Check for updates

OPEN ACCESS

EDITED BY Lei Zhang, Melbourne Sexual Health Centre (MSHC), Australia

REVIEWED BY

Luminiţa-Smaranda lancu, Grigore T. Popa University of Medicine and Pharmacy, Romania Zewen Kelvin Tuong, The University of Queensland, Australia

*CORRESPONDENCE Sarah C. Sasson Ssasson@kirby.unsw.edu.au

[†]These authors have contributed equally to this work

[‡]These authors share last authorship

SPECIALTY SECTION

This article was submitted to Viral Immunology, a section of the journal Frontiers in Immunology

RECEIVED 30 November 2022 ACCEPTED 03 February 2023 PUBLISHED 07 March 2023

CITATION

Hewavisenti RV, Arena J, Ahlenstiel CL and Sasson SC (2023) Human papillomavirus in the setting of immunodeficiency: Pathogenesis and the emergence of nextgeneration therapies to reduce the high associated cancer risk. *Front. Immunol.* 14:1112513. doi: 10.3389/fimmu.2023.1112513

COPYRIGHT

© 2023 Hewavisenti, Arena, Ahlenstiel and Sasson. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Human papillomavirus in the setting of immunodeficiency: Pathogenesis and the emergence of next-generation therapies to reduce the high associated cancer risk

Rehana V. Hewavisenti^{1†}, Joshua Arena^{1,2†}, Chantelle L. Ahlenstiel^{1,2‡} and Sarah C. Sasson^{1*‡}

¹Immunovirology and Pathogenesis Program, The Kirby Institute, The University of New South Wales, Sydney, NSW, Australia, ²UNSW RNA Institute, The University of New South Wales, Sydney, NSW, Australia

Human papillomavirus (HPV), a common sexually transmitted virus infecting mucosal or cutaneous stratified epithelia, is implicated in the rising of associated cancers worldwide. While HPV infection can be cleared by an adequate immune response, immunocompromised individuals can develop persistent, treatment-refractory, and progressive disease. Primary immunodeficiencies (PIDs) associated with HPV-related disease include inborn errors of GATA, EVER1/2, and CXCR4 mutations, resulting in defective cellular function. People living with secondary immunodeficiency (e.g. solidorgan transplants recipients of immunosuppression) and acquired immunodeficiency (e.g. concurrent human immunodeficiency virus (HIV) infection) are also at significant risk of HPV-related disease. Immunocompromised people are highly susceptible to the development of cutaneous and mucosal warts, and cervical, anogenital and oropharyngeal carcinomas. The specific mechanisms underlying high-risk HPVdriven cancer development in immunocompromised hosts are not well understood. Current treatments for HPV-related cancers include surgery with adjuvant chemotherapy and/or radiotherapy, with clinical trials underway to investigate the use of anti-PD-1 therapy. In the setting of HIV co-infection, persistent high-grade anal intraepithelial neoplasia can occur despite suppressive antiretroviral therapy, resulting in an ongoing risk for transformation to overt malignancy. Although therapeutic vaccines against HPV are under development, the efficacy of these in the setting of PID, secondary- or acquired- immunodeficiencies remains unclear. RNA-based therapeutic targeting of the HPV genome or mRNA transcript has become a promising next-generation therapeutic avenue. In this review, we summarise the current understanding of HPV pathogenesis, immune evasion, and malignant transformation, with a focus on key PIDs, secondary immunodeficiencies, and HIV infection. Current management and vaccine regimes are outlined in relation to HPVdriven cancer, and specifically, the need for more effective therapeutic strategies for immunocompromised hosts. The recent advances in RNA-based gene targeting

including CRISPR and short interfering RNA (siRNA), and the potential application to HPV infection are of great interest. An increased understanding of both the dysregulated immune responses in immunocompromised hosts and of viral persistence is essential for the design of next-generation therapies to eliminate HPV persistence and cancer development in the most at-risk populations.

KEYWORDS

human papillomavirus (HPV), human immunodeficiency virus (HIV), RNA interference, squamous cell carcinoma, transplant, primary immunodeficiencies, cancer, nanoparticles

1 Introduction

Approximately 15-20% of global cancers are associated with oncogenic viral infections (1, 2). These viruses adopt numerous mechanisms by which they evade immune responses and establish persistent infections resulting in malignant transformation (3). A common trait among oncogenic viruses is that only a small proportion of chronically-infected cells develop cancer (4). Viruses such as Human Papillomavirus (HPV), Epstein-Barr Virus (EBV), Hepatitis B virus (HBV), and Hepatitis C virus (HCV) are known to be implicated in the pathogenesis of human cancer (5). These viruses act as obligate intracellular parasites, where the microbe encodes proteins that affect cell development, apoptosis, and growth cycle. The virus 'reprograms' host cellular signaling, disrupting major checkpoints regulating proliferation, differentiation and cell death, and genomic integrity (3). Immunosurveillance, important in identifying and removing aberrant cells from the proliferative pool, is also negatively impacted (3). Additionally, cancer progression as a result of chronic viral infection is dependent on host factors, including local and systemic immunity, somatic mutations, immunosuppression, genetic predisposition, and environmental factors, such as carcinogen exposure.

HPV is a double-stranded DNA virus associated with cancers of the squamous epithelia of the cervix, oropharynx, and anogenital regions. There are currently ~200 identified HPV types, with many capable of causing a range of mucosal or cutaneous epithelial hyperplastic lesions (6). These types are divided into low-risk HPV (LR-HPV) and high-risk HPV (HR-HPV) groups depending on their likelihood of malignant progression (7, 8). Immunosuppressed individuals are at a heightened risk of developing HPV-associated lesions and cancers, making HPV a useful model for understanding viral-host interactions leading to epithelial-derived tumors. The most well-understood HPV-induced cancer is cervical cancer, with 99% being related to HR-HPV types (3). However, HPV-associated cancer affects many other mucosal sites, with 64–91% of vaginal, 40–50% of vulvar, 88–94% of anal, and 40–50% of penile cancers being HPVassociated (9).

Immunosurveillance plays a crucial role in initiating antigendependent responses in eliminating HPV infection and virally transformed cells. In immunocompetent hosts, 90% of anogenital HPV infections are cleared (10, 11). Innate immune cells such as natural killer (NK) cells, dendritic cells (DCs) and Langerhans cells (LCs) play an important role during the initial onset of infection, while HPV-specific CD8⁺ T cells target early viral proteins in infected cells (9, 12, 13). Conversely, HPV-associated malignancies have been commonly associated with at-risk population groups, including organ transplant recipients, systemic immunosuppressed patients, and people living with HIV (PLWH). Although HPV can evade immunity for long periods of time in immunocompetent hosts, infection is generally resolved. However, HPV infection can be persistent in immunocompromised individuals, with extensive HPV disease manifesting as non-regressive high-grade intraepithelial lesions (HSILs), leaving these individuals susceptible to HPV-associated malignancy formation (14–18).

Current therapeutic regimens for the treatment of HPV-associated cancers can include surgery, chemotherapy and/or radiotherapy. HPVassociated cancers are treated medically the same as HPV cancers, despite different biological origins. This approach is often insufficient and can be associated with poor survival rates (19). Globally, the 5-year survival rate of cervical carcinomas is 64%, penile squamous cell carcinomas (SCCs) is 47%, anal SCCs is 70%, rectal SCCs is 56%, and oropharyngeal SCC is 51% (20). In immunocompromised individuals, these survival statistics are lower. HIV-positive (HIV+) women with cervical cancer have a 3vear survival of 35% (21) whilst those with anal squamous cell carcinoma (ASCC) have a 5-year survival rate of 47% (22). Effective treatments for immunocompromised individuals are challenging due to the limited understanding of the varying immune responses to disease, which are highly dependent on host factors. A greater understanding of these underlying immune mechanisms in at-risk individuals is crucial for the future design of next-generation therapeutics and technologies targeting HPV. The prevention of persistent HPV infection and cancerous progression in at-risk groups is a priority globally. Here, we discuss the current understanding of HPV pathogenesis, key challenges in current clinical management, and the emergence of next-generation RNA therapeutics, focusing on at-risk populations (Figure 1).

2 HPV epidemiology and risk factors

2.1 HPV epidemiology

HPV is endemic globally, representing the most common sexually transmitted disease with 84% of sexually active women and 91% of sexually active men estimated to acquire the infection during their

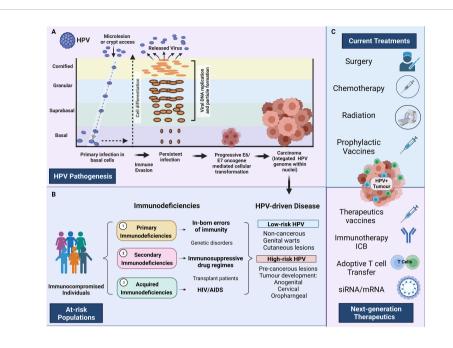


FIGURE 1

HPV pathogenesis and treatment in at-risk individuals. (A) HPV pathogenesis, persistent infection, and progression to cancer. Primary infection occurs when HPV gains access to basal cells through microlesions or damage to the skin. Upon viral replication, HPV can evade the immune response resulting in persistent disease. E6 and E7 oncogenes disrupt the cell cycle, which can result in persistent disease, cellular transformation and HPV-driven carcinoma. (B) Describes the three main immunodeficiencies 1) Primary immunodeficiencies relating to inborn errors of immunity, 2) Secondary immunodeficiencies relating to those under immunosuppressive drugs 3) Acquired immunodeficiencies relating to people living with HIV/AIDS. The major HPV pathologies caused by high-risk HPV and low-risk HPV strains are identified on the extreme right-hand side panel (HPV-driven disease) (C) Identifies the main treatments currently available and the next-generation therapeutics currently under investigation. Created with BioRender.com.

lifetime (23). HPV prevalence differs by age, with elevated rates of incidence amongst the 18-21 and 55–65 year-old cohorts. HPV is present in ~11.4% of the worldwide population, with the highest prevalence in Sub-Saharan Africa (24%), Eastern Europe (21%), and Latin America (16%) (24). In the period 1990-2012, the global incidence of HPV-associated malignancies reduced by 0.3%, but varied between geographic regions, with decreased incidence linked to strong HPV vaccination regiments in developed nations and increased incidence in countries with weaker healthcare infrastructure (25, 26) (Figure 2). Consequently, HPV infection is still implicated in the pathology of approximately 5% of all cancers (27).

2.2 Risk factors associated with HPV progression to cancerous lesions

HPV infection alone is not sufficient for the formation of HPVassociated cancers (28). Studies have aimed to identify the co-factors that predict and are associated with HPV infection progression to cancer, with major findings summarised below.

2.2.1 HPV type

HPV type is the primary factor associated with infectious progression to cancerous lesions. HR-HPV is a group 1 carcinogen and includes types 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, and 66 (29). HR-HPVs play a crucial role in the etiology and pathophysiology of malignancies of the cervix, anus, oropharynx, vulva, and penis (9, 30–34). LR-HPV genotypes include 6, 11, 40, 42, 43, 44, 54, 61, 70, 72,

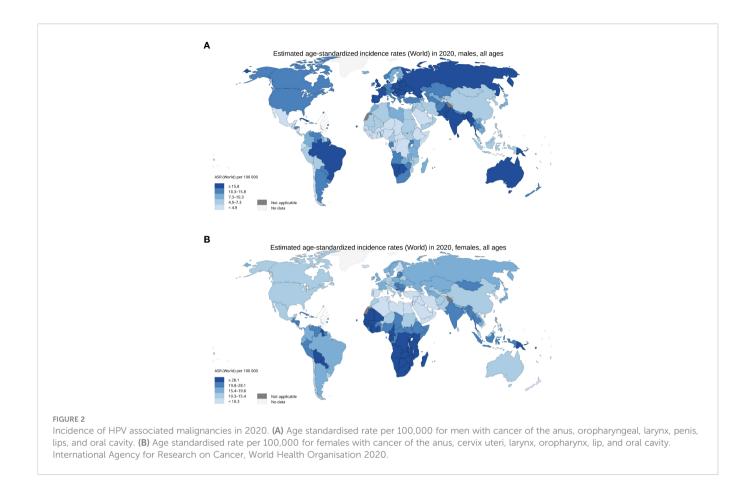
and 81, and can cause benign or low-grade cervical tissue changes and genital warts that seldom progress to cancerous lesions (35, 36). Of all the subtypes, only LR-HPVs 6 and 11 are classed as group 2B carcinogens (37).

2.2.2 Tobacco smoking

Tobacco smoke is classed as a group A carcinogen by the World Health Organisation, containing >4500 chemical compounds, with at least 70 oncogenic to humans (38). Inhalation and use of tobaccocontaining products have been significantly associated with the progression of HR-HPV infection to pre-cancerous and squamous cell carcinomas of the head and neck, cervix, anus, and penis (39–42). The precise mechanism by which tobacco smoke exacerbates the pathogenicity of HPV remains unknown, however recent studies suggest that tobacco comprises chemicals that enhance viral oncogenic expression in HPV-infected epithelium (43).

2.2.3 High parity

High parity has been suspected of playing a role in the pathophysiology of HPV-associated carcinogenesis. A 2002 International Agency for Research on Cancer (IARC) multicentric case-control study found a direct positive association between the number of full-term pregnancies and the onset of HPV-associated squamous cell carcinoma, but not adenocarcinoma or adenosquamous carcinoma (44). Similarly, a 2022 meta-analysis of 6685 parous women found a 2.65 higher odds ratio for the development of cancerous lesions existed compared to their null parity counterparts (45). It is hypothesized that increased levels of progesterone and estrogen during the final weeks of pregnancy alter tight junctions between



squamous and columnar epithelium, increasing the oncogenic activity of HPV (45).

2.2.4 Oral hormonal contraception

The increased uptake of oral hormonal contraceptive use amongst women has renewed interest in the potential impact of associated HPV infection progression. There is some evidence suggesting an association between the long-term use of oral hormonal contraception with an increased risk in the development of HPVassociated cervical and anogenital carcinomas (46). However, a 2022 systematic review of 7 prospective studies did not find supporting evidence to suggest a significant association between oral contraceptive use and an increased risk of HPV-associated malignancies (47). Further investigation into the relationship between oral hormonal contraceptives and HPV disease progression is required.

2.2.5 Diet

Dietary intake has been hypothesized to impact HPV clearance and resolution, though there remains no scientific consensus on the individual dietary components associated with resolution. Conflicting evidence surrounding the intake of vitamin A (48–50), B–Carotene (51–53) and folate (54–57) does not enable a determination as to their effect on the clinical resolution of HPV or cancerous lesion formation. Consequently, it remains uncertain as to the impact of diet in cases of HPV-associated cancers.

2.2.6 Immunodeficiencies

Immunodeficiency is one of the highest risk factors associated with severe clinical manifestations of HPV-associated cancers. Immunodeficiencies are categorized into three main groups; 1) Primary immunodeficiencies (PIDs), 2) Secondary immunodeficiencies, and 3) Acquired immunodeficiency e.g. HIV infection. PIDs are inborn errors of immunity (58) and correlate with a higher prevalence of HPVassociated conditions when compared to healthy controls (59). Secondary immunodeficiencies include states of malnutrition and immunosuppression. Immunosuppression therapies include regimens prescribed to organ transplant recipients to reduce the risk of graft rejection. Over 30 tumor types have been evident at an increased frequency in individuals following transplantation, with the most common being SCCs (60). Acquired immunodeficiencies, predominately HIV infection, remain a significant co-factor in the pathophysiology of HPV-associated malignancies.

3 HPV biology and pathogenesis

3.1 HPV genome

HPVs are small, ~8 kb double-stranded DNA viruses that contain eight open reading frames (ORFs): E1, E2, E4, E5, E6, E7, L1 and L2 (61). E1, E2, E4 and E6 maintain viral function, whilst L1 and L2 encode structural capsid proteins required for viral replication (62). Two major promoters regulate transcription within the HPV genome: the early promoter located in the long control region, which regulates the expression of E6 and E7; and the late promoter located within the E7 gene, which regulates the expression of E1, E2, E4, E5, L1 and L2 (63).

HPVs are classed phylogenetically into five distinct genealogical classes with α -papillomaviruses uniquely constituting the HR-HPV and LR-HPV subtypes, as described above (64). HPV subtypes and variants are declared based on the genetic distance between viral genomes (65). A new HPV subtype is declared when the L1 ORF differs from any other known HPV type by $\geq 10\%$. Within types, variants exhibit between 0.5-1% genome differentiation (66). As of 2022, over 200 different HPV subtypes have been identified (67). Variation amongst HR-HPV subtypes poses a significant risk to the development of a singular therapeutic to target global prevalence (65).

3.2 HPV pathogenesis and immune evasion

HPV oncogenes can contribute to viral replication and cellular aberrations. E6 and E7 are the two primary oncogenes that promote HPV-associated carcinogenesis. HPV-associated cancers are more likely to occur in combination with the aforementioned risk factors including smoking, immunocompromise, or genomic alterations of host cell DNA (3). The HR-HPV types most associated with cancerous formation (in combination with host factors) are HPV16 and HPV18 (35, 68).

Factors that influence HPV infection and immune evasion include host environmental, genetic, and immune elements. Minor damage to the epithelium allows HPV infection of the basal epithelium. HPV "reprograms" cellular machinery to replicate and reproduce. The virus is then released from terminally differentiated cells that slough off from the epithelial surface (69, 70). Although most immunocompetent individuals can clear the infection within 12 months, anti-HPV antibodies are only detected 6-12 months after infection. During this delayed adaptive response, HPV can adopt intrinsic mechanisms to evade the host immune surveillance (11, 71). This can be particularly problematic for immunocompromised hosts with limited immunosurveillance.

In persistent infection, viral oncogenic expression can result in alternations to the host genome, affecting the cell cycle, and yielding a small number of aberrant cells that can avoid immune controls. Oncogenic effects on the cell cycle result in intraepithelial neoplasia, which has been extensively studied in cervical cancer (72) and is also evident in HPV-driven anogenital disease (73). Following HPV epithelial infection and integration into the host genome, oncoproteins E6 and E7 inactivate tumor protein p53 and retinoblastoma suppressor protein (pRb) (74), resulting in cellular transformation. The E6 protein is associated with p53 degradation, resulting in uncontrolled cellular proliferation and tumor growth (75). The E7 protein degrades pRb which subsequently disrupts pRb interaction with the E2F family of transcription factors. This results in increased enzyme gene replication and cell division (76, 77). Therefore, the dysfunction of tumor suppressors p53 and pRb generates hallmark features of malignancy, including uncontrolled proliferation, impaired apoptosis, and chromosomal instability (69). E7 can also act as a mutator in the mitotic process resulting in mitotic abnormalities, such as unaligned or lagging chromosomes, and breaks in chromosomal structure, resulting in further destabilization of the host cell genome (78). Furthermore, HPV prevalence is not only due to persistent infection, HPV infection of one strain increases the risk of acquiring a new HPV infection. It has been reported that 20% of men infected with one type of HPV strain, exhibited a higher number of new HPV strains on follow-up. Therefore, an individual with more than one HPV strain has an increased risk of developing HPVassociated disease (79).

3.3 Immune responses in HPV clearance

During HPV infection, innate immune cells act as the first line of defense, with the recruitment of pro-inflammatory and cytotoxic immune cells from the adaptive immune system used to eliminate viral particles. This results in the activation of effector cells, interferon secretion, and the release of pro-inflammatory cytokines and chemokines. Antigen-presenting cells (APCs), such as LCs and DCs, identify viral particles *via* their toll-like receptors (TLRs). HPV gene expression is confined to the keratinocytes and resolution of infection requires antigen cross-presentation by APCs and cross-priming of naïve CD8⁺ T cells for subsequent infiltration of effector T cell responses (80). This expansion of HPV-specific T cells or chestrates the adaptive immune response. Additionally, B cells are important for producing neutralizing antibodies against HPV major capsid protein L1 and aid APCs to encourage cytotoxic CD8⁺ T cell proliferation.

T cells play crucial roles in preventing chronic infection. During initial infection, CD8^{+,} T cells target viral proteins E6 and E7. In subsequent infections, the immune memory of early antigen-specific T cells eliminates virally infected cells to prevent disease establishment. This has been demonstrated in cervical cancer, where a lack of HPV-oncogenic specific CD8⁺ T cells has been associated with high-grade cervical intraepithelial neoplasia (CIN) (81). Women with active HPV16 infection demonstrated a robust cytotoxic CD8⁺ T cells response and rapid HPV clearance. Both CD4⁺ and CD8⁺ T cell responses were associated with HPV-E6-related lesion regression, while E7 regression has been attributed solely to a $\mathrm{CD4}^{+}\ \mathrm{T}$ cell response (82–85). Therefore, E6 and E7 elicit different immune responses and recruit different T cell subsets, which could be highly dependent on HPV type. Although HPV16 and HPV18 are the predominant HPV types associated with high-grade neoplasia, E6 and E7 expression in other HR-HPV strains may elicit different immune responses, which are as equally severe in immunocompromised people and requires further investigation.

3.4 HPV progression to invasive cancer

A hallmark of HPV infection is its effectiveness in evading immune recognition. Viral replication is exclusively within the intraepithelial site with no induced cytolysis, viremia, cell death, or inflammation associated with replication and release (70, 86). Although effective viral clearance and HPV resolution are achieved by an effective cell-mediated immune response, in some cases, the HPV genome can lay dormant in cells evading immunosurveillance. Animal models have demonstrated that during lesion regression, when no active infection is evident, infected cells containing viral DNA persist in a latent state without viral gene expression. Although the virus is inactive, host factors including changes in hormone levels or immune suppression can cause HPV reactivation (70). HPV16 has a longer duration of persistence compared with other HPV types, which may contribute to its strong association with neoplastic development (87, 88). Failure to develop effective cell-mediated immune responses to control the infection results in high-grade intraepithelial neoplasia. Accumulation of genetic abnormalities can cause progression to invasive carcinoma. It has been shown that some cervical cancer patients lack antigen-specific T-cell responses, which may be due to an inability to recognize viral antigens. However, a clear link between HLA type and factors of host susceptibility to developing carcinoma requires further elucidation (12, 89, 90).

HPV can downregulate immune signaling pathways within keratinocytes, altering the innate and adaptive immune responses. This ceases the production of pro-inflammatory cytokines, such as type 1 interferons, that drive the inflammatory response and are important signals for the activation and migration of APCs. Transcriptomic analysis has shown oncogenic downregulation of genes with antiviral effects, such as IFIT1 and MX1, as well as those crucial for apoptosis (TRAIL and XAF1), IFN-y signaling (signal transducer and activator of transcription 1; STAT1) and pathogen recognition receptors (TLR3, RIG-I, MDA5) (91). HPV16 E6 expression disrupts Tyrosine kinase 2 (TYK2), an intracellular enzyme important for mediating immune signaling and inflammatory pathways essential for maintaining normal immune responses. This affects the Janus kinase (JAK)/STAT signaling pathway, which has been associated with cancer progression and metastatic development and is implicated in the development of cervical cancer. The STAT pathway is associated with essential cellular mechanisms including cell proliferation, invasion, survival, inflammation, and immunity (92). Induction of STAT3 in transformed keratinocytes drives the production of the cytokine IL-6 that, in a paracrine manner, causes STAT3-associated induction of CCL2 in monocytes. This potent monocyte-attracting chemokine skews the inflammatory microenvironment, resulting in a regulatory milieu by attracting anti-inflammatory macrophages and localized regulatory T cells (Tregs) (93, 94). High CCL2 expression has been associated with poor cervical cancer outcomes, while IL-6 production downregulates CCR7 expression on activated APCs. This inhibits migration-promoting chemokines essential for homing to regional lymph nodes (95).

E7 interferes with IFN-regulatory factor 1 induction, whilst both E6 and E7 have been shown to reduce E-cadherin surface levels. This inhibits the effective migration and abundance of LCs within the active lesion vicinity (96–99). Furthermore, CCL20 is an important chemoattractant produced by keratinocytes to recruit LCs *via* the CCR6 receptor. LCs are one of the most important tissue-resident APCs, initiating T cell priming during HPV infection. E6 and E7 have been shown to suppress the NF-κB pathway, implicated in chronic inflammation in cancer, specifically affecting transcription-factor dependent CCL20 induction, promoting APC recruitment into active lesion sites (100). This modulation of signaling pathways by HPV disrupts the cell cycle, resulting in an immunosuppressive and tumor-promoting environment. The developing HPV-driven tumor cells can upregulate immune checkpoint molecules, such as PD-1/ PDL1, and CTLA-4, which negatively regulate cytotoxic T cells, blocking anti-tumour specific responses. Overall, effective viral evasion strategies coupled with the disruption of crucial immunosurveillance pathways enable HPV to "hide" within host cells and skew the surrounding microenvironment towards immune tolerance.

4 HPV infection in primary, secondary and acquired immunodeficiencies

In immunocompromised individuals, persistent and extensive manifestations of HPV infection can result from an inadequate immune response. This includes the development of non-regressive lesions and/or progressive papillomas, leaving individuals in a precancerous state and at an elevated risk of developing HPV-driven malignancies (101, 102).

4.1 Primary immunodeficiencies

For individuals with PIDs, HPV infection can clinically manifest as benign warts, skin squamous cell carcinomas (SCC), or HPVdriven malignancies (103). PIDs with HPV clinical implications include epidermodysplasia verruciformis (EV), WHIM (warts, hypogammaglobulinemia, infections and myelokathexis) syndrome, DOCK8 mutations, GATA binding protein 2 (GATA2) mutations, and severe combined immunodeficiency (SCID) (103, 104). In immunocompetent individuals, B-HPVs, such as HPV5, HPV8, and HPV9, infect cutaneous sites but present as asymptomatic infections (105). In the setting of specific PIDs, clinical symptoms can be characterized by body surface involvement of cutaneous warts, including common warts (verrucae vulgaris), plantar warts (verrucae plantaris), and flat warts (verrucae plana) (103). Key PIDs associated with a high risk of progressive HPV disease are characterized by defects in NK cell and CD8⁺ T cell counts as well as CD4⁺ T cell lymphopenia. Although reported in detail (103, 104), key PIDs are summarised below.

4.1.1 Epidemodysplasia verruciformis

EV is characterized by mutations in the EVER1 and EVER2 genes. EVER has been implicated as a natural HPV barrier within keratinocytes and immune cells. EVER1/2 are zinc-transporting proteins expressed on immune cells including T cells, B cells, NK cells, and endothelial and myeloid cells (106). EVER1/2 encodes highly conserved membrane proteins in the zinc-transport complex, regulating zinc homeostasis and therefore limiting zinc availability to HPV-infected keratinocytes. Maintaining optimal zinc levels for cellular functions reduces viral replication and ensures an adequate anti-viral response (107). The specific roles of EVER1 and EVER2 in HPV-related disease focus on their interaction with zinc transporter 1 and its ability to drive activator protein 1 (AP-1) transcription activity. EVER dysfunction increases zinc concentration, activating AP-1, an important transcription factor for the HPV life cycle, aiding replication (107, 108). Zinc imbalance facilitates α -papillomavirus and B-papillomavirus pathogenesis, enhancing the expression of E6 and E7 (107). In EV, even LR-HPV can cause persistent disease leaving these individuals at high risk of HPV-driven SSC (109).

Tumour necrosis factor alpha (TNF-a) overproduction in EV individuals has been implicated in SCC development. TNF- α is a central cytokine important in host defense again viruses by triggering apoptosis with EVER proteins. EVER2 is a particularly important sensitizer to TNF- α -induced apoptosis. Contrary, the TNF- α signaling pathway can be harmful as it can switch from a proapoptotic to a pro-survival environment, thus promoting tumor progression, and metastasis (110). EVER2 is integral in preventing the pro-survival TNF- α signaling pathway by binding to TNFRassociated death domain protein (TRADD), preventing downstream pro-survival pathways and promoting TNF-\alpha-induced apoptosis (111). Host vulnerability to persistent disease is hypothesized to be due to defects in local TNF- α signaling (111), zinc transport dysfunction (108), and increased expression of transcription factors that enable HPV replication in the absence of functional EVER protein (107).

EVER is also implicated in keratinocyte participation in local inflammatory reactions within the skin. This is through local skin secretion and responses to growth factors, various cytokines such as TNF- α , IL-6, IL-19, and chemokines, such as TFG- β and IL-8 (107). T cells, macrophages, and keratinocytes secrete pro-inflammatory IL-6, enhancing inflammation in response to pathogens (112, 113) while IL-8 aids in the migration of circulating neutrophils into tissues (114). In EVER2 -/- keratinocytes, it has been reported that production of IL-6 was reduced while IL-8 levels increased compared to wildtype. The changes in signaling between keratinocytes and immune cells could affect the ability to clear persistent lesions (115), however further investigation into these mechanisms is required.

4.1.2 CXCR4 deficiency-WHIM syndrome

Warts, hypogammaglobulinemia, infections, myelokathexis (apoptosis of mature myeloid cells in the bone marrow) syndrome (WHIM) is associated with CXC chemokine receptor 4 (CXCR4) deficiency, with HPV-related disease being a major clinical feature. Manifestations include cutaneous warts typically on the hands, feet, and trunk, along with neutropenia, and in some cases lymphopenia (103). Papillomas can also develop in the genitalia and progress to neoplastic lesions and carcinoma (116, 117).

CXCR4 is a transmembrane receptor on leukocytes, endothelial cells, and stem cells, and is involved in important immune signaling pathways essential for HPV control (118). JAK2 and JAK3 interaction with CXCR4 activates the JAK/STAT pathway, which is important for immune regulation (119, 120). The interaction between CXCR4 and its ligand, stromal-derived factor-1 (SDF-1), is involved in multiple downstream signaling pathways important for chemotaxis, adhesion, and accumulation of immune cells to sites of inflammation. When CXCR4 is defective, a lack of SDF-1 signals disrupts leukocyte tracking to the affected site, allowing HPV to replicate and establish disease (121, 122). LCs and keratinocytes also express SDF-1 and CXCR4. CXCR4 gain of function mutation has been associated with an increase in cell immortalization, with increases in TNF- α expression (123, 124), driving HPV-driven carcinogenesis (103, 125).

4.1.3 Autosomal recessive hyper-IgE syndrome (DOCK8 mutation)

Dedicator of cytokines 8 (DOCK8) mutations cause an autosomal recessive combined immunodeficiency resulting in a range of hyper-IgE syndromes (126). People with DOCK8 deficiency are susceptible to cutaneous viral infections, including herpes simplex virus, HPV, molluscum contagiosum virus, and varicella-zoster virus. Clinical disease presents as flat and verrucous warts, which can be extensive, disfiguring, and often treatment-resistant (127). DOCK8 has been implicated in aiding leukocyte migration, with deficiency resulting in immunoregulation failure and subsequent HPV dissemination (128). Over time, lymphopenia progresses with age, affecting CD8⁺ and CD4⁺ T cells. In mice, DOCK8 deficiency impaired the suppression of thymic Tregs (129, 130). In vitro, CD8⁺ T cells from patients with DOCK8 deficiency had reduced proliferation to CD3/CD28 stimulation. Although rare, IFN- γ and TNF- α production can also be impaired (126). Multivariate cox regression analysis demonstrated DOCK8 was an independent positive factor in the survival of HPVpositive (HPV+) HNSCC, and positively correlated with immune cell infiltration levels (131). The expanded skin virome has been studied in DOCK8 immunodeficiency and it was found that even in the absence of clinical HPV warts, there was a high number of abundant reads (52% mean). This contrasted with other viruses such as molluscum contagiosum virus (MCV) here reads were increasingly low (0.4%) when clinical manifestations were absent. This biological distinction between HPV and MCV suggests that DOCK8-deficient individuals may be more susceptive to HPV carriage, and this may explain their susceptibility to the development of squamous cell cancers (132). The differences in the subclinical presence of HPV versus MCV suggest biological distinctions between these eukaryotic viruses, with patient skin demonstrating less susceptibility to MCV than to HPV. However, further investigation to address whether there is oncogenic potential HPV detected on the skin of DOCK8 deficient individuals is required.

4.1.4 Severe combined immunodeficiency (IL2RG or JAK3 deficiency)

Severe chronic HPV disease is associated with severe combined immunodeficiency (SCID) caused by IL2RG (interleukin 2 receptor subunit γ) or JAK3 deficiency. IL2RG and JAK3 are important for anti-viral immunity implicated in lymphocyte development, proliferation, and survival (133). Although hemopoietic stem-cell transplantation (HSCT) has been lifesaving for those with SCID, evidence of severe HPV disease and partial immunodeficiency are still evident up to 10 years post-HSCT. SCID solely associated with JAK3 deficiency is characterized by low NK cells, which can remain depleted following HSCT. This may have significant effects on the control of the earliest stages of HPV infection, where NK cell HPVtargeted cytotoxicity predominates (134). Keratinocytes from patients with IL2RG-deficient SCID have chemokine repertoire changes, resulting in an impaired ability to recruit immune cells (135). IL-2RG is a critical component of the IL-2 receptor and is shared among the receptors for IL-2, IL-4, IL-7, IL-9, IL-12, and IL-15. IL-2RG deficiency can impact HPV eradication and control, as it is shared among a broad array of cytokine receptors crucial for optimum innate and adaptive immunity. For example, myeloid development is highly reliant on IL-15, which in turn is dependent on IL2RG. IL-2RG deficiency impairs DCs and monocyte function, disrupting adequate signals to induce a pro-inflammatory immune response and recruit $CD8^+$ and $CD4^+$ T cells (136, 137). Individuals with these deficiencies are at a 50% risk of developing high-risk severe cutaneous warts post-HSCT (104, 138, 139). This suggests that HSCT does not completely revert immune defects, and a better understanding is required to tease out underlying mechanisms that allow HPV clinical disease to persist (140).

4.1.5 GATA2 mutations

GATA2 is a transcription factor important for hematopoiesis and stem cell progenitor maintenance (141-143). GATA2 deficiency results in the absence of major blood cell lineages, including B cells, DCs, monocytes and NK cells (141, 144). HPV is implicated in >75% of GATA2-deficient patients with 50% being at high risk of developing persistent infection and recurrent warts (140), subsequently progressing to malignant transformation. Although HSCT can have positive effects on restoring immunity, the progression of HPV prevalence and persistence post-HSCT is not well investigated, and only reverses the effects of GATA2 deficiency in approximately half of the patients (145). The clinical features of GATA2 deficiency are highly dependent on peripheral cell numbers with severe symptoms associated with more extreme cytopenia. T cells are not as affected by GATA2 mutation compared to other subsets, however, an inversion of the normal CD4:CD8 ratio, coupled with a reduction in naïve cells, is reflective of an immune repertoire of chronic viral infection.

4.1.6 Serine/threonine kinase 4 deficiency

STK4 homozygous mutations cause a deficiency associated with B cell lymphopenia and CD4 lymphopenia, particularly affecting naïve T cells, and signaling pathways important for T cell survival and death (103, 104). NK cell and CD8⁺ T cell levels remain at normal levels. STK4 deficiency results in lowering the transcription factor FOX01, which plays a significant role in T cell homeostasis, with patients showing progressive CD4⁺ T cell lymphopenia. This in turn affects downstream signaling pathways, impairing homing of naïve T cells by decreasing the IL-7 receptor, CCR7 and CD62L (103). In this condition, T cells are more prone to apoptosis and Treg development and maintenance are additionally affected (146). In immunocompetent individuals, STK4 is a tumor suppressor protein. STK4 is significantly decreased in HPV-associated cervical cancer via HPV E6 and E7 oncoproteins suppressing STK4 mRNA and increasing YAPdependent gene expression, which aids HPV replication and tumor progression. Therefore, defects in STK4 result in individuals being at an elevated risk of HPV-driven carcinogenesis (147).

4.2 Secondary and acquired immunodeficiencies

4.2.1 Transplant recipients

Immunosuppressive therapy improves long-term graft and patient survival in transplant recipients. However, immunosuppressive drug

regimens increase the cumulative occurrence of persistent HPV infection, including the development of pre-cancerous lesions and treatment-refractory cutaneous and anogenital warts. The development of HPV-associated warts in transplant patients has been linked to the duration of immunosuppression, with 50-90% of patients four to five years post-transplant developing warts (148). In HSCT, the development of HPV-driven cervical, vaginal, and vulvar warts leaves women at an increased risk of developing related cancer (149). A systematic review of anogenital HPV-associated cancer in solid organ transplant recipients found the highest increased standardized risk in cancers of the vulva and vagina (22.8), penis (15.7) and anus (4.9). Transplant patients with underlying HPV-associated infection and precancerous lesions were at the highest risk of developing cancer post-transplant (150).

The predominant body of evidence regarding persistent HPV infection transitioning to anogenital neoplasia and cancer is in renal transplant patients. Renal transplant studies have demonstrated that latent HPV reactivation occurring post-transplantation increased HR-HPV active infection by 27% (151-153). Additionally, prolonged CD4⁺ T cell lymphopenia has been reported in renal transplant patients that can persist for up to 10 years posttransplant, leaving individuals at an elevated risk of HPV disease persistence and allograft rejection (154, 155). Renal transplant recipients have a 65-100-fold increase in developing SCC compared to immunocompetent individuals (156, 157). In women who have undergone a renal transplant, a 5-fold increased risk of developing genital warts was observed compared to the control population (158). Within the first year of a renal transplant, the frequency of HR-HPV infection increased from 24% to 36% over a period of 6 months. The high HR-HPV viral load was positively associated with the development of HPV CIN, a cancer precursor (159). A case-control study reported that 20% of renal transplant patients were diagnosed with anal intraepithelial neoplasia (AIN) compared to 1% of controls (1). RT-PCR confirmed HPV16 prevalence in 47% of the transplant group versus 12% in the control group. Despite no evidence of anogenital disease in patients before transplant, AINs developed post-transplant in the setting of immunosuppressive therapy (160). AIN prevalence in ASCC is 20%, with established HPV infection being prevalent in 47% of transplant patients (160). Natural history studies of AIN in renal transplant recipients are predominately reported in HIV+ men who have sex with men (MSM), where more than 50% of lesions progressed from low to high-grade AIN over a two-to-four-year period (161). This response may be particular to renal transplant, with varied responses reported following other forms of transplant (162). Similarly, in a study of 1023 women following renal transplant in the Netherlands, there was a five-fold increased risk for cervical cancer, followed by a 41-fold increased risk in the vulvar and a 122-fold increased risk of anal cancer in the transplanted compared to the general population. In 91.7% of lesions, HR-HPV was detected with 54.5% being attributed to HPV16 (153).

For transplant recipients, viral oncogenicity is highly dependent on the level of required immunosuppression. The nature of immunosuppressive drugs is to block cytotoxic T cells to prevent organ rejection, however, this decrease in immunosurveillance can cause HPV reactivation and replication. *In vitro* and *in vivo* studies have shown that corticosteroids, calcineurin inhibitors (e.g. cyclosporine A) and antimetabolites (e.g. azathioprine) have a direct pro-oncogenic effect (163–165). Calcineurin inhibitors have been reported to be more potent immunosuppressors compared to other drugs such as antimetabolites, being associated with a higher risk of developing HPV disease. One study reported that ~36% of kidney transplant patients treated with cyclosporine, azathioprine, and prednisone, developed papillomatous lesions with 25% of these lesions attributed to HPV16 infection. Oral cavity lesions occurred in 14% of recipients treated with cyclosporine, which was not evident in those on an immunosuppressive regime without cyclosporine (166). In contrast, another study found a non-significant correlation between HPV status and cyclosporine use (167). There is minimal understanding of whether specific immunosuppressive drugs taken alone or in combination increase the likelihood of HR-HPV reactivation in transplant recipients. Improved knowledge around combination immunosuppressive drug regimens and their link to HPV reactivation and related diseases may lead to improved treatment protocols that reduce high-risk groups developing HPV disease.

4.2.2 HPV-driven malignancy in the setting of HIV co-infection

PLWH have an acquired immunodeficiency that places them at high risk of developing HPV-associated malignancies. Antiretroviral therapy (ART) reduces the incidence of many AIDS-defining cancers, including Kaposi's sarcoma caused by herpesvirus and EBV-driven non-Hodgkin's lymphoma. However, any beneficial effects in reducing the incidence of HPV-driven HSILs, persistent disease, and cancer risk are not well understood. PLWH are at a higher risk of developing many types of cancers compared to the general population (14, 18, 60, 168-171). While HIV+ women have significantly higher rates of cervical CIN when compared to uninfected women (172-174), HIV+ MSM have an increased risk of developing HPV-associated AINs (170, 175, 176). However, there is a limited understanding of HPV type and distribution. In Kenya and South Africa, HPV16/18 prevalence was not significantly different between HIV+ and HIV-negative (HIV-) women with cervical carcinoma (177). However, higher CD4⁺ T cell counts were negatively correlated to infection with multiple HPV types (177). CD4⁺ T cell counts of <200 cells/µL are strongly associated with HR-HPV infections as well as genital wart development (178-181). Therefore, even though the HPV distribution may not differ in the presence of HIV, CD4⁺ T cell lymphopenia may facilitate HR-HPV infection due to a lack of effective signaling to enable a robust cytotoxic response. While the oncogenic potential of HR-HPV is well-established, the impact of HIV co-infection on HPV and the downstream susceptibility to malignant transformation is not well understood (150).

The common underlying mechanisms of host susceptibility to HIV and HPV co-infection are centered around the disruption of epithelial integrity. It has been postulated that HIV and HPV co-infection occurs locoregionally within an individual, but not within the same cells. HIV infects CD4⁺ immune cells that can elicit disruptions to epithelial integrity, increasing the ability of HPV to reactivate or infect the host (182). Mucosal disruption is required for HIV virions to breach the mucosal barrier in the anogenital tract to target cells for viral replication, which can subsequently provide a pathway for HPV co-infection and progression to disease establishment and malignancy. It is hypothesized that mucosal epithelial cells upregulate inflammatory cytokines, such as TNF- α and TGF- β (183), and downregulate E-cadherin and tight junction proteins, allowing permeability and access for HIV and HPV to invade (184–186). Depending on the period between HIV establishment and HPV infection, the immune system, specifically CD4⁺ T cells, may already be affected and present at low numbers, increasing host susceptibility to persistent HPV infection.

4.2.2.1 HIV infection can increase host susceptibility to HPV invasion

While damage to epithelial integrity can result in susceptibility to HIV and HPV co-infection, active HIV infection may also increase susceptibility to HPV. A reduction in key innate molecules, including B-defensin-2 and thrombospondin, has been proposed due to the interaction between the two viruses, resulting in an increased risk of HPV-induced malignancies (187, 188). Defensins are important for viral inactivation, and the recruitment of T cells and neutrophils (189, 190). In CIN and invasive cervical SCC, B-defensin-2 is lower compared to the normal ectocervical epithelium (189). A similar pattern in thrombospondin levels is seen in invasive SCC (188). The HIV protein Tat is also implicated in increasing the gene expression of both HIV and other DNA viruses (191, 192). Disruption of tight junctions by HIV Tat and glycoprotein 120 (gp120), enables HPV to penetrate the basal layer of the oral epithelium (193). HIV Tat may increase the expression of E6 and E7 proteins, as well as E2, which is important for HPV replication (194) and has been described in HPVassociated cervical and oral SCC (195-197). Additionally, Tat can upregulate HPV-associated oncogenesis and reduce p53 protein levels, as well as re-activate dormant HPV (195, 198). Therefore, HIV results in pathogenesis that increases the likelihood of HPV disease in parallel.

Although many HIV-associated comorbidities have declined with ART, HPV disease burden remains high, specifically for anal and cervical SSC (199, 200). There is conflicting data on the impact of ART on HPV infection. Women living with advanced stages of HIV undergoing ART had a significant reduction in oncogenic HPV infection, prevalence, and incidence, whereas another study contradicted this outcome and suggested that ART was associated with increased lesions, warts, and oral HPV persistence (201, 202). Swiss patients (6%) undergoing highly active ART for four years, continued to have low CD4⁺ T cell counts (<200 cells/ μ L), with 50% unable to reach the clinically acceptable 500 cell/uL count (203). Prolonged ART in HIV-infected individuals may not improve HPVspecific immunity, resulting in HPV persistence and the development of neoplasia (204, 205). Additionally, although T cell reconstitution occurs, it may be functionally dysregulated. Such T cell dysregulation includes Treg dysfunction, poor antigen presentation by dendritic cells, as well as a skewing toward an inflammatory T helper 1 response (206).

4.2.2.2 Active HPV infection may increase the likelihood of HIV transmission in the host

While there is growing evidence that HIV infection can cause immune dysfunction resulting in opportunistic HPV disease, it has also been suggested that HPV can in turn increase the risk of HIV transmission. A meta-analysis of the effect of HPV infection on HIV susceptibility demonstrated that HPV infection is associated with a two-fold increased risk of HIV acquisition (207, 208). In sub-Saharan Africa, women with prior HPV cervical infection were at 2.4 times higher risk to acquire HIV than the control cohort (209), whilst a separate study found that HPV+MSM had a 3.5-fold increased risk of HIV seroconversion (210, 211). MSM with a CD4⁺ T cell count <200/ μ L had a higher risk of developing HSILs, with HIV+MSM having an increased risk of developing HPV-associated anal SCC.

Increased HIV susceptibility to an HPV-infected host has been linked to high HPV viral load, inflammatory cytokines at mucosal surfaces, mucosal barrier damage, CD4⁺ T cells in anogenital mucosa, and expression of neutrophil proteases. During HPV infection, lesion regression is marked by a high CD4⁺ T cell count within the stroma and epithelium, which potentially provides HIV with an opportunity to infect their target cell (212). Inflammatory mediators, such as MCP-1, IL-8 and LP-1, implicated in NK and T cells responses to wart regression, are important for HPV clearance but also associated with high HIV acquisition (213-215). Recruitment of neutrophil proteases and proinflammatory cytokines disrupts normal epithelial cell differentiation and epithelial barrier and integrity. Although HIV is known to primarily enter cells via CD4, HIV utilizes other common receptors, including CXCR4 (216) and CCR5 (212, 217). The HIV envelope glycoprotein binds to these receptors to facilitate HIV entry into cells, followed by membrane fusion and viral internalization where replication occurs (191). In the presence of HPV, a relative abundance of CD4⁺ T cells, DCs and macrophages that express CXCR4 and CCR5 provide HIV entry. DCs are known to capture HIV antigens and migrate to the lymph node for antigen presentation, however prolonged contact with autologous CD4⁺ T cells, can facilitate HIV infection (218). Therefore, these potential mechanisms and favorable environments during HPV infection can pave the way for successful HIV transmission.

5 HPV-driven cancers

5.1 HPV detection methods

Various detection methods are used to identify HPV to account for differential biomarkers and histopathological differences [Table 1; adapted from (219)]. Molecular detection methods and screening tools for specific HPV-driven cancer types have been previously summarised (220). In HPV-associated OPSCC (HPV+OPSCC), high levels of p16 and wild-type p53 are indicative of HPV infection. Immunostaining of p16 (associated with HPV-16) can be used as a marker to identify HPV-implicated cancers (221). This is of particular importance in OPSCC which can be caused by HPV or occur independently of HPV. HPV-negative OPSCC (HPV-OPSCC) can be identified through mutation in p53 and poor p16 expression due to deletion, mutation, or hypermethylation (222). In HPV +OPSCC, p16 overexpression can also be a result of other non-HPV-related pathways. Consequently, p16 staining is commonly used in conjunction with other methodologies to identify HPV+OPSCC accurately (223).

5.2 Predominant HPV-driven cancers in immunocompromised individuals

5.2.1 Cervical cancer

Cervical cancer is the fourth leading cause of cancer in women globally, accounting for 342 000 deaths worldwide in 2020 (224). The initial stages of cervical cancer can be asymptomatic prior to progression, where symptoms then include vaginal bleeding, pelvic pain, and dyspareunia (26). Immunocompromised individuals are at the greatest risk of developing non-regressive HPV-related lesions which can transition to cancer. Compared to uninfected women, women living with HIV have a two-fold increase in developing cervical cancer (26, 225, 226). In immunocompetent individuals, cervical cancer can take 15-20 years to develop, compared to 5-10 years in the setting of HIV co-infection. Cryotherapy or thermal ablation can be used to treat pre-cancerous lesions. HPV testing of women aged 30-65 years, with cervical screening occurring from the age of 21 or following the onset of sexual activity can aid in the early detection of HR-HPV genotypes and result in early intervention. If diagnosed with HIV, cervical cytology is recommended every six months from the year of diagnosis, with annual screening thereafter following no evidence of HPV infection. In solid organ transplant

TABLE 1 HI	PV methods	of detection	used to identify	HPV genotype.
------------	------------	--------------	------------------	---------------

Methods	Specificity	Sensitivity	Advantages and disadvantages	
Southern Blotting Assay	High	High	Ability to differentiate between episomal and integrated DNA. Not easily applied to FFPE samples	
ISH	High	Low	Ability to differentiate between episomal and integrated DNA. Low sensitivity	
HPV PCR	High	Low	Cost-effective Does not quantitate viral load or inform whether the virus is active (low sensitivity).	
Real-time PCR	High	High	Ability to differentiate between episomal and integrated DNA. Not easily applied to FFPE samples and is labor-intensive.	
Reverse Transcriptase PCR	High	High	High sensitivity <i>Time-consuming</i>	
p16 Immunostaining	Low	High	Identifies marker of transcriptionally active virus. Low specificity	
Signal Amplification Methods	High	Low	Methodology easily performed (user-friendly) May produce false positives (low sensitivity)	

10

patients, cervical cancer screening is also performed annually (227). A colposcopy, with or without a lesion biopsy, is conducted if abnormal cervical cells are evident. A cervical cancer diagnosis is made through histopathological examination with tumor size and disease metastasis defining the cancer stage (73).

5.2.2 Anal squamous cell carcinoma

HPV is detected in 80-90% of anal cancers, with HPV16 being the predominant type (228). The prevalence is highest in Australia, North America and regions of Europe (229). Like cervical cancer, anal carcinoma develops from lesions of varying cytological and histological severity, known as AIN. While low-grade AIN regressed in most immunocompetent individuals, high-grade AIN can progress to invasive cancer (230, 231). Persistent HR-HPV infection causes pre-cancerous lesions in immunocompromised individuals that do not regress due to inadequate adaptive immunity. These dysplastic lesions can progress to ASCC (232).

Despite the widespread availability of ART, the incidence of ASCC is continuing to increase (233). MSM and PLWH exhibit a 20-fold and 30-fold increased risk of ASCC, respectively (171), and HIV+ MSM have a 50-150-fold increased risk (232, 234, 235). In the natural history Study of the Prevention of Anal Cancer (SPANC), HIV+ and HIV- MSM (≥35 years) were enrolled to explore anal HPV infection epidemiology and cytological and historical abnormalities. HSIL incidence was strongly associated with younger age and persistent HR-HPV infection. Conversely, HSIL clearance was associated with smaller lesions, AIN 2, and a lack of HR-HPV infection (171). HPV16 and HPV18 showed the highest prevalence in MSM, however, HPV31, HPV22, and HPV58 also demonstrated increased incidence (>2.0/100 person-years). Women with a history of cervical dysplasia and transplant patients are also at an elevated risk. Renal transplant recipients have been shown to have a 10 times greater relative risk of developing ASCC than the general population (236).

ASCC screening involves high-risk populations undergoing anorectal exams where anal swabs can be collected for HPV genotyping and the identification of atypical cells. Screening aids in the early diagnosis and treatment of precancerous lesions prior to cancerous progression. Identification and characterization of earlystage cancer are facilitated by anal cytology and high-resolution anoscopy (HRA), which obtain directed tissue biopsies (176, 237). HRA is limited in its application due to restricted views in cases of uneven lesion topography, necessitating parallel use of digital anorectal exams. ASCC screening is not as established as cervical cancer screening, with little investigation into the effectiveness of routine surveillance and the treatment of high-grade AINs (60). With the rising prevalence of ASCC in immunocompromised individuals, additional research is required to develop effective screening strategies to accurately identify high-risk lesions in earlier stages where intervention mayimprove prognosis.

5.2.3 Oropharyngeal squamous cell carcinoma

Oropharyngeal SCC is associated with significant morbidity and mortality. The global, incidence of OPSCC in 2020 was 3,777,713 with

the highest prevalence in Asia, Europe, and North America (26). OPSCC affects the upper aerodigestive tract, including the tonsils, the base of the tongue, and the soft palate. Unlike most HNSSCs, which are associated with smoking and alcohol consumption, a biologically distinct subset of OPSCC arises from HPV. HPV+OPSCC represents 15-20% of HNSSC associated with HPV, with 95% of HPV+OPSCCs attributed to HPV16 (238). Over the past two decades, there has been an increase in HPV+OPSCCs, and a decrease in HPV-OPSCC prevalence due to lifestyle changes including a decrease in smoking rates (239-241). Compared to HPV-OPSCC, HPV+OPSCC occurs in younger individuals, with less exposure to alcohol and smoking, and a greater number of sexual partners. Immunodeficient individuals can also be at elevated risk, with PLWH demonstrating a two to six times increased risk of developing HPV-associated HNSCC (242). Immunosuppression is another major risk factor for SCC development, with organ transplant recipients having a 65 to 100fold increase in risk compared to the general population. Currently, there is no standard screening regime or treatment for HPV+OPSCC or oral cancer, with therapy based on tissue site rather than cancer pathophysiology. Like anal cancer, optimal screening and therapeutic regimens may depend on the underlying pathophysiological mechanisms. CD8⁺ tissue-resident T (CD8⁺ T_{RM}) cell populations in OPSCC are correlated with better patient survival and the underlying cellular and molecular mechanisms require further investigation (243). Further understanding of differences between HPV+OPSCC and HPV-OPSCC has the potential to inform novel host-directed therapies in this area.

5.2.4 Vulva cancer

VC accounts for 5% of gynecological cancers with HPV16 and HPV18 implicated in non-keratinizing vulva SCC (26). HPV-positive VC is diagnosed in young women, with immunosuppression being a significant risk factor (244) transplant patients have a 100-fold increase in VC incidence. Immunosuppressed patients develop VC precursor intraepithelial neoplasia, which displays hallmarks of severity i.e. multifocal lesions with extensive disease (245). In women living with HIV, vulvar intraepithelial neoplasia (VIN) development was 29 times more likely compared to the general population, with a 3.3-fold increase in the risk of persistent VIN post-treatment (246–248).

5.3 Clinical management of HPVdriven cancer

Clinical management of HPV-associated malignancies relates to the site of neoplasm. HPV-associated cancer treatment strategies are multimodal and can include monotherapies or combinations of traditional surgery, chemotherapy, radiotherapy, and targeted immune therapy (249, 250). Many patients with early nodal involvement require initial surgical removal and radiation therapy (251), whereas, in more advanced tumors, treatment is centered around radiochemotherapy (252). Therapeutic regimens for several malignancies are currently approached in the same manner irrespective of HPV status, and it is unclear if there is treatment equivalence in people who are co-infected with HPV and HIV (253). PLWH have shown improved outcomes from immune checkpoint inhibitor blockade (i.e. PD-1 inhibition) however, the detailed cellular and molecular mechanisms underlying this response are unclear.

The most comprehensive data for PD-1 inhibitors derive from the use of pembrolizumab (254) and nivolumab in HNSCC (255). In a KEYNOTE trial in active metastatic HNSCC, cetuximab (a drug that restricts tumor growth) with chemotherapy was compared with pembrolizumab monotherapy or pembrolizumab in combination with chemotherapy. In both pembrolizumab treatment groups, HNSCC exhibited improved overall survival when compared to cetuximab with chemotherapy. Patients on pembrolizumab monotherapy had a median overall survival of 14.9 months, compared to 10.7 months in the cetuximab with chemotherapy group. Similarly, patients on pembrolizumab with chemotherapy showed a median overall survival of 13 months versus 10.7 months in the cetuximab with the chemotherapy group. Despite these data suggesting the benefit of using pembrolizumab, HPV status was not reported among patients (256). In the KEYNOTE01 trial, using pembrolizumab in active HNSCC, HPV +HNSCC patients showed better progression-free survival (4 months) versus HPV-HNSCC (2 months) (255). A comprehensive summary of immunotherapies in HNSCC patients and differences in immune responses has been previously reviewed (249).

Similar clinical trials are ongoing in cervical cancer and ASCC. In cervical cancer, pembrolizumab was approved in 2018 based on a phase II study (KEYNOTE 158 -NCT02628067). This involved 98 patients with recurrent and/or metastatic cervical carcinomas (257), however, results did not show the same level of improvement as seen with HNSCC. Reports of the effects of pembrolizumab in cervical cancer are sparse and a better understanding is required amidst ongoing trials (KEYNOTE-826, phase III trial, NCT03635567) (258). Several clinical trials including NCT03233711, NCT02314169, and NCT02919969 have investigated PD-1 blockade in ASCC, and have demonstrated anti-tumor efficacy (259, 260). However, much of this work has not explored the efficacy of PD-1 blockade in the context of HPV and HIV coinfection. Further understanding of the molecular pathways underlying the responses to immune checkpoint blockade, resulting in tumor reduction is required (261).

The routine screening of pre-cancerous lesions remains the most effective evaluator of cancer development risk. Following cancer development, clinical management of HPV-related malignancies in immunocompromised people is multifactorial and dependent on cancer type and stage. In the ART era, alternative therapies, such as chemotherapy, have been increasingly offered alongside targeted therapy. A significant challenge is designing a treatment plan that will minimize the risk of leukopenia and immune dysfunction, increasing the risk of other opportunistic infections. For PLWH, ART is continued through cancer management to preserve adequate adaptive immunity. Surgery is useful if lesions are located in specific areas with well-defined margins and can involve electrodesiccation to destroy tumor cells. Chemotherapeutic drugs and HIV medication are metabolized through cytochrome p450 within the liver, with some chemotherapy agents regulating or inhibiting this enzyme which can in turn decrease treatment efficacy or increase toxicity. As with chemotherapy, ART can also interfere with the pharmacology of targeted therapy (262, 263).

6 Advances in HPV therapeutic strategies

6.1 Current conventional therapies and treatment plans

The 9-valent HPV (9vHPV) prophylactic vaccine targets HPV types 6, 11, 16, 18, 31, 33, 45, 52, and 58 (264) and has demonstrated efficacy in reducing HPV incidence in immunocompromised people (170). he 9vHPV vaccine covers the major HR-HPV and LR-HPV genotypes responsible for 70% of cervical cancers and 90% of other HPV-associated cancers (265, 266). Immunogenicity data suggest that a two-dose regimen with a 6-month interval is effective in immunocompetent individuals 16 years old and under, while a three-dose regime is recommended for those older than 16 years old (170, 267). Although a gender-neutral HPV vaccination regime inclusive of both boys and girls is beginning to be implemented in some upper-income countries (268), the major challenge lies in lower-income countries where vaccinating young girls is still a challenge due to limited resources. Furthermore, despite this new gender-neutral regime taking place, there is a significantly lower number of boys getting vaccinated, with 44% and 5% of boys being vaccinated in high-income and low/middle-income countries respectively. Nevertheless, prophylactic vaccines are not effective against established HPV infection and there is a need for a therapeutic HPV vaccine or specifically targeted immunotherapy for individuals that have already acquired HR-HPV.

For immunocompromised individuals, the long-term vaccine efficacy of prophylactic vaccines is poorly understood. Currently, no approved therapeutics targeting HPV are available for use postinfection. As the incidence of HPV rises in low-socioeconomic nations, there is a requirement for any novel HPV therapeutic (prophylactic or curative) to be cost-effective, globally distributable, and equally accessible whilst covering a large proportion of HR-HPV genotypes (269). As HPV poses a significant lifetime risk of associated cancer development within an immunocompromised host, a therapeutic capable of directly targeting HPV within infected cells or boosting the host's immune response to HPV is essential. Whilst current vaccination strategies utilize the L1/L2 capsid proteins as targets, this approach fails to account for limited L1/L2 expression in basal epithelial tissue where carcinogenesis occurs, preventing the initiation of a strong immune response (270). While prophylactic vaccination aims to prevent infection, therapeutic vaccines aim to generate robust cytotoxic lymphocyte responses against E6 and E7 viral proteins (269). Several different therapeutic vaccines are currently in development, including protein and peptide-based vaccines, live vector-based, DNA, and specific-cell-based vaccines, which have been discussed in detail elsewhere (271).

In contrast to prophylactics, host-directed therapies for HPVdriven cancer focus on the generation of host immune responses against antigens associated with cellular transformation. Such treatments include adoptive cell transfer, genetically engineered T cell therapy [T-cell receptors (TCRs) and chimeric antigen receptors (CARs)] (272, 273), and the use of immune checkpoint inhibitors (255, 258–260). Adoptive cell transfer, involves autologous antigenspecific CTL expansion *ex vivo*, with re-delivery to the patient's circulation (274). Although promising, the reintroduction of CTL to the host is highly dependent on immune and tumor cell signaling to maintain an effective immune response. This can result in varying levels of effectiveness. A major challenge of emerging immune or antiviralbased therapies targeting HPV is that a clear advantage over current treatment must be demonstrated whilst ensuring patient safety. Developing such therapeutics for use in immunocompromised patients has additional complexity, due to a suppressed immune response. Equal consideration for potential drug interactions with concurrent medications in both HIV individuals on ART and transplant patients on anti-rejection regimens is required. Therefore, a novel approach to the development of therapeutics is targeting HPVspecific proteins or genes, to prevent tumor establishment and cancer progression in hosts where the immune system is impaired.

A comprehensive list of therapeutic and immunotherapy-based developments has been previously summarised (269, 275), however minimal data is available for those that are immunocompromised. Despite major developments in therapeutic vaccinations and immunotherapies, a need to develop treatments that can cater to individuals that do not have an intact immune system persists. A thorough understanding of HPV-cellular interaction, virus-host interplay, and the tumor microenvironment's role in the development of immunotherapies that are specifically effective toward HPV-transformed cells is required (66). It is likely that for immunocompromised hosts, successful and effective treatment plans will be multi-pronged, focusing on aspects such as tumor microenvironment modulation, and specific alteration of HPV-infected cells to make them less resistant to therapeutic intervention.

6.2 RNA-based therapeutic strategies against HPV

Whilst therapeutic regimens exist in relation to HPV, all current therapeutics premise on a functional immune system. This excludes a small yet significant range of patients from obtaining adequate care. With the exception of mRNA-based vaccination (which necessitates an immune system by definition), RNA-based therapeutics hold the unique ability to operate in severely immunocompromised patient populations.

6.2.1 mRNA prophylactic vaccination

Next-generation mRNA-based vaccinations are a novel technology applicable to HPV that has shown significant promise in the fight against SARS-CoV-2 over the past two years and demonstrated safe and highly efficacious responses in typical and atypical patient populations (276, 277). Compared to traditional HPV vaccine candidates, mRNA vaccinations designed against HPV proteins lack any potential risk of causing disease, enable the regulation of immunogenicity, and evade anti-vector immunity (278). mRNA vaccination results in type I interferon (IFN-I) pathway activation and the production of pro-inflammatory cellular cytokines and chemokines, promoting a stronger immune response via increased APC stimulation (279). Cost remains the most significant hurdle to the widespread adoption of mRNA vaccination, as scalability and availability of suitable manufacturing facilities increase the cost per unit (280). Despite the challenges associated with initial cost, the flexibility offered by mRNA vaccination in response to novel pathogens makes them an appealing and viable avenue for therapeutic development. Investigation into the development of a single-dose prophylactic mRNA-based vaccination against HPV may also enable the greater prevention of new HPV incidences in developing regions by eliminating the need for two vaccine doses, as currently recommended by global governing bodies, reducing the load on local health infrastructure whilst improving intergovernmental responses to novel viral variants.

6.2.2 Zinc finger nucleases

Zinc finger nucleases (ZFN) are engineered, modifiable nucleases consisting of a Fokl DNA-cleaving domain bound to zinc finger proteins. Two complimentary ZFN join to form an active ZFN complex capable of recognizing sequence motifs \geq 24 base pairs (281). ZFN is programable by altering the amino acid sequence of each zinc finger, enabling site-directed double-stranded cleavage to suppress transcription (281).

ZFN has been demonstrated to be efficacious in the disruption of the HPV E7 protein expression *in vitro* and *in vivo*, recognizing HPVspecific genomic target sites and initiating double-stranded cleavage (282). It has been found that by targeting the E7 DNA region of HPV16+ (SiHa/CaSki) and HPV18+ (HeLa) cell lines, HPV-specific ZNF can effectively induce disruption of E7 oncogenes, leading to type-specific growth inhibition and apoptosis of HPV+ cells (282). ZFN have similarly shown promise in regulating HIV resistance *in vitro* and *in vivo* (283).

While ZFN provides numerous advantages including site-directed epigenetic modification, significant disadvantages arise. ZFN are difficult to construct, limited in target selection, and remain expensive in comparison to second-generation and third-generation genomic editing technologies, restricting their widespread adoption for use in HPV treatments in developing regions (284, 285).

6.2.3 Clustered regularly interspaced short palindromic repeat

Since its first reports in 1993, CRISPR systems have quickly developed to become one of the most powerful tools associated with the field of genomics (286). CRISPR-associated (Cas) proteins enable the precise epigenetic regulation of DNA through either insertion, deletion, or mutation of the genomic target. Many Cas proteins are nucleases that cleave the double-stranded DNA at the site of the genomic target, however, it is possible to affix various classes of effectors proteins (287). CRISPR/Cas9 specifically locates and binds a protospacer adjacent motif (PAM) within the host genome (288). An associated guide RNA interrogates the DNA strand to locate the complementary genomic target. Following the location of the specified target, the CRISPR/Cas system induces epigenetic modification to alter gene transcription and associated protein expression (289). Nuclease sites of the Cas protein cleave the DNA, which can be repaired via end joining, resulting in mutagenesis, or homology-directed repair (290).

CRISPR/Cas9 technology has shown significant promise in the suppression of virally expressed HPV E6/E7. CRISPR/Cas9 mediated frameshift knockout of the E7 oncogene has been shown to significantly inhibit aberrant cell proliferation of HPV18+ Hela and

HPV16+ SiHa-associated cancerous activity in vitro. In vivo confirmation using micelle delivery, CRISPR/Cas9 within xenografted mice demonstrated significant E7 knockout (291, 292). Additionally, CRISPR/Cas9 systems developed against E6 and E7 have been shown to restore the protein expression of p53 and phospho-pRB (292). Increased cellular levels p56 and phospho-pRB mitigate the hallmarks of HPV cancerous lesion progression by: (I) reducing cellular proliferation (II) ceasing DNA synthesis (III) halting cell cycle progression (IV) inducing apoptosis (V) down-regulating expression of E2F-1 and bcl-2 (293). These findings suggest that CRISPR/Cas9 systems hold the potential to strongly inhibit tumorigenesis post HPV genome integration and serve as a viable therapeutic avenue, although further investigation into unintended off-target effects is required for advancement to clinical trials. Further, the delivery of CRISPR/Cas9 therapeutics requires not just a gRNA molecule, but also the large Cas9 protein or mRNA, increasing the manufacturing complexity, cost and size of the delivery carrier.

6.2.4 Ribonucleic acid interference

Ribonucleic acid interference (RNAi) is a promising genesilencing technology that can be applied to HPV therapeutic research to restrict viral replication and improve host clearance. RNAi is a highly conserved, biological process that can operate *via* two distinct pathways in response to double-stranded ribonucleic acid (dsRNA) (1): *via* the targeting of transcribed messenger ribonucleic acid (mRNA) products within the cell cytoplasm, termed posttranscriptional gene silencing (PTGS); or 2) by directly targeting the gene promoter region located within the cell nucleus, in a process termed transcriptional gene silencing (TGS) (294). These mechanisms can be used to downregulate or impede gene expression for therapeutic application (Figure 3). Whilst numerous dsRNA duplex classes can operate *via* RNAi, short interfering ribonucleic acid (siRNA) is one class that can act *via* both the PTGS and TGS RNAi pathways (295, 296). siRNAs are 19-24 base pair (bp) double-stranded (ds) oligonucleotides, comprising of a 3'-5' active guide (antisense) strand and 5'-3' passive passenger (sense) strand (297).

PTGS-mediated siRNA targeting HPV oncogenic proteins E6/E7 significantly decreases the levels of mRNA transcripts associated with infection. Downregulation of viral gene expression has been demonstrated *in vitro* within the HPV16+ SiHa and CaSki, and HPV18+ Hela cell lines (296, 298, 299). Reduction in viral gene expression enables increased intercellular levels of p53 and pRb, which serve to mediate the effects of HPV. Findings obtained in commercial cell lines were replicated *in vivo* using nude mice, where treatment with PTGS-mediated siRNA targeting mRNA associated with E6 or E7 significantly reduced tumor size, weight and oncogenic expression (300–303). PTGS siRNA is highly effective, but restrictive as only a singular gene product can be targeted per engineered strand, compared to TGS-mediated promoter-targeted siRNA, which can target multiple gene products under the regulation of the same promoter.

TGS-mediated siRNA targeting viral gene promoters have similarly shown strong efficacy in reducing E6/E7 mRNA

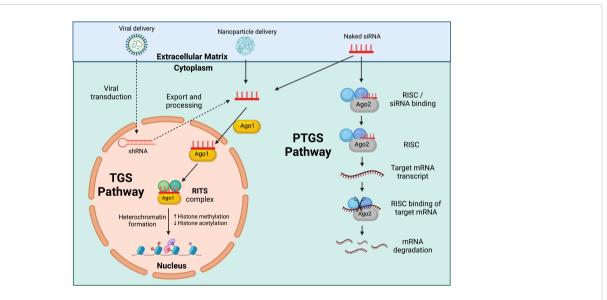


FIGURE 3

RNAi pathways. Post Transcriptional Gene Silencing (PTGS) inducing short interfering ribonucleic acid (siRNA) operate via the actions of the ribonucleic acid (RNA) induced silencing complex (RISC), a multiprotein nuclease complex comprised of Argonaute-2 (Ago-2), Dicer, TAR RNA binding protein, protein kinase interferon-inducible double stranded RNA dependent activator, and GW domain protein 182 in humans. The antisense strand binds Ago-2 and is incorporated into the RISC complex. The antisense strand guides the complex to the targeted mRNA product, where Ago-2 facilitates the cleavage of, and subsequent degradation of the messenger ribonucleic acid (mRNA). Transcriptional Gene Silencing (TGS) inducing siRNA operate via the actions of the RNA induced transcriptional silencing (RITS) complex, a multiprotein complex comprised of Argonaute-1 (Ago-1) and other, yet unidentified, proteins in humans. The antisense strand binds Ago-1 and is incorporated into the RISC complex into the cell nucleus and targets the gene promoter. Heritable epigenetic modifications result in compaction of chromatin structures surrounding the target site, preventing the transcription of gene products, and silencing expression. Created with BioRender.com.

expression using a single siRNA targeting the site of transcription initiation (304). TGS siRNA targeting the common HPV E6/E7 promoters p97 (HPV16) or p105 (HPV18) have shown significant down-regulation of mRNA associated with E6 and E7 *in vivo* (305– 307). Like PTGS siRNA, the reduction in viral gene expression enables increased intercellular levels of p53 and pRB, which serve to mediate the carcinogenic effects of HPV infection. A striking advantage of TGS-mediated siRNA as a therapeutic means of targeting HPV infection is the innate ability to target numerable gene products if regulated by a common gene promoter and the heritable repressive epigenetic modifications that are passed to daughter cells, resulting in longer and more durable silencing effect (304).

Central to the uptake of RNAi by the general populous are cost and availability. RNAi poses unique challenges to both. The costs associated with the research, development, and implementation of an RNAi platform are considerable (308). Although the manufacturing capability of RNA and the necessary delivery platform needed to deliver the inherently unstable RNA into the host cell (309) is expanding rapidly in response to the uptake of mRNA vaccinations. These necessary investments translate to high treatment costs. FDA approved siRNA therapeutic yearly list costs include: patisiran (Onpattro), 451,430 USD; GIVLAARI (givosiran), 575,000 USD; and AMVUTTRA[®] (Vutrisiran), 463,500 USD (310-312). These costs are considerable and would need to decrease in order to provide equitable access to RNAi-based therapeutics. Despite the initial costs, advancements in process design, delivery platform development, and RNA synthesis have enabled a steady decrease in the production costs associated with RNAi therapeutics, reducing costs and broadening the expected patient population (313). Significant to the promise of RNAi therapeutics is the unique adaptability to combat evolving viral pathogens and address several heritable genetic conditions (294, 309, 314). Following continued advancement in translational RNA research, improved access to RNAi therapeutics could enable widespread adoption and the subsequent treatment of numerable viral and genomic conditions currently without therapeutic options.

6.3 Delivery of siRNA or mRNA therapeutics and applications

Currently, siRNA delivery remains technically challenging on several fronts (315). RNAi delivery processes must prevent the destruction of the inherently unstable, naked RNA prior to entering the target cell and inducing silencing. Immune and metabolic factors dramatically limit delivery mechanisms as they must be able to overcome administrative, as well as vascular and cellular barriers (316). These challenges, therefore, require stabilizing modifications to siRNA sequences that can include 2'-O-methylation on the 3' terminal ribose and/or carriers to facilitate the functional delivery of RNAi therapeutics (317). Three delivery carriers that serve to overcome these factors are nanoparticle delivery, GalNAc conjugates, and lentiviral vector delivery (318).

Delivery through the employment of chemically modified multifunctional nanoparticles serves to encapsulate the negatively charged naked siRNA and effectively facilitate transportation to the target cell without degradation caused by host barriers (Figure 4) (319) At the time of publication, one therapeutic utilizing nanoparticle technology: Onpattro (Patisiran) has been approved by the US FDA for therapeutic use *via* intravenous infusion (320). Additionally, recent success surrounding the safe and effective implementation of nanoparticle-delivered RNA-based vaccinations in response to SARS-Cov-2 heightens the potential for nanoparticle delivery of genetic-based therapies (276, 277).

GalNac-siRNA conjugates are another form of siRNA delivery that bind to the Asialoglycoprotein receptor widely present in liver hepatocytes facilitating siRNA entry. Four siRNA utilizing GalNAc conjugates: Oxlumo (Lumasiran), GIVLAARI (givosiran), Leqvio[®] (inclisiran), and AMVUTTRA[®] (Vutrisiran), have been approved by the United States FDA for therapeutic administration, however, the potential for GalNac-siRNA applications outside targeting liver disorders is less likely (321–325).

Lentivirus vectors expressing short hairpin (sh)RNA, which match the siRNA target sequence and include a linking loop between the sense and antisense strands, similarly allowing for the

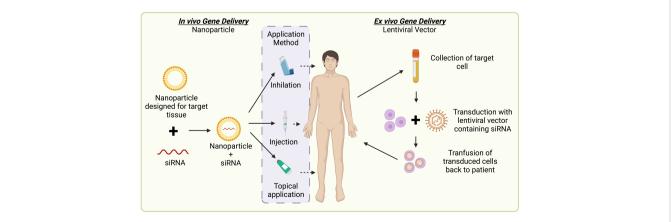


FIGURE 4

Comparative diagram of *in vivo / ex vivo* gene deliveries. *In-vivo* gene delivery via the use of nanoparticles. Conjugation of the nanoparticle designed for the target cell and the siRNA designed against the targeted genetic material are applied to the patient based of disease characteristics (i.e topical application for HPV infected cervical cells). *Ex-vivo* gene delivery via the use of lentiviral vectors requiring the collection of patient cells, transduction with the constructed lentiviral vector before re-introduction of the transduced patient cells back into the patient. Created with **BioRender.com**.

silencing of a targeted gene through the facilitated entry of targeted cells (326). The designed genetic material of the lentiviral genome can be transcribed to interfere with pathogenic viral RNA within the same cell, resulting in the downregulation of the targeted pathogenic viral protein (327). Lentiviral vector shRNA delivery uses an *ex vivo* gene therapy model, whereby patient cells are first isolated and transduced outside of the body prior to infusion back into the patient (Figure 4) (328). The use of lentiviral vector-mediated RNAi has been shown to reduce cellular proliferation and tumorigenesis of HPV-associated cancers and neuroblastoma *in vivo* and serves as a viable route for further inquiry (329, 330). However, there remains safety concerns with the lack of control of lentiviral integrations site into the host genome, and thus studies are ongoing to improve this aspect of lentiviral gene delivery (331).

7 Discussion

The epidemiological trends and biological mechanisms relating to HR-HPV and human carcinogenesis have been well documented. Additionally, there have been significant advancements made in preventative measures and therapeutic developments. Currently, the highest prevalence of HPV-related disease is in Sub-Saharan Africa, Eastern Europe, and Latin America (24, 25). Rates of cervical cancer are increasing and HPV+OPSCC are decreasing in western countries respectively. People living with immunodeficiencies are at a disproportionately high risk of persistent HPV infections and progressive disease compared to the general population.

Adequate immunosurveillance is crucial for viral elimination and preventing disease establishment. HPV-specific CD4⁺ and CD8⁺ T cells and inflammatory cytokines have been shown to negatively correlate with disease severity, with T cells being particularly important in targeting E6 and E7 epitopes (82). Where immunosurveillance is compromised, HPV disrupts critical signaling pathways such as JAK/ STAT, apoptotic (TRAIL and XAF1), and IFN- γ (STAT1) resulting in immune evasion.

Lymphopenia is a common feature in those with specific primary and secondary immunodeficiencies at high risk of HPV-related disease. Patients with PIDs predisposed to HPV-related disease (103, 104), are predominantly associated with a CD4⁺ T cell lymphopenia, a decrease in CD8⁺ T cells and NK cells, and impaired TCR signaling; all factors important for targeting and eliminating HPV. CD4⁺ T cell lymphopenia is also evident up to 10 years post-renal transplant, a long-term effect associated with accelerated renal allograft decline and a high risk of cancer development compared to non-CD4⁺ T cell lymphopenic patients (154, 155). Despite ART improving prolonged increases in immune function (332) in PLWH, there is still a proportion who do not clear their lesions and are at high risk of developing HPV-driven cancer.

Tissue-resident memory T cell subsets and their roles in those with immunodeficiencies also require further investigation. This may be crucial to understanding tissue-specific responses in at-risk populations. Many reports focus on T cell responses in peripheral blood to assess immune response, which singularly is not the most accurate measure of immune function or cancer development. High CD8⁺ T_{RM} cell numbers have been correlated with better patient prognosis in HPV+OPSCC compared to their peripheral counterparts

(243) and have shown to be a good prognostic indicator in many other malignancies including melanoma, ovarian and cervical cancer (333). There is a need to better understand tissue-specific responses to HPV, especially in immunocompromised individuals.

The clinical management of several malignancies remains consistent, irrespective of HPV status revealing a major limitation to current practices for poor responders and immunocompromised people. Currently, screening for HR-HPV detection or pre-cancerous lesions to aid in early diagnosis is only well established in cervical cancer compared to other HPV-driven cancers. While surgery can be combined with radiotherapy and chemotherapy, this approach is associated with substantial morbidity. In those established primary, secondary, and acquired immunodeficiency, opportunistic pathogens can establish disease. There is a need for more sophisticated, targeted treatments, especially for those with impaired immunity. Despite prophylactic vaccines showing high efficacy in the general population, long-term benefit in immunocompromised hosts is not well understood. Additionally, vaccine protection is limited to HR-HPVs naive to the host and therefore has major limitations as a preventative strategy. This is also applicable to HPV latency in infected cells. There is a need to address protection against LR-HPVs which can manifest severe clinical disease in immunodeficient individuals. There has been an acceleration in the investigation of cell-based therapies, immune checkpoint inhibitors, and therapeutic vaccines to address these limitations.

Host-directed therapies hold great promise for the treatment of patients with advanced-stage or relapsed HPV-associated cancers, however, there has been limited investigation into the efficacy of these in immunocompromised populations. Currently, PD-1/PD-L1 blockade has shown to improve overall survival in HPV+ individuals compared to chemotherapy, however, there is limited data on checkpoint inhibitor outcomes in specific at-risk populations. Furthermore, whether transplant patients and PLWH have different responses to checkpoint inhibition, regardless of HPV status, requires further elucidation. Several studies have shown that PD-1 inhibition (249) improves survival in HNSCC but has not been replicated in cervical cancer or ASCC, possibly suggesting divergent local tumor microenvironments at play (257, 259, 260). Additionally, the potential of therapeutic vaccines in combination with radiotherapy, chemotherapy or checkpoint inhibition requires further studies.

RNA-based therapeutic systems do not require an intact immune system (with the exception of mRNA vaccination) and hold great potential for the treatment of chronic pathological infections that resist traditional treatment approaches. RNA-associated downregulation of genes associated with HPV malignancies may inhibit viral genomic expression and reduce cellular proliferation by restoring normal intracellular p53 and pRb levels through the prevention or destruction of oncogenic RNA transcripts (334). Further development of RNA technologies against the early and late-stage promoters of the HPV genome could enable the silencing of multiple proteins simultaneously, reducing the likelihood of further cellular transformation. The disabling of viral replication and subsequent aberrant cellular development may facilitate the increased clearance of chronic HPV infection and result in improved clinical outcomes. Therefore, the investigation into RNA therapeutics serves as a logical route for next-generation antiviral therapeutics targeting HPV.

8 Conclusion

Given the inaccessibility of health infrastructure to provide sterilizing HPV vaccines in many parts of the world, and the existing high burden of HPV infection that can no longer be cured by prophylaxis, additional approaches are needed to treat the resulting pre-malignant and cancerous lesions. While there is a large body of research regarding the epidemiology of HPV-associated cancers in the immunocompromised population, it would be beneficial to investigate the effectiveness of currently available treatments in this sample patient population. There is a growing need to investigate the safety and immunogenicity of prophylactic HPV vaccines in PLWH. This will be important for determining the efficacy of these vaccines and the degree of risk of developing HPV-associated diseases. Although the use of ICB appears promising, particularly in regard to the use of PD-1 inhibitors for patients with HPV-driven cancers, treatment efficacy in immunocompromised individuals is not well understood. Given that people living with primary and secondary immunodeficiency have high rates of persistent, extensive, and malignant HPV-driven disease, such host-directed therapies may have divergent results from those seen in immunocompetent people. The rapid advancement of frontier gene-based RNA therapies, of which most do not require a functional immune system, may therefore play a pivotal role in eventually reducing the excess morbidity and mortality of HPVdriven disease in those most at-risk.

References

1. De Martel C, Georges D, Bray F, Ferlay J, Clifford GM. Global burden of cancer attributable to infections in 2018: A worldwide incidence analysis. *Lancet Global Health* (2020) 8(2):e180–e90. doi: 10.1016/S2214-109X(19)30488-7

2. Parkin DM. The global health burden of infection-associated cancers in the year 2002. Int J Cancer (2006) 118(12):3030-44. doi: 10.1002/ijc.21731

3. McLaughlin-Drubin ME, Munger K. Viruses associated with human cancer. Biochim Biophys Acta (2008) 1782(3):127-50. doi: 10.1016/j.bbadis.2007.12.005

4. Chinen J, Shearer WT. Secondary immunodeficiencies, including hiv infection. J Allergy Clin Immunol (2010) 125(2 Suppl 2):S195-203. doi: 10.1016/j.jaci.2009 .08.040

5. Liao JB. Cancer issue: Viruses and human cancer. Yale J Biol Med (2006) 79(3-4):115. doi: 10.3390/v6104047

6. De Villiers E-M, Fauquet C, Broker TR, Bernard H-U, zur Hausen H. Classification of papillomaviruses. *Virology* (2004) 324(1):17–27. doi: 10.1016/j.virol.2004.03.033

7. Shamanin V, Glover M, Rausch C, Proby C, Leigh IM, Zur Hausen H, et al. Specific types of human papillomavirus found in benign proliferations and carcinomas of the skin in immunosuppressed patients. *Cancer Res* (1994) 54(17):4610–3.

8. Shamanin V, Hausen HZ, Lavergne D, Proby CM, Leigh IM, Neumann C, et al. Human papillomavirus infections in nonmelanoma skin cancers from renal transplant recipients and nonimmunosuppressed patients. *JNCI: J Natl Cancer Institute* (1996) 88 (12):802–11. doi: 10.1093/jnci/88.12.802

9. Bosch FX, Broker TR, Forman D, Moscicki AB, Gillison ML, Doorbar J, et al. Comprehensive control of human papillomavirus infections and related diseases. *Vaccine* (2013) 31 Suppl 7(Suppl 7):H1–31. doi: 10.1016/j.vaccine.2013.10.003

10. Evander M, Edlund K, Gustafsson A, Jonsson M, Karlsson R, Rylander E, et al. Human papillomavirus infection is transient in young women: A population-based cohort study. *J Infect Dis* (1995) 171(4):1026–30. doi: 10.1093/infdis/171.4.1026

11. Ho GY, Bierman R, Beardsley L, Chang CJ, Burk RD. Natural history of cervicovaginal papillomavirus infection in young women. N Engl J Med (1998) 338 (7):423-8. doi: 10.1056/nejm199802123380703

12. De Jong A, van der Burg SH, Kwappenberg KM, van der Hulst JM, Franken KL, Geluk A, et al. Frequent detection of human papillomavirus 16 E2-specific T-helper immunity in healthy subjects. *Cancer Res* (2002) 62(2):472–9.

13. Woo YL, van den Hende M, Sterling JC, Coleman N, Crawford RA, Kwappenberg KM, et al. A prospective study on the natural course of low-grade squamous intraepithelial lesions and the presence of Hpv16 E2-, E6- and E7-specific T-cell responses. *Int J Cancer* (2010) 126(1):133–41. doi: 10.1002/ijc.24804

Author contributions

RH and JA reviewed the literature, created, and designed tables and figures, and wrote the manuscript. SS and CA reviewed the literature and revised and proofread the manuscript. All authors contributed to the article and approved the submitted version.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

14. Wang CJ, Sparano J, Palefsky JM. Human immunodeficiency Virus/Aids, human papillomavirus, and anal cancer. *Surg Oncol Clin N Am* (2017) 26(1):17–31. doi: 10.1016/j.soc.2016.07.010

15. Ellerbrock TV, Chiasson MA, Bush TJ, Sun XW, Sawo D, Brudney K, et al. Incidence of cervical squamous intraepithelial lesions in hiv-infected women. *Jama* (2000) 283(8):1031–7. doi: 10.1001/jama.283.8.1031

16. Halpert R, Fruchter RG, Sedlis A, Butt K, Boyce JG, Sillman FH. Human papillomavirus and lower genital neoplasia in renal transplant patients. *Obstet Gynecol* (1986) 68(2):251–8.

17. Brickman C, Palefsky JM. Human papillomavirus in the hiv-infected host: Epidemiology and pathogenesis in the antiretroviral era. *Curr HIV/AIDS Rep* (2015) 12 (1):6–15. doi: 10.1007/s11904-014-0254-4

18. Wieland U, Kreuter A, Pfister H. Human papillomavirus and immunosuppression. *Curr Probl Dermatol* (2014) 45:154–65. doi: 10.1159/000357907

19. Chu CS, Pfister DG. Opportunities and challenges: Human papillomavirus and cancer. J Natl Compr Cancer Netw (2017) 15(5S):726–9. doi: 10.6004/jnccn.2017.0085

20. Razzaghi H, Saraiya M, Thompson TD, Henley SJ, Viens L, Wilson R. Five-year relative survival for human papillomavirus-associated cancer sites. *Cancer* (2018) 124 (1):203–11. doi: 10.1002/cncr.30947

21. Dryden-Peterson S, Bvochora-Nsingo M, Suneja G, Efstathiou JA, Grover S, Chiyapo S, et al. Hiv infection and survival among women with cervical cancer. J Clin Oncol (2016) 34(31):3749–57. doi: 10.1200/jco.2016.67.9613

22. Bower M, Powles T, Newsom-Davis T, Thirlwell C, Stebbing J, Mandalia S, et al. Hiv-associated anal cancer: Has highly active antiretroviral therapy reduced the incidence or improved the outcome? *J Acquir Immune Defic Syndr* (2004) 37(5):1563–5. doi: 10.1097/00126334-200412150-00004

23. Chesson HW, Dunne EF, Hariri S, Markowitz LE. The estimated lifetime probability of acquiring human papillomavirus in the united states. *Sex Transm Dis* (2014) 41(11):660–4. doi: 10.1097/olq.00000000000193

24. Bruni L, Diaz M, Castellsagué X, Ferrer E, Bosch FX, de Sanjosé S. Cervical human papillomavirus prevalence in 5 continents: Meta-analysis of 1 million women with normal cytological findings. *J Infect Dis* (2010) 202(12):1789–99. doi: 10.1086/657321

25. Wu J, Xiao F, Zheng Y, Lin Y, Wang HL. Worldwide trend in human papillomavirus-attributable cancer incidence rates between 1990 and 2012 and Bayesian projection to 2030. *Cancer* (2021) 127(17):3172–82. doi: 10.1002/cncr.33628

26. Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: Globocan estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* (2021) 71(3):209–49. doi: 10.3322/caac.21660

27. National Cancer Intitute. *Hpv and cancer: National cancer intitute* (2023). Available at: https://www.cancer.gov/about-cancer/causes-prevention/risk/infectious-agents/hpv-and-cancer#:~:text=High%2Drisk%20HPVs%20cause%20about,HPV% 2Drelated%20cancer%20each%20year.

28. Schwartz RH. Acquisition of immunologic self-tolerance. Cell (1989) 57(7):1073-81. doi: 10.1016/0092-8674(89)90044-5

29. IARC Working Group on the Evaluation of Carcinogenic Risks to Humans. *Human papillomaviruses*. Lyon, France : Geneva, Switzerland: World Health Organization, International Agency for Research on Cancer (2007).

30. Lagheden C, Eklund C, Lamin H, Kleppe SN, Lei J, Elfström KM, et al. Nationwide comprehensive human papillomavirus (Hpv) genotyping of invasive cervical cancer. *Br J Cancer* (2018) 118(10):1377–81. doi: 10.1038/s41416-018-0053-6

31. Carlander AF, Jakobsen KK, Bendtsen SK, Garset-Zamani M, Lynggaard CD, Jensen JS, et al. A contemporary systematic review on repartition of hpv-positivity in oropharyngeal cancer worldwide. *Viruses* (2021) 13(7). doi: 10.3390/v13071326

32. Zhang J, Zhang Y, Zhang Z. Prevalence of human papillomavirus and its prognostic value in vulvar cancer: A systematic review and meta-analysis. *PloS One* (2018) 13(9):e0204162. doi: 10.1371/journal.pone.0204162

33. Olesen TB, Sand FL, Rasmussen CL, Albieri V, Toft BG, Norrild B, et al. Prevalence of human papillomavirus DNA and P16(Ink4a) in penile cancer and penile intraepithelial neoplasia: A systematic review and meta-analysis. *Lancet Oncol* (2019) 20(1):145–58. doi: 10.1016/s1470-2045(18)30682-x

34. Chaturvedi AK. Beyond cervical cancer: Burden of other hpv-related cancers among men and women. *J Adolesc Health* (2010) 46(4 Suppl):S20-6. doi: 10.1016/j.jadohealth.2010.01.016

35. Muñoz N, Bosch FX, de Sanjosé S, Herrero R, Castellsagué X, Shah KV, et al. Epidemiologic classification of human papillomavirus types associated with cervical cancer. *N Engl J Med* (2003) 348(6):518–27. doi: 10.1056/NEJMoa021641

36. Chelimo C, Wouldes TA, Cameron LD, Elwood JM. Risk factors for and prevention of human papillomaviruses (Hpv), genital warts and cervical cancer. J Infect (2013) 66(3):207–17. doi: 10.1016/j.jinf.2012.10.024

37. IARC Working Group on the Evaluation of Carcinogenic Risks to Humans. Human Papillomaviruses. Lyon (FR): International Agency for Research on Cancer; 2007. (IARC Monographs on the Evaluation of Carcinogenic Risks to Humans, No. 90.) 5, Summary of Data Reported and Evaluation. (2007). Available at https://www.ncbi.nlm. nih.gov/books/NBK321767/.

38. IARC Working Group on the Evaluation of Carcinogenic Risks to Humans. Personal Habits and Indoor Combustions. Lyon (FR): International Agency for Research on Cancer; 2012. (IARC Monographs on the Evaluation of Carcinogenic Risks to Humans, No. 100E.). *TOBACCO SMOKING* (2012). Available at: https://www.ncbi.nlm.nih.gov/books/NBK304395/.

39. Aguayo F, Muñoz JP, Perez-Dominguez F, Carrillo-Beltrán D, Oliva C, Calaf GM, et al. High-risk human papillomavirus and tobacco smoke interactions in epithelial carcinogenesis. *Cancers (Basel)* (2020) 12(8). doi: 10.3390/cancers12082201

40. Kumar R, Rai AK, Das D, Das R, Kumar RS, Sarma A, et al. Alcohol and tobacco increases risk of high risk hpv infection in head and neck cancer patients: Study from north-East region of India. *PloS One* (2015) 10(10):e0140700. doi: 10.1371/journal.pone.0140700

41. Appleby P, Beral V, Berrington de González A, Colin D, Franceschi S, Goodill A, et al. Carcinoma of the cervix and tobacco smoking: Collaborative reanalysis of individual data on 13,541 women with carcinoma of the cervix and 23,017 women without carcinoma of the cervix from 23 epidemiological studies. *Int J Cancer* (2006) 118 (6):1481–95. doi: 10.1002/ijc.21493

42. Grulich AE, Poynten IM, Machalek DA, Jin F, Templeton DJ, Hillman RJ. The epidemiology of anal cancer. Sex Health (2012) 9(6):504-8. doi: 10.1071/sh12070

43. Muñoz JP, Carrillo-Beltrán D, Aedo-Aguilera V, Calaf GM, León O, Maldonado E, et al. Tobacco exposure enhances human papillomavirus 16 oncogene expression *Via* Egfr/Pi3k/Akt/C-jun signaling pathway in cervical cancer cells. *Front Microbiol* (2018) 9:3022. doi: 10.3389/fmicb.2018.03022

44. Muñoz N, Franceschi S, Bosetti C, Moreno V, Herrero R, Smith JS, et al. Role of parity and human papillomavirus in cervical cancer: The iarc multicentric case-control study. *Lancet* (2002) 359(9312):1093–101. doi: 10.1016/s0140-6736(02)08151-5

45. Tekalegn Y, Sahiledengle B, Woldeyohannes D, Atlaw D, Degno S, Desta F, et al. High parity is associated with increased risk of cervical cancer: Systematic review and meta-analysis of case-control studies. *Womens Health (Lond)* (2022) 18:17455065221075904. doi: 10.1177/17455065221075904

46. Moreno V, Bosch FX, Muñoz N, Meijer CJ, Shah KV, Walboomers JM, et al. Effect of oral contraceptives on risk of cervical cancer in women with human papillomavirus infection: The iarc multicentric case-control study. *Lancet* (2002) 359(9312):1085–92. doi: 10.1016/s0140-6736(02)08150-3

47. Anastasiou E, McCarthy KJ, Gollub EL, Ralph L, van de Wijgert J, Jones HE. The relationship between hormonal contraception and cervical Dysplasia/Cancer controlling for human papillomavirus infection: A systematic review. *Contraception* (2022) 107:1–9. doi: 10.1016/j.contraception.2021.10.018

48. Sedjo RL, Roe DJ, Abrahamsen M, Harris RB, Craft N, Baldwin S, et al. Vitamin a, carotenoids, and risk of persistent oncogenic human papillomavirus infection. *Cancer Epidemiol Biomarkers Prev* (2002) 11(9):876–84.

49. Naresh A, Hagensee M, Myers L, Cameron J. Association of diet quality and dietary components with clinical resolution of hpv. *Nutr Cancer* (2021) 73(11-12):2579-88. doi: 10.1080/01635581.2020.1841251

50. La Vecchia C, Franceschi S, Decarli A, Gentile A, Fasoli M, Pampallona S, et al. Dietary vitamin a and the risk of invasive cervical cancer. *Int J Cancer* (1984) 34(3):319–22. doi: 10.1002/ijc.2910340306

51. Giuliano AR, Papenfuss M, Nour M, Canfield LM, Schneider A, Hatch K. Antioxidant nutrients: Associations with persistent human papillomavirus infection. *Cancer Epidemiol Biomarkers Prev* (1997) 6(11):917–23.

52. Palan PR, Chang CJ, Mikhail MS, Ho GY, Basu J, Romney SL. Plasma concentrations of micronutrients during a nine-month clinical trial of beta-carotene in women with precursor cervical cancer lesions. *Nutr Cancer* (1998) 30(1):46–52. doi: 10.1080/01635589809514639

53. De Vet HC, Knipschild PG, Willebrand D, Schouten HJ, Sturmans F. The effect of beta-carotene on the regression and progression of cervical dysplasia: A clinical experiment. J Clin Epidemiol (1991) 44(3):273–83. doi: 10.1016/0895-4356(91)90039-c

54. Piyathilake CJ, Macaluso M, Brill I, Heimburger DC, Partridge EE. Lower red blood cell folate enhances the hpv-16-Associated risk of cervical intraepithelial neoplasia. *Nutrition* (2007) 23(3):203–10. doi: 10.1016/j.nut.2006.12.002

55. Piyathilake CJ, Macaluso M, Chambers MM, Badiga S, Siddiqui NR, Bell WC, et al. Folate and vitamin B12 may play a critical role in lowering the hpv 16 methylationassociated risk of developing higher grades of cin. *Cancer Prev Res (Phila)* (2014) 7 (11):1128–37. doi: 10.1158/1940-6207.Capr-14-0143

56. Butterworth CE Jr, Hatch KD, Soong SJ, Cole P, Tamura T, Sauberlich HE, et al. Oral folic acid supplementation for cervical dysplasia: A clinical intervention trial. *Am J Obstet Gynecol* (1992) 166(3):803–9. doi: 10.1016/0002-9378(92)91337-a

57. Childers JM, Chu J, Voigt LF, Feigl P, Tamimi HK, Franklin EW, et al. Chemoprevention of cervical cancer with folic acid: A phase iii southwest oncology group intergroup study. *Cancer Epidemiol Biomarkers Prev* (1995) 4(2):155–9.

58. McCusker C, Upton J, Warrington R. Primary immunodeficiency. Allergy Asthma Clin Immunol (2018) 14(Suppl 2):61. doi: 10.1186/s13223-018-0290-5

59. Gernert M, Kiesel M, Fröhlich M, Renner R, Strunz PP, Portegys J, et al. High prevalence of genital human papillomavirus infection in patients with primary immunodeficiencies. *Front Immunol* (2021) 12:789345. doi: 10.3389/fimmu.2021.789345

60. Reusser NM, Downing C, Guidry J, Tyring SK. Hpv carcinomas in immunocompromised patients. J Clin Med (2015) 4(2):260-81. doi: 10.3390/jcm4020260

61. Zheng ZM, Baker CC. Papillomavirus genome structure, expression, and post-transcriptional regulation. *Front Biosci* (2006) 11:2286–302. doi: 10.2741/1971

62. Burk RD, Chen Z, Van Doorslaer K. Human papillomaviruses: Genetic basis of carcinogenicity. *Public Health Genomics* (2009) 12(5-6):281–90. doi: 10.1159/000214919

63. Taguchi A, Nagasaka K, Plessy C, Nakamura H, Kawata Y, Kato S, et al. Use of cap analysis gene expression to detect human papillomavirus promoter activity patterns at different disease stages. *Sci Rep* (2020) 10(1):17991. doi: 10.1038/s41598-020-75133-2

64. Egawa N, Doorbar J. The low-risk papillomaviruses. Virus Res (2017) 231:119–27. doi: 10.1016/j.virusres.2016.12.017

65. Burk RD, Harari A, Chen Z. Human papillomavirus genome variants. Virology (2013) 445(1-2):232-43. doi: 10.1016/j.virol.2013.07.018

66. Bernard HU, Burk RD, Chen Z, van Doorslaer K, zur Hausen H, de Villiers EM. Classification of papillomaviruses (Pvs) based on 189 pv types and proposal of taxonomic amendments. *Virology* (2010) 401(1):70–9. doi: 10.1016/j.virol.2010.02.002

67. Jendoubi-Ferchichi M, Satouri L, Ghoul F, Malek-Mellouli M, Derbel AM, Makni MK, et al. Phylogeny and classification of human papillomavirus (Hpv)16 and Hpv18 variants based on E6 and L1 genes in Tunisian women with cervical lesions. *Asian Pac J Cancer Prev* (2018) 19(12):3361–6. doi: 10.31557/apjcp.2018.19.12.3361

68. Lin C, Franceschi S, Clifford GM. Human papillomavirus types from infection to cancer in the anus, according to sex and hiv status: A systematic review and meta-analysis. *Lancet Infect Dis* (2018) 18(2):198–206. doi: 10.1016/s1473-3099(17)30653-9

69. zur Hausen H. Papillomaviruses and cancer: From basic studies to clinical application. *Nat Rev Cancer* (2002) 2(5):342–50. doi: 10.1038/nrc798

70. Doorbar J, Quint W, Banks L, Bravo IG, Stoler M, Broker TR, et al. The biology and life-cycle of human papillomaviruses. *Vaccine* (2012) 30:F55–70. doi: 10.1016/j.vaccine.2012.06.083

71. Carter JJ, Koutsky LA, Wipf GC, Christensen ND, Lee SK, Kuypers J, et al. The natural history of human papillomavirus type 16 capsid antibodies among a cohort of university women. J Infect Dis (1996) 174(5):927–36. doi: 10.1093/infdis/174.5.927

72. Kjær SK, Frederiksen K, Munk C, Iftner T. Long-term absolute risk of cervical intraepithelial neoplasia grade 3 or worse following human papillomavirus infection: Role of persistence. *JNCI: J Natl Cancer Institute* (2010) 102(19):1478–88. doi: 10.1093/jnci/djq356

73. Dunne EF, Park IU. Hpv and hpv-associated diseases. Infect Dis Clin North Am (2013) 27(4):765–78. doi: 10.1016/j.idc.2013.09.001

74. von Knebel Doeberitz M, Oltersdorf T, Schwarz E, Gissmann L. Correlation of modified human papilloma virus early gene expression with altered growth properties in C4-1 cervical carcinoma cells. *Cancer Res* (1988) 48(13):3780–6.

75. Crook T, Vousden KH, Tidy JA. Degradation of P53 can be targeted by hpv E6 sequences distinct from those required for P53 binding and trans-activation. *Cell* (1991) 67(3):547–56. doi: 10.1016/0092-8674(91)90529-8

76. Lipinski MM, Jacks T. The retinoblastoma gene family in differentiation and development. *Oncogene* (1999) 18(55):7873-82. doi: 10.1038/sj.onc.1203244

77. Gonzalez SL, Stremlau M, He X, Basile JR, Münger K. Degradation of the retinoblastoma tumor suppressor by the human papillomavirus type 16 E7 oncoprotein

is important for functional inactivation and is separable from proteasomal degradation of E7. J Virol (2001) 75(16):7583-91. doi: 10.1128/jvi.75.16.7583-7591.2001

78. Duensing S, Münger K. The human papillomavirus type 16 E6 and E7 oncoproteins independently induce numerical and structural chromosome instability. *Cancer Res* (2002) 62(23):7075–82. doi: 10.1073/pnas.170093297

79. Hernandez AL, Efird JT, Holly EA, Berry JM, Jay N, Palefsky JM. Incidence of and risk factors for type-specific anal human papillomavirus infection among hiv-positive msm. *AIDS (Lond Engl)* (2014) 28(9):1341. doi: 10.1097/QAD.00000000000254

 Bashaw AA, Leggatt GR, Chandra J, Tuong ZK, Frazer IH. Modulation of antigen presenting cell functions during chronic hpv infection. *Papillomavirus Res* (2017) 4:58– 65. doi: 10.1016/i.pvr.2017.08.002

81. van der Burg SH, Arens R, Ossendorp F, van Hall T, Melief CJM. Vaccines for established cancer: Overcoming the challenges posed by immune evasion. *Nat Rev Cancer* (2016) 16(4):219–33. doi: 10.1038/nrc.2016.16

82. Nakagawa M, Stites DP, Patel S, Farhat S, Scott M, Hills NK, et al. Persistence of human papillomavirus type 16 infection is associated with lack of cytotoxic T lymphocyte response to the E6 antigens. *J Infect Dis* (2000) 182(2):595–8. doi: 10.1086/315706

83. Bhat P, Bergot AS, Waterhouse N, Frazer IH. Human papillomavirus E7 oncoprotein expression by keratinocytes alters the cytotoxic mechanisms used by Cd8 T cells. *Oncotarget* (2018) 9(5):6015–27. doi: 10.18632/oncotarget.23210

84. Seresini S, Origoni M, Caputo L, Lillo F, Longhi R, Vantini S, et al. Cd4+ T cells against human papillomavirus-18 E7 in patients with high-grade cervical lesions associate with the absence of the virus in the cervix. *Immunology* (2010) 131(1):89–98. doi: 10.1111/j.1365-2567.2010.03277.x

85. Kim KH, Greenfield WW, Cannon MJ, Coleman HN, Spencer HJ, Nakagawa M. Cd4+ T-cell response against human papillomavirus type 16 E6 protein is associated with a favorable clinical trend. *Cancer Immunol Immunother* (2012) 61(1):63–70. doi: 10.1007/ s00262-011-1092-5

86. Stanley MA. Epithelial cell responses to infection with human papillomavirus. Clin Microbiol Rev (2012) 25(2):215–22. doi: 10.1128/CMR.05028-11

87. Koshiol JE, Schroeder JC, Jamieson DJ, Marshall SW, Duerr A, Heilig CM, et al. Time to clearance of human papillomavirus infection by type and human immunodeficiency virus serostatus. *Int J Cancer* (2006) 119(7):1623–9. doi: 10.1002/ijc.22015

88. Schiffman M, Rodriguez AC, Chen Z, Wacholder S, Herrero R, Hildesheim A, et al. A population-based prospective study of carcinogenic human papillomavirus variant lineages, viral persistence, and cervical neoplasia. *Cancer Res* (2010) 70(8):3159–69. doi: 10.1158/0008-5472.CAN-09-4179

89. Zoodsma M, Nolte IM, Te Meerman GJ, De Vries EG, van der Zee AG. Hla genes and other candidate genes involved in susceptibility for (Pre) neoplastic cervical disease. *Int J Oncol* (2005) 26(3):769–84. doi: 10.3892/ijo.26.3.769

90. Ades S, Koushik A, Duarte-Franco E, Mansour N, Arseneau J, Provencher D, et al. Selected class I and class ii hla alleles and haplotypes and risk of high-grade cervical intraepithelial neoplasia. *Int J Cancer* (2008) 122(12):2820–6. doi: 10.1002/ijc.23459

91. Stern PL. Harnessing immunity for therapy in human papillomavirus driven cancers. *Tumour Virus Res* (2021) 11:200212. doi: 10.1016/j.tvr.2021.200212

92. Gutiérrez-Hoya A, Soto-Cruz I. Role of the Jak/Stat pathway in cervical cancer: Its relationship with hpv E6/E7 oncoproteins. *Cells* (2020) 9(10). doi: 10.3390/cells9102297

93. Walch-Rückheim B, Pahne-Zeppenfeld J, Fischbach J, Wickenhauser C, Horn LC, Tharun L, et al. Stat3/Irf1 pathway activation sensitizes cervical cancer cells to chemotherapeutic drugs. *Cancer Res* (2016) 76(13):3872–83. doi: 10.1158/0008-5472.CAN-14-1306

94. Schröer N, Pahne J, Walch B, Wickenhauser C, Smola S. Molecular pathobiology of human cervical high-grade lesions: Paracrine Stat3 activation in tumor-instructed myeloid cells drives local mmp-9 expression. *Cancer Res* (2011) 71(1):87–97. doi: 10.1158/0008-5472.CAN-10-2193

95. Pahne-Zeppenfeld J, Schroer N, Wa Lch-Ruck Hei MB, Oldak M, Gorter A, Hegde S, et al. Cervical cancer cell-derived interleukin-6 impairs Ccr7-dependent migration of mmp-9 expressing dendritic cells. *Int J Cancer* (2014) 134:2061–73. doi: 10.1002/ijc.28549

96. Perea SE, Massimi P, Banks L. Human papillomavirus type 16 E7 impairs the activation of the interferon regulatory factor-1. *Int J Mol Med* (2000) 5(6):661–7. doi: 10.3892/ijmm.5.6.661

97. Um S-J, Rhyu J-W, Kim E-J, Jeon K-C, Hwang E-S, Park J-S. Abrogation of irf-1 response by high-risk hpv E7 protein in vivo. *Cancer Lett* (2002) 179(2):205–12. doi: 10.1016/S0304-3835(01)00871-0

98. Caberg J-HD, Hubert PM, Begon DY, Herfs MF, Roncarati PJ, Boniver JJ, et al. Silencing of E7 oncogene restores functional e-cadherin expression in human papillomavirus 16-transformed keratinocytes. *Carcinogenesis* (2008) 29(7):1441–7. doi: 10.1093/carcin/bgn145

99. Matthews K, Leong CM, Baxter L, Inglis E, Yun K, Bäckström BT, et al. Depletion of langerhans cells in human papillomavirus type 16-infected skin is associated with E6-mediated down regulation of e-cadherin. *J Virol* (2003) 77(15):8378–85. doi: 10.1128/JVI.77.15.8378-8385.2003

100. Karim R, Meyers C, Backendorf C, Ludigs K, Offringa R, van Ommen G-JB, et al. Human papillomavirus deregulates the response of a cellular network comprising of chemotactic and proinflammatory genes. *PloS One* (2011) 6(3):e17848. doi: 10.1371/ journal.pone.0017848

101. Tomaić V. Functional roles of E6 and E7 oncoproteins in hpv-induced malignancies at diverse anatomical sites. *Cancers (Basel)* (2016) 8(10). doi: 10.3390/ cancers8100095

102. Dickson EL, Vogel RI, Bliss RL, Downs LSJr. Multiple-type human papillomavirus (Hpv) infections: A cross-sectional analysis of the prevalence of specific types in 309,000 women referred for hpv testing at the time of cervical cytology. *Int J Gynecol Cancer* (2013) 23(7):1295–302. doi: 10.1097/IGC.0b013e31829e9fb4

103. Leiding JW, Holland SM. Warts and all: Human papillomavirus in primary immunodeficiencies. J Allergy Clin Immunol (2012) 130(5):1030-48. doi: 10.1016/j.jaci.2012.07.049

104. Béziat V. Human genetic dissection of papillomavirus-driven diseases: New insight into their pathogenesis. *Hum Genet* (2020) 139(6-7):919–39. doi: 10.1007/s00439-020-02183-x

105. Gupta R, Rady PL, Doan HQ, Tyring SK. Development of a B-hpv vaccine: Updates on an emerging frontier of skin cancer prevention. *J Clin Virol* (2020) 126:104348. doi: 10.1016/j.jcv.2020.104348

106. Ramoz N, Rueda LA, Bouadjar B, Montoya LS, Orth G, Favre M. Mutations in two adjacent novel genes are associated with epidermodysplasia verruciformis. *Nat Genet* (2002) 32(4):579–81. doi: 10.1038/ng1044

107. Lazarczyk M, Cassonnet P, Pons C, Jacob Y, Favre M. The ever proteins as a natural barrier against papillomaviruses: A new insight into the pathogenesis of human papillomavirus infections. *Microbiol Mol Biol Rev* (2009) 73(2):348–70. doi: 10.1128/MMBR.00033-08

108. Lazarczyk M, Pons C, Mendoza JA, Cassonnet P, Jacob Y, Favre M. Regulation of cellular zinc balance as a potential mechanism of ever-mediated protection against pathogenesis by cutaneous oncogenic human papillomaviruses. *J Exp Med* (2008) 205 (1):35–42. doi: 10.1084/jem.20071311

109. Patel AS, Karagas MR, Pawlita M, Waterboer T, Nelson HH. Cutaneous human papillomavirus infection, the Ever2 gene and incidence of squamous cell carcinoma: A case-control study. *Int J Cancer* (2008) 122(10):2377–9. doi: 10.1002/ijc.23377

110. Moore RJ, Owens DM, Stamp G, Arnott C, Burke F, East N, et al. Mice deficient in tumor necrosis factor-alpha are resistant to skin carcinogenesis. *Nat Med* (1999) 5 (7):828–31. doi: 10.1038/10552

111. Gaud G, Guillemot D, Jacob Y, Favre M, Vuillier F. Ever2 protein binds tradd to promote tnf-A-Induced apoptosis. *Cell Death Dis* (2013) 4(2):e499-e. doi: 10.1038/cddis.2013.27

112. Velazquez-Salinas I., Verdugo-Rodriguez A, Rodriguez LL, Borca MV. The role of interleukin 6 during viral infections. *Front Microbiol* (2019) 10:1057. doi: 10.3389/fmicb.2019.01057

113. Tanaka T, Narazaki M, Kishimoto T. Il-6 in inflammation, immunity, and disease. Cold Spring Harb Perspect Biol (2014) 6(10):a016295. doi: 10.1101/cshperspect.a016295

114. David JM, Dominguez C, Hamilton DH, Palena C. The il-8/Il-8r axis: A double agent in tumor immune resistance. Vaccines (Basel) (2016) 4(3). doi: 10.3390/vaccines4030022

115. Jiang WG, Sanders AJ, Ruge F, Harding KG. Influence of interleukin-8 (II-8) and il-8 receptors on the migration of human keratinocytes, the role of plc-T and potential clinical implications. *Exp Ther Med* (2012) 3(2):231–6. doi: 10.3892/etm.2011.402

116. McDermott DH, Murphy PM. Whim syndrome: Immunopathogenesis, treatment and cure strategies. Immunol Rev (2019) 287(1):91–102. doi: 10.1111/imr.12719

117. Kawai T, Malech HL. Whim syndrome: Congenital immune deficiency disease. Curr Opin Hematol (2009) 16(1):20-6. doi: 10.1097/MOH.0b013e32831ac557

118. Bianchi ME, Mezzapelle R. The chemokine receptor Cxcr4 in cell proliferation and tissue regeneration. *Front Immunol* (2020) 11:2109. doi: 10.3389/fimmu.2020.02109

119. Kucia M, Jankowski K, Reca R, Wysoczynski M, Bandura L, Allendorf DJ, et al. Cxcr4–Sdf-1 signalling, locomotion, chemotaxis and adhesion. *J Mol Histol* (2004) 35 (3):233–45. doi: 10.1023/b:hijo.0000032355.66152.b8

120. Vila-Coro AJ, Rodríguez-Frade JM, De Ana AM, Moreno-Ortíz MC, Martínez-AC, Mellado M. The chemokine sdf-l α triggers Cxcr4 receptor dimerization and activates the Jak/Stat pathway. *FASEB J* (1999) 13(13):1699–710. doi: 10.1096/fasebj.13.13.1699

121. Balabanian K, Lagane B, Pablos JL, Laurent L, Planchenault T, Verola O, et al. Whim syndromes with different genetic anomalies are accounted for by impaired Cxcr4 desensitization to Cxcl12. *Blood* (2005) 105(6):2449–57. doi: 10.1182/blood-2004-06-2289

122. Lagane B, Chow KY, Balabanian K, Levoye A, Harriague J, Planchenault T, et al. Cxcr4 dimerization and B-Arrestin-mediated signaling account for the enhanced chemotaxis to Cxcl12 in whim syndrome. *Blood J Am Soc Hematol* (2008) 112(1):34–44. doi: 10.1182/blood-2007-07-102103

123. Meuris F, Carthagena L, Jaracz-Ros A, Gaudin F, Cutolo P, Deback C, et al. The Cxcl12/Cxcr4 signaling pathway: A new susceptibility factor in human papillomavirus pathogenesis. *PloS Pathog* (2016) 12(12):e1006039. doi: 10.1371/journal.ppat.1006039

124. Zhao C, Lu X, Bu X, Zhang N, Wang W. Involvement of tumor necrosis factoralpha in the upregulation of Cxcr4 expression in gastric cancer induced by helicobacter pylori. *BMC Cancer* (2010) 10:419. doi: 10.1186/1471-2407-10-419

125. Moody C. Mechanisms by which hpv induces a replication competent environment in differentiating keratinocytes. Viruses (2017) 9(9). doi: 10.3390/v9090261

126. Zhang Q, Davis JC, Lamborn IT, Freeman AF, Jing H, Favreau AJ, et al. Combined immunodeficiency associated with Dock8 mutations. *New Engl J Med* (2009) 361(21):2046–55. doi: 10.1056/NEJMoa0905506

127. Chu EY, Freeman AF, Jing H, Cowen EW, Davis J, Su HC, et al. Cutaneous manifestations of Dock8 deficiency syndrome. *Arch Dermatol* (2012) 148(1):79–84. doi: 10.1001/archdermatol.2011.262

128. Zhang Q, Davis JC, Dove CG, Su HC. Genetic, clinical, and laboratory markers for Dock8 immunodeficiency syndrome. *Dis Markers* (2010) 29(3-4):131–9. doi: 10.3233/DMA-2010-0737

129. Singh AK, Eken A, Hagin D, Komal K, Bhise G, Shaji A, et al. Dock8 regulates fitness and function of regulatory T cells through modulation of il-2 signaling. *JCI Insight* (2017) 2(19). doi: 10.1172/jci.insight.94275

130. Randall KL, Law HD, Ziolkowski AF, Wirasinha RC, Goodnow CC, Daley SR. Dock8 deficiency diminishes thymic T-regulatory cell development but not thymic deletion. *Clin Trans Immunol* (2021) 10(1):e1236. doi: 10.1002/cti2.1236

131. Zhang Z, Bao Y, Zhou L, Ye Y, Fu W, Sun C. Dock8 serves as a prognostic biomarker and is related to immune infiltration in patients with hpv positive head and neck squamous cell carcinoma. *Cancer Control* (2021) 28:10732748211011951. doi: 10.1177/10732748211011951

132. Tirosh O, Conlan S, Deming C, Lee-Lin S-Q, Huang X, Barnabas BB, et al. Expanded skin virome in Dock8-deficient patients. *Nat Med* (2018) 24(12):1815–21. doi: 10.1038/s41591-018-0211-7

133. Lin JX, Leonard WJ. The common cytokine receptor Γ chain family of cytokines. Cold Spring Harb Perspect Biol (2018) 10(9). doi: 10.1101/cshperspect.a028449

134. Laffort C, Le Deist F, Favre M, Caillat-Zucman S, Radford-Weiss I, Debré M, et al. Severe cutaneous papillomavirus disease after haemopoietic stem-cell transplantation in patients with severe combined immune deficiency caused by common gammac cytokine receptor subunit or jak-3 deficiency. *Lancet* (2004) 363(9426):2051–4. doi: 10.1016/s0140-6736(04)16457-x

135. Nowak K, Linzner D, Thrasher AJ, Lambert PF, Di WL, Burns SO. Absence of Γ -chain in keratinocytes alters chemokine secretion, resulting in reduced immune cell recruitment. J Invest Dermatol (2017) 137(10):2120–30. doi: 10.1016/j.jid.2017.05.024

136. Mohamadzadeh M, Ariizumi K, Sugamura K, Bergstresser PR, Takashima A. Expression of the common cytokine receptor Γ chain by murine dendritic cells including epidermal langerhans cells. *Eur J Immunol* (1996) 26(1):156–60. doi: 10.1002/eji.1830260124

137. Kitashima DY, Kobayashi T, Woodring T, Idouchi K, Doebel T, Voisin B, et al. Langerhans cells prevent autoimmunity *Via* expansion of keratinocyte antigen-specific regulatory T cells. *EBioMedicine* (2018) 27:293–303. doi: 10.1016/j.ebiom.2017.12.022

138. Buckley RH. Transplantation of hematopoietic stem cells in human severe combined immunodeficiency: Longterm outcomes. *Immunol Res* (2011) 49(1-3):25-43. doi: 10.1007/s12026-010-8191-9

139. Bobby Gaspar H, Harwood C, Leigh I, Thrasher AJ. Severe cutaneous papillomavirus disease after haematopoietic stem-cell transplantation in patients with severe combined immunodeficiency. *Br J Haematol* (2004) 127(2):232–3. doi: 10.1111/j.1365-2141.2004.05176.x

140. Zampella J, Cohen B. Consideration of underlying immunodeficiency in refractory or recalcitrant warts: A review of the literature. *Skin Health Dis* (2022) 2(1): e98. doi: 10.1002/ski2.98

141. Spinner MA, Sanchez LA, Hsu AP, Shaw PA, Zerbe CS, Calvo KR, et al. Gata2 deficiency: A protean disorder of hematopoiesis, lymphatics, and immunity. *Blood* (2014) 123(6):809–21. doi: 10.1182/blood-2013-07-515528

142. West ES, Kingsbery MY, Mintz EM, Hsu AP, Holland SM, Rady PL, et al. Generalized verrucosis in a patient with Gata2 deficiency. *Br J Dermatol* (2014) 170 (5):1182–6. doi: 10.1111/bjd.12794

143. Ling K-W, Ottersbach K, Van Hamburg JP, Oziemlak A, Tsai F-Y, Orkin SH, et al. Gata-2 plays two functionally distinct roles during the ontogeny of hematopoietic stem cells. J Exp Med (2004) 200(7):871–82. doi: 10.1084/jem.20031556

144. Tsai F-Y, Orkin SH. Transcription factor gata-2 is required for Proliferation/ Survival of early hematopoietic cells and mast cell formation, but not for erythroid and myeloid terminal differentiation. *Blood J Am Soc Hematol* (1997) 89(10):3636–43. doi: 10.1182/blood.V89.10.3636

145. Parta M, Cole K, Avila D, Duncan L, Baird K, BB S, et al. Hematopoietic cell transplantation and outcomes related to human papillomavirus disease in Gata2 deficiency. *Transplant Cell Ther* (2021) 27(5):435.e1-.e11. doi: 10.1016/j.jtct.2020.12.028

146. Abdollahpour H, Appaswamy G, Kotlarz D, Diestelhorst J, Beier R, Schäffer AA, et al. The phenotype of human Stk4 deficiency. *Blood* (2012) 119(15):3450–7. doi: 10.1182/blood-2011-09-378158

147. Morgan EL, Patterson MR, Ryder EL, Lee SY, Wasson CW, Harper KL, et al. Microrna-18a targeting of the Stk4/Mst1 tumour suppressor is necessary for transformation in hpv positive cervical cancer. *PloS Pathog* (2020) 16(6):e1008624. doi: 10.1371/journal.ppat.1008624

148. Martelli-Marzagão F, Santos Junior G, Ogawa M, Enokihara M, Porro A, Tomimori J. Human papillomavirus detected in viral warts of renal transplant recipients. *Transplant Infect Dis* (2016) 18(1):37–43. doi: 10.1111/tid.12479

149. Shanis D, Anandi P, Grant C, Bachi A, Vyas N, Merideth MA, et al. Risks factors and timing of genital human papillomavirus (Hpv) infection in female stem cell transplant survivors: A longitudinal study. *Bone Marrow Transplant* (2018) 53(1):78–83. doi: 10.1038/bmt.2017.210

150. Grulich AE, Van Leeuwen MT, Falster MO, Vajdic CM. Incidence of cancers in people with Hiv/Aids compared with immunosuppressed transplant recipients: A metaanalysis. *Lancet* (2007) 370(9581):59–67. doi: 10.1016/s0140-6736(07)61050-2

151. Hinten F, Hilbrands LB, Meeuwis KA, IntHout J, Quint WG, Hoitsma AJ, et al. Reactivation of latent hpv infections after renal transplantation. *Am J Transplant* (2017) 17(6):1563–73. doi: 10.1111/ajt.14181

152. Meeuwis K, Hilbrands L, IntHout J, Slangen B, Hendriks I, Hinten F, et al. Cervicovaginal hpv infection in female renal transplant recipients: An observational, self-sampling based, cohort study. Am J Transplant (2015) 15(3):723–33. doi: 10.1111/ajt.13053

153. Meeuwis KA, Melchers WJ, Bouten H, van de Kerkhof PC, Hinten F, Quint WG, et al. Anogenital malignancies in women after renal transplantation over 40 years in a single center. *Transplantation* (2012) 93(9):914–22. doi: 10.1097/TP.0b013e318249b13d

154. Luque Y, Jamme M, Rabant M, DeWolf S, Noël LH, Thervet E, et al. Long-term Cd4 lymphopenia is associated with accelerated decline of kidney allograft function. *Nephrol Dial Transplant* (2016) 31(3):487–95. doi: 10.1093/ndt/gfv362

155. Ducloux D, Courivaud C, Bamoulid J, Vivet B, Chabroux A, Deschamps M, et al. Prolonged Cd4 T cell lymphopenia increases morbidity and mortality after renal transplantation. J Am Soc Nephrol (2010) 21(5):868–75. doi: 10.1681/asn.2009090976

156. Pfister H. Chapter 8: Human papillomavirus and skin cancer. J Natl Cancer Inst Monogr (2003) 31):52–6. doi: 10.1093/oxfordjournals.jncimonographs.a003483

157. Hartevelt MM, Bavinck JN, Kootte AM, Vermeer BJ, Vandenbroucke JP. Incidence of skin cancer after renal transplantation in the Netherlands. *Transplantation* (1990) 49(3):506–9. doi: 10.1097/00007890-199003000-00006

158. Larsen HK, Thomsen LT, Haedersdal M, Dehlendorff C, Schwartz Sørensen S, Kjaer SK. Risk of genital warts in renal transplant recipients–a registry-based, prospective cohort study. *Am J Transplant* (2019) 19(1):156–65. doi: 10.1111/ajt.15056

159. Cistjakovs M, Sultanova A, Jermakova O, Sokolovska L, Chapenko S, Lesina-Korne B, et al. Importance of high-risk human papillomavirus infection detection in female renal transplant recipients in the first year after transplantation. *Infect Dis Obstet Gynecol* (2018) 2018:9231031. doi: 10.1155/2018/9231031

160. Ogunbiyi O, Scholefield J, Raftery A, Smith J, Duffy S, Sharp F, et al. Prevalence of anal human papillomavirus infection and intraepithelial neoplasia in renal allograft recipients. J Br Surg (1994) 81(3):365–7. doi: 10.1002/bjs.1800810313

161. Palefsky JM, Holly EA, Ralston ML, Jay N, Berry JM, Darragh TM. High incidence of anal high-grade squamous intra-epithelial lesions among hiv-positive and hiv-negative homosexual and bisexual men. *Aids* (1998) 12(5):495–503. doi: 10.1097/00002030-199805000-00011

162. Patel HS, Silver AR, Northover JM. Anal cancer in renal transplant patients. Int J Colorectal Dis (2007) 22(1):1–5. doi: 10.1007/s00384-005-0023-3

163. Hojo M, Morimoto T, Maluccio M, Asano T, Morimoto K, Lagman M, et al. Cyclosporine induces cancer progression by a cell-autonomous mechanism. *Nature* (1999) 397(6719):530-4. doi: 10.1038/17401

164. Maluccio M, Sharma V, Lagman M, Vyas S, Yang H, Li B, et al. Tacrolimus enhances transforming growth factor-Beta1 expression and promotes tumor progression. *Transplantation* (2003) 76(3):597–602. doi: 10.1097/01.Tp.0000081399.75231.3b

165. Madeleine MM, Finch JL, Lynch CF, Goodman MT, Engels EA. Hpv-related cancers after solid organ transplantation in the united states. *Am J Transplant* (2013) 13 (12):3202–9. doi: 10.1111/ajt.12472

166. Bangash HK, Colegio OR. Management of non-melanoma skin cancer in immunocompromised solid organ transplant recipients. *Curr Treat Options Oncol* (2012) 13(3):354-76. doi: 10.1007/s11864-012-0195-3

167. Sindrewicz K, Kędzierska-Kapuza K, Jaworowska E, Ciechanowski K. Prevalence of human papillomavirus infection in the head and neck area of patients after kidney transplantation treated with immunosuppressive therapy. *Transplant Proc* (2020) 52 (8):2388–93. doi: 10.1016/j.transproceed.2020.01.112

168. Chaturvedi AK, Madeleine MM, Biggar RJ, Engels EA. Risk of human papillomavirus-associated cancers among persons with aids. *J Natl Cancer Inst* (2009) 101(16):1120–30. doi: 10.1093/jnci/djp205

169. Frisch M, Biggar RJ, Engels EA, Goedert JJ. Association of cancer with aidsrelated immunosuppression in adults. *Jama* (2001) 285(13):1736–45. doi: 10.1001/ jama.285.13.1736

170. Garland SM, Brotherton JML, Moscicki AB, Kaufmann AM, Stanley M, Bhatla N, et al. Hpv vaccination of immunocompromised hosts. *Papillomavirus Res* (2017) 4:35–8. doi: 10.1016/j.pvr.2017.06.002

171. Poynten IM, Jin F, Roberts JM, Templeton DJ, Law C, Cornall AM, et al. The natural history of anal high-grade squamous intraepithelial lesions in gay and bisexual men. *Clin Infect Dis* (2021) 72(5):853–61. doi: 10.1093/cid/ciaa166

172. Wright TCJr., Ellerbrock TV, Chiasson MA, Van Devanter N, Sun XW. Cervical intraepithelial neoplasia in women infected with human immunodeficiency virus: Prevalence, risk factors, and validity of papanicolaou smears. new York cervical disease study. *Obstet Gynecol* (1994) 84(4):591–7.

173. Ferenczy A, Coutlée F, Franco E, Hankins C. Human papillomavirus and hiv coinfection and the risk of neoplasias of the lower genital tract: A review of recent developments. *Cmaj* (2003) 169(5):431-4.

174. Duerr A, Kieke B, Warren D, Shah K, Burk R, Peipert JF, et al. Human papillomavirus-associated cervical cytologic abnormalities among women with or at risk of infection with human immunodeficiency virus. *Am J Obstet Gynecol* (2001) 184 (4):584–90. doi: 10.1067/mob.2001.111791

175. Wei F, Gaisa MM, D'Souza G, Xia N, Giuliano AR, Hawes SE, et al. Epidemiology of anal human papillomavirus infection and high-grade squamous intraepithelial lesions in 29 900 men according to hiv status, sexuality, and age: A collaborative pooled analysis of 64 studies. *Lancet HIV* (2021) 8(9):e531–43. doi: 10.1016/s2352-3018(21)00108-9

176. Cornall AM, Roberts JM, Molano M, Machalek DA, Phillips S, Hillman RJ, et al. Laser capture microdissection as a tool to evaluate human papillomavirus genotyping and methylation as biomarkers of persistence and progression of anal lesions. *BMJ Open* (2015) 5(8):e008439. doi: 10.1136/bmjopen-2015-008439

177. Vuyst HD, Ndirangu G, Moodley M, Tenet V, Estambale B, Meijer CJ, et al. Prevalence of human papillomavirus in women with invasive cervical carcinoma by hiv status in Kenya and south Africa. *Int J Cancer* (2012) 131(4):949–55. doi: 10.1002/ijc.26470

178. Denny LA, Franceschi S, de Sanjosé S, Heard I, Moscicki AB, Palefsky J. Human papillomavirus, human immunodeficiency virus and immunosuppression. *Vaccine* (2012) 30:F168–F74. doi: 10.1016/j.vaccine.2012.06.045

179. Lacey CJ. Therapy for genital human papillomavirus-related disease. *J Clin Virol* (2005) 32:82–90. doi: 10.1016/j.jcv.2004.10.020

180. Chaturvedi AK, Madeleine MM, Biggar RJ, Engels EA. Risk of human papillomavirus-associated cancers among persons with aids. *JNCI: J Natl Cancer Institute* (2009) 101(16):1120–30. doi: 10.1093/jnci/djp205

181. Mbulaiteye SM, Biggar RJ, Goedert JJ, Engels EA. Immune deficiency and risk for malignancy among persons with aids. *JAIDS J Acquired Immune Deficiency Syndromes* (2003) 32(5):527–33. doi: 10.1097/00126334-200304150-00010

182. Strickler HD, Burk RD, Fazzari M, Anastos K, Minkoff H, Massad LS, et al. Natural history and possible reactivation of human papillomavirus in human immunodeficiency virus-positive women. *JNCI: J Natl Cancer Institute* (2005) 97 (8):577–86. doi: 10.1093/jnci/dji073

183. Nazli A, Chan O, Dobson-Belaire WN, Ouellet M, Tremblay MJ, Gray-Owen SD, et al. Exposure to hiv-1 directly impairs mucosal epithelial barrier integrity allowing microbial translocation. *PloS Pathog* (2010) 6(4):e1000852. doi: 10.1371/journal.ppat.1000852

184. Perry I, Tselepis C, Hoyland J, Iqbal TH, Scott D, Sanders SA, et al. Reduced Cadherin/Catenin complex expression in celiac disease can be reproduced in vitro by cytokine stimulation. *Lab Invest* (1999) 79(12):1489–99.

185. Herfs M, Hubert P, Kholod N, Caberg JH, Gilles C, Berx G, et al. Transforming growth factor-B1-Mediated slug and snail transcription factor up-regulation reduces the density of langerhans cells in epithelial metaplasia by affecting e-cadherin expression. *Am J Pathol* (2008) 172(5):1391–402. doi: 10.2353/ajpath.2008.071004

186. Ye D, Ma I, Ma TY. Molecular mechanism of tumor necrosis factor-A modulation of intestinal epithelial tight junction barrier. *Am J Physiol Gastrointest Liver Physiol* (2006) 290(3):G496–504. doi: 10.1152/ajpgi.00318.2005

187. Klotman ME, Chang TL. Defensins in innate antiviral immunity. Nat Rev Immunol (2006) 6(6):447–56. doi: 10.1038/nri1860

188. Kodama J, Hashimoto I, Seki N, Hongo A, Yoshinouchi M, Okuda H, et al. Thrombospondin-1 and-2 messenger rna expression in invasive cervical cancer: Correlation with angiogenesis and prognosis. *Clin Cancer Res* (2001) 7(9):2826–31. doi: 10.3892/ijo.19.2.305

189. Hubert P, Herman L, Maillard C, Caberg JH, Nikkels A, Pierard G, et al. Defensins induce the recruitment of dendritic cells in cervical human papillomavirusassociated (Pre) neoplastic lesions formed in vitro and transplanted in vivo. *FASEB J* (2007) 21(11):2765–75. doi: 10.1096/fj.06-7646com

190. Yang D, Chertov O, Bykovskaia S, Chen Q, Buffo M, Shogan J, et al. B-defensins: Linking innate and adaptive immunity through dendritic and T cell Ccr6. *Science* (1999) 286(5439):525–8. doi: 10.1126/science.286.5439.525

191. Rice AP. The hiv-1 tat protein: Mechanism of action and target for hiv-1 cure strategies. Curr Pharm Des (2017) 23(28):4098–102. doi: 10.2174/1381612823666170704130635

192. Das AT, Harwig A, Berkhout B. The hiv-1 tat protein has a versatile role in activating viral transcription. J Virol (2011) 85(18):9506–16. doi: 10.1128/JVI.00650-11

193. Tugizov SM, Herrera R, Chin-Hong P, Veluppillai P, Greenspan D, Berry JM, et al. Hiv-associated disruption of mucosal epithelium facilitates paracellular penetration by human papillomavirus. *Virology* (2013) 446(1-2):378–88. doi: 10.1016/j.virol.2013.08.018

194. Tornesello ML, Buonaguro FM, Beth-Giraldo E, Giraldo G. Human immunodeficiency virus type 1 tat gene enhances human papillomavirus early gene expression. *Intervirology* (1993) 36(2):57–64. doi: 10.1159/000150322

195. Barillari G, Palladino C, Bacigalupo I, Leone P, Falchi M, Ensoli B. Entrance of the tat protein of hiv-1 into human uterine cervical carcinoma cells causes upregulation of hpv-E6 expression and a decrease in P53 protein levels. *Oncol Lett* (2016) 12(4):2389–94. doi: 10.3892/ol.2016.4921

196. Kim RH, Yochim JM, Kang MK, Shin KH, Christensen R, Park NH. Hiv-1 tat enhances replicative potential of human oral keratinocytes harboring hpv-16 genome. *Int J Oncol* (2008) 33(4):777–82. doi: 10.3892/ijo_00000064

197. Nyagol J, Leucci E, Omnis A, De Falco G, Tigli C, Sanseverino F, et al. The effects of hiv-1 tat protein on cell cycle during cervical carcinogenesis. *Cancer Biol Ther* (2006) 5 (6):684–90. doi: 10.4161/cbt.5.6.2907

198. Rayne F, Debaisieux S, Bonhoure A, Beaumelle B. Hiv-1 tat is unconventionally secreted through the plasma membrane. *Cell Biol Int* (2010) 34(4):409–13. doi: 10.1042/ cbi20090376

199. Silverberg MJ, Neuhaus J, Bower M, Gey D, Hatzakis A, Henry K, et al. Risk of cancers during interrupted antiretroviral therapy in the smart study. *Aids* (2007) 21 (14):1957–63. doi: 10.1097/QAD.0b013e3282ed6338

200. Biggar RJ, Chaturvedi AK, Goedert JJ, Engels EA. Aids-related cancer and severity of immunosuppression in persons with aids. *J Natl Cancer Institute* (2007) 99(12):962–72. doi: 10.1093/jnci/djm010

201. Minkoff H, Zhong Y, Burk RD, Palefsky JM, Xue X, Watts DH, et al. Influence of adherent and effective antiretroviral therapy use on human papillomavirus infection and squamous intraepithelial lesions in human immunodeficiency virus-positive women. J Infect Dis (2010) 201(5):681–90. doi: 10.1086/650467

202. Beachler DC, D'Souza G. Oral hpv infection and head and neck cancers in hiv-infected individuals. *Curr Opin Oncol* (2013) 25(5):503. doi: 10.1097/CCO.0b013e32836242b4

203. Kaufmann GR, Perrin I, Pantaleo G, Opravil M, Furrer H, Telenti A, et al. Cd4 Tlymphocyte recovery in individuals with advanced hiv-1 infection receiving potent antiretroviral therapy for 4 years: The Swiss hiv cohort study. *Arch Internal Med* (2003) 163(18):2187–95. doi: 10.1001/archinte.163.18.2187

204. Palefsky J. Biology of hpv in hiv infection. Adv Dental Res (2006) 19(1):99–105. doi: 10.1177/154407370601900120

205. King MD, Reznik DA, O'Daniels CM, Larsen NM, Osterholt D, Blumberg HM. Human papillomavirus-associated oral warts among human immunodeficiency virusseropositive patients in the era of highly active antiretroviral therapy: An emerging infection. *Clin Infect Dis* (2002) 34(5):641–8. doi: 10.1086/338637

206. Meys R, Gotch F, Bunker C. Human papillomavirus in the era of highly active antiretroviral therapy for human immunodeficiency virus: An immune reconstitution-associated disease? *Br J Dermatol* (2010) 162(1):6–11. doi: 10.1111/j.1365-2133.2009.09365.x

207. Lissouba P, Van de Perre P, Auvert B. Association of genital human papillomavirus infection with hiv acquisition: A systematic review and meta-analysis. *Sexually Transmitted Infect* (2013) 89(5):350-6. doi: 10.1136/sextrans-2011-050346

208. Houlihan CF, Larke NL, Watson-Jones D, Smith-McCune KK, Shiboski S, Gravitt PE, et al. Hpv infection and increased risk of hiv acquisition. a systematic review and meta-analysis. *AIDS (Lond Engl)* (2012) 26(17). doi: 10.1097/QAD.0b013e328358d908

209. Averbach SH, Gravitt PE, Nowak RG, Celentano DD, Dunbar MS, Morrison CS, et al. The association between cervical hpv infection and hiv acquisition among women in Zimbabwe. *AIDS (Lond Engl)* (2010) 24(7):1035. doi: 10.1097/qad.0b013e3283377973

210. Smith JS, Moses S, Hudgens MG, Parker CB, Agot K, Maclean I, et al. Increased risk of hiv acquisition among Kenyan men with human papillomavirus infection. *J Infect Dis* (2010) 201(11):1677–85. doi: 10.1086/652408

211. Chin-Hong PV, Husnik M, Cranston RD, Colfax G, Buchbinder S, Da Costa M, et al. Anal human papillomavirus infection is associated with hiv acquisition in men who have sex with men. *AIDS (London England)* (2009) 23(9):1135. doi: 10.1097/QAD.0b013e32832b4449

212. McKinnon LR, Nyanga B, Chege D, Izulla P, Kimani M, Huibner S, et al. Characterization of a human cervical Cd4+ T cell subset coexpressing multiple markers of hiv susceptibility. *J Immunol* (2011) 187(11):6032–42. doi: 10.4049/jimmunol.1101836

213. Arnold KB, Burgener A, Birse K, Romas L, Dunphy LJ, Shahabi K, et al. Increased levels of inflammatory cytokines in the female reproductive tract are associated with altered expression of proteases, mucosal barrier proteins, and an influx of hiv-susceptible target cells. *Mucosal Immunol* (2016) 9(1):194–205. doi: 10.1038/mi.2015.51

214. Masson L, Passmore J-AS, Liebenberg LJ, Werner L, Baxter C, Arnold KB, et al. Genital inflammation and the risk of hiv acquisition in women. *Clin Infect Dis* (2015) 61 (2):260–9. doi: 10.1093/cid/civ298

215. Liebenberg LJ, McKinnon LR, Yende-Zuma N, Garrett N, Baxter C, Kharsany A, et al. Hpv infection and the genital cytokine milieu in women at high risk of hiv acquisition. *Nat Commun* (2019) 10(1):1–12. doi: 10.1038/s41467-019-13089-2

216. Burger JA, Kipps TJ. Cxcr4: A key receptor in the crosstalk between tumor cells and their microenvironment. *Blood* (2006) 107(5):1761–7. doi: 10.1182/blood-2005-08-3182

217. Ansari AW, Heiken H, Moenkemeyer M, Schmidt RE. Dichotomous effects of c-c chemokines in hiv-1 pathogenesis. *Immunol Lett* (2007) 110(1):1–5. doi: 10.1016/j.imlet.2007.02.012

218. Koh WH, Lopez P, Ajibola O, Parvarchian R, Mohammad U, Hnatiuk R, et al. Hiv-captured dcs regulate T cell migration and cell-cell contact dynamics to enhance viral spread. *Iscience* (2020) 23(8):101427. doi: 10.1016/j.isci.2020.101427

219. Abreu AL, Souza RP, Gimenes F, Consolaro ME. A review of methods for detect human papillomavirus infection. Virol J (2012) 9:262. doi: 10.1186/1743-422x-9-262

220. Williams J, Kostiuk M, Biron VL. Molecular detection methods in hpv-related cancers. Front Oncol (2022) 12:864820. doi: 10.3389/fonc.2022.864820

221. Wiest T, Schwarz E, Enders C, Flechtenmacher C, Bosch FX. Involvement of intact Hpv16 E6/E7 gene expression in head and neck cancers with unaltered P53 status and perturbed prb cell cycle control. *Oncogene* (2002) 21(10):1510–7. doi: 10.1038/sj.onc.1205214

222. Worsham MJ, Chen KM, Tiwari N, Pals G, Schouten JP, Sethi S, et al. Finemapping loss of gene architecture at the Cdkn2b (P15ink4b), Cdkn2a (P14arf, P16ink4a), and mtap genes in head and neck squamous cell carcinoma. *Arch Otolaryngol Head Neck Surg* (2006) 132(4):409–15. doi: 10.1001/archotol.132.4.409

223. Shi W, Kato H, Perez-Ordonez B, Pintilie M, Huang S, Hui A, et al. Comparative prognostic value of Hpv16 E6 mrna compared with in situ hybridization for human oropharyngeal squamous carcinoma. *J Clin Oncol* (2009) 27(36):6213–21. doi: 10.1200/ jcc.2009.23.1670

224. World Health Organization. *Cervical cancer*. Geneva, Switzerland: World Health Organization (2022). Available at: https://www.who.int/news-room/fact-sheets/detail/cervical-cancer.

225. Stelzle D, Tanaka LF, Lee KK, Ibrahim Khalil A, Baussano I, Shah ASV, et al. Estimates of the global burden of cervical cancer associated with hiv. *Lancet Glob Health* (2021) 9(2):e161–e9. doi: 10.1016/s2214-109x(20)30459-9

226. De Vuyst H, Lillo F, Broutet N, Smith JS. Hiv, human papillomavirus, and cervical neoplasia and cancer in the era of highly active antiretroviral therapy. *Eur J Cancer Prev* (2008) 17(6):545–54. doi: 10.1097/CEJ.0b013e3282f75ea1

227. Kdigo. Clinical practice guideline for the care of kidney transplant recipients. Am J Transplant (2009) 9 Suppl 3:S1–155. doi: 10.1111/j.1600-6143.2009.02834.x

228. De Vuyst H, Clifford GM, Nascimento MC, Madeleine MM, Franceschi S. Prevalence and type distribution of human papillomavirus in carcinoma and intraepithelial neoplasia of the vulva, vagina and anus: A meta-analysis. *Int J Cancer* (2009) 124(7):1626–36. doi: 10.1002/ijc.24116

229. Alemany L, Saunier M, Alvarado-Cabrero I, Quirós B, Salmeron J, Shin HR, et al. Human papillomavirus DNA prevalence and type distribution in anal carcinomas worldwide. *Int J Cancer* (2015) 136(1):98–107. doi: 10.1002/ijc.28963

230. Elorza G, Saralegui Y, Enríquez-Navascués JM, Placer C, Velaz L. Anal intraepitelial neoplasia: A narrative review. *Rev Esp Enferm Dig* (2016) 108(1):31–9.

231. Stanley MA, Winder DM, Sterling JC, Goon PK. Hpv infection, anal intraepithelial neoplasia (Ain) and anal cancer: Current issues. *BMC Cancer* (2012) 12:398. doi: 10.1186/1471-2407-12-398

232. Machalek DA, Poynten M, Jin F, Fairley CK, Farnsworth A, Garland SM, et al. Anal human papillomavirus infection and associated neoplastic lesions in men who have sex with men: A systematic review and meta-analysis. *Lancet Oncol* (2012) 13(5):487–500. doi: 10.1016/s1470-2045(12)70080-3

233. Johnson LG, Madeleine MM, Newcomer LM, Schwartz SM, Daling JR. Anal cancer incidence and survival: The surveillance, epidemiology, and end results experience, 1973-2000. *Cancer* (2004) 101(2):281–8. doi: 10.1002/cncr.20364

234. van Leeuwen MT, Vajdic CM, Middleton MG, McDonald AM, Law M, Kaldor JM, et al. Continuing declines in some but not all hiv-associated cancers in Australia after widespread use of antiretroviral therapy. *Aids* (2009) 23(16):2183–90. doi: 10.1097/QAD.0b013e328331d384

235. Daling JR, Weiss NS, Hislop TG, Maden C, Coates RJ, Sherman KJ, et al. Sexual practices, sexually transmitted diseases, and the incidence of anal cancer. *N Engl J Med* (1987) 317(16):973–7. doi: 10.1056/nejm198710153171601

236. Adami J, Gäbel H, Lindelöf B, Ekström K, Rydh B, Glimelius B, et al. Cancer risk following organ transplantation: A nationwide cohort study in Sweden. *Br J Cancer* (2003) 89(7):1221–7. doi: 10.1038/sj.bjc.6601219

237. Berry JM, Palefsky JM, Jay N, Cheng SC, Darragh TM, Chin-Hong PV. Performance characteristics of anal cytology and human papillomavirus testing in patients with high-resolution anoscopy-guided biopsy of high-grade anal intraepithelial neoplasia. *Dis Colon Rectum* (2009) 52(2):239–47. doi: 10.1007/DCR.0b013e31819793d9

238. Farshadpour F, Konings S, Speel EJ, Hordijk GJ, Koole R, van Blokland M, et al. Human papillomavirus and oropharyngeal squamous cell carcinoma: A case-control study regarding tobacco and alcohol consumption. *Patholog Res Int* (2011) 2011:806345. doi: 10.4061/2011/806345

239. Chaturvedi AK, Zumsteg ZS. A snapshot of the evolving epidemiology of oropharynx cancers. *Cancer* (2018) 124(14):2893-6. doi: 10.1002/cncr.31383

240. Hong AM, Grulich AE, Jones D, Lee CS, Garland SM, Dobbins TA, et al. Squamous cell carcinoma of the oropharynx in Australian males induced by human papillomavirus vaccine targets. *Vaccine* (2010) 28(19):3269–72. doi: 10.1016/j.vaccine.2010.02.098

241. Lechner M, Liu J, Masterson L, Fenton TR. Hpv-associated oropharyngeal cancer: Epidemiology, molecular biology and clinical management. *Nat Rev Clin Oncol* (2022) 19 (5):306–27. doi: 10.1038/s41571-022-00603-7

242. Beachler DC, D'Souza G, Sugar EA, Xiao W, Gillison ML. Natural history of anal vs oral hpv infection in hiv-infected men and women. J Infect Dis (2013) 208(2):330–9. doi: 10.1093/infdis/jit170

243. Hewavisenti R, Ferguson A, Wang K, Jones D, Gebhardt T, Edwards J, et al. Cd103+ tumor-resident Cd8+ T cell numbers underlie improved patient survival in oropharyngeal squamous cell carcinoma. *J Immunother Cancer* (2020) 8(1). doi: 10.1136/jitc-2019-000452

244. Kutlubay Z, Engin B, Zara T, Tüzün Y. Anogenital malignancies and premalignancies: Facts and controversies. *Clin Dermatol* (2013) 31(4):362-73. doi: 10.1016/j.clindermatol.2013.01.003

245. Casolati E, Agarossi A, Valieri M, Ferrazzi E. Vulvar neoplasia in hiv positive women: A review. *Med Wieku Rozwoj* (2003) 7(4 Pt 1):487–93.

246. Frisch M, Biggar RJ, Goedert JJ. Human papillomavirus-associated cancers in patients with human immunodeficiency virus infection and acquired immunodeficiency syndrome. *J Natl Cancer institute* (2000) 92(18):1500–10. doi: 10.1093/jnci/92.18.1500

247. Abercrombie PD, Korn AP. Vulvar intraepithelial neoplasia in women with hiv. AIDS Patient Care STDS (1998) 12(4):251-4. doi: 10.1089/apc.1998.12.251

248. Korn AP, Abercrombie PD, Foster A. Vulvar intraepithelial neoplasia in women infected with human immunodeficiency virus-1. *Gynecol Oncol* (1996) 61(3):384–6. doi: 10.1006/gyno.1996.0161

249. Wang H, Zhao Q, Zhang Y, Zhang Q, Zheng Z, Liu S, et al. Immunotherapy advances in locally advanced and Recurrent/Metastatic head and neck squamous cell carcinoma and its relationship with human papillomavirus. *Front Immunol* (2021) 12:652054. doi: 10.3389/fimmu.2021.652054

250. Chang J-H, Wu C-C, Yuan KS-P, Wu AT, Wu S-Y. Locoregionally recurrent head and neck squamous cell carcinoma: Incidence, survival, prognostic factors, and treatment outcomes. *Oncotarget* (2017) 8(33):55600. doi: 10.18632/oncotarget.16340

251. Adelstein DJ, Ridge JA, Brizel DM, Holsinger FC, Haughey BH, O'Sullivan B, et al. Transoral resection of pharyngeal cancer: Summary of a national cancer institute head and neck cancer steering committee clinical trials planning meeting, November 6-7, 2011, Arlington, Virginia. *Head Neck* (2012) 34(12):1681–703. doi: 10.1002/hed.23136

252. Adelstein DJ, Li Y, Adams GL, Wagner HJr., Kish JA, Ensley JF, et al. An intergroup phase iii comparison of standard radiation therapy and two schedules of concurrent chemoradiotherapy in patients with unresectable squamous cell head and neck cancer. *J Clin Oncol* (2003) 21(1):92–8. doi: 10.1200/jco.2003.01.008

253. Kobayashi K, Hisamatsu K, Suzui N, Hara A, Tomita H, Miyazaki T. A review of hpv-related head and neck cancer. J Clin Med (2018) 7(9). doi: 10.3390/jcm7090241

254. Ferris RL, Blumenschein GJr., Fayette J, Guigay J, Colevas AD, Licitra L, et al. Nivolumab for recurrent squamous-cell carcinoma of the head and neck. *N Engl J Med* (2016) 375(19):1856–67. doi: 10.1056/NEJMoa1602252

255. Seiwert TY, Burtness B, Mehra R, Weiss J, Berger R, Eder JP, et al. Safety and clinical activity of pembrolizumab for treatment of recurrent or metastatic squamous cell carcinoma of the head and neck (Keynote-012): An open-label, multicentre, phase 1b trial. *Lancet Oncol* (2016) 17(7):956–65. doi: 10.1016/S1470-2045(16)30066-3

256. Burtness B, Harrington KJ, Greil R, Soulières D, Tahara M, de Castro G, et al. Pembrolizumab alone or with chemotherapy versus cetuximab with chemotherapy for recurrent or metastatic squamous cell carcinoma of the head and neck (Keynote-048): A randomised, open-label, phase 3 study. *Lancet* (2019) 394(10212):1915–28. doi: 10.1016/S0140-6736(19)32591-7

257. Ros W, Delord J, Perets R, Italiano A, Shapira-Frommer R, Manzuk L. Efficacy and safety of pembrolizumab in previously treated advanced cervical cancer: Results from the phase 2 keynote-158 study. *Early Phase Clin Stud Novel Immunother Oncol* (2019) 37:117. doi: 10.1200/JCO.18.01265

258. Colombo N, Dubot C, Lorusso D, Caceres MV, Hasegawa K, Shapira-Frommer R, et al. Pembrolizumab for persistent, recurrent, or metastatic cervical cancer. *N Engl J Med* (2021) 385(20):1856–67. doi: 10.1056/NEJMoa2112435

259. Marabelle A, Cassier PA, Fakih M, Kao S, Nielsen D, Italiano A, et al. Pembrolizumab for previously treated advanced anal squamous cell carcinoma: Results from the non-randomised, multicohort, multicentre, phase 2 keynote-158 study. *Lancet Gastroenterol Hepatol* (2022) 7(5):446–54. doi: 10.1016/s2468-1253(21)00382-4

260. Ott PA, Piha-Paul SA, Munster P, Pishvaian MJ, van Brummelen EMJ, Cohen RB, et al. Safety and antitumor activity of the anti-Pd-1 antibody pembrolizumab in patients with recurrent carcinoma of the anal canal. *Ann Oncol* (2017) 28(5):1036–41. doi: 10.1093/annonc/mdx029

261. Allouch S, Malki A, Allouch A, Gupta I, Vranic S, Al Moustafa AE. High-risk hpv oncoproteins and pd-1/Pd-L1 interplay in human cervical cancer: Recent evidence and future directions. *Front Oncol* (2020) 10:914. doi: 10.3389/fonc.2020.00914

262. Rubinstein PG, Aboulafia DM, Zloza A. Malignancies in Hiv/Aids: From epidemiology to the rapeutic challenges. Aids (2014) 28(4):453–65. doi: 10.1097/ qad.0000000000001

263. Deeken JF, Pantanowitz L, Dezube BJ. Targeted therapies to treat non-Aids-Defining cancers in patients with hiv on haart therapy: Treatment considerations and research outlook. *Curr Opin Oncol* (2009) 21(5):445–54. doi: 10.1097/CCO.0b013e32832f3e04

264. Joura EA, Ulied A, Vandermeulen C, Rua Figueroa M, Seppä I, Hernandez Aguado JJ, et al. Immunogenicity and safety of a nine-valent human papillomavirus vaccine in women 27-45 Years of age compared to women 16-26 Years of age: An open-label phase 3 study. *Vaccine* (2021) 39(20):2800–9. doi: 10.1016/j.vaccine. 2021.01.074

265. Gillison ML, Chaturvedi AK, Lowy DR. Hpv prophylactic vaccines and the potential prevention of noncervical cancers in both men and women. *Cancer* (2008) 113 (10 Suppl):3036–46. doi: 10.1002/cncr.23764

266. Markowitz LE, Dunne EF, Saraiya M, Lawson HW, Chesson H, Unger ER. Quadrivalent human papillomavirus vaccine: Recommendations of the advisory committee on immunization practices (Acip). *MMWR Recomm Rep* (2007) 56(Rr-2):1–24.

267. World Health Organization. Meeting of the strategic advisory group of experts on immunization, April 2014–conclusions and recommendations. *Weekly Epidemiol Rec* (2014) 89(21):221–36.

268. Bruni L, Saura-Lázaro A, Montoliu A, Brotons M, Alemany L, Diallo MS, et al. Hpv vaccination introduction worldwide and who and unicef estimates of national hpv immunization coverage 2010–2019. *Prev Med* (2021) 144:106399. doi: 10.1016/j.ypmed.2020.106399

269. Lin K, Doolan K, Hung C-F, Wu TC. Perspectives for preventive and therapeutic hpv vaccines. J Formos Med Assoc (2010) 109(1):4–24. doi: 10.1016/S0929-6646(10)60017-4

270. Ault KA. Effect of prophylactic human papillomavirus L1 virus-Like-Particle vaccine on risk of cervical intraepithelial neoplasia grade 2, grade 3, and adenocarcinoma in situ: A combined analysis of four randomised clinical trials. *Lancet* (2007) 369 (9576):1861–8. doi: 10.1016/s0140-6736(07)60852-6

271. Skeate JG, Woodham AW, Einstein MH, Da Silva DM, Kast WM. Current therapeutic vaccination and immunotherapy strategies for hpv-related diseases. *Hum Vaccin Immunother* (2016) 12(6):1418–29. doi: 10.1080/21645515.2015.1136039

272. Doran SL, Stevanović S, Adhikary S, Gartner JJ, Jia L, Kwong MLM, et al. T-Cell receptor gene therapy for human papillomavirus–associated epithelial cancers: A first-in-Human, phase I/Ii study. J Clin Oncol (2019) 37(30):2759–68. doi: 10.1200/jco.18.02424

273. Wang X, Sandberg ML, Martin AD, Negri KR, Gabrelow GB, Nampe DP, et al. Potent, selective cars as potential T-cell therapeutics for hpv-positive cancers. *J Immunother* (2021) 44(8):292–306. doi: 10.1097/cji.00000000000386

274. Houot R, Schultz LM, Marabelle A, Kohrt H. T-Cell-Based immunotherapy: Adoptive cell transfer and checkpoint inhibition. *Cancer Immunol Res* (2015) 3(10):1115–22. doi: 10.1158/2326-6066.Cir-15-0190

275. Messa L, Loregian A. Hpv-induced cancers: Preclinical therapeutic advancements. Expert Opin Investig Drugs (2022) 31(1):79–93. doi: 10.1080/13543784.2021.2010703

276. Baden LR, El Sahly HM, Essink B, Kotloff K, Frey S, Novak R, et al. Efficacy and safety of the mrna-1273 sars-Cov-2 vaccine. *N Engl J Med* (2021) 384(5):403–16. doi: 10.1056/NEJMoa2035389

277. Polack FP, Thomas SJ, Kitchin N, Absalon J, Gurtman A, Lockhart S, et al. Safety and efficacy of the Bnt162b2 mrna covid-19 vaccine. *N Engl J Med* (2020) 383(27):2603–15. doi: 10.1056/NEJMoa2034577

278. Pardi N, Hogan MJ, Porter FW, Weissman D. Mrna vaccines - a new era in vaccinology. Nat Rev Drug Discovery (2018) 17(4):261–79. doi: 10.1038/nrd.2017.243

279. Linares-Fernández S, Lacroix C, Exposito JY, Verrier B. Tailoring mrna vaccine to balance Innate/Adaptive immune response. *Trends Mol Med* (2020) 26(3):311–23. doi: 10.1016/j.molmed.2019.10.002

280. Rosa SS, Prazeres DMF, Azevedo AM, Marques MPC. Mrna vaccines manufacturing: Challenges and bottlenecks. *Vaccine* (2021) 39(16):2190-200. doi: 10.1016/j.vaccine.2021.03.038

281. Paschon DE, Lussier S, Wangzor T, Xia DF, Li PW, Hinkley SJ, et al. Diversifying the structure of zinc finger nucleases for high-precision genome editing. *Nat Commun* (2019) 10(1):1133. doi: 10.1038/s41467-019-08867-x

282. Ding W, Hu Z, Zhu D, Jiang X, Yu L, Wang X, et al. Zinc finger nucleases targeting the human papillomavirus E7 oncogene induce E7 disruption and a transformed phenotype in Hpv16/18-positive cervical cancer cells. *Clin Cancer Res* (2014) 20 (24):6495–503. doi: 10.1158/1078-0432.Ccr-14-0250

283. Perez EE, Wang J, Miller JC, Jouvenot Y, Kim KA, Liu O, et al. Establishment of hiv-1 resistance in Cd4+ T cells by genome editing using zinc-finger nucleases. *Nat Biotechnol* (2008) 26(7):808–16. doi: 10.1038/nbt1410

284. Gupta RM, Musunuru K. Expanding the genetic editing tool kit: Zfns, talens, and crispr-Cas9. J Clin Invest (2014) 124(10):4154–61. doi: 10.1172/jci72992

285. Mushtaq M, Bhat JA, Mir ZA, Sakina A, Ali S, Singh AK, et al. Crispr/Cas approach: A new way of looking at plant-abiotic interactions. *J Plant Physiol* (2018) 224-225:156–62. doi: 10.1016/j.jplph.2018.04.001

286. Mojica FJ, Juez G, Rodríguez-Valera F. Transcription at different salinities of haloferax mediterranei sequences adjacent to partially modified psti sites. *Mol Microbiol* (1993) 9(3):613–21. doi: 10.1111/j.1365-2958.1993.tb01721.x

287. Makarova KS, Koonin EV. Annotation and classification of crispr-cas systems. Methods Mol Biol (2015) 1311:47–75. doi: 10.1007/978-1-4939-2687-9_4

288. Gleditzsch D, Pausch P, Müller-Esparza H, Özcan A, Guo X, Bange G, et al. Pam Identification by crispr-cas effector complexes: Diversified mechanisms and structures. *RNA Biol* (2019) 16(4):504–17. doi: 10.1080/15476286.2018.1504546

289. Nakamura M, Gao Y, Dominguez AA, Qi LS. Crispr technologies for precise epigenome editing. Nat Cell Biol (2021) 23(1):11–22. doi: 10.1038/s41556-020-00620-7

290. Nambiar TS, Billon P, Diedenhofen G, Hayward SB, Taglialatela A, Cai K, et al. Stimulation of crispr-mediated homology-directed repair by an engineered Rad18 variant. *Nat Commun* (2019) 10(1):3395. doi: 10.1038/s41467-019-11105-z

291. Lao YH, Li M, Gao MA, Shao D, Chi CW, Huang D, et al. Hpv oncogene manipulation using nonvirally delivered Crispr/Cas9 or natronobacterium gregoryi argonaute. Adv Sci (Weinh) (2018) 5(7):1700540. doi: 10.1002/advs.201700540

292. Zhen S, Hua L, Takahashi Y, Narita S, Liu YH, Li Y. *In vitro* and in vivo growth suppression of human papillomavirus 16-positive cervical cancer cells by Crispr/Cas9. *Biochem Biophys Res Commun* (2014) 450(4):1422–6. doi: 10.1016/j.bbrc.2014.07.014

293. Pucci B, Kasten M, Giordano A. Cell cycle and apoptosis. *Neoplasia* (2000) 2 (4):291-9. doi: 10.1038/sj.neo.7900101

294. Ahlenstiel CL, Symonds G, Kent SJ, Kelleher AD. Block and lock hiv cure strategies to control the latent reservoir. *Front Cell Infect Microbiol* (2020) 10:424. doi: 10.3389/fcimb.2020.00424

295. Ahlenstiel C, Mendez C, Lim ST, Marks K, Turville S, Cooper DA, et al. Novel rna duplex locks hiv-1 in a latent state *Via* chromatin-mediated transcriptional silencing. *Mol Ther Nucleic Acids* (2015) 4(10):e261. doi: 10.1038/mtna.2015.31

296. Jiang M, Milner J. Selective silencing of viral gene expression in hpv-positive human cervical carcinoma cells treated with sirna, a primer of rna interference. *Oncogene* (2002) 21(39):6041-8. doi: 10.1038/sj.onc.1205878

297. Crooke ST, Witztum JL, Bennett CF, Baker BF. Rna-targeted therapeutics. *Cell Metab* (2018) 27(4):714–39. doi: 10.1016/j.cmet.2018.03.004

298. Butz K, Ristriani T, Hengstermann A, Denk C, Scheffner M, Hoppe-Seyler F. Sirna targeting of the viral E6 oncogene efficiently kills human papillomavirus-positive cancer cells. *Oncogene* (2003) 22(38):5938–45. doi: 10.1038/sj.onc.1206894

299. Jonson AL, Rogers LM, Ramakrishnan S, Downs LSJr. Gene silencing with sirna targeting E6/E7 as a therapeutic intervention in a mouse model of cervical cancer. *Gynecol Oncol* (2008) 111(2):356–64. doi: 10.1016/j.ygyno.2008.06.033

300. Tan TM, Ting RC. *In vitro* and in vivo inhibition of human papillomavirus type 16 E6 and E7 genes. *Cancer Res* (1995) 55(20):4599–605.

301. Hamada K, Sakaue M, Alemany R, Zhang WW, Horio Y, Roth JA, et al. Adenovirus-mediated transfer of hpv 16 E6/E7 antisense rna to human cervical cancer cells. *Gynecol Oncol* (1996) 63(2):219–27. doi: 10.1006/gyno.1996.0310

302. Alvarez-Salas LM, Arpawong TE, DiPaolo JA. Growth inhibition of cervical tumor cells by antisense oligodeoxynucleotides directed to the human papillomavirus type 16 E6 gene. *Antisense Nucleic Acid Drug Dev* (1999) 9(5):441–50. doi: 10.1089/oli.1.1999.9.441

303. Choo CK, Ling MT, Suen CK, Chan KW, Kwong YL. Retrovirus-mediated delivery of Hpv16 E7 antisense rna inhibited tumorigenicity of caski cells. *Gynecol Oncol* (2000) 78(3 Pt 1):293–301. doi: 10.1006/gyno.2000.5916

304. Weinberg MS, Morris KV. Transcriptional gene silencing in humans. Nucleic Acids Res (2016) 44(14):6505–17. doi: 10.1093/nar/gkw139

305. Graham SV. Human papillomavirus: Gene expression, regulation and prospects for novel diagnostic methods and antiviral therapies. *Future Microbiol* (2010) 5(10):1493–506. doi: 10.2217/fmb.10.107

306. Palanichamy JK, Mehndiratta M, Bhagat M, Ramalingam P, Das B, Das P, et al. Silencing of integrated human papillomavirus-16 oncogenes by small interfering rnamediated heterochromatization. *Mol Cancer Ther* (2010) 9(7):2114–22. doi: 10.1158/ 1535-7163.Mct-09-0977

307. Khan I, Zakaria MK, Kumar M, Mani P, Chattopadhyay P, Sarkar DP, et al. A novel placental like alkaline phosphatase promoter driven transcriptional silencing combined with single chain variable fragment antibody based virosomal delivery for neoplastic cell targeting [Corrected]. *J Transl Med* (2015) 13:254. doi: 10.1186/s12967-015-0602-1

308. Haussecker D. The business of rnai therapeutics in 2012. *Mol Ther Nucleic Acids* (2012) 1(2):e8. doi: 10.1038/mtna.2011.9

309. Chen X, Mangala LS, Rodriguez-Aguayo C, Kong X, Lopez-Berestein G, Sood AK. Rna interference-based therapy and its delivery systems. *Cancer Metastasis Rev* (2018) 37 (1):107–24. doi: 10.1007/s10555-017-9717-6

310. Massachi S, Epstein J, Hurd J, Bonkovsky HL. Cost savings with hemin versus givosiran for the treatment of patients with acute intermittent porphyria (Aip). J Med Econ (2020) 23(12):1441–9. doi: 10.1080/13696998.2020.1835306

311. Cadth Common Drug Reviews. Pharmacoeconomic review report: Patisiran (Onpattro): (Alnylam Netherlands bv): Indication: Treatment of polyneuropathy in adult patients with hereditary transthyretin-mediated amyloidosis. Ottawa (ON: Canadian Agency for Drugs and Technologies in Health (2019).

312. Alnylam. *Conference call to discuss fda approval of amvuttra* (2023). Available at: https://alnylampharmaceuticalsinc.gcs-web.com/static-files/d9054736-25e6-4c78-b409-2cab47c8a40c.

313. Sun G, Riggs AD. A simple and cost-effective approach for *In vitro* production of sliced sirnas as potent triggers for rnai. *Mol Ther Nucleic Acids* (2017) 8:345–55. doi: 10.1016/j.omtn.2017.07.008

314. Aguiar S, van der Gaag B, Cortese FAB. Rnai mechanisms in huntington's disease therapy: Sirna versus shrna. *Transl Neurodegener* (2017) 6:30. doi: 10.1186/s40035-017-0101-9

315. Alshaer W, Zureigat H, Al Karaki A, Al-Kadash A, Gharaibeh L, Hatmal MM, et al. Sirna: Mechanism of action, challenges, and therapeutic approaches. *Eur J Pharmacol* (2021) 905:174178. doi: 10.1016/j.ejphar.2021.174178

316. Tatiparti K, Sau S, Kashaw SK, Iyer AK. Sirna delivery strategies: A comprehensive review of recent developments. *Nanomaterials (Basel)* (2017) 7(4). doi: 10.3390/nano7040077

317. Ji L, Chen X. Regulation of small rna stability: Methylation and beyond. *Cell Res* (2012) 22(4):624–36. doi: 10.1038/cr.2012.36

318. Kelleher AD, Cortez-Jugo C, Cavalieri F, Qu Y, Glanville AR, Caruso F, et al. Rnai therapeutics: An antiviral strategy for human infections. *Curr Opin Pharmacol* (2020) 54:121–9. doi: 10.1016/j.coph.2020.09.011

319. Tam YY, Chen S, Cullis PR. Advances in lipid nanoparticles for sirna delivery. *Pharmaceutics* (2013) 5(3):498–507. doi: 10.3390/pharmaceutics5030498

320. Adams D, Gonzalez-Duarte A, O'Riordan WD, Yang CC, Ueda M, Kristen AV, et al. Patisiran, an rnai therapeutic, for hereditary transthyretin amyloidosis. *N Engl J Med* (2018) 379(1):11–21. doi: 10.1056/NEJMoa1716153

321. Scott LJ, Keam SJ. Lumasiran: First approval. Drugs (2021) 81(2):277-82. doi: 10.1007/s40265-020-01463-0

322. Ray KK, Wright RS, Kallend D, Koenig W, Leiter LA, Raal FJ, et al. Two phase 3 trials of inclisiran in patients with elevated ldl cholesterol. *N Engl J Med* (2020) 382 (16):1507–19. doi: 10.1056/NEJMoa1912387

323. Adams D, Tournev IL, Taylor MS, Coelho T, Planté-Bordeneuve V, Berk JL, et al. Efficacy and safety of vutrisiran for patients with hereditary transthyretin-mediated amyloidosis with polyneuropathy: A randomized clinical trial. *Amyloid* (2022), 1–9. doi: 10.1080/13506129.2022.2091985

324. Balwani M, Sardh E, Ventura P, Peiró PA, Rees DC, Stölzel U, et al. Phase 3 trial of rnai therapeutic givosiran for acute intermittent porphyria. *N Engl J Med* (2020) 382 (24):2289–301. doi: 10.1056/NEJMoa1913147

325. Springer AD, Dowdy SF. Galnac-sirna conjugates: Leading the way for delivery of rnai therapeutics. *Nucleic Acid Ther* (2018) 28(3):109-18. doi: 10.1089/nat.2018.0736

326. Li M, Rossi JJ. Lentiviral vector delivery of sirna and shrna encoding genes into cultured and primary hematopoietic cells. *Methods Mol Biol* (2008) 433:287–99. doi: 10.1007/978-1-59745-237-3_18

327. Schomber T, Kalberer CP, Wodnar-Filipowicz A, Skoda RC. Gene silencing by lentivirus-mediated delivery of sirna in human Cd34+ cells. *Blood* (2004) 103(12):4511–3. doi: 10.1182/blood-2003-07-2397

328. Kohn DB, Booth C, Shaw KL, Xu-Bayford J, Garabedian E, Trevisan V, et al. Autologous ex vivo lentiviral gene therapy for adenosine deaminase deficiency. N Engl J Med (2021) 384(21):2002–13. doi: 10.1056/NEJM0a2027675

329. Su L, Tian J, Sun J, Han N, Feng L, Yu B, et al. Lentivirus-mediated sirna knockdown of Sphk1 inhibits proliferation and tumorigenesis of neuroblastoma. *Onco Targets Ther* (2018) 11:7187–96. doi: 10.2147/ott.S180962

330. Gu W, Putral L, Hengst K, Minto K, Saunders NA, Leggatt G, et al. Inhibition of cervical cancer cell growth in vitro and in vivo with lentiviral-vector delivered short hairpin rna targeting human papillomavirus E6 and E7 oncogenes. *Cancer Gene Ther* (2006) 13(11):1023–32. doi: 10.1038/sj.cgt.7700971

331. Schenkwein D, Afzal S, Nousiainen A, Schmidt M, Ylä-Herttuala S. Efficient nuclease-directed integration of lentivirus vectors into the human ribosomal DNA locus. *Mol Ther* (2020) 28(8):1858–75. doi: 10.1016/j.ymthe.2020.05.019

332. Stock PG, Barin B, Murphy B, Hanto D, Diego JM, Light J, et al. Outcomes of kidney transplantation in hiv-infected recipients. *N Engl J Med* (2010) 363(21):2004–14. doi: 10.1056/NEJMoa1001197

333. Sasson SC, Gordon CL, Christo SN, Klenerman P, Mackay LK. Local heroes or villains: Tissue-resident memory T cells in human health and disease. *Cell Mol Immunol* (2020) 17(2):113–22. doi: 10.1038/s41423-019-0359-1

334. Gao C, Wu P, Yu L, Liu L, Liu H, Tan X, et al. The application of Crispr/Cas9 system in cervical carcinogenesis. *Cancer Gene Ther* (2022) 29(5):466–74. doi: 10.1038/ s41417-021-00366-w