer Res* (2004) 10:4349–56. doi: 10.1158/1078-0432.CCR-04-0059

203. Asada M, Ebihara S, Numachi Y, Okazaki T, Yamanda S, Ikeda K, et al. Reduced tumor growth in a mouse model of schizophrenia, lacking the dopamine transporter. *Int J Cancer* (2008) 123:511-8. doi: 10.1002/ijc.23562

204. Basu S, Sarkar C, Chakroborty D, Nagy J, Mitra RB, Dasgupta PS, et al. Ablation of peripheral dopaminergic nerves stimulates Malignant tumor growth by inducing vascular permeability factor/vascular endothelial growth factor-mediated angiogenesis. *Cancer Res* (2004) 64:5551–5. doi: 10.1158/0008-5472.CAN-04-1600

205. Hoeppner LH, Wang Y, Sharma A, Javeed N, Van Keulen VP, Wang E, et al. Dopamine D2 receptor agonists inhibit lung cancer progression by reducing angiogenesis and tumor infiltrating myeloid derived suppressor cells. *Mol Oncol* (2015) 9:270–81. doi: 10.1016/j.molonc.2014.08.008

206. Chakroborty D, Chowdhury UR, Sarkar C, Baral R, Dasgupta PS, Basu S. Dopamine regulates endothelial progenitor cell mobilization from mouse bone marrow in tumor vascularization. *J Clin Invest* (2008) 118:1380–9. doi: 10.1172/JCI33125

207. Miyazawa A, Fujiyoshi Y, Unwin N. Structure and gating mechanism of the acetylcholine receptor pore. *Nature* (2003) 423:949–55. doi: 10.1038/nature01748

208. Wu JCF, Chruscinski A, De Jesus Perez VA, Singh H, Pitsiouni M, Rabinovitch M, et al. Cholinergic modulation of angiogenesis: role of the 7 nicotinic acetylcholine receptor. *J Cell Biochem* (2009) 108:433–46. doi: 10.1002/jcb.22270

209. Arias HR, Richards VE, Ng D, Ghafoori ME, Le V, Mousa SA. Role of nonneuronal nicotinic acetylcholine receptors in angiogenesis. *Int J Biochem Cell Biol* (2009) 41:1441–51. doi: 10.1016/j.biocel.2009.01.013

210. Dom AM, Buckley AW, Brown KC, Egleton RD, Marcelo AJ, Proper NA. The α 7-nicotinic acetylcholine receptor and MMP-2/-9 pathway mediate the proangiogenic effect of nicotine in human retinal endothelial cells. *Invest Ophthalmol Vis Sci* (2011) 52:4428–38. doi: 10.1167/iovs.10-5461

211. Wong HPS, Yu L, Lam EKY, Tai EKK, Wu WKK, Cho C-H. Nicotine promotes colon tumor growth and angiogenesis through beta-adrenergic activation. *Toxicol Sci* (2007) 97:279–87. doi: 10.1093/toxsci/kfm060

212. Shin VY, Wu WKK, Chu K-M, Wong HPS, Lam EKY, Tai EKK. Nicotine induces cyclooxygenase-2 and vascular endothelial growth factor receptor-2 in association with tumor-associated invasion and angiogenesis in gastric cancer. *Mol Cancer Res* (2005) 3:607–15. doi: 10.1158/1541-7786.MCR-05-0106

213. Natori T, Sata M, Washida M, Hirata Y, Nagai R, Makuuchi M. Nicotine enhances neovascularization and promotes tumor growth. *Mol Cells* (2003) 16:143–6. doi: 10.1016/S1016-8478(23)13780-0

214. Heeschen C, Weis M, Aicher A, Dimmeler S, Cooke JP. A novel angiogenic pathway mediated by non-neuronal nicotinic acetylcholine receptors. J Clin Invest (2002) 110:527–36. doi: 10.1172/JCI0214676

215. Zdanowski R, Leśniak M, Karczmarczyk U, Saracyn M, Bilski M, Kiepura A, et al. The effects of isopropyl methylphosphono-fluoridate (IMPF) poisoning on tumor growth and angiogenesis in BALB/C mice. *Ann Transplant* (2018) 23:105–11. doi: 10.12659/AOT.906548

216. Tyagi N, Lominadze D, Gillespie W, Moshal KS, Sen U, Rosenberger DS, et al. Differential expression of gamma-aminobutyric acid receptor A (GABA(A)) and effects of homocysteine. *Clin Chem Lab Med* (2007) 45:1777–84. doi: 10.1515/CCLM.2007.342

217. Negri S, Faris P, Soda T, Moccia F. Endothelial signaling at the core of neurovascular coupling: The emerging role of endothelial inward-rectifier K(+) (K (ir)2.1) channels and N-methyl-d-aspartate receptors in the regulation of cerebral blood flow. *Int J Biochem Cell Biol* (2021) 135:105983. doi: 10.1016/j.biocel.2021.105983

218. Park SH, Kim B-R, Lee JH, Park ST, Lee S-H, Dong SM, et al. GABARBP down-regulates HIF-1 α expression through the VEGFR-2 and PI3K/mTOR/4E-BP1 pathways. Cell Signal (2014) 26:1506–13. doi: 10.1016/j.cellsig.2014.03.017

219. Schuller HM, Al-Wadei HAN, Ullah MF, Plummer HK. 3rd. Regulation of pancreatic cancer by neuropsychological stress responses: a novel target for intervention. *Carcinogenesis* (2012) 33:191-6. doi: 10.1093/carcin/bgr251

220. Fava G, Marucci L, Glaser S, Francis H, De Morrow S, Benedetti A, et al. gamma-Aminobutyric acid inhibits cholangiocarcinoma growth by cyclic AMP-dependent regulation of the protein kinase A/extracellular signal-regulated kinase 1/ 2 pathway. *Cancer Res* (2005) 65:11437–46. doi: 10.1158/0008-5472.CAN-05-1470

221. Roberts SS, Mendonça-Torres MC, Jensen K, Francis GL, Vasko V. GABA receptor expression in benign and Malignant thyroid tumors. *Pathol Oncol Res* (2009) 15:645–50. doi: 10.1007/s12253-009-9165-x

222. Mohammad-Zadeh LF, Moses L, Gwaltney-Brant SM. Serotonin: a review. J Vet Pharmacol Ther (2008) 31:187–99. doi: 10.1111/j.1365-2885.2008.00944.x

223. Zhao D, Qin L, Bourbon P-M, James L, Dvorak HF, Zeng H. Orphan nuclear transcription factor TR3/Nur77 regulates microvessel permeability by targeting endothelial nitric oxide synthase and destabilizing endothelial junctions. *Proc Natl Acad Sci U.S.A.* (2011) 108:12066–71.

224. Iwabayashi M, Taniyama Y, Sanada F, Azuma J, Iekushi K, Kusunoki H, et al. Role of serotonin in angiogenesis: induction of angiogenesis by sarpogrelate via endothelial 5-HT1B/Akt/eNOS pathway in diabetic mice. *Atherosclerosis* (2012) 220:337–42. doi: 10.1016/j.atherosclerosis.2011.10.042

225. Matsusaka S, Wakabayashi I. 5-Hydroxytryptamine as a potent migration enhancer of human aortic endothelial cells. *FEBS Lett* (2005) 579:6721-5. doi: 10.1016/j.febslet.2005.10.068

226. Qin L, Zhao D, Xu J, Ren X, Terwilliger EF, Parangi S, et al. The vascular permeabilizing factors histamine and serotonin induce angiogenesis through TR3/ Nur77 and subsequently truncate it through thrombospondin-1. *Blood* (2013) 121:2154–64. doi: 10.1182/blood-2012-07-443903

227. Yang M, Li K, Ng PC, Chuen CKY, Lau TK, Cheng YS, et al. Promoting effects of serotonin on hematopoiesis: ex vivo expansion of cord blood CD34+ stem/ progenitor cells, proliferation of bone marrow stromal cells, and antiapoptosis. *Stem Cells* (2007) 25:1800–6. doi: 10.1634/stemcells.2007-0048

228. Pakala R, Benedict CR. Effect of serotonin and thromboxane A2 on endothelial cell proliferation: effect of specific receptor antagonists. *J Lab Clin Med* (1998) 131:527–37. doi: 10.1016/S0022-2143(98)90061-0

229. Zamani A, Qu Z. Serotonin activates angiogenic phosphorylation signaling in human endothelial cells. *FEBS Lett* (2012) 586:2360-5. doi: 10.1016/ j.febslet.2012.05.047

230. Nocito A, Dahm F, Jochum W, Jang JH, Georgiev P, Bader M, et al. Serotonin regulates macrophage-mediated angiogenesis in a mouse model of colon cancer allografts. *Cancer Res* (2008) 68:5152–8. doi: 10.1158/0008-5472.CAN-08-0202

231. Asada M, Ebihara S, Yamanda S, Niu K, Okazaki T, Sora I, et al. Depletion of serotonin and selective inhibition of 2B receptor suppressed tumor angiogenesis by inhibiting endothelial nitric oxide synthase and extracellular signal-regulated kinase 1/2 phosphorylation. *Neoplasia* (2009) 11:408–17. doi: 10.1593/neo.81630

232. Chevalier S, Defoy I, Lacoste J, Hamel L, Guy L, Bégin LR, et al. Vascular endothelial growth factor and signaling in the prostate: more than angiogenesis. *Mol Cell Endocrinol* (2002) 189:169–79. doi: 10.1016/S0303-7207(01)00728-6

233. Heinrich E, Trojan L, Friedrich D, Voss M, Weiss C, Michel MS, et al. Neuroendocrine tumor cells in prostate cancer: evaluation of the neurosecretory products serotonin, bombesin, and gastrin - impact on angiogenesis and clinical follow-up. *Prostate* (2011) 71:1752–8. doi: 10.1002/pros.21392

234. Watts SW, Davis RP. 5-hydroxtryptamine receptors in systemic hypertension: an arterial focus. *Cardiovasc Ther* (2011) 29:54-67. doi: 10.1111/j.1755-5922.2010.00173.x

235. Speyer CL, Hachem AH, Assi AA, Johnson JS, DeVries JA, Gorski DH. Metabotropic glutamate receptor-1 as a novel target for the antiangiogenic treatment of breast cancer. *PloS One* (2014) 9:e88830. doi: 10.1371/journal.pone.0088830

236. Xia H, Zhao Y-N, Yu C-H, Zhao Y-L, Liu Y. Inhibition of metabotropic glutamate receptor 1 suppresses tumor growth and angiogenesis in experimental non-small cell lung cancer. *Eur J Pharmacol* (2016) 783:103–11. doi: 10.1016/j.ejphar.2016.04.053

237. Wen Y, Li J, Koo J, Shin S-S, Lin Y, Jeong B-S, Mehnert JM, et al. Activation of the glutamate receptor GRM1 enhances angiogenic signaling to drive melanoma progression. *Cancer Res* (2014) 74:2499–509. doi: 10.1158/0008-5472.CAN-13-1531

238. Speyer CL, Smith JS, Banda M, DeVries JA, Mekani T, Gorski DH. Metabotropic glutamate receptor-1: a potential therapeutic target for the treatment of breast cancer. *Breast Cancer Res Treat* (2012) 132:565–73. doi: 10.1007/s10549-011-1624-x

239. Ziche M, Morbidelli L, Pacini M, Geppetti P, Alessandri G, Maggi CA. Substance P stimulates neovascularization *in vivo* and proliferation of cultured endothelial cells. *Microvasc Res* (1990) 40:264–78. doi: 10.1016/0026-2862(90)90024-L

240. Mohammadi F, Javid H, Afshari AR, Mashkani B, Hashemy SI. Substance P accelerates the progression of human esophageal squamous cell carcinoma via MMP-2, MMP-9, VEGF-A, and VEGFR1 overexpression. *Mol Biol Rep* (2020) 47:4263–72. doi: 10.1007/s11033-020-05532-1

241. Tilan J, Kitlinska J, Neuropeptide Y. (NPY) in tumor growth and progression: Lessons learned from pediatric oncology. *Neuropeptides* (2016) 55:55–66. doi: 10.1016/j.npep.2015.10.005

242. Movafagh S, Hobson JP, Spiegel S, Kleinman HK, Zukowska Z. Neuropeptide Y induces migration, proliferation, and tube formation of endothelial cells bimodally via Y1, Y2, and Y5 receptors. *FASEB J* (2006) 20:1924–6. doi: 10.1096/fj.05-4770fje

243. Lee EW, Michalkiewicz M, Kitlinska J, Kalezic I, Switalska H, Yoo P, et al. Neuropeptide Y induces ischemic angiogenesis and restores function of ischemic skeletal muscles. *J Clin Invest* (2003) 111:1853–62. doi: 10.1172/JCI16929

244. Lee EW, Grant DS, Movafagh S, Zukowska Z. Impaired angiogenesis in neuropeptide Y (NPY)-Y2 receptor knockout mice. *Peptides* (2003) 24:99–106. doi: 10.1016/S0196-9781(02)00281-4

245. Medeiros PJ, Jackson DN. Neuropeptide Y Y5-receptor activation on breast cancer cells acts as a paracrine system that stimulates VEGF expression and secretion to promote angiogenesis. *Peptides* (2013) 48:106–13. doi: 10.1016/j.peptides.2013.07.029

246. Alasvand M, Rashidi B, Javanmard SH, Akhavan MM, Khazaei M. Effect of blocking of neuropeptide Y Y2 receptor on tumor angiogenesis and progression in normal and diet-induced obese C57BL/6 mice. *Glob J Health Sci* (2015) 7:69–78.

247. Chakroborty D, Goswami S, Fan H, Frankel WL, Basu S, Sarkar C. Neuropeptide Y, a paracrine factor secreted by cancer cells, is an independent regulator of angiogenesis in colon cancer. *Br J Cancer* (2022) 127:1440–9.

248. Tilan JU, Lu C, Galli S, Izycka-Swieszewska E, Earnest JP, Shabbir A, et al. Hypoxia shifts activity of neuropeptide Y in Ewing sarcoma from growth-inhibitory to growth-promoting effects. *Oncotarget* (2013) 4:2487–501.

249. Lu C, Everhart L, Tilan J, Kuo L, Sun C-CJ, Munivenkatappa RB, et al. Neuropeptide Y and its Y2 receptor: potential targets in neuroblastoma therapy. *Oncogene* (2010) 29:5630–42.

250. Czarnecka M, Trinh E, Lu C, Kuan-Celarier A, Galli S, Hong S-H, et al. Neuropeptide Y receptor Y5 as an inducible pro-survival factor in neuroblastoma: implications for tumor chemoresistance. *Oncogene* (2015) 34:3131–43.

251. Russell FA, King R, Smillie S-J, Kodji X, Brain SD. Calcitonin gene-related peptide: physiology and pathophysiology. *Physiol Rev* (2014) 94:1099-142. doi: 10.1152/physrev.00034.2013

252. Haegerstrand A, Dalsgaard CJ, Jonzon B, Larsson O, Nilsson J. Calcitonin generelated peptide stimulates proliferation of human endothelial cells. *Proc Natl Acad Sci* U.S.A. (1990) 87:3299–303.

253. Brain SD, Grant AD. Vascular actions of calcitonin gene-related peptide and adrenomedullin. *Physiol Rev* (2004) 84:903–34. doi: 10.1152/physrev.00037.2003

254. Dong Y-L, Reddy DM, Green KE, Chauhan MS, Wang H-Q, Nagamani M, et al. Calcitonin gene-related peptide (CALCA) is a proangiogenic growth factor in the human placental development. *Biol Reprod* (2007) 76:892–9. doi: 10.1095/biolreprod.106.059089

255. Toda M, Suzuki T, Hosono K, Kurihara Y, Kurihara H, Hayashi I, et al. Roles of calcitonin gene-related peptide in facilitation of wound healing and angiogenesis. *BioMed Pharmacother* (2008) 62:352–9. doi: 10.1016/j.biopha.2008.02.003

256. Zhang Q, Guo Y, Chen H, Jiang Y, Tang H, Gong P, et al. The influence of receptor activity-modifying protein-1 overexpression on angiogenesis in mouse brain capillary endothelial cells. *J Cell Biochem* (2019) 120:10087–96. doi: 10.1002/jcb.28292

257. Kurashige C, Hosono K, Matsuda H, Tsujikawa K, Okamoto H, Majima M. Roles of receptor activity-modifying protein 1 in angiogenesis and lymphangiogenesis during skin wound healing in mice. *FASEB J* (2014) 28:1237–47. doi: 10.1096/fj.13-238998

258. Fisher SA. Vascular smooth muscle phenotypic diversity and function. *Physiol Genomics* (2010) 42A:169–187. doi: 10.1152/physiolgenomics.00111.2010

259. Zahalka AH, Arnal-Estapé A, Maryanovich M, Nakahara F, Cruz CD, Finley LWS, et al. Adrenergic nerves activate an angio-metabolic switch in prostate cancer. *Science* (2017) 358:321–6. doi: 10.1126/science.aah5072

260. Renz BW, Takahashi R, Tanaka T, Macchini M, Hayakawa Y, Dantes Z, et al. β 2 adrenergic-neurotrophin feedforward loop promotes pancreatic cancer. *Cancer Cell* (2018) 33:75–90.e7. doi: 10.1016/j.ccell.2017.11.007

261. Thaker PH, Han LY, Kamat AA, Arevalo JM, Takahashi R, Lu C, et al. Chronic stress promotes tumor growth and angiogenesis in a mouse model of ovarian carcinoma. *Nat Med* (2006) 12:939–44. doi: 10.1038/nm1447

262. Yang EV, Kim S, Donovan EL, Chen M, Gross AC, Webster Marketon JI, et al. Norepinephrine upregulates VEGF, IL-8, and IL-6 expression in human melanoma tumor cell lines: implications for stress-related enhancement of tumor progression. *Brain Behav Immun* (2009) 23:267–75. doi: 10.1016/j.bbi.2008.10.005

263. Madden KS, Szpunar MJ, Brown EB. β -Adrenergic receptors (β -AR) regulate VEGF and IL-6 production by divergent pathways in high β -AR-expressing breast cancer cell lines. *Breast Cancer Res Treat* (2011) 130:747–58. doi: 10.1007/s10549-011-1348-y

264. Tan KS, Nackley AG, Satterfield K, Maixner W, Diatchenko L, Flood PM. Beta2 adrenergic receptor activation stimulates pro-inflammatory cytokine production in macrophages via PKA- and NF-kappaB-independent mechanisms. *Cell Signal* (2007) 19:251–60. doi: 10.1016/j.cellsig.2006.06.007

265. He J-J, Zhang W-H, Liu S-L, Chen Y-F, Liao C-X, Shen Q-Q, et al. Activation of β -adrenergic receptor promotes cellular proliferation in human glioblastoma. *Oncol Lett* (2017) 14:3846–52. doi: 10.3892/ol.2017.6653

266. Seegers HC, Hood VC, Kidd BL, Cruwys SC, Walsh DA. Enhancement of angiogenesis by endogenous substance P release and neurokinin-1 receptors during neurogenic inflammation. *J Pharmacol Exp Ther* (2003) 306:8–12. doi: 10.1124/jpet.103.050013

267. Momen Razmgah M, Ghahremanloo A, Javid H, AlAlikhan A, Afshari A-R, Hashemy SI. The effect of substance P and its specific antagonist (aprepitant) on the expression of MMP-2, MMP-9, VEGF, and VEGFR in ovarian cancer cells. *Mol Biol Rep* (2022) 49:9307–14. doi: 10.1007/s11033-022-07771-w

268. Rodriguez PL, Jiang S, Fu Y, Avraham S, Avraham HK. The proinflammatory peptide substance P promotes blood-brain barrier breaching by breast cancer cells through changes in microvascular endothelial cell tight junctions. *Int J Cancer* (2014) 134:1034–44. doi: 10.1002/ijc.28433

269. Leroux A, Paiva Dos Santos B, Leng J, Oliveira H, Amédée J. Sensory neurons from dorsal root ganglia regulate endothelial cell function in extracellular matrix remodelling. *Cell Commun Signal* (2020) 18:162. doi: 10.1186/s12964-020-00656-0

270. Costa PAC, Silva WN, Prazeres PHDM, Picoli CC, Guardia GDA, Costa AC, et al. Chemogenetic modulation of sensory neurons reveals their regulating role in melanoma progression. *Acta Neuropathol Commun* (2021) 9:183. doi: 10.1186/s40478-021-01273-9

271. Hu B, Lv X, Chen H, Xue P, Gao B, Wang X, et al. Sensory nerves regulate mesenchymal stromal cell lineage commitment by tuning sympathetic tones. *J Clin Invest* (2020) 130:3483–98.

272. Fitzgerald PJ. Norepinephrine release may play a critical role in the Warburg effect: an integrative model of tumorigenesis. *Neoplasma* (2020) 67:947–57.

273. Chakroborty D, Sarkar C, Basu B, Dasgupta PS, Basu S. Catecholamines regulate tumor angiogenesis. *Cancer Res* (2009) 69:3727–30.

274. Montgomery MK, Kim SH, Dovas A, Zhao HT, Goldberg AR, Xu W, et al. Glioma-induced alterations in neuronal activity and neurovascular coupling during disease progression. *Cell Rep* (2020) 31:107500.

275. Chow DS, Horenstein CI, Canoll P, Lignelli A, Hillman EMC, Filippi CG, et al. Glioblastoma induces vascular dysregulation in nonenhancing peritumoral regions in humans. *AJR Am J Roentgenol* (2016) 206:1073–81.

276. Stackhouse TL, Mishra A. Neurovascular coupling in development and disease: focus on astrocytes. Front Cell Dev Biol (2021) 9:702832. doi: 10.3389/fcell.2021.702832

277. Brown LS, Foster CG, Courtney J-M, King NE, Howells DW, Sutherland BA. Pericytes and neurovascular function in the healthy and diseased brain. *Front Cell Neurosci* (2019) 13:282. doi: 10.3389/fncel.2019.00282

278. Pak RW, Hadjiabadi DH, Senarathna J, Agarwal S, Thakor NV, Pillai JJ, et al. Implications of neurovascular uncoupling in functional magnetic resonance imaging (fMRI) of brain tumors. *J Cereb Blood Flow Metab* (2017) 37:3475–87. doi: 10.1177/0271678X17707398

279. Watkins S, Robel S, Kimbrough IF, Robert SM, Ellis-Davies G, Sontheimer H. Disruption of astrocyte-vascular coupling and the blood-brain barrier by invading glioma cells. *Nat Commun* (2014) 5:4196. doi: 10.1038/ncomms5196

280. Farin A, Suzuki SO, Weiker M, Goldman JE, Bruce JN, Canoll P. Transplanted glioma cells migrate and proliferate on host brain vasculature: a dynamic analysis. *Glia* (2006) 53:799–808. doi: 10.1002/glia.20334

281. Liebner S, Fischmann A, Rascher G, Duffner F, Grote EH, Kalbacher H, et al. Claudin-1 and claudin-5 expression and tight junction morphology are altered in blood vessels of human glioblastoma multiforme. *Acta Neuropathol* (2000) 100:323–31. doi: 10.1007/s004010000180

282. Mauffrey P, Tchitchek N, Barroca V, Bemelmans A-P, Firlej V, Allory Y, et al. Progenitors from the central nervous system drive neurogenesis in cancer. *Nature* (2019) 569:672–8. doi: 10.1038/s41586-019-1219-y

283. Magnon C, Hall SJ, Lin J, Xue X, Gerber L, Freedland SJ, et al. Autonomic nerve development contributes to prostate cancer progression. *Science* (2013) 341:1236361. doi: 10.1126/science.1236361

284. Lu R, Fan C, Shangguan W, Liu Y, Li Y, Shang Y, et al. Neurons generated from carcinoma stem cells support cancer progression. *Signal Transduct Target Ther* (2017) 2:16036. doi: 10.1038/sigtrans.2016.36

285. Zhang D, Park D, Zhong Y, Lu Y, Rycaj K, Gong S, et al. Stem cell and neurogenic gene-expression profiles link prostate basal cells to aggressive prostate cancer. *Nat Commun* (2016) 7:10798. doi: 10.1038/ncomms10798