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## Multifaceted involvements of Paneth cells in various diseases within intestine and systemically

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Serving as the guardians of small intestine, Paneth cells (PCs) play an important role in intestinal homeostasis maintenance. Although PCs uniquely exist in intestine under homeostasis, the dysfunction of PCs is involved in various diseases not only in intestine but also in extraintestinal organs, suggesting the systemic importance of PCs. The mechanisms under the participation of PCs in these diseases are multiple as well. The involvements of PCs are mostly characterized by limiting intestinal bacterial translocation in necrotizing enterocolitis, liver disease, acute pancreatitis and graft-vs-host disease. Risk genes in PCs render intestine susceptible to Crohn's disease. In intestinal infection, different pathogens induce varied responses in PCs, and toll-like receptor ligands on bacterial surface trigger the degranulation of PCs. The increased level of bile acid dramatically impairs PCs in obesity. PCs can inhibit virus entry and promote intestinal regeneration to alleviate COVID-19. On the contrary, abundant IL-17A in PCs aggravates multi-organ injury in ischemia/ reperfusion. The pro-angiogenic effect of PCs aggravates the severity of portal hypertension. Therapeutic strategies targeting PCs mainly include PC protection, PC-derived inflammatory cytokine elimination, and substituting AMP treatment. In this review, we discuss the influence and importance of Paneth cells in both intestinal and extraintestinal diseases as reported so far, as well as the potential therapeutic strategies targeting PCs.

#### KEYWORDS

Paneth cell, Crohn's disease, necrotizing enterocolitis, liver disease, acute pancreatitis, COVID-19, therapeutic strategy

## Introduction

Paneth cells (PCs) were first discovered by Gustav Schwalbe in 1872 (1) and named by Josef Paneth in 1887 (2). Acting as a unique type of intestinal epithelial cells, PCs are derived from adjacent intestinal stem cells (ISCs) and located at the base of epithelial crypt region. In small intestine, the differentiation of PCs is conducted under the condition of

Notch signaling off and Wnt signaling on in ISCs (3). PCs can be identified by the presence of cytoplasmic abundant granules mainly including antibacterial peptides (AMPs), interleukin (IL)-17A, tumor necrosis factor (TNF)- $\alpha$  and CD95 ligand (4-6). There are many identified AMPs such as  $\alpha$ -defensin, lysozyme, and regenerating islet-derived  $3\alpha$  (REG3 $\alpha$ , REG3 $\gamma$  in mice) in PCs. PCs possess abundant endoplasmic reticulum (ER) and trans-Golgi network to realize their highly secretory nature (7). As a key component of intestinal innate immune, the functional PCs are of great importance for intestinal homeostasis. Abundant AMPs secreted by PCs control the balance of host-microbiota interactions within small intestine (8). The defects in PCs or AMP expression could lead to microbiota disorders and mucosal penetration by intestinal bacteria (9, 10). In addition, PCs support the functions of ISCs by providing several factors such as Wnt3a, epidermal growth factor (EGF), and metabolites (11-13). PCs can manipulate intestinal epithelial apoptosis by releasing CD95 ligand (14).

Considering that PCs maintain the health of intestine in multiple manners, PC dysfunction is generally involved in intestinal disorders and even diseases such as Crohn's disease (CD) and necrotizing enterocolitis (NEC) (15). The impaired unfolded protein response (UPR) and autophagy turn PCs into an origin of intestinal inflammation (16). Furthermore, the mutations in several UPR- and autophagy-related genes in PCs such as autophagy related 16 like 1 (ATG16L1) and X-box-binding protein 1 (XBP1) are identified risk factors for CD (17). Additionally, NEC pathogenesis is associated with intestinal bacterial translocation (18).

Although the presence of PCs is mainly limited in small intestine under intestinal homeostasis, PCs participate in the pathogenesis of extraintestinal diseases in addition to intestinal diseases. Large amounts of extraintestinal diseases such as liver diseases, acute pancreatitis (AP) and graft-vs-host disease (GVHD) involve the decreases in PC number and AMP expression (19–21). PC defects result in visceral hypersensitivity that induced by the expansion of intestinal *Escherichia coli*, implying the susceptibility to diseases after PC disruption (22). Ischemia/reperfusion (IR)induced multi-organ injury is mediated by the IL-17A secreted by PCs (6). Here we provide an overview of the influence and importance of PCs on various diseases within intestine and other bodily organs, as well as potential therapeutic strategies targeting PCs in these diseases.

#### PCs in inflammatory bowel disease

Inflammatory bowel disease (IBD) is a severe intestinal disease in the 21st century all over the world (23). IBD is divided into two types, ulcerative colitis (UC) and CD (24). UC is a chronic and continuous disease impacting the colon (25), whereas CD is a transmural disease occurring anywhere in the gastrointestinal tract (from mouth to anus) (26). PC abnormalities are observed in 20%-50% of CD patients and are more prevalent in pediatric CD patients than adult CD patients (27, 28). The presence of PC abnormalities is used to forecast the recurrence of CD after surgery (29). Under healthy condition, mouse PCs are limited in ileum, while human PCs normally exist in ileum as well as sporadically in cecum and ascending colon (30). The decreased expressions of  $\alpha$ -defensins (HD5 and HD6) are observed in ileal PCs of CD patients, which is attributed to the diminished Wnt ligands (enhancers of AMP expression) in monocytes and partly in ileum, highlighting the multiple regulations of PCs in CD (31).

Since PCs exhibit continuous AMP synthesis and release, functional mitochondria in PCs are required to provide energy. Recent studies have reported that active CD is associated with mitochondrial abnormalities in PCs, thus impairing PC function (32, 33). The importance of mitochondrial homeostasis in PCs is further confirmed by the fact that the level of Prohibitin 1 (PHB1), a major component protein of the inner mitochondrial membrane, is down-regulated in the mucosal biopsies from CD patients (34), and *Phb1* deficiency in PCs triggers PC defects and spontaneous ileitis in mice (35).

In addition to small intestine, PCs are occasionally observed at other sites under pathological condition, such as stomach and colon, and the phenomenon is called PC metaplasia (36). Both UC and CD patients display the occurrence of abundant metaplastic PC along the whole colon (30). It is generally accepted that metaplastic PCs tend to protect the intestine from infections. Metaplastic PCs secrete several AMPs into colonic lumen, such as  $\alpha$ -defensins, lysozyme, sPLA2 and intelectin-2 (ITLN2), which is considered as a host defense response to IBD (37-40). The activities of AMPs in colon are associated with the degree of intestinal injury in IBD. Notably, colonic expression of HD5 is significantly higher in CD than in UC, indicating that HD5 may be a potential biomarker in IBD diagnosis, a complicated process with 30% misdiagnosis (40). However, lysozyme derived from metaplastic PCs has been proved to be detrimental to colon. Lysozyme-processed and nonprocessed Ruminococcus gnavus (a CD-associated pathobiont) induced distinct immune responses in colon (41). Proinflammatory responses are triggered by Ruminococcus gnavus after lysozyme processing, whereas the transfer of Ruminococcus gnavus to Lyz1 knockout mice contributes to a type 2 immune response promoting intestinal epithelial repair (41). Considering that both CD and UC increase the risk of colorectal cancer (42), experimental studies on the role of colonic metaplastic PCs in this event should be conducted since PCs can secrete Wnt3a and EGF that might promote cancerization in intestine.

There are many identified risk genes of CD in PCs such as *ATG16L1*, *NOD2*, and *XBP1* (16, 17). Most of these risk genes are associated with the normality of autophagy and UPR, and ATG16L1 has been the most well-studied target so far. CD patients homozygous for the *ATG16L1* risk allele exhibit granule disruption and mitochondria degeneration in PCs (43). Besides, *ATG16L1* mutations also lead to ER stress in PCs as demonstrated by the enhanced levels of GPR78 and pEIF2 $\alpha$  (44). The mechanism under elevated ER stress caused by *ATG16L1* deficiency is the impaired removal of IRE $\alpha$ , an ER stress sensor (45). The protective role of ATG16L1 is further validated by the increased susceptibility to bacteria-induced inflammation in *ATG16L1*-mutated mice (46). *S. typhimurium*-induced ER stress triggers ATG16L1-mediated secretory autophagy, thus limiting bacterial penetration (47).

ATG16L1 is recruited to the plasma membrane at the bacterial entry site by NOD2 (48). In addition to autophagy, AMP expression and lysozyme sorting are also regulated by NOD2, suggesting the crucial role of NOD2 in the pathogenesis of CD (49, 50). In Caco-2 cells and the ileum of CD patients, the abnormality of NOD2 is associated with the reduced expression of  $\alpha$ -defensins, but not lysozyme (50, 51). *ATG16L1/XBP1* knockout mice develop higher level of intestinal inflammation than mice with *ATG16L1* or *XBP1* deletion, pointing out the compensatory interaction between autophagy and UPR (16). The loss of XBP1 and pEIF2 $\alpha$ , two key components of UPR, impairs PC homeostasis as well (52, 53). These findings suggest that autophagy and UPR in PCs may be potential therapeutic targets for CD.

CD is associated with the decreased expression of caspase-8 and the increased occurrence of necroptosis in PCs (54, 55). PC necroptosis leading to PC loss may be the reason for the reduced AMP expression mentioned above in CD. The expression of mixed lineage kinase domain-like protein (MLKL), the executor of necroptosis, is positively correlated to disease activity of CD ileitis (56). Abundant expression of receptor-interacting protein 3 (RIP3), another key hub of necroptosis, is observed in PCs from both humans and mice (55). Caspase-8-deleted PCs undergo necroptosis in mice without any treatment, and necrostatin-1 (Nec-1, an inhibitor of RIP1-mediated necroptosis) rescues the PC necroptosis induced by TNF- $\alpha$  (55). CD patients with X-linked inhibitor of apoptosis protein (XIAP) mutations display fewer PCs than normal CD patients (57). In XIAP knockout mice, PC loss is rescued by RIP3 silencing and Nec-1 administration intraperitoneally, suggesting that PC necroptosis is the reason for PC loss (57). In addition, ATG16L1 also suppresses necroptosis through maintaining mitochondrial functions in TNF-α-treated intestinal organoids (58). The elevated level of IFN- $\lambda$  is detected in serum and inflamed ileum of CD patients, and it is mainly located at ileal PCs. IFN- $\lambda$  treatment enhances MLKL expression, thus rendering PCs sensitive to necroptosis (56). PC necroptosis not only weakens the function of PCs, but also might trigger the release of inflammatory medium such as ATP and mitochondrial DNA, thus aggravating inflammation. Strategies to control PC necroptosis may be developed to prevent CD ileitis.

CD can induce robust cell apoptosis in crypt regions of ileum (59). Recent study has demonstrated that PCs acting as phagocytes remove the apoptotic cells in intestinal crypts, avoiding the occurrence of inflammation (60). Therefore, PC loss in CD may weaken the engulfment and removal of apoptotic cells in crypts, which requires further experimental demonstration. The accumulation of apoptotic cells in crypts may also lead to inflammation, thus impairing ISC niche and aggravating CD.

In addition to genetic factors, environmental risk factors such as smoking, western diet and, alcohol play an important role in CD pathogenesis (61). *ATG16L1*-mutated CD patients display more abnormal PCs after smoking (62). The treatments of these environmental risk factors in mice lead to PC dysfunction in manners of microbiota alterations, ER stress induction and AMP inhibition (63–65). Activating transcription factor 4 (ATF4) is down-regulated in the inflamed intestine from CD and UC patients (66). ATF4 is responsible for the uptake of glutamine promoting AMP expression, suggesting the importance of alimentary supplementation in IBD (66). However, the involvements of PCs in CD induced by environmental risk factors are poorly understood and still requires further investigation in CD patients. The detailed information about the association between PCs and CD is provided by Wehkamp and Stange (67).

#### PCs in necrotizing enterocolitis

NEC is a common gastrointestinal disease with devastating disorders and contributes to high morbidity and mortality in preterm infants, and it is characterized by pneumatosis intestinalis mediated by bacteria-derived gas (68). The pathogenesis of NEC is attributed to intestinal injury and damage that induced by bacterial permeation across the undeveloped epithelial mucosa (18). The decrease or absence of PCs is observed in inflamed intestine from NEC infants, indicating the possibility that PC defects could allow bacterial invasion in NEC (69-71). In addition, PC-derived EGF seems to be beneficial to alleviate NEC since it can reduce intestinal autophagy and NEC incidence in rats (72, 73). Dithizone (a selective destroyer of PCs)mediated PC loss in combination with acute Klebsiella pneumoniae infection induces severe intestinal injury similar to human NEC in immature mice (74). Notably, this method is only applied to postnatal day 14-16 (P14-P16) mice rather than P5 and P28 mice. The limitation may attribute to the fact that PC-dependent innate immunity is essential for the immature intestine of P14-P16 mice (75), highlighting the importance of PCs in preventing NEC. The immature intestine of P5 mice without PCs is protected by cathelinrelated antimicrobial peptide (CRAMP), and P28 mice possess mature small intestine (75). In this mouse NEC model, dithizone and Klebsiella pneumoniae can be substituted with diphtheria toxin/ PC-DTR mice and other bacteria (Klebsiella Zea mays and Bacillus cereus) respectively (18). PC deletion-induced NEC model mice display the increased Enterobacteriaceae species participating in human NEC development, as well as the decreased Helicobacteraceae species in cecum (10).

Antibiotic is generally used to treat infection. However, the prolonged antibiotic exposure to preterm infants elevates the NEC incidence (76, 77). Subsequent study conducted with neonatal mice has demonstrated that intraperitoneal antibiotic treatment after birth (P1-P10) reduces PC number in crypts, and *Klebsiella pneumoniae* infection at P14 triggers NEC-like intestinal injury in mice after 10-day antibiotic treatment, suggesting the necessity to control antibiotic use in newborns (78). Efforts to protect intestinal PCs in infants must be conducted to prevent NEC attack.

### PCs in intestinal infection

PCs residing in crypt bottoms are sensitive to intestinal microorganisms mainly including bacteria, virus and parasite. Intestinal bacteria directly stimulate the expression of AMPs *via* toll-like receptor (TLR)-MyD88 axis (79). There are multiple TLR ligands in microorganisms, and their abilities to trigger

degranulation of PCs are different. Polyinosinic-polycytidylic acid (TLR3 agonist) and CpG-oligodeoxynucleotide (TLR9 agonist) treatments induce dramatic degranulation of PCs in mice, while LPS (TLR4 agonist) and flagellin (TLR5 agonist)-induced degranulation is dilatory and TNF- $\alpha$ -dependent (80). This degranulation process of PCs also occurs in intestinal organoids (81). Under infectious conditions, lysozyme secretion is achieved by secretory autophagy, an alternative secretion approach, rather than degranulation (47). Functional PCs are required for the maintenance of intestinal health, and PC defects render intestine susceptible to microorganism infections (57, 82, 83).

Intestinal microorganisms have various effects on PCs, highlighting the importance of host-microorganism interactions in PC development. Enterotoxigenic *Escherichia coli* and *Salmonella typhimurium* infections lead to the increases in PC number and AMP expression (84, 85). The expansion of PC attributes to Wnt signaling activation (84) mediated by *Salmonella* protein AvrA (86). *Listeria monocytogenes* inhibits intestinal Notch signaling to facilitate PC differentiation (87). In *Clostridium difficile* infection, intestinal epithelial Stat5 signaling activates Wnt/ $\beta$ -catenin signaling in ISCs, thus promoting PC differentiation and intestinal regeneration (88). In human chronic gastritis induced by *Helicobacter pylori* infection,  $\alpha$ -defensins secreted by metaplastic PCs in stomach have a bactericidal effect on *Helicobacter pylori*. These findings suggest that bacterial infections lead to PC activation rather than PC defects.

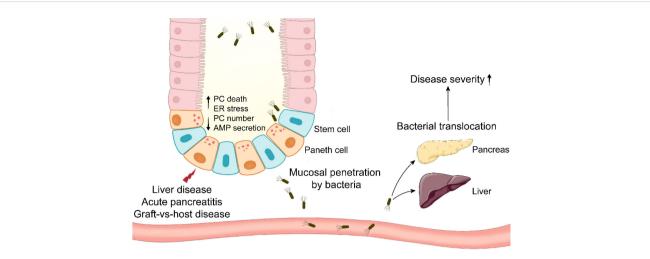
Compared with bacterial infections, viral infections tend to impair PCs. Early simian immunodeficiency virus (SIV) is localized in close proximity to PCs after infection (4). PCs express pro-inflammatory IL- $1\beta$  impairing intestinal epithelial barrier in response to SIV infection, which precedes the IFN antiviral response, suggesting that PCs may play an important role in amplifying intestinal inflammation (4). Transmissible gastroenteritis virus-infected piglets exhibit PC mitochondria damage that further impairs Notch signaling and ISC functions (89). As for parasite, helminth infection doubles PC number in mice (90), and Toxoplasma gondii infection results in PC mitochondria damage and PC death depending on mTORC1 signaling (91). In consideration of the antibacterial and ISCsupporting roles of PCs, targeting PCs may be a practical strategy to alleviate viral and parasitic infections.

#### PCs in liver disease

Liver disease represents one of the major causes of human being death in the world (92). The incidences of nonalcoholic fatty liver disease (NAFLD) and alcohol-related liver disease (ALD) are increasing these years, which results in the morbidity of liver cirrhosis (LC) and even cancer (93–95). The mouse NAFLD model is established with high-fat diet (HFD) treatment, and NAFLD mice exhibit the down-regulated expression of AMPs (19). The reduction of AMPs leads to the emergence of LPSpositive cells in small intestinal mucosa and liver (19). PC disruption induced by vitamin D deficiency increases the abundance of ileal *Helicobacter hepaticus* (a known hepatic pathogen) and bacterial translocation, which worsens hepatic steatosis and inflammation during HFD treatment (96). Oral administration of DEFA5 reduces the abundance of *Helicobacter hepaticus* in ileum and resolves hepatic steatosis and inflammation (96). However, these results are insufficient to demonstrate the role of PCs in NAFLD since vitamin D has extensive functions *in vivo* and vitamin D deficiency can affect other organs in addition to intestine. Contradictorily, PC deletion caused by dithizone alleviates HFD-induced hepatic lipid accumulation by upregulating the abundance of *Bacteroides* (97). *Bacteroides* promotes the biosynthesis of L-methionine and tetrahydrofolate alleviating hepatic steatosis (98, 99). These inverse results suggest that more studies should be conducted to confirm the role of PCs in NAFLD.

Mouse alcoholic hepatitis (AH) model is established with alcohol gavage, whereas the effect of alcohol on PCs is varied among different parts of gastrointestinal tract. The patients with massive alcohol stimulation display the enhanced expression of HD5 and HD6 in metaplastic PCs as well as the activation of Wnt signaling in antrum (100). Chronic alcohol treatment dramatically boosts the number of PCs in the proximal small intestine (63). However, this effect of alcohol is reversed in ileum since the reduced PC granules and AMP expression are observed in the ileum from alcohol-treated mice (101). In addition, AH patients exhibit the elevated plasmatic REG3a level which is further boosted in AH patients died within 30 days (102). REG3a secreted by PCs can maintain intestinal barrier integrity, and its translocation to blood indicates the changed intestinal permeability. Plasmatic REG3 $\alpha$ level is correlated with AH severity, hepatic bacterial translocation and inflammation in AH patients, suggesting that REG3 $\alpha$  could be regarded as a potential biomarker of AH (102). The involvements of PCs in AH are further confirmed by the facts that  $\alpha$ -defensin deficiency and zinc deprivation aggravates intestinal disorders and hepatic inflammation in alcohol-treated mice.

LC is another relatively grievous liver disease. LC patients exhibit the decreased expression of  $\alpha$ -defensins and increased plasmatic level of LPS (103). Notably, the  $\alpha$ -defensin expression is lower in decompensated LC patients than in compensated LC patients (103). The abnormality of  $\alpha$ -defensin leads to hepatic bacterial translocation in LC patients (104). Similarly, hepatic bacterial translocation and the reduced AMP expression are observed in LC rats (105). Recent study has reported that PC disruption is associated with the impaired production of hepatic 25-hydroxyl vitamin D in LC mice, and the loss of intestinal vitamin D receptor aggravates the severity of LC (106). These studies highlight the importance of intestine-liver axis in systematic homeostasis maintenance. HD5 administration and fecal microbiota transplantation mitigate the severe symptoms in LC mice (106). In addition to these major liver diseases, the involvements of PCs in liver are also verified by PC metaplasia in uncommon cystadenomas of liver and extrahepatic bile ducts (107). The decreased AMP expression and enhanced bacterial translocation occur in rats with acute liver failure as well (108). These findings suggest that PCs can secrete AMPs limiting bacterial translocation to alleviate liver diseases (Figure 1), highlighting a potential role of PCs in the prevention of liver disease.



#### FIGURE 1

Paneth cells in systemic diseases involving bacterial translocation. Paneth cells are impaired in systemic diseases such as liver disease, acute pancreatitis, and graft-vs-host disease. Paneth cell disruption results in mucosal penetration by intestinal bacteria. The bacterial translocation to systemic organs aggravates the disease severity eventually.

### PCs in acute pancreatitis

AP is a potentially lethal disease characterized by its unpredictability and high incidence (109). The meta-analysis has revealed that the global incidence of AP has increased in the late 20th and the early 21st century, particularly in North America and Europe (110). AP patients exhibit the defects in PC number and AMP expression, and similar phenomenon is observed in experimental AP mice and rats (20, 111). In AP rats, intestinal microbiota disorders and PC disruption may be the reasons for the compromised intestinal epithelial barriers, and there is a negative correlation between Escherichia-Shigella level and lysozyme expression (111). Hypertriglyceridemia (HTG) is a common risk factor of AP, and it results in approximately 10% incidence of AP attack (112). HTG worsens the intestinal mucosa permeability and AMP expression in AP rats. To examine the role of PCs in AP pathogenesis, dithizone is utilized to disturb PCs in AP mice and rats. Dithizone-induced PC disruption in AP rats triggers severe inflammation through several mechanisms such as ER stress activation, intestinal microbiota alteration and short-chain fatty acids reduction (113, 114). Longterm (2 weeks) deletion of PCs by dithizone exacerbates the inflammation in pancreas and ileum from AP mice, and it leads to an increase in the pathogenic Helicobacter and a decrease in the probiotic Blautia (20). Although TNF- $\alpha$  from dithizone-treated PCs has been proved beneficial to intestinal cell proliferation, PCs themselves seem to be more important to intestinal homeostasis in AP. Notably, lysozyme administration mitigates the disorders in pancreas, ileal mucosa and intestinal microbiota in AP mice with dithizone treatment. After receiving transplant of feces from lysozymetreated mice, antibiotic-treated AP mice exhibit the improved symptoms in pancreas and ileum, while transplant of feces from dithizone-treated mice has a reverse effect on the symptoms, suggesting the involvement of PCs in AP via intestinal microbiota regulation (20). These studies suggest that stabilizing PCs and microbiota could be a feasible strategy for the therapy of AP (Figure 1).

## PCs in graft-vs-host disease

After allogeneic hematopoietic stem cell transplantation, GVHD is a frequent complication due to the immune reaction of allogeneic T cells in transplant against host antigens (115). The allogeneic T cells instinctively attack host intestinal cells, mainly including ISC, goblet cells and PCs (115). The reduced PC number in GVHD patients is correlated with clinical severity and nonrelapse mortality (21). Similar to AH, the level of serum REG3 $\alpha$ , a specific GVHD biomarker, is boosted and also correlated with nonrelapse mortality in GVHD patients, whereas the expression of REG3 $\alpha$  in small intestine is reduced (116). Therefore, whether there are metaplastic PCs in other sites such as stomach and colon of GVHD patients should be examined in the future work. The patients with severe GVHD (stage 2-4) display lower expression of  $\alpha$ -defensins and REG3 $\alpha$  in small intestine, while higher expression of them in large intestine, compared to mild GVHD patients (stage 0-1) (117). In GVHD, recipient single nucleotide polymorphisms (SNPs) in DEFA5 (gene for HD5) are involved in GVHD pathogenesis. It is identified that DEFA5 rs4415345G and rs4610776A can effectively prevent GVHD stage 2-4 (118). DEFA5 rs4415345G elevates the abundance of intestinal Odoribacter splanchnicus (a butyric acid-producing bacterium), which may decrease the incidence of GVHD stage 2-4 (119). Odoribacter splanchnicus can inhibit the production of inflammatory cytokines (120), suggesting the fact that DEFA5 rs4415345G possesses strong anti-inflammatory activity. GVHD mice exhibit the decreases in AMP expression and fecal cryptdin-1 level (121, 122). The reduced  $\alpha$ -diversity and abundance of *Escherichia coli* are observed in GVHD mice (121, 122). In addition, the level of Escherichia coli is enhanced in MLN and liver from GVHD mice, and antibiotic treatment significantly alleviates the severity of GVHD (122) (Figure 1).

In view of the crucial role of PCs in GVHD, treatments targeting PCs may be effective methods to attenuate GVHD severity. This

hypothesis is confirmed by direct REG3 $\gamma$  or cryptdin-4 supplementation and IL-22 treatment (in a REG3 $\gamma$ -dependant manner) (116, 123). R-Spondin1, a Wnt signaling agonist, can promote PC differentiation to relieve GVHD (123). A decrease in glucagon-like peptide 2 (GLP-2) derived from intestinal L cells is observed in GVHD patients and mice, and teduglutide (a GLP-2 agonist) treatment facilitates PC regeneration, thus benefiting AMP expression and microbiota control against GVHD (124). In contrast to the enhanced level of IFN- $\lambda$  inducing PC necroptosis in IBD, IFN- $\lambda$  in GVHD has no effect on PCs, suggesting the ambiguous role of IFN- $\lambda$  on PCs under different conditions (125). The novel therapeutic strategies on PCs or AMPs could be effectively applied in GVHD treatment.

#### PCs in diabetes

Diabetes serves as a widespread danger to public health, and the morbidity of diabetes is dramatically ascending all over the world. Diabetes is associated with insufficient insulin secretion and insulin resistance (126). Increasing evidence has demonstrated that diabetic patients are susceptible to intestinal pathogen infections (127-129). However, the alterations of PC-derived AMPs in diabetic mice are different in several studies. On one hand, streptozotocin (STZ)induced diabetes impairs AMP expression in both proximal and distal small intestine, thus leading to the increased bacterial burden and lowered bactericidal activity in intestine (130). The deficiency of endogenous insulin may inhibit AMP expression in diabetic mice since exogenous insulin treatment restores AMP expression (130). On the other hand, STZ-treated mice display enhanced mRNA and protein levels of lysozyme, which attributes to the impaired signal transduction of Notch1/NICD in the small intestine (131). PC number is boosted in small intestine from diabetic mice, and the mechanisms involve the inactivation of Notch/Hes1 signal pathway in ISCs and the activation of insulin receptor-A isoform in PCs (130-132). Further investigation has revealed that the number of Lgr5 positive ISCs is increased in STZ-induced diabetic mice, and Lgr5 positive ISCs isolated from diabetic mice can differentiate into larger proportion of PC lineage compared to those isolated from control mice (133). In addition, insulin resistance induced by S961, an effective antagonist of insulin receptor, impairs AMP expression and granule integrity in PCs, thus leading to the enhanced intestinal permeability and the occurrence of low-degree inflammation (134). Although diabetes involves PCs, the role of PCs in diabetes is still indistinct and requires massive investigation in clinical trials and mice with PC deletion or AMP treatment.

#### PCs in obesity

Obesity has become a worldwide epidemic during the last few decades. The current prevalence of obesity in America is 18.5% among youth and 39.6% among adults (135), and that in Europe is 15.3% to 25.6% among youth (136). Obese people tend to exhibit PC abnormality (64, 137). HFD-induced obesity triggers PC defects, microbiota composition alterations and low-grade intestinal

inflammation in mice, and the alterations of PCs and microbiota occur prior to intestinal inflammation (138).

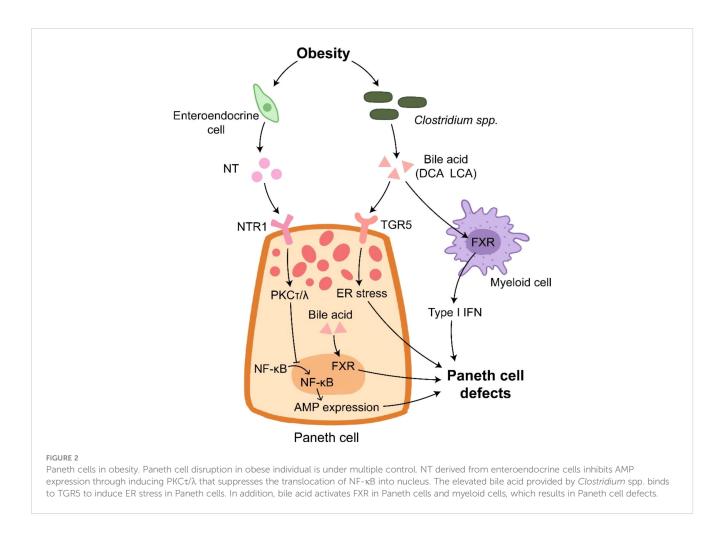
PCs possess abundant ER and high protein biosynthesis activity to support their highly secretory nature, and PCs are susceptible to ER stress triggered by the accumulation of misfolded or unfolded proteins in ER (139). ER stress can induce UPR activation facilitating the restoration of ER homeostasis. Obese people display the reduced protein levels of HD5 and lysozyme and the elevated gene expression of them (137). Notably, the activated UPR and ER stress are present in the jejunal PCs from obese people, and UPR activation is negatively correlated with lysozyme level, suggesting that the impaired protein biosynthesis of AMPs is the reason for the discordance between AMP protein level and mRNA expression (137).

Bile acid plays an important role in HFD-induced PC defects. HFD treatment enhances the level of bile acid, and bile acid can bind to G protein-coupled bile acid receptor (TGR5) highly expressed on the membrane of PCs (140). The elevated bile acid induce ER stress to impair PC functions and AMP expression, which can be rescued by pretreatment with ER stress inhibitor 4PBA or bile acid binder cholestyramine (140). Certain bacteria, such as *Clostridium* spp., are major sources of deoxycholic acid (DCA) and lithocholic acid (LCA) that are enriched in the ileum of western diet or HFD-treated mice (64, 140). *Clostridium*-mediated DCA production activates farnesoid X receptor (FXR) pathways in PCs and myeloid cells, which leads to PC defects (64).

Plasma neurotensin (NT), an enteroendocrine cell-derived hormone, is also involved in HFD-induced PC defects since NT deficiency alleviates the impaired PC function in HFD-treated mice (141). NT binding to NT receptor 1 (NTR1) activates PKC $\tau/\lambda$  to inhibit the nuclear translocation of p65, thus impairing AMP expression. Besides, the deletion of intestinal epithelial insulin receptor decreases the elevated AMP mRNA expression induced by HFD treatment, but it has no effect on the number of lysozyme positive cells in jejunum (142). These findings suggest the impaired function of PC under obese condition (Figure 2). HD5 treatment reduces circulating cholesterol and fatty acids in obese mice (143), implying the feasibility that utilizes AMP as a complemental method for obesity therapy.

#### PCs in ischemia reperfusion

IR is a serious condition of prolonged inadequate organic blood supply and subsequent sudden restoration of blood flow, and IR causes catastrophic and deadly injury to many organs such as intestine, liver and kidney (144, 145). Approximately 30% deaths of ischemic patients attribute to IR injury (146). UPR activation and PC apoptosis induced by ER stress are observed during intestinal IR injury in humans and rats, and PC disruption by dithizone further exacerbates intestinal epithelial permeability and inflammation in the intestine of IR rats (147). IR injury dramatically induces PC degranulation, and IL-17A in PC granules is responsible for multiorgan injury during IR. IL-17A neutralization or PC deletion protects organs from inflammation and injury induced by IR (6, 148). The detrimental role of PC-derived IL-17A in IR is achieved



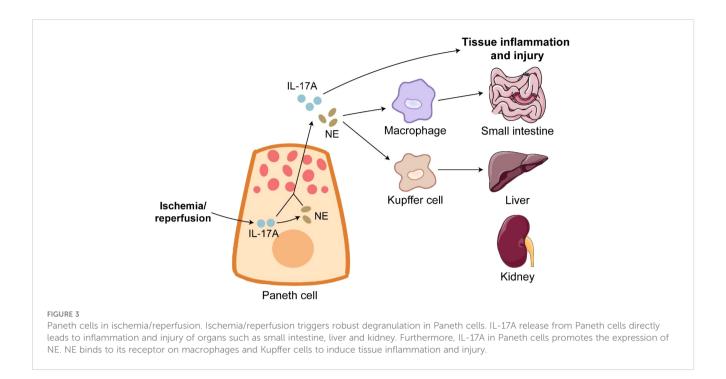
by macrophage-mediated transportation (149). PC hyperplasia induced by *TLR9* deletion worsens multi-organ inflammation and injury after IR in an IL-17A-dependant manner (150, 151). As expected, intravenous treatment of IL-17A neutralizing antibody effectively alleviates IR-induced severe inflammation and injury in *TLR9* knockout mice (150). The expression of tyrosine hydroxylase, a key enzyme of norepinephrine (NE) synthesis, is detected in human and mouse PC. NE release is driven by IL-17A in PCs (152). NE activates intestinal macrophages and Kupffer cells to damage multiple organs after IR, and the block of  $\alpha$ -adrenergic receptor significantly alleviates IR-induced injury in mice (152). The modulation of PC-derived IL-17A and NE could have therapeutic value for the treatment of IR-mediated systemic complications (Figure 3).

### PCs in COVID-19

COVID-19 caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) has rapidly risen to a threatening and lethal epidemic worldwide (153). Although COVID-19 is a respiratory disease characterized by cough and severe pneumonia, gastrointestinal dysbiosis such as diarrhea and abdominal pain

also occurs in COVID-19 patients (154, 155). SARS-CoV-2 infection inhibits ZO-3 and claudin-1 expression to impair intestinal epithelial integrity (156). PCs express certain genes related to SARS-CoV-2 entry, such as angiotensin-converting enzyme 2 (ACE2) and serine protease transmembrane protease 2 (TMPRSS2), thus rendering PCs susceptible to SARS-CoV-2 infection (157). SARS-CoV-2 mainly targets enterocytes and PCs in the intestine (156). SARS-CoV-2 infection increases PC number in the small intestine of rhesus macaques at 7-10 dpi (158). PCs exhibit the activated gene expression of factors related to cell cytoskeleton organization and epithelial cell differentiation at 3-7 dpi, which may contribute to the enhanced expressions of ZO-1 and claudin-1 at 10 dpi, suggesting the important role of PCs in intestinal epithelial repair during SARS-CoV-2 infection (158) (Figure 4). However, PC-mediated epithelial repair leads to rectal viral shedding accelerating viral transmission (158). In addition, aging PCs disturb intestinal ISC functions, thus indirectly impairing the differentiation of M cells (159). This may explain why the olds are more susceptible to COVID-19 than the youths (160).

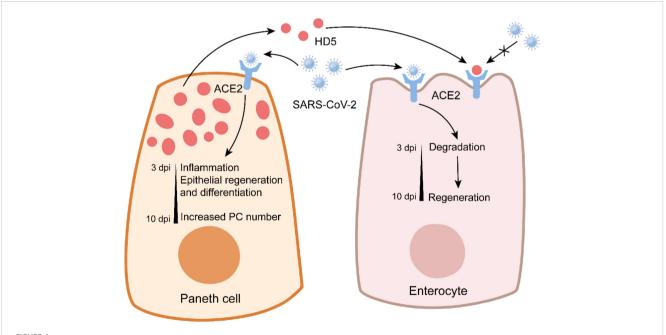
HD5 plays a critical role in the inhibition of SARS-CoV-2 infection (Figure 4). As the most abundant  $\alpha$ -defensin secreted by PCs, HD5 can bind to the ligand-binding domain of ACE2 with a



high affinity (161). In Caco-2 cells, HD5 pretreatment decreases SARS-CoV-2 binding to ACE2, suggesting the competition for ACE2 between HD5 and SARS-CoV-2 (161). It is worth noting that HD5 has no protective effect on SARS-CoV-2 infection when provided post-infection or as precursor form (162). These studies suggest the beneficial role of PCs in COVID-19. More information about the effect of AMPs (from PCs or not) on SARS-CoV-2 is recently discussed by Ali et al. in detail (163).

## PCs in portal hypertension

Portal hypertension (PH) is associated with the increased portal pressure induced by elevated resistance of blood stream (164). Blocking intestinal angiogenesis has a beneficial effect on the alleviation of PH (165). The reduced intestinal angiogenesis and PC number in GF mice imply a potential association between PCs and PH (166). PC deletion by dithizone dramatically decreases the



#### FIGURE 4

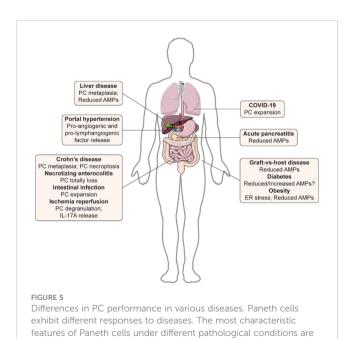
Paneth cells in COVID-19. In Paneth cells, SARS-CoV-2 infection triggers the expression of genes related to inflammation and epithelial repair, and it enhances the number of Paneth cells at later stage. In enterocytes, SARS-CoV-2 infection leads to the conversion from degradation to regeneration over time. Notably, Paneth cell-derived HD5 binds to ACE2 to prevent SARS-CoV-2 infection.

angiogenesis in small intestinal homeostasis (167). Mouse PH model is established with partial portal vein ligation (PPVL). Hassan et al. firstly reported that portal pressure and portosystemic shunts are weakened in PC-deleted mice after PPVL (168). PC disruption weakens intestinal and mesenteric angiogenesis in PPVL mice, which attributes to the reduced expression of angiogenic genes (168). Furthermore, intestinal microbial signals are responsible for the induction of angiogenesis of endothelial cells (168). In addition to angiogenesis, lymphangiogenesis is also supported by PCs (169). During PPVL, PCs secrete lymphangiogenesis in response to intestinal and mesenteric lymphangiogenesis in response to intestinal microbial signals (169). These findings suggest that PC could be a potential target for therapeutic interventions of PH.

# Therapeutic strategies targeting PCs in various diseases

As mentioned above, PC defects are involved in many diseases within intestine and systemically, which worsens the severity of diseases. However, PC performance can be different in these diseases (Figure 5), suggesting that the therapeutic strategies should be also flexible. Therapeutic strategies targeting PCs mainly include three aspects: PC protection, PC-derived inflammatory cytokine elimination, and substituting AMP treatment.

PC protection is aimed to restore PC homeostasis affected by many factors such as mitochondrial abnormalities, ER stress, and cell death. Mitochondrial abnormalities are observed in PCs of CD patients (33). Mito-Tempo (a mitochondrial-targeted antioxidant) treatment improves the inflammatory response, metabolism,



apoptotic, and epithelial barrier function in the ileal biopsies from CD patients (33). ER stress in PCs is generally induced in many diseases, which leads to the abnormality of PCs (170). Efforts to inhibit ER stress could be conducted to alleviate disease severity. In obese individuals, bile acid is also an alternative target due to its ability to trigger ER stress in PCs (140). Excessive ER stress can induce PC apoptosis (171), thus lessening the number of PCs. Besides, several risk factors in CD, such as IFN- $\lambda$  and *ATG16L1*, are able to induce PC necroptosis (56, 58). Uncontrolled cell death of PCs not only leads to the loss of PCs directly, but also induces the occurrence of intestinal inflammation. Therefore, methods to preserve the functional PCs could be an effective therapy, such as the administration of drug inhibiting necroptosis and alleviating

Although PCs play an important role in intestinal homeostasis, the functional PCs seem to be detrimental to health in some cases. IL-17A and IL-1 $\beta$  are two pro-inflammatory cytokines in the granules of PCs. IR triggers the degranulation of PCs and the subsequent release of IL-17A responsible for multi-organ injury (6, 148). IL-17A neutralization or PC deletion effectively attenuates IRinduced injury (148), suggesting the feasibility of blocking the production of IL-17A in IR. In SIV infection, IL-1B production in PCs is rapidly conducted, and it is prior to AMP expression in PCs and type 1 IFN response in intestinal mucosa, suggesting that PCderived IL-1 $\beta$  may be the key origin of inflammation (4). The presences of activated IL-1 $\beta$  and caspase-1 (a key component of pyroptosis) is also observed in PCs after irradiation (172). However, whether IL-1ß neutralization or PC pyroptosis inhibition could weaken the amplification of intestinal inflammation in these diseases still remains unclear.

Since PCs can regulate the composition of intestinal microbiota via AMPs, these diseases impairing PCs generally accompany with the disorders in microbiota. For example, the abundances of pathogenic bacteria Ruminococcus gnavus and adherent-invasive Escherichia coli are elevated in ileal lumen from CD patients (41, 173), and the increased pathogenic bacteria Helicobacter and the reduced probiotic bacteria Blautia are observed in the ileocecum of AP mice (20). To improve the disordered microbiota and disease severity, fecal microbiota transplantation has been proved feasible according to the results from research on AP mice (20). PC disruption in combination of microbiota disorders results in bacterial translocation aggravating the severity of NEC, AH and AP (68, 101, 112). In most diseases involving PC disruption, PC-derived AMP treatment significantly alleviates the symptoms of diseases, suggesting the importance of PC-derived AMPs in controlling intestinal microbiota. Furthermore, utilization of antibiotic to destroy the intestinal microbiota is another effective strategy on GVHD treatment (122). However, antibiotic is not applicable to NEC since antibiotic-induced PC disruption renders intestine susceptible to NEC attack in newborns (78).

## Conclusion

ER stress.

PCs are known as the guardians of the small intestine. In this paper, we introduce the multifaceted involvements of PCs in intestinal and extraintestinal diseases. Large amounts of systemic

summarized in the figure

diseases such as CD, NEC and COVID-19, are associated with PC disruption. PCs are involved in these diseases *via* various mechanisms. PCs possess numerous risk genes related to CD, such as *ATG16L1* and *XBP1*. In addition, PC-mediated limitation of intestinal bacterial translocation is of importance for the prevention or alleviation of diseases. However, the role of PCs in IR is different from other diseases. The presence of PC-derived IL-17A aggravates the multi-organ injury induced by IR, and the removal of IL-17A or PCs has a protective effect in IR. In COVID-19, PCs can inhibit the entry of virus and promote intestinal regeneration to resist SARS-CoV-2 infection. Except for IR, AMP treatment has a beneficial effect on all diseases involving PCs. All in all, strategies to stabilize PCs could be developed to effectively intervene these diseases within intestine and systemically.

#### Author contributions

Writing - original draft: CC. Visualization: XW, LL, and HW. Writing - review & editing: CC and JP. All authors contributed to the article and approved the submitted version.

## References

1. Schwalbe G. Beiträge zur kenntniss der drüsen in den darmwandungen, in's besondere der brunner'schen drüsen. *Archiv für mikroskopische Anatomie* (1872) 8 (1):92–140. doi: 10.1007/BF02955835

2. Paneth J. Ueber die secernirenden zellen des dünndarm-epithels. Archiv für mikroskopische Anatomie (1887) 31(1):113–91. doi: 10.1007/BF02955706

3. Beumer J, Clevers H. Cell fate specification and differentiation in the adult mammalian intestine. *Nat Rev Mol Cell Biol* (2021) 22(1):39–53. doi: 10.1038/s41580-020-0278-0

4. Hirao LA, Grishina I, Bourry O, Hu WK, Somrit M, Sankaran-Walters S, et al. Early mucosal sensing of siv infection by paneth cells induces il-1 beta production and initiates gut epithelial disruption. *PloS Pathog* (2014) 10(8):e1004311. doi: 10.1371/ journal.ppat.1004311

 Seno H, Sawada M, Fukuzawa H, Morita-Fujisawa Y, Takaishi S, Hiai H, et al. Involvement of tumor necrosis factor alpha in intestinal epithelial cell proliferation following paneth cell destruction. *Scandinavian J Of Gastroenterol* (2002) 37(2):154–60. doi: 10.1080/003655202755416803

6. Lee HT, Kim M, Kim JY, Brown KM, Ham A, D'Agati VD, et al. Critical role of interleukin-17a in murine intestinal ischemia-reperfusion injury. *Am J Of Physiology-Gastrointestinal And Liver Physiol* (2013) 304(1):G12-25. doi: 10.1152/ajpgi.00201.2012

7. Clevers HC, Bevins CL. Paneth cells: Maestros of the small intestinal crypts. Annu Rev Of Physiol (2013) 75:289–311. doi: 10.1146/annurev-physiol-030212-183744

8. Cardoso MH, Meneguetti BT, Oliveira-Junior NG, Macedo MLR, Franco OL. Antimicrobial peptide production in response to gut microbiota imbalance. *Peptides* (2022) 157:170865. doi: 10.1016/j.peptides.2022.170865

9. Vaishnava S, Behrendt CL, Ismail AS, Eckmann L, Hooper LV. Paneth cells directly sense gut commensals and maintain homeostasis at the intestinal hostmicrobial interface. *Proc Natl Acad Sci USA* (2008) 105(52):20858–63. doi: 10.1073/ pnas.0808723105

10. Lueschow SR, Stumphy J, Gong H, Kern SL, Elgin TG, Underwood MA, et al. Loss of murine paneth cell function alters the immature intestinal microbiome and mimics changes seen in neonatal necrotizing enterocolitis. *PloS One* (2018) 13(10): e0204967. doi: 10.1371/journal.pone.0204967

11. Sato T, van Es JH, Snippert HJ, Stange DE, Vries RG, van den Born M, et al. Paneth cells constitute the niche for Lgr5 stem cells in intestinal crypts. *Nature* (2011) 469(7330):415-+. doi: 10.1038/nature09637

12. Rodríguez-Colman MJ, Schewe M, Meerlo M, Stigter E, Gerrits J, Pras-Raves M, et al. Interplay between metabolic identities in the intestinal crypt supports stem cell function. *Nature* (2017) 543(7645):424–7. doi: 10.1038/nature21673

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## Conflict of interest

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13. Igarashi M, Guarente L. Mtorc1 and Sirt1 cooperate to foster expansion of gut adult stem cells during calorie restriction. *Cell* (2016) 166(2):436–50. doi: 10.1016/j.cell.2016.05.044

14. Moller P, Walczak H, Riedl S, Strater J, Krammer PH. Paneth cells express high levels of Cd95 ligand transcripts - a unique property among gastrointestinal epithelia. *Am J Of Pathol* (1996) 149(1):9–13.

15. Cray P, Sheahan BJ, Dekaney CM. Secretory sorcery: Paneth cell control of intestinal repair and homeostasis. *Cell And Mol Gastroenterol And Hepatol* (2021) 12 (4):1239–50. doi: 10.1016/j.jcmgh.2021.06.006

16. Adolph TE, Tomczak MF, Niederreiter L, Ko HJ, Böck J, Martinez-Naves E, et al. Paneth cells as a site of origin for intestinal inflammation. *Nature* (2013) 503 (7475):272-6. doi: 10.1038/nature12599

17. Yang E, Shen J. The roles and functions of paneth cells in crohn's disease: A critical review. *Cell Proliferation* (2021) 54(1):e12958. doi: 10.1111/cpr.12958

18. White JR, Gong H, Pope B, Schlievert P, McElroy SJ. Paneth-Cell-Disruption-Induced necrotizing enterocolitis in mice requires live bacteria and occurs independently of Tlr4 signaling. *Dis Models Mech* (2017) 10(6):727-36. doi: 10.1242/ dmm.028589

19. Nakanishi T, Fukui H, Wang X, Nishiumi S, Yokota H, Makizaki Y, et al. Effect of a high-fat diet on the small-intestinal environment and mucosal integrity in the gutliver axis. *Cells* (2021) 10(11):3168. doi: 10.3390/cells10113168

20. Fu Y, Mei Q, Yin N, Huang Z, Li B, Luo S, et al. Paneth cells protect against acute pancreatitis *Via* modulating gut microbiota dysbiosis. *Msystems* (2022) 7(3):e0150721. doi: 10.1128/msystems.01507-21

21. Levine JE, Huber E, Hammer STG, Harris AC, Greenson JK, Braun TM, et al. Low paneth cell numbers at onset of gastrointestinal graft-Versus-Host disease identify patients at high risk for nonrelapse mortality. *Blood* (2013) 122(8):1505–9. doi: 10.1182/blood-2013-02-485813

22. Riba A, Olier M, Lacroix-Lamande S, Lencina C, Bacquie V, Harkat C, et al. Paneth cell defects induce microbiota dysbiosis in mice and promote visceral hypersensitivity. *Gastroenterology* (2017) 153(6):1594-606. doi: 10.1053/j.gastro.2017.08.044

23. Kaplan GG, Ng SC. Understanding and preventing the global increase of inflammatory bowel disease. *Gastroenterology* (2017) 152(2):313–21. doi: 10.1053/j.gastro.2016.10.020

24. Ng SC, Shi HY, Hamidi N, Underwood FE, Tang W, Benchimol EI, et al. Worldwide incidence and prevalence of inflammatory bowel disease in the 21st century: A systematic review of population-based studies. *Lancet* (2017) 390 (10114):2769–78. doi: 10.1016/s0140-6736(17)32448-0

25. Kaenkumchorn T, Wahbeh G. Ulcerative colitis: Making the diagnosis. Gastroenterol Clinics North America (2020) 49(4):655-69. doi: 10.1016/j.gtc.2020.07.001

26. Roda G, Chien Ng S, Kotze PG, Argollo M, Panaccione R, Spinelli A, et al. Crohn's disease. Nat Rev Dis Primers (2020) 6(1):22. doi: 10.1038/s41572-020-0156-2

27. Stappenbeck TS, McGovern DPB. Paneth cell alterations in the development and phenotype of crohn's disease. *Gastroenterology* (2017) 152(2):322-6. doi: 10.1053/j.gastro.2016.10.003

28. Liu T-C, Gurram B, Baldridge MT, Head R, Vy L, Luo C, et al. Paneth cell defects in crohn's disease patients promote dysbiosis. *JCI Insight* (2016) 1(8):e86907. doi: 10.1172/jci.insight.86907

29. VanDussen KL, Liu T-C, Li D, Towfic F, Modiano N, Winter R, et al. Genetic variants synthesize to produce paneth cell phenotypes that define subtypes of crohn's disease. *Gastroenterology* (2014) 146(1):200–9. doi: 10.1053/j.gastro.2013.09.048

30. Simmonds N, Furman M, Karanika E, Phillips A, Bates AWH. Paneth cell metaplasia in newly diagnosed inflammatory bowel disease in children. *BMC Gastroenterol* (2014) 14:93. doi: 10.1186/1471-230x-14-93

31. Courth LF, Ostaff MJ, Mailaender-Sanchez D, Malek NP, Stange EF, Wehkamp J. Crohn's disease-derived monocytes fail to induce paneth cell defensins. *Proc Natl Acad Sci USA* (2015) 112(45):14000–5. doi: 10.1073/pnas.1510084112

32. Khaloian S, Rath E, Hammoudi N, Gleisinger E, Blutke A, Giesbertz P, et al. Mitochondrial impairment drives intestinal stem cell transition into dysfunctional paneth cells predicting crohn's disease recurrence. *Gut* (2020) 69(11):1939–51. doi: 10.1136/gutjnl-2019-319514

33. Alula KM, Jackson DN, Smith AD, Kim DS, Turner K, Odstrcil E, et al. Targeting mitochondrial damage as a therapeutic for ileal crohn's disease. *Cells* (2021) 10(6):1349. doi: 10.3390/cells10061349

34. Theiss AL, Idell RD, Srinivasan S, Klapproth J-M, Jones DP, Merlin D, et al. Prohibitin protects against oxidative stress in intestinal epithelial cells. *FASEB J* (2007) 21(1):197–206. doi: 10.1096/fj.06-6801com

35. Jackson DN, Panopoulos M, Neumann WL, Turner K, Cantarel BL, Thompson-Snipes L, et al. Mitochondrial dysfunction during loss of prohibitin 1 triggers paneth cell defects and ileitis. *Gut* (2020) 69(11):1928–38. doi: 10.1136/gutjnl-2019-319523

36. Singh R, Balasubramanian I, Zhang L, Gao N. Metaplastic paneth cells in extraintestinal mucosal niche indicate a link to microbiome and inflammation. *Front In Physiol* (2020) 11:280. doi: 10.3389/fphys.2020.00280

37. Nonnecke EB, Castillo PA, Johansson MEV, Hollox EJ, Shen B, Lonnerdal B, et al. Human intelectin-2 (Itln2) is selectively expressed by secretory paneth cells. *FASEB J* (2022) 36(3):e22200. doi: 10.1096/fj.202101870R

38. Fahlgren A, Hammarstrom S, Danielsson A, Hammarstrom ML. Increased expression of antimicrobial peptides and lysozyme in colonic epithelial cells of patients with ulcerative colitis. *Clin And Exp Immunol* (2003) 131(1):90–101. doi: 10.1046/ j.1365-2249.2003.02035.x

39. Haapamaki MM, Gronroos JM, Nurmi H, Alanen K, Kallajoki M, Nevalainen TJ. Gene expression of group ii phospholipase A2 in intestine in ulcerative colitis. *Gut* (1997) 40(1):95–101. doi: 10.1136/gut.40.1.95

40. Williams AD, Korolkova OY, Sakwe AM, Geiger TM, James SD, Muldoon RL, et al. Human alpha defensin 5 is a candidate biomarker to delineate inflammatory bowel disease. *PloS One* (2017) 12(8):e0189551. doi: 10.1371/journal.pone.0179710

41. Yu S, Balasubramanian I, Laubitz D, Tong K, Bandyopadhyay S, Lin X, et al. Paneth cell-derived lysozyme defines the composition of mucolytic microbiota and the inflammatory tone of the intestine. *Immunity* (2020) 53(2):398–416. doi: 10.1016/j.immuni.2020.07.010

42. Wan Q, Zhao R, Xia L, Wu Y, Zhou Y, Wang Y, et al. Inflammatory bowel disease and risk of gastric, small bowel and colorectal cancer: A meta-analysis of 26 observational studies. J Of Cancer Res And Clin Oncol (2021) 147(4):1077-87. doi: 10.1007/s00432-020-03496-0

43. Cadwell K, Liu JY, Brown SL, Miyoshi H, Loh J, Lennerz JK, et al. A key role for autophagy and the autophagy gene Atg16l1 in mouse and human intestinal paneth cells. *Nature* (2008) 456(7219):259–63. doi: 10.1038/nature07416

44. Deuring JJ, Fuhler GM, Konstantinov SR, Peppelenbosch MP, Kuipers EJ, de Haar C, et al. Genomic Atg1611 risk allele-restricted paneth cell er stress in quiescent crohn's disease. *Gut* (2014) 63(7):1081–91. doi: 10.1136/gutjnl-2012-303527

45. Tschurtschenthaler M, Adolph TE, Ashcroft JW, Niederreiter L, Bharti R, Saveljeva S, et al. Defective Atg16l1-mediated removal of Ire1 alpha drives crohn's disease-like ileitis. J Of Exp Med (2017) 214(2):401-22. doi: 10.1084/jem.20160791

46. Lassen KG, Kuballa P, Conway KL, Patel KK, Becker CE, Peloquin JM, et al. Atg1611 T300a variant decreases selective autophagy resulting in altered cytokine signaling and decreased antibacterial defense. *Proc Natl Acad Sci USA* (2014) 111 (21):7741-6. doi: 10.1073/pnas.1407001111

47. Bel S, Pendse M, Wang Y, Li Y, Ruhn KA, Hassell B, et al. Paneth cells secrete lysozyme *Via* secretory autophagy during bacterial infection of the intestine. *Science* (2017) 357(6355):1047–51. doi: 10.1126/science.aal4677

48. Iida T, Yokoyama Y, Wagatsuma K, Hirayama D, Nakase H. Impact of autophagy of innate immune cells on inflammatory bowel disease. *Cells* (2019) 8 (1):7. doi: 10.3390/cells8010007

49. Wang H, Zhang X, Zuo Z, Zhang Q, Pan Y, Zeng B, et al. Rip2 is required for Nod2-mediated lysozyme sorting in paneth cells. *J Of Immunol* (2017) 198(9):3729–36. doi: 10.4049/jimmunol.1601583

50. Tan G, Zeng B, Zhi F-C. Regulation of human enteric alpha-defensins by Nod2 in the paneth cell lineage. *Eur J Of Cell Biol* (2015) 94(1):60–6. doi: 10.1016/ j.ejcb.2014.10.007

51. Wehkamp J, Harder J, Weichenthal M, Schwab M, Schaffeler E, Schlee M, et al. Nod2 (Card15) mutations in crohn's disease are associated with diminished mucosal alpha-defensin expression. *Gut* (2004) 53(11):1658–64. doi: 10.1136/gut.2003.032805

52. Cao SS, Wang M, Harrington JC, Chuang B-M, Eckmann L, Kaufman RJ. Phosphorylation of Eif2a is dispensable for differentiation but required at a posttranscriptional level for paneth cell function and intestinal homeostasis in mice. *Inflammatory Bowel Dis* (2014) 20(4):712–22. doi: 10.1097/mib.00000000000001

53. Kaser A, Lee A-H, Franke A, Glickman JN, Zeissig S, Tilg H, et al. Xbp1 links er stress to intestinal inflammation and confers genetic risk for human inflammatory bowel disease. *Cell* (2008) 134(5):743–56. doi: 10.1016/j.cell.2008.07.021

54. Pierdomenico M, Negroni A, Stronati L, Vitali R, Prete E, Bertin J, et al. Necroptosis is active in children with inflammatory bowel disease and contributes to heighten intestinal inflammation. *Am J Of Gastroenterol* (2014) 109(2):279–87. doi: 10.1038/ajg.2013.403

55. Guenther C, Martini E, Wittkopf N, Amann K, Weigmann B, Neumann H, et al. Caspase-8 regulates tnf-Alpha-Induced epithelial necroptosis and terminal ileitis. *Nature* (2011) 477(7364):335–9. doi: 10.1038/nature10400

56. Guenther C, Ruder B, Stolzer I, Dorner H, He G-W, Chiriac MT, et al. Interferon lambda promotes paneth cell death *via* Stat1 signaling in mice and is increased in inflamed ileal tissues of patients with crohn's disease. *Gastroenterology* (2019) 157 (5):1310–22. doi: 10.1053/j.gastro.2019.07.031

57. Strigli A, Gopalakrishnan S, Zeissig Y, Basic M, Wang J, Schwerd T, et al. Deficiency in X-linked inhibitor of apoptosis protein promotes susceptibility to microbial triggers of intestinal inflammation. *Sci Immunol* (2021) 6(65):eabf7473. doi: 10.1126/sciimmunol.abf7473

58. Matsuzawa-Ishimoto Y, Shono Y, Gomez LE, Hubbard-Lucey VM, Cammer M, Neil J, et al. Autophagy protein Atg16l1 prevents necroptosis in the intestinal epithelium. J Of Exp Med (2017) 214(12):3687–705. doi: 10.1084/jem.20170558

59. Dirisina R, Katzman RB, Goretsky T, Managlia E, Mittal N, Williams DB, et al. P53 and puma independently regulate apoptosis of intestinal epithelial cells in patients and mice with colitis. *Gastroenterology* (2011) 141(3):1036–45. doi: 10.1053/j.gastro.2011.05.032

60. Shankman LS, Fleury ST, Evans WB, Penberthy KK, Arandjelovic S, Blumberg RS, et al. Efferocytosis by paneth cells within the intestine. *Curr Biol* (2021) 31 (11):2469–76. doi: 10.1016/j.cub.2021.03.055

 Ramos GP, Papadakis KA. Mechanisms of disease: Inflammatory bowel diseases. Mayo Clinic Proc (2019) 94(1):155–65. doi: 10.1016/j.mayocp.2018.09.013

62. Liu T-C, Kern JT, VanDussen KL, Xiong S, Kaiko GE, Wilen CB, et al. Interaction between smoking and Atg16l1(T300a) triggers paneth cell defects in crohn's disease. *J Of Clin Invest* (2018) 128(11):5110–22. doi: 10.1172/jci120453

63. Gyongyosi B, Cho Y, Lowe P, Calenda CD, Iracheta-Vellve A, Satishchandran A, et al. Alcohol-induced il-17a production in paneth cells amplifies endoplasmic reticulum stress, apoptosis, and inflammasome-Il-18 activation in the proximal small intestine in mice. *Mucosal Immunol* (2019) 12(4):930–44. doi: 10.1038/s41385-019-0170-4

64. Liu T-C, Kern JT, Jain U, Sonnek NM, Xiong S, Simpson KF, et al. Western Diet induces paneth cell defects through microbiome alterations and farnesoid X receptor and type I interferon activation. *Cell Host Microbe* (2021) 29(6):988–1001. doi: 10.1016/j.chom.2021.04.004

65. Berkowitz L, Pardo-Roa C, Salazar GA, Salazar-Echegarai F, Miranda JP, Ramirez G, et al. Mucosal exposure to cigarette components induces intestinal inflammation and alters antimicrobial response in mice. *Front In Immunol* (2019) 10:2289. doi: 10.3389/fimmu.2019.02289

66. Hu X, Deng J, Yu T, Chen S, Ge Y, Zhou Z, et al. Atf4 deficiency promotes intestinal inflammation in mice by reducing uptake of glutamine and expression of antimicrobial peptides. *Gastroenterology* (2019) 156(4):1098–111. doi: 10.1053/j.gastro.2018.11.033

67. Wehkamp J, Stange EF. An update review on the paneth cell as key to ileal crohn's disease. *Front In Immunol* (2020) 11:646. doi: 10.3389/fmmu.2020.00646

68. Thanert R, Keen EC, Dantas G, Warner BB, Tarr PI. Necrotizing enterocolitis and the microbiome: Current status and future directions. J Of Infect Dis (2021) 223: S257–63. doi: 10.1093/infdis/jiaa604

69. McElroy SJ, Prince LS, Weitkamp J-H, Reese J, Slaughter JC, Polk DB. Tumor necrosis factor receptor 1-dependent depletion of mucus in immature small intestine: A potential role in neonatal necrotizing enterocolitis. *Am J Of Physiology-Gastrointestinal And Liver Physiol* (2011) 301(4):G656–G66. doi: 10.1152/ajpgi.00550.2010

70. McElroy SJ, Underwood MA, Sherman MP. Paneth cells and necrotizing enterocolitis: A novel hypothesis for disease pathogenesis. *Neonatology* (2013) 103 (1):10-20. doi: 10.1159/000342340

71. Coutinho HB, da Mota HC, Coutinho VB, Robalinho TI, Furtado AF, Walker E, et al. Absence of lysozyme (Muramidase) in the intestinal paneth cells of newborn infants with necrotising enterocolitis. *J Of Clin Pathol* (1998) 51(7):512–4. doi: 10.1136/jcp.51.7.512

72. Grave GD, Nelson SA, Walker WA, Moss RL, Dvorak B, Hamilton FA, et al. New therapies and preventive approaches for necrotizing enterocolitis: Report of a research planning workshop. *Pediatr Res* (2007) 62(4):510–4. doi: 10.1203/PDR.0b013e318142580a

73. Maynard AA, Dvorak K, Khailova L, Dobrenen H, Arganbright KM, Halpern MD, et al. Epidermal growth factor reduces autophagy in intestinal epithelium and in the rat model of necrotizing enterocolitis. *Am J Of Physiology-Gastrointestinal And Liver Physiol* (2010) 299(3):G614–G22. doi: 10.1152/ajpgi.00076.2010

74. Zhang C, Sherman MP, Prince LS, Bader D, Weitkamp J-H, Slaughter JC, et al. Paneth cell ablation in the presence of klebsiella pneumoniae induces necrotizing enterocolitis (Nec)-like injury in the small intestine of immature mice. *Dis Models Mech* (2012) 5(4):522–32. doi: 10.1242/dmm.009001

75. Lueschow SR, McElroy SJ. The paneth cell: The curator and defender of the immature small intestine. *Front Immunol* (2020) 11:587. doi: 10.3389/fimmu.2020.00587

76. Kuppala VS, Meinzen-Derr J, Morrow AL, Schibler KR. Prolonged initial empirical antibiotic treatment is associated with adverse outcomes in premature infants. J Of Pediatr (2011) 159(5):720–5. doi: 10.1016/j.jpeds.2011.05.033

77. Cantey JB, Pyle AK, Wozniak PS, Hynan LS, Sanchez PJ. Early antibiotic exposure and adverse outcomes in preterm, very low birth weight infants. *J Of Pediatr* (2018) 203:62–7. doi: 10.1016/j.jpeds.2018.07.036

78. Chaaban H, Patel MM, Burge K, Eckert JV, Lupu C, Keshari RS, et al. Early antibiotic exposure alters intestinal development and increases susceptibility to necrotizing enterocolitis: A mechanistic study. *Microorganisms* (2022) 10(3):519. doi: 10.3390/microorganisms10030519

79. Menendez A, Willing BP, Montero M, Włodarska M, So CC, Bhinder G, et al. Bacterial stimulation of the tlr-Myd88 pathway modulates the homeostatic expression of ileal paneth cell alpha-defensins. *J Of Innate Immun* (2013) 5(1):39–49. doi: 10.1159/000341630

80. Rumio C, Sommariva M, Sfondrini L, Palazzo M, Morelli D, Vigano L, et al. Induction of paneth cell degranulation by orally administered toll-like receptor ligands. *J Of Cell Physiol* (2012) 227(3):1107–13. doi: 10.1002/jcp.22830

81. Yokoi Y, Nakamura K, Yoneda T, Kikuchi M, Sugimoto R, Shimizu Y, et al. Paneth cell granule dynamics on secretory responses to bacterial stimuli in enteroids. *Sci Rep* (2019) 9:2710. doi: 10.1038/s41598-019-39610-7

82. Wang Y, He K, Sheng B, Lei X, Tao W, Zhu X, et al. The rna helicase Dhx15 mediates wnt-induced antimicrobial protein expression in paneth cells. *Proc Natl Acad Sci USA* (2021) 118(4):e2017432118. doi: 10.1073/pnas.2017432118

83. Burger E, Araujo A, Lopez-Yglesias A, Rajala MW, Geng L, Levine B, et al. Loss of paneth cell autophagy causes acute susceptibility to toxoplasma gondii-mediated inflammation. *Cell Host Microbe* (2018) 23(2):177–90. doi: 10.1016/j.chom.2018.01.001

84. Rodriguez NRM, Eloi MD, Huynh A, Dominguez T, Lam AHC, Carcamo-Molina D, et al. Expansion of paneth cell population in response to enteric salmonella enterica serovar typhimurium infection. *Infection And Immun* (2012) 80(1):266–75. doi: 10.1128/iai.05638-11

85. Yang X, Xiao Z, Liu F, Chen S, Tang W, Zhang D, et al. Enterotoxigenic escherichia coli infection alters intestinal immunity in mice. *Mol Med Rep* (2016) 14 (1):825–30. doi: 10.3892/mmr.2016.5302

86. Liu X, Lu R, Wu S, Sun J. Salmonella regulation of intestinal stem cells through the Wnt/Beta-catenin pathway. *FEBS Lett* (2010) 584(5):911–6. doi: 10.1016/j.febslet.2010.01.024

87. Zhou C, Zhang Y, Bassey A, Huang J, Zou Y, Ye K. Expansion of intestinal secretory cell population induced by listeria monocytogenes infection: Accompanied with the inhibition of notch pathway. *Front In Cell And Infection Microbiol* (2022) 12:793335. doi: 10.3389/fcimb.2022.793335

88. Liu R, Moriggl R, Zhang D, Li H, Karns R, Ruan H-B, et al. Constitutive Stat5 activation regulates paneth and paneth-like cells to control clostridium difficile colitis. *Life Sci alliance* (2019) 2(2):e201900296. doi: 10.26508/lsa.201900296

89. Wu A, Yu B, Zhang K, Xu Z, Wu D, He J, et al. Transmissible gastroenteritis virus targets paneth cells to inhibit the self-renewal and differentiation of Lgr5 intestinal stem cells *Via* notch signaling. *Cell Death Dis* (2020) 11(1):40. doi: 10.1038/s41419-020-2233-6

90. Khatun A, Sakurai M, Okada K, Sakai Y, Morimoto M. Detection of alphadefensin in eosinophils in helminth-infected mouse model. *J Of Veterinary Med Sci* (2018) 80(12):1887–94. doi: 10.1292/jvms.18-0601

91. Araujo A, Safronova A, Burger E, Lopez-Yglesias A, Giri S, Camanzo ET, et al. Ifn-gamma mediates paneth cell death *Via* suppression of mtor. *eLife* (2021) 10:e60478. doi: 10.7554/eLife.60478

92. Wang R, Tang R, Li B, Ma X, Schnabl B, Tilg H. Gut microbiome, liver immunology, and liver diseases. *Cell Mol Immunol* (2021) 18(1):4–17. doi: 10.1038/ s41423-020-00592-6

93. Younossi Z, Anstee QM, Marietti M, Hardy T, Henry L, Eslam M, et al. Global burden of nafld and Nash: Trends, predictions, risk factors and prevention. *Nat Rev Gastroenterol Hepatol* (2018) 15(1):11–20. doi: 10.1038/nrgastro.2017.109

94. Seitz HK, Bataller R, Cortez-Pinto H, Gao B, Gual A, Lackner C, et al. Alcoholic liver disease. *Nat Rev Dis Primers* (2018) 4(1):16. doi: 10.1038/s41572-018-0014-7

95. Wong MCS, Huang JLW, George J, Huang J, Leung C, Eslam M, et al. The changing epidemiology of liver diseases in the Asia-pacific region. *Nat Rev Gastroenterol Hepatol* (2019) 16(1):57–73. doi: 10.1038/s41575-018-0055-0

96. Su D, Nie Y, Zhu A, Chen Z, Wu P, Zhang L, et al. Vitamin d signaling through induction of paneth cell defensins maintains gut microbiota and improves metabolic disorders and hepatic steatosis in animal models. *Front In Physiol* (2016) 7:498. doi: 10.3389/fphys.2016.00498 97. Zhang S, Tun HM, Zhang D, Chau H-T, Huang F-Y, Kwok H, et al. Alleviation of hepatic steatosis: Dithizone-related gut microbiome restoration during paneth cell dysfunction. *Front Microbiol* (2022) 13:813783. doi: 10.3389/fmicb.2022.813783

98. Qiao S, Bao L, Wang K, Sun S, Liao M, Liu C, et al. Activation of a specific gut bacteroides-Folate-Liver axis benefits for the alleviation of nonalcoholic hepatic steatosis. *Cell Rep* (2020) 32(6):108005. doi: 10.1016/j.celrep.2020.108005

99. Pacana T, Cazanave S, Verdianelli A, Patel V, Min H-K, Mirshahi F, et al. Dysregulated hepatic methionine metabolism drives homocysteine elevation in dietinduced nonalcoholic fatty liver disease. *PloS One* (2015) 10(8):e0136822. doi: 10.1371/ journal.pone.0136822

100. Ostaff MJ, Schaefer C, Courth L, Stebe SRD, Ott G, Stange EF, et al. Chronic heavy alcohol use is associated with upregulated paneth cell antimicrobials in gastric mucosa. *Clin And Trans Gastroenterol* (2015) 6(7):e103. doi: 10.1038/ctg.2015.26

101. Zhong W, Wei X, Hao L, Lin T-D, Yue R, Sun X, et al. Paneth cell dysfunction mediates alcohol-related steatohepatitis through promoting bacterial translocation in mice: Role of zinc deficiency. *Hepatology* (2020) 71(5):1575–91. doi: 10.1002/hep.30945

102. Yang J, Syed F, Xia Y, Sanyal AJ, Shah VH, Chalasani N, et al. Blood biomarkers of intestinal epithelium damage regenerating islet-derived protein 3 alpha and trefoil factor 3 are persistently elevated in patients with alcoholic hepatitis. *Alcoholism-Clinical And Exp Res* (2021) 45(4):720–31. doi: 10.1111/acer.14579

103. Tsiaoussis GI, Papaioannou EC, Kourea EP, Assimakopoulos SF, Theocharis GI, Petropoulos M, et al. Expression of -defensins, Cd20+B-lymphocytes, and intraepithelial Cd3+T-lymphocytes in the intestinal mucosa of patients with liver cirrhosis: Emerging mediators of intestinal barrier function. *Digestive Dis Sci* (2018) 63 (10):2582–92. doi: 10.1007/s10620-018-5146-9

104. Kaliannan K. Compromise of -defensin function in liver cirrhosis facilitates the toxic relationship between gut permeability and endotoxemia. *Digestive Dis Sci* (2018) 63(10):2492–4. doi: 10.1007/s10620-018-5197-y

105. Teltschik Z, Wiest R, Beisner J, Nuding S, Hofmann C, Schoelmerich J, et al. Intestinal bacterial translocation in rats with cirrhosis is related to compromised paneth cell antimicrobial host defense. *Hepatology* (2012) 55(4):1154–63. doi: 10.1002/hep.24789

106. Wu P, Zhang R, Luo M, Zhang T, Pan L, Xu S, et al. Impaired 25-hydroxylation of vitamin d in liver injury suppresses intestinal paneth cell defensins, leading to gut dysbiosis and liver fibrogenesis. *Am J Of Physiology-Gastrointestinal And Liver Physiol* (2020) 319(6):G685–G95. doi: 10.1152/ajpgi.00021.2020

107. Albores-Saavedra J, Carlos Cordova-Ramon J, Chable-Montero F, Dorantes-Heredia R, Henson DE. Cystadenomas of the liver and extrahepatic bile ducts morphologic and immunohistochemical characterization of the biliary and intestinal variants. *Ann Of Diagn Pathol* (2015) 19(3):124–9. doi: 10.1016/j.anndiagpath.2015.03.001

108. Chen S, Li X, Li M, Mei Q, Huang J, Wu Z, et al. Mucosal expression of defensin-5, soluble phospholipase A2 and lysozyme in the intestine in a rat model of acute liver failure and its relationship to intestinal bacterial translocation. *Gastroenterologia Y Hepatologia* (2020) 43(6):293–300. doi: 10.1016/j.gastrohep. 2020.01.004

109. Szatmary P, Grammatikopoulos T, Cai W, Huang W, Mukherjee R, Halloran C, et al. Acute pancreatitis: Diagnosis and treatment. *Drugs* (2022) 82(12):1251–76. doi: 10.1007/s40265-022-01766-4

110. Iannuzzi JP, King JA, Leong JH, Quan J, Windsor JW, Tanyingoh D, et al. Global incidence of acute pancreatitis is increasing over time: A systematic review and meta-analysis. *Gastroenterology* (2022) 162(1):122–34. doi: 10.1053/j.gastro.2021.09.043

111. Chen J, Huang C, Wang J, Zhou H, Lu Y, Lou L, et al. Dysbiosis of intestinal microbiota and decrease in paneth cell antimicrobial peptide level during acute necrotizing pancreatitis in rats. *PloS One* (2017) 12(4):e0176583. doi: 10.1371/journal.pone.0176583

112. Valdivielso P, Ramirez-Bueno A, Ewald N. Current knowledge of hypertriglyceridemic pancreatitis. *Eur J Of Internal Med* (2014) 25(8):689–94. doi: 10.1016/j.ejim.2014.08.008

113. Liu L, Guo Y, Zheng J, Lu Y, Shen Y, Huang C, et al. Paneth cell ablation increases the small intestinal injury during acute necrotizing pancreatitis in rats. *Mol Med Rep* (2019) 20(1):473–84. doi: 10.3892/mmr.2019.10274

114. Guo Y, Huang C, Liu L, Fu X, Lu Y, Zheng J, et al. Paneth cell ablation aggravates pancreatic and intestinal injuries in a rat model of acute necrotizing pancreatitis after normal and high-fat diet. *Mediators Of Inflammation* (2019) 2019:8474523. doi: 10.1155/2019/8474523

115. Li J, Zhang X, Chen Y, Zheng Q, Zhao M, Jiang H. A promising insight: The potential influence and therapeutic value of the gut microbiota in gi gvhd. *Oxid Med And Cell Longevity* (2022) 2022:2124627. doi: 10.1155/2022/2124627

116. Zhao D, Kim Y-H, Jeong S, Greenson JK, Chaudhry MS, Hoepting M, et al. Survival signal Reg3 alpha prevents crypt apoptosis to control acute gastrointestinal graft-Versus-Host disease. *J Of Clin Invest* (2018) 128(11):4970–9. doi: 10.1172/jci99261

117. Weber D, Frauenschlaeger K, Ghimire S, Peter K, Panzer I, Hiergeist A, et al. The association between acute graft-Versus-Host disease and antimicrobial peptide expression in the gastrointestinal tract after allogeneic stem cell transplantation. *PloS One* (2017) 12(9):e0185265. doi: 10.1371/journal.pone.0185265

118. Rashidi A, Shanley R, Yohe SL, Thyagarajan B, Curtsinger J, Anasetti C, et al. Recipient single nucleotide polymorphisms in paneth cell antimicrobial peptide genes and acute graft-Versus-Host disease: Analysis of bmt ctn-0201 and-0901 samples. *Br J Of Haematology* (2018) 182(6):887–94. doi: 10.1111/bjh.15492

119. Rashidi A, Herman A, Gomes ALC, Peled JU, Jenq RR, Brereton DG, et al. An alpha-defensin gene single nucleotide polymorphism modulates the gut microbiota and may alter the risk of acute graft-Versus-Host disease. *Br J Of Haematol* (2020) 189 (5):926–30. doi: 10.1111/bjh.16458

120. Schirmer M, Smeekens SP, Vlamakis H, Jaeger M, Oosting M, Franzosa EA, et al. Linking the human gut microbiome to inflammatory cytokine production capacity. *Cell* (2016) 167(4):1125–36. doi: 10.1016/j.cell.2016.10.020

121. Eriguchi Y, Nakamura K, Hashimoto D, Shimoda S, Shimono N, Akashi K, et al. Decreased secretion of paneth cell -defensins in graft-Versus-Host disease. *Transplant Infect Dis* (2015) 17(5):702-6. doi: 10.1111/tid.12423

122. Eriguchi Y, Takashima S, Oka H, Shimoji S, Nakamura K, Uryu H, et al. Graft-Versus-Host disease disrupts intestinal microbial ecology by inhibiting paneth cell production of alpha-defensins. *Blood* (2012) 120(1):223–31. doi: 10.1182/blood-2011-12-401166

123. Hayase E, Hashimoto D, Nakamura K, Noizat C, Ogasawara R, Takahashi S, et al. R-Spondin1 expands paneth cells and prevents dysbiosis induced by graft-Versus-Host disease. *J Of Exp Med* (2017) 214(12):3507–18. doi: 10.1084/jem.20170418

124. Norona J, Apostolova P, Schmidt D, Ihlemann R, Reischmann N, Taylor G, et al. Glucagon-like peptide 2 for intestinal stem cell and paneth cell repair during graft-Versus-Host disease in mice and humans. *Blood* (2020) 136(12):1442–55. doi: 10.1182/ blood.2020005957

125. Henden AS, Koyama M, Robb RJ, Forero A, Kuns RD, Chang K, et al. Ifnlambda therapy prevents severe gastrointestinal graft-Versus-Host disease. *Blood* (2021) 138(8):722–37. doi: 10.1182/blood.2020006375

126. Edgerton DS, Kraft G, Smith M, Farmer B, Williams PE, Coate KC, et al. Insulin's direct hepatic effect explains the inhibition of glucose production caused by insulin secretion. *JCI Insight* (2017) 2(6):e91863. doi: 10.1172/jci.insight.91863

127. Gosiewski T, Salamon D, Szopa M, Sroka A, Malecki MT, Bulanda M. Quantitative evaluation of fungi of the genus candida in the feces of adult patients with type 1 and 2 diabetes - a pilot study. *Gut Pathog* (2014) 6(1):43. doi: 10.1186/s13099-014-0043-z

128. Telzak EE, Greenberg MSZ, Budnick LD, Singh T, Blum S. Diabetes-mellitus a newly described risk factor for infection from salmonella-enteritidis. *J Of Infect Dis* (1991) 164(3):538–41. doi: 10.1093/infdis/164.3.538

129. Oikarinen M, Tauriainen S, Honkanen T, Oikarinen S, Vuori K, Kaukinen K, et al. Detection of enteroviruses in the intestine of type 1 diabetic patients. *Clin And Exp Immunol* (2008) 151(1):71–5. doi: 10.1111/j.1365-2249.2007.03529.x

130. Yu T, Yang H-S, Lu X-J, Xia Z-S, Ouyang H, Shan T-D, et al. Association of bactericidal dysfunction of paneth cells in streptozocin-induced diabetic mice with insulin deficiency. *Med Sci Monitor* (2016) 22:3062–72. doi: 10.12659/msm.897270

131. Min X-H, Yu T, Qing Q, Yuan Y-H, Zhong W, Chen G-C, et al. Abnormal differentiation of intestinal epithelium and intestinal barrier dysfunction in diabetic mice associated with depressed Notch/Nicd transduction in Notch/Hes1 signal pathway. *Cell Biol Int* (2014) 38(10):1194–204. doi: 10.1002/cbin.10323

132. Ulanet DB, Ludwig DL, Kahn CR, Hanahan D. Insulin receptor functionally enhances multistage tumor progression and conveys intrinsic resistance to igf-1r targeted therapy. *Proc Natl Acad Sci USA* (2010) 107(24):10791–8. doi: 10.1073/ pnas.0914076107

133. Zhong X-Y, Yu T, Zhong W, Li J-Y, Xia Z-S, Yuan Y-H, et al. Lgr5 positive stem cells sorted from small intestines of diabetic mice differentiate into higher proportion of absorptive cells and paneth cells in vitro. *Dev Growth Differ* (2015) 57 (6):453–65. doi: 10.1111/dgd.12226

134. Gueddouri D, Cauzac M, Fauveau V, Benhamed F, Charifi W, Beaudoin L, et al. Insulin resistance per Se drives early and reversible dysbiosis-mediated gut barrier impairment and bactericidal dysfunction. *Mol Metab* (2022) 57:101438. doi: 10.1016/j.molmet.2022.101438

135. Hales CM, Fryar CD, Carroll MD, Freedman DS, Ogden CL. Trends in obesity and severe obesity prevalence in us youth and adults by sex and age, 2007-2008 to 2015-2016. Jama-Journal Of Am Med Assoc (2018) 319(16):1723–5. doi: 10.1001/jama.2018.3060

136. Garrido-Miguel M, Cavero-Redondo I, Alvarez-Bueno C, Rodriguez-Artalejo F, Moreno LA, Ruiz JR, et al. Prevalence and trends of overweight and obesity in European children from 1999 to 2016 a systematic review and meta-analysis. JAMA Pediatr (2019) 173(10):e192430. doi: 10.1001/jamapediatrics.2019.2430

137. Hodin CM, Verdam FJ, Grootjans J, Rensen SS, Verheyen FK, Dejong CHC, et al. Reduced paneth cell antimicrobial protein levels correlate with activation of the unfolded protein response in the gut of obese individuals. *J Of Pathol* (2011) 225 (2):276–84. doi: 10.1002/path.2917

138. Guo X, Li J, Tang R, Zhang G, Zeng H, Wood RJ, et al. High fat diet alters gut microbiota and the expression of paneth cell-antimicrobial peptides preceding changes of circulating inflammatory cytokines. *Med Inflamm* (2017) 2017;9474896. doi: 10.1155/2017/9474896

139. Wallaeys C, Garcia-Gonzalez N, Libert C. Paneth cells as the cornerstones of intestinal and organismal health: A primer. *EMBO Mol Med* (2022) 15(2):e16427. doi: 10.15252/emmm.202216427

140. Zhou H, Zhou S-Y, Gillilland M, Li J-Y, Lee A, Gao J, et al. Bile acid toxicity in paneth cells contributes to gut dysbiosis induced by high-fat feeding. *JCI Insight* (2020) 5(20):e138881. doi: 10.1172/jci.insight.138881

141. Li J, Li X, Song J, Yan B, Rock SA, Jia J, et al. Absence of neurotensin attenuates intestinal dysbiosis and inflammation by maintaining Mmp7/Alpha-defensin axis in diet-induced obese mice. *FASEB J* (2020) 34(6):8596–610. doi: 10.1096/fj.201902374RR

142. Andres SF, Santoro MA, Mah AT, Keku JA, Bortvedt AE, Blue RE, et al. Deletion of intestinal epithelial insulin receptor attenuates high-fat diet-induced elevations in cholesterol and stem, enteroendocrine, and paneth cell mrnas. *Am J Of Physiology-Gastrointestinal And Liver Physiol* (2015) 308(2):G100–G11. doi: 10.1152/ajpgi.00287.2014

143. Larsen IS, Fritnn AM, Carl CS, Agerhulm M, Daingaard MTF, Holm JB, et al. Human paneth cell alpha-Defensin-5 treatment reverses dyslipidemia and improves glucoregulatory capacity in diet-induced obese mice. *Am J Of Physiology-Endocrinology And Metab* (2019) 317(1):E42–52. doi: 10.1152/ajpendo.00019.2019

144. Iliodromitis EK, Andreadou I, Iliodromitis KE, Dagres N. Ischemic and postischemic conditioning of the myocardium in clinical practice: Challenges, expectations and obstacles. *Cardiology* (2014) 129(2):117–25. doi: 10.1159/000362499

145. Zerva K, Sanidas E, Tzanis G, Chantziara V, Papadopoulos D, Barbetseas J. Effect of thrombolytic therapy in plaque inflammation and structural remodeling in acute myocardial infarction patients. *Hellenic J Of Cardiol* (2017) 58(6):446–9. doi: 10.1016/j.hjc.2017.06.002

146. Kumar K, Singh N, Jaggi AS, Maslov L. Clinical applicability of conditioning techniques in ischemia-reperfusion injury: A review of the literature. *Curr Cardiol Rev* (2021) 17(3):306–18. doi: 10.2174/1573403x16999200817170619

147. Grootjans J, Hodin CM, de Haan J-J, Derikx JPM, Rouschop KMA, Verheyen FK, et al. Level of activation of the unfolded protein response correlates with paneth cell apoptosis in human small intestine exposed to Ischemia/Reperfusion. *Gastroenterology* (2011) 140(2):529–39. doi: 10.1053/j.gastro.2010.10.040

148. Park SW, Kim M, Brown KM, D'Agati VD, Lee HT. Paneth cell-derived interleukin-17a causes multiorgan dysfunction after hepatic ischemia and reperfusion injury. *Hepatology* (2011) 53(5):1662–75. doi: 10.1002/hep.24253

149. Park SW, Kim M, Kim JY, Ham A, Brown KM, Mori-Akiyama Y, et al. Paneth cell-mediated multiorgan dysfunction after acute kidney injury. *J Of Immunol* (2012) 189(11):5421–33. doi: 10.4049/jimmunol.1200581

150. Han SJ, Li H, Kim M, D'Agati V, Lee HT. Intestinal toll-like receptor 9 deficiency leads to paneth cell hyperplasia and exacerbates kidney, intestine, and liver injury after Ischemia/Reperfusion injury. *Kidney Int* (2019) 95(4):859–79. doi: 10.1016/j.kint.2018.10.035

151. Han SJ, Kim M, Novitsky E, D'Agati V, Lee HT. Intestinal Tlr9 deficiency exacerbates hepatic ir injury *Via* altered intestinal inflammation and short-chain fatty acid synthesis. *FASEB J* (2020) 34(9):12083–99. doi: 10.1096/fj.202000314R

152. Han SJ, Kim M, D'Agati VD, Lee HT. Norepinephrine released by intestinal paneth cells exacerbates ischemic aki. *Am J Of Physiology-Renal Physiol* (2020) 318(1): F260–F72. doi: 10.1152/ajprenal.00471.2019

153. Baloch S, Baloch MA, Zheng T, Pei X. The coronavirus disease 2019 (Covid-19) pandemic. *Tohoku J Of Exp Med* (2020) 250(4):271–8. doi: 10.1620/tjem.250.271

154. Kariyawasam JC, Jayarajah U, Riza R, Abeysuriya V, Seneviratne SL. Gastrointestinal manifestations in covid-19. *Trans Of R Soc Of Trop Med And Hygiene* (2021) 115(12):1362–88. doi: 10.1093/trstmh/trab042

155. Zoghi G, Moosavy SH, Yavarian S, HasaniAzad M, Khorrami F, Brojeni MS, et al. Gastrointestinal implications in covid-19. *BMC Infect Dis* (2021) 21(1):1135. doi: 10.1186/s12879-021-06824-y

156. Yamada S, Noda T, Okabe K, Yanagida S, Nishida M, Kanda Y. Sars-Cov-2 induces barrier damage and inflammatory responses in the human ipsc-derived intestinal epithelium. J Of Pharmacol Sci (2022) 149(3):139–46. doi: 10.1016/j.jphs.2022.04.010

157. Pearce SC, Suntornsaratoon P, Kishida K, Al-Jawadi A, Guardia J, Nadler I, et al. Expression of sars-Cov-2 entry factors, electrolyte, and mineral transporters in different mouse intestinal epithelial cell types. *Physiol Rep* (2021) 9(21):e15061. doi: 10.14814/phy2.15061

158. Zheng H, Chen Y, Li J, Li H, Zhao X, Li J, et al. Longitudinal analyses reveal distinct immune response landscapes in lung and intestinal tissues from sars-Cov-2-Infected rhesus macaques. *Cell Rep* (2022) 39(8):111014. doi: 10.1016/j.celrep.2022.110864

159. Donaldson DS, Shih BB, Mabbott NA. Aging-related impairments to m cells in peyer's patches coincide with disturbances to paneth cells. *Front Immunol* (2021) 12:761949. doi: 10.3389/fimmu.2021.761949

160. Docherty AB, Harrison EM, Green CA, Hardwick HE, Pius R, Norman L, et al. Features of 20133 uk patients in hospital with covid-19 using the isaric who clinical characterisation protocol: Prospective observational cohort study. *Bmj-British Med J* (2020) 369:m1985. doi: 10.1136/bmj.m1985

161. Wang C, Wang S, Li D, Wei D-Q, Zhao J, Wang J. Human intestinal defensin 5 inhibits sars-Cov-2 invasion by cloaking Ace2. *Gastroenterology* (2020) 159(3):1145–7. doi: 10.1053/j.gastro.2020.05.015

162. Xu C, Wang A, Marin M, Honnen W, Ramasamy S, Porter E, et al. Human defensins inhibit sars-Cov-2 infection by blocking viral entry. *Viruses-Basel* (2021) 13 (7):1246. doi: 10.3390/v13071246

163. Ali W, Elsahn A, Ting DSJ, Dua HS, Mohammed I. Host defence peptides: A potent alternative to combat antimicrobial resistance in the era of the covid-19 pandemic. *Antibiotics-Basel* (2022) 11(4):475. doi: 10.3390/antibiotics11040475

164. Mauro E, Gadano A. What's new in portal hypertension? *Liver Int* (2020) 40:122–7. doi: 10.1111/liv.14366

165. Deng W, Duan M, Qian B, Zhu Y, Lin J, Zheng L, et al. Nadph oxidase 1/4 inhibition attenuates the portal hypertensive syndrome *via* modulation of mesenteric angiogenesis and arterial hyporeactivity in rats. *Clinics And Res In Hepatol And Gastroenterol* (2019) 43(3):255–65. doi: 10.1016/j.clinre.2018.10.004

166. Moghadamrad S, McCoy KD, Geuking MB, Saegesser H, Kirundi J, Macpherson AJ, et al. Attenuated portal hypertension in germ-free mice: Function of bacterial flora on the development of mesenteric lymphatic and blood vessels. *Hepatology* (2015) 61(5):1685–95. doi: 10.1002/hep.27698

167. Berger JN, Gong H, Good M, McElroy SJ. Dithizone-induced paneth cell disruption significantly decreases intestinal perfusion in the murine small intestine. J Of Pediatr Surg (2019) 54(11):2402–7. doi: 10.1016/j.jpedsurg.2019.02.021

168. Hassan M, Moghadamrad S, Sorribas M, Muntet SG, Kellmann P, Trentesaux C, et al. Paneth cells promote angiogenesis and regulate portal hypertension in response to microbial signals. *J Of Hepatol* (2020) 73(3):628–39. doi: 10.1016/j.jhep.2020.03.019

169. Hassan M, Juanola O, Keller I, Nanni P, Wolski W, Martinez-Lopez S, et al. Paneth cells regulate lymphangiogenesis under control of microbial signals during experimental portal hypertension. *Biomedicines* (2022) 10(7):1503. doi: 10.3390/ biomedicines10071503

170. Marciniak SJ, Chambers JE, Ron D. Pharmacological targeting of endoplasmic reticulum stress in disease. *Nat Rev Drug Discov* (2022) 21(2):115–40. doi: 10.1038/ s41573-021-00320-3

171. Zong S, Liu T, Wan F, Chen P, Luo P, Xiao H. Endoplasmic reticulum stress is involved in cochlear cell apoptosis in a cisplatin-induced ototoxicity rat model. *Audio Neuro-Otol* (2017) 22(3):160–8. doi: 10.1159/000480346

172. Gorbunov NV, Garrison BR, Kiang JG. Response of crypt paneth cells in the small intestine following total-body gamma-irradiation. *Int J Of Immunopathology And Pharmacol* (2010) 23(4):1111–23. doi: 10.1177/039463201002300415

173. Mann EA, Saeed SA. Gastrointestinal infection as a trigger for inflammatory bowel disease. *Curr Opin Gastroenterol* (2012) 28(1):24-9. doi: 10.1097/MOG.0b013e32834c453e