#### Check for updates

#### **OPEN ACCESS**

EDITED BY Maria Sole Chimenti, Rheumatology, Allergology and Clinical Immunology University of Rome Tor Vergata, Italy

REVIEWED BY Patricia Bozzetto Ambrosi, Université Paris Cité, France Runze Qiu, Nanjing Medical University, China

\*CORRESPONDENCE Chao Zhao ostuald78@qfnu.edu.cn

SPECIALTY SECTION This article was submitted to Autoimmune and Autoinflammatory Disorders: Autoinflammatory Disorders, a section of the journal Frontiers in Immunology

RECEIVED 17 September 2022 ACCEPTED 28 October 2022 PUBLISHED 14 November 2022

#### CITATION

Wu A, Zhao C, Mou S, Li S, Cui X and Zhang R (2022) Integrated analysis identifies the IL6/JAK/STAT signaling pathway and the estrogen response pathway associated with the pathogenesis of intracranial aneurysms. *Front. Immunol.* 13:1046765. doi: 10.3389/fimmu.2022.1046765

#### COPYRIGHT

© 2022 Wu, Zhao, Mou, Li, Cui and Zhang. This is an open-access article distributed under the terms of the Creative Commons Attribution License

(CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms. Integrated analysis identifies the IL6/JAK/STAT signaling pathway and the estrogen response pathway associated with the pathogenesis of intracranial aneurysms

Aihong Wu<sup>1</sup>, Chao Zhao<sup>2,3</sup>\*, Shanling Mou<sup>4</sup>, Shengjun Li<sup>3</sup>, Xinchun Cui<sup>3</sup> and Ronghua Zhang<sup>4</sup>

<sup>1</sup>Library, Qufu Normal University, Rizhao, Shandong, China, <sup>2</sup>Department of Neurosurgery, The Affiliated Rizhao People's Hospital of Jining Medical University, Rizhao, Shandong, China, <sup>3</sup>School of Computer Science, Qufu Normal University, Rizhao, Shandong, China, <sup>4</sup>Department of Laboratory, The Affiliated Rizhao People's Hospital of Jining Medical University, Rizhao, Shandong, China

**Objective:** We intended to identify the potential key biomarker and pathways that correlated with infiltrating immune cells during the pathogenesis of intracranial aneurysms (IA), to develop a diagnostic model, and to predict therapeutic drugs.

**Methods:** Three datasets containing intracranial aneurysm tissue samples and normal artery control samples from Gene Expression Omnibus (GEO) were included. Gene-set variation analysis(GSVA) and gene set enrichment analysis (GSEA) were conducted to find the significant differentially expressed pathways in IA formation. The least absolute shrinkage and selection operator (LASSO) regression and the multivariate logistic regression analysis were performed to identify the characteristic genes in the IL6/JAK/STAT signaling pathway (ISP) and the estrogen response pathway (ERP). A diagnostic model was constructed. xCell was used to identify immune cell types in IA pathogenesis. We used the weighted gene co-expression network analysis (WGCNA) algorithm to explore the correlations between the key modules and the four traits. Potential therapeutic drugs were investigated in Enrichr and Drugbank database.

**Results:** The ISP is significant positively correlated with IA onset. The biological function of the ISP is positively correlated with that of the ERP, and is significantly associated with immune cells activities. *CSF2RB, FAS, IL6, PTPN1, STAT2, TGFB1* of the ISP gene set and *ALDH3A2, COX6C, IGSF1, KRT18, MICB, NPY1R* of the ERP gene set were proved to be the characteristic genes. The *STAT2* gene can be the potential biomarker of IA onset. The immune score of IA samples was significantly higher than the controls. The *STAT2* gene expression is associated with infiltration of immune cells. The WGCNA results were

consistent with our finds. Acetaminophen can be a potential therapeutic drug for IA targeting *STAT2*.

**Conclusions:** We identified that the ISP was one of the most significant positively correlated pathways in IA onset, and it was activated in this process concordant with the ERP and immune responses. Except for beneficial effects, complex and multiple roles of estrogen may be involved in IA formation. STAT2 could be a potential biomarker and a promising therapeutic target of IA pathogenesis.

KEYWORDS

intracranial aneurysm, GSVA, IL6/JAK/STAT signaling pathway, estrogen, immune microenviroment, WGCNA

## Introduction

Intracranial aneurysm (IA) has been a life-threatening disease by now. According to angiographic studies, the prevalence of intracranial aneurysms in the general population ranges from 2.0 to 6.0 percent (1, 2). The mortality after IA rupture is about 30%, with a disability rate of 36.7% in surviving patients (3). The current treatment options for IA, endovascular coiling and surgical clipping, are invasive and somewhat risky. The molecular mechanisms of pathogenesis of IAs remain poorly elucidated. Noninvasive treatments have not been identified and implemented in clinical practice. Clarification of the mechanisms may contribute to the early diagnosis of IA and the development of potential safe and effective drug treatments.

The clinical data reveal that familial history of IA, high blood pressure, cigarette smoking, alcohol consumption, and female sex have been identified as risk factors of IA formation (4). Females have a significantly higher incidence of IA than males, and it is even higher in familial cases of IA (5–7), whereas the majority of risk factors, such as cigarette smoking, hypertension, atherosclerosis, and alcohol consumption, are more prevalent in men (8–10). This evidence implies that female sex steroids such as estrogen may play an important role in IA onset.

The recruitment and infiltration of immune cells have been identified as a critical stage in the occurrence of IA (11). For example, infiltrating macrophage can impair intracranial arterial wall integrity and contribute to IA formation (12). The tumor necrosis factor  $\alpha$  (TNF- $\alpha$ )/NF- $\kappa$ B pathway may be involved in the development of IAs (13).

In this study, we sought to find the most significant differentially expressed pathway (DEP) that was associated with IA formation *via* the gene set variation analysis (GSVA) algorithm, and explored its correlation with the estrogen response pathway. We discovered the characteristic genes in these two pathways and the infiltration pattern of immune cells. A diagnostic model was constructed to distinguish IAs from controls using bioinformatics analysis. Potential drugs of IA were also detected.

The workflow chart is Figure 1.

## Materials and methods

#### Gene expression datasets and data processing

Three transcriptome-wide gene expression profiles of GSE122897, GSE75436 and GSE15629, were downloaded from Gene Expression Omnibus (GEO) (14). The RNA sequencing dataset GSE122897 contains 44 IA tissue samples and 16 control tissue samples (cerebral arteries), which was based on the GPL16791 (Illumina HiSeq 2500 Homo sapiens) platform. Two IA samples in GSE122897, GSM3487858 and GSM3487892, were rejected as their statuses were "unknown". As an outlier, GSM3487895 was also omitted according to our previous research (15). A total of 41 IA samples and 16 control samples were included in the analysis. Raw read counts profile of GSE122897 was processed by "DESeq2" package for library normalization and independent filtering.

The microarray datasets GSE75436 included data from 15 IA tissues and 15 matched superficial temporal artery tissues of

Abbreviations: IA, Intracranial Aneurysm; DEP, differentially expressed pathway; GSVA, Gene Set Variation Analysis; GSEA, Gene Set Enrichment Analysis; ISP, IL6/JAK/STAT Signaling Pathway; ERP, Estrogen Response Pathway; GO, Gene Ontology; BP, Biological Process; MF, Molecular Function; KEGG, Kyoto Encyclopedia of Genes and Genomes; LR, LASSO Regression; MLR, Multivariate Logistic Regression; ROC, Receiver Operator Characteristic; WGCNA, Weighted Gene Co-expression Network Analysis; NSAID, Nonsteroidal Anti-inflammatory Drug.



them and served as independently external validation. The microarray dataset GSE15629 provided a gene expression profile of 14 IA samples (8 ruptured and 6 unruptured) and 5 controls (middle meningeal arteries). The ruptured and unruptured IA samples were combined as IA tissue samples. All data used in the study were obtained from the GEO, and hence, ethics approval and informed consent were not required.

# Identification of the associated pathways with IA formation

We downloaded HALLMARK gene sets from the MSigDB database (http://www.gsea-msigdb.org/gsea/msigdb/index.jsp) to perform the GSVA algorithm using the R package "GSEABase". The "h.all.v7.5.1.symbols.gmt" profile was used

for this analysis. To reveal the enriched pathways within 50 HALLMARK gene sets, we used the "GSVA" package to evaluate enrichment score and assign pathway activity conditions between control and IA samples in GSE122897, GSE75436 and GSE15629 with a Gaussian distribution. Then, the limma package was also used to find the DEPs. Adjusted p value< 0.05 and |logFC|>0.3 were set as criteria. Gene set enrichment analysis (GSEA) was used to analyze the selected pathway activity in GSE122897 and GSE75436.

## Functional correlation analysis

The IL6/JAK/STAT signaling pathway gene set (M5897) and the estrogen response pathway gene sets (M5906 and M5907) were downloaded from the MSigDB database. The GSVA scores were used to investigate the relationship between the IL6/JAK/ STAT signaling pathway (ISP) geneset and the estrogen response pathway (ERP) geneset based on expression profile data of GSE122897 and GSE75436. Gene Ontology (GO) analysis was performed by R package "clusterProfiler". The significantly different Biological Process (BP) and Molecular function (MF) terms were screened with the criterion adjusted p value <0.05. The results were visualized by R package "enrichplot" and "ggplot2".

# Selection of characteristic genes in ISP and ERP

The least absolute shrinkage and selection operator (LASSO) regression was performed to identify the characteristic genes with the highest IA predictive values of ISP and ERP based on expression profile data of GSE122897. Moreover, the predictive ability was further evaluated by multivariate logistic regression analysis with "rms" package using GSE122897 dataset.

### Validation of predictive models

The receiver operator characteristic (ROC) analyses were used to evaluate the capability of the characteristic genes to distinguish IA patients with controls using the "pROC" package based on GSE75436 dataset. The ROC curves were plotted by the "ggplot2" package, and the area under the curve (AUC) was also calculated. The calibration curve was conducted to analyze the accuracy of the predictive model during IA occurrence. Nomogram was established to illustrate the results of multivariate logistic analyses for the characteristic genes in ISP using the "rms" package.

# Correlation analysis between characteristic genes

Spearman correlation analysis was performed to explore the correlation between *STAT2* and the characteristic genes of the estrogen response pathway in GSE122897 and GSE75436.

# Correlation analysis between biomarker and immune cell infiltrations

xCell (16) was implemented to analyze the infiltration levels of immune cells on the basis of the expression profiling of GSE122897, and portrayals of cellular heterogeneity landscape for IA tissue expression profiles were acquired. Cutoff for significance were p value < 0.05. Correlation matrix of immune cells and the characteristic genes in ISP were constructed by The SangerBox 3.0 (http://vip.sangerbox.com/home.html).

### Construction of weighted gene coexpression network

We used the whole gene transcriptome expression profile of GSE122897 to perform weighted gene co-expression network analysis (WGCNA). Topology network of WGCNA was generated by "WGCNA" R package (17). To transform the adjacency matrix to a topological overlap matrix, the soft-threshold power ( $\beta$ ) was eight when 0.85 was chosen as the correlation coefficient threshold with a scale-free R<sup>2</sup> above 0.85 and a slope near 1. The minimum number of genes in modules was 80, and the cut height threshold was 0.3 for module detection. We produced a module-trait relationships chart and searched for the key modules that were closely related to group (IA or controls), immuneScore, ERP and ISP. The correlations among the eigengenes of the modules and the four traits were used to identify key modules.

We obtained the module significance (MS) resulted from calculating the average absolute gene significance (GS) of all the genes involved in the module. In the WGCNA, a module with high correlation score and p value <0.05 among all modules is defined as the key one and employed for further analysis.

### Functional enrichment analysis

Genes in the key module of group indicated a significant correlation with traits of IA occurrence. Hub genes in WGCNA were defined as those with a gene significance (|GS|) >0.4 and a module membership (|MM|) >0.8. Hub genes were selected to perform GO and Kyoto Encyclopedia of Genes and Genomes (KEGG) analyses. GO and KEGG pathway analyses were conducted to explore their biological functions utilizing the R package " clusterProfiler " (18). GO terms and KEGG pathways were explored with adjusted *p* value <0.05 were set as the cutoff and were visualized by the R package "GOplot" (19).

### Screening potential therapeutic drugs

The Enrichr (https://amp.pharm.mssm.edu/Enrichr/) and Drugbank platform (https://go.drugbank.com/) were used for drug screening of STAT2. Access of the DSigDB database was acquired using Enrichr. Potential therapeutic agents were determined by the adj. *p* values and the abundance of acting on STAT2 and co-expression genes. All experimentally validated drug-target information is displayed in Drugbank. We selected the drugs that can downregulate the expression of *STAT2* mRNA in it.

### Statistical analysis

The Wilcoxon test was applied to compare the difference of continuous variables between the two groups. All analyses were conducted using R4.1.0. software. The p value or adjusted p value< 0.05 were considered statistically significant. R packages of limma, WGCNA, ggplot2, export, clusterProfiler, rms, GSEABase, GSVA, ROCR, etc. were used in this study.

## Results

#### Associated pathways with IA formation

The GSVA was conducted to analyze the significant DEPs between IAs and controls. The geneset with logFC >0.3 or logFC<-0.3 was defined accordingly as an upregulated or downregulated geneset of the three datasets (Supplementary Table 1). Three common pathways were significant positively correlated with IA onset according to GSVA results, which are HALLMARK\_ALLOGRAFT\_REJECTION, HALLMARK\_ INFLAMMATORY\_RESPONSE and HALLMARK\_IL6\_JAK\_ STAT3\_SIGNALING. We selected the IL6/JAK/STAT signaling geneset in our research as it has been mentioned in our previous work (15). In GSE122897 and GSE75436, the ISP is activated by GSEA. (Figures 2A, B)

### Functional correlation analysis

The GSVA is also performed to quantify the ISP and ERP scores of all samples. The ISP is positively correlated with the ERP in GSE122897 and GSE75436, as is shown in Figures 2C, D. To explore the physiological and pathological function of ISP, GO BP and GO MF enrichment analysis were performed with the GSE122897 dataset. The results manifested the ISP were mainly related to biological processes of immune cells, such as T cell activation, leukocyte adhesion, response to molecule of bacterial origin, response to biotic stimulus, regulation of response to cytokine stimulus, regulation of cytokine-mediated signaling pathway; molecular functions of immune response, such as cytokine and immune receptor activity, cytokine and its receptor binding, cytokine and chemokine activity, chemokine and CXCR chemokine receptor binding. (Figures 2E, F). These findings show that the function of ISP is significantly associated with immune cells activities.

# Selection of characteristic genes in ISP and ERP

In order to investigate the characteristic genes of the ISP and the ERP in IA formation, we obtained 87 genes of ISP and 299 genes of ERP from MSidDB. These two gene sets were used for LASSO regression with gene expression data of GSE122897 to select the valuable predictive genes.

Firstly, we retained 6 genes within ISP with lambda = 0.0681, including *CSF2RB*, *FAS*, *IL6*, *PTPN1*, *STAT2*, *TGFB1* (Figures 3A, B). Multivariate logistic regression was performed to further identify the pivotal genes to simplify the model. The *p* value of *STAT2* was <0.05 in the logistic regression model, implying that *STAT2* has a greater contribution to the model than the other genes and might be an independent risk factor of IA occurrence (Supplementary Table 2). The expression levels of *STAT2* were significantly increased in IA samples compared with control samples in GSE122897 and GSE75436 (Supplementary Image 1A, 1B). Therefore, we constructed a diagnostic model based on *STAT2*.

Next, we obtained 7 genes within ERP with lambda = 0.124, including *ALDH3A2*, *COX6C*, *GAB2*, *IGSF1*, *KRT18*, *MICB*, *NPY1R* (Figures 3C, D). Multivariate logistic regression was also performed; whereas, the *p* value of each gene was >0.05 in this logistic regression model (Supplementary Table 3). The expression level of *GAB2* was not significantly different between IAs and controls in GSE75436 and it was omitted in this model. The expression patterns of the other six characteristic genes were the same in GSE122897 as in GSE75436 (Supplementary Image 1C, 1D).

# Diagnostic efficacy and validation of the diagnostic model of IA formation

Nomogram based on the characteristic genes of ISP was constructed as diagnostic tools for IA formation. In the nomogram, each characteristic gene corresponded to a score, and the total score was calculated by adding the scores for all characteristic genes. The total points corresponded to different risks of IA onset. (Figure 3G)

The ROC curves were plotted and the AUC was calculated to distinguish IA samples from controls. The AUC of *STAT2* were greater than 0.8 in GSE75436 (Figure 3E). The AUCs of *ALDH3A2*, *COX6C*, *IGSF1*, *KRT18*, *MICB*, *NPY1R* (Supplementary Image 2A) and the combination of them (Supplementary Image 2B) were greater than 0.7 in GSE75436. These results indicated a moderate diagnostic efficacy of these genes as new biomarkers.

The calibration curve demonstrated that the predictive model of *STAT2* enabled an accurate estimation during the progression of IA occurrence (Figure 3F). Nomogram was shown in Figure 3H.These analyses indicated that the *STAT2* gene can predict the onset of IA.

# Correlation analysis between characteristic genes

Our research displayed that *KRT18*, *MICB* were positively correlated with *STAT2* (*p* value <0.05), and *ALDH3A2*, *IGSF1*, *NPY1R* were negatively correlated with *STAT2* (*p* value <0.05) in



function (MF) enrichment analysis.

GSE122897 and in GSE75436; whereas *COX6C* did not show significant correlation with *STAT2* in GSE75436. (Figures 4A, B)

# Correlation analysis between biomarker and immune cell infiltrations

xCell was used to identify immune and stromal cell types that could play roles in IA pathogenesis. Using the GSE122897 expression data, xCell generated enrichment scores for various cell types that were shown in (Supplementary Image 2C). The immune score of IA samples was significantly higher than the controls, while there were no significant differences between the stromal scores of IA tissues and controls (Figure 4C). This result indicated that the immune microenvironment played a more important role in IA formation than the stromal microenvironment. There were 16 cell types with significant differences in IA tissues vs. controls in the GSE122897 dataset as shown in Figure 4D. The correlation of *STAT2* and infiltrating cells was recorded in Supplementary Data Sheet 1 and visualized in Figure 4E. The scores of lymphatic (ly) endothelial cells, endothelial cells, microvascular (mv) endothelial cells, osteoblasts, neurons, CD4 central memory T cells (Tcm) and eosinophils were significantly decreased, while T-helper cells 2 (Th2 cells), fibroblasts, pericytes and mesenchymal stem cells were significantly enriched.

### Gene co-expression networks

In this study, the cohort of GSE122897 consisted of 41 IA tissues and 16 cerebral artery controls. The sample clustering



Selecting and validating characteristic genes (A, B) Investigation of the characteristic genes of the IL6/JAK/STAT signaling pathway (ISP) by the least absolute shrinkage and selection operator (LASSO) regression. (C, D) Investigation of the characteristic genes of the estrogen response pathway (ERP) by LASSO regression. (E) Nomogram based on the characteristic genes of ISP. (F) Validation of the diagnostic efficacy of STAT2 for intracranial aneurysm (IA) by the receiver operator characteristic (ROC)analysis. (G) Predictive accuracy of STAT2 for IA by the calibration curve. (H) Nomogram based on STAT2 as diagnostic tool for IA formation.

dendrograms of four traits (group, immuneScore, ERP and ISP) were shown in Figure 5A. In the WGCNA, we correlated each module with traits in the GSE122897 dataset by calculating the MS for each module-trait correlation. When 0.85 was used as the correlation coefficient threshold, the soft power  $\beta$  was selected as 8 (Figures 5B, D). Twenty-three co-expression modules were constructed by using the average linkage hierarchical clustering algorithm (Figure 5E). The red module had significant positive correlations with the four traits, whereas the blue module had significant negative correlations with these traits (Figure 5C). These results indicated strong associations among these traits. Seventy-six genes in the red module were identified as hub genes based on the cut-off criteria (|MM| > 0.8 and |GS| > 0.4) (Supplementary Data Sheet 2) (Figure 5F). In the blue module,

171 hub genes were identified (Supplementary Data Sheet3) (Figure 5G).

# Functional enrichment analysis of the hub genes

The GO BP enrichment analysis showed that the hub genes in the red module were mainly enriched in cellular response to biotic stimulus, regulation of cytokine-mediated signaling pathway, intrinsic apoptotic signaling pathway in response to endoplasmic reticulum stress, etc. MF enrichment analysis showed that these genes play an important role in protease binding. Protein processing in endoplasmic reticulum is the only



The correlations of *STAT2*, the estrogen response pathway (ERP) and the immune infiltrating cells (**A**, **B**) The correlations of *STAT2* and the characteristic genes of the ERP in GSE122897 and GSE75436. (**C**) The immune score and the stromal score of intracranial aneurysm (IA) samples compared with controls. (**D**) The infiltrating cells of IA samples compared with controls. (**E**) The correlation of *STAT2* and the infiltrating cells. ns=no significance, \*p<0.05, \*\*p<0.001.

pathway resulted from KEGG analysis (adj. p value <0.05). (Figure 6A)

The GO BP enrichment analysis revealed that the hub genes in the blue module were mainly enriched in response to endoplasmic reticulum stress, regulation of ion transmembrane transport. MF enrichment analysis showed that these genes play an important role in cadherin and coenzyme binding. Protein processing in endoplasmic reticulum and HIF-1 signaling pathway are enriched in KEGG analysis (adj. *p* value <0.05). (Figure 6B). The functions of hub genes in the red module and blue module are related to cellular substance metabolism.

### Screening of potential therapeutic drugs

Enrichr platform analysis found that suloctidil HL60 UP, prenylamine HL60 UP, terfenadine HL60 UP and acetaminophen CTD 00005295 etc. have been identified as drugs that regulate action for both *STAT2* and co-expression genes (Supplementary Data Sheet 4). Acetaminophen was approved to downregulate the expression of *STAT2* mRNA in Drugbank database (Supplementary Data Sheet 5). Acetaminophen can be a potential therapeutic drug for IA targeting *STAT2*.



#### FIGURE 5

Weighted gene co-expression network analysis (A) Clustering dendrogram of total genes related to intracranial aneurysm (IA) onset. (B, C) Softthreshold power analysis implemented to obtain the scale-free fit index of the network topology. (D) Identification of modules. (E) Heatmap shows the relationships between different modules and traits. (F) Scatter plot of module eigengenes in the red module. (G) Scatter plot of module eigengenes in the blue module.

## Discussion

The activation of ISP can stimulate cell proliferation, differentiation, migration and apoptosis in various diseases, e.g., cardiovascular diseases, colorectal cancer and myelofibrosis (20–22). It is well established that the ISP acts as a critical regulator of inflammatory processes during the pathogenesis of atherosclerosis and arterial hypertension, and its members are constitutively expressed in the vessel wall and transfer intracellular signaling events of various receptor families, e.g., that of cytokines, growth factors and vasoactive peptides (23). However, no study has reported the role of ISP in IA onset so far. Previously, we found that the ISP correlated with *SLC2A12*, a potential biomarker in IA occurrence (15). In the present study, the role of ISP in IA formation

was explored for the first time using bioinformatics analysis. It was significantly activated in IA formation by GSEA, and is significantly associated with immune cells activities. Then, we constructed a one-gene signature of *STAT2* and generated the correlation networks of *STAT2* with characteristic genes of ERP and infiltrating immune cells.

In this research, we found the ERP was activated in IA formation together with the ISP and immune response. But the roles of estrogen in IA formation remain controversial nowadays. Generally, the majority of present studies suggest a beneficial effect of estrogen in IA pathogenesis (24, 25). In a rat model of IA, the incidence of IA is significantly higher in ovariectomized females than in males (24). A clinical meta-analysis study conducted by Vlak MH, et al. found that the female/male prevalence ratio (PR)



of IA was 1.1 in populations with mean age of 50 years old, while this PR went up to 2.2 in those older than 50 years old (25). The decline of estrogen concentration in peri- and post-menopause periods can favor the formation of IAs.

However, it is well known that women are predisposed to IA (26). Female gender is an independent risk factor for the formation of new IAs (8). The preponderance of IAs between women and men starts after the first two decades of life (25). The role of female sex steroids such as estrogen may contribute to the sex disparities that exist in the presentation of IA. Moreover, estrogen can enhance immune response resulting in many acquired immunity and autoimmune diseases (27). Except for the beneficial effect of estrogen, the complex and multiple roles of estrogen may be involved in IA formation. In our research, the ISP, an immune-related pathway, was activated in IA formation, and the ERP was positively correlated with the ISP. Then we deduced that the ERP was positively correlated to this progress.

In addition, the WGCNA results were consistent with the above findings. The four traits, group, immuneScore, ERP and ISP, shared the same red module in WGCNA with significant positive correlations and the same blue module in WGCNA with significant negative correlations.

Vascular inflammation/immune responses were involved in IA formation, with neutrophil and macrophage infiltration,

oxidative stress, fragmentation of the internal elastic lamina, and degradation of the extracellular matrix by metalloproteinase (28, 29). Estrogen is implicated in immune responses (30). The estrogen receptors (ERs) are expressed by many immunological cells including T cells, B cells, macrophages, and dendritic cells (DCs), suggesting a direct effect of estrogens on immune responses. Estrogen and ERs can alleviate the inflammation of arterial cell wall matrix and apoptotic pathways (31-33). Gprotein coupled estrogen receptor (GPER) is a member of ERs, and it can induce apoptosis of vascular endothelial cells (34). The dysfunction of vascular endothelial cells is the initial part of IA formation (35, 36). Whereas, the other two members of ERs, ER $\alpha$  and ER $\beta$ , show a protective effect on IA (37). We speculate that there is a functional balance between ERs during the initiation of IA. Our research can shed light on the renewed understanding of the roles of estrogen involved in IA formation.

Studies manifested that estrogen deficiency leads to endothelial dysfunction and inflammation (38). And then, hemodynamic stress on the endothelial cells of the cerebral arteries triggers the pathogenesis of IAs, causing prolonged and excess inflammation in the vessel wall (39). The inflammation cellular infiltrations were discovered to occur in the IA fundus in 1966 (40). Since then, a series of experimental reports and animal studies have supported the role of inflammation in the pathogenesis of IA. The infiltration

of immune cells including macrophages, T cells and Mast cells is a characteristic of IA (41–43). Cytokines are powerful mediators of the immune response and are reported to be involved in the pathogenesis of IA, especially IL1 $\beta$ , IL6, and tumor necrosis factor- $\alpha$  (TNF $\alpha$ ) (44–46). Our research indicated that the immune microenvironment played a more important role in IA formation than the stromal microenvironment. Infiltrating immune cells included the B cells, CD4+ memory T cells, macrophages, monocytes, Th2 cells. These findings corroborated the previous researches.

To the best of our knowledge, the function of *STAT2* in IA pathogenesis has not been studied by now. We identified that *STAT2* was a potential biomarker with moderate diagnostic efficacy and an accurate estimation to distinguish IA samples from normal controls. *STAT2* may be a promising therapeutic target for the prevention of IA pathogenesis. Acetaminophen was approved to a potential drug targeting *STAT2* in Drugbank database. Nonsteroidal anti-inflammatory drugs (NSAIDs) are currently being investigated as a potential pharmaceutical treatment for patients with IA (47). Hasan DM reported acetylsalicylic acid, a member of NSAIDs, can attenuate inflammation in the walls of human cerebral aneurysms (48). More pharmacological research in this field will pave the way for the prevention and treatment of IA.

Limitations exist in this study. First, no experimental validation was performed in our research. Obtaining test samples (aneurysm domes) are getting more challenging at this stage. Compared with the microsurgical clipping, the endovascular coiling of IAs (ruptured and unruptured) shows more positive outcomes (49, 50). The endovascular treatment has become popular for the treatment of IAs and continues to take momentum around the world. Even in the open operation of IA, the key point is to clip the neck of IA but not to remove the dome of it (obtaining test samples). More exposure of microsurgical view may lead to more complications. Obtaining biological samples need more time and multicenter cooperation. We think animal models with endogenously induced IA formation will supplement reliable data in the future (15). Second, the control samples in this study included external carotid arteries. External and internal carotid arteries may differ at the transcriptome level. Third, the estrogen response pathway gene set was a combination of M5906 (early estrogen response) and M5907 (late estrogen response) from the MSigDB database. This combination may cause variations in the results.

# Conclusions

In the present study we identified that the ISP was one of the most significant positively correlated pathways with IA onset. The ISP, ERP and immune response were all activated together in this process. Except for the beneficial effect, complex and multiple roles of estrogen may be involved in IA formation. STAT2 could be a potential biomarker and a promising therapeutic target for the pathogenesis of IA.

# Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material. Further inquiries can be directed to the corresponding author.

## Author contributions

CZ designed the study. SL, XC and RZ performed the data analyses. AW and SM wrote the manuscript. CZ revised the manuscript. All authors contributed to the article and approved the submitted version.

## Funding

This study was supported by the Key Research and Development Program of Rizhao City (2021ZDYF020205).

## Acknowledgments

We especially thank Dr. Shipeng Guo (Department of Endocrine Breast Surgery, The First Affiliated Hospital of Chongqing Medical University, Chongqing, China) for his guidance on bioinformatics and conceptualization.

# Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

# Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/ fimmu.2022.1046765/full#supplementary-material

SUPPLEMENTARY FIGURE 1

Expression patterns of characteristic genes (A) The expression levels of *STAT2* in GSE122897. (B) The expression levels of *STAT2* in GSE75436. (C) The expression patterns of the characteristic genes of the estrogen response pathway (ERP) in GSE122897. (D) The expression patterns of the characteristic genes of the ERP in GSE75436.

#### SUPPLEMENTARY FIGURE 2

Discrimination ability of the six characteristic genes of the estrogen response pathway (ERP) and cell type enrichment result from xCell (A)

# References

1. Rinkel GJ, Djibuti M, Algra A, van Gijn J. Prevalence and risk of rupture of intracranial aneurysms: a systematic review. *Stroke* (1998) 29:251–6. doi: 10.1161/01.str.29.1.251

2. Hurford R, Taveira I, Kuker W, Rothwell PMOxford Vascular Study Phenotyped Cohort. Prevalence, predictors and prognosis of incidental intracranial aneurysms in patients with suspected TIA and minor stroke: a population-based study and systematic review. J Neurol Neurosurg Psychiatry (2021) 92:542–8. doi: 10.1136/jnnp-2020-324418

3. Tawk RG, Hasan TF, D'Souza CE, Peel JB, Freeman WD. Diagnosis and treatment of unruptured intracranial aneurysms and aneurysmal subarachnoid hemorrhage. *Mayo Clin Proc* (2021) 96:1970-2000. doi: 10.1016/j.mayocp.2021.01.005

4. Bakker MK, van der Spek RAA, van Rheenen W, Morel S, Bourcier R, Hostettler IC, et al. Genome-wide association study of intracranial aneurysms identifies 17 risk loci and genetic overlap with clinical risk factors. *Nat Genet* (2020) 52:1303–13. doi: 10.1038/s41588-020-00725-7

5. Leblanc R. Familial cerebral aneurysms. a bias for women. Stroke (1996) 27:1050-4. doi: 10.1161/01.str.27.6.1050

6. Francis SE, Tu J, Qian Y, Avolio AP. A combination of genetic, molecular and haemodynamic risk factors contributes to the formation, enlargement and rupture of brain aneurysms. *J Clin Neurosci* (2013) 20:912–8. doi: 10.1016/j.jocn.2012.12.003

7. Sijtsma LC, Rinkel GJ, Ruigrok YM. Sex-related clustering of intracranial aneurysms within families. *Stroke* (2015) 46:1107-9. doi: 10.1161/STROKEAHA.115.008798

8. Steiner T, Juvela S, Unterberg A, Jung C, Forsting M, Rinkel G, et al. European Stroke organization guidelines for the management of intracranial aneurysms and subarachnoid haemorrhage. *Cerebrovasc Dis* (2013) 2:93–112. doi: 10.1159/000346087

9. Kappelle LJ, Eliasziw M, Fox AJ, Barnett HJ. Small, unruptured intracranial aneurysms and management of symptomatic carotid artery stenosis. north American symptomatic carotid endarterectomy trial group. *Neurology* (2000) 2:307–9. doi: 10.1212/wnl.55.2.307

10. Etminan N, Dörfler A, Steinmetz H. Unruptured intracranial aneurysmspathogenesis and individualized management. *Dtsch Arztebl Int* (2020) 14:235–42. doi: 10.3238/arztebl.2020.0235

11. Signorelli F, Sela S, Gesualdo L, Chevrel S, Tollet F, Pailler-Mattei C, et al. Hemodynamic stress, inflammation, and intracranial aneurysm development and rupture: a systematic review. *World Neurosurg* (2018) 115:234–44. doi: 10.1016/j.wneu.2018.04.143

12. Berge J, Blanco P, Rooryck C, Boursier R, Marmat G, Gariel F, et al. Understanding flow patterns and inflammatory status in intracranial aneurysms: Towards a personalized medicine. *J Neuroradiol* (2016) 43:141–7. doi: 10.1016/j.neurad.2015.09.005

13. Aoki T, Fukuda M, Nishimura M, Nozaki K, Narumiya S. Critical role of TNF-alpha-TNFR1 signaling in intracranial aneurysm formation. *Acta Neuropathol Commun* (2014) 2:34. doi: 10.1186/2051-5960-2-34

14. Barrett T, Wilhite SE, Ledoux P, Evangelista C, Kim IF, Tomashevsky M, et al. NCBI GEO: archive for functional genomics datasets-update. *Nucleic Acids Res* (2013) 41:D991–5. doi: 10.1093/nar/gks1193

15. Zhao C, Ma Z, Shang J, Cui X, Liu J, Shi R, et al. Bioinformatics analysis reveals potential biomarkers associated with the occurrence of intracranial aneurysms. *Sci Rep* (2022) 12:13282. doi: 10.1038/s41598-022-17510-7

16. Aran D, Hu Z, Butte AJ. xCell: digitally portraying the tissue cellular heterogeneity landscape. *Genome Biol* (2017) 18:220. doi: 10.1186/s13059-017-1349-1

17. Langfelder P, Horvath S. WGCNA: an r package forweighted correlation network analysis. *BMC Bioinf* (2008) 9:559. doi: 10.1186/1471-2105-9-559

18. Yu G, Wang LG, Han Y, He QY. ClusterProfiler: an r package for comparing biological themes among gene clusters. *OMICS* (2012) 16:284–7. doi: 10.1089/omi.2011.0118

The AUCs of the six characteristic genes of the ERP in GSE75436. **(B)** The AUC of the combination of the six characteristic genes of the ERP in GSE75436 **(C)** Cell type enrichment result from xCell.

19. Walter W, Sanchez-Cabo F, Ricote M. GOplot: an r package for visually combining expression data with functional analysis. *Bioinformatics* (2015) 31:2912–4. doi: 10.1093/bioinformatics/btv300

20. Baldini C, Moriconi FR, Galimberti S, Libby P, De Caterina R. The JAK-STAT pathway: an emerging target for cardiovascular disease in rheumatoid arthritis and myeloproliferative neoplasms. *Eur Heart J* (2021) 42:4389–400. doi: 10.1093/eurheartj/ehab447

21. Cui C, Cheng X, Yan L, Ding H, Guan X, Zhang W, et al. Downregulation of TfR1 promotes progression of colorectal cancer *via* the JAK/STAT pathway. *Cancer Manag Res* (2019) 11:6323–41. doi: 10.2147/CMAR.S198911

22. Levavi H, Hoffman R, Marcellino BK. JAK inhibitors in the treatment of myelofibrosis. Clin Adv Hematol Oncol (2022) 20:456-67.

23. Grote K, Luchtefeld M, Schieffer B. JANUS under stress-role of JAK/STAT signaling pathway in vascular diseases. *Vascul Pharmacol* (2005) 43:357–63. doi: 10.1016/j.vph.2005.08.021

24. Oka M, Ono I, Shimizu K, Kushamae M, Miyata H, Kawamata T, et al. The bilateral ovariectomy in a female animal exacerbates the pathogenesis of an intracranial aneurysm. *Brain Sci* (2020) 10:335. doi: 10.3390/brainsci10060335

25. Vlak MH, Algra A, Brandenburg R, Rinkel GJ. Prevalence of unruptured intracranial aneurysms, with emphasis on sex, age, comorbidity, country, and time period: a systematic review and meta-analysis. *Lancet Neurol* (2011) 10:626–36. doi: 10.1016/S1474-4422(11)70109-0

26. Krzyzewski RM, Klis KM, Kucala R, Polak J, Kwinta BM, Starowicz-Filip A, et al. Intracranial aneurysm distribution and characteristics according to gender. *Br J Neurosurg* (2018) 32:541–3. doi: 10.1080/02688697.2018.1518514

27. Moulton VR. Sex hormones in acquired immunity and autoimmune disease. Front Immunol (2018) 9:2279. doi: 10.3389/fimmu.2018.02279

28. Frösen J, Cebral J, Robertson AM, Aoki T. Flow-induced, inflammationmediated arterial wall remodeling in the formation and progression of intracranial aneurysms. *Neurosurg Focus* (2019) 47:E21. doi: 10.3171/2019.5.FOCUS19234

29. Aoki T, Nishimura M. The development and the use of experimental animal models to study the underlying mechanisms of CA formation. *J BioMed Biotechnol* (2011) 2011:535921. doi: 10.1155/2011/535921

30. Cutolo M, Capellino S, Sulli A, Serioli B, Secchi ME, Villaggio B, et al. Estrogens and autoimmune diseases. *Ann N Y Acad Sci* (2006) 1089:538–47. doi: 10.1196/annals.1386.043

31. Krause DN, Duckles SP, Pelligrino DA. Influence of sex steroid hormones on cerebrovascular function. J Appl Physiol (1985) (2006) 101:1252–61. doi: 10.1152/japplphysiol.01095.2005

32. Knowlton AA, Lee AR. Estrogen and the cardiovascular system. *Pharmacol Ther* (2012) 1:54–70. doi: 10.1016/j.pharmthera.2012.03.007

33. Resanovic I, Rizzo M, Zafirovic S, Bjelogrlic P, Perovic M, Savic K, et al. Anti-atherogenic effects of 17 $\beta$ -estradiol. Horm Metab Res (2013) 10:701–8. doi: 10.1055/s-0033-1343478

34. Gros R, Ding Q, Liu B, Chorazyczewski J, Feldman RD. Aldosterone mediates its rapid effects in vascular endothelial cells through GPER activation. *Am J Physiol Cell Physiol* (2013) 304:C532–40. doi: 10.1152/ajpcell.00203.2012

35. Kaneko N, Mashiko T, Namba K, Tateshima S, Watanabe E, Kawai K. A patient-specific intracranial aneurysm model with endothelial lining: a novel *in vitro* approach to bridge the gap between biology and flow dynamics. *J neurointerv Surg* (2018) 10:306–9. doi: 10.1136/neurintsurg-2017-013087

36. Aoki T, Nishimura M, Matsuoka T, Yamamoto K, Furuyashiki T, Kataoka H, et al. PGE (2) -EP (2) signalling in endothelium is activated by haemodynamic stress and induces cerebral aneurysm through an amplifying loop via NF-κB. Br J Pharmacol (2011) 163:1237–49. doi: 10.1111/j.1476-5381.2011.01358.x

37. Maekawa H, Tada Y, Yagi K, Miyamoto T, Kitazato KT, Korai M, et al. Bazedoxifene, a selective estrogen receptor modulator, reduces cerebral aneurysm rupture in ovariectomized rats. *J Neuroinflamm* (2017) 14:197. doi: 10.1186/ s12974-017-0966-7 38. Tabuchi S. Relationship between postmenopausal estrogen deficiency and aneurysmal subarachnoid hemorrhage. *Behav Neurol* (2015) 2015:720141. doi: 10.1155/2015/720141

39. Kataoka H. Molecular mechanisms of the formation and progression of intracranial aneurysms. *Neurol Med Chir (Tokyo)* (2015) 55:214–29. doi: 10.2176/nmc.ra.2014-0337

40. Crompton MR. Mechanism of growth and rupture in cerebral berry an eurysms. Br Med J (1966) 1:1138–42. doi: 10.1136/bmj.1.5496.1138

41. Kataoka K, Taneda M, Asai T, Kinoshita A, Ito M, Kuroda R. Structural fragility and inflammatory response of ruptured cerebral aneurysms. a comparative study between ruptured and unruptured cerebral aneurysms. *Stroke* (1999) 3:1396–401. doi: 10.1161/01.str.30.7.1396

42. Frösen J, Piippo A, Paetau A, Kangasniemi M, Niemelä M, Hernesniemi J, et al. Remodeling of saccular cerebral artery aneurysm wall is associated with rupture: histological analysis of 24 unruptured and 42 ruptured cases. *Stroke* (2004) 35:2287–93. doi: 10.1161/01.STR.0000140636.30204.da

43. Ollikainen E, Tulamo R, Frösen J, Lehti S, Honkanen P, Hernesniemi J, et al. Mast cells, neovascularization, and microhemorrhages are associated with saccular intracranial artery aneurysm wall remodeling. *J Neuropathol Exp Neurol* (2014) 9:855–64. doi: 10.1097/NEN.00000000000105

44. Zhang HF, Zhao MG, Liang GB, Song ZQ, Li ZQ. Expression of proinflammatory cytokines and the risk of intracranial aneurysm. *Inflammation* (2013) 6:1195–200. doi: 10.1007/s10753-013-9655-6 45. Niu PP, Wang X, Xu YM. Association of interleukin-6 signaling and creactive protein with intracranial aneurysm: A mendelian randomization and genetic correlation study. *Front Genet* (2021) 12:679363. doi: 10.3389/ fgene.2021.679363

46. Wang J, Wei L, Lu H, Zhu Y. Roles of inflammation in the natural history of intracranial saccular aneurysms. *J Neurol Sci* (2021) 424:117294. doi: 10.1016/j.jns.2020.117294

47. Fisher CL, Demel SL. Nonsteroidal anti-inflammatory drugs: A potential pharmacological treatment for intracranial aneurysm. *Cerebrovasc Dis Extra* (2019) 1:31–45. doi: 10.1159/000499077

48. Hasan DM, Chalouhi N, Jabbour P, Dumont AS, Kung DK, Magnotta VA, et al. Evidence that acetylsalicylic acid attenuates inflammation in the walls of human cerebral aneurysms: preliminary results. *J Am Heart Assoc* (2013) 2: e000019. doi: 10.1161/JAHA.112.000019

49. Brinjikji W, Rabinstein AA, Nasr DM, Lanzino G, Kallmes DF, Cloft HJ. Better outcomes with treatment by coiling relative to clipping of unruptured intracranial aneurysms in the united states, 2001-2008. *AJNR Am J Neuroradiol* (2011) 6:1071–75. doi: 10.3174/ajnr.A2453

50. Molyneux A, Kerr R, Stratton I, Sandercock P, Clarke M, Shrimpton J, et al. International subarachnoid aneurysm trial (ISAT) of neurosurgical clipping versus endovascular coiling in 2143 patients with ruptured intracranial aneurysms: a randomised trial. *Lancet* (2002) 9342:1267-74. doi: 10.1016/s0140-6736(02)11314-6