



The Emerging Jamboree of Transformative Therapies for Autoimmune Diseases

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Standard treatments for autoimmune and autoinflammatory disorders rely mainly on immunosuppression. These are predominantly symptomatic remedies that do not affect the root cause of the disease and are associated with multiple side effects. Immunotherapies are being developed during the last decades as more specific and safer alternatives to small molecules with broad immunosuppressive activity, but they still do not distinguish between disease-causing and protective cell targets and thus, they still have considerable risks of increasing susceptibility to infections and/or malignancy. Antigen-specific approaches inducing immune tolerance represent an emerging trend carrying the potential to be curative without inducing broad immunosuppression. These therapies are based on antigenic epitopes derived from the same proteins that are targeted by the autoreactive T and B cells, and which are administered to patients together with precise instructions to induce regulatory responses capable to restore homeostasis. They are not personalized medicines, and they do not need to be. They are precision therapies exquisitely targeting the disease-causing cells that drive pathology in defined patient populations. Immune tolerance approaches are truly transformative options for people suffering from autoimmune diseases.

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INTRODUCTION

Autoimmune diseases (AID) such as multiple sclerosis (MS), rheumatoid arthritis (RA), type 1 diabetes (T1D), and about 100 other conditions (<https://www.aarda.org/diseaselist/>), develop when certain body tissues are attacked by their own immune system. Such attack is orchestrated by autoantigen-specific T cells, generally restricted to defined major histocompatibility complex (MHC) molecules, that drive pathogenic effector T (T_{eff}) cell and humoral responses (1). AID affect 5–10% of the global population and their incidence is increasing, particularly in women, who are 2–10 times more likely to develop AID than men (2). AID are chronic debilitating disorders that often associate with autoinflammatory processes and are accompanied by many comorbidities resulting in significant shortening of life expectancy. A major factor driving chronicity of these diseases and poor quality of life of the affected patients is the limited availability of transformative therapies.

Traditional treatments for AID still rely predominantly on the use of broad cytotoxic or immune suppressive chemicals (3), e.g., azathioprine, calcineurin inhibitors, corticosteroids,

cyclophosphamide, leflunomide, methotrexate, mycophenolate, etc. These agents are effective in many situations and thus, they still remain a first treatment option and/or the standard of care. Recently, the Janus kinase (JAK) inhibitors have emerged as a new type of low molecular weight compounds that inhibit intracellular signal transduction of cytokine receptors. The cytokine selectivity of JAK inhibitors depends on the specificity of the different JAK family members, nonetheless, all of them inhibit the signaling of a large group of cytokines. JAK inhibitors are used for the treatment of multiple diseases (4). However, as long-term treatments, they may expose the patient to potential life-threatening opportunistic infections and risk of malignancies (4–6). Consequently, there is a clear medical need for effective, safe and specific treatments for AID. This review will discuss some of the existing and developing immunotherapy and precision medicine approaches aiming to ameliorate and/or cure AID.

THE REVOLUTION OF BIOLOGICS

About three decades ago, immunotherapy, mostly based on monoclonal antibodies (mAb), emerged as an alternative to limit the side effects associated with classical broad immune suppression (7, 8). Because of their better perceived benefit-risk profile compared to conventional therapies and their ability to control disease, the use of biologics has become widespread. This trend is expected to reinforce in the near future, due to the emergence of biosimilars and the continuous development of new biologic modalities.

Currently approved biologics target proinflammatory cytokines and mediators of inflammation and eliminate, block the functionality or the trafficking capabilities of specific leukocyte populations (Table 1). As these biologics are exquisitely selective for their targets, they are believed to have lower toxicity and better safety profiles than the classical small chemical molecules that have dominated the pharmaceutical industry in the past century. However, despite the fine specificity of these biologics for soluble molecules or cellular receptors, these treatments are not exempt of side effects. For example, as they suppress immunocompetent cell subsets, and block the action of soluble factors involved in the autoimmune attack, they also interfere with immune mechanisms that are essential in host resistance to a multitude of pathogens and thus, they increase the risk to acquire opportunistic infections, which present important complications and a potential cause of morbidity (5, 9, 10).

In the following sections we briefly review the major mode of action (MoA) of approved immunotherapeutic biologics as well as some products currently in development.

Inhibiting Cytokine Signaling

Tissue damage associated with AID can be mediated by a range of proinflammatory cytokines produced by cells of the innate and adaptive immune system (7, 11). Initially, a shared cytokine framework was considered to define highly conserved mechanisms of inflammation in human AID. It was thought that neutralization of one of multiple redundant nodes would suffice to disrupt the inflammatory process in a large variety of human inflammatory diseases. Clinical trials testing the

efficacy of various novel cytokine or cytokine receptor inhibitors revealed however a rather different reality and pointed toward the existence of a cytokine hierarchy, which could define a disease taxonomy (7, 11). Drugs aiming to block cytokine signaling are used in a large number of AID, mostly to dampen the deleterious inflammatory milieu (Table 1A).

Amongst the first biologics targeting pro inflammatory cytokines, which have deeply influenced the management of several AID are the TNF- α blockers (7, 12). Patients with diverse AID such as psoriasis (Pso), psoriatic arthritis (PsA), RA, Crohn's disease (CD), ulcerative colitis (UC), ankylosing spondylitis (AS), juvenile idiopathic arthritis (JIA), but not MS, have benefitted from TNF- α inhibition (7). On the flip side, TNF- α blockers carry the potential to reactivate opportunistic intracellular pathogens such as *Mycobacterium tuberculosis* from dormancy leading to resurgence of tuberculosis (13, 14). Surprisingly, inhibition of IL-1 has provided limited efficacy in rheumatic diseases, but it has shown great effects in autoinflammatory conditions mediated by inflammasome activation (7, 15, 16). Antibodies targeting the IL-6 receptor have been successful in RA, but they displayed limited or no effect in other chronic inflammatory conditions (7).

Additional biologics targeting other proinflammatory cytokines (IL-12, IL-17, IL-23) have progressively emerged and are becoming the standard of care in many inflammatory conditions or AID (17, 18). Initial mAb targeting this cytokine axis, like Ustekinumab, were directed to the p40 protein, which can associate with both, p35 to form the heterodimeric cytokine IL-12, or with p19 to generate IL-23 (19). Clinical trials with Ustekinumab supported its registration for Pso, at a time that this disease was still largely considered as a T_{H1} disease. Soon thereafter, it was realized that most AID could share or be exclusively of T_{H17} origin and that targeting specifically IL-17A or IL-23 could be a more selective treatment for many of these conditions (18, 20). At that time, the most advanced immunotherapeutic in clinical trials was the IL-17A specific mAb Secukinumab, which was originally aimed to be a treatment for RA, based on the initial association of IL-17 with osteoclastogenesis (21, 22). Thus, Secukinumab was tested and proved to be highly efficacious in Pso (23). Subsequent trials with IL-17 and IL-23 specific mAbs have highlighted the relevance of IL-17A blockade and provided support demonstrating a major role for the IL-23-IL-17 axis in the pathophysiology of this disease (18). In addition, and in contrast to anti-TNF- α therapy, the composite of clinical, animal and *in vitro* data accumulated with anti-IL-17A therapy indicates a low risk for mycobacterial infection (24–26).

The examples described above illustrate how blockade of key cytokine nodes regulating the differentiation and effector responses of pathogenic cell populations can be very effective ameliorating systemic and local inflammation. However, they are only optimally efficacious in certain dermatologic and rheumatologic conditions and some diseases are still looking for the ideal treatment. For example, in the case of systemic lupus erythematosus (SLE), only an anti-B-cell activating factor (-BAFF) mAb has shown a moderate efficacy in some patients (27), whereas other indications like MS have not yet clearly benefited from targeted cytokine blockade. Anifrolumab, a

TABLE 1 | Immunotherapy drugs approved for the treatment of autoimmune diseases^a.

	Target	Drug name	Biologic modality	First approval	Brand name	Approved indication	Company	
Inhibiting cytokine signaling	BAFF/Blys	Belimumab	mAb	2011	Benlysta	SLE	GSK	
	IL-1 β	Anakinra	Cytokine receptor antagonist	2001	Kineret	CAPS, RA, ScS, SD	Sobi	
		Rilonacept	Cytokine receptor antagonist	2008	Arcalyst	CAPS, FCAS, Gout, MWS	Regeneron	
		Canakinumab	mAb	2009	Ilaris	CAPS, FCAS, FMF, HDS, JIA, MWS, TNFR-APS	Novartis	
	IL-6	Siltuximab	mAb	2014	Sylvant	MCD	Janssen (J&J)	
	IL-6R	Tocilizumab	mAb	2009	Actemra	CRS, GCA, JIA, RA, SD, SS	Chugai/Genentech (Roche)	
	IL-12/23	Sarilumab	mAb	2017	Kevzara	RA	Sanofi	
		Ustekinumab	mAb	2009	Stelara	CD, pPso, PsA, Pso	Janssen (J&J)	
	IL-17A	Secukinumab	mAb	2015	Cosentyx	AS, pPso, Pso, PsA	Novartis	
		Ixekizumab	mAb	2016	Taltz	AS, pPso, PsA, Pso	Eli Lilly	
	IL-17RA	Brodalumab	mAb	2017	Siliq/Kyntheum	Pso	Ortho/Leo Pharma	
	IL-23	Guselkumab	mAb	2017	Tremfya	pPso, Pso,	Janssen (J&J)	
		Tildrakizumab	mAb	2018	Ilumya/Ilumetri	pPso, Pso,	Sun Pharma/Almirall	
	TNF- α	Risankizumab	mAb	2019	Skyrizi	pPso, Pso,	Abbvie	
		Infliximab biosimilars	Infliximab	mAb	1998	Remicade	AS, BD, BP, CD, CelD, CgD, GCA, IBD, JIA, KD, NSrc, Pcd, PF, pPso, PsA, Pso, PV, PyG, RA, SAPHO, SJS, SpA, SPD, Src, TEN, UC, Uve	Janssen (J&J)
			Infliximab biosimilars	mAb	2016	Inflectra		Pfizer
			Infliximab biosimilars	mAb	2017	Flixabi/Renflexis		Samsung/Biogen
			Infliximab biosimilars	mAb	2017	Ixifi		MSD
			Infliximab biosimilars	mAb	2018	Zessly		Pfizer
			Infliximab biosimilars	mAb	2018	Humira		Sandoz (Novartis)
			Infliximab biosimilars	mAb	2002	Humira		Abbvie
			Infliximab biosimilars	mAb	2014	Exemptia		Zydus Cadila
			Infliximab biosimilars	mAb	2016	Adfrar		Torrent
Infliximab biosimilars			mAb	2016	Amjevita/Solymbic		Pharmaceutic.	
Infliximab biosimilars			mAb	2016	Imraldi		Amgen	
Infliximab biosimilars	mAb	2017	Cyltezo		Biogen			
Certolizumab Fab' Ab	Certolizumab	Pegylated Fab' Ab	2017	Hadlima		Boehringer Ingelheim		
			2017	Hadlima		Samsung		
			2018	Mabura		Hetero		
			2018	Hyrimoz		Sandoz (Novartis)		
			2018	CinnaRA		CinnaGen		
			2018	Hulio		Fujifilm Kyowa Kirin		
			2019	Idacio		Fresenius Kabi		
			2008	Cimzia	AS, CD, pPsO, PsA, RA, SpA	UCB		
			2009	Simponi	AS, PsA, RA, SpA, UC	Janssen (J&J)		

(Continued)

TABLE 1 | Continued

	Target	Drug name	Biologic modality	First approval	Brand name	Approved indication	Company
		Etanercept	Soluble receptor antagonist TNFR2_Fc	1998	Embrel	AS, AzD, BD, BP, CgS, CwP, Hct, JIA, JRA, MAS, pPso, PsA, Pso, PV, RA, SAPHO, SD, SpA, Uve	Amgen, Pfizer, Takeda
		Etanercept biosimilars	Soluble receptor antagonist TNFR2_Fc	2015	Benepali		Samsung
				2016	Erelzi		Sandoz (Novartis)
				2016	Eticovo		Samsung
B							
Targeting leukocyte subsets	CD20	Rituximab	mAb	1997	Rithuxan/Mabthera	CLL*, DLBCL*, FL*, MCL*, NHL* BP, ES, FSG, GwP, ITP, MPA, PV, RA	Genentech (Roche)
		Rituximab biosimilars	mAb	2015	Zytus		Aryogen
				2017	Truxima/Blitima/ Ritemvia/ Rituzena		Celltrion Healthcare
				2018	Reditux		Dr Reddy's Labs
				2015	Maball		Hetero Healthcare
				2013	Mabtas		Intas Pharma
				2013	Novex		EleaPhonix
				2015	RituxiRel		Reliance
				2017	Rixathon/Riximyo		Sandoz (Novartis)
		Ocrelizumab	mAb	2017	Ocrevus	MS	Roche
		Ofatumumab	mAb	2009	Arzerra	CLL*	Novartis
	CD52	Alemtuzumab		2013	Lemtrada/Campath	CLL* MS, RA	Sanofi
C							
Preventing tissue homing	CD11a	Efalizumab	mAb	2003	Raptiva	pPso, Pso (withdrawn in 2009)	Genentech (Roche)
	Integrin α 4 chain	Natalizumab	mAb	2004	Tysabri	CD, MS	Merck Serono Biogen
	Integrin α 4 β 7 chain	Vedolizumab	mAb	2014	Entyvio	CD, UC	Takeda
D							
Intervening with immune checkpoints	CD2	Alefacept	Soluble receptor antagonist LFA3_Fc	2003	Amevive	pPso, Pso (discontinued in 2011)	Biogen/Astellas
	CD28	Abatacept	Soluble receptor antagonist CTLA-4_Fc	2011	Orencia	JIA, PsA, RA	BMS

AS, Ankylosing Spondylitis; AzD, Alzheimer's Disease; BD, Behcet's Disease; BP, Bullous Pemphigoid; CAPS, Cryopyrin-Associated Periodic Syndromes; CD, Crohn's Disease; CelD, Celiac disease; CgS, Cogan's Syndrome; CRS, Cytokine Release Syndrome; ES, Evan's Syndrome; FCAS, Familial Cold Autoinflammatory Syndrome; FMF, Familial Mediterranean Fever; FSG, Focal Segmental Glomerulosclerosis; GCA, Giant Cell Arteritis; Gout, Gout; GwP, Granulomatosis with Polyangiitis; Hct, Histiocytosis; HDS, Hyperimmunoglobulin D Syndrome; IBD, Inflammatory Bowel Disease; ITP, Idiopathic Thrombocytopenic Purpura; JIA, Juvenile Idiopathic Arthritis; JRA, Juvenile Rheumatoid Arthritis; KD, Kawasaki Disease; MAS, Macrophage Activation Syndrome; MCD, Multicentric Castleman's Disease; MPA, Microscopic polyangiitis; MWS, Muckle Wells Syndrome; NSrc, Neurosarcooidosis; Pcd, Polychondritis; PF, Pulmonary fibrosis; pPso, Plaque Psoriasis; PsA, Psoriatic Arthritis; Pso, Psoriasis; PV, Pemphigus vulgaris; PyG, Pyoderma Gangrenosum; RA, Rheumatoid Arthritis; RS, Reiter's Syndrome; SAPHO, SAPHO Syndrome; ScS, Schnitzler Syndrome; SD, Still's Disease; SjS, Sjögren Syndrome; SLE, Systemic lupus erythematosus; SpA, Spondyloarthritis; SPD, Subcorneal pustular dermatosis; Src, Sarcoidosis; SS, Systemic sclerosis; TEN, Toxic epidermal necrolysis; TNFR-APS, Tumor Necrosis Factor Receptor Associated Periodic Syndrome; UC, Ulcerative Colitis; Uve, Uveitis.

*Cancer indications: CLL, Chronic Lymphocytic Leukemia; DLBCL, Diffuse Large B-Cell Lymphoma; FL, Follicular Lymphoma; MCL, Mantle cell Lymphoma; NHL, Non-Hodgkin's Lymphoma.

°Sources: www.ema.europa.eu, www.drugbank.ca, www.fda.gov, www.gabionline.net/biosimilars and web sites from pharmaceutical companies.

human mAb to type I IFN receptor did not meet primary endpoints in an initial phase 3 trial with SLE patients. However, the drug is being reevaluated by astrazeneca.com in a subsequent study (NCT02446899) that uses different efficacy criteria.

Overall, cytokine antagonism can result in dramatic and sometimes sustained clinical responses, particularly if used at the early stages of the disease. However, such approaches may not constitute a definite cure, as they usually do not induce robust and prolonged immune regulatory mechanisms. The limitations of these therapies could be explained by the known redundancy of the cytokine pathways and/or by the differential hierarchy exerted by these cytokines in particular conditions (7, 11). Today, it is generally accepted that a better understanding of the AID endotypes will be required to select the best medication for each single patient (7, 11).

Targeting Leukocyte Subsets

A category of biologics frequently used for the treatment of AID includes mAbs targeting specific leukocyte subsets, aiming to eliminate or inactivate these cell populations (Table 1B). This approach developed following the initial success with therapies designed for the treatment of lymphoproliferative disorders. Among the best examples of the class are the mAbs directed to the B cell receptor CD20 (i.e., Rituximab, Ocrelizumab, and Ofatumumab) that induce B cell depletion (28). Rituximab was originally developed to eliminate B-cell non-Hodgkin lymphomas (29). Subsequently, it was found to be efficacious in a series of AID through depleting autoantigen-specific B cells that i) could develop into antibody secreting cells or ii) could exert immune accessory functions such as antigen presentation or cytokine release (30, 31). Despite anti-CD20 mAb therapy causing severe B cell immunodeficiency, this approach has been validated and is currently used for the treatment of several AID such as RA, pemphigus vulgaris (PV) and MS (32).

Another registered mAb in this category is Alemtuzumab, which is an antibody directed to CD52, a receptor broadly expressed on mature leukocytes (33). Alemtuzumab was originally registered for the treatment of B-cell chronic lymphocytic leukemia under the trade name of Campath and, subsequently, relaunched as Lemtrada to treat severe cases of relapsing remitting MS (34, 35). Because of the broad expression of CD52, Alemtuzumab drives cell depletion of both B and T cells, which implies significant safety concerns after extended use.

Although not yet registered, Teplizumab is an investigational product that has been in development for many years. Teplizumab consists of a humanized, Fc receptor silenced, anti-CD3 mAb, which was derived from the mouse Orthoclone, OKT3 mAb (36). The antibody was initially intended to prevent transplant rejection, however the mouse origin and the strong ability to crosslink Fc receptors of the original product resulted in strong side effects (37). Rather than acting as a strong depleting antibody, it seems that one important MoA of Teplizumab involves the inactivation of T cells, driving them into exhausted phenotypes (38). Recently, Teplizumab has shown to delay the onset of T1D for 2 years in patients at risk of developing disease (stage 2) (39). Maintenance in

these patients, mostly pediatrics, in stages that precede overt disease is considered a major improvement in the field, but it is not curative. Because the patients were only exposed to a single (14 days) treatment course, there are high expectations that repeated treatment could provide stage 2 T1D patients with a longer delay in transitioning into the overt phase of the disease (stage 3).

All drugs mentioned above, have shown efficacy in some patient populations. However, they do not distinguish pathogenic vs. beneficial cells and thus, their degree of efficacy correlates with their immunosuppressant potential.

Preventing Tissue Homing

A way to limit general immunosuppression has been to focus the therapeutic intervention at the mechanisms controlling leukocyte migration, avoiding the recruitment of the T_{eff} cells and preventing their activation at inflammatory sites (Table 1C). One of the first biologics using this MoA was Efalizumab, which was approved in 2003, under the trade name of Raptiva, to treat Pso (40, 41). Efalizumab, binds to integrin α_L (CD11a), which together with CD18 constitutes lymphocyte function-associated antigen 1 (LFA-1) (42). Efalizumab blocks the binding of LFA-1 to intercellular adhesion molecule-1 (ICAM-1), thereby (i) preventing the migration of T cells into inflamed tissue and (ii) inhibiting T cell activation by antigen presenting cells. Raptiva was discontinued in 2009 based on its association with the development of progressive multifocal leukoencephalopathy (PML), a rare and usually fatal disease caused by reactivation of the human polyomavirus 2 (John Cunningham or JC virus) in the central nervous system (43, 44).

Tissue migration has also been targeted using mAbs that block the α_4 (CD49d) and β_7 integrin families. α_4 integrins play a critical role in the adhesive interactions of lymphocytes with endothelial cells required for cell extravasation and migration to sites of inflammation (42). Integrin α_4 form heterodimers with integrin β_1 (CD29) or β_7 to generate very late antigen-4 (VLA-4) or lymphocyte Peyer patch adhesion molecule (LPAM), respectively (42). VLA-4 interacts with endothelial vascular cell adhesion molecule 1 (VCAM-1/CD106) whereas LPAM has a preference for the endothelial receptor mucosal vascular addressin cell adhesion molecule 1 (MAdCAM-1) (42). Integrin β_7 can also dimerize with integrin α_E (CD103) to form the heterodimeric receptor $\alpha_E\beta_7$, which interacts specifically with E-cadherin on epithelial cells (42). There are currently two marketed mAbs directed to α_4 integrins: Natalizumab which binds specifically to the α_4 integrin and Vedolizumab that interacts with an epitope displayed by the heterodimer of $\alpha_4\beta_7$ (45). Natalizumab was first approved for the treatment of MS (45) and subsequently, for the treatment of moderate to severe CD (46, 47). However, because prolonged therapy with Natalizumab may lead to JC virus reactivation (48–51), blocking more specifically the trafficking of leukocytes to the gut through preventing interactions with MAdCAM-1 might be a safer treatment to intervene in ulcerative colitis (UC) and CD (52). The aspiration to develop safer biotherapeutics for inflammatory bowel disease (IBD) has triggered the advancement of further additional antibodies

specific for (i) $\alpha_4\beta_7$ epitopes (i.e., Abrilumab by Amgen¹) (53), (ii) integrin β_7 (Etrolizumab, by <https://www.gene.com> / <https://www.roche.com>) (54) that will prevent interaction of $\alpha_4\beta_7$ and $\alpha_E\beta_7$ with MAdCAM-1 and E-cadherin, respectively or (iii) endothelial MAdCAM-1 (Ontamalimab by <https://www.shire.com> / <https://www.takeda.com>) (55) that should further increase tissue selectivity by selective blocking gut-specific integrin interaction (**Figure 1**).

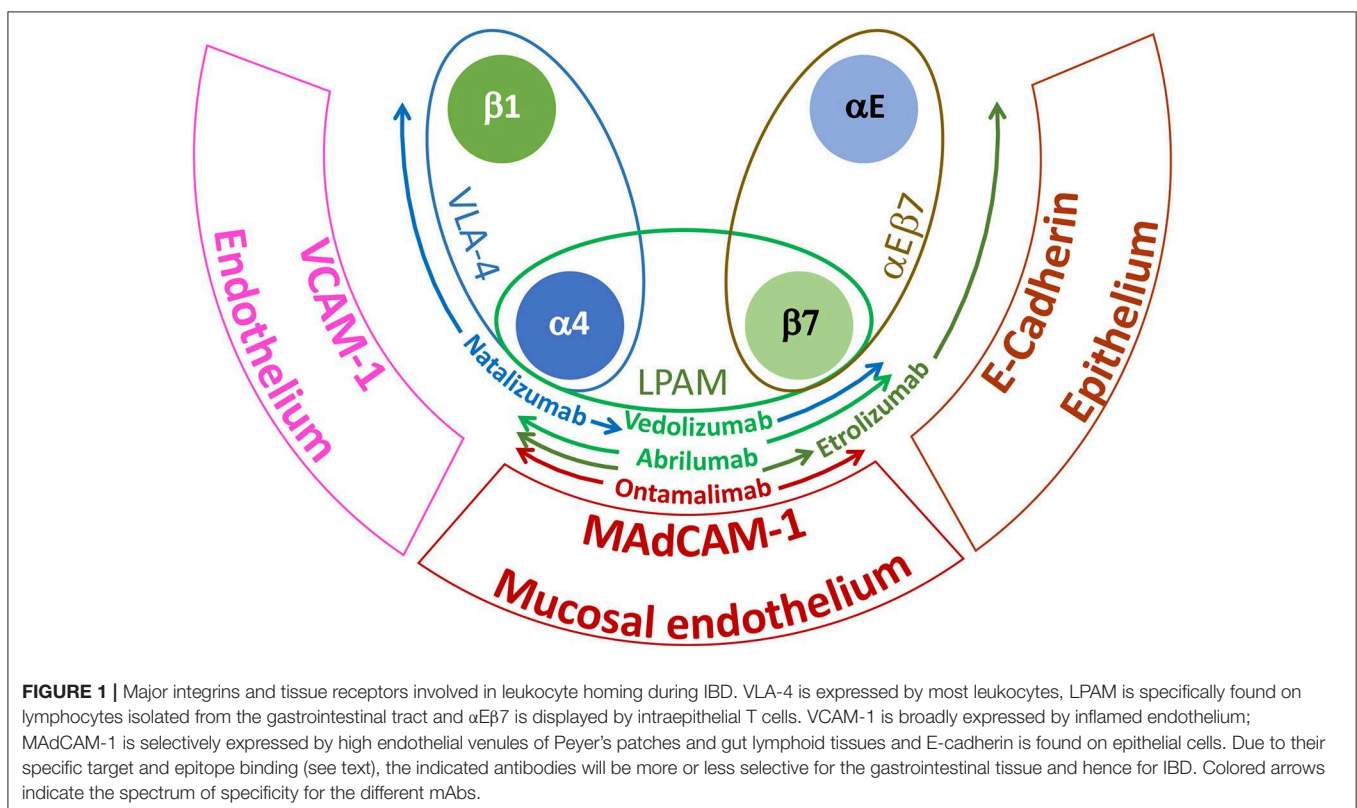
In addition to integrins, mAbs directed to chemokines or chemokine receptors are also being considered. Many of these ligand receptor pairs have been associated with inflammatory and autoimmune processes, however their redundancy proved a real challenge for therapeutic intervention using small molecules (56). Current immunotherapy approaches aim to antagonize migratory receptors preferentially expressed by T_{H1} and T_{H17} cell subpopulations, such as CXCR3 (57–60) and CCR6 (60, 61). Most of the mAbs, currently in development, do not only block migration but mediate the killing of the target cells by antibody-dependent cellular cytotoxicity (ADCC) and/or complement-dependent cytotoxicity (CDC).

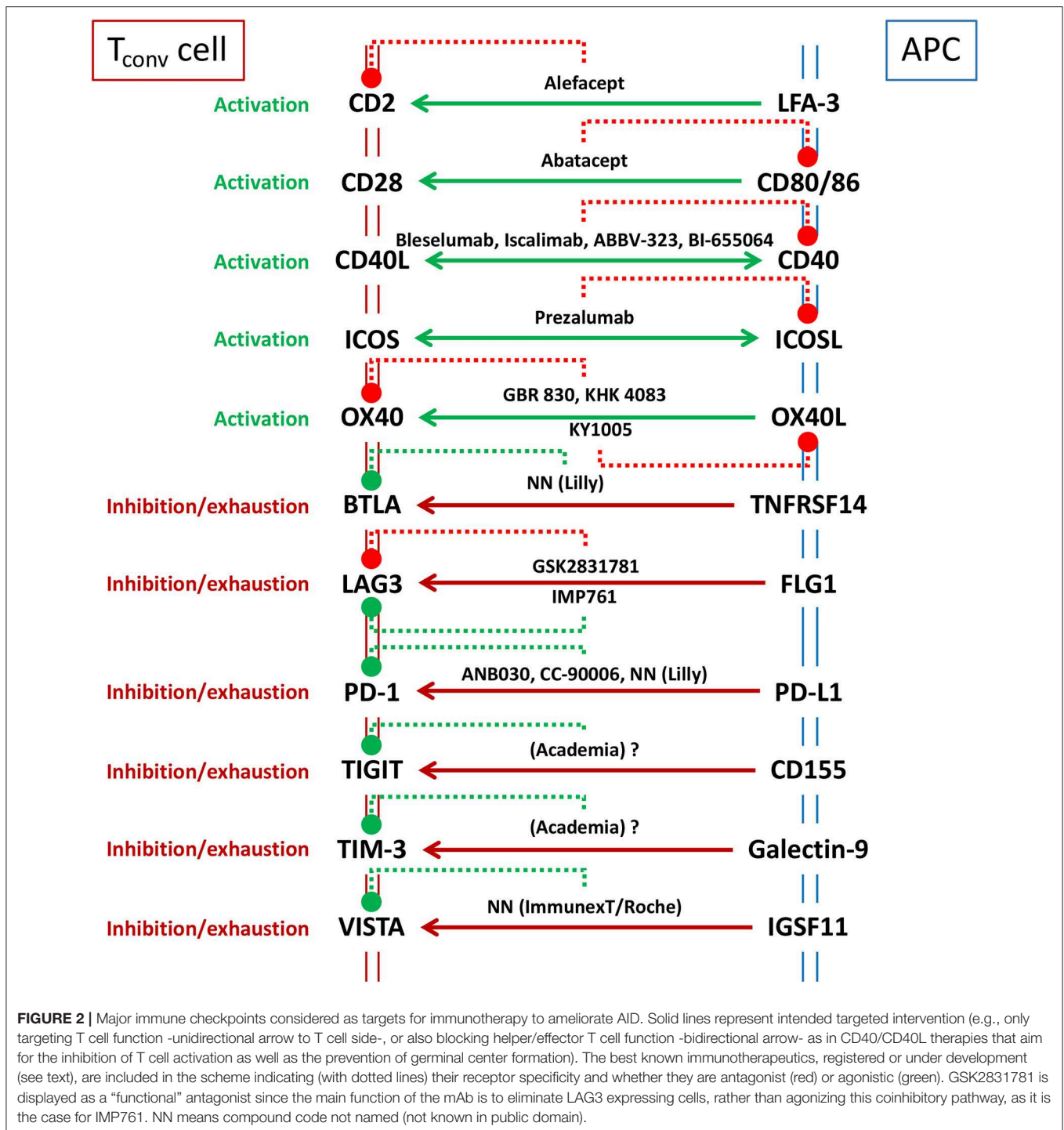
In general, these approaches have a very sound rationale. On the other hand, they suffer from the limitation that not all autoimmunity-causing T cells can be mapped using a particular integrin or chemokine receptor expression and therefore, since they still retain immunosuppressive potential, the utility of these biologics is limited to some special situations.

¹<https://www.amgen.com>

Intervening With Immune Checkpoints

Immune checkpoints are a diverse class of receptor pairs, predominantly expressed on the surface of T cells and interacting partners, that regulate T cell activation (62, 63). Engagement of these receptors at the T cell side could be costimulatory or coinhibitory. These receptors do not act alone, they rather amplify or diminish the signals initiated by the recognition of cognate peptide-major histocompatibility complex (p-MHC) upon engagement of specific antigen-receptors on T cells in the context of a large array of cell-cell interaction bridges (63). Thus, the outcome of T cell activation does not depend on a single receptor pair but on the overall balance of a multitude of costimulatory and coinhibitory signals that occurs simultaneously during the engagement of T cells with APC or target cells. For example, costimulatory receptor bridging may dominate such balance during early phases of infection, but the equilibrium will be tilted toward coinhibitory signaling once the immune response becomes effective and pathogen loads are diminished. Such switch is designed to avoid exacerbated reactions and unnecessary host tissue damage. Unfortunately, many tumors do exploit these coinhibitory immune checkpoint pathways to elude immune surveillance. The awareness of this mechanism has sparked the use of antagonist mAbs (particularly involving cytotoxic T-lymphocyte-associated protein 4 (CTLA-4/CD152) and programmed cell death protein 1 (PD-1/CD279) or its ligand programmed death-ligand 1 (PD-L1/CD274) that prevents inhibitory receptor-counterreceptor bridging without triggering coinhibitory signals and thus, releases T cell inhibition and enhances anti-tumor





immunity (62, 64). On the other hand, immunotherapeutics based on the antagonism of costimulatory immune checkpoints or on the agonism of coinhibitory counterparts would have therapeutic potential for the treatment of AID. A graphic summary of the biologics directed to immune checkpoints, already registered or under clinical development is presented in **Figure 2**.

Antagonizing Costimulatory Checkpoints

Approved examples for the antagonist of costimulation include fusion proteins consisting of extracellular domains of lymphocyte function-associated antigen 3 (LFA-3/CD58) and CTLA-4, linked to human immunoglobulin Fc domains, and are represented by Alefacept (trade name Amevive) (65) and Abatacept (trade name Orencia) (66), respectively (**Table 1D**).

CD2 on T cell surfaces interacts with LFA-3/CD58 during antigen presentation and thus, blockade of this costimulatory pathway using the soluble CD2 ligand, Alefacept, prevents efficient T cell activation (67). In addition, because the Fc domain of Alefacept is a wild type human IgG1, the molecule also mediates killing of memory T cells (68). Alefacept was approved for Pso (69) in the USA and a few other countries (but not in the EU market) and it was discontinued voluntarily by the manufacturer shortly after launch. Recently, Alefacept has been tested in a clinical trial for T1D. The study involved a short treatment period and showed signs of efficacy without reporting major adverse effects (70).

CD28 is a classical costimulatory molecule expressed on the surface of all naïve and most memory T cells (71). Upon T cell receptor engagement, CD28 binds to its APC counter-receptors CD80 (B7-1) and CD86 (B7-2) providing costimulatory signals required for efficient T cell activation (72). CTLA-4 is normally induced on the surface of T cells following activation. Once it is expressed, CTLA-4 also binds to CD80 and CD86 with much higher affinity than CD28, removing costimulation and providing coinhibitory signals that terminate T cell activation (73). Soluble CTLA-4_Ig (Abatacept) is a fusion molecule that uses an engineered Fc-silent domain, and thus, it inhibits CD28-mediated T cell activation without triggering cytotoxicity to target cells (74). Abatacept is an approved drug for RA (75). A drug with similar MoA, Belatacept (trade name Nulojix) has been developed specifically for the prevention of transplant rejection (76). Belatacept differs from Abatacept in two amino-acid substitutions (L104E and A29Y) (77). These modifications result in a more powerful blockade of T cell activation, especially by blocking the CD86–CD28 interaction, which in general is considered to precede CD80–CD28 due to the constitutive high abundance of CD86 on APC surfaces (78).

Interestingly, agonists of CD28 were also considered in the past for the treatment of AID, based on the observed potential to expand regulatory T (T_{reg}) cells (79). However, TGN1412, a CD28 super-agonist mAb, induced a polyclonal T cell activation with a dramatic cytokine release syndrome during the first clinical trial (80, 81). TGN1412 essentially promoted T cell stimulation without simultaneous antigen-specific T cell receptor (TcR) engagement (80, 82). This unfortunate event illustrates the fact that the balance between T_{reg} and T_{eff} cell responses in laboratory animals and humans and their response to super-agonists can be significantly different (80, 83).

The CD40/CD40L costimulatory pathways have been considered since long as a potential site of intervention to prevent transplant rejection and to treat AID (84). Initial efforts focused on antagonizing CD40L (CD154), since this receptor is induced on the surface of activated T cells, which was considered an advantage over targeting the counterreceptor CD40 that is broadly expressed on APC and other cell types, i.e., endothelial cells (85). Unfortunately, clinical trials with two independent drug candidates and preclinical results with additional compounds revealed thromboembolic events, related to the expression of CD40L and Fc γ R2 on platelets (86), and resulted in the discontinuation of these projects (87, 88). A new wave of modified antibodies and CD40L binding proteins that do not induce platelet aggregation is currently under

development (84). Alternatively, there is a series of mAbs that block CD40 without causing depletion or activation of the target cells, including Bleselumab (<https://www.kyowakirin.com>), Iscalimab (<https://www.novartis.com/>), BI-655064 (<https://www.boehringer-ingenelheim.com> in collaboration with <https://www.abbvie.com>), ABBV-323 (<https://www.abbvie.com>), ch5D12, and FFP104 (84). Particularly, Iscalimab (CFZ533) is an investigational compound that has shown to prolong the durability of transplanted kidneys and improve long-term outcomes for kidney transplant (89). Iscalimab is a fully human mAb directed to CD40. It blocks CD40-CD40L (CD154) pathway interaction without causing depletion of APC or other CD40-bearing cells. Iscalimab has shown to be effective in patients with primary Sjögren syndrome (SjS) (90) and in a subgroup of Graves' disease (GD) patients (91) and it is being explored in several other autoimmune conditions (92).

Inducible T-cell costimulator (ICOS/CD278) is another costimulatory immune checkpoint expressed on T cell surfaces, which interacts with ICOS-ligand (ICOS-L/CD275) on APC. ICOS is induced in naïve T cells upon encounter with specific antigen. Afterwards, it is expressed, and upregulated following activation, on broad subsets of antigen-experienced T cells including T_{h1} , T_{h2} , T_{h17} , and T_{reg} cells (93). On the other hand, ICOS has been mostly implicated with follicular helper T (T_{fh}) cell responses, particularly supporting survival and immunoglobulin switch of B cells in germinal centers (94–96). These characteristics drove the development and clinical investigation of Prezalumab (formerly AMG557). Prezalumab is an IgG2 mAb directed to ICOS-L that prevents productive T_{fh} -B cell interactions. Prezalumab was originally tested in Pso and cutaneous lupus erythematosus by Amgen¹. These projects were discontinued, but clinical testing resumed in collaboration with AstraZeneca² for SjS, although recently AstraZeneca also halted development following poor results in a phase 2 study. AMG577 has now been re-engineered into a bispecific mAb (MEDI0700, previously AMG570) that simultaneously targets BAFF and ICOS-L (97).

Additional costimulatory immune checkpoints like OX40 (TNFRSF4/CD134), the T cell counterpart of OX40L (TNFSF4/CD252) induced during late T cell activation phases has been considered by <https://www.glenmarkpharma.com/> (anti-OX40, GBR 830), <https://www.kyowakirin.com/> (anti-OX40, KHK 4083) and <https://www.kymab.com/> (anti-OX40L, KY1005) for the treatment of ulcerative colitis, although the emphasis of these products is to target T_{h2} mediated allergic disease.

Agonizing Coinhibitory Checkpoints

The alternative concept, namely agonizing immune inhibitory checkpoints, to treat AID has also been proposed in the literature in the past years (63, 98–102) and the efficacy of several coinhibitory agonists has been demonstrated preclinically *in vitro* using human peripheral blood mononuclear cells (PBMC) and *in vivo* using rodent models of AID (103–106). Among others, B- and T-lymphocyte attenuator (BTLA/CD272) (107), lymphocyte-activation gene 3 (LAG-3/CD223) (107, 108), T cell

²<https://www.astrazeneca.com/>

immunoglobulin and mucin-domain containing-3 (TIM-3) (107, 108), T cell immunoreceptor with Ig and ITIM domains (TIGIT) (107–114), PD1/PD-L1 (63, 101) and V-domain Ig suppressor of T cell activation (VISTA) (107, 115, 116) are emerging as very attractive target candidates for agonistic immune checkpoint intervention in AID.

BTLA has been described to inhibit T cell responses following engagement of the TNF receptor superfamily member 14 (TNFRSF14) which is also known as the herpes virus entry mediator (HVEM). A low expression of HVEM has been clinically associated to increased SLE disease activity and an elevated IFN gene signature (117). These results suggest that inefficient BTLA engagement could favor SLE and accordingly, <https://www.lilly.com/> is developing an agonistic antibody to BTLA.

LAG3 is a receptor primarily found on activated T and NK cells (118) which originally was reported to interact with MHC-II molecules (119) and, more recently, has been described as a functional receptor for fibrinogen-like protein 1 (FGL1) (120). LAG3 has been implicated as a marker of exhaustion on CD8⁺ T cells (121) and as a negative regulator of T cell activation and effector function (119, 122), in addition of contributing to suppression when expressed on T_{reg} cell surfaces (123). LAG3 is being targeted in the oncology field, with compounds that aim to remove the unresponsive state of tumor specific T cells. However, it is also viewed as a candidate target for immunotherapy to treat AID. <https://www.immutep.com> is a biotechnology company exclusively focusing on therapeutics intervening with the LAG3 pathway. In addition of having several compounds in development for cancer, this team is working on two mAbs for AID. The most advanced compound, IMP731, is based on a T cell-depleting murine antibody, chimeric with human IgG1, that has been licensed to <https://www.gsk.com> (GSK2831781) and is currently in phase 2 for UC. In addition, <https://www.immutep.com> is developing by its own a non-depleting agonistic anti-LAG3, humanized IgG4 mAb (IMP761) for AID.

PD-1/PD-L1 is a prototype target for cancer. Antagonist mAbs to PD-1, like Pembrolizumab, Nivolumab, and Cemiplimab or to PD-L1, like Atezolizumab, Avelumab, or Durvalumab are very successful drugs treating a variety of tumors and thus, the complementary option of developing agonists counterparts for the treatment of AID has been considered by a few companies. Among those, <https://www.anaptysbio.com/> has internally ANB030, and in partnership CC-90006 (<https://www.celgene.com/> now part of <https://www.bms.com/>), agonistic anti-PD-1 projects to intervene with inflammatory diseases and Pso, respectively. <https://www.lilly.com/> is also reporting the development of an agonist mAb to PD-1 for the treatment of AID.

TIGIT is a receptor absent on naïve T cells, which is induced following activation and then, remains constitutively present on subsets of memory T cells (109). TIGIT binds predominantly to the poliovirus receptor (PVR/CD155) on APCs (124). TIGIT shares this binding pattern with an additional coinhibitory receptor, T cell activation, increased late expression (TACTILE/CD96) (125), and with the costimulatory receptor DNAX accessory molecule-1 (DNAM-1/CD226) (126). CD226 is broadly expressed among all T cell subsets, although at lower

intensities in the naïve populations, whereas CD96 displays also broad but low-density expression on all T cells. CD155 on the other hand is present on all APC surfaces, but it is also found in endothelial and epithelial surfaces, particularly under inflammatory conditions. The highest affinities of these receptor interactions are found between TIGIT with CD155, followed by those between CD226 with CD155, and thereafter, but weakly, by those of CD96 with CD155 (127). Furthermore, there is another inhibitory molecule on T cells, namely, poliovirus receptor related immunoglobulin domain containing (PVRIG/CD112R), which interacts with the APC counterreceptor poliovirus receptor-related 2 (PVRR2/CD112) (128), that is also a weak binder for TIGIT and perhaps also for CD226. Overall, based on receptor abundance and affinity interactions the major players on this immune checkpoint cluster are CD155 on the APC and CD226 and TIGIT on the T cells (114). Such triad is reminiscent of the CD80/86 interactions with CD28 and CTLA-4 that tightly controls the activation of naïve (T_n) and central memory (T_{cm}) T cells. Thus, intervention with CD226/TIGIT immune checkpoints might be particularly efficient to prevent the effector functions of the CD28⁻ memory T cells (129) that are abundant in AID (130). However, there is little public information on the development of therapeutic anti-TIGIT mAbs for indications outside oncology.

TIM-3 was identified screening hybridomas from rats immunized with mouse T cells against established mouse T_{h1} and T_{h2} cell clones, followed by an expression cloning approach (131). The exercise identified TIM-3 as a molecule selectively expressed by mouse IFN- γ producing T_{h1} and T_{c1} cells but not by the T_{h2} counterparts. This pattern has been confirmed in human and extended to T_{h17} cells, that also showed TIM-3 expression although at lower abundance (132). A series of experiments in mouse models, including the use of its ligand Galectin-9 (133) demonstrated that engagement of TIM-3 ameliorate disease by elimination of T_{h1} cells (132). In addition to Galectin-9, several ligands capable of interacting with TIM-3 have been described (100) but Galectin-9 is probably the most studied due to its broad expression in the tumor environment where it could promote inflammation or facilitate tumor escape (134). The abundance of TIM-3 has been found to be particularly low on T cells isolated from cerebrospinal fluid of MS patients (135, 136) and from peripheral blood of RA (137) or Pso (138) subjects. Altogether, this would indicate that agonistic anti-TIM-3 mAbs might be effective eliminating disease-causing T cells. However, there are no reports in the public domain indicating drug development projects using this approach.

VISTA (also known as B7-H5) is a member of the B7 family of immunoregulatory molecules present on the surfaces of hematopoietic cells. It is expressed on myeloid cells as well as on NK cells and naïve and memory T cells but absent on B cells (139). It was originally cloned in the context of gene expression studies that compared resting vs. activated mouse CD25⁺ T_{reg} cells (115). The molecule was described to function as both, a ligand and a receptor and attributed to have immunosuppressive function based on *in vivo* studies with gene-deficient mice (139). VISTA expression was also abundant in tumor microenvironments where it facilitated tumor evasion (140) and thus antagonist anti-VISTA mAbs were considered

for several malignancies. Recently, immunoglobulin superfamily member 11 (IGSF11; also known as V-set and immunoglobulin domain containing 3, VSIG3) was described to be a ligand for VISTA (141) and reported to inhibit T cell activation (142). Furthermore, antagonistic and agonistic antibodies to mouse VISTA, have been shown to enhance (143) or prevent (144) lupus, respectively, in experimental rodent models. These data support the development of agonistic anti-VISTA therapeutics for the treatment of AID. In line with these observations, <http://immunext.com> has an agonist anti-VISTA mAb project partnered with <https://www.roche.com> to intervene in AID.

Whereas, some immunotherapeutics blocking costimulatory immune checkpoints have shown efficacy in certain autoimmune conditions, the complementary intervention aiming to agonize the coinhibitory receptors seems to be lagging behind. One possible explanation could be the special challenges associated to these therapeutics that often require non-cytotoxic crosslinking of the target and necessitate to adapt their MoA depending on whether they interact with T_{eff} or T_{reg} cells.

THE HOPE OF CELL THERAPIES

The capability to counterbalance antigen-specific immune activation is critical for effective and precise immune function without inducing collateral tissue damage (145–147). T_{reg} cells expressing transcription factor forkhead box p3 (FOXP3) are a small but essential subset of lymphoid cells which are able to contract activated immune responses and maintain immune system homeostasis, thereby preventing inflammation and AID. Indeed, patients with systemic or organ-specific AID have compromised T_{reg} cell numbers and/or function (145, 146). Hence, a promising way to restrain autoimmune responses, is to expand the pools of patient's T_{reg} cells (145, 146). However, it is not always clear which subset of T_{reg} cells can or should be expanded, as an increasing diversity of immunosuppressive lymphocytes have been described (145, 146). T_{reg} cells are generally divided into two major subsets: thymus-derived cells, that normally react to self-antigens, and peripherally derived cells, that preferentially respond to “environmental” antigens (145). Peripheral $CD4^{+}$ T_{reg} cells are classically distinguished on the basis of their constitutive CD25 (IL-2 receptor α chain) expression which parallels FOXP3 expression and immunosuppressive function (145). Another type of regulatory T cell subset, called regulatory type 1 (T_{r1}) cell, does not constitutively show high expression of CD25 and FOXP3, but is characterized by surface CD49b and LAG3 and by their ability to produce abundant IL-10 and TGF- β 1 with little or no IFN- γ (146, 148–150). These cells are responsible for dampening host-reactive T cells responses after hematopoietic stem cell transplantation (151–153) and for the suppression of effector autoimmune T cell responses (154–156). Furthermore, immunosuppressive, TGF- β 1-producing, $CD4^{+}$ type 3 helper T (T_{h3}) cells as well as $CD8^{+}$ T_{reg} cells and IL-10 producing regulatory B (B_{reg}) cells have also been documented (146).

T_{reg} cells have been considered as living drugs for the treatment of autoimmune and inflammatory diseases (146) as

they have been used to reconstitute patients with AID after *ex vivo* expansion. To this end, T_{reg} cells need to show consistent immunosuppression potential, prolonged *in vivo* survival and stability of the phenotype (146). Nevertheless, T_{reg} cells have shown phenotypic plasticity, potentially related to the diverging contextual conditions they encounter in different tissues (e.g., cytokine milieu, availability of antigen, TcR affinity etc.). The level of immunoregulation likely will need to be customized for each AID and inflammatory disorder (146), and to be balanced against unwanted, non-specific immunosuppression, that would increase the risk for opportunistic infections or malignancies. Because the $CD25^{+}$ $FOXP3^{+}$ T_{reg} cells are very amenable to *in vitro* expansion, they have been used in several clinical trials, including treatment of T1D patients (146, 157–159). In some cases, the infused T_{reg} cells remained stable and detectable for 1 year. In other cases, the effects were rather transient and did not provide a therapeutic benefit (146). One possible explanation for this limited success could be related to the fact that the large numbers of transferred cells were not selected on the basis of any antigen-specificity and therefore, they probably harbored only very small amount of T_{reg} cells specific for the antigens involved in the autoimmune process.

$FOXP3^{+}$ T_{reg} cells have constitutive expression of CD25 in marked contrast to conventional T cells that only express this protein following activation. This difference has prompted the search for therapeutic products based on modified and/or low dose IL-2. The rationale behind this is that a low dose of IL-2, especially when the cytokine is modified to increase its binding to CD25 and diminish that to CD122 (β chain of the IL-2 receptor) will engage IL-2 receptors (expressing the $\alpha/\beta/\gamma$ IL-2 receptor chains) on T_{reg} cells and promote their expansion without activating T and NK cells (expressing β/γ IL-2 receptor chains) (160). Indeed, low-dose IL-2 has shown a satisfactory safety profile in human and is currently being tested in several AID (160). To increase the circulating half-life of these products, the muteins are generally grafted into an irrelevant mAb or fused to an immunoglobulin Fc portion (160). Alternatively, some investigators have proposed the use of complexes of IL-2/anti-IL-2 mAb that also increase the half-life of circulating IL-2 and release the cytokine in the proximity of high affinity $\alpha/\beta/\gamma$ IL-2 receptors (161).

It is expected that the *in vivo* expanded polyclonal T_{reg} cell populations will contain a sizable amount of autoantigen-specific T_{reg} cells, as consequence of the ongoing chronic response, but these therapeutic principles are not designed to favor any antigen specificity and thus, the same risks of inducing immunosuppression that were associated to the polyclonal T_{reg} cell transfer may also apply for this situation. Thus, finding approaches to promote an optimal balance between disease- and pathogen-specific T_{reg} cell activation continues to be a critical challenge for the clinical development of this cytokine therapy (160). A potential solution to avoid broad immunosuppression could be a localized delivery of IL-2. <https://pandiontx.com> is currently using such strategy by fusing an IL-2 mutein to the Fc chains of a mAbs directed to MadCAM. This engineered biologic is expected to focus IL-2 on endothelial surfaces of

the gastrointestinal tract and hence, expand locally T_{reg} cells and provide a benefit for patients with IBD. Furthermore, a recent investigation has shown that chemical inhibition of the cyclin-dependent kinase 8 (CDK8) and CDK19, or knockdown/knockout of the *CDK8* or *CDK19* gene, can induce FOXP3⁺ T_{reg} cells in mice (162). The conversion which resulted from the release of FOXP3 repression, was TGF- β 1-independent, not affected by inflammatory cytokines and provides a new opportunity to expand T_{reg} cells *in vivo* (162).

Polyclonal and antigen specific T_{r1} cells have been also used in clinical trials (163–166). However, as the CD25⁺ counterparts, they have shown limited survival capacity *in vivo* (164). This limitation has prompted alternative designs based on the generation of conventional antigen-specific CD4⁺ T cells that are converted into T_{r1} cells by lentiviral transduction of *IL10*, and an additional gene, which could be included in a bidirectional vector to provide a convenient marker for purification (167, 168). Alternatively, <https://www.sangamo.com> is trying to solve the issue of antigen selectivity by developing regulatory-chimeric antigen receptor T (CAR- T_{reg}) cells (169–172). Their original target indication is transplant rejection and thus, they aim to generate CAR-T cells using donor-human leukocyte antigens (-HLA) binding elements (i.e., variable fragments of anti-HLA mAbs) engineered in a chimeric molecule that express intracellular domains of CD28 and CD3 molecules, which are transfected into highly purified T_{reg} cells isolated from the host patient (173–176).

THE PROMISE OF IMMUNE TOLERANCE

Despite the recent advances in the immunotherapy field, there is still a significant correlation between the increased efficacy of these novel medicines with the undesired risk of infections. This relationship is understandable, since none of the existing approaches is exclusively directed to the specific triggers of the disease. At most, immunotherapy principles target cell subsets or soluble factors that are predominantly involved in the autoimmune attack, but nevertheless are also needed to fight pathogen invasion. Thus, the necessity to develop truly transformative therapies, that specifically control and provide curative potential for AID, without broad immunosuppression remains actual, and is prompting a resurgence of the long-sought immune tolerance field.

Induction of immune tolerance to self-antigens occurs naturally in the thymus and bone marrow during T and B cell ontogeny (177, 178). These processes are not completely efficacious, since they permit the development and subsequent export to periphery of lymphocytes with low avidity for autoantigens (179). Escaping autoreactive clonotypes are normally silent in the body (180, 181), but they can be awakened when their thresholds of activation are reduced; i.e., due to cross-reactivity with pathogen or commensal organisms in the context of immunological danger that may be favored by the host expression of AID-predisposing genetic variants (182). Specific intervention in autoantigen-specific immune function, to enhance or restore the mechanisms of immune tolerance,

represents an excellent opportunity to fulfill the huge medical need existing in AID therapy (183). Such intervention could be, in principle, attempted targeting either the central or peripheral sites where immune tolerance is orchestrated.

Central Tolerance

Induction of central tolerance, i.e., by resetting the immune system, is a possible option, although associated with very harsh procedures (184, 185). This notion developed following reports of patients that underwent hematopoietic stem cell transplantation (HSCT) to treat hematological malignancies and showed remissions of coincident AID (186, 187). HSCT following extensive immune ablation restores the immune system and provides long-term remission rates in some AID patients (188). However, the risk-benefit ratio of these therapies has to be carefully and individually evaluated, since the procedures expose the patients to temporary iatrogenic severe immune deficiency and are, overall, associated with unacceptable mortality (188, 189).

Peripheral Tolerance

Immune tolerance approaches directed to the peripheral immune system seem to be more amenable from the safety and practical point of view. However, these strategies are still in preclinical phases of development or have not yet demonstrated conclusive long-term efficacy in the clinic. The landscape of peripheral immune tolerance approaches has been recently reviewed (190), including some points of view on how to maximize their development and smooth progression to the clinic (183). Thus, this section will only comment on the different mechanisms of action of a few representative therapies without entering into the details of the specific products.

Therapeutic principles to induce peripheral immune tolerance have been historically grouped into (i) induction of clonal deletion and/or anergy or (ii) promotion of active regulatory mechanisms.

Deletional approaches might work well in situations where there is a limited immunogenic repertoire, as it the case of immune responses to immunotherapy products (191). However, these will face major challenges in the context of AID, which are driven by a complex autoimmune repertoire that is almost impossible to completely determine. The rationale for deletion approaches was triggered by a concept of hierarchy in the development of autoimmune responses, which postulates the existence of major immunodominant epitopes that initiate the immune response and precede broad epitope spreading (192–195). Eliminating or inactivating the T cell clones directed to these primary disease-related epitopes was suggested to suffice in aborting further disease progression and stopping autoimmunity in mice (196, 197). Indeed, there is evidence that reactivity to certain antigens, i.e., insulin or glutamic acid decarboxylase (GAD) in T1D, is often observed before the autoimmune response expand to other autoantigens (198). However, it would be too risky to assume that eliminating a very restricted antigen repertoire will be sufficient to halt the entire complex autoimmune response. This could be particularly difficult in the human population, where the MHC, although strongly

associated to certain AID, is still very diverse and thus, allows presentation of multiple autoantigen epitopes in the context of different binding registers (197) that might even derive from post-translationally modified or hybrid peptides (199). It is also reasonable to expect that less immunodominant epitopes could drive AID once the stronger reactivities would be eliminated, provided this could be achieved completely.

The alternative options to promote regulatory responses have classically selected one or a few discrete autoantigen epitopes. The choice has been determined by the specific indication

(i.e., aiming to induce tolerance to therapeutic substances with high immunogenic potential) or by the conviction that truly regulatory responses directed to a single/discrete epitope/s involved in a given autoimmune disorder, are necessary and sufficient to control all disease causing reactivities in a bystander manner (183, 200). Therapeutics based on bystander regulation are intrinsically more feasible since they only require the knowledge of some disease associated autoantigens, which is available for most of the AID. On the other hand, it has to be noted that strictly classifying immune tolerance approaches

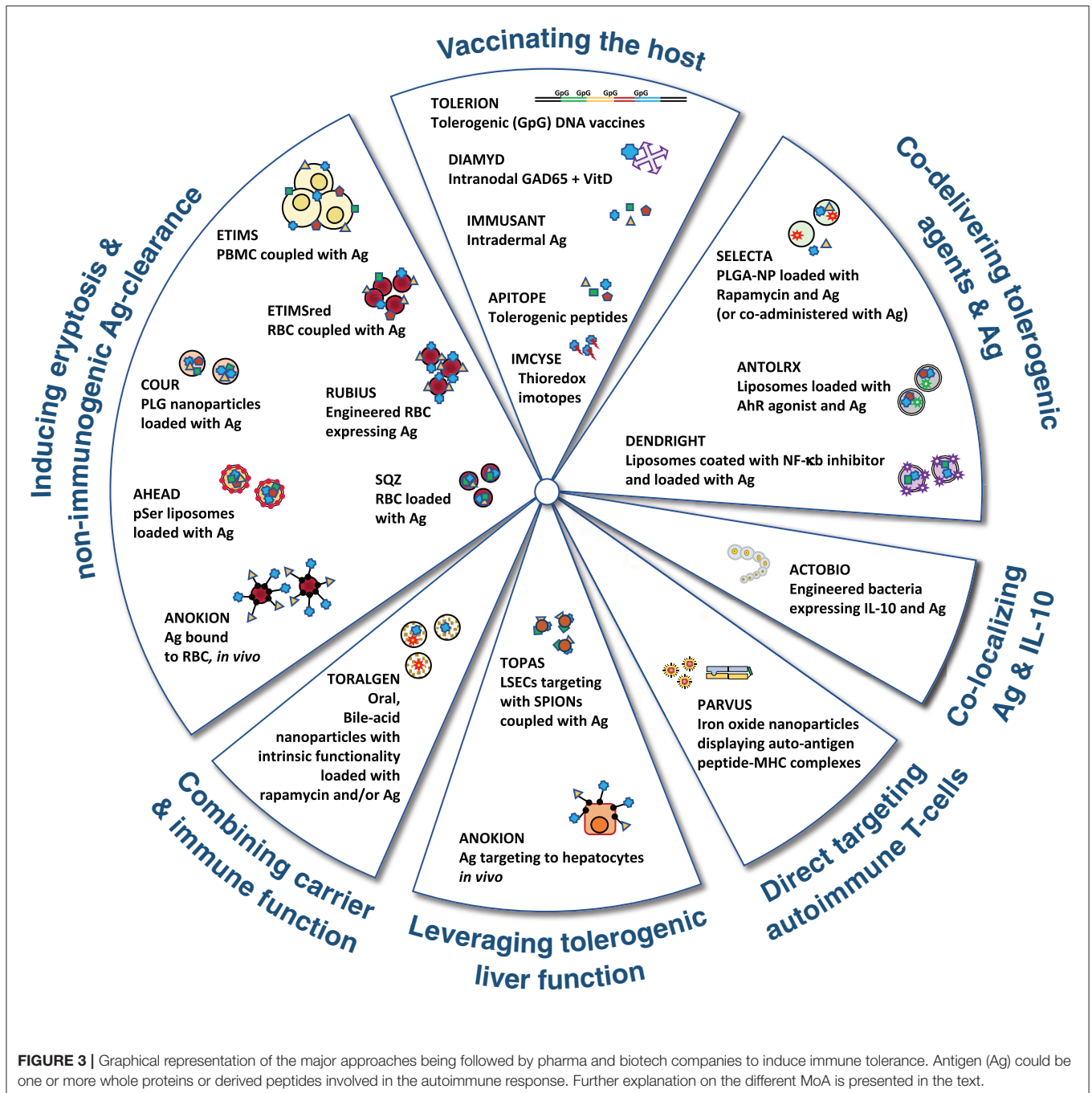


FIGURE 3 | Graphical representation of the major approaches being followed by pharma and biotech companies to induce immune tolerance. Antigen (Ag) could be one or more whole proteins or derived peptides involved in the autoimmune response. Further explanation on the different MoA is presented in the text.

into deletion or regulatory type is rather artificial. Often such choice is guided by the original scientific concept that prompted the therapeutic intervention. However, *in vivo*, these mechanisms probably overlap to a certain extent and immune tolerance might result from the concerted action of several different MoA (201, 202). A schematic representation of different approaches used to induce peripheral immune tolerance is depicted in **Figure 3**.

Inducing Eryptosis and Non-immunogenic Antigen-Clearance

Induction of tolerogenic responses that deleted clonotypic T cells were induced by coupling specific antigens to syngeneic lymphoid cells already four decades ago (203). Currently, the believe is that tolerogenic responses mediated by peptide antigens coupled to cells do not rely exclusively on induction of anergy or clonal deletion but on the parallel induction and expansion of regulatory T cells, which can also exert suppression, in the context of the affected organ, in a bystander manner (201, 204). These concepts have inspired a series of therapeutic approaches aiming to promote immune tolerance in AID patients. For example, the group led by Roland Martin infused MS patients with autologous PBMC, which were previously coupled with a cocktail of seven myelin peptides (Establish Tolerance in MS; ETIMS) (205). The same group, <http://nims-zh.ch/etimsred.html>, has initiated a subsequent clinical trial (ETIMSred) replacing PBMC by autologous red blood cells (RBC) (206). Both trials demonstrated feasibility of the approach and were safe and well-tolerated. Furthermore, *ex vivo* studies using PBMC from treated patients showed some reductions in recall T cell responses to myelin peptides (205, 206). Another scientific group has used a sortase-A mediated reaction to covalently link autoantigen peptides to mouse RBC and to, following re-infusion of these modified RBC, demonstrate protection toward experimental autoimmune encephalomyelitis (EAE) and T1D disease (207).

Several biotechnology companies are currently developing additional therapeutic products that use RBC as a carrier for autoantigen delivery: <https://www.rubiustx.com> is developing engineered RBC from autologous hematopoietic precursor cells that express AID-related antigens on their surfaces, and <http://sqzbiotech.com> is loading antigens into RBC by temporary disrupting the membrane of these cells, during a forced passage through microfluidic devices that contain antigen cocktails (208). These approaches are based on the principle of eryptosis (209), which ensures an orderly elimination of senescent or damaged RBC that is accompanied of regulatory instructions supporting the induction of anergic and/or tolerogenic responses (210). In this line, and taking advantage of the scrambling processes occurring in the cell membranes during apoptosis (211), scientists at <http://www.aheadtherapeutics.com> are developing a nanotechnology based on phosphatidylserine liposomes. These liposomes that additionally encapsulate autoantigens, are taken up by APC as if they would be apoptotic cells and thus, the cargo is presented to T cells in a tolerogenic manner (212, 213). A further strategy, built on existing physiologic mechanisms for clearing apoptotic debris, is pursued at <https://www.courpharma.com>. In this case the approach is based on proprietary

biodegradable poly[lactide-co-glycolide] (PLG) microparticles (circa 500-nm diameter) that encapsulate autoantigens and are functionalized on their surfaces to enhance their uptake by the mononuclear phagocyte system (214). It seems that the uptake of these microparticles for subsequent cargo processing and presentation to T cells in a tolerogenic manner (215, 216) is mediated by the scavenger receptor MARCO (217), which is abundant on the surface of marginal zone macrophages (218).

All the technologies indicated above require an *in vitro* or *ex vivo* loading of the antigenic peptide into/on the carrier cells or particles before transferring (generally by i.v. injection) the drug substance to patients with AID. An alternative to this laborious process is being considered at <https://anokion.com>. Their approach consists on the direct, *in vivo*, targeting of autoantigens to RBC. This intervention is accomplished by conjugating the selected proteins to a synthetic peptide (ERY1) that is specific, and displays high affinity, for glycophorin-A (GYPA), which is a surface molecule exclusively expressed on erythrocyte surfaces (219). In the case of small peptide antigens, the autoantigen focusing to RBC is accomplished by fusing these peptides to a single-chain Fv (scFv) antibody fragment (TER119) that is also directed to GYPA (219). Initially, the administration of these GYPA-targeting biologics was reported to induce tolerance to the linked antigens in mice, via deletion and/or anergy of cognate T cells (191, 219). More recently, these authors have also shown preclinical evidence for T_{reg} cell induction using the same principles (220).

Vaccinating the Host

Development of immune tolerance has also been attempted by direct administration of the antigen, both in the form of proteins or peptides or by using DNA vectors encoding these proteins. DNA vaccination is the approach selected at <https://tolerion.bio>. The company is developing DNA vaccines for intramuscular delivery using plasmids encoding different autoantigen epitopes that have the immunostimulatory CpG motifs replaced by immunoinhibitory GpG (221, 222).

Several approaches, mostly initiated by academic groups, aiming to induce immune tolerance using whole protein autoantigens or derived peptides have reached the clinic and although the approaches were shown to be safe, their insufficient efficacy did not support progression beyond phase 1 or 2 trials. These were recently reviewed by Serra and Santamaria elsewhere (190) and thus here only a few examples of biotech companies that remain actively engaged in clinical trials are discussed. <https://www.diamyd.com> is among the pioneers in treating T1D patients with a disease autoantigen. The initial intervention which included pediatric/juvenile patients consisted of multiple daily s.c. injections of glutamic acid decarboxylase 65-kilodalton isoform protein formulated in alum adjuvant (GAD-Alum). The treatment was well-tolerated but did not meet the overall expectations (223, 224). The group has continued with their immune tolerance program combining the original product GAD-Alum with immunomodulators such as Vit D and/or changing the route of administration (i.e., intranodal injections). <https://apitope.com> has based their immune tolerant products on the selection of peptides (apitopes) that can elicit T cell responses

without any further processing by the APC (225). The group has shown encouraging results in MS (226) and is pursuing additional AID indications. Another approach, using cocktails of immunogenic peptides delivered intradermally in the absence of adjuvants has been developed at <http://www.immusant.com>. The main focus of the company was on celiac disease (CeD) and the team delivered an encouraging phase 1 trial (227), which was safe and well-tolerated. However, the company has announced recently the discontinuation of the ongoing phase 2 study after reviewing the results of an interim analysis.

In most cases, the therapeutic intervention for the protein and epitope-based therapies involves the production of IL-10 and often, this signature is also associated to increased frequencies of T_{reg} cells. In some cases, the vaccination has been proposed to induce a switch from inflammatory T_{H1} profile toward anti-inflammatory types. In other cases, it is postulated that the antigen presentation of the immunogenic epitopes is performed by immature APC that deliver tolerogenic instructions to the cognate T cells and skew them into T_{reg} cells. Additional MoA can be induced using vaccination approaches. For example, <http://imcyse.com> is aiming to induce cytotoxic/cytolytic T cell responses to APC displaying disease related pMHC on their surfaces, *in vivo*. The therapy is based on s.c. administration of modified autoantigen peptides that are flanked by thioredoxin-like motives to augment pMHC-TcR interactions and promote the differentiation of cognate CD4⁺ T cells into cytolytic T cells. Specific killing of pMHC-loaded APC by the expanded “cytolytic killers” also induces the death of other disease-related T cells engaged with the same APC. Furthermore, such action prevents the activation of any disease-specific T cell by active removal of the APC source (228). A phase 1 trial in recent onset of T1D patients has been reported by the company with no safety concerns.

Co-delivering Tolerogenic Agents and Antigen

Delivering autoantigens alone is probably the simpler way to induce tolerance. However, such method rely considerable on the selection of the right peptide and potential adjuvant, as well as on the dose, frequency and route of administration, since ultimately the immune system of the treated patients, that is poised to promote effector responses, would need to interpret this new source of antigen and mount a regulatory response. This difficulty to predict the *in vivo* fate of the immunogenic proteins has triggered further therapies that deliver the autoantigens together with specific instructions to promote regulatory responses. One of those include the loading of the immunogenic cargo into autologous “tolerogenic” APC. Frequently, dendritic cells (DC) are differentiated from peripheral blood monocytes in the presence of modulatory agents that skew these cells toward anti-inflammatory/tolerogenic phenotypes or that remove some costimulatory elements before reinfusion into patients (190). Alternatively, and to avoid laborious cell therapy processes, the autoantigens could be delivered using nanoparticles that simultaneously incorporate compounds known to drive T_{reg} cell differentiation. Among others, rapamycin (229), aryl hydrocarbon receptor (AhR) agonists (230–232), and inhibitors of nuclear factor kappa-light-chain-enhancer of activated B cells

(NF-κB) (233, 234) have been described to induce tolerogenic phenotypes on DC.

<https://www.selectabio.com> is developing a platform based on polylactic acid (PLA) and poly(lactic-co-glycolic acid) (PLGA) nanoparticles that incorporate rapamycin and autoantigen, or only rapamycin if the autoantigen is co-injected (s.c.) with the tolerogenic nanoparticle (235–239). The company has successfully completed phase 2 trials to induce tolerance to exogenous administered uricase in patients with gout and is advancing in other indications. <https://antolrx.com> has chosen to deliver the AhR ligand, 2-(1'H-indole-3'-carbonyl)-thiazole-4-carboxylic acid methyl ester (ITE), together with autoantigen. Initially the two components were intended to be co-delivered by simultaneous coupling to the surface of gold nanoparticles (240, 241). Following a collaboration with <https://www.pfizer.com> the team has switched to liposomes as the preferred carrier vehicle (242). <https://www.dendright.com.au> on the other hand, utilizes a platform that incorporates NF-κB inhibitors in the lipid bilayer of liposomes and autoantigens in the hydrophilic core (243). The company has completed recently a phase 1 trial in RA patients using DEN-181; a liposome product containing calcitriol and a collagen II peptide (244).

Co-localizing Antigen and IL-10

In addition of delivering antigens, nanomedicine approaches have also aimed to deliver anti-inflammatory cytokines to reverse ongoing immune responses. For example, TGF-β1 and IL-2 have been encapsulated into PLGA nanoparticles to promote T_{reg} cell responses in mice (245). On the other hand, based on many reports using human and mouse cells *in vitro* as well as mouse models of disease, IL-10 has emerged as a key cytokine for dampening autoimmune responses. Such knowledge prompted its use in clinical trials for Crohn's Disease. Unfortunately, systemic IL-10 treatment to AID patients was not effective and rather resulted in considerable side effects (246, 247). These results encouraged the consideration to target IL-10 delivery to the inflammatory site minimizing systemic exposure. Scientists have explored different ways to locally deliver IL-10. For example, different densities of DEC-205 ligands for targeting, ovalbumin-encapsulating, nanoparticles to DC have been used to regulate IL-10 production by these cells (248). Alternative, other authors have encapsulated plasmids encoding IL-10 into poly[epsilon-caprolactone] (PCL) large (200 nm) nanoparticles for oral delivery and treatment of IBD in mice. These plasmids were released in the intestinal tract, endocytosed by cells of the intestinal lumen and subsequently, transcribed for local IL-10 expression (249). Gut expression of IL-10 is also the strategy followed by <https://actobio.com>, a biotechnology company that has opted for the use of genetically modified bacteria instead of nanoparticles. Particularly, they have replaced the thymidylate synthase gene of *Lactococcus lactis* by a synthetic sequence encoding human IL-10 (250). This company has successfully tested the safety of the approach, including biologic containment in a phase 1 clinical trial (251). The same group is also developing other products based on *L. lactis*, encoding nanobodies to human TNF-α (252) and is investing on immune tolerance programs for CeD (253) and T1D (254–256), by

providing simultaneous bacterial expression of the autoantigen and the anti-inflammatory cytokine. In the case of T1D, best preclinical data was always observed when the oral treatment with the engineered bacteria was complemented with systemic (i.v.) treatment with anti-CD3 mAb (254–256). Such preclinical evidence, together with the recent report on human T1D therapy using Teplizumab (39), suggests that a combination of these two agents would be a good approach to intervene in human T1D.

Direct Targeting of Autoimmune T Cells

A completely different strategy for the induction of immune tolerance is followed by <https://parvustx.com>. The platform of this biotech company is based on the development of Navacims, consisting of iron oxide nanoparticles that display large density arrays of pMHC. These pMHC are selected on the basis of known disease-related T cell epitopes that associate with MHC class II molecules displayed by patients suffering from AID. The Navacims engage directly microclusters of TcR on cognate, antigen experienced, CD4⁺ T cells in the absence of any costimulatory signal and instruct these cells to become T_{r1} cells (257, 258). These T_{r1} cells expand *in vivo* and are subsequently activated exclusively in the attacked tissue and its draining lymph nodes, where they locally suppress all disease-causing T_{eff} cell responses with no systemic effects. Efficacy of Navacims is mediated by IL-10, IL-21, and TGF-β1 and also involves functions of B_{reg} cells. Navacims have been shown to prevent disease progression in pre-diabetic non-obese diabetic (NOD) mice and reverse disease in already diabetic NOD mice (257). Mice treated with Navacims remain capable of mounting immune responses to pathogens and to develop antibody responses to disease unrelated antigens following vaccination (257). Human-specific Navacims expand human T_{reg} cells in NOD- *Scid*-common gamma chain deficient (NSG) mice reconstituted with PBMC from T1D patients (257). Furthermore, the induction of Navacim-specific T_{reg} cells provides a biomarker (using tetramers made of the same pMHC included in the Navacim) to guide human dosing and to decide timing of repeat treatment course (“booster” regimen) for maintenance of immune tolerance. A Navacim project for T1D is being developed in collaboration with <https://www.novartis.com>. Additional Navacim tool compounds have shown efficacy in mouse models of MS, RA, primary biliary cholangitis (PBC), primary sclerosing cholangitis (PSC) and autoimmune hepatitis (AIH) (257, 259). Navacims to treat CeLD, IBD, and autoimmune liver diseases are pursued together with <https://www.gene.com>. Navacims display exquisite selectivity for the organ and disease being targeted. However, to ensure a large patient coverage, it would be required to develop more than one product per disease. This could be easily accomplished in situations like T1D where disease is tightly associated to certain HLA DRB1 loci. Conditions with more diverse DRB1 associations might require the use of oligomorphic HLA loci, such as DRB3, DRB4, or DRB5 (259).

Leveraging Tolerogenic Liver Function

A feature of most types of particles, that is also shared by apoptotic cells, is their accumulation in the liver. The liver is the largest solid organ of the human body and it is generally regarded as a fundamental metabolic organ. However, the liver

is also a core component of the immune system that has key relevance removing pathogens and exogenous antigens from the systemic circulation and promoting tolerance toward these agents (260–262). The liver is strategically positioned at a crossroad between the intestine and systemic circulation. It receives blood supply from the hepatic artery containing naïve and memory lymphocytes and from the portal vein, which is rich in food and microbial antigens. These gut-derived constituents are efficiently cleared by cells of the hepatic reticuloendothelial system such as fenestrated liver sinusoidal endothelial cells (LSECs), Kupffer cells, and DC, which also present them to passenger T cells in a tolerogenic context. Hepatic APC secrete abundant TGF-β1 and IL-10, express PD-L1 and contribute to the differentiation of T_{reg} cells (260–262). Thus, the liver immune system favors induction and maintenance of tolerance over immunity, avoiding immune responses to harmless foreign antigens form the diet and this function is believed to be largely mediated by LSECs (263–268). Based on these observations, <https://topas-therapeutics.com> is developing a nanomedicine platform aiming to focus autoantigens to LSECs. This company generates super paramagnetic iron oxide nanoparticles (SPION) of about 10 nm diameter that are conjugated with up to 100 autoantigen peptides (269). The small size of these particles is a feature for LSEC targeting. LSECs display poor phagocytic capabilities and preferentially take up soluble macromolecules and small particles (270). Electron microscopy of liver sections from mice treated with SPIONs have been shown how these nanoparticles localize into the endosomal compartment of LSECs (269). Treatment with SPIONs coated with myelin basic protein (MBP) or myelin oligodendrocyte glycoprotein (MOG) peptide protected mice from EAE and improved disease scores in mice with established disease (269). Interestingly, disease protection was also achieved in splenectomized mice, indicating that the spleen was not required for the induction of the regulatory response although it played a role in maintenance of tolerance (269). The relevance of hepatic APC promoting immune tolerance has triggered a new research line at <https://anokion.com>. Adding to the erythrocyte binding technology described above, further binding principles based on N-acetyl galactosamine (GalNAc) or N-acetylglucosamine (GluNAc) that target receptors expressed by hepatic APC are now described (271). Binding and internalization of galactose (Gal) or N-acetyl galactosamine (GalNAc)-terminating glycoproteins by hepatocytes is often mediated by the C-type lectin receptor, asialo-glycoprotein receptor (ASGPR) (272), which is a feature often used in gene therapy approach to deliver the therapeutic principles to hepatic cells (272). Interestingly, scientists at the Baylor Institute for Immunology Research have demonstrated that focusing of antigens to DC using antibodies directed to a type of ASGPR expressed on these cells is a very effective method to generate antigen-specific IL-10-producing T cells with suppressive capabilities (273).

Combining Carrier and Immune Function

The liver is intimately linked to the gut and overall intestinal tract and the composition of the microbiome represents another node of crucial relevance preventing or inducing autoimmunity (274). The gut microbiota can control gut inflammation by preventing

or promoting T_{reg} cell responses (275). There are also reports demonstrating that, based on molecular mimicry, $CD8^+$ T cell responses against microbial antigens that protect mouse and human hosts against colitogenic insults are also linked to T1D (276). It is conceivable to think that many more cases of cross-reactivity between human and microbiota antigens exist, and that evolutionary regulatory responses that prevent exacerbated responses to commensal bacteria also support tolerance to self-antigens. Conversely, mechanisms promoting the elimination of certain microbiota could result in autoimmunity in patients with a predisposing HLA. Furthermore, metabolites produced in the gut by the host or microbiome may also play an important role controlling homeostasis at distant sites. A good example of metabolic interaction between host and microbiome is provided by the production of bile acids (BA), which are initially produced as primary BA from cholesterol in the liver and subsequently, modified into secondary BA in the intestine by commensal bacterial (277). Both types of BA are capable on interacting with, farnesoid-X-receptor (FXR) and G protein-coupled bile acid receptor 1 (GPBAR1), although the preferences of primary BA are for FXR and secondary BA prefer GPBAR1 (277). Because the function of different BA receptors could be immune counterregulatory the balance of the different BA species due to alterations in microbiota composition or following exogenous administration or one or other type of agonist could be very important maintaining immune homeostasis (278). In this regard, the platform announced by <https://toralgen.com> that proposes the delivery of multiple types of cargo using oral nanoparticles made of polymerized ursodeoxycholic acid (pUDCA) (279), represents an advance toward the use of vehicle nanocarriers with intrinsic therapeutic function.

SAFETY ASPECTS

In this review, we have discussed three major types of therapies aiming to ameliorate AID, namely biologic immunotherapy, cell (and gene) therapy, and immune tolerance. All of these emerged with the intention to close the therapeutic gap resulting from the limited availability of medicines capable of delivering a substantial clinical benefit in the absence of adverse effects (280). Biologics were pioneers in this attempt and represent today a major part of the established drugs used for autoinflammatory and AID. These drugs have an exquisite selectivity for their targets, which is a major difference over the classical low molecular weight remedies. Nevertheless, they target immunocompetent cell subsets, soluble factors, or pathways, which are essential in host resistance to pathogens, and therefore they also carry the risk of increasing the susceptibility to opportunistic infections. This problem turned out to be a major concern for the development of the first TNF- α blockers (281), and it is still today a significant cause of morbidity for many immunotherapies (9, 10, 282, 283). Moreover, in the context of obtaining an adequate immunological balance, TNF- α blockers intended for the treatment of RA have elicited SLE (284–286). Over the years, the scientific community has learned that immunotherapy products do not represent

universal solutions for all patient populations and that their indiscriminate use could be detrimental for some of them. Different drugs have shown to be efficacious in some diseases, but of limited benefit in others, which is in line with the recent views classifying autoinflammatory and autoimmune disorders according to hierarchic cytokine pathway maps (7, 11). Furthermore, the existence of multiple endotypes within a given disease is recognized (287, 288), and thus, there is a need to accurately diagnose the appropriate disease subtype and provide the right treatment to the right patient at the right time (289).

In addition to the specific target effects, therapeutic proteins, particularly when they contain non-human sequences and/or are repeatedly administered, can be immunogenic and promote host anti-drug antibody (ADA) responses. The clinical utility of immunotherapeutics may be impacted by these ADA, since they could neutralize the therapeutic effects of the medicine and/or induce hypersensitivity reactions in the treated patient (290, 291). Strategies to mitigate these risks included switching to a different molecule with similar targeting capabilities, engineering the product to eliminate as much as possible the non-human sequences (i.e., mouse antibody framework), reducing the chance to generate protein aggregates, eliminating Fc receptor binding ability or inducing immune tolerance. Most of these solutions are already incorporated during the design of the new drugs, however when these predictions fail and ADA responses occur, the removal of ADAs could be a real challenge and could result in a discontinuation of the product.

As discussed in this review, cell therapies intended to induce regulatory responses could be polyclonal (antigen agnostic) or antigen-specific. Polyclonal T_{reg} cells will be able to efficiently suppress T_{eff} cells of various specificities (bystander suppression), however they will also indiscriminately affect the host defense response (dominant suppression) and therefore, the same risks of infections related to some of the biologics will apply (145, 146). Antigen-specific cell therapies will be safer. However, if they result from genetic engineering of the regulatory cell populations other risks associated to uncontrolled expansion and extensive bystander suppression may develop. One extreme of non-specific immune dysregulation, could be severe acute immunosuppressive effects mediated by cytokine release syndrome of CAR- T_{reg} cells, resembling the immunostimulatory outcomes observed with the complementary effector CAR-T cell therapies in oncology (292). Furthermore, T_{reg} cells may still display a considerable phenotypic plasticity and consequently, there is a risk that the inflammatory conditions ongoing in the treated patients (e.g., cytokine milieu, availability of antigen, concomitant therapy) affect the phenotype of the transferred cells, reverting them into subsets with potential to exacerbate disease. Thus, the level of required immunosuppression likely will need to be customized for each autoimmune and autoinflammatory disorder and to be balanced against unwanted, non-specific immunosuppression (146).

Some of the approaches currently followed to induce immune tolerance were inspired by the antigen desensitization protocols established for the treatment of allergy. These therapies aimed

originally to switch the ratios of allergen-specific IgE toward the non-anaphylactogenic IgG4 class, rather than searching for a true induction of tolerance that should have dampened all antibody responses (293). To achieve such goal, atopic patients were exposed, over long periods (months to years), to gradually increasing doses of allergen. Such process implied the administration of the whole allergen which represented a high risk of anaphylaxis. In general, autoimmune responses are not associated to IgE, and the administration of autoantigen protein or peptides used in the immune tolerance protocols does not favor the induction of autoantigen-specific IgE reactions. However, IgG-mediated anaphylactic responses could develop upon repeated exposure of the antigen (294) as detected for some therapeutic antibodies (295) and thus, these could also evolve during antigen-specific tolerance protocols that require repeated administration of proteins or peptides alone or attached to the surfaces of nanoparticles. Furthermore, therapies that expose patients to the same immunogenic agents involved in the autoimmune attack have the inherent risk of provoking disease exacerbation rather than protection. The challenge is particularly high since these safety concerns may not be optimally extrapolated from preclinical animal studies, even from non-human primates, because both clinical intervention and potential disease exacerbation are strongly linked to predisposing MHC alleles/haplotypes that are specific to human. Therefore, careful design for first in human, phase 1 trials, is essential and it should pay increased attention to the selection of dose and frequency of administration. Moreover, the status of the individuals in phase 1 studies is also critical, since choosing healthy individuals displaying HLA alleles/haplotypes not associated with the AID might be irrelevant to obtain any preliminary indication of target engagement and/or efficacy for the drug candidates under evaluation. On the other hand, choosing healthy individuals expressing HLA alleles/haplotypes related to the disease might expose them to un-ethical drug exposure that might trigger unwanted immune responses. Such argumentation would support the selection of patients from the very onset of clinical testing. However, an acceptable approach would be to opt for patients with only subclinical disease or for patients with long-lasting disease. Both situations offer advantages and inconveniences for clinical evaluation and the choice will be determined by the MoA of the test drug and specific trial design. These studies would benefit decisively of having exploratory and clinically accepted biomarkers that can predict the outcome of the trial. In addition, a cautious clinical approach that takes into account the specific MoA of each therapy will be required to advance the different programs.

CONCLUSION

This review has discussed some of the existing and developing immunotherapy and precision medicine approaches aiming to ameliorate and/or cure AID. The established immunotherapies for AID are the less specific options whereas highly sophisticated antigen-specific alternatives are still under development. Immunotherapy is particularly useful for autoinflammatory conditions, where the antigenic trigger is still elusive, or the

disease is mediated by multifactorial activation of several inflammatory pathways like those found in cryopyrin-associated periodic syndrome. These therapies are also very efficient in some selected AID, like Pso where they can deliver extraordinary efficacy results with almost absence of adverse effects. Unfortunately, immunotherapies for all AID do not exist and, when they are available, they still rely heavily on immunosuppression that is associated with increased susceptibility to opportunistic infections. Cell therapies represent a bridge between immunotherapy and immune tolerance since these living drugs have the potential to adapt to the host environment and produce a repertoire of factors and immune functions of benefit for the affected patient. However, in most cases they are not specific and/or involve logistically complicated development processes that are patient-personalized. The emerging immune tolerance alternatives do have the potential to address all the missing aspects of the former therapies and to provide a truly transformative and game changing option for the treatment of AID.

FUTURE PERSPECTIVES

It is expected that immunotherapies for AID will continue to evolve. They will (i) incorporate new target specificities, i.e., immune checkpoints, (ii) increase in sophistications to enhance efficacy and reduce adverse effects, i.e., utilizing bispecific molecules and selectively using them in optimal disease endotypes, and (iii) gain acceptance as first line treatment option, i.e., with increased use of biosimilars. Immunotherapeutics are expected to be the next current standard of care and to significantly raise the requirements for clinical improvement of newly developed medicines. Cell-based personalized therapies might find their niche in the near future for the treatment of AID. However, their development processes will require substantial improvements to accelerate production, combine efficacy features, incorporate safety switches and reduce costs to ensure broad availability to all patients in need. The current assortment of immune tolerance approaches makes it difficult to predict which of them will provide the most effective MoA. Hence, it is anticipated that one or several of these strategies will succeed in delivering the long-awaited curative solution. "...rien n'est plus imminent que l'impossible, et que ce qu'il faut toujours prévoir, c'est l'imprévu" (296).

AUTHOR CONTRIBUTIONS

JC, CRe, CRa, LR, DP, and MK contributed to the writing of the manuscript, are accountable for its content, and approved its publication.

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REFERENCES

- Gutierrez-Arcelus M, Rich SS, Raychaudhuri S. Autoimmune diseases - connecting risk alleles with molecular traits of the immune system. *Nat Rev Genet.* (2016) 17:160–74. doi: 10.1038/nrg.2015.33
- Cooper GS, Stroehla BC. The epidemiology of autoimmune diseases. *Autoimmun Rev.* (2003) 2:119–25. doi: 10.1016/S1568-9972(03)00006-5
- Nash P, Clegg DO. Psoriatic arthritis therapy: NSAIDs and traditional DMARDs. *Ann Rheum Dis.* (2005) 64(Suppl 2):ii74–7. doi: 10.1136/ard.2004.030783
- Jamilloux Y, Jammal El T, Vuitton L, Gerfaud-Valentin M, Kerever S, Sève P. JAK inhibitors for the treatment of autoimmune and inflammatory diseases. *Autoimmun Rev.* (2019) 18:102390. doi: 10.1016/j.autrev.2019.102390
- Winthrop KL. Serious infections with antirheumatic therapy: are biologicals worse? *Ann Rheum Dis.* (2006) 65(Suppl. 3):iii54–7. doi: 10.1136/ard.2006.058503
- Winthrop KL. The emerging safety profile of JAK inhibitors in rheumatic disease. *Nat Rev Rheumatol.* (2017) 13:234–43. doi: 10.1038/nrrheum.2017.23
- Schett G, Elewaut D, McInnes IB, Dayer J-M, Neurath MF. How cytokine networks fuel inflammation: toward a cytokine-based disease taxonomy. *Nat Med.* (2013) 19:822–4. doi: 10.1038/nm.3260
- Smolen JS, landewé R, Bijlsma J, Burmester G, Chatzidionysiou K, Dougados M, et al. EULAR recommendations for the management of rheumatoid arthritis with synthetic and biological disease-modifying antirheumatic drugs: 2016 update. *Ann Rheum Dis.* (2017) 76:960–77. doi: 10.1136/annrheumdis-2016-210715
- Woodrick RS, Ruderman EM. Safety of biologic therapy in rheumatoid arthritis. *Nat Rev Rheumatol.* (2011) 7:639–52. doi: 10.1038/nrrheum.2011.145
- Fishman JA. Infection in organ transplantation. *Am J Transplant.* (2017) 17:856–79. doi: 10.1111/ajt.14208
- McInnes IB, Buckley CD, Isaacs JD. Cytokines in rheumatoid arthritis - shaping the immunological landscape. *Nat Rev Rheumatol.* (2016) 12:63–8. doi: 10.1038/nrrheum.2015.171
- Monaco C, Nanchahal J, Taylor P, Feldmann M. Anti-TNF therapy: past, present and future. *Int Immunol.* (2015) 27:55–62. doi: 10.1093/intimm/dxu102
- Baddley JW, Cantini F, Goletti D, Gómez-Reino JJ, Mylonakis E, San-Juan R, et al. ESCMID Study Group for Infections in Compromised Hosts (ESGICH) consensus document on the safety of targeted and biological therapies: an infectious diseases perspective (soluble immune effector molecules [I]: anti-tumor necrosis factor- α agents). *Clin Microbiol Infect.* 24(Suppl. 2):S10–20. doi: 10.1016/j.cmi.2017.12.025
- Godfrey MS, Friedman LN. Tuberculosis and biologic therapies: anti-tumor necrosis factor- α and beyond. *Clin Chest Med.* (2019) 40:721–39. doi: 10.1016/j.ccm.2019.07.003
- Lachmann HJ, Kone-Paut I, Kuemmerle-Deschner JB, Leslie KS, Hachulla E, Quartier P, et al. Use of canakinumab in the cryopyrin-associated periodic syndrome. *N Engl J Med.* (2009) 360:2416–25. doi: 10.1056/NEJMoa0810787
- Swanson KV, Deng M, Ting JPY. The NLRP3 inflammasome: molecular activation and regulation to therapeutics. *Nat Rev Immunol.* (2019) 19:477–89. doi: 10.1038/s41577-019-0165-0
- Tsai Y-C, Tsai T-F. Anti-interleukin and interleukin therapies for psoriasis: current evidence and clinical usefulness. *Ther Adv Musculoskelet Dis.* (2017) 9:277–94. doi: 10.1177/1759720X17735756
- McGeachy MJ, Cua DJ, Gaffen SL. The IL-17 family of cytokines in health and disease. *Immunity.* (2019) 50:892–906. doi: 10.1016/j.immuni.2019.03.021
- Koutruba N, Emer J, Lebwohl M. Review of ustekinumab, an interleukin-12 and interleukin-23 inhibitor used for the treatment of plaque psoriasis. *Ther Clin Risk Manag.* (2010) 6:123–41. doi: 10.2147/TCRM.S5599
- Neurath MF. Current and emerging therapeutic targets for IBD. *Nat Rev Gastroenterol Hepatol.* (2017) 14:269–78. doi: 10.1038/nrgastro.2016.208
- Kotake S, Udagawa N, Takahashi N, Matsuzaki K, Itoh K, Ishiyama S, et al. IL-17 in synovial fluids from patients with rheumatoid arthritis is a potent stimulator of osteoclastogenesis. *J Clin Invest.* (1999) 103:1345–52. doi: 10.1172/JCI15703
- Sato K, Suematsu A, Okamoto K, Yamaguchi A, Morishita Y, Kadono Y, et al. Th17 functions as an osteoclastogenic helper T cell subset that links T cell activation and bone destruction. *J Exp Med.* (2006) 203:2673–82. doi: 10.1084/jem.20061775
- Hueber W, Patel DD, Dryja T, Wright AM, Koroleva I, Bruin G, et al. Effects of AIN457, a fully human antibody to interleukin-17A, on psoriasis, rheumatoid arthritis, and uveitis. *Sci Transl Med.* (2010) 2:52ra72. doi: 10.1126/scitranslmed.3001107
- van de Kerkhof PCM, Griffiths CEM, Reich K, Leonardi CL, Blauvelt A, Tsai T-F, et al. Secukinumab long-term safety experience: a pooled analysis of 10 phase II and III clinical studies in patients with moderate to severe plaque psoriasis. *J Am Acad Dermatol.* (2016) 75:83–98.e4. doi: 10.1016/j.jaad.2016.03.024
- Segueni N, Tritto E, Bourigault M-L, Rose S, Erard F, Le Bert M, et al. Controlled *Mycobacterium tuberculosis* infection in mice under treatment with anti-IL-17A or IL-17F antibodies, in contrast to TNF α neutralization. *Sci Rep.* (2016) 6:36923–17. doi: 10.1038/srep36923
- Kammüller M, Tsai T-F, Griffiths CE, Kapoor N, Kolattukudy PE, Brees D, et al. Inhibition of IL-17A by secukinumab shows no evidence of increased *Mycobacterium tuberculosis* infections. *Clin Transl Immunol.* (2017) 6:e152. doi: 10.1038/cti.2017.34
- Jackson SW, Davidson A. BAFF inhibition in SLE-Is tolerance restored? *Immunol Rev.* (2019) 292:102–19. doi: 10.1111/imr.12810
- Du FH, Mills EA, Mao-Draayer Y. Next-generation anti-CD20 monoclonal antibodies in autoimmune disease treatment. *Auto Immun Highlights.* (2017) 8:12. doi: 10.1007/s13317-017-0100-y
- Rehwal U, Schulz H, Reiser M, Sieber M, Staak JO, Morschhauser F, et al. Treatment of relapsed CD20+ hodgkin lymphoma with the monoclonal antibody rituximab is effective and well tolerated: results of a phase 2 trial of the German hodgkin lymphoma study group. *Blood.* (2003) 101:420–4. doi: 10.1182/blood.V101.2.420
- Boster A, Ankeny DP, Racke MK. The potential role of B cell-targeted therapies in multiple sclerosis. *Drugs.* (2010) 70:2343–56. doi: 10.2165/11585230-000000000-00000
- Blüml S, McKeever K, Ettinger R, Smolen J, Herbst R. B-cell targeted therapeutics in clinical development. *Arthritis Res Ther.* (2013) 15(Suppl. 1):S4. doi: 10.1186/ar3906
- Gürçan HM, Keskin DB, Stern JNH, Nitzberg MA, Shekhani H, Ahmed AR. A review of the current use of rituximab in autoimmune diseases. *Int Immunopharmacol.* (2009) 9:10–25. doi: 10.1016/j.intimp.2008.10.004
- Ginaldi L, De Martinis M, Matutes E, Farahat N, Morilla R, Dyer MJ, et al. Levels of expression of CD52 in normal and leukemic B and T cells: correlation with *in vivo* therapeutic responses to Campath-1H. *Leuk Res.* (1998) 22:185–91. doi: 10.1016/S0145-2126(97)00158-6
- Cohen JA, Coles AJ, Arnold DL, Confavreux C, Fox EJ, Hartung H-P, et al. Alemtuzumab versus interferon beta 1a as first-line treatment for patients with relapsing-remitting multiple sclerosis: a randomised controlled phase 3 trial. *Lancet.* (2012) 380:1819–28. doi: 10.1016/S0140-6736(12)61769-3

35. Coles AJ, Twyman CL, Arnold DL, Cohen JA, Confavreux C, Fox EJ, et al. Alectuzumab for patients with relapsing multiple sclerosis after disease-modifying therapy: a randomised controlled phase 3 trial. *Lancet*. (2012) 380:1829–39. doi: 10.1016/S0140-6736(12)61768-1
36. Kung P, Goldstein G, Reinherz EL, Schlossman SF. Monoclonal antibodies defining distinctive human T cell surface antigens. *Science*. (1979) 206:347–9. doi: 10.1126/science.314668
37. Norman DJ, Leone MR. The role of OKT3 in clinical transplantation. *Pediatr Nephrol*. (1991) 5:130–6. doi: 10.1007/BF00852870
38. Long SA, Thorpe J, DeBerg HA, Gersuk V, Eddy JA, Harris KM, et al. Partial exhaustion of CD8T cells and clinical response to teplizumab in new-onset type 1 diabetes. *Sci Immunol*. (2016) 1:eaa17793. doi: 10.1126/sciimmunol.aai7793
39. Herold KC, Bundy BN, Long SA, Bluestone JA, Dimeglio LA, Dufort MJ, et al. An Anti-CD3 antibody, teplizumab, in relatives at risk for type 1 diabetes. *N Engl J Med*. (2019) 381:603–13. doi: 10.1056/NEJMoa1902226
40. Gottlieb A, Krueger JG, Bright R, Ling M, Lebwohl M, Kang S, et al. Effects of administration of a single dose of a humanized monoclonal antibody to CD11a on the immunobiology and clinical activity of psoriasis. *J Am Acad Dermatol*. (2000) 42:428–35. doi: 10.1016/S0190-9622(00)90214-7
41. Gottlieb AB, Hamilton T, Caro I, Kwon P, Compton PG, Leonardi CL, et al. Long-term continuous efalizumab therapy in patients with moderate to severe chronic plaque psoriasis: updated results from an ongoing trial. *J Am Acad Dermatol*. (2006) 54(4 Suppl. 1):S154–63. doi: 10.1016/j.jaad.2005.12.018
42. Takada Y, Ye X, Simon S. The integrins. *Genome Biol*. (2007) 8:215. doi: 10.1186/gb-2007-8-5-215
43. Kothary N, Diak I-L, Brinker A, Bezabeh S, Avigan M, Dal Pan G. Progressive multifocal leukoencephalopathy associated with efalizumab use in psoriasis patients. *J Am Acad Dermatol*. (2011) 65:546–51. doi: 10.1016/j.jaad.2010.05.033
44. Schwab N, Ulzheimer JC, Fox RJ, Schneider-Hohendorf T, Kieseier BC, Monoranu CM, et al. Fatal PML associated with efalizumab therapy: insights into integrin $\alpha\text{L}\beta\text{2}$ in JC virus control. *Neurology*. (2012) 78:458–67. doi: 10.1212/WNL.0b013e3182478d4b
45. Polman CH, O'Connor PW, Havrdova E, Hutchinson M, Kappos L, Miller DH, et al. A randomized, placebo-controlled trial of natalizumab for relapsing multiple sclerosis. *N Engl J Med*. (2006) 354:899–910. doi: 10.1056/NEJMoa044397
46. Ghosh S, Goldin E, Gordon FH, Malchow HA, Rask-Madsen J, Rutgeerts P, et al. Natalizumab for active crohn's disease. *N Engl J Med*. (2003) 348:24–32. doi: 10.1056/NEJMoa020732
47. Sandborn WJ, Colombel J-F, Enns R, Feagan BG, Hanauer SB, Lawrance IC, et al. Natalizumab induction and maintenance therapy for crohn's disease. *N Engl J Med*. (2005) 353:1912–25. doi: 10.1056/NEJMoa043335
48. Van Assche G, Van Ranst M, Sciort R, Dubois B, Vermeire S, Noman M, et al. Progressive multifocal leukoencephalopathy after natalizumab therapy for Crohn's disease. *N Engl J Med*. (2005) 353:362–8. doi: 10.1056/NEJMoa051586
49. Langer-Gould A, Atlas SW, Green AJ, Bollen AW, Pelletier D. Progressive multifocal leukoencephalopathy in a patient treated with natalizumab. *N Engl J Med*. (2005) 353:375–81. doi: 10.1056/NEJMoa051847
50. Kleinschmidt-DeMasters BK, Tyler KL. Progressive multifocal leukoencephalopathy complicating treatment with natalizumab and interferon beta-1a for multiple sclerosis. *N Engl J Med*. (2005) 353:369–74. doi: 10.1056/NEJMoa051782
51. Tan CS, Koralnik IJ. Progressive multifocal leukoencephalopathy and other disorders caused by JC virus: clinical features and pathogenesis. *Lancet Neurol*. (2010) 9:425–37. doi: 10.1016/S1474-4422(10)70040-5
52. Bamias G, Clark DJ, Rivera-Nieves J. Leukocyte traffic blockade as a therapeutic strategy in inflammatory bowel disease. *Curr Drug Targets*. (2013) 14:1490–500. doi: 10.2174/13894501113149990158
53. Sandborn WJ, Cyrille M, Hansen MB, Feagan BG, Loftus EV, Rogler G, et al. Efficacy and safety of abirumab in a randomized, placebo-controlled trial for moderate-to-severe ulcerative colitis. *Gastroenterology*. (2019) 156:946–57.e18. doi: 10.1053/j.gastro.2018.11.035
54. Lichnog C, Klabunde S, Becker E, Fuh F, Tripal P, Atreya R, et al. Cellular mechanisms of etrolizumab treatment in inflammatory bowel disease. *Front Pharmacol*. (2019) 10:39. doi: 10.3389/fphar.2019.00039
55. Reinisch W, Sandborn WJ, Danese S, Hébuterne X, Salzberg B, Klopocka M, et al. Long-term safety and efficacy on the anti-mucosal addressin cell adhesion molecule-1 (MADCAM-1) antibody SHP647 in ulcerative colitis: an open-label extension study (TURANDOT II). *Inflamm Bowel Dis*. (2019) 25:S347. doi: 10.1093/ibd/izy393.041
56. Schall TJ, Proudfoot AEI. Overcoming hurdles in developing successful drugs targeting chemokine receptors. *Nat Rev Immunol*. (2011) 11:355–63. doi: 10.1038/nri2972
57. Dai Z, Xing L, Cerise J, Wang EHC, Jabbari A, de Jong A, et al. CXCR3 blockade inhibits t cell migration into the skin and prevents development of alopecia areata. *J Immunol*. (2016) 197:1089–99. doi: 10.4049/jimmunol.1501798
58. He S, Cao Q, Qiu Y, Mi J, Zhang JZ, Jin M, et al. A new approach to the blocking of alloreactive T cell-mediated graft-versus-host disease by *in vivo* administration of anti-CXCR3 neutralizing antibody. *J Immunol*. (2008) 181:7581–92. doi: 10.4049/jimmunol.181.11.7581
59. Richmond JM, Masterjohn E, Chu R, Tedstone J, Youd ME, Harris JE. CXCR3 depleting antibodies prevent and reverse vitiligo in mice. *J Invest Dermatol*. (2017) 137:982–5. doi: 10.1016/j.jid.2016.10.048
60. Robert R, Juglair L, Lim EX, Ang C, Wang CJH, Ebert G, et al. A fully humanized IgG-like bispecific antibody for effective dual targeting of CXCR3 and CCR6. *PLoS ONE*. (2017) 12:e0184278. doi: 10.1371/journal.pone.0184278
61. Robert R, Ang C, Sun G, Juglair L, Lim EX, Mason LJ, et al. Essential role for CCR6 in certain inflammatory diseases demonstrated using specific antagonist and knockin mice. *JCI Insight*. (2017) 2:e94821. doi: 10.1172/jci.insight.94821
62. Andrews LP, Yano H, Vignali DAA. Inhibitory receptors and ligands beyond PD-1, PD-L1 and CTLA-4: breakthroughs or backups. *Nat Immunol*. (2019) 20:1425–34. doi: 10.1038/s41590-019-0512-0
63. Huang C, Zhu H-X, Yao Y, Bian Z-H, Zheng Y-J, Li L, et al. Immune checkpoint molecules. Possible future therapeutic implications in autoimmune diseases. *J Autoimmun*. (2019) 104:102333. doi: 10.1016/j.jaut.2019.102333
64. Postow MA, Callahan MK, Wolchok JD. Immune checkpoint blockade in cancer therapy. *J Clin Oncol*. (2015) 33:1974–82. doi: 10.1200/JCO.2014.59.4358
65. Jenneck C, Novak N. The safety and efficacy of alefacept in the treatment of chronic plaque psoriasis. *Ther Clin Risk Manag*. (2007) 3:411–20.
66. Edward M, Vital PE. Abatacept in the treatment of rheumatoid arthritis. *Ther Clin Risk Manag*. (2006) 2:365–75. doi: 10.2147/tcrm.2006.2.4.365
67. Miller GT, Hochman PS, Meier W, Tizard R, Bixler SA, Rosa MD, et al. Specific interaction of lymphocyte function-associated antigen 3 with CD2 can inhibit T cell responses. *J Exp Med*. (1993) 178:211–22. doi: 10.1084/jem.178.1.211
68. Cooper JC, Morgan G, Harding S, Subramanyam M, Majeau GR, Moulder K, et al. Alefacept selectively promotes NK cell-mediated deletion of CD45R0+ human T cells. *Eur J Immunol*. (2003) 33:666–75. doi: 10.1002/eji.200323586
69. Ellis CN, Krueger GG, Alefacept Clinical Study Group. Treatment of chronic plaque psoriasis by selective targeting of memory effector T lymphocytes. *N Engl J Med*. (2001) 345:248–55. doi: 10.1056/NEJM200107263450403
70. Rigby MR, Harris KM, Pinckney A, Dimeglio LA, Rendell MS, Felner EI, et al. Alefacept provides sustained clinical and immunological effects in new-onset type 1 diabetes patients. *J Clin Invest*. (2016) 125:3285–96. doi: 10.1172/JCI81722
71. Beyersdorf N, Kerkau T, Hünig T. CD28 co-stimulation in T-cell homeostasis: a recent perspective. *Immunotargets Ther*. (2015) 4:111–22. doi: 10.2147/ITT.S61647
72. Eesensten JH, Helou YA, Chopra G, Weiss A, Bluestone JA. CD28 Costimulation: from mechanism to therapy. *Immunity*. (2016) 44:973–88. doi: 10.1016/j.immuni.2016.04.020
73. Schildberg FA, Klein SR, Freeman GJ, Sharpe AH. Coinhibitory pathways in the B7-CD28 ligand-receptor family. *Immunity*. (2016) 44:955–72. doi: 10.1016/j.immuni.2016.05.002

74. Davis PM, Abraham R, Xu L, Nadler SG, Suchard SJ. Abatacept binds to the Fc receptor CD64 but does not mediate complement-dependent cytotoxicity or antibody-dependent cellular cytotoxicity. *J Rheumatol.* (2007) 34:2204–10.
75. Blair HA, Deeks ED. Abatacept: a review in rheumatoid arthritis. *Drugs.* (2017) 77:1221–33. doi: 10.1007/s40265-017-0775-4
76. Vincenti F, Rostaing L, Grinyo J, Rice K, Steinberg S, Gaité L, et al. Belatacept and long-term outcomes in kidney transplantation. *N Engl J Med.* (2016) 374:333–43. doi: 10.1056/NEJMoa1506027
77. Ippoliti G, D'Armini AM, Lucioni M, Marjeh M, Viganò M. Introduction to the use of belatacept: a fusion protein for the prevention of posttransplant kidney rejection. *Biologics.* (2012) 6:355–62. doi: 10.2147/BTT.S27565
78. Slavik JM, Hutchcroft JE, Bierer BE. CD80 and CD86 are not equivalent in their ability to induce the tyrosine phosphorylation of CD28. *J Biol Chem.* (1999) 274:3116–24. doi: 10.1074/jbc.274.5.3116
79. Lin C-H, Hünig T. Efficient expansion of regulatory T cells *in vitro* and *in vivo* with a CD28 superagonist. *Eur J Immunol.* (2003) 33:626–38. doi: 10.1002/eji.200323570
80. Hünig T. The storm has cleared: lessons from the CD28 superagonist TGN1412 trial. *Nat Rev Immunol.* (2012) 12:317–8. doi: 10.1038/nri3192
81. Horvath C, Andrews L, Baumann A, Black L, Blanset D, Cavagnaro J, et al. Storm forecasting: additional lessons from the CD28 superagonist TGN1412 trial. *Nat Rev Immunol.* (2012) 12:740. doi: 10.1038/nri3192-c1
82. Tacke M, Hanke G, Hanke T, Hünig T. CD28-mediated induction of proliferation in resting T cells *in vitro* and *in vivo* without engagement of the T cell receptor: evidence for functionally distinct forms of CD28. *Eur J Immunol.* (1997) 27:239–47. doi: 10.1002/eji.1830270136
83. Beyersdorf N, Gaupp S, Balbach K, Schmidt J, Toyka KV, Lin C-H, et al. Selective targeting of regulatory T cells with CD28 superagonists allows effective therapy of experimental autoimmune encephalomyelitis. *J Exp Med.* (2005) 202:445–55. doi: 10.1084/jem.20051060
84. Karnell JL, Rieder SA, Ettinger R, Kolbeck R. Targeting the CD40-CD40L pathway in autoimmune diseases: humoral immunity and beyond. *Adv Drug Deliv Rev.* (2019) 141:92–103. doi: 10.1016/j.addr.2018.12.005
85. Elgueta R, Benson MJ, de Vries VC, Wasiuk A, Guo Y, Noelle RJ. Molecular mechanism and function of CD40/CD40L engagement in the immune system. *Immunol Rev.* (2009) 229:152–72. doi: 10.1111/j.1600-065X.2009.00782.x
86. Langer F, Ingersoll SB, Amirkhosravi A, Meyer T, Siddiqui FA, Ahmad S, et al. The role of CD40 in CD40L- and antibody-mediated platelet activation. *Thromb Haemost.* (2005) 93:1137–46. doi: 10.1160/TH04-12-0774
87. Kawai T, Andrews D, Colvin RB, Sachs DH, Cosimi AB. Thromboembolic complications after treatment with monoclonal antibody against CD40 ligand. *Nat Med.* (2000) 6:114. doi: 10.1038/72162
88. Schuler W, Bigaud M, Brinkmann V, di Padova F, Geisse S, Gram H, et al. Efficacy and safety of ABI793, a novel human anti-human CD154 monoclonal antibody, in cynomolgus monkey renal allotransplantation. *Transplantation.* (2004) 77:717–26. doi: 10.1097/01.TP.0000116563.72763.83
89. Cordoba F, Wiczorek G, Audet M, Roth L, Schneider MA, Kunkler A, et al. A novel, blocking, fc-silent anti-CD40 monoclonal antibody prolongs nonhuman primate renal allograft survival in the absence of B cell depletion. *Am J Transplant.* (2015) 15:2825–36. doi: 10.1111/ajt.13377
90. Farag A, Papas A, Fisher B, Zeher M, Ng W-F, Bombardieri M, et al. Novel anti-cd40 monoclonal antibody CFZ533 in patients with primary sjogren syndrome: a phase iia double-blind, placebo-controlled randomized trial. *Oral Surg Oral Med Oral Pathol Oral Radiol.* (2018) 126:e203–4. doi: 10.1016/j.oooo.2018.05.036
91. Kahaly GJ, Stan MN, Frommer L, Gergely P, Colin L, Amer A, et al. A novel anti-cd40 monoclonal antibody, iscalimab, for control of graves hyperthyroidism—a proof-of-concept trial. *J Clin Endocrinol Metab.* (2020) 105:dgz013. doi: 10.1210/clinem/dgz013
92. Espié P, He Y, Koo P, Sickert D, Dupuy C, Chokoté E, et al. First-in-human clinical trial to assess pharmacokinetics, pharmacodynamics, safety, and tolerability of iscalimab, an anti-CD40 monoclonal antibody. *Am J Transplant.* (2020) 20:463–73. doi: 10.1111/ajt.15661
93. Simpson TR, Quezada SA, Allison JP. Regulation of CD4 T cell activation and effector function by inducible costimulator (ICOS). *Curr Opin Immunol.* (2010) 22:326–32. doi: 10.1016/j.coi.2010.01.001
94. Dong C, Juedes AE, Temann UA, Shresta S, Allison JP, Ruddle NH, et al. ICOS co-stimulatory receptor is essential for T-cell activation and function. *Nature.* (2001) 409:97–101. doi: 10.1038/35051100
95. McAdam AJ, Greenwald RJ, Levin MA, Chernova T, Malenkovich N, Ling V, et al. ICOS is critical for CD40-mediated antibody class switching. *Nature.* (2001) 409:102–5. doi: 10.1038/35051107
96. Tafuri A, Shahinian A, Bladt F, Yoshinaga SK, Jordana M, Wakeham A, et al. ICOS is essential for effective T-helper-cell responses. *Nature.* (2001) 409:105–9. doi: 10.1038/35051113
97. Zhang M, Lee F, Knize A, Jacobsen F, Yu S, Ishida K, et al. Development of an ICOSL and BAFF bispecific inhibitor AMG 570 for systemic lupus erythematosus treatment. *Clin Exp Rheumatol.* (2019) 37:906–14.
98. Joller N, Peters A, Anderson AC, Kuchroo VK. Immune checkpoints in central nervous system autoimmunity. *Immunol Rev.* (2012) 248:122–39. doi: 10.1111/j.1600-065X.2012.01136.x
99. Ford ML, Adams AB, Pearson TC. Targeting co-stimulatory pathways: transplantation and autoimmunity. *Nat Rev Nephrol.* (2013) 10:14–24. doi: 10.1038/nrneph.2013.183
100. Anderson AC, Joller N, Kuchroo VK. Lag-3, Tim-3, and TIGIT: co-inhibitory receptors with specialized functions in immune regulation. *Immunity.* (2016) 44:989–1004. doi: 10.1016/j.immuni.2016.05.001
101. Zhang Q, Vignali DAA. Co-stimulatory and co-inhibitory pathways in autoimmunity. *Immunity.* (2016) 44:1034–51. doi: 10.1016/j.immuni.2016.04.017
102. Paluch C, Santos AM, Anzilotti C, Cornall RJ, Davis SJ. Immune checkpoints as therapeutic targets in autoimmunity. *Front Immunol.* (2018) 9:2306. doi: 10.3389/fimmu.2018.02306
103. Joller N, Lozano E, Burkett PR, Patel B, Xiao S, Zhu C, et al. Treg cells expressing the coinhibitory molecule TIGIT selectively inhibit proinflammatory Th1 and Th17 cell responses. *Immunity.* (2014) 40:569–81. doi: 10.1016/j.immuni.2014.02.012
104. Kurtulus S, Sakuishi K, Ngiew S-F, Joller N, Tan DJ, Teng MWL, et al. TIGIT predominantly regulates the immune response via regulatory T cells. *J Clin Invest.* (2015) 125:4053–62. doi: 10.1172/JCI81187
105. Zhao W, Dong Y, Wu C, Ma Y, Jin Y, Ji Y. TIGIT overexpression diminishes the function of CD4 T cells and ameliorates the severity of rheumatoid arthritis in mouse models. *Exp Cell Res.* (2016) 340:132–8. doi: 10.1016/j.yexcr.2015.12.002
106. Ben Nasr M, Tezza S, D'Addio F, Mameli C, Usulli V, Maestroni A, et al. PD-L1 genetic overexpression or pharmacological restoration in hematopoietic stem and progenitor cells reverses autoimmune diabetes. *Sci Transl Med.* (2017) 9:eaam7543. doi: 10.1126/scitranslmed.aam7543
107. Le Mercier I, Lines JL, Noelle RJ. Beyond CTLA-4 and PD-1, the generation z of negative checkpoint regulators. *Front Immunol.* (2015) 6:418. doi: 10.3389/fimmu.2015.00418
108. Joller N, Kuchroo VK. Tim-3, Lag-3, and TIGIT. *Curr Top Microbiol Immunol.* (2017) 410:127–56. doi: 10.1007/82_2017_62
109. Joller N, Hafler JP, Brynedal B, Kassam N, Spoerl S, Levin SD, et al. Cutting edge: TIGIT Has T Cell-intrinsic inhibitory functions. *J Immunol.* (2011) 186:1338–42. doi: 10.4049/jimmunol.1003081
110. Lozano E, Dominguez-Villar M, Kuchroo V, Hafler DA. The TIGIT/CD226 axis regulates human T cell function. *J Immunol.* (2012) 188:3869–75. doi: 10.4049/jimmunol.1103627
111. Dixon KO, Schorer M, Nevin J, Etninan Y, Amoozgar Z, Kondo T, et al. Functional Anti-TIGIT antibodies regulate development of autoimmunity and antitumor immunity. *J Immunol.* (2018) 200:3000–7. doi: 10.4049/jimmunol.1700407
112. Riquelme P, Haarer J, Kammler A, Walter L, Tomiuk S, Ahrens N, et al. TIGIT+ iTregs elicited by human regulatory macrophages control T cell immunity. *Nat Commun.* (2018) 9:2858. doi: 10.1038/s41467-018-05167-8
113. Wang FF, Wang Y, Wang L, Wang TS, Bai YP. TIGIT expression levels on CD4+ T cells are correlated with disease severity in patients with psoriasis. *Clin Exp Dermatol.* (2018) 43:675–82. doi: 10.1111/ced.13414
114. Lucca LE, Axisa P-P, Singer ER, Nolan NM, Dominguez-Villar M, Hafler DA. TIGIT signaling restores suppressor function of Th1 Tregs. *JCI Insight.* (2019) 4:3263. doi: 10.1172/jci.insight.124427

115. Wang L, Rubinstein R, Lines JL, Wasiek A, Ahonen C, Guo Y, et al. VISTA, a novel mouse Ig superfamily ligand that negatively regulates T cell responses. *J Exp Med.* (2011) 208:577–92. doi: 10.1084/jem.20100619
116. Lines JL, Pantazi E, Mak J, Sempere LF, Wang L, O'Connell S, et al. VISTA is an immune checkpoint molecule for human T cells. *Cancer Res.* (2014) 74:1924–32. doi: 10.1158/0008-5472.CAN-13-1504
117. Linnik M, Godzik A, Jaroszewski L, Ware C, Vendel A. Clinical evidence supporting therapeutic potential of activating the immune checkpoint receptor BTLA in SLE - ACR meeting abstracts. *Arthritis Rheumatol.* (2019) 71:A2576.
118. Triebel F, Jitsukawa S, Baixeras E, Roman-Roman S, Genevec C, Viegas-Pequignot E, et al. LAG-3, a novel lymphocyte activation gene closely related to CD4. *J Exp Med.* (1990) 171:1393–405. doi: 10.1084/jem.171.5.1393
119. Huard B, Tournier M, Hercend T, Triebel F, Faure F. Lymphocyte-activation gene 3/major histocompatibility complex class II interaction modulates the antigenic response of CD4+ T lymphocytes. *Eur J Immunol.* (1994) 24:3216–21. doi: 10.1002/eji.1830241246
120. Wang J, Sanmamed MF, Datar I, Su TT, Ji L, Sun J, et al. Fibrinogen-like protein 1 is a major immune inhibitory ligand of LAG-3. *Cell.* (2019) 176:334–47.e12. doi: 10.1016/j.cell.2018.11.010
121. Blackburn SD, Shin H, Haining WN, Zou T, Workman CJ, Polley A, et al. Coregulation of CD8+ T cell exhaustion by multiple inhibitory receptors during chronic viral infection. *Nat Immunol.* (2009) 10:29–37. doi: 10.1038/ni.1679
122. Workman CJ, Dugger KJ, Vignali DAA. Cutting edge: molecular analysis of the negative regulatory function of lymphocyte activation gene-3. *J Immunol.* (2002) 169:5392–5. doi: 10.4049/jimmunol.169.10.5392
123. Huang C-T, Workman CJ, Flies D, Pan X, Marson AL, Zhou G, et al. Role of LAG-3 in regulatory T cells. *Immunity.* (2004) 21:503–13. doi: 10.1016/j.immuni.2004.08.010
124. Stengel KF, Harden-Bowles K, Yu X, Rouge L, Yin J, Comps-Agrar L, et al. Structure of TIGIT immunoreceptor bound to poliovirus receptor reveals a cell-cell adhesion and signaling mechanism that requires cis-trans receptor clustering. *Proc Natl Acad Sci USA.* (2012) 109:5399–404. doi: 10.1073/pnas.1120606109
125. Meyer D, Seth S, Albrecht J, Maier MK, Pasquier du L, Ravens I, et al. CD96 interaction with CD155 via its first Ig-like domain is modulated by alternative splicing or mutations in distal Ig-like domains. *J Biol Chem.* (2009) 284:2235–44. doi: 10.1074/jbc.M807698200
126. Tahara-Hanaoka S. Functional characterization of DNAM-1 (CD226) interaction with its ligands PVR (CD155) and nectin-2 (PRR-2/CD112). *INTIMM.* (2004) 16:533–8. doi: 10.1093/intimm/dxh059
127. Dougall WC, Kurtulus S, Smyth MJ, Anderson AC. TIGIT and CD96: new checkpoint receptor targets for cancer immunotherapy. *Immunol Rev.* (2017) 276:112–20. doi: 10.1111/immr.12518
128. Zhu Y, Paniccia A, Schulick AC, Chen W, Koenig MR, Byers JT, et al. Identification of CD112R as a novel checkpoint for human T cells. *J Exp Med.* (2016) 213:167–76. doi: 10.1084/jem.20150785
129. Mahnke YD, Brodie TM, Sallusto F, Roederer M, Lugli E. The who's who of T-cell differentiation: human memory T-cell subsets. *Eur J Immunol.* (2013) 43:2797–809. doi: 10.1002/eji.201343751
130. Mou D, Espinosa J, Lo DJ, Kirk AD. CD28 negative T cells: is their loss our gain? *Am J Transplant.* (2014) 14:2460–6. doi: 10.1111/ajt.12937
131. Monney L, Sabatos CA, Gaglia JL, Ryu A, Waldner H, Chernova T, et al. Th1-specific cell surface protein Tim-3 regulates macrophage activation and severity of an autoimmune disease. *Nature.* (2002) 415:536–41. doi: 10.1038/415536a
132. Anderson AC, Anderson DE. TIM-3 in autoimmunity. *Curr Opin Immunol.* (2006) 18:665–9. doi: 10.1016/j.coi.2006.09.009
133. Zhu C, Anderson AC, Schubart A, Xiong H, Imitola J, Khoury SJ, et al. The Tim-3 ligand galectin-9 negatively regulates T helper type 1 immunity. *Nat Immunol.* (2005) 6:1245–52. doi: 10.1038/ni1271
134. Chou F-C, Chen H-Y, Kuo C-C, Sytwu H-K. Role of galectins in tumors and in clinical immunotherapy. *Int J Mol Sci.* (2018) 19:430. doi: 10.3390/ijms19020430
135. Koguchi K, Anderson DE, Yang L, O'Connor KC, Kuchroo VK, Hafler DA. Dysregulated T cell expression of TIM3 in multiple sclerosis. *J Exp Med.* (2006) 203:1413–8. doi: 10.1084/jem.20060210
136. Yang L, Anderson DE, Kuchroo J, Hafler DA. Lack of TIM-3 immunoregulation in multiple sclerosis. *J Immunol.* (2008) 180:4409–14. doi: 10.4049/jimmunol.180.7.4409
137. Liu Y, Shu Q, Gao L, Hou N, Zhao D, Liu X, et al. Increased Tim-3 expression on peripheral lymphocytes from patients with rheumatoid arthritis negatively correlates with disease activity. *Clin Immunol.* (2010) 137:288–95. doi: 10.1016/j.clim.2010.07.012
138. Kanai Y, Satoh T, Igawa K, Yokozeki H. Impaired expression of Tim-3 on Th17 and Th1 cells in psoriasis. *Acta Derm Venereol.* (2012) 92:367–71. doi: 10.2340/00015555-1285
139. Nowak EC, Lines JL, Varn FS, Deng J, Sarde A, Mabaera R, et al. Immunoregulatory functions of VISTA. *Immunol Rev.* (2017) 276:66–79. doi: 10.1111/immr.12525
140. Zhang M, Pang H-J, Zhao W, Li Y-F, Yan L-X, Dong Z-Y, et al. VISTA expression associated with CD8 confers a favorable immune microenvironment and better overall survival in hepatocellular carcinoma. *BMC Cancer.* (2018) 18:511. doi: 10.1186/s12885-018-4435-1
141. Yang W, Padkjær SB, Wang J, Sun Z, Shan B, Yang L, et al. Construction of a versatile expression library for all human single-pass transmembrane proteins for receptor pairings by high throughput screening. *J Biotechnol.* (2017) 260:18–30. doi: 10.1016/j.jbiotec.2017.08.023
142. Wang J, Wu G, Manick B, Hernandez V, Renelt M, Erickson C, et al. VSIG-3 as a ligand of VISTA inhibits human T-cell function. *Immunology.* (2019) 156:74–85. doi: 10.1111/imm.13001
143. Sergeant PA, Plummer SF, Pettus J, Mabaera R, DeLong JK, Pechenick DA, et al. Blocking the VISTA pathway enhances disease progression in (NZB × NZW) F1 female mice. *Lupus.* (2018) 27:210–6. doi: 10.1177/0961203317716322
144. Han X, Vesely MD, Yang W, Sanmamed MF, Badri T, Alawa J, et al. PD-1H (VISTA)-mediated suppression of autoimmunity in systemic and cutaneous lupus erythematosus. *Sci Transl Med.* (2019) 11:eaax1159. doi: 10.1126/scitranslmed.aax1159
145. Sharabi A, Tsokos MG, Ding Y, Malek TR, Klatzmann D, Tsokos GC. Regulatory T cells in the treatment of disease. *Nat Rev Drug Discov.* (2018) 17:823–44. doi: 10.1038/nrd.2018.148
146. Ferreira LMR, Muller YD, Bluestone JA, Tang Q. Next-generation regulatory T cell therapy. *Nat Rev Drug Discov.* (2019) 18:749–69. doi: 10.1038/s41573-019-0041-4
147. Campbell C, Rudensky A. Roles of regulatory T cells in tissue pathophysiology and metabolism. *Cell Metab.* (2020) 31:18–25. doi: 10.1016/j.cmet.2019.09.010
148. Levings MK, Sangregorio R, Galbiati F, Squadrone S, de Waal Malefyt R, Roncarolo MG. IFN- α and IL-10 induce the differentiation of human type 1 T regulatory cells. *J Immunol.* (2001) 166:5530–9. doi: 10.4049/jimmunol.166.9.5530
149. Roncarolo M-G, Bacchetta R, Bordignon C, Narula S, Levings MK. Type 1 T regulatory cells. *Immunol Rev.* (2001) 182:68–79. doi: 10.1034/j.1600-065X.2001.1820105.x
150. Levings MK, Sangregorio R, Sartirana C, Moschin AL, Battaglia M, Orban PC, et al. Human CD25+CD4+ T suppressor cell clones produce transforming growth factor beta, but not interleukin 10, and are distinct from type 1 T regulatory cells. *J Exp Med.* (2002) 196:1335–46. doi: 10.1084/jem.20021139
151. Roncarolo MG, Bacchetta R. T cell repertoire and tolerance after fetal stem cell transplantation. *Bone Marrow Transplant.* (1992) 9(Suppl 1):127–8.
152. Bacchetta R, Bigler M, Touraine JL, Parkman R, Tovo PA, Abrams J, et al. High levels of interleukin 10 production *in vivo* are associated with tolerance in SCID patients transplanted with HLA mismatched hematopoietic stem cells. *J Exp Med.* (1994) 179:493–502. doi: 10.1084/jem.179.2.493
153. Groux H, Bigler M, de Vries JE, Roncarolo MG. Interleukin-10 induces a long-term antigen-specific anergic state in human CD4+ T cells. *J Exp Med.* (1996) 184:19–29. doi: 10.1084/jem.184.1.19
154. Battaglia M, Stabellini A, Draghici E, Migliaiava B, Gregori S, Bonifacio E, et al. Induction of tolerance in type 1 diabetes via both CD4+CD25+ T regulatory cells and T regulatory type 1 cells. *Diabetes.* (2006) 55:1571–80. doi: 10.2337/db05-1576

155. Gianfrani C, Levings MK, Sartirana C, Mazzarella G, Barba G, Zanzi D, et al. Gliadin-specific type 1 regulatory T cells from the intestinal mucosa of treated celiac patients inhibit pathogenic T cells. *J Immunol.* (2006) 177:4178–86. doi: 10.4049/jimmunol.177.6.4178
156. Roncarolo M-G, Gregori S, Bacchetta R, Battaglia M, Gagliani N. The Biology of T Regulatory type 1 cells and their therapeutic application in immune-mediated diseases. *Immunity.* (2018) 49:1004–19. doi: 10.1016/j.immuni.2018.12.001
157. Bluestone JA, Buckner JH, Fitch M, Gitelman SE, Gupta S, Hellerstein MK, et al. Type 1 diabetes immunotherapy using polyclonal regulatory T cells. *Sci Transl Med.* (2015) 7:315ra189. doi: 10.1126/scitranslmed.aad4134
158. Marek-Trzonkowska N, Mysliwiec M, Iwazkiewicz-Grzes D, Gliwinski M, Derkowska I, Zalinska M, Zielinski M, et al. Factors affecting long-term efficacy of T regulatory cell-based therapy in type 1 diabetes. *J Transl Med.* (2016) 14:332–11. doi: 10.1186/s12967-016-1090-7
159. Romano M, Fanelli G, Albany CJ, Giganti G, Lombardi G. Past, present, and future of regulatory T cell therapy in transplantation and autoimmunity. *Front Immunol.* (2019) 10:43. doi: 10.3389/fimmu.2019.00043
160. Abbas AK, Trotta E, R Simeonov D, Marson A, Bluestone JA. Revisiting IL-2: biology and therapeutic prospects. *Sci Immunol.* (2018) 3:eaat1482. doi: 10.1126/sciimmunol.aat1482
161. Boyman O, Kovar M, Rubinstein MP, Surh CD, Sprent J. Selective stimulation of T cell subsets with antibody-cytokine immune complexes. *Science.* (2006) 311:1924–7. doi: 10.1126/science.1122927
162. Akamatsu M, Mikami N, Ohkura N, Kawakami R, Kitagawa Y, Sugimoto A, et al. Conversion of antigen-specific effector/memory T cells into Foxp3-expressing Treg cells by inhibition of CDK8/19. *Sci Immunol.* (2019) 4:eaaw2707. doi: 10.1126/sciimmunol.aaw2707
163. Bacchetta R, Lucarelli B, Sartirana C, Gregori S, Lupo Stanghellini MT, Miqueu P, et al. Immunological outcome in haploidentical-HSC transplanted patients treated with IL-10-energized donor t cells. *Front Immunol.* (2014) 5:16. doi: 10.3389/fimmu.2014.00016
164. Desreumaux P, Foussat A, Allez M, Beaugerie L, Hébuterne X, Bouhnik Y, et al. Safety and efficacy of antigen-specific regulatory T-cell therapy for patients with refractory Crohn's disease. *Gastroenterology.* (2012) 143:1207–17.e1–2. doi: 10.1053/j.gastro.2012.07.116
165. Geissler EK. The ONE Study compares cell therapy products in organ transplantation: introduction to a review series on suppressive monocyte-derived cells. *Transplant Res.* (2012) 1:1–3. doi: 10.1186/2047-1440-1-11
166. Mfarrej B, Tresoldi E, Stabilini A, Paganelli A, Caldara R, Secchi A, et al. Generation of donor-specific Tr1 cells to be used after kidney transplantation and definition of the timing of their *in vivo* infusion in the presence of immunosuppression. *J Transl Med.* (2017) 15:1–12. doi: 10.1186/s12967-017-1133-8
167. Andolfi G, Foustieri G, Rossetti M, Magnani CF, Jofra T, Locafaro G, et al. Enforced IL-10 expression confers type 1 regulatory T cell (Tr1) phenotype and function to human CD4(+) T cells. *Mol Ther.* (2012) 20:1778–90. doi: 10.1038/mt.2012.71
168. Gregori S, Roncarolo M-G. Engineered T regulatory type 1 cells for clinical application. *Front Immunol.* (2018) 9:233. doi: 10.3389/fimmu.2018.00233
169. Elinav E, Waks T, Eshhar Z. Redirection of regulatory T Cells with predetermined specificity for the treatment of experimental colitis in mice. *Gastroenterology.* (2008) 134:2014–24. doi: 10.1053/j.gastro.2008.02.060
170. Elinav E, Adam N, Waks T, Eshhar Z. Amelioration of colitis by genetically engineered murine regulatory T cells redirected by antigen-specific chimeric receptor. *Gastroenterology.* (2009) 136:1721–31. doi: 10.1053/j.gastro.2009.01.049
171. Fransson M, Piras E, Burman J, Nilsson B, Essand M, Lu B, et al. CAR/FoxP3-engineered T regulatory cells target the CNS and suppress EAE upon intranasal delivery. *J Neuroinflammation.* (2012) 9:112. doi: 10.1186/1742-2094-9-112
172. Zhang Q, Lu W, Liang C-L, Chen Y, Liu H, Qiu F, et al. Chimeric Antigen Receptor (CAR) treg: a promising approach to inducing immunological tolerance. *Front Immunol.* (2018) 9:2359. doi: 10.3389/fimmu.2018.02359
173. MacDonald KG, Hoeppli RE, Huang Q, Gillies J, Luciani DS, Orban PC, et al. Alloantigen-specific regulatory T cells generated with a chimeric antigen receptor. *J Clin Invest.* (2016) 126:1413–24. doi: 10.1172/JCI82771
174. Noyan F, Zimmermann K, Hardtke-Wolenski M, Knoefel A, Schulte E, Geffers R, et al. Prevention of allograft rejection by use of regulatory T cells with an MHC-Specific chimeric antigen receptor. *Am J Transplant.* (2017) 17:917–30. doi: 10.1111/ajt.14175
175. Boardman DA, Philippos C, Fruhwirth GO, Ibrahim MAA, Hannen RF, Cooper D, et al. Expression of a chimeric antigen receptor specific for donor HLA class I enhances the potency of human regulatory t cells in preventing human skin transplant rejection. *Am J Transplant.* (2017) 17:931–43. doi: 10.1111/ajt.14185
176. Pierini A, Iliopoulou BP, Peiris H, Pérez-Cruz M, Baker J, Hsu K, et al. T cells expressing chimeric antigen receptor promote immune tolerance. *JCI Insight.* (2017) 2:e92865. doi: 10.1172/jci.insight.92865
177. Edry E, Melamed D. Receptor editing in positive and negative selection of B lymphopoiesis. *J Immunol.* (2004) 173:4265–71. doi: 10.4049/jimmunol.173.7.4265
178. McCaughy TM, Wilken MS, Hogquist KA. Thymic emigration revisited. *J Exp Med.* (2007) 204:2513–20. doi: 10.1084/jem.20070601
179. Yu W, Jiang N, Ebert PJR, Kidd BA, Müller S, Lund PJ, et al. Clonal deletion prunes but does not eliminate self-specific $\alpha\beta$ CD8(+) T Lymphocytes. *Immunity.* (2015) 42:929–41. doi: 10.1016/j.immuni.2015.05.001
180. Sakaguchi S, Sakaguchi N, Asano M, Itoh M, Toda M. Immunologic self-tolerance maintained by activated T cells expressing IL-2 receptor alpha-chains (CD25). Breakdown of a single mechanism of self-tolerance causes various autoimmune diseases. *J Immunol.* (1995) 155:1151–64.
181. Rice JS, Newman J, Wang C, Michael DJ, Diamond B. Receptor editing in peripheral B cell tolerance. *Proc Natl Acad Sci USA.* (2005) 102:1608–13. doi: 10.1073/pnas.0409217102
182. Mackay IR, Leskovsek NV, Rose NR. Cell damage and autoimmunity: a critical appraisal. *J Autoimmun.* (2008) 30:5–11. doi: 10.1016/j.jaut.2007.11.009
183. Carballido JM, Santamaria P. Taming autoimmunity: translating antigen-specific approaches to induce immune tolerance. *J Exp Med.* (2019) 216:247–50. doi: 10.1084/jem.20182287
184. Swart JF, Delemarre EM, van Wijk F, Boelens J-J, Kuball J, van Laar JM, et al. Haematopoietic stem cell transplantation for autoimmune diseases. *Nat Rev Rheumatol.* (2017) 13:244–56. doi: 10.1038/nrrheum.2017.7
185. Malmegrim KCR, Lima-Júnior JR, Arruda LCM, de Azevedo JTC, de Oliveira GLV, Oliveira MC. Autologous hematopoietic stem cell transplantation for autoimmune diseases: from mechanistic insights to biomarkers. *Front Immunol.* (2018) 9:2602. doi: 10.3389/fimmu.2018.02602
186. Roubenoff R, Jones RJ, Karp JE, Stevens MB. Remission of rheumatoid arthritis with the successful treatment of acute myelogenous leukemia with cytosine arabinoside, daunorubicin, and m-AMSA. *Arthritis Rheum.* (1987) 30:1187–90. doi: 10.1002/art.1780301017
187. Eedy DJ, Burrows D, Bridges JM, Jones FG. Clearance of severe psoriasis after allogeneic bone marrow transplantation. *BMJ.* (1990) 300:908. doi: 10.1136/bmj.300.6729.908
188. Atkins HL, Bowman M, Allan D, Anstee G, Arnold DL, Bar-Or A, et al. Immunoablation and autologous haemopoietic stem-cell transplantation for aggressive multiple sclerosis: a multicentre single-group phase 2 trial. *Lancet.* (2016) 388:576–85. doi: 10.1016/S0140-6736(16)30169-6
189. Bryant A, Atkins H, Pringle CE, Allan D, Anstee G, Bence-Bruckler I, et al. Myasthenia gravis treated with autologous hematopoietic stem cell transplantation. *JAMA Neurol.* (2016) 73:652–8. doi: 10.1001/jamaneurol.2016.0113
190. Serra P, Santamaria P. Antigen-specific therapeutic approaches for autoimmunity. *Nat Biotechnol.* (2019) 37:238–51. doi: 10.1038/s41587-019-0015-4
191. Lorentz KM, Kontos S, Diaceri G, Henry H, Hubbell JA. Engineered binding to erythrocytes induces immunological tolerance to *E. coli asparaginase*. *Sci Adv.* (2015) 1:e1500112. doi: 10.1126/sciadv.1500112
192. Lehmann PV, Forsthuber T, Miller A, Sercarz EE. Spreading of T-cell autoimmunity to cryptic determinants of an autoantigen. *Nature.* (1992) 358:155–7. doi: 10.1038/358155a0
193. McRae BL, Vanderlugt CL, Dal Canto MC, Miller SD. Functional evidence for epitope spreading in the relapsing pathology of experimental autoimmune encephalomyelitis. *J Exp Med.* (1995) 182:75–85. doi: 10.1084/jem.182.1.75

194. Yu M, Johnson JM, Tuohy VK. A predictable sequential determinant spreading cascade invariably accompanies progression of experimental autoimmune encephalomyelitis: a basis for peptide-specific therapy after onset of clinical disease. *J Exp Med.* (1996) 183:1777–88. doi: 10.1084/jem.183.4.1777
195. Tuohy VK, Yu M, Weinstock-Guttman B, Kinkel RP. Diversity and plasticity of self recognition during the development of multiple sclerosis. *J Clin Invest.* (1997) 99:1682–90. doi: 10.1172/JCI119331
196. Moudgil KD, Sercarz EE. Crypticity of self antigenic determinants is the cornerstone of a theory of autoimmunity. *Discov Med.* (2005) 5:378–82.
197. Vanderlugt CL, Miller SD. Epitope spreading in immune-mediated diseases: implications for immunotherapy. *Nat Rev Immunol.* (2002) 2:85–95. doi: 10.1038/nri724
198. Ziegler A-G, Nepom GT. Prediction and pathogenesis in type 1 diabetes. *Immunity.* (2010) 32:468–78. doi: 10.1016/j.immuni.2010.03.018
199. James EA, Pietropaolo M, Mamula MJ. Immune recognition of β -Cells: neopeptides as key players in the loss of tolerance. *Diabetes.* (2018) 67:1035–42. doi: 10.2337/dbi17-0030
200. Wraith D. Antigen-specific immunotherapy. *Nature.* (2016) 530:422–3. doi: 10.1038/nature17300
201. Herndon JM, Stuart PM, Ferguson TA. Peripheral deletion of antigen-specific T cells leads to long-term tolerance mediated by CD8+ cytotoxic cells. *J Immunol.* (2005) 174:4098–104. doi: 10.4049/jimmunol.174.7.4098
202. Gurung P, Kucaba TA, Schoenberger SP, Ferguson TA, Griffith TS. TRAIL-expressing CD8+ T cells mediate tolerance following soluble peptide-induced peripheral T cell deletion. *J Leukoc Biol.* (2010) 88:1217–25. doi: 10.1189/jlb.0610343
203. Miller SD, Wetzig RP, Claman HN. The induction of cell-mediated immunity and tolerance with protein antigens coupled to syngeneic lymphoid cells. *J Exp Med.* (1979) 149:758–73. doi: 10.1084/jem.149.3.758
204. Miller SD, Turley DM, Podojil JR. Antigen-specific tolerance strategies for the prevention and treatment of autoimmune disease. *Nat Rev Immunol.* (2007) 7:665–77. doi: 10.1038/nri2153
205. Lutterotti A, Yousef S, Sputek A, Stürner KH, Stellmann J-P, Breiden P, et al. Antigen-specific tolerance by autologous myelin peptide-coupled cells: a phase I trial in multiple sclerosis. *Sci Transl Med.* (2013) 5:188ra75. doi: 10.1126/scitranslmed.3006168
206. Lutterotti A, Ludersdorfer TH, Docampo MJ, Hohmann M, Selles Moreno C, Hayward-Koennecke H, et al. ECTRIMS 2019 – late breaking news abstracts. *Mult Scler.* (2019) 25:890–938. doi: 10.1177/1352458519869496
207. Pishesha N, Bilate AM, Wibowo MC, Huang N-J, Li Z, Deshycka R, et al. Engineered erythrocytes covalently linked to antigenic peptides can protect against autoimmune disease. *Proc Natl Acad Sci USA.* (2017) 114:3157–62. doi: 10.1073/pnas.1701746114
208. Ramakrishnan A, Lee T, Moore LJ, Suarez AV, Moore F, Bernstein H. 1743-P: engineering erythrocytes with the SQZ cell therapy platform to enhance immunotolerance. *Diabetes.* (2019) 68(Suppl. 1):1743–P. doi: 10.2337/db19-1743-P
209. Lang F, Lang E, Föller M. Physiology and pathophysiology of eryptosis. *Transfus Med Hemother.* (2012) 39:308–14. doi: 10.1159/000342534
210. Cremel M, Guérin N, Horand F, Banz A, Godfrin Y. Red blood cells as innovative antigen carrier to induce specific immune tolerance. *Int J Pharm.* (2013) 443:39–49. doi: 10.1016/j.ijpharm.2012.12.044
211. Lang F, Gulbins E, Lang PA, Zappulla D, Föller M. Ceramide in suicidal death of erythrocytes. *Cell Physiol Biochem.* (2010) 26:21–8. doi: 10.1159/000315102
212. Pujol-Autonell I, Serracant-Prat A, Cano-Sarabia M, Ampudia RM, Rodriguez-Fernandez S, Sanchez A, et al. Use of autoantigen-loaded phosphatidylserine-liposomes to arrest autoimmunity in type 1 diabetes. *PLoS ONE.* (2015) 10:e0127057. doi: 10.1371/journal.pone.0127057
213. Rodriguez-Fernandez S, Pujol-Autonell I, Brioso F, Perna-Barrull D, Cano-Sarabia M, Garcia-Jimeno S, et al. Phosphatidylserine-liposomes promote tolerogenic features on dendritic cells in human type 1 diabetes by apoptotic mimicry. *Front Immunol.* (2018) 9:253. doi: 10.3389/fimmu.2018.00253
214. Hunter Z, McCarthy DP, Yap WT, Harp CT, Getts DR, Shea LD, et al. A biodegradable nanoparticle platform for the induction of antigen-specific immune tolerance for treatment of autoimmune disease. *ACS nano.* (2014) 8:2148–60. doi: 10.1021/nn405033r
215. Prasad S, Neef T, Xu D, Podojil JR, Getts DR, Shea LD, et al. Tolerogenic Ag-PLG nanoparticles induce tregs to suppress activated diabetogenic CD4 and CD8 T cells. *J Autoimmun.* (2018) 89:112–24. doi: 10.1016/j.jaut.2017.12.010
216. Jamison BL, Neef T, Goodspeed A, Bradley B, Baker RL, Miller SD, et al. Nanoparticles containing an insulin-ChgA hybrid peptide protect from transfer of autoimmune diabetes by shifting the balance between effector T Cells and regulatory T Cells. *J Immunol.* (2019) 203:48–57. doi: 10.4049/jimmunol.1900127
217. Arredouani MS, Palecanda A, Koziel H, Huang Y-C, Imrich A, Sulahian TH, et al. MARCO is the major binding receptor for unopsonized particles and bacteria on human alveolar macrophages. *J Immunol.* (2005) 175:6058–64. doi: 10.4049/jimmunol.175.9.6058
218. Getts DR, Martin AJ, McCarthy DP, Terry RL, Hunter ZN, Yap WT, et al. Microparticles bearing encephalitogenic peptides induce T-cell tolerance and ameliorate experimental autoimmune encephalomyelitis. *Nat Biotechnol.* (2012) 30:1217–24. doi: 10.1038/nbt.2434
219. Kontos S, Kourits IC, Dane KY, Hubbell JA. Engineering antigens for *in situ* erythrocyte binding induces T-cell deletion. *Proc Natl Acad Sci USA.* (2013) 110:E60–8. doi: 10.1073/pnas.1216353110
220. Grimm AJ, Kontos S, Diaceri G, Quaglia-Thermes X, Hubbell JA. Memory of tolerance and induction of regulatory T cells by erythrocyte-targeted antigens. *Sci Rep.* (2015) 5:15907. doi: 10.1038/srep15907
221. Roep BO, Solvason N, Gottlieb PA, Abreu JRF, Harrison LC, Eisenbarth GS, et al. Plasmid-encoded proinsulin preserves C-peptide while specifically reducing proinsulin-specific CD8⁺ T cells in type 1 diabetes. *Sci Transl Med.* (2013) 5:191ra82. doi: 10.1126/scitranslmed.3006103
222. Bar-Or A, Vollmer T, Antel J, Arnold DL, Bodner CA, Campagnolo D, et al. Induction of antigen-specific tolerance in multiple sclerosis after immunization with DNA encoding myelin basic protein in a randomized, placebo-controlled phase 1/2 trial. *Arch Neurol.* (2007) 64:1407–15. doi: 10.1001/archneur.64.10.nct70002
223. Ludvigsson J, Faresjö M, Hjorth M, Axelsson S, Chéramy M, Pihl M, et al. GAD treatment and insulin secretion in recent-onset type 1 diabetes. *N Engl J Med.* (2008) 359:1909–20. doi: 10.1056/NEJMoa0804328
224. Ludvigsson J, Chéramy M, Axelsson S, Pihl M, Akerman L, Casas R, et al. GAD-treatment of children and adolescents with recent-onset type 1 diabetes preserves residual insulin secretion after 30 months. *Diabetes Metab Res Rev.* (2014) 30:405–14. doi: 10.1002/dmrr.2503
225. Streeter HB, Rigden R, Martin KF, Scolding NJ, Wraith DC. Preclinical development and first-in-human study of ATX-MS-1467 for immunotherapy of MS. *Neural Neuroimmunol Neuroinflamm.* (2015) 2:e93. doi: 10.1212/NXL.0000000000000093
226. Chataway J, Martin K, Barrell K, Sharrack B, Stolt P, Wraith DC, et al. Effects of ATX-MS-1467 immunotherapy over 16 weeks in relapsing multiple sclerosis. *Neurology.* (2018) 90:e955–62. doi: 10.1212/WNL.00000000000005118
227. Goel G, King T, Daveson AJ, Andrews JM, Krishnarajah J, Krause R, et al. Epitope-specific immunotherapy targeting CD4-positive T cells in coeliac disease: two randomised, double-blind, placebo-controlled phase 1 studies. *Lancet Gastroenterol Hepatol.* (2017) 2:479–93. doi: 10.1016/S2468-1253(17)30110-3
228. Carlier VA, VanderElst L, Janssens W, Jacquemin MG, Saint-Remy J-MR. Increased synapse formation obtained by T cell epitopes containing a CxxC motif in flanking residues convert CD4+ T cells into cytolytic effectors. *PLoS ONE.* (2012) 7:e45366. doi: 10.1371/journal.pone.0045366
229. Turnquist HR, Raimondi G, Zahorchak AF, Fischer RT, Wang Z, Thomson AW. Rapamycin-conditioned dendritic cells are poor stimulators of allogeneic CD4+ T cells, but enrich for antigen-specific Foxp3+ T regulatory cells and promote organ transplant tolerance. *J Immunol.* (2007) 178:7018–31. doi: 10.4049/jimmunol.178.11.7018
230. Hauben E, Gregori S, Draghici E, Migliavacca B, Olivieri S, Woisetschlager M, et al. Activation of the aryl hydrocarbon receptor promotes allograft-specific tolerance through direct and dendritic cell-mediated effects on regulatory T cells. *Blood.* (2008) 112:1214–22. doi: 10.1182/blood-2007-08-109843
231. Quintana FJ, Murugaiyan G, Farez MF, Mitsdoerffer M, Tukupah A-M, Burns EJ, et al. An endogenous aryl hydrocarbon receptor ligand acts on dendritic

- cells and T cells to suppress experimental autoimmune encephalomyelitis. *Proc Natl Acad Sci USA*. (2010) 107:20768–73. doi: 10.1073/pnas.1009201107
232. Nguyen NT, Kimura A, Nakahama T, Chinen I, Masuda K, Nohara K, et al. Aryl hydrocarbon receptor negatively regulates dendritic cell immunogenicity via a kynurenine-dependent mechanism. *Proc Natl Acad Sci USA*. (2010) 107:19961. doi: 10.1073/pnas.1014465107
 233. Giannoukakis N, Bonham CA, Qian S, Zhou Z, Peng L, Harnaha J, et al. Prolongation of cardiac allograft survival using dendritic cells treated with NF- κ B decoy oligodeoxyribonucleotides. *Mol Ther*. (2000) 1:430–7. doi: 10.1006/mthe.2000.0060
 234. Yang J, Bernier SM, Ichim TE, Li M, Xia X, Zhou D, et al. LF15-0195 generates tolerogenic dendritic cells by suppression of NF- κ B signaling through inhibition of IKK activity. *J Leukoc Biol*. (2003) 74:438–47. doi: 10.1189/jlb.1102582
 235. Kishimoto TK, Ferrari JD, LaMothe RA, Kolte PN, Griset AP, O'Neil C, et al. Improving the efficacy and safety of biologic drugs with tolerogenic nanoparticles. *Nat Nanotech*. (2016) 11:890–9. doi: 10.1038/nnano.2016.135
 236. Maldonado RA, LaMothe R, Ferrari D, Griset A, O'Neil C, Johnston L, et al. Inducing immunological tolerance with synthetic nanoparticles. *J Immunol*. (2017) 198:73.10.
 237. LaMothe RA, Kolte PN, Vo T, Ferrari JD, Gelsing TC, Wong J, et al. Tolerogenic nanoparticles induce antigen-specific regulatory T Cells and provide therapeutic efficacy and transferrable tolerance against experimental autoimmune encephalomyelitis. *Front Immunol*. (2018) 9:281. doi: 10.3389/fimmu.2018.00281
 238. Meliani A, Boisgerault F, Hardet R, Marmier S, Collaud F, Ronzitti G, et al. Antigen-selective modulation of AAV immunogenicity with tolerogenic rapamycin nanoparticles enables successful vector re-administration. *Nat Commun*. (2018) 9:4098–13. doi: 10.1038/s41467-018-06621-3
 239. Kishimoto TK, Maldonado RA. Nanoparticles for the induction of antigen-specific immunological tolerance. *Front Immunol*. (2018) 9:230. doi: 10.3389/fimmu.2018.00230
 240. Yeste A, Nadeau M, Burns EJ, Weiner HL, Quintana FJ. Nanoparticle-mediated codelivery of myelin antigen and a tolerogenic small molecule suppresses experimental autoimmune encephalomyelitis. *Proc Natl Acad Sci USA*. (2012) 109:11270–5. doi: 10.1073/pnas.1120611109
 241. Yeste A, Takenaka MC, Mascanfroni ID, Nadeau M, Kenison JE, Patel B, et al. Tolerogenic nanoparticles inhibit T cell-mediated autoimmunity through SOCS2. *Sci Signal*. (2016) 9:ra61. doi: 10.1126/scisignal.aad0612
 242. Kenison-White J, Jhaveri A, Tezza S, Nowakowska D, Sherr D, Quintana FJ. The aryl hydrocarbon receptor as a regulator of CNS autoimmunity. *J Immunol*. (2019) 202(1 Suppl.):193.13.
 243. Yekollu SK, Thomas R, O'Sullivan B. Targeting curcumin to inflammatory dendritic cells inhibits NF- κ B and improves insulin resistance in obese mice. *Diabetes*. (2011) 60:2928–38. doi: 10.2337/db11-0275
 244. Sonigra A, Nel H, Ramnoruth N, Talekar M, Tesiram J, Stuurman F, et al. A phase I, randomized, double-blind, placebo-controlled, single center, single-dose escalation to investigate the safety, tolerability, and pharmacodynamics of subcutaneously administered den-181 in adult patients with ACPA+ rheumatoid arthritis on stable methotrexate. *Arthritis Rheumatol*. (2019) 71(Suppl 10):1–5362.
 245. McHugh MD, Park J, Uhrich R, Gao W, Horwitz DA, Fahmy TM. Paracrine co-delivery of TGF- β and IL-2 using CD4-targeted nanoparticles for induction and maintenance of regulatory T cells. *Biomaterials*. (2015) 59:172–81. doi: 10.1016/j.biomaterials.2015.04.003
 246. Fedorak RN, Gangl A, Elson CO, Rutgeerts P, Schreiber S, Wild G, et al. Recombinant human interleukin 10 in the treatment of patients with mild to moderately active Crohn's disease. The Interleukin 10 inflammatory bowel disease cooperative study group. *Gastroenterology*. (2000) 119:1473–82. doi: 10.1053/gast.2000.20229
 247. Tilg H, Van Montfrans C, Van den Ende A, Kaser A, van Deventer SJH, Schreiber S, et al. Treatment of Crohn's disease with recombinant human interleukin 10 induces the proinflammatory cytokine interferon γ . *Gut*. (2002) 50:191–5. doi: 10.1136/gut.50.2.191
 248. Bandyopadhyay A, Fine RL, Demento S, Bockenstedt LK, Fahmy TM. The impact of nanoparticle ligand density on dendritic-cell targeted vaccines. *Biomaterials*. (2011) 32:3094–105. doi: 10.1016/j.biomaterials.2010.12.054
 249. Bhavsar MD, Amiji MM. Oral IL-10 gene delivery in a microsphere-based formulation for local transfection and therapeutic efficacy in inflammatory bowel disease. *Gene Ther*. (2008) 15:1200–9. doi: 10.1038/gt.2008.67
 250. Braat H, Peppelenbosch MP, Hommes DW. Interleukin-10-based therapy for inflammatory bowel disease. *Expert Opin Biol Ther*. (2003) 3:725–31. doi: 10.1517/14712598.3.5.725
 251. Braat H, Rottiers P, Hommes DW, Huyghebaert N, Remaut E, Remon J-P, et al. A phase I trial with transgenic bacteria expressing interleukin-10 in Crohn's disease. *Clin Gastroenterol Hepatol*. (2006) 4:754–9. doi: 10.1016/j.cgh.2006.03.028
 252. Vandenbroucke K, de Haard H, Beirnaert E, Dreier T, Lauwereys M, Huyck L, et al. Orally administered *L. lactis* secreting an anti-tnf nanobody demonstrate efficacy in chronic colitis. *mucosal immunol*. (2010) 3:49–56. doi: 10.1038/mi.2009.116
 253. Huijbregtse IL, Marietta EV, Rashtak S, Koning F, Rottiers P, David CS, et al. Induction of antigen-specific tolerance by oral administration of *Lactococcus lactis* delivered immunodominant DQ8-restricted gliadin peptide in sensitized nonobese diabetic Abo Dq8 transgenic mice. *J Immunol*. (2009) 183:2390–6. doi: 10.4049/jimmunol.0802891
 254. Takiishi T, Korf H, Van Belle TL, Robert S, Grieco FA, Caluwaerts S, et al. Reversal of autoimmune diabetes by restoration of antigen-specific tolerance using genetically modified *Lactococcus lactis* in mice. *J Clin Invest*. (2012) 122:1717–25. doi: 10.1172/JCI60530
 255. Robert S, Gysemans C, Takiishi T, Korf H, Spagnuolo I, Sebastiani G, et al. Oral delivery of Glutamic Acid Decarboxylase (GAD)-65 and IL10 by *Lactococcus lactis* reverses diabetes in recent-onset NOD mice. *Diabetes*. (2014) 63:2876–87. doi: 10.2337/db13-1236
 256. Takiishi T, Cook DP, Korf H, Sebastiani G, Mancarella F, Cunha JPMCM, et al. reversal of diabetes in NOD mice by clinical-grade proinsulin and IL-10-secreting *Lactococcus lactis* in combination with low-dose anti-CD3 depends on the induction of Foxp3-positive T cells. *Diabetes*. (2017) 66:448–59. doi: 10.2337/db15-1625
 257. Clemente-Casares X, Blanco J, Ambalavanan P, Yamanouchi J, Singha S, Fandos C, et al. Expanding antigen-specific regulatory networks to treat autoimmunity. *Nature*. (2016) 530:434–40. doi: 10.1038/nature16962
 258. Singha S, Shao K, Yang Y, Clemente-Casares X, Solé P, Clemente A, et al. Peptide-MHC-based nanomedicines for autoimmunity function as T-cell receptor microclustering devices. *Nat Nanotech*. (2017) 12:701–10. doi: 10.1038/nnano.2017.56
 259. Umeshappa CS, Singha S, Blanco J, Shao K, Nanjundappa RH, Yamanouchi J, et al. Suppression of a broad spectrum of liver autoimmune pathologies by single peptide-MHC-based nanomedicines. *Nat Commun*. (2019) 10:2150. doi: 10.1038/s41467-019-09893-5
 260. Weiler-Normann C, Rehermann B. The liver as an immunological organ. *J Gastroenterol Hepatol*. (2004) 19:S279–83. doi: 10.1111/j.1440-1746.2004.03687.x
 261. Thomson AW, Knolle PA. Antigen-presenting cell function in the tolerogenic liver environment. *Nat Rev Immunol*. (2010) 10:753–66. doi: 10.1038/nri2858
 262. Bogdanos DP, Gao B, Gershwin ME. Liver immunology. *Compr Physiol*. (2013) 3:567–98. doi: 10.1002/cphy.c120011
 263. Limmer A, Ohl J, Kurts C, Ljunggren H-G, Reiss Y, Groettrup M, et al. Efficient presentation of exogenous antigen by liver endothelial cells to CD8+ T cells results in antigen-specific T-cell tolerance. *Nat Med*. (2000) 6:1348–54. doi: 10.1038/82161
 264. Diehl L, Schurich A, Grochtmann R, Hegenbarth S, Chen L, Knolle PA. Tolerogenic maturation of liver sinusoidal endothelial cells promotes B7-homolog 1-dependent CD8+ T cell tolerance. *Hepatology*. (2008) 47:296–305. doi: 10.1002/hep.21965
 265. Schildberg FA, Hegenbarth SI, Schumak B, Scholz K, Limmer A, Knolle PA. Liver sinusoidal endothelial cells veto CD8 T cell activation by antigen-presenting dendritic cells. *Eur J Immunol*. (2008) 38:957–67. doi: 10.1002/eji.200738060
 266. Kruse N, Neumann K, Schrage A, Derkow K, Schott E, Erben U, et al. Priming of CD4+ T cells by liver sinusoidal endothelial cells induces CD25low forkhead box protein 3- regulatory T cells suppressing autoimmune hepatitis. *Hepatology*. (2009) 50:1904–13. doi: 10.1002/hep.23191

267. Neumann K, Rudolph C, Neumann C, Janke M, Amsen D, Scheffold A. Liver sinusoidal endothelial cells induce immunosuppressive IL-10-producing Th1 cells via the notch pathway. *Eur J Immunol.* (2015) 45:2008–16. doi: 10.1002/eji.201445346
268. Wohlleber D, Knolle PA. The role of liver sinusoidal cells in local hepatic immune surveillance. *Clin Transl Immunol.* (2016) 5:e117. doi: 10.1038/cti.2016.74
269. Carambia A, Freund B, Schwinge D, Bruns OT, Salmen SC, Ittrich H, et al. Nanoparticle-based autoantigen delivery to Treg-inducing liver sinusoidal endothelial cells enables control of autoimmunity in mice. *J Hepatol.* (2015) 62:1349–56. doi: 10.1016/j.jhep.2015.01.006
270. Poisson J, Lemoine S, Boulanger C, Durand F, Moreau R, Valla D, et al. Liver sinusoidal endothelial cells: physiology and role in liver diseases. *J Hepatol.* (2017) 66:212–27. doi: 10.1016/j.jhep.2016.07.009
271. Wilson DS, Damo M, Hirose S, Raczy MM, Brüggel K, Diaceri G, et al. Synthetically glycosylated antigens induce antigen-specific tolerance and prevent the onset of diabetes. *Nat Biomed Eng.* (2019) 3:817–29. doi: 10.1038/s41551-019-0424-1
272. Hu J, Liu J, Yang D, Lu M, Yin J. Physiological roles of asialoglycoprotein receptors (ASGPRs) variants and recent advances in hepatic-targeted delivery of therapeutic molecules via ASGPRs. *Protein Pept Lett.* (2014) 21:1025–30. doi: 10.2174/0929866521666140626102429
273. Li D, Romain G, Flamar A-L, Duluc D, Dullaers M, Li X-H, et al. Targeting self- and foreign antigens to dendritic cells via DC-ASGPR generates IL-10-producing suppressive CD4+ T cells. *J Exp Med.* (2012) 209:109–21. doi: 10.1084/jem.20110399
274. Li B, Selmi C, Tang R, Gershwin ME, Ma X. The microbiome and autoimmunity: a paradigm from the gut-liver axis. *Cell Mol Immunol.* (2018) 15:595–609. doi: 10.1038/cmi.2018.7
275. Strober W. Impact of the gut microbiome on mucosal inflammation. *Trends Immunol.* (2013) 34:423–30. doi: 10.1016/j.it.2013.07.001
276. Hebbandi Nanjundappa R, Ronchi F, Wang J, Clemente-Casares X, Yamanouchi J, Sokke Umeshappa C, et al. A gut microbial mimic that hijacks diabetogenic autoreactivity to suppress colitis. *Cell.* (2017) 171:655–67.e17. doi: 10.1016/j.cell.2017.09.022
277. Fiorucci S, Biagioli M, Zampella A, Distrutti E. Bile acids activated receptors regulate innate immunity. *Front Immunol.* (2018) 9:1853. doi: 10.3389/fimmu.2018.01853
278. Shapiro H, Kolodziejczyk AA, Halstuch D, Elinav E. Bile acids in glucose metabolism in health and disease. *J Exp Med.* (2018) 215:383–96. doi: 10.1084/jem.20171965
279. Samstein RM, Perica K, Balderrama F, Look M, Fahmy TM. The use of deoxycholic acid to enhance the oral bioavailability of biodegradable nanoparticles. *Biomaterials.* (2008) 29:703–8. doi: 10.1016/j.biomaterials.2007.10.026
280. Doran MF, Crowson CS, Pond GR, O'Fallon WM, Gabriel SE. Frequency of infection in patients with rheumatoid arthritis compared with controls: a population-based study. *Arthritis Rheum.* (2002) 46:2287–93. doi: 10.1002/art.10524
281. Fernández-Ruiz M, Aguado JM. Risk of infection associated with anti-TNF- α therapy. *Expert Rev Anti Infect Ther.* (2018) 16:939–56. doi: 10.1080/14787210.2018.1544490
282. Winthrop KL, Novosad SA, Baddley JW, Calabrese L, Chiller T, Polgreen P, et al. Opportunistic infections and biologic therapies in immune-mediated inflammatory diseases: consensus recommendations for infection reporting during clinical trials and postmarketing surveillance. *Ann Rheum Dis.* (2015) 74:2107–16. doi: 10.1136/annrheumdis-2015-207841
283. Winthrop KL, Mariette X, Silva JT, Benamu E, Calabrese LH, Dumusc A, et al. ESCMID Study Group for Infections in Compromised Hosts (ESGICH) consensus document on the safety of targeted and biological therapies: an infectious diseases perspective (Soluble immune effector molecules [II]: agents targeting interleukins, immunoglobulins and complement factors). *Clin Microbiol Infect.* 24(Suppl. 2):S21–40. doi: 10.1016/j.cmi.2018.02.002
284. Vaglio A, Grayson PC, Fenaroli P, Gianfreda D, Boccaletti V, Ghiggeri GM, et al. Drug-induced lupus: traditional and new concepts. *Autoimmun Rev.* (2018) 17:912–8. doi: 10.1016/j.autrev.2018.03.016
285. Shovman O, Tamar S, Amital H, Watad A, Shoenfeld Y. Diverse patterns of anti-TNF- α -induced lupus: case series and review of the literature. *Clin Rheumatol.* (2018) 37:563–8. doi: 10.1007/s10067-017-3884-2
286. Shoenfeld Y, Ehrenfeld M, Perry O. The kaleidoscope of autoimmunity - from genes to microbiome. *Clin Immunol.* (2019) 199:1–4. doi: 10.1016/j.clim.2018.12.003
287. Karsdal MA, Bay-Jensen A-C, Henriksen K, Christiansen C, Genant HK, Chamberlain C, et al. Rheumatoid arthritis: a case for personalized health care? *Arthritis Care Res.* (2014) 66:1273–80. doi: 10.1002/acr.22289
288. Karsdal MA, Christiansen C, Ladel C, Henriksen K, B KV, Bay-Jensen AC. Osteoarthritis - a case for personalized health care? *Osteoarthritis Cartilage.* (2014) 22:7–16. doi: 10.1016/j.joca.2013.10.018
289. Hey SP, Kesselheim AS. Countering imprecision in precision medicine. *Science.* (2016) 353:448–9. doi: 10.1126/science.aaf5101
290. Sauna ZE, Lagassé D, Pedras-Vasconcelos J, Golding B, Rosenberg AS. Evaluating and mitigating the immunogenicity of therapeutic proteins. *Trends Biotechnol.* (2018) 36:1068–84. doi: 10.1016/j.tibtech.2018.05.008
291. Dingman R, Balu-Iyer SV. Immunogenicity of protein pharmaceuticals. *J Pharm Sci.* (2019) 108:1637–54. doi: 10.1016/j.xphs.2018.12.014
292. Oved JH, Barrett DM, Teachey DT. Cellular therapy: immune-related complications. *Immunol Rev.* (2019) 290:114–26. doi: 10.1111/imr.12768
293. Rachid R, Umetsu DT. Immunological mechanisms for desensitization and tolerance in food allergy. *Semin Immunopathol.* (2012) 34:689–702. doi: 10.1007/s00281-012-0333-9
294. Reber LL, Hernandez JD, Galli SJ. The pathophysiology of anaphylaxis. *J Allergy Clin Immunol.* (2017) 140:335–48. doi: 10.1016/j.jaci.2017.06.003
295. Cheifetz A, Smedley M, Martin S, Reiter M, Leone G, Mayer L, et al. The incidence and management of infusion reactions to infliximab: a large center experience. *Am J Gastroenterol.* (2003) 98:1315–24. doi: 10.1111/j.1572-0241.2003.07457.x
296. Hugo V. (1890) *Les Misérables*. Paris: Émile Testard Et Cie, Éditeurs.

Conflict of Interest: All authors are employees of Novartis and are engaged in drug development and/or translational medicine activities aiming to develop new therapeutics for AID. Novartis has a research and development collaboration with Parvus Therapeutics in the field of T1D.

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