



NF- κ B Pathway in Autoinflammatory Diseases: Dysregulation of Protein Modifications by Ubiquitin Defines a New Category of Autoinflammatory Diseases

Ivona Aksentijevich* and Qing Zhou

Inflammatory Disease Section, National Human Genome Research Institute, Bethesda, MD, USA

OPEN ACCESS

Edited by:

José Hernández-Rodríguez,
Hospital Clinic of Barcelona, Spain

Reviewed by:

Jordi Yagüe,
Hospital Clinic of Barcelona, Spain
Kyeong Kyu Kim,
Sungkyunkwan University,
South Korea

*Correspondence:

Ivona Aksentijevich
aksentii@mail.nih.gov

Specialty section:

This article was submitted
to Inflammation,
a section of the journal
Frontiers in Immunology

Received: 22 December 2016

Accepted: 21 March 2017

Published: 19 April 2017

Citation:

Aksentijevich I and Zhou Q (2017)
NF- κ B Pathway in Autoinflammatory
Diseases: Dysregulation of Protein
Modifications by Ubiquitin Defines a
New Category of Autoinflammatory
Diseases.
Front. Immunol. 8:399.
doi: 10.3389/fimmu.2017.00399

Autoinflammatory diseases are caused by defects in genes that regulate the innate immunity. Recently, the scope of autoinflammation has been broadened to include diseases that result from dysregulations in protein modifications by the highly conserved ubiquitin (Ub) peptides. Thus far these diseases consist of linear ubiquitin chain assembly complex (LUBAC) and OTULIN deficiencies, and haploinsufficiency of A20. The LUBAC is critical for linear ubiquitination of key signaling molecules in immune response pathways, while deubiquitinase enzymes, OTULIN and TNFAIP3/A20, reverse the effects of ubiquitination by hydrolyzing linear (Met1) and Lys63 (K63) Ub moieties, respectively, from conjugated proteins. Consequently, OTULIN or A20-deficient cells have an excess of Met1 or K63 Ub chains on NEMO, RIPK1, and other target substrates, which lead to constitutive activation of the NF- κ B pathway. Mutant cells produce elevated levels of many proinflammatory cytokines and respond to therapy with cytokine inhibitors. Patients with an impairment in LUBAC stability have compromised NF- κ B responses in non-immune cells such as fibroblasts, while their monocytes are hyperresponsive to IL-1 β . Discoveries of germline mutations in enzymes that regulate protein modifications by Ub define a new category of autoinflammatory diseases caused by upregulations in the NF- κ B signaling. The primary aim of this review is to summarize the latest developments in our understanding of the etiology of autoinflammation.

Keywords: TNFAIP3/A20, linear ubiquitin chain assembly complex, OTULIN, haploinsufficiency of A20, LUBAC deficiency, otulipenia/otulin-related autoinflammatory syndrome

INTRODUCTION

Autoinflammatory diseases are a diverse group of inherited conditions characterized by early-onset systemic inflammation and are accompanied by a range of organ-specific manifestations. The genetic etiology involves abnormalities in molecules such as inflammasomes, cytokine inhibitors, cytokine receptors, enzymes, and proteasome complex. The excessive secretion of proinflammatory cytokines can lead to chronic morbidity and may be life-threatening. Therapies with cytokine inhibitors are efficacious in most patients; however, considering the high cost of biologics there is a need to develop more affordable treatment options for these lifelong conditions. The discovery of germline mutations

linked to autoinflammation offer an opportunity to search for new therapeutic targets.

UBIQUITIN (Ub) PATHWAY

Posttranslational protein modification by ubiquitination (also known as ubiquitylation) is critical for the regulation of many biological processes including DNA repair, endocytosis, transcription, protein degradation, and preservation of cellular homeostasis. Ubiquitination involves the attachment of evolutionarily conserved 76-aa Ub molecules to target proteins in the form of a monomer or polymers (Ub chains). The type of conjugation determines the fate of the modified protein by directing protein localization and regulating protein interactions, activity, and degradation (1).

The ubiquitination process is initiated by the attachment of a single Ub molecule to a target protein through a three-step enzymatic pathway that includes Ub-activating enzymes (E1), Ub-conjugating enzymes (E2), and Ub-ligating enzymes (E3) (2). Ub chains are generated by the sequential addition of Ub monomers through one of seven lysine residues that serve as a linker, thus there are seven types of Ub Lys-linkages. Ub can be also conjugated through an N-terminal methionine residue (Met1 linkage; linear linkage). Lys11-, Lys48-, Lys63-, and Met1-linked chains are the best known and most studied. Significance of four other Ub chains Lys6-, Lys27-, Lys29-, and Lys33- is poorly understood. In addition, there is increasing evidence that more than one linkage type can exist on modified proteins either in the form of mixed (hybrid) Ub chains or branched Ub chains, and this may provide a new layer of complexity to the Ub-mediated modifications (2, 3). Ub proteins are also subject to modifications by acetylation, phosphorylation, and ubiquitin-like (UBL) molecules (SUMO or NEDD8) (4). Ub-conjugated proteins are recognized by Ub sensors, Ub-binding proteins (Ub receptors) that can translate each linkage type of Ub modifications into specific functional outcomes. For example, proteins conjugated with Lys (K11) or Lys48 (K48) Ub chains are targeted for proteasomal degradation *via* the ubiquitin–proteasome system (UPS) (5). In addition, Lys11 Ub chains have a role in cell cycle control and may have other functions in the context of mixed Ub chains (6). Lys63 (K63) Ub chains are involved in cell signaling and are essential for DNA damage response (7). Linear (Met1) Ub chains regulate a wide range of immune signaling pathways (8–10). Ubiquitination is a highly dynamic and reversible process whereby Ub chains are removed from modified substrates by a class of enzymes known as deubiquitylases or deubiquitinases (DUBs) (11). There are close to 100 proteases that have DUB activity with different degrees of specificity for Ub linkages. Several DUBs, including A20, OTULIN, CYLD, and Cezanne, function as negative regulators of NF- κ B signaling (12).

Alterations in various components of the Ub–proteasome machinery have been linked to many human conditions including immune diseases. Recently, deregulations in the UPS were reported in patients with autoinflammatory disorders including chronic atypical neutrophilic dermatosis with lipodystrophy and elevated temperature (13), linear ubiquitin chain assembly complex (LUBAC) deficiency (14, 15), haploinsufficiency of A20

(HA20) (16), and otulipenia/otulin-related autoinflammatory syndrome (ORAS) (17, 18). This review will primarily focus on two diseases caused by malfunction in DUB enzymes, TNFAIP3/A20 and OTULIN, which are known to hydrolyze Lys63- and Met1-linked Ub chains, respectively. In both conditions, HA20 and otulipenia/ORAS, a defect in DUB activity results in excessive ubiquitination and increased activity of key signaling molecules in the canonical NF- κ B pathway. LUBAC-associated diseases will be briefly discussed in the context of LUBAC–OTULIN interactions.

LINEAR Ub CHAINS IN IMMUNE SIGNALING

Linear ubiquitin chain assembly complex (LUBAC)-mediated Met1 ubiquitination has emerged pivotal for regulation of innate and adaptive immune responses and regulation of cell death (9, 19). The E3 ligase complex, LUBAC, has been shown to maintain the stability of the TNF receptor 1 (TNFR1), TLRs, IL-1R, CD40, RLR, and inflammasome receptor signaling complexes (RSCs). Upon stimulation with proinflammatory signals, LUBAC is recruited to attach linear Ub chains to target substrates such as IKK (NEMO), RIPK1, RIPK2, IRAKs, MyD88, and ASC (8, 20, 21). Attachment of linear Ub chains is critical for the assembly of RSCs. LUBAC depletion leads to attenuation of NF- κ B and the mitogen-activated protein kinases (MAPK)-mediated signaling and increases cell death.

Linear ubiquitin chain assembly complex consists of HOIP (HOIL-1 interacting protein; *RNF31*) and two accessory proteins: HOIL-1 (heme-oxidized IRP2 ubiquitin ligase 1; *RBCK1*) and SHARPIN (SHANK-interacting protein like 1; *SIPL1*) (Figure 1). The catalytic subunit HOIP is auto-inhibited until the complex is fully assembled. Depletion of any subunits greatly reduces the stability of LUBAC in cells (14, 15). As the immune responses must be tightly regulated to avoid chronic inflammation, LUBAC activity is counter-regulated by the specific DUB OTULIN (also known as *gumby*; *FAM105B*).

OTULIN is a highly conserved protease with a specific activity to hydrolyze linear (Met1)-linked Ub chains (22) (Figure 1). OTULIN interacts with the N-terminal PUB domain of HOIP *via* an evolutionarily conserved PUB-interacting motif (23). Loss of HOIP–OTULIN interaction reduces OTULIN capacity to restrict LUBAC-induced NF- κ B activation (24). A recent study showed that the activity of LUBAC is also negatively regulated by its interaction with tumor necrosis factor receptor-associated factor 1 (TRAF1). TRAF1 directly interacts with LUBAC to interfere with the activation of IKK/NEMO. Reduced expression of TRAF1 could explain the association of susceptibility alleles in *TRAF1* with rheumatoid arthritis (RA) and other autoimmune diseases (25).

The importance of the linear ubiquitination in regulation of inflammatory pathways has been demonstrated in murine models. Genetic ablation of the catalytic HOIP subunit results in embryonic lethality at day 10.5 due to TNF-mediated endothelial cell death and vascular abnormalities (26). Mice deficient for non-catalytic subunits have variable degrees of inflammation manifesting with chronic proliferative dermatitis in the case

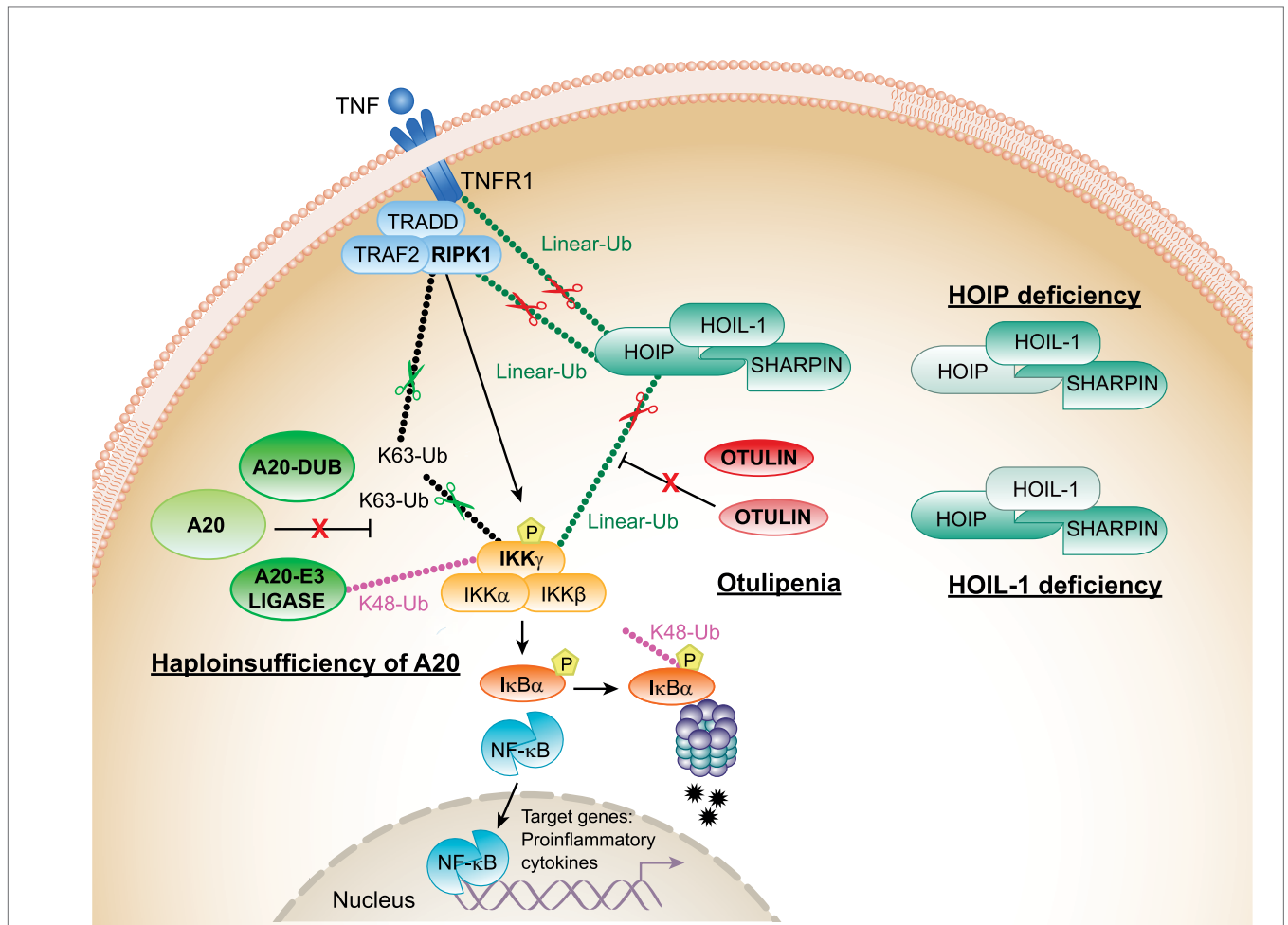


FIGURE 1 | Proposed mechanisms of pathogenesis in haploinsufficiency of A20 (HA20), otulipenia/otulin-related autoinflammatory syndrome (ORAS), and linear ubiquitin chain assembly complex (LUBAC) deficiencies. The canonical NF- κ B pathway is regulated both by K63 (Lys63)-linked and linear (Met1)-linked ubiquitin (Ub) chains. RIPK1 is the central adaptor for assembly of the TNFR1 receptor signaling complex and is a predominant target for ubiquitination by K63 and linear Ub chains. Polyubiquitylated RIPK1 mediates recruitment of IKK complex that is also target for ubiquitination. The activated IKK complex phosphorylates inhibitor of kappa B ($I\kappa$ B α) and targets $I\kappa$ B α for proteasome-mediated degradation. Linear Ub chains are added to RIPK1 and IKK γ by LUBAC. A20 and OTULIN negatively regulate the NF- κ B pathway by cleaving K63 and linear Ub chains from target molecules, RIPK1 and IKK γ . In addition, A20 through its E3 ligase activity adds K48 Ub chains to IKK γ (and TRAF6, not shown in the figure) targeting them for proteasomal degradation. Decreased expression of A20 in patients with HA20 or OTULIN in patients with otulipenia/ORAS will lead to activation of the NF- κ B pathway, increased expression of proinflammatory transcripts in immune cells, and systemic inflammation. Decreased expression of LUBAC complex subunits, in patients who carry mutations in HOIP or HOIL-1, results in inhibition of the NF- κ B pathway in fibroblasts and B-cells causing immunodeficiency. However, their monocytes hyperproduce proinflammatory cytokines. TNFR1, TNF receptor 1; TRADD, TNFR1-associated death domain protein; RIPK1, the death domain-containing protein kinase receptor-interacting protein 1; IKK γ /NEMO, inhibitor of nuclear factor kappa B kinase subunit gamma.

of SHARPIN-deficient mice (*cpdm*) (27, 28) or with no overt inflammation in mice lacking HOIL-1 (21). Skin inflammation in Sharpin-KO mice is largely dependent on the TNFR1-induced apoptosis (19, 29, 30). LUBAC activity is also important for proper B and T cell development, activation, and maintenance of adaptive immune responses (31, 32). Genetic loss of *Otulin* (*gumby/gumby*) causes embryonic lethality (E12.5–E14) due to compromised angiogenesis and defects in neuronal development (33). Tamoxifen-induced *Otulin* deficiency in immune cells results in an acute severe multiorgan inflammatory phenotype (18). Targeted ablation of *Otulin* in myeloid cells leads to chronic

inflammation with features of autoimmunity, while *Otulin* deficiency in adaptive immune cells does not produce overt phenotype (18). Together, these data show critical and cell-specific function of OTULIN in the maintenance of immune homeostasis.

LUBAC DEFICIENCIES

Patients with defects in the LUBAC components develop immunodeficiency, autoinflammation, muscular amylopectinosis, and die in early childhood (Tables 1 and 2). HOIL-1 and HOIP deficiencies are recessively inherited diseases caused by mutations

TABLE 1 | Comparison of genetics and mechanisms of disease in otulipenia, haploinsufficiency of A20 (HA20), and linear ubiquitin chain assembly complex (LUBAC) deficiencies.

		Otulipenia/ORAS	HA20
Gene	Gene name	<i>OTULIN (FAM105B)</i>	<i>TNFAIP3</i>
	Exons	7 exons	9 exons
	Chromosome	Chr.5	Chr.6
Protein	Protein name	OTULIN	A20
	Protein length	352aa	790aa
	Protein domains	PUB-interacting motif domain, ovarian tumor (OTU) domain	OTU domain, 7 ZnF domains
	Protein function	Met1 linear deubiquitinase (DUB)	K63 DUB
	Involved pathway	NF- κ B	NF- κ B, NLRP3
	Substrate molecules	NEMO, TNF receptor 1 (TNFR1), RIPK1, ASC	NEMO, RIPK1, TRAF6, pro IL-1 β
Genetics	Inheritance	Recessive	Dominant
	Type of mutations	Loss-of-function mutations (missense, INDELS)	Loss-of-function mutations (stop gain mutation, missense, INDELS)
	Frequency of mutant alleles	Rare or novel	Novel
	Location of the mutations	OTU domain (linear DUB activity)	OTU domain (k63 DUB activity) or ZnF4 domain
	Number of mutations	Biallelic (compound heterozygous or homozygous)	Heterozygous
Mechanisms	Mechanism	Loss-of-function (reduced protein expression)	Haploinsufficiency (50% decrease in protein expression)
	Protein Interactions	Instability of LUBAC	Decreased association with TNFR1, TRAF2, and RIPK1
	Effect of mutant proteins	Impaired linear deubiquitination of NEMO, TNFR1, RIPK1, ASC	Impaired K63 deubiquitination of NEMO, RIPK1, and TRAF6
	Involved pathway	Increased signaling in NF- κ B and mitogen-activated protein kinases (MAPK) pathways	Increased activity of NF- κ B, MAPKs, and NLRP3
	Cytokines	IL-1 β , TNF, IL-6, IL-12, IL-18, IFN γ	IL-1 β , TNF, IL-6, IL-9, IL-17, IL-18, IFN γ
		HOIL-1 deficiency	HOIP deficiency
Gene	Gene name	<i>RBCK1</i>	<i>RNF31</i>
	Exons	12 exons	21 exons
	Chromosome	Chr.20	Chr.14
Protein	Protein name	HOIL-1	HOIP
	Protein length	510aa	1,072aa
	Protein domains	Ubiquitin-like (UBL), novel zinc finger (NZF), RING1, IBR, RING2	PUB, ZnF, NZF1, NZF2, UBA, RING1, IBR, RING2, LDD
	Protein function	Subunit of the LUBAC	Catalytic subunit of the LUBAC
	Involved pathway	NF- κ B signaling pathway	NF- κ B signaling pathway
Mutations	Type of mutations	Loss-of-function mutations (stop gain mutation, INDELS)	Loss-of-function mutations (missense)
	Frequency of mutant alleles	Rare or novel	Novel
	Location of the mutations	UBL domain (interacts with HOIP UBA domain), NZF domain (ubiquitin binding)	PUB domain (interacts with OTULIN)
	Number of mutations	Biallelic (compound heterozygous or homozygous)	Biallelic (homozygous)
Mechanisms	Inheritance	Recessive	Recessive
	Mechanism	Loss-of-function (decreased protein expression, instability of LUBAC, impaired linear ubiquitination)	Loss-of-function (decreased protein expression, instability of LUBAC, impaired linear ubiquitination)
	Effect of mutant proteins	Defect in linear ubiquitination NEMO, RIPK1, IRAK-1	Defect in linear ubiquitination
	Involved pathway	Impaired NF- κ B activation in fibroblasts, increased NF- κ B activity in monocytes	Impaired NF- κ B activation in fibroblasts and CD40L stimulated B cells, increased NF- κ B activity in monocytes
	Cytokines	Impaired expression of IL-6 in fibroblasts upon IL-1 β and TNF stimulation; hyperproduction of IL-6 upon IL-1 β stimulated monocytes; high serum IL-1, IL-6	Hyperproduction of IL-6 in IL-1 β stimulated monocytes

that either create truncating proteins or affect a highly conserved PUB domain of HOIP (14, 15) (**Figure 2**). Pathogenic mutations in one subunit destabilize the expression of the entire LUBAC complex. As LUBAC is important for activation of immune signaling, stimulated patient fibroblasts and B cells fail to upregulate NF- κ B activity, which manifests in recurrent bacterial infections. In contrast to immunodeficient fibroblasts, peripheral blood mononuclear cells (PBMCs) of HOIP and HOIL-1-deficient

patients are highly responsive to IL-1 stimulation and produce high levels of proinflammatory cytokines IL-6 and MIP-1 α . One patient was noted to have severe T cell lymphopenia and impaired antibody production. Muscular amylopectinosis/myopathy appear to be unrelated to a defect of linear ubiquitination in immune cells and its mechanism remains to be investigated. Abnormalities in the lymphatic system have been observed in the HOIP-deficient patient and HOIP-deficient mice, which suggests

TABLE 2 | Clinical manifestations in patients with otulipenia, haploinsufficiency of A20 (HA20), and linear ubiquitin chain assembly complex deficiencies.

Clinical manifestations	Otulipenia (17)		HA20 (16, 35–37)				HOIL-1 deficiency (14)		HOIP deficiency (15)	
	Yes or no	Patients (n = 3)	Yes or no	Patients (16) (n = 11)	Patients (37) (n = 6)	Patients (36) (n = 3)	Patients (35) (n = 1)	Yes or no	Patients (n = 3)	Yes or no (n = 1)
Early age onset	Yes (1–4.5 months)	3/3	Yes (7 months–16 years)	11/11	6/6	3/3	1/1	Yes	3/3	Yes
Fevers	Yes (fever lasting 2–3 weeks)	3/3	Yes	2/11	3/6	1/3	1/1	Yes	3/3	Yes
Skin rash	Yes (erythematous with skin nodules, pustular rash)	3/3	Yes (erythematous papules, folliculitis, skin abscesses)	4/11	6/6	0/3	1/1	Yes (eczema; erythroderma, desquamative dermatitis)	1/3; 1/3	No
CNS	No	1/3	Yes (CNS vasculitis, chorea, migraine)	2/11	/	/	0/1	No	0/3	No
GI	Yes (abdominal pain, diarrhea)	1/3	Yes (colitis)	4/11	1/6	1/3	0/1	Yes (abdominal pain, diarrhea, vomiting, blood and mucus in the stools)	3/3	Yes (recurrent episodes of fatty diarrhea, intestinal lymphangiectasia)
Arthritis Arthralgias Myalgias	Yes (arthralgias, myalgias)	3/3	Yes (arthralgia, polyarthritis)	6/11	/	2/3	0/1	No	0/3	Yes
Elevated CRP, ESR	Yes	3/3	Yes	6/6	1/1	/	1/1	Yes	3/3	Yes
Immunodeficiency	No obvious primary immunodeficiency	2/3	Yes (IgG2 and 4 subclass deficiency, low anti-polysaccharide antibodies lymphopenia)	2/11	/	/	1/1	Yes (recurrent bacterial infections, memory B-cell deficiency, and hyper-IgA syndrome)	3/3	Yes (recurrent viral and bacterial infections, lymphopenia, antibody deficiency, hypogammaglobulinemia)
Oral ulcers	No	0/3	Yes	11/11	6/6	2/3	0/1	Yes	1/3	Yes
Genital ulcers	No	0/3	Yes	10/11	6/6	1/3	0/1	No	0/3	No
Ophtho	No	0/3	Yes (uveitis, retinal vasculitis)	3/11	/	1/3	/	No	0/3	No
Pathergy	No	0/3	Yes	3/11	/	/	/	No	0/3	No
Autoantibodies	No	0/3	Yes (RNP, ANA, lupus anticoagulant)	5/11	/	/	1/1	No	0/3	No
Panniculitis	Yes	3/3	No	0/11	0/6	0/3	0/1	No	0/3	No
Failure to thrive	Yes	3/3	No	0/11	0/6	0/3	0/1	Yes	3/3	Yes
Lipodystrophy	Yes	3/3	No	0/11	0/6	0/3	0/1	No	0/3	No
Lymphadenopathy	Yes	2/3	Yes	0/11	0/6	0/3	1/1	Yes	2/3	Yes
Systemic lymphangiectasia	No	0/3	No	0/11	0/6	0/3	0/1	No	0/3	Yes

(Continued)

TABLE 2 | Continued

Clinical manifestations	Otulipenia (17)		HA20 (16, 35–37)		HOIL-1 deficiency (14)		HOIP deficiency (15)	
	Yes or no	Patients (n = 3)	Patients (16) (n = 1)	Patients (37) (n = 6)	Patients (36) (n = 3)	Patients (35) (n = 1)	Patients (n = 3)	Yes or no (n = 1)
Weakness in lower extremities	No	0/3	0/11	0/6	0/3	0/1	2/3	Yes
Cardiomyopathy	No	0/3	0/11	0/6	0/3	0/1	3/3	Yes
Respiratory distress	No	0/3	0/11	0/6	0/3	0/1	1/3	Yes
Amylopectinosis	No	0/3	0/11	0/6	0/3	0/1	3/3	Yes
Systemic edema	No	0/3	0/11	0/6	0/3	0/1	0/3	Yes
Effective treatment	anti-TNF, anti-IL-1	2/3; 1/3	1/11; 4/11; 2/11	0/6; 0/6; 1/6	0/6; 0/6; 0/6	0/1; 0/1; 0/1	2/3; 1/3	Naproxen, antibiotic prophylaxis, IVIG

/, not reported.
Source of data: Ref. (14–18, 35–37).

that HOIP, independently of other LUBAC components, may have a function in the regulation of angiogenesis. LUBAC appears to be important for toll-like receptor 3 (TLR3)-mediated innate immune responses to influenza A virus infection, by enabling TLR3-mediated activation of NF-κB, MAPK, and IRF3 (34).

In summary, the identification of patients with LUBAC deficiencies showed the critical role of LUBAC in regulation of immune responses.

OTULIPENIA/ORAS

Recessively inherited loss-of-function mutations in the linear (Met1)-specific DUB OTULIN have been linked to the early-onset severe inflammatory disease, named otulipenia/ORAS (Table 1) (17, 18). Patients present with prolonged recurrent fevers, joint swelling, GI inflammation/diarrhea, and failure to thrive (Table 2; Figure 3). The cutaneous manifestations include painful erythematous rash with skin nodules, lipodystrophy, and episodes of pustular rash in one patient (Figure 3B). Skin biopsy showed evidence for neutrophilic dermatitis, mixed type panniculitis, and vasculitis of small and medium-sized blood vessels (Figure 3C) (17). In contrast to patients with the LUBAC deficiency, OTULIN-deficient patients have no obvious immunodeficiency, although some of them suffered from iatrogenic infections induced by immunosuppressive therapies. Initial analyses showed adequate specific antibody responses to vaccines, adequate T and B cell proliferative responses, normal levels of immunoglobulins, and normal T, B, and NK cell counts.

The four patients identified carry novel homozygous mutations in the *FAM105B* gene that encodes OTULIN. Heterozygote carriers are asymptomatic, which suggests that reduced protein expression of OTULIN may not be critical for maintenance of immune homeostasis. OTULIN is a 352-residue protein that consists of N-terminal LUBAC-binding domain and C-terminal ovarian tumor (OTU) domain. Disease-associated mutations affect the OTU domain and binding of OTULIN to linear Ub chains (Figure 2).

OTULIN functions as a negative regulator of the canonical NF-κB pathway and as such is essential for resolving inflammation (Table 1; Figure 1). Mutant OTULIN proteins cannot restrict the accumulation of Met1 Ub chains on target substrates IKK/NEMO, RIPK1, and ASC. Patients' mononuclear leukocytes and fibroblasts have a strong inflammatory signature as evidenced by increased degradation of IκBα and increased phosphorylation of IKKα/IKKβ and IκBα, the hallmarks of the activated NF-κB pathway. Mutant cells overproduce many proinflammatory cytokines, including cytokines associated with activation of adaptive immune cells, and therapy with TNF inhibitors is very effective in controlling disease activity. Tamoxifen-induced *Otulin* ablation in murine immune cells (CreERT2-*Otulin*^{LacZ/flox} chimeras) resembles the phenotype described in patients with otulipenia/ORAS including responsiveness to therapy with TNF inhibitors (18).

In summary, combined data from human and murine studies suggest that deficiency of the linear DUB, OTULIN, leads to amplification of the Met1 Ub regulated signaling in the canonical NF-κB pathway, most notably in myeloid cells.

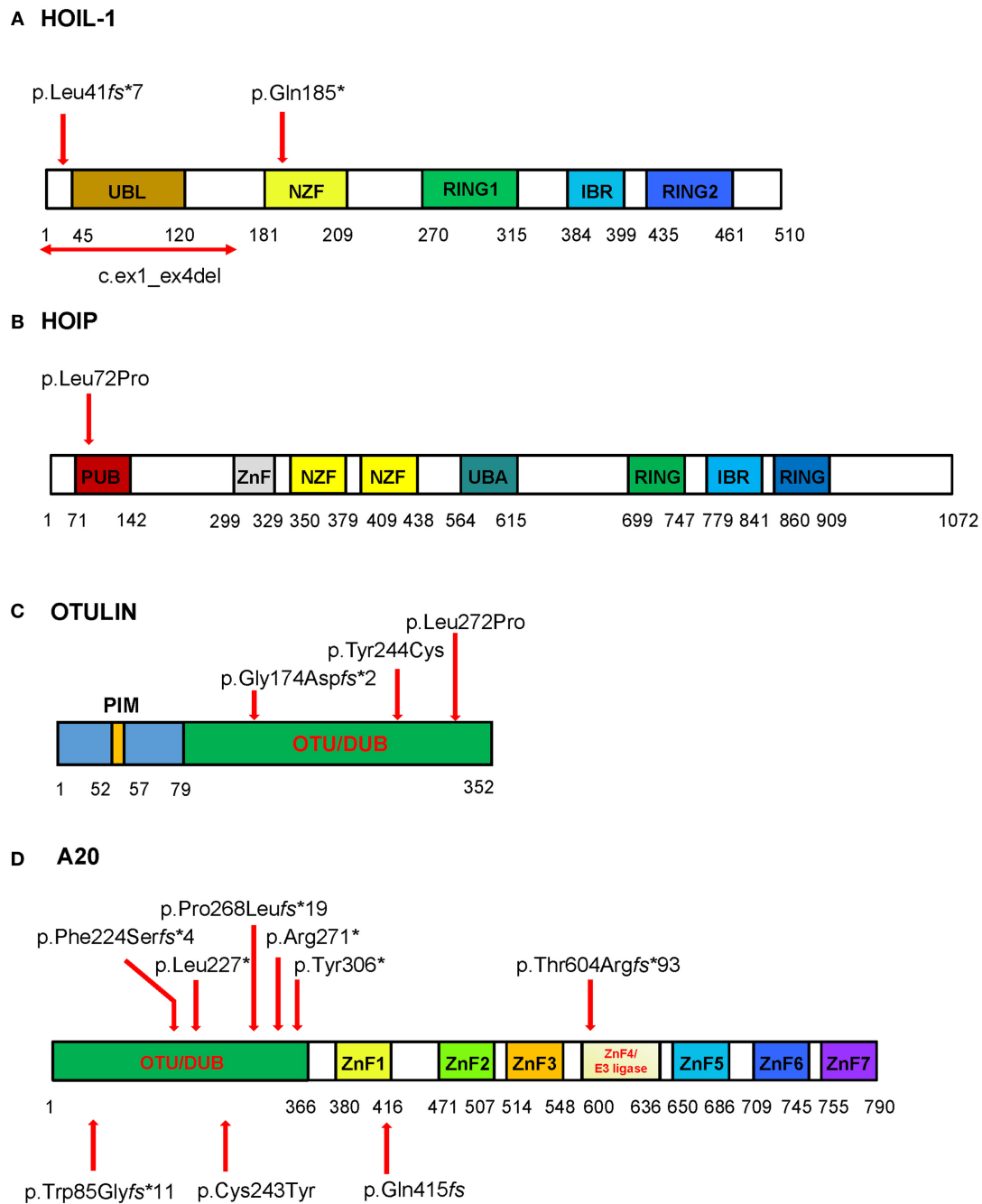


FIGURE 2 | Schematic of the domains and locations of mutations in respective proteins HOIL-1, HOIP, OTULIN, and TNFAIP3/A20. The domains identified are depicted as boxes. The locations of the mutations are indicated with red arrows. **(A)** HOIL-1 contains the ubiquitin-like (UBL), NPL4 zinc-fingers (NZF) domain, really interesting new gene (RING) domain, and in-between RING (IBR) domain. The mutations are located in UBL and NZF domains. The UBL domain is required for linear ubiquitin chain assembly complex (LUBAC) formation and linear ubiquitination. **(B)** HOIP has PNGase/UBA or UBX (PUB), zinc finger (ZnF), NZF, ubiquitin-associated (UBA), RING domain, and IBR domain. The homozygous disease-associated mutation is located in the PUB domain that is critical for interaction with OTULIN and stability of LUBAC. **(C)** OTULIN consists of N-terminal LUBAC-binding PUB-interacting motif (PIM) and C-terminal ovarian tumor (OTU) domain that mediates deubiquitinase (DUB) activity of OTULIN (79–352aa). All three mutations are located in the OTU domain. **(D)** The DUB activity of A20 is mediated by the OTU domain, and the ZnF domains mediate A20 ubiquitin (Ub) E3 ligase activity, binding to Lys63-linked Ub chains and dimerization. Mutations reported by Zhou et al. are shown on the top of the diagram, while three mutations reported in Japanese patients are shown below the diagram. To stay consistent with the Human Genome Variation Society nomenclature the reported p.Gln415fs (35) should be described as p.Lys417Serfs*4. The four nucleotide deletion in the repeat sequence (reported as c.1245_1248del) should be assigned as the most 3' position in the repeat, i.e., c.1249_1252del; therefore, the proposed nomenclature for the mutation is p.Lys417Serfs*4.

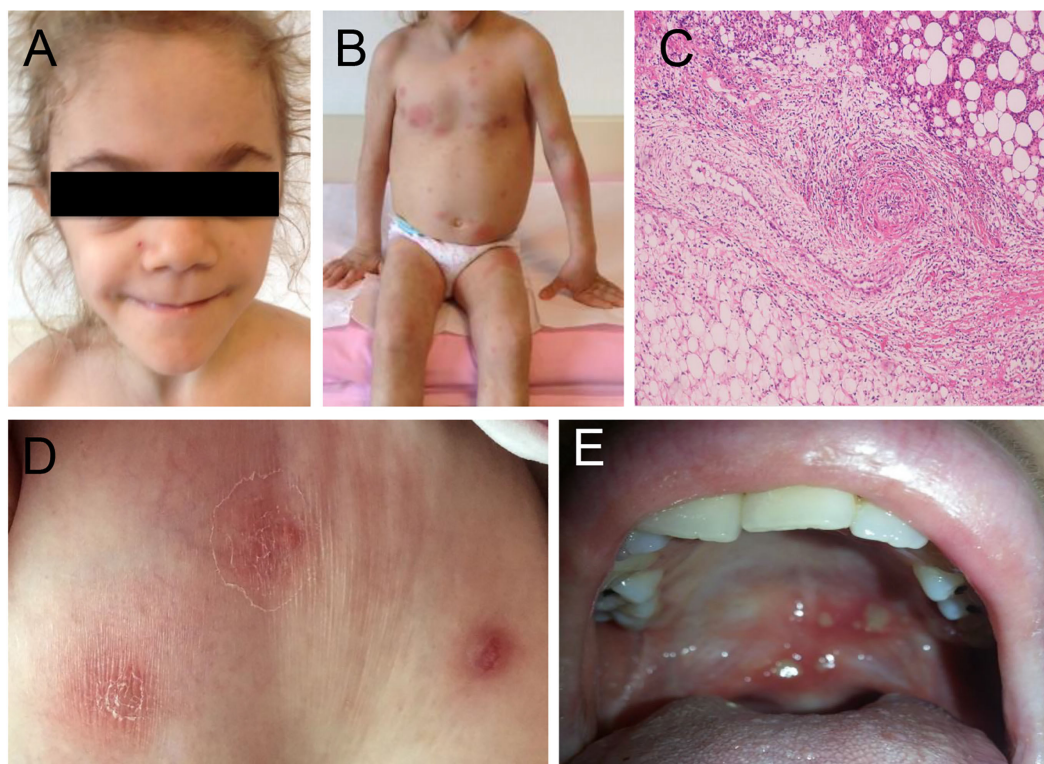


FIGURE 3 | Clinical manifestations of the patients with haploinsufficiency of A20 (HA20) and otulipenia/otulin-related autoinflammatory syndrome.

(A) Prominent fat loss (lipodystrophy) and **(B)** erythematous skin lesions and subcutaneous nodules in a patient with otulipenia. **(C)** Skin biopsy showed dense inflammatory infiltrate throughout the subcutaneous lobules (right upper corner of the image) and subcutaneous lobular atrophy or lipodystrophy (similar findings have been reported in chronic atypical neutrophilic dermatosis with lipodystrophy and elevated temperature patients) on the left lower corner of the image. The middle part of the image showed vasculitis affecting a medium-sized artery, characterized by dense intramural and perivascular inflammation with endothelial proliferation and vascular occlusion. Adjacent the affected artery is a medium-sized vein (left lower) showing mild inflammation of the vessel wall. **(D)** Dermal abscesses in a patient with HA20. **(E)** Recurrent aphthous (oral) ulcers in a patient with HA20.

Lys63-LINKED Ub CHAINS IN IMMUNE SIGNALING

Lys(K)63-Ub modification was first described as a mechanism for the activation of the canonical NF- κ B pathway (38). K63-linked Ub chains are generated following cell stimulation with inflammatory cytokines, and they function as a scaffold for the formation of receptor signaling complexes. Molecules such as NEMO/IKK, TRAF6, and RIPK1 are ubiquitinated both by linear and K63 Ub chains, which suggest a substantial regulatory redundancy in immune signaling (39–42). TNFAIP3/A20 restricts inflammatory responses *via* its dual yet synergistic functions: its deubiquitinase activity by hydrolyzing K63 linkages and its E3 ligase activity by conjugating substrates with K48 Ub chains to target them for proteasomal degradation (43, 44) (**Figure 1**). A20 is also subject to regulations as it undergoes posttranslational phosphorylation (45) and is cleaved by mucosa-associated lymphoid tissue lymphoma translocation protein 1 (46). In tumor cells, A20 acts as a tumor-suppressor gene and is frequently inactivated by somatic mutations and deletions in diffuse large B-cell and Hodgkin lymphomas (47, 48).

The *TNFAIP3* gene is highly conserved and intolerant to genetic variations, in particular to loss-of-function mutations. Common low-penetrance mostly non-coding variants in *TNFAIP3* have been associated with many autoimmune diseases including systemic lupus erythematosus (SLE) (49–51), RA (52), psoriasis (53), type 1 diabetes (54), celiac disease (55), coronary artery disease (56), inflammatory bowel disease (57), and more recently with protection to allergy and asthma (58, 59). Given the potent anti-inflammatory function of A20, these susceptibility alleles are predicted to decrease A20 expression and function, although that has been experimentally demonstrated only for a single non-coding variant associated with SLE. The dinucleotide functional variant downstream of *TNFAIP3*, TT > A, likely alters the binding of transcription factors in response to proinflammatory signals (51, 60).

Genetic ablation of A20 leads to spontaneous inflammation in mice with a range of cell-specific phenotypes. A20-deficient (A20^{-/-}) mice exhibit multiorgan inflammation, cachexia, and early lethality (61). Although A20 was initially described as required for termination of TNF-induced signals, the excessive inflammation observed in double-deficient mice, A20-TNF or

A20-TNFR1, suggested that A20 might be critical for the regulation of TNF-independent signals including the termination of TLR-induced activity of NF- κ B (62). Cell-specific deletions of A20 resemble human autoimmune diseases, from a mild autoimmune phenotype in *Tnfaip3 Cd19* (B-cell) KO mice to severe spontaneous inflammation in mice with A20-deficient dendritic cells. Loss of A20 in macrophages mimics human RA, although the phenotype appears to be TNF-independent. Deficiency of A20 in keratinocytes leads to hyperkeratosis, while loss of A20 in intestinal epithelial cells causes DSS-induced TNF-dependent colitis (63). Aging heterozygous mice (A20^{+/-}) develop spontaneous autoantibodies. In summary, multiple murine models with cell-specific ablation of A20 demonstrate closer approximation of human diseases than the complete knock-out mice.

This past year, Zhou et al. reported 11 patients from 6 families with a new dominantly inherited autoinflammatory disease, termed haploinsufficiency of A20, characterized by childhood-onset episodic fevers, arthralgia/arthritis, oral and/or genital ulcers, skin pathergy, GI, and ocular inflammation (16) (**Table 2; Figures 3D,E**). These symptoms resemble Behcet's disease (BD). One patient was initially diagnosed with SLE and presented with CNS vasculitis and idiopathic thrombocytopenic purpura. Subsequently, two families of Japanese ancestry diagnosed with entero-BD and one Japanese patient diagnosed with autoimmune lymphoproliferative syndrome (ALPS) were reported to carry novel LOF mutations in the gene (**Figure 2**) (35–37). A patient with ALPS presented with fevers, bilateral cervical lymphadenopathy, extensive skin rash, and massive hepatosplenomegaly. The patient's immunophenotyping revealed an increased percentage of DNT cells and a decreased number of IgM memory B cells, which is characteristic of ALPS. However, unlike in ALPS the patient's central memory (TCM), naïve, TEMRA, and effector memory (TEM) subpopulations of CD³⁺CD⁸⁺ cells were normal (35).

TNFAIP3/A20 is a 790-residue protein that consists of an amino-terminal OTU followed by seven zinc finger (ZnF) domains. HA20-associated mutations create truncated mutant proteins, and most of them are located in the OTU domain (**Figure 2**). In addition to its OTU domain-mediated DUB activity, A20 can downregulate IKK activation by blocking IKK phosphorylation (64). Two pathogenic mutations have been identified in the ZnF domains 1 and 4. The ZnF4 domain is essential for A20 E3 ligase activity and dimerization (65).

Similar to patients with otulipenia/ORAS, mutant A20 cells have enhanced NF- κ B activity as demonstrated by increased phosphorylation of IKK α / β and increased degradation of I κ B α . Stimulated patient PBMCs and fibroblasts failed to hydrolyze K63 Ub chains from NEMO/IKK, RIPK1, and TNFR1 (16). Accumulation of K63 Ub proteins on these molecules triggers activity of the NF- κ B and the MAPK pathways (**Table 1; Figure 1**). In murine models, A20/*Tnfaip3* was shown to downregulate the activity of NLRP3 inflammasome (40, 66). Zhou et al. demonstrated constitutive NLRP3 activity in PBMCs of HA20 patients. Stimulated PBMCs and serum samples of HA20 patients have highly elevated levels of many proinflammatory cytokines produced by myeloid cells (IL-1, TNF, IL-6, IL-18, and IP-10)

and T cells (IL-9, IL-17, and IFN γ). Therapies with cytokine inhibitors, anti-TNF, or anti-IL-1, have been very efficient in suppressing systemic inflammation.

Taken together, human genetic studies and murine models of A20 deficiency provide strong evidence that the reduced expression of A20 is associated with a range of inflammatory phenotypes.

CONCLUSION

Maintenance of immune homeostasis is a highly balanced act that requires coordinated action of many proteins to allow optimal and efficient immune responses. Discovery of otulipenia/ORAS, HA20, and LUBAC-associated diseases has reiterated the importance of ubiquitination in regulation of immune signaling and revealed cell-specific functions of these proteins.

Despite similar function of OTULIN and A20 in restricting immune responses, the DUB activity of A20 appears to be less critical than the one of OTULIN (22). This may explain a milder inflammatory phenotype in HA20 than in patients with otulipenia. In addition, patients with otulipenia have more profound protein deficiency (less than 50%) than the patients with HA20 who retain one functional allele of the gene (50% protein deficiency).

Given the importance of the ubiquitination in cellular physiology, the UPS system has elicited a significant interest for drug development. The list of human diseases related to abnormalities in UPS has been steadily increasing and includes neurodegenerative diseases, cancer, and immune diseases. Ub-mediated protein degradation is critical for homeostasis in aging neuronal cells (67). Deficiency of A20 has been linked to lymphomas, and its most aggressive subtype is associated with constitutive activation of NF- κ B. Reconstitution of A20 in mutant cell lines induced apoptosis and suppressed tumor growth (47). In immune diseases, A20 and OTULIN might be new therapeutic targets for development of immunomodulatory drugs that can potentially increase or stabilize their expression.

A key challenge for finding effective drugs will be in developing cell-based therapies. The ubiquitination process is regulated at multiple levels: generation, recognition, and removal. Targeting more components of the Ub–proteasome pathway may provide new opportunities for therapeutic exploitation and drug discovery (68, 69).

AUTHOR CONTRIBUTIONS

IA reviewed the literature and wrote the manuscript. QZ reviewed the literature to make figures and tables and helped with writing the manuscript.

ACKNOWLEDGMENTS

We would like to thank Dr. C. Richard Lee from National Cancer Institute for his help with interpretation of the dermal histology slide and Ms. Julia Fececs from NHGRI for her help with the figures.

REFERENCES

- Komander D, Rape M. The ubiquitin code. *Annu Rev Biochem* (2012) 81:203–29. doi:10.1146/annurev-biochem-060310-170328
- Ye Y, Rape M. Building ubiquitin chains: E2 enzymes at work. *Nat Rev Mol Cell Biol* (2009) 10:755–64. doi:10.1038/nrm2780
- Emmerich CH, Ordureau A, Strickson S, Arthur JSC, Pedrioli PGA, Komander D, et al. Activation of the canonical IKK complex by K63/M1-linked hybrid ubiquitin chains. *Proc Natl Acad Sci U S A* (2013) 110:15247–52. doi:10.1073/pnas.1314715110
- Swatek KN, Komander D. Ubiquitin modifications. *Cell Res* (2016) 26:399–422. doi:10.1038/cr.2016.39
- Hershko A, Ciechanover A. The ubiquitin system. *Annu Rev Biochem* (1998) 67:425–79. doi:10.1146/annurev.biochem.67.1.425
- Bremm A, Komander D. Emerging roles for Lys11-linked polyubiquitin in cellular regulation. *Trends Biochem Sci* (2011) 36:355–63. doi:10.1016/j.tibs.2011.04.004
- Chen ZJJ. Ubiquitination in signaling and activation of IKK. *Immunol Rev* (2012) 246:95–106. doi:10.1111/j.1600-065X.2012.01108.x
- Fiil BK, Gyrd-Hansen M. Met1-linked ubiquitination in immune signalling. *FEBS J* (2014) 281:4337–50. doi:10.1111/febs.12944
- Iwai K, Fujita H, Sasaki Y. Linear ubiquitin chains: NF-kappa B signalling, cell death and beyond. *Nat Rev Mol Cell Biol* (2014) 15:503–8. doi:10.1038/nrm3836
- Shimizu Y, Taraborrelli L, Walczak H. Linear ubiquitination in immunity. *Immunol Rev* (2015) 266:190–207. doi:10.1111/imr.12309
- Komander D, Clague MJ, Urbe S. Breaking the chains: structure and function of the deubiquitinases. *Nat Rev Mol Cell Biol* (2009) 10:550–63. doi:10.1038/nrm2731
- Harhaj EW, Dixit VM. Regulation of NF-kappa B by deubiquitinases. *Immunol Rev* (2012) 246:107–24. doi:10.1111/j.1600-065X.2012.01100.x
- Liu Y, Ramot Y, Torreló A, Paller AS, Si N, Babay S, et al. Mutations in proteasome subunit β 5 type 8 cause chronic atypical neutrophilic dermatosis with lipodystrophy and elevated temperature with evidence of genetic and phenotypic heterogeneity. *Arthritis Rheum* (2012) 64:895–907. doi:10.1002/art.33368
- Boisson B, Laplantine E, Prando C, Giliani S, Israelsson E, Xu ZH, et al. Immunodeficiency, autoinflammation and amylopectinosis in humans with inherited HOIL-1 and LUBAC deficiency. *Nat Immunol* (2012) 13:1178. doi:10.1038/ni.2457
- Boisson B, Laplantine E, Dobbs K, Cobat A, Tarantino N, Hazen M, et al. Human HOIP and LUBAC deficiency underlies autoinflammation, immunodeficiency, amylopectinosis, and lymphangiectasia. *J Exp Med* (2015) 212:939–51. doi:10.1084/jem.20141130
- Zhou Q, Wang H, Schwartz DM, Stoffels M, Park YH, Zhang Y, et al. Loss-of-function mutations in TNFAIP3 leading to A20 haploinsufficiency cause an early-onset autoinflammatory disease. *Nat Genet* (2016) 48:67–73. doi:10.1038/ng.3459
- Zhou Q, Yu XM, Demirkaya E, Deutch N, Stone D, Tsai WXL, et al. Biallelic hypomorphic mutations in a linear deubiquitinase define otulipenia, an early-onset autoinflammatory disease. *Proc Natl Acad Sci U S A* (2016) 113:10127–32. doi:10.1073/pnas.1612594113
- Damgaard RB, Walker JA, Marco-Casanova P, Morgan NV, Titheradge HL, Elliott PR, et al. The deubiquitinase OTULIN is an essential negative regulator of inflammation and autoimmunity. *Cell* (2016) 166:1215. doi:10.1016/j.cell.2016.07.019
- Gerlach B, Cordier SM, Schmukle AC, Emmerich CH, Rieser E, Haas TL, et al. Linear ubiquitination prevents inflammation and regulates immune signalling. *Nature* (2011) 471:591. doi:10.1038/nature09816
- Rahighi S, Ikeda F, Kawasaki M, Akutsu M, Suzuki N, Kato R, et al. Specific recognition of linear ubiquitin chains by NEMO is important for NF-kappa B activation. *Cell* (2009) 136:1098–109. doi:10.1016/j.cell.2009.03.007
- Tokunaga F, Sakata S, Saeki Y, Satomi Y, Kirisako T, Kamei K, et al. Involvement of linear polyubiquitylation of NEMO in NF-kappa B activation. *Nat Cell Biol* (2009) 11:123–32. doi:10.1038/ncb1821
- Keusekotten K, Elliott PR, Glockner L, Fiil BK, Damgaard RB, Kulathu Y, et al. OTULIN antagonizes LUBAC signaling by specifically hydrolyzing Met1-linked polyubiquitin. *Cell* (2013) 153:1312–26. doi:10.1016/j.cell.2013.05.014
- Schaeffer V, Akutsu M, Olma MH, Gomes LC, Kawasaki M, Dikic I. Binding of OTULIN to the PUB domain of HOIP controls NF-kappa B signaling. *Mol Cell* (2014) 54:349–61. doi:10.1016/j.molcel.2014.03.016
- Elliott PR, Nielsen SV, Marco-Casanova P, Fiil BK, Keusekotten K, Mailand N, et al. Molecular basis and regulation of OTULIN-LUBAC interaction. *Mol Cell* (2014) 54:335–48. doi:10.1016/j.molcel.2014.03.018
- Abdul-Sater AA, Edilova MI, Clouthier DL, Mbanwi A, Kremmer E, Watts TH. The signaling adaptor TRAF1 negatively regulates toll-like receptor signaling and this underlies its role in rheumatic disease. *Nat Immunol* (2017) 18:26–35. doi:10.1038/ni.3618
- Peltzer N, Rieser E, Taraborrelli L, Draber P, Darding M, Pemaute B, et al. HOIP deficiency causes embryonic lethality by aberrant TNFR1-mediated endothelial cell death. *Cell Rep* (2014) 9:153–65. doi:10.1016/j.celrep.2014.08.066
- HogenEsch H, Janke S, Boggess D, Sundberg JP. Absence of Peyer's patches and abnormal lymphoid architecture in chronic proliferative dermatitis (cpdm/cpdm) mice. *J Immunol* (1999) 162:3890–6.
- Potter CS, Wang Z, Silva KA, Kennedy VE, Stearns TM, Burzenski L, et al. Chronic proliferative dermatitis in Sharpin null mice: development of an autoinflammatory disease in the absence of B and T lymphocytes and IL4/IL13 signaling. *PLoS One* (2014) 9:e85666. doi:10.1371/journal.pone.0085666
- Kumari S, Redouane Y, Lopez-Mosqueda J, Shiraishi R, Romanowska M, Lutzmayr S, et al. Sharpin prevents skin inflammation by inhibiting TNFR1-induced keratinocyte apoptosis. *Elife* (2014) 3. doi:10.7554/eLife.03422
- Rickard JA, Anderton H, Etemadi N, Nachbur U, Darding M, Peltzer N, et al. TNFR1-dependent cell death drives inflammation in Sharpin-deficient mice. *Elife* (2014) 3. doi:10.7554/eLife.03464
- Sasaki Y, Sano S, Nakahara M, Murata S, Kometani K, Aiba Y, et al. Defective immune responses in mice lacking LUBAC-mediated linear ubiquitination in B cells. *EMBO J* (2013) 32:2463–76. doi:10.1038/emboj.2013.184
- Okamura K, Kitamura A, Sasaki Y, Chung DH, Kagami S, Iwai K, et al. Survival of mature T cells depends on signaling through HOIP. *Sci Rep* (2016) 6:36135. doi:10.1038/srep36135
- Rivkin E, Almeida SM, Ceccarelli DF, Juang YC, MacLean TA, Srikumar T, et al. The linear ubiquitin-specific deubiquitinase gumby regulates angiogenesis. *Nature* (2013) 498:318. doi:10.1038/nature12296
- Zinngrebe J, Rieser E, Taraborrelli L, Peltzer N, Hartwig T, Ren H, et al. LUBAC deficiency perturbs TLR3 signaling to cause immunodeficiency and autoinflammation. *J Exp Med* (2016) 213:2671–89. doi:10.1084/jem.20160041
- Takagi M, Ogata S, Ueno H, Yoshida K, Yeh T, Hoshino A, et al. Haploinsufficiency of TNFAIP3 (A20) by germline mutation is involved in autoimmune lymphoproliferative syndrome. *J Allergy Clin Immunol* (2016). doi:10.1016/j.jaci.2016.09.038
- Ohnishi H, Kawamoto N, Seishima M, Ohara O, Fukao T. A Japanese family case with juvenile onset Behcet's disease caused by TNFAIP3 mutation. *Allergol Int* (2016) 66:146–8. doi:10.1016/j.alit.2016.06.006
- Shigemura T, Kaneko N, Kobayashi N, Kobayashi K, Takeuchi Y, Nakano N, et al. Novel heterozygous C243Y A20/TNFAIP3 gene mutation is responsible for chronic inflammation in autosomal-dominant Behcet's disease. *RMD Open* (2016) 2:e000223. doi:10.1136/rmdopen-2015-000223
- Chen ZJ, Parent L, Maniatis T. Site-specific phosphorylation of I κ B kinase by a novel ubiquitination-dependent protein kinase activity. *Cell* (1996) 84:853–62. doi:10.1016/S0092-8674(00)81064-8
- Chen J, Chen ZJ. Regulation of NF-kappaB by ubiquitination. *Curr Opin Immunol* (2013) 25:4–12. doi:10.1016/j.coi.2012.12.005
- Duong BH, Onizawa M, Oses-Prieto JA, Advincula R, Burlingame A, Malynn BA, et al. A20 restricts ubiquitination of pro-interleukin-1beta protein complexes and suppresses NLRP3 inflammasome activity. *Immunity* (2015) 42:55–67. doi:10.1016/j.immuni.2014.12.031
- Hadian K, Griesbach RA, Dornauer S, Wanger TM, Nagel D, Metlitzky M, et al. NF-kappaB essential modulator (NEMO) interaction with linear and lys-63 ubiquitin chains contributes to NF-kappaB activation. *J Biol Chem* (2011) 286:26107–17. doi:10.1074/jbc.M111.233163
- Coornaert B, Carpentier I, Beyaert R. A20: central gatekeeper in inflammation and immunity. *J Biol Chem* (2009) 284:8217–21. doi:10.1074/jbc.R800032200
- Catrysse L, Vereecke L, Beyaert R, van Loo G. A20 in inflammation and autoimmunity. *Trends Immunol* (2014) 35:22–31. doi:10.1016/j.it.2013.10.005

44. Vereecke L, Beyaert R, van Loo G. The ubiquitin-editing enzyme A20 (TNFAIP3) is a central regulator of immunopathology. *Trends Immunol* (2009) 30:383–91. doi:10.1016/j.it.2009.05.007
45. Hutti JE, Turk BE, Asara JM, Ma A, Cantley LC, Abbott DW. IkappaB kinase beta phosphorylates the K63 deubiquitinase A20 to cause feedback inhibition of the NF-kappaB pathway. *Mol Cell Biol* (2007) 27:7451–61. doi:10.1128/MCB.01101-07
46. Coornaert B, Baens M, Heynincx K, Bekaert T, Haegman M, Staal J, et al. T cell antigen receptor stimulation induces MALT1 paracaspase-mediated cleavage of the NF-kappaB inhibitor A20. *Nat Immunol* (2008) 9:263–71. doi:10.1038/ni1561
47. Compagno M, Lim WK, Grunn A, Nandula SV, Brahmachary M, Shen Q, et al. Mutations of multiple genes cause deregulation of NF-kappaB in diffuse large B-cell lymphoma. *Nature* (2009) 459:717–21. doi:10.1038/nature07968
48. Schmitz R, Hansmann ML, Bohle V, Martin-Subero JI, Hartmann S, Mechttersheimer G, et al. TNFAIP3 (A20) is a tumor suppressor gene in Hodgkin lymphoma and primary mediastinal B cell lymphoma. *J Exp Med* (2009) 206:981–9. doi:10.1084/jem.20090528
49. Graham RR, Cotsapas C, Davies L, Hackett R, Lessard CJ, Leon JM, et al. Genetic variants near TNFAIP3 on 6q23 are associated with systemic lupus erythematosus. *Nat Genet* (2008) 40:1059–61. doi:10.1038/ng.200
50. Musone SL, Taylor KE, Lu TT, Nititham J, Ferreira RC, Ortmann W, et al. Multiple polymorphisms in the TNFAIP3 region are independently associated with systemic lupus erythematosus. *Nat Genet* (2008) 40:1062–4. doi:10.1038/ng.202
51. Adrianto I, Wen F, Templeton A, Wiley G, King JB, Lessard CJ, et al. Association of a functional variant downstream of TNFAIP3 with systemic lupus erythematosus. *Nat Genet* (2011) 43:253–8. doi:10.1038/ng.766
52. Thomson W, Barton A, Ke X, Eyre S, Hinks A, Bowes J, et al. Rheumatoid arthritis association at 6q23. *Nat Genet* (2007) 39:1431–3. doi:10.1038/ng.2007.32
53. Nair RP, Duffin KC, Helms C, Ding J, Stuart PE, Goldgar D, et al. Genome-wide scan reveals association of psoriasis with IL-23 and NF-kappaB pathways. *Nat Genet* (2009) 41:199–204. doi:10.1038/ng.311
54. Fung EY, Smyth DJ, Howson JM, Cooper JD, Walker NM, Stevens H, et al. Analysis of 17 autoimmune disease-associated variants in type 1 diabetes identifies 6q23/TNFAIP3 as a susceptibility locus. *Genes Immun* (2009) 10:188–91. doi:10.1038/gene.2008.99
55. Trynka G, Zhernakova A, Romanos J, Franke L, Hunt KA, Turner G, et al. Coeliac disease-associated risk variants in TNFAIP3 and REL implicate altered NF-kappaB signalling. *Gut* (2009) 58:1078–83. doi:10.1136/gut.2008.169052
56. Boonyasrisawat W, Eberle D, Bacci S, Zhang YY, Nolan D, Gervino EV, et al. Tag polymorphisms at the A20 (TNFAIP3) locus are associated with lower gene expression and increased risk of coronary artery disease in type 2 diabetes. *Diabetes* (2007) 56:499–505. doi:10.2337/db06-0946
57. Lodolce JP, Kolodziej LE, Rhee L, Kariuki SN, Franek BS, McGreal NM, et al. African-derived genetic polymorphisms in TNFAIP3 mediate risk for autoimmunity. *J Immunol* (2010) 184:7001–9. doi:10.4049/jimmunol.1000324
58. Li X, Ampleford EJ, Howard TD, Moore WC, Torgerson DG, Li H, et al. Genome-wide association studies of asthma indicate opposite immunopathogenesis direction from autoimmune diseases. *J Allergy Clin Immunol* (2012) 130:861–8.e7. doi:10.1016/j.jaci.2012.04.041
59. Schuijs MJ, Willart MA, Vergote K, Gras D, Deswarte K, Ege MJ, et al. Farm dust and endotoxin protect against allergy through A20 induction in lung epithelial cells. *Science* (2015) 349:1106–10. doi:10.1126/science.aac6623
60. Nititham J, Taylor KE, Gupta R, Chen H, Ahn R, Liu J, et al. Meta-analysis of the TNFAIP3 region in psoriasis reveals a risk haplotype that is distinct from other autoimmune diseases. *Genes Immun* (2015) 16:120–6. doi:10.1038/gene.2014.75
61. Lee EG, Boone DL, Chai S, Libby SL, Chien M, Lodolce JP, et al. Failure to regulate TNF-induced NF-kappaB and cell death responses in A20-deficient mice. *Science* (2000) 289:2350–4. doi:10.1126/science.289.5488.2350
62. Boone DL, Turer EE, Lee EG, Ahmad RC, Wheeler MT, Tsui C, et al. The ubiquitin-modifying enzyme A20 is required for termination of toll-like receptor responses. *Nat Immunol* (2004) 5:1052–60. doi:10.1038/ni1110
63. Martin F, Dixit VM. A20 edits ubiquitin and autoimmune paradigms. *Nat Genet* (2011) 43:822–3. doi:10.1038/ng.916
64. Skaug B, Chen J, Du F, He J, Ma A, Chen ZJ. Direct, noncatalytic mechanism of IKK inhibition by A20. *Mol Cell* (2011) 44:559–71. doi:10.1016/j.molcel.2011.09.015
65. Lu TT, Onizawa M, Hammer GE, Turer EE, Yin Q, Damko E, et al. Dimerization and ubiquitin mediated recruitment of A20, a complex deubiquitinating enzyme. *Immunity* (2013) 38:896–905. doi:10.1016/j.immuni.2013.03.008
66. Vande Walle L, Van Opdenbosch N, Jacques P, Fossoul A, Verheugen E, Vogel P, et al. Negative regulation of the NLRP3 inflammasome by A20 protects against arthritis. *Nature* (2014) 512:69–73. doi:10.1038/nature13322
67. Atkin G, Paulson H. Ubiquitin pathways in neurodegenerative disease. *Front Mol Neurosci* (2014) 7:63. doi:10.3389/fnmol.2014.00063
68. Zhu J, Zhuang T, Yang H, Li X, Liu H, Wang H. Atypical ubiquitin ligase RNF31: the nuclear factor modulator in breast cancer progression. *BMC Cancer* (2016) 16:538. doi:10.1186/s12885-016-2575-8
69. Yang Y, Kelly P, Shaffer AL III, Schmitz R, Yoo HM, Liu X, et al. Targeting non-proteolytic protein ubiquitination for the treatment of diffuse large B cell lymphoma. *Cancer Cell* (2016) 29:494–507. doi:10.1016/j.ccell.2016.03.006

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The reviewer, JY, and handling Editor declared their shared affiliation, and the handling Editor states that the process nevertheless met the standards of a fair and objective review.

Copyright © 2017 Aksentijevich and Zhou. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.