



OPEN ACCESS

EDITED BY

Mirza S. Baig,
Indian Institute of Technology Indore, India

REVIEWED BY

Bettina Grötsch,
University Hospital Erlangen, Germany
Hua Ren,
East China Normal University, China

*CORRESPONDENCE

Yunhua Zhou
✉ hzhuster@outlook.com
Xiangjie Liu
✉ liuxiangjie1968@126.com

SPECIALTY SECTION

This article was submitted to
Inflammation,
a section of the journal
Frontiers in Immunology

RECEIVED 11 November 2022

ACCEPTED 30 December 2022

PUBLISHED 18 January 2023

CITATION

Gao Z, Gao Z, Zhang H, Hou S, Zhou Y and
Liu X (2023) Targeting STING: From
antiviral immunity to treat osteoporosis.
Front. Immunol. 13:1095577.
doi: 10.3389/fimmu.2022.1095577

COPYRIGHT

© 2023 Gao, Gao, Zhang, Hou, Zhou and
Liu. This is an open-access article distributed
under the terms of the [Creative Commons
Attribution License \(CC BY\)](#). The use,
distribution or reproduction in other
forums is permitted, provided the original
author(s) and the copyright owner(s) are
credited and that the original publication in
this journal is cited, in accordance with
accepted academic practice. No use,
distribution or reproduction is permitted
which does not comply with these terms.

Targeting STING: From antiviral immunity to treat osteoporosis

Zhonghua Gao¹, Zhongguo Gao², Hao Zhang¹, Shoubo Hou³,
Yunhua Zhou^{4*} and Xiangjie Liu^{1*}

¹Department of Geriatrics, Liyuan Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, Hubei, China, ²Department of Medical Laboratory Technology, School of Biomedical Engineering, Hubei University of Medicine, Shiyan, Hubei, China, ³Department of General Practice, General Hospital of Central Theater Command, Wuhan, Hubei, China, ⁴Department of Wound Repair Surgery, Liyuan Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, Hubei, China

The cGAS-STING signaling pathway can trigger innate immune responses by detecting dsDNA from outside or within the host. In addition, the cGAS-STING signaling pathway has emerged as a critical mediator of the inflammatory response and a new target for inflammatory diseases. STING activation leads to dimerization and translocation to the endoplasmic reticulum Golgi intermediate compartment or Golgi apparatus catalyzed by TBK1, triggers the production of IRF3 and NF- κ B and translocates to the nucleus to induce a subsequent interferon response and pro-inflammatory factor production. Osteoporosis is a degenerative bone metabolic disease accompanied by chronic sterile inflammation. Activating the STING/IFN- β signaling pathway can reduce bone resorption by inhibiting osteoclast differentiation. Conversely, activation of STING/NF- κ B leads to the formation of osteoporosis by increasing bone resorption and decreasing bone formation. In addition, activation of STING inhibits the generation of type H vessels with the capacity to osteogenesis, thereby inhibiting bone formation. Here, we outline the mechanism of action of STING and its downstream in osteoporosis and discuss the role of targeting STING in the treatment of osteoporosis, thus providing new ideas for the treatment of osteoporosis.

KEYWORDS

osteoporosis, STING, IFN- β , NF- κ B, type H vessels

1 Introduction

The stimulator of interferon genes (STING, also known as MITA, MPYS, ERIS, and TMEM173) is a pattern recognition receptor (PRR) that recognizes nucleic acids of pathogenic microorganisms or cell membrane components, among others (1). It acts as the first line of defense of cells against pathogenic invasion. Initially found in the endoplasmic reticulum (ER) membrane of the innate immune cells and later also found to be expressed in T cells and other cells, it recognizes released DNA and triggers innate immune activation with essential functions in infection, inflammation, and cancer (2). STING, a necessary protein of natural immunity, plays a crucial role in antiviral immunity by activating nuclear factor-

kappa B (NF- κ B) and interferon regulatory factor 3 (IRF3) and producing type I interferon (IFN-I) independently of Toll-like receptors (TLRs, another type of PRR) (2).

The cGAS-STING in the innate immune response is vital in defending against pathogenic microbial invasion (3). In addition to its antiviral immune function, STING can cause inflammatory and autoimmune diseases (4). Activation of STING causes the transcription of inflammatory genes and increases pro-inflammatory cytokines. The increase of overpowering pro-inflammatory factors then causes inflammatory and autoimmune diseases (5). Therefore, STING is also the inflammatory protein that triggers chronic inflammation (6). The cGAS-STING pathway mediates the cellular inflammatory response and thus plays a crucial role in the pathogenesis of inflammatory diseases such as ischemic myocardial infarction (MI), nonalcoholic steatohepatitis (NASH), traumatic brain injury (TBI), and silicosis (7). A chronic low-grade inflammatory state accompanies aging. The cGAS-STING pathway can also induce the senescence-associated secretory phenotype (SASP) through the accumulation of cytoplasmic DNA during aging, which leads to the development of aging-related diseases (8).

Hundreds of millions worldwide are affected by bone-related diseases such as osteoporosis, degenerative disc disease, and rheumatoid arthritis. Osteoporosis is an age-related degenerative disease of bone, mainly due to changes in the bone microenvironment and structural degeneration, resulting in reduced bone density (9). It seriously endangers patients' quality of life and lives due to the extreme risk of fractures and others and causes a

substantial financial burden on society. Osteoporosis is also a sterile inflammatory disease characterized by the activation of NF- κ B at the molecular level, which promotes osteoclast-mediated bone resorption and inhibits osteoblast-induced bone formation (10). Notably, STING can act as an upstream of NF- κ B, stimulating its activation and transcription, thus mediating pro-inflammatory effects and playing a role in the pathogenesis of osteoporosis (Figure 1). IFN- β is also a downstream target of STING. However, unlike NF- κ B, although IFN- β is induced by STING activation in osteoclasts, it can inhibit osteoclast activation through negative feedback (11). In addition, STING can act on vascular endothelial cells (ECs) to regulate the formation of type H vessels, which can control bone formation. STING activation can impair their formation and thus affect bone formation (12) (Table 1). Therefore, the role of STING in osteoporosis deserves further investigation to determine how to target STING for osteoporosis treatment.

2 cGAS-STING pathway

The cGAS-STING pathway is a significant component of the host's innate immune response to viral infection. The cGAS senses pathogenic DNA to activate STING to modulate the type I interferon response to trigger a natural immune response. Herpes simplex virus 1 (HSV-1) is a double-stranded DNA virus sensed by the cGAS to activate STING and induce innate antiviral immunity (22). Similarly, other DNA viruses, such as HIV and CMV, can trigger cGAS-STING

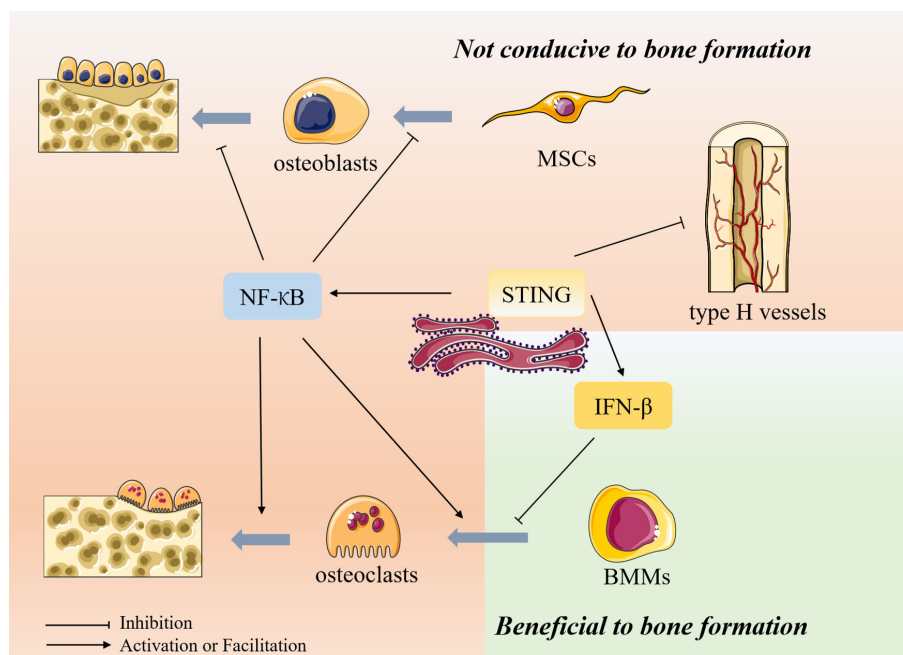


FIGURE 1

The role of STING in bone metabolism Bone metabolism is mainly composed of osteoblast-mediated bone formation and osteoclast-mediated bone resorption. In addition, type H vessels also can induce bone formation and thus participate in bone remodeling. Osteoblasts are differentiated from mesenchymal stem cells (MSCs), whereas osteoclasts are differentiated from bone marrow macrophages. STING can act as an upstream of NF- κ B, stimulating its activation and transcription and thus exerting biological effects. NF- κ B inhibits the differentiation of MSCs toward osteogenesis and inhibits osteoblast activity, thus inhibiting bone formation. In osteoclasts, NF- κ B can promote osteoclast production and activity, thereby promoting bone resorption. IFN- β is also a downstream target of STING. However, unlike NF- κ B, although IFN- β is induced in osteoclasts after activation by STING, it can inhibit osteoclast activation through its negative feedback, thereby inhibiting bone resorption. In addition, STING can inhibit the formation of type H vessels, inhibiting bone formation.

TABLE 1 Summary of research on targeting STING and its signaling pathways in bone metabolism.

Disease	Interventions	Mechanism & Target	Model	Effects
OA	Deficient in STING (13)	Inhibiting STING / IFN-I signaling	DNase II ^{-/-} / IFNAR ^{-/-} double-knockout arthritis mice	Inhibiting abnormal bone formation
	Itaconate (14)	Inhibiting STING/NF-κB axis, Promoting M2 polarization in macrophages	OA mouse	Inhibiting chondrocyte senescence and ECM degeneration, Attenuating osteoarthritis
IVDD	Lipopolysaccharide (LPS) (15)	Activating cGAS/STING pathway	vertebral inflammation-induced caudal IVDD (VI-IVDD) rat	Building a novel model of VI-IVDD
	STING knock-down (16)	Inhibiting cGAS/STING pathway	puncture-induced IVDD rat	Alleviating IVDD development
	Epigallocatechin-3-Gallate (17)	Inhibiting cGAS/STING/ NLRP3 pathway	H ₂ O ₂ -Treated NP cells	Protecting NP cells from apoptosis
Bone Loss (including OP)	STING agonists (18) (DMXAA, ADU-S100)	Activating STING/IFN-I signaling	Lewis lung carcinoma or breast cancer -induced bone loss mice	Reducing bone loss
	CDNs (19)	Activating STING/IFN-β signaling	RANKL-induced BMMs, calvarial implantation mouse	Inhibiting osteoclast differentiation and bone resorption
	Tmem173(STING) overexpression (20)	Overexpressing STING	RANKL-induced BMMs	Inhibiting osteoclast differentiation
	RTA-408 (21)	Inhibiting STING /NF-κB signaling	OVX-induced bone loss	Attenuating osteoclastogenesis
Bone Fracture, Bone Defect	2',3'-cGAMP (12)	Activating STING	Bone fracture and defect mice	Inhibiting type H vessel formation, Delaying bone healing
	STING inhibitors (12) (C-176 and H-151)	Inhibiting STING		Enhancing type H vessel formation, Accelerating bone healing

(23, 24). STING also plays a vital role in the immune response induced by RNA viruses (25). RNA viruses, like dengue virus and SARS-CoV-2, have no DNA and cannot induce cGAS autonomously. However, these RNA viruses can activate the cGAS-STING pathway by triggering intracellular mitochondrial stress damage to release mitochondrial DNA (mtDNA), thereby generating antiviral immunity (26). In addition, cGAS can sense bacterial DNA and the host's DNA, such as senescent apoptotic cells, extracellular vesicles, and chromatin fragments (27). Thus, the cGAS-STING pathway is critical in many disease processes, including autoimmune diseases, inflammatory diseases, degenerative diseases, and cancer (5, 28).

STING is a PRR on the ER that does not bind directly to DNA. Pathogenic microbes and damaged host cells can release free double-stranded DNA (dsDNA) (29). Then dsDNA is recognized by the cytoplasmic DNA sensor, the cyclic GMP-AMP synthase (cGAS) (30). The dsDNA binding to cGAS triggers the conversion of ATP and GTP to cGAMP (2',3'-cyclic GMP-AMP) (31). The cGAMP is canonical cyclic dinucleotides (CDNs) that bind and activate STING (32). CDNs are essential second messengers produced by cyclic dinucleotide synthase, which is widely distributed and can trigger from various cellular signaling cascades, as well as being an activating ligand for STING (33, 34). The binding of cGAMP to STING triggers STING conformational transition, dimerization, and translocation to the endoplasmic reticulum-Golgi intermediate compartment (ERGIC) and Golgi apparatus (Golgi) (35). Then STING dimers recruit TBK1, which phosphorylates STING and induces IRF3 (36). STING also leads to NF-κB activation. IRF3 and NF-κB are translocated to the nucleus to induce the production of IFN-I and other cytokines involved in the host immune response (37) (Figure 2).

Classical STING activation induces the critical transcription factor IRF3 *via* the cGAS-STING pathway, which promotes IFN-I secretion and activates NF-κB to trigger pro-inflammatory cytokines. In recent years, atypical patterns of STING activation have also been identified. Keratinocytes generate an innate immune response within hours of etoposide-induced DNA damage, which involves the DNA sensing adapter STING but is not dependent on cGAS (38). And this non-canonical STING signaling predominantly activates NF-κB rather than IRF3, which induces IFN-I production. This also provides another way of thinking for future STING research.

Although it has been reported that cytoplasmic DNA-mediated STING-dependent inflammatory response requires activation of NF-κB *via* TBK1 (39), at the same time, activation of TBK1 can also cause IFN-β production. However, investigators have found that selective activation of NF-κB can occur in the cGAS-STING pathway, while a parallel path blocks activation of the IRF3/IFN system (40). In late 2019, SARS-CoV-2 emerged as a highly infectious coronavirus that causes a human respiratory disease called COVID-19. SARS-CoV-2 infection can cause respiratory symptoms ranging from mild to severe, resulting in lasting lung damage or death (41). One of the hallmarks of severe COVID-19 is low levels of IFN-I and high levels of expression of inflammatory cytokines or chemokines such as IL-6 and tumor necrosis factor (TNF) (42–44). This unbalanced immune response fails to limit viral transmission and leads to severe systemic symptoms. Specific activation of NF-κB and blockade of IRF3 nuclear translocation occurs in SARS-CoV-2 infected cells, and STING-targeted drugs can attenuate this NF-κB response (44). This NF-κB response is induced by mtDNA released from cellular oxidative stress injury (44, 45). MtDNA mediates the activation of

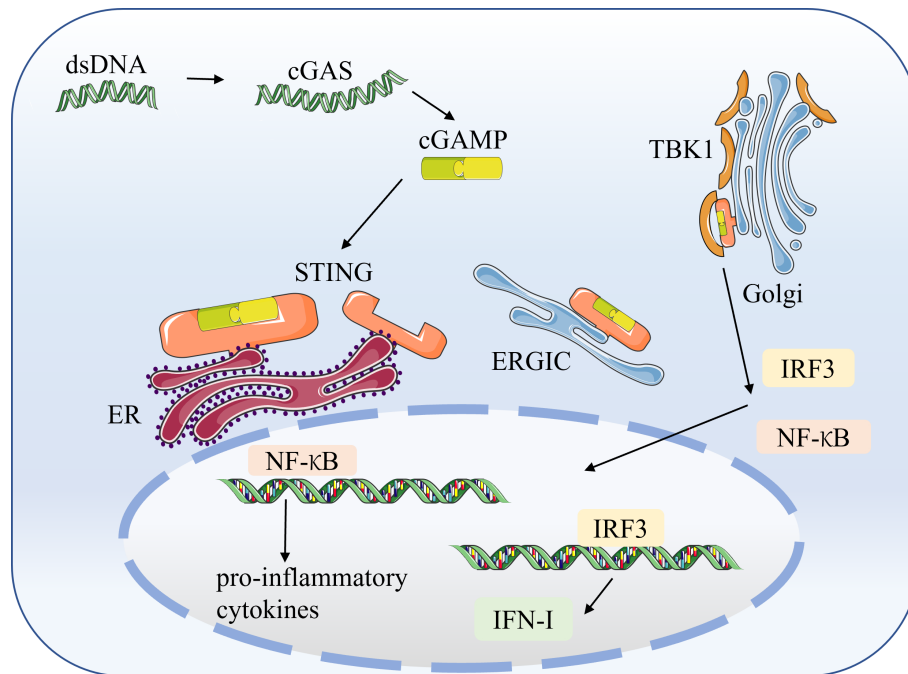


FIGURE 2

The cGAS-STING signaling pathway STING is a pattern recognition receptor (PRR) on the endoplasmic reticulum (ER) that does not bind directly to DNA. Pathogenic microorganisms and damaged host cells can release double-stranded DNA (dsDNA). The cytoplasmic DNA sensor, cyclic GMP-AMP synthase (cGAS), recognizes dsDNA and catalyzes the synthesis of cGAMP from ATP and GTP. The cGAMP binds to STING, triggering STING conformational transitions, dimerization, and translocation to the endoplasmic reticulum-Golgi intermediate compartment (ERGIC) and the Golgi apparatus (Golgi). The dimerized STING recruits TBK1, which induces the production of IRF3 and NF-κB. Subsequently, IRF3 and NF-κB translocate to the nucleus to induce the production of IFN-I and pro-inflammatory factors.

cGAS-STING. Furthermore, cancer studies found that the classical NF-κB pathway in the cGAS-STING pathway enhances anti-tumor effects by promoting IFN-I expression. In contrast, the non-classical NF-κB pathway impedes anti-tumor effects by decreasing IFN-I expression (40). Activation of STING triggers NF-κB activation that can be generated independently of IFN-I.

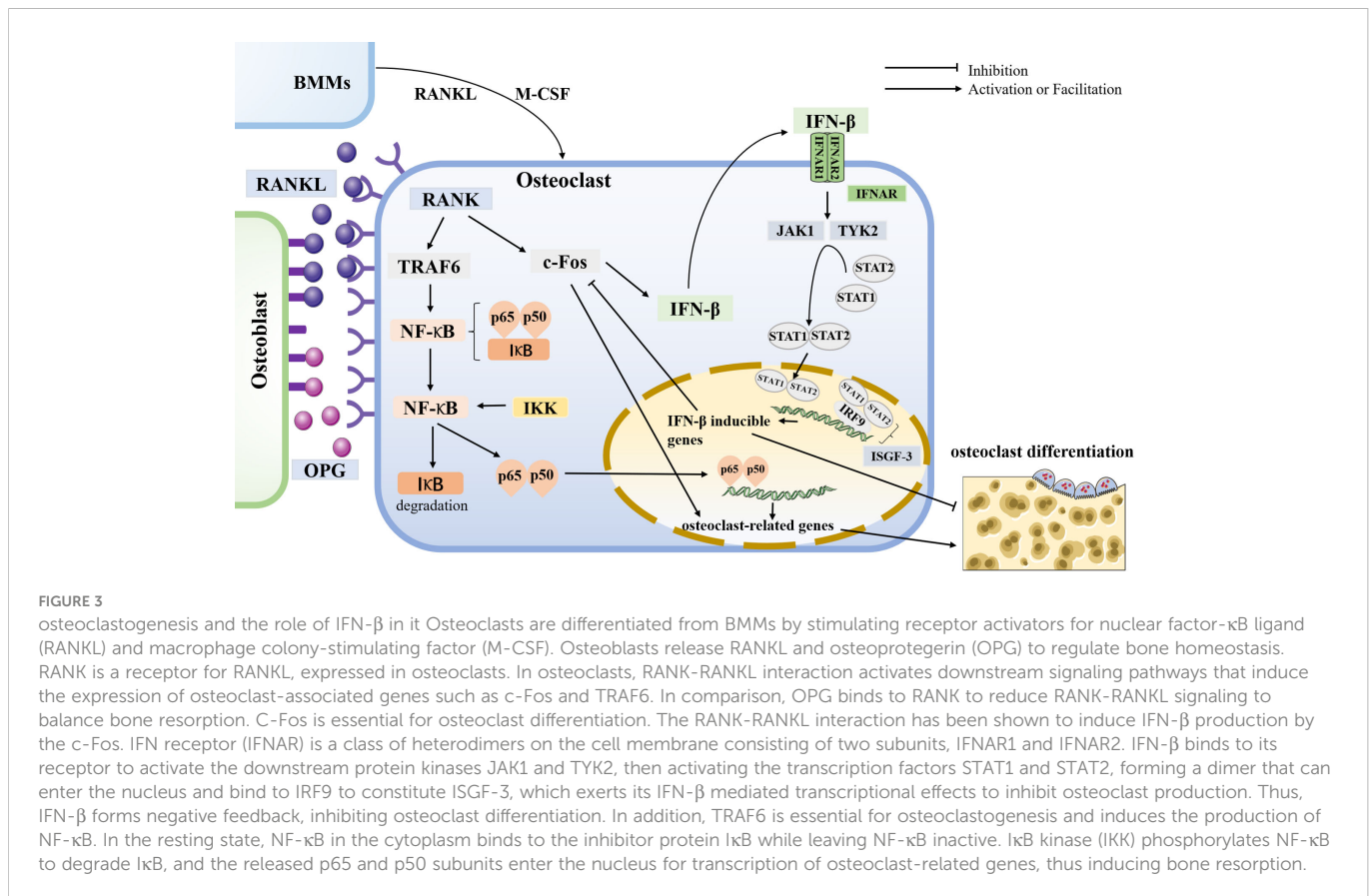
3 STING/IFN-β and osteoporosis

Bone, the body's central axis system, provides physical support and protection, is involved in calcium metabolism and endocrine regulation, and promotes the hematopoietic system in the bone marrow (10). In response to normal wear and mechanical forces as well as the aging process, bone in the adult skeleton undergoes continuous remodeling in which damaged or failing microscopic parts of the bone are removed by osteoclasts and subsequently replaced by new bone laid down by osteoblasts (46). Bone remodeling is a continuous dynamic process that includes bone formation and bone resorption activities, generally in balance, thus maintaining bone homeostasis (47). Bone homeostasis depends on the functional balance between bone-forming cells (osteoblasts) and bone-resorbing cells (osteoclasts) (48). Disruption of bone homeostasis is the frequent pathophysiological mechanism of bone metabolic diseases (49). Excessive osteoclast activity can lead to bone diseases such as osteoporosis, Paget's disease, and rheumatoid arthritis.

Osteoclast differentiation is initiated by bone marrow macrophages (BMMs) through stimulation of receptor activators of nuclear factor-κB ligand (RANKL) and macrophage colony-stimulating factor (M-CSF) (50). Osteoblasts release RANKL and osteoprotegerin (OPG) to regulate bone homeostasis. RANK, the receptor of RANKL, is expressed in osteoclasts. Furthermore, RANK-RANKL interaction activates downstream signaling pathways such as NF-κB, MAPK, and AKT, thereby inducing the expression of osteoclast-associated genes, including c-Fos and NFATc1 (51). Additional studies have shown that osteoclastogenesis generates reactive oxygen species (ROS), and these ROS can induce the activation of downstream signaling pathways, such as NF-κB and MAPK, which also play a role in osteoclast differentiation and bone resorption (52). Conversely, OPG binds to RANK to reduce RANK-RANKL signaling, thereby balancing bone resorption (53). C-Fos is essential for osteoclast differentiation, and lack of c-Fos can lead to osteosclerosis. It interacts with NFATc1, then activates multiple target genes for osteoclast function, triggering a transcriptional regulatory cascade (54). RANKL interactions have been shown to induce IFN-β production through the induction of c-Fos genes (11)(Figure 3).

3.1 Relationship between IFN-β and osteoporosis

IFN-β belongs to type I interferons. The human body produces three known types of interferons: type I, type II, and type III (55).



Type I interferons are mainly IFN- α and IFN- β , secreted by innate immune cells. Type II interferons, IFN- γ , is mainly produced by activated T cells. Type III interferons include IFN- λ , whose known distribution and function are minimal. Type I interferons are mainly produced by surface or internal receptors of innate immune cells (TLRs, NLRs, RLRs, and cGAS) binding to specific antigens from outside or inside the host (56). IFN receptors (IFNAR) are a class of heterodimers located on the cell membrane and consist of two subunits, IFNAR1 and IFNAR2, and widely distributed, including monocytes, macrophages, B cells, T cells, epithelial cells, endothelial cells, and tumor cells (57). Ligand receptor binding activates downstream protein kinases JAK1 and TYK2, and kinase activation activates cytoplasmic transcription factors STAT1 and STAT2, forming a dimer that enters the nucleus to assist IRF9 in transcribing some downstream effector genes (56). Type I interferons can play a biological role in antiviral and immunomodulatory, inhibiting specific cell growth and proliferation and killing tumor cells (58, 59). Therefore, interferon therapy has been used to treat common viral diseases such as hepatitis (60) and various cancers (61).

When osteoclasts are induced to produce IFN- β , the binding of IFN- β to its bioreceptor activates ISGF-3 (formed by the aggregation of STAT1, STAT2, and IRF9) *via* the classical JAK/STAT pathway, initiating a signal transduction cascade (62). Then, c-Fos will be inhibited, leading to the inhibition of osteoclast production and activity (11). Thereby, IFN- β forms negative feedback of its own (Figure 3). Thus, IFN- β also plays a vital role in regulating bone homeostasis. In addition, osteoclasts express

iNOS and release NO, and the NO produced by this pathway also acts as a negative feedback signal to limit RANKL-stimulated osteoclastogenesis (63). In iNOS-deficient bone marrow cells, RANKL-induced NO production was inhibited, leading to an increase in the number of terminally differentiated osteoblasts (64). Direct administration of IFN- β in RAW264.7 cells stimulated iNOS expression in the absence of RANKL, thereby upregulating NO expression. NO, like IFN- β , inhibited osteoclast differentiation. These results suggest that IFN- β may be a key mediator of iNOS-derived NO induction by RANKL in developing osteoclasts and that iNOS can mediate the inhibitory effect of IFN- β on osteoclasts (65).

Another interaction of IFN- β involved in the regulation of bone homeostasis is 4-1BBL with 4-1BB. 4-1BB, also known as CD137, is similar to RANK and is a member of the same TNF receptor family, encoded by the TNFRSF9 gene. Upon its activation, antigen-presenting cells, such as dendritic cells, B cells, and macrophages, express 4-1BBL (66). Osteoclast precursors can express 4-1BB and 4-1BBL after exposure to RANKL (67). In BMMs co-stimulated by M-CSF and RANKL, 4-1BBL mRNAs are upregulated (68). In the animal model, 4-1BB knockout mice also showed increased bone mass compared to the wild group. The number of osteoclasts was significantly reduced in the presence of immobilized recombinant 4-1BB (4-1BB-Fc). 4-1BB can induce the binding activity of IRF3, and IRF3 is activated by 4-1BB stimulation, which induces IFN- β (11, 69, 70). It is not difficult to speculate that the decrease in osteoclast activity caused by 4-1BB should be due to the inhibition of c-Fos expression by IFN- β .

3.2 Targeting STING/IFN- β in osteoporosis

As a critical signal transduction molecule involved in the innate immune response, STING, triggered by cytoplasmic DNA from pathogens and hosts, can induce type I interferon and pro-inflammatory cytokine secretion, defend against viral and intracellular bacterial infections, and regulate the spontaneous anti-tumor immune response *in vivo*. Targeted STING is a new tool for immunotherapy. In addition to immune or oncological diseases, the role of STING in bone metabolic diseases has been the focus of attention in recent years. DNase II is a nuclease that degrades dsDNA. Lack of DNase II causes DNA accumulation in cells and produces several cytokines, including type I IFN (71). Mice lacking DNase II and IFNAR were able to develop distal aggressive inflammatory arthritis (72). However, this arthritis was eliminated in the absence of STING (71, 73). Surprisingly, the arthritis model (DNase II^{-/-}/IFNAR^{-/-} double-knockout mice) showed aberrant accumulation of bone in both long bones and the spleen at sites of local DNA accumulation (13). STING deficiency inhibited bone accumulation (13), revealing a potential role of the STING pathway in bone, although the exact mechanism is unclear.

Patients with advanced cancer often suffer from severe pain due to bone metastases and bone destruction with osteolytic lesions (74). Agonists of the immunomodulator STING have significantly protected against pain (75), bone destruction (13, 19), and local tumor burden (76). One of its effects is alleviating cancerous bone pain by regulating osteoclast function in the tumor microenvironment to prevent local bone destruction, which depends on host-intrinsic STING/IFN- β signaling. Bone metastases in patients with cancer produce osteolytic bone destruction due to tumor-induced osteoclast formation and activation (77). Bone loss was significantly reduced in Lewis lung carcinoma (LLC) or breast cancer mice treated with DMXAA and ADU-S100, two different STING agonists, similar to zoledronic acid (ZA) (18). In contrast, the reversal effect of DMXAA on bone loss was eliminated in the STING knockout group of mice. Thus, the inhibitory effect of STING agonists on bone resorption is dependent on STING. Systemic administration of STING agonists also promotes a robust IFN-I response in the systemic and bone cancer tumor microenvironment. DMXAA treatment does not prevent bone destruction in IFNAR1-deficient mice. IFNAR is required for IFN signaling (57). Thus, the protective effects of STING agonists against cancerous bone destruction require IFN-I signaling (18).

STING, also known as Tmem173, has been shown to inhibit osteoclast differentiation and activity by regulating IFN- β production. It inhibits the expression of osteoclast-specific genes and related enzymes and downregulates the activation of osteoclast-specific transcription factors (20). CDNs are symbiotic bacterial-derived second messengers in the intestine that regulates bacterial survival, colonization, and biofilm formation and have immunomodulatory activity by inducing type I interferon expression by macrophages through the STING signaling pathway (34, 78). CDNs dose-dependently inhibit M-CSF and RANKL-induced differentiation of bone marrow macrophages to osteoclasts and induce phosphorylation of TBK1 and IRF3, representative features of STING activation (19). In contrast, inhibition of osteoclast differentiation was reversed in STING knockdown BMMs. These suggest that the STING signaling

pathway plays a crucial role in CDNs-mediated inhibition of osteoclast differentiation. In addition, CDNs increased the expression of IFN- β , a member of the IFN-I family, which has also been identified as a typical negative regulator of RANKL-induced osteoclast differentiation (79, 80). RANKL induces IFN- β expression *via* c-Fos (81). In turn, IFN- β binds to IFNAR on the membrane, which activates ISGF-3 and prevents RANKL-induced c-Fos expression from inhibiting osteoclast differentiation (11). The inhibitory effect of CDNs on osteoclast differentiation was absent in the presence of antibodies blocking IFNAR, and no inhibitory effect was observed in knockout IFNAR macrophages (19). These also confirm that CDNs induce phosphorylation of STAT1, which mediates IFNAR signaling. Experiments performed with a mouse cranial implant model showed that CDNs inhibit RANKL-induced bone resorption (19). These results suggest STING induces IFN- β to inhibit osteoclast differentiation and bone resorption.

It is well known that IFN-I response is a weapon against viruses. IFN-I is induced during STING-mediated immune responses. IFN-I can also be stimulated by the osteoclast-specific gene c-Fos and ultimately inhibits osteoclast production and activation through IFNAR transmission (11, 82). STING regulates the inhibitory effect of IFN-I on osteoclasts, and the knockdown of STING reverses this effect (79). Knockdown of STING can offset this effect (19). Therefore, targeting STING/IFN- β to increase IFN- β expression and thereby inhibit osteoclast bone resorption is expected to be a new approach to treating osteoporosis. In addition, interferon therapy has been used in the clinic. And IFN- β has a relatively good clinical tolerability and safety profile. However, it faces several tests when using IFN- β to treat bone metabolic diseases, including osteoporosis. First, as with other protein drugs, treatment with interferon results in the production of neutralizing antibodies in the patient (83). Second, effective delivery of the drug to the bone microenvironment is another challenge in using IFN- β for treating bone metabolic diseases (84). Targeting STING to increase the level of interferon in the body may solve these problems faced by treatment with interferon alone. In addition, the inhibitory effect of IFN- β on osteoclasts is mainly due to its negative feedback mechanism. However, the action of IFN- β is also inhibited by another kind of negative feedback. Suppressors of cytokine signaling (SOCS)-1 and SOCS-3 in response to RANKL can act as inhibitory factors that significantly inhibit IFN- β signaling (85). Thus IFN- β -mediated inhibition of osteoclastogenesis has a potential counteracting pathway. It may be the same challenge for targeting STING/IFN- β with interferon therapy alone.

4 STING/NF- κ B and osteoporosis

Past studies have indicated that the cGAS-STING pathway is a key component of the innate immune response as a host defense against multiple pathogens. At the same time, sustained STING activity may lead to fatal inflammatory diseases (39). The continuous secretion of pro-inflammatory cytokines enhances tissue destruction and impairs the organism's homeostasis, thus affecting functional integrity. NF- κ B is a downstream target of STING signaling and can be activated by it. NF- κ B is a ubiquitous transcription factor activated by various stimuli, including infection, inflammation, and oxidative stress (86). The aging process is

accompanied by a chronic and persistent inflammatory state (87). NF- κ B is also a hub of the aging process, promoting transcription and expression of various genes associated with inflammation and can regulate inflammatory signaling during aging induced by oxidative stress (88). NF- κ B is associated with many age-related diseases and inflammatory diseases (89), including Alzheimer's disease (90), diabetes mellitus (91), cancer (92), and autoimmune and inflammatory diseases (93). Activation of NF- κ B signaling was found in senescent ARPE-19, and NF- κ B was confirmed to be a downstream target of STING in oxidative stress-induced senescent retinal pigment epithelium (RPE) (94). In microgliomas, polyglutamine binding protein 1 (PQBP1) activates cGAS-STING by interacting with sensing extrinsic tau 3R/4R proteins (95). Activation of the PQBP1-cGAS-STING pathway leads to nuclear translocation of NF- κ B and expression of inflammatory genes, resulting in brain inflammation and cognitive dysfunction in mice. Psoriasis, a chronic inflammatory skin disease, is associated with innate and adaptive immune responses. STING antagonist H-151 ameliorates psoriasis by inhibiting STING/NF- κ B-mediated inflammation (96).

Inflammation is also closely associated with bone metabolism diseases, including osteoporosis (97), osteoarthritis(OA) (98), intervertebral disc degeneration(IVDD) (99), bone lysis (100), and spondyloarthritis (101). STING upregulation was found to be associated with the development of IVDD. And vertebral inflammation mediated by activation of the cGAS/STING molecular pathway is a novel form of animal model used to induce disc degeneration (15). Excessive accumulation of ROS can lead to DNA damage, which activates the cGAS/STING pathway (102). ROS-induced DNA damage is thought to be one of the leading causes of nucleus pulposus (NP) cell degeneration during IVDD progression (103). Moreover, the knockdown of STING expression can attenuate ROS-induced disc degeneration (16). Similarly, pharmacological inhibition of STING also protects NP cells from inflammation-induced apoptosis (17). Moreover, the process of OA is also accompanied by increased expression of STING and NF- κ B, and exogenous supplementation with itaconate can inhibit the STING/NF- κ B signaling pathway to alleviate the progression of OA (14). Osteoporosis, an age-related disease of bone metabolism, is also an inflammatory disease. Elevated levels of NF- κ B can also be found in osteoporosis models. Aging-associated bone loss is characterized by decreased bone formation and increased bone resorption, and it is often referred to as senile osteoporosis (104). Aging is a biological process characterized by changes in the redox state of the organism and inflammatory responses induced by oxidative stress (105). Oxidative stress can release mtDNA (106), which can act as an upstream of the cGAS-STING signaling pathway and activate STING (107). The activation and transduction of STING are crucial in the development and progression of aging-related diseases (108, 109). Therefore the application of targeting STING/NF- κ B in osteoporosis is worth exploring.

4.1 NF- κ B

NF- κ B is one of the best-characterized transcription factors that regulate inflammation and innate and adaptive immune responses

(110). Activation of NF- κ B signaling leads to the production of various inflammatory cytokines, chemokines, adhesion molecules, transcription factors, and antimicrobial effector molecules that initiate and mimic inflammatory responses and coordinate the immediate host response to pathogens and tissue damage. The NF- κ B transcription factor family includes five members p50 (NF- κ B1), p52 (NF- κ B2), RelA (p65), RelB, and c-Rel (111). All NF- κ B subunits have a structurally conserved N-terminal sequence spanning 300 amino acid residues called the Rel homology structural domain (RHD) (112). The RHD is responsible for DNA binding, dimerization, and nuclear translocation of NF- κ B subunits, which can divide into three structural components - the N-terminal structural domain (NTD), the dimerization structural domain (DD), and the nuclear localization sequence (NLS) polypeptide - all of which mediate the various activities of the RHD and subsequent NF- κ B signaling (110, 113, 114). Subunits RelA, RelB, and c-Rel are produced as mature proteins. In contrast, the p50 and p52 subunits are produced by the precursor proteins (113).

In the resting state, NF- κ B subunits bind to I κ B proteins, inhibiting their activity and maintaining NF- κ B subunits in an inactive state (115). In turn, I κ B kinase (IKK) can degrade these inhibitory proteins (116). Once activated by upstream signaling cascades, phosphorylated IKK degrades the I κ B protein and releases the subunits of the NF- κ B. Then these subunits go into the nucleus as dimers and participate in the transcription of various target genes (10). For example, the functional subunits p65 and p50 enter the nucleus and bind to target genes, producing large amounts of inflammatory mediators, and the gene products further activate NF- κ B, causing an expanded cascade of uncontrolled inflammatory responses. More and more NF- κ B target genes have been identified (117), including various cytokines such as interleukin (IL) and TNF, interferons, and antiapoptotic proteins, such as BIRC2, BIRC3, and BCL2L1.

4.2 Relationship between NF- κ B and osteoporosis

Bone homeostasis is necessary for the maintenance of normal bone function. Bone homeostasis is maintained by bone remodeling mediated by osteoblasts and osteoclasts, which are responsible for bone formation and resorption (48). Osteoblasts and osteoclasts are the essential cells that regulate bone homeostasis. NF- κ B is the master transcription factor that regulates the inflammatory response and bone remodeling process (118, 119). Chronic inflammation induces excess pro-inflammatory cytokines, disrupting homeostasis (120). These result in abnormal bone remodeling, including osteosclerotic and osteolytic lesions (121).

Pro-inflammatory cytokines driven by NF- κ B are powerful signals to regulate bone homeostasis (122). Elevated expression of TNF, IL-1, IL-6, and IL-7 has been found in various chronic inflammatory bone diseases, including osteoarthritis (123), osteoporosis (124), and periodontal disease (125). These pro-inflammatory cytokines are all produced by macrophages, lymphocytes, osteoblasts, and bone marrow stromal cells under the regulation of NF- κ B and stimulate NF- κ B signaling in target cells, which further serves to amplify inflammation (126). Osteoclasts are specialized cells of the monocyte-macrophage lineage responsible for

bone resorption. In contrast, osteoblasts are differentiated from mesenchymal stem cells to osteogenesis and are responsible for establishing new bone. NF- κ B has an essential role in osteoblasts and osteoclasts, thus affecting bone regulation.

4.2.1 Role of NF- κ B in bone resorption

NF- κ B signaling is directly involved in the differentiation and activation of osteoclasts responsible for bone resorption (127). The binding of RANKL to RANK triggers a complex and unique signaling cascade that controls lineage commitment and activation of osteoclasts (128). Activating NF- κ B signaling in osteoclasts is essential for their differentiation and activation (54). TNF receptor-associated factor (TRAF) proteins are cytoplasmic adaptor proteins that bind to various receptors of the TNF receptor (TNFR) superfamily. An essential role of TRAFs in RANK-RANKL signaling is inducing NF- κ B (51). Among TRAFs, TRAF6 is the most critical adaptor of RANK-RANKL-induced osteoclastogenesis (129). Genetic experiments have shown that TRAF6 is required for osteoclast formation and activation (130). Like mice lacking NF- κ B p50 and p52 subunits (131), TRAF6-deficient mice develop severe osteoporosis (132).

Usually, NF- κ B in the cytoplasm is bound to the inhibitory protein I κ B while keeping NF- κ B in a resting state. While various stimuli lead to the activation of IKK, which leads to the degradation of I κ B bound to the NF- κ B subunits, the released NF- κ B enters the nucleus as a homodimer or a heterodimer and activates transcription, thus exerting biological effects. For example, released p65 and p50 subunits enter the nucleus for transcription of osteoclast-related genes, thus inducing bone resorption (21). IKK is a complex of three subunits, IKK α (also known as IKK1), IKK β (also known as IKK2), and IKK γ (also known as NEMO). IKK β is required for osteolysis *in vitro* and *in vivo*, and the knockdown of IKK β can lead to bone loss in mice (133). While IKK α is required for RANK ligand-induced osteoclast formation *in vitro*, it is not required *in vivo* (134). Thus, targeting IKK can regulate the NF- κ B activity of osteoclasts and prevent bone loss, providing a new idea for the treatment of osteoporosis (135). In conclusion, NF- κ B is an essential mediator of osteoclastogenesis (136), which leads to excessive bone resorption and osteoporosis. Pharmacotherapy can inhibit RANKL-mediated osteoclastic formation by targeting the NF- κ B pathway to attenuate inflammatory factors and ROS production and can reduce bone loss *in vivo* in ovariectomized (OVX) model (137).

4.2.2 Role of NF- κ B in bone formation

Osteoblasts derived from mesenchymal stem cells (MSCs) are responsible for bone formation. NF- κ B activity is suppressed in mature osteoblasts, so NF- κ B activation in osteoblasts inhibits bone formation (136). NF- κ B activation occurs in bone trabeculae of naturally aging mice (138). Increased NF- κ B activity was found in MSCs isolated from aged mice compared to young mice, and inhibition of the NF- κ B pathway partially rescued the reduction in osteogenesis in aged MSCs (139). And increased RANKL and reduced OPG expression was observed in aged MSCs, which resulted in increased RANKL/OPG ratio and osteoclast activation. Chronic NF- κ B activation has also been shown to impair the differentiation of MSCs along the osteogenic pathway and osteoblast-mediated bone formation (140).

In the absence of NF- κ B activation, prolonged c-Jun N-terminal kinase (JNK) activation, which regulates FOSL1 (also known as Fra1)

expression, contributes to bone formation (126, 141). Mice specifically lacking IKK- β in osteoblasts exhibit increased bone mass, mainly because reduced NF- κ B activity by IKK- β deficiency increases JNK activity and Fra1 expression, ultimately leading to increased bone formation to maintain bone mass in OVX mice (142). Fra1 is an important transcription factor involved in bone matrix formation (143). Chronic inflammation can inhibit bone formation. For example, the pro-inflammatory factor TNF- α inhibits osteoblast differentiation (144), but the IKK inhibitor BAY11-7082 rescues the TNF- α -induced inhibition of osteoblast differentiation by inhibiting NF- κ B (145). Thus, inhibition of osteoblast NF- κ B can promote bone formation. A decrease in NF- κ B activity in osteoblasts leads to an increase in bone formation (146). The NF- κ B inhibitor, S1627, upregulates the mRNA of osteoblast-specific genes (such as type I collagen and alkaline phosphatase) to increase osteoblast differentiation and bone formation *in vitro* (147). Moreover, it can increase bone formation to repair bone defects in a mouse cranial defect model and alleviate osteoporosis in the OVX mouse model. Therefore, targeting NF- κ B could provide a novel and effective therapeutic strategy for osteoporosis and other inflammatory bone diseases.

4.3 Targeting STING/NF- κ B in osteoporosis

Excessive accumulation of ROS leading to redox imbalance and overactive osteoclasts is associated with the progression of osteoporosis. The process of osteoclastogenesis is accompanied by the production of ROS, which plays a role in osteoclast differentiation and bone resorption (52). In addition, ROS-induced mtDNA release induces inflammation through the activation of cGAS/STING (148). ROS can induce NF- κ B through the RANKL/RANK cascade reaction, which is further involved in osteoclastogenesis (149). In addition to promoting type I interferon production in the innate immune response (150), STING can also act as an NF- κ B upstream of NF- κ B, stimulating its production.

In the past, it was generally considered that NF- κ B activation *via* STING is exclusively dependent on TBK1. However, studies have now demonstrated that TBK1 is dispensable for NF- κ B, although TBK1 and its kinase activity are essential for STING-dependent IRF3 activation and INF-I. In fact, TBK1 and IKK redundantly drive NF- κ B activation when the IFN-I reaction is triggered by TBK1 and its homolog, IKK (151). Inhibition by TBK1/IKK kinase indicates that IRF3 activation highly depends on TBK1 kinase activity. In contrast, NF- κ B sensitivity to TBK1/IKK kinase inhibition is significantly reduced, and TBK1 was dispensable for NF- κ B activation downstream of STING *in vitro* and *in vivo* (151). So, NF- κ B production can be activated by STING through a non-classical pathway, a process that is independent of IFN- β (44). In addition, non-classical STING signaling activates NF- κ B pathways mainly through K63-mediated ubiquitination and has no effect on IFN-I (38).

CDNs can inhibit osteoclast differentiation by inducing IFN- β through STING signaling (19), suggesting that activation of STING can inhibit osteoclast differentiation through IFN- β . A sustained activation state of STING can cause a series of inflammatory responses in the organism. NF- κ B, acting as a pro-inflammatory gene, is involved in osteoclastogenesis as another downstream of

STING. In contrast to NF- κ B, nuclear factor erythroid2-related factor 2 (Nrf2), a critical antioxidant molecule, has been shown to inhibit osteoclast formation and bone resorption by reducing ROS (152). In addition, Nrf2 negatively regulates STING signaling (153).

RTA-408 was found to act as an activator of Nrf2 that inhibits STING expression and subsequent NF- κ B activation but does not affect IFN- β expression (21). RTA-408 inhibits RANKL-induced K63 ubiquitination of STING by suppressing the interaction between STING and the E3 ubiquitin ligase TRAF6. As a downstream of STING, NF- κ B was also inhibited by RTA-408, mainly by suppressing I κ B α protein degradation, preventing p65 from translocating to the nucleus and thus rendering NF- κ B inactive. Overexpression of STING rescued the inhibitory effect of RTA-408 on NF- κ B signaling and osteoclastogenesis. *In vivo* experiments showed that RTA-408 attenuated osteoclastogenesis-induced bone loss in C57BL/6 mice by inhibiting STING-mediated NF- κ B (21). Thus, inhibition of STING-dependent NF- κ B signaling could inhibit osteoclastogenesis and reduce bone loss. Targeting STING/NF- κ B may be a promising pathway for the future treatment of osteoporosis.

5 STING/type H vessels and osteoporosis

In recent years, it has been found that in addition to osteogenic and osteoclastic effects, angiogenesis also plays a vital role in bone homeostasis in the mammalian skeletal system. Type H vessels that can induce bone formation have been discovered recently and are named for their high expression of EMCN and CD31 (154). Angiogenesis, the development of new blood vessels from pre-existing vessels, is closely associated with osteogenesis during skeletal development and bone remodeling. Blood vessels provide bone tissue with essential nutrients, oxygen, growth factors, and hormones and play a crucial role in the regulation of bone formation (155).

5.1 Relationship between type H vessels and osteoporosis

Osteogenesis is linked to angiogenesis (156). The close spatial and temporal link between osteogenesis and angiogenesis has been termed “angiogenesis-osteogenesis coupling” (157). Type H vessels are located near the epiphyseal growth plate, the epiphyseal periosteum, and the endosteum. Type H vessels are densely surrounded by osteoprogenitors expressing Osterix, a potent promoter of bone formation (158). These osteoprogenitor cells can differentiate into osteoblasts and osteocytes. Under aging conditions, osteoblasts are significantly reduced in the long bones of mice (159), which is associated with a decrease in Type H vessels and reduced bone mass (160). The abundance of Type H vessels is an essential indicator of bone loss in elderly subjects and patients with osteoporosis (161).

PDGF-BB is a chemotactic and mitogenic factor of the PDGF family, produced by hematopoietic stem cells (162). It is essential for promoting the migration, proliferation, and differentiation of various mesenchymal cell types, such as endothelial progenitor cells and mesenchymal stem cells, to promote angiogenesis and osteogenesis (163, 164). PDGF-BB enhanced type H vessels and bone formation

during bone plastination and remodeling. The concentration of PDGF-BB was decreased in the OVX mouse model (165). Pharmacological stimulation was able to secrete PDGF-BB to stimulate H-type angiogenesis, thereby promoting osteogenesis to prevent bone loss in OVX mice (166). Glucocorticoids reduce vascularity and blood flow to the bone, causing osteonecrosis and bone loss (167, 168). Glucocorticoid-induced osteoporosis (GIO) is also common osteoporosis. In GIO mouse models, glucocorticoids inhibit PDGF-BB secretion by pre-osteoblasts, inhibiting Type H vessels and reducing osteogenic capacity (169). And L-235, a cathepsin K inhibitor, prevents bone loss by inhibiting osteoclast-inducing bone resorption while maintaining PDGF-BB secreted by preosteoclasts preserving Type H vessels (169).

HIF-1 α is a transcription factor that mediates the cellular response to an altered oxygen environment and controls angiogenesis (170). HIF-1 α plays a crucial role in bone formation, regeneration plays a key role in bone formation and regeneration, and its expression and activity are regulated by hypoxia. Oxygen is required for the high metabolic demand of osteoblasts. Therefore, osteoblasts and nearby ECs may increase HIF-1 α expression during relative hypoxia during osteogenesis (155). And ECs express HIF-1 α at high levels in young mice, which decreases with age and is associated with a decrease in ECs and age-dependent bone loss. Activation of hypoxic signaling in ECs increased the number of Type H vessels and enhanced angiogenesis and osteogenesis (171). EC-specific deletion of HIF-1 α resulted in a significant decrease in osteoblast producers and was associated with reduced trabecular formation. Thus, HIF-1 α signaling is vital in regulating type H vascular abundance and couples angiogenesis to osteogenesis (172). Tetramethylpyrazine activates the AMPK/mTOR/HIF-1 α signaling pathway to induce type H vessel angiogenesis and improve bone homeostasis in aging mice (173). This provides an additional therapeutic target for the treatment of age-related osteoporosis.

5.2 Targeting STING/type H vessels in osteoporosis

Type H vessels also play an indelible role in bone remodeling. Osteoblasts, osteoclasts, and periosteal cells interact with vascular endothelial cells. The STING signaling pathway may act directly on these cells, thereby affecting the angiogenic process. Several studies have shown an association between STING and angiogenesis. STING is expressed in endothelial cells, and cGAMP leads to the activation of cGAS-STING in endothelial cells (174, 175). STING-associated vascular disease (SAVI), with onset in infancy, is an autoinflammatory disease caused by mutations in STING function, which can cause vascular and pulmonary syndromes and cause systemic inflammatory responses (176, 177).

Previous studies have shown that STING can affect angiogenesis in multiple ways. In the zebrafish xenograft model, exogenous administration of cGAMP can activate STING-dependent STAT3, leading to the inhibition of tumor vascular proliferation and migration (178). Palmitic acid (PA) induces mtDNA into the cytoplasm by inducing mitochondrial damage, activating the cGAS-STING-IRF3 signaling (179). Activation of the cGAS-STING-IRF3 pathway dysregulates the Hippo-YAP pathway and inhibits angiogenesis. In

addition, STING-IRF3 can trigger endothelial inflammation in response to PA-induced mitochondrial damage (180). In retinal microangiopathy, mtDNA drives inflammation of microvascular endothelial cells *via* the cGAS-STING signaling pathway (181). Inflammation in the physiological state is a protective mechanism for tissue damage and the basis for tissue repair and regeneration. Nevertheless, an excessive inflammatory response can impair the integrity of the tissue and its function. The persistent inflammatory state of the vascular endothelium leads to impaired angiogenesis and poor bone healing, which affects bone reconstruction (182).

Activation of the STING signaling pathway impairs angiogenesis, including type H vessels. In addition, the prolonged inflammatory response stimulated by the STING pathway delays the bone healing process. Activation of STING inhibits angiogenesis both *in vitro* and *in vivo* and slows the bone healing process *in vivo* (12). Conversely, inhibition of STING accelerated bone healing by enhancing type H vessel formation during coupled osteogenesis (12). Therefore, targeting STING to enhance type H vessel formation and thus promote osteogenesis provides a new idea for the treatment of osteoporosis.

6 Conclusions

Osteoporosis is a bone metabolic disease and an aseptic inflammatory disease. In recent years, bone immunology has become a hot research topic in bone metabolic diseases by studying the functional interactions between the skeletal and immune systems, including various cytokines and transcription factors that affect both systems, to explore new immunological therapeutic avenues for bone metabolic diseases. STING is the core of natural immunity and a new target for immunotherapy. The core of IFN- β treatment for osteoporosis is also inseparable from bone immunology. With the discovery that IFN- β can play a unique role in regulating bone homeostasis, targeting the STING/IFN- β signaling pathway is also emerging as a potential therapeutic tool for osteoporosis. However, the STING pathway has a dual role in bone metabolism. In addition to its immune function, STING can also act as an inflammatory protein to induce NF- κ B, thereby mediating the development and progression of various inflammatory diseases, including osteoporosis. But in studies on osteoclastic inhibition by STING/IFN- β , the effect of another STING downstream signaling molecule, NF- κ B, had rarely been considered, which is a drawback of related studies. If NF- κ B is not disturbed in the activation of STING/IFN- β pathway to inhibit osteoclastic resorption, it will be a more rigorous and appropriate choice. The classical STING/NF- κ B pathway, the cGAS-STING pathway, is dependent on the activation of TBK1, which not only activates NF- κ B but also mediates the production of IFN- β by activating IRF3. In contrast, the non-classical STING/NF- κ B pathway blocks the production of IRF3/IFN- β in parallel. Inhibition of NF- κ B activation and, thus, osteoclast differentiation by targeting STING without affecting the level of IFN- β has been shown to alleviate bone loss in the OVX mice. Therefore, targeting the STING/NF- κ B pathway is also expected to be a new therapy for osteoporosis. In addition, activation of STING leads to a prolonged inflammatory response that delays revascularization and inhibits the production of type H vessels, which are closely related to osteogenesis and can induce bone formation. Targeting STING/type H

vessels also promotes bone reconstruction and osteogenesis by promoting type H vessel formation. To enhance bone formation and inhibit bone resorption, a STING inhibitor would be a reasonable choice if it were designed to specifically target NF- κ B rather than IFN- β .

Overall, STING has a unique role in osteoporosis. The drugs commonly used in clinical to treat osteoporosis are mainly bisphosphonates, which inhibit bone resorption, and calcitonin and estrogen drugs, which can also promote osteoblastogenesis, but they all have many side effects. Therefore, how to safely and effectively treat osteoporosis remains a challenge to tackle. Targeted STING has been applied in antiviral immunotherapy and does represent a rather promising therapeutic option in the new field of treatment of osteoporosis. Targeted STING not only has the ability to directly inhibit osteoclast-mediated bone resorption and promote osteoblast-mediated bone formation but also can promote type H vascular angiogenesis to indirectly enhance the osteogenic effect. However, the network of regulatory pathways involved in STING is also very complex, and the decrease in IFN- β production-mediated bone resorption due to STING activation is in contradiction with the increase in bone resorption and decrease in bone formation mediated by NF- κ B activation and type H vascular inhibition. Targeting STING as a therapeutic option for osteoporosis requires balancing these conflicting biological effects. Therefore, there is still much room for exploration of the STING pathway in bone metabolism.

Author contributions

ZG (1st author) conceived and drafted the manuscript. HZ, SH, YZ, and XL discussed the concepts of the manuscript. ZG (1st author) and ZG (2nd author) drew the figures. YZ and XL reviewed and revised the manuscript. All authors contributed to the article and approved the submitted version.

Funding

This study was supported by the National Key R&D Program of China (2018YFC2002000).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

- Cheng Z, Dai T, He X, Zhang Z, Xie F, Wang S, et al. The interactions between cGAS-STING pathway and pathogens. *Signal Transduct Target Ther* (2020) 5:91. doi: 10.1038/s41392-020-0198-7
- Ishikawa H, Barber GN. STING is an endoplasmic reticulum adaptor that facilitates innate immune signalling. *Nature* (2008) 455:674–8. doi: 10.1038/nature07317
- Xia P, Wang S, Gao P, Gao G, Fan Z. DNA Sensor cGAS-mediated immune recognition. *Protein Cell* (2016) 7:777–91. doi: 10.1007/s13238-016-0320-3
- Zhou R, Xie X, Li X, Qin Z, Wei C, Liu J, et al. The triggers of the cGAS-STING pathway and the connection with inflammatory and autoimmune diseases. *Infect Genet Evol* (2020) 77:104094. doi: 10.1016/j.meegid.2019.104094
- Skopelja-Gardner S, An J, Elkouf KB. Role of the cGAS-STING pathway in systemic and organ-specific diseases. *Nat Rev Nephrol* (2022) 18:558–72. doi: 10.1038/s41581-022-00589-6
- Li T, Chen ZJ. The cGAS-cGAMP-STING pathway connects DNA damage to inflammation, senescence, and cancer. *J Exp Med* (2018) 215:1287–99. doi: 10.1084/jem.20180139
- Wan D, Jiang W, Hao J. Research advances in how the cGAS-STING pathway controls the cellular inflammatory response. *Front Immunol* (2020) 11:615. doi: 10.3389/fimmu.2020.00615
- Paul BD, Snyder SH, Bohr VA. Signaling by cGAS-STING in neurodegeneration, neuroinflammation, and aging. *Trends Neurosci* (2021) 44:83–96. doi: 10.1016/j.tins.2020.10.008
- Gao Z, Chen Z, Xiong Z, Liu X. Ferroptosis - a new target of osteoporosis. *Exp Gerontol* (2022) 165:111836. doi: 10.1016/j.exger.2022.111836
- Lin T-H, Pajarinen J, Lu L, Nabeshima A, Cordova LA, Yao Z, et al. NF- κ B as a therapeutic target in inflammatory-associated bone diseases. *Adv Protein Chem Struct Biol* (2017) 107:117–54. doi: 10.1016/bs.apcsb.2016.11.002
- Abraham AK, Ramanathan M, Weinstock-Guttman B, Mager DE. Mechanisms of interferon- β effects on bone homeostasis. *Biochem Pharmacol* (2009) 77:1757–62. doi: 10.1016/j.bcp.2009.01.007
- Chen X, He W, Sun M, Yan Y, Pang Y, Chai G. STING inhibition accelerates the bone healing process while enhancing type h vessel formation. *FASEB J* (2021) 35:e21964. doi: 10.1096/fj.202100069RR
- Baum R, Sharma S, Organ JM, Jakobs C, Hornung V, Burr DB, et al. STING contributes to abnormal bone formation induced by deficiency of DNase II in mice. *Arthritis Rheumatol* (2017) 69:460–71. doi: 10.1002/art.39863
- Ni L, Lin Z, Hu S, Shi Y, Jiang Z, Zhao J, et al. Itaconate attenuates osteoarthritis by inhibiting STING/NF- κ B axis in chondrocytes and promoting M2 polarization in macrophages. *Biochem Pharmacol* (2022) 198:114935. doi: 10.1016/j.bcp.2022.114935
- Su Q, Cai Q, Li Y, Ge H, Zhang Y, Zhang Y, et al. A novel rat model of vertebral inflammation-induced intervertebral disc degeneration mediated by activating cGAS/STING molecular pathway. *J Cell Mol Med* (2021) 25:9567–85. doi: 10.1111/jcmm.16898
- Guo Q, Zhu D, Wang Y, Miao Z, Chen Z, Lin Z, et al. Targeting STING attenuates ROS induced intervertebral disc degeneration. *Osteoarthritis Cartilage* (2021) 29:1213–24. doi: 10.1016/j.joca.2021.04.017
- Tian Y, Bao Z, Ji Y, Mei X, Yang H. Epigallocatechin-3-Gallate protects H2O2-induced nucleus pulposus cell apoptosis and inflammation by inhibiting cGAS/Sting/NLRP3 activation. *Drug Des Devel Ther* (2020) 14:2113–22. doi: 10.2147/DDDT.S251623
- Wang K, Donnelly CR, Jiang C, Liao Y, Luo X, Tao X, et al. STING suppresses bone cancer pain via immune and neuronal modulation. *Nat Commun* (2021) 12:4558. doi: 10.1038/s41467-021-24867-2
- Kwon Y, Park O-J, Kim J, Cho J-H, Yun C-H, Han SH. Cyclic dinucleotides inhibit osteoclast differentiation through STING-mediated interferon- β signaling. *J Bone Miner Res* (2019) 34:1366–75. doi: 10.1002/jbmr.3701
- Ch C, Is P JP, Ky Y HJ, J K, Ys J. Transmembrane protein 173 inhibits RANKL-induced osteoclast differentiation. *FEBS Lett* (2015) 589:836–841. doi: 10.1016/j.febslet.2015.02.018
- Sun X, Xie Z, Hu B, Zhang B, Ma Y, Pan X, et al. The Nrf2 activator RTA-408 attenuates osteoclastogenesis by inhibiting STING dependent NF- κ B signaling. *Redox Biol* (2020) 28:101309. doi: 10.1016/j.redox.2019.101309
- Huang J, You H, Su C, Li Y, Chen S, Zheng C. Herpes simplex virus 1 tegument protein VP22 abrogates cGAS/STING-mediated antiviral innate immunity. *J Virol* (2018) 92:e00841–18. doi: 10.1128/JVI.00841-18
- Lahaye X, Gentili M, Silvain A, Conrad C, Picard L, Jouve M, et al. NONO detects the nuclear HIV capsid to promote cGAS-mediated innate immune activation. *Cell* (2018) 175:488–501.e22. doi: 10.1016/j.cell.2018.08.062
- Wilski NA, Stotesbury C, Del Casale C, Montoya B, Wong E, Sigal LJ, et al. STING sensing of murine cytomegalovirus alters the tumor microenvironment to promote antitumor immunity. *J Immunol* (2020) 204:2961–72. doi: 10.4049/jimmunol.1901136
- Fan YM, Zhang YL, Luo H, Mohamud Y. Crosstalk between RNA viruses and DNA sensors: Role of the cGAS-STING signalling pathway. *Rev Med Virol* (2022) 32:e2343. doi: 10.1002/rmv.2343
- Sun B, Sundström KB, Chew JJ, Bist P, Gan ES, Tan HC, et al. Dengue virus activates cGAS through the release of mitochondrial DNA. *Sci Rep* (2017) 7:3594. doi: 10.1038/s41598-017-03932-1
- Hopfner K-P, Hornung V. Molecular mechanisms and cellular functions of cGAS-STING signalling. *Nat Rev Mol Cell Biol* (2020) 21:501–21. doi: 10.1038/s41580-020-0244-x
- Kwon J, Bakhom SF. The cytosolic DNA-sensing cGAS-STING pathway in cancer. *Cancer Discovery* (2020) 10:26–39. doi: 10.1158/2159-8290.CD-19-0761
- Kato K, Omura H, Ishitani R, Nureki O. Cyclic GMP-AMP as an endogenous second messenger in innate immune signaling by cytosolic DNA. *Annu Rev Biochem* (2017) 86:541–66. doi: 10.1146/annurev-biochem-061516-044813
- Chen Q, Sun L, Chen ZJ. Regulation and function of the cGAS-STING pathway of cytosolic DNA sensing. *Nat Immunol* (2016) 17:1142–9. doi: 10.1038/ni.3558
- Liu H, Moura-Alves P, Pei G, Mollenkopf H-J, Hurwitz R, Wu X, et al. cGAS facilitates sensing of extracellular cyclic dinucleotides to activate innate immunity. *EMBO Rep* (2019) 20:e46293. doi: 10.15252/embr.201846293
- Zhang X, Shi H, Wu J, Zhang X, Sun L, Chen C, et al. Cyclic GMP-AMP containing mixed phosphodiester linkages is an endogenous high-affinity ligand for STING. *Mol Cell* (2013) 51:226–35. doi: 10.1016/j.molcel.2013.05.022
- Annibal A, Ripa R, Ballhysa E, Latza C, Hochhard N, Antebi A. Mass spectrometric characterization of cyclic dinucleotides (CDNs). *in vivo. Anal Bioanal Chem* (2021) 413:6457–68. doi: 10.1007/s00216-021-03628-6
- Jenal U, Reinders A, Lori C. Cyclic di-GMP: Second messenger extraordinaire. *Nat Rev Microbiol* (2017) 15:271–84. doi: 10.1038/nrmicro.2016.190
- Taguchi T, Mukai K, Takaya E, Shindo R. STING operation at the ER/Golgi interface. *Front Immunol* (2021) 12:646304. doi: 10.3389/fimmu.2021.646304
- Zhou R, Zhang Q, Xu P. TBK1, a central kinase in innate immune sensing of nucleic acids and beyond. *Acta Biochim Biophys Sin (Shanghai)* (2020) 52:757–67. doi: 10.1093/abbs/gmaa051
- Liu Y, Lu X, Qin N, Qiao Y, Xing S, Liu W, et al. STING, a promising target for small molecular immune modulator: A review. *Eur J Med Chem* (2021) 211:113113. doi: 10.1016/j.ejmech.2020.113113
- Dunphy G, Flannery SM, Almine JF, Connolly DJ, Paulus C, Jönsson KL, et al. Non-canonical activation of the DNA sensing adaptor STING by ATM and IFI16 mediates NF- κ B signaling after nuclear DNA damage. *Mol Cell* (2018) 71:745–760.e5. doi: 10.1016/j.molcel.2018.07.034
- Abe T, Barber GN. Cytosolic-DNA-mediated, STING-dependent proinflammatory gene induction necessitates canonical NF- κ B activation through TBK1. *J Virol* (2014) 88:5328–41. doi: 10.1128/JVI.00037-14
- Hou Y, Liang H, Rao E, Zheng W, Huang X, Deng L, et al. Non-canonical NF- κ B antagonizes STING sensor-mediated DNA sensing in radiotherapy. *Immunity* (2018) 49:490–503.e4. doi: 10.1016/j.immuni.2018.07.008
- Wiersinga WJ, Rhodes A, Cheng AC, Peacock SJ, Prescott HC. Pathophysiology, transmission, diagnosis, and treatment of coronavirus disease 2019 (COVID-19): A review. *JAMA* (2020) 324:782–93. doi: 10.1001/jama.2020.12839
- Mehta P, McAuley DF, Brown M, Sanchez E, Tattersall RS, Manson JJ. HLH across speciality collaboration, UK. COVID-19: consider cytokine storm syndromes and immunosuppression. *Lancet* (2020) 395:1033–4. doi: 10.1016/S0140-6736(20)30628-0
- Wen W, Su W, Tang H, Le W, Zhang X, Zheng Y, et al. Immune cell profiling of COVID-19 patients in the recovery stage by single-cell sequencing. *Cell Discovery* (2020) 6:31. doi: 10.1038/s41421-020-0168-9
- Neufeldt CJ, Cerikan B, Cortese M, Frankish J, Lee J-Y, Plociennikowska A, et al. SARS-CoV-2 infection induces a pro-inflammatory cytokine response through cGAS-STING and NF- κ B. *Commun Biol* (2022) 5:45. doi: 10.1038/s42003-021-02983-5
- Berthelot J-M, Lioté F. COVID-19 as a STING disorder with delayed over-secretion of interferon-beta. *EBioMedicine* (2020) 56:102801. doi: 10.1016/j.jebiom.2020.102801
- Boyce BF, Li J, Xing L, Yao Z. Bone remodeling and the role of TRAF3 in osteoclastic bone resorption. *Front Immunol* (2018) 9:2263. doi: 10.3389/fimmu.2018.02263
- Siddiqui JA, Partridge NC. Physiological bone remodeling: Systemic regulation and growth factor involvement. *Physiol (Bethesda)* (2016) 31:233–45. doi: 10.1152/physiol.00061.2014
- Kim J-M, Lin C, Stavre Z, Greenblatt MB, Shim J-H. Osteoblast-osteoclast communication and bone homeostasis. *Cells* (2020) 9:E2073. doi: 10.3390/cells9092073
- Feng X, McDonald JM. Disorders of bone remodeling. *Annu Rev Pathol* (2011) 6:121–45. doi: 10.1146/annurev-pathol-011110-130203
- Boyle WJ, Simonet WS, Lacey DL. Osteoclast differentiation and activation. *Nature* (2003) 423:337–42. doi: 10.1038/nature01658
- Wada T, Nakashima T, Hiroshi N, Penninger JM. RANKL-RANK signaling in osteoclastogenesis and bone disease. *Trends Mol Med* (2006) 12:17–25. doi: 10.1016/

j.molmed.2005.11.007

52. Agidigbi TS, Kim C. Reactive oxygen species in osteoclast differentiation and possible pharmaceutical targets of ROS-mediated osteoclast diseases. *Int J Mol Sci* (2019) 20:E3576. doi: 10.3390/ijms20143576
53. Boyce BF, Xing L. The RANKL/RANK/OPG pathway. *Curr Osteoporos Rep* (2007) 5:98–104. doi: 10.1007/s11914-007-0024-y
54. Yao Z, Getting SJ, Locke IC. Regulation of TNF-induced osteoclast differentiation. *Cells* (2021) 11:132. doi: 10.3390/cells11010132
55. Walter MR. The role of structure in the biology of interferon signaling. *Front Immunol* (2020) 11:606489. doi: 10.3389/fimmu.2020.606489
56. Peignier A, Parker D. Impact of type I interferons on susceptibility to bacterial pathogens. *Trends Microbiol* (2021) 29:823–35. doi: 10.1016/j.tim.2021.01.007
57. Zanin N, Viaris de Lesegno C, Lamaze C, Blouin CM. Interferon receptor trafficking and signaling: Journey to the cross roads. *Front Immunol* (2020) 11:615603. doi: 10.3389/fimmu.2020.615603
58. McNab F, Mayer-Barber K, Sher A, Wack A, O'Garra A. Type I interferons in infectious disease. *Nat Rev Immunol* (2015) 15:87–103. doi: 10.1038/nri3787
59. Dunn GP, Koebel CM, Schreiber RD. Interferons, immunity and cancer immunooediting. *Nat Rev Immunol* (2006) 6:836–48. doi: 10.1038/nri1961
60. Tang LSY, Covert E, Wilson E, Kottitil S. Chronic hepatitis b infection: A review. *JAMA* (2018) 319:1802–13. doi: 10.1001/jama.2018.3795
61. Stiff A, Carson W. Investigations of interferon-lambda for the treatment of cancer. *J Innate Immun* (2015) 7:243–50. doi: 10.1159/000370113
62. Michalska A, Blaszczyk K, Wesoly J, Bluysen HAR. A positive feedback amplifier circuit that regulates interferon (IFN)-stimulated gene expression and controls type I and type II IFN responses. *Front Immunol* (2018) 9:1135. doi: 10.3389/fimmu.2018.01135
63. Sunyer T, Rothe L, Kirsch D, Jiang X, Anderson F, Osdoby P, et al. Ca²⁺ or phorbol ester but not inflammatory stimuli elevate inducible nitric oxide synthase messenger ribonucleic acid and nitric oxide (NO) release in avian osteoclasts: autocrine NO mediates Ca²⁺-inhibited bone resorption. *Endocrinology* (1997) 138:2148–62. doi: 10.1210/endo.138.5.5144
64. Brandt ML, Hukkanen M, Umeda T, Moradi-Bidhendi N, Bianchi S, Gross SS, et al. Bidirectional regulation of osteoclast function by nitric oxide synthase isoforms. *Proc Natl Acad Sci U.S.A.* (1995) 92:2954–8. doi: 10.1073/pnas.92.7.2954
65. Zheng H, Yu X, Collin-Osdoby P, Osdoby P. RANKL stimulates inducible nitric oxide synthase expression and nitric oxide production in developing osteoclasts. An autocrine negative feedback mechanism triggered by RANKL-induced interferon-beta via NF-kappaB that restrains osteoclastogenesis and bone resorption. *J Biol Chem* (2006) 281:15809–20. doi: 10.1074/jbc.M513225200
66. Kwon B, Moon CH, Kang S, Seo SK, Kwon BS. 4-1BB: still in the midst of darkness. *Mol Cells* (2000) 10:119–26. doi: 10.1007/s10059-000-0119-0
67. J Y, Oj P, Yj L, Hm J, Km W YC. The 4-1BB ligand and 4-1BB expressed on osteoclast precursors enhance RANKL-induced osteoclastogenesis via bi-directional signaling. *Eur J Immunol* (2008) 38:1598–1690. doi: 10.1002/eji.200737650
68. Shin H-H, Lee E-A, Kim S-J, Kwon BS, Choi H-S. A signal through 4-1BB ligand inhibits receptor for activation of nuclear factor-kappaB ligand (RANKL)-induced osteoclastogenesis by increasing interferon (IFN)-beta production. *FEBS Lett* (2006) 580:1601–6. doi: 10.1016/j.febslet.2006.01.091
69. Sarkar SN, Peters KL, Elco CP, Sakamoto S, Pal S, Sen GC. Novel roles of TLR3 tyrosine phosphorylation and PI3 kinase in double-stranded RNA signaling. *Nat Struct Mol Biol* (2004) 11:1060–7. doi: 10.1038/nsmb847
70. Aksoy E, Vanden Berghe W, Detienne S, Amraoui Z, Fitzgerald KA, Haegeman G, et al. Inhibition of phosphoinositide 3-kinase enhances TRIF-dependent NF-kappa B activation and IFN-beta synthesis downstream of toll-like receptor 3 and 4. *Eur J Immunol* (2005) 35:2200–9. doi: 10.1002/eji.200425801
71. Ahn J, Gutman D, Saijo S, Barber GN. STING manifests self DNA-dependent inflammatory disease. *Proc Natl Acad Sci U.S.A.* (2012) 109:19386–91. doi: 10.1073/pnas.1215006109
72. Kawane K, Ohtani M, Miwa K, Kizawa T, Kanbara Y, Yoshioka Y, et al. Chronic polyarthritis caused by mammalian DNA that escapes from degradation in macrophages. *Nature* (2006) 443:998–1002. doi: 10.1038/nature05245
73. Baum R, Sharma S, Carpenter S, Li Q-Z, Busto P, Fitzgerald KA, et al. Cutting edge: AIM2 and endosomal TLRs differentially regulate arthritis and autoantibody production in DNase II-deficient mice. *J Immunol* (2015) 194:873–7. doi: 10.4049/jimmunol.1402573
74. Weilbaecher KN, Guise TA, McCauley LK. Cancer to bone: a fatal attraction. *Nat Rev Cancer* (2011) 11:411–25. doi: 10.1038/nrc3055
75. Donnelly CR, Jiang C, Andriessen AS, Wang K, Wang Z, Ding H, et al. STING controls nociception via type I interferon signalling in sensory neurons. *Nature* (2021) 591:275–80. doi: 10.1038/s41586-020-03151-1
76. Corrales L, Glickman LH, McWhirter SM, Kanne DB, Sivick KE, Katibah GE, et al. Direct activation of STING in the tumor microenvironment leads to potent and systemic tumor regression and immunity. *Cell Rep* (2015) 11:1018–30. doi: 10.1016/j.celrep.2015.04.031
77. Brodowicz T, O'Byrne K, Manegold C. Bone matters in lung cancer. *Ann Oncol* (2012) 23:2215–22. doi: 10.1093/annonc/mds009
78. Margolis SR, Wilson SC, Vance RE. Evolutionary origins of cGAS-STING signaling. *Trends Immunol* (2017) 38:733–43. doi: 10.1016/j.it.2017.03.004
79. Jin L, Hill KK, Filak H, Mogan J, Knowles H, Zhang B, et al. MPYS is required for IFN response factor 3 activation and type I IFN production in the response of cultured phagocytes to bacterial second messengers cyclic-di-AMP and cyclic-di-GMP. *J Immunol* (2011) 187:2595–601. doi: 10.4049/jimmunol.1100088
80. Xiong Q, Zhang L, Ge W, Tang P. The roles of interferons in osteoclasts and osteoclastogenesis. *Joint Bone Spine* (2016) 83:276–81. doi: 10.1016/j.jbspin.2015.07.010
81. Feng X. RANKing intracellular signaling in osteoclasts. *IUBMB Life* (2005) 57:389–95. doi: 10.1080/15216540500137669
82. Yim HY, Park C, Lee YD, Arimoto K-I, Jeon R, Baek SH, et al. Elevated response to type I IFN enhances RANKL-mediated osteoclastogenesis in Usp18-knockout mice. *J Immunol* (2016) 196:3887–95. doi: 10.4049/jimmunol.1501496
83. Basu A, Yang K, Wang M, Liu S, Chintala R, Palm T, et al. Structure-function engineering of interferon-beta-1b for improving stability, solubility, potency, immunogenicity, and pharmacokinetic properties by site-selective mono-PEGylation. *Bioconjug Chem* (2006) 17:618–30. doi: 10.1021/bc050322y
84. Weinstock-Guttman B, Hong J, Santos R, Tamaño-Blanco M, Badgett D, Patrick K, et al. Interferon-beta modulates bone-associated cytokines and osteoclast precursor activity in multiple sclerosis patients. *Mult Scler* (2006) 12:541–50. doi: 10.1177/1352458506070605
85. Hayashi T, Kaneda T, Toyama Y, Kumegawa M, Hakeda Y. Regulation of receptor activator of NF-kappa B ligand-induced osteoclastogenesis by endogenous interferon-beta (INF-beta) and suppressors of cytokine signaling (SOCS): the possible counteracting role of SOCS- in IFN-beta-inhibited osteoclast formation. *J Biol Chem* (2002) 277:27880–6. doi: 10.1074/jbc.M203836200
86. Oeckinghaus A, Hayden MS, Ghosh S. Crosstalk in NF-kB signaling pathways. *Nat Immunol* (2011) 12:695–708. doi: 10.1038/ni.2065
87. He S, Sharpless NE. Senescence in health and disease. *Cell* (2017) 169:1000–11. doi: 10.1016/j.cell.2017.05.015
88. Chung HY, Lee EK, Choi YJ, Kim JM, Kim DH, Zou Y, et al. Molecular inflammation as an underlying mechanism of the aging process and age-related diseases. *J Dent Res* (2011) 90:830–40. doi: 10.1177/0022034510387794
89. Salminen A, Huuskonen J, Ojala J, Kauppinen A, Kaarniranta K, Suuronen T. Activation of innate immunity system during aging: NF-kB signaling is the molecular culprit of inflamm-aging. *Ageing Res Rev* (2008) 7:83–105. doi: 10.1016/j.arr.2007.09.002
90. Seo E-J, Fischer N, Efferth T. Phytochemicals as inhibitors of NF-kB for treatment of alzheimer's disease. *Pharmacol Res* (2018) 129:262–73. doi: 10.1016/j.phrs.2017.11.030
91. Olefsky JM, Glass CK. Macrophages, inflammation, and insulin resistance. *Annu Rev Physiol* (2010) 72:219–46. doi: 10.1146/annurev-physiol-021909-135846
92. Zinatizadeh MR, Schock B, Chalbatani GM, Zarandi PK, Jalali SA, Miri SR, et al. (NF-kB) signaling in cancer development and immune diseases. *Genes Dis* (2021) 8:287–97. doi: 10.1016/j.gendis.2020.06.005
93. Barnabei L, Laplantine E, Mbongo W, Rieux-Laucat F, Weil R. NF-kB: At the borders of autoimmunity and inflammation. *Front Immunol* (2021) 12:716469. doi: 10.3389/fimmu.2021.716469
94. Chen Q, Tang L, Zhang Y, Wan C, Yu X, Dong Y, et al. STING up-regulates VEGF expression in oxidative stress-induced senescence of retinal pigment epithelium via NF-kB/HIF-1 α pathway. *Life Sci* (2022) 293:120089. doi: 10.1016/j.lfs.2021.120089
95. Jin M, Shiwaku H, Tanaka H, Obita T, Ohuchi S, Yoshioka Y, et al. Tau activates microglia via the PQBP1-cGAS-STING pathway to promote brain inflammation. *Nat Commun* (2021) 12:6565. doi: 10.1038/s41467-021-26851-2
96. Pan Y, You Y, Sun L, Sui Q, Liu L, Yuan H, et al. The STING antagonist h-151 ameliorates psoriasis via suppression of STING/NF-kB-mediated inflammation. *Br J Pharmacol* (2021) 178:4907–22. doi: 10.1111/bph.15673
97. Mundy GR. Osteoporosis and inflammation. *Nutr Rev* (2007) 65:S147–151. doi: 10.1111/j.1753-4887.2007.tb00353.x
98. Goldring MB, Otero M. Inflammation in osteoarthritis. *Curr Opin Rheumatol* (2011) 23:471–8. doi: 10.1097/BOR.0b013e328349c2b1
99. Lyu F-J, Cui H, Pan H, Mc Cheung K, Cao X, Iatridis JC, et al. Painful intervertebral disc degeneration and inflammation: from laboratory evidence to clinical interventions. *Bone Res* (2021) 9:7. doi: 10.1038/s41413-020-00125-x
100. Haynes DR. Bone lysis and inflammation. *Inflammation Res* (2004) 53:596–600. doi: 10.1007/s00011-004-1303-z
101. Briot K, Roux C. Inflammation, bone loss and fracture risk in spondyloarthritis. *RMD Open* (2015) 1:e000052. doi: 10.1136/rmdopen-2015-000052
102. Smith JA. STING, the endoplasmic reticulum, and mitochondria: Is there a crowd or a conversation? *Front Immunol* (2020) 11:611347. doi: 10.3389/fimmu.2020.611347
103. Saberi M, Zhang X, Mobasher A. Targeting mitochondrial dysfunction with small molecules in intervertebral disc aging and degeneration. *Geroscience* (2021) 43:517–37. doi: 10.1007/s11357-021-00341-1
104. Qadir A, Liang S, Wu Z, Chen Z, Hu L, Qian A. Senile osteoporosis: The involvement of differentiation and senescence of bone marrow stromal cells. *Int J Mol Sci*

(2020) 21:E349. doi: 10.3390/ijms21010349

105. Rea IM, Gibson DS, McGilligan V, McNeerlan SE, Alexander HD, Ross OA. Age and age-related diseases: Role of inflammation triggers and cytokines. *Front Immunol* (2018) 9:586. doi: 10.3389/fimmu.2018.00586

106. Sharma P, Sampath H. Mitochondrial DNA integrity: Role in health and disease. *Cells* (2019) 8:100. doi: 10.3390/cells8020100

107. Liu Z, Wang M, Wang X, Bu Q, Wang Q, Su W, et al. XBP1 deficiency promotes hepatocyte pyroptosis by impairing mitophagy to activate mtDNA-cGAS-STING signaling in macrophages during acute liver injury. *Redox Biol* (2022) 52:102305. doi: 10.1016/j.redox.2022.102305

108. Zhong W, Rao Z, Rao J, Han G, Wang P, Jiang T, et al. Aging aggravated liver ischemia and reperfusion injury by promoting STING-mediated NLRP3 activation in macrophages. *Aging Cell* (2020) 19:e13186. doi: 10.1111/ace1.13186

109. Zhang D, Liu Y, Zhu Y, Zhang Q, Guan H, Liu S, et al. A non-canonical cGAS-STING-PERK pathway facilitates the translational program critical for senescence and organ fibrosis. *Nat Cell Biol* (2022) 24:766–82. doi: 10.1038/s41556-022-00894-z

110. Huxford T, Hoffmann A, Ghosh G. Understanding the logic of I κ B: NF- κ B signaling in structural terms. *Curr Top Microbiol Immunol* (2011) 349:1–24. doi: 10.1007/82_2010_99

111. Hayden MS, Ghosh S. Shared principles in NF- κ B signaling. *Cell* (2008) 132:344–62. doi: 10.1016/j.cell.2008.01.020

112. Ghosh S, May MJ, Kopp EB. NF- κ B and rel proteins: Evolutionarily conserved mediators of immune responses. *Annu Rev Immunol* (1998) 16:225–60. doi: 10.1146/annurev.immunol.16.1.225

113. Courtis G, Gilmore TD. Mutations in the NF- κ B signaling pathway: implications for human disease. *Oncogene* (2006) 25:6831–43. doi: 10.1038/sj.onc.1209939

114. Leibowitz SM, Yan J. NF- κ B pathways in the pathogenesis of multiple sclerosis and the therapeutic implications. *Front Mol Neurosci* (2016) 9:84. doi: 10.3389/fnmol.2016.00084

115. Napetschnig J, Wu H. Molecular basis of NF- κ B signaling. *Annu Rev Biophys* (2013) 42:443–68. doi: 10.1146/annurev-biophys-083012-130338

116. Mercurio F, Zhu H, Murray BW, Shevchenko A, Bennett BL, Li J, et al. IKK-1 and IKK-2: cytokine-activated I κ B kinase essential for NF- κ B activation. *Science* (1997) 278:860–6. doi: 10.1126/science.278.5339.860

117. Ghosh S, Karin M. Missing pieces in the NF- κ B puzzle. *Cell* (2002) 109 Suppl:S81–96. doi: 10.1016/s0092-8674(02)00703-1

118. Hadjidakis DJ, Androulakis II. Bone remodeling. *Ann New York Acad Sci* (2006) 1092:385–96. doi: 10.1196/annals.1365.035

119. Lawrence T. The nuclear factor NF- κ B pathway in inflammation. *Cold Spring Harb Perspect Biol* (2009) 1:a001651. doi: 10.1101/cshperspect.a001651

120. Adamopoulos IE. Inflammation in bone physiology and pathology. *Curr Opin Rheumatol* (2018) 30:59–64. doi: 10.1097/BOR.0000000000000449

121. Theoleyre S, Wittrant Y, Tat SK, Fortun Y, Redini F, Heymann D. The molecular triad OPG/RANK/RANKL: involvement in the orchestration of pathophysiological bone remodeling. *Cytokine Growth Factor Rev* (2004) 15:457–75. doi: 10.1016/j.cytogfr.2004.06.004

122. Cheng T, Zhang X. NF κ B gene silencing inhibits wear particles-induced inflammatory osteolysis. *Med Hypotheses* (2008) 71:727–9. doi: 10.1016/j.mehy.2008.07.003

123. Wang T, He C. Pro-inflammatory cytokines: The link between obesity and osteoarthritis. *Cytokine Growth Factor Rev* (2018) 44:38–50. doi: 10.1016/j.cytogfr.2018.10.002

124. Fujita T, Matsui T, Nakao Y, Shiozawa S, Imai Y. Cytokines and osteoporosis. *Ann N Y Acad Sci* (1990) 587:371–5. doi: 10.1111/j.1749-6632.1990.tb00178.x

125. Hegde R, Awan KH. Effects of periodontal disease on systemic health. *Dis Mon* (2019) 65:185–92. doi: 10.1016/j.disamonth.2018.09.011

126. Krum SA, Chang J, Miranda-Carboni G, Wang C-Y. Novel functions for NF κ B: inhibition of bone formation. *Nat Rev Rheumatol* (2010) 6:607–11. doi: 10.1038/nrrheum.2010.133

127. Asagiri M, Takayanagi H. The molecular understanding of osteoclast differentiation. *Bone* (2007) 40:251–64. doi: 10.1016/j.bone.2006.09.023

128. Boyce BF, Xing L. Functions of RANKL/RANK/OPG in bone modeling and remodeling. *Arch Biochem Biophys* (2008) 473:139–46. doi: 10.1016/j.abb.2008.03.018

129. Yamamoto M, Gohda Y, Akiyama T, Inoue J-I. TNF receptor-associated factor 6 (TRAF6) plays crucial roles in multiple biological systems through polyubiquitination-mediated NF- κ B activation. *Proc Jpn Acad Ser B Phys Biol Sci* (2021) 97:145–60. doi: 10.2183/pjab.97.009

130. Darnay BG, Ni J, Moore PA, Aggarwal BB. Activation of NF- κ B by RANK requires tumor necrosis factor receptor-associated factor (TRAF) 6 and NF- κ B-inducing kinase. *Identification novel TRAF6 interaction motif. J Biol Chem* (1999) 274:7724–31. doi: 10.1074/jbc.274.12.7724

131. Iotsova V, Caamaño J, Loy J, Yang Y, Lewin A, Bravo R. Osteopetrosis in mice lacking NF- κ B1 and NF- κ B2. *Nat Med* (1997) 3:1285–9. doi: 10.1038/nm1197-1285

132. Lomaga MA, Yeh WC, Sarosi I, Duncan GS, Furlonger C, Ho A, et al. TRAF6 deficiency results in osteopetrosis and defective interleukin-1, CD40, and LPS signaling. *Genes Dev* (1999) 13:1015–24. doi: 10.1101/gad.13.8.1015

133. Ruocco MG, Karin M. IKK[β] as a target for treatment of inflammation induced bone loss. *Ann Rheum Dis* (2005) 64 Suppl 4:iv81–85. doi: 10.1136/ard.2005.042721

134. Ruocco MG, Maeda S, Park JM, Lawrence T, Hsu L-C, Cao Y, et al. I κ B kinase (IKK)[β], but not IKK[α], is a critical mediator of osteoclast survival and is required for inflammation-induced bone loss. *J Exp Med* (2005) 201:1677–87. doi: 10.1084/jem.20042081

135. Ruocco MG, Karin M. Control of osteoclast activity and bone loss by IKK subunits: new targets for therapy. *Adv Exp Med Biol* (2007) 602:125–34. doi: 10.1007/978-0-387-72009-8_16

136. Boyce BF, Yao Z, Xing L. Functions of nuclear factor κ B in bone. *Ann N Y Acad Sci* (2010) 1192:367–75. doi: 10.1111/j.1749-6632.2009.05315.x

137. Lee S-Y, Lee K-S, Yi SH, Kook S-H, Lee J-C. Acteoside suppresses RANKL-mediated osteoclastogenesis by inhibiting c-fos induction and NF- κ B pathway and attenuating ROS production. *PLoS One* (2013) 8:e80873. doi: 10.1371/journal.pone.0080873

138. Yu B, Chang J, Liu Y, Li J, Kevork K, Al-Hezaimi K. Wnt4 signaling prevents skeletal aging and inflammation by inhibiting nuclear factor- κ B. *Nat Med* (2014) 20:1009–1017. doi: 10.1038/nm.3586

139. Lin T-H, Gibon E, Loi F, Pajarinen J, Córdova LA, Nabeshima A, et al. Decreased osteogenesis in mesenchymal stem cells derived from the aged mouse is associated with enhanced NF- κ B activity. *J Orthop Res* (2017) 35:281–8. doi: 10.1002/jor.23270

140. Lin T, Tamaki Y, Pajarinen J, Waters HA, Woo DK, Yao Z, et al. Chronic inflammation in biomaterial-induced periprosthetic osteolysis: NF- κ B as a therapeutic target. *Acta Biomater* (2014) 10:1–10. doi: 10.1016/j.actbio.2013.09.034

141. Liu J, Lin A. Role of JNK activation in apoptosis: a double-edged sword. *Cell Res* (2005) 15:36–42. doi: 10.1038/sj.cr.7290262

142. Jimi E, Takakura N, Hiura F, Nakamura I, Hirata-Tsuchiya S. The role of NF- κ B in physiological bone development and inflammatory bone diseases: Is NF- κ B inhibition “Killing two birds with one stone”? *Cells* (2019) 8:1636. doi: 10.3390/cells8121636

143. Eferl R, Hoebertz A, Schilling AF, Rath M, Karreth F, Kenner L, et al. The fos-related antigen fra-1 is an activator of bone matrix formation. *EMBO J* (2004) 23:2789–99. doi: 10.1038/sj.emboj.7600282

144. Gilbert L, He X, Farmer P, Boden S, Kozlowski M, Rubin J, et al. Inhibition of osteoblast differentiation by tumor necrosis factor- α . *Endocrinology* (2000) 141:3956–64. doi: 10.1210/endo.141.11.7739

145. Yamazaki M, Fukushima H, Shin M, Katagiri T, Takahashi D, Takahashi T, et al. Tumor necrosis factor α represses bone morphogenetic protein (BMP) signaling by interfering with the DNA binding of smads through the activation of NF- κ B. *J Biol Chem* (2009) 284:35987–35995. doi: 10.1074/jbc.M109.070540

146. Chang J, Wang Z, Tang E, Fan Z, McCauley L, Franceschi R, et al. Inhibition of osteoblastic bone formation by nuclear factor- κ B. *Nat Med* (2009) 15:682–9. doi: 10.1038/nm.1954

147. Alles N, Soysa NS, Hayashi J, Khan M, Shimoda A, Shimokawa H, et al. Suppression of NF- κ B increases bone formation and ameliorates osteopenia in ovariectomized mice. *Endocrinology* (2010) 151:4626–34. doi: 10.1210/en.2010-0399

148. Quan Y, Xin Y, Tian G, Zhou J, Liu X. Mitochondrial ROS-modulated mtDNA: A potential target for cardiac aging. *Oxid Med Cell Longev* (2020) 2020:9423593. doi: 10.1155/2020/9423593

149. Gloire G, Legrand-Poels S, Piette J. NF- κ B activation by reactive oxygen species: fifteen years later. *Biochem Pharmacol* (2006) 72:1493–505. doi: 10.1016/j.bcp.2006.04.011

150. Burdette DL, Monroe KM, Sotelo-Troha K, Iwig JS, Eckert B, Hyodo M, et al. STING is a direct innate immune sensor of cyclic di-GMP. *Nature* (2011) 478:515–8. doi: 10.1038/nature10429

151. Balka KR, Louis C, Saunders TL, Smith AM, Calleja DJ, D’Silva DB, et al. TBK1 and IKK ϵ act redundantly to mediate STING-induced NF- κ B responses in myeloid cells. *Cell Rep* (2020) 31:107492. doi: 10.1016/j.celrep.2020.03.056

152. Sun Y-X, Xu A-H, Yang Y, Li J. Role of Nrf2 in bone metabolism. *J BioMed Sci* (2015) 22:101. doi: 10.1186/s12929-015-0212-5

153. Olgagnier D, Brandt AM, Gunderstoft C, Villadsen NL, Krapp C, Thielke AL, et al. Nrf2 negatively regulates STING indicating a link between antiviral sensing and metabolic reprogramming. *Nat Commun* (2018) 9:3506. doi: 10.1038/s41467-018-05861-7

154. Zhang J, Pan J, Jing W. Motivating role of type h vessels in bone regeneration. *Cell Prolif* (2020) 53:e12874. doi: 10.1111/cpr.12874

155. Peng Y, Wu S, Li Y, Crane JL. Type h blood vessels in bone modeling and remodeling. *Theranostics* (2020) 10:426–36. doi: 10.7150/thno.34126

156. Yang Y-Q, Tan Y-Y, Wong R, Wenden A, Zhang L-K, Rabie ABM. The role of vascular endothelial growth factor in ossification. *Int J Oral Sci* (2012) 4:64–8. doi: 10.1038/ijos.2012.33

157. Grosso A, Burger MG, Lunger A, Schaefer DJ, Banfi A, Di Maggio N. It takes two to tango: Coupling of angiogenesis and osteogenesis for bone regeneration. *Front Bioeng*

Biotechnol (2017) 5:68. doi: 10.3389/fbioe.2017.00068

158. Zhou X, Kunkel G, Zhang Z, Deng JM, Behringer RR, et al. The novel zinc finger-containing transcription factor osterix is required for osteoblast differentiation and bone formation. *Cell* (2002) 108:17–29. doi: 10.1016/s0092-8674(01)00622-5
159. Farr JN, Khosla S. Cellular senescence in bone. *Bone* (2019) 121:121–33. doi: 10.1016/j.bone.2019.01.015
160. Zhu Y, Ruan Z, Lin Z, Long H, Zhao R, Sun B, et al. The association between CD31hiEmcnhi endothelial cells and bone mineral density in Chinese women. *J Bone Miner Metab* (2019) 37:987–95. doi: 10.1007/s00774-019-01000-4
161. Wang L, Zhou F, Zhang P, Wang H, Qu Z, Jia P, et al. Human type h vessels are a sensitive biomarker of bone mass. *Cell Death Dis* (2017) 8:e2760. doi: 10.1038/cddis.2017.36
162. Wang H, Yin Y, Li W, Zhao X, Yu Y, Zhu J, et al. Over-expression of PDGFR- β promotes PDGF-induced proliferation, migration, and angiogenesis of EPCs through PI3K/Akt signaling pathway. *PLoS One* (2012) 7:e30503. doi: 10.1371/journal.pone.0030503
163. Ball SG, Shuttleworth CA, Kielty CM. Mesenchymal stem cells and neovascularization: role of platelet-derived growth factor receptors. *J Cell Mol Med* (2007) 11:1012–30. doi: 10.1111/j.1582-4934.2007.00120.x
164. Andrae J, Gallini R, Betsholtz C. Role of platelet-derived growth factors in physiology and medicine. *Genes Dev* (2008) 22:1276–312. doi: 10.1101/gad.1653708
165. Xie H, Cui Z, Wang L, Xia Z, Hu Y, Xian L, et al. PDGF-BB secreted by preosteoclasts induces angiogenesis during coupling with osteogenesis. *Nat Med* (2014) 20:1270–8. doi: 10.1038/nm.3668
166. Huang J, Yin H, Rao S-S, Xie P-L, Cao X, Rao T, et al. Harmine enhances type h vessel formation and prevents bone loss in ovariectomized mice. *Theranostics* (2018) 8:2435–46. doi: 10.7150/thno.22144
167. Jiang Y, Liu C, Chen W, Wang H, Wang C, Lin N. Tetramethylpyrazine enhances vascularization and prevents osteonecrosis in steroid-treated rats. *BioMed Res Int* (2015) 2015:315850. doi: 10.1155/2015/315850
168. Pufe T, Scholz-Ahrens KE, Franke ATM, Petersen W, Mentlein R, Varoga D, et al. The role of vascular endothelial growth factor in glucocorticoid-induced bone loss: evaluation in a minipig model. *Bone* (2003) 33:869–76. doi: 10.1016/j.bone.2003.08.002
169. Yang P, Lv S, Wang Y, Peng Y, Ye Z, Xia Z, et al. Preservation of type h vessels and osteoblasts by enhanced preosteoclast platelet-derived growth factor type BB attenuates glucocorticoid-induced osteoporosis in growing mice. *Bone* (2018) 114:1–13. doi: 10.1016/j.bone.2018.05.025
170. Pugh CW, Ratcliffe PJ. Regulation of angiogenesis by hypoxia: Role of the HIF system. *Nat Med* (2003) 9:677–84. doi: 10.1038/nm0603-677
171. Kusumbe AP, Ramasamy SK, Adams RH. Coupling of angiogenesis and osteogenesis by a specific vessel subtype in bone. *Nature* (2014) 507:323–8. doi: 10.1038/nature13145
172. Riddle RC, Khatri R, Schipani E, Clemens TL. Role of hypoxia-inducible factor-1 α in angiogenic-osteogenic coupling. *J Mol Med (Berl)* (2009) 87:583–90. doi: 10.1007/s00109-009-0477-9
173. Gao B, Lin X, Jing H, Fan J, Ji C, Jie Q, et al. Local delivery of tetramethylpyrazine eliminates the senescent phenotype of bone marrow mesenchymal stromal cells and creates an anti-inflammatory and angiogenic environment in aging mice. *Aging Cell* (2018) 17:e12741. doi: 10.1111/acel.12741
174. Demaria O, De Gassart A, Coso S, Gestermann N, Di Domizio J, Flatz L, et al. STING activation of tumor endothelial cells initiates spontaneous and therapeutic antitumor immunity. *Proc Natl Acad Sci U.S.A.* (2015) 112:15408–13. doi: 10.1073/pnas.1512832112
175. Baris AM, Fraile-Bethencourt E, Anand S. Nucleic acid sensing in the tumor vasculature. *Cancers (Basel)* (2021) 13:4452. doi: 10.3390/cancers13174452
176. Liu Y, Jesus AA, Marrero B, Yang D, Ramsey SE, Sanchez GAM, et al. Activated STING in a vascular and pulmonary syndrome. *N Engl J Med* (2014) 371:507–18. doi: 10.1056/NEJMoa1312625
177. Crow YJ, Casanova J-L. STING-associated vasculopathy with onset in infancy—a new interferonopathy. *N Engl J Med* (2014) 371:568–71. doi: 10.1056/NEJMe1407246
178. Jiang X, Liu G, Hu Z, Chen G, Chen J, Lv Z. cGAMP inhibits tumor growth in colorectal cancer metastasis through the STING/STAT3 axis in a zebrafish xenograft model. *Fish Shellfish Immunol* (2019) 95:220–6. doi: 10.1016/j.fsi.2019.09.075
179. Yuan L, Mao Y, Luo W, Wu W, Xu H, Wang XL, et al. Palmitic acid dysregulates the hippo-YAP pathway and inhibits angiogenesis by inducing mitochondrial damage and activating the cytosolic DNA sensor cGAS-STING-IRF3 signaling mechanism. *J Biol Chem* (2017) 292:15002–15. doi: 10.1074/jbc.M117.804005
180. Mao Y, Luo W, Zhang L, Wu W, Yuan L, Xu H, et al. STING-IRF3 triggers endothelial inflammation in response to free fatty acid-induced mitochondrial damage in diet-induced obesity. *Arterioscler Thromb Vasc Biol* (2017) 37:920–9. doi: 10.1161/ATVBAHA.117.309017
181. Guo Y, Gu R, Gan D, Hu F, Li G, Xu G. Mitochondrial DNA drives noncanonical inflammation activation via cGAS-STING signaling pathway in retinal microvascular endothelial cells. *Cell Commun Signal* (2020) 18:172. doi: 10.1186/s12964-020-00637-3
182. Huang J, Han Q, Cai M, Zhu J, Li L, Yu L, et al. Effect of angiogenesis in bone tissue engineering. *Ann BioMed Eng* (2022) 50:898–913. doi: 10.1007/s10439-022-02970-9