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Self-assembled nanoparticles: A new platform for revolutionizing therapeutic cancer vaccines

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Cancer vaccines have had some success in the past decade. Based on in-depth analysis of tumor antigen genomics, many therapeutic vaccines have already entered clinical trials for multiple cancers, including melanoma, lung cancer, and head and neck squamous cell carcinoma, which have demonstrated impressive tumor immunogenicity and antitumor activity. Recently, vaccines based on selfassembled nanoparticles are being actively developed as cancer treatment, and their feasibility has been confirmed in both mice and humans. In this review, we summarize recent therapeutic cancer vaccines based on self-assembled nanoparticles. We describe the basic ingredients for self-assembled nanoparticles, and how they enhance vaccine immunogenicity. We also discuss the novel design method for self-assembled nanoparticles that pose as a promising delivery platform for cancer vaccines, and the potential in combination with multiple therapeutic approaches.

KEYWORDS

cancer vaccines, nanoparticles, adjuvant, self-assembled, immunotherapies

1 Introduction

Cancer vaccines, typically given through exogenous administration of selected tumor antigens combined with adjuvants that activate dendritic cells (DCs), ultimately aim to stimulate the immune system against tumor antigens. In recent years, the success of therapeutic cancer vaccines has dramatically revolutionized cancer treatments and has provided more alternative approaches to patients with cancer (1, 2). Several clinical trials have also confirmed the potential of this strategy, including those for melanoma, lung cancer, and prostate cancer (3-5). With the discovery of neoantigen target being continually advanced to improve the identification of immunogenic neoepitopes that can be recognized by CD8+ T cells, personalized therapeutic cancer vaccines have been considered in clinical investigation (6). The potential of therapeutic cancer vaccines in tumor immunotherapy has led to growing interest in developing more effective vaccine strategies to achieve tumor regression. However, recent years have seen excellent advances in selfassembled nanoparticle (SANP) platforms for therapeutic cancer vaccines. SANPs can evoke the epitope-specific B/T-cell immune responses through epitope folding similar to microorganisms that can protect payloads from enzymatic degradation, deliver to lymphoid organs, and target the tumor (7, 8). Additionally, antigen loading is also increased with the improved performance of SANPs. Because of the advantages from SANPs, they have naturally become the focus of current vaccine research and have appeared in various medical fields.

As early as the 1950s, rod-shaped particles have been found in tobacco mosaic virus (TMV); this is one of the earliest discoveries in history about self-assembled particles (9). In 1981, Drexler proposed the molecular engineering of proteins, being the monomeric building blocks, through self-assembly (10). Since then, materials based on self-assembled peptides such as nanofibers have been produced (11). Indeed, with the development of nanoparticle (NP) technology, including poly(lactic-coglycolic acid) (PLGA) (12), polymersomes (13), and liposomes (14), the design and manufacture of selfassembled nanovaccines are an empirical process. SANPs are composed of multi-component homologous NPs through noncovalent bonds or weak covalent bond interactions aimed at achieving a stable and balanced state (15). The assembly of different or similar molecules driven by intermolecular forces, including van der Waals interactions, electrostatic interactions, hydrogen bonding, coordination, hydration forces, and solvation interactions, occurs if there is equilibrium between them (11, 16). While these forces are indeed weak when considered individually, mixtures of different interaction types can result in structural and chemical stabilization, and these conformations formed by the free assembly of molecules are easy to accept. Of course, the modified module can also be selected as the substrate, which can control the direction of self-assembly to a certain extent and can directly act between the molecules or the module by external stimuli, including pH, temperature, solvent polarity, electromagnetic radiation, and light, to guide self-assembly (17). NPs designed through synthetic self-assembled technology (SANPs) can be given certain physiological functions. They can be used as platforms to display the arrangement of specific immunogens and orderly matrices to mimic the folding and complex structure of natural microbial surfaces (18). Therefore, SANPs can offer potential advantages such as improved stability due to the co-delivery of an antigen with adjuvant, and constant release and/or specific activation of the immune system (19). To a certain extent, we believe that the limitations of clinical application caused by the defects of the conventional vaccine platforms can be alleviated by SANPs.

The development of SANPs has led to many vaccine platforms and drug delivery systems (17), including broad platforms such as peptides/polymers (20). Generally, antigen loading is chemically defined, and SANPs based on peptide antigens are linked to hydrophobic carriers like peptide, lipids, and polymers. Of course, we also introduce a new way of antigen loading here. In this review, we discuss the characteristics of the internal structure of vaccines based on various self-assembled NPs, such as polymers and lipids for inducing anticancer T-cell immunity, and some SANP-based delivery platforms. Additionally, we also discuss current challenges of therapeutic cancer vaccines and their potential applications when combined with other treatments for overcoming tumor resistance and promoting clinical efficacy.

2 The design of SANPs

There are two ways to achieve nanotechnology: top-down and bottom-up (21). The top-down method dissociates NPs from larger structures while maintaining their original composition and properties; the bottom-up method uses assembly or self-assembly to design NPs at the basic molecular level. To design an appropriate antigen carrier that delivers the screened pathogen epitopes and can induce a corresponding response in animal immunity studies model (22, 23). For example, in the PGS-co-PEG NPs system, Ankur and his colleagues synthesized the PGS-co-PEG polymer through a two- step polycondensation reaction (24). The first step, polycondensation reaction of PEG and sebacic acid under stirring condition. In the second step, glycerol was added to obtain a block copolymer of PGSco-PEG (pre-polymer) with different ratio of PEG segments.. The size of NPs can be controlled by controlling the molar ratio of PEG, thereby synthesizing precise nanostructures. To a certain extent, the size of NPs determines the binding and activation of their membrane proteins and receptors (25). Different immune cells have their corresponding SANPs from different dimensions. SANPs of size > 100 nm are mostly absorbed by local antigen-presenting cells (APCs) at the site of injection, as well as tend to accumulate in highly permeable tumor tissue, when employed in prophylactic and therapeutic immunotherapy (26). In contrast, smaller nanostructures (40-50 nm) tend to enter lymphatic vessels and reach lymph nodes (LNs); they can be internalized with greater efficiency by LN-resident DCs, triggering an effective and long-lasting immune response against tumor (26, 27) (Figure 1C). The peptide self-assembly process and the size of its production are strongly influenced by amino acid sequences and physicochemical indicators, such as pH (from acidic to slightly alkaline), temperature (4-95°C), metal ion concentrations (such as calcium, potassium, magnesium and sodium), and salt concentrations (28). SANPs can become structurally stable if the environmental parameters are obtained (26). The self-assembled form can be achieved by adjusting the concentration of metal ions. Changing the MgCl₂ concentration, for example, could encourage carbonic anhydrase (BCA) linked with P114 peptide (BCA-P114) NPs to self-assemble (26). NPs can be obtained by using MgCl₂ and Tris-HCl as low as 5 mM and 10 mM at neutral pH. The maximum size of these NPs can be formed when MgCl₂ concentration is 25 mM. When the concentration of MgCl₂ exceeds 25 mM, the particle size decreases (28). In another study (29), a self-assembled nanovaccine based on synthetic peptide conjugation of IKVAV-PA and OVA shows that different concentrations have an effect on vaccine characteristics including size, zeta potential, and encapsulation efficacy. Therefore, optimization of the formulation that is used for chemical reaction is a critical step for producing the best-performing SANP.

To summarize, one of the important functions of SANPs as tumor antigen carriers is to effectively deliver tumor antigens to LNs



FIGURE 1

Strategies of SANPs: (A) SANPs based on multiple materials. (B) During the synthesis of SANPs, antigens or other immune activators can be loaded onto SANPs by chemical modification or physical packaging. Nucleic acids and antigens can be physically embedded in liposome and polymer SANPs; bacterial membrane can wrap around SANPs' surface; virus glycoprotein and peptides are loaded into viral particles or peptide chains through autonomous synthesis, as well as antigens. (C) SANP-based nanovaccines targeting SANPs were injected intravenously into the bloodstream and circulate throughout the body to activate dendritic cells and recruit T cells around lymph nodes and tumor tissue. (D) Tumor-specific T-cell induction: SANPs can induce cytotoxic T lymphocytes by activating peripheral dendritic cells (DCs) that then present antigens to T cells in lymph nodes, or SANPs can directly target lymph nodes for induction within lymph nodes. Nanovaccines deliver antigens and adjuvants to DCs, which subsequently upregulate co-stimulatory molecules and present the processed antigen to T cells *via* MHC-II molecules. DCs can directly activate T cells y MHC-bound antigens and costimulatory molecules present on cell-derived membranes on the surface of SANPs.

and then induce durable and effective specific-tumor immunity and immune memory. Aside from the fact that the encapsulation process protects antigens from the effect of phagocytosis or protease degradation, facilitates longer circulation time, and can generate effective immune responses (30–32), modularization with surface-attached antigens often elicit superior immune responses in comparison to encapsulated antigens perhaps owing to intracellular processing (33). Thus, composition, size, shape, and surface characteristics are key factors in ensuring stable delivery and inducing an immune response. Better biocompatible materials need to be selected to prevent their rapid removal. Particle size, as well as surface area (related to the load of the antigen), plays a critical role in DC phagocytosis and immune induction, which can be determined by controlling reaction conditions. During the design, attention should be paid to the control of chemical reaction conditions, while avoiding the infiltration of impurities, which may cause toxicity or damage the stability of the structure.

3 Several SANPs for vaccine design

We can classify SANPs for vaccine design from multiple perspectives. From a general point of view, we can classify them according to the method of self-assembly: dynamic self-assembly and static self-assembly (15). According to the constituent elements, SANPs can be divided into protein-based biological entities and non-protein-derived synthetic particles (23). In this review, we briefly introduce several vaccine designs based on the different components of SANPs (Figures 1A, B; Table 1).

3.1 Polymer-based SANPs

Polymer NPs are one of the earliest NPs used as drug delivery systems. They are often used as a material for packaging drugs like nanocapsules and nanospheres because they are non-toxic and have good biocompatibility. They can also be biodegraded into harmless alcohols and other low-molecular-weight products (21, 56). Polymers that have been developed for drug delivery can be classified into synthetic polymers and natural polymers. Common synthetic polymer NPs include polylactic acid (PLA), chitosan, polylactide glycolide acid (PLGA), polyglutamic acid (PGA), and polyethylene glycol (PEG), which are all typical polymers (11). Natural polymers such as albumin have been used to carry paclitaxel and can be used to treat metastatic cancer in the clinic (21, 57). The variety and adaptability of polymer carriers increase their therapeutic efficacy by controlling drug release, long cycle characteristics, tissue or cell targeting, and stimuli responsiveness (58).

Chemical modification of the polymer system helps to prolong the retention time in the blood, reduce widespread distribution, and target tissue or specific cell surface antigens with targeted ligands (peptides, aptamers, antibodies/antibody fragments, and small molecules). NPs based on PLGA are currently one of the most widely studied adjuvants for the effective delivery of antigens, and PLGA is a biodegradable copolymer composed of lactic acid and glycolic acid linked through ester bonds (58). DSPE-PEG2000-Gal was synthesized in an NHS-ester crosslinking reaction, which can be modified to the surface of PLGA to combine the advantages of biodegradable polymeric NPs and biomimetic liposomes, by using a water-in-oil-in-water (W/O/W) double-emulsion solvent evaporation method. PLGA polyester can be degraded from

TABLE 1 Summary of SANPs that have been explored as delivery platforms for vaccines.

NP type	Pathogen/disease	Antigen/adjuvant	Animal model	Reference
Polymer	Cancer	OVA	BALB/c mice	(34)
	HPV-related cancers	human papillomavirus (HPV) E7 protein-derived peptide	Mice	(35)
	Cancer	Ovalbumin	Female C57BL/6 mice	(36)
	Cancer	MUC1	Mice	(37)
	Cancer	R837	BALB/c mice	(38)
	Cancer	TLR7/8	C57BL/6 mice	(39)
VLPs	Breast cancer (BC)	Aberrantly glycosylated mucin-1	Mice	(40)
	Breast cancer	HER2 ECO	Mice	(41)
	Breast cancer	IGF-1R	Mice	(42)
	Cancer	Influenza virus, hemagglutinin, neuraminidase	BALB/c mice	(36)
	Cancer	HPV16 E6	C57BL/6 mice	(43)
	Pancreatic cancer	hMSLN	C57BL/6 mice	(44)
	Pancreatic cancer	mMSLN	C57BL/6 mice	(45)
	Melanoma	Gp100	C57BL/6 mice	(46)
	Melanoma	LCMV-TT830-843	C57BL/6 mice	(47)
Self-assembled peptides	Cancer	Antigen peptide SL8 (SIIN-FEKL), O antigen polysaccharides (OPS)	BALB/c mice, Cynomolgus monkeys	(36)
	HER2+ cancer	HER2	Mice	(48)
	Cervical cancer	HPV-L2	Mice	(49)
	Glioblastoma	EGFRvIII, PADRE, OVA	Mice	(50)
Liposomes	Cancer	_	Murine melanoma model	(51)
	Cancer	HER-2/neu-P5	BALB/c mice	(52)
	Cancer	Mycolic acid	C57BL/6 mice	(53)
	Pancreatic tumor	CpG-DNA	C57BL/6 mice	(54)
	Melanoma	OVA RNA	C57BL/6	(55)

certain substances *in vivo* into lactic acid and glycolic acid so that efficiently delivered the SLI the internal immunogen can be released continuously; in this way, SIINFEKL-specific CD8⁺ cells are stimulated multiple times to avoid the need for multiple

the internal immunogen can be released continuously; in this way, cells are stimulated multiple times to avoid the need for multiple vaccinations (59, 60). For example, Po et al. designed OVA antigenloaded PLGA NPs, glycosylated poly(lactic-co-glycolic acid) NPs loaded with the SIINFEKL peptide (OVA) as a tumor-specific antigen, and CpG oligodeoxynucleotide (CpG) as an adjuvant for an effective DC-targeted cancer vaccine (61). The biodistribution and antitumor efficiency of the SANP vaccine had a longer retention time compared with other groups. Geoffrey and his colleagues linked TLR-7/8a to HPMA-based polymers particles (Poly-7/8a) with high agonist density (8–10 mol% TLR-7/8a) and induced substantially higher LN cytokine production [which led to a ~400-fold higher level of TLR-7/8a (AUC) in draining LNs] compared with unformulated small-molecule TLR-7/8a (SM 7/ 8a) (62).

3.2 Lipid-based SANPs

The versatility and plasticity of a liposome-based delivery system are the main reasons why liposomes can act as vehicles to directly deliver drugs to cancer cells (63, 64). Liposomes are natural or artificial biodegradable phospholipid bilayer vesicles surrounded by an amphiphilic lipid bilayer with an internal water core (14, 65). It has been proven that liposomes have a good lifespan in the blood, allowing them to accumulate in lesions where blood vessels are damaged (65). This is conducive to the specific targeting of the antigen or cargo attached to the surface of the carrier to the disease site, resulting in a powerful therapeutic effect. In vaccine designs, antigenic peptide determinants can be encapsulated in lipid bilayers, and the immunogenic peptides can be protected by lipid bilayers and not degraded by enzymes (60). In addition to being a delivery agent, it also acts as a potent immune adjuvant to induce the innate immunity of the immune system (66).

One of the most classic cationic liposomes is 1,2-dioleoyl-3trimethylammonium-propane (DOTAP) (67). It is a cationic monounsaturated phospholipid that has been found to activate DCs, thereby promoting endocytosis and antigen delivery in APCs, leading to MAP kinase, ERK activation, and induction of chemokines (68). Reactive oxygen species (ROS) were induced by cationic DOTAP liposomes from mouse bone marrow DCs (BMDCs), and ROS were involved in the expression of DOTAPinduced costimulatory molecule CD86/CD80, indicating the antitumor activity of ROS for vaccine induction (69). Recently, cationic liposomes have been shown by several studies to be promising vaccine delivery platforms for therapeutic cancer vaccines because of their ability to increase the immunogenicity of antigen-based vaccines (70-72). The liposome-based synthetic long peptide (SLP) vaccine has been shown to efficiently induce functional antigen-specific CD8+ and CD4+ T cells and to effectively eliminate tumor in two different tumor-bearing mouse models (71, 72). Heuts et al. investigated cationic liposomes loaded with 15 SIINFEKL T-cell epitope-containing SLPs; cationic liposomes

efficiently delivered the SLPs to DCs that subsequently activated SIINFEKL-specific CD8⁺ T cells, indicating the improved immunological activity of the SLPs (73). In addition to vaccines based on peptide antigens, liposomes have also been used to deliver RNA-based vaccines (74). Jinjin et al. developed an LN targeting lipid (113-O12B), which was used to deliver an OVA-encoding mRNA cancer vaccine against a melanoma mouse model. In addition to exhibiting better expression in LNs compared with ALC-0315 (a synthetic lipid used in the COVID-19 vaccine Comirnaty), the vaccine also promoted mRNA expression in APCs, but the potential hepatotoxicity needs to be verified (75). Matthias et al. found that incorporation of different adjuvants (LPS) will improve the potency of the lipid-based SANP mRNA vaccine, resulting in B16 F10 melanoma tumor shrinkage and extended survival of the tumor-bearing mouse (76).

3.3 Peptides-based SANPs

Most of the vaccines based on biological materials have been extensively studied. Peptide-based vaccines can cause a more concentrated immune response over protein sequences or inactivated viruses (77). The primary structure and site modification of peptide nanomaterials are based on amino acid design (78), which can be designed into self-assembled peptide nanostructures with unique secondary structures (79). The distribution of charges around amino acids will affect the performance of the nanoscaffold and the speed of self-assembly (80, 81). Compared to other types of vaccine platforms, selfassembled peptides are less toxic than polymer-based SANPs (82), and they are generally considered to have higher stability than lipidbased SANPs (83). The self-assembled supramolecular structures can be perfect modules for constructing nanofibers, nanotubes, nanocapsules, and nanomicelles (84). Self-assembled peptides can be coupled with peptide epitopes without affecting their respective characteristics; finally, immunogenic, biocompatible, stable, and selfadjuvanted vaccines can be designed. In recent years, due to their unique chemical, physical, and biological properties, self-assembled peptides have received increasing attention from scientists. Geoffrey et al. developed a vaccine platform (SNP-7/8a) that is chemically programmed to self-assemble into homogeneously sized (~20 nm) NPs, based on charge-modified peptide-TLR-7/8a conjugates (85), which targets patient-specific neoantigens' physicochemical variability and presents challenges in manufacturing personalized optimal cancer vaccines to induce anticancer T cells. This approach promotes T-cell immunity by promoting the uptake and activation of APCs through the precise loading of multiple peptide neoantigens linked to TLR-7/8a (adjuvant) in NPs. Wei et al. developed a selfassembled peptide system as a novel adjuvant that can efficiently deliver two antigens (MAGE/NY-ESO-1) (86). The SANPs not only widened the response range within the same molecule but also significantly prolonged the plasma half-life of single antigenic peptides, which can be a broad-spectrum candidate for effective breast cancer therapy.

3.4 Virus-like particles and SANPs

Similar to self-assembling peptides, virus-like particles (VLPs) are a complex structure composed of viral structural proteins that have the ability to self-assemble when they are recombined and expressed (87, 88). The main difference between VLPs and native viruses is that they lack viral genetic material, so they cannot replicate or induce infection (89). Therefore, VLPs are one of the safest candidates for attenuated or inactivated pathogen-based vaccines. VLP epitopes have repetitiveness and high density, which is the main reason why they can cause a strong immune response (90). This also shows that it has the potential for effective recognition, cell absorption, and processing of the host immune system (91). Its granularity further consolidates this advantage, and its proper size (less than 40 nm) is particularly suitable for obtaining DCs (92).

In the past 10 years, several VLP-based preventive vaccines have been approved for marketing, including human papillomavirus (HPV) and hepatitis B virus (HBV). VLPs based on HPV are formed by a single structural protein, and there are also more complex VLPs, such as the VLPs of the Reoviridae family composed of two to four different proteins (93, 94). When selecting a VLP production platform, it is necessary to fully consider the modification requirements of the protein and select an appropriate expression system. Common production platforms include Escherichia coli, yeast, insect cells, and mammalian cells (87). Consider safety or production times and costs, plant-derived viruses such as cowpea mosaic virus (CPMV), tobacco mosaic virus, and potato virus X (PVX) were successfully used as a carrier/ platform to present foreign epitopes (95, 96). Sourabh et al. developed CPMV and PVX as vaccine platforms against HER2⁺ malignancies (48). CPMV generated a stronger selective induction of cytokines and chemokines than empty CPMV, a VLP composed of CPMV capsid without nucleic acids, in naïve mouse splenocytes (95). Valeria et al. evaluated the function of bacteriophage MS2 VLP, which was used to display an extracellular loop of xCT protein. In a metastatic breast cancer model, the VLP-based vaccine reduces tumor metastasis by eliciting a strong antibody response and is well-tolerated (97).

3.5 Other SANPs

Polymeric lipid hybrid nanoparticles (PLHNs) are an emerging carrier platform, which can overcome the limitations of liposomes and polymeric NPs. The biodegradable polymeric matrix core has a larger surface area and more stability than liposomes, as well as a controlled release, with the lipid layer surrounding the polymeric core being a highly biocompatible shell (98). Although this self-assembly method can obtain an optimal conformation, the gap between synthetic immunotherapy and endogenous immunity has been the reason for imprecise and inefficient treatment. The strategy of using natural biomembranes to camouflage NPs to mimic the properties and functions of biological interfaces has been widely explored. For example,

Chen and colleagues developed a cancer vaccine based on PLGA and a hybrid membrane, which was from the fusion of E. coli cytoplasmic membranes and an autologous tumor membrane (99). The hybrid membrane NPs can induce DC maturation, eliciting strong tumor-specific immune responses. Wang et al. developed a bacterial outer-membrane vesicle-cancer cell hybrid membrane and successfully tested it on HPDA NPs. The immune activation properties derived from the source membrane can rapidly stimulate DC maturation in LNs (100). Existing selfassembled nanovaccines that can be biosynthesized in vivo have been investigated. Pan and colleagues developed a Nano-B5 platform that can be synthesized in vivo and is a biosynthetic self-assembled nanovaccine based on a whole protein (101). They used genetic engineering technology to express a fusion module monomer in the periplasmic space of *E. coli* DH5 α cells, in which the protein could be folded correctly. The monomer consists of B5 subunits of cholera toxin (CTB) as an adjuvant internal module, and a C-terminal trimer peptide is linked through a connecting peptide (GGSG). The antigen part is made up of O antigen polysaccharides (OPS) formed by linking the pentameric sugar unit oligosaccharide substrate produced by the cell. They can selfassemble into NPs with a size of approximately 25-50 nm in the cytoplasm without affecting their unique properties.

4 The self-adjuvanticity of SANPs

The regulatory mechanism of adjuvants is related to the direct or indirect stimulation of APCs (60). The maturation of DCs stimulated by adjuvants and the immune response to host antigens are two major prerequisites for therapeutic cancer vaccines to work (102) (Figure 1D). DCs are an important messenger between CD4+ T helper cells and CD8+ T cells, and are associated with triggering further adaptive immune responses (103). The mechanistic routes of adjuvants were summarized by Schijns (1): adjuvants facilitate antigen uptake, presence, and transport by antigen-capturing and -processing cells; (2) adjuvants prolong the time of antigen storage and release; (3) adjuvants activate innate immune cells to release cytokines through targeting the pattern recognition receptor (PRR); and (4) adjuvants mimic danger signals to stimulate the APCs (104). Tolllike receptors (TLRs) are a popular target for adjuvant research, because of their involvement in identifying pathogen-associated molecular patterns (PAMPs) (60). In the process of the adaptive immune response, adjuvants can fully stimulate CD4+ T cells, but the initiation of CD8+ T cells requires more complex immunological activities (60).

Conventional adjuvants like aluminum salt have a certain effect on inducing T helper 2 (Th2) cell responses, not Th1 or cytotoxic T lymphocytes. Therefore, it is assumed that this type of adjuvant would fail to activate the immune response required to cause tumor killing (105). Live-attenuated viral vaccines generally do not need an adjuvant because they can mimic a natural infection. Therefore, virus-like NPs can be regarded as an exogenous antigen to be presented by MHC class II molecules, which can also combine with

MHC class I molecules by cross-presentation to activate humoral and cellular immunity (106). In general, self-adjuvanticity of SANPs relies on the formulation characteristics of the particulate's structure. Because SANPs have a bio-particulate structure that is usually taken up efficiently by DCs, these SANPs with a perfect phenotype are often preferred as adjuvants of cancer vaccines and have been applied in multiple immunotherapy programs involving vaccines. A variety of SANPs have been reported to induce powerful antigen-specific T-cell responses through targeting TLRs. Yoshikawa et al. demonstrated that poly-y-glutamic acid could activate APCs and induce a potent antigen-specific T-cell response through the TLR4 and MyD88-dependent signaling pathway (107). Mesa et al. found that TLR4 on DCs can be activated by small-sized proteoliposomes (VSSP), which are produced by the hydrophobic interaction of GM3 ganglioside with the meningococcal outer-membrane protein complex (108). Additionally, Luo et al. developed heterocyclic lipid SANPs with self-adjuvanticity that could activate type I interferon-stimulated genes by inducing STING activation but not the TLR or MAVS pathway (109). The potent T-cell activation with checkpoint inhibition showed great synergy with 100% survival over 60 days in a TC-1 tumor model. Furthermore, maintaining safety and reducing the vaccine dose are important factors for cancer patients who can benefit from the advancement of SANP technology.

Notably, the potential of SANPs can act both as delivery systems for vaccine antigens and as immunomodulators as previously reported (110). Several in situ vaccine (ISV) approaches have been suggested, with vaccine regimens including SANPs, Flt3L, and TLR agonists (111-113). However, the development of nanomedicine has fully demonstrated the advantages of SANP with both physiological activity and physical properties. A semiconducting polymer nano-immunomodulator (SPNI) has been previously reported (114), which is self-assembled by a polymer conjugated with a TLR7 agonist via an acid-labile linker. The semiconducting polymer contributed to tumor eradication and immunogenic cancer cell death through a near-infrared (NIR) absorbing to exert photodynamic effects. This way, the synergistic action of released immunogenic factors and acidic TME-activated TLR7 agonist can not only kill tumor cells directly but also serve as an in situ generated cancer vaccine to evoke strong antitumor activities. This SANP-based design refreshes the acquired route and presentation of traditional vaccines and improved therapeutic cancer vaccines.

5 Self-assembled nanoparticle platforms are a promising strategy in future cancer therapy

Reports in recent years have shown us a variety of strategies that try to improve the immunogenicity of vaccines by using the powerful immune stimulation of SANPs in the tumor microenvironment. However, the great obstacles to the production of the specific killing effect of cancer vaccines include the genomic and phenotypic heterogeneity of cancer and the immunosuppressive effect of the tumor microenvironment. Furthermore, intertumoral and intratumoral heterogeneity provided a basis for therapeutic resistance (115). The advent of personalized therapeutic cancer vaccines provides a strategy by targeting personal tumor antigens (neoantigen), and the success of the neoantigen platform was achieved by tumor mutational burden (TMB), which is the total number of mutations per coding area of a tumor genome (116, 117). Specifically, the clinical significance from immune checkpoint inhibitor therapy has also been shown to correlate with the higher TMB, within several tumors including NSCLC, small cell lung cancer, melanoma, and colorectal cancer (118–123). It is reasonable to assume that high TMB induces high densities of neoantigen-specific tumor-infiltrating lymphocytes, leading to tumor cell secretion of IFN- γ and upregulation of PD-L1, while their relationship across the entire human cancer spectrum remains unclear (124, 125). If the combination of the SANP platform and a neoantigen is feasible, then it will bring unexpected results. However, considerable challenges to this approach include the cost and time required to achieving this high degree of personalization.

5.1 SANPs combined with other treatments

In established cancers, therapeutic vaccines will require cotreatments to overcome immune evasion and to become fully effective. However, in clinical practical applications, these drugs have failed to eliminate immunosuppressive cells (126-130) and may have inadvertently created immune evasion of tumor cells in the tumor microenvironment (131-134), resulting in suboptimal therapeutic cancer vaccine efficacy. Nanotechnology interventions have overcome the limitations of current conventional chemotherapy, including poor biological distribution, cancer cell resistance, and severe systemic side effects, dramatically changing the treatment of cancer. The properties of these delivery systems have been adjusted to enhance the stability of delivery to tumors; for example, the hydrophilicity of NP surfaces provides longer cycle times through stealth, positively charged surfaces that can enhance the internalization of cancer cells. Delivery systems based on SANPs are commonly used to deliver anticancer drugs, gene drugs, targeted drugs, and stimulus-responsive drugs (26). Peptide self-assembled fibers are popular materials for anti-cancer drug delivery systems. Compared with traditional cancer treatment, peptide hydrogels can slowly and directly release chemotherapeutic drugs to cancer tissues. For example, Li and colleagues successfully developed NPs through the self-assembly of hyaluronic acid (HA)-cystaminecholesterol (HSC) conjugates, in which IR780 was simultaneously incorporated (HSCI NPs) (135). After cellular uptake, HSCI NPs are broken down by the reaction of cystamine with overexpressed GSH. The released IR780 will induce a fluorescent "on" transition,

which can be used to effectively image the tumor site. After irradiation with an 808-nm laser, programmed photoactive therapy (PPAT) can be realized, and the ROS generated therein will generate photodynamic therapy (PDT).

One of the main advantages of NP-based delivery systems is that multiple drugs with similar or synergistic effects can be encapsulated in a single nanoformulation (136). However, NPs that enter the systemic circulation by intravenous injection usually absorb the majority of macrophages through the mechanism of phagocytosis (137). To avoid similar recognition, the stealth substance PEG can be wrapped in the outer layer of the NP to improve the efficiency of drug delivery to target cells and tissues (138). However, people are worried about the production of PEG antibodies. Of course, this can be solved by using other substances instead of PEG, such as zwitterionic polymers (139). Akhilesh et al. synthesized PGS and PEG copolymer elliptical NP PGS-co-PEG by chemical synthesis based on the elastic properties and biocompatibility of polyglycerol sebacate (PGS) (13). Subsequently, bovine serum albumin was used as a model protein to encapsulate and self-assemble using the nanoprecipitation method. The encapsulation rates of PGS-20PEG and PGS-40PEG were 88.5% and 91%, respectively. As the outer surface of NPs, PEG provides good stability and stealth properties, which play an important role in improving pharmacokinetics, drug delivery efficiency, and drug targeting.

5.2 The prospect of combination immunotherapy

5.2.1 PD-1 blockers

The common goal of cancer vaccines and other immunotherapies is to stimulate effective antigen-specific immunity. The combined use of cancer vaccines and other immunotherapies also has a large experimental and theoretical basis. Immune checkpoint inhibitors (ICIs) such as PD-1 blockers have been shown to have exceptional effectiveness against solid tumors by preventing PD-1/PD-L1 binding; however, the monotherapy approach is not sufficient and most patients either do not respond or eventually relapse. ICIs have emerged as a breakthrough approach in cancer therapy due to the exposure of the tumor microenvironment to immunosuppression and provided a strong rationale for the combination with vaccines.

For advanced tumors, they are generally more resistant to ICIs and vaccination alone has been ineffective for invasive cancer. A randomized clinical trial has demonstrated the safety and immunogenicity of this treatment regimen, and the contribution of vaccination to the tumoricidal effects of PD-1 inhibition has also been confirmed, including that for advanced melanoma, non-small cell lung cancer, or bladder cancer (140). Recently, a therapeutic HPV-16 SLP vaccine combined with the PD1 inhibitor in patients with HPV-16-positive cancer was shown with only apparent additive effects from each agent without increased immune adverse events, relative to PD-1 monotherapy (141). Based on the

discovery of the synergy of the two treatment modalities, we can combine two routes of administration into one treatment system or fuse them into a single expression system. Le et al. modified an anti-PDL1 antibody to NP surfaces, which involve adjuvant-loaded NPs that were prepared by entrapping imiquimod (IQ) in photoresponsive polydopamine nanoparticles (IQ/PNs) (142). Following NIR irradiation, mice treated with PDL1Ab-IQ/PNs not only resulted in primary tumor ablation, but also completely prevented secondary tumor growth at distant sites, with 100% survival for up to 150 days. Recently, researchers from New York have modified chimeric antigen receptor T cells (CAR-T) to secrete PD-1 blocked single-chain variable fragments (scFv); these scFvsecreting CAR-T cells play paracrine and autocrine roles to improve the antitumor activity of CAR-T cells and bystander tumor-specific T cells in syngeneic and xenogeneic mouse models of clinically relevant PD-L1⁺ hematological and solid tumors (143). These examples illustrate the complexity of tumor resistance to vaccines and immunotherapies, and highlight how multiple modalities will be required for therapeutic vaccines or other immunomodulatory therapies to overcome suppressive TME.

5.2.2 Chimeric antigen receptor T-cell therapy

Directly inducing the immune response of tumor lesions or a targeted effector T-cell response can be achieved through vaccination approaches combined with adoptive cell therapy. Due to the remarkable success of CAR-T cell therapy in hematological malignancies but not in solid tumors, the combination with nanotechnology may be more attractive (144, 145). Conventional NPs can boost CAR-T therapy by emerging as carriers for CAR-T to enhance targeting, or as a tool to enhance transfection efficiencies of CAR gene. However, SANPs can provide more potential strategies that could be used to modify CAR-T cells or as a booster vaccine to overcome the existing challenges in solid tumors. Natnaree et al. reported an approach in which cross-linked multilamellar vesicles (cMLV) could be covalently attached to CAR-T cells to deliver the A2aR-specific small-molecule antagonist SCH-58261 and the cMLVs NPs without affecting the effector and viability of CAR-T cells (146). In addition, Chenjun et al. previously established selfassembled multivalent CAR-like aptamer NPs, which can activate T cells while targeting B16 mouse melanoma tumor cells (147). The predictable result is the increase in durability of tumors and even the increase in efficacy of CAR-T against solid tumors. Towards this end, the combination of therapeutic CAR-T cell therapy with vaccines, such as DC vaccine, RNA vaccines, or novel approaches, was tentatively developed. The main argument of this treatment plan is synergy after immune activation. Reinhard et al. developed a CAR-T cell-amplifying RNA vaccine with lipid-based SANPs (148), by inducing DC natively displayed CLDN6, a tetraspanin membrane protein that is involved in tight junction formation (149), to promote cognate and selective expansion of CLDN6-CAR-T cells. However, this vaccine did not directly participate in the antitumor immune response in this process. Additionally, codelivering CAR-specific ligands that boost CAR-T cell numbers and functionality in vivo with low toxicity decorate the APCs in the

LN and provide critical priming signals to the CAR T cells (150). In general, it is necessary to pay attention to the crucial problems and obstacles of clinical progress, concentrate on resources and promote the understanding of the inconclusive interactions of the immune system, and develop optimal treatment schemes for cancer patients.

6 The challenge of SANPs in cancer vaccine

Today, most of the existing production processes of selfassembled nanovaccines are synthetic methods or semibiological technologies based on certain specific cells (101). The potential advantages of these strategies include reduced production time and cost. However, chemical synthesis and semibiosynthesis will limit the preventive and therapeutic effects of vaccines, which must face the difficulty of displaying versatile antigens on these proteinaceous NPs, especially glycan antigens with complex structures. Of course, the toxicity of individual materials is still present. A series of local events are generally considered as tissue or cell response continuum, such as injury by injection or implantation, acute inflammation, chronic inflammation, formation of granulation tissue, severe cytotoxicity, foreign body reaction, and fibrosis following implantation of microspheres (59, 151, 152). The method of using protein-based fully biosynthetic vaccines is still being explored, and this strategy still faces many difficulties related to antigen display. The preparation process of some SANPs used for antigen and drug delivery involves a series of chemical reactions; interestingly, the molecular interaction between the drug and the carrier does not break its inherent activity with the self-assembly process (17, 153, 154), which is conducive to the carrier and cargo fully playing their respective roles in vivo.

7 Conclusion

Future research on therapeutic cancer vaccines will focus not only on individualization of antigen for every tumor, but even more on the proper vaccine platform to maximize their impact. Breakthroughs in antigen delivery platforms have increased our capacity to personalize vaccines. Several supramolecular assembled programmable nanomedicines reported in recent years to improve immunotherapy efficiency and the emerging technology have made it possible to dissect the TME in-depth (113, 155, 156). To further inform the design of future treatment platforms, mechanistic analyses can be collected from previously failed and some clinically significant vaccine trials. Many ongoing studies using a range of different vaccine delivery platforms and combination therapies to alleviate tumor resistance have the ultimate goal of inducing effective, long-lasting, tumor-specific immunity in cancer patients.

SANPs composed of each material have specific advantages. For example, polymers have good stability and excellent biocompatibility, and peptide NPs are expected to achieve comprehensive biosynthesis in the future. The high loading, low toxicity, and high biocompatibility of most SANPs make them more valuable in vaccine development than traditional adjuvant vaccines. At the same time, vaccines based on various NPs also have shown good results in animal models, which provides a huge impetus for the development of many vaccines against complex diseases in the future. Although many satisfactory experimental results have been produced in the field of self-assembled nanovaccines, the current understanding of the physical and chemical characteristics of nanomaterials and their interaction with physiological systems is limited, and the optimal synthesis, synthesis of nanomaterials, and chemical modification are unclear (157). To provide better protection, a self-assembled nanovaccine must be able to induce a series of immune responses. Although existing functionalized NPs can be internalized by immune cells, the effect of the sites of many functional proteins for the uptake of APCs is still unknown (158). It should be recognized that the idea of carrying antigens on the surface of SANPs to induce immune responses proves to be promising in the least. In the future, we are expected to load more antigens on the surface of multivalent NPs to drive specific and strong CD4+ T-cell and/or CD8+ T-cell responses. If an NP platform can resolve distribution barriers in the future, the age of enlightenment of nanotherapeutics is nearing its end.

Author contributions

TS and FM conceived the structure of this review. TS wrote and edited the manuscript. MS collected and prepared the related figures. FM and CL reviewed and made revisions to the manuscript. All authors contributed to the article and approved the submitted version.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

1. Tan AC, Goubier A, Kohrt HE. A quantitative analysis of therapeutic cancer vaccines in phase 2 or phase 3 trial. *J Immunother Cancer* (2015) 3:48. doi: 10.1186/ s40425-015-0093-x

2. Tanne JH. FDA Approves prostate cancer "vaccine". *BMJ* (2010) 340:c2431. doi: 10.1136/bmj.c2431

3. Filaci G, Fenoglio D, Nole F, Zanardi E, Tomasello L, Aglietta M, et al. Telomerase-based GX301 cancer vaccine in patients with metastatic castration-resistant prostate cancer: a randomized phase II trial. *Cancer Immunol Immunother* (2021) 70(12):3679–92. doi: 10.1007/s00262-021-03024-0

4. Ding Z, Li Q, Zhang R, Xie L, Shu Y, Gao S, et al. Personalized neoantigen pulsed dendritic cell vaccine for advanced lung cancer. *Signal Transduct Target Ther* (2021) 6 (1):26. doi: 10.1038/s41392-020-00448-5

5. Ott PA, Hu Z, Keskin DB, Shukla SA, Sun J, Bozym DJ, et al. An immunogenic personal neoantigen vaccine for patients with melanoma. *Nature* (2017) 547 (7662):217–21. doi: 10.1038/nature22991

6. Blass E, Ott PA. Advances in the development of personalized neoantigen-based therapeutic cancer vaccines. *Nat Rev Clin Oncol* (2021) 18(4):215–29. doi: 10.1038/ s41571-020-00460-2

7. Ni Q, Zhang F, Liu Y, Wang Z, Yu G, Liang B, et al. A bi-adjuvant nanovaccine that potentiates immunogenicity of neoantigen for combination immunotherapy of colorectal cancer. Sci Adv (2020) 6(12):eaaw6071. doi: 10.1126/sciadv.aaw6071

8. Tostanoski LH, Jewell CM. Engineering self-assembled materials to study and direct immune function. *Adv Drug Delivery Rev* (2017) 114:60–78. doi: 10.1016/j.addr.2017.03.005

9. Takahashi WN, Ishii M. An abnormal protein associated with tobacco mosaic virus infection. *Nature* (1952) 169(4297):419-20. doi: 10.1038/169419a0

10. Drexler KE. Molecular engineering: An approach to the development of general capabilities for molecular manipulation. *Proc Natl Acad Sci USA* (1981) 78(9):5275–8. doi: 10.1073/pnas.78.9.5275

11. Negahdaripour M, Golkar N, Hajighahramani N, Kianpour S, Nezafat N, Ghasemi Y. Harnessing self-assembled peptide nanoparticles in epitope vaccine design. *Biotechnol Adv* (2017) 35(5):575–96. doi: 10.1016/j.biotechadv.2017.05.002

12. Ilyinskii PO, Roy CJ, O'Neil CP, Browning EA, Pittet LA, Altreuter DH, et al. Adjuvant-carrying synthetic vaccine particles augment the immune response to encapsulated antigen and exhibit strong local immune activation without inducing systemic cytokine release. *Vaccine* (2014) 32(24):2882–95. doi: 10.1016/j.vaccine.2014.02.027

13. Desai P, Venkataramanan A, Schneider R, Jaiswal MK, Carrow JK, Purwada A, et al. Self-assembled, ellipsoidal polymeric nanoparticles for intracellular delivery of therapeutics. J BioMed Mater Res A (2018) 106(7):2048–58. doi: 10.1002/jbm.a.36400

14. Taylor TM, Davidson PM, Bruce BD, Weiss J. Liposomal nanocapsules in food science and agriculture. *Crit Rev Food Sci Nutr* (2005) 45(7-8):587–605. doi: 10.1080/10408390591001135

15. Whitesides GM, Grzybowski B. Self-assembly at all scales. Science (2002) 295 (5564):2418-21. doi: 10.1126/science.1070821

16. Arango-Restrepo A, Barragan D, Rubi JM. Self-assembling outside equilibrium: emergence of structures mediated by dissipation. *Phys Chem Chem Phys* (2019) 21 (32):17475–93. doi: 10.1039/C9CP01088B

17. Varma LT, Singh N, Gorain B, Choudhury H, Tambuwala MM, Kesharwani P, et al. Recent advances in self-assembled nanoparticles for drug delivery. *Curr Drug Deliv* (2020) 17(4):279–91. doi: 10.2174/1567201817666200210122340

18. Caston JR, Carrascosa JL. The basic architecture of viruses. Subcell Biochem (2013) 68:53–75. doi: 10.1007/978-94-007-6552-8_2

19. Zottig X, Cote-Cyr M, Arpin D, Archambault D, Bourgault S. Protein supramolecular structures: From self-assembly to nanovaccine design. *Nanomater* (*Basel*) (2020) 10(5). doi: 10.3390/nano10051008

20. Lv S, Song K, Yen A, Peeler DJ, Nguyen DC, Olshefsky A, et al. Well-defined mannosylated polymer for peptide vaccine delivery with enhanced antitumor immunity. *Adv Healthc Mater* (2022) 11(9):e2101651. doi: 10.1002/adhm.202101651

21. Ahlawat J, Henriquez G, Narayan M. Enhancing the delivery of chemotherapeutics: Role of biodegradable polymeric nanoparticles. *Molecules* (2018) 23(9):2157. doi: 10.3390/molecules23092157

22. Charlton Hume HK, Lua LHL. Platform technologies for modern vaccine manufacturing. *Vaccine* (2017) 35(35 Pt A):4480–5. doi: 10.1016/j.vaccine.2017.02.069

23. Pan J, Cui Z. Self-assembled nanoparticles: Exciting platforms for vaccination. *Biotechnol J* (2020) 15(12):e2000087. doi: 10.1002/biot.202000087

24. Patel A, Gaharwar AK, Iviglia G, Zhang H, Mukundan S, Mihaila SM, et al. Highly elastomeric poly(glycerol sebacate)-co-poly(ethylene glycol) amphiphilic block copolymers. *Biomaterials* (2013) 34(16):3970-83. doi: 10.1016/j.biomaterials.2013.01.045

25. Jiang W, Kim BY, Rutka JT, Chan WC. Nanoparticle-mediated cellular response is size-dependent. *Nat Nanotechnol* (2008) 3(3):145–50. doi: 10.1038/nnano.2008.30

26. Delfi M, Sartorius R, Ashrafizadeh M, Sharifi E, Zhang Y, De Berardinis P, et al. Self-assembled peptide and protein nanostructures for anti-cancer therapy: Targeted delivery, stimuli-responsive devices and immunotherapy. *Nano Today* (2021) 38:101119. doi: 10.1016/j.nantod.2021.101119

27. Manolova V, Flace A, Bauer M, Schwarz K, Saudan P, Bachmann MF. Nanoparticles target distinct dendritic cell populations according to their size. *Eur J Immunol* (2008) 38(5):1404–13. doi: 10.1002/eji.200737984

28. Shanbhag BK, Liu C, Haritos VS, He L. Understanding the interplay between self-assembling peptides and solution ions for tunable protein nanoparticle formation. *ACS Nano* (2018) 12(7):6956–67. doi: 10.1021/acsnano.8b02381

29. Yang Y, Ge S, Song Z, Zhao A, Zhao L, Hu Z, et al. A novel self-assembled epitope peptide nanoemulsion vaccine targeting nasal mucosal epithelial cell for reinvigorating CD8(+) T cell immune activity and inhibiting tumor progression. *Int J Biol Macromol* (2021) 183:1891–902. doi: 10.1016/j.ijbiomac.2021.05.158

30. Ma T, Liu Y, Cheng J, Liu Y, Fan W, Cheng Z, et al. Liposomes containing recombinant e protein vaccine against duck tembusu virus in ducks. *Vaccine* (2016) 34 (19):2157–63. doi: 10.1016/j.vaccine.2016.03.030

31. Teng X, Tian M, Li J, Tan S, Yuan X, Yu Q, et al. Immunogenicity and protective efficacy of DMT liposome-adjuvanted tuberculosis subunit CTT3H vaccine. *Hum Vaccin Immunother* (2015) 11(6):1456–64. doi: 10.1080/21645515.2015.1037057

32. Taki A, Smooker P. Small wonders-the use of nanoparticles for delivering antigen. Vaccines (Basel) (2015) 3(3):638-61. doi: 10.3390/vaccines3030638

33. Rao M, Wassef NM, Alving CR, Krzych U. Intracellular processing of liposomeencapsulated antigens by macrophages depends upon the antigen. *Infect Immun* (1995) 63(7):2396–402. doi: 10.1128/iai.63.7.2396-2402.1995

34. Jahan ST, Sadat SM, Haddadi A. Design and immunological evaluation of anti-CD205-tailored PLGA-based nanoparticulate cancer vaccine. *Int J Nanomed* (2018) 13:367–86. doi: 10.2147/JJN.S144266

35. Liu TY, Hussein WM, Jia Z, Ziora ZM, McMillan NA, Monteiro MJ, et al. Selfadjuvanting polymer-peptide conjugates as therapeutic vaccine candidates against cervical cancer. *Biomacromolecules* (2013) 14(8):2798–806. doi: 10.1021/bm400626w

36. Polla Ravi S, Shamiya Y, Chakraborty A, Elias C, Paul A. Biomaterials, biological molecules, and polymers in developing vaccines. *Trends Pharmacol Sci* (2021) 42 (10):813–28. doi: 10.1016/j.tips.2021.07.006

37. Jyoti K, Jain S, Katare OP, Katyal A, Chandra R, Madan J. Non-small cell lung cancer tumour antigen, MUC-1 peptide-loaded non-aggregated poly (lactide-co-glycolide) nanoparticles augmented cellular uptake in mouse professional antigenpresenting cells: optimisation and characterisation. *J Microencapsul* (2020) 37(1):14–28. doi: 10.1080/02652048.2019.1692943

38. Yang R, Xu J, Xu L, Sun X, Chen Q, Zhao Y, et al. Cancer cell membrane-coated adjuvant nanoparticles with mannose modification for effective anticancer vaccination. *ACS Nano* (2018) 12(6):5121–9. doi: 10.1021/acsnano.7b09041

39. Kim H, Niu L, Larson P, Kucaba TA, Murphy KA, James BR, et al. Polymeric nanoparticles encapsulating novel TLR7/8 agonists as immunostimulatory adjuvants for enhanced cancer immunotherapy. *Biomaterials* (2018) 164:38–53. doi: 10.1016/j.biomaterials.2018.02.034

40. Campbell K, Young VL, Donaldson BC, Woodall MJ, Shields NJ, Walker GF, et al. Delivering two tumour antigens survivin and mucin-1 on virus-like particles enhances anti-tumour immune responses. *Vaccines (Basel)* (2021) 9(5):463. doi: 10.3390/vaccines9050463

41. Palladini A, Thrane S, Janitzek CM, Pihl J, Clemmensen SB, de Jongh WA, et al. Virus-like particle display of HER2 induces potent anti-cancer responses. *Oncoimmunology* (2018) 7(3):e1408749. doi: 10.1080/2162402X.2017.1408749

42. Salazar-Gonzalez JA, Ruiz-Cruz AA, Bustos-Jaimes I, Moreno-Fierros L. Expression of breast cancer-related epitopes targeting the IGF-1 receptor in chimeric human parvovirus B19 virus-like particles. *Mol Biotechnol* (2019) 61(10):742–53. doi: 10.1007/s12033-019-00198-y

43. Jemon K, Young V, Wilson M, McKee S, Ward V, Baird M, et al. An enhanced heterologous virus-like particle for human papillomavirus type 16 tumour immunotherapy. *PloS One* (2013) 8(6):e66866. doi: 10.1371/journal.pone.0066866

44. Zhang S, Yong LK, Li D, Cubas R, Chen C, Yao Q. Mesothelin virus-like particle immunization controls pancreatic cancer growth through CD8+ T cell induction and reduction in the frequency of CD4+ foxp3+ ICOS- regulatory T cells. *PloS One* (2013) 8 (7):e68303. doi: 10.1371/journal.pone.0068303

45. Cubas R, Zhang S, Li M, Chen C, Yao Q. Chimeric Trop2 virus-like particles: a potential immunotherapeutic approach against pancreatic cancer. *J Immunother* (2011) 34(3):251–63. doi: 10.1097/CJI.0b013e318209ee72

46. Cheng K, Du T, Li Y, Qi Y, Min H, Wang Y, et al. Dual-Antigen-Loaded hepatitis b virus core antigen virus-like particles stimulate efficient immunotherapy against melanoma. ACS Appl Mater Interfaces (2020) 12(48):53682–90. doi: 10.1021/acsami.0c16012

47. Mohsen MO, Heath MD, Cabral-Miranda G, Lipp C, Zeltins A, Sande M, et al. Vaccination with nanoparticles combined with micro-adjuvants protects against cancer. *J Immunother Cancer* (2019) 7(1):114. doi: 10.1186/s40425-019-0587-z

48. Shukla S, Myers JT, Woods SE, Gong X, Czapar AE, Commandeur U, et al. Plant viral nanoparticles-based HER2 vaccine: Immune response influenced by differential transport, localization and cellular interactions of particulate carriers. *Biomaterials* (2017) 121:15–27. doi: 10.1016/j.biomaterials.2016.12.030

49. Zhai L, Peabody J, Pang YS, Schiller J, Chackerian B, Tumban E. A novel candidate HPV vaccine: MS2 phage VLP displaying a tandem HPV L2 peptide offers similar protection in mice to gardasil-9. *Antiviral Res* (2017) 147:116–23. doi: 10.1016/j.antiviral.2017.09.012

50. Wu Y, Norberg PK, Reap EA, Congdon KL, Fries CN, Kelly SH, et al. A supramolecular vaccine platform based on alpha-helical peptide nanofibers. ACS Biomater Sci Eng (2017) 3(12):3128–32. doi: 10.1021/acsbiomaterials.7b00561

51. Lee A, Dadhwal S, Gamble A, Hook S. Liposomes with cyclodextrin channels and polyethyleneimine (PEI) improves cytoplasmic vaccine delivery and induces anticancer immune activity in mice. J Liposome Res (2022) 32(1):22–31. doi: 10.1080/ 08982104.2020.1831016

52. Rastakhiz S, Yazdani M, Shariat S, Arab A, Momtazi-Borojeni AA, Barati N, et al. Preparation of nanoliposomes linked to HER2/neu-derived (P5) peptide containing MPL adjuvant as vaccine against breast cancer. *J Cell Biochem* (2018) 120 (2):1294–303. doi: 10.1002/jcb.27090

53. Yoshino T, Miyazaki J, Kojima T, Kandori S, Shiga M, Kawahara T, et al. Cationized liposomal keto-mycolic acids isolated from mycobacterium bovis bacillus calmette-guerin induce antitumor immunity in a syngeneic murine bladder cancer model. *PloS One* (2019) 14(1):e0209196. doi: 10.1371/journal.pone.0209196

54. Park S, Kim D, Wu G, Jung H, Park JA, Kwon HJ, et al. A peptide-CpG-DNAliposome complex vaccine targeting TM4SF5 suppresses growth of pancreatic cancer in a mouse allograft model. *Onco Targets Ther* (2018) 11:8655–72. doi: 10.2147/ OTT.S186606

55. Sayour EJ, De Leon G, Pham C, Grippin A, Kemeny H, Chua J, et al. Systemic activation of antigen-presenting cells *via* RNA-loaded nanoparticles. *Oncoimmunology* (2017) 6(1):e1256527. doi: 10.1080/2162402X.2016.1256527

56. Bonifacio BV, Silva PB, Ramos MA, Negri KM, Bauab TM, Chorilli M. Nanotechnology-based drug delivery systems and herbal medicines: a review. *Int J Nanomed* (2014) 9:1–15. doi: 10.2147/IJN.S52634

57. Ensign LM, Cone R, Hanes J. Oral drug delivery with polymeric nanoparticles: the gastrointestinal mucus barriers. *Adv Drug Delivery Rev* (2012) 64(6):557–70. doi: 10.1016/j.addr.2011.12.009

58. Chen G, Zhao B, Ruiz EF, Zhang F. Advances in the polymeric delivery of nucleic acid vaccines. *Theranostics* (2022) 12(9):4081–109. doi: 10.7150/thno.70853

59. Cui C, Stevens VC, Schwendeman SP. Injectable polymer microspheres enhance immunogenicity of a contraceptive peptide vaccine. *Vaccine* (2007) 25(3):500–9. doi: 10.1016/j.vaccine.2006.07.055

60. Azmi F, Ahmad Fuaad AA, Skwarczynski M, Toth I. Recent progress in adjuvant discovery for peptide-based subunit vaccines. *Hum Vaccin Immunother* (2014) 10 (3):778–96. doi: 10.4161/hv.27332

61. Chou PY, Lin SY, Wu YN, Shen CY, Sheu MT, Ho HO. Glycosylation of OVA antigen-loaded PLGA nanoparticles enhances DC-targeting for cancer vaccination. *J Control Release* (2022) 351:970–88. doi: 10.1016/j.jconrel.2022.10.002

62. Lynn GM, Laga R, Darrah PA, Ishizuka AS, Balaci AJ, Dulcey AE, et al. *In vivo* characterization of the physicochemical properties of polymer-linked TLR agonists that enhance vaccine immunogenicity. *Nat Biotechnol* (2015) 33(11):1201–10. doi: 10.1038/nbt.3371

63. Schwendener RA. Liposomes as vaccine delivery systems: a review of the recent advances. *Ther Adv Vaccines* (2014) 2(6):159–82. doi: 10.1177/2051013614541440

64. Tran S, DeGiovanni PJ, Piel B, Rai P. Cancer nanomedicine: a review of recent success in drug delivery. *Clin Transl Med* (2017) 6(1):44. doi: 10.1186/s40169-017-0175-0

65. Musacchio T, Torchilin VP. Recent developments in lipid-based pharmaceutical nanocarriers. *Front Biosci (Landmark Ed).* (2011) 16(4):1388–412. doi: 10.2741/3795

66. Chen WC, Huang L. Non-viral vector as vaccine carrier. Adv Genet (2005) 54:315–37. doi: 10.1016/S0065-2660(05)54013-6

67. Simberg D, Weisman S, Talmon Y, Barenholz Y. DOTAP (and other cationic lipids): chemistry, biophysics, and transfection. *Crit Rev Ther Drug Carrier Syst* (2004) 21(4):257–317. doi: 10.1615/CritRevTherDrugCarrierSyst.v21.i4.10

68. Yan W, Chen W, Huang L. Mechanism of adjuvant activity of cationic liposome: phosphorylation of a MAP kinase, ERK and induction of chemokines. *Mol Immunol* (2007) 44(15):3672–81. doi: 10.1016/j.molimm.2007.04.009

69. Yan W, Chen W, Huang L. Reactive oxygen species play a central role in the activity of cationic liposome based cancer vaccine. *J Control Release* (2008) 130(1):22–8. doi: 10.1016/j.jconrel.2008.05.005

70. Korsholm KS, Hansen J, Karlsen K, Filskov J, Mikkelsen M, Lindenstrom T, et al. Induction of CD8+ T-cell responses against subunit antigens by the novel cationic liposomal CAF09 adjuvant. *Vaccine* (2014) 32(31):3927–35. doi: 10.1016/j.vaccine.2014.05.050

71. Varypataki EM, Benne N, Bouwstra J, Jiskoot W, Ossendorp F. Efficient eradication of established tumors in mice with cationic liposome-based synthetic long-peptide vaccines. *Cancer Immunol Res* (2017) 5(3):222–33. doi: 10.1158/2326-6066.CIR-16-0283

72. Varypataki EM, van der Maaden K, Bouwstra J, Ossendorp F, Jiskoot W. Cationic liposomes loaded with a synthetic long peptide and poly(I:C): a defined adjuvanted vaccine for induction of antigen-specific T cell cytotoxicity. *AAPS J* (2015) 17(1):216–26. doi: 10.1208/s12248-014-9686-4

73. Heuts J, Varypataki EM, van der Maaden K, Romeijn S, Drijfhout JW, van Scheltinga AT, et al. Cationic liposomes: A flexible vaccine delivery system for physicochemically diverse antigenic peptides. *Pharm Res* (2018) 35(11):207. doi: 10.1007/s11095-018-2490-6

74. Sayour EJ, Mendez-Gomez HR, Mitchell DA. Cancer vaccine immunotherapy with RNA-loaded liposomes. *Int J Mol Sci* (2018) 19(10):2890. doi: 10.3390/ ijms19102890

75. Chen J, Ye Z, Huang C, Qiu M, Song D, Li Y, et al. Lipid nanoparticle-mediated lymph node-targeting delivery of mRNA cancer vaccine elicits robust CD8(+) T cell response. *Proc Natl Acad Sci USA* (2022) 119(34):e2207841119. doi: 10.1073/pnas.2207841119

76. Oberli MA, Reichmuth AM, Dorkin JR, Mitchell MJ, Fenton OS, Jaklenec A, et al. Lipid nanoparticle assisted mRNA delivery for potent cancer immunotherapy. *Nano Lett* (2017) 17(3):1326–35. doi: 10.1021/acs.nanolett.6b03329

77. Ghaffari-Nazari H, Tavakkol-Afshari J, Jaafari MR, Tahaghoghi-Hajghorbani S, Masoumi E, Jalali SA. Improving multi-epitope long peptide vaccine potency by using a strategy that enhances CD4+ T help in BALB/c mice. *PloS One* (2015) 10(11):e0142563. doi: 10.1371/journal.pone.0142563

78. Ulijn RV, Smith AM. Designing peptide based nanomaterials. Chem Soc Rev (2008) 37(4):664-75. doi: 10.1039/b609047h

79. Mandal D, Nasrolahi Shirazi A, Parang K. Self-assembly of peptides to nanostructures. Org Biomol Chem (2014) 12(22):3544–61. doi: 10.1039/C4OB00447G

80. Jun S, Hong Y, Imamura H, Ha BY, Bechhoefer J, Chen P. Self-assembly of the ionic peptide EAK16: the effect of charge distributions on self-assembly. *Biophys J* (2004) 87(2):1249–59. doi: 10.1529/biophysj.103.038166

81. Gelain F, Horii A, Zhang S. Designer self-assembling peptide scaffolds for 3-d tissue cell cultures and regenerative medicine. *Macromol Biosci* (2007) 7(5):544–51. doi: 10.1002/mabi.200700033

82. Hossen S, Hossain MK, Basher MK, Mia MNH, Rahman MT, Uddin MJ. Smart nanocarrier-based drug delivery systems for cancer therapy and toxicity studies: A review. J Adv Res (2019) 15:1–18. doi: 10.1016/j.jare.2018.06.005

83. Doll TA, Raman S, Dey R, Burkhard P. Nanoscale assemblies and their biomedical applications. J R Soc Interface (2013) 10(80):20120740. doi: 10.1098/ rsif.2012.0740

84. Qi GB, Gao YJ, Wang L, Wang H. Self-assembled peptide-based nanomaterials for biomedical imaging and therapy. *Adv Mater* (2018) 30(22):e1703444. doi: 10.1002/adma.201703444

85. Lynn GM, Sedlik C, Baharom F, Zhu Y, Ramirez-Valdez RA, Coble VL, et al. Peptide-TLR-7/8a conjugate vaccines chemically programmed for nanoparticle selfassembly enhance CD8 T-cell immunity to tumor antigens. *Nat Biotechnol* (2020) 38 (3):320–32. doi: 10.1038/s41587-019-0390-x

86. Shi W, Qiu Q, Feng Z, Tong Z, Guo W, Zou F, et al. Design, synthesis and immunological evaluation of self-assembled antigenic peptides from dual-antigen targets: a broad-spectrum candidate for an effective antibreast cancer therapy. *J Immunother Cancer* (2021) 9(6):e002523. doi: 10.1136/jitc-2021-002523

87. Fuenmayor J, Godia F, Cervera L. Production of virus-like particles for vaccines. *N Biotechnol* (2017) 39(Pt B):174–80. doi: 10.1016/j.nbt.2017.07.010

88. Kang SM, Pushko P, Bright RA, Smith G, Compans RW. Influenza virus-like particles as pandemic vaccines. *Curr Top Microbiol Immunol* (2009) 333:269–89. doi: 10.1007/978-3-540-92165-3_14

89. Tan M, Jiang X. Subviral particle as vaccine and vaccine platform. *Curr Opin Virol* (2014) 6:24–33. doi: 10.1016/j.coviro.2014.02.009

90. Grgacic EV, Anderson DA. Virus-like particles: passport to immune recognition. *Methods* (2006) 40(1):60-5. doi: 10.1016/j.ymeth.2006.07.018

91. Charlton Hume HK, Vidigal J, Carrondo MJT, Middelberg APJ, Roldao A, Lua LHL. Synthetic biology for bioengineering virus-like particle vaccines. *Biotechnol Bioeng* (2019) 116(4):919–35. doi: 10.1002/bit.26890

92. Fifis T, Gamvrellis A, Crimeen-Irwin B, Pietersz GA, Li J, Mottram PL, et al. Size-dependent immunogenicity: therapeutic and protective properties of nano-vaccines against tumors. *J Immunol* (2004) 173(5):3148–54. doi: 10.4049/jimmunol.173.5.3148

93. Abdoli A, Soleimanjahi H, Fotouhi F, Teimoori A, Pour Beiranvand S, Kianmehr Z. Human papillomavirus Type16- L1 VLP production in insect cells. *Iran J Basic Med Sci* (2013) 16(8):891–5.

94. Lua LH, Connors NK, Sainsbury F, Chuan YP, Wibowo N, Middelberg AP. Bioengineering virus-like particles as vaccines. *Biotechnol Bioeng* (2014) 111(3):425–40. doi: 10.1002/bit.25159

95. Mao C, Beiss V, Fields J, Steinmetz NF, Fiering S. Cowpea mosaic virus stimulates antitumor immunity through recognition by multiple MYD88-dependent toll-like receptors. *Biomaterials* (2021) 275:120914. doi: 10.1016/j.biomaterials.2021.120914

96. Evtushenko EA, Ryabchevskaya EM, Nikitin NA, Atabekov JG, Karpova OV. Plant virus particles with various shapes as potential adjuvants. *Sci Rep* (2020) 10 (1):10365. doi: 10.1038/s41598-020-67023-4

97. Rolih V, Caldeira J, Bolli E, Salameh A, Conti L, Barutello G, et al. Development of a VLP-based vaccine displaying an xCT extracellular domain for the treatment of metastatic breast cancer. *Cancers (Basel)* (2020) 12(6):1492. doi: 10.3390/cancers12061492

98. Jose C, Amra K, Bhavsar C, Momin M, Omri A. Polymeric lipid hybrid nanoparticles: Properties and therapeutic applications. *Crit Rev Ther Drug Carrier Syst* (2018) 35(6):555–88. doi: 10.1615/CritRevTherDrugCarrierSyst.2018024751

99. Chen L, Qin H, Zhao R, Zhao X, Lin L, Chen Y, et al. Bacterial cytoplasmic membranes synergistically enhance the antitumor activity of autologous cancer vaccines. *Sci Transl Med* (2021) 13(601):eabc2816. doi: 10.1126/scitranslmed.abc2816

100. Wang D, Liu C, You S, Zhang K, Li M, Cao Y, et al. Bacterial vesicle-cancer cell hybrid membrane-coated nanoparticles for tumor specific immune activation and photothermal therapy. *ACS Appl Mater Interfaces* (2020) 12(37):41138–47. doi: 10.1021/acsami.0c13169

101. Pan C, Wu J, Qing S, Zhang X, Zhang L, Yue H, et al. Biosynthesis of selfassembled proteinaceous nanoparticles for vaccination. *Adv Mater* (2020) 32(42): e2002940. doi: 10.1002/adma.202002940

102. Nijen Twilhaar MK, Czentner L, van Nostrum CF, Storm G, den Haan JMM. Mimicking pathogens to augment the potency of liposomal cancer vaccines. *Pharmaceutics* (2021) 13(7):eabc2816. doi: 10.3390/pharmaceutics13070954

103. Banchereau J, Briere F, Caux C, Davoust J, Lebecque S, Liu YJ, et al. Immunobiology of dendritic cells. *Annu Rev Immunol* (2000) 18:767-811. doi: 10.1146/annurev.immunol.18.1.767

104. Schijns VE. Immunological concepts of vaccine adjuvant activity. *Curr Opin Immunol* (2000) 12(4):456–63. doi: 10.1016/S0952-7915(00)00120-5

105. Zhang R, Wang C, Guan Y, Wei X, Sha M, Yi M, et al. Manganese salts function as potent adjuvants. *Cell Mol Immunol* (2021) 18(5):1222–34. doi: 10.1038/s41423-021-00669-w

106. Donaldson B, Lateef Z, Walker GF, Young SL, Ward VK. Virus-like particle vaccines: immunology and formulation for clinical translation. *Expert Rev Vaccines* (2018) 17(9):833-49. doi: 10.1080/14760584.2018.1516552

107. Uto T, Akagi T, Yoshinaga K, Toyama M, Akashi M, Baba M. The induction of innate and adaptive immunity by biodegradable poly(gamma-glutamic acid) nanoparticles *via* a TLR4 and MyD88 signaling pathway. *Biomaterials* (2011) 32 (22):5206–12. doi: 10.1016/j.biomaterials.2011.03.052

108. Mesa C, De Leon J, Rigley K, Fernandez LE. Very small size proteoliposomes derived from neisseria meningitidis: an effective adjuvant for Th1 induction and dendritic cell activation. *Vaccine* (2004) 22(23-24):3045-52. doi: 10.1016/j.vaccine.2004.02.010

109. Luo M, Wang H, Wang Z, Cai H, Lu Z, Li Y, et al. A STING-activating nanovaccine for cancer immunotherapy. *Nat Nanotechnol* (2017) 12(7):648–54. doi: 10.1038/nnano.2017.52

110. Zhao L, Seth A, Wibowo N, Zhao CX, Mitter N, Yu C, et al. Nanoparticle vaccines. Vaccine (2014) 32(3):327-37. doi: 10.1016/j.vaccine.2013.11.069

111. Shetab Boushehri MA, Lamprecht A. TLR4-based immunotherapeutics in cancer: A review of the achievements and shortcomings. *Mol Pharm* (2018) 15 (11):4777–800. doi: 10.1021/acs.molpharmaceut.8b00691

112. Kawashita Y, Deb NJ, Garg M, Kabarriti R, Alfieri A, Takahashi M, et al. An autologous *in situ* tumor vaccination approach for hepatocellular carcinoma. 1. Fl3 ligand gene transfer increases antitumor effects of a radio-inducible suicide gene therapy in an ectopic tumor model. *Radiat Res* (2014) 182(2):191–200. doi: 10.1667/RR13594.1

113. Zhang Y, Ma S, Liu X, Xu Y, Zhao J, Si X, et al. Supramolecular assembled programmable nanomedicine as *In situ* cancer vaccine for cancer immunotherapy. *Adv Mater* (2021) 33(7):e2007293. doi: 10.1002/adma.202007293

114. Liu J, He S, Luo Y, Zhang Y, Du X, Xu C, et al. Tumor-Microenvironment-Activatable polymer nano-immunomodulator for precision cancer photoimmunotherapy. *Adv Mater* (2022) 34(8):e2106654. doi: 10.1002/adma.202106654

115. Kwok M, Fritsch EF, Wu CJ. Cancer and COVID-19: On the quest for effective vaccines. *Blood Cancer Discovery* (2021) 2(1):13–8. doi: 10.1158/2643-3230.BCD-20-0205

116. Saxena M, van der Burg SH, Melief CJM, Bhardwaj N. Therapeutic cancer vaccines. *Nat Rev Cancer*. (2021) 21(6):360–78. doi: 10.1038/s41568-021-00346-0

117. Yarchoan M, Albacker LA, Hopkins AC, Montesion M, Murugesan K, Vithayathil TT, et al. PD-L1 expression and tumor mutational burden are independent biomarkers in most cancers. *JCI Insight* (2019) 4(6):e126908. doi: 10.1172/jci.insight.126908

118. Hellmann MD, Callahan MK, Awad MM, Calvo E, Ascierto PA, Atmaca A, et al. Tumor mutational burden and efficacy of nivolumab monotherapy and in combination with ipilimumab in small-cell lung cancer. *Cancer Cell* (2018) 33 (5):853–61.e4. doi: 10.1016/j.ccell.2018.04.001

119. Le DT, Uram JN, Wang H, Bartlett BR, Kemberling H, Eyring AD, et al. PD-1 blockade in tumors with mismatch-repair deficiency. *N Engl J Med* (2015) 372 (26):2509–20. doi: 10.1056/NEJMoa1500596

120. Hellmann MD, Nathanson T, Rizvi H, Creelan BC, Sanchez-Vega F, Ahuja A, et al. Genomic features of response to combination immunotherapy in patients with advanced non-Small-Cell lung cancer. *Cancer Cell* (2018) 33(5):843–52.e4. doi: 10.1016/j.ccell.2018.03.018

121. Van Allen EM, Miao D, Schilling B, Shukla SA, Blank C, Zimmer L, et al. Genomic correlates of response to CTLA-4 blockade in metastatic melanoma. *Science* (2015) 350(6257):207–11. doi: 10.1126/science.aad0095

122. Snyder A, Makarov V, Merghoub T, Yuan J, Zaretsky JM, Desrichard A, et al. Genetic basis for clinical response to CTLA-4 blockade in melanoma. *N Engl J Med* (2014) 371(23):2189–99. doi: 10.1056/NEJM0a1406498

123. Rizvi NA, Hellmann MD, Snyder A, Kvistborg P, Makarov V, Havel JJ, et al. Cancer immunology. mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. *Science* (2015) 348(6230):124–8. doi: 10.1126/science.aaa1348

124. Garcia-Diaz A, Shin DS, Moreno BH, Saco J, Escuin-Ordinas H, Rodriguez GA, et al. Interferon receptor signaling pathways regulating PD-L1 and PD-L2 expression. *Cell Rep* (2017) 19(6):1189–201. doi: 10.1016/j.celrep.2017.04.031

125. Spranger S, Spaapen RM, Zha Y, Williams J, Meng Y, Ha TT, et al. Upregulation of PD-L1, IDO, and t(regs) in the melanoma tumor microenvironment is driven by CD8(+) T cells. *Sci Transl Med* (2013) 5(200):200ra116. doi: 10.1126/ scitranslmed.3006504

126. Finke JH, Rini B, Ireland J, Rayman P, Richmond A, Golshayan A, et al. Sunitinib reverses type-1 immune suppression and decreases T-regulatory cells in renal cell carcinoma patients. *Clin Cancer Res* (2008) 14(20):6674–82. doi: 10.1158/1078-0432.CCR-07-5212

127. Vermeij R, Leffers N, Hoogeboom BN, Hamming IL, Wolf R, Reyners AK, et al. Potentiation of a p53-SLP vaccine by cyclophosphamide in ovarian cancer: a single-arm phase II study. *Int J Cancer.* (2012) 131(5):E670–80. doi: 10.1002/ijc.27388

128. Borch TH, Engell-Noerregaard L, Zeeberg Iversen T, Ellebaek E, Met O, Hansen M, et al. mRNA-transfected dendritic cell vaccine in combination with metronomic cyclophosphamide as treatment for patients with advanced malignant melanoma. *Oncoimmunology* (2016) 5(9):e1207842. doi: 10.1080/2162402X.2016.1207842

129. Camisaschi C, Filipazzi P, Tazzari M, Casati C, Beretta V, Pilla L, et al. Effects of cyclophosphamide and IL-2 on regulatory CD4+ T cell frequency and function in melanoma patients vaccinated with HLA-class I peptides: impact on the antigenspecific T cell response. *Cancer Immunol Immunother* (2013) 62(5):897–908. doi: 10.1007/s00262-013-1397-7

130. Dijkgraaf EM, Santegoets SJ, Reyners AK, Goedemans R, Nijman HW, van Poelgeest MI, et al. A phase 1/2 study combining gemcitabine, pegintron and p53 SLP vaccine in patients with platinum-resistant ovarian cancer. *Oncotarget* (2015) 6 (31):32228–43. doi: 10.18632/oncotarget.4772

131. Dehlin M, Andersson S, Erlandsson M, Brisslert M, Bokarewa M. Inhibition of fms-like tyrosine kinase 3 alleviates experimental arthritis by reducing formation of dendritic cells and antigen presentation. *J Leukoc Biol* (2011) 90(4):811–7. doi: 10.1189/ jlb.1110640

132. Sevko A, Sade-Feldman M, Kanterman J, Michels T, Falk CS, Umansky L, et al. Cyclophosphamide promotes chronic inflammation-dependent immunosuppression and prevents antitumor response in melanoma. *J Invest Dermatol* (2013) 133(6):1610–9. doi: 10.1038/jid.2012.444

133. van Dongen M, Savage ND, Jordanova ES, Briaire-de Bruijn IH, Walburg KV, Ottenhoff TH, et al. Anti-inflammatory M2 type macrophages characterize metastasized and tyrosine kinase inhibitor-treated gastrointestinal stromal tumors. *Int J Cancer.* (2010) 127(4):899–909. doi: 10.1002/ijc.25113

134. Rini BI, Stenzl A, Zdrojowy R, Kogan M, Shkolnik M, Oudard S, et al. IMA901, a multipeptide cancer vaccine, plus sunitinib versus sunitinib alone, as first-line therapy for advanced or metastatic renal cell carcinoma (IMPRINT): a multicentre, open-label, randomised, controlled, phase 3 trial. *Lancet Oncol* (2016) 17(11):1599–611. doi: 10.1016/S1470-2045(16)30408-9

135. Li X, Wang X, Zhao C, Shao L, Lu J, Tong Y, et al. From one to all: selfassembled theranostic nanoparticles for tumor-targeted imaging and programmed photoactive therapy. *J Nanobiotechnol* (2019) 17(1):23. doi: 10.1186/s12951-019-0450-x

136. Mainini F, De Santis F, Fuca G, Di Nicola M, Rivoltini L, Eccles M. Nanobiotechnology and immunotherapy: Two powerful and cooperative allies against cancer. *Cancers (Basel)* (2021) 13(15):3765. doi: 10.3390/cancers13153765

137. Aderem A, Underhill DM. Mechanisms of phagocytosis in macrophages. Annu Rev Immunol (1999) 17:593–623. doi: 10.1146/annurev.immunol.17.1.593

138. Suk JS, Xu Q, Kim N, Hanes J, Ensign LM. PEGylation as a strategy for improving nanoparticle-based drug and gene delivery. *Adv Drug Delivery Rev* (2016) 99 (Pt A):28–51. doi: 10.1016/j.addr.2015.09.012

139. Li B, Yuan Z, Zhang P, Sinclair A, Jain P, Wu K, et al. Zwitterionic nanocages overcome the efficacy loss of biologic drugs. *Adv Mater* (2018) 30(14):e1705728. doi: 10.1002/adma.201705728

140. Ott PA, Hu-Lieskovan S, Chmielowski B, Govindan R, Naing A, Bhardwaj N, et al. A phase ib trial of personalized neoantigen therapy plus anti-PD-1 in patients with advanced melanoma, non-small cell lung cancer, or bladder cancer. *Cell* (2020) 183 (2):347–62.e24. doi: 10.1016/j.cell.2020.08.053

141. Massarelli E, William W, Johnson F, Kies M, Ferrarotto R, Guo M, et al. Combining immune checkpoint blockade and tumor-specific vaccine for patients with incurable human papillomavirus 16-related cancer: A phase 2 clinical trial. *JAMA Oncol* (2019) 5(1):67–73. doi: 10.1001/jamaoncol.2018.4051

142. Le QV, Suh J, Choi JJ, Park GT, Lee JW, Shim G, et al. *In situ* nanoadjuvantassembled tumor vaccine for preventing long-term recurrence. *ACS Nano* (2019) 13 (7):7442–62. doi: 10.1021/acsnano.9b02071

143. Rafiq S, Yeku OO, Jackson HJ, Purdon TJ, van Leeuwen DG, Drakes DJ, et al. Targeted delivery of a PD-1-blocking scFv by CAR-T cells enhances anti-tumor efficacy in vivo. *Nat Biotechnol* (2018) 36(9):847–56. doi: 10.1038/nbt.4195

144. Sterner RC, Sterner RM. CAR-T cell therapy: current limitations and potential strategies. *Blood Cancer J* (2021) 11(4):69. doi: 10.1038/s41408-021-00459-7

145. Nawaz W, Xu S, Li Y, Huang B, Wu X, Wu Z. Nanotechnology and immunoengineering: How nanotechnology can boost CAR-T therapy. *Acta Biomater* (2020) 109:21–36. doi: 10.1016/j.actbio.2020.04.015

146. Siriwon N, Kim YJ, Siegler E, Chen X, Rohrs JA, Liu Y, et al. CAR-T cells surface-engineered with drug-encapsulated nanoparticles can ameliorate intratumoral T-cell hypofunction. *Cancer Immunol Res* (2018) 6(7):812–24. doi: 10.1158/2326-6066.CIR-17-0502

147. Bai C, Gao S, Hu S, Liu X, Li H, Dong J, et al. Self-assembled multivalent aptamer nanoparticles with potential CAR-like characteristics could activate T cells and inhibit melanoma growth. *Mol Ther Oncol* (2020) 17:9–20. doi: 10.1016/j.omto.2020.03.002

148. Reinhard K, Rengstl B, Oehm P, Michel K, Billmeier A, Hayduk N, et al. An RNA vaccine drives expansion and efficacy of claudin-CAR-T cells against solid tumors. *Science* (2020) 367(6476):446–53. doi: 10.1126/science.aay5967

149. Turksen K, Troy TC. Claudin-6: a novel tight junction molecule is developmentally regulated in mouse embryonic epithelium. *Dev Dyn* (2001) 222 (2):292–300. doi: 10.1002/dvdy.1174

150. Ma L, Dichwalkar T, Chang JYH, Cossette B, Garafola D, Zhang AQ, et al. Enhanced CAR-T cell activity against solid tumors by vaccine boosting through the chimeric receptor. *Science* (2019) 365(6449):162–8. doi: 10.1126/science.aav8692

151. Zhang C, Ma Y, Zhang J, Kuo JC, Zhang Z, Xie H, et al. Modification of lipidbased nanoparticles: An efficient delivery system for nucleic acid-based immunotherapy. *Molecules* (2022) 27(6):1943. doi: 10.3390/molecules27061943

152. Shive MS, Anderson JM. Biodegradation and biocompatibility of PLA and PLGA microspheres. *Adv Drug Delivery Rev* (1997) 28(1):5–24. doi: 10.1016/s0169-409x(97)00048-3

153. Goodsell DS, Olson AJ. Structural symmetry and protein function. Annu Rev Biophys Biomol Struct (2000) 29:105–53. doi: 10.1146/annurev.biophys. 29.1.105

154. Larson N, Greish K, Bauer H, Maeda H, Ghandehari H. Synthesis and evaluation of poly(styrene-co-maleic acid) micellar nanocarriers for the delivery of tanespimycin. *Int J Pharm* (2011) 420(1):111–7. doi: 10.1016/j.ijpharm.2011. 08.011

155. Chang R, Zou Q, Zhao L, Liu Y, Xing R, Yan X. Amino-Acid-Encoded supramolecular photothermal nanomedicine for enhanced cancer therapy. *Adv Mater* (2022) 34(16):e2200139. doi: 10.1002/adma.202200139

156. Xu Y, Wang Y, Yang Q, Liu Z, Xiao Z, Le Z, et al. A versatile supramolecular nanoadjuvant that activates NF-kappaB for cancer immunotherapy. *Theranostics* (2019) 9(11):3388–97. doi: 10.7150/thno.34031

157. Walkey CD, Olsen JB, Guo H, Emili A, Chan WC. Nanoparticle size and surface chemistry determine serum protein adsorption and macrophage uptake. *J Am Chem Soc* (2012) 134(4):2139–47. doi: 10.1021/ja2084338

158. Morris C, Glennie SJ, Lam HS, Baum HE, Kandage D, Williams NA, et al. A modular vaccine platform combining self-assembled peptide cages and immunogenic peptides. *Adv Funct Mater* (2019) 29(8):1807357. doi: 10.1002/adfm.201807357