Check for updates

OPEN ACCESS

EDITED BY Shengjun Wang, Jiangsu University Affiliated People's Hospital, China

REVIEWED BY Yuqian Chang, Fourth Military Medical University, China Michael Hickey, Monash University, Australia

*CORRESPONDENCE Jia-An Zhang jazhang618@126.com

SPECIALTY SECTION

This article was submitted to Autoimmune and Autoinflammatory Disorders: Autoinflammatory Disorders, a section of the journal Frontiers in Immunology

RECEIVED 19 August 2022 ACCEPTED 24 October 2022 PUBLISHED 03 November 2022

CITATION

Zhou X-Y, Chen K and Zhang J-A (2022) Mast cells as important regulators in the development of psoriasis. *Front. Immunol.* 13:1022986. doi: 10.3389/fimmu.2022.1022986

COPYRIGHT

© 2022 Zhou, Chen and Zhang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Mast cells as important regulators in the development of psoriasis

Xu-Yue Zhou, Kun Chen and Jia-An Zhang*

Institute of Dermatology, Jiangsu Key Laboratory of Molecular Biology for Skin Diseases and STIs, Chinese Academy of Medical Science and Peking Union Medical College, Nanjing, China

Psoriasis is a chronic inflammatory immune skin disease mediated by genetic and environmental factors. As a bridge between innate and adaptive immunity, mast cells are involved in the initiation, development, and maintenance of psoriasis by interactions and communication with a variety of cells. The current review describes interactions of mast cells with T cells, Tregs, keratinocytes, adipocytes, and sensory neurons in psoriasis to emphasize the important role of mast cell-centered cell networks in psoriasis.

KEYWORDS

mast cell, psoriasis, T cell, cell network, inflammation

Introduction

Psoriasis is a chronic-relapsing inflammatory autoimmune skin disorder mediated by complex interactions between genetic and environmental factors. Psoriasis can be triggered by injury or infection, and is characterized by the abnormal proliferation and differentiation of keratinocytes and infiltration of inflammatory cells, including T cells, mast cells (MCs), neutrophils, and macrophages. Although the pathogenesis of psoriasis is not fully understood, the dysfunction of innate and adaptive immune, especially the IL-23/IL-17 pathogenic axis, is now considered to play key roles in the development of psoriasis (1).

MCs are tissue-resident immune cells. Upon maturation, they are primarily located in barrier tissues, such as the skin, respiratory tract, and intestinal mucosa. The skin is highly enriched in MCs, which account for 8% of the total number of cells in the dermis, located near the epidermis and the subcutaneous vascular system and nerves (2). As local tissue sentinels, MCs are often initially activated by external environmental stimuli or pathogen invasion and subsequently release a variety of mediators, including *de novo* synthesized cytokines and chemokines and pre-stored histamine, proteases, leukotrienes, and prostaglandins in cytoplasmic granules, thus exerting immunomodulatory functions or driving allergic responses (3). MCs are key initiators and regulators of innate and adaptive immunity, and their complex communication with other cells is involved in the maintenance of barrier function and immune homeostasis (4). Several studies have confirmed the important role of MCs in regulating epidermal barrier function and maintaining immune homeostasis in the skin, including roles in wound healing and processes related to skin aging (5, 6). Moreover, MCs are required in allergeninduced skin inflammation. DC immunoreceptor (DCIR) is upregulated in skin MCs of patients with atopic dermatitis and mediates allergen binding and uptake (7). Thus, MCs have a strategic position in host defense, the response to allergens, and immune homeostasis.

Mast cells in psoriasis

Toruniowa et al. found that the number and degranulation of MCs were higher in the early stages of psoriasis lesions, especially in newly formed psoriatic lesions, than in mature psoriatic lesions. The Koebner phenomenon refers to the occurrence of new psoriatic lesions in the area of healthy skin following traumatic injury to patients with psoriasis. There is a consistent trend towards an increase in MCs in Koebner's isomorphic response (8). The number of MCs in psoriatic lesions is reduced by successful treatment with anthralin, psoralen plus ultraviolet A irradiation (PUVA), or cyclosporine (9–11).

FceRI is a high-affinity IgE receptor that is constitutively expressed on MCs. IgE can recognize exogenous antigens (Ags), facilitate antigen presentation via FceRI, trigger the degranulation of MCs and the release of multiple mediators, and regulate innate and acquired immunity. Several reports have shown that total serum IgE levels are significantly elevated in patients with psoriasis and that IgE⁺ FccRI⁺ cells are significantly increased in psoriatic lesions and decrease with the improvement of the disease (12, 13). A non-classical IgE response may occur in psoriasis. MCs can directly bind to immature dendritic cells (DCs), induce increased expression levels of the costimulators CD86, CD80, and CD40 on the DC surface, indicating DC maturation, and promote the release of T cell-modulating cytokines, such as IFN-y, IL-12, IL-6, and TGF- β , enhancing the ability of DCs to induce the proliferation of CD4+ T cells and Th1 and Th17 polarization (14). Increased IL-17A induces IgG memory B cells to produce IgE by class-switch recombination, forming immune complexes containing soluble IgE with psoriasis-associated autoantigens; these are presented to plasmacytoid (p) DCs via FccRI, activating pDCs to trigger inflammatory cytokine responses, including IFN-α, IL-6, IL-8, and TNF, involved in the development and maintenance of inflammatory processes in psoriatic lesions (15-17). Future studies are needed to determine how IgE-FceRI interactions precisely affect the activation of multiple immune cells in psoriatic skin lesions. Tryptases and chymases are classical mediators secreted by activated MCs and contribute to the persistence of inflammation by activating many innate and adaptive immune responses via protease-activated receptor 2

(PAR-2) (18). Serum tryptase levels are elevated in psoriasis (19). Tryptases can induce the production and release of multiple proinflammatory cytokines that directly cause and amplify psoriasis-associated inflammatory processes, such as IL-1 β , IL-8, TNF α , and IL-6 production (20), and activate matrix metalloproteinases (MMP) to destroy various matrix components associated with the development of psoriatic arthritis (21). Pso p27 is an autoantigen detectable in MCs of psoriatic lesions and is thought to be produced by the noncanonical cleavage of MC chymases (22).

CIBERSORT is an analytical tool that uses a deconvolution algorithm to estimate the composition and abundance of various types of immune cells in samples based on gene-expression data (23). Several recent studies have used the CIBERSORT algorithm to assess immune cell infiltration in psoriasis. CIBERSORT uses a leukocyte gene signature matrix termed LM22 as the input matrix of reference gene expression signatures, with 547 genes to distinguish 22 human hematopoietic cell phenotypes; it defines activated MC as MCs activated by IgE receptors (24). Su et al. found fewer resting MCs in psoriatic lesions than in normal control samples (25). Zhang et al. found that tyrosine kinase receptor (c-Kit) and tryptase double-positive activated MCs were enriched in psoriatic skin and were progressively reduced in number after etanercept treatment, while resting MCs were nearly absent in psoriatic lesions. The significant upregulation of the MC activator IL-1B and the downregulation of FAM124B and FAM174B, identified as the specific genes in resting MCs, further support the important role of MC activation in the progression of psoriasis. In addition, a decrease in the total number of MCs and an increase in the proportion of activated MCs in psoriatic lesions have been reported (26). However, Liu et al. used the CIBERSORT method to evaluate immune cell infiltration based on hypermethylated genes in psoriasis and found a significant increase in resting MCs (27). The difference in results between studies may be related to the different stages of psoriatic lesions in the datasets. Gong et al. did not detect a difference in activated MCs between severe psoriasis and mild psoriasis, while resting MCs were decreased and correlated with disease severity (28). The authors suggested that this may be related to the fact that the gene expression profile used by the CIBERSORT algorithm for deconvolution mainly reflects IgEactivated MCs, while MC activation in psoriasis may be mediated by a non-IgE-dependent pathway.

Contribution of mast cells to the immunopathogenesis of psoriasis

T cells

The role of T cells in psoriasis is well-established. The functional interactions between MCs and T cells have been

10.3389/fimmu.2022.1022986

confirmed by several studies. The local density and activation of MCs are increased in a variety of T cell-mediated inflammatory diseases, such as rheumatoid arthritis, multiple sclerosis, and Crohn's disease, and the co-localization of activated MCs and T cells has been reported in inflammatory tissues (29–32).

MCs show surface expression of major histocompatibility complex (MHC) I and II molecules that directly contact and activate T lymphocytes by interacting with T cell receptor (TCR), thereby inducing the antigen-specific clonal expansion of T cells (33, 34). Activated MCs act synergistically with IL-1 β released in response to toll-like receptor (TLR)/FccRI receptor to enhance Th17 cell responses (35). In a hypercholesterolemic environment, MCs are capable of presenting Ags and effectively modulate CD4⁺ T effector cells to a proatherogenic Th1 phenotype within atherosclerotic plaques (36). IFN- γ -triggered mouse MCs can present Ags to CD4⁺ T cells, resulting in functional immunological synapses (37). Similarly, IFN- γ induces the expression of HLA-DR, HLA-DM, CD80, and CD40 on primary human MCs, which activate T cells by taking up and processing Ags and acting as antigen-presenting cells in vitro (38). In addition, the increased surface delivery of HLA class II and peptides induced by the degranulation of MCs further promotes T cell proliferation and creates a feed-forward loop of T cell-MC cross-activation (39).

MCs are a major source of IL-22 in patients with psoriasis (40). Gaudenzio et al. have shown that they act as inflammatory amplifiers in inflammatory lesions of patients with psoriasis. IFN- γ mediates the establishment of synaptic contacts between MCs and CD4⁺ T cells and induces their proliferation, leading to the induction of Th22 and IL-22⁺ IFN- γ^+ Th cell subpopulations by the release of IL-6 and TNF α . Approximately 30% of unbound CD4⁺ T cells in psoriatic skin are IL-22⁺, while up to 60% of CD4⁺ T cells in contact with MCs express IL-22 (41). Mashiko et al. found that approximately 89% of c-Kit⁺ FccRI⁺ MCs in psoriatic lesions produced IL-22, whereas only about 1% of CD3⁺ T cells produced IL-22. c-Kit⁺ cells also produced IL-17, and most IL-17-producing c-Kit⁺ FccRI⁺ MCs co-expressed IL-22 (42).

MCs can transport contents, including micro-RNAs and MHC, to distant antigen-presenting cells *via* secretory extracellular vesicles. Phospholipase A2 (PLA₂) is highly expressed in psoriatic lesions and produces cutaneous neolipid antigens for recognition by CD1a-reactive T cells. Cheung et al. found that the IFN- α -induced release of exosomes from MCs transferred PLA2 to adjacent CD1a hyper-expressing Langerhans cells, leading to the production of neolipid antigens and subsequent recognition by lipid-specific CD1areactive T cells, which induced the production of IL-22 and IL-17A (43).

It is currently believed that primary human MCs do not produce IL-17A by themselves; instead, exogenous IL-17A is captured and stored in specialized intracellular vesicles *via* receptor-mediated endocytosis. (44). Thus, in disease states, mast cells may function as sentinel cells by rapidly releasing preformed bioactive IL-17A to promote host defense and/or tissue inflammation. Eliasse et al. found that MCs are the primary source of IL-17 in early acne which relies on contact between activated $CD4^+$ T cells and MCs, while in the post-inflammatory phase, neutrophils are recruited and amplify the production of IL-17 (45). Chen et al. demonstrated that synovial tissue-resident MCs in peripheral spondyloarthritis can act as IL-17A-loaded sentinel cells that release IL-17A to exacerbate tissue inflammation (46).

Recent studies have shown that neutrophil extracellular traps (NETs) play an important role in psoriasis. Lambert et al. have identified the connection between NETs and Th17 responses in psoriasis. Mutations in the TRAF3IP2 gene in psoriasis promotes the ability of NET to induce Th17 cells in peripheral blood mononuclear cells and the secretion of IL-17A (47). Abundant polymorphonuclear neutrophils can release RNA, DNA, and cathelicidin (LL-37) via NETs to form nucleic acid-LL37 complexes, contributing to the release of TLR-mediated cytokines and chemokines; this also form a vicious cycle, promoting the release of NETs and exacerbating inflammation (48). As a chemoattractant for MCs, LL37 activates innate immune cells and disrupts innate tolerance to self-nucleic acids, resulting MC degranulation and the release of inflammatory mediators (49). In addition to neutrophils, several functions of other innate immune cells, such as MCs, macrophages, and eosinophils, in the form of extracellular traps (ETs) have gradually been revealed. The main components of MC extracellular traps (MCETs) are DNA, histones, and granule proteins, such as tryptase and LL-37. A variety of stimulants can induce MCET formation, such as IL-23 and IL-1 β (50). MCs can exert antimicrobial activity by the release of MCET (von 51). In human psoriatic lesions, IL-17 is released at low levels by MCETosis, and IL-1 β and IL-23 can rapidly induce MC degranulation and MCETosis in human skin, leading to the release of IL-17 and other MC products (50).

In addition, MCs can be activated by micro-vesicles released by T cells, thereby generating a response at the site of inflammation without contact with T cells (52). MCs can take up activated T cell-derived micro-vesicles. Shefler et al. demonstrated that MC-derived IL-24 induces the phosphorylation of signal transducer and activator of transcription-3 (STAT3) in keratinocytes of psoriasis after treatment with T cell-derived micro-vesicles (53).

Tregs

Skin-resident regulatory T cells (Tregs) plays an important role in the maintenance of immune homeostasis. Tregs are mostly composed of CD4⁺CD25⁺FoxP3⁺ T cells, which can control the activity of other effector immune cells and maintain autoimmune tolerance by direct contact or the secretion of suppressive cytokines, such as IL-10 and transforming growth factor (TGF)- β . The dysregulation of these cells is closely associated with chronic inflammatory diseases (54). Tregs control the progression of skin inflammation by inhibiting the infiltration of CD4⁺ T cells, which produce granulocyte-macrophage colony-stimulating factor (GM-CSF), in the skin (55). In patients with psoriasis, dermal Tregs are dysfunctional, characterized by an inability to inhibit the proliferation of normal CD4⁺CD25⁻ responder T cells (Tresps) (56). The treatment of psoriasis may increase the number and activity of Tregs (57).

The interaction between Tregs and MCs has a bidirectional nature and is critical for immunosuppression in various diseases. Activated MCs inhibit Treg function *via* the OX40/OX40L axis in an IL-6-dependent manner, thereby promoting Th17 cell differentiation (58). MC-derived chymase downregulates immune tolerance-related cytokines, such as IL-10, TGF- β 1, and IL-17A, thereby inhibiting Treg function. Chymase inhibitors effectively reduce inflammation and result in significant increases in the expression of immune tolerance-related cytokines, FoxP3 and Tregs (59). IL-10 and TGF- β secreted by Tregs can downregulate FccRI expression in MCs, thereby inhibiting MC maturation and degranulation (60).

Bovenschen et al. observed that CD4 ⁺ CD25 ⁺ Foxp3 ⁺ Tregs were more frequently located in the dermis than in the epidermis in patients with plaque psoriasis (61). The Koebner phenomenon occurs at sites of skin barrier damage in patients with psoriasis, and the use of tape stripping to remove the stratum corneum provides an *in vivo* model for skin barrier damage (62). Suttle et al. used tape stripping to induce the Koebner isomorphic response on normal skin of patients with psoriasis and found a significant decrease in the number of FoxP3⁺ Tregs in biopsies and an decreased number of apparent morphological contacts between tryptase⁺ MCs and FoxP3⁺ Tregs within 3–7 d in the Koebner-positive group (63). Thus, MCs may play a role in the development of the isomorphic response by crosstalk with Tregs to regulate the expression of related inhibitory factors.

Keratinocytes

A growing number of recent studies have demonstrated that keratinocytes in psoriasis interact with immune cells to form a feed-forward inflammatory loop. They act as a trigger and amplifier of the inflammatory response by recruiting and activating immune cells, thereby triggering local inflammation, which plays important roles in the initiation, maintenance, and relapse phases of psoriasis (64).

Stem cell factor (SCF), which binds to c-Kit, is critical for the development, differentiation, and survival of MCs as well as recruitment, maturation, and migration in tissues. Keratinocytes represent pivotal cellular sources that trigger MC maturation and function in the skin; therefore, the homeostasis of resident skin MCs may be regulated in part by epidermal keratinocytes. Elevated serum SCF levels and significant enrichment of SCF in lesional keratinocytes were found in patients with psoriasis (65). The skin microbiota can extend into the dermis and establish physical contact with various cells beneath the basement membrane. The large amount of lipoteichoic acid (LTA) contained in the cell wall of Gram-positive bacteria triggers the production of SCF from adjacent keratinocytes via the TLR signaling pathway, thereby recruiting and regulating MCs in the dermis (66). Cho et al. further found that the expression of SCF is increased in the epidermis of IMQ-induced psoriasis-like lesions. The stimulation of rhIL-17 significantly improved the secretion of SCF in HaCaT cells and positively regulated MC proliferation (67). On the other hand, dexamethasone-activated MCs can promote keratinocyte proliferation via keratinocyte growth factor (KGF) in vitro, thereby disrupting skin barrier homeostasis (68).

Stressed keratinocytes can initiate the activation and recruitment of immune cells by the release of alarmins, including thymic stromal lymphopoietin (TSLP) or IL-33 (69). It has been suggested that herpes simplex virus type 2 (HSV-2)infected keratinocytes contribute to the MC-mediated antiviral host defense via the IL-33/ST2 axis (70). IL-33, a member of the IL-1 cytokine family, regulates MC apoptosis and proinflammatory mediator release by binding to the ST2 receptor. Chymotrypsin and tryptases secreted by MCs facilitate the production of shorter, more mature, and more active IL-33 (4, 71). After release, IL-33 can diffuse from the epidermis into the dermis, triggering ST2-positive cells, including MCs, since the gene expression levels of IL-33 and histidine decarboxylase are significantly increased in the skin of patients with psoriasis (72, 73). The co-administration of IL-33 with the peptide substance P (SP) significantly increases the expression and secretion of TNF and VEGF in MCs, further promoting angiogenesis and increasing inflammation in psoriasis (73, 74). TSLP is a pro-allergic cytokine expressed by keratinocytes and elevated in psoriatic lesions. MCs regulate TSLP production in keratinocytes via the tryptase/PAR-2 axis and are thereby involved in the development of inflammatory skin diseases (75). Vasoactive intestinal peptide (VIP)-positive nerve fibers in the dermis, dermal-epidermal junction, and epidermis are higher in patients with psoriasis than in undamaged and healthy skin. Th1 cytokines (i.e., TNF- α and IFN- γ) and VIP released by MC in the dermis induce the production of IL-6, SCF, and VEGF by keratinocytes, resulting in increased vascular permeability and amplification of the inflammatory condition of the skin (76).

Adipocytes

The neuro-endocrine-immune system is an integrated regulatory network with an important role in organismal homeostasis. A complex triangular relationship among adipocytes, sympathetic nerves, and immune cells has been described (77). Neuro-immuno-metabolism is a promising but mysterious new area in the pathogenesis of psoriasis.

The sympathetic nervous system releases norepinephrine locally in adipose tissue (AT), thereby promoting lipolysis, and immune cells within adipose tissue are involved in the pathophysiology of obesity and its comorbidities. Innate immune cells promote insulin resistance (IR) in AT, which can act as a large secretory organ, producing substantially amounts of pro-inflammatory cytokines and adipokines that recruit relevant immune cells to drive systemic inflammation. AT-resident cells are significantly altered during obesity, with the accumulation of neutrophils, pro-inflammatory M1 macrophages, and MCs but a reduction in some cell types, e.g., Th2, Treg, and eosinophils (78). Some researchers have reported the accumulation of MCs in the AT of obese individuals. The number of MCs in subcutaneous AT is increased in individuals with metabolic syndrome and is positively correlated with IR and glucose levels (79, 80).

The effects of obesity on adipocytes mainly include the upregulation of pro-inflammatory adipokines, such as leptin and resistin, and the downregulation of anti-inflammatory adipokines, such as adiponectin (81). Obesity is one of the most common complications of psoriasis and an independent risk factor for the development of psoriasis (82). The levels of leptin and resistin are increased in patients with psoriasis and are positively correlated with disease severity, while the levels of lipocalin are decreased (83–85).

In a mouse model, IMQ-induced psoriasis-like skin inflammation was attenuated by a leptin deficiency, whereas leptin stimulation not only directly induced the proinflammatory phenotype in keratinocytes but also indirectly affected keratinocytes by inducing the secretion of proinflammatory cytokines from dermal fibroblasts (86, 87). Leptin can induce the production of IL-6 in keratinocytes by reducing the expression of caveolin-1 (88). MCs in AT are adjacent to leptin-producing adipocytes and are involved in the development of low-grade inflammation in obese individuals (89). A leptin deficiency in MCs facilitates the acquisition of an anti-inflammatory phenotype and polarizes macrophages toward M2 without affecting T cell differentiation (90). In addition, leptin can act as a paracrine or autocrine regulator of MC activity during inflammation by triggering the release of mediators, including cysteinyl leukotrienes (cysLTs) and chemokine CCL2 (91, 92).

Resistin exerts proinflammatory effects *via* the NF-kB signaling pathway by stimulating the production of cytokines, such as IL-6, IL-12, and TNF- α , thereby promoting the proliferation of keratinocytes and the recruitment of T cells to the skin and participating in the inflammatory process in psoriasis (93). Lipocalin has anti-inflammatory effects on keratinocytes *in vitro* by the inhibition of TNF- α . In psoriasis mouse models, a lack of lipocalin aggravates skin inflammation and the excessive infiltration of $\gamma\delta T$ cells producing IL-17 in the dermis (94). In addition, lipocalin induces the switch of MCs to an anti-inflammatory phenotype, mediating the production of IL-10 *via* the PI3K and p38 pathways, thus participating in the regulation of inflammation (91).

Sensory neurons

It has recently been shown that in addition to roles in sensory signaling, sensory neurons are involved in the regulation of immune responses and associated inflammation *via* the secretion of various neuropeptides, such as SP, nerve growth factor (NGF), CGRP, VIP, and somatostatin. MCs can be activated by the stimulation of neuropeptides and the close anatomical proximity and functional communication of MC with sensory nerves has been detected in the intestine, skin, and respiratory tract (95, 96).

MC-neuron communication may contribute to the pathogenesis of non-allergic skin diseases, such as psoriasis and rosacea, and may play a critical role in the development of neurogenic inflammation, pain, and pruritus. In the skin, MCs enhance TRPV1 sensitivity or lower its activation threshold by releasing nociceptive and pruritogenic mediators, such as histamine, prostaglandins, and leukotrienes, and initiate reciprocal communication with specific nociceptors on sensory nerve fibers (97). Nerve fibers release neuropeptides to activate MCs by a feedback mechanism. In psoriatic lesions, the level of SP, nerve density, total number and degranulation of MCs, and contacts between MCs and sensory nerves are elevated (98). Nociceptors can drive IL-23-mediated psoriasis-like skin inflammation (99). Psoriasis is regulated by the brain-skin axis. The HPA axis is involved in the development of psoriasis by the release of CRH, ACTH, and glucocorticoids, and these regulate the skin response to stress and the topical immune response (100). MCs express CRHR1 in proximity of psoriatic plaques, and elevated CRH in psoriasis induces the degranulation of MCs. By contrast, the secretion of IL-6 by MCs can also induce the CRH secretion by the activation of the HPA axis by a feedback mechanism (101).

Mas-related G-protein-coupled receptors (Mrgprs) belong to the G protein-coupled receptor (GPCR) family and are mostly expressed by small-diameter sensory neurons in the peripheral

nervous system. MRGPRX2 is a human Mrgpr and Mrgprb2 is the mouse ortholog of MRGPRX2. Human MCs are the only cells other than dorsal root ganglia that express MRGPRX2. MRGPRX2 can be activated by basic secretagogues, such as SP, somatostatin, compound 48/80, and mastoparan, which induce the activation of MCs by an IgE-independent pathway (102, 103). In the skin, MCs activated by MrgprB2/X2 can communicate locally with nearby peripheral neurons by releasing granules (104). IL-33 activates the JNK, p38, and NFκB pathways in cutaneous MCs and synergistically stimulates the release of IL-8, TNF- α , CCL1, and CCL2 from MCs with MRGPRX2 (105). Bidirectional communication between MCs and neurons initiates and potentially increases inflammation as well as pruritus and pain perception in psoriasis. A better understanding of these interactions and the development of high-affinity MRGPRX2 inhibitors hold the promise of unlocking a new generation of therapeutic targets for neurogenic inflammation and non-histamine pruritus complicated by psoriasis.

Conclusion

In this review, we highlight the key role of MCs in psoriasis. MCs are involved in the initiation, development, and maintenance phases of psoriasis, with clear changes in the number and function of these cells, especially in the early stages of psoriasis. As a bridge between innate and adaptive immunity, MCs are involved in the regulation of inflammation and immune homeostasis in psoriasis by interacting with a variety of cells, including T cells, Tregs, neutrophils, DCs, keratinocytes, adipocytes, and sensory nerve cells, forming a complex cellular network (Figure 1).

Although our understanding of the role of MCs in the pathogenesis of psoriasis has improved considerably, relatively little is known about the mechanisms by which various factors regulate the specific functions of MCs. Studying the mechanisms underlying MC-immune cell interactions may provide a basis for restoring MC homeostasis and contribute to the treatment of psoriasis.



FIGURE 1

Overview of mast cell involvement in psoriasis. When exposed to external stresses, stimulated keratinocytes can promote angiogenesis and amplify inflammation by releasing alarmins to trigger ST2-positive MCs secreting TNF and VEGF, etc. The skin microbiota induces keratinocytes to produce more SCF, which bind to the c-kit and are involved in the recruitment and regulation of dermal MCs. MCs can activate T lymphocytes and strengthen Th17 and Th22 responses by interaction with TCR, secretion of extracellular vesicles, and formation of MCETs. MCs also regulate Treg function through the OX40/OX40L axis. Activated MCs were found to increase the sensitivity of TRPV1 in peripheral neurons by releasing nociceptive and pruritogenic mediators. While the stimulated nerve fibers release CRH and neuropeptides, which induce the activation of MCS *via* CRH-R1 and MRGPRX2 on the MC surface in a feedback manner. In addition, nerve fibers release norepinephrine into adipose tissue to regulate the secretion of leptin, resistin, and lipocalin in adipocytes, which promote the conversion of MCS to a proinflammatory phenotype, thus constituting a neuroendocrine-immune loop that is involved in the development of psoriasis.

Author contributions

X-YZ, J-AZ and KC contributed to conception and design of the study. X-YZ wrote the first draft of the manuscript. J-AZ and KC wrote sections of the manuscript. All authors contributed to manuscript revision, read, and approved the submitted version.

Funding

This study is supported by grants from National Natural Science Foundation of China (No. 82273552, 82203947 and 82073445), Natural Science Foundation of Jiangsu Province (No. BK20210049), CAMS Innovation Fund for Medical Sciences (CIFMS-2021-I2M-1-001) and Medical Scientific Research Project of Jiangsu Provincial Health Commission (M2022113).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

1. Griffiths CEM, Armstrong AW, Gudjonsson JE, Barker J. Psoriasis. Lancet (2021) 397:1301-15. doi: 10.1016/S0140-6736(20)32549-6

2. Chang TM, Yang TY, Huang HC. Nicotinamide mononucleotide and coenzyme Q10 protects fibroblast senescence induced by particulate matter preconditioned mast cells. *Int J Mol Sci* (2022) 23:7539. doi: 10.3390/ijms23147539

3. Mukai K, Tsai M, Saito H, Galli SJ. Mast cells as sources of cytokines, chemokines, and growth factors. *Immunol Rev* (2018) 282:121–50. doi: 10.1111/ imr.12634

4. Lefrançais E, Duval A, Mirey E, Roga S, Espinosa E, Cayrol C, et al. Central domain of IL-33 is cleaved by mast cell proteases for potent activation of group-2 innate lymphoid cells. *Proc Natl Acad Sci USA* (2014) 111:15502–7. doi: 10.1073/pnas.1410700111

5. Wulff BC, Parent AE, Meleski MA, Dipietro LA, Schrementi ME, Wilgus TA. Mast cells contribute to scar formation during fetal wound healing. *J Invest Dermatol* (2012) 132:458–65. doi: 10.1038/jid.2011.324

 Abdel Hafez SMN. Age related changes in the dermal mast cells and the associated changes in the dermal collagen and cells: A histological and electron microscopy study. Acta Histochem (2019) 121:619–27. doi: 10.1016/j.acthis.2019.05.004

7. Luo X, Chen J, Yang H, Hu X, Alphonse MP, Shen Y, et al. Dendritic cell immunoreceptor drives atopic dermatitis by modulating oxidized CaMKII-involved mast cell activation. *JCI Insight* (2022) 7:e152559. doi: 10.1172/jci.insight.152559

8. Toruniowa B, Jabłońska S. Mast cells in the initial stages of psoriasis. Arch Dermatol Res (1988) 280:189-93. doi: 10.1007/BF00513956

9. Yamamoto T, Matsuuchi M, Irimajiri J, Otoyama K, Nishioka K. Topical anthralin for psoriasis vulgaris: evaluation of 70 Japanese patients. *J Dermatol* (2000) 27:482-5. doi: 10.1111/j.1346-8138.2000.tb02212.x

10. Stellato C, De Paulis A, Ciccarelli A, Cirillo, et al. Anti-inflammatory effect of cyclosporin a on human skin mast cells. *J Invest Dermatol* (1992) 98:800–4. doi: 10.1111/1523-1747.ep12499960

11. Töyry S, Fräki J, Tammi R. Mast cell density in psoriatic skin. the effect of PUVA and corticosteroid therapy. *Arch Dermatol Res* (1988) 280:282–5. doi: 10.1007/BF00440601

12. Colas L, Magnan A, Brouard S. Immunoglobulin e response in health and disease beyond allergic disorders. *Allergy* (2022) 77(6):1700–18. doi: 10.1111/all.15230

13. Ding Y, Yi X, Yu N. Serum IgE levels are increased in patients with generalized pustular psoriasis. *Clin Exp Dermatol* (2013) 38:549–52. doi: 10.1111/ced.12086

14. Dudeck A, Suender CA, Kostka SL, Von Stebut E, Maurer M. Mast cells promote Th1 and Th17 responses by modulating dendritic cell maturation and function. *Eur J Immunol* (2011) 41:1883–93. doi: 10.1002/eji.201040994

15. Yan KX, Huang Q, Fang X, Zhang ZH, Han L, Gadaldi K, et al. IgE and FccRI are highly expressed on innate cells in psoriasis. *Br J Dermatol* (2016) 175:122–33. doi: 10.1111/bjd.14459

16. Kasumagic-Halilovic E. Total serum immunoglobulin e levels in patients with psoriasis. *Mater Sociomed* (2020) 32:105–7. doi: 10.5455/msm.2020.32.105-107

17. Luo L, Luo Y, Xu J, Zhu R, Wu J, Liu X, et al. Heterogeneous origin of IgE in atopic dermatitis and psoriasis revealed by b cell receptor repertoire analysis. *Allergy* (2022) 77:559–68. doi: 10.1111/all.15173

18. Redhu D, Franke K, Aparicio-Soto M, Kumari V, Pazur K, Illerhaus A, et al. Mast cells instruct keratinocytes to produce thymic stromal lymphopoietin: Relevance of the tryptase/protease-activated receptor 2 axis. *J Allergy Clin Immunol* (2022) 149:2053–2061.e6. doi: 10.1016/j.jaci.2022.01.029

19. Paolino G, Di Nicola ,MR, Currado M, Brianti P, Mercuri SR. Serum tryptase levels in patients with psoriasis: a pilot study for a possible useful biomarker. *Clin Exp Dermatol* (2022) 47:178–9. doi: 10.1111/ced.14874

20. Malamud V, Vaaknin A, Abramsky O, Mor M, Burgess LE, Ben-Yehudah A, et al. Tryptase activates peripheral blood mononuclear cells causing the synthesis and release of TNF-alpha, IL-6 and IL-1 beta: possible relevance to multiple sclerosis. *J Neuroimmunol* (2003) 138:115–22. doi: 10.1016/S0165-5728(03)00090-0

21. Shin K, Nigrovic PA, Crish J, Boilard E, Mcneil HP, Larabee KS, et al. Mast cells contribute to autoimmune inflammatory arthritis via their tryptase/heparin complexes. J Immunol (2009) 182:647–56. doi: 10.4049/jimmunol.182.1.647

22. Lysvand H, Hagen L, Klubicka L, Slupphaug G, Iversen OJ. Psoriasis pathogenesis - pso p27 is generated from SCCA1 with chymase. *Biochim Biophys Acta* (2014) 1842:734–8. doi: 10.1016/j.bbadis.2014.02.005

23. Newman AM, Liu CL, Green MR, Gentles AJ, Feng W, Xu Y, et al. Robust enumeration of cell subsets from tissue expression profiles. *Nat Methods* (2015) 12:453–7. doi: 10.1038/nmeth.3337

24. Chtanova T, Newton R, Liu SM, Weininger L, Young TR, Silva DG, et al. Identification of T cell-restricted genes, and signatures for different T cell responses, using a comprehensive collection of microarray datasets. *J Immunol* (2005) 175:7837–47. doi: 10.4049/jimmunol.175.12.7837

25. Su W, Wei Y, Huang B, Ji J. Identification of hub genes and immune infiltration in psoriasis by bioinformatics method. *Front Genet* (2021) 12:606065. doi: 10.3389/fgene.2021.606065

26. Zhang Y, Shi Y, Lin J, Li X, Yang B, Zhou J. Immune cell infiltration analysis demonstrates excessive mast cell activation in psoriasis. *Front Immunol* (2021) 12:773280. doi: 10.3389/fimmu.2021.773280

27. Liu Y, Cui S, Sun J, Yan X, Han D. Identification of potential biomarkers for psoriasis by DNA methylation and gene expression datasets. *Front Genet* (2021) 12:722803. doi: 10.3389/fgene.2021.722803

28. Gong X, Wang W. Profiles of innate immune cell infiltration and related core genes in psoriasis. *BioMed Res Int* (2021) 2021:6656622. doi: 10.1155/2021/6656622

29. Kim KW, Kim BM, Won JY, Min HK, Lee KA, Lee SH, et al. Regulation of osteoclastogenesis by mast cell in rheumatoid arthritis. *Arthritis Res Ther* (2021) 23:124. doi: 10.1186/s13075-021-02491-1

30. Gregory GD, Robbie-Ryan M, Secor VH, Sabatino JJ, Brown MA. Mast cells are required for optimal autoreactive T cell responses in a murine model of multiple sclerosis. *Eur J Immunol* (2005) 35:3478–86. doi: 10.1002/eji.200535271

31. Bilotta S, Arbogast J, Schart N, Frei M, Lorentz A. Resveratrol treatment prevents increase of mast cells in both murine OVA enteritis and IL-10(-/-) colitis. *Int J Mol Sci* (2022) 23:1213. doi: 10.3390/ijms23031213

32. Russi AE, Walker-Caulfield ME, Guo Y, Lucchinetti CF, Brown MA. Meningeal mast cell-T cell crosstalk regulates T cell encephalitogenicity. J Autoimmun (2016) 73:100–10. doi: 10.1016/j.jaut.2016.06.015

33. Dudeck J, Medyukhina A, Fröbel J, Svensson CM, Kotrba J, Gerlach M, et al. Mast cells acquire MHCII from dendritic cells during skin inflammation. *J Exp Med* (2017) 214:3791–811. doi: 10.1084/jem.20160783

34. Stelekati E, Bahri R, D'orlando O, Orinska Z, Mittrücker HW, Langenhaun R, et al. Mast cell-mediated antigen presentation regulates CD8+ T cell effector functions. *Immunity* (2009) 31:665–76. doi: 10.1016/j.immuni.2009.08.022

35. Suurmond J, Habets KL, Dorjée AL, Huizinga TW, Toes RE. Expansion of Th17 cells by human mast cells is driven by inflammasome-independent IL-1β. J Immunol (2016) 197:4473-81. doi: 10.4049/jimmunol.1502640

36. Kritikou E, van der Heijden T, Swart M, van Duijn J, Slütter B, Wezel A, et al. Hypercholesterolemia induces a mast cell-CD4(+) T cell interaction in atherosclerosis. *J Immunol* (2019) 202:1531–9. doi: 10.4049/jimmunol.1800648

37. Gaudenzio N, Espagnolle N, Mars LT, Liblau R, Valitutti S, Espinosa E. Cellcell cooperation at the T helper cell/mast cell immunological synapse. *Blood* (2009) 114:4979–88. doi: 10.1182/blood-2009-02-202648

38. Suurmond J, van Heemst ,J, van Heiningen J, Dorjée AL, Schilham MW, van der Beek FB, et al. Communication between human mast cells and CD4(+) T cells through antigen-dependent interactions. *Eur J Immunol* (2013) 43:1758–68. doi: 10.1002/eji.201243058

39. Lotfi-Emran S, Ward BR, Le QT, Pozez AL, Manjili MH, woodfolk JA, et al. Human mast cells present antigen to autologous CD4(+) T cells. J Allergy Clin Immunol (2018) 141:311–321.e10. doi: 10.1016/j.jaci.2017.02.048

40. Mashiko S, Edelmayer RM, Bi Y, Olson LM, Wetter JB, Wang J, et al. Persistence of inflammatory phenotype in residual psoriatic plaques in patients on effective biologic therapy. *J Invest Dermatol* (2020) 140:1015–1025.e4. doi: 10.1016/j.jid.2019.09.027

41. Gaudenzio N, Laurent C, Valitutti S, Espinosa E. Human mast cells drive memory CD4+ T cells toward an inflammatory IL-22+ phenotype. J Allergy Clin Immunol (2013) 131:1400–7.e11. doi: 10.1016/j.jaci.2013.01.029

42. Mashiko S, Bouguermouh S, Rubio M, Baba N, Bissonnette R, Sarfati M. Human mast cells are major IL-22 producers in patients with psoriasis and atopic dermatitis. J Allergy Clin Immunol (2015) 136:351–9.e1. doi: 10.1016/j.jaci.2015.01.033

43. Cheung KL, Jarrett R, Subramaniam S, Salimi M, Gutowska-Owsiak D, Chen YL, et al. Psoriatic T cells recognize neolipid antigens generated by mast cell phospholipase delivered by exosomes and presented by CD1a. *J Exp Med* (2016) 213:2399–412. doi: 10.1084/jem.20160258

44. Noordenbos T, Blijdorp I, Chen S, Stap J, Mul E, Cañete JD, et al. Human mast cells capture, store, and release bioactive, exogenous IL-17A. *J Leukoc Biol* (2016) 100:453–62. doi: 10.1189/jlb.3HII215-542R

45. Eliasse Y, Leveque E, Garidou L, Battut L, Mckenzie B, Nocera T, et al. IL-17 (+) mast Cell/T helper cell axis in the early stages of acne. *Front Immunol* (2021) 12:740540. doi: 10.3389/fimmu.2021.740540

46. Chen S, Noordenbos T, Blijdorp I, Van Mens L, Ambarus CA, Vogels E, et al. Histologic evidence that mast cells contribute to local tissue inflammation in peripheral spondyloarthritis by regulating interleukin-17A content. *Rheumatol (Oxford)* (2019) 58:617–27. doi: 10.1093/rheumatology/key331

47. Lambert S, Hambro CA, Johnston A, Stuart PE, Tsoi LC, Nair RP, et al. Neutrophil extracellular traps induce human Th17 cells: Effect of psoriasisassociated TRAF3IP2 genotype. *J Invest Dermatol* (2019) 139:1245–53. doi: 10.1016/j.jid.2018.11.021

48. Herster F, Bittner Z, Archer NK, Dickhöfer S, Eisel D, Eigenbrod T, et al. Neutrophil extracellular trap-associated RNA and LL37 enable self-amplifying inflammation in psoriasis. *Nat Commun* (2020) 11:105. doi: 10.1038/s41467-019-13756-4

49. Agrahari G, Sah SK, Nguyen CT, Choi SS, Kim HY, Kim TY. Superoxide dismutase 3 inhibits LL-37/KLK-5-Mediated skin inflammation through modulation of EGFR and associated inflammatory cascades. *J Invest Dermatol* (2020) 140:656–665.e8. doi: 10.1016/j.jid.2019.08.434

50. Lin AM, Rubin CJ, Khandpur R, Wang JY, Riblett M, Yalavarthi S, et al. Mast cells and neutrophils release IL-17 through extracellular trap formation in psoriasis. *J Immunol* (2011) 187:490–500. doi: 10.4049/jimmunol.1100123

51. Köckritz-Blickwede MVON, Goldmann O, Thulin P, Heinemann K, Norrby-Teglund A, Rohde M, et al. Phagocytosis-independent antimicrobial activity of mast cells by means of extracellular trap formation. *Blood* (2008) 111:3070–80. doi: 10.1182/blood-2007-07-104018

52. Shefler I, Salamon P, Reshef T, Mor A, Mekori YA. T Cell-induced mast cell activation: a role for microparticles released from activated T cells. *J Immunol* (2010) 185:4206–12. doi: 10.4049/jimmunol.1000409

53. Shefler I, Pasmanik-Chor M, Kidron D, Mekori YA, Hershko AY. T Cellderived microvesicles induce mast cell production of IL-24: relevance to inflammatory skin diseases. *J Allergy Clin Immunol* (2014) 133:217–24.e1-3. doi: 10.1016/j.jaci.2013.04.035

54. Moreau JM, Velegraki M, Bolyard C, Rosenblum MD, Li Z. Transforming growth factor- β 1 in regulatory T cell biology. *Sci Immunol* (2022) 7:eabi4613. doi: 10.1007/s10157-017-1516-4

55. Hartwig T, Zwicky P, Schreiner B, Yawalkar N, Cheng P, Navarini A, et al. Regulatory T cells restrain pathogenic T helper cells during skin inflammation. *Cell Rep* (2018) 25:3564–3572.e4. doi: 10.1016/j.celrep.2018.12.012

56. Sugiyama H, Gyulai R, Toichi E, Garaczi E, Shimada S, Stevens SR, et al. Dysfunctional blood and target tissue CD4+CD25high regulatory T cells in psoriasis: mechanism underlying unrestrained pathogenic effector T cell proliferation. *J Immunol* (2005) 174:164–73. doi: 10.4049/jimmunol.174.1.164

57. Priyadarssini M, Chandrashekar L, Rajappa M. Effect of methotrexate monotherapy on T-cell subsets in the peripheral circulation in psoriasis. *Clin Exp Dermatol* (2019) 44:491–7. doi: 10.1111/ced.13795

58. Piconese S, Gri G, Tripodo C, Musio S, Gorzanelli A, Frossi B, et al. Mast cells counteract regulatory T-cell suppression through interleukin-6 and OX40/ OX40L axis toward Th17-cell differentiation. *Blood* (2009) 114:2639–48. doi: 10.1182/blood-2009-05-220004

59. Liu WX, Wang Y, Sang LX, Zhang S, Wang T, Zhou F, et al. Chymase inhibitor TY-51469 in therapy of inflammatory bowel disease. *World J Gastroenterol* (2016) 22:1826–33. doi: 10.3748/wjg.v22.i5.1826

60. Kashyap M, Thornton AM, Norton SK, Barnstein B, Macey M, Brenzovich J, et al. Cutting edge: CD4 T cell-mast cell interactions alter IgE receptor expression and signaling. J Immunol (2008) 180:2039–43. doi: 10.4049/jimmunol.180.4.2039

61. Bovenschen HJ, van Vlijmen-Willems IM, van de Kerkhof PC, van Erp PE. Identification of lesional CD4+ CD25+ Foxp3+ regulatory T cells in psoriasis. *Dermatology* (2006) 213:111–7. doi: 10.1159/000093849

62. Hendriks AG, Keijsers RR, Seyger MM, van Erp PE, van de Kerkhof PC. Are newly discovered drivers of immune-mediated skin disorders expressed in normal skin regenerating from standardized surface injury? *Dermatology* (2014) 228:255– 60. doi: 10.1159/000358008

63. Suttle MM, Ritanen RL, Harvima IT. Increase in FoxP3-positive cells and their contacts with mast cells in köbner-negative patients with psoriasis after tapestripping. *Acta Derm Venereol* (2019) 99:103–4. doi: 10.2340/00015555-3023

64. Ni X, Lai Y. Crosstalk between keratinocytes and immune cells in inflammatory skin diseases. *Explor Immunol* (2021) 1:418–31. doi: 10.37349/ei.2021.00028

65. Yamamoto T, Katayama I, Nishioka K. Possible contribution of stem cell factor in psoriasis vulgaris. *J Dermatol Sci* (2000) 24:171–6. doi: 10.1016/S0923-1811(00)00095-5

66. Wang Z, Mascarenhas N, Eckmann L, Miyamoto Y, Sun X, Kawakami T, et al. Skin microbiome promotes mast cell maturation by triggering stem cell factor production in keratinocytes. *J Allergy Clin Immunol* (2017) 139:1205–1216.e6. doi: 10.1016/j.jaci.2016.09.019

67. Cho KA, Park M, Kim YH, Woo SY. Th17 cell-mediated immune responses promote mast cell proliferation by triggering stem cell factor in keratinocytes. *Biochem Biophys Res Commun* (2017) 487:856–61. doi: 10.1016/j.bbrc.2017.04.141

68. Cho KA, Kim HJ, Kim YH, Park M, Woo SY. Dexamethasone promotes keratinocyte proliferation by triggering keratinocyte growth factor in mast cells. *Int Arch Allergy Immunol* (2019) 179:53–61. doi: 10.1159/000494624

69. Zeng F, Chen H, Chen L, Mao J, Cai S, Xiao Y, et al. An autocrine circuit of IL-33 in keratinocytes is involved in the progression of psoriasis. *J Invest Dermatol* (2021) 141:596–606.e7. doi: 10.1016/j.jid.2020.07.027

 Aoki R, Kawamura T, Goshima F, Ogawa Y, Nakae S, Moriishi K, et al. The alarmin IL-33 derived from HSV-2-Infected keratinocytes triggers mast cellmediated antiviral innate immunity. *J Invest Dermatol* (2016) 136:1290–2. doi: 10.1016/j.jid.2016.01.030

71. Wang JX, Kaieda S, Ameri S, Fishgal N, Dwyer D, Dellinger A, et al. IL-33/ ST2 axis promotes mast cell survival *via* BCLXL. *Proc Natl Acad Sci USA* (2014) 111:10281-6. doi: 10.1073/pnas.1404182111

72. Hueber AJ, Alves-Filho JC, Asquith DL, Michels C, Millar NL, Reilly JH, et al. IL-33 induces skin inflammation with mast cell and neutrophil activation. *Eur J Immunol* (2011) 41:2229–37. doi: 10.1002/eji.201041360

73. Theoharides TC, Zhang B, Kempuraj D, Tagen M, Vasiadi M, Angelidou A, et al. IL-33 augments substance p-induced VEGF secretion from human mast cells and is increased in psoriatic skin. *Proc Natl Acad Sci U.S.A.* (2010) 107:4448–53. doi: 10.1073/pnas.1000803107

74. Taracanova A, Alevizos M, Karagkouni A, Weng Z, Norwitz E, Conti P, et al. SP and IL-33 together markedly enhance TNF synthesis and secretion from human mast cells mediated by the interaction of their receptors. *Proc Natl Acad Sci USA* (2017) 114:E4002–e4009. doi: 10.1073/pnas.1524845114

75. Redhu D, Franke K, Aparicio-Soto M, Kumari V, Pazur K, Illerhaus A, et al. Mast cells instruct keratinocytes to produce thymic stromal lymphopoietin: Relevance of the tryptase/protease-activated receptor 2 axis. *J Allergy Clin Immunol* (2022) 149(6):2053–61. doi: 10.1016/j.jaci.2022.01.029

76. Kakurai M, Demitsu T, Umemoto N, Kobayashi Y, Inoue-Narita T, Fujita N, et al. Vasoactive intestinal peptide and inflammatory cytokines enhance vascular endothelial growth factor production from epidermal keratinocytes. *Br J Dermatol* (2009) 161:1232–8. doi: 10.1111/j.1365-2133.2009.09439.x

77. Larabee CM, Neely OC, Domingos AI. Obesity: a neuroimmunometabolic perspective. Nat Rev Endocrinol (2020) 16:30-43. doi: 10.1038/s41574-019-0283-6

78. Oishi Y, Manabe I. Integrated regulation of the cellular metabolism and function of immune cells in adipose tissue. *Clin Exp Pharmacol Physiol* (2016) 43:294–303. doi: 10.1111/1440-1681.12539

79. Divoux A, Moutel S, Poitou C, Lacasa D, Veyrie N, Aissat A, et al. Mast cells in human adipose tissue: link with morbid obesity, inflammatory status, and diabetes. *J Clin Endocrinol Metab* (2012) 97:E1677-85. doi: 10.1210/jc.2012-1532

80. Gurung P, Moussa K, Adams-HUET B, Devaraj S, Jialal I. Increased mast cell abundance in adipose tissue of metabolic syndrome: relevance to the proinflammatory state and increased adipose tissue fibrosis. *Am J Physiol Endocrinol Metab* (2019) 316:E504–e509. doi: 10.1152/ajpendo.00462.2018

81. Fantuzzi G. Adipose tissue, adipokines, and inflammation. J Allergy Clin Immunol (2005) 115:911-9. doi: 10.1016/j.jaci.2005.02.023

82. Xu C, Ji J, Su T, Wang HW, Su ZL, et al. The Association of Psoriasis and Obesity: Focusing on IL-17A-Related Immunological Mechanisms. *Int J Dermatol Venereol* (2021) 4:116–21. doi: 10.1097/JD9.00000000000155

83. Xue K, Liu H, Jian Q, Liu B, Zhu D, Zhang M, et al. Leptin induces secretion of pro-inflammatory cytokines by human keratinocytes *in vitro*-a possible reason for increased severity of psoriasis in patients with a high body mass index. *Exp Dermatol* (2013) 22:406–10. doi: 10.1111/exd.12162

84. Huang H, Shen E, Tang S, Tan X, Guo X, Wang Q, et al. Increased serum resistin levels correlate with psoriasis: a meta-analysis. *Lipids Health Dis* (2015) 14:44. doi: 10.1186/s12944-015-0039-9

85. Kyriakou A, Patsatsi A, Sotiriadis D, Goulis DG. Serum leptin, resistin, and adiponectin concentrations in psoriasis: A meta-analysis of observational studies. *Dermatology* (2017) 233:378–89. doi: 10.1159/000481882

86. Stjernholm T, Ommen P, Langkilde A, Johansen C, Iversen L, Rosada C, et al. Leptin deficiency in mice counteracts imiquimod (IMQ)-induced psoriasislike skin inflammation while leptin stimulation induces inflammation in human keratinocytes. *Exp Dermatol* (2017) 26:338–45. doi: 10.1111/exd.13149

87. Ommen P, Stjernholm T, Kragstrup T, Raaby L, Johansen C, Stenderup K, et al. The role of leptin in psoriasis comprises a proinflammatory response by the dermal fibroblast. *Br J Dermatol* (2016) 174:187–90. doi: 10.1111/bjd.13969

88. Watanabe Y, Yamaguchi Y, Takamura N, Komitsu N, Aihara M. Leptin induces interleukin-6 production in keratinocytes *via* decreased expression of caveolin-1: a possible link between obesity and psoriatic inflammation. *Br J Dermatol* (2020) 183:768–70. doi: 10.1111/bjd.19133

89. Żelechowska P, Agier J, Kozłowska E, Brzezińska-Błaszczyk E. Mast cells participate in chronic low-grade inflammation within adipose tissue. *Obes Rev* (2018) 19:686–97. doi: 10.1111/obr.12670

90. Zhou Y, Yu X, Chen H, Sjöberg S, Roux J, Zhang L, et al. Leptin deficiency shifts mast cells toward anti-inflammatory actions and protects mice from obesity and diabetes by polarizing M2 macrophages. *Cell Metab* (2015) 22:1045–58. doi: 10.1016/j.cmet.2015.09.013

91. Żelechowska P, Brzezińska-Błaszczyk E, Wiktorska M, Różalska S, Wawrocki S, Kozłowska E, et al. Adipocytokines leptin and adiponectin function as mast cell activity modulators. *Immunology* (2019) 158:3–18. doi: 10.1111/imm.13090

92. Żelechowska P, Wiktorska M, różalska S, Stasikowska-Kanicka O, Wągrowska-Danilewicz M, Agier J, et al. Leptin receptor is expressed by tissue mast cells. *Immunol Res* (2018) 66:557–66. doi: 10.1007/s12026-018-9029-0

93. Silswal N, Singh AK, Aruna B, Mukhopadhyay S, Ghosh S, Ehtesham NZ. Human resistin stimulates the pro-inflammatory cytokines TNF-alpha and IL-12 in macrophages by NF-kappaB-dependent pathway. *Biochem Biophys Res Commun* (2005) 334:1092–101. doi: 10.1016/j.bbrc.2005.06.202

94. Shibata S, Tada Y, Hau CS, Mitsui A, Kamata M, Asano Y, et al. Adiponectin regulates psoriasiform skin inflammation by suppressing IL-17 production from $\gamma\delta$ -T cells. *Nat Commun* (2015) 6:7687. doi: 10.1038/ncomms8687

95. Stassen M, Hartmann AK, Delgado SJ, Dehmel S, Braun A. Mast cells within cellular networks. *J Allergy Clin Immunol* (2019) 144:S46–s54. doi: 10.1016/j.jaci.2019.01.031

96. Aroni K, Tsagroni E, Kavantzas N, Patsouris E, Ioannidis E. A study of the pathogenesis of rosacea: how angiogenesis and mast cells may participate in a complex multifactorial process. *Arch Dermatol Res* (2008) 300:125–31. doi: 10.1007/s00403-007-0816-z

97. Voisin T, Perner C, Messou MA, Shiers S, Ualiyeva S, Kanaoka Y, et al. The CysLT(2)R receptor mediates leukotriene C(4)-driven acute and chronic itch. *Proc Natl Acad Sci U.S.A.* (2021) 118:e2022087118. doi: 10.1073/pnas.2022087118

98. Nakamura M, Toyoda M, Morohashi M. Pruritogenic mediators in psoriasis vulgaris: comparative evaluation of itch-associated cutaneous factors. *Br J Dermatol* (2003) 149:718–30. doi: 10.1046/j.1365-2133.2003.05586.x

99. Riol-Blanco L, Ordovas-Montanes J, Perro M, Naval E, Thiriot A, Alvarez D, et al. Nociceptive sensory neurons drive interleukin-23-mediated psoriasiform skin inflammation. *Nature* (2014) 510:157–61. doi: 10.1038/nature13199

 Marek-Jozefowicz I, Czajkowski R, Borkowska A, Nedoszytko B, ŻmijewskiMA, et al. The brain-skin axis in psoriasis-psychological, psychiatric, hormonal, and dermatological aspects. Int J Mol Sci (2022) 23:669. doi: 10.3390/ijms23020669

101. Tagen M, Stiles L, Kalogeromitros D, Gregoriou S, Kempuraj D, Makris M, et al. Skin corticotropin-releasing hormone receptor expression in psoriasis. *J Invest Dermatol* (2007) 127:1789–91. doi: 10.1038/sj.jid.5700757

102. Mcneil BD, Pundir P, Meeker S, Han L, Undem BJ, Kulka M, et al. Identification of a mast-cell-specific receptor crucial for pseudo-allergic drug reactions. *Nature* (2015) 519:237-41. doi: 10.1038/nature14022

103. Green DP, Limjunyawong N, Gour N, Pundir P, Dong X. And pain. Neuron (2019) 101:412-420.e3. doi: 10.1016/j.neuron.2019.01.012

104. Gaudenzio N, Sibilano R, Marichal T, Starkl P, Reber LL, Cenac N, et al. Different activation signals induce distinct mast cell degranulation strategies. *J Clin Invest* (2016) 126:3981–98. doi: 10.1172/JCI85538

105. Franke K, Wang Z, Zuberbier T, Babina M. Cytokines stimulated by IL-33 in human skin mast cells: Involvement of NF- κ B and p38 at distinct levels and potent Co-operation with FcεRI and MRGPRX2. *Int J Mol Sci* (2021) 22:3580. doi: 10.3390/ijms22073580