



Inhibition of Inflammatory Changes in Human Myometrial Cells by Cell Penetrating Peptide and Small Molecule Inhibitors of NF κ B

Leo R. I. Gurney^{1*}, Julie Taggart¹, Wing-Chiu Tong¹, Arwyn T. Jones², Stephen C. Robson³ and Michael J. Taggart^{1*}

¹ Institute of Genetic Medicine, Newcastle University, Newcastle upon Tyne, United Kingdom, ² School of Pharmacy and Pharmaceutical Sciences, Cardiff University, Cardiff, United Kingdom, ³ Institute of Cellular Medicine, Newcastle University, Newcastle upon Tyne, United Kingdom

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*Correspondence:

Leo R. I. Gurney
leo.gurney@doctors.org.uk
Michael J. Taggart
michael.taggart@ncl.ac.uk

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Complications arising from Preterm Birth are the leading causes of neonatal death globally. Current therapeutic strategies to prevent Preterm Birth are yet to demonstrate success in terms of reducing this neonatal disease burden. Upregulation of intracellular inflammatory pathways in uterine cells, including those involving nuclear factor kappa-B (NF κ B), have been causally linked to both human term and preterm labor, but the barrier presented by the cell membrane presents an obstacle to interventions aimed at dampening these inflammatory responses. Cell penetrating peptides (CPPs) are novel vectors that can traverse cell membranes without the need for recognition by cell surface receptors and offer the ability to deliver therapeutic cargo internal to cell membranes. Using a human uterine cell culture inflammatory model, this study aimed to test the effectiveness of CPP-cargo delivery to inhibit inflammatory responses, comparing this effect with a small molecule inhibitor (Sc514) that has a similar intracellular target of action within the NF κ B pathway (the IKK complex). The CPP Penetratin, conjugated to rhodamine, was able to enter uterine cells within a 60 min timeframe as assessed by live confocal microscopy, this phenomena was not observed with the use of a rhodamine-conjugated inert control peptide (GC(GS)₄). Penetratin CPP conjugated to an IKK-inhibitory peptide (Pen-NBD) demonstrated ability to inhibit both the IL1 β -induced expression of the inflammatory protein COX2 and dampen the expression of a bespoke array of inflammatory genes. Truncation of the CPP vector rendered the CPP-cargo conjugate much less effective, demonstrating the importance of careful vector selection. The small molecule inhibitor Sc514 also demonstrated ability to inhibit COX2 protein responses and a broad down-regulatory effect on uterine cell inflammatory gene expression. These results support the further exploration of either CPP-based or small molecular treatment strategies to dampen gestational cell inflammatory responses in the context of preterm birth. The work underlines both the importance of careful selection of CPP vector-cargo combinations and basic testing over a broad time and concentration range to ensure effective responses. Further work should demonstrate the effectiveness of CPP-linked cargos to dampen alternative pathways of inflammation linked to Preterm Birth such as MAP Kinase or AP1.

Keywords: preterm birth, nuclear factor kappa B, cell penetrating peptide, tocolytic, myometrial cell

INTRODUCTION

Preterm birth, or birth before 37 weeks completed gestation as defined by the World Health Organization, is the main cause of neonatal death in developed countries and presents an enormous global problem (1). Being born too soon can confer significant clinical deficits throughout life, leading to neuro-developmental disorders such as cerebral palsy, learning impairment and visual disorders; with such problems being more likely to occur with greater frequency and severity at earlier gestations of birth (2).

Preterm birth is a syndrome best understood as the final endpoint of several possible pathophysiological events. It can be initiated by an array of processes including: uterine over-distension, utero-placental hemorrhage or ischaemia, maternal stress, cervical insufficiency, and inflammation with or without clinically apparent infection (3). Parallel to this, there is evidence to suggest that the processes that drive spontaneous human labor, whether at term or preterm, have an inflammatory basis. Macrophage and neutrophil numbers in human fetal membranes, myometrium and cervix are increased in tissue derived from laboring patients (4, 5) and such invasion leads to increases in local cytokine and chemokine production including increases in IL1 β , IL8, and TNF α (6). These changes may precipitate, or contribute to, a sequence of uterine activation processes leading to labor (7). Furthermore, invasion of the decidua by these cell types occurs in animal models of preterm birth and precedes the onset of labor (8); although whether this invasion is a cause or consequence of labor remains to be fully defined (9). The inflammatory milieu provoked within the uterine environment has been reported to upregulate varied pro-inflammatory pathways in uterine and placental cells including signaling pathways involving p38 MAP kinase (p38 MAPK) and Activator Protein 1 (AP1) (10, 11). Most importantly, upregulation of intracellular pathways involving the transcription factor nuclear factor κ B (NF κ B) are suggested to play a central role in the sequelae of inflammation-associated preterm birth (12).

NF κ B proteins are a family of five structurally related transcription factors named p65 (Rel A), RelB, c-Rel, p50 (NF κ B-1), and p52 (NF κ B-2) present in nearly all mammalian cells that play a ubiquitous role in inflammatory and infectious responses (13). All NF κ B proteins exist in the form of heterogeneous dimers, the most commonly described of which is p65/p50 which is known to activate the expression of pro-inflammatory genes within the uterus (14). Within the canonical NF κ B pathway interaction between cell surface receptors and lipopolysaccharide (LPS), or pro-inflammatory cytokines such as IL-1 β , TNF α , or IL6 activates the I κ B kinase complex (IKK complex) (13). This kinase complex consists of two catalytic subunits (IKK α & IKK β) each containing a six amino acid segment LDWSWL, known as the Nemo Binding Domain (NBD), which forms the basis of their binding site with the regulatory NF κ B essential modulator (NEMO) scaffold protein. Activation of IKK leads to the targeted phosphorylation of I κ B α allowing for the ubiquitination and subsequent degradation of this anchoring protein by the proteasome. This step allows it to translocate to the nucleus of the cell, a process enhanced by the IKK-dependent

phosphorylation of p65. Once in the cell nucleus, NF κ B binds to target gene promoters regulating the expression of inflammatory genes including those that transcribe IL1 β , TNF α , IL6, IL8, CXCL2, MMP 9, and COX2 (15). Due to the presence of κ B binding sites on the I κ B α gene promoter, activation of NF κ B leads to the rapid resynthesis of I κ B α which dissociates NF κ B from DNA complexes and is shuttled out of the nucleus in an inactive complex (16).

Amongst the evidence supporting the role of NF κ B activation in labor are observations from *in vitro* studies on human myometrial cells: activation of NF κ B has been shown to promote the expression of the inducible prostaglandin synthase enzyme cyclooxygenase 2 (COX2) leading to subsequent increases in prostaglandin production in these cells (17). Prostaglandins E2 and F2 α promote uterine contractions and their increased production within reproductive tissues is associated with the onset of human labor (18), thus increases in COX2 expression are thought to correspond to both inflammatory and contractile responses in the myometrium during human labor.

Myometrial cell NF κ B activation also promotes the increased production of pro-inflammatory cytokines including IL-6 and IL-8 (19), matrix metalloproteinases (20), and up-regulates the expression of mRNA encoding genes associated with labor including the oxytocin receptor and gap junction proteins (21, 22).

Agents aimed at the acute prevention of preterm birth are a class of drugs referred to as tocolytics. Despite their use in more than 3000 clinical trials over 60 years, tocolytic agents have yet to demonstrate significant improvements in neonatal outcome and their use is frequently associated with an unacceptably high frequency of unwanted sequelae (23). This leaves an urgent need for the exploration of new therapeutic strategies aimed at targeting the molecular pathways whose upregulation is linked to preterm birth.

Peptides targeting protein-protein interactions that regulate cellular processes are gaining increasing traction as therapeutic entities that target a number of diseases. As biological molecules they offer very high selectivity and specificity and are relatively cheap to manufacture (24). A major barrier to the development of new peptides as pharmaceuticals is presented by the cell membrane: the lipid bilayer can prevent the passage of therapeutics from extracellular space to intracellular targets that often lie within the cytosol of a cell. To overcome this obstacle requires a vector system that can deliver cargo to the cell cytosol either directly through the plasma membrane or through utilizing endocytosis as a portal to cytoplasm before mediating endocytic escape processes to reach the cytosol (25).

Cell Penetrating Peptides (CPPs) offer an attractive solution to this drug delivery puzzle: they are characterized as short peptides, usually <30 amino acids length, that have the ability to cross cell membranes without the need for recognition by cell surface receptors (26). CPPs have been shown to deliver cargo efficiently at low doses to a diverse range of cell types and a number of studies in varied clinical fields have confirmed the potential of CPP-cargo conjugates as therapeutic agents (27). This has led to several CPP-based therapies being tested in phase 3 clinical trials for a diverse range of inflammatory conditions (28). Despite this,

the effectiveness of CPP-linked therapy in gestational cells has yet to be examined in detail.

Amongst the broad array of cargoes that can be conjugated to CPPs and delivered intracellularly are molecules with the capability to block NF κ B-dependent signaling (15). The best described CPP-cargo conjugate with NF κ B inhibitory ability is the Nemo Binding Domain (NBD) peptide: an 11 amino acid residue peptide which was designed to span the NBD and therefore disrupt the interaction between the three IKK subunits within the NF κ B canonical pathway, with the effect of inhibiting the inflammatory ligand-dependant activation of NF κ B (29). The NBD peptide, conjugated to varied CPP vectors, has been shown to down-regulate elicited NF κ B responses and thus improve physiological endpoints of inflammation both *in vitro* and in animal models of diseases as varied as Muscular Dystrophy and Parkinson's disease (30–32).

Small molecule inhibitors targeting several levels of the NF κ B pathway have been reported including those blocking the phosphorylation of I κ B α via IKK complex inhibition or preventing proteasome degradation of this molecule (33). Inhibitors have also been demonstrated that prevent NF κ B localization to the nucleus or block post translational modifications of p65. The advantages of using small molecule drug inhibition include low immunogenicity and the possibility of attaining high oral bioavailability with the aim of inhibiting specific signaling pathways. However, high concentrations may be required to achieve therapeutic benefit; they may be poor at crossing biological barriers, and are often substrates for swift removal via drug efflux proteins (34).

The non-peptidic inhibitor Sc514 represents an established small molecule compound targeting NF κ B mediated effects. It exerts action via inhibition of the IKK β portion of the IKK complex and therefore putatively offers discrete inhibition of the NF κ B pathway via a mechanism similar to the NBD peptide (35). It has demonstrated ability to significantly reduce LPS-stimulated TNF α secretion in placentally-derived human primary cells (36), however it has yet to be broadly tested in a uterine cell setting. Thus, the use of this drug allows examination of the efficacy of a small molecule approach to inhibit NF κ B-related inflammatory responses in myometrial cells, as well as offering a comparison with the CPP-cargo inhibitory approach presented by the NBD peptide.

With the intention of broadening the scope of anti-inflammatory candidate agents available to be tested for their potential as treatments for preterm birth, the aims of this study were to: (i) first define the ability of CPPs to enter human myometrial cells and deliver the NBD peptide and (ii) subsequent to this, examine the effectiveness of CPP-NBD peptide conjugates in dampening NF κ B related signaling compared to Sc514 in human myometrial cells.

MATERIALS AND METHODS

Subjects & Samples

Ethical approval was obtained from Newcastle and North Tyneside Research Ethics Committee (10/H0906/71) to perform

research on samples collected as part of the Newcastle Utero-placental Tissue Bank. Human myometrial tissue was obtained from patients following written informed consent from non-laboring women with uncomplicated, singleton pregnancies undergoing elective Cesarean section at term (≥ 37 weeks gestation). Myometrial muscle strip biopsies approximately 1 \times 1 cm in size were taken from the upper portion of the lower segment uterine incision and placed in tissue collection buffer.

Biopsies were excluded from women with underlying medical or obstetric disease, women on any current medication, those with a body mass index outside range 20–35 kg/m², or who gave birth to a baby with weight below the 10th percentile or above the 90th percentile. Separate myometrial biopsies from 46 patients were included in the study.

Preparation of Myometrial Cells

Human myometrial biopsies were micro-dissected under a light microscope to isolate myometrial tissue from any remaining decidua. In a microbiological safety cabinet, tissue was cut into small fragments before adding 10 mls warmed hanks balanced salt solution containing 10 mg each of collagenase 1A and XI (Sigma Aldrich, C7657, C9891) plus 20 mg bovine serum albumin (Sigma Aldrich, A6003). This tissue digestion mix was placed in an orbital shaker at 110 rpm for approximately 40 min at 37°C. The sample was then triturated, filtered through a 70 μ m cell strainer into 10ml warmed media and centrifuged at 1,000 rpm (89 \times g) for 5 min. The supernatant was discarded and cell pellet re-suspended in GlutaMAX cell culture media (Life Technologies 61965) containing 10% FCS and Penicillin/Streptomycin until 80–90% confluent (37).

Cell Penetrating Peptides

CPP's were custom synthesized commercially and purchased from either EZ Biolabs (USA) or Abingdon Health Laboratory (UK) services. Rhodamine fluorophore cargo (excitation 561 nm, emission 617 nm) was labeled at the N-terminal end of either Penetratin or Pen(43-56) CPP. NBD or inactive NBD mutant cargo was conjugated at the C-terminal end of either Penetratin or Pen(43-56) CPP. Amino hexanoic acid (Aca) was used as a linker between CPP and cargo. The amino acid structures of the peptides used in experimentation are detailed in **Table 1**.

To test specificity of COX2 responses to the NBD peptide, the effect of a Penetratin-conjugated mutant version of the NBD peptide: Pen NBD (Mut), was examined. This peptide contains the substitution of two tryptophan (W) amino acid residues to alanine (A) thus rendering it unable to span the NBD and interfere with IKK related signaling toward NF κ B (29).

To further investigate the importance of CPP vector structure for the delivery of biologically active cargo a truncated form of Penetratin CPP was conjugated to the NBD peptide: Pen(43-56)-NBD (38).

Live Cell Confocal Microscopy

Primary human myometrial cells at passage P ≤ 4 were transferred into 4 or 8-well microscope Ibidi μ -Slides (Thistle Scientific, IB-80441, IB-80821) and grown up to 80–90% confluency. Cells

TABLE 1 | The nomenclature and amino acid structure of peptides used for experimentation.

Peptide name	Structure
Rhodamine labeled Penetratin	Rhodamine-RQIKIWFQNRMRMKWKK
Rhodamine labeled Pen-NBD	Rhodamine-RQIKIWFQNRMRMKWKKTALDWSWLQTE
Rhodamine labeled Pen(43-56) - NBD	Rhodamine-RQIKIWFQNRMRMKW-Aca- TALDWSWLQTE
Rhodamine labeled GS ₄ (GC)	Rhodamine - GSGSGSGSGC
Pen-NBD	RQIKIWFQNRMRMKWKKTALDWSWLQTE-NH ₂
Pen-NBD Mutant	RQIKIWFQNRMRMKWKKTALDASALQTE-NH ₂
NBD	TALDWSWLQTE
Pen (43-56)-NBD	Ac-RQIKIWFQNRMRMKW-Aca-TALDWSWLQTE-NH ₂
Pen (43-56)-NBD Mutant	Ac-RQIKIWFQNRMRMKW-Aca-TALDASALQTE-NH ₂

Rhodamine fluorophore cargo (excitation 561 nm, emission 617 nm) is labeled at the N-terminal end of Penetratin or Pen(43-56) CPP. NBD or inactive NBD mutant cargo is conjugated at the C-terminal end. Acyl (Ac), Aminohexanoic acid (Aca), Amide (NH₂).

were washed in phosphate buffered saline and changed to serum-reduced media (0.1%FCS) for 18–24 h (39). Cells were then loaded with 1 μ M Hoechst nuclear dye (Life Technologies, 33342) for 1 h and washed with PBS before transfer to the confocal microscope where they were maintained within a temperature-controlled chamber maintained at 37°C, 5%CO₂ on the stage of the inverted microscope.

Initial images were collected to assess baseline cellular auto-fluorescence prior to the addition of varying concentrations of fluorophore-CPP/fluorophore- CPP-Cargo/fluorophore-control peptide combinations (see **Table 1**) that were ready made up in serum-reduced cell media. Further images were captured 60 min following application. During imaging cells were placed in phenol red free Dulbecco modified eagle medium (Life Technologies, 21063) to reduce background glare.

To make a thorough assessment of fluorescent uptake throughout whole cells, all images were captured as a series of slices in the Z plane (range 3–10 slices per area selected, each slice 0.54 μ m apart) with the 40x objective using live cell spinning disk confocal microscopy (Andor Revolution XD coupled to an Olympus IX-81 inverted microscope; Andor Belfast UK). All images were digitally recorded with IQ2 software (Andor, Belfast, UK).

Cell Stimulation Experiments for Subsequent Protein or mRNA Extraction

Primary human myometrial cells at passage \leq P4 were split equally between 6 or 12 well-plates and grown to 80–90% confluency. At this point media was changed to serum-reduced (0.1% FCS) media for 18 to 24 h before discarding from each well. Five hundred microliters of fresh media (0.1% FCS) was produced containing: Pen-NBD, Pen(43-56)-NBD, Sc514 (Tocris bioscience, 3318); or controls (equivalent volume DMSO, Pen-NBD Mutant, NBD alone) at indicated concentrations. The inhibitor/control media was added to wells as a pre-incubation

step. After 1 h, 10 ng/ml IL1 β (Peprotech, 200-01B), or equivalent volume DMSO vehicle, was added to each well.

For western blotting experiments cells were washed with PBS before being lysed using sucrose cell lysis buffer (62.5 mM Tris-HCl pH6.8, 2% SDS, 10% saccharose) containing 20 μ l/ml protease inhibitor (Sigma Aldrich, P1860) and 5 μ l/ml phosphatase inhibitor (Sigma Aldrich P2745) at times of 0/15/60/120/240 minute from cytokine stimulation.

For RNA array experiments, 4 h following cytokine stimulation cells were washed with PBS before RNA was extracted using the RNeasy Mini Kit (Qiagen, 74101) according to the manufacturers' protocol.

Western Immunoblotting

Cell lysate samples were sonicated and underwent Lowry assay to determine overall protein concentration before dilution in an equal volume of 2x Laemmli buffer (Tris pH6.8 250 mM; SDS 4% w/v; Glycerol 10% v/v; β -mercaptoethanol; bromophenol blue). Ten micrograms of sample protein per lane was loaded onto a 1.5 mm 10% sodium dodecyl sulfate poly-acrylamide gel (SDS-PAGE) before undergoing separation by electrophoresis, gels then underwent electrophoretic transfer to a methanol-activated polyvinylidene difluoride (PVDF) membrane.

Following a blocking step of washing membranes in 5% non-fat dry milk in Tris-buffered saline with 0.1% Tween-20 (TBS-T) for 1 h at room temperature, membranes were incubated overnight at 4°C with primary anti-COX-2 antibody 1:500 (Cayman laboratories, CAY160112), anti-I κ B- α antibody 1:500 (Santa Cruz, C21 Sc371) anti-phospho-P65 antibody 1:2000 (Cell Signaling, 3013), or anti-P65 antibody 1:15,000 (Santa Cruz, Sc372) in 1% non-fat dry milk with TBS-T. After three washing steps with TBS-T, secondary antibody incubation occurred for 1 h at room temperature. Polyclonal horseradish peroxidase conjugated goat anti-mouse immunoglobulin 1:3000 (DAKO, P0447) in 1% non-fat dry milk was used in conjunction with COX2 antibody and a polyclonal horseradish peroxidase conjugated goat anti-rabbit immunoglobulin 1:5000 in 1% non-fat dry milk (DAKO, P0448) was used for I κ B- α , phospho-P65 and P65 antibody.

After further TBS-T wash, enhanced chemiluminescent (ECL) reagent (Fisher Scientific, 12316992) was applied to the PVDF membranes for 5 min. Membranes were dried, placed in development cassettes and developed manually onto photographic film in a dark room. Equal loading of proteins was assessed by staining of the PVDF membrane with Napthol Blue Black Reagent (0.1% napthol blue black, 10% methanol, 2% acetic acid) to detect actin protein (43kDa). Developed films were densitometrically scanned using UMAX PowerLook III (UMAX) and quantification performed with Bio Image Intelligent Quantifier 2 software (Bioimage systems).

Measurement of mRNA Expression

Following extraction of RNA (as per above), gene expression was assessed via the measurement of mRNA transcripts across a panel of selected genes using RT qPCR arrays (Qiagen, Custom RT2 profiler PCR arrays, 330171). All work was carried out in a ribonuclease free environment with ribonuclease free equipment.

TABLE 2 | Genes used in RT2 profiler PCR array plates.

Gene grouping	Gene name
Labor associated genes	OXTR/MMP 9/ MMP19/ TIMP1/ GJA1/GJB2
Inflammatory Genes	IL1A/IL1B/TNFA/IL6/IL8/ICAM1/ SOCS3 /IL1R1/ IL1R2/IL4R/CXCL2/ CXCL1/CXCL6/CCL2
NFκB pathway	NFKB1/NFKB2/ RELA/NFKBIA/NFKBIZ
Prostaglandin production	PTGER3/PLA2G2A/ PTGES/PTGS2
G protein receptors	GPR37/GPR34/RGS 10
Novel genes	S100A9/ S100A8/STAT1/ FOXO1/ ZEB1/ LILRA5/ SPINK5/FOSB/JUN/TRIB1
Housekeeping genes	GAPDH/ACTB/B2M

Measurement of the RNA content in each sample was undertaken using a nanodrop spectrophotometer (ND-1000, Labtech), with suitable RNA purity considered to be a 260/280 ratio of >2.1. 0.5 µg of total RNA from each sample was added to the genomic DNA elimination mix from RT2 first strand kit (Qiagen, 330404) reverse transcription was then carried out according the manufacturers' instructions.

cDNA as synthesized from 0.5 µg of total RNA from each original sample was added to the SYBR Green Master Mix (Qiagen, 330171). Twenty-five microliters of this final mix was then added to each well of a 96 well-array plate, with each well-containing a different primer for a gene of interest or control. The plate was sealed with optical film, centrifuged at 1,000 rpm (89 × g) for 1 min before inserting the plate into the PCR cyclor (Step One Plus, Applied Biosciences) according to a protocol of enzyme activation hot start at 95°C for 10 min, followed by 40 cycles of denaturing at 95°C for 15 s and extension at 60°C for 60 s.

Fluorogenic data was collected via the FAM channel and the cycle threshold (Ct) values were calculated by applying a threshold limit that represented the exponential phase of amplification. To ensure comparability of gene expression between different array plates the same threshold limit was applied to all experiments (Ct 0.116). This was calculated as a mean threshold value from the first 6 arrays and corresponded to the exponential phase of the amplification curve for all subsequent experiments.

These array plates were created bespoke for the authors by Qiagen and the genes selected for examination are demonstrated in **Table 2**. The approach taken to gene selection is described within the results section below.

Analysis of mean Ct values and quality control were evaluated by exporting mean Ct values for each gene on the RT2 array plate to the following data analysis website supported by Qiagen: <https://dataanalysis.sabiosciences.com/pcr/arrayanalysis.php>.

Gene expression was normalized to three reference housekeeping genes β2-microglobulin (B2M), GAPDH and ACTB, and results calculated using the $\Delta\Delta C_T$ method.

For all qPCR gene array experiments raw Ct values are included in the **Supplemental Data 1**.

Statistical Analysis

Prism 7.0 (GraphPad) software was used to perform statistical analysis. For western blotting data statistical analysis was

performed on raw optical densitometry values. One or two-way (time/dose) ANOVA with *post-hoc* corrections were performed to compare differences between groups, as indicated in figure legends. For initial mRNA expression experiments comparing untreated and IL1β treated samples (**Figure 8**) *P*-values were calculated based on a Student's *t*-test of the replicate $2^{\Delta\Delta C_t}$ values of each gene. For subsequent mRNA experiments comparing inhibitor groups (**Table 3** and **Supplemental Figure 3**) one-way ANOVA with Bonferroni *post-hoc* corrections were performed to compare differences between groups. Statistical significance was assumed at $P < 0.05$.

For all experimental conditions: each n represents a biological replicate i.e., each n is a separate experiment using cell cultures grown from myometrial biopsy of a new patient.

RESULTS

Penetratin CPP Effectively Delivers Fluorescent Cargo to Human Myometrial Cells

Using the well-characterized Penetratin CPP, we initially investigated whether human myometrial cells were accessible to the CPP-mediated delivery of rhodamine fluorophore.

Figure 1 displays representative confocal microscope images of live uterine smooth muscle cells 1 h after application of rhodamine fluorescent cargo conjugated to either Penetratin (CPP), Pen-NBD, or to Pen(43-56)-NBD. The delivery of fluorescent cargo was assessed by comparison to that of a rhodamine-conjugated control peptide purported to not have cell penetrating ability [GS₄ (GC)] (40) which can be viewed in **Figure 1D**.

At 1, 3, and 10 µM the rhodamine fluorophore was successfully delivered internal to the cells within 60 min of application, with an increase in intensity of uptake with increasing concentration (**Figures 1A–C**). By comparison the inert peptide GS₄ (GC) at 10 µM did not appreciably deliver the rhodamine fluorophore after 60 min (**Figure 1D**). This indicates the ability of CPPs to deliver cargo to human myometrial cells.

Following 60 min of cellular application, the CPP-fluorophore conjugates (**Figure 1A**) and also the CPP-cargo-fluorophore conjugates (**Figures 1B,C**) largely demonstrated a punctate pattern of labeling indicative of endosomal labeling. This is best demonstrated in the image of Rhodamine-Pen-NBD at 10 µM (Row B, third picture from left). It can be noted that this punctate labeling forms larger accumulations with the use of Pen(43-56)-NBD (**Figure 1C**), this may be attributed to alteration of the CPP-cargo amino acid structure, a phenomenon referred to in the discussion below.

The CPP-Cargo Conjugate Pen-NBD Inhibits Cytokine-Activated COX2 Expression

Initial experiments determined that CPP use was not toxic to myometrial cells (**Supplementary Figure 4**). Penetratin conjugated to the NBD peptide (Pen-NBD) was applied to myometrial cell cultures in increasing concentrations 1 h prior to addition of 10 ng/ml of IL1β. As demonstrated

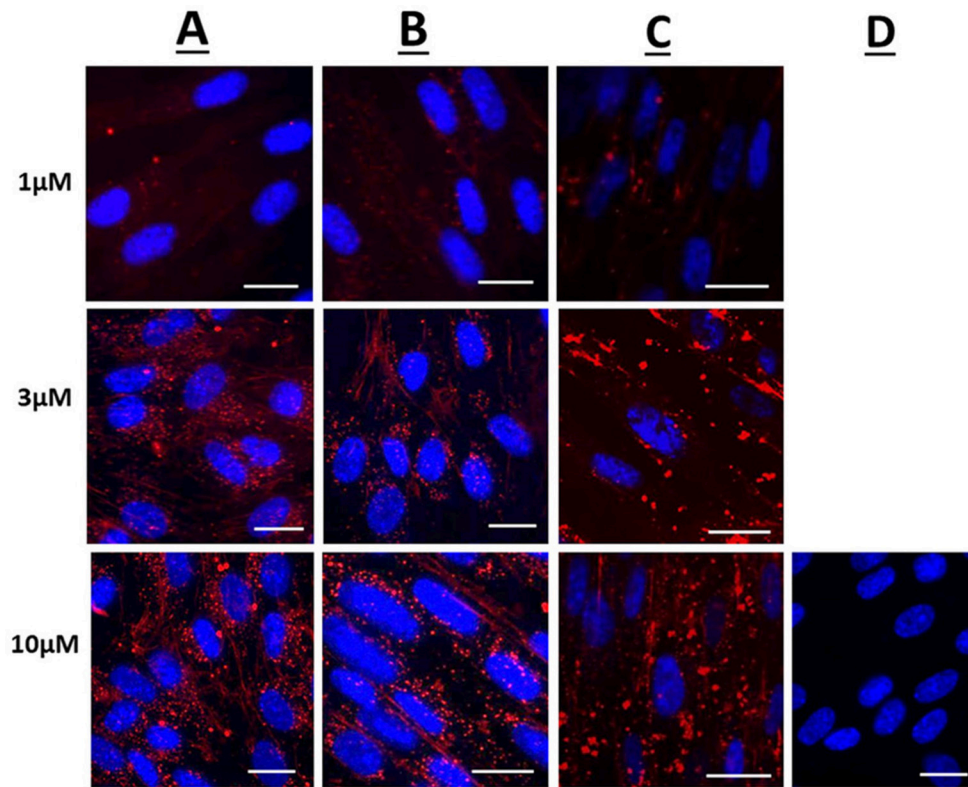


FIGURE 1 | Confocal microscopy images of live human uterine smooth muscle cells following application of rhodamine-conjugated CPPs or CPP-cargo combinations. Images captured 60 min following addition of either 1, 3, or 10 μM concentrations of (A) Rhodamine—Penetratin, (B) Rhodamine—Pen-NBD, or (C) Rhodamine—Pen (43-56)-NBD. (D) Displays cells imaged 60 min following addition of 10 μM control peptide Rhodamine - GS4 (GC). Scale bars 20 μm . Cells are maintained in serum deprived media within a temperature-controlled chamber (37°C, 5%CO₂). To demonstrate intra-cellularity of uptake, images are taken from the center of a Z-stack of 3–10 slices (0.54 μm apart) intended to capture the full depth of the cell. Images are representative of 3 independent experiments.

in the Western blot portrayed in **Figure 2**: in the absence of inhibitor (IL1 β alone) induction of the protein enzyme COX2 is evident at 2 h and continues to increase at 4 h following cytokine application. Pre-incubation of 50 and 100 μM Pen-NBD led to significant inhibition of these cytokine-stimulated increases in COX2 protein signal at both 2 h (50 μM $p = 0.01$; 100 μM $p = 0.003$) and 4 h (50 μM $p = 0.049$; 100 μM $p = 0.0003$).

Control experiments were performed to test the specificity of COX2 responses to the NBD peptide. The inhibitory effect of Pen-NBD on IL1 β -induced COX2 protein expression was compared with a Penetratin-conjugated mutant version of the NBD peptide: Pen-NBD (Mut), and NBD peptide alone (without conjugation to CPP). **Figure 3** displays a representative Western blot with scatter graphs demonstrating three independent experiments to compare COX2 protein responses to IL1 β following the pre-incubation with either Pen-NBD mutant or wild type peptide. It demonstrates that the mutant peptide does not diminish IL1 β -stimulated COX2 induction when used at identical concentrations to the wild type peptide.

Application of non-conjugated NBD peptide had no inhibitory effect on cytokine-stimulated COX2 protein increases, nor did application of CPP alone (**Supplementary Figure 1**), thus indicating the requirement of CPP-conjugation to NBD for the inhibitory effect to be evident.

In Human Myometrial Cells Pen-NBD Does Not Prevent the Cytokine-Stimulated Degradation of I κ B α but Does Diminish Phosphorylation of P65 Following IL1 β Exposure

It was important to assess the effect of CPP-based inhibition on protein expression changes within the canonical NF κ B pathway subsequent to IL1 β stimulation. Within myometrial cells, increases in the phosphorylated form of the NF κ B subunit p65 protein are observed at 15 min due to the action of the activated IKK complex (**Supplementary Figure 2**), these responses gradually diminish over the tested 4 h time frame but do not return to normal, suggestive of increased background NF κ B activity following cytokine stimulation in human uterine cells over the time frame examined. Additionally, within 15 min of exposure to IL1 β , there is both phosphorylation of the inhibitory I κ B α protein and degradation of its native form with full return of the native form of this inhibitory protein occurring between 60 min and 2 h.

At all concentrations tested, Pen-NBD did not prevent the cytokine-induced degradation of I κ B α (**Figure 4A**). However, a significant reduction of P65 protein phosphorylation 15 min following application of IL1 β was

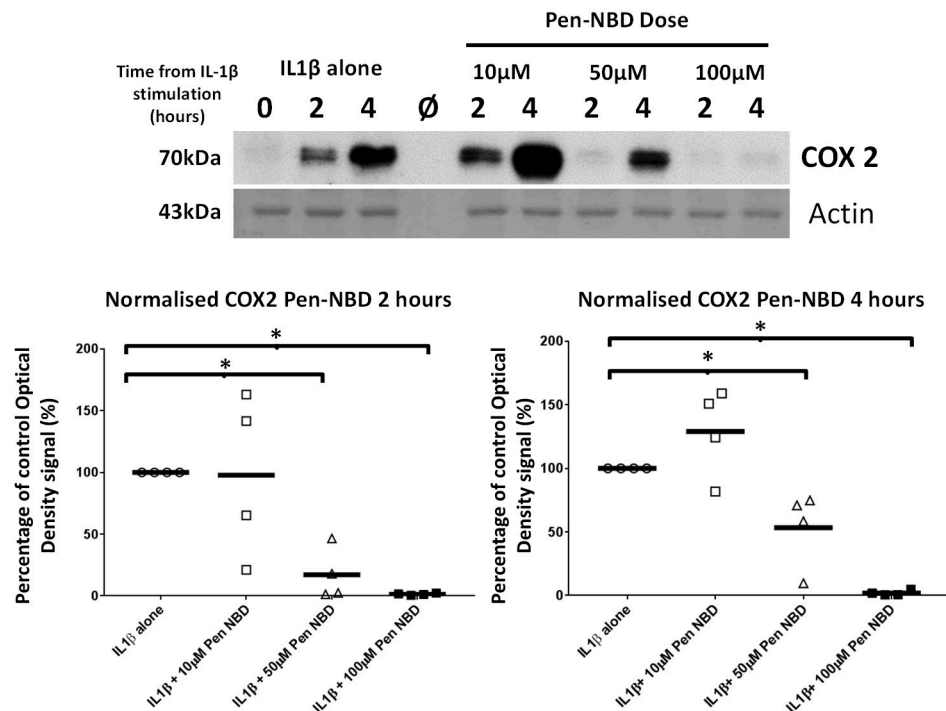


FIGURE 2 | Effect of Pen-NBD peptide on IL1 β -stimulated COX2 protein increases. **(Upper)** Representative Western blot demonstrating COX2 protein increase at 2 and 4 h following application of 10 ng/ml IL1 β alone or with pre-incubation for 1 h with indicated concentrations of Pen-NBD peptide prior to IL1 β addition. Actin expression displayed as loading control. \emptyset = no protein loaded. **(Lower)** Scatter plot demonstrating optical density values of COX2 protein signal at 2 and 4 h time points comparing IL1 β alone or Pen-NBD plus IL1 β experiments. Data presented has been normalized to control signal to demonstrate inhibitory effect. *Significant difference from raw optical density values between IL1 β alone and IL1 β plus Pen-NBD groups ($n = 4$, one-way ANOVA with Bonferroni *post-hoc* correction).

observed with the use of 100 μ M Pen-NBD ($p = 0.025$) (Figure 4B).

The NBD Peptide Conjugated to the Truncated Penetratin Vector Pen(43-56) Is Capable of COX2 Inhibition but Is Less Efficacious Than Pen-NBD and Does Not Influence IKK-Dependent Protein Alterations Within the NF κ B Pathway

To test the effect of alterations of the CPP vector on the efficacy of NBD cargo protein-protein interactions, experiments were performed using the NBD peptide conjugated to a truncated arrangement of the CPP Penetratin, to form Pen(43-56)-NBD. This CPP-peptide fusion has previously demonstrated ability to block IL1 β -induced NF κ B activity in HeLa cell lines (38), but has not been examined previously in a gestational cell setting.

Applied to human myometrial cells 100 μ M Pen (43-56)-NBD was able to inhibit IL1 β -induced COX2 protein increases at 4 h ($p = 0.0045$) (Figure 5A). Such inhibition was not seen with the use of lower doses (data not shown). This CPP-cargo conjugation did not inhibit I κ B α protein degradation or P65 protein phosphorylation (Figures 5B,C).

The Small Molecule Inhibitor Sc514 Exerts Inhibitory Effect on COX2 Protein Induction Via Alteration of IKK-Dependent Events Within the NF κ B Pathway

Sc514 at 50 μ M concentration was applied to myometrial cells 1 h prior to IL1 β stimulation. Significant reduction in COX2 protein expression with the use of this inhibitor was observed at 4 h ($p = 0.001$). IL1 β -induced degradation of I κ B α was inhibited ($p = 0.009$) as was the phosphorylation of P65 protein ($p = 0.001$) (Figures 6A–C).

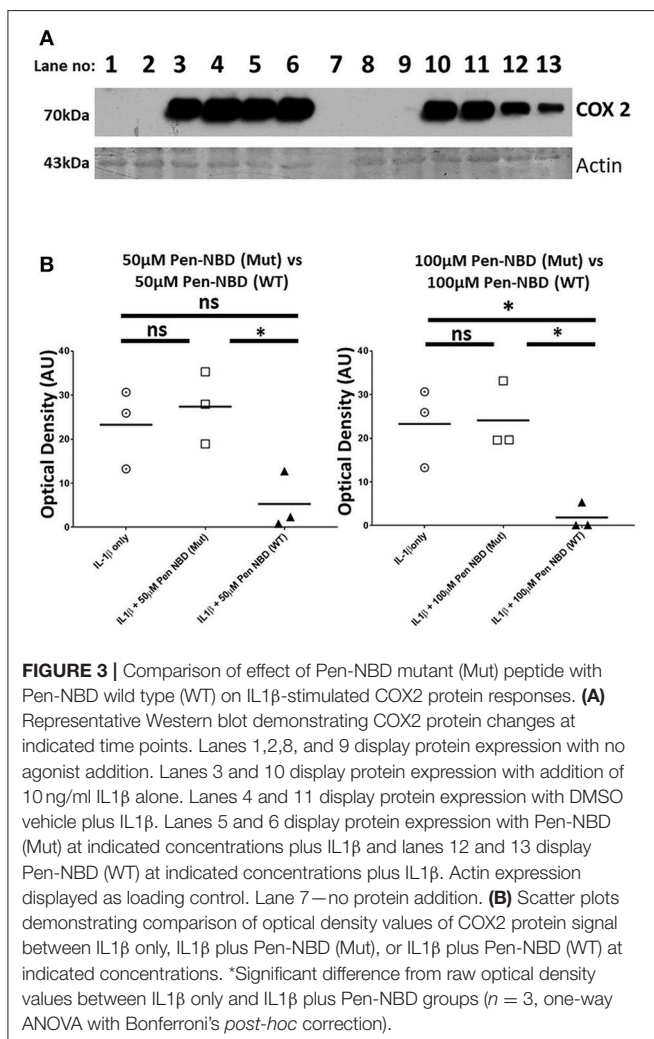
IL1 β Promotes Inflammation- and Labor-Related Gene Expression Changes in Human Myometrial Cells

To further demonstrate the response of myometrial cells to inflammatory stimulation and to compare the ability of CPP based and small molecule inhibitory strategies to these responses a bespoke q-PCR array panel of 42 genes was used. The genes examined in this study are listed in Table 2. The approach to gene selection for this panel was multifactorial: a number of genes encoding for proteins associated with the physiological events of uterine contraction, cervical dilation and membrane rupture that occur during human labor, termed labor associated genes, were selected (41); also, for comparison, a collection of

genes encoding for G-proteins involved in maintaining uterine quiescence (42). A number of pro inflammatory genes, including genes encoding for proteins in the NF κ B pathway (16), and genes involved in the production of prostaglandins (18) were chosen to aid understanding of the potential mechanism of action of inhibitors used in this study. A selection of novel genes that demonstrate altered expression in the presence of human labor were also included in the array (43).

Across eight independent samples a series of gene expression changes were observed in myometrial cells 4 h following exposure to 10 ng/ml IL1 β . These changes included genes involved in human labor, inflammatory processes, NF κ B signaling, and prostaglandin expression as demonstrated in the heat map displayed in **Figure 7**.

The volcano plot presented in **Figure 8** demonstrates those genes exhibiting significant expression changes with greater than 2-fold difference between IL1 β -treated and untreated samples. The genes displaying the largest expression changes within the panel were inflammatory cytokines/chemokines and genes associated with the NF κ B family.



CPP-Conjugated and Small Molecule Inhibitors of NF κ B Dampen the IL1 β -Stimulated Expression of Different Groups of Genes in Myometrial Cells

Having identified expression changes within the gene array panel following stimulation by IL1 β , it was important to examine how this expression pattern would be altered using a CPP-based or small molecule inhibitory strategy. It was also of interest to observe how alterations to vector structure may affect the efficacy of CPP-cargo inhibition.

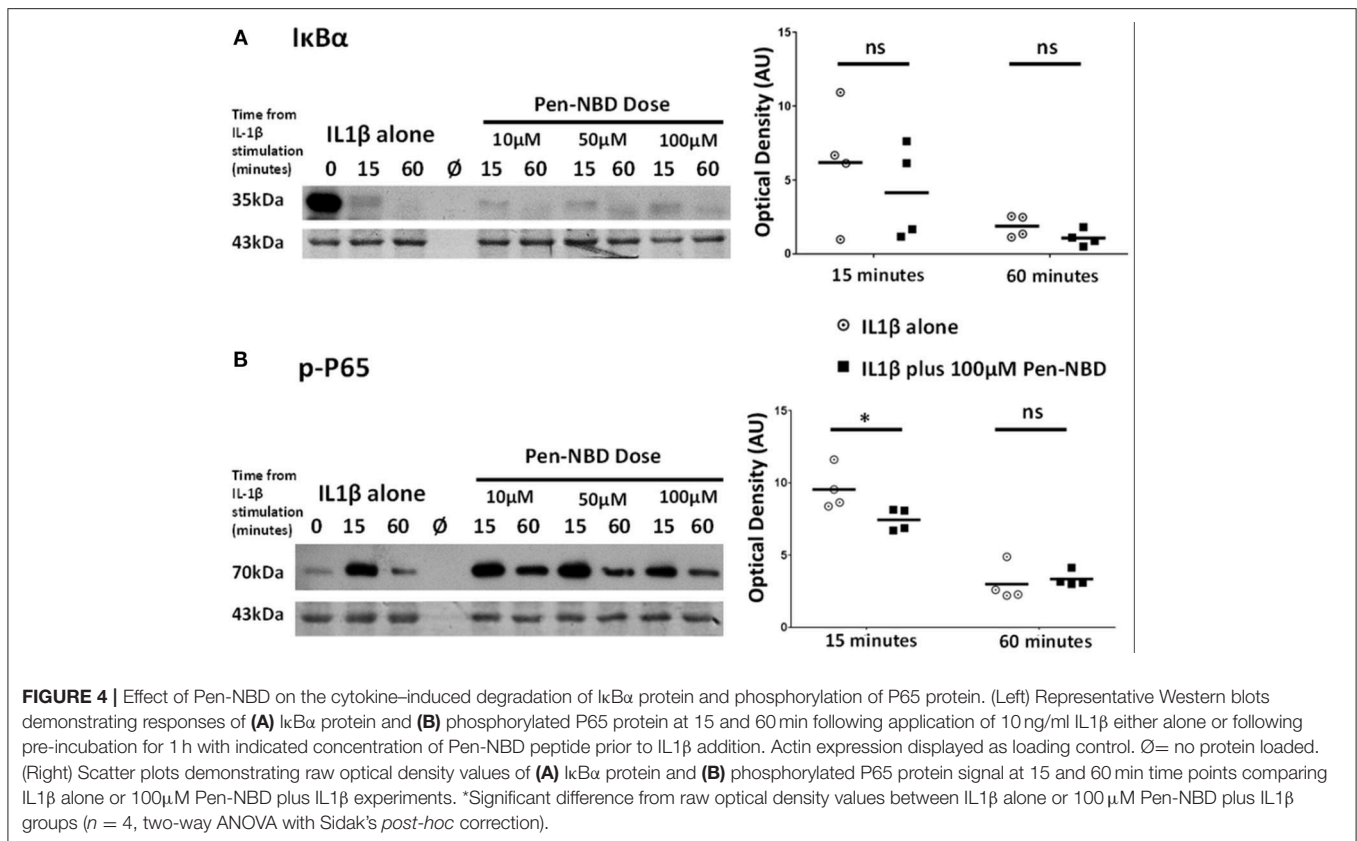
Application of either Pen-NBD, Pen(43-56)-NBD or Sc514, 1 h prior to cytokine exposure, significantly inhibited the IL1 β -stimulated expression of many genes within the array panel. The pattern of inhibition can be observed in both **Table 3** and the scatter graphs demonstrating fold expression changes presented within the **Supplementary Data (Supplementary Figures 3A–E)**.

Of note, Pen-NBD and Sc514 had broad and similar anti-inflammatory effects and both approaches significantly inhibited NF κ B family gene expression and prostaglandin pathway expression. For the labor associated genes: Pen-NBD, but not Sc514, inhibited the cytokine-induced expression of the oxytocin

TABLE 3 | Effect of CPP-based or small molecule inhibitors on IL1 β induced gene expression changes in myometrial cells.

Gene group	Gene name	Inhibitor		
		Pen-NBD	Pen(43-56)-NBD	Sc514
Labor associated genes	OXTR	↓ ($p = 0.01$)	–	–
	MMP9	–	–	↓ ($p = 0.01$)
Inflammatory cytokines	IL1A	↓ ($p \leq 0.0001$)	↓ ($p = 0.003$)	↓ ($p = 0.001$)
	IL1B	↓ ($p \leq 0.0001$)	↓ ($p = 0.034$)	↓ ($p = 0.004$)
	TNF	–	–	↓ ($p = 0.0004$)
	IL16	↓ ($p = 0.016$)	–	↓ ($p = 0.0004$)
	IL18	↓ ($p = 0.023$)	–	↓ ($p = 0.0007$)
	ICAM1	–	–	↓ ($p = 0.04$)
Inflammatory chemokines	CXCL2	–	–	↓ ($p = 0.0013$)
	CXCL6	↓ ($p \leq 0.0001$)	↓ ($p \leq 0.0001$)	–
	CCL2	↓ ($p \leq 0.0001$)	↓ ($p = 0.0003$)	↓ ($p = 0.01$)
NF κ B family genes	NFKB1	↓ ($p = 0.0039$)	–	↓ ($p = 0.001$)
	NFKB2	↓ ($p = 0.01$)	–	↓ ($p = 0.045$)
	RELA	↓ ($p = 0.0049$)	–	–
	NFKBIZ	–	–	↓ ($p = 0.008$)
Prostaglandin pathway genes	PTGES	↓ ($p \leq 0.0001$)	↓ ($p = 0.003$)	–
	COX2	↓ ($p = 0.0007$)	–	↓ ($p = 0.01$)

Taken from the 42 gene array, the table demonstrates only the genes with significantly altered expression patterns between control (IL1 β alone) or treatment (IL1 β plus either 100 μ M Pen-NBD/100 μ M Pen(43-56)-NBD/50 μ M Sc514) groups. ↓ indicates significantly altered gene expression in presence of inhibitor compared with IL1 β alone. – indicates no significant change between groups. Scatter graphs demonstrating changes for each gene displayed can be found in **Supplementary Figure 3**.



receptor gene OXTR. In contrast, Sc514, but not Pen-NBD inhibited the gene encoding for the matrix metalloproteinase MMP 9.

Use of the truncated CPP vector Pen (43-56) combined with NBD demonstrated much more limited ability to inhibit cytokine induced changes in gene expression and did not affect the expression of NFκB pathway, prostaglandin, or labor associated genes. This further demonstrates the importance of appropriate vector selection for any treatment strategy involving CPP-Cargo delivery.

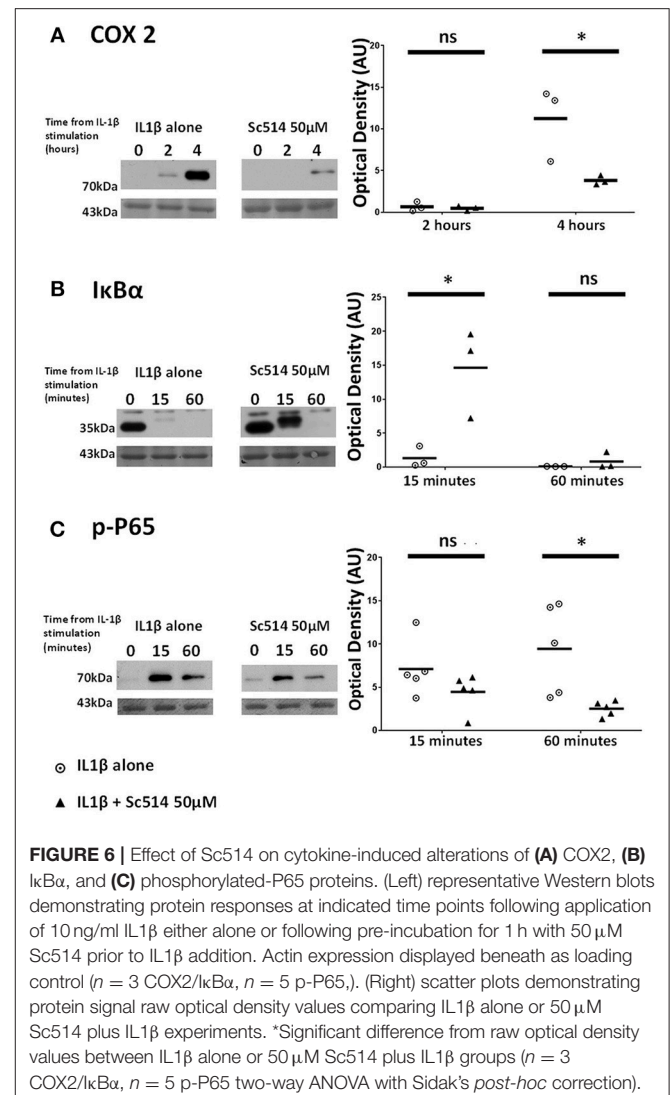
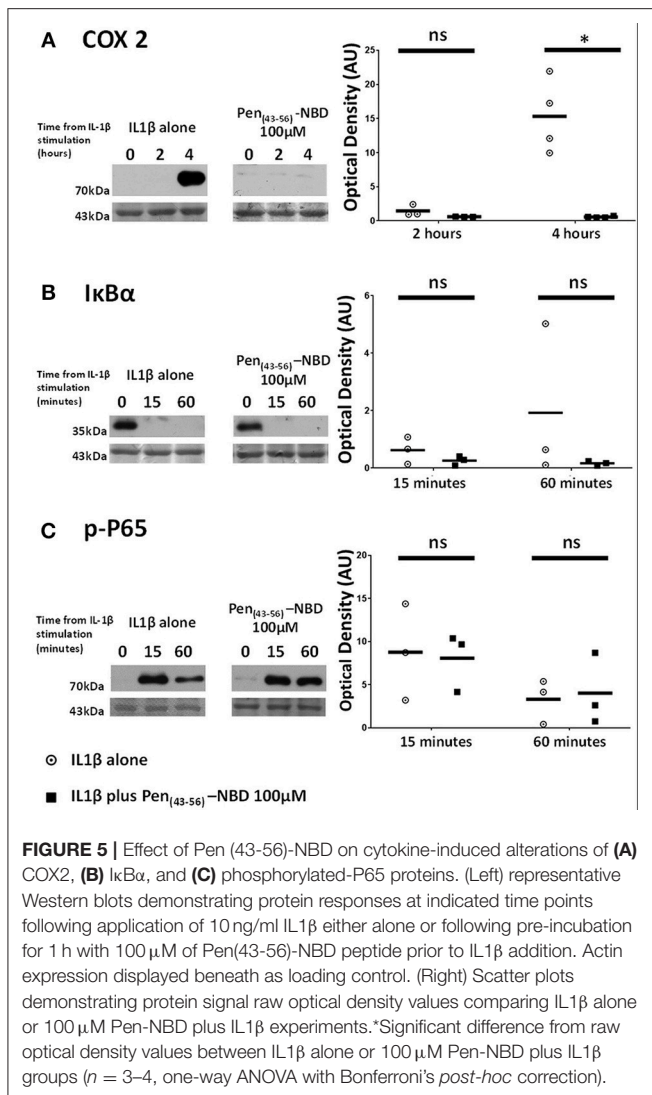
DISCUSSION

Within the reproductive literature a substantial body of evidence places intracellular NFκB activation centrally amongst the molecular pathways associated with inflammatory preterm birth (12, 44). With this in mind, a novel approach was adopted here to investigate inhibition of inflammatory signaling in human myometrial cells. We compared two classes of agent targeting the IKK complex upstream within the NFκB pathway, aiming to diminish NFκB-dependent uterine cell inflammatory responses: CPP's conjugated to the NBD peptide and the small molecule inhibitor Sc514.

The CPP-cargo construct, Pen-NBD, was used with the aim of avoiding the need for receptor-ligand interactions on the cellular membrane and thus to precisely deliver bioactive cargo

to the intended intracellular target. COX2 is an inducible rate-limiting enzyme in the production of prostaglandins in uterine cells. Prostaglandins are vital mediators in the induction of laboring processes and potent uterotonins that can initiate uterine contractions therefore the demonstration in this study that CPP mediated delivery of the NBD peptide inhibits the IL1β-induced expression of COX2 is of importance. A previous study examining the effect of the NBD peptide in a sheep model found it to be broadly ineffective at dampening inflammatory responses in sheep fetal membranes at 10 μM (45). This work supports that conclusion, but also emphasizes the importance of basic testing of novel compounds over a broad time and concentration range as inhibition of inflammatory responses was only observed at concentrations greater than or equal to 50 μM within the experiments presented here.

Hundreds of CPP sequences have now been discovered that could be linked to the NBD peptide. Even amongst well-characterized CPPs such as Penetratin or TAT differences exist within the literature as to the precise amino acid structure used (46), and differences in the position of key amino acid residues or charged groups within the CPP vector-cargo conjugate can alter the efficiency of delivery, with the possibility that this may in turn affect the biological effectiveness and targeting of the peptide (47, 48). In order to explore this effect of vector alteration on the overall efficacy of CPP-cargo combination, this study also examined NBD delivery and biological effect when combined to a truncated form of Penetratin CPP (Pen 43-56) that has



two lysine amino acid residues removed from the C-terminal end of the peptide (49). Conjugated to rhodamine Pen(43-56)-NBD demonstrated ability to enter myometrial cells within a similar time-dose range to Pen-NBD, but it was notable that the intracellular appearances of fluorophore were larger, possibly due to a clumping phenomenon secondary to altered charge and pH properties of the truncated CPP-cargo construct. Pen(43-56)-NBD also required increased concentrations to inhibit cytokine-induced increases in COX2 protein when compared to the non-truncated form and did not prevent alterations of NF κ B pathway proteins downstream of IKK complex activation. Additionally, pen(53-46)-NBD was less effective in attenuating IL1 β -induced gene expression changes compared with Pen-NBD peptide. Thus, careful selection of the CPP vector used, consideration of vector-cargo interactions and investigation of mechanisms of action are vital considerations for work involving peptide-based therapies.

The small molecule inhibitor Sc514 was chosen to compare this CPP-based therapeutic approach as it has a putatively very similar mechanism of action focussed on interaction with IKK

complex to prevent its activation within the canonical NF κ B pathway. As with Pen-NBD, Sc514 demonstrated ability to inhibit cytokine-induced COX2 production and p65 phosphorylation. However, whereas Pen-NBD did not prevent the IL1 β -dependent degradation of I κ B α , Sc514 demonstrated ability to inhibit degradation of this protein. Therefore, the ability of Sc514 to prevent the degradation of this protein in comparison to the NBD cargo may relate to subtle differences in mechanism of action. The IKK β subunit is primarily responsible for the phosphorylation of I κ B α from which subsequent degradation occurs and Sc514 has previously demonstrated specificity toward IKK β inhibition (35), whereas NBD peptide blocks association between NEMO and the other subunit portions of the IKK complex (29). Thus, the inability of Pen-NBD to prevent I κ B α degradation in myometrial cells could either be due to an incapacity to dampen IKK β -specific responses or to the actions of alternative IKK complexes within the cell that can lack Nemo Binding Domain (50).

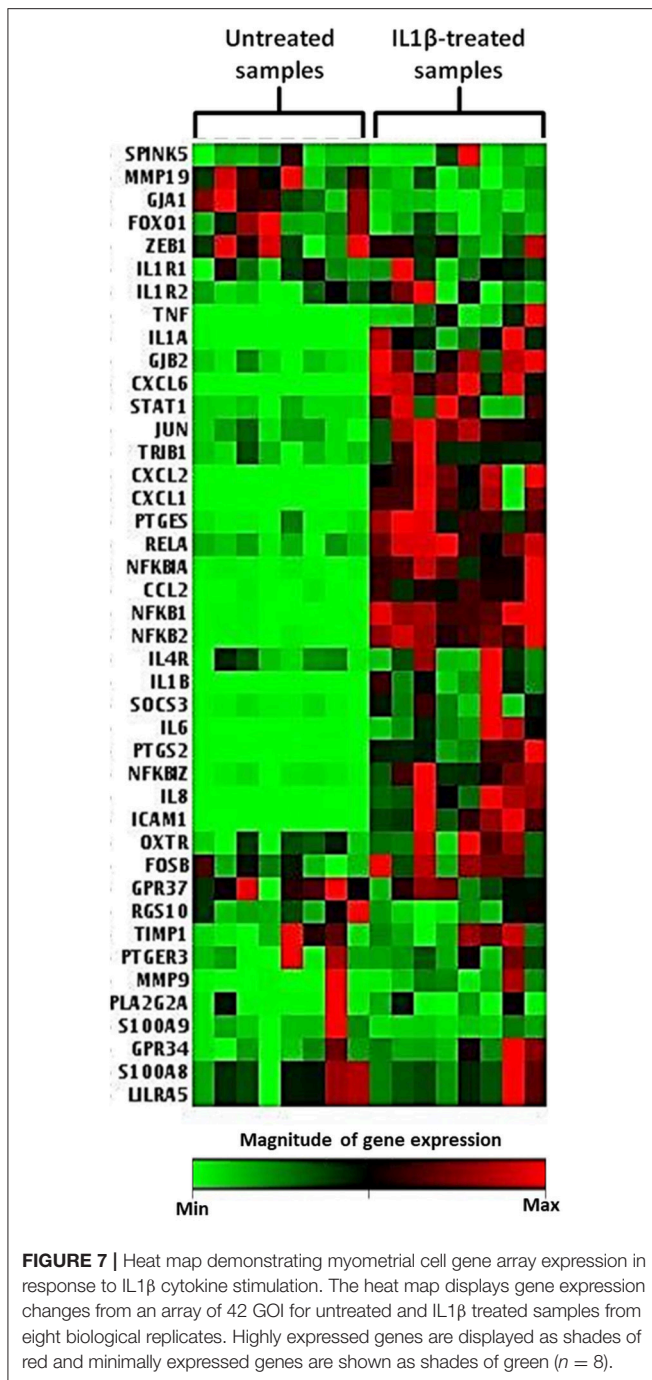


FIGURE 7 | Heat map demonstrating myometrial cell gene array expression in response to IL1 β cytokine stimulation. The heat map displays gene expression changes from an array of 42 GOI for untreated and IL1 β treated samples from eight biological replicates. Highly expressed genes are displayed as shades of red and minimally expressed genes are shown as shades of green ($n = 8$).

To examine the wider anti-inflammatory effects of both CPP-cargo and small molecular inhibitory approaches the expression of a bespoke array of genes was used to examine myometrial gene expression changes in response to cytokine stimulation, and, subsequently, the effect of the inhibitory approaches to dampen such responses. Application of IL1 β to myometrial cells produced many gene expression changes that have been previously observed in comparisons between human laboring and non-laboring samples including upregulation of NF κ B pathway components, cytokines such as IL6 and IL8 and genes

with specific associations with human labor such as PTGS2 and OXTR (51, 52).

Sc514 and Pen-NBD had similar anti-inflammatory effects dampening the cytokine-provoked transcription of NF κ B family genes and key cytokines such as IL6 and IL8. There were interesting variations in this effect with Pen-NBD demonstrating inhibition of OXTR, the gene transcribing for oxytocin receptor protein; whereas Sc514 inhibited the cytokine-induced upregulation of the matrix metalloproteinase enzyme gene MMP9. Such variations suggest that with future work to clarify the detailed mechanistic processes behind the varied presentations of preterm birth, different inhibitory approaches could be taken dependent on individual biological situations. It also gives rise to the possibility that in situations where a broad insult is responsible for the onset of preterm birth, more than one agent may be required to prevent this outcome.

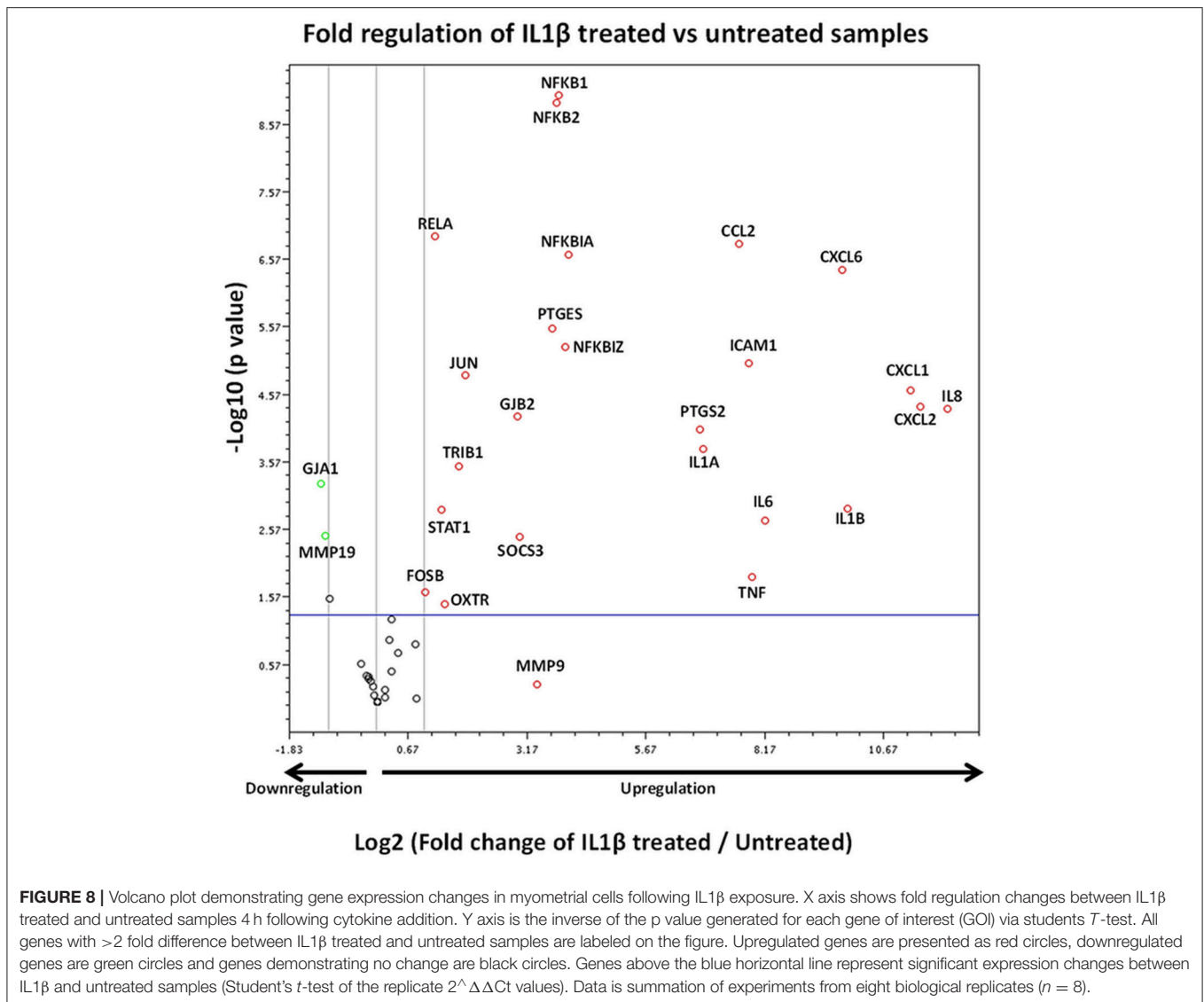
LIMITATIONS

A consistent limitation of fundamental biological studies investigating several outcomes of multiple interventions when using primary sourced human material is tissue availability. This can be further exacerbated when studies refrain from using cells at high passage (>4) to minimize potential deviation from original cellular phenotype. Within such practical and temporal constraints, the final numbers used in some experiments within this study are low; however, the authors feel this does not detract from the innovative nature of the study, and offers incentive for more detailed investigation of the uses of CPP based therapies and Sc514 as inflammatory and potentially contractile inhibitors for use in the field of pregnancy research.

CONCLUSION

Due to the shortage of candidate agents aimed at the prevention of preterm birth available for clinical testing, a number of approaches have been investigated with the intention of dampening inflammatory-mediated signaling in uterine cells (53, 54). However, most of these, outside of anti-infection strategies, rely upon inhibiting plasmalemmal receptor engagement with cognate ligands. This study presents a novel approach to addressing this problem by establishing that a biologically active cargo, intracellularly delivered by a CPP, can inhibit inflammation-related signaling in human myometrial cells in a similar manner to that of the potent cell-permeable small molecule inhibitor Sc514.

CPPs have the potential to deliver a diverse range of cargo to cells thus the proof of concept demonstrated here that CPP-cargo constructs enter human uterine smooth muscle cells and dampen inflammatory pathways in a targeted fashion is encouraging for the further pursuit of strategies involving varied CPP-cargo constructs that could include targeting alternative inflammatory pathways associated with preterm birth such as MAP kinase and AP1 (10, 11). It is well-established that many protein-protein interactions are classed as non-druggable and cannot be inhibited with small molecule drugs or large proteins and this



offers an opportunity for investigating more peptide therapeutics delivered by CPPs.

Although further extensive preclinical testing of these therapeutic strategies is required to establish safety and efficacy over a broad concentration range; the authors envisage the future usage of such agents in an acute setting for pregnant patients presenting with the clinical phenotype of regular contractions before 37 weeks of completed gestation with the intention of both delaying birth to reduce the impact of prematurity on the fetus, and ameliorating the effect of inflammatory insult to both mother and fetus. Treatment delivery could be either through local application to gestational tissues via vaginal pessary; or techniques are evolving quickly for tissue and/or cell-specific delivery of biologically active mediators, including those directed toward uteroplacental environments (55, 56), enabling the consideration of systemic usage. Thus, there is much promise for extending in the near future the use of Pen-NBD,

other CPP-cargo conjugates and Sc514 to preclinical models of preterm birth.

AUTHOR CONTRIBUTIONS

LG produced data through experimentation and analyzed and presented data, he also wrote the manuscript and finalized the final paper for submission. JT produced data through experimentation and analyzed and presented data. W-CT provided assistance and advice regarding the use of Sc514. AJ and his group provided collaborative supervision for the use of cell penetrating peptides and live cell microscopy and edited the manuscript. SR provided clinical supervision and oversight throughout the research project, helped facilitate the collection and use of human samples and edited the manuscript. MT oversaw and provided supervision for the whole project and in depth editing and direction for the manuscript.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fimmu.2018.02966/full#supplementary-material>

REFERENCES

- Blencowe H, Cousens S, Chou D, Oestergaard M, Say LA, Moller B, et al. Born Too Soon: The global epidemiology of 15 million preterm births. *Reprod Health* (2013) 10(Suppl. 1): S2. doi: 10.1186/1742-4755-10-S1-S2
- Costeloe KL, Hennessy EM, Haider S, Stacey F, Marlow N, Draper ES. Short term outcomes after extreme preterm birth in England: comparison of two birth cohorts in 1995 and 2006 (the EPICure studies). *BMJ* (2012) 345:e7976. doi: 10.1136/bmj.e7976
- Romero R, Espinoza J, Kusanovic JP, Gotsch F, Hassan S, Erez O, et al. The preterm parturition syndrome. *BJOG* (2006) 113 (Suppl. 3):17–42. doi: 10.1111/j.1471-0528.2006.01120.x
- Thomson A, Telfer J, Young A, Campbell S, Stewart C, Cameron I, et al. Leukocytes infiltrate the myometrium during human parturition: further evidence that labour is an inflammatory process. *Hum Reprod*. (1999) 14:229–236. doi: 10.1093/humrep/15.1.229
- Osman I, Young A, Ledingham MA, Thomson AJ, Jordan F, Greer IA, Norman JE. Leukocyte density and pro-inflammatory cytokine expression in human fetal membranes, decidua, cervix and myometrium before and during labour at term. *Mol. Hum. Reprod*. (2003) 9:41–5. doi: 10.1093/molehr/gag001
- Gomez-Lopez N, Estrada-Gutierrez G, Jimenez-Zamudio L, Vega-Sanchez R, Vadillo-Ortega F. Fetal membranes exhibit selective leukocyte chemotactic activity during human labor. *J Reprod Immunol*. (2009) 80:122–31. doi: 10.1016/j.jri.2009.01.002
- Mitchell BF, Taggart MJ. Are animal models relevant to key aspects of human parturition? *Am J Physiol Regul Integr Comp Physiol*. (2009) 297:R525–45. doi: 10.1152/ajpregu.00153.2009
- Hamilton S, Oomomian Y, Stephen G, Shynlova O, Tower CL, Garrod A, et al. Macrophages infiltrate the human and rat decidua during term and preterm labor: evidence that decidual inflammation precedes labor. *Biol Reprod* (2012) 39:31–9. doi: 10.1095/biolreprod.111.095505
- Singh N, Herbert B, Sooranna GR, Orsi NM, Edey L, Dasgupta T, et al. Is myometrial inflammation a cause or a consequence of term human labour? *J Endocrinol*. (2017) 235:69–83. doi: 10.1530/JOE-17-0318
- Lappas M, Permezel M, Rice GE. Mitogen-activated protein kinase proteins regulate LPS-stimulated release of pro-inflammatory cytokines and prostaglandins from human gestational tissues. *Placenta* (2007) 28:936–45. doi: 10.1016/j.placenta.2007.02.009
- MacIntyre DA, Lee YS, Migale R, Herbert BR, Waddington SN, Peebles D, et al. Activator protein 1 is a key terminal mediator of inflammation-induced preterm labor in mice. *FASEB J*. (2014) 28:2358–68. doi: 10.1096/fj.13-247783
- Lindstrom TM, Bennett PR. The role of nuclear factor kappa B in human labour. *Reproduction* (2005) 130:569–81. doi: 10.1530/rep.1.00197
- Perkins ND. Integrating cell-signalling pathways with NF-kappaB and IKK function. *Nat Rev Mol Cell Biol*. (2007) 8:49–62. doi: 10.1038/nrm2083
- Cookson VJ, Chapman NR. NF-kappaB function in the human myometrium during pregnancy and parturition. *Histol Histopathol*. (2010) 25:945–56. doi: 10.14670/HH-25.945
- Orange JS, May MJ. Cell penetrating peptide inhibitors of nuclear factor-kappa B. *Cell Mol Life Sci*. (2008) 65:3564–91. doi: 10.1007/s00018-008-8222-z
- Hayden MS, Ghosh S. Signaling to NF-kappaB. *Genes Dev*. (2004) 18:2195–224. doi: 10.1101/gad.1228704
- Belt AR, Baldassare JJ, Molnar M, Romero R, Hertelendy F. The nuclear transcription factor NF-kappaB mediates interleukin-1beta-induced expression of cyclooxygenase-2 in human myometrial cells. *Am J Obstet Gynecol*. (1999) 181:359–66. doi: 10.1016/S0002-9378(99)70562-4
- Gibb W. The role of prostaglandins in human parturition. *Ann Med*. (1998) 30:235–41. doi: 10.3109/07853899809005850
- Soloff MS, Cook DL Jr, Jeng YJ, Anderson GD. In situ analysis of interleukin-1-induced transcription of cox-2 and il-8 in cultured human myometrial cells. *Endocrinology* (2004) 145:1248–54. doi: 10.1210/en.2003-1310
- Choi SJ, Oh S, Kim JH, Roh CR. Changes of nuclear factor kappa B (NF-kappaB), cyclooxygenase-2 (COX-2) and matrix metalloproteinase-9 (MMP-9) in human myometrium before and during term labor. *Eur J Obstet Gynecol Reprod Biol*. (2007) 132:182–8. doi: 10.1016/j.ejogrb.2006.07.024
- Lye SJ, Nicholson BJ, Mascarenhas M, MacKenzie L, Petrocelli T. Increased expression of connexin-43 in the rat myometrium during labor is associated with an increase in the plasma estrogen:progesterone ratio. *Endocrinology* (1993) 132:2380–6. doi: 10.1210/endo.132.6.8389279
- Terzidou V, Lee Y, Lindstrom T, Johnson M, Thornton S, Bennett PR. Regulation of the human oxytocin receptor by nuclear factor-kappaB and CCAAT/enhancer-binding protein-beta. *J Clin Endocrinol Metab*. (2006) 91:2317–26. doi: 10.1210/jc.2005-2649
- Haas DM, Caldwell DM, Kirkpatrick P, McIntosh JJ, Welton NJ. Tocolytic therapy for preterm delivery: systematic review and network meta-analysis. *BMJ* (2012) 345:e6226. doi: 10.1136/bmj.e6226
- Fosgerau K, Hoffmann T. Peptide therapeutics: current status and future directions. *Drug Discov Today* (2015) 20:122–8. doi: 10.1016/j.drudis.2014.10.003
- Copolovici DM, Langel K, Eriste E, Langel U. Cell-penetrating peptides: design, synthesis, and applications. *ACS Nano* (2014) 8:1972–94. doi: 10.1021/nn4057269
- Jones AT, Sayers EJ. Cell entry of cell penetrating peptides: tales of tails wagging dogs. *J Control Release* (2012) 161:582–91. doi: 10.1016/j.jconrel.2012.04.003
- Marqus S, Pirogova E, Piva TJ. Evaluation of the use of therapeutic peptides for cancer treatment. *J Biomed Sci*. (2017) 24:21. doi: 10.1186/s12929-017-0328-x
- Guidotti G, Brambilla L, Rossi D. Cell-penetrating peptides: from basic research to clinics. *Trends Pharmacol Sci*. (2017) 38:406–24. doi: 10.1016/j.tips.2017.01.003
- May MJ, D'Acquisto F, Madge LA, Glockner J, Pober JS, Ghosh S. Selective inhibition of NF-kappaB activation by a peptide that blocks the interaction of NEMO with the IkappaB kinase complex. *Science* (2000) 289:1550–4. doi: 10.1126/science.289.5484.1550
- Reay DP, Yang M, Watchko JF, Daoud M, O'Day TL, Rehman KK, et al. Systemic delivery of NEMO binding domain/IKKgamma inhibitory peptide to young mdx mice improves dystrophic skeletal muscle histopathology. *Neurobiol Dis*. (2011) 43:598–608. doi: 10.1016/j.nbd.2011.05.008
- McCorkell KA, May MJ. NEMO-binding domain peptide inhibition of inflammatory signal-induced NF-kappaB activation *in vivo*. *Methods Mol Biol*. (2015) 1280:505–25. doi: 10.1007/978-1-4939-2422-6_30

32. Roy A, Mondal S, Kordower JH, Pahan K. Attenuation of microglial RANTES by NEMO-binding domain peptide inhibits the infiltration of CD8 T cells in the nigra of hemiparkinsonian monkey. *Neuroscience* (2015) 302:36–46. doi: 10.1016/j.neuroscience.2015.03.011
33. Gupta SC, Sundaram C, Reuter S, Aggarwal BB. Inhibiting NF-kappaB activation by small molecules as a therapeutic strategy. *Biochim Biophys Acta* (2010) 1799:775–87. doi: 10.1016/j.bbagr.2010.05.004
34. Mandery K, Glaeser H, Fromm MF. Interaction of innovative small molecule drugs used for cancer therapy with drug transporters. *Br J Pharmacol.* (2012) 165:345–62. doi: 10.1111/j.1476-5381.2011.01618.x
35. Kishore N, Sommers C, Mathialagan S, Guzova J, Yao M, Hauser S, et al. A selective IKK-2 inhibitor blocks NF-kappa B-dependent gene expression in interleukin-1 beta-stimulated synovial fibroblasts. *J Biol Chem.* (2003) 278:32861–71. doi: 10.1074/jbc.M211439200
36. De Silva D, Mitchell MD, Keelan JA. Inhibition of choriodecidual cytokine production and inflammatory gene expression by selective I-kappaB kinase (IKK) inhibitors. *Br J Pharmacol.* (2010) 160:1808–22. doi: 10.1111/j.1476-5381.2010.00839.x
37. Karolczak-Bayatti M, Sweeney M, Cheng J, Edey L, Robson SC, Ulrich SM, et al., (2011). Acetylation of heat shock protein 20 (Hsp20) regulates human myometrial activity. *J Biol Chem.* 286:34346–55. doi: 10.1074/jbc.M111.278549
38. Khaja KRP. Comparison of functional protein transduction domains using the NEMO binding domain peptide. *Pharmaceuticals* (2010) 3:110–24. doi: 10.3390/ph3010110
39. Khammanit R, Chantakru S, Kitiyanant Y, Saikhun J. Effect of serum starvation and chemical inhibitors on cell cycle synchronization of canine dermal fibroblasts. *Theriogenology* (2008) 70:27–34. doi: 10.1016/j.theriogenology.2008.02.015
40. Sayers EJ, Cleal K, Eissa NG, Watson P, Jones AT. Distal phenylalanine modification for enhancing cellular delivery of fluorophores, proteins and quantum dots by cell penetrating peptides. *J Control Release* (2014) 195:55–62. doi: 10.1016/j.jconrel.2014.07.055
41. Challis JRG, Matthews SG, Gibb W, Lye SJ. Endocrine and paracrine regulation of birth at term and preterm. *Endocr Rev.* (2000) 21:514–50. doi: 10.1210/er.21.5.514
42. Webster SJ, Waite SL, Cookson VJ, Warren A, Khan R, Gandhi SV, et al. Regulation of GTP-binding protein (Galphas) expression in human myometrial cells: a role for tumor necrosis factor in modulating Galphas promoter acetylation by transcriptional complexes. *J Biol Chem.* (2013) 288:6704–16. doi: 10.1074/jbc.M112.440602
43. Chan YW, van den Berg H. A, Moore JD, Quenby S, Blanks AM. Assessment of myometrial transcriptome changes associated with spontaneous human labour by high-throughput RNA-seq. *Exp Physiol.* (2014) 99:510–24. doi: 10.1113/expphysiol.2013.072868
44. Sakowicz A. The role of NFkappaB in the three stages of pregnancy - implantation, maintenance, and labour: a review article. *BJOG* (2018) 125:1379–87. doi: 10.1111/1471-0528.15172
45. Stinson LE, Ireland DJ, Kemp MW, Payne MS, Stock SJ, Newnham JP, et al. Effects of cytokine-suppressive anti-inflammatory drugs on inflammatory activation in *ex vivo* human and ovine fetal membranes. *Reproduction* (2014) 147:313–20. doi: 10.1530/REP-13-0576
46. Gautam A, Singh H, Tyagi A, Chaudhary K, Kumar R, Kapoor P, et al. CPPsite: a curated database of cell penetrating peptides. *Database* (2012) 2012:bas015. doi: 10.1093/database/bas015
47. Wender PA, Mitchell DJ, Pattabiraman K, Pelkey ET, Steinman L, Rothbard JB. The design, synthesis, and evaluation of molecules that enable or enhance cellular uptake: Peptoid molecular transporters. *Proc Natl Acad Sci USA.* (2000) 97:13003–8. doi: 10.1073/pnas.97.24.13003
48. Christiaens B, Grooten J, Reusens M, Joliot A, Goethals M, Vandekerckhove J, et al., (2004). Membrane interaction and cellular internalization of penetratin peptides. *Eur J Biochem.* 271:1187–97. doi: 10.1111/j.1432-1033.2004.04022.x
49. Fischer PM, Zhelev NZ, Wang S, Melville JE, Fähræus R, Lane DP. Structure-activity relationship of truncated and substituted analogues of the intracellular delivery vector Penetratin. *J Peptide Res.* (2000) 55:163–72. doi: 10.1034/j.1399-3011.2000.00163.x
50. Israël A. The IKK Complex, a Central Regulator of NF-κB Activation. *Cold Spring Harb Perspect Biol.* (2010) 2:a000158. doi: 10.1101/cshperspect.a000158
51. Bollopragada S, Youssef R, Jordan F, Greer I, Norman J, Nelson S. Term labor is associated with a core inflammatory response in human fetal membranes, myometrium, and cervix. *Am J Obst Gynecol.* (2009) 200:104.e101–104.e111. doi: 10.1016/j.ajog.2008.08.032
52. Mittal P, Romero R, Tarca AL, Gonzalez J, Draghici S, Xu Y, et al. Characterization of the myometrial transcriptome and biological pathways of spontaneous human labor at term. *J Perinat Med.* (2010) 38:617–43. doi: 10.1515/jpm.2010.097
53. Ng PY, Ireland DJ, Keelan JA. Drugs to block cytokine signaling for the prevention and treatment of inflammation-induced preterm birth. *Front Immunol.* (2015) 6:166. doi: 10.3389/fimmu.2015.00166
54. Garcia-Flores V, Romero R, Miller D, Xu Y, Done B, Veerapaneni C, et al. Inflammation-induced adverse pregnancy and neonatal outcomes can be improved by the immunomodulatory peptide exendin-4. *Front Immunol.* (2018) 9:1291. doi: 10.3389/fimmu.2018.01291
55. King A, Ndifon C, Lui S, Widdows K, Kotamraju VR, Agemy L, et al. Tumor-homing peptides as tools for targeted delivery of payloads to the placenta. *Sci Adv.* (2016) 2:e1600349. doi: 10.1126/sciadv.1600349
56. Beards F, Jones LE, Charnock J, Forbes K, Harris LK. Placental homing peptide-microRNA inhibitor conjugates for targeted enhancement of intrinsic placental growth signaling. *Theranostics* (2017) 7:2940–55. doi: 10.7150/thno.18845

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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