



Recent Advances in Immune Cell Therapy for Glioblastoma

Xianhui Kang^{1,2†}, Yiyang Zheng^{3†}, Wandong Hong^{4†}, Xixi Chen³, Huiting Li³,
Baojun Huang³, Zhenyang Huang³, Hongli Tang^{1*} and Wujun Geng^{1*}

¹ Department of Anesthesiology, The First Affiliated Hospital of Wenzhou Medical University, Wenzhou, China, ² Department of Anesthesiology, The First Affiliated Hospital, College of Medicine, Zhejiang University, Hangzhou, China, ³ First School of Clinical Medicine, Wenzhou Medical University, Wenzhou, China, ⁴ Department of Gastroenterology and Hepatology, The First Affiliated Hospital of Wenzhou Medical University, Wenzhou, China

OPEN ACCESS

Edited by:

Xiaoxing Xiong,
Renmin Hospital of Wuhan University,
China

Reviewed by:

Lin Sun,
First Affiliated Hospital of Chongqing
Medical University, China
Hamilton Shay,
New York Presbyterian Queens Cornell
University, American Samoa

*Correspondence:

Wujun Geng
gengwujun@126.com
Hongli Tang
tanghongliok@126.com

[†]These authors have contributed
equally to this work

Specialty section:

This article was submitted to
Cancer Immunity
and Immunotherapy,
a section of the journal
Frontiers in Immunology

Received: 17 April 2020

Accepted: 11 September 2020

Published: 21 October 2020

Citation:

Kang X, Zheng Y, Hong W, Chen X,
Li H, Huang B, Huang Z, Tang H and
Geng W (2020) Recent Advances in
Immune Cell Therapy for Glioblastoma.
Front. Immunol. 11:544563.
doi: 10.3389/fimmu.2020.544563

Glioblastoma (GBM) is the most malignant form of astrocytoma with short survival and a high recurrence rate and remains a global problem. Currently, surgery, chemotherapy, radiotherapy, and other comprehensive treatments are the main treatment modalities, but patients still have a poor prognosis mainly due to the infiltrative growth of GBM and the protective effect of the blood–brain barrier on tumor cells. Therefore, immunotherapy is expected to be a good option for GBM. In the immune system, different cells play varying roles in the treatment of GBM, so understanding the roles played by various immune cells in treating GBM and considering how to combine these effects to maximize the efficacy of these cells is important for the selection of comprehensive and optimal treatment plans and improving GBM prognosis. Therefore, this study reviews the latest research progress on the role of various types of immune cells in the treatment of GBM.

Keywords: glioblastoma, immunotherapy, immune cell, advances, mechanism

INTRODUCTION

Glioblastoma

Glioblastoma (GBM) is a rare tumor that is one of the most fatal and difficult-to-treat malignancies. Currently, the primary treatment for GBM is still based on surgery, and patients usually have a poor prognosis and poor quality of life (1). The tumor is subcortical, and most grow throughout the supratentorial cerebral hemispheres. It exhibits infiltrative growth, often invades several lobes and deep structures, and has been shown to affect the contralateral cerebral hemisphere through the corpus callosum with the frontal lobe being the most common site of occurrence (2). GBM grows rapidly with 70% to 80% of patients dying of GBM within 3 to 6 months after diagnosis and a 1-year survival rate of only 10%.

Immune Cell Therapy

There is a large body of literature demonstrating that immunotherapy is important for the treatment of GBM. Chimeric antigen receptor (CAR) T cell therapy can directly and accurately identify, localize to, and kill cancer cells. Natural killer (NK) cells control GBM expansion and inhibit tumor progression; dendritic cells (DCs) play a role in GBM immune recognition, and other immune cells play an adjuvant role in radiotherapy and chemotherapy treatments for GBM.

The main feature of GBM metastasis is extensive local invasion, which is different from the rarer event of systemic metastasis. Therefore, cancer immunotherapy at this stage focuses more on the use of immune cells to inhibit the metastasis of GBM.

Advantages of Immune Cell Therapy

Some immune cells with recognition functions can distinguish themselves from nonself cells (which present nonself antigens), providing a great opportunity for the use of immune cells to specifically recognize and kill cancer cells.

Although the metastatic spread of GBM is extremely rare, GBMs can grow in areas of the brain that are hard to access surgically. Compared with the gross removal of tissue *via* surgical resection, the specific recognition and killing ability of immune cells is more likely to remove only the cancer cells, which has great advantages over less specific treatment modalities.

Disadvantages of Immunotherapy

GBM is highly susceptible to recurrence, and most recurrent tumors have been subjected to genotoxic stress from radiotherapy and/or chemotherapy and are, thus, more immunogenic than untreated tumors (3). However, because recurrent gliomas often engage in antigen escape after immunotherapy, it is difficult to perform immunotherapy on these tumors.

CHANGES IN ASSOCIATED IMMUNE SYSTEM AFTER GBM DEVELOPMENT

Because GBM occurs in the brain, the immunosuppression of GBM involves both the tumor itself and the unique immune characteristics of the brain. The interactions of glioma stem cells (GSCs) and the tumor microenvironment play vital roles in

promoting the malignant growth of GBMs. A schematic illustrating the immunosuppressive microenvironment in GBM is shown in **Figure 1**.

Brain Autoimmune Properties

The blood–brain barrier (BBB) is an important line of defense for brain immunity. The BBB is an astrocyte-supported network of tight junctions on the endothelium that prevents the diffusion of hydrophilic macromolecules into the CNS while allowing the entry of small hydrophobic molecules and the active transport of glucose and nutrients (4).

The Immune Microenvironment of GBM Glioma Vasculature

The vasculature within gliomas shows upregulated protein expression of the macromolecules periostin and tenascin C (TNC), which can prevent T cells from moving into glioma-associated vessels and prevent their migration into the brain parenchyma (5).

Upregulation of Immunosuppressive Molecules (Immune Checkpoints)

Immune checkpoints are small molecules present on the cell surface of T lymphocytes that maintain immune homeostasis. Some immune checkpoint genes, such as CTLA-4, PD-1, LAG3, TIM, and BTLA, mediate inhibitory signals, thereby inhibiting T cell activity (6). The expression of CTLA-4 and PD-1 in GBM often rises immensely, which suppresses immunity (3).

Soluble Factors (e.g., Cytokines and Growth Factors)

The soluble factors TGF β , IL-10, and prostaglandin 50 were the earliest immunosuppressive mediators identified in GBM patients. TGF- β TME and IL-10 cause microglia to lose their MHC expression (5).

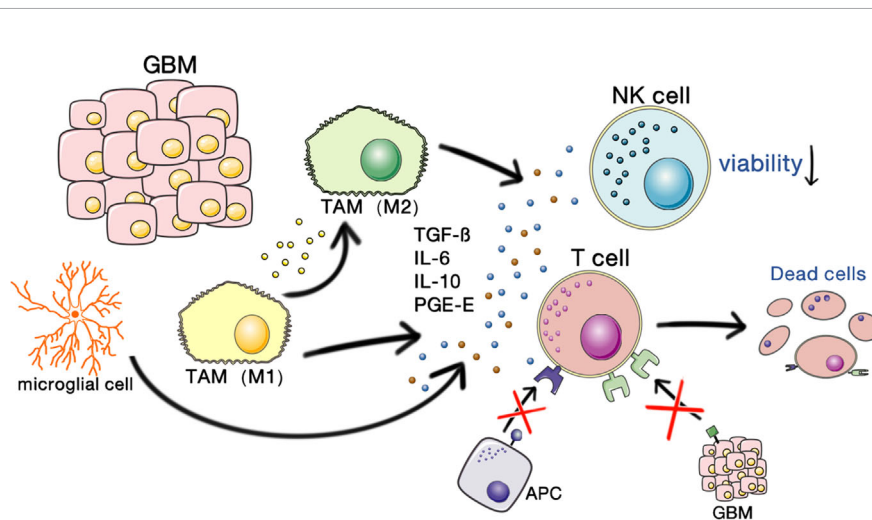


FIGURE 1 | Immunosuppressive microenvironment of GBM. GBM-associated macrophages and microglia secrete inhibitory cytokines, which decrease NK cell activity and T cell-mediated apoptosis and inhibit the binding and killing effects of T cells on antigen-presenting cells and GBM cells. This allows the tumor to escape the immune-killing effects of NK cells and T cells.

Tumor-Associated Immunosuppressive Cells

GBM is characterized by the infiltration of microglia and peripherally recruited macrophages, whereas lymphocytic infiltration is usually low (7). Tumor-associated macrophages (TAMs) secrete inhibitory cytokines, such as interleukin-6 (IL-6), IL-10, transforming growth factor β (TGF- β), and prostaglandin-E, which inhibit NK cell activity and the activation and proliferation of T cells and induce T cell apoptosis, thereby downregulating the expression of MHC and changing TAMs to the M2 phenotype, resulting in immunosuppression (3).

IMMUNE CELL THERAPY FOR GBM

Role of NK Cells in the Treatment of GBM

NK cells are the first natural line of defense against infection and antitumor immunity, and their surface inhibitory receptors recognize MHC class I molecules on the surface of normal somatic cells. When somatic cells are mutated (e.g., GBM), MHC class I expression on their surface is lost, and NK cells initiate a killing effect.

NK cells are persistent in targeting tumor cells and are difficult to escape, and current studies focus on mimicking NK cell activity to replicate their attacking and immune-killing effects (8).

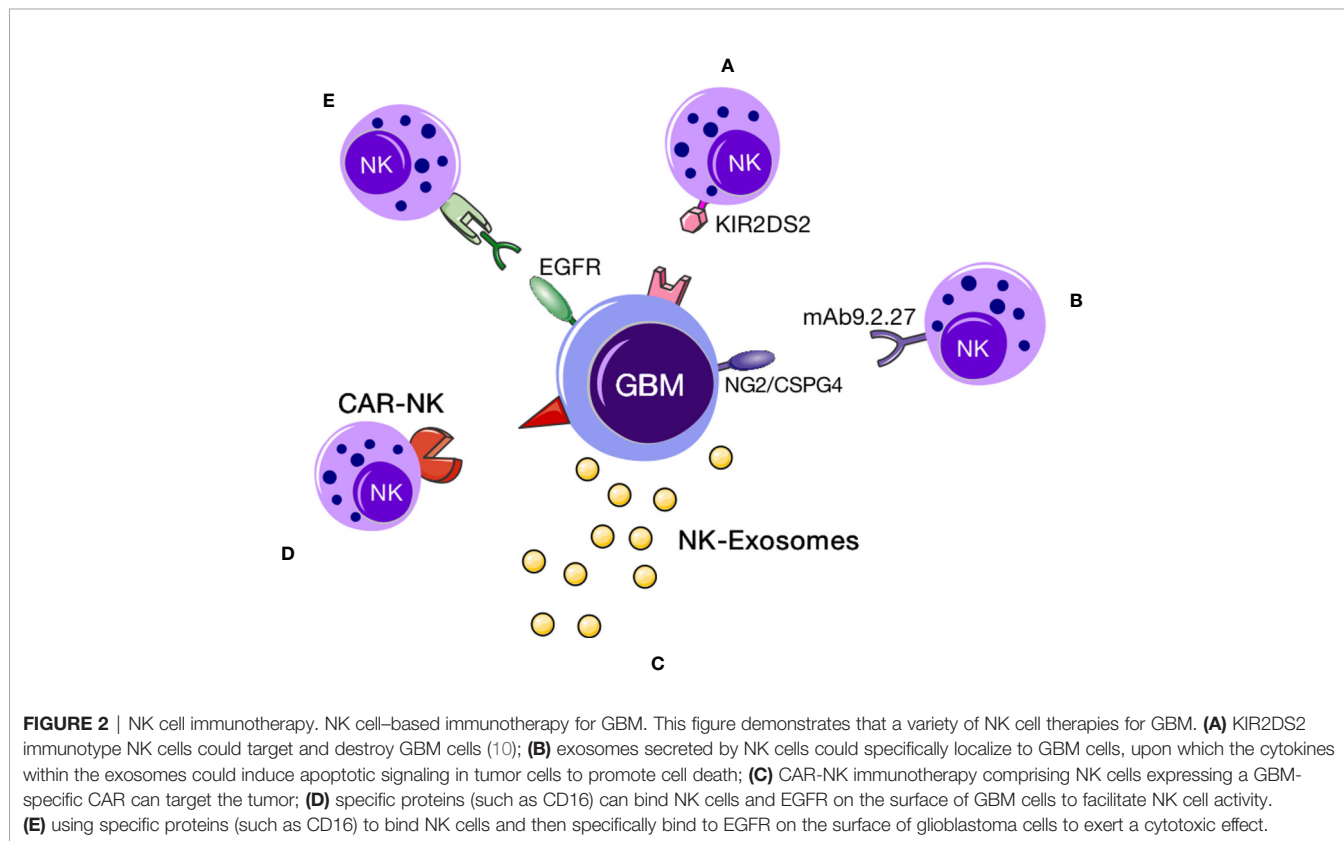
The applications of NK cell therapy for GBM can be summarized as follows: 1. direct use of NK cells to kill GBM cells, 2. combined immune cell therapy regimens comprising NK

cells and immune checkpoint inhibitors or drugs targeting immune-related genes or specific antibodies targeting proteins that protect against immunosuppression of NK cells, and 3. chimeric antigen-modified NK (CAR-NK) cell therapy (9). Images of NK cell-based immunotherapies are shown in **Figure 2**.

NK cells prevent systemic metastasis of GBM. If NK cells are transplanted into a GBM model, GBM death can be directly induced (11), but the difficulty of this method lies in the uncertainty of the transplantation process of the NK cells. It has been suggested that GBM occurs due to cytomegalovirus infection interfering with the immune response of NK cells (12).

Yvon et al. (13) propose an immunotherapy approach for GBM using NK cells derived from cord blood, but this method is similarly limited by immunosuppressive cytokines in the tumor microenvironment. In addition, there have been some studies proposing combination immunotherapies related to NK cells. For example, by using the addition of the immune checkpoint inhibitor PD-1 combined antibody, one group found that decreased PD-1 activity could promote the massive infiltration of NK cells and T cells as well as inhibit tumor progression (14). However, none of these combination immunotherapies have been studied with regard to the specific mechanism of action of other cells beyond NK cells to help kill GBM.

Systemic metastasis of GBM is rare thanks to the innate immunity of NK cells, and studying the combination of radiotherapy, immune cells, and immune checkpoint inhibitors is beneficial to improve the treatment of GBM. Based on the



above studies, although a large amount of experimental data show that the above methods can increase immune cell infiltration, there are still many problems in the clinical application of this regimen. Therefore, the use of NK cells in the brain to kill GBM still has a series of problems, the most important and challenging of which is to prevent the inhibition of the cytotoxic effects of NK cells. NK cells are functionally inhibited in the GBM tumor microenvironment. Kozłowska et al. (15) show that GBM stem cells are highly susceptible to NK-mediated killing, but after differentiation of these stem cells, anergic NK cells fail to control GBM tumor growth because of the release of IL-6 and IL-8.

CAR-NK therapy is one of the latest developments in treating GBM. One group of investigators found that ErbB2 protein expression was elevated in a large proportion of GBM samples and used ErbB2/HER2-specific NK cells to target GBM (16), proposing sustainably expanded “CAR-NK” cells—human NK cells that express ErbB2-specific chimeric antigen receptors. The *in vitro* and *in vivo* effects of these CAR-NK cells on GBM cell culture and orthotopic GBM xenograft models as well as the therapeutic effects of NK-92/5.28 cells on endogenous antitumor immunity were also confirmed. Murakami et al. (17) also propose a method comprising a novel NK cell line carrying a chimeric immune antigen receptor (CAR-KHYG-1) to target epidermal growth factor receptor variant III (EGFRvIII) and induce antitumor effects in GBM cells. Han et al. (18) reveal that CAR-redirectioned NK cells effectively target wt EGFR and EGFRvIII to treat GBM and demonstrate that intracranial application of NK-92-EGFR-CAR cells can effectively inhibit tumor growth, which is a prospective clinical strategy for the treatment of GBM.

In conclusion, owing to the large number of studies on targeted NK cell therapy for GBM in progress, it seems that this treatment modality has a good chance of becoming a full-fledged immunotherapy regimen. However, most of these studies have not made any profound breakthroughs, and the safety and efficacy of adoptive immunotherapy with CAR-NK cells need to be further assessed in clinical trials. Thus, treating GBM with NK cells still has a long way to go.

Dendritic Cells

DC vaccines have been administered clinically for the treatment of GBM, but the results remain unsatisfactory. Pellegatta et al. (19) propose that DC immunotherapy for GBM might be associated with NK cells, that DC vaccines induced significant and sustained NK cell activation, and that the increase in their response had a significant correlation with prolonged patient survival. Dusoswa et al. (20) selected Siglec-9 ligands highly expressed on GBM extracellular vesicles and modified these vesicles with a receptor to promote Siglec binding on the vesicle surface, thereby achieving efficient targeting of adjuvant DCs to GBM and enhancing their potential as anticancer vaccines.

Vaccines are based on DCs containing peptides that represent one or more specific tumor antigens or whole lysates as a source of multiple antigens. However, factors such as the immunosuppressive microenvironment, lack of appropriate specific epitopes, and cancer immunoeediting may limit their efficacy (21).

The activation of DCs can be driven by GBM stem cells and a mixture of monocytes, such as CD34-, CD45-, and CD56-positive cells from allogeneic umbilical cord blood (UCB) (22). Eiraku et al. (23) study the interaction of DCs with CD8+ T cells as well as with V γ 9 γ δ T cells and V α 24NKT cells. Immunocyte therapy based on DCs interacting with GBM lysate (24) is also a promising treatment for GBM.

A large body of literature has indicated that, because DC vaccines themselves are less toxic and do not have many adverse effects, they may become a new hope for the treatment of GBM.

Tumor-Associated Macrophages

Only a few studies have shown that macrophages directly play an immune-related role in the treatment of GBM, and Sun et al. (25) find that inhibiting Romo1 in combination with anti-PD-1 immunotherapy significantly improved the prognosis of GBM patients and particularly enhanced the function of macrophages.

Hallmark indicators of genetic alterations in GBM are amplification of EGFR and EGFRvIII, and investigators have proposed a pathway in which EGFR in combination with EGFRvIII induces macrophage infiltration by upregulating the expression of the chemokine CCL2 (26).

Most existing studies on TAMs have focused on the secretion of cytokines in the GBM microenvironment, promoting GBM progression. Herting et al. (27) find in their study that coculturing TAMs derived from bone marrow with primary GBM cells promoted the upregulation of the cytokine IL-1, which is detrimental to the tumor-killing effect of NK cells and T cells.

TAMs promote the growth of GBM by secreting pleiotropic phosphorus and promoting PTPRZ1 signaling in GBM stem cells (28). In addition, a similar study indicated that Wnt-induced signaling protein 1 (WISP1) secreted from GBM stem cells promotes the survival of both GBM stem cells and TAMs (phagocytes) to establish a tumorigenic microenvironment (29).

In addition, research on macrophages has not been consistent. Here, we simply provide examples of the following recent studies. Macrophage-associated cytokines are used as prognostic indicators of GBM, and increased IL-6 levels predict poor prognosis (30); Wei et al. (31) find that osteopontin (OPN) is an important chemokine for recruiting macrophages into GBM. Cui et al. (32) reported the importance of macrophage-associated immunosuppression in GBM angiogenesis. Although these discoveries have not been fully elaborated upon, they provide new ideas for the treatment of GBM with macrophages, indicating that macrophages play multiple roles and are expected to be applied in other aspects.

In fact, there are both advantages and disadvantages of the use of macrophages in immunotherapy. We found that many studies have proven that macrophages have an adverse effect on the prognosis of GBM. Therefore, we believe that how to apply macrophages in the future to maintain their advantages and avoiding their disadvantages will become a new research focus.

Mast Cells

Põlajeva et al. (33) propose that the accumulation of mast cells (MCs) in GBM tumors might be related to the levels of stem cell

factor and the chemokine CXCL12; Attarha et al. (34) demonstrate that MCs respond to multiple signals in a glioma grade-dependent manner to infiltrate mouse and human gliomas and induce the differentiation of glioma cells. Roy et al. (35) use the degree of recruitment of MCs as a potential biomarker for grading GBM.

APPLICATION OF IMMUNE CELL THERAPY FOR GBM

The GBM immunotherapy category includes adoptive T cell immunotherapy, CAR-T immunotherapy, DC tumor vaccines, immune checkpoint blockade, monoclonal antibodies, and cytokine therapy.

Adoptive T Cell Immunotherapy

Adoptive lymphocyte transfer (ALT) is an antigen-specific treatment during which either tumor-infiltrating lymphocytes (TILs) are obtained from tumor specimens or T cells are isolated from peripheral blood mononuclear cells (PBMCs), expanded *in vitro* against tumor antigens, and systemically applied or directly injected into the tumor site (36). Schuessler et al. (37) report the successful expansion of cytomegalovirus-specific T cells from 13 of 19 patients with recurrent GBM; moreover, 4 of the 10 patients who completed the treatment remained tumor-free during the study period.

Currently, multiple clinical trials have used ALT therapy in GBM patients (NCT01082926, NCT00331526, NCT01588769, NCT00003185, and NCT00730613), and these studies have confirmed the safety and feasibility of ALT therapy (38).

More recently, there has been progress in a clinical trial involving adoptive cellular immunotherapies (ACT), which has shown that CMV-specific ACT can effectively delay or even prevent the recurrence of GBM, which indicates that a favorable T cell gene signature is associated with the improvement in therapeutic efficacy and prolonged survival (39).

CAR-T Immunotherapy

CAR-T immunotherapy is a precisely targeted therapy for the treatment of tumors, which transduces a CAR into T cells to create CAR-T cells (40), after which they are expanded to large numbers *in vitro* and then re injected into the patient, prompting B cells to produce antibodies and specifically recognize antigens, which, in turn, kill the tumors (41).

CAR-T immunotherapy has the capacity to cross the BBB and can safely and effectively reach tumor cells that cannot be accessed surgically (42). Brown et al. (38) treated a patient with recurrent GBM by using CAR-T cells and found that IL13R-2 was a useful immunotherapeutic target in GBM. Although this therapy has been recognized by many patients, and 4 categories have been clinically approved in China, it is still not considered a conventional treatment.

After reviewing many studies on the subject, we deduced that CAR-T therapy has not made a breakthrough in the treatment of solid tumors in recent years, and there are many unsolved issues,

especially in terms of CAR-T cells entering the microenvironment of solid tumors, such as GBM, maintaining their viability and ability to rapidly and accurately identify tumor cells, and overcoming immunosuppression. Therefore, if the issue of tumor microenvironment inhibition of the CAR-T cell therapeutic effects can be solved at this stage, it will have a great impact on the immunotherapy of solid tumors.

Dendritic Cell Vaccines

The production of DC vaccines includes isolating DCs from patients, loading the cells with tumor antigens, culturing the DCs with cytokines to induce maturation, and re injecting the cells back into the body (43). At present, vaccines are broadly divided into three categories according to different antigens: tumor-associated antigens (TAAs), tumor-specific antigens (TSAs), and tumor lysates (44).

1. *TAAs are ubiquitous but are expressed at higher levels in tumor cells than in healthy cells, so TAA vaccines are easy to develop and have good targeting.* At present, the clinical application of TAA-based DC vaccines is limited, mainly due to the following reasons: 1. There are few known TAAs, 2. the consistency of TAA expression marks its own limitations, and 3. TAA vaccines may not induce the best immune response due to immune tolerance (44). Wen et al. (45) discovered that ICT-107 vaccination in patients with newly diagnosed GBM developed good tolerance and significantly improved survival by 2.2 months.
2. *TSAs are unique to tumor cells and, unlike TAAs in tumor cells and normal cells, are usually proteins encoded by mutated genes in tumors.* They are relatively fixed in different types of cancer and patients and can be used as targets for immunotherapy (43). TSA-based DC vaccines may generate an intense targeted inflammatory response against tumor cells while avoiding potential autoimmune responses in other tissues (44). Rindopepimut (46) (Celldex Therapeutics, Hampton, New Jersey, USA), a TSA vaccine, has shown clinical benefits and significant efficacy in phase II clinical trials. However, the phase III clinical trial was terminated early because it was thought that the patients in the study might not reach their primary endpoint.
3. *Vaccination in combination with tumor lysates is usually delivered by autologous lysate-pulsed DCs, which are usually collected from the patient a few days before surgical resection, incubated with the resected tumor lysate, and then reintroduced back into patients via postoperative multiple vaccinations in combination with standardized radiotherapy administration to target residual tumor cells* (44). The phase III trial by Liao et al. (47) demonstrated that the addition of DCVax-L to the standard of care for GBM patients is feasible and safe and prolongs survival. The combination of tumor antigens and α -GalCer in anticancer vaccines can efficiently induce long-lasting immunity by activating iNK T cells (48).

Therefore, the latest research direction of DC vaccines at this stage focuses on combining tumor lysates with DCs, and the

mechanism of tumor lysate vaccines on GBM is very likely to be the combined action of multiple immune cells.

PROSPECT OF IMMUNOTHERAPY FOR GBM

Different Combination Regimens

Immunotherapy for GBM has not been used in a wide range of direct treatments due to its immaturity and our incomplete understanding of the mechanisms, so the main treatment modalities after diagnosis confirmation are surgical resection, radiotherapy, chemotherapy, and various comprehensive treatments. Conventional treatment is maximal gross resection of the tumor followed by radiotherapy and chemotherapy, and maintenance therapy with temozolomide (TMZ) is started 4 weeks after completion of the chemoradiotherapy cycle. This treatment regimen is known the standard of care (SOC). The common chemotherapeutic agent axitinib has been shown by Stephanie Du Four et al. to have a favorable effect on immune function (49).

There are currently several main GBM vaccines available:

1. autologous monocyte vaccine, 2. peptide-based tumor vaccine, 3. nucleotide-based tumor vaccine, and 4. cell line-based tumor vaccine. Each of them requires coculture of the corresponding cells with surgically resected tumor cells under different conditions to achieve immunization against tumor cells. Therefore, it is also necessary to obtain a sufficient number of tumor cells at the time of surgery.

CRS-T is a genetically modified chimera-switched receptor T cell therapy targeting PD-1. After intravenous infusion of CSR-T cells, the levels of IFN- γ and IL-6 in peripheral blood increase with the number of reinfused cells, and local intracranial injection of CSR-T cells is often more effective than intravenous injection (50).

Immunotherapy with DC vaccines has been associated with adverse effects of immunotherapy in newly diagnosed patients, and increases in tumor-specific immune responses after vaccination, including immune cell proliferation and cytokine production, can be detected (51). For patients with recurrent GBM, elevated levels of chemoresistance-associated peptides (CAPs) and/or cytoplasmic accumulation can be observed in fusion cells generated after cervical implantation of autologous glioma cells and DCs, and a specific immune response to CAPs can also be observed, which promotes an antitumor response in patients (52).

Cytokine-induced killer (CIK) cells are nonhistocompatibility (MHC)-restricted lymphotoxic cells that can be produced from PBMCs under the induction of interferon (IFN)- γ , IL-2, and CD3 monoclonal antibodies (CD3 mAb) and have a high proliferation rate and antitumor activity. Chemoradiation with CIK cells plus the standard radiotherapy-TMZ regimen shows no significant difference in survival but improves progression-free survival compared with that of TMZ alone; unfortunately, no evidence of improved overall survival was found (53).

Rindopepimut (also known as CDX-110) is a peptide-based vaccine against EGFRvIII, an EGFR variate with a deletion mutation, and the addition of rindopepimut to a standardized

course did not increase survival in patients with newly diagnosed GBM in clinical trials. Further exploration of the effect of immunotherapy in future treatment combinations containing rindopepimut may be required (54).

Different vaccines are injected in different ways, and some cellular vaccines can be injected directly into the surgical cavity. After surgical resection of GBM, patients undergoing the SOC followed by local injection of an immune-stimulating oligonucleotide containing an unmethylated cytosine-guanosine motif (CpG-ODN) immediately around the surgical cavity were more likely to develop fever and postoperative hematoma after surgery than patients who received the SOC alone with similar incidences of other adverse events. Overall, this vaccine did not improve survival in patients with newly diagnosed GBM (55).

In addition, vaccines made from oncolytic viruses are also under investigation. The oncolytic virus aglatimagene besadenovec (AdV-tk) combined with valacyclovir constitutes gene-mediated cytotoxic immunotherapy (GMCI). When combined with the SOC, GMCI may stimulate immune responses in participants with GTR and subtotal resection. However, if the residual tumor burden is too large, the tumor-mediated immunosuppressive effect may mask the effect of GMCI (56).

At present, most immunotherapies can only be performed after patients undergo surgical resection, which is a limitation of immunotherapy. However, as a brain tumor, GBM is very likely to grow in sites that are not suitable for surgical resection and are more likely to recur, and treatment with immune cells or vaccines can achieve the destruction of tumors at sites that are not easily accessible *via* surgery; therefore, if the field of immunotherapy can mature, this treatment modality may replace surgical resection.

Role of Exosomes in Tumor Immunology

Exosomal vesicles naturally released by tumor cells transmit some molecules to target cells and act as an intercellular signaling pathway between the donor cytoplasm and the lumen of target cells. For example, NK cells release exosomes that express typical NK markers (CD56, etc.), killer proteins, and stimulate antitumor activity (57, 58). Exosomes carrying different RNAs and proteins can be measured in patients as biomarkers to assess disease onset and progression (59).

Barile et al. (60) demonstrate that GBM stem cells could secrete exosomes carrying active vascular endothelial growth factor A (VEGF-A), which could protect VEGF-A from cytokines, proteases, etc., in the tumor microenvironment and maintain the vascular niche. Non-GBM-stem-cell-derived VEGF-A changes with tumor size and treatment, whereas GBM-stem-cell-derived exosomes may be continuously produced and released and cross the BBB.

Skog et al. (61) and Brennan et al. (62) report that EGFRvIII could be detected in the serum exosomes of GBM patients with high diagnostic sensitivity, but EGFRvIII was present in only approximately 25% of GBM patients. Kai et al. (63) demonstrate that PTRF was also present on the exosomal membrane and that PTRF overexpression increased exosome secretion, resulting in

increased rates of tumor formation and receptor proliferation *in vitro* and *in vivo*.

A study by Liu et al. (48) shows that, in a DC vaccine, the codelivery of tumor-derived exosomes with α -galactosylceramide (α -GalCer) could effectively improve the tumor microenvironment by balancing the release of immunosuppressive factors and immunostimulatory factors. Zhu et al. (64) find that exosomes secreted by NK cells could specifically localize to GBM tumors and that NK-Exos contain FasL, perforin, granzyme B, and TNF- α , thereby inducing proapoptotic signals and triggering cell death in tumors. Chen et al. (65) propose that exosomes could interact with target cells with barrier cells. It is a new promising therapeutic agent for the treatment of refractory GBM.

Exosomal transfer of miRNAs or miRNA inhibitors to tumor cells has emerged as a new approach to deliver miRNAs that can target cancer. MiRNA-21 is overexpressed in GBM, which can improve the proliferation and malignant metastatic behavior of tumor cells (66, 67). Sponge constructs are designed to bind to their complementary miRNAs (one or more) or their seed sequences, thereby preventing miRNA binding to their biological targets.

GBM cells use exosomes to communicate with the tumor microenvironment and promote their proliferation, invasion, and metastasis (68). One of the major challenges of exosome-based therapeutic approaches is the low productivity of exosomes. Therefore, effective methods that can produce exosomes on a large scale are needed. Watson et al. (69) propose that the use of hollow fiber bioreactors could increase the yield of exosomes by 5- to 10-fold.

Although exosomes have been a research hot spot in recent years, the still-limited data on the use of exosomes as a targeted therapy for GBM is far from mature. There is no denying the fact that using GBM-derived and -targeted exosomes is a very good idea, but more research and clinical trials are needed to truly determine their effect in patients, which can take years. Thus, it is difficult to judge whether this approach is worth pursuing, but at this stage, we are optimistic that ongoing clinical trials will provide a preliminary basis for the use of exosomes in immune therapy.

Research Focuses and Prospects of Other Immune Cell Therapies

One popular direction is the implementation of combined antigen immunotherapies with the latest focus on CAR-Ts and “bispecific T cell engagers” (BiTEs) (70, 71), that is, using antibodies such as BiTEs to bind both T cells and tumor cells and create a molecular bridge that induces T cells to kill GBM cells.

REFERENCES

- Alexander BM, Cloughesy TF. Adult Glioblastoma. *J Clin Oncol* (2017) 35 (21):2402–9. doi: 10.1200/JCO.2017.73.0119
- Chaichana KL, Parker SL, Olivi A, Quiñones-Hinojosa A. Long-term seizure outcomes in adult patients undergoing primary resection of malignant brain astrocytomas. *Clinical article. J Neurosurg* (2009) 111(2):282–92. doi: 10.3171/2009.2.JNS081132
- Lim M, Xia YX, Bettegowda C, Weller M. Current state of immunotherapy for glioblastoma. *Nat Rev Clin Oncol* (2018) 15(7):422–42. doi: 10.1038/s41571-018-0003-5
- Jackson CM, Choi J, Lim M. Mechanisms of immunotherapy resistance: lessons from glioblastoma. *Nat Immunol* (2019) 20(9):1100–9. doi: 10.1038/s41590-019-0433-y
- Brown NF, Carter TJ, Ottaviani D, Mulholland P. Harnessing the immune system in glioblastoma. *Br J Cancer* (2018) 119(10):1171–81. doi: 10.1038/s41416-018-0258-8

In addition, following CAR-T therapy, a series of CAR immunotherapies have been investigated, such as CAR-NK cells (72) and CAR-macrophages. In the field of cancer immunotherapy, CAR-T cell therapy and PD-1 inhibitors are the most high-profile members, but in addition to T cells, the functions of NK cells and macrophages should not be ignored.

SUMMARY

In summary, it is of great significance to explore the mechanisms of action of immunotherapy in depth for the treatment of GBM and to specifically study these mechanisms of action in each type of immune cell. In addition, immunotherapy has incomparable advantages to surgical treatment, radiotherapy, and chemotherapy.

We believe that the most valuable applications of immunotherapy for the treatment of GBM are CAR-T cell therapy and DC vaccines. These two methods have a large body of preliminary data and have undergone clinical trials and applications. Although there may be some issues, any problems that arise are expected to be resolved in follow-up studies. Therefore, in the process of GBM occurrence and expansion, the use of appropriate treatment options and attempts to use immunotherapy are conducive to not only the broad killing of cancer cells but also mediating the occurrence and expansion rate of GBM. This activity may strongly contribute to the treatment of GBM and even other types of cancer.

AUTHOR CONTRIBUTIONS

XK, HT, and WG contributed to the conception and design of the study. XK, YZ, WG, XC, and WH wrote the first draft of the manuscript. HL, BH, and ZH wrote sections of the manuscript. All authors contributed to the article and approved the submitted version.

FUNDING

This study was funded by the National Natural Science Foundation of China (81774109, 81973620), the Natural Science Foundation of Zhejiang Province (Y19H310028), the Wenzhou Science and Technology Project (ZY2019015), and the Zhejiang Public Welfare Technology Research Plan (LGD20H290002).

6. Chang LS, Barroso-Sousa R, Tolaney SM, Hodi FS, Kaiser UB, Min L. Endocrine Toxicity of Cancer Immunotherapy Targeting Immune Checkpoints. *Endocr Rev* (2019) 40(1):17–65. doi: 10.1210/er.2018-00006
7. Luoto S, Hermelo I, Vuorinen EM, Hannus P, Kesseli J, Nykter M, et al. Computational Characterization of Suppressive Immune Microenvironments in Glioblastoma. *Cancer Res* (2018) 78(19):5574–85. doi: 10.1158/0008-5472.CAN-17-3714
8. Golán I, Rodríguez de la FL, Costoya JA. NK Cell-Based Glioblastoma Immunotherapy. *Cancers (Basel)* (2018) 10(12):1–32. doi: 10.3390/cancers10120522
9. Burger MC, Zhang CC, Harter PN, Romanski A, Strassheimer F, Senft C, et al. CAR-Engineered NK Cells for the Treatment of Glioblastoma: Turning Innate Effectors Into Precision Tools for Cancer Immunotherapy. *Front Immunol* (2019) 10:2683. doi: 10.3389/fimmu.2019.02683
10. Gras NA, Kmiecik J, Leiss L, Zekowski M, Engelsens A, Bruserud Ø., et al. NK cells with KIR2DS2 immunogenotype have a functional activation advantage to efficiently kill glioblastoma and prolong animal survival. *J Immunol* (2014) 193(12):6192–206. doi: 10.4049/jimmunol.1400859
11. Lee SJ, Kang WY, Yoon Y, Jin JY, Song HJ, Her JH, et al. Natural killer (NK) cells inhibit systemic metastasis of glioblastoma cells and have therapeutic effects against glioblastomas in the brain. *BMC Cancer* (2015) 15:1–28. doi: 10.1186/s12885-015-2034-y
12. Dominguez-Valentin M, Gras NA, Rahman AM, Kumar S, Retière C, Ulvestad E, et al. Identification of a Natural Killer Cell Receptor Allele That Prolongs Survival of Cytomegalovirus-Positive Glioblastoma Patients. *Cancer Res* (2016) 76(18):5326–36. doi: 10.1158/0008-5472.CAN-16-1162
13. Yvon ES, Burga R, Powell A, Cruz CR, Fernandes R, Barese C, et al. Cord blood natural killer cells expressing a dominant negative TGF- β receptor: Implications for adoptive immunotherapy for glioblastoma. *Cytotherapy* (2017) 19(3):408–18. doi: 10.1016/j.jcyt.2016.12.005
14. Shevtsov M, Pitkin E, Ischenko A, Stangl S, Khachatryan W, Galibin O, et al. Ex vivo Hsp70-Activated NK Cells in Combination With PD-1 Inhibition Significantly Increase Overall Survival in Preclinical Models of Glioblastoma and Lung Cancer. *Front Immunol* (2019) 10:454. doi: 10.3389/fimmu.2019.00454
15. Kozłowska AK, Tseng H-C, Kaur K, Topchyan P, Inagaki A, Bui VT, et al. Resistance to cytotoxicity and sustained release of interleukin-6 and interleukin-8 in the presence of decreased interferon- γ after differentiation of glioblastoma by human natural killer cells. *Cancer Immunol Immunother* (2016) 65(9):1085–97. doi: 10.1007/s00262-016-1866-x
16. Zhang CC, Burger MC, Jennewein L, Gensler S, Schönfeld K, Zeiner P, et al. ErbB2/HER2-Specific NK Cells for Targeted Therapy of Glioblastoma. *J Natl Cancer Inst* (2016) 108(5):1–12. doi: 10.1093/jnci/djv375
17. Murakami T, Nakazawa T, Natsume A, Nishimura F, Nakamura M, Matsuda R, et al. Novel Human NK Cell Line Carrying CAR Targeting EGFRvIII Induces Antitumor Effects in Glioblastoma Cells. *Anticancer Res* (2018) 38(9):5049–56. doi: 10.21873/anticancer.12824
18. Han JF, Chu JH, Keung CW, Zhang JY, Wang YW, Cohen JB, et al. CAR-Engineered NK Cells Targeting Wild-Type EGFR and EGFRvIII Enhance Killing of Glioblastoma and Patient-Derived Glioblastoma Stem Cells. *Sci Rep* (2015) 5:11483. doi: 10.1038/srep11483
19. Pellegatta S, Eoli M, Cuccarini V, Anghileri E, Pollo B, Pessina S, et al. Survival gain in glioblastoma patients treated with dendritic cell immunotherapy is associated with increased NK but not CD8 T cell activation in the presence of adjuvant temozolomide. *Oncoimmunology* (2018) 7(4):e1412901. doi: 10.1080/2162402X.2017.1412901
20. Dusoswa SA, Horrevorts SK, Ambrosini M, Kalay H, Paauw NJ, Nieuwland R, et al. Glycan modification of glioblastoma-derived extracellular vesicles enhances receptor-mediated targeting of dendritic cells. *J Extracell Vesicles* (2019) 8(1):1648995. doi: 10.1080/20013078.2019.1648995
21. Finocchiaro G, Pellegatta S. Immunotherapy with dendritic cells loaded with glioblastoma stem cells: from preclinical to clinical studies. *Cancer Immunol Immunother* (2016) 65(1):101–9. doi: 10.1007/s00262-015-1754-9
22. Yurtsever A, Haydaroglu A, Biray AC, Gunduz C, Oktar N, Dalbasti T, et al. Assessment of genetic markers and glioblastoma stem-like cells in activation of dendritic cells. *Hum Cell* (2013) 26(3):105–13. doi: 10.1007/s13577-013-0065-8
23. Eiraku Y, Terunuma H, Yagi M, Deng X, Nicol AJ, Nieda M. Dendritic cells cross-talk with tumour antigen-specific CD8 T cells, V γ 9 δ T cells and V α 24NKT cells in patients with glioblastoma multiforme and in healthy donors. *Clin Exp Immunol* (2018) 194(1):54–66. doi: 10.1111/cei.13185
24. Nava S, Lisini D, Pogliani S, Dossena M, Bersano A, Pellegatta S, et al. Safe and Reproducible Preparation of Functional Dendritic Cells for Immunotherapy in Glioblastoma Patients. *Stem Cells Transl Med* (2015) 4(10):1164–72. doi: 10.5966/sctm.2015-0091
25. Sun G, Cao Y, Qian C, Wan Z, Zhu J, Guo J, et al. Romo1 is involved in the immune response of glioblastoma by regulating the function of macrophages. *Aging (Albany NY)* (2020) 12:1114–27. doi: 10.18632/aging.102648
26. An Z, Knobbe-Thomsen CB, Wan X, Fan QW, Reifemberger G, Weiss WA. Correction: EGFR Cooperates with EGFRvIII to Recruit Macrophages in Glioblastoma. *Cancer Res* (2019) 79(21):5681. doi: 10.1158/0008-5472.CAN-19-2800
27. Herting CJ, Chen Z, Maximov V, Duffy A, Szulzewsky F, Shayakhmetov DM, et al. Tumour-associated macrophage-derived interleukin-1 mediates glioblastoma-associated cerebral oedema. *Brain* (2019) 142(12):3834–51. doi: 10.1093/brain/awz331
28. Shi Y, Ping YF, Zhou W, He ZC, Chen C, Bian BSJ, et al. Tumour-associated macrophages secrete pleiotrophin to promote PTPRZ1 signalling in glioblastoma stem cells for tumour growth. *Nat Commun* (2017) 8:15080. doi: 10.1038/ncomms15080
29. Tao W, Chu C, Zhou W, Huang Z, Zhai K, Fang X, et al. Dual Role of WISP1 in maintaining glioma stem cells and tumor-supportive macrophages in glioblastoma. *Nat Commun* (2020) 11(1):3015. doi: 10.1038/s41467-020-16827-z
30. Hori T, Sasayama T, Tanaka K, Koma YII, Nishihara M, Tanaka H, et al. Tumor-associated macrophage related interleukin-6 in cerebrospinal fluid as a prognostic marker for glioblastoma. *J Clin Neurosci* (2019) 68:281–9. doi: 10.1016/j.jocn.2019.07.020
31. Wei J, Marisetty A, Schrand B, Gabrusiewicz K, Hashimoto Y, Ott M, et al. Osteopontin mediates glioblastoma-associated macrophage infiltration and is a potential therapeutic target. *J Clin Invest* (2019) 129(1):137–49. doi: 10.1172/JCI121266
32. Cui X, Morales RTT, Qian W, Wang H, Gagner JP, Dolgalev I, et al. Hacking macrophage-associated immunosuppression for regulating glioblastoma angiogenesis. *Biomaterials* (2018) 161:164–78. doi: 10.1016/j.biomaterials.2018.01.053
33. Pölajeva J, Sjösten AM, Lager N, Kastemar M, Waern I, Alafuzoff I, et al. Mast cell accumulation in glioblastoma with a potential role for stem cell factor and chemokine CXCL12. *PLoS One* (2011) 6(9):e25222. doi: 10.1371/journal.pone.0025222
34. Attarha S, Roy A, Westermark B, Tchougounova E. Mast cells modulate proliferation, migration and stemness of glioma cells through downregulation of GSK3 β expression and inhibition of STAT3 activation. *Cell Signal* (2017) 37:81–92. doi: 10.1016/j.cellsig.2017.06.004
35. Roy A, Attarha S, Weishaupt H, Edqvist PH, Swartling FJ, Bergqvist M, et al. Serglycin as a potential biomarker for glioma: association of serglycin expression, extent of mast cell recruitment and glioblastoma progression. *Oncotarget* (2017) 8(15):24815–27. doi: 10.18632/oncotarget.15820
36. Farber SH, Elsamadicy AA, Atik AF, Suryadevara CM, Chongsathidkiet P, Fecci PE, et al. The Safety of available immunotherapy for the treatment of glioblastoma. *Expert Opin Drug Saf* (2017) 16(3):277–87. doi: 10.1080/14740338.2017.1273898
37. Schuessler A, Smith C, Beagley L, Boyle GM, Rehan S, Matthews K, et al. Autologous T-cell therapy for cytomegalovirus as a consolidative treatment for recurrent glioblastoma. *Cancer Res* (2014) 74(13):3466–76. doi: 10.1158/0008-5472.CAN-14-0296
38. Brown CE, Alizadeh D, Starr R, Weng L, Wagner JR, Naranjo A, et al. Regression of Glioblastoma after Chimeric Antigen Receptor T-Cell Therapy. *N Engl J Med* (2016) 375(26):2561–9. doi: 10.1056/NEJMoa1610497
39. Smith C, Lineburg KE, Martins JP, Ambalathingal G, Neller MA, Morrison B, et al. Autologous CMV-specific T cells are a safe adjuvant immunotherapy for primary glioblastoma multiforme. *J Clin Invest* (2020). doi: 10.1172/JCI138649
40. Srivastava S, Riddell SR. Engineering CAR-T cells: Design concepts. *Trends Immunol* (2015) 36(8):494–502. doi: 10.1016/j.it.2015.06.004

41. Bagley SJ, Desai AS, Linette GP, June CH, O'Rourke DM. CAR T-cell therapy for glioblastoma: recent clinical advances and future challenges. *Neuro-oncology* (2018) 20(11):1429–38. doi: 10.1093/neuonc/now032
42. Filley AC, Henriquez M, Dey M. CART Immunotherapy: Development, Success, and Translation to Malignant Gliomas and Other Solid Tumors. *Front Oncol* (2018) 8:453. doi: 10.3389/fonc.2018.00453
43. Kong Z, Wang Y, Ma W. Vaccination in the immunotherapy of glioblastoma. *Hum Vaccin Immunother* (2018) 14(2):255–68. doi: 10.1080/21645515.2017.1388481
44. Wilcox JA, Ramakrishna R, Magge R. Immunotherapy in Glioblastoma. *World Neurosurg* (2018) 116:518–28. doi: 10.1016/j.wneu.2018.04.020
45. Wen PY, Reardon DA, Armstrong TS, Phuphanich S, Aiken RD, Landolfi JC, et al. A Randomized Double-Blind Placebo-Controlled Phase II Trial of Dendritic Cell Vaccine ICT-107 in Newly Diagnosed Patients with Glioblastoma. *Clin Cancer Res* (2019) 25(19):5799–807. doi: 10.1158/1078-0432.CCR-19-0261
46. Elsamadicy AA, Chongsathidkiet P, Desai R, Woroniecka K, Farber SH, Fecci PE, et al. Prospect of rindopepimut in the treatment of glioblastoma. *Expert Opin Biol Ther* (2017) 17(4):507–13. doi: 10.1080/14712598.2017.1299705
47. Liau LM, Ashkan K, Tran DD, Campian JL, Trusheim JE, Cobbs CS, et al. First results on survival from a large Phase 3 clinical trial of an autologous dendritic cell vaccine in newly diagnosed glioblastoma. *J Transl Med* (2018) 16(1):142. doi: 10.1186/s12967-018-1507-6
48. Liu H, Chen L, Liu J, Meng HX, Zhang R, Ma L, et al. Co-delivery of tumor-derived exosomes with alpha-galactosylceramide on dendritic cell-based immunotherapy for glioblastoma. *Cancer Lett* (2017) 411:182–90. doi: 10.1016/j.canlet.2017.09.022
49. Du FS, Maenhout SK, Bentejn D, De KB, Duerinck J, Thielemans K, et al. Disease progression in recurrent glioblastoma patients treated with the VEGFR inhibitor axitinib is associated with increased regulatory T cell numbers and T cell exhaustion. *Cancer Immunol Immunother* (2016) 65(6):727–40. doi: 10.1007/s00262-016-1836-3
50. Guo JX, Wu CX, Wang PF, Li ZJ, Han S, Jin W, et al. Bioactivity and safety of chimeric switch receptor T cells in glioblastoma patients. *Front Biosci (Landmark Ed)* (2019) 24:1158–66. doi: 10.2741/4772
51. Inogés S, Tejada S, Lopez-Diaz de Cerio Ascensión, Gállego PLJ, Espinós J, Idoate MA, et al. A phase II trial of autologous dendritic cell vaccination and radiochemotherapy following fluorescence-guided surgery in newly diagnosed glioblastoma patients. *J Transl Med* (2017) 15(1):104. doi: 10.1186/s12967-017-1202-z
52. Akasaki Y, Kikuchi T, Homma S, Koido S, Ohkusa T, Tasaki T, et al. Phase I/II trial of combination of temozolomide chemotherapy and immunotherapy with fusions of dendritic and glioma cells in patients with glioblastoma. *Cancer Immunol Immunother* (2016) 65(12):1499–509. doi: 10.1007/s00262-016-1905-7
53. Kong DS, Nam DH, Kang SH, Lee JW, Chang JH, Kim JH, et al. Phase III randomized trial of autologous cytokine-induced killer cell immunotherapy for newly diagnosed glioblastoma in Korea. *Oncotarget* (2017) 8(4):7003–13. doi: 10.18632/oncotarget.12273
54. Weller M, Butowski N, Tran DD, Recht LD, Lim M, Hirte H, et al. Rindopepimut with temozolomide for patients with newly diagnosed, EGFRvIII-expressing glioblastoma (ACT IV): a randomised, double-blind, international phase 3 trial. *Lancet Oncol* (2017) 18(10):1373–85. doi: 10.1016/S1470-2045(17)30517-X
55. Ursu R, Carpentier A, Metellus P, Lubrano V, Laigle-Donadey F, Capelle L, et al. Intracerebral injection of CpG oligonucleotide for patients with de novo glioblastoma-A phase II multicentric, randomised study. *Eur J Cancer* (2017) 73:30–7. doi: 10.1016/j.ejca.2016.12.003
56. Wheeler LA, Manzanera AG, Bell SD, Cavaliere R, McGregor JM, Grecula JC, et al. Phase II multicenter study of gene-mediated cytotoxic immunotherapy as adjuvant to surgical resection for newly diagnosed malignant glioma. *Neuro-oncology* (2016) 18(8):1137–45. doi: 10.1093/neuonc/now002
57. Parolini I, Federici C, Raggi C, Lugini L, Palleschi S, De MA, et al. Microenvironmental pH is a key factor for exosome traffic in tumor cells. *J Biol Chem* (2009) 284(49):34211–22. doi: 10.1074/jbc.M109.041152
58. Shoaie-Hassani A, Hamidieh AA, Behfar M, Mohseni R, Mortazavi-Tabatabaei SA, Asgharzadeh S. NK Cell-derived Exosomes From NK Cells Previously Exposed to Neuroblastoma Cells Augment the Antitumor Activity of Cytokine-activated NK Cells. *J Immunother* (2017) 40(7):265–76. doi: 10.1097/CJI.0000000000000179
59. Treps L, Perret R, Edmond S, Ricard D, Gavard J. Glioblastoma stem-like cells secrete the pro-angiogenic VEGF-A factor in extracellular vesicles. *J Extracell Vesicles* (2017) 6(1):1359479. doi: 10.1080/20013078.2017.1359479
60. Barile L, Vassalli G. Exosomes: Therapy delivery tools and biomarkers of diseases. *Pharmacol Ther* (2017) 174:63–78. doi: 10.1016/j.pharmthera.2017.02.020
61. Skog J, Würdinger T, van RS, Meijer DH, Gainche L, Sena-Estevés M, et al. Glioblastoma microvesicles transport RNA and proteins that promote tumour growth and provide diagnostic biomarkers. *Nat Cell Biol* (2008) 10(12):1470–6. doi: 10.1038/ncb1800
62. Brennan CW, Verhaak RGW, McKenna A, Campos B, Nourbakhsh H, Salama SR, et al. The somatic genomic landscape of glioblastoma. *Cell* (2013) 155(2):462–77. doi: 10.1016/j.cell.2013.09.034
63. Huang K, Fang C, Yi K, Liu X, Qi H, Tan Y, et al. The role of PTRF/Cavin1 as a biomarker in both glioma and serum exosomes. *Theranostics* (2018) 8(6):1540–57. doi: 10.7150/thno.22952
64. Zhu L, Kalimuthu S, Gangadaran S, Gangadaran P, Oh JM, Lee HW, Baek SH, et al. Exosomes Derived From Natural Killer Cells Exert Therapeutic Effect in Melanoma. *Theranostics* (2017) 7(10):2732–45. doi: 10.7150/thno.18752
65. Chen CC, Liu L, Ma F, Wong CW, Guo XE, Chacko JV, et al. Elucidation of Exosome Migration across the Blood-Brain Barrier Model In Vitro. *Cell Mol Bioeng* (2016) 9(4):509–29. doi: 10.1007/s12195-016-0458-3
66. Monfared H, Jahangard Y, Nikkha M, Mirnajafi-Zadeh J, Mowla SJ. Potential Therapeutic Effects of Exosomes Packed With a miR-21-Sponge Construct in a Rat Model of Glioblastoma. *Front Oncol* (2019) 9:782. doi: 10.3389/fonc.2019.00782
67. Chai C, Song LJ, Han SY, Li XQ, Li M. MicroRNA-21 promotes glioma cell proliferation and inhibits senescence and apoptosis by targeting SPRY1 via the PTEN/PI3K/AKT signaling pathway. *CNS Neurosci Ther* (2018) 24(5):369–80. doi: 10.1111/cns.12785
68. Kucharzewska P, Christianson HC, Welch JE, Svensson KJ, Fredlund E, Ringnér M, et al. Exosomes reflect the hypoxic status of glioma cells and mediate hypoxia-dependent activation of vascular cells during tumor development. *Proc Natl Acad Sci U S A* (2013) 110(18):7312–7. doi: 10.1073/pnas.1220998110
69. Watson DC, Bayik D, Srivatsan A, Bergamaschi C, Valentin A, Niu G, et al. Efficient production and enhanced tumor delivery of engineered extracellular vesicles. *Biomaterials* (2016) 105:195–205. doi: 10.1016/j.biomaterials.2016.07.003
70. Choi BD, Yu X, Castano AP, Bouffard AA, Schmidts A, Larson RC, et al. CAR-T cells secreting BiTEs circumvent antigen escape without detectable toxicity. *Nat Biotechnol* (2019) 37(9):1049–58. doi: 10.1038/s41587-019-0192-1
71. Abbott RC, Cross RS, Jenkins MR. Finding the Keys to the CAR: Identifying Novel Target Antigens for T Cell Redirection Immunotherapies. *Int J Mol Sci* (2020) 21(2):1–16. doi: 10.3390/ijms21020515
72. Li Ye, Hermanson DL, Moriarity BS, Kaufman DS. Human iPSC-Derived Natural Killer Cells Engineered with Chimeric Antigen Receptors Enhance Anti-tumor Activity. *Cell Stem Cell* (2018) 23(2):181–92.e5. doi: 10.1016/j.stem.2018.06.002

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Kang, Zheng, Hong, Chen, Li, Huang, Huang, Tang and Geng. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.