



The New Frontier of Host-Directed Therapies for *Mycobacterium avium* Complex

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Mycobacterium avium complex (MAC) is an increasingly important cause of morbidity and mortality, and is responsible for pulmonary infection in patients with underlying lung disease and disseminated disease in patients with AIDS. MAC has evolved various virulence strategies to subvert immune responses and persist in the infected host. Current treatment for MAC is challenging, requiring a combination of multiple antibiotics given over a long time period (for at least 12 months after negative sputum culture conversion). Moreover, even after eradication of infection, many patients are left with residual lung dysfunction. In order to address similar challenges facing the management of patients with tuberculosis, recent attention has focused on the development of novel adjunctive, host-directed therapies (HDTs), with the goal of accelerating the clearance of mycobacteria by immune defenses and reducing or reversing mycobacterial-induced lung damage. In this review, we will summarize the evidence supporting specific adjunctive, HDTs for MAC, with a focus on the repurposing of existing immune-modulatory agents targeting a variety of different cellular pathways. We also highlight areas meriting further investigation.

Keywords: nontuberculous mycobacteria (NTM), *Mycobacterium avium* complex, host-directed therapy, *Mycobacterium tuberculosis*, drug repurposing

OPEN ACCESS

Edited by:

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Specialty section:

This article was submitted to
Vaccines and Molecular Therapeutics,
a section of the journal
Frontiers in Immunology

Received: 29 October 2020

Accepted: 14 December 2020

Published: 22 January 2021

Citation:

Crilly NP, Ayeh SK and Karakousis PC
(2021) The New Frontier of Host-
Directed Therapies for *Mycobacterium*
avium Complex.
Front. Immunol. 11:623119.
doi: 10.3389/fimmu.2020.623119

INTRODUCTION

Nontuberculous mycobacteria (NTM), including organisms of the *Mycobacterium avium* complex (MAC), represent a significant and growing threat to human health worldwide. Since the beginning of the AIDS epidemic in the 1980s, the prevalence of MAC infection has increased substantially worldwide (1). MAC is widely distributed in the environment, including in water and soil, and is transmitted via inhalation into the respiratory tract and via ingestion into the GI tract (2). The most common clinical syndromes caused by MAC are pulmonary infection in patients with underlying lung disease, as well as disseminated disease in the severely immunocompromised (3, 4). A recent review of MAC pulmonary disease worldwide reported a five-year all-cause mortality rate of 27% (5).

In addition to the virulence factors common to all mycobacteria, MAC possesses several unique features which may contribute to pathogenesis. For example, MAC demonstrates increased resistance

to phagosome-lysosome fusion and oxidative damage in murine macrophages, suggesting a unique ability to survive within activated macrophages (6). MAC can escape from macrophages undergoing apoptosis and survive extracellularly, evading the cytotoxic response necessary to eliminate intracellular bacteria (7). MAC also expresses several unique glycopeptidolipids, which may modulate macrophage signaling cascades, thereby preventing an effective inflammatory response (8).

Treatment of MAC is challenging. Current treatment recommendations vary depending on the underlying conditions, severity of disease, and *in vitro* susceptibility profile. Macrolide-susceptible pulmonary disease is generally treated with a three-drug regimen, which includes a macrolide, ethambutol and a rifamycin, for at least 12 months after negative sputum-culture conversion (9). MAC often exhibits resistance to first-line antibiotics, and *in vitro* susceptibility testing for non-macrolide drugs has poor correlation with clinical efficacy. MAC pulmonary infection can present as cavitary disease with long-term respiratory sequelae. A milder form of the disease, which manifests as fibronodular bronchiectasis has a slower progression, but has been linked to increased mortality (10).

In the face of the increasing prevalence, high mortality, and treatment challenges associated with MAC infections, new therapeutic options are urgently needed. A promising avenue of research is that of host-directed therapies (HDTs). HDTs are adjuncts to antimicrobial therapy, differing from the latter in that they target host processes rather than the pathogen itself. The goal of HDTs is to boost protective immune responses, especially those inhibited or otherwise modified by the pathogen, and prevent excessive pathological inflammation (11, 12). Unlike novel antibacterial agents, they also confer the advantage of not contributing to drug resistance or cross-resistance to conventional antibiotics (12). Although HDTs are an active area of investigation in the therapy of tuberculosis (TB), as well as many non-mycobacterial infectious diseases (11–15), there has been a relative dearth of research into the potential of HDTs as adjunctive therapies for disease caused by MAC (16).

In the current review, we summarize HDT agents which are currently under investigation for MAC disease, as well as other HDTs and potentially targetable host pathways, which have not been investigated directly for MAC, but which show promise for future research.

IMPROVEMENT OF ANTIMYCOBACTERIAL IMMUNITY

Enhancing Autophagy: mTOR Inhibitors

Autophagy is a key self-degradative process in which the cytoplasmic contents of a cell are taken up by autophagosomes, trafficked to the lysosome, and digested (17). Although basal levels of this process occur in every cell, stress conditions, such as nutrient deficiency or pathogen infection, induce autophagy as a way of establishing homeostasis (18, 19). Autophagy plays a role in multiple physiological and pathological pathways, including the clearance of mycobacteria and other intracellular pathogens (17).

Initiation of autophagy is dependent on the Unc-51-like kinase-1 (ULK1) complex. This initiator complex is, in turn, regulated by the master regulator of autophagy, mammalian target of rapamycin (mTOR). mTOR plays a critical role in cellular metabolism, promoting anabolism and suppressing catabolic processes, such as autophagy (20). mTOR signaling is complex and can be activated or inhibited by a wide variety of molecules and signaling pathways. Nutrient states, particularly amino acid levels at the cellular level, serve as the main signal for mTOR activation. In nutrient-rich states, mTOR exerts an inhibitory effect on the ULK1 complex, leading to suppression of autophagy (21). Because of its important role in metabolism and cell growth, mTOR inhibition is a therapeutic target for a number of diseases, including autoimmune disorders and cancer (22). Rapamycin and other analogs directly inhibit mTOR activity, and vitamin D blocks upstream signaling to activate mTOR (22, 23). During *Mycobacterium tuberculosis* (Mtb) infection, the activation of both intracellular or extracellular surface pattern recognition receptors (PRRs) by certain unique Mtb-associated molecules, such as lipomannan, lipoarabinomannan, phthiocerol dimycocerosate (PDIM), lipoproteins, mycolic acid and Mtb DNA/RNA, induces autophagy (24, 25). Given that autophagy plays an important role in mycobacterial clearance, and MAC can survive intracellularly by blocking phagosome-lysosome fusion, enhancing autophagy through inhibition of the mTOR pathway appears to be an attractive HDT strategy (26, 27).

To date, there has been little research on targeting autophagy to improve host control of MAC infection. Early *et al.* reported that induction of autophagy by lactoferrin increases MAC killing by macrophages and renders the bacteria more susceptible to ethambutol, suggesting that autophagy is worthy of further investigation as an HDT target (28). Although they have not been studied in the context of MAC infection, mTOR inhibitors have been explored as HDTs for Mtb, with mixed results (29). Most data from *in vitro* studies have suggested that mTOR inhibition may result in enhanced intracellular killing of Mtb, however there is also some contrasting evidence to suggest that induction of autophagy results in increased Mtb growth, especially in the context of Mtb/HIV co-infection (30, 31). Vitamin D, an upstream inhibitor of mTOR signaling, also has shown some promise as an HDT for TB, although clinical trials do not show a consistent benefit, and it has not been investigated specifically against MAC (32).

Aside from autophagy, mTOR is involved in multiple metabolic and immunological pathways, which could affect mycobacterial pathogenesis and immunity. As a whole, the role of mTOR and autophagy in MAC infection remains largely unexplored, and further research is required to evaluate its suitability as an HDT target.

Blocking the PD-1/PD-L1 Pathway: Anti-PD-1/PD-L1 Therapy

The Programmed Cell Death Protein-1 (PD-1) and its ligand, PD-L1, are the major components of the PD-1/PD-L1 pathway, an immune checkpoint, which regulates peripheral immune tolerance and suppresses inflammation (33). PD-1 is expressed

on multiple cell types, including activated T cells, B cells, natural killer cells, and macrophages. PD-L1 is expressed on nonlymphoid cells. Binding of PD-1 to PD-L1 inhibits proliferation and effector functions of T and B cells, preventing self-reactivity (34). PD-L1 is highly expressed on tumor cells and virus-infected cells, conferring resistance to cell-mediated immunity. PD-L1 is also expressed on macrophages and plays a role in regulating immunosuppressive and pro-inflammatory activity. PD-L1 signaling in tumor-associated macrophages induces an immunosuppressive phenotype (35). Recently, the PD-1/PD-L1 pathway has become the subject of extensive research in cancer immunotherapy, as PD-1/PD-L1 antibody blockade has demonstrated efficacy in inducing cell-mediated immunity against multiple cancer types. Treatment of tumor-associated macrophages with anti-PD-L1 antibodies confers a pro-inflammatory phenotype, with increased expression of inducible nitric oxide synthase (iNOS), MHC II, TNF- α , and CD40 (36, 37). This is particularly important, since TNF- α and iNOS are critical effector mechanisms in the killing of intracellular mycobacteria, including MAC by macrophages (38). In patients with MAC pulmonary disease, expression of PD-1 by CD4 T cells is directly correlated with disease severity (39). An analysis of peripheral blood mononuclear cell (PBMC) function in such patients found that expression of PD-1 and PD-L1 were increased in lymphocytes of infected patients, which correlated with increased lymphocyte apoptosis compared to lymphocytes from healthy controls (40). Treatment of PBMCs obtained from MAC patients with anti-PD-1 and PD-L1 antibodies resulted in increased IFN- γ production and reduced T-cell apoptosis compared to PBMCs from healthy controls (40). These data suggest that PD-1/PDL-1 therapy could rescue immune cells from an immunosuppressive phenotype, allowing an improved immune response against MAC.

Although anti-PD-1 therapy may hold promise for treatment of MAC, there is some evidence that PD-1 is necessary for mycobacterial immunity, particularly against Mtb. Thus, mice deficient in PD-1 are more susceptible to Mtb infection (41). In Mtb granulomas, PD-1 is expressed in stable, cellular granulomas, but not in caseating ones, suggesting that it plays a role in granuloma maintenance. In a three-dimensional cell culture model, PD-1 inhibition led to increased Mtb growth, possibly due to excessive TNF- α expression (42).

The potential of anti-PD-1/PDL-1 therapy to improve the immune response to MAC remains to be investigated, both *in vitro* and *in vivo*. As anti-PD-1/PD-L1 therapy becomes more common in cancer therapy, retrospective analyses of its effect on patient susceptibility to MAC disease and clinical outcomes following MAC therapy may be useful.

Heme Oxygenase Inhibition

Heme oxygenase (HO-1) is an antioxidant enzyme that catalyzes the conversion of heme into carbon monoxide, biliverdin and iron (43, 44). Apart from its role in cytoprotection, HO-1 has been shown to regulate cell proliferation, differentiation, and apoptosis (44). The induction of pulmonary HO-1 is associated

with TB disease (45), suggesting its potential utility as a diagnostic biomarker. Although its role in TB pathogenesis is not fully understood, experimental data in Mtb-infected mice have shown that lung bacterial loads decrease following HO-1 inhibition by the metalloporphyrin, SnPPIX (45). The same study found that a combination of an HO-1 inhibitor, SnPPIX and antimycobacterial therapy enhanced T-cell-dependent pathogen clearance. Clinical data have shown that plasma HO-1 levels decline following successful TB treatment (46).

As in the case of Mtb infection, HO-1 has been found to be elevated during MAC infection in BALB/c mice (47). Consistent with a host protective role in resisting MAC infection, mycobacterial burden in the liver, lungs and spleen was significantly higher and the disease was more likely to be disseminated in mice with HO-1 deficiency compared to HO-1 homozygous or heterozygous mice (47, 48). Further investigation is required to determine how HO-1 activity is regulated during MAC infection, and whether HO-1 inhibition is a promising HDT in the context of MAC.

IFN- γ Therapy

IFN- γ plays a significant role in immunity against Mycobacterium infections. In contrast to type I IFNs (α and β), which are made by virus-infected cells, IFN- γ is produced by activated T cells, NK cells, and macrophages, leading to the activation of phagocytes, stimulation of antigen presentation to T cells, and regulation of several other cellular functions, including proliferation, apoptosis, and cell adhesion (49). In particular, IFN- γ induces the expression of iNOS (50) and the respiratory burst enzyme NADPH-dependent phagocyte oxidase (51), thereby enhancing the mycobactericidal activity of macrophages. Mice with mutations in the IFN- γ receptor have been shown to have increased susceptibility to intracellular pathogens (52). Pre-treatment of intestinal and peritoneal-derived macrophages with IFN- γ produced both bactericidal and bacteriostatic activity against MAC following infection of these cells (53, 54). Although *in vivo* treatment of beige and Swiss-Webster mice with recombinant murine IFN- γ did not alter the course of visceral MAC infection (55), the bactericidal activity of clofazimine against MAC was enhanced in beige mice pre-treated with IFN- γ (54).

Mutations in the IFN- γ receptor gene or anti-IFN- γ autoantibodies confer increased susceptibility to disseminated NTM infections in humans (56–58). IFN- α , which, like IFN- γ , signals through STAT1, activating many common downstream effector genes, has shown some promise in treating patients with IFN- γ signaling defects and disseminated mycobacterial disease (59). In a study of 7 patients with disseminated MAC infection, subcutaneous administration of IFN- γ , in combination with conventional medical treatment, resulted in improvement in symptoms, and pathological and radiological findings, and also reduced the need for medical procedures, such as paracentesis following 8 weeks of treatment (60). Aerosolized IFN- γ has shown some promise in treating patients with TB and idiopathic pulmonary fibrosis, and is worthy of study in patients with pulmonary MAC (61).

PREVENTION OF EXCESSIVE AND PATHOLOGICAL INFLAMMATION

Suppressing Excessive TNF- α Activation: Anti-TNF Antibodies

Tumor necrosis factor alpha (TNF- α) is a pro-inflammatory cytokine which is upregulated during MAC and Mtb infection and plays an essential role in antimycobacterial immunity (62). During mycobacterial infection, T cells, macrophages, and dendritic cells produce TNF- α in response to multiple signaling pathways (63). TNF- α signaling is complex, and the cytokine serves multiple functions, including in the formation and maintenance of granulomas, as evidenced by the observation that mice deficient in TNF- α or receiving anti-TNF- α therapy produce defective granulomas following mycobacterial infection (64, 65). TNF- α also promotes killing of intracellular mycobacteria by macrophages, as the TNF blockers adalimumab and infliximab suppressed phagosome maturation in primary human PBMCs in the presence or absence of IFN- γ (66). Moreover, TNF- α serves macrophage antimicrobial functions by activating reactive oxygen and nitrogen species (67). Treatment with anti-TNF- α antibody has been associated with decreased resistance to MAC infection in mice (68).

Although TNF- α is required for an effective immune response, excessive TNF- α production has deleterious pathological effects. Thus, when its production is properly regulated, TNF- α induces apoptosis of Mtb-infected cells by recruiting Fas-associated protein with death domain (FADD) and subsequent activation of effector caspases and signal-regulating kinase 1 (ASK1), thus favoring mycobacterial clearance (63, 69, 70). However, when produced in excessive amounts, TNF- α results in necrosis of Mtb-infected macrophages and hyperinflammation through activation of serine/threonine-protein (RIP)1/3 kinases and mitochondrial reactive oxygen species (ROS) production (70–72). TNF- α also induces necroptosis, a highly inflammatory form of cell death, which could contribute to pathological inflammation (73).

Because of its roles in mycobacterial immunity and pathology, TNF- α has been a focus of HDT investigation. Multiple anti-TNF antibodies and TNF soluble receptors have been approved for use in humans to block TNF- α activity, and are primarily used to treat autoinflammatory conditions, such as rheumatoid arthritis. TNF blockers have shown some promise as HDTs for mycobacterial infections. Combined use of the TNF- α receptor inhibitor etanercept with antibiotics decreased the lung burden of Mtb and reduced TB-associated lung pathology in infected mice compared to antibiotics alone (74). However, the role of anti-TNF therapy in clinical cases of mycobacterial infection is controversial. Patients receiving anti-TNF therapy are at increased risk for developing disease due to Mtb and MAC (75–77). After a diagnosis of TB or MAC disease is made, anti-TNF therapy is usually halted at least until anti-mycobacterial therapy has been initiated and the infection is under control. On the other hand, there are several reports of TB patients experiencing clinical exacerbation upon discontinuation of anti-TNF treatment, and improvement of disease following its reinstatement (78–80).

In addition, a subset of MAC-infected patients show favorable outcomes if anti-TNF therapy is maintained throughout treatment (76). However, it is uncertain in these cases whether anti-TNF therapy contributed as an adjunctive HDT or by ameliorating the underlying autoimmune disease.

The roles of TNF- α in mycobacterial immunity and disease are complex, and the therapeutic potential and risk of inhibiting TNF- α function during MAC infection require further investigation. Given the relatively long half-lives of most TNF blockers relative to antibiotics, there is concern over sudden stoppage of all treatment by patients, resulting in the unopposed anti-TNF activity and possible worsening of infection (81). Since TNF- α interacts with multiple other signaling pathways, further research is also needed to identify other cytokines which, if targeted in tandem with TNF- α , could hold promise as HDTs.

Broad Suppression of Inflammation: Nonsteroidal Anti-Inflammatory Drugs and Corticosteroids

Excessive and chronic inflammation is an important factor in the progression of mycobacterial disease (82). Thus, the broad inhibition of the inflammatory response by non-steroidal anti-inflammatory drugs (NSAIDs) or corticosteroids is an attractive HDT strategy. NSAIDs have been well-studied as adjunctive therapies for TB, with a protective effect, both in animal models and in human disease, when used in conjunction with antibiotics (83). There are multiple proposed mechanisms for these effects. NSAIDs suppress the excessive recruitment of neutrophils to granulomas, which can be responsible for destructive inflammation (84, 85). By reducing prostaglandin E2 (PGE2) expression, NSAIDs also inhibit phagocytosis and killing of mycobacteria during late TB (86). NSAIDs have anti-thrombotic effects, which may prevent the hypercoagulable state occasionally observed with severe TB (87, 88). Despite their relatively well-characterized role as an adjunctive therapy for TB, there has been little research into NSAIDs as HDTs for MAC. The NSAID diclofenac sodium modulates multiple cytokines in MAC-stimulated macrophages but does not improve bacterial clearance by macrophages or infected mice (89). Although NSAIDs can prevent destructive inflammation, they might also inhibit an effective immune response. This is especially concerning for MAC, since an immunocompromised state is a major risk factor for disseminated MAC disease (10). NSAIDs have not been causally linked to MAC disease, but long-term NSAID use has been identified as a possible predisposing factor in at least one case (90).

Corticosteroids are some of the earliest HDTs used for mycobacterial disease and may be useful in treating patients with late-stage and extrapulmonary TB (91, 92). In particular, short-term steroid use, by reducing inflammation caused by antibiotic-mediated killing of mycobacteria and accompanying increased intracranial pressure, has been shown to improve mortality by as much as 25% in patients with tuberculous meningitis (93). Similar to NSAIDs, the beneficial effect of corticosteroids is primarily attributed to the suppression of pathological inflammation. Corticosteroids exert their anti-inflammatory effects through a variety of mechanisms, including

by inducing transcription of anti-inflammatory genes, such as annexin-1, IL-10 and $\text{I}\kappa\text{B-}\alpha$ (inhibitor of NF- κB), by direct interacting with NF- κB , AP-1 and other immunomodulatory transcription factors, inhibiting maturation and differentiation of antigen presentation cells with reduced sensitivity to T cell regulation, and promoting the formation of macrophages with anti-inflammatory properties (94).

The use of corticosteroids as an HDT for MAC disease is somewhat controversial, due to their immunosuppressive effects and the lack of controlled studies (95–97). Although there is a significant body of research on the use of corticosteroids in reducing inflammation due to a variety of infectious diseases, their specific role as an adjunctive HDT for MAC disease has not been studied. Further research is required to understand the effects of corticosteroids on MAC infection on the molecular, cellular, and organismal level, to determine whether their use is justified or contraindicated in specific stages of MAC disease.

MULTIPLE MECHANISMS OF ACTION

Targeting Lipid Metabolism and Inducing Autophagy: Statins

The 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase inhibitors (statins) are a class of lipid-lowering medications, which have shown promise as HDTs for TB (98). PBMCs from patients with familial hypercholesterolemia receiving statin therapy demonstrate resistance to *ex vivo* Mtb infection compared to those from untreated donors (99). Adjunctive therapy with simvastatin enhanced the bactericidal activity of the first-line anti-mycobacterial regimen in a mouse model of chronic TB and shortened the duration of curative treatment in a murine model of TB relapse (100, 101). Consistent with a class effect of statins, pravastatin adjunctive therapy showed a dose-dependent reduction in bacillary lung burden and decreased lung inflammation in conjunction with front-line chemotherapy in a mouse model of chronic TB (102). Mechanistically, statins reduce the formation of lipid droplets in foamy macrophages, which may serve as a nutrient source for intracellular Mtb and contribute to antibiotic tolerance (99, 103). However, the primary HDT mechanism of action of statins likely involves the promotion of phagosome maturation and autophagy, thereby improving clearance of Mtb by infected macrophages (99). Statins enhance autophagy of Mtb-infected macrophages by blocking mTORC1, activating AMP-activated protein kinase (AMPK) and favoring nuclear translocation of transcription factor EB (TFEB) (104). Although the role of lipid-laden, foamy macrophages in MAC pathogenesis is less well understood than in TB, morphologically similar phenotypes have also been described in MAC-infected macrophages, and it is possible that statins could have similar HDT effects (105).

Activation of AMPK and Potentiation of Macrophage Effector Function: Metformin

Multiple studies have found that use of the anti-hyperglycemic drug metformin reduces the risk of TB and improves clinical outcomes in patients with diabetes mellitus (106, 107).

Experimental evidence indicates that metformin has multiple host-directed effects, which may promote clearance of MAC. The drug enhances mycobacterial killing in human PBMCs by promoting autophagy and phagosome-lysosome fusion, as well as by selectively increasing mitochondrial ROS production (108). Metformin has a dose-dependent inhibitory effect on intracellular replication of mycobacteria through activation of the adenosine monophosphate-activated protein kinase (AMPK) signaling pathway (109). Metformin also suppresses TNF- α expression in human monocytes (110). In Mtb-infected mice, metformin adjunctive therapy is associated with reduced chronic lung inflammation, enhanced immune responses, and improved efficacy of antibiotics (111, 112). In contrast, Dutta *et al.* showed that adjunctive therapy with human-equivalent doses of metformin did not enhance the bactericidal or sterilizing activities of the first line antitubercular regimen in Mtb-infected BALB/c mice (111). Given the widespread use of metformin and the high prevalence of MAC disease, retrospective analyses of the effect of metformin on MAC microbiological and clinical outcomes would be useful to gauge its promise as an adjunctive HDT for MAC.

Immunomodulation and Antimicrobial Properties: Clavanin-MO

Clavanin-MO is a naturally occurring antimicrobial peptide which possesses immunomodulatory properties (113). Both *in vitro* and *in vivo*, clavanin-MO stimulates production of inflammatory mediators, including IFN- γ , granulocyte-macrophage-stimulating factor, and monocyte chemoattractant protein-1, while suppressing the pro-inflammatory cytokines IL-12 and TNF- α (113). Clavanin-MO protects animal models from infection by both gram-positive and gram-negative bacteria (113). Although clavanin-MO has not been tested against mycobacteria, its immunomodulatory effects could potentially improve the immune response against MAC while blocking pathological inflammation, especially since it affects both IFN- γ and TNF- α , which are targets of other promising HDTs.

Potentiation of Macrophage Effector Function and Antimicrobial Activity: Thioridazine

Thioridazine is a neuroleptic drug, which has both direct antimycobacterial and host-directed effects (114, 115). The drug acts directly against Mtb by inhibiting antibiotic efflux pumps, thereby enhancing antibiotic susceptibility *in vitro* (116). Thioridazine also affects the host by inhibiting mammalian efflux pumps in the macrophage, leading to acidification of the phagosome and improving mycobacterial clearance (114, 117). Although its efficacy as an adjunctive therapy in murine models of chronic TB is controversial (118, 119), thioridazine was found to reduce the emergence of isoniazid-resistant mutants in Mtb-infected mouse lungs following co-administration with the standard anti-TB regimen (120). Thioridazine has been suggested as an adjunctive therapy for MAC, but research in this area has been limited (121–123). A short course of thioridazine and moxifloxacin was sufficient to clear MAC from infected monocytes (122). However, the pharmacokinetics of

thioridazine may prevent it from reaching effective concentrations in the lung, thus limiting its clinical utility in MAC pulmonary disease (121, 123).

HDTs WITH UNKNOWN OR POORLY UNDERSTOOD MECHANISMS OF ACTION

Poloxamer CRL-1072

Poloxamer CRL-1072 is a surfactant which makes mycobacteria more susceptible to some antibiotics, possibly through disruption of mycobacterial surface lipids (124). Its effects are especially pronounced in macrophages and mice compared to broth culture, suggesting that it has an effect on the host response to mycobacterial infection (124). The mechanisms of action of CRL-1072 are poorly understood. The surfactant induces production of nitric oxide in cultured human macrophages, leading to improved clearance of MAC (125). In addition, CRL-1072 induces production of IL-8 in human macrophages, a chemotactic factor which attracts neutrophils and T cells to the site of infection (126). To date, there has been little research on CRL-1072, and much remains unknown about its potential as an HDT. An important consideration is that, as a surfactant, CRL-1072 would likely have to be delivered topically to the lungs via inhalation. There is precedent for inhaled therapies for MAC with the recently FDA-approved Amikacin Liposome Inhalation Suspension (ALIS) (127).

Picolinic Acid

Picolinic acid is a degradation product of L-tryptophan with metal-chelating properties (128). An oral formulation, chromium(III) picolinate is safe and available as a nutritional supplement (129–131). Experimentally, it has both antimicrobial and host-directed effects against MAC. Specifically, picolinic acid potentiates the antimicrobial effects of clarithromycin, rifampicin, and some fluoroquinolones against both extracellular and intracellular MAC, suggesting that it has direct antimicrobial activity, which may be due to its iron-chelating properties (132). When used together with IFN- γ , picolinic acid also triggers apoptosis of MAC-infected mouse macrophages, thereby inhibiting intracellular mycobacterial growth (133, 134). Picolinic acid may also increase expression of TNF- α and interleukin-1, improving macrophage effector function (135). On the other hand, picolinic acid does not upregulate production of β -defensin-1, free fatty acids, or reactive oxygen and nitrogen intermediates (136). Therefore, its potentiation of macrophage effector functions remains poorly understood.

HDT TARGET PATHWAYS FOR FUTURE INVESTIGATION

HIF-1 α

Hypoxia-inducible factor-1 alpha (HIF-1 α) is a key regulator of cellular metabolism in hypoxic environments and is involved in the immune response, even under normoxic conditions (137).

HIF-1 α is thought to play an important role in immunity to mycobacterial infection. In zebrafish, stabilization of HIF-1 α protects against *M. marinum* infection (138). The protective effect is related to upregulation of IL-1 β in macrophages, which results in increased nitric oxide production by neutrophils (139). There is also evidence that HIF-1 α plays multiple roles in the macrophage response to Mtb infection by mediating IFN- γ -dependent genes, regulating immune effectors, shifting metabolism to aerobic glycolysis, and blocking excessive inflammation (140–142). In general, HIF-1 α promotes a pro-inflammatory state, which may improve mycobacterial clearance early in infection, but also induces pathological inflammation and immune exhaustion during chronic infection.

HIF-1 α has not been well-studied in the context of MAC infection. However, research on other mycobacteria suggests that HIF-1 α is a double-edged sword. Whereas induction of HIF-1 α promotes a pro-inflammatory state, which may improve mycobacterial clearance early during the course of infection, it can also lead to pathological inflammation and immune exhaustion during chronic infection (140, 143). Targeting the HIF-1 α pathway (and its timing) as an HDT strategy for MAC remains to be investigated.

Broadly Protective HDT Targets Against Intracellular Pathogens

A recent study screened FDA-approved drugs to identify HDT targets with broad protection against multiple intracellular pathogens (14). Three targets were identified which broadly protect THP-1 cells from intracellular bacteria: antagonizing G protein receptor (GPCR) signaling, interfering with intracellular calcium signaling, and disrupting membrane cholesterol distribution (14). Although mycobacteria have been shown to manipulate G-protein-coupled receptors to suppress epithelial signaling pathways (144) and to inhibit intracellular calcium signaling, leading to reduced phagosome-lysosome fusion and increased mycobacterial survival within human macrophages (145), these cellular pathways have not been directly targeted by therapies, and represent an area of potential future investigation.

CONCLUSIONS

Although HDTs represent a promising tool to improve MAC clinical outcomes, they have been the subject of little research to date. Looking to the future, there are several major challenges and opportunities in MAC HDT research which remain to be met. Two specific research needs are a better understanding of MAC pathophysiology to identify HDT targets, and improved model systems to allow investigation of potential HDTs.

An improved understanding of the host-pathogen interactions during MAC disease could reveal additional HDT targets. To date, the majority of HDTs against MAC fall into two general categories: improving immune effector function or modulating pathologic inflammation. The mechanism of several HDTs are not completely understood. A better mechanistic understanding of their function could improve our knowledge of MAC pathophysiology and identify new pathways to be targeted by

HDTs. For example, the efficacy of statins in improving TB clinical outcomes suggests that the metabolism of mycobacterial-infected cells may be a promising area of investigation (102).

A lack of *in vitro* and *in vivo* experimental models of MAC infection has been a major barrier to research. Current model systems are not standardized, and do not always yield replicable or clinically useful results (146). Cell cultures cannot entirely recapitulate a disease which involves long-term, complex interactions between multiple cell types, tissues and organs, while murine models of NTM differ from human disease in their immune responses and granuloma structure, and generally do not sustain chronic infection unless immune suppression is induced (147). These deficiencies are especially important for investigating HDTs, which may target complex or human-specific pathways. Recent advances in model systems will inform future HDT research. *In silico* models could identify promising HDTs prior to the expense and difficulty of *in vitro* and *in vivo* experimentation. Recent developments in organoid models promise to allow better *in vitro* investigation of complex pathways involving interactions between multiple cell types and the extracellular matrix. For example, a three-dimensional granuloma model has recently been developed for Mtb and could be a valuable tool for investigating HDTs if adapted for MAC (148).

Finally, there is an unexplored need to investigate the use of HDTs in combination. To date, most studies have examined a

particular HDT in isolation or in combination with antibiotics. Investigation of HDTs with potentially complementary mechanisms could identify therapeutic combinations that have a greater effect than the sum of their parts.

MAC is an emerging infectious disease of particular concern due to its rising prevalence, resistance to frontline antibiotics, and associated chronic morbidity and mortality (1, 5, 10). HDTs against MAC represent a promising but underexplored avenue of research, which could hold great potential in improving microbiological and clinical outcomes.

AUTHOR CONTRIBUTIONS

NC and PK conceived the work. NC, SA, and PK wrote the manuscript. All authors contributed to the article and approved the submitted version.

FUNDING

This work was supported by NIH/NIAID grants UH3AI122309 and K24AI143447 to PK. The funders have no role in the content of this manuscript.

REFERENCES

1. Brode SK, Daley CL, Marras TK. The epidemiologic relationship between tuberculosis and non-tuberculous mycobacterial disease: a systematic review. *Int J Tuberc Lung Dis* (2014) 18:1370–7. doi: 10.5588/ijtld.14.0120
2. Lande L, George J, Plush T. Mycobacterium avium complex pulmonary disease: new epidemiology and management concepts. *Curr Opin Infect Dis* (2018) 31:199–207. doi: 10.1097/QCO.0000000000000437
3. Karakousis PC, Moore RD, Chaisson RE. Mycobacterium avium complex in patients with HIV infection in the era of highly active antiretroviral therapy. *Lancet Infect Dis* (2004) 4:557–65. doi: 10.1016/S1473-3099(04)01130-2
4. Chen C, Chen H, Chou C, Huang C, Lai C, Hsueh P. Pulmonary infection caused by nontuberculous mycobacteria in a medical center in Taiwan, 2005–2008. *Diagn Microbiol Infect Dis* (2012) 72:47–51. doi: 10.1016/j.diagmicrobio.2011.09.009
5. Diel R, Lipman M, Hoefsloot W. High mortality in patients with Mycobacterium avium complex lung disease: a systematic review. *BMC Infect Dis* (2018) 18:206–x. doi: 10.1186/s12879-018-3113-x
6. Gomes MS, Paul S, Moreira AL, Appelberg R, Rabinovitch M, Kaplan G. Survival of Mycobacterium avium and Mycobacterium tuberculosis in acidified vacuoles of murine macrophages. *Infect Immun* (1999) 67:3199–206. doi: 10.1128/IAI.67.7.3199-3206.1999
7. Early J, Fischer K, Bermudez LE. Mycobacterium avium uses apoptotic macrophages as tools for spreading. *Microb Pathog* (2011) 50:132–9. doi: 10.1016/j.micpath.2010.12.004
8. Rocco JM, Irani VR. Mycobacterium avium and modulation of the host macrophage immune mechanisms. *Int J Tuberc Lung Dis* (2011) 15:447–52. doi: 10.5588/ijtld.09.0695
9. Daley CL, Iaccarino JM, Lange C, Cambau E, Wallace RJ, Andrejak C, et al. Treatment of Nontuberculous Mycobacterial Pulmonary Disease: An Official ATS/ERS/ESCMID/IDSA Clinical Practice Guideline: Executive Summary. *Clin Infect Dis* (2020) 71:e1–e36. doi: 10.1093/cid/ciaa241
10. Griffith DE, Aksamit T, Brown-Elliott BA, Catanzaro A, Daley C, Gordin F, et al. An Official ATS/IDSA Statement: Diagnosis, Treatment, and Prevention of Nontuberculous Mycobacterial Diseases. *Am J Respir Crit Care Med* (2007) 175:367–416. doi: 10.1164/rccm.200604-571ST
11. Palucci I, Delogu G. Host Directed Therapies for Tuberculosis: Futures Strategies for an Ancient Disease. *Chemotherapy* (2018) 63:172–80. doi: 10.1159/000490478
12. Zumla A, Rao M, Wallis RS, Kaufmann SH, Rustomjee R, Mwaba P, et al. Host-directed therapies for infectious diseases: current status, recent progress, and future prospects. *Lancet Infect Dis* (2016) 16:47. doi: 10.1016/S1473-3099(16)00078-5
13. Kolloli A, Subbian S. Host-Directed Therapeutic Strategies for Tuberculosis. *Front Med* (2017) 4:171. doi: 10.3389/fmed.2017.00171
14. Czyż DM, Podluri L, Jain-Gupta N, Riley SP, Martinez JJ, Steck TL, et al. Host-Directed Antimicrobial Drugs with Broad-Spectrum Efficacy against Intracellular Bacterial Pathogens. *mBio* (2014) 5:e01534–14. doi: 10.1128/mBio.01534-14
15. Frank DJ, Horne DJ, Dutta NK, Shaku MT, Madensein R, Hawn TR, et al. Remembering the Host in Tuberculosis Drug Development. *J Infect Dis* (2019) 219:1518–24. doi: 10.1093/infdis/jiy712
16. Bento CM, Gomes MS, Silva T. Looking beyond Typical Treatments for Atypical Mycobacteria. *Antibiot (Basel)* (2020) 9:18. doi: 10.3390/antibiotics9010018
17. Mizushima N. Autophagy: process and function. *Genes Dev* (2007) 21:2861–73. doi: 10.1101/gad.1599207
18. Boya P, Reggiori F, Codogno P. Emerging regulation and functions of autophagy. *Nat Cell Biol* (2013) 15:713–20. doi: 10.1038/ncb2788
19. Ohsumi Y. Historical landmarks of autophagy research. *Cell Res* (2014) 24:9–23. doi: 10.1038/cr.2013.169
20. Kim J, Guan K. mTOR as a central hub of nutrient signalling and cell growth. *Nat Cell Biol* (2019) 21:63–71. doi: 10.1038/s41556-018-0205-1
21. Saxton RA, Sabatini DM. mTOR Signaling in Growth, Metabolism, and Disease. *Cell* (2017) 168:960–76. doi: 10.1016/j.cell.2017.02.004
22. Zheng Y, Jiang Y. mTOR Inhibitors at a Glance. *Mol Cell Pharmacol* (2015) 7:15–20.
23. Lisse TS, Hewison M. Vitamin D. *Cell Cycle (Georgetown Tex)* (2011) 10:1888–9. doi: 10.4161/cc.10.12.15620

24. Delgado M, Singh S, De Haro S, Master S, Ponpuak M, Dinkins C, et al. Autophagy and Pattern Recognition Receptors in Innate Immunity. *Immunol Rev* (2009) 227:189–202. doi: 10.1111/j.1600-065X.2008.00725.x
25. Oh JE, Lee HK. Pattern Recognition Receptors and Autophagy. *Front Immunol* (2014) 5:300. doi: 10.3389/fimmu.2014.00300
26. Frehel C, de Chastellier C, Lang T, Rastogi N. Evidence for inhibition of fusion of lysosomal and prelysosomal compartments with phagosomes in macrophages infected with pathogenic *Mycobacterium avium*. *Infect Immun* (1986) 52:252–62. doi: 10.1128/IAI.52.1.252-262.1986
27. Crowle AJ, Dahl R, Ross E, May MH. Evidence that vesicles containing living, virulent *Mycobacterium tuberculosis* or *Mycobacterium avium* in cultured human macrophages are not acidic. *Infect Immun* (1991) 59:1823–31. doi: 10.1128/IAI.59.5.1823-1831.1991
28. Silva T, Moreira AC, Nazmi K, Moniz T, Vale N, Rangel M, et al. Lactoferricin Peptides Increase Macrophages' Capacity To Kill *Mycobacterium avium*. *mSphere* (2017) 2:301. doi: 10.1128/msphere.00301-17
29. Singh P, Subbian S. Harnessing the mTOR Pathway for Tuberculosis Treatment. *Front Microbiol* (2018) 9:70. doi: 10.3389/fmicb.2018.00070
30. Andersson A, Andersson B, Lorell C, Raffetseder J, Larsson M, Blomgran R. Autophagy induction targeting mTORC1 enhances *Mycobacterium tuberculosis* replication in HIV co-infected human macrophages. *Sci Rep* (2016) 6:28171. doi: 10.1038/srep28171
31. Floto RA, Sarkar S, Perlstein EO, Kampmann B, Schreiber SL, Rubinsztein DC. Small Molecule Enhancers of Rapamycin-Induced TOR Inhibition Promote Autophagy, Reduce Toxicity in Huntington's Disease Models and Enhance Killing of *Mycobacteria* by Macrophages. *Autophagy* (2007) 3:620–2. doi: 10.4161/auto.4898
32. Hawn TR, Matheson AI, Maley SN, Vandal O. Host-Directed Therapeutics for Tuberculosis: Can We Harness the Host? *Microbiol Mol Biol Rev* (2013) 77:608–27. doi: 10.1128/mmr.00032-13
33. Hu J, Zhang W, Zuo W, Tan H, Bai W. Inhibition of the PD-1/PD-L1 signaling pathway enhances innate immune response of alveolar macrophages to *mycobacterium tuberculosis* in mice. *Pulm Pharmacol Ther* (2020) 60:101842. doi: 10.1016/j.pupt.2019.101842
34. Okazaki T, Chikuma S, Iwai Y, Fagarasan S, Honjo T. A rheostat for immune responses: the unique properties of PD-1 and their advantages for clinical application. *Nat Immunol* (2013) 14:1212–8. doi: 10.1038/ni.2762
35. Hartley GP, Chow L, Ammons DT, Wheat WH, Dow SW. Programmed Cell Death Ligand 1 (PD-L1) Signaling Regulates Macrophage Proliferation and Activation. *Cancer Immunol Res* (2018) 6:1260–73. doi: 10.1158/2326-6066.CIR-17-0537
36. Xiong H, Mittman S, Rodriguez R, Moskalenko M, Pacheco-Sanchez P, Yang Y, et al. Anti-PD-L1 Treatment Results in Functional Remodeling of the Macrophage Compartment. *Cancer Res* (2019) 79:1493–506. doi: 10.1158/0008-5472.CAN-18-3208
37. Zhang Y, Du W, Chen Z, Xiang C. Upregulation of PD-L1 by SPP1 mediates macrophage polarization and facilitates immune escape in lung adenocarcinoma. *Exp Cell Res* (2017) 359:449–57. doi: 10.1016/j.yexcr.2017.08.028
38. Bekker LG, Freeman S, Murray PJ, Ryffel B, Kaplan G. TNF- α controls intracellular mycobacterial growth by both inducible nitric oxide synthase-dependent and inducible nitric oxide synthase-independent pathways. *J Immunol* (2001) 166:6728–34. doi: 10.4049/jimmunol.166.11.6728
39. Shu C, Pan S, Feng J, Wang J, Chan Y, Yu C, et al. The Clinical Significance of Programmed Death-1, Regulatory T Cells and Myeloid Derived Suppressor Cells in Patients with Nontuberculous *Mycobacteria*-Lung Disease. *J Clin Med* (2019) 8:736. doi: 10.3390/jcm8050736
40. Shu C, Wang J, Wu M, Wu C, Lai H, Lee L, et al. Attenuation of lymphocyte immune responses during *Mycobacterium avium* complex-induced lung disease due to increasing expression of programmed death-1 on lymphocytes. *Sci Rep* (2017) 7:42004. doi: 10.1038/srep42004
41. Tousif S, Singh Y, Prasad DVR, Sharma P, Kaer LV, Das G. T Cells from Programmed Death-1 Deficient Mice Respond Poorly to *Mycobacterium tuberculosis* Infection. *PLoS One* (2011) 6:19864. doi: 10.1371/journal.pone.0019864
42. Tezera LB, Bielecka MK, Ogongo P, Walker NF, Ellis M, Garay-Baquero DJ, et al. Anti-PD-1 immunotherapy leads to tuberculosis reactivation via dysregulation of TNF- α . *Elife* (2020) 9:52668. doi: 10.7554/eLife.52668
43. Scharn CR, Collins AC, Nair VR, Stamm CE, Marciano DK, Graviss EA, et al. Heme Oxygenase-1 Regulates Inflammation and *Mycobacterium tuberculosis* Survival in Human Macrophages during *Mycobacterium tuberculosis* Infection. *J Immunol* (2016) 196:4641–9. doi: 10.4049/jimmunol.1500434
44. Grochot-Przeczek A, Dulak J, Jozkowicz A. Haem oxygenase-1: non-canonical roles in physiology and pathology. *Clin Sci (Lond)* (2012) 122:93–103. doi: 10.1042/CS20110147
45. Costa DL, Namasivayam S, Amaral EP, Arora K, Chao A, Mittereder LR, et al. Pharmacological Inhibition of Host Heme Oxygenase-1 Suppresses *Mycobacterium tuberculosis* Infection In Vivo by a Mechanism Dependent on T Lymphocytes. *mBio* (2016) 7:1675. doi: 10.1128/mbio.01675-16
46. Rockwood N, Costa DL, Amaral EP, Du Bruyn E, Kubler A, Gil-Santana L, et al. *Mycobacterium tuberculosis* Induction of Heme Oxygenase-1 Expression Is Dependent on Oxidative Stress and Reflects Treatment Outcomes. *Front Immunol* (2017) 8:542. doi: 10.3389/fimmu.2017.00542
47. Silva-Gomes S, Appelberg R, Larsen R, Soares MP, Gomes MS. Heme Catabolism by Heme Oxygenase-1 Confers Host Resistance to *Mycobacterium tuberculosis*. *Infection Immun* (2013) 81:2536–45. doi: 10.1128/iai.00251-13
48. Saunders BM, Cooper AM. Restraining mycobacteria: Role of granulomas in mycobacterial infections. *Immunol Cell Biol* (2000) 78:334–41. doi: 10.1046/j.1440-1711.2000.00933.x
49. Reljic R. IFN- γ therapy of tuberculosis and related infections. *J Interferon Cytokine Res* (2007) 27:353–64. doi: 10.1089/jir.2006.0103
50. Kamijo R, Harada H, Matsuyama T, Bosland M, Gerecitano J, Shapiro D, et al. Requirement for transcription factor IRF-1 in NO synthase induction in macrophages. *Science* (1994) 263:1612–5. doi: 10.1126/science.7510419
51. Cassatella MA, Bazzoni F, Flynn RM, Dusi S, Trinchieri G, Rossi F. Molecular basis of interferon- γ and lipopolysaccharide enhancement of phagocyte respiratory burst capability. Studies on the gene expression of several NADPH oxidase components. *J Biol Chem* (1990) 265:20241–6.
52. Huang S, Hendriks W, Althage A, Hemmi S, Bluethmann H, Kamijo R, et al. Immune response in mice that lack the interferon- γ receptor. *Science* (1993) 259:1742–5. doi: 10.1126/science.8456301
53. Hsu N, Young LS, Bermudez LE. Response to stimulation with recombinant cytokines and synthesis of cytokines by murine intestinal macrophages infected with the *Mycobacterium avium* complex. *Infect Immun* (1995) 63:528–33. doi: 10.1128/IAI.63.2.528-533.1995
54. Gomez-Flores R, Tucker SD, Kansal R, Tamez-Guerra R, Mehta RT. Enhancement of antibacterial activity of clofazimine against *Mycobacterium avium*-*Mycobacterium intracellulare* complex infection induced by IFN- γ is mediated by TNF- α . *J Antimicrob Chemother* (1997) 39:189–97. doi: 10.1093/jac/39.2.189
55. Squires KE, Murphy WF, Madoff LC, Murray HW. Interferon- γ and *Mycobacterium avium*-intracellular infection. *J Infect Dis* (1989) 159:599–600. doi: 10.1093/infdis/159.3.599
56. Wu U, Holland SM. Host susceptibility to non-tuberculous mycobacterial infections. *Lancet Infect Dis* (2015) 15:968–80. doi: 10.1016/S1473-3099(15)00089-4
57. Glosli H, Stray-Pedersen A, Brun AC, Holtmon LW, Tønjum T, Chappier A, et al. Infections due to various atypical mycobacteria in a Norwegian multiplex family with dominant interferon- γ receptor deficiency. *Clin Infect Dis* (2008) 46:23. doi: 10.1086/525855
58. Jouanguy E, Altare F, Lamhamedi S, Revy P, Emile JF, Newport M, et al. Interferon- γ -receptor deficiency in an infant with fatal bacille Calmette-Guérin infection. *N Engl J Med* (1996) 335:1956–61. doi: 10.1056/NEJM199612263352604
59. Bax HI, Freeman AF, Ding L, Hsu AP, Marciano B, Kristosturyan E, et al. Interferon alpha treatment of patients with impaired interferon gamma signaling. *J Clin Immunol* (2013) 33:991–1001. doi: 10.1007/s10875-013-9882-5
60. Holland SM, Eisenstein EM, Kuhns DB, Turner ML, Fleisher TA, Strober W, et al. Treatment of refractory disseminated nontuberculous mycobacterial infection with interferon gamma. A preliminary report. *N Engl J Med* (1994) 330:1348–55. doi: 10.1056/NEJM199405123301904
61. Smaldone GC. Repurposing of gamma interferon via inhalation delivery. *Adv Drug Deliv Rev* (2018) 133:87–92. doi: 10.1016/j.addr.2018.06.004
62. Appelberg R. Protective Role of Interferon Gamma, Tumor Necrosis Factor Alpha and Interleukin-6 in *Mycobacterium tuberculosis* and *M. avium*

- Infections. *Immunobiology* (1994) 191:520–5. doi: 10.1016/S0171-2985(11)80458-4
63. Dorhoi A, Kaufmann SHE. Tumor necrosis factor alpha in mycobacterial infection. *Semin Immunol* (2014) 26:203–9. doi: 10.1016/j.smim.2014.04.003
 64. Kindler V, Sappino A, Grau GE, Piguat P, Vassalli P. The inducing role of tumor necrosis factor in the development of bactericidal granulomas during BCG infection. *Cell* (1989) 56:731–40. doi: 10.1016/0092-8674(89)90676-4
 65. Kaneko H, Yamada H, Mizuno S, Udagawa T, Kazumi Y, Sekikawa K, et al. Role of tumor necrosis factor-alpha in Mycobacterium-induced granuloma formation in tumor necrosis factor-alpha-deficient mice. *Lab Invest* (1999) 79:379–86.
 66. Harris J, Hope JC, Keane J. Tumor Necrosis Factor Blockers Influence Macrophage Responses to Mycobacterium tuberculosis. *J Infect Dis* (2008) 198:1842–50. doi: 10.1086/593174
 67. Blaser H, Dostert C, Mak TW, Brenner D. TNF and ROS Crosstalk in Inflammation. *Trends Cell Biol* (2016) 26:249–61. doi: 10.1016/j.tcb.2015.12.002
 68. Flynn JL, Goldstein MM, Chan J, Triebold KJ, Pfeffer K, Lowenstein CJ, et al. Tumor necrosis factor-alpha is required in the protective immune response against Mycobacterium tuberculosis in mice. *Immunity* (1995) 2:561–72. doi: 10.1016/1074-7613(95)90001-2
 69. Keane J, Balcewicz-Sablinska MK, Remold HG, Chupp GL, Meek BB, Fenton MJ, et al. Infection by Mycobacterium tuberculosis promotes human alveolar macrophage apoptosis. *Infect Immun* (1997) 65:298–304. doi: 10.1128/IAI.65.1.298-304.1997
 70. Vassalli P. The Pathophysiology of Tumor Necrosis Factors. *Annu Rev Immunol* (1992) 10:411–52. doi: 10.1146/annurev.iy.10.040192.002211
 71. Kundu M, Pathak SK, Kumawat K, Basu S, Chatterjee G, Pathak S, et al. A TNF- and c-Cbl-dependent FLIP(S)-degradation pathway and its function in Mycobacterium tuberculosis-induced macrophage apoptosis. *Nat Immunol* (2009) 10:918–26. doi: 10.1038/ni.1754
 72. Keane J, Remold HG, Kornfeld H. Virulent Mycobacterium tuberculosis strains evade apoptosis of infected alveolar macrophages. *J Immunol* (2000) 164:2016–20. doi: 10.4049/jimmunol.164.4.2016
 73. Declercq W, Vanden Berghe T, Vandenabeele P. RIP kinases at the crossroads of cell death and survival. *Cell* (2009) 138:229–32. doi: 10.1016/j.cell.2009.07.006
 74. Skerry C, Harper J, Klunk M, Bishai WR, Jain SK. Adjunctive TNF inhibition with standard treatment enhances bacterial clearance in a murine model of necrotic TB granulomas. *PLoS One* (2012) 7:e39680. doi: 10.1371/journal.pone.0039680
 75. Winthrop KL, Chang E, Yamashita S, Iademarco MF, LoBue PA. Nontuberculous Mycobacteria Infections and Anti-Tumor Necrosis Factor- α Therapy. *Emerging Infect Dis* (2009) 15:1556–61. doi: 10.3201/eid1510.090310
 76. Yoo J, Jo K, Kang B, Kim MY, Yoo B, Lee C, et al. Mycobacterial diseases developed during anti-tumor necrosis factor- α therapy. *Eur Respir J* (2014) 44:1289–95. doi: 10.1183/09031936.00063514
 77. Wallis RS. Tumour necrosis factor antagonists: structure, function, and tuberculosis risks. *Lancet Infect Dis* (2008) 8:601–11. doi: 10.1016/S1473-3099(08)70227-5
 78. Wallis RS, van Vuuren C, Potgieter S. Adalimumab treatment of life-threatening tuberculosis. *Clin Infect Dis* (2009) 48:1429–32. doi: 10.1086/598504
 79. Arend SM, Leyten EMS, Franken WPJ, Huisman EM, van Dissel JT. A patient with de novo tuberculosis during anti-tumor necrosis factor-alpha therapy illustrating diagnostic pitfalls and paradoxical response to treatment. *Clin Infect Dis* (2007) 45:1470–5. doi: 10.1086/522993
 80. Garcia Vidal C, Rodríguez Fernández S, Martínez Lacasa J, Salavert M, Vidal R, Rodríguez Carballeira M, et al. Paradoxical response to antituberculous therapy in infliximab-treated patients with disseminated tuberculosis. *Clin Infect Dis* (2005) 40:756–9. doi: 10.1086/427941
 81. Yew WW. Clinically significant interactions with drugs used in the treatment of tuberculosis. *Drug Saf* (2002) 25:111–33. doi: 10.2165/00002018-200225020-00005
 82. Ehlers S. Immunity to tuberculosis: a delicate balance between protection and pathology. *FEMS Immunol Med Microbiol* (1999) 23:149–58. doi: 10.1111/j.1574-695X.1999.tb01234.x
 83. Kroesen VM, Gröschel MI, Martinson N, Zumla A, Maeurer M, der Werf v, et al. Non-Steroidal Anti-inflammatory Drugs As Host-Directed Therapy for Tuberculosis: A Systematic Review. *Front Immunol* (2017) 8:772. doi: 10.3389/fimmu.2017.00772
 84. Vilaplana C, Marzo E, Tapia G, Diaz J, Garcia V, Cardona P. Ibuprofen therapy resulted in significantly decreased tissue bacillary loads and increased survival in a new murine experimental model of active tuberculosis. *J Infect Dis* (2013) 208:199–202. doi: 10.1093/infdis/jit152
 85. Marzo E, Vilaplana C, Tapia G, Diaz J, Garcia V, Cardona P. Damaging role of neutrophilic infiltration in a mouse model of progressive tuberculosis. *Tuberc (Edinb)* (2014) 94:55–64. doi: 10.1016/j.tube.2013.09.004
 86. Ivanyi J, Zumla A. Nonsteroidal antiinflammatory drugs for adjunctive tuberculosis treatment. *J Infect Dis* (2013) 208:185–8. doi: 10.1093/infdis/jit153
 87. Schafer AI. Effects of nonsteroidal antiinflammatory drugs on platelet function and systemic hemostasis. *J Clin Pharmacol* (1995) 35:209–19. doi: 10.1002/j.1552-4604.1995.tb04050.x
 88. Schoeman J, Mansvelt E, Springer P, van Rensburg AJ, Carlini S, Fourie E. Coagulant and fibrinolytic status in tuberculous meningitis. *Pediatr Infect Dis J* (2007) 26:428–31. doi: 10.1097/01.inf.0000261126.60283.cf
 89. Sano C, Shimizu T, Sato K, Kawachi H, Kawahara S, Tomioka H. Therapeutic Effects of Benzoxazinorifamycin KRM-1648 Administered Alone or in Combination with a Half-Sized Secretory Leucocyte Protease Inhibitor or the Nonsteroidal Anti-Inflammatory Drug Diclofenac Sodium against Mycobacterium avium Complex Infection in Mice. *Antimicrob Agents Chemother* (1999) 43:360–4. doi: 10.1128/AAC.43.2.360
 90. Yang D, Chang W, Cheng M, Lai J, Chang D, Chen C. Peripheral arthritis caused by Mycobacterium avium-intracellulare in a patient with ankylosing spondylitis. *J Clin Rheumatol* (2009) 15:323–4. doi: 10.1097/RHU.0b013e3181bbcbfb
 91. Dolecek R. [Certain steroids in the treatment of pulmonary tuberculosis]. *Cas Lek Cesk* (1951) 90:1160–3.
 92. Critchley JA, Young F, Orton L, Garner P. Corticosteroids for prevention of mortality in people with tuberculosis: a systematic review and meta-analysis. *Lancet Infect Dis* (2013) 13:223–37. doi: 10.1016/S1473-3099(12)70321-3
 93. Prasad K, Singh MB, Ryan H. Corticosteroids for managing tuberculous meningitis. *Cochrane Database Syst Rev* (2016) 2016:2244. doi: 10.1002/14651858.CD002244.pub4
 94. Schutz C, Davis AG, Sossen B, Lai RP, Ntsekhe M, Harley YX, et al. Corticosteroids as an adjunct to tuberculosis therapy. *Expert Rev Respir Med* (2018) 12:881–91. doi: 10.1080/17476348.2018.1515628
 95. Goetz MB. Are corticosteroids useful adjunctive agents in the treatment of disseminated Mycobacterium avium complex infection associated with human immunodeficiency virus infection? *Clin Infect Dis* (1998) 26:687–8. doi: 10.1086/514598
 96. Dorman SE, Heller HM, Basgoz NO, Sax PE. Adjunctive corticosteroid therapy for patients whose treatment for disseminated Mycobacterium avium complex infection has failed. *Clin Infect Dis* (1998) 26:682–6. doi: 10.1086/514597
 97. Kobashi Y, Matsushima T. Clinical analysis of pulmonary Mycobacterium avium complex disease in association with corticosteroid treatment. *J Infect Chemother* (2003) 9:68–74. doi: 10.1007/s10156-002-0216-4
 98. Yang C. Advancing host-directed therapy for tuberculosis. *Microb Cell* (2017) 4:105–7. doi: 10.15698/mic2017.03.565
 99. Parihar SP, Guler R, Khutlang R, Lang DM, Hurdal R, Mhlanga MM, et al. Statin therapy reduces the mycobacterium tuberculosis burden in human macrophages and in mice by enhancing autophagy and phagosome maturation. *J Infect Dis* (2014) 209:754–63. doi: 10.1093/infdis/jit550
 100. Skerry C, Pinn ML, Bruiners N, Pine R, Gennaro ML, Karakousis PC. Simvastatin increases the in vivo activity of the first-line tuberculosis regimen. *J Antimicrob Chemother* (2014) 69:2453–7. doi: 10.1093/jac/dku166
 101. Dutta NK, Bruiners N, Pinn ML, Zimmerman MD, Prideaux B, Dartois V, et al. Statin adjunctive therapy shortens the duration of TB treatment in mice. *J Antimicrob Chemother* (2016) 71:1570–7. doi: 10.1093/jac/dkw014
 102. Dutta NK, Bruiners N, Zimmerman MD, Tan S, Dartois V, Gennaro ML, et al. Adjunctive Host-Directed Therapy With Statins Improves Tuberculosis-Related Outcomes in Mice. *J Infect Dis* (2020) 221:1079–87. doi: 10.1093/infdis/jiz517

103. Peyron P, Vaubourgeix J, Poquet Y, Levillain F, Botanch C, Bardou F, et al. Foamy Macrophages from Tuberculous Patients' Granulomas Constitute a Nutrient-Rich Reservoir for *M. tuberculosis* Persistence. *PLoS Pathog* (2008) 4:204. doi: 10.1371/journal.ppat.1000204
104. Bruiners N, Dutta N, Guerrini V, Salamon H, Yamaguchi KD, Karakousis PC, et al. The anti-tubercular activity of simvastatin is mediated by cholesterol-driven autophagy via the AMPK-mTORC1-TFEB axis. *J Lipid Res* (2020) 61:1617–28. doi: 10.1194/jlr.RA120000895
105. Caire-Brändli I, Papadopoulos A, Malaga W, Marais D, Cnaan S, Thilo L, et al. Reversible Lipid Accumulation and Associated Division Arrest of *Mycobacterium avium* in Lipoprotein-Induced Foamy Macrophages May Resemble Key Events during Latency and Reactivation of Tuberculosis. *Infection Immun* (2014) 82:476–90. doi: 10.1128/iai.01196-13
106. Zhang M, He J. Impacts of metformin on tuberculosis incidence and clinical outcomes in patients with diabetes: a systematic review and meta-analysis. *Eur J Clin Pharmacol* (2020) 76:149–59. doi: 10.1007/s00228-019-02786-y
107. Degner NR, Wang J, Golub JE, Karakousis PC. Metformin Use Reverses the Increased Mortality Associated With Diabetes Mellitus During Tuberculosis Treatment. *Clin Infect Dis* (2018) 66:198–205. doi: 10.1093/cid/cix819
108. Lachmandas E, Eckold C, Böhme J, Koeken, Valerie ACM, Marzuki MB, et al. Metformin Alters Human Host Responses to *Mycobacterium tuberculosis* in Healthy Subjects. *J Infect Dis* (2019) 220:139–50. doi: 10.1093/infdis/jiz064
109. Yew WW, Chang KC, Chan DP, Zhang Y. Metformin as a host-directed therapeutic in tuberculosis: Is there a promise? *Tubercul (Edinb)* (2019) 115:76–80. doi: 10.1016/j.tube.2019.02.004
110. Arai M, Uchiba M, Komura H, Mizuochi Y, Harada N, Okajima K. Metformin, an antidiabetic agent, suppresses the production of tumor necrosis factor and tissue factor by inhibiting early growth response factor-1 expression in human monocytes in vitro. *J Pharmacol Exp Ther* (2010) 334:206–13. doi: 10.1124/jpet.109.164970
111. Dutta NK, Pinn ML, Karakousis PC. Metformin Adjunctive Therapy Does Not Improve the Sterilizing Activity of the First-Line Antitubercular Regimen in Mice. *Antimicrob Agents Chemother* (2017) 61:652. doi: 10.1128/aac.00652-17
112. Singhal A, Jie L, Kumar P, Hong GS, Leow MK, Paleja B, et al. Metformin as adjunct antituberculosis therapy. *Sci Transl Med* (2014) 6:263ra159. doi: 10.1126/scitranslmed.3009885
113. Silva ON, de la Fuente-Núñez C, Haney EF, Fensterseifer ICM, Ribeiro SM, Porto WF, et al. An anti-infective synthetic peptide with dual antimicrobial and immunomodulatory activities. *Sci Rep* (2016) 6:35465. doi: 10.1038/srep35465
114. Amaral L, Viveiros M. Thioridazine: A Non-Antibiotic Drug Highly Effective, in Combination with First Line Anti-Tuberculosis Drugs, against Any Form of Antibiotic Resistance of *Mycobacterium tuberculosis* Due to Its Multi-Mechanisms of Action. *Antibiot (Basel)* (2017) 6:3. doi: 10.3390/antibiotics6010003
115. Dutta NK, Karakousis PC. Thioridazine for treatment of tuberculosis: promises and pitfalls. *Tubercul (Edinb)* (2014) 94:708–11. doi: 10.1016/j.tube.2014.09.001
116. Machado D, Couto I, Perdigão J, Rodrigues L, Portugal I, Baptista P, et al. Contribution of Efflux to the Emergence of Isoniazid and Multidrug Resistance in *Mycobacterium tuberculosis*. *PLoS One* (2012) 7:e34538. doi: 10.1371/journal.pone.0034538
117. Machado D, Pires D, Perdigão J, Couto I, Portugal I, Martins M, et al. Ion Channel Blockers as Antimicrobial Agents, Efflux Inhibitors, and Enhancers of Macrophage Killing Activity against Drug Resistant *Mycobacterium tuberculosis*. *PLoS One* (2016) 11:e0149326. doi: 10.1371/journal.pone.0149326
118. Dutta NK, Pinn ML, Karakousis PC. Sterilizing activity of thioridazine in combination with the first-line regimen against acute murine tuberculosis. *Antimicrob Agents Chemother* (2014) 58:5567–9. doi: 10.1128/AAC.03408-14
119. Dutta NK, Pinn ML, Zhao M, Rudek MA, Karakousis PC. Thioridazine lacks bactericidal activity in an animal model of extracellular tuberculosis. *J Antimicrob Chemother* (2013) 68:1327–30. doi: 10.1093/jac/dkt037
120. Dutta NK, Pinn ML, Karakousis PC. Reduced emergence of isoniazid resistance with concurrent use of thioridazine against acute murine tuberculosis. *Antimicrob Agents Chemother* (2014) 58:4048–53. doi: 10.1128/AAC.02981-14
121. Ruth MM, Pennings LJ, Koeken, Valerie ACM, Schildkraut JA, Hashemi A, et al. Thioridazine Is an Efflux Pump Inhibitor in *Mycobacterium avium* Complex but of Limited Clinical Relevance. *Antimicrob Agents Chemother* (2020) 64:181. doi: 10.1128/AAC.00181-20
122. Srivastava S, Deshpande D, Sherman CM, Gumbo T. A 'shock and awe' thioridazine and moxifloxacin combination-based regimen for pulmonary *Mycobacterium avium*-intracellular complex disease. *J Antimicrob Chemother* (2017) 72:i43–7. doi: 10.1093/jac/dkx308
123. Deshpande D, Srivastava S, Musuka S, Gumbo T. Thioridazine as Chemotherapy for *Mycobacterium avium* Complex Diseases. *Antimicrob Agents Chemother* (2016) 60:4652–8. doi: 10.1128/AAC.02985-15
124. Jagannath C, Emanuele MR, Hunter RL. Activities of Poloxamer CRL-1072 against *Mycobacterium avium* in Macrophage Culture and in Mice. *Antimicrob Agents Chemother* (1999) 43:2898–903. doi: 10.1128/AAC.43.12.2898
125. Jagannath C, Sepulveda E, Actor JK, Luxem F, Emanuele MR, Hunter RL. Effect of poloxamer CRL-1072 on drug uptake and nitric-oxide-mediated killing of *Mycobacterium avium* by macrophages. *Immunopharmacology* (2000) 48:185–97. doi: 10.1016/s0162-3109(00)00203-4
126. Jagannath C, Pai S, Actor JK, Hunter RL. CRL-1072 enhances antimycobacterial activity of human macrophages through interleukin-8. *J Interferon Cytokine Res* (1999) 19:67–76. doi: 10.1089/107999099314432
127. Daley CL, Olivier KN. ALIS (Amikacin Liposome Inhalation Suspension): The Beginning of a Wonderland? *Am J Respir Crit Care Med* (2018) 198:1473–5. doi: 10.1164/rccm.201810-1901ED
128. Evans GW, Johnson PE. Characterization and quantitation of a zinc-binding ligand in human milk. *Pediatr Res* (1980) 14:876–80. doi: 10.1203/00006450-198007000-00007
129. Ganguly R, Sahu S, Ohanyan V, Haney R, Chavez RJ, Shah S, et al. Oral chromium picolinate impedes hyperglycemia-induced atherosclerosis and inhibits proatherogenic protein TSP-1 expression in STZ-induced type 1 diabetic ApoE^{-/-} mice. *Sci Rep* (2017) 7:45279. doi: 10.1038/srep45279
130. Vincent JB. Chromium: celebrating 50 years as an essential element? *Dalton Trans* (2010) 39:3787–94. doi: 10.1039/b920480f
131. Stout MD, Nyska A, Collins BJ, Witt KL, Kissling GE, Malarkey DE, et al. Chronic Toxicity and Carcinogenicity Studies of Chromium Picolinate Monohydrate Administered in Feed to F344/N Rats and B6C3F1 Mice for 2 Years. *Food Chem Toxicol* (2009) 47:729–33. doi: 10.1016/j.fct.2009.01.006
132. Cai S, Sato K, Shimizu T, Yamabe S, Hiraki M, Sano C, et al. Antimicrobial activity of picolinic acid against extracellular and intracellular *Mycobacterium avium* complex and its combined activity with clarithromycin, rifampicin and fluoroquinolones. *J Antimicrob Chemother* (2006) 57:85–93. doi: 10.1093/jac/dki418
133. Pais TF, Appelberg R. Induction of *Mycobacterium avium* growth restriction and inhibition of phagosome-endosome interactions during macrophage activation and apoptosis induction by picolinic acid plus IFN γ . *Microbiol (Reading)* (2004) 150:1507–18. doi: 10.1099/mic.0.26815-0
134. Pais TF, Appelberg R. Macrophage control of mycobacterial growth induced by picolinic acid is dependent on host cell apoptosis. *J Immunol* (2000) 164:389–97. doi: 10.4049/jimmunol.164.1.389
135. Blasi E, Mazzolla R, Pitzurra L, Barluzzi R, Bistoni F. Protective effect of picolinic acid on mice intracerebrally infected with lethal doses of *Candida albicans*. *Antimicrob Agents Chemother* (1993) 37:2422–6. doi: 10.1128/aac.37.11.2422
136. Tomioka H, Shimizu T, Tatano Y. Effects of picolinic acid on the antimicrobial functions of host macrophages against *Mycobacterium avium* complex. *Int J Antimicrob Agents* (2007) 29:460–4. doi: 10.1016/j.ijantimicag.2006.12.010
137. Hellwig-Bürgel T, Stiehl DP, Wagner AE, Metzén E, Jelkmann W. Review: hypoxia-inducible factor-1 (HIF-1): a novel transcription factor in immune reactions. *J Interferon Cytokine Res* (2005) 25:297–310. doi: 10.1089/jir.2005.25.297
138. Elks PM, Brizee S, van der Vaart M, Walmsley SR, van Eeden FJ, Renshaw SA, et al. Hypoxia inducible factor signaling modulates susceptibility to mycobacterial infection via a nitric oxide dependent mechanism. *PLoS Pathog* (2013) 9:e1003789. doi: 10.1371/journal.ppat.1003789
139. Ogryzko NV, Lewis A, Wilson HL, Meijer AH, Renshaw SA, Elks PM. Hif-1 α -Induced Expression of IL-1 β Protects against Mycobacterial Infection in Zebrafish. *J Immunol* (2019) 202:494–502. doi: 10.4049/jimmunol.1801139

140. Braverman J, Sogi KM, Benjamin D, Nomura DK, Stanley SA. HIF-1 α Is an Essential Mediator of IFN- γ -Dependent Immunity to Mycobacterium tuberculosis. *J Immunol* (2016) 197:1287–97. doi: 10.4049/jimmunol.1600266
141. Braverman J, Stanley SA. Nitric Oxide Modulates Macrophage Responses to Mycobacterium tuberculosis Infection through Activation of HIF-1 α and Repression of NF- κ B. *J Immunol* (2017) 199:1805–16. doi: 10.4049/jimmunol.1700515
142. Resende M, Ferreira CM, Barbosa AM, Cardoso MS, Sousa J, Saraiva M, et al. Myeloid HIF-1 α regulates pulmonary inflammation during experimental Mycobacterium tuberculosis infection. *Immunology* (2020) 159:121–9. doi: 10.1111/imm.13131
143. Guan R, Wang J, Li Z, Ding M, Li D, Xu G, et al. Sodium Tanshinone IIA Sulfonate Decreases Cigarette Smoke-Induced Inflammation and Oxidative Stress via Blocking the Activation of MAPK/HIF-1 α Signaling Pathway. *Front Pharmacol* (2018) 9:263. doi: 10.3389/fphar.2018.00263
144. Alaridah N, Lutay N, Tenland E, Rönnholm A, Hallgren O, Puthia M, et al. Mycobacteria Manipulate G-Protein-Coupled Receptors to Increase Mucosal Rac1 Expression in the Lungs. *J Innate Immun* (2017) 9:318–29. doi: 10.1159/000453454
145. Malik ZA, Denning GM, Kusner DJ. Inhibition of Ca(2+) signaling by Mycobacterium tuberculosis is associated with reduced phagosome-lysosome fusion and increased survival within human macrophages. *J Exp Med* (2000) 191:287–302. doi: 10.1084/jem.191.2.287
146. Andr jak C, Almeida DV, Tyagi S, Converse PJ, Ammerman NC, Grosset JH. Characterization of mouse models of Mycobacterium avium complex infection and evaluation of drug combinations. *Antimicrob Agents Chemother* (2015) 59:2129–35. doi: 10.1128/AAC.04841-14
147. Bernut A, Herrmann J, Ordway D, Kremer L. The Diverse Cellular and Animal Models to Decipher the Physiopathological Traits of Mycobacterium abscessus Infection. *Front Cell Infect Microbiol* (2017) 7:100. doi: 10.3389/fcimb.2017.00100
148. Tezera LB, Bielecka MK, Chancellor A, Reichmann MT, Shammari BA, Brace P, et al. Dissection of the host-pathogen interaction in human tuberculosis using a bioengineered 3-dimensional model. *Elife* (2017) 6:21283. doi: 10.7554/eLife.21283

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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