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The role of tumor immune microenvironment in chordoma: promising immunotherapy strategies

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Chordoma is a rare malignant bone tumor with limited therapeutic options, which is resistant to conventional chemotherapy and radiotherapy, and targeted therapy is also shown with little efficacy. The long-standing delay in researching its mechanisms of occurrence and development has resulted in the dilemma of no effective treatment targets and no available drugs in clinical practice. In recent years, the role of the tumor immune microenvironment in driving tumor growth has become a hot and challenging topic in the field of cancer research. Immunotherapy has shown promising results in the treatment of various tumors. However, the study of the immune microenvironment of chordoma is still in its infancy. In this review, we aim to present a comprehensive reveal of previous exploration on the chordoma immune microenvironment and propose promising immunotherapy strategies for chordoma based on these characteristics.

KEYWORDS

chordoma, tumor immune environment, immune-excluded, immune checkpoint inhibitor (ICI), immunotherapy

1 Introduction

Chordoma is a rare, slow-growing, low-to-moderate malignant bone tumor that originates from residual embryonic notochord. The common sites of chordoma are axial bones, with the sacrum accounting for over 50% of primary sacral tumors (1). In adults, 50% of chordoma involve the sacrococcygeal region, 35% occur in the skull base, and 15% are found in other parts of the spine. The incidence of chordoma is low, with only 0.08 cases per 10,000 people (2), presenting a male predominance, commonly found in 50- to 60-year-old male patients; however, chordomas located in the skull base tend to occur at younger ages, even in adolescents and children. The median survival time for patients with chordoma is 6.29 years, with overall survival rates of 67.6% at 5 years and 39.9% at 10 years (3). Although chordoma are slow-growing, their robust local invasiveness leads to a high rate of local recurrence after surgery. The local recurrence rate has become the strongest

predictor of mortality for patients with chordoma (4). The advanced chordoma often metastasize to multiple organs in the body, such as the lungs, bones, soft tissues, lymph nodes, and liver. Once distant metastases occur, the median survival time drops to less than 12 months (5).

Chordoma is resistant to chemotherapy (6, 7) and exhibits some degree of resistance to radiation therapy, which may be because of the low proliferation pool of cells (8), with no effective treatment for chordoma. Additionally, high doses of radiation are limited due to the low tolerance of adjacent tissues such as the spinal cord, rectum, bladder, and brainstem to the therapeutic dose for chordoma.

A wealth of evidence has emerged revealing how the functionality of the tumor immune microenvironment (TIME) determines its integral and indispensable role in tumor initiation and progress (9). Immunotherapy by targeting TIME and potentiating host anti-tumor immune responses has gradually become a hot topic in the field of cancer treatment and has been clinically applied in various types of tumors (10, 11). Immunotherapy is expected to improve the prognosis of patients with chordoma. However, there has been no review comprehensively sketching the TIME of chordoma by far; in this review, we aim to investigate the immune microenvironment studies of chordoma and propose rational and effective immunotherapeutic strategies for the patients with chordoma.

2 The basic research on chordoma

2.1 The role of brachyury in chordoma

Brachyury, a T-box family transcription factor, plays a critical role in the development of the notochord during embryogenesis and is only minimally expressed in adult tissues, such as the pituitary gland, thyroid, and testes (12). Brachyury not only is used as a diagnostic marker for chordoma, but also serves as a key driver in the development and progression of chordoma. Sharifnia et al. found that brachyury is highly enriched in multiple chordoma cell lines and mediate the overexpression of genes involved in malignant progression, such as extracellular matrix (ECM) regulation and EGFR-related signaling pathways, by binding to super enhancers in chordoma (13). Experimental evidence has demonstrated that knockdown of brachyury expression by shRNA can inhibit the progression of chordoma (14, 15). However, similar to other transcription factors, brachyury is difficult to target with drugs, and currently there are no targeted drugs or small-molecule inhibitors available for brachyury. Clinical trials of a brachyury-targeting vaccine (GI-6301) have also been declared ineffective (16).

2.2 Non-coding RNAs in chordoma progression

In recent years, more and more studies have shown that noncoding RNAs (ncRNAs) play an important role in the occurrence and development of various diseases by regulating gene expression (17, 18). Currently, research on ncRNAs in chordoma mainly focuses on miRNA and lncRNA (19–21). For example, Zhang et al. found that miR-16-5p can inhibit chordoma proliferation, migration, and invasion by regulating downstream Smad3 expression (21). Lou et al. further revealed that circTLK1 can regulate the biological functions of miR-16-5p through sponge adsorption, thus forming a circTLK1/miR-16-5p/Smad3 positive feedback loop to promote chordoma invasion, migration, and epithelial-mesenchymal transition (22). Revealing the competing endogenous RNA (ceRNA) network in chordoma can help to better explore the developmental mechanisms of chordoma. Furthermore, the ceRNA network also plays an important role in chemotherapy resistance, tumor prognostic markers, and other aspects of various tumors (23, 24), which require further investigation in chordoma.

2.3 Research on signaling pathways in chordoma

Current research has shown that multiple mutations or overexpression of receptor tyrosine kinases (RTKs) occur in chordoma tissue and cell lines. Currently, the dysregulated signaling pathways in chordoma mainly focus on platelet-derived growth factor receptor (PDGFR) (25), vascular endothelial growth factor receptor (VEGFR), epidermal growth factor receptor (EGFR) (26), human epidermal growth factor receptor 2 (HER2/neu) (26), phosphatidylinositol 3-kinase (PI3K)/protein kinase B (Akt)/ mammalian target of rapamycin (mTOR) (27), fibroblast growth factor (FGF)/mitogen-activated protein kinase kinase (MEK)/ extracellular signal-regulated kinase (ERK) (28), hepatocyte growth factor receptor (HGFR, c-Met) (29), and insulin-like growth factor 1 receptor (IGF-1R)-related signaling pathways (30). Among these pathways, multiple pathways can mediate the expression of brachyury, which may be a key pathway driving the occurrence and development of chordoma (Figure 1). In addition to these signaling pathways, scattered studies have also revealed other signaling pathways involved in chordoma. For example, Shihabi et al. (31) constructed a patient-derived organoid model of chordoma and conducted high-throughput drug screening, and found that the NF-KB pathway and MUC1 were dysregulated in chordoma. Corresponding molecular targeted therapy clinical trials have been conducted for the relevant RTKs and signaling pathways in advanced and recurrent cases of chordoma.

3 Tumor immune microenvironment

3.1 Immune cells interacted with chordoma cells

Immune cells are the cellular basis for immunotherapy, and understanding the composition and functional status of immune cells in the tumor microenvironment (TME) is crucial. Various immune cells in the body can exert anti-tumor effects, among which cytotoxic T cells (CTLs, CD8⁺ T cells) are considered to be the main effector cells. CD8⁺ T cells can be activated and recruited into the



The basic research on chordoma. Chordoma studies mainly focus on the signaling pathway, brachyury, and non-coding RNAs. Various receptors and signaling pathways are dysregulated in chordoma; using imatinib, dastinib, and erotinib can inhibit the corresponding signaling pathways for targeted therapy. Brachyury plays a driving role in chordoma, causing a variety of malignant genes upregulated by binding to super enhancers. Non-coding RNAs are also dysregulated and lead to tumor progression.

TME to exert anti-tumor effects by releasing granules and performs (32). In addition to CTLs, NK cells, macrophages, DCs, and other immune cells can also exert certain anti-tumor effects. However, there exist abnormally complex interactions between tumor cells and the immune system. Tumor cells can create an immunosuppressive microenvironment by secreting a large number of immunosuppressive cytokines (such as IL-10, TGF β , and PGE2), recruiting immunosuppressive cells such as regulatory T cells (Tregs), activating negative regulatory pathways (CD80/CTLA4 and PD-1/PD-L1), and overexpressing immune regulatory enzymes such as indoleamine 2,3-dioxygenase (33), resulting in immune cell exhaustion or dysfunction, and affecting the normal anti-tumor function of the immune system.

T lymphocytes are the most important immune cells that exert anti-tumor effects and there are multiple cell subtypes, including CTLs, Th cells, and Tregs, all of which play important roles in the TME (34). T cells can recognize tumor cells by binding to major histocompatibility complex I/II (MHC-I/II) molecules on the surface of tumor cells via T-cell receptors (TCRs) on their own cell membrane. The human MHC molecules are also known as HLAs, where HLA-I molecules present endogenous antigens to CD8⁺ T cells, and HLA-II molecules present exogenous antigens to CD4⁺ T cells (35). Tumor antigens are presented to T cells on the surface of HLA-I molecules, recognized by TCRs, and then activate T cells to exert their effects. Patel et al. (36) performed immunohistochemistry staining for HLA-I molecules, including HLA-A, HLA-B/C, and B2-chain on 24 chordoma samples, and found that 21 out of 24 samples showed expression defects of at least one HLA-I molecule subunit. This to some extent indicates the existence of immune escape mechanisms in chordoma and alerts us to the limitations of various HLA-dependent immunomodulatory therapies. Furthermore, CTLs need to migrate into the tumor site and infiltrate into tumor tissue in order to interact with tumor cells (37). Therefore, a better understanding of the infiltration characteristics of CTLs in solid tumors is crucial for subsequent tumor immunotherapy.

Tumor-associated macrophages (TAMs) are a type of immune cells that play important roles in the TME, and can affect tumor cell proliferation, angiogenesis, and ECM formation, thus promoting or inhibiting tumor progression (38). TAMs are usually classified into two subtypes: M1 and M2 macrophages. M1 macrophages are activated by Th1 cytokine interferon-gamma (IFN- γ) or lipopolysaccharides (LPS) and have an anti-tumor effect through complement-mediated phagocytosis of tumor cells. In contrast, M2 macrophages are activated by Th2 cytokines interleukin-4/13 (IL-4/ IL-13), and play roles in fibrosis, anti-inflammatory response, and tumor progression (39-41). Macrophages are highly plastic, and can polarize into M0 subtype before differentiating into M1 or M2 subtype depending on the presence of specific cytokines (42). M2 macrophages have been shown to be associated with poor prognosis in various cancers such as colorectal cancer and bladder cancer, and promote tumor progression through mechanisms such as enhancing tumor cell proliferation, invasion, and angiogenesis (43-46). Currently, there are various therapeutic strategies targeting TAMs in the TME, such as inhibiting macrophage polarization towards the M2 subtype and enhancing macrophage phagocytosis by targeting the CD47/SIRPa signaling pathway. CD47 is a protein widely expressed on the surface of many tumor cells, and studies have shown that tumor cells overexpressing CD47 can interact with SIRPa on the surface of macrophages or dendritic cells, leading to activation of downstream SHP-1 and SHP-2 phosphatases, and inhibition of macrophage phagocytosis, thereby promoting immune escape (47). High expression of CD47 has been associated with poor prognosis in acute myeloid leukemia, ovarian cancer, and glioma (48, 49). Targeting CD47 has been shown to enhance macrophage phagocytosis and inhibit tumor growth in

animal models of ovarian cancer, small cell lung cancer, breast cancer, and other cancers (48, 50).

DCs are important antigen-presenting cells (APCs) that belong to the mononuclear phagocyte system (MPS) along with monocytes and macrophages (51). There are two main subtypes of DCs, plasmacytoid DCs (pDCs) and classical/traditional DCs (cDCs) (52). pDCs are capable of producing high levels of type I interferons and play important roles in regulating innate and adaptive immunity. Type I interferons are considered a double-edged sword in tumor immunity, as they can provide inflammatory signals to activate the immune system, but can also stimulate the inhibitory feedback of immune and tumor cells (53). In addition to type I interferons, pDCs secrete multiple cytokines that can stimulate the activation of DCs and macrophages, enhance the cytotoxicity of NK cells and CD8⁺ T cells, as well as promote the formation of immunosuppressive TME by recruiting Tregs and inducing the expression of immunosuppressive molecules (53, 54). cDCs can be divided into two subgroups, cDC1s and cDC2s (55). cDC1s can present MHC-I molecule antigens to CTLs, and cDCs can secrete cytokines such as CXCL9 and CXCL10 to recruit effector T cells and NK cells to the tumor site, as well as secrete cytokines such as IL-12 and CCL5 to maintain the function of effector T cells (56-58). cDC2s mainly present MHC-II molecule antigens to CD4⁺ T cells (56); when $CD4^+$ T cells migrate to lymph nodes, cDC2s can activate CD4⁺ T cells (59). Therefore, the relationship between DCs and T lymphocytes is closely related in tumor immune regulation, but there is still much to be explored in terms of their communication mechanisms.

NK cells are an important subset of lymphocytes in innate immunity that exert anti-tumor effects without the need for antigen stimulation. As a cytotoxic lymphocyte, NK cells can kill tumor cells through both the release of cytotoxic granules and antibodydependent cellular cytotoxicity (ADCC) and engagement of death ligand receptors. Therefore, NK cells can exert their effects in the early stages of tumor development. NK cells not only secrete proinflammatory cytokines such as IFN- γ , TNF, and granulocytemacrophage colony-stimulating factor (GM-CSF), but also secrete immunosuppressive factors such as TGF- β and IL-4 (60). Currently, some therapies targeting NK cells are in the exploration phase, such as transferring NK cells into tumor patients, CAR-NK cell therapy, and cytokine therapy.

3.2 Tumor immune microenvironment classification

Based on the infiltration of CD3⁺ and CD8⁺ T cells in the tumor center and invasive margin of the TME, the immune score system has been introduced to classify tumors into "cold" and "hot" types (61). Hot tumors are characterized by a large amount of infiltrating T cells that are activated, while cold tumors mainly exhibit a lack of T-cell infiltration (62, 63). Currently, the immune score system has been widely applied in various types of cancer, and its predictive accuracy for patient prognosis in colorectal cancer has surpassed that of the traditional TNM staging system (64). Therefore, the evaluation of tumor prognosis has shifted from a tumor-centric system to an immune-centric system. The widespread infiltration of CD3⁺ and CD8⁺ T cells is considered a hallmark of immune recognition initiated, but the immune microenvironment of tumors is complex. With the development of various research techniques, further exploration of the number, type, spatial distribution, and activation state of immune cells in the microenvironment has revealed significant heterogeneity in the composition and phenotype of tumor-infiltrating lymphocytes (TILs) (65-67). Some studies have identified the existence of CD8⁺FOXP3⁺ regulatory T cells' subtype using single-cell sequencing and multiple immunofluorescence (mIF) techniques (68, 69). Although CD8⁺ T cells generally exhibit cytotoxicity, CD8⁺FOXP3⁺ Treg cells exhibit immunosuppressive characteristics in certain diseases. However, their function in tumor immunity still requires further exploration, and therefore, relying solely on the infiltration of CD3⁺ and CD8⁺ T cells to evaluate the immune status of tumors is incomplete.

Therefore, more new and detailed classifications of the TIME have emerged, and exploring the TIME characteristics of different tumor types can provide more personalized and rational treatment strategies (70). Through the comprehensive analysis of the immune microenvironment characteristics of multiple tumors, Galon et al. proposed a new immunological classification, which mainly divides the TIME into the following four types: immune-infiltrated/hot tumors, immune-excluded tumors, immune-suppressed tumors, and immune-desert/cold tumors (70, 71).

The immune-excluded phenotype of tumors is characterized by a typical spatial distribution pattern, where a large number of T cells are confined to the periphery of the tumor and unable to infiltrate into the tumor parenchyma or interact with tumor cells to exert their anti-tumor effects. The immune-excluded TME is composed of various mechanisms, such as vascular structure and fibroblasts that can form a physical barrier to impede T-cell infiltration (72). Additionally, there are numerous soluble molecules in the microenvironment that can suppress T-cell chemotaxis into the tumor parenchyma (73), and when T cells come into contact with tumor cells, it can trigger immune-inhibitory signaling pathways such as PD-1/PD-L1, creating a dynamic exclusion mechanism. Furthermore, some researchers have proposed that hypoxia may be a triggering factor that induces the above mechanisms in tumors (74), resulting in the immune-excluded phenotype through remodeling of the ECM, modulation of TME metabolism, and upregulation of immune checkpoint such as CD47/SIRP α (75–78).

The difference between the immune-suppressed and immuneexcluded phenotypes lies in that a large number of immune cells are not restricted to the stromal regions surrounding the tumor, but rather their infiltration and expansion are limited by the immunosuppressive microenvironment (71), resulting in less immune cell infiltration in the tumor parenchyma. For example, research has revealed that low expression of IL-15 in colorectal cancer can inhibit the proliferation of B cells and T cells and increase the risk of tumor recurrence (79).

3.3 The exploration for chordoma TIME

Chordoma is a rare malignant bone tumor and there is limited basic research on its immune aspects. Literature on this subject only began to emerge in 2015 (80, 81), when Mathios examined PD-L1 expression in 10 samples of chordoma. This study opened the door to exploring the immune microenvironment of chordoma and revealed that PD-L1 was negative in chordoma cells, but was expressed in macrophages and T cells (80). Subsequent studies expanded the sample to 78 tissue specimens and further investigated PD-L1 expression, as well as its relationship with clinical characteristics of patients. Unlikethe findings of Mathios, these studies showed that PD-L1 was positively expressed in tumor cells and was associated with poor prognosis of patients with chordoma (81, 82). However, the infiltration of immune cells was subjectively judged only based on the histological characteristics of the cells at the time, and analyzing the infiltration of individual immune cells or the expression of PD-L1 in tumor tissue alone could not fully elucidate the TIME characteristics of chordoma. In 2016, Inagaki used immunohistochemistry to explore the immune infiltration characteristics of various bone tumors, including two cases of chordoma. The study analyzed the infiltration of various immune cells and revealed that macrophages were the major cell subtype, followed by T cells and dendritic cells (DCs), in multiple bone tumors (83).

Different subgroups of immune cells often play different roles, as M1 and M2 macrophages even have opposite functions in many diseases. Therefore, analysis of cell types needs to delve into various subtypes. Zou analyzed two subtypes of T cells (CD8⁺ TILs and Foxp3⁺ TILs) in chordoma in 2018 and found that the ratio of CD8⁺ TILs to Foxp3⁺ TILs was 3.3 times and related to the patient's overall survival (OS) (84). This team then developed an immune scoring system based on the distribution of CD3⁺ and CD8⁺ TILs in the tumor and invasive edge in the following year, which could serve as a prognostic factor for the patient's local recurrence-free survival (LRFS) and OS (85). In addition, the team used mIF to analyze the expression of PD-1, CD3, CD8, CD20, and FOXP3 in 2020, and validated the results by flow cytometry (86). They constructed an immune scoring system based on the density of interstitial CD8⁺ TILs, interstitial Foxp3⁺ TILs, tumor area Foxp3⁺ TILs, and tumor area PD-1⁺ TILs, which showed correlations with patient LRFS and OS. This is the first time that an objective experimental technique, different from immunohistochemistry and H&E staining, which require subjective interpretation, has been used in exploring the immune microenvironment of chordoma.

In a large-scale sarcoma study conducted in 2020 (1,242 cases, including over 27 types of sarcoma and 28 cases of chordoma), M1 and M2 subtypes of macrophages were analyzed for the first time. The results showed that the number of macrophages infiltrating the tumors was higher than that of lymphocytes in almost all sarcomas, and the infiltration of M2 macrophages was dominant. Among them, chordoma had the highest macrophage/TIL ratio among the various types of sarcoma, exceeding 10 times, and the M2/M1 ratio

was 1.5 times. The expression of the macrophage-related immune checkpoint CD47/SIRP was also extremely high in chordoma, with CD47 expressed on all tumor cells in 82% of cases, and SIRP⁺ macrophage infiltration present in 71% of cases (87).

In addition to the PD-1/PD-L1 pathway of T-cell immune checkpoints, other immune checkpoints have also been studied, which can induce T-cell apoptosis by binding to their corresponding ligands. In 2019, Zhou (88) revealed that TIM3⁺ TIL infiltration is associated with invasion of chordoma. In 2020, He (89) analyzed 32 specimens of chordoma and found that CTLA-4 showed positive expression on the surface of tumor cells and TILs in all cases.

Both the immune surveillance function of the body to clear foreign invaders and T-cell therapy using immune checkpoint inhibitors (ICIs) rely on the T cells' ability to exert cytotoxic effects. The premise for T cells to exert their effector function is to recognize the human leukocyte antigen-I (HLA-I) molecules on the surface of tumor cells and the tumor antigens. However, studies have found that HLA-I expression is downregulated in 21 out of 24 cases of chordoma (36), indicating that the cytotoxic ability of T cells may be limited in chordoma, and immunotherapy relying on T cells may have limited efficacy.

3.4 Overall immune landscape of chordoma

In recent years, single-cell sequencing technology (scRNA-seq) has made great strides. Unlike traditional transcriptome sequencing technologies, scRNA-seq can comprehensively reveal gene expression at the single-cell level. Currently, it has been widely applied in various research studies. In 2022, more objective and comprehensive exploration of chordoma was conducted through single-cell sequencing and mIF (90, 91) (Figure 2). Duan et al. first used scRNA-seq to depict the gene expression profile of chordoma and analyze tumor cells, immune cells, and fibroblasts in the microenvironment (91). This marks the beginning of a highthroughput era for exploring the immune microenvironment of chordoma. Duan found that mononuclear phagocytes were the most prominent infiltrating immune cell (14.0%). Based on the gene expression characteristics of macrophages, M2 macrophages were widely infiltrated and may play a critical role in chordoma angiogenesis. T cells and NK cells (11.7%) only expressed a small amount of immune checkpoint, suggesting that various ICIs targeting T and NK cells may have poor efficacy in chordoma.

Owing to the rarity of chordoma, there is currently a limited number of studies on its immunemicroenvironment and the number of samples involved is relatively small. In some cases, the conclusions may even be conflicting. Therefore, we are still unable to make further judgments on the overall status of the immune microenvironment of chordoma. Further research is needed to provide a more comprehensive and in-depth description of the immune microenvironment of chordoma.



4 Immunotherapy strategies for chordoma

4.1 Immune checkpoint inhibitor therapy

Immune checkpoint pathways are inhibitory signaling pathways that exist within the immune system. ICI therapy works by blocking the immune checkpoint pathway, and releasing the "brakes" on immune cells to restore their anti-tumor function. Under normal physiological conditions, immune checkpoints are designed to prevent overactivation of immune cells. The emergence of tumors is a result of mutations in normal cells. Immune checkpoint on the surface of immune cells can bind to corresponding receptors on the surface of tumor cells and cause immune dysfunction through various pathways, such as inhibition of T-cell proliferation, survival, cytokine secretion, and other effector functions, leading to immune escape.

Currently, the most extensively studied checkpoint pathways in the field of cancer research are programmed death protein 1 (PD-1/ CD279) and its ligands PD-L1/CD274/B7-H1 and PD-L2/CD273/ B7-DC, cytotoxic T lymphocyte-associated antigen 4 (CTLA-4), Tcell immunoglobulin and mucin domain 3 (TIM3), and lymphocyte activation gene 3 (LAG3), among others (92).

PD-1 and PD-L1 both belong to the immunoglobulin superfamily type I transmembrane proteins. PD-1 is expressed on the surface of various immune cells such as T cells, B cells, NK cells, and DC cells (93), while PD-L1 is mainly expressed on the surface of tumor cells and tumor-infiltrating lymphocytes. The binding of PD-L1 on tumor cells with PD-1 on T cells can cause a conformational change in PD-1, leading to the phosphorylation of the cytoplasmic immune receptor tyrosine-based inhibitory motif (ITIM) and immune receptor tyrosine-based switch motif (ITSM). The phosphorylated tyrosine-based motifs subsequently recruit SHP-2 and SHP-1 proteins, which weaken T-cell activation signals and inhibit T-cell activation. In addition, many studies have focused on the expression regulation mechanisms of PD-L1 at multiple levels. For example, at the gene level, Roemer et al. (94) found that 97% of classical Hodgkin lymphoma cases had mutations in the PD-L1/PD-L2 gene locus. At the protein level, Burr et al. (95) found that the protein CMTM6 can maintain the stable expression of PD-L1 on the cell membrane by binding to it, and participate in inhibiting T-cell activation and promoting tumor progression.

Currently, studies revealed that PD-L1 expression in chordoma is associated with tumor metastasis and tumor staging, and is a poor prognostic factor for both LRFS and OS. However, there is no consistent opinion among studies when analyzing the expression of PD-L1 on tumor cells and immune cells. This may be due to the inadequate sample used in the studies, but to some extent, it shows the potential for targeted PD-1/PD-L1 therapy. The expression and regulatory mechanisms of immune checkpoints on the surface of chordoma still need to be explored in depth. Currently, several drugs targeting the PD-1/PD-L1 pathway in chordoma are undergoing clinical trials (96, 97) (Table 1). Pembrolizumab and nivolumab are monoclonal antibodies targeting PD-1 and have demonstrated robust efficacy in areas such as melanoma and nonsmall cell lung cancer. Pembrolizumab has become the most widely used drug in ICI therapy for chordoma (98). It has been shown that DCs and macrophages in the chordoma microenvironment highly express regulators of PD-L1, such as AXL, TLR3, and CD40, suggesting that our treatment targeting the PD-1/PD-L1 axis in combination with blockade of the above molecules may provide better therapeutic efficacy (99).

In addition, studies have shown that the expression levels of CTLA-4 and TIM3 are both high in chordoma, and their expression is related to the invasiveness and recurrence of chordoma (88). In addition to immune checkpoints on T cells, the role of immune checkpoints on other immune cells is gradually being revealed. TAMs infiltrate heavily in chordoma and are mainly M2 macrophages, which participate in the formation of the immunosuppressive microenvironment of chordoma. CD47,

| Year | Immunotherapy strategy | Registration number | Sample of chordoma | Sample of all patients | Study phase | Drug |
|------|---------------------------------|------------------------|--------------------|------------------------|----------------|---|
| 2023 | ICI | NCT03012620 | 34 | 97 | Phase 2 | Pembrolizumab |
| 2022 | ICI | NCT02815995 | 5 | 57 | Phase 2 | Durvalumab (anti-PD-L1) + tremelimumab (anti-CTLA-4) |
| 2021 | Tumor vaccine | NCT04134312 | 10 | 13 | Phase 1 | MVA-BN-brachyury-TRICOM |
| 2021 | Tumor vaccine + radiotherapy | NCT02383498 | 24 | 24 | Phase 2 | GI-6301 + radiotherapy |
| 2020 | Tumor vaccine | NCT03349983 | 3 | 10 | Phase 1 | BN-brachyury (MVA-brachyury + FPV- brachyury) |
| 2017 | ICI | NCT01772004 | 1 | 53 | Phase 1 | Avelumab (anti-PD-L1) |
| 2017 | Tumor vaccine | NCT02179515 | 13 | 38 | Phase 1 | MVA-brachyury-TRICOM |
| 2015 | Tumor vaccine | NCT01519817 | 11 | 34 | Phase 1 | GI-6301 |

TABLE 1 Immunotherapy clinical trial for chordoma.

SIRPα, and TIM3 are all expressed on TAMs, and chordoma CD47 expression shows the highest degree of expression among various sarcomas (87, 91), However, whether it can be used as a prognostic factor and therapeutic target for chordoma is currently unknown.

To date, the ICI therapy for chordoma is mainly limited to targeting PD-1/PD-L1, while research on other target points is relatively scarce and only limited to preclinical stages, which seriously hinders the selection of ICI therapy strategies for chordoma. Moreover, the current ICI therapy strategy only benefits a subset of patients and most patients do not show sustained remission. In addition, immune-related adverse events (IRAEs) such as myocarditis and colitis have been observed in some patients (100). This may be due to the fact that anti-tumor immune regulation requires the complex TME to exert its effects (101), and there are heterogeneities in the expression of immune checkpoints, differences in the distribution of immune cells, and interference from other inhibitory pathways in different individuals. Therefore, it is urgent to better understand the status and regulatory mechanisms of the chordoma microenvironment to guide the safer and more effective use of ICI therapy strategies (102, 103).

4.2 Adoptive cell therapy

Adoptive cell therapy (ACT) is one type of cellular immunotherapy, and it involves the transfer of immune cells infused into patients where they can mediate strong anti-tumor responses, resulting in tumor regression (104). Because of the HLA expression defects in chordoma cells, some T cell-dependent immunotherapy strategies may be limited. ACT involves the infusion of modified immune cells into patients, and unlike T cells that rely on the combination of TCR and tumor cell surface HLA molecules, ACT can specifically recognize the tumor cell surface antigen to exert immune effects and attack tumor cells to achieve anti-tumor goals. This may provide an effective treatment strategy for chordoma patients with downregulated HLA molecules. Among them, chimeric antigen receptor T-cell therapy (CAR-T) is currently a hot spot in immunotherapy due to its strong specificity and multiple advantages.

CAR is a cell surface receptor synthesized artificially *in vitro*, mainly composed of a single-chain variable fragment (scFv) of monoclonal antibodies in the extracellular domain, a CD3 ζ molecule in the intracellular domain, and a transmembrane domain that couples them together. The extracellular domain of scFv does not rely on HLA molecules on the surface of tumor cells to present antigens, but specifically recognizes the surface antigen of tumor cells, thereby activating the signal structure domain of the intracellular domain.

By isolating T cells from patients and genetically modifying them using virus vectors and other methods in vitro, T cells can stably express CARs with specificity, which are then expanded and infused into patients. With the advancement of technology and research, second-, third-, and fourth-generation CARs have emerged. Second-generation CARs enhance the effect of CAR-T cells by adding a co-stimulatory structure such as CD28, CD137/4-1BB to the intracellular domain. Third-generation CARs contain two co-stimulatory domains linked to CD3ζ, which can enhance and prolong T-cell activation time (105). Fourth-generation CAR-T cells, also known as TRUCK, not only increase the co-stimulatory structure domain but can also locally secrete cytokines such as IL-12 after targeting tumor cells, enhance T-cell activation, recruit immune cells to attack other tumor cells not recognized by CAR-T cells, and regulate vascular generation to improve the immunosuppressive microenvironment of solid tumors (106).

The CAR-T cells recognize tumor antigens similar to antigenantibody reactions and have stronger specificity compared to other immunotherapies. Currently, CAR-T cell therapy has shown good efficacy in the treatment of hematologic malignancies, and several CD19-targeted CAR-T cell products such as Kymriah and Yescarta have been approved by the FDA for the treatment of B-cell acute lymphoblastic leukemia and diffuse large B-cell lymphoma (107, 108). However, the efficacy of CAR-T therapy in solid tumors is not as significant as in hematologic malignancies, mainly due to the immunosuppressive microenvironment of solid tumors, which limits

the migration of CAR-T cells to tumor sites. Even if they migrate to the tumor site, the surrounding immunosuppressive cells and cytokines can also affect the further effectiveness of CAR-T cells (109). Therefore, current studies not only focus on the structure of CAR on immune cells but also attempt to explore how to modify immune cells to enhance immunotherapy and reduce toxicity. For example, by increasing the receptor for chemokine on CAR-T cells, immune cells can infiltrate into the tumor stroma more rapidly (110, 111), knocking out the inhibitory receptors on the cell surface to inhibit early immune cell exhaustion (112), and embedding dual CAR targeting two antigens on the same immune cell to avoid the occurrence of immune escape caused by the loss of a single antigen (113).

However, CAR-T therapy has shown slow progress in solid tumors, especially in rare tumors like chordoma. Currently, there is only one article on CAR-T cell therapy for chordoma, in which Long et al. (114) constructed CAR-T cells targeting B7-H3 and demonstrated anti-tumor efficacy in vitro. However, the positivity rate of B7-H3 expression in chordoma is only 16%, suggesting that CAR-T cells targeting B7-H3 may only have anti-tumor efficacy in a few patients. CAR-T therapy relies primarily on identifying special targets, and ideal tumor antigens should have high specificity and only be expressed in tumor cells. However, little is known about specific antigens for chordoma at present.

4.3 Tumor vaccine therapy

A tumor vaccine therapy involves administering low doses of inactivated antigens to activate the immune response in the body of patients (115). Brachyury is a transcription factor found in multiple tumors that can promote EMT. It is expressed in almost all patients with chordoma, and chordoma-targeted tumor vaccines are developed based on this, including the Yeast-Brachyury (GI-6301) vaccine (16, 116), the MVA-Brachyury-TRICOM vaccine constructed using the modified vaccinia Ankara (MVA) carrier, and subsequent modified vaccines (117-119). Clinical benefits have been demonstrated in chordoma patients receiving tumor vaccines, and trials have found that pre-treatment with ICI or radiotherapy can enhance the therapeutic efficacy of vaccines, providing a potential new treatment approach for chordoma patients. However, unfortunately, a phase II trial of radiotherapy combined with the Yeast-Brachvury vaccine (GI-6301) in patients with advanced unresectable chordoma showed no complete response (CR) and the efficacy was not satisfactory, leading to termination of the clinical trial before reaching the endpoint (16) (Table 1).

4.4 Breaking the immunosuppressive microenvironment

Overall, "hot tumors" often exhibit better responses to immunotherapy, so many studies are trying to convert "cold tumors" into "hot tumors" to enhance their therapeutic effects, which is a new direction of current research. According to existing data, although we do not have a clear understanding of the specific cell phenotypes and infiltration proportions in the microenvironment of chordoma, numerous studies have shown that immune cells, especially T cells, mainly infiltrate the fibrous septa in the stroma of chordoma, exhibiting immune-excluded phenotypes (Figure 3). Therefore, breaking their immune suppression from multiple directions based on their spatial distribution characteristics may open up new therapeutic strategies for chordoma.



FIGURE 3

The immune escape mechanisms of chordoma and immunotherapy strategies. Chordoma was shown with immune-excluded microenvironment, the fibroblasts impede the infiltration of immune cells. Multiple immune checkpoints were upregulated on the tumor cells' and TIL cells' surfaces, which inhibit the immune response effect. The current immunotherapy strategies are mainly focused on immune checkpoint inhibitor therapy, with ACT and tumor vaccine also emerging

Cancer-associated fibroblasts (CAFs) are the main cell component of tumor stroma and are currently a hot target for tumor therapy, and research has shown that activation of the TGF-B pathway may be the molecular mechanism underlying the immune exclusion phenotype. TGF- β is a typical immunosuppressive molecule that can act on surface receptors of fibroblasts, indirectly affecting the localization and migration of immune cells in tumors by increasing the expression of immunosuppressive cytokines and regulating various ECM components such as collagen, α -SMA, and versican, thereby forming an immuneexcluded phenotype that affects the occurrence, development, and metastasis of tumors (120). It has been revealed that both TGF- β and its multiple receptors are highly expressed in chordoma, where TGF- β is mainly secreted by microenvironmental fibroblasts and macrophages, which can both drive tumor cell EMT progression and enhance their own malignant ability, and also regulate the immune microenvironment by converting CD4⁺ T cells into Treg cells, regulating NK cells and other functions.

Inhibiting CAF activation includes the use of inhibitors of focal adhesion kinase (FAK), fibroblast growth factor receptor (FGFR), and TGF- β (121–123), as well as downstream pathways that regulate the TME by inhibiting the activation of fibroblasts, including the use of angiotensin receptor blockers and degradation of hyaluronic acid produced by fibroblasts (124, 125), which are all expected to have a regulatory effect on the TME. Tumor progression in chordoma has been found to be inhibited by blocking the TGF- β pathway (126), but the regulatory effects on the fibroblast in TME remain to be further explored.

5 Conclusion

As a rare malignant bone tumor, the research of chordoma has been slow. Macrophages and T cells were dominated in the chordoma microenvironment, which are likely to be the crucial effector cells in tumor immune regulation and may be the therapeutic targets for chordoma immunotherapy. HLA-I is downregulated in chordoma, suggesting that T cell-dependent immunotherapy strategy may be impeded. Various classic immune checkpoints and immuneexcluded phenotype were progressively revealed, which may be the essential mechanisms of chordoma immune escape.

6 Future directions

The limited basic research of chordoma has severely restricted the choice of clinical treatment strategies. Previous studies have focused mainly on the driving mechanisms of tumor cells themselves, but tumor progression is not limited to the tumor cells alone. The exploration of the immune microenvironment of chordoma started in 2015, and by summarizing the findings so far, we found that macrophages and T cells were the most highly infiltrated in chordoma and may be the crucial effector cells in tumor immune regulation. A variety of immunosuppressive checkpoints such as PD-1/PD-L1, CD47/SIRP α , TIM3, and CTLA4 are expressed on the surface of both chordoma cells and immune cells, and ICIs are the

most widely performed strategy in the field of chordoma immunotherapy. It is worthwhile to further investigate the molecular regulation mechanisms of immune checkpoints and whether the dual-target immunotherapy can achieve better efficacy.

However, HLA-I is downregulated on the surface of chordoma cells, which is a prerequisite for T cells to exert tumor-killing effects, suggesting that T cell-dependent immunotherapy strategy may be impeded. Deciphering the intricate mechanism of HLA downregulation is a direction worthy of further investigation, and whether CAR-T therapy, which does not depend on the identification of TCR-HLA, can be a promising approach to immune-mediated chordoma treatment. Similar to tumor vaccines, the efficacy is highly dependent on the specificity of the tumor antigens; therefore, specific chordoma tumor neoantigens need to be immediately identified.

At the same time, we found that chordoma exhibits an immuneexcluded phenotype, with a large number of immune cells infiltrating the mesenchymal region and preventing it from exerting an immune response. Exploring the genetic differences between parenchymal immune cells and mesenchymal immune cells may provide novel therapeutic targets for chordoma and alter its immune-excluded phenotype.

Brachyury and various signaling pathways are revealed to be associated with the malignant capacity of chordoma cells, but their functions on chordoma immune microenvironment remains to be unclear. For example, the VEGF and TGF- β pathways are dysregulated in chordoma, whether inhibiting the VEGF pathway could modulate the vascular structure of the microenvironment and whether targeting TGF- β could inhibit its effect on fibroblasts, which creates a tough physical barrier. Thus, altering the immune-excluded phenotype will hopefully improve the efficacy of various immunotherapeutic strategies.

Author contributions

JX: Data curation, Writing – original draft, Writing – review & editing. QS: Data curation, Writing – original draft, Writing – review & editing. BW: Data curation, Writing – original draft. TJ: Supervision, Writing – original draft. WG: Supervision, Writing – review & editing. TR: Funding acquisition, Supervision, Writing – review & editing. Tr: Funding acquisition, Supervision, Writing – review & editing.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

1. Cheng EY, Ozerdemoglu RA, Transfeldt EE, Thompson RC Jr. Lumbosacral chordoma. Prognostic factors and treatment *Spine* (1999) 24(16):1639–45 doi: 10.1097/00007632-199908150-00004

2. McMaster ML, Goldstein AM, Bromley CM, Ishibe N, Parry DM. Chordoma: incidence and survival patterns in the United States, 1973-1995. *Cancer causes control CCC* (2001) 12(1):1–11. doi: 10.1023/a:1008947301735

3. Pan Y, Lu L, Chen J, Zhong Y, Dai Z. Analysis of prognostic factors for survival in patients with primary spinal chordoma using the SEER Registry from 1973 to 2014. *J orthopaedic Surg Res* (2018) 13(1):76. doi: 10.1186/s13018-018-0784-3

4. Boriani S, Chevalley F, Weinstein JN, Biagini R, Campanacci L, De Iure F, et al. Chordoma of the spine above the sacrum. Treatment and outcome in 21 cases. *Spine* (1996) 21(13):1569–77. doi: 10.1097/00007632-199607010-00017

5. Catton C, O'Sullivan B, Bell R, Laperriere N, Cummings B, Fornasier V, et al. Chordoma: long-term follow-up after radical photon irradiation. *Radiotherapy Oncol J Eur Soc Ther Radiol Oncol* (1996) 41(1):67–72. doi: 10.1016/s0167-8140(96)91805-8

6. Azzarelli A, Quagliuolo V, Cerasoli S, Zucali R, Bignami P, Mazzaferro V, et al. Chordoma: natural history and treatment results in 33 cases. *J Surg Oncol* (1988) 37 (3):185–91. doi: 10.1002/jso.2930370311

 York JE, Kaczaraj A, Abi-Said D, Fuller GN, Skibber JM, Janjan NA, et al. Sacral chordoma: 40-year experience at a major cancer center. *Neurosurgery* (1999) 44(1):74– 9. doi: 10.1097/00006123-199901000-00041

8. Walcott BP, Nahed BV, Mohyeldin A, Coumans J-V, Kahle KT, Ferreira MJ. Chordoma: current concepts, management, and future directions. *Lancet Oncol* (2012) 13(2):e69–76. doi: 10.1016/s1470-2045(11)70337-0

9. Pitt JM, Marabelle A, Eggermont A, Soria JC, Kroemer G, Zitvogel L. Targeting the tumor microenvironment: removing obstruction to anticancer immune responses and immunotherapy. *Ann Oncol Off J Eur Soc Med Oncol* (2016) 27(8):1482–92. doi: 10.1093/annonc/mdw168

10. Lu W, Yu W, He J, Liu W, Yang J, Lin X, et al. Reprogramming immunosuppressive myeloid cells facilitates immunotherapy for colorectal cancer. *EMBO Mol Med* (2021) 13(1):e12798. doi: 10.15252/emmm.202012798

11. Liu Y, Xun Z, Ma K, Liang S, Li X, Zhou S, et al. Identification of a tumour immune barrier in the HCC microenvironment that determines the efficacy of immunotherapy. *J Hepatol* (2023) 78(4):770–82. doi: 10.1016/j.jhep.2023.01.011

 Sheppard HE, Dall'Agnese A, Park WD, Shamim MH, Dubrulle J, Johnson HL, et al. Targeted brachyury degradation disrupts a highly specific autoregulatory program controlling chordoma cell identity. *Cell Rep Med* (2021) 2(1):100188. doi: 10.1016/ j.xcrm.2020.100188

13. Sharifnia T, Wawer MJ, Chen T, Huang QY, Weir BA, Sizemore A, et al. Small-molecule targeting of brachyury transcription factor addiction in chordoma. *Nat Med* (2019) 25(2):292–300. doi: 10.1038/s41591-018-0312-3

14. Presneau N, Shalaby A, Ye H, Pillay N, Halai D, Idowu B, et al. Role of the transcription factor T (brachyury) in the pathogenesis of sporadic chordoma: a genetic and functional-based study. *J Pathol* (2011) 223(3):327–35. doi: 10.1002/path.2816

15. Shah SR, David JM, Tippens ND, Mohyeldin A, Martinez-Gutierrez JC, Ganaha S, et al. Brachyury-YAP regulatory axis drives stemness and growth in cancer. *Cell Rep* (2017) 21(2):495–507. doi: 10.1016/j.celrep.2017.09.057

16. DeMaria PJ, Bilusic M, Park DM, Heery CR, Donahue RN, Madan RA, et al. Randomized, double-blind, placebo-controlled phase II study of yeast-brachyury vaccine (GI-6301) in combination with standard-of-care radiotherapy in locally advanced, unresectable chordoma. *oncologist* (2021) 26(5):e847–e58. doi: 10.1002/onco.13720

17. Liu Y, Liu X, Lin C, Jia X, Zhu H, Song J, et al. Noncoding RNAs regulate alternative splicing in Cancer. J Exp Clin Cancer Res CR (2021) 40(1):11. doi: 10.1186/s13046-020-01798-2

18. Han B, Xi W, Hong Y, Gu L, Chao Y, Li L, et al. Mutual regulation of noncoding RNAs and RNA modifications in psychopathology: Potential therapeutic targets for psychiatric disorders? *Pharmacol Ther* (2022) 237:108254. doi: 10.1016/j.pharmthera.2022.108254

19. Zhang K, Liu Z, Wang Z, Zhou Z, Shao X, Hua X, et al. Long Non-coding RNA MDFIC-7 promotes chordoma progression through modulating the miR-525-5p/ARF6 axis. *Front Oncol* (2021) 11:743718. doi: 10.3389/fonc.2021.743718

20. Gong F, Wang X, Sun Q, Su X, Hu X, Liu B. Long non-coding RNA LINC00525 interacts with miR-31-5p and miR-125a-5p to act as an oncogenic molecule in spinal chordoma. *Biochem Biophys Res Commun* (2021) 536:80–7. doi: 10.1016/j.bbrc.2020.12.042

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21. Zhang H, Yang K, Ren T, Huang Y, Tang X, Guo W. miR-16-5p inhibits chordoma cell proliferation, invasion and metastasis by targeting Smad3. *Cell Death Dis* (2018) 9(6):680. doi: 10.1038/s41419-018-0738-z

22. Lou J, Zhang H, Huang Q, Chen C, Wang W, Niu J, et al. The Malignancy of chordomas is enhanced via a circTLK1/miR-16-5p/Smad3 positive feedback axis. *Cell Death Discovery* (2023) 9(1):64. doi: 10.1038/s41420-023-01332-1

23. Chen Y, Huang X, Zhu K, Li C, Peng H, Chen L, et al. LIMD2 is a prognostic and predictive marker in patients with esophageal cancer based on a ceRNA network analysis. *Front Genet* (2021) 12:774432. doi: 10.3389/fgene.2021.774432

24. Li S, Liu F, Zheng K, Wang W, Qiu E, Pei Y, et al. CircDOCK1 promotes the tumorigenesis and cisplatin resistance of osteogenic sarcoma via the miR-339-3p/IGF1R axis. *Mol Cancer* (2021) 20(1):161. doi: 10.1186/s12943-021-01453-0

25. Tamborini E, Miselli F, Negri T, Lagonigro MS, Staurengo S, Dagrada GP, et al. Molecular and biochemical analyses of platelet-derived growth factor receptor (PDGFR) B, PDGFRA, and KIT receptors in chordomas. *Clin Cancer Res an Off J Am Assoc Cancer Res* (2006) 12(23):6920–8. doi: 10.1158/1078-0432.Ccr-06-1584

26. Weinberger PM, Yu Z, Kowalski D, Joe J, Manger P, Psyrri A, et al. Differential expression of epidermal growth factor receptor, c-Met, and HER2/neu in chordoma compared with 17 other Malignancies. *Arch otolaryngology-head Neck Surg* (2005) 131 (8):707–11. doi: 10.1001/archotol.131.8.707

27. Presneau N, Shalaby A, Idowu B, Gikas P, Cannon SR, Gout I, et al. Potential therapeutic targets for chordoma: PI3K/AKT/TSC1/TSC2/mTOR pathway. *Br J Cancer* (2009) 100(9):1406–14. doi: 10.1038/sj.bjc.6605019

28. Hu Y, Mintz A, Shah SR, Quinones-Hinojosa A, Hsu W. The FGFR/MEK/ERK/ brachyury pathway is critical for chordoma cell growth and survival. *Carcinogenesis* (2014) 35(7):1491–9. doi: 10.1093/carcin/bgu014

29. Lohberger B, Scheipl S, Heitzer E, Quehenberger F, de Jong D, Szuhai K, et al. Higher cMET dependence of sacral compared to clival chordoma cells: contributing to a better understanding of cMET in chordoma. *Sci Rep* (2021) 11(1):12466. doi: 10.1038/s41598-021-92018-0

30. Zhang H, Yang K, Ren T, Huang Y, Liang X, Yu Y, et al. miR-100-5p inhibits Malignant behavior of chordoma cells by targeting IGF1R. *Cancer Manage Res* (2020) 12:4129–37. doi: 10.2147/cmar.S252185

31. Al Shihabi A, Davarifar A, Nguyen HTL, Tavanaie N, Nelson SD, Yanagawa J, et al. Personalized chordoma organoids for drug discovery studies. *Sci Adv* (2022) 8(7): eabl3674. doi: 10.1126/sciadv.abl3674

32. Farhood B, Najafi M, Mortezaee K. CD8(+) cytotoxic T lymphocytes in cancer immunotherapy: A review. J Cell Physiol (2019) 234(6):8509-21. doi: 10.1002/jcp.27782

33. Beatty GL, Gladney WL. Immune escape mechanisms as a guide for cancer immunotherapy. *Clin Cancer Res an Off J Am Assoc Cancer Res* (2015) 21(4):687–92. doi: 10.1158/1078-0432.Ccr-14-1860

34. Ostroumov D, Fekete-Drimusz N, Saborowski M, Kühnel F, Woller N. CD4 and CD8 T lymphocyte interplay in controlling tumor growth. *Cell Mol Life Sci CMLS* (2018) 75(4):689–713. doi: 10.1007/s00018-017-2686-7

35. Horton R, Wilming L, Rand V, Lovering RC, Bruford EA, Khodiyar VK, et al. Gene map of the extended human MHC. *Nat Rev Genet* (2004) 5(12):889–99. doi: 10.1038/nrg1489

36. Patel SS, Nota SP, Sabbatino F, Nielsen GP, Deshpande V, Wang X, et al. Defective HLA class I expression and patterns of lymphocyte infiltration in chordoma tumors. *Clin orthopaedics related Res* (2020) 479(6):1373–82. doi: 10.1097/corr.000000000001587

37. Corgnac S, Boutet M, Kfoury M, Naltet C, Mami-Chouaib F. The emerging role of CD8(+) tissue resident memory T (T(RM)) cells in antitumor immunity: A unique functional contribution of the CD103 integrin. *Front Immunol* (2018) 9:1904. doi: 10.3389/fimmu.2018.01904

38. Pan Y, Yu Y, Wang X, Zhang T. Tumor-associated macrophages in tumor immunity. Front Immunol (2020) 11:583084. doi: 10.3389/fimmu.2020.583084

39. Goerdt S, Orfanos CE. Other functions, other genes: alternative activation of antigenpresenting cells. *Immunity* (1999) 10(2):137-42. doi: 10.1016/s1074-7613(00)80014-x

40. Mantovani A, Sica A, Sozzani S, Allavena P, Vecchi A, Locati M. The chemokine system in diverse forms of macrophage activation and polarization. *Trends Immunol* (2004) 25(12):677–86. doi: 10.1016/j.it.2004.09.015

41. Biswas SK, Mantovani A. Macrophage plasticity and interaction with lymphocyte subsets: cancer as a paradigm. *Nat Immunol* (2010) 11(10):889–96. doi: 10.1038/ni.1937

42. Tarique AA, Logan J, Thomas E, Holt PG, Sly PD, Fantino E. Phenotypic, functional, and plasticity features of classical and alternatively activated human macrophages. *Am J Respir Cell Mol Biol* (2015) 53(5):676–88. doi: 10.1165/rcmb.2015-0012OC

43. Martínez VG, Rubio C, Martínez-Fernández M, Segovia C, López-Calderón F, Garín MI, et al. BMP4 induces M2 macrophage polarization and favors tumor progression in bladder cancer. *Clin Cancer Res an Off J Am Assoc Cancer Res* (2017) 23(23):7388–99. doi: 10.1158/1078-0432.Ccr-17-1004

44. Mantovani A, Marchesi F, Malesci A, Laghi L, Allavena P. Tumour-associated macrophages as treatment targets in oncology. *Nat Rev Clin Oncol* (2017) 14(7):399–416. doi: 10.1038/nrclinonc.2016.217

45. Liang ZX, Liu HS, Wang FW, Xiong L, Zhou C, Hu T, et al. LncRNA RPPH1 promotes colorectal cancer metastasis by interacting with TUBB3 and by promoting exosomes-mediated macrophage M2 polarization. *Cell Death Dis* (2019) 10(11):829. doi: 10.1038/s41419-019-2077-0

46. Cao J, Dong R, Jiang L, Gong Y, Yuan M, You J, et al. LncRNA-MM2P identified as a modulator of macrophage M2 polarization. *Cancer Immunol Res* (2019) 7(2):292–305. doi: 10.1158/2326-6066.Cir-18-0145

47. Weiskopf K. Cancer immunotherapy targeting the CD47/SIRP α axis. Eur J Cancer (2017) 76:100–9. doi: 10.1016/j.ejca.2017.02.013

48. Willingham SB, Volkmer JP, Gentles AJ, Sahoo D, Dalerba P, Mitra SS, et al. The CD47-signal regulatory protein alpha (SIRPa) interaction is a therapeutic target for human solid tumors. *Proc Natl Acad Sci United States America* (2012) 109(17):6662–7. doi: 10.1073/pnas.1121623109

49. Majeti R, Chao MP, Alizadeh AA, Pang WW, Jaiswal S, Gibbs KDJr., et al. CD47 is an adverse prognostic factor and therapeutic antibody target on human acute myeloid leukemia stem cells. *Cell* (2009) 138(2):286–99. doi: 10.1016/j.cell.2009.05.045

50. Weiskopf K, Jahchan NS, Schnorr PJ, Cristea S, Ring AM, Maute RL, et al. CD47-blocking immunotherapies stimulate macrophage-mediated destruction of small-cell lung cancer. *J Clin Invest* (2016) 126(7):2610–20. doi: 10.1172/jci81603

51. Guilliams M, Ginhoux F, Jakubzick C, Naik SH, Onai N, Schraml BU, et al. Dendritic cells, monocytes and macrophages: a unified nomenclature based on ontogeny. *Nat Rev Immunol* (2014) 14(8):571–8. doi: 10.1038/nri3712

52. Naik SH, Sathe P, Park HY, Metcalf D, Proietto AI, Dakic A, et al. Development of plasmacytoid and conventional dendritic cell subtypes from single precursor cells derived in vitro and in vivo. *Nat Immunol* (2007) 8(11):1217–26. doi: 10.1038/ni1522

53. Snell LM, McGaha TL, Brooks DG. Type I interferon in chronic virus infection and cancer. Trends Immunol (2017) 38(8):542–57. doi: 10.1016/j.it.2017.05.005

54. Koucký V, Bouček J, Fialová A. Immunology of plasmacytoid dendritic cells in solid tumors: A brief review. *Cancers* (2019) 11(4):470. doi: 10.3390/cancers11040470

55. Schlitzer A, Sivakamasundari V, Chen J, Sumatoh HR, Schreuder J, Lum J, et al. Identification of cDC1- and cDC2-committed DC progenitors reveals early lineage priming at the common DC progenitor stage in the bone marrow. *Nat Immunol* (2015) 16(7):718–28. doi: 10.1038/ni.3200

56. Mildner A, Jung S. Development and function of dendritic cell subsets. *Immunity* (2014) 40(5):642-56. doi: 10.1016/j.immuni.2014.04.016

57. Spranger S, Dai D, Horton B, Gajewski TF. Tumor-residing batf3 dendritic cells are required for effector T cell trafficking and adoptive T cell therapy. *Cancer Cell* (2017) 31(5):711–23.e4. doi: 10.1016/j.ccell.2017.04.003

58. Ruffell B, Chang-Strachan D, Chan V, Rosenbusch A, Ho CM, Pryer N, et al. Macrophage IL-10 blocks CD8+ T cell-dependent responses to chemotherapy by suppressing IL-12 expression in intratumoral dendritic cells. *Cancer Cell* (2014) 26 (5):623–37. doi: 10.1016/j.ccell.2014.09.006

59. Qiu Q, Lin Y, Ma Y, Li X, Liang J, Chen Z, et al. Exploring the emerging role of the gut microbiota and tumor microenvironment in cancer immunotherapy. *Front Immunol* (2020) 11:612202. doi: 10.3389/fimmu.2020.612202

60. Abel AM, Yang C, Thakar MS, Malarkannan S. Natural killer cells: development, maturation, and clinical utilization. *Front Immunol* (2018) 9:1869. doi: 10.3389/fimmu.2018.01869

61. Angell H, Galon J. From the immune contexture to the Immunoscore: the role of prognostic and predictive immune markers in cancer. *Curr Opin Immunol* (2013) 25 (2):261–7. doi: 10.1016/j.coi.2013.03.004

62. Binnewies M, Roberts EW, Kersten K, Chan V, Fearon DF, Merad M, et al. Understanding the tumor immune microenvironment (TIME) for effective therapy. *Nat Med* (2018) 24(5):541–50. doi: 10.1038/s41591-018-0014-x

63. Duan Q, Zhang H, Zheng J, Zhang L. Turning Cold into Hot: Firing up the Tumor Microenvironment. *Trends Cancer* (2020) 6(7):605–18. doi: 10.1016/j.trecan.2020.02.022

64. Mlecnik B, Tosolini M, Kirilovsky A, Berger A, Bindea G, Meatchi T, et al. Histopathologic-based prognostic factors of colorectal cancers are associated with the state of the local immune reaction. *J Clin Oncol Off J Am Soc Clin Oncol* (2011) 29 (6):610–8. doi: 10.1200/jco.2010.30.5425

65. Li Z, Wang H, Dong R, Man J, Sun L, Qian X, et al. Single-cell RNA-seq reveals characteristics of Malignant cells and immune microenvironment in subcutaneous panniculitis-like T-cell lymphoma. *Front Oncol* (2021) 11:611580. doi: 10.3389/ fonc.2021.611580

66. Giraud J, Chalopin D, Blanc JF, Saleh M. Hepatocellular carcinoma immune landscape and the potential of immunotherapies. *Front Immunol* (2021) 12:655697. doi: 10.3389/fimmu.2021.655697

67. Papalexi E, Satija R. Single-cell RNA sequencing to explore immune cell heterogeneity. *Nat Rev Immunol* (2018) 18(1):35–45. doi: 10.1038/nri.2017.76

68. Zheng L, Qin S, Si W, Wang A, Xing B, Gao R, et al. Pan-cancer single-cell landscape of tumor-infiltrating T cells. *Sci (New York NY)* (2021) 374(6574):abe6474. doi: 10.1126/science.abe6474

69. Berry S, Giraldo NA, Green BF, Cottrell TR, Stein JE, Engle EL, et al. Analysis of multispectral imaging with the AstroPath platform informs efficacy of PD-1 blockade. *Science* (2021) 372(6547):eaba2609. doi: 10.1126/science.aba2609

70. Desbois M, Wang Y. Cancer-associated fibroblasts: Key players in shaping the tumor immune microenvironment. *Immunol Rev* (2021) 302(1):241–58. doi: 10.1111/ imr.12982

71. Galon J, Bruni D. Approaches to treat immune hot, altered and cold tumours with combination immunotherapies. *Nat Rev Drug Discovery* (2019) 18(3):197–218. doi: 10.1038/s41573-018-0007-y

72. Salerno EP, Bedognetti D, Mauldin IS, Deacon DH, Shea SM, Pinczewski J, et al. Human melanomas and ovarian cancers overexpressing mechanical barrier molecule genes lack immune signatures and have increased patient mortality risk. Onco Immunol (2016) 5(12):e1240857. doi: 10.1080/2162402x.2016.1240857

73. Garris CS, Arlauckas SP, Kohler RH, Trefny MP, Garren S, Piot C, et al. Successful anti-PD-1 cancer immunotherapy requires T cell-dendritic cell crosstalk involving the cytokines IFN- γ and IL-12. *Immunity* (2022) 55(9):1749. doi: 10.1016/j.immuni.2022.07.021

74. Pietrobon V, Marincola FM. Hypoxia and the phenomenon of immune exclusion. J Trans Med (2021) 19(1):9. doi: 10.1186/s12967-020-02667-4

75. Monteran L, Erez N. The dark side of fibroblasts: cancer-associated fibroblasts as mediators of immunosuppression in the tumor microenvironment. *Front Immunol* (2019) 10:1835. doi: 10.3389/fimmu.2019.01835

76. Borriello L, Nakata R, Sheard MA, Fernandez GE, Sposto R, Malvar J, et al. Cancer-associated fibroblasts share characteristics and protumorigenic activity with mesenchymal stromal cells. *Cancer Res* (2017) 77(18):5142–57. doi: 10.1158/0008-5472.Can-16-2586

77. Michaels AD, Newhook TE, Adair SJ, Morioka S, Goudreau BJ, Nagdas S, et al. CD47 blockade as an adjuvant immunotherapy for resectable pancreatic cancer. *Clin Cancer Res an Off J Am Assoc Cancer Res* (2018) 24(6):1415–25. doi: 10.1158/1078-0432.Ccr-17-2283

78. Samanta D, Park Y, Ni X, Li H, Zahnow CA, Gabrielson E, et al. Chemotherapy induces enrichment of CD47(+)/CD73(+)/PDL1(+) immune evasive triple-negative breast cancer cells. *Proc Natl Acad Sci United States America* (2018) 115(6):E1239–e48. doi: 10.1073/pnas.1718197115

79. Mlecnik B, Bindea G, Angell HK, Sasso MS, Obenauf AC, Fredriksen T, et al. Functional network pipeline reveals genetic determinants associated with in situ lymphocyte proliferation and survival of cancer patients. *Sci Trans Med* (2014) 6 (228):228ra37. doi: 10.1126/scitranslmed.3007240

80. Mathios D, Ruzevick J, Jackson CM, Xu H, Shah SR, Taube JM, et al. PD-1, PD-L1, PD-L2 expression in the chordoma microenvironment. *J neuro-oncology* (2015) 121 (2):251–9. doi: 10.1007/s11060-014-1637-5

81. Feng Y, Shen J, Gao Y, Liao Y, Cote G, Choy E, et al. Expression of programmed cell death ligand 1 (PD-L1) and prevalence of tumor-infiltrating lymphocytes (TILs) in chordoma. *Oncotarget* (2015) 6(13):11139–49. doi: 10.18632/oncotarget.3576

82. Zou MX, Peng AB, Lv GH, Wang XB, Li J, She XL, et al. Expression of programmed death-1 ligand (PD-L1) in tumor-infiltrating lymphocytes is associated with favorable spinal chordoma prognosis. *Am J Trans Res* (2016) 8(7):3274–87. doi: 10.13140/RG.2.1.1007.0649

83. Inagaki Y, Hookway E, Williams KA, Hassan AB, Oppermann U, Tanaka Y, et al. Dendritic and mast cell involvement in the inflammatory response to primary Malignant bone tumours. *Clin sarcoma Res* (2016) 6:13. doi: 10.1186/s13569-016-0053-3

84. Zou MX, Guo KM, Lv GH, Huang W, Li J, Wang XB, et al. Clinicopathologic implications of CD8(+)/Foxp3(+) ratio and miR-574-3p/PD-L1 axis in spinal chordoma patients. *Cancer immunology immunotherapy CII* (2018) 67(2):209–24. doi: 10.1007/s00262-017-2080-1

85. Zou MX, Lv GH, Wang XB, Huang W, Li J, Jiang Y, et al. Clinical impact of the immune microenvironment in spinal chordoma: immunoscore as an independent favorable prognostic factor. *Neurosurgery* (2019) 84(6):E318–e33. doi: 10.1093/neuros/ nyy274

86. Zou MX, Pan Y, Huang W, Zhang TL, Escobar D, Wang XB, et al. A four-factor immune risk score signature predicts the clinical outcome of patients with spinal chordoma. *Clin Trans Med* (2020) 10(1):224–37. doi: 10.1002/ctm2.4

87. Dancsok AR, Gao D, Lee AF, Steigen SE, Blay JY, Thomas DM, et al. Tumorassociated macrophages and macrophage-related immune checkpoint expression in sarcomas. *Oncoimmunology* (2020) 9(1):1747340. doi: 10.1080/2162402x.2020.1747340

88. Zhou J, Jiang Y, Zhang H, Chen L, Luo P, Li L, et al. Clinicopathological implications of TIM3(+) tumor-infiltrating lymphocytes and the miR-455-5p/Galectin-9 axis in skull base chordoma patients. *Cancer immunology immunotherapy CII* (2019) 68(7):1157–69. doi: 10.1007/s00262-019-02349-1

 He G, Liu X, Pan X, Ma Y, Liu X. Cytotoxic T lymphocyte antigen-4 (CTLA-4) expression in chordoma and tumor-infiltrating lymphocytes (TILs) predicts prognosis of spinal chordoma. Clin Trans Oncol Off Publ Fed Spanish Oncol Societies Natl Cancer Institute Mexico (2020) 22(12):2324–32. doi: 10.1007/s12094-020-02387-7 90. Lopez DC, Robbins YL, Kowalczyk JT, Lassoued W, Gulley JL, Miettinen MM, et al. Multi-spectral immunofluorescence evaluation of the myeloid, T cell, and natural killer cell tumor immune microenvironment in chordoma may guide immunotherapeutic strategies. *Front Oncol* (2022) 12:1012058. doi: 10.3389/fonc.2022.1012058

91. Duan W, Zhang B, Li X, Chen W, Jia S, Xin Z, et al. Single-cell transcriptome profiling reveals intra-tumoral heterogeneity in human chordomas. *Cancer immunol immunother CII* (2022) 71(9):2185–95. doi: 10.1007/s00262-022-03152-1

92. Memon H, Patel BM. Immune checkpoint inhibitors in non-small cell lung cancer: A bird's eye view. Life Sci (2019) 233:116713. doi: 10.1016/j.lfs.2019.116713

93. Sharpe AH, Pauken KE. The diverse functions of the PD1 inhibitory pathway. Nat Rev Immunol (2018) 18(3):153-67. doi: 10.1038/nri.2017.108

94. Roemer MG, Advani RH, Ligon AH, Natkunam Y, Redd RA, Homer H, et al. PD-L1 and PD-L2 genetic alterations define classical hodgkin lymphoma and predict outcome. *J Clin Oncol Off J Am Soc Clin Oncol* (2016) 34(23):2690–7. doi: 10.1200/ jco.2016.66.4482

95. Burr ML, Sparbier CE, Chan YC, Williamson JC, Woods K, Beavis PA, et al. CMTM6 maintains the expression of PD-L1 and regulates anti-tumour immunity. *Nature* (2017) 549(7670):101–5. doi: 10.1038/nature23643

96. Thanindratarn P, Dean DC, Nelson SD, Hornicek FJ, Duan Z. Advances in immune checkpoint inhibitors for bone sarcoma therapy. J Bone Oncol (2019) 15:100221. doi: 10.1016/j.jbo.2019.100221

97. D'Angelo SP, Richards AL, Conley AP, Woo HJ, Dickson MA, Gounder M, et al. Pilot study of bempegaldesleukin in combination with nivolumab in patients with metastatic sarcoma. *Nat Commun* (2022) 13(1):3477. doi: 10.1038/s41467-022-30874-8

98. Blay JY, Chevret S, Le Cesne A, Brahmi M, Penel N, Cousin S, et al. Pembrolizumab in patients with rare and ultra-rare sarcomas (AcSé Pembrolizumab): analysis of a subgroup from a non-randomised, open-label, phase 2, basket trial. *Lancet Oncol* (2023) 24(8):892–902. doi: 10.1016/s1470-2045(23)00282-6

99. Zhang Q, Fei L, Han R, Huang R, Wang Y, Chen H, et al. Single-cell transcriptome reveals cellular hierarchies and guides p-EMT-targeted trial in skull base chordoma. *Cell Discovery* (2022) 8(1):94. doi: 10.1038/s41421-022-00459-2

100. Bishop AJ, Amini B, Lin H, Raza SM, Patel S, Grosshans DR, et al. Immune checkpoint inhibitors have clinical activity in patients with recurrent chordoma. *J immunotherapy* (2022) 45(8):374–8. doi: 10.1097/cji.00000000000431

101. Darvin P, Toor SM, Sasidharan Nair V, Elkord E. Immune checkpoint inhibitors: recent progress and potential biomarkers. *Exp Mol Med* (2018) 50(12):1–11. doi: 10.1038/s12276-018-0191-1

102. Page DB, Postow MA, Callahan MK, Allison JP, Wolchok JD. Immune modulation in cancer with antibodies. *Annu Rev Med* (2014) 65:185–202. doi: 10.1146/annurev-med-092012-112807

103. Dolladille C, Ederhy S, Sassier M, Cautela J, Thuny F, Cohen AA, et al. Immune checkpoint inhibitor rechallenge after immune-related adverse events in patients with cancer. *JAMA Oncol* (2020) 6(6):865–71. doi: 10.1001/jamaoncol.2020.0726

104. Laskowski T, Rezvani K. Adoptive cell therapy: Living drugs against cancer. J Exp Med (2020) 217(12):e20200377. doi: 10.1084/jem.20200377

105. June CH, O'Connor RS, Kawalekar OU, Ghassemi S, Milone MC. CAR T cell immunotherapy for human cancer. *Science* (2018) 359(6382):1361–5. doi: 10.1126/ science.aar6711

106. Chmielewski M, Abken H. TRUCKs: the fourth generation of CARs. *Expert* Opin Biol Ther (2015) 15(8):1145–54. doi: 10.1517/14712598.2015.1046430

107. Fournier C, Martin F, Zitvogel L, Kroemer G, Galluzzi L, Apetoh L. Trial Watch: Adoptively transferred cells for anticancer immunotherapy. *Oncoimmunology* (2017) 6(11):e1363139. doi: 10.1080/2162402x.2017.1363139

108. Neelapu SS, Locke FL, Bartlett NL, Lekakis LJ, Miklos DB, Jacobson CA, et al. Axicabtagene ciloleucel CAR T-cell therapy in refractory large B-cell lymphoma. *New Engl J Med* (2017) 377(26):2531–44. doi: 10.1056/NEJMoa1707447

109. Ma S, Li X, Wang X, Cheng L, Li Z, Zhang C, et al. Current progress in CAR-T cell therapy for solid tumors. *Int J Biol Sci* (2019) 15(12):2548–60. doi: 10.7150/ ijbs.34213

110. Kershaw MH, Wang G, Westwood JA, Pachynski RK, Tiffany HL, Marincola FM, et al. Redirecting migration of T cells to chemokine secreted from tumors by genetic modification with CXCR2. *Hum Gene Ther* (2002) 13(16):1971–80. doi: 10.1089/10430340260355374

111. Craddock JA, Lu A, Bear A, Pule M, Brenner MK, Rooney CM, et al. Enhanced tumor trafficking of GD2 chimeric antigen receptor T cells by expression of the chemokine receptor CCR2b. *J immunotherapy* (2010) 33(8):780–8. doi: 10.1097/CJI.0b013e3181ee6675

112. Kloss CC, Lee J, Zhang A, Chen F, Melenhorst JJ, Lacey SFK, et al. Dominantnegative TGF- β Receptor enhances PSMA-targeted human CAR T cell proliferation and augments prostate cancer eradication. *Mol Ther J Am Soc Gene Ther* (2018) 26 (7):1855–66. doi: 10.1016/j.ymthe.2018.05.003

113. Yang M, Tang X, Zhang Z, Gu L, Wei H, Zhao S, et al. Tandem CAR-T cells targeting CD70 and B7-H3 exhibit potent preclinical activity against multiple solid tumors. *Theranostics* (2020) 10(17):7622–34. doi: 10.7150/thno.43991

114. Long C, Li G, Zhang C, Jiang T, Li Y, Duan X, et al. B7-H3 as a target for CAR-T cell therapy in skull base chordoma. *Front Oncol* (2021) 11:659662. doi: 10.3389/ fonc.2021.659662

115. Sahin U, Türeci Ö. Personalized vaccines for cancer immunotherapy *Science* (2018) 359(6382):1355–60 doi: 10.1126/science.aar7112

116. Heery CR, Singh BH, Rauckhorst M, Marté JL, Donahue RN, Grenga I, et al. Phase I trial of a yeast-based therapeutic cancer vaccine (GI-6301) targeting the transcription factor brachyury. *Cancer Immunol Res* (2015) 3(11):1248–56. doi: 10.1158/2326-6066.Cir-15-0119

117. Heery CR, Palena C, McMahon S, Donahue RN, Lepone LM, Grenga I, et al. Phase I study of a poxviral TRICOM-based vaccine directed against the transcription factor brachyury. *Clin Cancer Res an Off J Am Assoc Cancer Res* (2017) 23(22):6833–45. doi: 10.1158/1078-0432.Ccr-17-1087

118. DeMaria PJ, Lee-Wisdom K, Donahue RN, Madan RA, Karzai F, Schwab A, et al. Phase 1 open-label trial of intravenous administration of MVA-BN-brachyury-TRICOM vaccine in patients with advanced cancer. *J immunother Cancer* (2021) 9(9): e003238. doi: 10.1136/jitc-2021-003238

119. Collins JM, Donahue RN, Tsai YT, Manu M, Palena C, Gatti-Mays ME, et al. Phase I trial of a modified vaccinia ankara priming vaccine followed by a fowlpox virus boosting vaccine modified to express brachyury and costimulatory molecules in advanced solid tumors. *oncologist* (2020) 25(7):560–e1006. doi: 10.1634/ theoncologist.2019-0932

120. Barrett RL, Puré E. Cancer-associated fibroblasts and their influence on tumor immunity and immunotherapy. *eLife* (2020) 9:e57243. doi: 10.7554/eLife.57243

121. de Gramont A, Faivre S, Raymond E. Novel TGF- β inhibitors ready for prime time in onco-immunology. Oncoimmunology (2017) 6(1):e1257453. doi: 10.1080/2162402x.2016.1257453

122. Nishina T, Takahashi S, Iwasawa R, Noguchi H, Aoki M, Doi T. Safety, pharmacokinetic, and pharmacodynamics of erdafitinib, a pan-fibroblast growth factor receptor (FGFR) tyrosine kinase inhibitor, in patients with advanced or refractory solid tumors. *Investigational New Drugs* (2018) 36(3):424–34. doi: 10.1007/s10637-017-0514-4

123. Lin HM, Lee BY, Castillo L, Spielman C, Grogan J, Yeung NK, et al. Effect of FAK inhibitor VS-6063 (defactinib) on docetaxel efficacy in prostate cancer. *Prostate* (2018) 78(4):308–17. doi: 10.1002/pros.23476

124. Thompson CB, Shepard HM, O'Connor PM, Kadhim S, Jiang P, Osgood RJ, et al. Enzymatic depletion of tumor hyaluronan induces antitumor responses in preclinical animal models. *Mol Cancer Ther* (2010) 9(11):3052–64. doi: 10.1158/1535-7163.Mct-10-0470

125. Chauhan VP, Martin JD, Liu H, Lacorre DA, Jain SR, Kozin SV, et al. Angiotensin inhibition enhances drug delivery and potentiates chemotherapy by decompressing tumour blood vessels. *Nat Commun* (2013) 4:2516. doi: 10.1038/ncomms3516

126. Halvorsen SC, Benita Y, Hopton M, Hoppe B, Gunnlaugsson HO, Korgaonkar P, et al. Transcriptional profiling supports the notochordal origin of chordoma and its dependence on a TGF β 1-TBXT network. *Am J Pathol* (2023) 193(5):532–47. doi: 10.1016/j.ajpath.2023.01.014