



OPEN ACCESS

EDITED AND REVIEWED BY
Francesco Pezzella,
University of Oxford, United Kingdom

*CORRESPONDENCE

Vijay Kumar
✉ vij_tox@yahoo.com

†PRESENT ADDRESS

Vijay Kumar,
Department of Surgery, Laboratory of
Tumor Immunology and Immunotherapy,
Morehouse School of Medicine, Atlanta,
GA, United States

RECEIVED 17 September 2023

ACCEPTED 25 September 2023

PUBLISHED 05 October 2023

CITATION

Kumar V (2023) Editorial: Manipulation of
immune-vascular crosstalk in solid tumors.
Front. Immunol. 14:1295953.
doi: 10.3389/fimmu.2023.1295953

COPYRIGHT

© 2023 Kumar. This is an open-access
article distributed under the terms of the
[Creative Commons Attribution License
\(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use, distribution or
reproduction in other forums is permitted,
provided the original author(s) and the
copyright owner(s) are credited and that
the original publication in this journal is
cited, in accordance with accepted
academic practice. No use, distribution or
reproduction is permitted which does not
comply with these terms.

Editorial: Manipulation of immune-vascular crosstalk in solid tumors

Vijay Kumar*†

Department of Interdisciplinary Oncology, School of Medicine, Stanley S. Scott Cancer Center,
Louisiana State University Health Science Center (LSUHSC), New Orleans, LA, United States

KEYWORDS

tumor immune microenvironment, tumor immunology, solid tumor, tumor vascular
networks, tumor immunity, immunotherapy, immune check inhibitor (ICI)

Editorial on the Research Topic

Manipulation of immune-vascular crosstalk in solid tumors

Advances in modern sciences such as molecular biology, genetics, and immunology have equipped us to eradicate different infectious diseases (smallpox, polio) known from ancient times. However, despite these advances, cancer is the number one killer of humans. For example, in 2023, 1,958,310 new cancer cases and 609,820 cancer deaths may occur only in the United States (1). Immune-checkpoint inhibitors (ICIs) are the latest breakthrough in cancer treatment with existing chemo and radiotherapies, which are not without side effects (2–5). Furthermore, ICIs usually benefit less than 15% of cancer patients with adverse events in a large number of patients (6). Chimeric antigen receptor (CAR) T cells (especially autologous CAR-T cells) targeting tumor-associated antigen (TAAs) has transformed the treatment of many hematological cancers (CD19 CAR T cells for leukemia) and also have a high therapeutic potential for solid tumors (7–10). Besides toxic (immune and nervous system-mediated) events associated with CAR-T cells-based immunotherapies, another hurdle in their success in treating solid cancers is resistance development and cancer cell escape in the immunosuppressive tumor microenvironment (TME) or tumor immune microenvironment (TIME) (11–13). Furthermore, allogenic CAR-T cell immunotherapy may induce severe graft-versus-host disease (GVHD) and can be easily eliminated by the immune system (9). Therefore, we have to move forward to find alternative approaches to target cancers with minimum or no side/adverse events.

Blood vascular and lymphoid systems are critical for nutrition supply and immune cell infiltration in the TME or TIME (14, 15). The immune and vascular cross-talk is also fully consistent with the seed (cancer cell) and soil (specific organ environment) hypothesis given by Stephen Paget in 1899, approximately 150 years back, to explain conditions creating an environment for metastasis (16, 17). For example, tumor metastasis depends on several factors promoting cancer cell growth, proliferation, survival, immune escape, nutrition supply, and local and distant tissue invasion. Blood vascular endothelium (VE) and lymphatic endothelium (LE) differ due to their different roles that also depend on the specific tissue environment (18–20). Of note, many cancers, such as breast cancer and melanoma, preferentially spread via lymphatics (21). Furthermore, primary tumors, like blood vasculature, also modify lymphatic vessels and draining lymph nodes (dLNs) by

lymph-angiogenesis, incorporating myeloid cells into lymphatics and delivering exosome or extracellular vesicles (EVs) to dLNs for generating pretumoral niches for cancer cell metastasis (15, 22–25).

A strong connection between blood vasculature and lymphatics has emerged that can change tumor biology and immunology than previously expected (26–30). Furthermore, the cross-talk between endothelial cells, immune cells, and immune checkpoints in the TME is emerging with a therapeutic potential (31). For example, a combination of anti-angiogenic therapy and ICIs normalizes vascular-immune cross-talk to increase the antitumor immunity (32). Therefore, understanding immune-vascular cross-talk in the TME or TIME of solid cancers is critical for designing better therapies. The current Research Topic of the thematic issue with four articles focusses on this less explored area of tumor immunology and vascularization.

For example, the research article by Yang et al. in this Research Topic has explored that the efficacy of ICIs such as anti-programmed cell death protein-1 (PD-1 or CD279) antibodies increases in combination with the anti-angiogenesis drug, lenvatinib (a multi-receptor tyrosine kinase inhibitor, including vascular endothelial growth factor receptor-1 (VEGFR1), VEGFR2, and VEGFR3) against hepatocellular carcinoma (HCC). The increased efficacy of the ICI with lenvatinib occurred due to the normalization of the tumor vasculature and increased antitumor immune cell infiltration in the TME/TIME. For example, at adequate doses, lenvatinib increases the integrity among endothelial cells and prevents vascular leakage. Lenvatinib treatment maintains the endothelial cell integrity by forming the neuropilin-1 (NRP-1)-platelet-derived growth factor receptor- β (PDGFR- β) complex, activating a novel cdc-2 related protein kinase 1 (Crkl)-C3G (a guanine nucleotide exchange factor for Ras-associated protein 1 or Rap1, which is a small GTPase)-Rap1 signaling pathway in endothelial cells. The details of Rap-1 signaling in cancer are discussed elsewhere (33, 34). The NRP-1/PDGFR- β complex also promotes the interaction between endothelial cells and pericytes by inducing tyrosine-phosphorylation in pericytes. Thus, normovascularization by the lenvatinib in the TME increased the antitumor activity of the ICIs to suppress HCC.

Another study in the Research Topic by Liu et al., has indicated that blocking the interaction between microRNA (miR)-27a and VE-cadherin with Blockmir CD5-2, a novel oligonucleotide-based miR-27a inhibitor inhibits angiogenesis and its combination with ICIs (anti-PD-1s) significantly reduce the HCC tumor size. CD5-2 takes care of leaky blood vessels and reduces tumor hypoxia, and ICIs increase the infiltration of antitumor immune cells in the TME or TIME. Thus, combining anti-angiogenesis approach and ICIs proves beneficial to the host to decrease the tumor burden. Therefore, different approaches combining anti-angiogenesis and ICIs are emerging to target solid cancer for developing a field called tumor vasculoimmunology, where tumor vasculature and immune environment would be studied and targeted together for lowering the tumor burden and metastasis.

Furthermore, the review article by Dianat-Moghadam et al., discusses about failure of anti-angiogenesis therapies as anticancer drugs when used alone as they completely deplete vascularization,

inducing hypoxia, drug resistance, and tumor recurrence along with negatively impacting chemotherapies and immune cell infiltration in the TME/TIME. They have further discussed different challenges to target tumor vasculature and its normalization for improving cancer immunotherapies. The authors have further discussed that nanomedicine-based tumor vasculature targeting and normalization has a bright future to act as potent antitumor molecules to enhance the efficacy of existing immunotherapies such as ICIs.

I have mentioned earlier that VE or vascular endothelial cells (VECs) differ from LE or lymphatic endothelial cells (LECs) and vice versa. The fourth article in the Research Topic by Viudez-Pareja et al. discuss the immunomodulatory properties of LE in the TME/TIME. LE or LECs have unique properties to suppress or potentiate the antitumor immune response. For example, tumor-associated lymphangiogenesis can promote tumor dissemination and metastasis, as seen in breast cancers and melanoma. Viudez-Pareza et al., discuss the immunomodulatory properties of LE within the TME/TIME of primary tumors and tumor-dLNs. They have further discussed emerging approaches to target tumor LE or LECs to enhance antitumor immune response. Hence, understating the cross-talk between LE, VE, and immune response or cells in the TME/TIME may increase the efficacy of currently available immunotherapies, such as ICIs. Additionally, it will be interesting to study the impact of immune-vascular cross-talk targeting on cellular immunotherapies, including chimeric antigen receptor-T (CAR-T) cells. Therefore, immune-vascular cross-talk has a bright future in understanding the TIME and enhancing the efficacies of current immunotherapies.

Author contributions

VK: Conceptualization, Writing – original draft, Writing – review & editing.

Conflict of interest

The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The author declared that they were an editorial board member of Frontiers, at the time of submission. This had no impact on the peer review process and the final decision.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

- Siegel RL, Miller KD, Wagle NS, Jemal A. Cancer statistics, 2023. *CA: A Cancer J Clin* (2023) 73:17–48. doi: 10.3322/caac.21763
- Postow MA, Callahan MK, Wolchok JD. Immune checkpoint blockade in cancer therapy. *J Clin Oncol* (2015) 33:1974–82. doi: 10.1200/JCO.2014.59.4358
- Robert C. A decade of immune-checkpoint inhibitors in cancer therapy. *Nat Commun* (2020) 11:3801. doi: 10.1038/s41467-020-17670-y
- Martins F, Sofiya L, Sykietis GP, Lamine F, Maillard M, Fraga M, et al. Adverse effects of immune-checkpoint inhibitors: epidemiology, management and surveillance. *Nat Rev Clin Oncol* (2019) 16:563–80. doi: 10.1038/s41571-019-0218-0
- Sullivan RJ, Weber JS. Immune-related toxicities of checkpoint inhibitors: mechanisms and mitigation strategies. *Nat Rev Drug Discov* (2022) 21:495–508. doi: 10.1038/s41573-021-00259-5
- Munn LL, Jain RK. Vascular regulation of antitumor immunity. *Science* (2019) 365:544–5. doi: 10.1126/science.aaw7875
- D'Aloia MM, Zizzari IG, Sacchetti B, Pierelli L, Alimandi M. CAR-T cells: the long and winding road to solid tumors. *Cell Death Dis* (2018) 9:282. doi: 10.1038/s41419-018-0278-6
- Rodriguez-Garcia A, Palazon A, Noguera-Ortega E, Powell DJ Jr., Guedan S. CAR-T cells hit the tumor microenvironment: strategies to overcome tumor escape. *Front Immunol* (2020) 11:1109. doi: 10.3389/fimmu.2020.01109
- Depil S, Duchateau P, Grupp SA, Mufti G, Poirot L. 'Off-the-shelf' allogeneic CAR T cells: development and challenges. *Nat Rev Drug Discov* (2020) 19:185–99. doi: 10.1038/s41573-019-0051-2
- Newick K, O'Brien S, Moon E, Albelda SM. CAR T cell therapy for solid tumors. *Annu Rev Med* (2017) 68:139–52. doi: 10.1146/annurev-med-062315-120245
- Donnadieu E, Duprè L, Pinho LG, Cotta-de-Almeida V. Surmounting the obstacles that impede effective CAR T cell trafficking to solid tumors. *J Leukoc Biol* (2020) 108:1067–79. doi: 10.1002/JLB.1MR0520-746R
- Grinda T, Brouard J, Tran D, Rubio MT. Mechanisms of resistance and escape to CAR-T cells. *Bull Cancer* (2021) 108:S128–s140. doi: 10.1016/j.bulcan.2021.09.002
- Labanieh L, Majzner RG, Mackall CL. Programming CAR-T cells to kill cancer. *Nat BioMed Eng* (2018) 2:377–91. doi: 10.1038/s41551-018-0235-9
- Schaaf MB, Garg AD, Agostinis P. Defining the role of the tumor vasculature in antitumor immunity and immunotherapy. *Cell Death Dis* (2018) 9:115. doi: 10.1038/s41419-017-0061-0
- Takeda A, Salmi M, Jalkanen S. Lymph node lymphatic endothelial cells as multifaceted gatekeepers in the immune system. *Trends Immunol* (2023) 44:72–86. doi: 10.1016/j.it.2022.10.010
- Fidler IJ. The pathogenesis of cancer metastasis: the 'seed and soil' hypothesis revisited. *Nat Rev Cancer* (2003) 3:453–8. doi: 10.1038/nrc1098
- Akhtar M, Haider A, Rashid S, Al-Nabet A. Paget's "Seed and soil" Theory of cancer metastasis: an idea whose time has come. *Adv Anat Pathol* (2019) 26:69–74. doi: 10.1097/PAP.0000000000000219
- Amatschek S, Kriehuber E, Bauer W, Reininger B, Meraner P, Wolpl A, et al. Blood and lymphatic endothelial cell-specific differentiation programs are stringently controlled by the tissue environment. *Blood* (2007) 109:4777–85. doi: 10.1182/blood-2006-10-053280
- Podgrabsinska S, Braun P, Velasco P, Kloos B, Pepper MS, Skobe M. Molecular characterization of lymphatic endothelial cells. *Proc Natl Acad Sci USA* (2002) 99:16069–74. doi: 10.1073/pnas.242401399
- Pepper MS, Skobe M. Lymphatic endothelium: morphological, molecular and functional properties. *J Cell Biol* (2003) 163:209–13. doi: 10.1083/jcb.200308082
- Leong SP, Pissas A, Scarato M, Gallon F, Pissas MH, Amore M, et al. The lymphatic system and sentinel lymph nodes: conduit for cancer metastasis. *Clin Exp Metastasis* (2022) 39:139–57. doi: 10.1007/s10585-021-10123-w
- Li YL, Hung WC. Reprogramming of sentinel lymph node microenvironment during tumor metastasis. *J BioMed Sci* (2022) 29:84. doi: 10.1186/s12929-022-00868-1
- Leong SP, Naxerova K, Keller L, Pantel K, Witte M. Molecular mechanisms of cancer metastasis via the lymphatic versus the blood vessels. *Clin Exp Metastasis* (2022) 39:159–79. doi: 10.1007/s10585-021-10120-z
- Karpanen T, Alitalo K. Molecular biology and pathology of lymphangiogenesis. *Annu Rev Pathol* (2008) 3:367–97. doi: 10.1146/annurev.pathmechdis.3.121806.151515
- Trisko J, Fleck J, Kau S, Oesterreicher J, Holthöner W. Lymphatic and blood endothelial extracellular vesicles: A story yet to be written. *Life* (2022) 12. doi: 10.3390/life12050654
- Lampejo AO, Ghavimi SAA, Hägerling R, Agarwal S, Murfee WL. Lymphatic/blood vessel plasticity: motivation for a future research area based on present and past observations. *Am J Physiol Heart Circ Physiol* (2023) 324:H109–h121. doi: 10.1152/ajpheart.00612.2022
- Martinez-Corral I, Makinen T. Regulation of lymphatic vascular morphogenesis: Implications for pathological (tumor) lymphangiogenesis. *Exp Cell Res* (2013) 319:1618–25. doi: 10.1016/j.yexcr.2013.01.016
- Kahn BM, Lucas A, Alur RG, Wengyn MD, Schwartz GW, Li J, et al. The vascular landscape of human cancer. *J Clin Invest* (2021) 131. doi: 10.1172/JCI136655
- Takakura N. Vascular reconstitution in the tumor for more effective tumor immunotherapy. *Cancer Sci* (2021) 112:1348–56. doi: 10.1111/cas.14854
- Mpekris F, Voutouri C, Baish JW, Duda DG, Munn LL, Stylianopoulos T, et al. Combining microenvironment normalization strategies to improve cancer immunotherapy. *Proc Natl Acad Sci* (2020) 117:3728–37. doi: 10.1073/pnas.1919764117
- Fang J, Lu Y, Zheng J, Jiang X, Shen H, Shang X, et al. Exploring the crosstalk between endothelial cells, immune cells, and immune checkpoints in the tumor microenvironment: new insights and therapeutic implications. *Cell Death Dis* (2023) 14:586. doi: 10.1038/s41419-023-06119-x
- Lee WS, Yang H, Chon HJ, Kim C. Combination of anti-angiogenic therapy and immune checkpoint blockade normalizes vascular-immune crosstalk to potentiate cancer immunity. *Exp Mol Med* (2020) 52:1475–85. doi: 10.1038/s12276-020-00500-y
- Zhang YL, Wang RC, Cheng K, Ring BZ, Su L. Roles of Rap1 signaling in tumor cell migration and invasion. *Cancer Biol Med* (2017) 14:90–9. doi: 10.20892/j.issn.2095-3941.2016.0086
- Gloerich M, Bos JL. Regulating Rap small G-proteins in time and space. *Trends Cell Biol* (2011) 21:615–23. doi: 10.1016/j.tcb.2011.07.001