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# HDAC6 inhibition: a significant potential regulator and therapeutic option to translate into clinical practice in renal transplantation

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Histone deacetylase 6 (HDAC6), an almost exclusively cytoplasmic enzyme, plays an essential role in many biological processes and exerts its deacetylationdependent/independent effects on a variety of target molecules, which has contributed to the flourishing growth of relatively isoform-specific enzyme inhibitors. Renal transplantation (RT) is one of the alternatively preferred treatments and the most cost-effective treatment approaches for the great majority of patients with end-stage renal disease (ESRD). HDAC6 expression and activity have recently been shown to be increased in kidney disease in a number of studies. To date, a substantial amount of validated studies has identified HDAC6 as a pivotal modulator of innate and adaptive immunity, and HDAC6 inhibitors (HDAC6i) are being developed and investigated for use in arrays of immune-related diseases, making HDAC6i a promising therapeutic candidate for the management of a variety of renal diseases. Based on accumulating evidence, HDAC6i markedly open up new avenues for therapeutic intervention to protect against oxidative stress-induced damage, tip the balance in favor of the generation of tolerance-related immune cells, and attenuate fibrosis by inhibiting multiple activations of cell profibrotic signaling pathways. Taken together, we have a point of view that targeting HDAC6 may be a novel approach for the therapeutic strategy of RT-related complications, including consequences of ischemia-reperfusion injury, induction of immune tolerance in transplantation, equilibrium of rejection, and improvement of chronic renal graft interstitial fibrosis after transplantation in patients. Herein, we will elaborate on the unique function of HDAC6, which focuses on therapeutical mechanism of action related to immunological events with a general account of the tantalizing potential to the clinic.

#### KEYWORDS

epigenetics, HDAC6, selective HDAC6 inhibitors, molecular regulation, immunoregulation, renal transplantation, ameliorate outcomes



### 1 Introduction

Progressing to end-stage renal disease (ESRD), many chronic kidney disease (CKD) patients require renal replacement therapies (1). The preferred therapy and the most cost-effective treatment approach for the majority of patients with ESRD are actually renal transplantation (RT) (2). Actually, despite ongoing advances in RT, there are still emerging challenges related to immunosuppressive

therapy in this field, including ongoing consequences of ischemia reperfusion injury (IRI) (3), ideal goal of immune tolerance (4), equilibrium of rejection (5), and improvement of chronic interstitial fibrosis (6) in patients after transplantation. All these posttransplant conditions are related to the prominent function of the immune system and are responsible for the fate of the renal graft (7). Therefore, optimal management of RT is imperative for improvement of global public health problems. Improving long-term outcomes is now a major focus of RT, with much effort being conducted (8). As advances have been made in both the surgical techniques and immunosuppression (IS) aspects of transplantation, the long-term complications related with RT have been of increasing importance, especially as transplant recipients live longer (9). The induction immunosuppressive regimens currently known to delay the progression of these complications are not effective enough, nor are the triple maintenance regimens (9). Therefore, therapeutic candidates needed to mitigate the development of these clinical dilemmas are under investigation, and histone deacetylase inhibitors (HDACis) may be identified as such important therapeutic applications (10–12).

In eukaryotic cells, histone acetyltransferases (HATs) and histone deacetylases (HDACs) are key enzymes that play an essential role in modifying chromatin structure and thereby modulate gene expression levels, although their possible potential roles in determining cell fate has yet to be fully elucidated (13). They form part of multiprotein complexes that include other proteins known to be involved in activating/repressing transcription, along with antagonistic proteases that play a regulatory role in balancing HAT and HDAC in nucleosomes (14). HATs neutralize the positive charge of histone tails by transferring acetyl groups from acetyl-CoA to the  $\epsilon$ -NH2 group of lysine residue side-chains, thereby making chromatin looser and promoting gene expression. Nevertheless, deacetylation is associated with gene silencing; to facilitate histone

deacetylation and balance the acetylating activities of HAT in chromatin remodeling, HDACs remove acetyl groups from hyperacetylated histones, thereby balancing HATs where the positive charge on the surface of the recombinant lysine increases the binding affinity to the negatively charged surface of DNA (14, 15).

As a large family of transcriptional repressors, HDACs play an extremely important role in the epigenetic regulation of a wide range of cellular processes, chromatin structure, gene expression, and physiological functions in various renal diseases (16). Based on homology to yeast HDACs, biological function, and DNA sequence similarity, the family of HDAC isoforms consists of at least 18 characterized members to date, which can be grouped into four classes (Figure 1) (17): class I (HDAC1-HDAC3, and HDAC8), class II (HDAC4-HDAC7 and HDAC9-HDAC10), class III (SIRT1-SIRT7), and class IV (HDAC11). Class I, II, and IV HDACs are all  $Zn_2$  +-dependent deacetylases, while class III HDACs are known as nicotinamide adenine dinucleotide (NAD+)-dependent sirtuins (SIRT1-7).

Within the chemical classes of HDAC-inhibiting agents, some HDAC-inhibiting agents act against most HDAC isoforms non-specifically, while others act selectively against a specific HDAC class or isoform (17). Pan HDACis have demonstrated prominent efficacy in treating allergy (18), cancer (19), and inflammatory and autoimmune diseases (20). HDACis have been shown to initiate a remarkable



#### FIGURE 1

Schematic depiction of the different isoforms of HDAC. N, N-terminus; C, C-terminus; Blue lines depict the length of the protein; Yellow bars depict catalytic domains; Orange bars depict nuclear localization sequences (NLS); Brown bars depict nuclear export sequences (NES1, NES2); Green bar depicts Ser-Glu containing a tetradecapeptide repeating domain (SE14); Purple bar depicts a zinc-finger ubiquitin-binding domain (ZnF-UBD).

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immunosuppressive effect and prolong graft survival in studies evaluating the effect of HDACis on renal injury after RT. Recently, the administered TsA alone doubled the fully the major histocompatability complex (MHC)-mismatched murine cardiac allograft survival carrier, while a subtherapeutic course of rapamycin (RPM) merely extended allograft survival by an additional 2–3 days. Of note, a short course of HDACi therapy was shown to be synergistic with low-dose RPM therapy, promoting permanent cardiac allograft survival and the development of Treg-dependent donor-specific allograft tolerance, as also was the case with islet allografts (21). Regarding mechanism, HDACi therapy increased histone 3 acetylation within the graft, confirming the expected effect of HDACi on protein expression within cardiac allografts, and most of the clear results are associated with increased Foxp3Treg cells within the graft (22).

Nevertheless, their profiles of non-selective HDACi possess a main disadvantage, as it is accompanied by some undesirable effects, including anorexia, nausea, vomiting, diarrhea, thrombocytopenia, neutropenia, anemia, and alteration of serum biochemistry profiles, which severely limit their clinical utility in chronic disease indications (21). In this context, increasing efforts are being made to identify isoform-selective HDAC inhibitors as epigenetic therapeutics (22). A wealth of evidence supported that HDAC subtype-selective inhibitors with fewer adverse effects compared with Pan HDAC inhibitors could be applicable to treat chronic conditions; the evolving trend is the identification of HDAC isoenzyme-selective inhibitors with immunomodulatory function, as well as an elevated safety profile (22). Based on a comprehensive analysis of previous studies, it is more advisable that the HDAC subtype may represent a novel therapeutic target for RT, with many isoform-selective HDAC inhibitors showing positive efficiency (10). Herein, we pay special attention to the role of HDAC6 in the progression of kidney diseases and the tantalizing promise of its inhibitor (23), which could facilitate the advancement of HDAC6-targeted therapeutic methods for complications of RT.

# 2 Introduction of histone deacetylase 6 and selective histone deacetylase 6 inhibitors

# 2.1 Characterization of histone deacetylase 6

Among epigenetic regulation processes of HDACs, HDAC6, a cytoplasmic class IIb HDAC, stands out for its nearly all deacetylation in the cytoplasm and for its catalytic activity involving both dependent and independent mechanisms, which paved an enormously therapeutic potential avenue for identifying its substrates, together with developing highly selective enzyme inhibitors (24). HDAC6 displays important roles in many biological processes and potentially regulates several cellular functions, including cycle regulation, DNA repair, actin-dependent cell motility, and tumor metastasis, respectively (25). Importantly, HDAC6 not only displays a variety of non-histone substrates

such as alpha-tubulin (26), heat shock protein 90 (HSP90) (27) and cortactin (28), Ku70 (29), RIG-I (30), p53 (31), STAT3 (32), NF- $\kappa$ B (33), FoxO1 (34), and  $\beta$ -catenin (35), as well as regulate protein degradation, but also plays its mutual effect with several proteins that modulate its deacetylase activity through interactions between proteins.

Uniquely, the HDAC6 isoform displays two functional deacetylase domains in tandem, designated DAC1 and DAC2, respectively, DD1 and DD2, along with a C-terminus hydrolase-like zinc finger domain with conserved regions rich in cysteine and histidine for ubiquitin binding (ZnF-UBP), which is specifically binded to mono- and polyubiquitin chains (Figure 1) (36). DD2 is in charge of its deacetylation activity on histones and  $\alpha$ -tubulin, according to experimental data using purified HDAC6 (37). Additionally, depending on their acetylation status, HDAC6 shuttle between the cytoplasm and the nucleus, allowing them to interact with miscellaneous cytoplasmic proteins and polyubiquitins, thereby having an influence on cell migration and proliferation, and, together with misfolded proteins, induced cell stress, as well as the processing or degradation of protein aggregates (25).

Meanwhile, the elevated expression and activity of HDAC6 have confirmation of participating in various kidney diseases including acute kidney injury (AKI) (38), polycystic kidney disease (39), lupus nephritis (40), renal cancer (41), and hypertensive nephropathy (42). With enhanced efficacy and less toxicity, selective HDAC6 inhibitors (HDAC6i) are growingly in demand as powerful and promising treatments for multiple pathological manifestations in the kidney including glomerulosclerosis and nephritis, tubulointerstitial inflammation, and renal interstitial fibrosis, which have shown beneficial effects for previous examination using experimental animal models (23).

Collectively, as the specific cytoplasmic localization and the wide range of protein substrates, together with aberrant HDAC6 activity in increased fibrosis (42) and inflammation (43) of kidney, HDAC6 inhibitors have indeed effectively mitigated the progression of immune-related complications in the kidney, indicating that targeting HDAC6 may offer a novel and attractive therapeutic method for complications associated with RT. In this review, we endeavor to have a summary of the role of HDAC6 in biological processes relevant to RT diseases and highlight the effects of HDAC6i and the mechanisms underlying their protection, including IRI, tolerance, rejection, and fibrosis, which would improve recipients' clinical outcomes and hold great promise as therapeutic agents.

# 2.2 Selective histone deacetylase 6 inhibitor

The majority of hydroxamate-based inhibitors have been shown to be highly effective in inhibiting HDAC6, which are able to form a bidentate complex with zinc ions. The flourishing growth and refinement of crystallization technology have enabled the design of novel HDAC6i with significant therapeutic value. From a

structural point of view, HDAC6i are typically composed of three main components: the zinc-binding group (ZBG), which is located at the bottom of the HDAC6 catalytic cavity coordinated with Zn+, by means of an unusual monodentate coordination geometry, or a canonical bidentate Zn<sub>2</sub>+ coordination geometry (44). With the selectivitydetermining region, a strong hydrophobic interaction is established by the capping group, "L1 loop pocket," which occupies the large surface area of the HDAC6 molecule. The linker group binds to HDAC6 via hydrogen bonds and bridges between the ZBG and the cap group (44). Moreover, earlier studies have validated that HDAC6 catalytic pocket flexibility allowed protein to undergo structural changes with accommodation of the cap part of the ligands under investigation (45). Currently, the most widely investigated inhibitors of selective small molecule HDAC6 mainly include Rocilinostat (ACY-1215), Citarinostat (ACY-241), Tubacin, CKD-506, Tubastatin A(TA), BML-281, and LTB2 (46). The most successful anti-tumor HDAC6 inhibitors to date are ACY-1215 (47) and ACY-241 (48), which are oral selective HDAC6i under clinical investigation for the treatment of refractory multiple myeloma (MM). Subsequently, a series of carbazole hydroxamic acids with alkyl and alkylaryl linker groups were synthesized by Butler et al. in accordance with a wider and shallower channel of HDAC6. Meanwhile, with an IC50 of 15 nM against HDAC6, TA was found to have anti-HDAC6 function (49). A potent and selective inhibitor of the HDAC6 enzyme is hydroxamic acid (M808), with improved selectivity and solubility over the structural analog ACY-1215 (50). Table 1 summarizes some representative HDAC6i, together with their inhibitory functions, currently in clinical trials (disease), reported related diseases, the pathomechanisms of complication therapy of RTrelated complications.

# 3 Histone deacetylase 6 modulates immune response

The immune system falls into two general types, which are innate and adaptive (66). Neutrophils, macrophages, dendritic cells

TABLE 1 A summary of representative HDAC6 is 1–21 along with their inhibitory activities, original targets (disease), mechanisms of therapy for RT related complications and respective limitations.

No./ Name	Structure	IC50	Currenty in Clinical Trials (Disease)	Reported related dis- eases	Mechanism of therapy for RT related complications
1 Tubacin	off of of the	4nM	HDAC6 (Anticancer) (51)	1. Polycystic kidney disease (39).	1. Amelioration of learning and memory deficits.
2 TA	C N HN-OH	15 nM	HDAC6 (NDs) (49)	1. Rhabdomyolysis- induced nephropathy (52) 2. Ischemia- and Cisplatin-Induced AKI (53)	2. Inhibit the mitosis of Teffs.
3 MPT0G2	HO-H	0.291 μM	HDAC6 (Alzheimer's disease, AD) (54)	Not reported so far	Not reported so far.
4 ACY-7	H N N O H N O H	1.7nM	HDAC6 (Multiple myeloma, MM) (55)	1. Polycystic kidney disease (56) 2. SLE (57)	<ol> <li>Attenuate IL-1β production.</li> <li>Increased Tregs.</li> <li>Decrease pro-B cell differentiation, Bcl-2:Bax ratio.</li> <li>Suppression of T cells.</li> </ol>
5 ACY- 1215, Ricolino stat	N N N N NHOH	4.7nM	HDAC6 (MM) (47)	<ol> <li>UUO-induced</li> <li>fibrotic kidneys</li> <li>(58)</li> <li>Polycystic kidney</li> <li>disease (59)</li> </ol>	<ol> <li>Suppress Th2 cytokines.</li> <li>Reduce differentiation of Th17 cells.</li> <li>Modulate CD8 T-cell activation and functions.</li> <li>Decrease M1 and cytokines.</li> </ol>

(Continued)

#### TABLE 1 Continued

No./ Name	Structure	IC50	Currenty in Clinical Trials (Disease)	Reported related dis- eases	Mechanism of therapy for RT related complications
6 CKD-5	Not discover	Not discovered.	HDAC6 (Rheumatoid Arthritis, RA) (60)	1. Systemic lupus erythematosus (SLE) (61)	<ol> <li>Decrease levels of IFN-γ, IL- 1β, IL-4, IL-6, IP-10, MCP-1, and CCL4 in the kidney extracts of mice with lupus nephritis</li> <li>Repress innate immune systems such as production of TNF-α and type1 interferons</li> <li>RegulatE T cell- and macrophage-mediated peripheral immune responses.</li> <li>Downregulate the expression of IFN-γ and IL-17A.</li> <li>Downregulate the levels of pro-inflammatory cytokines</li> </ol>
7 Nexturastat A (NexA	H N OH	5.02nM	HDAC6 (MM) (62)	Not reported so far.	<ol> <li>Suppresse the mRNA levels of pro-inflammatory cytokines IL-1β and IL-6</li> <li>Decrease the anti-inflammatory phenotype of macrophages and down-regulation of immunosuppressive proteins</li> </ol>
8 Citarinostat (ACY-241)		2.6 nM	HDAC6 (MM) (63)	Not reported so far.	Not reported so far.
9 Benzamide (HPOB)		56 nM	HDAC6 (Antitumor) (64)	Not reported so far.	3. Not reported so far.
10 BML- 281 CAY10603	Not the second s	0.002 nM	HDAC6 1. (non-small cell lung cancer, NSCLC)) (65)	Not reported so far.	Not reported so far.

(DCs), natural killer cells, gamma and gamma delta cells and other innate lymphoid cells, and complement system members, and mediators of inflammation, are among the different cell types involved in the innate immune system (67), which acts on most immunological events following RT with an important role and recognize immune antigens that are fast and lasting through patternrecognition receptorsin a very short time frame (68). Through migration and maturation in the kidney, DCs are involved in inducing and regulating innate and adaptive immunity (69). Generally considered to be antigen-presenting cells (APCs) and a core participant for all immune responses, DCs are not only particularly effective immunogens in inflammation but also critical for inducing and maintaining self-tolerance in the context of stable immune status (69). Furthermore, selective HDAC6i is also necessary for the recovery of innate immune cells in the bone marrow, followed by reducing stress and immune atrophy and apoptosis (70). Macrophages, as the key effector modulators of the innate immune system, play a predominant role in inflammatory initiation, resolution, tissue repair, and regeneration (71). Recent work has presented that monocytes and macrophages directly recognize allorecognition (72), which is independent of lymphoid cells and induces the maturation of APC; thus, the initiation and maintenance of an adaptive allogeneic immune response is possible.

Rapid response of innate immune cells develop an immunological memory, whereas a delayed response is seen in adaptive immune cells and can be days away from full development (73). One of the most vigorous elements of the immune system is the adaptive immunity system, where the main immunoreactions are generated by T and B cells, whose products are of crucial importance for immune protection against pathogens and inflammation (67). In the process of T-cell receptor (TCR) recognition of Th cells, CD4 is the receptor and participator of the Th cell TCR for antigen recognition. CD8, a leukocyte differentiation antigen, is used to assist the TCR to recognize antigens and take part in the signal transduction of T-cell activation. Based on the subsets, B cells play a diverse role toward organ allografts, either enhancing or suppressing immunity (67). What is interesting is that the communication between the innate and adaptive immune responses during RT is bidirectional, as it is now clear to see that activating the adaptive immune response can be the cause of kidney damage induced by cellular and molecular components of innate immunity (74).

Relying on the cytoskeleton to undergo constant reorganization, including acetylation or deacetylation, HDAC6 influences a wide range of cellular functions and cytoplasm shape, such as cell signaling, activation, survival, motility, and protein degradation

(24, 25, 75), which ultimately has an impact on cell function. Various studies have since shown that HDAC6 activity regulation, which is expressed in both the nucleus and the cytoplasm, have identified a significantly critical role for the behavior in innate and adaptive immunity (76), together with modulating the inflammatory genes (24, 28). In clinical RT, activation of the innate immune response can be triggered by certain elements (77). Brain death itself may accelerate the systemic production and release of proinflammatory cytokines (monocyte chemotactic peptide-1 and interleukin-6) in donation after brain death (DBD), contributing to activating innate immune pathways, such as recruiting and activating monocytes in multiple organs, including the kidneys (78). Based on previous and current studies, HDAC6i conduct suppression of excessive inflammation by inhibiting DCs, with prevention of damage to kidney function. Through enabling macrophages to the selective recognition and elimination of allogeneic targets, HDAC6 deficiency or HDACi also produces a protective immune response. HDAC6 therefore has the critical role in regulation of innate immune responses, and HDAC6 inhibition can sustain the ability to ameliorate the inflammatory response at a later stage. A previous study reported that HDAC6 affects the activation of antigen-dependent CD4 T cells (67), and another revealed that HDAC6 plays a role in CD8 T-cell cytotoxic function (79). In addition to suppressing B- and T-cell activation, HDAC6i also attenuated T-cell differentiation into several other subtypes known to be regulated in immune-related diseases, both in vivo and in vitro (80). There were pieces of evidence showing that abnormal B-cell differentiation was associated with a less severe form of kidney disease, which could be corrected by selective HDAC6 inhibition (81).

Inhibition of HDAC6 therefore has pleiotropic effects on the immune system. In the future, for current clinical relevance, therapeutics targeting HDAC6 may have particular potential for the early intervention strategy of RT-related diseases. Beyond that, the effects of later intervention with HDAC6 inhibition, starting after the onset of RT-related disease, are more important to investigate. These are discussed in detail in the following sections.

## 4 Role of histone deacetylase 6 and potential of histone deacetylase 6 inhibitors as treatments for RT complications

# 4.1 Mechanism of histone deacetylase 6 in renal IRI

IRI, a common posttransplant event, increases the risk of delayed graft function (DGF), setting the stage for graft rejection and long-term graft loss (3). Accumulating evidence indicates that early warning signs of danger linked to IRI spell innate immune activation, together with promoting graft rejection and inhibiting tolerance occurrence (78, 82). Innate and adaptive immune cells such as neutrophils, including DCs, macrophages, and lymphocyte

subsets, have been shown to be involved in the pathological processes of IRI, resulting in the release of potent cytokines and chemokines, not least certain subpopulations involved in the repair process (82, 83). Despite decades of research into the mechanism of IRI, no clinically approved maneuvers to mitigate this process have been the mainstay of clinical practice for the modification of the negative influence of IRI associated with both short- and long-term outcomes of renal transplant recipients. As such, we focused on the critical role of HDAC6 in IRI, with the aim of identifying potential targets for preventive therapeutics (Figure 2).

Also, it is now evident that HDAC6 deacetylates non-histone proteins to take part in broader signaling and transcriptional pathways (27-35). These targets including Hsp70, Hsp90, and heat shock factor protein-1 (HSF-1), known elements and highly expressed during IRI, were acetylated during stress situation, leading to the nuclear translocation and induction of several genes (84, 85). Numerous studies have demonstrated that HDAC6 inhibition is protective in preclinical models of myocardial infarction (86) and IRI (53). Furthermore, HDAC6i have also been the investigated subject in some acute injury models and have presented a positive protective effect. For example, TA improved long-term survival in a lethal sepsis model (86) and ameliorated stroke-related brain infarction and functional deficits (87). It has already been demonstrated that inhibition of HDAC6 can attenuate kidney injury in a 5/6 nephrectomy model of CKD and in rhabdomyolysis- or cisplatin-induced models of AKI (52).

There are several mechanisms by which HDAC6 inhibition may promote a good protective effect during renal IRI. Among these, more obvious data indicated that HDAC6 is a crucial regulator of macrophages (88). There is increasing evidence that macrophages are of paramount importance in the development and progression of renal inflammation and may be mediators of inflammation through surface-expressed receptors (71). Upon activation, macrophages release proteolytic enzymes and inflammatory cytokines such as TNF- $\alpha$ , TGF- $\beta$ , IL-1 $\beta$ , and IFN- $\gamma$  (71). After IRI, monocytes infiltrate the renal interstitial space and differentiate into proinflammatory macrophages, known as M1, in response to the initial kidney injury, and subsequently switch to a form called M2, which promotes renal repair (71, 72, 89). Overexpression of HDAC6 results in a spontaneous proinflammatory macrophage response, whereas HDAC6 deficiency has been reported to suppress the immune response during inflammation by ameliorating the monocyte-derived macrophage infiltration and a heavy influx of circulating monocytes (71). Stimulation of macrophages with lipopolysaccharide results in translocation of HDAC6 and cortactin from the cytosol to the periphery, with promotion of filopodial protrusion and enhancement of microtubule acetylation around the microtubule hub, all of which are abolished by HDAC6 deficiency or HDAC6i. Previous studies have mentioned that inhibition of HDAC6 by TA and M808 gave rise to effective alleviation of expression of multiple cytokines and chemokines, as well as elevated macrophage migration and infiltration into the injured kidney in a murine rheumatoid arthritis (RA) model and a monocyte (monocytic cell line U937) migration assay (43, 52). An experimental study confirmed that TA treatment significantly hampered the production of proinflammatory cytokines like



MCP-1, TNF- $\alpha$ , and IL-6 by monocytes in rhabdomyolysis-induced AKI (52, 53). Moreover, provided further corroborating evidence, another study supported that TA remarkably enhanced the phagocytic capacity of RAW264.7 macrophages (90). In conclusion, these findings implicate that HDAC6i may represent a potent target for the treatment of macrophage-associated immune diseases with great promise.

DCs can facilitate harmful immune activations, but they are also responsible for beneficial functions in protecting kidney from IRI (83). Serving as a "bridge" between the innate immune system and adaptive immune systems, DCs are a section of the inflammatory response following IRI in the kidney (74, 91). After exposure to an antigen during IRI, DCs migrate to the damaged tissue where they expose antigens to adaptive immune cells (83). Current studies tend to suggest that HIF-1a triggers the maturation of DCs and subsequently damages renal function (92). Furthermore, DCs are thought to have important implications in the early pathophysiology of IRI development within the first 24 h after IRI (74). During the post-IRI period, intra-renal DCs can activate NKT cells and amplify the innate immune response (83). All DC subsets interact with the various effector T cells (Teff), including CD4 T cells, CD8 T cells, and Tregs, induced by IRI procedures, although the classification remains controversial, according to different effects (69). The presence of the DC population was known to be reduced to basal levels in the NextA combination arms from a previous study (93). Recently, it has been suggested that TNF-a, which is also secreted by chlamydia-infected DCs, induces HDAC6 (94). In agreement with these observations, LPS was also found to strongly upregulate the expression of HDAC6 in APCs. On the contrary, HDAC6 inhibition has an effect on the downstream effects of TNF- $\alpha$ . Furthermore, the therapeutic regimen of DCs with selective HDAC6i CKD-506 mitigated the progression of T-cell activation *via* diminishing the production and release of an amount of immunosuppressive cytokine, IL-10, on the basis of the IL-10 colitis model (95).

As described earlier, the renal protection elicited by HDAC6 blockage is associated with the restraint of inflammation and the decreased oxidative stress (96). A previous study investigated that blocking HDAC6 with TA could interfere with oxidative stress triggered in the mouse model of rhabdomyolysis-induced AKI, which effectively lessens malondialdehyde (MDA) levels, preserves the expression of superoxide dismutase (SOD), two representative biomarkers of oxidative stress in the kidney (52). The production of Reactive oxygen species (ROS) and release of cytokines are all cellular events released during reperfusion that result in the renal injury from donor and subsequent generation of DGF (3, 82). Overexpression of HDAC6 during IRI upregulates the activity of Nicotinamide Adenine Dinucleotide Phosphate (NADPH) oxidase with ascending generation of ROS (97). The generated ROS fuels the activation of the NF-KB and AP-1 mechanism pathways and then transcribed and produced proinflammatory cytokines (98). Several reports have reported that HDAC6 inhibition with TA dampens the expression of ROS with subsequent transcription of the NF-KB and AP-1 mechanism pathways, conducted in vitro and in vivo studies, including various chronic murine colitis with the IL-10 KO mouse and adoptive transfer models (99). By upregulating the expression and activity of NADPH oxidase, studies of HDAC6 overexpression in macrophages significantly triggering the generation of an inflammatory cascade and ACY-1215 decreased the inflammatory response in LPS-induced RAW264.7

macrophages (100). Another preliminary work has shown that HDAC6 participated in genotoxic stress induced by DNA damage agents (101). Taken together, HDAC6i markedly protects against oxidative stress-induced damage, which is of particular importance for immune cells exposed to high ROS concentrations (99, 102). All these findings lend strong support to the fact that HDAC6i represents a therapeutic potential for treating renal IRI.

# 4.2 The role of histone deacetylase 6 in equilibrium between rejection and tolerance after transplantation

The ultimate goal of transplantation is to achieve donor-specific tolerance, where recipients would enjoy long-term graft survival without the need for lifelong immunosuppressive drugs (103). Despite undeniable advances in RT techniques and IS, immunological rejection is the major cause of graft dysfunction and ultimately graft loss (104). There is compelling evidence that common immunosuppressants have no significant effect on long-term outcome of solid organ transplantation (SOT) because the prevention of chronic graft rejection is less effective with these drugs. Meanwhile, transplant recipients are at significant risk of side effects, infection, and malignancy due to lifelong systemic immunosuppression (105). Many efforts have been made to achieve graft tolerance, and researchers have attempted numerous approaches to induce

immune tolerance toward the donor graft in transplant recipients (4, 106). Given that both rejection and tolerance are influenced by the innate and adaptive immune systems, the ideal strategy would be to regulate B cells, DCs, and macrophages in a concerted manner, rather than targeting T cells alone, all of which lead to both the acute and chronic rejection response (107). This leads to the following result: understanding how the immune system works in tolerance and rejection of transplanted organs is of vital importance for the development of the improved therapeutics and better results for the recipient. HDAC6 has emerged as a hopeful policy in recent times to modulate antidonor immune reactions, allowing minimization of immunosuppressive drugs and induction of tolerance (50, 108). The hypothesis is supported by several lines of evidence that HDAC6 deacetylase activity is emerging as a key regulator of the immune response, the targeted inhibition of which has the potential to reduce the risk of organ rejection after transplantation (23, 40, 43). In this chapter, we describe the role of HDAC6 in the many immune cells and different mechanisms participating in the rejection (Figure 3A) and tolerance (Figure 3B) and discuss the popular and promising new HDAC6i approaches employed to treat transplant recipients.

# 4.2.1 Histone deacetylase 6 inhibitor-based therapies on dendritic cells

DCs are potent APCs that play important and diverse roles in initiating and regulating antigen-specific immune responses by capturing, processing, and presenting antigens (69). Acute and



#### FIGURE 3

Mechanisms of graft tolerance (A) and rejection (B) consist of complex processes involving the induction of proinflammatory APCs and tolerogenic DCs, leading to a different microenvironment. (1) HDAC6 promotes the expression of T-bet in the Th1 master regulating the ratio of Th1 to Th2, which also controls the population of CD4 cells, and the CD4T-bet/CD4T-GATA-3 ratio was significantly reduced in the HDAC6i group. in the HDAC6i group. (2) Acetylation promotes Foxp3 dimerisation, DNA binding and transcriptional activity, and expansion of the Treg population, and is ultimately associated with immune tolerance. The effects are likely to be related to increased Foxp3 acetylation and consequent expansion of Tregs. (3) HDAC6 inhibitor upregulates the Treg cell population through the STAT3/IL-10 tolerogenic axis to overcome immune tolerance and tilt the balance towards T-cell immunity. HDAC6 improved the expression levels of TF Rora in Th17, IL-17A secretion in Th17 cells and IL-4 levels secreted by Tregs, Th2 induced, as well as the differentiation of TH17/Treg imbalance. HDAC6i has therapeutic effects on the pathology of acute rejection by targeting the HDAC6/HIF-1a/ROR gt/Th17 signalling axis.

chronic allograft rejection is induced by DC-lymphocyte interaction with the assistance of other immune cells (109). During the process of graft migration and maturation to induce and regulate adaptive immunity, DCs can adopt different phenotypes-rejection-related DCs cause acute or chronic graft injury, whereas tolerogenic DCs help to prevent kidney damage by suppressing the overwhelming immune response. Through the production of cytokines (including IL-10, IL-12, and IL-23), DCs also play an essential role in the modulation of innate and adaptive immunity (69, 109). Of note, HDAC6 of specific APC genes plays a critical determinant of the chromatin substrate, which has been shown to be involved in immune synapse formation and immune response regulation (94). Disruption of HDAC6 leads to the induction of proinflammatory APCs (secreting the proinflammatory cytokines IFNy, IL-6, IL-1β, IL-12, and IL-23) and tolerogenic DCs (secreting the anti-inflammatory cytokine IL-10), which are critical for T-cell activation and induction of tolerance, respectively (94). The heterogeneity of the DCs and the states of their activation provides researchers with a greater range of options to define and manipulate the immune response researchers with a greater range of options to define and manipulate the immune response through these specialized leukocytes (110). The cytokine IL-10 and its immunosuppressive effects played a more important role in the generation of tolerant APCs. Expanding on these properties, Chromatin Immunoprecipitation (ChIP) analysis has also presented that HDAC6 regulates STAT3/IL-10 signaling activity, leading to increased antigen presentation (94). Together, previous work presented that HDAC6 activity is of paramount importance in the immune system by regulating DC-derived cytokines, which are of paramount importance in the modulation of the polarization of T cells, the activation of NK cells, and inflammation. HDAC6 thus opens up new avenues for therapeutic intervention to tip the balance in favor of the generation of APCs that are able to induce both up- and downregulation of important surface molecules.

# 4.2.2 Dependence of T cells on histone deacetylase 6 activation

Directional regulation of Th cell subset differentiation and induction of immune deviation is beneficial for inhibition of graft rejection and the induction of transplant tolerance (111). In the periphery, the goal of tolerance is to inhibit mature alloreactive T and B cells through mechanisms such as deletion, anergy, and regulation (112). HDAC6 is paramount to regulate the immune system, among other functions. In a murine model of cecal ligation and puncture (CLP), TA has been proven to alter the blood cell composition, along with lymphocyte population restoration (84). Previous studies have also shown that CKD-506 directly suppressed the proliferation of pathogenic lymphocytes (88). As clearly demonstrated in an immunization study, overexpression of HDAC6 contributes to increased T-cell migration and chemotactic capacity (113), whereas HDAC6 deficiency results in impaired CD8 T-cell functions. It has been demonstrated that the hydroxamic acid HDAC6 inhibitor ACY-1215 acts directly on T cells to inhibit antigen-specific CD8 T-cell differentiation and

proliferation during skin inflammation (114). It is important to note that the development and function of CD4 and CD8 T cells were invested to be normal in CD4-specific Hdac6 knockout mice (80). In addition, qRT-PCR results showed that ACY-1215 also suppressed the upregulation of IL-2 and IFN- $\gamma$  expression induced by CD8 T cells with anti-CD3/CD28 antibody stimulation in the Contact hypersensitivity (CHS) model (115). The results also presented that the CD4 T-cell proliferation and cytokine production were decreased (115). As mentioned in a previous work, phenotypic analyses of the effector T-cell (Teff) indicated an increased apoptosis of T-acute lymphoblastic leukemia (ALL) cells, following reduced Notch3 expression in the HDAC6 silencing group by specific short hairpin ribonucleic acid vector (shRNA) (116).

Hence, a regulation in T-bet expression of the Th1 master regulator and the ratio of Th1 to Th2 in the spleen may have resulted in the enhancement in rejected graft presentations (104). Analysis of the master regulator of Th1 and Th2 by CD4 cells indicated that the CKD506 group was less prone to express the Th1 differentiation factor T-bet and showed decreased levels of the Th1 cytokine IL-2 in a mouse model of systemic lupus erythematosus (61). In addition, the Th1/Th2 ratio, which controls CD4 cells, and the CD4T-bet/CD4T-GATA-3 ratio were significantly reduced in the CKD506 group compared to the vehicle group (61). It is known from previous work that T-cell migration and chemotaxis were increased in T-cell overexpression of HDAC6 (113). Another finding suggested that CKD-506 attenuated the clinical manifestations of EAE in mice by suppressing T-cell infiltration into the spinal cord and reducing the levels of Th1 cell-related inflammatory cytokines in the peripheral blood and spinal cord of mice (117). The evidence upon all data leads to a just conclusion that HDAC6 might make absolute sense in the activation of Teff cells and in their functions and that HDAC6i might be favorable in the treatment of Teff cell-mediated diseases, including rejection.

# 4.2.3 Dependence of Treg cells on histone deacetylase 6 activation

Tregs are active participants in the regulation of immune system homeostasis, and alteration of their phenotype is involved in the process of tolerance induction by suppressing naive and subsequent T cell responses, producing TGF-B or inhibiting Teff cells, thereby preventing allograft rejection (118, 119). The pharmacological function of Treg suppression in pathological conditions is an increasingly vibrant area of research. How Tregs therapy is used in SOT is specifically related to the goal of achieving tolerance, along with the reduction or elimination of immunosuppressants, as well as tissue repair and resistance to rejection (120, 121). On another level, established autoimmune diseases can be reversed or even cured by Tregs therapies, which may be effective in controlling autoimmune diseases in animal models (118). Nevertheless, a crucial issue related to Treg therapy is to ensure that the administration of Tregs is precarious and that there may be a decline in the suppressive effect in the body over time (120). Increased Treg proliferation and function exerted by selective HDAC6 inhibitors probably an extremely reasonable regimen for

the therapeutic strategy of the organ transplantation, particularly in tolerance.

A previous study reported that Foxp3 transcription factor, a key transcription factor involved in Treg development and activity, was deacetylated under the regulation of HDAC6, and HDAC6 knockout or HDAC6i enhanced the suppressive effect of Treg (122, 123). Previous reports indicate that HDAC6 was presented at a level several times higher in the Tregs than in the Tcon cells, and HDAC6-knockout mice are immunocompetent and inclined to be greater in vitro/vivo suppression than WT Tregs (124). Another considerable phenomenon associated with transplant models is that a strong determinant of Treg-dependent resistance is the presence or absence of HDAC6 within Tregs to allograft rejection, underscoring the significance of HDAC6 as a treatment strategy for Treg modulation (108). Preliminary work by Yang J et al. presents that there are Hdac6 KO Treg downregulate five MHC class II molecules, manifesting that HDAC6 could help the expression of MHC class II molecules among Treg (125). The published study reasonably concluded that the HDAC6-specific inhibitor promotes the suppressive activity of Treg in the model of fully MHC incompatible skin allograft rejection (108). In a recent study of a cardiac allograft model, the presence or absence of the HDAC6 molecule alone could determine acute rejection or longterm engraftment (121). In addition, long-term graft survival was also observed in HDAC6-/- mice, but not in WT mice, when recipients in fully immunocompetent hosts received a subtherapeutic dose of rapamycin (126).

Beier and colleagues provided evidence to support the previous study that HDAC6 is involved in promoting other transcription factor activities necessary for Treg development and activation, such as cyclic adenosine monophosphate response element-binding protein (125). More evidence, furthermore, suggested that HDAC6 deficiency took part in the stabilizing the acetylated form of IL-2, which mediates STAT5, thereby promoting its transcriptional activity (125). In terms of other molecular mechanisms, previous studies have confirmed that Foxp3 complexes with HSP70, with increased HSP70 promoting Treg activity and efficacy under stress conditions, and subsequent inhibition of HSP70 impairing Treg activity and efficacy (127). Interestingly, there is strong evidence that HDAC6 or HSP90 inhibitors produce similar results in models of inflammation and autoimmunity, that is, HDAC6i or HSP90i can rescue Tregs population and HSP90-dependent enhancement of Treg function with HDAC6i (128). Taken together, HDAC6 is an important regulator of Treg plasticity that determines the fate of plastic Tregs, which promotes expression of Th subset regulators in Tregs and regulates plasticity and heterogeneity of Tregs. Enhancement of the suppressive function of Tregs with HDAC6i is an extremely potential target for the achievement of tolerance as well as for helping prevent graft rejection.

# 4.2.4 The divergent roles of histone deacetylase 6 in Th17 cells

There is accumulating evidence to suggest that Th1 cells result in the progress of both acute and chronic rejection of the allograft following transplantation of a variety of organs, including the kidney (129). Th17 cells, a subset of T lymphocytes, are known to be involved in the pathogenesis of many autoimmune diseases through the production of proinflammatory cytokines such as IL-17, which is known to regulate many inflammatory diseases (130, 131). Preliminary findings have presented that Th17 cells and their secretion of IL-17 are involved not solely in the course of acute rejection but also in the development of iTreg differentiationassociated tolerance induction (132). Interestingly, accumulating evidence exhibits that the ultimate development of TH17 has a closely reciprocal relationship with Tregs fate and arises from a common T-cell precursor (133). Additionally, a study of systemic autoimmune disease suggested that the transfer of Tregs enhanced IL-17 production in in vivo (133). There is a range of evidence that the favorable approach of HDAC6i inhibits the activity of Th17 cells in lung allografts and ultimately presents a protective effect (134). However, other results have shown that HDAC6 improved the expression levels of TF Tbx21 in Th1, together with expression of TF Rora in Th17 and IL-4 levels secreted by Tregs induced by Th2 (133).

Available data confirm that differentiated Tregs, probably through their TGF- $\beta$  production, contribute to the promotion of the Th17 cell differentiation fate of CD4 T cells, with IL-6 levels determining the transcriptional balance between Foxp3 and RORyt in the presence of TGF- $\beta$  (135). A previous study showed that Th17 and IL-17+  $\gamma\delta$  T cells were depleted in recipients following adoptive transfer of induced Treg cells (iTregs) and attenuated the pathological manifestation of acute rejection by suppressing Th17 cell accumulation in lung allograft recipients (132, 135). Previous data demonstrated that ACY-1215 decreased IL-17A secretion in Th17 cell culture system, as well as the Th17 cell differentiation, which was confirmed in the experimental autoimmune thyroiditis (EAT) mouse model (134). Also, a recent study exhibited that the HDAC6-specific inhibitor TA showed the same result in an orthotopic mouse lung transplantation model, demonstrating the protective role of HDAC6 in lung allografts through the suppression of Th17 cells (136). It is presented that TA suppressed the differentiation and function of Th17 cells and ultimately improved the attenuation of acute rejection of lung allografts and prolonged survival (136). Furthermore, a dramatically increased rate of Tregs to CD4 T cells in both allograft and spleen isolation in the TA-treated group is also noted, as well as an obviously increased IL-10 level of Tregs in the TA-treated group in the animal lung transplantation experiment. From another perspective, HDAC6 depletion restored the suppressive activity of Tregs in models of autoimmunity and inflammation (136). The above results correlate well with previous reports showing that HDAC6 inhibition/deficiency enhances Treg function and inhibits TH17 lineage commitment, thereby readjusting Th17 and Treg cell fractions. In allograft recipients, all reliable evidence suggests that HDAC6i therapy had a general and substantial effect on the fractions of Th17 and Tregs.

Other mechanistic studies have mainly involved HIF, the role of an essential regulator in T-cell fate determination. It has been reported that HIF-1, the transcription factor in response to hypoxia (137), actually regulates the balance of TH17/Treg (138). It has been demonstrated that high HIF-1 $\alpha$  expression was

observed in lung allograft transplantation and that HIF-1 $\alpha$  is required for Th17 cell development (136). HIF-1 $\alpha$  KO mice were protected from acute rejection in the lung allograft and manifested an apparently reduced infiltration of Th17 cells. Consistent with previous presentations, high HIF-1a expression of T cells can be induced by hypoxic and normoxic conditions (137). A previous study has identified an important target through which HIF-1α regulates Th17 cell expansion and function, such as the HIF-1αdependent RORyt pathway, as reduced RORyt expression was observed in HIF-1 $\alpha$ -deficient mice (139). Other studies have reported that decreased RORyt levels are of greatest value for HIF-1α, resulting in downregulation of Th17 cells and attenuated acute rejection of lung allografts (132). A recent study demonstrated that TA downregulated HIF-1 transcriptional activity of nucleus pulposus cells and the protein expression level, and in the TAtreated lung transplant group, reduced HIF-1α expression may be in relation to the improvement of inflammation responses (140). Collectively, there is significant evidence that HDAC6i, TA, has therapeutic effects on the pathological lesions of acute rejection in the lung allograft models by targeting the HDAC6/HIF-1 $\alpha$ /ROR  $\gamma$ t/ Th17-signaling axis. Given the above evidence of plasticity between TH17 and Treg programs, attention to manipulating the balance of these cell lineage decisions may suggest novel strategies for treating RT fields associated with TH17/Treg imbalance.

### 4.2.5 The roles of histone deacetylase 6 in B cells

Playing an essential role in the immunological response, B cells can promote rejection through the production of antibodies, proinflammatory cytokine expression, and presenting antigens to T-cells, which leads to the activation and differentiation of T cells (105). B cells also evolve into antibody-secreting cells in which the antibodies act as opsonins to facilitate activation of DCs and T-cell responses, which can also produce IL-10 and TGF-B to induce Treg differentiation, thereby reducing the severity of inflammatory diseases (140, 141). It has long been known that the expression level in B cells and splenic B cells was significantly increased in diseased MRL/MpJ-Faslpr (MRL/lpr) mice (84, 142). It was previously reported that pre-B cells treated with ACY-738 were able to suppress pre-B cell proliferation by increasing Bax expression and reducing the Bcl-2:Bax ratio, thereby supporting a proapoptotic environment in vitro (143). Furthermore, ACY-738 treatment restored early pre-B-cell proportions but reduced late pre-B-cell proportions in the bone marrow during SLE in NZB/W mice (57). It is worth noting that treatment with HDAC6i upregulates alpha-tubulin acetylation and reduces the activity of cytoplasmic HDACs of B cells and restores the percentage of B lymphocytes (142). Based on the study that Hsp90 hyperacetylation has been proven to reduce Hsp90 function; hyperacetylated  $\alpha$ tubulin and Hsp90 of B cells were observed in mice depleted of HDAC6 and those treated with HDAC6 small interfering Ribonucleic Acid (siRNA) (144). All these findings indicate that HHDAC6 may have a critical role in the dysregulation of B-cell development during rejection, that restored HDAC6 expression contributes to immunopathogenesis, and that HDAC6i appears to be advantageous in the prevention of rejection through the acetylation of key signaling and transcription factors involved in the activation of the immune system. Altogether, the limited success of these reported strategies has provided novel insights on HDACi6 plasticity into improvement to prevent allograft rejection, which will eventually be beneficial in inducing the future development of immune tolerance in clinical trials.

# 4.3 Mechanism and histone deacetylase 6 of renal graft fibrosis

The progression and development of renal injury after RT ultimately contributes to renal interstitial fibrosis in chronic renal allograft injury, which is also the major precipitating factor in the decline of renal function (145). Several factors, including IRI, acute rejection, various types of infection, and chronic allograft dysfunction, can lead to inflammation in the recipient's kidney, which is a major consequence of the development of chronic hypoxia, ultimately leading to renal fibrosis (6). Emerging evidence implicates HDAC6 in renal fibrogenesis (146) (Figure 4).

Accumulating studies from past and present have reported the significant role of HDAC6 implicated in inflammatory and fibrotic disorders in a model of inflammation in a deacetylase activitydependent manner, which we also mentioned in the previous IRI part. A previous study demonstrated that genetic knockdown and pharmacological HDAC6i regulated inflammatory cytokine production in response to multiple pathogen stimuli such as LPS, Clostridium difficile Toxin A, and HIV-1 Tat. This supports previous findings that HDAC6 overexpression induces ROS generation and inflammatory cytokine secretion (147). It was previously shown that HDAC6i also attenuated LPS-induced macrophage activation and production of proinflammatory cytokines, including TNF- $\alpha$ , IL-1 $\beta$ , and IL-6, while reducing acetylated  $\alpha$ -tubulin (148). In line with the changes in inflammatory cytokines, there was a significant attenuation of nicotine-induced macrophage pyroptosis by HDAC6 deficiency by TA or siRNA, confirmed by decreased protein expression levels of NLRP3, cleaved caspase1, IL-1β, IL-18, and LDH (149). In addition, administration of HDAC6i by TA effectively suppressed the production of MPO, TNF-a, and IL-6 in the cisplatin-induced AKI, as well as the prevention of damage to kidney function, together with the survival improvement in a rodent model of subsequent CLP (150). Given the known effect of regulating the expression of inflammatory cytokines, HDAC6 also modulates other proinflammatory factors such as monocyte chemoattractant protein-1 (MCP-1), which is thought to play an important role in monocyte recruitment during acute inflammation (75). Studies have also shown that monocyte/macrophage migration into the peritoneal cavity is reduced by the loss of HDAC6 (89).

It has been observed that TubA could reduce the increased expression and localization of connective tissue growth factor (CTGF), a well-known pro-fibrotic factor, in kidney tissue sections after ANG treatment (98). The mRNA and protein levels of fibrosis marker genes (fibronectin, collagen type I, and collagen type III) were significantly increased in renal fibrosis in hypertensive mice compared to the control sham group, which



#### FIGURE 4

Schematic illustration of the anti-fibrotic property of HDAC6i in renal transplantation. Injured tissue or activated immune cells secrete profibrotic factors that induce fibroblast differentiation into myofibroblasts. Myofibroblasts actively synthesise extracellular matrix. HDAC6i negatively regulates fibrosis. (1) In the initial stage of the pro-fibrotic process, intra-graft inflammation, an integral part of the host defence mechanism in response to injury, is activated. Through modulation of p38 MAPK, NF-kB and AP-1 pathways, several pro-inflammatory, pro-fibrotic cytokines and adhesion molecules (MMP-1, MMP-2, MMP-9, MCP-1) are secreted by tubular cells with consequent recruitment of inflammatory infiltrates (lymphocytes, macrophages, DCs, neutrophils) that may facilitate the recruitment of new interstitial mononuclear cells. (2) Macrophages are a major source of TGF-b, which modulates the production of IL-1 $\beta$ , TNF- $\alpha$ , IFN-b $\gamma$  and IL-10 and induces myofibroblast differentiation and the production of ECM proteins. (3) Damaged mitochondria mediate apoptosis and necrosis with the consequent release of DAMPs, which activate the NLRP3 inflammasome and participate in the prosis of fibrosis. (4) ROS accumulation via IRI, NADPH oxidase and mitochondrial dysfunction contribute to EMT. (5) Inhibition of HDAC6 attenuated EGFR, TGF-b1 induced EMT markers, which is accompanied by HDAC6-dependent deacetylation of a -tubulin. (6) The underlying mechanism by which HDAC6 inhibition can attenuate fibrosis by inhibiting profibrotic genes, multiple activations of cell pro-fibrotic signalling pathways and inflammation. Dashed arrows indicate indirect pro-fibrotic effects.

was blocked by TubA treatments or HDAC6 siRNA. Similarly, in response to ANG stimulation, immunohistochemistry showed that TubA administration alleviated FSP1 expression, a specific hallmark of renal fibrosis, in the interstitial region (52). Emerging research has shown that collagen type 1 is a novel substrate for acetylation and deacetylation by p300 and HDAC6, respectively. Moreover, knockdown of the p300 transcriptional coactivator reduced collagen type I acetylation, in contrast to TubA, which ultimately modulates collagen expression (52).

A wealth of experimental evidence supports the diverse role of HDAC6 in the regulation of key transcription factors of inflammatory signaling (98). A previous finding indicated that HDAC6 modulates inflammatory expression, which may result from modulation of the p38 MAPK, NF- $\kappa$ B and AP-1 pathways, or perhaps other underlying pathways (99). Activation of the NF- $\kappa$ B pathway, a canonical inflammatory pathway, is engaged in the development of AKI and is responsible for the transcriptional induction of a variety of proinflammatory cytokines and a number of chemokines (98). Studies in mouse models of inflammation have implicated the overexpression of HDAC6 in the abnormal activation of inflammatory signaling pathways, including the NF- $\kappa$ B pathway, in cisplatin-induced AKI (53, 95). The level of NF- $\kappa$ B p65 phosphorylation was significantly promoted in the cytoplasm and reduced in the nucleus by

pharmaceutical HDAC6i with TA (99). Confocal microscopy showed that TA apparently blocked nicotine-induced p65 nuclear translocation (151). In agreement with previous reports, it has also been shown that HDAC6 inhibition by TA pretreatment restores site-specific acetylation of p65 and reduces its nuclear translocation in another model. In addition, p65 has been implicated in NLRP3 transcription and macrophage pyroptosis (151). Taken together, it can be concluded that HDAC6 inhibition by TA attenuates cisplatin-induced inflammatory cytokine expression in AKI and nicotine-induced macrophage pyroptosis by reducing NF- $\kappa$ B phosphorylation (151, 152).

Increasingly, previous research has indicated that persistently high expression of the phosphorylated epidermal growth factor receptor (EGFR) signaling pathway is also involved in the fibrotic kidney, suggesting that the EGFR signaling pathway also plays a key role in mediating angiotensin II–induced renal fibrosis. ACY-1215 treatment also attenuates renal fibrosis by inhibiting Unilateral Ureteral Obstruction (UUO)-induced phosphorylation of EGFR and its downstream signaling protein molecules (98). TGF- $\beta$ , the most important marker of interstitial fibrosis, is of prime importance in synthesis and degradation of epithelial– mesenchymal transition (EMT) by inducing transcription of collagen genes and inducing EMT of tubular epithelial cells, leading to interstitial fibrosis and chronic allograft dysfunction

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(153). In addition, from another work on idiopathic pulmonary fibrosis, it is known that inhibition of HDAC6 attenuated TGF- $\beta$ -induced EMT markers released from the complex by macrophages and activated by inflammation, accompanied by HDAC6-dependent deacetylation of  $\alpha$ -tubulin (154). Mechanistically, HDAC6 is shown to influence epigenetic histone modification, leading to renal fibrosis and TGF- $\beta$ 1-dependent nuclear accumulation of Smad3-dependent fibrotic genes, and inhibition of HDAC6 attenuated renal fibrosis and inflammation (58).

Collectively, selective HDAC6i can be involved in attenuating fibrosis by inhibiting profibrotic genes, multiple activations of cell pro-fibrotic signaling pathways, and inflammation. The favorable safety profile of HDAC6i and works presenting HDAC6 in multiple inflammatory and fibrotic diseases make sure the pathological change of selective HDAC inhibitors in the therapeutic strategy of RT-related fibrosis.

### 5 Limitations and future perspectives

With the elucidated mechanism of RT-related complications, HDAC6 has attracted a great deal of attention as a potential drug target. As the mechanism of RT-related complications induced by immune and non-immune injury has been elucidated, HDAC6 has attracted wide attention as a potential drug target. Due to its unique cytoplasmic localization, HDAC6 isoform regulates acetylation status of  $\alpha$ -tubulin, HSP90, TGF- $\beta$ , and peroxiredoxins. As an emerging class of drugs, selective HDAC6 inhibitors can act on many pathological links of various immune-related and renal diseases, such as oxidative stress injury, inflammatory response, apoptosis, B-cell and T-cell activation, Th cell differentiation, and macrophage responses, and have been shown to be beneficial in preventing disease progression. However, existing HDAC6 inhibitors also have some shortcomings, such as tubacin, which is useful as a research tool but is not suitable for clinical treatment due to its highly lipophilic properties, rapid metabolism in the body, and cumbersome synthesis (155). Although many contemporary HDAC6 inhibitors are efficacious in the immune system, only the DD2 domain has been well targeted in all small-molecule HDAC6 inhibitors, leading to off-target effects in pharmacological inhibitors due to limited selectivity (156). The absence of another ubiquitinbinding zinc-finger domain could result in discrepancies between experimental observations made with pharmacological inhibitors and genetic knockout approaches, the latter targeting both catalytic domains. It will be interesting to explore whether other targets contribute to HDAC6's role in inflammation and inflammatory disorders. Additionally, some HDAC6 inhibitors do not display high selectivity and may inhibit related HDACs, and the newly discovered HDAC6 inhibitors are still in the experimental stage (157, 158). Additionally, it should be mentioned that although there is limited literature on RT directly, the conclusions in other organ transplantations, fibrosis and IRI models can strongly prove the effectiveness and feasibility of HDAC6 application in the field of RT. Thus, further basic research in preclinical models is necessary for the identification of novel high-potency HDAC6 inhibitors with superior selectivity. Finally, regarding its role in immune responsivity, HADC6 may be involved in the recurrence of primary kidney diseases in RT; however, additional studies are needed to confirm this hypothesis.

### 6 Conclusion

A substantial amount of validated studies has identified HDAC6 as a pivotal modulator of inflammation and a promising therapeutic candidate for the management of a variety of renal diseases. Even more significantly, small-molecule inhibitors of HDAC6 have not been accompanied by severe toxic effects. The highly selective HDAC6 inhibitor TA has been evaluated in the therapeutic areas of neurodegenerative diseases, cancer, and COPD with no apparent adverse effects. Over the past few decades, HDAC6 has been identified as a relatively attractive target for the development of drugs to treat a wide range of diseases. These studies have significantly improved our understanding of HDAC6 function in the kidney and support the potential evaluation of HDAC inhibitors in kidney transplant patients. In view of the innovation of these inhibitors, more work is needed to substantiate their role as a promising intervention and to address unknowns before clinical translation.

Collectively, the favorable profile of HDAC6 inhibitors and the identification of HDAC6 in a series of inflammatory disorders offer compelling opportunities for the advancement of selective HDAC inhibitors in the management of RT-associated complications. Important evidence on the HDAC6 apparatus particularly involved in the pathogenesis of renal complications is encapsulated in the present review through modulation of innate and adaptive immune responses, which focuses on therapeutically relevant avenues with considerable translational promise to the clinic.

### Author contributions

Q-qZ and SC mainly contributed to the conception, the design, and the writing of the manuscript. Q-qZ wrote, prepared, reviewed, and edited the manuscript; W-jZ reviewed, revised, and commented on this article and funding acquisition; Q-qZ and SC supported the final draft editing and revised the manuscript critically for final acceptance for publication. SC supported and supervised the overall design of the article. Q-qZ conceived of all figures. All authors contributed to the article and approved the submitted version.

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### Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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### References

1. Wouk N. End-stage renal disease: medical management. Am Fam Phys (2021) 104(5):493-9.

2. Marroquin CE. Patient selection for kidney transplant. Surg Clin North Am (2019) 99(1):1-35. doi: 10.1016/j.suc.2018.09.002

3. Zhao H, Alam A, Soo AP, George AJT, Ma D. Ischemia-reperfusion injury reduces long term renal graft survival: mechanism and beyond. *EBioMedicine* (2018) 28:31-42. doi: 10.1016/j.ebiom.2018.01.025

4. Rickert CG, Markmann JF. Current state of organ transplant tolerance. *Curr Opin Organ Transplant* (2019) 24(4):441–50. doi: 10.1097/MOT.000000000000670

5. Erpicum P, Bonvoisin C, Grosch S, Bovy C, Pinto Coelho P, Detry O, et al. Diagnostic et prise en charge thérapeutique du rejeten transplantation rénale [Diagnosis and management of renal allograft rejection]. *Rev Med Liege* (2022) 77(5-6):338-44.

6. Granata S, Benedetti C, Gambaro G, Zaza G. Kidney allograft fibrosis: what we learned from latest translational research studies. *J Nephrol* (2020) 33(6):1201–11. doi: 10.1007/s40620-020-00726-z

7. Lu J, Zhang Y, Sun J, Huang S, Wu W, Tan J. The immune cell landscape in renal allografts. *Cell Transplant* (2021) 30:963689721995458. doi: 10.1177/0963689721995458

8. Foroncewicz B, Mucha K, Florczak M, Szymańska A, Ciszek M, Durlik M, et al. Long-term outcome of renal transplantation: a 10-year follow-up of 765 recipients. *Pol Arch Intern Med* (2019) 129(7-8):476–83. doi: 10.20452/pamw.14914

9. Banas B, Krämer BK, Krüger B, Kamar N, Undre N. Long-term kidney transplant outcomes: role of prolonged-release tacrolimus. *Transplant Proc* (2020) 52(1):102–10. doi: 10.1016/j.transproceed.2019.11.003

10. Levine MH, Wang Z, Bhatti TR, Wang Y, Aufhauser DD, McNeal S, et al. Classspecific histone/protein deacetylase inhibition protects against renal ischemia reperfusion injury and fibrosis formation. *Am J Transplant* (2015) 15(4):965–73. doi: 10.1111/ajt.13106

11. Hadden MJ, Advani A. Histone deacetylase inhibitors and diabetic kidney disease. *Int J Mol Sci* (2018) 19(9):2630. doi: 10.3390/ijms19092630

12. Liu H. The roles of histone deacetylases in kidney development and disease. Clin Exp Nephrol (2021) 25(3):215–23. doi: 10.1007/s10157-020-01995-5

13. Yang XJ, Seto E. HATs and HDACs: from structure, function and regulation to novel strategies for therapy and prevention. *Oncogene* (2007) 26(37):5310-8. doi: 10.1038/sj.onc.1210599

14. Kornberg RD. Chromatin structure: a repeating unit of histones and DNA. *Science* 184(1974):868–71. doi: 10.1126/science.184.4139.868

15. Sadoul K, Boyault C, Pabion M, Khochbin S. Regulation of protein turnover by acetyltransferases and deacetylases. *Biochimie* (2008) 90:306–12. doi: 10.1016/j.biochi.2007.06.009

16. Brilli LL, Swanhart LM, de Caestecker MP, Hukriede NA. HDAC inhibitors in kidney development and disease. *Pediatr Nephrol* (2013) 28(10):1909–21. doi: 10.1007/s00467-012-2320-8

17. Dekker FJ, Haisma HJ. Histone acetyl transferases as emerging drug targets. *Drug Discovery Today* (2009) 14:942-8. doi: 10.1016/j.drudis.2009.06.008

18. Islam R, Dash D, Singh R. Intranasal curcumin and sodium butyrate modulates airway inflammation and fibrosis *via* HDAC inhibition in allergic asthma. *Cytokine* (2022) 149:155720. doi: 10.1016/j.cyto.2021.155720

19. Hagelkruys A, Sawicka A, Rennmayr M, Seiser C. The biology of HDAC in cancer: the nuclear and epigenetic components. *Handb Exp Pharmacol* (2011) 206:13–37. doi: 10.1007/978-3-642-21631-2\_2

20. Hamminger P, Rica R, Ellmeier W. Histone deacetylases as targets in autoimmune and autoinflammatory diseases. *Adv Immunol* (2020) 147:1–59. doi: 10.1016/bs.ai.2020.06.001

21. Grozinger CM, Schreiber SL. Deacetylase enzymes: biological functions and the use of small-molecule inhibitors. *Chem Biol* (2002) 9:3–16. doi: 10.1016/S1074-5521 (02)00092-3

22. Roche J, Bertrand P. Inside HDACs with more selective HDAC inhibitors. Eur J Med Chem (2016) 121:451-83. doi: 10.1016/j.ejmech.2016.05.047

23. Ke B, Chen Y, Tu W, Ye T, Fang X, Yang L. Inhibition of HDAC6 activity in kidney diseases: a new perspective. *Mol Med* (2018) 24(1):33. doi: 10.1186/s10020-018-0027-4

24. Gupta PK, Reid RC, Liu L, Lucke AJ, Broomfield SA, Andrews MR, et al. Inhibitors selective for HDAC6 in enzymes and cells. *Bioorg Med Chem Lett* (2010) 20:7067–70. doi: 10.1016/j.bmcl.2010.09.100 organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

25. Lee JH, Mahendran A, Yao Y, Ngo L, Venta-perez G, Choy ML, et al. Development of a histone deacetylase 6 inhibitor and its biological effects. *Proc Natl Acad Sci USA* (2013) 110:15704-9. doi: 10.1073/pnas.1313893110

26. Li L, Yang XJ. Tubulin acetylation: responsible enzymes, biological functions and human diseases. *Cell Mol Life Sci* (2015) 72(22):4237–55. doi: 10.1007/s00018-015-2000-5

27. Du Y, Yang X, Li Z, Le W, Hao Y, Song Y, et al. HDAC6-mediated Hsp90 deacetylation reduces aggregation and toxicity of the protein alpha-synuclein by regulating chaperone-mediated autophagy. *Neurochem Int* (2021) 149:105141. doi: 10.1016/j.neuint.2021.105141

28. Zhang X, Yuan Z, Zhang Y, Yong S, Salas-Burgos A, Koomen J, et al. HDAC6 modulates cell motility by altering the acetylation level of cortactin. *Mol Cell* (2007) 27:197–213. doi: 10.1016/j.molcel.2007.05.033

29. Boucherat O, Chabot S, Paulin R, Trinh I, Bourgeois A, Potus F, et al. HDAC6: a novel histone deacetylase implicated in pulmonary arterial hypertension. *Sci Rep* (2017) 7(1):4546. doi: 10.1038/s41598-017-04874-4

30. Hada M, Kwok RP. Regulation of ku70-bax complex in cells. J Cell Death (2014) 7:11–3. doi: 10.4137/JCD.S13695

31. Ryu HW, Shin DH, Lee DH, Choi J, Han G, Lee KY, et al. HDAC6 deacetylates p53 at lysines 381/382 and differentially coordinates p53-induced apoptosis. *Cancer Lett* (2017) 391:162–71. doi: 10.1016/j.canlet.2017.01.033

32. Keremu A, Aimaiti A, Liang Z, Zou X. Role of the HDAC6/STAT3 pathway in regulating PD-L1 expression in osteosarcoma cell lines. *Cancer Chemother Pharmacol* (2019) 83(2):255–64. doi: 10.1007/s00280-018-3721-6

33. Imai Y, Hirano M, Kobayashi M, Futami M, Tojo A. HDAC inhibitors exert anti-myeloma effects through multiple modes of action. *Cancers (Basel)* (2019) 11 (4):475. doi: 10.3390/cancers11040475

34. Qiu W, Wang B, Gao Y, Tian Y, Tian M, Chen Y, et al. Targeting histone deacetylase 6 reprograms interleukin-17-producing helper T cell pathogenicity and facilitates immunotherapies for hepatocellular carcinoma. *Hepatology* (2020) 71 (6):1967–1987. doi: 10.1002/hep.30960

35. Yan X, Qu X, Liu B, Zhao Y, Xu L, Yu S, et al. Autophagy-induced HDAC6 activity during hypoxia regulates mitochondrial energy metabolism through the  $\beta$ -Catenin/COUP-TFII axis in hepatocellular carcinoma cells. Front Oncol (2021) 11:742460. doi: 10.3389/fonc.2021.742460

36. Osko JD, Christianson DW. Structural basis of catalysis and inhibition of HDAC6 CD1, the enigmatic catalytic domain of histone deacetylase 6. *Biochemistry* (2019) 58(49):4912-24. doi: 10.1021/acs.biochem.9b00934

37. Zou H, Wu Y, Navre M, Sang B-C. Characterization of the two catalytic domains in histone deacetylase 6. *Biochem Biophys Res Commun* (2006) 341:45–50. doi: 10.1016/ j.bbrc.2005.12.144

38. Hyndman KA. Histone deacetylases in kidney physiology and acute kidney injury. Semin Nephrol (2020) 40(2):138-47. doi: 10.1016/j.semnephrol.2020.01.005

39. Cebotaru L, Liu Q, Yanda MK, Boinot C, Outeda P, Huso DL, et al. Inhibition of histone deacetylase 6 activity reduces cyst growth in polycystic kidney disease. *Kidney Int* (2016) 90(1):90–9. doi: 10.1016/j.kint.2016.01.026

40. Vieson MD, Gojmerac AM, Khan D, Dai R, van Duzer JH, Mazitschek R, et al. Treatment with a selective histone deacetylase 6 inhibitor decreases lupus nephritis in NZB/W mice. *Histol Histopathol* (2017) 32(12):1317–32. doi: 10.14670/HH-11-885

41. Zhang Z, Cao Y, Zhao W, Guo L, Liu W. HDAC6 serves as a biomarker for the prognosis of patients with renal cell carcinoma. *Cancer Biomark* (2017) 19(2):169–75. doi: 10.3233/CBM-160298

42. Choi SY, Ryu Y, Kee HJ, Cho SN, Kim GR, Cho JY, et al. Tubastatin a suppresses renal fibrosis *via* regulation of epigenetic histone modification and Smad3-dependent fibrotic genes. *Vascul Pharmacol* (2015) 72:130–40. doi: 10.1016/j.vph.2015.04.006

43. Vishwakarma S, Iyer LR, Muley M, Singh PK, Shastry A, Saxena A, et al. Tubastatin, a selective histone deacetylase 6 inhibitor shows anti-inflammatory and anti-rheumatic effects. *Int Immunopharmacol* (2013) 16(1):72–8. doi: 10.1016/j.intimp.2013.03.016

44. Kumari S, Carmona AV, Tiwari AK, Trippier PC. Amide bond bioisosteres: strategies, synthesis, and successes. *J Med Chem* (2020) 63(21):12290–358. doi: 10.1021/acs.jmedchem.0c00530

45. Ho TCS, Chan AHY, Ganesan A. Thirty years of HDAC inhibitors: 2020 insight and hindsight. J Med Chem (2020) 63(21):12460-84. doi: 10.1021/acs.jmedchem.0c00830

46. Zhang XH, Qin-Ma, Wu HP, Khamis MY, Li YH, Ma LY, et al. A review of progress in histone deacetylase 6 inhibitors research: structural specificity and functional diversity. J Med Chem (2021) 64(3):1362–91. doi: 10.1021/acs.jmedchem.0c01782

47. Santo L, Hideshima T, Kung AL, Tseng JC, Tamang D, Yang M, et al. Preclinical activity, pharmacodynamic, and pharmacokinetic properties of a selective HDAC6 inhibitor, ACY-1215, in combination with bortezomib in multiple myeloma. *Blood* (2012) 119(11):2579–89. doi: 10.1182/blood-2011-10-387365

48. Huang P, Almeciga-Pinto I, Jarpe M, van Duzer JH, Mazitschek R, Yang M, et al. Selective HDAC inhibition by ACY-241 enhances the activity of NSC 125973 in solid tumor models. *Oncotarget* (2017) 8(2):2694–707. doi: 10.18632/oncotarget.13738

49. Butler KV, Kalin J, Brochier C, Vistoli G, Langley B, Kozikowski AP. Rational design and simple chemistry yield a superior, neuroprotective HDAC6 inhibitor, tubastatin a. J Am Chem Soc (2010) 132:10842–10846. doi: 10.1021/ja102758v

50. Park JK, Shon S, Yoo HJ, Suh DH, Bae D, Shin J, et al. Inhibition of histone deacetylase 6 suppresses inflammatory responses and invasiveness of fibroblast-like-synoviocytes in inflammatory arthritis. *Arthritis Res Ther* (2021) 23(1):177. doi: 10.1186/s13075-021-02561-4

51. Carbajal A, Chesta ME, Bisig CG, Arce CA. A novel method for purification of polymerizable tubulin with a high content of the acetylated isotype. *Biochem J* (2013) 449(3):643–8. doi: 10.1042/BJ20121439

52. Shi Y, Xu L, Tang J, Fang L, Ma S, Ma X, et al. Inhibition of HDAC6 protects against rhabdomyolysis-induced acute kidney injury. *Am J Physiol Renal Physiol* (2017) 312(3):F502–15. doi: 10.1152/ajprenal.00546.2016

53. Shi L, Song Z, Li C, Deng F, Xia Y, Huang J, et al. HDAC6 inhibition alleviates ischemia- and cisplatin-induced acute kidney injury by promoting autophagy. *Cells* (2022) 11(24):3951. doi: 10.3390/cells11243951

54. Fan SJ, Huang FI, Liou JP, Yang CR. The novel histone de acetylase 6 inhibitor, MPT0G211, ameliorates tau phosphorylation and cognitive deficits in an alzheimer's disease model. *Cell Death Dis* (2018) 9(6):655. doi: 10.1038/s41419-018-0688-5

55. Mithraprabhu S, Khong T, Jones SS, Spencer A. Histone deacetylase (HDAC) inhibitors as single agents induce multiple myeloma cell death principally through the inhibition of class I HDAC. *Br J Haematol* (2013) 162(4):559–62. doi: 10.1097/TA.00000000000897

56. Lorenzo Pisarello M, Masyuk TV, Gradilone SA, Masyuk AI, Ding JF, Lee PY, et al. Combination of a histone deacetylase 6 inhibitor and a somatostatin receptor agonist synergistically reduces hepatorenal cystogenesis in an animal model of polycystic liver disease. *Am J Pathol* (2018) 188(4):981–94. doi: 10.1016/j.ajpath.2017.12.016

57. Regna NL, Vieson MD, Luo XM, Chafin CB, Puthiyaveetil AG, Hammond SE, et al. Specific HDAC6 inhibition by ACY-738 reduces SLE pathogenesis in NZB/W mice. *Clin Immunol* (2016) 162:58–73. doi: 10.1016/j.clim.2015.11.007

58. Chen X, Yu C, Hou X, Li J, Li T, Qiu A, et al. Histone deacetylase 6 inhibition mitigates renal fibrosis by suppressing TGF- $\beta$  and EGFR signaling pathways in obstructive nephropathy. *Am J Physiol Renal Physiol* (2020) 319(6):F1003–14. doi: 10.1152/ajprenal.00261.2020

59. Yanda MK, Liu Q, Cebotaru L. An inhibitor of histone deacetylase 6 activity, ACY-1215, reduces cAMP and cyst growth in polycystic kidney disease. *Am J Physiol Renal Physiol* (2017) 313(4):F997–F1004. doi: 10.1152/ajprenal.00186.2017

60. Lee J, Hong EC, Jeong H, Hwang JW, Kim H, Bae EK, et al. A novel histone deacetylase 6-selective inhibitor suppresses synovial inflammation and joint destruction in a collagen antibody-induced arthritis mouse model. *Int J Rheumatol Dis* (2015) 18:514–23. doi: 10.1016/j.tcb.2008.04.003

61. Choi EW, Song JW, Ha N, Choi YI, Kim S. CKD-506, a novel HDAC6-selective inhibitor, improves renal outcomes and survival in a mouse model of systemic lupus erythematosus. *Sci Rep* (2018) 8(1):17297. doi: 10.1038/s41598-018-35602-1

62. Bergman JA, Woan K, Perez-Villarroel P, Villagra A, Sotomayor EM, Kozikowski AP. Selective histone deacetylase 6 inhibitors bearing substituted urea linkers inhibit melanoma cell growth. *J Med Chem* (2012) 55(22):9891–9899. doi: 10.1111/ajt.16425

63. Richardson PG, Moreau P, Laubach JP, Maglio ME, Lonial S, San-Miguel J. Deacetylase inhibitors as a novel modality in the treatment of multiple myeloma. *Pharmacol Res* (2017) 117:185–91. doi: 10.1016/j.phrs.2016.11.020

64. Li ZY, Li QZ, Chen L, Chen BD, Zhang C, Wang X, et al. HPOB, an HDAC6 inhibitor, attenuates corticosterone-induced injury in rat adrenal pheochromocytoma PC12 cells by inhibiting mitochondrial GR translocation and the intrinsic apoptosis pathway. *Neurochem Int* (2016) 99:239–51. doi: 10.1016/j.neuint.2016.08.004

65. Wang Z, Tang F, Hu P, Wang Y, Gong J, Sun S, et al. HDAC6 promotes cell proliferation and confers resistance to gefitinib in lung adenocarcinoma. *Oncol Rep* (2016) 36(1):589–97. doi: 10.3892/or.2016.4811

66. Rengarajan J, Szabo SJ, Glimcher LH. Transcriptional regulation of Th1/Th2 polarization. *Immunol Today* (2000) 21(10):479-83. doi: 10.1016/S0167-5699(00) 01712-6

67. Parkin J, Cohen B. An overview of the immune system. Lancet (2001) 357 (9270):1777-89. doi: 10.1016/S0140-6736(00)04904-7

68. Cucchiari D, Podestà MA, Ponticelli C. The critical role of innate immunity in kidney transplantation. *Nephron* (2016) 132(3):227–37. doi: 10.1159/000444267

69. Lin J, Wang H, Liu C, Cheng A, Deng Q, Zhu H, et al. Dendritic cells: versatile players in renal transplantation. *Front Immunol* (2021) 12:654540. doi: 10.3389/fmmu.2021.654540

70. Zhao T, Li Y, Liu B, Pan B, Cheng X, Georgoff P, et al. Inhibition of histone deacetylase 6 restores innate immune cells in the bone marrow in a lethal septic model. *J Trauma Acute Care Surg* (2016) 80(1):34–40; discussion 40-1. doi: 10.1097/TA.00000000000897

71. Tang PM, Nikolic-Paterson DJ, Lan HY. Macrophages: versatile players in renal inflammation and fibrosis. *Nat Rev Nephrol* (2019) 15(3):144–58. doi: 10.1038/s41581-019-0110-2

72. Li J, Li C, Zhuang Q, Peng B, Zhu Y, Ye Q, et al. The evolving roles of macrophages in organ transplantation. J Immunol Res (2019) 2019:5763430. doi: 10.1155/2019/5763430

73. Yatim KM, Lakkis FG. A brief journey through the immune system. *Clin J Am Soc Nephrol* (2015) 10(7):1274–81. doi: 10.2215/CJN.10031014

74. Gill RG, Lin CM. Linking innate immunity and chronic antibody-mediated allograft rejection. *Curr Opin Organ Transplant* (2019) 24(6):694–8. doi: 10.1097/MOT.00000000000708

75. Valenzuela-Fernandez A, Cabrero JR, Serrador JM, Sanchez-Madrid F. HDAC6: a key regulator of cytoskeleton, cell migration and cell-cell interactions. *Trends Cell Biol* (2008) 18:291–7. doi: 10.7150/thno.25317

76. Maharaj K, Powers JJ, Ren J, Liao X, Vieson MD, Chen M, et al. Selective HDAC6 inhibition decreases early stage of lupus nephritis by down-regulating both innate and adaptive immune responses. *Clin Exp Immunol* (2018) 191(1):19–31. doi: 10.1111/cei.13046

77. Ochando J, Ordikhani F, Boros P, Jordan S. The innate immune response to allotransplants: mechanisms and therapeutic potentials. *Cell Mol Immunol* (2019) 16 (4):350–6. doi: 10.1038/s41423-019-0216-2

78. Savoye E, Legeai C, Branchereau J, Gay S, Riou B, Gaudez F, et al. Optimal donation of kidney transplants after controlled circulatory death. *Am J Transplant* (2021) 21(7):2424–36. doi: 10.1111/ajt.16425

79. Núñez-Andrade N, Iborra S, Trullo A, Moreno-Gonzalo O, Calvo E, Catalán E, et al. HDAC6 regulates the dynamics of lytic granules in cytotoxic T lymphocytes. *J Cell Sci* (2016) 129(7):1305–11. doi: 10.1242/jcs.180885

80. Yan B, Liu Y, Bai H, Chen M, Xie S, Li D, et al. HDAC6 regulates IL-17 expression in T lymphocytes: implications for HDAC6-targeted therapies. *Theranostics* (2017) 7:1002–09. doi: 10.3390/nano10010161

81. Do A, Reid RC, Lohman RJ, Sweet MJ, Fairlie DP, Iyer A. An HDAC6 inhibitor confers protection and selectively inhibits b-cell infiltration in DSS-induced colitis in mice. *J Pharmacol Exp Ther* (2017) 360(1):140–51. doi: 10.1124/jpet.116.236711

82. Nieuwenhuijs-Moeke GJ, Pischke SE, Berger SP, Sanders JSF, Pol RA, Struys MMRF, et al. Ischemia and reperfusion injury in kidney transplantation: relevant mechanisms in injury and repair. *J Clin Med* (2020) 9(1):253. doi: 10.3390/jcm9010253

83. Denecke C, Tullius SG. Innate and adaptive immune responses subsequent to ischemia-reperfusion injury in the kidney. *Prog Urol* (2014) 24 Suppl 1:S13–9. doi: 10.1016/S1166-7087(14)70058-2

84. Deng Q, Zhao T, Pan B, Dennahy IS, Duan X, Williams AM, et al. Protective effect of tubastatin a in CLP-induced lethal sepsis. *Inflammation* (2018) 41:2101–2109. doi: 10.1007/s10753-018-0853-0

85. Xiong B, Li M, Xiang S, Han L. A1AR-mediated renal protection against ischemia/reperfusion injury is dependent on HSP27 induction. *Int Urol Nephrol* (2018) 50(7):1355-63. doi: 10.1007/s11255-018-1797-x

86. Li Y, Zhao T, Liu B, Halaweish I, Mazitschek R, Duan X, et al. Inhibition of histone deacetylase 6 improves long-term survival in a lethal septic model. *J Trauma Acute Care Surg* (2015) 78(2):378–85. doi: 10.1097/TA.00000000000510

87. Li J, Zhao Y, Shi J, Ren Z, Chen F, Tang W. Histone deacetylase 6 interference protects mice against experimental stroke-induced brain injury *via* activating Nrf2/HO-1 pathway. *Anim Cells Syst (Seoul)* (2019) 23(3):192–9. doi: 10.1080/19768354.2019.1601132

88. Knox T, Sahakian E, Banik D, Hadley M, Palmer E, Noonepalle S, et al. Selective HDAC6 inhibitors improve anti-PD-1 immune checkpoint blockade therapy by decreasing the anti-inflammatory phenotype of macrophages and down-regulation of immunosuppressive proteins in tumor cells. *Sci Rep* (2019) 9(1):6136. doi: 10.1038/ s41598-019-42237-3

89. Yan B, Xie S, Liu Y, Liu W, Li D, Liu M, et al. Histone deacetylase 6 modulates macrophage infiltration during inflammation. *Theranostics* (2018) 8(11):2927-38. doi: 10.7150/thno.25317

90. Zhang F, Yu S, Chai Q, Wang J, Wu T, Liu R, et al. HDAC6 contributes to human resistance against mycobacterium tuberculosis infection *via* mediating innate immune responses. *FASEB J* (2021) 35(11):e22009. doi: 10.1096/fj.202100614R

91. Requião-Moura LR, Durão Junior Mde S, Matos AC, Pacheco-Silva A. Ischemia and reperfusion injury in renal transplantation: hemodynamic and immunological paradigms. *Einstein (Sao Paulo)* (2015) 13(1):129–35. doi: 10.1590/S1679-45082015RW3161

92. Rama I, Bruene B, Torras J, Koehl R, Cruzado JM, Bestard O, et al. Hypoxia stimulus: an adaptive immune response during dendritic cell maturation. *Kidney Int* (2008) 73(7):816–25. doi: 10.1038/sj.ki.5002792

93. Ledezma DK, Balakrishnan PB, Cano-Mejia J, Sweeney EE, Hadley M, Bollard CM, et al. Indocyanine green-nexturastat a-PLGA nanoparticles combine photothermal and epigenetic therapy for melanoma. *Nanomater (Basel)* (2020) 10 (1):161. doi: 10.3390/nano10010161

94. Cheng F, Lienlaf M, Wang HW, Perez-Villarroel P, Lee C, Woan K, et al. A novel role for histone deacetylase 6 in the regulation of the tolerogenic STAT3/IL-10 pathway in APCs. *J Immunol* (2014) 193(6):2850–62. doi: 10.4049/jimmunol.1302778

95. Tavares MT, Shen S, Knox T, Hadley M, Kutil Z, Bařinka C, et al. Synthesis and pharmacological evaluation of selective histone deacetylase 6 inhibitors in melanoma models. ACS Med Chem Lett (2017) 8(10):1031–6. doi: 10.1021/acsmedchemlett.7b00223

96. Zheng Y, Chen Y, Lu X, Weng Q, Dai G, Yu Y, et al. Inhibition of histone deacetylase 6 by tubastatin a attenuates the progress of osteoarthritis *via* improving mitochondrial function. *Am J Pathol* (2020) 190(12):2376–86. doi: 10.1016/j.ajpath.2020.08.013

97. Parira T, Figueroa G, Granado S, Napuri J, Castillo-Chabeco B, Nair M, et al. Trichostatin a shows transient protection from chronic alcohol-induced reactive oxygen species (ROS) production in human monocyte-derived dendritic cells. J Alcohol Drug Depend (2018) 6(4):316. doi: 10.4172/2329-6488.1000316

98. Lee JW, Lee SM, Chun J, Im JP, Seo SK, Ha N, et al. Novel histone deacetylase 6 inhibitor CKD-506 inhibits NF- $\kappa$ B signaling in intestinal epithelial cells and macrophages and ameliorates acute and chronic murine colitis. *Inflammation Bowel Dis* (2020) 26(6):852–62. doi: 10.1093/ibd/izz317

99. Youn GS, Lee KW, Choi SY, Park J. Overexpression of HDAC6 induces proinflammatory responses by regulating ROS-MAPK-NF-κB/AP-1 signaling pathways in macrophages. *Free Radic Biol Med* (2016) 97:14–23. doi: 10.1016/j.freeradbiomed.2016.05.014

100. Zhang WB, Yang F, Wang Y, Jiao FZ, Zhang HY, Wang LW, et al. Inhibition of HDAC6 attenuates LPS-induced inflammation in macrophages by regulating oxidative stress and suppressing the TLR4-MAPK/NF- $\kappa$ B pathways. *BioMed Pharmacother* (2019) 117:109166. doi: 10.1016/j.biopha.2019.109166

101. Namdar M, Perez G, Ngo L, Marks PA. Selective inhibition of histone deacetylase 6 (HDAC6) induces DNA damage and sensitizes transformed cells to anticancer agents. *Proc Natl Acad Sci USA* (2010) 107:20003-8. doi: 10.1016/j.jaci.2014.10.002

102. Wang J, Zhao L, Wei Z, Zhang X, Wang Y, Li F, et al. Inhibition of histone deacetylase reduces lipopolysaccharide-induced-inflammation in primary mammary epithelial cells by regulating ROS-NF- $\kappa$ B signaling pathways. *Int Immunopharmacol* (2018) 56:230–4. doi: 10.1016/j.intimp.2018.01.039

103. Chandran S, Feng S. Current status of tolerance in kidney transplantation. *Curr* Opin Nephrol Hypertens (2016) 25(6):591–601. doi: 10.1097/MNH.00000000000269

104. Humar A, Kerr S, Gillingham KJ, Matas AJ. Features of acute rejection that increase risk for chronic rejection. *Transplantation* (1999) 68(8):1200–3. doi: 10.1097/00007890-199910270-00023

105. Galliford J, Game DS. Modern renal transplantation: present challenges and future prospects. *Postgrad Med J* (2009) 85(1000):91–101. doi: 10.1136/pgmj.2008.070862

106. Radi ZA, Wynn TA. Opinion on immune tolerance therapeutic development. Toxicol Pathol (2020) 48(6):712–7. doi: 10.1177/0192623320948805

107. Cornell LD, Smith RN, Colvin RB. Kidney transplantation: mechanisms of rejection and acceptance. *Annu Rev Pathol* (2008) 3:189–220. doi: 10.1146/annurev.pathmechdis.3.121806.151508

108. Ellis JD, Neil DA, Inston NG, Jenkinson E, Drayson MT, Hampson P, et al. Inhibition of histone deacetylase 6 reveals a potent immunosuppressant effect in models of transplantation. *Transplantation* (2016) 100(8):1667–74. doi: 10.1097/ TP.00000000001208

109. Young JW, Merad M, Hart DN. Dendritic cells in transplantation and immune-based therapies. *Biol Blood Marrow Transplant* (2007) 13(1 Suppl 1):23–32. doi: 10.1016/j.bbmt.2006.10.023

110. Radomski N, Kägebein D, Liebler-Tenorio E, Karger A, Rufer E, Tews BA, et al. Mito-xenophagic killing of bacteria is coordinated by a metabolic switch in dendritic cells. *Sci Rep* (2017) 7(1):3923. doi: 10.1038/s41598-017-04142-5

111. Kim IK, Bedi DS, Denecke C, Ge X, Tullius SG. Impact of innate and adaptive immunity on rejection and tolerance. *Transplantation* (2008) 86(7):889–94. doi: 10.1097/TP.0b013e318186ac4a

112. Balzar S. Self-centered function of adaptive immunity in regulation of immune responses and in tolerance. J Immunol Res (2021) 2021:7507459. doi: 10.1155/2021/7507459

113. Cabrero JR, Serrador JM, Barreiro O, Mittelbrunn M, Naranjo-Suárez S, Martín-Cófreces N, et al. Lymphocyte chemotaxis is regulated by histone deacetylase 6, independently of its deacetylase activity. *Mol Biol Cell* (2006) 17:3435–3445. doi: 10.1126/scisignal.2002873

114. Tsuji G, Okiyama N, Villarroel VA, Katz SI. Histone deacetylase 6 inhibition impairs effector CD8 T-cell functions during skin inflammation. *J Allergy Clin Immunol* (2015) 135(5):1228–39. doi: 10.1016/j.jaci.2014.10.002

115. Laino AS, Betts BC, Veerapathran A, Dolgalev I, Sarnaik A, Quayle SN, et al. HDAC6 selective inhibition of melanoma patient T-cells augments anti-tumor characteristics. *J Immunother Cancer* (2019) 7(1):33. doi: 10.1097/TP.00000000000526

116. Aldana-Masangkay GI, Rodriguez-Gonzalez A, Lin T, Ikeda AK, Hsieh YT, Kim YM, et al. Tubacin suppresses proliferation and induces apoptosis of acute lymphoblastic leukemia cells. *Leukemia lymphoma* (2011) 52(8):1544–55. doi: 10.1128/MCB.05155-11

117. Bae D, Lee JY, Ha N, Park J, Baek J, Suh D, et al. CKD-506: a novel HDAC6selective inhibitor that exerts therapeutic effects in a rodent model of multiple sclerosis. *Sci Rep* (2021) 11(1):14466. doi: 10.1038/s41598-021-93232-6 118. Martin-Moreno PL, Tripathi S, Chandraker A. Regulatory T cells and kidney transplantation. *Clin J Am Soc Nephrol* (2018) 13(11):1760–4. doi: 10.2215/CJN.01750218

119. Lee JH, Kim HS, Jang SW, Lee GR. Histone deacetylase 6 plays an important role in TGF- $\beta$ -induced murine treg cell differentiation by regulating cell proliferation. Sci Rep (2022) 12(1):22550. doi: 10.1038/s41598-022-27230-7

120. Sakaguchi S, Mikami N, Wing JB, Tanaka A, Ichiyama K, Ohkura N. Regulatory T Cells and Human Disease. *Annu Rev Immunol* (2020) 38:541–566. doi: 10.1146/annurev-immunol-042718-041717

121. Romano M, Tung SL, Smyth LA, Lombardi G. Treg therapy in transplantation: a general overview. *Transpl Int* (2017) 30(8):745–53. doi: 10.1111/tri.12909

122. Giganti G, Atif M, Mohseni Y, Mastronicola D, Grageda N, Povoleri GA, et al. Treg cell therapy: how cell heterogeneity can make the difference. *Eur J Immunol* (2021) 51(1):39–55. doi: 10.1002/eji.201948131

123. Kalin JH, Butler KV, Akimova T, Hancock WW, Kozikowski AP. Secondgeneration histone deacetylase 6 inhibitors enhance the immunosuppressive effects of Foxp3 + T-regulatory cells. J Med Chem (2012) 55(2):639–51. doi: 10.1021/jm200773h

124. Doñas C, Fritz M, Manríquez V, Tejón G, Bono MR, Loyola A, et al. Trichostatin a promotes the generation and suppressive functions of regulatory T cells. *Clin Dev Immunol* (2013) 2013:679804. doi: 10.1155/2013/679804

125. Beier UH, Wang L, Han R, Akimova T, Liu Y, Hancock WW. Histone deacetylases 6 and 9 and sirtuin-1 control Foxp3+ regulatory T cell function through shared and isoform-specific mechanisms. *Sci Signal* (2012) 5(229):ra45. doi: 10.1126/scisignal.2002873

126. Xu K, Yang WY, Nanayakkara GK, Shao Y, Yang F, Hu W, et al. GATA3, HDAC6, and BCL6 regulate FOXP3+ treg plasticity and determine treg conversion into either novel antigen-presenting cell-like treg or Th1-treg. *Front Immunol* (2018) 9:45. doi: 10.3389/fimmu.2018.00045

127. Zhou W, Zhou X, Gaowa S, Meng Q, Zhan Z, Liu J, et al. The critical role of induced CD4+ FoxP3+ regulatory cells in suppression of interleukin-17 production and attenuation of mouse orthotopic lung allograft rejection. *Transplantation* (2015) 99:1356–64. doi: 10.1007/s13238-011-1114-2

128. de Zoeten EF, Wang L, Butler K, Beier UH, Akimova T, Sai H, et al. Histone deacetylase 6 and heat shock protein 90 control the functions of Foxp3+ T-regulatory cells. *Mol Cell Biol* (2011) 31:2066–78. doi: 10.3390/jcm6020013

129. Qiu W, Wang B, Gao Y, Tian Y, Tian M, Chen Y, et al. Targeting histone deacetylase 6 reprograms interleukin-17-Producing helper T cell pathogenicity and facilitates immunotherapies for hepatocellular carcinoma. *Hepatology* (2020) 71 (6):1967–87. doi: 10.1002/hep.30960

130. Yasuda K, Takeuchi Y, Hirota K. The pathogenicity of Th17 cells in autoimmune diseases. *Semin Immunopathol* (2019) 41(3):283-97. doi: 10.1007/s00281-019-00733-8

131. Zhang M, Xu M, Wang K, Li L, Zhao J. Effect of inhibition of the JAK2/STAT3 signaling pathway on the Th17/IL-17 axis in acute cellular rejection after heart transplantation in mice. *J Cardiovasc Pharmacol* (2021) 77(5):614–20. doi: 10.1097/FJC.00000000001007

132. Shilling RA, Wilkes DS. Role of Th17 cells and IL-17 in lung transplant rejection. *Semin Immunopathol* (2011) 33:129–34. doi: 10.1016/j.clim.2015.11.007

133. Valmori D, Raffin C, Raimbaud I, Ayyoub M. Human RORgammat+ TH17 cells preferentially differentiate from naive FOXP3+Treg in the presence of lineagespecific polarizing factors. *Proc Natl Acad Sci U S A* (2010) 107:19402–7. doi: 10.18632/ oncotarget.5167

134. Chang Q, Yin D, Li H, Du X, Wang Z, Liu Y, et al. HDAC6-specific inhibitor alleviates hashimoto's thyroiditis through inhibition of Th17 cell differentiation. *Mol Immunol* (2022), 149:39–47. doi: 10.1016/j.molimm.2022.05.004

135. Kimura A, Kishimoto T. IL-6: regulator of Treg/Th17 balance. Eur J Immunol (2010) 40:1830–5. doi: 10.1042/CS20160084

136. Zhou W, Yang J, Saren G, Zhao H, Cao K, Fu S, et al. HDAC6-specific inhibitor suppresses Th17 cell function *via* the HIF-1 $\alpha$  pathway in acute lung allograft rejection in mice. *Theranostics* (2020) 10(15):6790–805. doi: 10.7150/thno.44961

137. Semenza GL. Hypoxia-inducible factor 1 (HIF-1) pathway. *Sci STKE* (2007) 2007(407):cm8. doi: 10.1126/stke.4072007cm8

138. Bollinger T, Gies S, Naujoks J, Feldhoff L, Bollinger A, Solbach W, et al. HIF-1alpha- and hypoxia-dependent immune responses in human CD4+CD25high T cells and T helper 17 cells. J Leukoc Biol (2014) 96:305–12. doi: 10.1189/jlb.3A0813-426RR

139. Tsun A, Chen Z, Li B. Romance of the three kingdoms: RORgammat allies with HIF1alpha against FoxP3 in regulating T cell metabolism and differentiation. *Protein Cell* (2011) 2:778–81. doi: 10.1016/j.jss.2015.02.070

140. Ellis JS, Braley-Mullen H. Mechanisms by which b cells and regulatory T cells influence development of murine organ-specific autoimmune diseases. *J Clin Med* (2017) 6(2):13. doi: 10.3390/jcm6020013

141. Chong AS. B cells as antigen-presenting cells in transplantation rejection and tolerance. *Cell Immunol* (2020) 349:104061. doi: 10.1016/j.cellimm.2020.104061

142. Ren J, Catalina MD, Eden K, Liao X, Read KA, Luo X, et al. Selective histone deacetylase 6 inhibition normalizes b cell activation and germinal center formation in a model of systemic lupus erythematosus. *Front Immunol* (2019) 10:2512. doi: 10.3389/fimmu.2019.02512

143. Jia YJ, Liu ZB, Wang WG, Sun CB, Wei P, Yang YL, et al. HDAC6 regulates microRNA-27b that suppresses proliferation, promotes apoptosis and target MET in diffuse large b-cell lymphoma. *Leukemia* (2018) 32(3):703–11. doi: 10.1038/leu.2017.299

144. Ganguly S, Home T, Yacoub A, Kambhampati S, Shi H, Dandawate P, et al. Targeting HSF1 disrupts HSP90 chaperone function in chronic lymphocytic leukemia. *Oncotarget* (2015) 6(31):31767–79. doi: 10.18632/oncotarget.5167

145. Franzin R, Stasi A, Fiorentino M, Stallone G, Cantaluppi V, Gesualdo L, et al. Inflammaging and complement system: a link between acute kidney injury and chronic graft damage. *Front Immunol* (2020) 11:734. doi: 10.3389/fimmu.2020.00734

146. Batchu SN, Brijmohan AS, Advani A. The therapeutic hope for HDAC6 inhibitors in malignancy and chronic disease. *Clin Sci* (2016) 130:987–1003. doi: 10.1016/bs.mie.2019.06.028

147. Zheng K, Jiang Y, He Z, Kitazato K, Wang Y. Cellular defence or viral assist: the dilemma of HDAC6. J Gen Virol (2017) 98(3):322–37. doi: 10.1099/jgv.0.000679

148. Youn GS, Park JK, Lee CY, Jang JH, Yun SH, Kwon HY, et al. MicroRNA-22 negatively regulates LPS-induced inflammatory responses by targeting HDAC6 in macrophages. *BMB Rep* (2020) 53(4):223–8. doi: 10.5483/BMBRep.2020.53.4.209

149. Magupalli VG, Negro R, Tian Y, Hauenstein AV, Di Caprio G, Skillern W, et al. HDAC6 mediates an aggresome-like mechanism for NLRP3 and pyrin inflammasome activation. *Science* (2020) 369(6510):eaas8995. doi: 10.1126/science.aas8995

150. Cheng X, Liu Z, Liu B, Zhao T, Li Y, Alam HB. Selective histone deacetylase 6 inhibition prolongs survival in a lethal two-hit model. *J Surg Res* (2015) 197(1):39–44. doi: 10.1016/j.jss.2015.02.070

151. Xu S, Chen H, Ni H, Dai Q. Targeting HDAC6 attenuates nicotine-induced macrophage pyroptosis *via* NF-κB/NLRP3 pathway. *Atherosclerosis* (2021) 317:1–9. doi: 10.1016/j.atherosclerosis.2020.11.021

152. Hao Y, Guo F, Huang Z, Feng Y, Xia Z, Liu J, et al. 2-methylquinazoline derivative 23BB as a highly selective histone deacetylase 6 inhibitor alleviated cisplatininduced acute kidney injury. *Biosci Rep* (2020) 40(1):BSR20191538. doi: 10.1042/ BSR20191538

153. Gu YY, Liu XS, Huang XR, Yu XQ, Lan HY. TGF- $\beta$  in renal fibrosis: triumphs and challenges. Future Med Chem (2020) 12(9):853–66. doi: 10.4155/fmc-2020-0005

154. Saito S, Zhuang Y, Shan B, Danchuk S, Luo F, Korfei M, et al. Tubastatin ameliorates pulmonary fibrosis by targeting the TGF $\beta$ -PI3K-Akt pathway. *PloS One* (2017) 12(10):e0186615. doi: 10.1371/journal.pone.0186615

155. Benoy Vanden BP, Jarpe M, VanDamme Robberecht Van DenBosch L. Development of improved HDAC6 inhibitors as pharmacological therapy for axonalcharcot-marie-tooth disease. *Neurotherapeutics* (2017) 14(2):417\_1428. doi: 10.1021/jm301098e

156. Osko JD, Christianson DW. Methods for the expression, purification, and crystallization of histone deacetylase 6-inhibitor complexes. *Methods Enzymol* (2019) 626:447–74. doi: 10.1016/bs.mie.2019.06.028

157. Bertos NR, Gilquin B, Chan GK, Yen TJ, Khochbin S, Yang XJ. Role of the tetradecapeptide repeat domain of human histone deacetylase 6 in cytoplasmic retention. *J Biol Chem* (2004) 279:48246-54. doi: 10.1016/j.neuint.2016.08.004

158. Vergani B, Sandrone G, Marchini M, Ripamonti C, Cellupica E, Galbiati E, et al. Novel benzohydroxamate-based potent and selective histone deacetylase 6 (HDAC6) inhibitors bearing a pentaheterocyclic scaffold: design, synthesis, and biological evaluation. *J Med Chem* (2019) 62(23):10711-39. doi: 10.1021/asc.jmedchem.9b01194