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*CORRESPONDENCE Shuqi Qiu drqiusq@163.com Hailiang Zhao drzhaohl@163.com Jiangqi Liu drliujq@163.com

[†]These authors have contributed equally to this work

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Tolerogenic dendritic cells and TLR4/IRAK4/NF-kB signaling pathway in allergic rhinitis

Chenglin Kang^{1,2,3†}, Xiaomei Li^{3†}, Peng Liu¹, Yue Liu¹, Yuan Niu⁴, Xianhai Zeng^{1,2}, Hailiang Zhao^{1,2*}, Jiangqi Liu^{1,2*} and Shuqi Qiu^{1,2*}

¹Department of Graduate and Scientific Research, Zunyi Medical University Zhuhai Campus, Zhuhai, China, ²Department of Otolaryngology, Longgang E.N.T Hospital and Shenzhen Key Laboratory of E.N.T, Institute of E.N.T Shenzhen, Shenzhen, China, ³Department of Otolaryngology, Second People's Hospital of Gansu Province, Lanzhou, China, ⁴Department of Neurology, Second People's Hospital of Gansu Province, Lanzhou, China

Dendritic cells (DCs), central participants in the allergic immune response, can capture and present allergens leading to allergic inflammation in the immunopathogenesis of allergic rhinitis (AR). In addition to initiating antigen-specific immune responses, DCs induce tolerance and modulate immune homeostasis. As a special type of DCs, tolerogenic DCs (tolDCs) achieve immune tolerance mainly by suppressing effector T cell responses and inducing regulatory T cells (Tregs). TolDCs suppress allergic inflammation by modulating immune tolerance, thereby reducing symptoms of AR. Activation of the TLR4/IRAK4/NF- κ B signaling pathway contributes to the release of inflammatory cytokines, and inhibitors of this signaling pathway induce the production of tolDCs to alleviate allergic inflammatory responses. This review focuses on the relationship between tolDCs and TLR4/IRAK4/NF- κ B signaling pathway with AR.

KEYWORDS

tolerogenic dendritic cells, TLR4/IRAK4/NF- κ B signaling pathway, immune response, immune tolerance, allergic rhinitis

1 Introduction

Dendritic cells (DCs) are bridges between innate and adaptive immune responses, they not only act as key immunomodulatory factors driving T cell initiation and activation, but also have tolerogenic functions that promote immune tolerance (1-3). Whether DCs exhibit immunogenicity or tolerance depends on their different subsets of them and the different stimuli to which they are exposed (4). Exposure of DCs to allergens leads to enhanced priming, which further mediates allergic inflammatory responses (5, 6). However, repeated low-dose exposure to allergens has the potential to make DCs tolerant (7). Tolerogenic DCs (tolDCs) are DCs with immunomodulatory functions that slow the allergic response by killing T cells (7). This would be beneficial in the treatment of AR.

DCs play an integral role in the pathogenesis and treatment of allergic rhinitis (AR). DCs recognize pathogen-associated molecular patterns (PAMPs) or lipopolysaccharide (LPS) through Toll-like receptor 4 (TLR4) expressed on their membrane to trigger signaling cascades (2, 6, 8). TLR4/nuclear factor kappa-B (NF- κ B) regulates the balance of T helper type 1 (Th1) and T helper type 17 (Th17) by affecting the maturation and migration of DCs, which in turn affects the development of AR (9). The TLR4/interleukin-1 receptor-associated kinase 4 (IRAK4)/NF- κ B signaling pathway eventually leads to the production of pro-inflammatory cytokines that initiate inflammatory and immune responses (10). Blocking the propagation of the TLR4/IRAK4/NF- κ B signaling pathway with tolDCs may become a therapeutic target for AR.

2 Overview of DCs

DCs can be categorized into conventional DCs (cDCs) and plasmacytoid DCs (pDCs) according to their morphological features and function (2, 11). pCDs, primarily found in lymphoid organs, are characterized by the production of large amounts of type I interferon (IFN) after recognition of foreign nucleic acids (2, 12). In addition to producing IFN, pDCs also secrete pro-inflammatory cytokines and chemokines such as interleukin-6 (IL-6), IL-12, CXC-chemokine ligand 8 (CXCL8), CC-chemokine ligand 3 (CCL3) and more, and these chemokines attract immune cells to sites of inflammation (13).. MHC class II molecules as well as costimulatory molecules CD40, CD80, and CD86 can be expressed by pDCs, although not as efficiently as cDCs (13). cCDs consist of two main subsets: cDC1 and cDC2, which are present in almost all tissues (11, 14–16). cDC1 and cDC2 normally exert their roles in the priming of CD8 T cells and CD4 T cells, cDC1 primarily presents antigens to CD8 T cells, while cDC2 preferentially initiates various immune responses of CD4 T cells (11, 17). The locations of cDC1 and cDC2 in lymphoid and nonlymphoid organs vary, which affects their interactions with other immune cells and potentially antigens to which they are exposed (15).

Both pDCs and cDCs play important roles in immune regulation. The regulatory function of DCs depends on their activation state, which might impact their ability to induce immunity or tolerance (2, 16). The activity of DCs is closely related to the presence of immunosuppressive factors (18). Upon encountering danger signals, DCs are activated and costimulatory molecules on their surface are upregulated, followed by the production of chemokines and cytokines (1). Inflammatory cytokines and chemokines induce allergic inflammatory responses in the pathogenesis of AR. Allergens are taken up by immature DCs (imDCs) and presented to naïve T cells, which induce DCs maturation, and mature DCs (mDCs) promote adaptive immune responses by inducing effector T cells (18, 19). ImDCs induce immune tolerance by decreasing CD40 expression and increasing IL-10 expression (20). In this procedure, imDCs primarily capture allergens while mDCs mainly act as APCs (21, 22). The imDCs can produce immunogenic, pro-inflammatory mDCs as well as semimature DCs that have the potential to acquire tolerogenic functions (23).

In addition to participating in the immune response, DCs are important modulators of central and peripheral tolerance (24). The inability of newly formed T and B lymphocytes to respond to selfantigens is known as central tolerance (25). DCs maintain central tolerance by regulating the negative selection of self-antigens (26). Increased synthesis of indoleamine-2,3-dioxygenase (IDO)1, upregulated expression of FasL, and induction and maintenance of T cell incompetence by programmed cell death ligand 1 (PDL-1) can promote peripheral tolerance in DCs (26, 27). In contrast to peripheral tolerance, the function of DCs in central tolerance appears to be very limited and may be limited to promoting tolerance to a small subset of self-antigens (28). The induction and maintenance of immune tolerance in DCs is critical for the development of inflammatory diseases. IL-10 and TGF- β have strong anti-inflammatory effects and can induce tolerance of DCs, while IL-1 and IFN-a have obvious pro-inflammatory effects and can promote the activation of DCs (Figure 1) (29). The presence or absence of pro-inflammatory cytokines seems to be decisive in the induction of immunity or tolerance, respectively (4). Both cDCs and pDCs have distinct roles in inducing immune tolerance (30). pDCs maintain immune tolerance by secreting IL-10 and other immunosuppressive mediators, inducing regulatory T cells (Tregs) and inhibiting the secretion of Th2 cells (12, 31). cDCs can induce immune tolerance by initiating T cells and can also lead to peripheral tolerance by inducing T cell incompetence or deletion (30). IDO1 also helps to maintain the tolerance of cDCs (16, 27). The migration pathways and functions of pDCs and cDCs differ due to differences in the expression of chemokines and chemokine receptors (9). The functions of different subtypes of DCs vary and are regulated by environmental factors, and changes in the external environment have an impact on the balance between their tolerance and immunity (32).

3 TolDCs

TolDCs are generally regarded as a type of steady-state semimature DCs, including most imDCs and some cells with advanced maturation states (23, 33, 34). With the ability to re-establish immune tolerance, tolDCs are a specific subset of DCs that overexpress tolerance markers and release tolerance cytokines while underexpressing T cells and costimulatory molecules (35, 36). They can be subdivided into induced tolerogenic DCs (itDCs) and natural tolerogenic DCs (ntDCs) (9, 37). DCs that promote immune responses in response to some inducing signals (agonists, physiological conditions, drugs) to acquire tolerogenic functions are known as itDCs, and ntDCs refer to DCs that inherently promote T cell tolerance (including T cell anergy, T cell depletion, and peripheral Treg cells transformation) in the absence of specific extrinsic signals (37). ItDCs favor the maintenance of homeostasis under pro-inflammatory conditions, while ntDCs contribute to the establishment of tolerance under homeostatic conditions (9). ItDCs and ntDCs are not separate populations, they overlap and may collaborate within organizations (23). Modulation of DC-induced



tolerogenic functions that induce Tregs to suppress inflammation in the presence of inhibitory factors such as IL-10 and TGF-β. Activation of DCs highly expresses MHC class II and costimulatory molecules (CD80/CD86 and CD40), while tolerance of DCs lowly expresses them.

tolerance can influence the immune response, both for itDCs and ntDCs (37).

TolDCs promote tolerance by facilitating the induction and expansion of different subsets of regulatory lymphocytes (38). They can acquire tolerogenic functions by inducing Tregs proliferation, autoreactive T cell incompetence and apoptosis, and interact with naïve T cells to promote tolerance (4, 39, 40). Treg cells or allergenspecific type 1 regulatory T (Tr1) cells also induce tolDCs to regulate immune responses (41). Both tolDCs and imDCs induce Tregs, but the former are more stable and do not produce proinflammatory cytokines (33). With plasticity functions, tolDCs play important roles in maintaining homeostasis and regulating inflammation (39). Inhibitory receptor signaling is one of the essential factors for tolDCs to suppress pro-inflammatory immune responses and induce immune tolerance (42). Immunosuppressive cytokines secreted by them can induce differentiation of Tregs to further mediate tolerogenic immune responses (24, 43). TGF- β , a cytokine with immunosuppressive functions, regulates the function of tolDCs by promoting Tregs expansion and impairing the differentiation, activation and proliferation of CD4 and CD8 T cells (44). IL-10 downregulates DC expression of MHC class II and costimulatory molecules and reverses the effects of pro-inflammatory cytokines, and IL-10induced toIDCs release higher levels of IL-10 (37, 45). IL-10 also upregulates FasL and PDL-1 expression on tolDCs and reduces inflammatory responses by inhibiting NF- κ B (34, 44). TGF- β /IL-10 signaling plays a key role in toIDCs-mediated Tregs amplification (46, 47).

TolDCs modulate immune tolerance by regulating the release of TGF- β , IL-10, IL-35, and granzyme B from regulatory B cells

(Bregs) (44). IL-10 can also be secreted by Tregs and Tr1 cells, which are produced by toIDCs-induced naïve CD4 T cells (33, 48). Along with increasing IDO1 expression, toIDCs induced T cell apoptosis via the Fas/FasL pathway (27, 38). In addition, tolDCs release other cytokines to modulate T cell and Treg activity, and increase the expression and release of immunomodulatory molecules to promote the development of tolerance (38, 45). Perforin-expressing DCs, an important population of tolDCs, can restrict autoreactive T cells (2). Retinoic acid, directly secreted by tolDCs, inhibits effector T cells and induces Tregs and Bregs differentiation (49). $CD103^+$ DCs, which produce TGF β and retinoic acid, can induce the differentiation of naïve T cells into Tregs and Tr1 cells, thus further regulating the function of tolDCs (50). Heme oxygenase-1 (HO-1) regulates tolDCs function by actively inhibiting T cell responses. High expression of HO-1 favors the tolerance capacity of tolDCs, while when HO-1 is blocked, tolDCs lose their immunomodulatory effect (51). Vitamin D, estrogen, IL-27 and many others can also induce tolDCs to regulate immune tolerance (Figure 2) (33). Because they express low levels of costimulatory molecules and high levels of inhibitory receptors, tolDCs are beneficial in reducing inflammation and immune responses (18, 24).

4 TLR4/IRAK4/NF-κB signaling pathway

TLRs are 1 class of transmembrane proteins synthesized in the endoplasmic reticulum (52, 53). TLRs, sensing damage-associated molecular patterns (DAMPs) as well as PAMPs, can initiate



signaling in both innate and adaptive immune pathways (54, 55). Up to now, 10 TLRs have been identified in humans and 12 in mice (56). Hyperactivation of TLR family member TLR4 triggers the production of inflammatory factors, which is associated with a variety of diseases (57). TLR4 triggers inflammation in various microbial infections, cancer, and autoimmune diseases (58). TLR4 induces pro-inflammatory responses to invading pathogens and plays a crucial role in allergic inflammation (59). In the presence of LPS-binding protein (LBP) and CD14, TLR4 binds to LPS with the help of co-receptor myeloid differentiation protein 2 (MD-2) (53, 59). The surroundings have an effect on the bond between them (60). Stimulated activation of TLR4 consists of two major intracellular signaling pathways: the myeloid differentiation primary response 88 (MyD88)-dependent pathway and the tollinterleukin-1 receptor (TIR) structural domain-containing adapterinduced IFN- β (TRIF) pathway (54). These two signaling pathways lead to the production of two sets of pro-inflammatory cytokines. Interaction of MyD88 and TIR homology domain-containing adaptor protein (TIRAP) further leads to activation of IRAK4, accompanied by activation of IRAK1 and IRAK2 (59, 61-63). IRAK4 is the most upstream kinase in this pathway and is directly related to MyD88 (61). IRAK4 plays a decisive role in the TIR signaling pathway (64). Activated IRAK4 is recruited to TNFreceptor associated factor 6 (TRAF6), which then further activates IKB kinase (IKK) signaling via transforming growth factor-activated kinase 1 (TAK1), ultimately leading to NF-KB activation and expression of other pro-inflammatory cytokines (59). These inflammatory factors will drive the inflammatory response, leading to hyperactivation of the immune system. While the TRIF pathway is mediated by TRIF and TRIF-related adaptor molecule (TRAM) to activate type 1 IFN genes and delayed NF- κ B via IFN regulatory factor 3 (IRF-3) (54). These pathways will have an impact on the balance of inflammatory cytokines (Figure 3). Notably, TLR4 is the only TLR that relies on both MyD88 and TRIF pathways, and the TLR4/MyD88 signaling pathway is associated with AR (65, 66).

5 TolDCs and TLR4/IRAK4/NF-κB signaling pathway

DCs are one of the most important immune cells that express TLR4 (67). There are a large number of TLR4 receptors on the surface of DCs (68). Activation of the TLR4 signaling pathway promotes the maturation of DCs (67). The expression of costimulatory molecules CD80/CD86 on DCs is also affected by TLR4 (69). Meanwhile, TLR4 stimulates B cells to produce IL-10, which can further induce toIDCs (70). TLR4 expressed on DCs regulates immune tolerance by releasing IDO1, which further regulates adaptive immune responses by promoting immune suppression and tolerance (27, 67, 71). TLR4 ligation in LPSprimed DCs induced higher levels of IDO1 and aryl hydrocarbon receptor (AhR), further inducing the production of tolDCs (71). Cvetkovic et al. (72) showed that excretory-secretion products (ES L1) released by trichinella spiralis larvae induce the production of toIDCs via TLR4. Kim et al. (73) demonstrated that mycobacterium avium subspecies hominissuis (MAH) infection promotes the generation of tolDCs under the influence of TLR4 signaling. Han et al. (74) showed that minocycline may induce toIDCs by blocking the suppressor of cytokine signaling 1 (SOCS1)/TLR4/NF-κB



signaling pathway. The above studies suggest that TLR4 may be a target for the action of tolDCs.

LPS primarily signals through TLR4 and thus may be involved in MyD88, IRAK4 and NF-KB signaling. As a cell wall component of all Gram-negative bacteria, LPS largely contributes to the induction of tolDCs by upregulating anti-inflammatory cytokines such as IL-10 and TGF- β (53, 75, 76). IL-10, which induces the generation of tolDCs, can inhibit IRAK4, TRAF6 and IRAK1 on the TLR4/IRAK4/NF-KB signaling pathway, thereby inhibiting MyD88-dependent TLR4 signaling (75). IL-10 also inhibits IKK, NF- κ B P65/P50 activity (77). With the function of inducing toIDCs, IL-37 can combine with IL-18R α to reduce the expression of MyD88, IRAK4 and TRAF6, which further leads to the reduction of NF-KB expression (78). MiRNA-155, miRNA-146, let-7 can regulate toIDCs via different signaling molecules, among which miR-146 can directly target IRAK1, TRAF6 to negatively regulate the TLR/MyD88/NF-KB pathway, and mi-155 can directly target TRAF6 (79, 80). Apoptotic cells induced the production of tolDCs to inhibit the transduction of the TLR4/NF-KB signaling pathway (81). Atorvastatin-induced toIDCs reduce inflammatory cell infiltration and inhibit oxidative stress via the TLR4/NF-ĸB signaling pathway (82).

NF-κB can be activated through canonical and non-canonical signaling pathways (83). Activation of the non-canonical NF-κB signaling pathway may be more effective in stimulating peripheral tolerance than the canonical NF-κB signaling pathway, which

primarily responds to pro-inflammatory signals, and the noncanonical NF- κ B pathway in tolDCs may treat inflammatory diseases (84). Inhibition of core transcription factor pathways such as NF- κ B produces tolDCs, which further interfere with NF- κ B signaling by increasing IL-10 (76, 85). The tolDCs phenotype is promoted by NF- κ B p50, which negatively affects the survival of DCs and their ability to effectively activate T cells (75). The expression of tolerance-promoting molecules such as IDO1 can be enhanced and the expression of pro-inflammatory cytokines such as IFN β , IL-1 β , and IL-18 can be reduced during the accumulation of p50 in the nuclei of tolDCs (27, 75). The above studies demonstrated that tolDCs could inhibit different targets on the TLR4/IRAK4/NF- κ B signaling pathway according to different induction signals. This will facilitate the development of different targeted drugs.

6 TolDCs and AR

DCs trigger allergic inflammation or contribute to immune tolerance to sensitizing allergens at different maturation stages, locations, and environments (86). The nature and level of inhaled allergens, route of administration, and changes in the local microenvironment all have an impact on the function of different subsets of DCs (5). Factors such as the type of antigen, the presence of danger signals in the microenvironment, and the genetic background of the host determine whether DCs produce strong Th2-driven allergic responses or acquire tolerance (5). TolDCs achieve suppressive function by suppressing T cell inflammation or activating Tregs (26). Induction of tolDCs by delivery of antibodies bearing the antigen has been shown to be highly efficient in ameliorating the disease process in a range of mouse models (4). Co-delivery of tolerogenic drugs and antigens into nanoparticles has been reported to promote the production of tolDCs (87).

TolDCs are beneficial in suppressing allergic inflammation and relieving allergic symptoms in AR, and may be a potential therapeutic target for AR. Suppression of allergic immune responses by tolDCs inducing allergen-specific blocking antibodies, immunosuppressive cytokines, Tregs, and Bregs is an alternative way to treat AR (88). Induction of tolDCs producing IDO1 promotes immune tolerance in allergic inflammation (27, 29). Upregulation of toIDCs and Tregs favors allergen-specific IgG production and induces immune tolerance, which may inhibit IgE activity and basophil activation (86). The protective allergenspecific IgG4 produced by Tregs competes with IgE for allergen binding to prevent IgE-mediated allergic reactions (52). TolDCs with high expression of IL-10 inhibit Th1 differentiation and limit effector T cell function, thereby suppressing allergic inflammation and promoting allergen-specific tolerance (41). Cui et al. (89) showed that activated AhR could induce the production of tolDCs to differentiate naïve T cells into Treg cells and inhibit Th17 cell differentiation, regulating the balance between Treg and Th17 cells. Activated AhR also induces toIDCs to generate Tr1 cells to regulate the balance of Th1 and Th17 (2). Min et al. (90) used LPS to activate bone marrow-derived DCs to induce the production of tolDCs in a mouse model in vivo, and they found that induced such tolDCs increased the number of Tregs in the lungs of ovalbumininduced asthmatic mice, which could help to attenuate the Th2mediated allergic immune response and treat allergic asthma. It is worth noting that the dosages of LPS need to be strictly controlled when inducing toIDCs, because the maturation of DCs by LPS is highly dose-dependent and also depends on the type of LPS used. Higher dosages induce an inflammatory DC phenotype rather than a tolerogenic one. Hong et al. (91) modulated immune tolerance by inducing toIDCs, which promotes the differentiation of Tregs thereby alleviating allergic reactions to food in mice. Sun et al. (46) demonstrated that Tregs are involved in the anti-inflammatory activity of tolDCs and that the adaptive transfer of tolDCs suppresses allergic airway inflammation. Liu et al. (92) showed that the protein disulfide isomerase (PDI) produced by house dust mites (HDMs) induces tolDCs to produce Tregs to promote immune tolerance, which helps to alleviate airway allergic inflammation. Liu et al. (93) showed that tolDCs and Tregs were suppressed in the AR nasal mucosa compared with those in the non-AR nasal mucosa, which in turn suggested that large amounts of tolDCs are beneficial in the treatment of AR. Sublingual immunotherapy (SLIT) of AR can also induce the production of toIDCs to further alleviate allergic inflammation (94). All these findings fully demonstrate the definitive efficacy of tolDCs in the treatment of allergic diseases. This will promote the application of tolDCs in the treatment of AR.

7 TLR4/IRAK4/NF-κB signaling pathway and AR

With abundant leucine repeats, TLRs regulate Th1/Th2 immune balance through DCs, mast cells and Tregs (52, 60). The adaptive immune response can be skewed toward Th1 by TLRs, which can cause DCs maturation and T cell activation (60). A variety of inflammatory cytokines and chemokines are released when TLRs activate DCs (95). Activation of DCs by allergens results in Th1/Th2 imbalance, which contributes to the development of AR. Genetic factors, environmental factors, and allergens themselves all influence the role of the TLRs in AR (52). TLRs agonists with well-defined immunomodulatory properties, favoring anti-allergic T lymphocyte responses and also increasing IL-10 production to prevent Th1 and Th17 responses (37, 96). TLR4 can initiate, exacerbate, or prevent allergic diseases (66). Generally, TLR4 is expressed at low levels and is upregulated once activated by allergens or other factors (52). TLR4 protein expression levels are elevated in the nasal mucosa of individuals with AR (96). Their elevated expression contributes to TLR4/MyD88 signaling to enhance inflammatory cell generation (66). MyD88 is a protein that articulates with TLR4 in the cytoplasm, which further mediates downstream signaling pathways.

IRAK4, a serine/threonine kinase, is an intermediate in the TLR4/ NF-KB signaling pathway that transduces signals from TLR4 by bridging MyD88 (62, 97). IRAK4 is the only kinase in the IRAK family whose activity has been shown to be required to initiate signaling, and IRAK4 inhibitors will block all MyD88-dependent signaling (63). IRAK4 kinase has an important role in defense against infection in vivo, and its activity is a prerequisite for the establishment of an innate immune response, the loss of which will lead to exacerbation of the infection (98). Korppi et al. (99) showed that IRAK4 may play a role in allergic diseases, where IRAK4 rs4251513, rs4251559, and rs1461567 single nucleotide polymorphisms (SNPs) were associated with serum immunoglobulin E (IgE) levels. Staschke et al. (100) showed that IRAK4 kinase can regulate Th17 differentiation, thereby favoring the treatment of Th17-mediated inflammatory diseases, and their findings suggest that IRAK4 is a promising target for the treatment of Th17 cell-mediated inflammatory diseases. While Th17 cells are strongly associated with AR (101). Because of the important role of IRAK4 in triggering allergic inflammation, IRAK4 inhibitors could be targets for anti-inflammatory drugs (102, 103). Deletion or inactivation of IRAK4 attenuates the development of inflammation (104). Activation of IRAK4 transmits inflammatory signals to NF-KB in the nucleus via TRAF6 and IKKs.

NF-κB was originally thought to be a transcription factor that regulates immunoglobulin gene expression, and its optimal function may be to regulate the development and activation of the immune system (105). Further research has revealed that NF-κB is a key regulator of innate and adaptive immune responses (106). NF-κB is involved in signaling regulation of multiple pathways. Activation of NF-κB induces the secretion of many proinflammatory mediators leading to an inflammatory response as well as the activation of immune cells (107). DCs can recognize allergic inflammation and propagate pro-inflammatory signals through NF-κB, which can also influence the occurrence and development of allergic inflammation by regulating the differentiation and maturation of T cells (106). Increased NF- κ B activity induces IgE synthesis, and decreased NF- κ B activity suppresses allergic inflammatory responses (108, 109).

Multiple drugs can treat AR via TLR4/IRAK4/NF- κ B signaling pathway. Wu et al. (110) showed that probiotics can ameliorate allergic inflammation through the TLR4/NF- κ B signaling pathway. Dong et al. (111) showed that Luteolin could treat AR by improving Th1/Th2 imbalance and reducing inflammation via the TLR4/NF- κ B signaling pathway. Li et al. (112) showed that apigenin could attenuate the inflammatory response in AR through the TLR4/ MyD88/NF- κ B signaling pathway. Liu et al. (113) showed that microRNA-345-5p could alleviate allergic inflammation in AR mice through the TLR4/NF- κ B signaling pathway. These studies amply demonstrate the inseparable relationship between the TLR4/NF- κ B signaling pathway and AR. We can choose the best target on this signaling pathway to intervene in AR depending on the needs.

8 Conclusions and future outlook

The immune response of DCs in AR towards immunity or tolerance is essential for the treatment of AR. We can induce DCs with tolerance function on demand, and inducing tolDCs to act on specific targets is expected to be a scalable immunotherapy for AR. Knocking down the expression of costimulatory molecules and MHC class II with inhibitors of the TLR4/IRAK4/NF- κ B signaling pathway to induce tolDCs is a feasible approach. NF- κ B inhibitors that induce tolDCs are already in clinical trials (45). We can also combine the immunogenicity and tolerability of DCs as needed for better clinical translation. The purpose of inducing tolDCs is to suppress unwanted immune responses in the long term (24). DC-targeting strategies reduce the risk of extensive immunosuppression (4). Current studies have shown that the application of DCs immunotherapy is safe and well tolerated (114).

Despite the great potential of toIDCs-based immunotherapies, the mechanism of their immunomodulatory activity is unclear (18), and further studies and a full understanding of their function in immunosuppression are needed. Are toIDCs phenotypically and functionally stable in a pro-inflammatory environment? Whether toIDCs have a stable and long-lasting effect in treating AR via the TLR4/IRAK4/NF- κ B signaling pathway? This is something we need to explore further. Although there are many ways for inducing toIDCs, there are still substantial knowledge gaps to be filled in the application of toIDCs to treat AR via the TLR4/IRAK4NF- κ B signaling pathway. Easier, shorter cycles and long-term tolerance are our goals. A better understanding of the pathogenesis of AR will contribute to new ways of treating AR. We hope to use toIDCs to

References

1. Balan S, Saxena M, Bhardwaj N. Dendritic cell subsets and locations. Int Rev Cell Mol Biol (2019) 348:1-68. doi: 10.1016/bs.ircmb.2019.07.004 develop drugs without toxic side effects to treat AR via the TLR4/ IRAK4NF-κB signaling pathway in the future.

In conclusion, the potential of tolDCs applied to the treatment of AR is enormous.

Author contributions

CK: Writing – original draft. XL: Writing – original draft. PL: Writing – review & editing. YL: Writing – review & editing. YN: Writing – review & editing. XZ: Writing – review & editing. HZ: Supervision, Writing – review & editing. JL: Funding acquisition, Supervision, Writing – review & editing. SQ: Supervision, Writing – review & editing.

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Conflict of interest

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^{2.} Takenaka MC, Quintana FJ. Tolerogenic dendritic cells. Semin Immunopathol (2017) 39(2):113-20. doi: 10.1007/s00281-016-0587-8

3. Waisman A, Lukas D, Clausen BE, Yogev N. Dendritic cells as gatekeepers of tolerance. *Semin Immunopathol* (2017) 39(2):153-63. doi: 10.1007/s00281-016-0583-z

4. Castenmiller C, Keumatio-Doungtsop BC, van Ree R, de Jong EC, van Kooyk Y. Tolerogenic immunotherapy: targeting DC surface receptors to induce antigen-specific tolerance. *Front Immunol* (2021) 12:643240. doi: 10.3389/fimmu.2021.643240

5. Morianos I, Semitekolou M. Dendritic cells: critical regulators of allergic asthma. Int J Mol Sci (2020) 21(21):7930. doi: 10.3390/ijms21217930

6. Bousquet J, Anto JM, Bachert C, Baiardini I, Bosnic-Anticevich S, Walter Canonica G, et al. Allergic rhinitis. *Nat Rev Dis Primers*. (2020) 6(1):95. doi: 10.1038/s41572-020-00227-0

7. Luckey U, Maurer M, Schmidt T, Lorenz N, Seebach B, Metz M, et al. T cell killing by tolerogenic dendritic cells protects mice from allergy. *J Clin Invest.* (2011) 121 (10):3860–71. doi: 10.1172/JCI45963

8. Hollingsworth JW, Free ME, Li Z, Andrews LN, Nakano H, Cook DN. Ozone activates pulmonary dendritic cells and promotes allergic sensitization through a Tolllike receptor 4-dependent mechanism. *J Allergy Clin Immunol* (2010) 125(5):1167–70. doi: 10.1016/j.jaci.2010.03.001

9. Feng M, Zhou S, Yu Y, Su Q, Li X, Lin W. Regulation of the migration of distinct dendritic cell subsets. *Front Cell Dev Biol* (2021) 9:635221. doi: 10.3389/ fcell.2021.635221

10. Kuzmich NN, Sivak KV, Chubarev VN, Porozov YB, Savateeva-Lyubimova TN, Peri F. TLR4 signaling pathway modulators as potential therapeutics in inflammation and sepsis. *Vaccines* (2017) 5(4):34. doi: 10.3390/vaccines5040034

11. Eisenbarth SC. Dendritic cell subsets in T cell programming: location dictates function. Nat Rev Immunol (2019) 19(2):89-103. doi: 10.1038/s41577-018-0088-1

12. Reizis B. Plasmacytoid dendritic cells: development, regulation, and function. *Immunity* (2019) 50(1):37–50. doi: 10.1016/j.immuni.2018.12.027

13. Swiecki M, Colonna M. The multifaceted biology of plasmacytoid dendritic cells. Nat Rev Immunol (2015) 15(8):471–85. doi: 10.1038/nri3865

14. Murphy TL, Murphy KM. Dendritic cells in cancer immunology. Cell Mol Immunol (2022) 19(1):3–13. doi: 10.1038/s41423-021-00741-5

15. Hilligan KL, Ronchese F. Antigen presentation by dendritic cells and their instruction of CD4+ T helper cell responses. *Cell Mol Immunol* (2020) 17(6):587–99. doi: 10.1038/s41423-020-0465-0

16. Gargaro M, Scalisi G, Manni G, Briseño CG, Bagadia P, Durai V, et al. Indoleamine 2,3-dioxygenase 1 activation in mature cDC1 promotes tolerogenic education of inflammatory cDC2 *via* metabolic communication. *Immunity* (2022) 55 (6):1032–50.e14. doi: 10.1016/j.immuni.2022.05.013

17. Binnewies M, Mujal AM, Pollack JL, Combes AJ, Hardison EA, Barry KC, et al. Unleashing type-2 dendritic cells to drive protective antitumor CD4(+) T cell immunity. *Cell* (2019) 177(3):556–71.e16. doi: 10.1016/j.cell.2019.02.005

18. Nam JH, Lee JH, Choi SY, Jung NC, Song JY, Seo HG, et al. Functional ambivalence of dendritic cells: tolerogenicity and immunogenicity. *Int J Mol Sci* (2021) 22(9):4430. doi: 10.3390/ijms22094430

19. Sabado RL, Bhardwaj N. Dendritic cell immunotherapy. Ann N Y Acad Sci (2013) 1284:31-45. doi: 10.1111/nyas.12125

20. Yu H, Tian Y, Wang Y, Mineishi S, Zhang Y. Dendritic cell regulation of graftvs.-host disease: immunostimulation and tolerance. *Front Immunol* (2019) 10:93. doi: 10.3389/fimmu.2019.00093

21. Tiberio L, Del Prete A, Schioppa T, Sozio F, Bosisio D, Sozzani S. Chemokine and chemotactic signals in dendritic cell migration. *Cell Mol Immunol* (2018) 15 (4):346–52. doi: 10.1038/s41423-018-0005-3

22. Worbs T, Hammerschmidt SI, Förster R. Dendritic cell migration in health and disease. Nat Rev Immunol (2017) 17(1):30–48. doi: 10.1038/nri.2016.116

23. Maldonado RA, von Andrian UH. How tolerogenic dendritic cells induce regulatory T cells. *Adv Immunol* (2010) 108:111–65. doi: 10.1016/B978-0-12-380995-7.00004-5

24. Passeri L, Marta F, Bassi V, Gregori S. Tolerogenic dendritic cell-based approaches in autoimmunity. Int J Mol Sci (2021) 22(16):8415. doi: 10.3390/ijms22168415

25. Hadeiba H, Butcher EC. Thymus-homing dendritic cells in central tolerance. Eur J Immunol (2013) 43(6):1425–9. doi: 10.1002/eji.201243192

26. Devi KS, Anandasabapathy N. The origin of DCs and capacity for immunologic tolerance in central and peripheral tissues. *Semin Immunopathol* (2017) 39(2):137–52. doi: 10.1007/s00281-016-0602-0

27. Merlo LMF, Peng W, Mandik-Nayak L. Impact of IDO1 and IDO2 on the B cell immune response. *Front Immunol* (2022) 13:886225. doi: 10.3389/fimmu.2022.886225

28. Ganguly D, Haak S, Sisirak V, Reizis B. The role of dendritic cells in autoimmunity. *Nat Rev Immunol* (2013) 13(8):566–77. doi: 10.1038/nri3477

29. Horwitz DA, Fahmy TM, Piccirillo CA, La Cava A. Rebalancing immune homeostasis to treat autoimmune diseases. *Trends Immunol* (2019) 40(10):888–908. doi: 10.1016/j.it.2019.08.003

30. Audiger C, Rahman MJ, Yun TJ, Tarbell KV, Lesage S. The importance of dendritic cells in maintaining immune tolerance. *J Immunol* (2017) 198(6):2223–31. doi: 10.4049/jimmunol.1601629

31. Li S, Wu J, Zhu S, Liu YJ, Chen J. Disease-associated plasmacytoid dendritic cells. *Front Immunol* (2017) 8:1268. doi: 10.3389/fimmu.2017.01268 32. Shortman K, Liu YJ. Mouse and human dendritic cell subtypes. Nat Rev Immunol (2002) 2(3):151–61. doi: 10.1038/nri746

33. Xie ZX, Zhang HL, Wu XJ, Zhu J, Ma DH, Jin T. Role of the immunogenic and tolerogenic subsets of dendritic cells in multiple sclerosis. *Mediators Inflamm* (2015) 2015:513295. doi: 10.1155/2015/513295

34. Sim WJ, Ahl PJ, Connolly JE. Metabolism is central to tolerogenic dendritic cell function. *Mediators Inflamm* (2016) 2016:2636701. doi: 10.1155/2016/2636701

35. Deak P, Knight HR, Esser-Kahn A. Robust tolerogenic dendritic cells *via* push/ pull pairing of toll-like-receptor agonists and immunomodulators reduces EAE. *Biomaterials* (2022) 286:121571. doi: 10.1016/j.biomaterials.2022.121571

36. Sahar A, Nicorescu I, Barran G, Paterson M, Hilkens CMU, Lord P. Tolerogenic dendritic cell reporting: Has a minimum information model made a difference? *PeerJ* (2023) 11:e15352. doi: 10.7717/peerj.15352

37. Iberg CA, Hawiger D. Natural and induced tolerogenic dendritic cells. *J Immunol* (2020) 204(4):733–44. doi: 10.4049/jimmunol.1901121

38. Ochando J, Ordikhani F, Jordan S, Boros P, Thomson AW. Tolerogenic dendritic cells in organ transplantation. *Transpl Int* (2020) 33(2):113-27. doi: 10.1111/tri.13504

39. Funes SC, Manrique de Lara A, Altamirano-Lagos MJ, Mackern-Oberti JP, Escobar-Vera J, Kalergis AM. Immune checkpoints and the regulation of tolerogenicity in dendritic cells: Implications for autoimmunity and immunotherapy. *Autoimmun Rev* (2019) 18(4):359–68. doi: 10.1016/j.autrev.2019.02.006

40. Hu J, Wan Y. Tolerogenic dendritic cells and their potential applications. *Immunology* (2011) 132(3):307-14. doi: 10.1111/j.1365-2567.2010.03396.x

41. Schülke S. Induction of interleukin-10 producing dendritic cells as a tool to suppress allergen-specific T helper 2 responses. *Front Immunol* (2018) 9:455. doi: 10.3389/fimmu.2018.00455

42. Fucikova J, Palova-Jelinkova L, Bartunkova J, Spisek R. Induction of tolerance and immunity by dendritic cells: mechanisms and clinical applications. *Front Immunol* (2019) 10:2393. doi: 10.3389/fimmu.2019.02393

43. Cifuentes-Rius A, Desai A, Yuen D, Johnston APR, Voelcker NH. Inducing immune tolerance with dendritic cell-targeting nanomedicines. *Nat Nanotechnol.* (2021) 16(1):37–46. doi: 10.1038/s41565-020-00810-2

44. Marín E, Cuturi MC, Moreau A. Tolerogenic dendritic cells in solid organ transplantation: where do we stand? *Front Immunol* (2018) 9:274. doi: 10.3389/fimmu.2018.00274

45. Zhuang Q, Cai H, Cao Q, Li Z, Liu S, Ming Y. Tolerogenic dendritic cells: the pearl of immunotherapy in organ transplantation. *Front Immunol* (2020) 11:552988. doi: 10.3389/fimmu.2020.552988

46. Sun W, Wei JW, Li H, Wei FQ, Li J, Wen WP. Adoptive cell therapy of tolerogenic dendritic cells as inducer of regulatory T cells in allergic rhinitis. *Int Forum Allergy Rhinol* (2018) 8(11):1291–9. doi: 10.1002/alr.22217

47. Durham SR, Shamji MH. Allergen immunotherapy: past, present and future. *Nat Rev Immunol* (2023) 23(5):317–28. doi: 10.1038/s41577-022-00786-1

48. Raffin C, Vo LT, Bluestone JA. T(reg) cell-based therapies: challenges and perspectives. Nat Rev Immunol (2020) 20(3):158-72. doi: 10.1038/s41577-019-0232-6

49. Morante-Palacios O, Fondelli F, Ballestar E, Martínez-Cáceres EM. Tolerogenic dendritic cells in autoimmunity and inflammatory diseases. *Trends Immunol* (2021) 42 (1):59–75. doi: 10.1016/j.it.2020.11.001

50. Yu W, Freeland DMH, Nadeau KC. Food allergy: immune mechanisms, diagnosis and immunotherapy. *Nat Rev Immunol* (2016) 16(12):751-65. doi: 10.1038/nri.2016.111

51. Zhao Y, Jia Y, Wang L, Chen S, Huang X, Xu B, et al. Upregulation of heme oxygenase-1 endues immature dendritic cells with more potent and durable immunoregulatory properties and promotes engraftment in a stringent mouse cardiac allotransplant model. *Front Immunol* (2018) 9:1515. doi: 10.3389/fmmu.2018.01515

52. Wenger M, Grosse-Kathoefer S, Kraiem A, Pelamatti E, Nunes N, Pointner L, et al. When the allergy alarm bells toll: The role of Toll-like receptors in allergic diseases and treatment. *Front Mol Biosci* (2023) 10:1204025. doi: 10.3389/fmolb.2023.1204025

53. Fitzgerald KA, Kagan JC. Toll-like receptors and the control of immunity. *Cell* (2020) 180(6):1044–66. doi: 10.1016/j.cell.2020.02.041

54. Bruno K, Woller SA, Miller YI, Yaksh TL, Wallace M, Beaton G, et al. Targeting toll-like receptor-4 (TLR4)-an emerging therapeutic target for persistent pain states. *Pain* (2018) 159(10):1908–15. doi: 10.1097/j.pain.00000000001306

55. Arora S, Ahmad S, Irshad R, Goyal Y, Rafat S, Siddiqui N, et al. TLRs in pulmonary diseases. Life Sci (2019) 233:116671. doi: 10.1016/j.lfs.2019.116671

56. Duan T, Du Y, Xing C, Wang HY, Wang RF. Toll-like receptor signaling and its role in cell-mediated immunity. *Front Immunol* (2022) 13:812774. doi: 10.3389/fimmu.2022.812774

57. Zhang Y, Liang X, Bao X, Xiao W, Chen G. Toll-like receptor 4 (TLR4) inhibitors: Current research and prospective. *Eur J Med Chem* (2022) 235:114291. doi: 10.1016/j.ejmech.2022.114291

58. Mahish C, De S, Chatterjee S, Ghosh S, Keshry SS, Mukherjee T, et al. TLR4 is one of the receptors for Chikungunya virus envelope protein E2 and regulates virus induced pro-inflammatory responses in host macrophages. *Front Immunol* (2023) 14:1139808. doi: 10.3389/fimmu.2023.1139808

59. Ciesielska A, Matyjek M, Kwiatkowska K. TLR4 and CD14 trafficking and its influence on LPS-induced pro-inflammatory signaling. *Cell Mol Life Sci* (2021) 78 (4):1233–61. doi: 10.1007/s00018-020-03656-y

60. Chen RX, Dai MD, Zhang QZ, Lu MP, Wang ML, Yin M, et al. TLR signaling pathway gene polymorphisms, gene-gene and gene-environment interactions in allergic rhinitis. *J Inflammation Res* (2022) 15:3613–30. doi: 10.2147/JIR.S364877

61. Wang L, Ferrao R, Li Q, Hatcher JM, Choi HG, Buhrlage SJ, et al. Conformational flexibility and inhibitor binding to unphosphorylated interleukin-1 receptor-associated kinase 4 (IRAK4). *J Biol Chem* (2019) 294(12):4511-9. doi: 10.1074/jbc.RA118.005428

62. Pereira M, Durso DF, Bryant CE, Kurt-Jones EA, Silverman N, Golenbock DT, et al. The IRAK4 scaffold integrates TLR4-driven TRIF and MYD88 signaling pathways. *Cell Rep* (2022) 40(7):111225. doi: 10.1016/j.celrep.2022.111225

63. Sabnis RW. Novel tricyclic heteroaryl compounds as IRAK4 inhibitors for treating cancer, autoimmune and inflammatory diseases. *ACS Med Chem Lett* (2022) 13(3):336–7. doi: 10.1021/acsmedchemlett.2c00060

64. Seumen CHT, Grimm TM, Hauck CR. Protein phosphatases in TLR signaling. *Cell Commun Signal* (2021) 19(1):45. doi: 10.1186/s12964-021-00722-1

65. Squillace S, Salvemini D. Toll-like receptor-mediated neuroinflammation: relevance for cognitive dysfunctions. *Trends Pharmacol Sci* (2022) 43(9):726–39. doi: 10.1016/j.tips.2022.05.004

66. Shalaby KH, Al Heialy S, Tsuchiya K, Farahnak S, McGovern TK, Risse PA, et al. The TLR4-TRIF pathway can protect against the development of experimental allergic asthma. *Immunology* (2017) 152(1):138–49. doi: 10.1111/imm.12755

67. Bruning EE, Coller JK, Wardill HR, Bowen JM. Site-specific contribution of Tolllike receptor 4 to intestinal homeostasis and inflammatory disease. *J Cell Physiol* (2021) 236(2):877–88. doi: 10.1002/jcp.29976

68. Ye Z, Zhong L, Zhu S, Wang Y, Zheng J, Wang S, et al. The P-selectin and PSGL-1 axis accelerates atherosclerosis *via* activation of dendritic cells by the TLR4 signaling pathway. *Cell Death Dis* (2019) 10(7):507. doi: 10.1038/s41419-019-1736-5

69. Pufnock JS, Cigal M, Rolczynski LS, Andersen-Nissen E, Wolfl M, McElrath MJ, et al. Priming CD8+ T cells with dendritic cells matured using TLR4 and TLR7/8 ligands together enhances generation of CD8+ T cells retaining CD28. *Blood* (2011) 117 (24):6542–51. doi: 10.1182/blood-2010-11-317966

70. Boldison J, Da Rosa LC, Davies J, Wen L, Wong FS. Dendritic cells license regulatory B cells to produce IL-10 and mediate suppression of antigen-specific CD8 T cells. *Cell Mol Immunol* (2020) 17(8):843–55. doi: 10.1038/s41423-019-0324-z

71. Salazar F, Awuah D, Negm OH, Shakib F, Ghaemmaghami AM. The role of indoleamine 2,3-dioxygenase-aryl hydrocarbon receptor pathway in the TLR4-induced tolerogenic phenotype in human DCs. *Sci Rep* (2017) 7:43337. doi: 10.1038/srep43337

72. Cvetkovic J, Ilic N, Gruden-Movsesijan A, Tomic S, Mitic N, Pinelli E, et al. DC-SIGN signalling induced by Trichinella spiralis products contributes to the tolerogenic signatures of human dendritic cells. *Sci Rep* (2020) 10(1):20283. doi: 10.1038/s41598-020-77497-x

73. Kim WS, Yoon JH, Shin MK, Shin SJ. Infection of Dendritic Cells With Mycobacterium avium subspecies hominissuis Exhibits a Functionally Tolerogenic Phenotype in Response to Toll-Like Receptor Agonists *via* IL-10/Cox2/PGE2/EP2 Axis. *Front Microbiol* (2019) 10:1795. doi: 10.3389/fmicb.2019.01795

74. Han X, Wei Q, Xu RX, Wang S, Liu XY, Guo C, et al. Minocycline induces tolerance to dendritic cell production probably by targeting the SOCS1/ TLR4/NF- κ B signaling pathway. *Transpl Immunol* (2023) 79:101856. doi: 10.1016/j.trim.2023.101856

75. Steimle A, Frick JS. Molecular mechanisms of induction of tolerant and tolerogenic intestinal dendritic cells in mice. *J Immunol Res* (2016) 2016:1958650. doi: 10.1155/2016/1958650

76. Klaska IP, Muckersie E, Martin-Granados C, Christofi M, Forrester JV. Lipopolysaccharide-primed heterotolerant dendritic cells suppress experimental autoimmune uveoretinitis by multiple mechanisms. *Immunology* (2017) 150(3):364–77. doi: 10.1111/imm.12691

77. Hubo M, Trinschek B, Kryczanowsky F, Tuettenberg A, Steinbrink K, Jonuleit H. Costimulatory molecules on immunogenic versus tolerogenic human dendritic cells. *Front Immunol* (2013) 4:82. doi: 10.3389/fimmu.2013.00082

78. Dinarello CA, Nold-Petry C, Nold M, Fujita M, Li S, Kim S, et al. Suppression of innate inflammation and immunity by interleukin-37. *Eur J Immunol* (2016) 46 (5):1067–81. doi: 10.1002/eji.201545828

 Zheng J, Jiang HY, Li J, Tang HC, Zhang XM, Wang XR, et al. MicroRNA-23b promotes tolerogenic properties of dendritic cells in *vitro* through inhibiting Notch1/ NF-κB signalling pathways. *Allergy* (2012) 67(3):362–70. doi: 10.1111/j.1398-9995.2011.02776.x

80. Kim SJ, Diamond B. Modulation of tolerogenic dendritic cells and autoimmunity. *Semin Cell Dev Biol* (2015) 41:49–58. doi: 10.1016/j.semcdb.2014.04.020

81. Trahtemberg U, Mevorach D. Apoptotic cells induced signaling for immune homeostasis in macrophages and dendritic cells. *Front Immunol* (2017) 8:1356. doi: 10.3389/fimmu.2017.01356

82. Wang Q, Chen Z, Guo J, Peng X, Zheng Z, Chen H, et al. Atorvastatin-induced tolerogenic dendritic cells improve cardiac remodeling by suppressing TLR-4/NF- κ B activation after myocardial infarction. *Inflammation Res* (2023) 72(1):13–25. doi: 10.1007/s00011-022-01654-3

83. Sun SC. The non-canonical NF-κB pathway in immunity and inflammation. Nat Rev Immunol (2017) 17(9):545-58. doi: 10.1038/nri.2017.52

84. van Delft MA, Huitema LF, Tas SW. The contribution of NF-κB signalling to immune regulation and tolerance. *Eur J Clin Invest.* (2015) 45(5):529–39. doi: 10.1111/eci.12430

85. Jha A, Ahad A, Mishra GP, Sen K, Smita S, Minz AP, et al. SMRT and NCoR1 fine-tune inflammatory versus tolerogenic balance in dendritic cells by differentially regulating STAT3 signaling. *Front Immunol* (2022) 13:910705. doi: 10.3389/fimmu.2022.910705

86. Wang W, Yin J. Is it worthy to take full-course immunotherapy for allergic rhinitis? About efficacy biomarker of allergen immunotherapy. *Scand J Immunol* (2020) 91(1):e12817. doi: 10.1111/sji.12817

87. Stead SO, Kireta S, McInnes SJP, Kette FD, Sivanathan KN, Kim J, et al. Murine and non-human primate dendritic cell targeting nanoparticles for in vivo generation of regulatory T-cells. *ACS nano.* (2018) 12(7):6637–47. doi: 10.1021/acsnano.8b01625

88. Drazdauskaitė G, Layhadi JA, Shamji MH. Mechanisms of allergen immunotherapy in allergic rhinitis. *Curr Allergy Asthma Rep* (2020) 21(1):2. doi: 10.1007/s11882-020-00977-7

89. Cui X, Ye Z, Wang D, Yang Y, Jiao C, Ma J, et al. Aryl hydrocarbon receptor activation ameliorates experimental colitis by modulating the tolerogenic dendritic and regulatory T cell formation. *Cell Biosci* (2022) 12(1):46. doi: 10.1186/s13578-022-00780-z

90. Min Z, Zeng Y, Zhu T, Cui B, Mao R, Jin M, et al. Lipopolysaccharide-activated bone marrow-derived dendritic cells suppress allergic airway inflammation by ameliorating the immune microenvironment. *Front Immunol* (2021) 12:595369. doi: 10.3389/fimmu.2021.595369

91. Hong J, Xiao X, Gao Q, Li S, Jiang B, Sun X, et al. Co-delivery of allergen epitope fragments and R848 inhibits food allergy by inducing tolerogenic dendritic cells and regulatory T cells. *Int J Nanomedicine*. (2019) 14:7053–64. doi: 10.2147/IJN.S215415

92. Liu X, Wang Y, Chen D, Ji S, Yang LT, Huang Q, et al. Dust-mite-derived protein disulfide isomerase suppresses airway allergy by inducing tolerogenic dendritic cells. *J Biol Chem* (2021) 296:100585. doi: 10.1016/j.jbc.2021.100585

93. Liu T, Liang X, Li TL, Ma J, Yang JF, Yang PC. Staphylococcal enterotoxin B compromises the immune tolerant status in the airway mucosa. *Clin Exp Allergy* (2012) 42(3):375–82. doi: 10.1111/j.1365-2222.2011.03869.x

94. Moingeon P. Update on immune mechanisms associated with sublingual immunotherapy: practical implications for the clinician. J Allergy Clin Immunol Pract (2013) 1(3):228-41. doi: 10.1016/j.jaip.2013.03.013

95. Napolitani G, Rinaldi A, Bertoni F, Sallusto F, Lanzavecchia A. Selected Toll-like receptor agonist combinations synergistically trigger a T helper type 1-polarizing program in dendritic cells. *Nat Immunol* (2005) 6(8):769–76. doi: 10.1038/ni1223

96. Kirtland ME, Tsitoura DC, Durham SR, Shamji MH. Toll-like receptor agonists as adjuvants for allergen immunotherapy. *Front Immunol* (2020) 11:599083. doi: 10.3389/fimmu.2020.599083

97. Li Q, Li R, Yin H, Wang S, Liu B, Li J, et al. Oral IRAK4 inhibitor BAY-1834845 prevents acute respiratory distress syndrome. *BioMed Pharmacother*. (2022) 153:113459. doi: 10.1016/j.biopha.2022.113459

98. Pattabiraman G, Murphy M, Agliano F, Karlinsey K, Medvedev AE. IRAK4 activity controls immune responses to intracellular bacteria Listeria monocytogenes and Mycobacterium smegmatis. *J Leukoc Biol* (2018) 104(4):811–20. doi: 10.1002/JLB.2A1117-449R

99. Korppi M, Teräsjärvi J, Lauhkonen E, Törmänen S, He Q, Nuolivirta K. Interleukin-1 receptor-associated kinase-4 gene variation may increase postbronchiolitis asthma risk. *Acta Paediatr* (2021) 110(3):952–8. doi: 10.1111/apa.15607

100. Staschke KA, Dong S, Saha J, Zhao J, Brooks NA, Hepburn DL, et al. IRAK4 kinase activity is required for Th17 differentiation and Th17-mediated disease. J Immunol (2009) 183(1):568–77. doi: 10.4049/jimmunol.0802361

101. Ueda S, Miura K, Kawasaki H, Ogata S, Yamasaki N, Miura S, et al. Th17dependent nasal hyperresponsiveness is mitigated by steroid treatment. *Biomolecules* (2022) 12(5):674. doi: 10.3390/biom12050674

102. Sabnis RW. Novel IRAK4 inhibitors for treating asthma. ACS Med Chem Lett (2022) 13(8):1219–20. doi: 10.1021/acsmedchemlett.2c00324

103. Otto G. IRAK4 inhibitor attenuates inflammation. Nat Rev Rheumatol (2021) 17(11):646. doi: 10.1038/s41584-021-00699-8

104. Winkler A, Sun W, De S, Jiao A, Sharif MN, Symanowicz PT, et al. The interleukin-1 receptor-associated kinase 4 inhibitor PF-06650833 blocks inflammation in preclinical models of rheumatic disease and in humans enrolled in a randomized clinical trial. *Arthritis Rheumatol* (2021) 73(12):2206–18. doi: 10.1002/art.41953

105. Sun SC, Liu ZG. A special issue on NF- κB signaling and function. Cell Res (2011) 21(1):1–2. doi: 10.1038/cr.2011.1

106. Hayden MS, Ghosh S. NF-κB in immunobiology. Cell Res (2011) 21(2):223–44. doi: 10.1038/cr.2011.13

107. Yu H, Lin L, Zhang Z, Zhang H, Hu H. Targeting NF-κB pathway for the therapy of diseases: mechanism and clinical study. *Signal Transduct Target Ther* (2020) 5(1):209. doi: 10.1038/s41392-020-00312-6

108. Yanagihara Y, Basaki Y, Ikizawa K, Kajiwara K, Koshio T, Akiyama K. Involvement of nuclear factor-kappa B activation in IgE synthesis in human B cells. J Allergy Clin Immunol (1996) 98(6 Pt 2):S224–9. doi: 10.1016/S0091-6749(96)70070-2

109. Piao CH, Fan YJ, Nguyen TV, Song CH, Chai OH. Mangiferin alleviates ovalbumin-induced allergic rhinitis via nrf2/HO-1/NF-κB signaling pathways. Int J Mol Sci (2020) 21(10):3415. doi: 10.3390/ijms21103415

110. Wu Z, Mehrabi Nasab E, Arora P, Athari SS. Study effect of probiotics and prebiotics on treatment of OVA-LPS-induced of allergic asthma inflammation and pneumonia by regulating the TLR4/NF-kB signaling pathway. *J Transl Med* (2022) 20 (1):130. doi: 10.1186/s12967-022-03337-3

111. Dong J, Xu O, Wang J, Shan C, Ren X. Luteolin ameliorates inflammation and Th1/ Th2 imbalance via regulating the TLR4/NF-κB pathway in allergic rhinitis rats. *Immunopharmacol Immunotoxicol.* (2021) 43(3):319–27. doi: 10.1080/08923973.2021.1905659 112. Li H, Zhao H, Zhao H. Apigenin attenuates inflammatory response in allergic rhinitis mice by inhibiting the TLR4/MyD88/NF- κ B signaling pathway. *Environ Toxicol* (2023) 38(2):253–65. doi: 10.1002/tox.23699

113. Liu J, Jiang Y, Han M, Jiang L, Liang D, Li S, et al. MicroRNA-345-5p acts as an anti-inflammatory regulator in experimental allergic rhinitis via the TLR4/NF- κ B pathway. Int Immunopharmacol. (2020) 86:106522. doi: 10.1016/j.intimp.2020.106522

114. Ness S, Lin S, Gordon JR. Regulatory dendritic cells, T cell tolerance, and dendritic cell therapy for immunologic disease. *Front Immunol* (2021) 12:633436. doi: 10.3389/fimmu.2021.633436