



ADGRE1 (EMR1, F4/80) Is a Rapidly-Evolving Gene Expressed in Mammalian Monocyte-Macrophages

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The F4/80 antigen, encoded by the *Adgre1* locus, has been widely-used as a monocyte-macrophage marker in mice, but its value as a macrophage marker in other species is unclear, and has even been questioned. ADGRE1 is a seven transmembrane G protein-coupled receptor with an extracellular domain containing repeated Epidermal Growth Factor (EGF)-like calcium binding domains. Using a new monoclonal antibody, we demonstrated that ADGRE1 is a myeloid differentiation marker in pigs, absent from progenitors in bone marrow, highly-expressed in mature granulocytes, monocytes, and tissue macrophages and induced by macrophage colony-stimulating factor (CSF1) treatment *in vivo*. Based upon these observations, we utilized RNA-Seq to assess the expression of *ADGRE1* mRNA in bone marrow or monocyte-derived macrophages (MDM) and alveolar macrophages from 8 mammalian species including pig, human, rat, sheep, goat, cow, water buffalo, and horse. *ADGRE1* mRNA was expressed by macrophages in each species, with inter-species variation both in expression level and response to lipopolysaccharide (LPS) stimulation. Analysis of the RNA-Seq data also revealed additional exons in several species compared to current Ensembl annotations. The ruminant species and horses appear to encode a complete duplication of the 7 EGF-like domains. In every species, Sashimi plots revealed evidence of exon skipping of the EGF-like domains, which are highly-variable between species and polymorphic in humans. Consistent with these expression patterns, key elements of the promoter and a putative enhancer are also conserved across all species. The rapid evolution of this molecule and related ADGRE family members suggests immune selection and a role in pathogen recognition.

Keywords: macrophage, monocyte, bone marrow, F4/80, porcine, ADGRE1/EMR1, adhesion G protein-coupled receptor E1

INTRODUCTION

The F4/80 antigen, first identified by Austyn and Gordon (1), was rapidly adopted as a marker for cells of the mononuclear phagocyte lineage in mice (2). F4/80 expression varies amongst mouse mononuclear populations, being very low or absent from osteoclasts, macrophages of T cell areas and marginal zone, lung alveolar macrophages, and the majority of classical dendritic cells [reviewed in (3)]. High expression of F4/80 has been proposed as a marker for populations of mouse tissue macrophages that derive from embryonic progenitors and renew independently of blood monocytes (4). The isolation of F4/80 cDNA (5) revealed that it encodes a large extracellular domain containing multiple Epidermal Growth Factor (EGF)-like calcium-binding domains, linked to a seven-transmembrane domain characteristic of G protein coupled receptors. The mouse gene encoding the F4/80 antigen was given the name *Emr1* (EGF module-containing mucin-like receptor) based upon homology to a previously-identified human cDNA (6). Subsequent studies identified related receptors (*EMR2*, *EMR3*, *EMR4*, *CD97*) collectively called EGF-TM7 proteins (5). These receptors are a subfamily of adhesion G protein coupled receptors (ADGRE) (7) and *Emr1* has been renamed *Adgre1*. Despite the high level of expression and lineage restriction in mouse mononuclear phagocytes, the knockout of *Adgre1* in the mouse germ line produced few phenotypic impacts, apart from apparent dysregulation of autoimmune responses and generation of regulatory T cells (8). One possible explanation for the lack of phenotype in mutant mice is the existence of a closely-related gene, *Adgre4* (*EMR4*), which is also expressed in mouse monocyte-macrophage lineage cells (9, 10). *Adgre1* mRNA is also highly-expressed by rat macrophages (11, 12).

Rodent macrophages differ significantly from humans in both constitutive and inducible gene expression profiles (13–15). In humans, ADGRE1 has been proposed as an eosinophil-specific marker. Monoclonal antibodies produced against the human protein bound specifically to eosinophils, and *ADGRE1* mRNA was apparently enriched in these cells compared to mononuclear phagocytes (16, 17). F4/80 antibody also binds to mouse eosinophils (18, 19). *Adgre1* mRNA in mice is also expressed by neutrophils, although the protein is not present on the cell surface (20). The apparent lack of *ADGRE1* in human monocyte-macrophages is not likely to be compensated by *ADGRE4* which was annotated as a pseudogene, due to a one base deletion which alters the reading frame before the transmembrane domains. The *ADGRE4* transcript, if expressed, could potentially encode a secreted extracellular domain, with the open reading frame encoding the seven-transmembrane domain retained intact and containing a start codon, and so is potentially translated separately (21). However, by contrast to rodents, the human genome contains two additional members of the EGF-TM7 family, *ADGRE2* and *ADGRE3*, which are both highly-expressed in myeloid cells (22). The receptors encoded

by these genes might fulfill functions mediated by ADGRE1 in rodents.

We have previously characterized peripheral blood monocytes and macrophages in the domestic pig and shown that pigs resemble humans in their patterns of constitutive and inducible gene expression (23–25). Pigs are increasingly recognized as a superior predictive model for the innate immune system and the pathology of human disease (26). We showed previously that *ADGRE1* mRNA is highly-expressed in several macrophage-rich pig tissues, and strongly-induced in the liver, associated with macrophage infiltration, following treatment with macrophage colony-stimulating factor (CSF1) (27). CSF1 treatment of mice was shown to increase the level of F4/80 on individual macrophages (28) and *Adgre1* mRNA was also strongly-induced in the liver of rats treated with CSF1 (12). Here we describe the production and characterization of an anti-pig ADGRE1 monoclonal antibody. The antibody binds strongly to monocytes and granulocytes in bone marrow and blood, and to tissue macrophages.

Our laboratory has also generated RNA-Seq expression profiles of CSF1-stimulated macrophages from rats and from multiple large animal species, including pig, sheep, goat, cow, water buffalo, and horse (29, 30). There are numerous human macrophage RNA-Seq datasets available in the public domain. Based upon the macrophage-enriched expression in the pig, we (re)examined the expression and complex alternative splicing of *ADGRE1* in each of these species. We conclude that *ADGRE1* is highly-expressed and regulated in macrophages in large animals, but that both the protein-coding sequence and the transcriptional regulation is divergent amongst species. We discuss the rapid evolution of these genes in mammals and speculate on their possible function.

MATERIALS AND METHODS

Ethics Statement

Approval for this research was obtained from The Roslin Institute and University of Edinburgh Protocols and Ethics Committees. All experiments were carried out under the authority of a UK Home Office Project Licence under the regulations of the Animals (Scientific Procedures) Act 1986.

Primary Cell Isolation and Culture

Isolation of primary pig bone marrow, peripheral blood mononuclear cells (PBMCs) and alveolar macrophages was carried out as described previously (24, 25) and all cells were cultured in full RPMI-1640 medium containing 10% HI-FCS (Sigma), 1 mM Glutamax, 100 U/ml penicillin and 100 µg/ml streptomycin (Invitrogen). Primary pig bone marrow cell cultures were supplemented with 10⁴ Units/ml rh-CSF1 to produce bone marrow-derived macrophages (BMDM) as described elsewhere (24).

In-vivo Treatment With pCSF1-Fc Recombinant Protein

Large white pigs were treated with pCSF1-Fc recombinant protein as previously described (27). In summary 8.5 week old

Abbreviations: ADGRE1, Adhesion G protein coupled receptor E1; BMDM, bone marrow-derived macrophages; CSF1, colony-stimulating factor 1; EGF, epidermal growth factor; HI-FCS, heat inactivated fetal calf serum; MDM, monocyte-derived macrophages; PBS-T, PBS containing 0.05% Tween-20.

animals were injected subcutaneously on three consecutive days with pCSF1-Fc (0.75 mg/kg; $n = 6$) or PBS control ($n = 5$). Blood was taken and all animals weighed 4 days prior to the first injections. Animals were euthanized by captive bolt following sedation with ketamine and azaperone, following which cells of interest were isolated and frozen for future analysis. Cells from a subset of these animals were utilized herein.

Construction of *Sus scrofa* ADGRE1 Chimeric Expression Vector

The full-length cDNA sequence of *Sus scrofa* ADGRE1 (Ensembl *S. scrofa* 10.2) was amplified using the primers fwd 5'-GGTCCTCACTCAATCTGCAAG-3' and rev 5'-GGAACAGCATTTTGGAAAGC-3', then cloned into pGEM[®]-T Easy vector (Promega). The cDNA encoding the four N-terminal extracellular EGF-like domains (1,082 bp) was extracted from the pGEM[®]-T Easy vector by PCR then cloned in frame into the EcoRI-EcoRV site of the pFUSE-hIgG1-Fc2 vector (InvivoGen) and sequenced by Edinburgh Genomics. We note that subsequent to our study, there are now sequences for pig ADGRE1 (GACC01000182.1/JAA53625.1) and ADGRE4 (GACC01000180.1/JAA53627.1) mRNA and proteins in the TSA archives.

Recombinant Protein Production

HEK293T cells grown in DMEM media supplemented with 10% Ig-depleted HI-FCS (Sigma), 1 mM Glutamax, 100 U/ml penicillin and 100 µg/ml streptomycin (Invitrogen) were transiently transfected with pADGRE1-pFUSE-hIgG1-Fc2 DNA at a ratio of 1:1 with Lipofectamine 2000 (Life Technologies) following manufacturer's instructions. Four days post-transfection, the supernatant was harvested, centrifuged to remove debris and filter sterilized. Soluble protein was then purified using a HiTrap Protein G column (GE Healthcare) and desalted using Slide-A-Lyzer[™] G2 Dialysis Cassettes 1–3 mL 10 K MWCO (Life Technologies). The recombinant protein was used as an immunogen for monoclonal antibody (mAb) production.

Monoclonal Antibody Production

BALB/c mice were purchased from Charles River Laboratories (UK). Hybridoma production was performed as previously described (31). In brief, three BALB/c female mice received a total of three subcutaneous immunisations with 50 µg of pADGRE1-hIgG1-Fc2 recombinant protein and TiterMax Gold adjuvant (Sigma) at least 14 days apart. Immune response was determined by screening sera collected from all mice prior to immunization and after the second and subsequent injections by indirect ELISA. A final intraperitoneal immunization of 50 µg protein in PBS without adjuvant was given and 4 days later splenocytes were harvested.

Splenocytes were fused with Sp2/0-Ag14 mouse myeloma cells (32) at a ratio of 5:1 using polyethylene glycol 1500 (Sigma), following standard procedures. Following fusion, fused hybridoma cells were grown in RPMI-1640 media supplemented with 10% IgG-depleted HI-FCS, 1 mM Glutamax, HAT Media Supplement Hybri-Max[™] (Sigma), Hybridoma Fusion and

Cloning Supplement (HFCS, Roche), recombinant mouse IL-6 (Sigma), 100 U/ml penicillin, and 100 µg/ml streptomycin. After seven to 10 days cells were switched to RPMI-1640 containing 10% IgG-depleted HI-FCS, 1 mM/l Glutamax, HT Media Supplement Hybri-Max, mouse IL-(6.25 ng/µl)6, 100 U/ml penicillin and 100 µg/ml streptomycin. An indirect ELISA was performed to identify hybridomas producing antibodies against porcine ADGRE1. Positive wells were expanded and subcloned to single cell colonies by limiting dilution. Supernatant was collected and purified by Protein G HiTrap column, with desalting carried out using Slide-A-Lyzer[™] G2 Dialysis Cassettes 1–3 mL 10 K MWCO.

The mAbs were isotyped using IsoStrip Mouse Monoclonal Antibody Isotyping Kit (Roche) according to manufacturer's instructions.

Detection of Pig ADGRE1 Protein by Indirect ELISA

Indirect ELISA was performed as previously described (33). In brief, microplates were coated with 50 µg of 1 µg/ml pADGRE1-pFUSE-hIgG1 or Human IgG1-Fc recombinant protein (R&D Systems) diluted in coating buffer and incubated at 4°C overnight. The following day plates were blocked in PBS, 1% BSA, 1% horse serum for 1 h, followed by three washes with PBS-0.05% Tween20. Fifty microliters of pre- or post-immune serum diluted 1:200 in PBS, neat supernatant from hybridoma cloning plates or 1 µg/ml purified mAb in PBS was added to individual wells and incubated for 1 h at room temperature. Following a further three washes in PBS-0.05% Tween20 50 µl of horse anti-mouse IgG-HRP (Cell Signaling) diluted 1:5,000 in PBS was added and incubated for 1 h at room temperature. After three further washes peroxidase activity was visualized using TMB Substrate (BD Biosciences). The reaction was stopped with 2 N hydrochloric acid. A Multiskan Ascent spectrophotometer was used to read absorbance at 450 nm.

Flow Cytometry

One million cells per sample were first blocked using PBS, 2% normal horse serum for 15 min on ice, then incubated with either neat supernatant for initial screening or a 1:100 dilution of Alexa-Fluor-647 conjugated purified mAb for 30 min on ice. Conjugation was performed using a commercially available kit (Molecular Probes) according to manufacturer's instructions. Signal was detected on unconjugated supernatant by incubation with anti-mouse IgG-allophycocyanin (1:400; BioLegend) for 30 min on ice. Following 3 washes in PBS, cells were re-suspended in 0.1% Sytox Blue (Invitrogen) in PBS for analysis using a Fortessa LSR flow cytometer (BD Biosciences). Double staining included mouse anti-pig CD14-FITC (1:50; BioRad), mouse anti-pig CD16-PE (1:200, BioRad), mouse anti-pig CD163-PE (1:100, BioRad) and mouse anti-pig SIRP-alpha-PE (1:400, Southern Biotech). Relevant isotype controls were included in all experiments. Data collection was performed using Diva software (BD Biosciences) and analysis using FlowJo v10 software. Welch's *t*-tests were performed on data and results are presented as treatment group means ± SE. All statistical analyses were

performed using GraphPad Prism 6.0 (GraphPad Software). A $p < 0.05$ was considered statistically significant.

Immunohistochemistry for ADGRE1 and CD163

Tissues were placed in phosphate buffered saline solution pH7 and mounted in OCT. Frozen sections (10 μm) were cut and mounted on Superfrost slides, then dried for 24 h at 4°C prior to use. The slides were fixed for 10 min in ice cold methanol then washed 2 \times 5 min in PBS. To block endogenous peroxidase activity the slides were incubated in 0.3% hydrogen peroxide (H_2O_2) in PBS then washed 2 \times 5 min in PBS. The slides were incubated for 1 h in a humidified chamber at room temperature in Tris-buffered saline with 20% normal goat serum and 5% BSA, then washed 2 \times 5 min in PBS. Immunostaining was performed using primary mAbs against mouse anti-pig ADGRE1 (ROS-4E12-3E6, 2.4 ng/ μL) and mouse anti-human CD163 (2 ng/ μL , BioRad); control slides had no primary antibody added. Slides were incubated overnight at 4°C then washed 2 \times 5 min in PBS. The primary antibodies were detected using anti-mouse IgG polymer (Vector MP-7402) incubated at room temperature for 1 h then washed 2 \times 5 min in PBS. The signal was detected using DAB peroxidase substrate (Vector Laboratories SK-4100) then washed 2 \times 5 min in water. Sections were counterstained with haematoxylin and eosin using a Leica Autostainer XL. Slides were then mounted, and microscopic analysis performed using the NanoZoomer-XR (Hamamatsu).

Sources of RNA-Seq Data

RNA-Seq data was previously generated by our lab for BMDM, or monocyte-derived macrophages (MDM) of rat, cattle, water buffalo, sheep, goat, pig and horse. All RNA-Seq libraries were paired-end, the read length for rat was 90 bp, for goat was 75 bp and for the remainder of the species was 124–126 bp. RNA-Seq data was also available for a subset of these species (sheep, water buffalo and goat) from alveolar macrophages. The sheep data was published as part of a comprehensive transcriptional atlas (34). A subset of data from the remaining species was previously analyzed in a comparative study of the regulation of genes involved in arginine metabolism and nitric oxide production (35), which generated gene-level expression estimates, as transcripts per million (TPM), using Kallisto v0.43.0 (36). The primary RNA-Seq data is available in the European Nucleotide Archive (ENA) using NCBI BioProject IDs PRJEB21180 (water buffalo), PRJEB19199 (sheep), PRJEB22535 (cattle, unpublished), PRJEB23119 (pig), PRJEB24920 (horse), PRJEB22553 (rat, unpublished), and PRJEB23196 (goat). Human alveolar macrophage, microglia, non-classical monocyte, and classical monocyte datasets were obtained from, respectively, Pinilla-Vera et al. (37); 10 RNA-Seq libraries within NCBI BioProject PRJNA428090, representing 6 untreated and 4 LPS-treated samples), Galatro et al. (38); 39 RNA-Seq libraries within NCBI BioProject PRJNA387182), Williams et al. (39); 17 RNA-Seq libraries within NCBI BioProject PRJNA339968), and

Mirsafian et al. (40); 1 RNA-Seq library within NCBI BioProject PRJNA264020).

RESULTS

Production and Characterization of a Monoclonal Antibody Against Pig ADGRE1

Bone marrow-derived macrophages and alveolar macrophages from the pig were previously profiled as part of a transcriptional atlas project (41). However, at the time of development of the microarray platform used, the *ADGRE1* locus was not annotated. The predicted pig cDNA was truncated at the 5' end and missed the start codon. The longest *ADGRE1* cDNA for the pig currently in Ensembl (version 91) predicts a protein of 939 amino acids with an extracellular domain containing 7 EGF-like calcium-binding domains, as in the mouse. The predicted amino acid sequence is only around 60% identical to mouse and almost all the variation resides in these EGF-like domains. The longest human cDNA in Ensembl encodes 6 EGF-like domains. Cross-species alignment of mouse and pig cDNA with the human genome reveals that there is an additional candidate exon encoding a seventh EGF-like domain in the human genome (not shown). We have confirmed the expression of this exon in human macrophage RNA-Seq data as discussed below. Including this exon, the longest human ADGRE1 protein is predicted to be 940 amino acids. A recent study (42) utilized RNA-Seq from pig alveolar macrophages to improve the annotation of genes within the pig immunome. Within that project, the alveolar macrophage data were used to assemble two alternative pig *ADGRE1* cDNAs. As part of the ongoing annotation of the pig genome, we have produced RNA-Seq data from pig BMDM grown in CSF1 and stimulated with LPS (29).

Based upon the gene expression data, and emerging evidence that pigs provide a useful model for the study of human macrophage biology (26), we decided to make an antibody against the pig protein, with the expectation that it could (a) enable studies of *ADGRE1* function in a relevant model, (b) confirm the expression of the ADGRE1 protein in macrophages, and (c) potentially cross-react with human ADGRE1. Previous antibodies against human *ADGRE1* were produced in hamsters, against a cell-expressed ADGRE1-CD97 fusion protein (16) and in mice, against a full length ADGRE1 extracellular domain-Fc fusion protein (17). As an immunogen, we constructed cDNA, expressed and purified a pig ADGRE1-Fc fusion protein, immunized mice, and screened hybridomas for specific binding to the fusion protein, but not to the human IgG recombinant tag. From amongst 29 primary hybridomas, eight were cloned, and each produced clones that bound specifically to the immunogen. One mAb, ROS-4E12-3E6, was selected for further detailed characterization. The remaining hybridoma supernatants were screened for cross-reactivity against human and cattle blood monocytes and sheep alveolar macrophages, but none were cross-reactive to any other species. This is likely a reflection of the extensive sequence divergence between the species.

Expression of ADGRE1 by Pig Leukocytes

We first examined surface expression of ADGRE1 in PBMC. **Figure 1A** shows that mAb ROS-4E12-3E6 bound to monocytes, identified in this case by their size and granularity profile. In PBMCs, ADGRE1 was highly-expressed on the entire CD14⁺⁺ monocyte population (**Figure 1B**). By contrast to humans, where the gene is duplicated, CD16 is not regulated in monocyte subsets in pigs and is uniformly high on the CD14⁺⁺ population (23). Accordingly, ADGRE1 was uniform on all CD16⁺⁺ cells (**Figure 1C**). The ADGRE1-negative, CD16^{lo} cells are most likely natural killer cells. The haptoglobin receptor, CD163, provides a marker for monocyte differentiation in pigs that varies inversely with CD14, with CD163^{hi} cells resembling intermediate monocytes in humans (23). Here, ADGRE1 expression did not vary with the level of expression of CD163 (**Figure 1D**).

The differentiation of the non-classical monocyte subset in other species depends upon CSF1 (43) and in mice, F4/80 is also induced further in tissue macrophages following CSF1 treatment (28). We previously developed a porcine CSF1-Fc fusion protein, with an enhanced greater circulating half-life (44). Injection of this protein into mice (44) or pigs (27) led to a substantial increase in blood monocyte and tissue macrophage numbers. Analysis of a subset of samples stored from the previous pig study confirmed that CSF1-Fc treatment significantly increased the percentage of ADGRE1⁺/CD14⁺ blood monocytes (**Figure 1E**), as well as the proportion of ADGRE1⁺/CD16⁺⁺ (**Figure 1F**) and ADGRE1⁺/CD163⁺⁺ monocytes (**Figure 1G**) within the PBMC fraction, when compared to cells from control animals. This result is consistent with the monocytosis described previously and supports the use of ADGRE1 as a pig monocyte marker. The median level of surface ADGRE1 was also increased on the monocytes from CSF1-Fc treated pigs (**Figure 1H**). In mice, the F4/80 antigen is expressed at very low levels on isolated alveolar macrophages (45). By contrast, and consistent with gene expression profiles reported by Dawson et al. (42), the level of cell surface ADGRE1 was uniformly exceptionally high on pig alveolar macrophages (**Figure 1I**). The identity of these cells as alveolar macrophages was confirmed by the co-expression of high levels of SIRP-alpha (CD172a), which is highly-expressed by monocytes and macrophages in the pig (23).

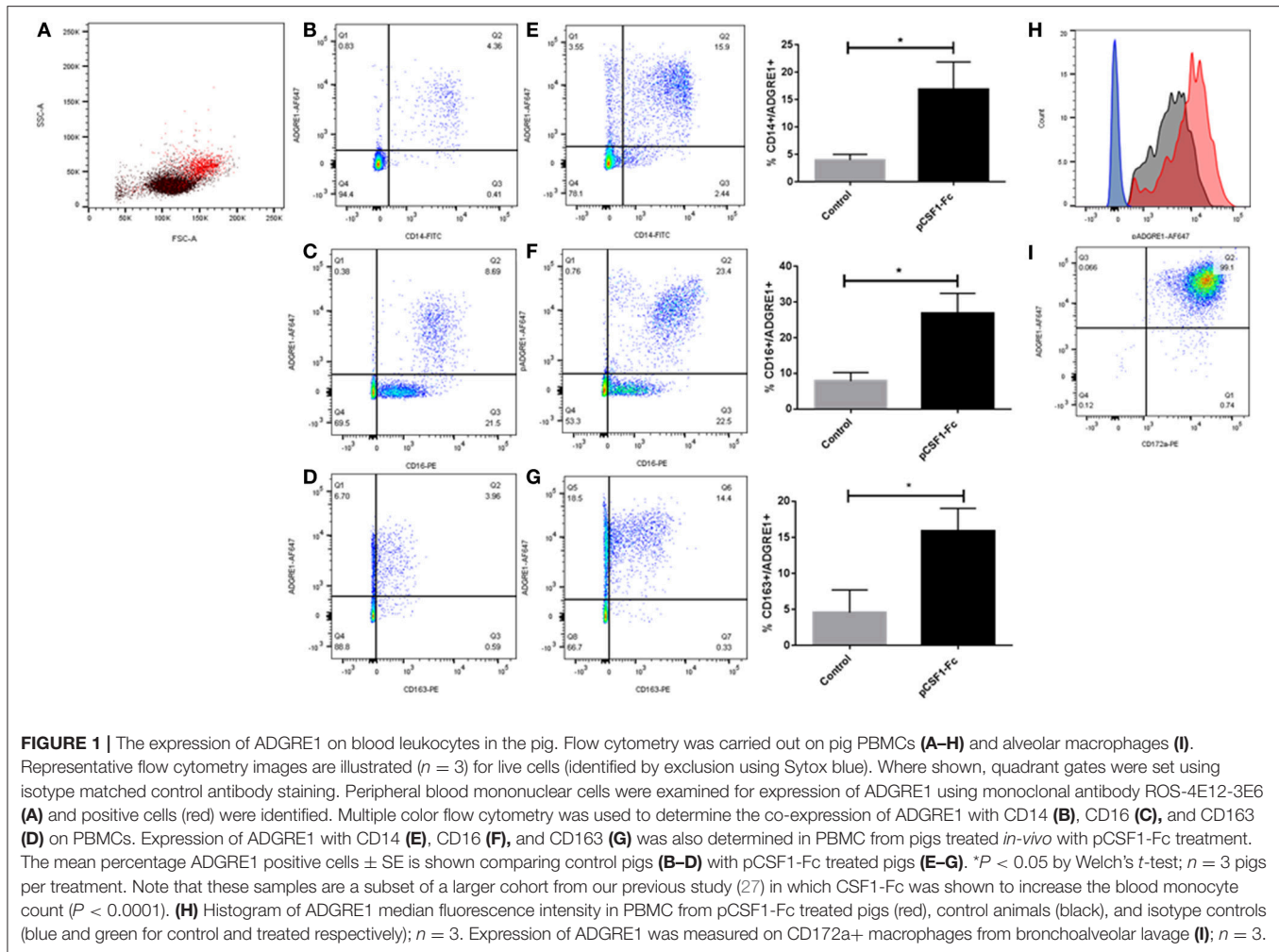
These initial observations indicated that ADGRE1 is highly-expressed on the surface of monocytes and macrophages in pigs, but with a distinct pattern compared to rodents. We next considered whether ADGRE1 was widely distributed amongst myeloid cell lineages, and whether it might provide a useful differentiation marker. **Figure 2A** shows that ADGRE1 was detected on presumptive granulocytes in bone marrow (SSC^{hi}). In the bone marrow mononuclear cell fraction, a minor population of larger cells was ADGRE1^{hi}, presumably the resident bone marrow macrophages, whereas expression was low and variable on small mononuclear cells. **Figure 2D** shows the fluorescence histogram, indicating relative uniformity of expression of ADGRE1 amongst the positive cells in unstimulated bone marrow. When bone marrow cells were cultivated in rhCSF1, the ADGRE1⁺/SSC^{hi} granulocyte

population largely disappeared within 3 days (**Figure 2B**), consistent with the short half-life of granulocytes in culture. At the same time, there was an evident expansion of an SSC^{lo}, presumptive macrophage population, that had higher levels of ADGRE1 (**Figure 2E**). After 7 days in rhCSF1 the population of bone marrow cells, largely comprised of adherent macrophages as previously described (24), was entirely SSC^{lo} and ADGRE1⁺ (**Figure 2C**). At this time, the level of expression of ADGRE1 on individual cells was heterogeneous (**Figure 2F**).

To confirm the expression of ADGRE1 on resident tissue macrophages, we stained sections of liver and lung. Compared to the negative control (**Figure 3A**) and CD163 positive control (**Figure 3B**) sections there was abundant expression of ADGRE1 on lung macrophages (**Figure 3C**), including stellate interstitial macrophages and more rounded, presumptive alveolar macrophages. *ADGRE1* mRNA was previously detected in expression profiles of whole pig liver and was massively induced by CSF1-Fc treatment (27). Compared to negative (**Figure 3D**) and CD163 (**Figure 3E**) positive sections, high levels of expression of ADGRE1 was also detected on stellate Kupffer cells in pig liver (**Figure 3F**), showing the same portal-centrilobular gradient observed in mouse liver (46).

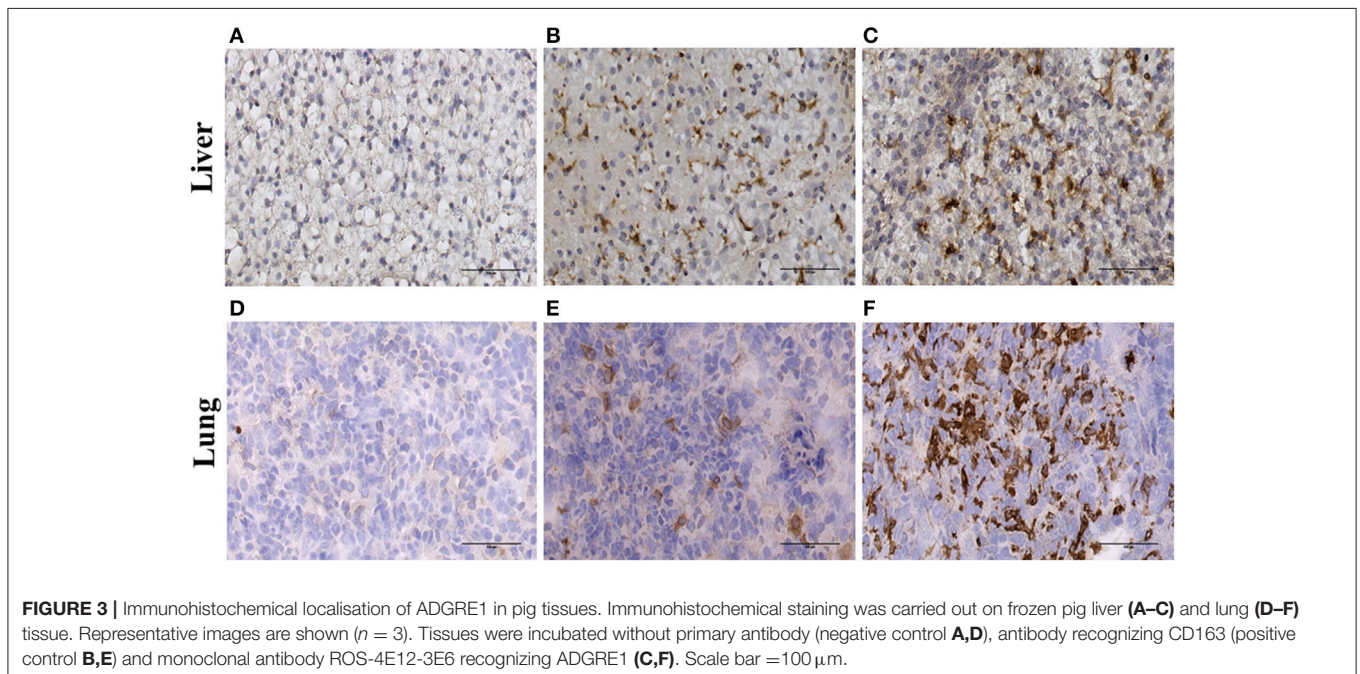
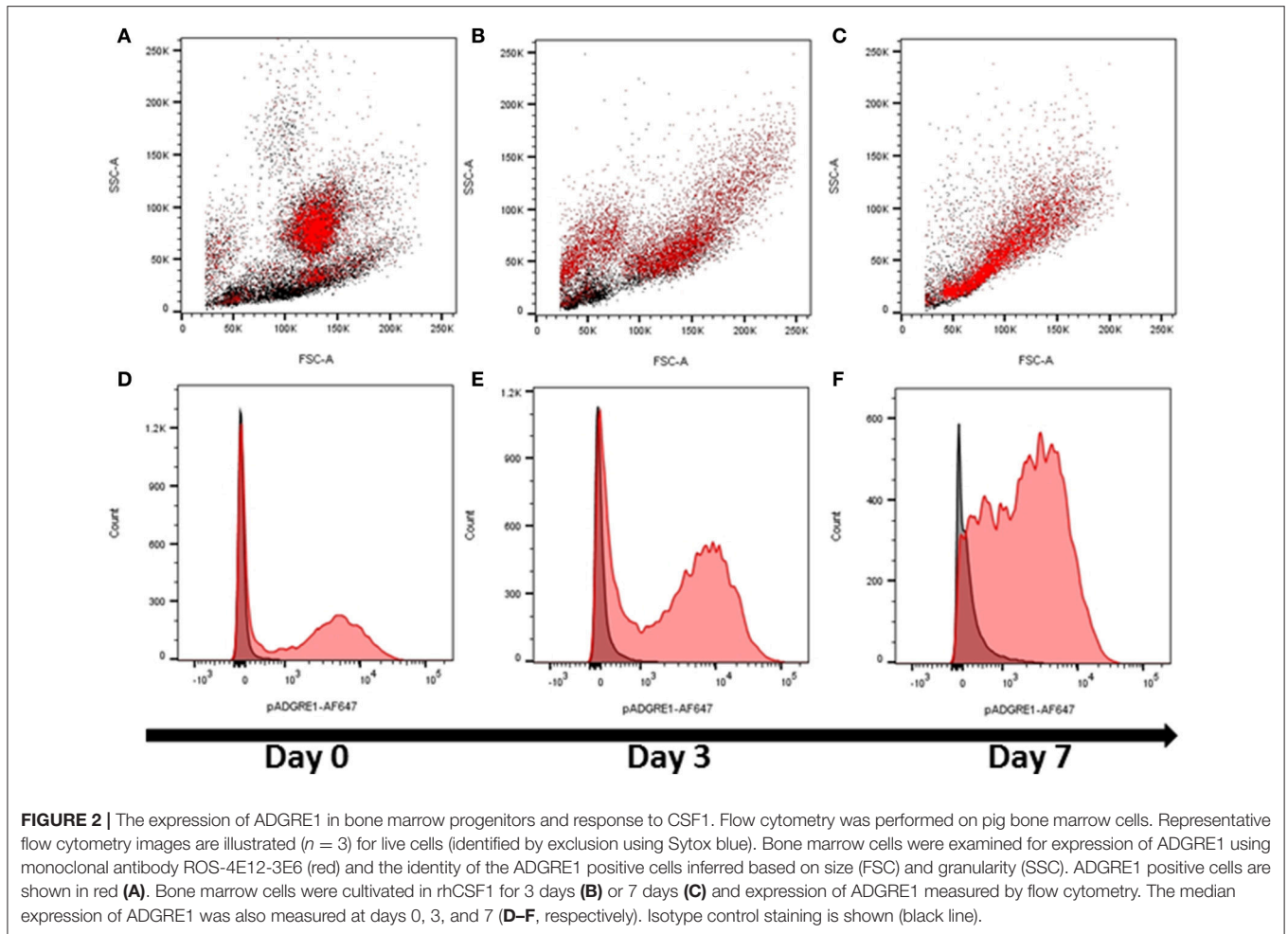
The Regulated Expression of *ADGRE1* mRNA in Human Macrophages

Two previous reports used RT-PCR to determine the expression of *ADGRE1* in human myeloid cell populations, and concluded that the mRNA was highly-expressed in eosinophils, and low or undetectable in other leukocyte populations (16, 17). This conclusion was at odds with the detection of the mRNA in mononuclear cells in the original cloning of human *EMRI* cDNA (6). Given the expression of *ADGRE1* in the pig, we concluded that either pigs are fundamentally different from humans, or the previous studies of *ADGRE1* expression in humans were incorrect. To address these two possibilities, we examined a range of data available in the public domain. An expression atlas of human cell populations generated based upon public domain microarray data indicates that *ADGRE1* mRNA is expressed at similar levels in neutrophils, monocytes, and alveolar macrophages [(47); see data on www.biogps.org]. The FANTOM5 consortium used cap-analysis of gene expression (CAGE, genome-scale 5' end tag sequencing) to analyze monocyte subsets, monocyte-derived macrophages, activated monocytes and macrophages, and many other myeloid populations (48, 49). The data can be visualized on the ZENBU viewer (<http://fantom.gsc.riken.jp/zenbu/>). This confirms *ADGRE1* is expressed in human eosinophils, but the level of expression was no higher than in CD16⁺⁺ monocytes. Transcription of *ADGRE1* in all the human myeloid lineages appears to be a differentiation marker, as expected, since mRNA was not detected in CD34⁺ stem cells or committed myeloid progenitors. Data from Maiga et al. (50) also indicate the high expression of *ADGRE1* in monocytes and relative absence from progenitors. The FANTOM5 data also reveal that *ADGRE1* was repressed in MDM grown in CSF1,



relative to the higher level of expression in blood monocytes but was strongly induced in MDM relatively late in the time course by LPS. Indeed, the highest expression of ADGRE1 detected in the FANTOM5 dataset was observed in LPS-stimulated cells (30). Comparative analysis of monocytes, MDM and isolated microglia (38) by RNA-Seq confirmed both monocyte expression and relative down-modulation in MDM. The increased expression of *ADGRE1* mRNA in CD16⁺⁺ human monocytes detected by CAGE is confirmed in three published microarray datasets (51, 52). A further marker, SLAN, dissects the CD16⁺⁺ monocyte subpopulation still further, with *ADGRE1* mRNA further elevated in the SLAN-positive population (53). *ADGRE1* mRNA was very highly expressed in human alveolar macrophages (54, 55) and isolated placental macrophages (56) enriched in the macrophage fraction isolated from human adipose tissue (57) and was readily detected in RNA-Seq data derived from human peritoneal macrophages (58). Taken together, all the evidence indicates that *ADGRE1* is robustly expressed in the human macrophage lineage and is not eosinophil-restricted. In humans, the related gene *ADGRE4* is located immediately adjacent to the *ADGRE1* locus. In mice, this gene, also known as FIRE

(9, 10), is located around 1.5 mb downstream of *ADGRE1* on chromosome 17. Human *ADGRE4* is annotated as a pseudogene, based upon the analysis of a cDNA that contains a 1 bp deletion relative to the *ADGRE4* gene in other primates, which shifts the reading frame before the transmembrane domain (21, 59). The FANTOM5 data places the transcription start site (TSS) around 7 kb upstream of the published cDNA. Since *ADGRE4* mRNA in other species (mouse, rat, dog, and pig) is around 50-100 bp longer at the 5' end than human, and since there is one fewer annotated exon compared to mouse, it seems likely that there is an alternative 5' non-coding exon. *ADGRE4* CAGE tags were even more enriched than *ADGRE1* in CD16⁺⁺ monocytes, compared to CD14⁺⁺ monocytes. In common with many other monocyte differentiation markers, the CD14⁺/CD16⁺ intermediate monocytes expressed *ADGRE4* at intermediate levels (49). Hence, *ADGRE4* is a novel marker for “non-classical” monocytes in humans. The human genome contains two other genes, *ADGRE2* and *ADGRE3*, related to *ADGRE1* and *ADGRE4* and located in the same region of chromosome 19. Consistent with previous reports (60, 61) in the FANTOM5 data, *ADGRE2* shows a very similar pattern of



regulation in human monocytes and MDM to *ADGRE1*, whereas *ADGRE3* is more highly-expressed and restricted to granulocytes, neutrophils and eosinophils.

***ADGRE1*, *ADGRE2*, *ADGRE3*, and *ADGRE4* Expression in Macrophages of Other Species**

As part of the generation of transcriptional atlases for several mammalian species, and a comparative analysis of macrophage-expressed genes (29) we have produced deep RNA-Seq data for rat, sheep, goat, cattle, water buffalo, pig, and horse macrophages produced by the cultivation of bone marrow cells in CSF1 using methods originally developed for the pig (24). We utilized these data to assess the expression of *ADGRE1* and *ADGRE4* and the response to LPS, in each of these species. **Table S1** summarizes the expression of *ADGRE1* and the related *ADGRE4* mRNA in BMDM, and the response to LPS, for each of the species examined, including human data from published RNA-Seq profiles. Where available, additional data is shown for isolated alveolar macrophages and for monocyte-derived macrophages. Several findings emerge from these data. Firstly, the highest level of *ADGRE1* mRNA expression detected was in rat BMDM, but *ADGRE1* was robustly expressed in macrophages in all of the species. Secondly, the level and pattern of expression was variable, even between ruminant species. For example, *ADGRE1* was highly-expressed in BMDM from sheep, 10-fold lower in cattle and water buffalo (but LPS-inducible as in human macrophages) and very low in goat. Where we have data available *ADGRE1* was highly expressed in alveolar macrophages in every large animal species. Of the species examined, only the pig had high expression of *ADGRE4* in BMDM. Like the mouse, the rat has only three members of the ADGRE family, *Adgre1*, *Adgre4*, and *Adgre5* (62) and *Adgre1* mRNA was very highly-expressed in rat BMDM. However, by contrast to mouse, *Adgre4* was barely-detectable in these cells.

ADGRE2 and *ADGRE3* are not well annotated in livestock species, and clear orthology relationships are unclear. For example, in Ensembl build 11.1, there are now 4 *ADGRE2*-like (ENSSSCG00000036342, ENSSSCG00000021675, ENSSSCG00000013788, ENSSSCG00000013785) and 2 *ADGRE3*-like (ENSSSCG00000013787, ENSSSCG00000026402) genes. In Ensembl, at this time *ADGRE2*- and *ADGRE3*-like genes are annotated as many-to-many orthologs with the other livestock species. We have included expression estimates for these transcripts in pig macrophages in **Table S1**. Only the two *ADGRE3*-like genes have significant expressions, and then 10-fold lower than *ADGRE1* or *ADGRE4*.

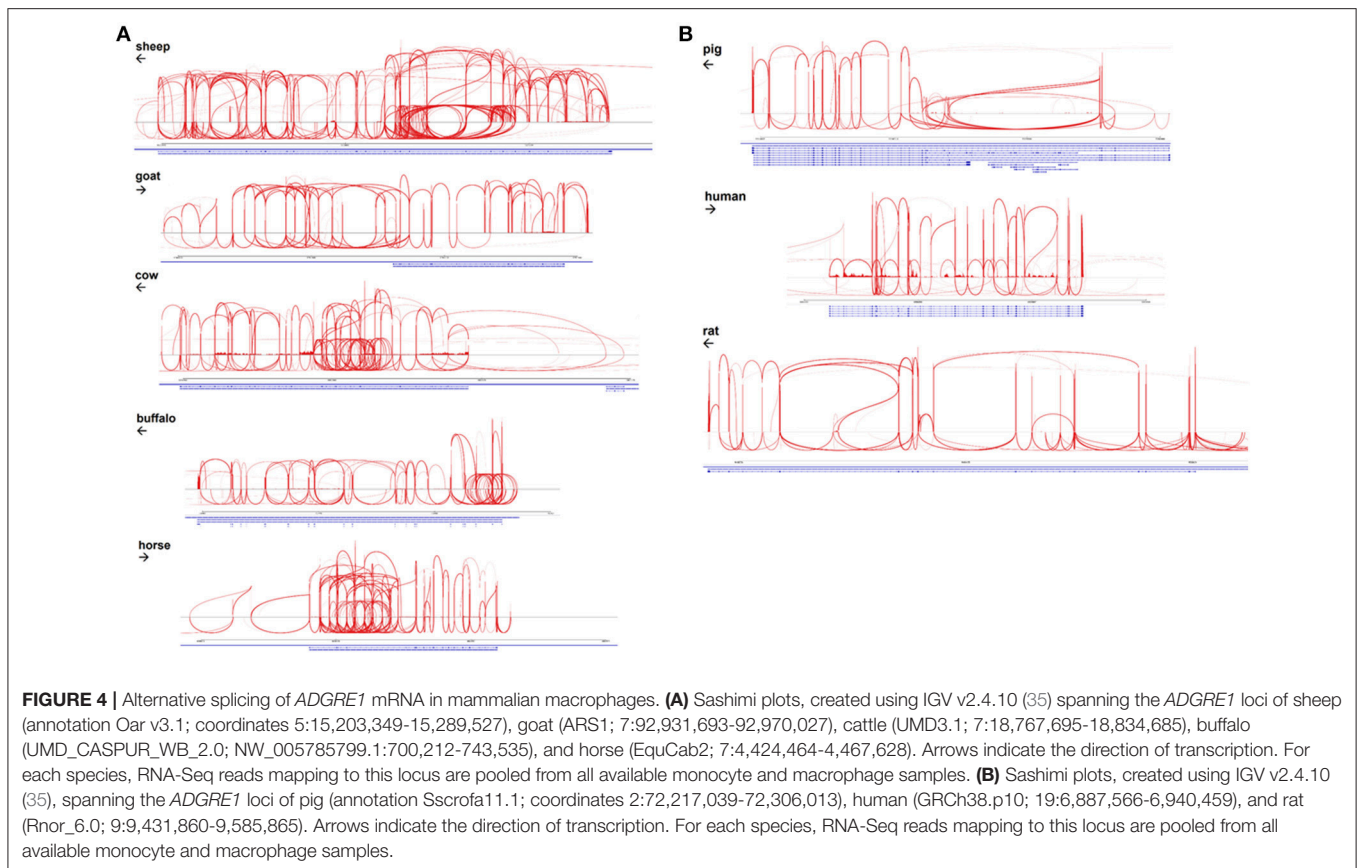
Evidence of Species-Specific Exon Skipping in *ADGRE1*

The EGF-like calcium binding domains of *ADGRE1* are encoded by single exons and can potentially be alternatively spliced. McKnight and Gordon (63) described 5 isoforms of mouse, and 4 isoforms of human *ADGRE1*, in which variable numbers of EGF-like domains are expressed, from none to 7 in mouse, and 1 to 6 in human. These authors did not report the expression of

the seventh EGF-like domain in humans. With the deep RNA-Seq data available for multiple species we examined the extent of alternative splicing in each species by aggregating available macrophage datasets, as shown in Sashimi plots in **Figures 4A,B**. These data indicate that the pig and human genomes, like the rat, encode a maximum of 7 EGF-like transmembrane domains. The accurate mapping of the RNA-Seq outputs to exons, and especially analysis of splice junctions, is complicated by the presence of an internal duplication in each of these species. The longest predicted cDNA for any species in Ensembl is for sheep, but the RNA-Seq data indicates the existence of a previously unannotated exon (**Figure 4B**). Even with the mapping limitations imposed by the repeated structure of the gene, the Sashimi plots indicate that in each species, every exon encoding an EGF-like domain can probably be bypassed by an exon skipping event. Accordingly, the potential number of isoforms may be large, although limitations of the datasets, including varying read lengths, mean that additional testing is required to increase statistical confidence in this hypothesis. The level of heterogeneity suggested is, however, supported by the broad band detected on the northern blot analysis of mouse macrophages when mouse *Adgre1* cDNA was originally cloned, with the apparent size ranging over at least 1 kb (5).

Analysis of the Promoter and Enhancer of *Adgre1*

A number of alternative isoforms originally described for human and mouse *ADGRE1* appear to encode N-terminally truncated proteins (63). However, a single major TSS region for *ADGRE1* is indicated from the FANTOM5 CAGE data from mouse and human macrophages, and we have confirmed the TSS region in CAGE data generated from pig BMDM (64) and from water buffalo (LL, RY, DAH, unpublished observations). Based upon the conserved expression in macrophages, we anticipated that regulatory elements would also be conserved. Accordingly, we extracted genomic sequences from Ensembl to compare the predicted transcription start sites and promoters for all of the species examined. The promoter in all species is TATA-less, and purine-rich, conforming to the pattern seen in many myeloid promoters in which the macrophage-specific transcription factor PU.1 is a key determinant of transcription initiation (65). The precise start site is conserved between the three species for which we have CAGE data (DAH, unpublished observations). Based upon these data, we can align the promoter sequences of mouse, rat, human, horse, pig, cattle, goat, sheep, and water buffalo as shown in **Figure 5A**. The promoter contains a perfectly-conserved binding site for C/EBP transcription factors, and for the Maf and MafB transcription factors which have both been shown, through the use of knockout mutations, to control F4/80 (*Adgre1*) expression in mice (66, 67). Analysis of the mouse *Adgre1* promoter revealed the presence of at least 9 purine-rich motifs containing the consensus GGAA core binding sites for PU.1 (68). As in humans, *ADGRE1* promoters from each of the large animals are purine-rich, but with a smaller number of the core GGAA motifs than observed in the mouse promoter, and with considerable variation in their location (**Figure 5A**).



The FANTOM5 human CAGE datasets also reveal the presence of an enhancer within intron 2, consistent with a previously-published ChIP-seq analysis of human monocyte subsets (49). This element is conserved at the sequence level in large animal genomes (Figure 5B) and contains an identical motif to the PU.1/IRF8 responsive element of the human IFN β 1 locus (69), which could contribute to the LPS-inducible expression of the gene. In overview, the conservation of regulatory elements is consistent with macrophage-specific expression in all of the species examined.

DISCUSSION

In this study, we have combined the generation of a new monoclonal antibody against pig ADGRE1 with detailed informatic analysis based upon RNA-Seq to gain further insight into the biology of ADGRE1 in mammals. The data in Figures 1–3 show clearly that *ADGRE1* is highly-expressed by pig monocytes, tissue macrophages and granulocytes and inducible during monocyte-macrophage differentiation from progenitors in the bone marrow. We demonstrated the utility of this new antibody for analysis of the response to agonists in pigs, such as the novel pig CSF1-Fc reagent (27, 44). ADGRE1 will provide a useful marker for dissection of infectious disease models in pigs, especially those of the lung where the pig is clearly more human-like (70).

The location of ADGRE1 expression in the pig revealed two major differences from the mouse. Mouse granulocytes express *Adgre1* mRNA, but do not translate the protein (20) whereas in pigs, ADGRE1 was highly-expressed on the surface of granulocytes in bone marrow. Consistent with the protein expression (45), *Adgre1* mRNA is very low in mouse alveolar macrophages (71) and we have found the same at the mRNA level in rats (CP, DAH, manuscript in preparation). By contrast, ADGRE1 protein was very highly-expressed in both alveolar and interstitial macrophages in the pig, and at least at the mRNA level, this pattern appears to be conserved in all the large animals including humans.

We summarized evidence from multiple datasets and platforms that the expression pattern we have dissected in pigs is shared with humans and other large animals. *ADGRE1* mRNA is not, as previously suggested, restricted in its expression to eosinophils or even enriched in those cells in humans. Indeed, the gene is a marker for differentiation of the CD16⁺⁺ monocyte subset in human peripheral blood and is also highly-expressed in many tissue macrophage populations, notably those of the lung. Expression of *ADGRE1* in macrophages in all large animals is consistent with the conservation of promoter and enhancer elements across species. Nevertheless, where *Adgre1* mRNA and protein were induced by CSF1 in rodent macrophages, in the large animals *ADGRE1* mRNA expression in macrophages was down-regulated, compared to

for evolution of transcriptional regulatory mechanisms within selected macrophage populations.

The two previous reports of specific *ADGRE1* mRNA and protein expression in eosinophils are difficult to reconcile with our analysis. Both reports used the same set of PCR primers, amplifying a short product from bases 1,408 to 1,490 in the cDNA sequence. To explain the apparent selective binding of anti-ADGRE1 antibodies to eosinophils, there is the formal possibility that the *ADGRE1* protein is selectively translated in these cells. Alternatively, the antibodies may cross-react with related calcium-binding EGF domain-containing molecules such as ADGRE3 which are actually strongly enriched in granulocytes, or with a specific glycoform of ADGRE1 produced by eosinophils. Whatever the explanation, it would clearly be worthwhile to make and test additional anti-human ADGRE1 reagents.

Aside from differences in gene expression profiles, ADGRE1 also varies greatly at the protein sequence level between species, including differences in the numbers of the EGF-like calcium binding domains. Within a species, the EGF-like domains also differ in sequence from each other. The mouse *Adgre1* cDNA described originally encoded 7 EGF-like calcium binding domains. Subsequent analysis of both mouse cDNAs identified at least 5 variants differing in the inclusion of individual domains. These included variants that lack exon 4, which contains the likely site for glycosaminoglycan attachment (5). Human *ADGRE2* shows evidence of both alternative splicing of the EGF-like domains and regulated glycosylation (60). There is also some evidence of alternative splicing of the human *ADGRE1* transcript. Sequencing of multiple full-length mouse macrophage cDNAs by the FANTOM consortium (72) identified further variants, apparently containing any combination of the EGF-like domains and including a possible secreted isoform. The longest cloned human *ADGRE1* cDNA in Ensembl encodes 6 EGF-like calcium binding domains, but our analysis indicates that there are actually 7 domains, as in rodents and pigs. By contrast to these species, the longest predicted cDNA encoding ADGRE1 in other species are considerably larger. The longest Ensembl sheep cDNA encodes 1,259 amino acids and 13 calcium binding EGF-like domains but the RNA-Seq data suggests there is at least one additional exon. The predicted protein for the goat is only 905 amino acids, which is clearly incorrect since the longest Ensembl-predicted sheep cDNA maps in its entirety to the goat genome (not shown). The longest Ensembl-predicted cattle protein is 1,148 amino acids and contains 12 predicted EGF-like calcium binding domains. However, the RNA-Seq data indicates the existence of at least two additional exons and the entire sheep cDNA maps to the cattle genome. Longer forms of ADGRE1 are also predicted by Ensembl for cat (1,101 amino acids) and dog (1,162 amino acids), but here again, the entire sheep cDNA maps contiguously to the dog and cat genome (not shown). In overview, the longer forms in ruminants, horses, cats, and dogs probably encode 14 EGF-like domains arising from a duplication of the 7-domain structure of rodent, pig and human. Five of the additional domains contain the acidic motif (most commonly CEDxDEC) implicated in calcium binding which is present in 5 of the 7 EGF-like domains of mouse *Adgre1* (5). Splicing analysis of the RNA-Seq data derived from BMDM (Figures 4A,B) from each

of these species reveals that as in mouse, human and pig, each of the EGF-like domains, encoded by single exons, in frame, exhibit evidence of exon-skipping, with the potential to produce numerous different isoforms. The protein sequence of ADGRE1 is highly divergent amongst species. For example, mice and pigs have only 58% identity, and a dN/dS ratio of only 0.24 (obtained via Ensembl using methods detailed at https://www.ensembl.org/info/genome/compara/homology_method.html). Almost all coding differences reside in the extracellular domain. In the EGF-like domains, only the structural and calcium-binding motifs are conserved. This may imply that there is no evolutionary/functional constraint on the sequence of these domains other than preservation of structure. The human exome database (<http://exac.broadinstitute.org>) reveals that *ADGRE1* (*EMR1*) is also polymorphic in human populations, with numerous non-synonymous protein-coding variants with high allele frequencies. Interestingly, a missense splice donor site, with a population allele frequency of 0.38, might explain the apparent lack of one exon in the longest current human cDNA. Alternatively, the rapid evolution of protein-coding sequences, and gene duplication, has long been recognized as a feature of immune-associated genes (73). Both within species polymorphism and accelerated between species divergence of innate immune genes was highlighted in detailed analysis of the pig immunome (74).

Hamann et al. (7) reviewed the evidence that members of the adhesion G protein coupled receptor family [*ADGRE1*, *ADGRE2*, *ADGRE3*, *ADGRE4*, and *CD97* (aka *ADGRE5*)] are modulators of immune cell function. Analysis of the function of this gene family has been constrained by the lack of knowledge of likely ligands. The only evidence comes from ADGRE2 which was reported to bind selectively to the proteoglycan chondroitin sulfate (75). Interaction with various glycosaminoglycans (GAG) at portals of entry is a very common feature of microbial pathogenesis (76). Conceivably, host GAG might act as opsonins to bind pathogens to innate immune cells through the ADGRE family. In the case of ADGRE1, no ligand has been identified, but the pig ADGRE1 fusion protein we used as an immunogen in the current study will facilitate future binding studies. The diversification of the extracellular domain through evolutionary selection and splicing, and high expression in lung macrophages in pigs, humans and other large animals, support the hypothesis that ADGRE1 has evolved in response to selection by pathogens, and participates in the host defense. A recent report demonstrated that cross-linking with anti-ADGRE2 antibody produced a pro-inflammatory signal in a human monocytic cell line (77). To our knowledge, the widely-used anti-mouse ADGRE1 antibody, F4/80, has not been shown to generate a detectable signal when added to macrophages. The availability of anti-ADGRE1 antibodies for the pig will enable further functional studies in this species that may translate to humans and other species.

AUTHOR CONTRIBUTIONS

LW, LL, and SB contributed equally to the study and carried out experimental work, data and sequence analysis. AR, RY, ZL,

MM, CM, KS, EC, KI, and CP contributed to experimental work and data analysis. DH conceived the idea and secured funding for the study. JH and DH supervised the experimental work and contributed to data analysis. All authors contributed to the manuscript.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fimmu.2018.02246/full#supplementary-material>

REFERENCES

- Austyn JM, Gordon S. F4/80, a monoclonal antibody directed specifically against the mouse macrophage. *Eur J Immunol.* (1981) 11:805–15. doi: 10.1002/eji.1830111013
- Hume DA, Ross IL, Himes SR, Sasmono RT, Wells CA, Ravasi T. The mononuclear phagocyte system revisited. *J Leukoc Biol.* (2002) 72:621–7. doi: 10.1189/jlb.72.4.621
- Gordon S, Hamann J, Lin HH, Stacey M. F4/80 and the related adhesion-GPCRs. *Eur J Immunol.* (2011) 41:2472–6. doi: 10.1002/eji.201141715
- Schulz C, Gomez Perdiguero E, Chorro L, Szabo-Rogers H, Cagnard N, Kierdorf K, et al. A lineage of myeloid cells independent of Myb and hematopoietic stem cells. *Science* (2012) 336:86–90. doi: 10.1126/science.1219179
- McKnight AJ, Macfarlane AJ, Dri P, Turley L, Willis AC. Molecular cloning of F4/80, a murine macrophage-restricted cell surface glycoprotein with homology to the G-protein-linked transmembrane 7 hormone receptor family. *J Biol Chem.* (1996) 271:486–9. doi: 10.1074/jbc.271.1.486
- Baud V, Chissoe SL, Viegas-Pequignot E, Diriong S, N'Guyen VC, Roe BA, et al. EMR1, an unusual member in the family of hormone receptors with seven transmembrane segments. *Genomics* (1995) 26:334–44. doi: 10.1016/0888-7543(95)80218-B
- Hamann J, Aust G, Arac D, Engel FB, Formstone C, Fredriksson R, et al. International Union of Basic and Clinical Pharmacology. XCIV. Adhesion G protein-coupled receptors. *Pharmacol Rev.* (2015) 67:338–67. doi: 10.1124/pr.114.009647
- Lin HH, Faunce DE, Stacey M, Terajewicz A, Nakamura T, Zhang-Hoover J, et al. The macrophage F4/80 receptor is required for the induction of antigen-specific efferent regulatory T cells in peripheral tolerance. *J Exp Med.* (2005) 201:1615–25. doi: 10.1084/jem.20042307
- Caminschi I, Lucas KM, O'Keefe MA, Hochrein H, Laabi Y, Kontgen F, et al. Molecular cloning of F4/80-like-receptor, a seven-span membrane protein expressed differentially by dendritic cell and monocyte-macrophage subpopulations. *J Immunol.* (2001) 167:3570–6. doi: 10.4049/jimmunol.167.7.3570
- Stacey M, Chang GW, Sanos SL, Chittenden LR, Stubbs L, Gordon S, et al. EMR4, a novel epidermal growth factor (EGF)-TM7 molecule up-regulated in activated mouse macrophages, binds to a putative cellular ligand on B lymphoma cell line A20. *J Biol Chem.* (2002) 277:29283–93. doi: 10.1074/jbc.M204306200
- Dobyns AE, Goyal R, Carpenter LG, Freeman TC, Longo LD, Yellon SM. Macrophage gene expression associated with remodeling of the prepartum rat cervix: microarray and pathway analyses. *Plos ONE* (2015) 10:e0119782. doi: 10.1371/journal.pone.0119782
- Pridans C, Sauter KA, Irvine KM, Davis GM, Lefevre, L, Raper A, et al. Macrophage colony-stimulating factor increases hepatic macrophage content, liver growth, and lipid accumulation in neonatal rats. *Am J Physiol Gastrointest Liver Physiol.* (2018) 314:G388–98. doi: 10.1152/ajpgi.00343.2017
- Irvine KM, Andrews MR, Fernandez-Rojo MA, Schroder K, Burns CJ, Su S, et al. Colony-stimulating factor-1 (CSF-1) delivers a proatherogenic signal to human macrophages. *J Leukoc Biol.* (2009) 85:278–88. doi: 10.1189/jlb.0808497
- Jubb AW, Young RS, Hume DA, Bickmore WA. Enhancer turnover is associated with a divergent transcriptional response to glucocorticoid in mouse and human macrophages. *J Immunol.* (2016) 196:813–22. doi: 10.4049/jimmunol.1502009
- Schroder K, Irvine KM, Taylor MS, Bokil NJ, Le Cao KA, Masterman KA, et al. Conservation and divergence in Toll-like receptor 4-regulated gene expression in primary human versus mouse macrophages. *Proc Natl Acad Sci USA.* (2012) 109:E944–53. doi: 10.1073/pnas.1110156109
- Hamann J, Koning N, Pouwels W, Ulfman LH, van Eijk M, Stacey M, et al. EMR1, the human homolog of F4/80, is an eosinophil-specific receptor. *Eur J Immunol.* (2007) 37:2797–802. doi: 10.1002/eji.20073[]7553
- Legrand F, Tomasevic N, Simakova O, Lee C-C, Wang Z, Raffeld M, et al. The eosinophil surface receptor epidermal growth factor-like module containing mucin-like hormone receptor 1 (EMR1): a novel therapeutic target for eosinophilic disorders. *J Allergy Clin Immunol.* (2014) 133:1439–47, 1447 e1–8. doi: 10.1016/j.jaci.2013.11.041
- Fukushima A, Ishida W, Ojima A, Kajisako M, Sumi T, Yamada J, et al. Participation of CD11b and F4/80 molecules in the conjunctival eosinophilia of experimental allergic conjunctivitis. *Int Arch Allergy Immunol.* (2010) 151:129–36. doi: 10.1159/000236002
- McGarry MP, Stewart CC. Murine eosinophil granulocytes bind the murine macrophage-monocyte specific monoclonal antibody F4/80. *J Leukoc Biol.* (1991) 50:471–8. doi: 10.1002/jlb.50.5.471
- Sasmono RT, Ehrnsperger A, Cronau SL, Ravasi T, Kandane R, Hickey MJ, et al. Mouse neutrophilic granulocytes express mRNA encoding the macrophage colony-stimulating factor receptor (CSF-1R) as well as many other macrophage-specific transcripts and can transdifferentiate into macrophages *in vitro* in response to CSF-1. *J Leukoc Biol.* (2007) 82:111–23. doi: 10.1189/jlb.1206713
- Caminschi I, Vandenabeele S, Sofi M, McKnight AJ, Ward N, Brodnicki TC, et al. Gene structure and transcript analysis of the human and mouse EGF-TM7 molecule, FIRE. *DNA Seq.* (2006) 17:8–14. doi: 10.1080/10425170500355737

22. Lin HH, Hsiao CC, Pabst C, Hebert J, Schoneberg T, Hamann J. Adhesion GPCRs in Regulating Immune Responses and Inflammation. *Adv Immunol.* (2017) 136:163–201. doi: 10.1016/bs.ai.2017.05.005
23. Fairbairn L, Kapetanovic R, Beraldi D, Sester DP, Tuggle CK, Archibald AL, et al. Comparative analysis of monocyte subsets in the pig. *J Immunol.* (2013) 190:6389–96. doi: 10.4049/jimmunol.1300365
24. Kapetanovic R, Fairbairn L, Beraldi D, Sester DP, Archibald AL, Tuggle CK, et al. Pig bone marrow-derived macrophages resemble human macrophages in their response to bacterial lipopolysaccharide. *J Immunol.* (2012) 188:3382–94. doi: 10.4049/jimmunol.1102649
25. Kapetanovic R, Fairbairn L, Downing A, Beraldi D, Sester DP, Freeman TC, et al. The impact of breed and tissue compartment on the response of pig macrophages to lipopolysaccharide. *BMC Genomics* (2013) 14:581. doi: 10.1186/1471-2164-14-581
26. Fairbairn L, Kapetanovic R, Sester DP, Hume DA. The mononuclear phagocyte system of the pig as a model for understanding human innate immunity and disease. *J Leukoc Biol.* (2011) 89:855–71. doi: 10.1189/jlb.1110607
27. Sauter KA, Waddell LA, Lisowski ZM, Young R, Lefevre L, Davis GM, et al. Macrophage colony-stimulating factor (CSF1) controls monocyte production and maturation and the steady-state size of the liver in pigs. *Am J Physiol Gastrointest Liver Physiol.* (2016) 311:G533–47. doi: 10.1152/ajpgi.00116.2016
28. Hume DA, Pavli P, Donahue RE, Fidler IJ. The effect of human recombinant macrophage colony-stimulating factor (CSF-1) on the murine mononuclear phagocyte system *in vivo*. *J Immunol.* (1988) 141:3405–9.
29. Young R, Bush SJ, Lefevre L, McCulloch MEB, Lisowski ZM, Muriuki C, et al. Species differences in the expression of inducible nitric oxide synthase (NOS2) and arginase in macrophages are associated with gain and loss of regulatory elements. *Immunohorizons* (2018) 2:27–37. doi: 10.4049/immunohorizons.1700073
30. Baillie JK, Arner E, Daub C, De Hoon M, Itoh M, Kawaji H, Lassmann T, et al. Analysis of the human monocyte-derived macrophage transcriptome and response to lipopolysaccharide provides new insights into genetic aetiology of inflammatory bowel disease. *PLoS Genet.* (2017) 13:e1006641. doi: 10.1371/journal.pgen.1006641
31. Garcia-Morales C, Rothwell L, Moffat L, Garceau V, Balic A, Sang H, Kaiser P, et al. Production and characterisation of a monoclonal antibody that recognises the chicken CSF1 receptor and confirms that expression is restricted to macrophage-lineage cells. *Dev Comp Immunol.* (2014) 42:278–85. doi: 10.1016/j.dci.2013.09.011
32. Shulman M, Wilde CD, Kohler G. A better cell line for making hybridomas secreting specific antibodies. *Nature* (1978) 276:269–70. doi: 10.1038/276269a0
33. Rothwell L, Hamblin A, Kaiser P. Production and characterisation of monoclonal antibodies specific for chicken interleukin-2. *Vet Immunol Immunopathol.* (2001) 83:149–60. doi: 10.1016/S0165-2427(01)00391-9
34. Clark EL, Bush SJ, McCulloch MEB, Farquhar IL, Young R, Lefevre L, et al. A high resolution atlas of gene expression in the domestic sheep (*Ovis aries*). *PLoS Genet.* (2017) 13:e1006997. doi: 10.1371/journal.pgen.1006997
35. Robinson JT, Thorvaldsdottir H, Winckler W, Guttman M, Lander ES, Getz G, et al. Integrative genomics viewer. *Nat Biotechnol.* (2011) 29:24–6. doi: 10.1038/nbt.1754
36. Bray NL, Pimentel H, Melsted P, Pachter L. Near-optimal probabilistic RNA-seq quantification. *Nat Biotechnol.* (2016) 34:525–7. doi: 10.1038/nbt.3519
37. Pinilla-Vera M, Xiong Z, Zhao Y, Zhao J, Donahoe MP, Barge S, et al. Full spectrum of LPS activation in alveolar macrophages of healthy volunteers by whole transcriptomic profiling. *PLoS ONE* (2016) 11:e0159329. doi: 10.1371/journal.pone.0159329
38. Galatro TF, Holtman IR, Lerario AM, Vainchtein ID, Brouwer N, Sola PR, et al. Transcriptomic analysis of purified human cortical microglia reveals age-associated changes *Nat Neurosci.* (2017) 20:1162–71. doi: 10.1038/nn.4597
39. Williams CR, Baccarella A, Parrish JZ, Kim CC. Empirical assessment of analysis workflows for differential expression analysis of human samples using RNA-Seq. *BMC Bioinf.* (2017) 18:38. doi: 10.1186/s12859-016-1457-z
40. Mirsafian H, Manda SS, Mitchell CJ, Sreenivasamurthy S, Ripen AM, Mohamad SB, et al. Long non-coding RNA expression in primary human monocytes. *Genomics* (2016) 108:37–45. doi: 10.1016/j.ygeno.2016.01.002
41. Freeman TC, Ivens A, Baillie JK, Beraldi D, Barnett MW, Dorward D, et al. A gene expression atlas of the domestic pig. *BMC Biol.* (2012) 10:90. doi: 10.1186/1741-7007-10-90
42. Dawson HD, Smith AD, Chen C, Urban JF Jr. An in-depth comparison of the porcine, murine and human inflammasomes; lessons from the porcine genome and transcriptome. *Vet Microbiol.* (2017) 202:2–15. doi: 10.1016/j.vetmic.2016.05.013
43. MacDonald KP, Palmer JS, Cronau S, Seppanen E, Olver S, Raffelt NC, et al. An antibody against the colony-stimulating factor 1 receptor depletes the resident subset of monocytes and tissue- and tumor-associated macrophages but does not inhibit inflammation. *Blood* (2010) 116:3955–63. doi: 10.1182/blood-2010-02-266296
44. Gow DJ, Sauter KA, Pridans C, Moffat L, Sehgal A, Stutchfield BM, et al. Characterisation of a novel Fc conjugate of macrophage colony-stimulating factor *Mol Ther.* (2014) 22:1580–92. doi: 10.1038/mt.2014.112
45. Gordon S, Pludemann A, Martinez Estrada F. Macrophage heterogeneity in tissues: phenotypic diversity and functions. *Immunol Rev.* (2014) 262:36–55. doi: 10.1111/imr.12223
46. Hume DA, Robinson AP, MacPherson GG, Gordon S. The mononuclear phagocyte system of the mouse defined by immunohistochemical localization of antigen F4/80. Relationship between macrophages, Langerhans cells, reticular cells, and dendritic cells in lymphoid and hematopoietic organs. *J Exp Med.* (1983) 158:1522–36.
47. Mabbott NA, Baillie JK, Brown H, Freeman TC, Hume DA. An expression atlas of human primary cells: inference of gene function from coexpression networks. *BMC Genomics* (2013) 14:632. doi: 10.1186/1471-2164-14-632
48. FANTOM Consortium and the RIKEN PMI and CLST (DGT), Forrest AR, Kawaji H, Rehli M, Baillie JK, de Hoon MJ, et al. A promoter-level mammalian expression atlas. *Nature* (2014) 507:462–70. doi: 10.1038/nature13182
49. Schmid C, Renner K, Peter K, Eder R, Lassmann T, Balwierz P, et al. Transcription and enhancer profiling in human monocyte subsets. *Blood* (2014) 123:e90–9. doi: 10.1182/blood-2013-02-484188
50. Maiga A, Lemieux S, Pabst C, Lavalley VP, Bouvier M, Sauvageau G, et al. Transcriptome analysis of G protein-coupled receptors in distinct genetic subgroups of acute myeloid leukemia: identification of potential disease-specific targets. *Blood Cancer J.* (2016) 6:e431. doi: 10.1038/bcj.2016.36
51. Ancuta P, Liu KY, Misra V, Wacleche VS, Gosselin A, Zhou X, et al. Transcriptional profiling reveals developmental relationship and distinct biological functions of CD16+ and CD16- monocyte subsets. *BMC Genomics* (2009) 10:403. doi: 10.1186/1471-2164-10-403
52. Ingersoll MA, Spanbroek R, Lottaz C, Gautier EL, Frankenberger M, Hoffmann R, et al. Comparison of gene expression profiles between human and mouse monocyte subsets *Blood* (2010) 115:e10–9. doi: 10.1182/blood-2009-07-235028
53. Hofer TP, Zawada AM, Frankenberger M, Skokann K, Satz AA, Gesierich W, et al. slan-defined subsets of CD16-positive monocytes: impact of granulomatous inflammation and M-CSF receptor mutation. *Blood* (2015) 126:2601–10. doi: 10.1182/blood-2015-06-651331
54. Madore AM, Perron S, Turmel V, Laviolette M, Bissonnette EY, Laprise C. Alveolar macrophages in allergic asthma: an expression signature characterized by heat shock protein pathways. *Hum Immunol.* (2010) 71:144–50. doi: 10.1016/j.humimm.2009.11.005
55. Woodruff PG, Koth LL, Yang YH, Rodriguez MW, Favoreto S, Dolganov GM, et al. A distinctive alveolar macrophage activation state induced by cigarette smoking. *Am J Respir Crit Care Med.* (2005) 172:1383–92. doi: 10.1164/rccm.200505-686OC
56. Ben Amara A, Gorvel L, Baulan K, Derain-Court J, Buffat C, Verollet CL, et al. Placental macrophages are impaired in chorioamnionitis, an infectious pathology of the placenta. *J Immunol.* (2013) 191:5501–14. doi: 10.4049/jimmunol.1300988
57. Khazen W, M'bika JP, Tomkiewicz C, Benelli C, Chany C, Achour A, et al. Expression of macrophage-selective markers in human and rodent adipocytes. *FEBS Lett.* (2005) 579:5631–4. doi: 10.1016/j.febslet.2005.09.032
58. Irvine KM, Banh X, Gadd VL, Wojcik KK, Ariffin JK, Jose S, et al. CRIG-expressing peritoneal macrophages are associated with disease severity in patients with cirrhosis and ascites. *JCI Insight* (2016) 1:e86914. doi: 10.1172/jci.insight.86914

59. Hamann J, Kwakkenbos MJ, de Jong EC, Heus H, Olsen AS, van Lier RA. Inactivation of the EGF-TM7 receptor EMR4 after the Pan-Homo divergence. *Eur J Immunol.* (2003) 33:1365–71. doi: 10.1002/eji.200323881
60. Chang GW, Davies JQ, Stacey M, Yona S, Bowdish DM, Hamann J, et al. CD312, the human adhesion-GPCR EMR2, is differentially expressed during differentiation, maturation, and activation of myeloid cells. *Biochem Biophys Res Commun.* (2007) 353:133–138. doi: 10.1016/j.bbrc.2006.11.148
61. Matmati M, Pouwels W, van Bruggen R, Jansen M, Hoek RM, Verhoeven AJ, et al. The human EGF-TM7 receptor EMR3 is a marker for mature granulocytes. *J Leukoc Biol.* (2007) 81:440–8. doi: 10.1189/jlb.0406276
62. Hamann J. The EGF-TM7 family of the rat. *Immunogenetics* (2004) 56:679–81. doi: 10.1007/s00251-003-0625-2
63. McKnight AJ, Gordon S. The EGF-TM7 family: unusual structures at the leukocyte surface. *J Leukoc Biol.* (1998) 63:271–80.
64. Robert C, Kapetanovic R, Beraldi D, Watson M, Archibald AL, Hume DA. Identification and annotation of conserved promoters and macrophage-expressed genes in the pig genome. *BMC Genomics* (2015) 16:970. doi: 10.1186/s12864-015-2111-2
65. Ross IL, Yue X, Ostrowski MC, Hume DA. Interaction between PU.1 and another Ets family transcription factor promotes macrophage-specific Basal transcription initiation. *J Biol Chem.* (1998) 273:6662–9.
66. Moriguchi T, Hamada M, Morito N, Terunuma T, Hasegawa K, Zhang C, et al. MafB is essential for renal development and F4/80 expression in macrophages. *Mol Cell Biol.* (2006) 26:5715–27. doi: 10.1128/MCB.00001-06
67. Nakamura M, Hamada M, Hasegawa K, Kusakabe M, Suzuki H, Greaves DR, et al. c-Maf is essential for the F4/80 expression in macrophages *in vivo*. *Gene* (2009) 445:66–72. doi: 10.1016/j.gene.2009.06.003
68. O'Reilly D, Addley M, Quinn C, MacFarlane AJ, Gordon S, McKnight AJ, et al. Functional analysis of the murine *Emr1* promoter identifies a novel purine-rich regulatory motif required for high-level gene expression in macrophages. *Genomics* (2004) 84:1030–40. doi: 10.1016/j.ygeno.2004.08.016
69. Li P, Wong JJ-Y, Sum C, Sin W-X, Ng KQ, Koh MB, et al. IRF8 and IRF3 cooperatively regulate rapid interferon- β induction in human blood monocytes. *Blood* (2011) 117:2847–54. doi: 10.1182/blood-2010-07-294272
70. Meurens F, Summerfield A, Nauwynck H, Saif L, Gerdt V. The pig: a model for human infectious diseases. *Trends Microbiol.* (2012) 20:50–7. doi: 10.1016/j.tim.2011.11.002
71. Mass E, Ballesteros I, Farlik M, Halbritter F, Gunther P, Crozet L, et al. Specification of tissue-resident macrophages during organogenesis. *Science* (2016) 353:aaf4238. doi: 10.1126/science.aaf4238
72. Carninci P, Kasukawa T, Katayama S, Gough J, Frith MC, Maeda N, et al. The transcriptional landscape of the mammalian genome. *Science* (2005) 309:1559–63. doi: 10.1126/science.1112014
73. Trowsdale J, Parham P. Mini-review: defense strategies and immunity-related genes. *Eur J Immunol.* (2004) 34:7–17. doi: 10.1002/eji.200324693
74. Dawson HD, Loveland JE, Pascal G, Gilbert JG, Uenishi H, Mann KM, et al. Structural and functional annotation of the porcine immunome. *BMC Genomics* (2013) 14:332. doi: 10.1186/1471-2164-14-332
75. Burzava ALS, Jasieniak M, Cockshell MP, Bonder CS., Harding FJ., Griesser HJ, et al. Affinity binding of EMR2 expressing cells by surface-grafted chondroitin sulfate B. *Biomacromolecules* (2017) 18:1697–704. doi: 10.1021/acs.biomac.6b01687
76. Aquino RS, Park PW. Glycosaminoglycans and infection. *Front Biosci.* (2016) 21:1260–77. doi: 10.2741/4455
77. I KY, Huang YS, Hu CH, Tseng WY, Cheng CH, Stacey M, Gordon S, et al. Activation of adhesion GPCR EMR2/ADGRE2 induces macrophage differentiation and inflammatory responses via G α_{16} /Akt/MAPK/NF-kappaB signaling pathways. *Front Immunol.* (2017) 8:73. doi: 10.3389/fimmu.2017.00373

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