



Studying autism in rodent models: reconciling endophenotypes with comorbidities

Andrew Argyropoulos¹, Krista L. Gilby¹ and Elisa L. Hill-Yardin²*

¹ Department of Medicine, The University of Melbourne, Parkville, VIC, Australia

² Department of Physiology, The University of Melbourne, Parkville, VIC, Australia

Edited by:

Charles Claudianos, University of Queensland, Australia

Reviewed by:

Wah Chin Boon, Florey Neuroscience Institutes, Australia

Thomas Burne, Queensland Brain Institute, Australia

*Correspondence:

Elisa L. Hill-Yardin, Enteric Neuroscience Laboratory, Department of Physiology, The University of Melbourne, Cnr of Grattan Street and Royal Parade, Parkville, VIC 3010, Australia
e-mail: elhill@unimelb.edu.au

Autism spectrum disorder (ASD) patients commonly exhibit a variety of comorbid traits including seizures, anxiety, aggressive behavior, gastrointestinal problems, motor deficits, abnormal sensory processing, and sleep disturbances for which the cause is unknown. These features impact negatively on daily life and can exaggerate the effects of the core diagnostic traits (social communication deficits and repetitive behaviors). Studying endophenotypes relevant to both core and comorbid features of ASD in rodent models can provide insight into biological mechanisms underlying these disorders. Here we review the characterization of endophenotypes in a selection of environmental, genetic, and behavioral rodent models of ASD. In addition to exhibiting core ASD-like behaviors, each of these animal models display one or more endophenotypes relevant to comorbid features including altered sensory processing, seizure susceptibility, anxiety-like behavior, and disturbed motor functions, suggesting that these traits are indicators of altered biological pathways in ASD. However, the study of behaviors paralleling comorbid traits in animal models of ASD is an emerging field and further research is needed to assess altered gastrointestinal function, aggression, and disorders of sleep onset across models. Future studies should include investigation of these endophenotypes in order to advance our understanding of the etiology of this complex disorder.

Keywords: autism, epilepsy, sleep, motor deficits, aggression, sensory, gastrointestinal function, anxiety

Studying endophenotypes in rodent models of Autism spectrum disorder (ASD) can offer insights into the heterogeneity and underlying biological causes of this complex disorder. Patients with ASD demonstrate a high degree of variability in both the severity of core diagnostic symptoms (social communication deficits alongside repetitive behaviors) and in the nature and strength of a range of associated comorbidities. If comorbid traits associated with ASD are integral to the disorder we expect that many of these traits will present in animal models. However, despite the prevalence of comorbidities in patients, studies in animal models to date have largely focused on characterizing core behavioral traits. Here we review findings from salient rodent models of ASD identifying endophenotypes that parallel core ASD deficits in combination with one or more comorbid traits commonly reported in patients.

ASD: COMORBID TRAITS

Comorbid traits in ASD include seizures, heightened aggression, and anxiety disorders as well as gastrointestinal problems, altered sensory processing, motor deficits, and sleep disorders (Table 1). While treatment of these issues can significantly improve quality of life for patients and their families, the biological mechanisms underlying these symptoms and their co-expression are generally unknown in the context of ASD.

Current estimates for the prevalence of epilepsy in ASD patients range between 8 and 25% (Hara, 2007; Jeste, 2011; Sansa et al., 2011; Woolfenden et al., 2012). Recent meta-analysis data show

that epilepsy is more common in ASD patients with an intellectual disability (21.5 vs. 8%; Woolfenden et al., 2012). When epilepsy and abnormal EEG data are compared within the general ASD population, 15% of ASD subjects have an epilepsy diagnosis whereas a larger proportion (24.6%) shows interictal epileptiform EEG abnormalities during sleep (Ekinici et al., 2010). Other reports reveal that as many as 25% of ASD patients have comorbid epilepsy, and that 45.5% show non-seizure-related EEG abnormalities (Parmeggiani et al., 2010). Furthermore, one third (34%) of patients with idiopathic ASD have treatment resistant epilepsy (Sansa et al., 2011).

Aggressive behavior and elevated anxiety are frequently reported in children and adolescents with ASD. Caregiver surveys suggest that as many as 68% of ASD patients show episodes of aggression toward them (Kanne and Mazurek, 2011). Pouw et al. (2013) found that aggression behaviors in ASD are most likely due to a relative impairment in the understanding of the emotions of others. It is also estimated that 40% of ASD patients have at least one anxiety disorder (van Steensel et al., 2011). Specific phobias, obsessive compulsive disorder, and social anxiety disorder are most frequently observed.

A significant proportion of patients with ASD also suffer from gastrointestinal problems (42–90%); with constipation, chronic diarrhea, abnormal stool patterns, and stomach cramps frequently reported (Parracho et al., 2005; Valicenti-McDermott et al., 2006; Ibrahim et al., 2009; Buie et al., 2010; Wang et al., 2011a). Alterations in gastrointestinal function in the context

Table 1 | Clinical comorbidities commonly associated with ASD.

Domain	Comorbid symptoms
Epilepsy	High prevalence of epilepsy (8–25%) and EEG abnormalities (46%) in ASD patients (Amiet et al., 2008; Parmeggiani et al., 2010; Jeste, 2011) High rate of treatment resistant epilepsy in idiopathic autism (34%) (Dudova et al., 2011; Sansa et al., 2011)
Heightened aggression	Approximately 70% of ASD patients exhibit aggression toward caregivers (Kanne and Mazurek, 2011) Reactive aggression correlates with impairments in emotional regulation in children with ASD but not in typically developing children (Pouw et al., 2013)
Anxiety	40% of ASD cases associated with at least one comorbid anxiety disorder (van Steensel et al., 2011)
Gastrointestinal disturbances	Up to 90% ASD patients have chronic GI problems, most commonly constipation, also abdominal pain, diarrhea, and bloating (Parracho et al., 2005; Ibrahim et al., 2009; Buie et al., 2010)
Sensory	Tactile: heightened sensitivity to vibration and thermal pain in palm and forearm (Blakemore et al., 2006; Cascio et al., 2008) Auditory: atypical change detection of auditory stimuli (Gomot et al., 2006; Kwakye et al., 2011) Visual: superior performance in detail oriented tasks, deficits in motion perception (Dakin and Frith, 2005; Latham et al., 2013; Robertson et al., 2013) Altered olfaction and taste in high-functioning ASD patients (Bennetto et al., 2007; Dudova et al., 2011)
Motor impairment	Delays in gross and fine motor domains (Jeste, 2011) Deficits in motor planning, coordination, and gait (Rinehart et al., 2006; Jeste, 2011)
Sleep	Sleep disturbances (quality, quantity, latency to sleep) found in 40–80% of children and adolescents with ASD (Allik et al., 2006; Malow et al., 2006; Jeste, 2011) Sleep onset problems and night waking common in 2- to 5-year-olds with ASD (Krakowiak et al., 2008)

Comorbid traits observed in patients with ASD are heterogeneous and include enhanced seizure susceptibility, heightened aggression, anxiety, gastrointestinal (GI) disturbances, altered sensory and motor function, and sleep disorders.

of ASD are thought to be linked to the effects of anxiety and thereby mediated via CNS function; however investigations into mechanisms involving the enteric nervous system have not been reported.

By far the most common changes associated with ASD are those related to sensory processing which are present in over 90% of individuals diagnosed with ASD (Leekam et al., 2007). Patients with Asperger Syndrome show significantly higher sensitivity to high frequency tactile stimuli compared to control subjects (Cascio et al., 2008). Abnormalities in tactile sensitivity, as well as hypersensitivity to hot and cold stimuli have also been reported in adults with ASD (Blakemore et al., 2006). Auditory processing deficits related to the discrimination of temporally separated tones (Kwakye et al., 2011) and impaired odor detection thresholds (Bennetto et al., 2007; Dudova et al., 2011) have been documented in patients with high-functioning autism as well as subtle impairments in identifying tastes (Bennetto et al., 2007). Interestingly, aberrant motion perception can occur alongside superior visual processing performance in detail oriented tasks, highlighting the potential complexity of sensory changes in ASD patients (reviewed in Dakin and Frith, 2005; also see Latham et al., 2013; Robertson et al., 2013).

Motor abnormalities occur in 60–80% of individuals with ASD and include hypotonia, apraxia, and subtle gait anomalies (see Geschwind, 2009 for review). Abnormal fine and gross motor function, as well as delayed motor learning, dyspraxia, and postural abnormalities are also commonly reported in ASD patients (reviewed in Jeste, 2011). Finally, difficulties initiating

sleep, frequent night time waking, and insomnia are frequently reported in children with ASD (Allik et al., 2006; Malow et al., 2006; Krakowiak et al., 2008; Jeste, 2011).

The systematic analysis of traits in animal models corresponding to patient comorbidities can potentially provide insight into the underlying biological mechanisms of ASD. Such outcomes may lead to the design of new therapies and benefits to patients.

ANIMAL MODELS OF ASD

Over the last decade, a substantial number of rodent models of ASD have been generated (reviewed in Silverman et al., 2010a; Peca et al., 2011; Penagarikano et al., 2011; Wang et al., 2011b; Schmeisser et al., 2012; Won et al., 2012) and demonstrate face validity by replicating behavioral traits relevant to ASD. Well-characterized social and communication assessment paradigms and tests for the presence of repetitive behaviors exist for rodent models of ASD (Silverman et al., 2010a). In addition, a battery of tests is available to determine the presence of potential comorbidities including anxiety-like and aggressive behaviors, seizures, disrupted motor activity, sleep dysfunction, and sensory processing deficits (Crawley, 2007) as well as assays for gastrointestinal motility dysfunction (Roberts et al., 2007) in these models. Here we outline findings derived from investigations using these tests (Table 2) and highlight areas requiring further research (Table 3).

Animal models are discussed in three groups; (i) models with acquired behaviors resulting from environmental insult, (ii) models expressing a human genetic mutation associated with ASD,

Table 2 | Endophenotypes identified in rodent models relevant to comorbid features of ASD.

Domain	Model	Behavior
Seizure susceptibility	VPA	↑ Sensitivity to PTZ (Sobrian and Nandedkar, 1986) and electroshock-induced seizures (Kim et al., 2011)
	PPA	↑ Susceptibility to kindling with repeated intracerebroventricular infusions (MacFabe et al., 2007)
	Shank3B ^{-/-}	Occasional handling-induced seizures (Peca et al., 2011)
	CNTNAP2	Handling-induced seizures common in adults (Penagarikano et al., 2011)
	FAST	↑ Sensitivity to kindling and chemoconvulsant-induced seizures (McIntyre et al., 1999; Xu et al., 2004; Gilby et al., 2005)
	EL	Handling-induced seizures (Todorova et al., 1999)
	BALB/c	↑ Audiogenic seizures (Morin et al., 1994; Banko et al., 1997)
	C58/J	↑ Sensitivity to PTZ-induced seizures (Nutt and Lister, 1988)
Aggression	Shank2 ^{-/-}	↑ Aggression in home cages although no change in resident-intruder test (Schmeisser et al., 2012)
	FAST	↑ (Reinhart et al., 2004)
	BALB/c	↑ (Brodkin, 2007; Velez et al., 2010)
Anxiety-like behavior	VPA	↑ (Mice) (Markram et al., 2008)
	Shank3B ^{-/-}	↑ (Peca et al., 2011)
	Shank2 ^{-/-}	↑ (Schmeisser et al., 2012; Won et al., 2012)
	FAST	↑ Fear-potentiated startle (Anisman et al., 2000)
	BALB/c	↑ (Brodkin, 2007)
	BTBR	↑ Under some conditions (McFarlane et al., 2008; Pobbe et al., 2011)
Gastro-intestinal disturbances	BALB/c	Altered intestinal motility compared to C57BL/6 mice in response to serotonin antagonists (Neal et al., 2009)
Sensory	VPA	↓ PPI, ↑ tactile sensitivity (Schneider and Przewlocki, 2005), ↓ olfactory (Schneider and Przewlocki, 2005; Rouillet et al., 2010) and pain (Markram et al., 2008) sensitivity
	PPA	↓ Sensorimotor function (increased tendency to slip/fall during beam task; Shultz et al., 2009)
	NL3 ^{R451C}	↓ Acoustic startle at high decibel levels (Chadman et al., 2008)
	Shank3B ^{-/-}	↓ PPI (Peca et al., 2011)
	CNTNAP2	↑ Pain and olfactory sensitivity (Penagarikano et al., 2011)
	FAST	↓ Acoustic startle (Anisman et al., 2000)
	BTBR	↓ Thermal response (Silverman et al., 2010b)
	Motor	NL3 ^{R451C}
Shank3 ^{e4-9}		Mild motor impairments (Wang et al., 2011b)
CNTNAP2		Slight ↑ motor coordination (↑ latency to fall from rotarod Penagarikano et al., 2011)
EL		Delays in visuomotor development (McFadyen-Leussis and Heinrichs, 2005)
Sleep	VPA	Abnormal circadian rhythms (Tsujino et al., 2007)

Endophenotypes relevant to enhanced seizure susceptibility, altered sensory function, and anxiety-like behavior were observed across environmental, monogenetic, and phenotype first models. However, each model was assessed for only a subset of the endophenotypes listed and further research is required to clarify full endophenotypic profiles. VPA, rodents administered valproate.

and (iii) naturally occurring rodent strains that demonstrate behavioral endophenotypes relevant to ASD.

ENVIRONMENTAL MODELS

Autism spectrum disorder-like features exhibited by environmental rodent models are generally elicited in response to an overt insult or developmental challenge, such as exposure to toxins resulting in altered neurological development.

Valproate models

During pregnancy, maternal exposure to the first generation antiepileptic drug valproate has been shown to significantly

increase the risk of ASD in children (Rasalam et al., 2005; Meador et al., 2006; Bromley et al., 2008). Valproate is a short-chain fatty acid and is thought to reduce neuronal excitability primarily by increasing concentrations of the inhibitory neurotransmitter GABA and modulating voltage-gated sodium channels (Chapman et al., 1982; Rogawski and Loscher, 2004). In both mice and rats, exposure to valproate during gestation via intraperitoneal injection or orally with food produces deficits in social interaction and repetitive behaviors (Schneider and Przewlocki, 2005; Wagner et al., 2006; Rouillet et al., 2010; Kim et al., 2011). These animals also show reduced sensitivity to pain (Markram et al., 2008) and olfactory cues (Schneider and

Table 3 | An overview of endophenotypes assayed in rodent models of ASD.

	Seizure susceptibility	Aggression	Anxiety	Gastrointestinal	Sensory	Motor coordination	Sleep
VPA	↑		↑				
PPA	↑				↓	↓	
NL3 ^{R451C}					↓	↑	
Shank2		↑	↑				
Shank3	↑		↑		↓	↓	
CNTNAP2	↑				↑	↑	
EL	↑					↓	
C58/J	↑						
BALB/c	↑	↑	↑				
BTBR					↓		
FAST	↑	↑	↑		↓		

Increase

Decrease

No change

Not tested

Mixed/complex

Seizure susceptibility, sensory function, motor coordination, and anxiety-like behaviors are most commonly tested across models. Aggressive behavior, gastrointestinal function, and sleep cycles are generally understudied. Dual colored cells: formal aggression testing in *Shank2*^{-/-} mice did not yield data suggesting abnormal aggressive behavior, however excessive aggression was observed in home cages; PPA rats had impaired sensorimotor abilities when tested using the beam task but showed no change in swim speed in other assays. VPA, rodents administered valproate.

Przewlocki, 2005; Roulet et al., 2010), increased tactile sensitivity (Schneider and Przewlocki, 2005), and diminished acoustic pre-pulse inhibition, a test commonly used to index abnormalities in sensorimotor gating (Schneider and Przewlocki, 2005; Markram et al., 2008; Gandal et al., 2010; Roulet et al., 2010). Valproate-exposed adult rats show increased levels of anxiety-like behaviors (Markram et al., 2008) and a reduced threshold for electroshock (Kim et al., 2011) and pentylenetetrazole (PTZ)-induced seizures (Sobrian and Nandedkar, 1986). These rats also show altered circadian rhythms characterized by frequent arousal during the light/sleep phase (Tsujino et al., 2007; Tables 2 and 3).

Propionic acid model

The gut microbiota have been suggested to play a role in the etiology of ASD (Mulle et al., 2013). Potential mechanisms contributing to ASD phenotypes are unknown, however excess toxin-producing bacteria have been identified in patients with ASD (Parracho et al., 2005) and increased levels of short-chain fatty acids (such as propionic acid; PPA) produced by enteric bacteria have been studied in rats (MacFabe et al., 2007). In rodent models, administration of the endogenous short-chain fatty acids butyric acid (Thomas et al., 2010), sodium acetate (Shultz et al., 2008, 2009), and PPA directly into the cerebral ventricles produces endophenotypes relevant to ASD (MacFabe et al., 2007, 2011; Shultz et al., 2008, 2009; Thomas et al., 2010). Acute intracerebral ventricular infusion of PPA in rats reduces sociability and learning and also produces sensorimotor impairments (Shultz et al., 2009). This paradigm also results in reduced cognitive flexibility during reversal learning (MacFabe et al., 2011). Furthermore, repeated intraventricular PPA infusion leads to increased susceptibility to kindling-induced seizures and stereotypic behavior (MacFabe et al., 2007, 2011; Shultz et al., 2009; Tables 2 and 3).

A small number of ASD patients (5%) show mitochondrial dysfunction along with altered levels of various metabolites suggestive of altered fatty acid processing (Frye et al., 2013). Further investigation to assess the effects of both PPA and valproate on gastrointestinal function (i.e., following oral administration) is needed (see Table 3), as the short-chain fatty acid receptor (GPR43) expressed by some mucosal enteroendocrine cells may play a role (Karakci et al., 2006). The effects of orally administered PPA in particular would be of interest and would serve to strengthen construct validity of this model.

GENETIC MODELS

Many gene mutations associated with ASD code for proteins involved in the formation and maintenance of synapses (Sudhof, 2008; Betancur et al., 2009; Bourgeron, 2009; Chakrabarti et al., 2009; Betancur, 2011; Geschwind, 2011). Here we review findings from monogenic mouse models expressing mutations in four genes modulating synaptic function; the neuroligin-3^{R451C} (NL3^{R451C}) mice (Tabuchi et al., 2007; Chadman et al., 2008) two models expressing specific mutations in the Shank3B/ProSAP2 gene [Shank3B knockout mice and Shank3B^{e4-9} partial knockout mice (Peca et al., 2011; Wang et al., 2011b)], as well as two SHANK2 knockout models (Schmeisser et al., 2012; Won et al., 2012) and the contactin associated protein-like 2/Neurexin IV (CNTNAP2/NRXN4; Penagarikano et al., 2011) knockout mouse model (Table 2). Electrophysiological studies in these mice report altered glutamatergic and GABAergic synaptic function (Tabuchi et al., 2007; Etherton et al., 2009, 2011; Peca et al., 2011; Wang et al., 2011b; Schmeisser et al., 2012; Won et al., 2012). Each of these models also expresses strong ASD behavioral endophenotypes suggesting a role for these genes in shaping core behaviors relevant to ASD diagnosis. However, it is not well established whether these animal models replicate comorbid traits observed in patients.

Neurologin-3^{R451C} mice

Neurologins are adhesion molecules which interact with a range of post-synaptic scaffolding proteins including Shank3 and CNTNAP2 and bind to members of the presynaptic neuroligin family across the synaptic cleft (Sudhof, 2008; Krueger et al., 2012; Verpelli and Sala, 2012). Mutations in the neurologin family of post-synaptic adhesion molecules were implicated in ASD after a spontaneous point mutation in the gene encoding NL3 was identified in two brothers with ASD; one with comorbid epilepsy (Jamain et al., 2003). Mice expressing the NL3^{R451C} mutation show a subtle reduction in pup distress calls (on post-natal day 8) and reduced acoustic startle (Chadman et al., 2008). Under some conditions and on some genetic backgrounds, NL3^{R451C} mice also show impaired social interaction (Tabuchi et al., 2007; Etherton et al., 2011). Delays in meeting developmental milestones (e.g., slower righting reflexes), which may appear as motor deficits early in development, have also been observed in these mice (Chadman et al., 2008). However, adult NL3^{R451C} mice showed better motor coordination in the accelerating rotarod test compared with wild type littermates (Chadman et al., 2008).

Shank3-related models

The Shank (SH3 and multiple ankyrin repeat domains) gene family (also known as Proline-rich synapse-associated proteins; ProSAPs) contains three members; Shank1-3 that code for post-synaptic scaffolding proteins involved in the recruitment of several receptors and proteins (including the neurologins and neuroligins) to the excitatory post-synaptic membrane (Irie et al., 1997; Meyer et al., 2004; Baron et al., 2006; Hayashi et al., 2009; Arons et al., 2012). Rare microdeletions within the 22q13 locus (containing Shank3) are associated with intellectual disability, speech delay, and ASD (Nesslinger et al., 1994; Bonaglia et al., 2006; Durand et al., 2007). Mutations in Shank2 are also associated with ASD (Berkel et al., 2010; Kumar, 2010). Two different genetic models in which Shank3 is altered; Shank3B^{-/-} (Peca et al., 2011) and Shank3^{e4-9} (Wang et al., 2011b) in addition to two recently reported Shank2 knockout models (Schmeisser et al., 2012; Won et al., 2012) demonstrate core and comorbid traits relevant to ASD. A third model in which one full length copy of Shank3 is deleted shows core ASD endophenotypes; however the expression of secondary/comorbid features outlined here has not been investigated in these mice (Bozdagi et al., 2010). Shank3B^{-/-} mice lacking the Shank3 α and β isoforms show increased repetitive behavior (self-injurious grooming) and reduced interaction with a stranger mouse as well as occasional handling-induced seizures (Peca et al., 2011 and reviewed in Herbert, 2011). Shank3^{e4-9} mice (in which exons 4–9 are deleted) show core ASD-like deficits including social impairments, repetitive behaviors, and altered communication (i.e., less complex vocalization patterns), with learning deficits and mild motor abnormalities also evident (Wang et al., 2011b). In addition to a role as a structural protein in the central nervous system, Shank3 is present at enteric nervous system synapses (Huett et al., 2009). The enteric nervous system controls gastrointestinal motility and mucous secretion and therefore gene mutations leading to changes in synaptic function (including many ASD candidate genes) may also affect gastrointestinal function (Gershon and Ratcliffe, 2004). The Shank3 mouse models of ASD are

therefore excellent candidates for investigating effects of ASD-associated gene mutations on gastrointestinal motility. Shank2 knockout mice demonstrate abnormal vocal and social behaviors, and increased grooming behaviors. Hyperactivity (e.g., repetitive jumping) and anxiety-like behaviors have also been reported in these mice (Schmeisser et al., 2012; Won et al., 2012). Schmeisser et al. (2012) detected no change in aggressive behaviors in Shank2 knockout mice using a resident-intruder assay. Despite this negative result, a high level of aggression between Shank2 knockout males was observed in home cages (Schmeisser et al., 2012).

CNTNAP2 mice

Genetic ablation of the contactin associated protein-like 2 (CNTNAP2) gene, a member of the neuroligin transmembrane protein superfamily (also known as CASPR2 and Neuroligin IV), results in ASD-like deficits in social interaction and stereotypic behaviors in mice (Penagarikano et al., 2011). In addition, CNTNAP2 knockout mice show hyperactivity, impaired nest building, and frequent handling-induced seizures after 6 months of age (Penagarikano et al., 2011). The CNTNAP2 gene has been associated with ASD and a recessive form of epilepsy (Strauss et al., 2006). These mice exhibit sensory endophenotypes including hyper-reactivity to thermal sensory stimuli and superior performance in the buried food test, an assay for olfactory function (Penagarikano et al., 2011). CNTNAP2 knockout mice also showed slightly improved motor coordination on the rotarod compared to wild type littermates. Perhaps surprisingly, the atypical antipsychotic risperidone (prescribed to treat aggression and irritability in some cases of ASD) reversed nest building deficits as well as locomotor hyperactivity in these mice (Penagarikano et al., 2011), demonstrating predictive validity in this model (Table 2).

Behavioral analyses in transgenic mouse models of ASD confirm that a range of proteins regulating synaptic function are likely to be integral to this disorder. Most studies involving genetic models have investigated one or two endophenotypes relevant to patient core and comorbid traits (Tables 2 and 3). However, to better understand the relationship between these traits a focus on assessing the more subtle secondary endophenotypes is required. Seizure susceptibility, gastrointestinal function, sleep cycles, and aggressive behaviors remain to be investigated in the majority of these genetic models of ASD (Table 3). Still, the presence of endophenotypes relevant to comorbid traits of ASD in each of these genetic models suggests that at least some of these traits may be associated with the core behavioral features of the disorder.

PHENOTYPE FIRST MODELS

Interplay between genomic and non-genomic influences (e.g., maternal effects) is almost certainly involved in the symptom heterogeneity associated with ASD. To further understand their relative degree of contribution, animal models in which clinically relevant endophenotypes occur “naturally” are of great interest. There are currently several rodent models developed via breeding processes alone that exhibit measurable endophenotypes relevant to the diagnostic criteria and comorbid traits associated with ASD. These animal models include the FAST/SLOW rats and the C58/J, BALB/c, BtBR T + tf/J (BTBR), and epileptic-like (EL) mice (Tables 2 and 3).

FAST/SLOW rats and EL mice

The FAST and SLOW rat strains were derived from parent populations of Long Evans Hooded and Wistar rats using selective breeding processes based on relative seizure susceptibility in the amygdala kindling model (Racine et al., 1999). This process ultimately produced a seizure-prone (FAST) and seizure-resistant (SLOW) strain. FAST rats have since proven highly seizure-prone in both the kindling model and in chemoconvulsant (e.g., pilocarpine, kainate) seizure-induction models (McIntyre et al., 1999; Xu et al., 2004; Gilby et al., 2007; Gilby and O'Brien, 2013). EL mice, like FAST rats, were also created via selective breeding based on relative seizure susceptibility and originated from the non-epileptic DDY mouse strain (Meidenbauer et al., 2011). EL mice typically exhibit handling-induced seizures by postnatal day 50–60 (Todorova et al., 1999). Remarkably, the breeding processes used to create heightened seizure sensitivity in both colonies simultaneously produced robust, comorbid ASD-like traits. Both FAST rats and EL mice exhibit significant social impairment (Reinhart et al., 2004, 2006; Gilby et al., 2007; Lim et al., 2007; Turner et al., 2007) and repetitive behaviors (e.g., overgrooming, self-injurious scratching, and/or myoclonic jumping; Gilby, 2008; Meidenbauer et al., 2011) alongside delays in social, physical, and visuomotor development (McFadyen-Leussis and Heinrichs, 2005), learning deficits, impulsivity, and hyperactivity in various testing paradigms (Anisman and McIntyre, 2002; McFadyen-Leussis and Heinrichs, 2005; Azarbar et al., 2010). FAST rats are also more aggressive than their comparison (SLOW) strain (Reinhart et al., 2004, 2006) and show reduced acoustic startle but enhanced fear conditioning (Anisman et al., 2000). Thus, FAST rats and EL mice offer a similar endophenotypic profile relevant to core and comorbid symptoms observed in ASD.

C58/J mice

C58/J mice naturally exhibit ASD-like traits including poor sociability (Moy et al., 2008; Ryan et al., 2010), relative learning deficits, hyperactivity (Moy et al., 2008), and stereotypic behaviors (i.e., jumping and flipping; Ryan et al., 2010). Interestingly, C58/J mice also demonstrate a reduced threshold for PTZ-induced seizures (Nutt and Lister, 1988). However, in contrast to the ASD-like developmental delays observed in FAST and EL animals, C58/J mice meet developmental milestones earlier than their comparison strain (C57BL/6J; Ryan et al., 2010).

BALB/c and BTBR mice

The BALB/c and BTBR mouse strains exhibit core ASD traits in the form of impaired social interaction and repetitive behaviors (i.e., overgrooming and/or excessive marble burying; Brodtkin, 2007; Shoji and Kato, 2009; Pearson et al., 2011). BTBR mice also demonstrate increased social anxiety-like behavior (Pobbe et al., 2011) although anxiety responses to novel situations are inconsistent (McFarlane et al., 2008). BTBR mice are less reactive to thermal (hotplate) stimuli than the C57BL/6J standard strain (Silverman et al., 2010b), suggesting subtle sensory changes exist in this model. In addition, several BALB/c substrains displaying distinct behavioral phenotypes offer particular strengths for comorbidity investigation. BALB/cJ mice exhibit altered gastrointestinal function (Neal et al., 2009) and are highly aggressive (Velez

et al., 2010) while the epilepsy-prone (EP) BALB/c substrain is susceptible to audiogenic seizures (Morin et al., 1994; Banko et al., 1997). Notably, BTBR and BALB models have a high incidence of corpus callosal agenesis and severely reduced hippocampal commissural volumes (Wahlsten et al., 2003), which may be relevant to reports of reduced corpus callosal volumes in ASD patients (Anderson et al., 2011).

The characterization of ASD-relevant traits in these “natural” models is a relatively new initiative. Still, the documented commonalities thus far are striking; particularly the co-expression of repetitive behaviors and impaired social interaction together with heightened seizure sensitivity (**Table 3**). Finally, while we are aware that a few studies have investigated aggression and sensory processing in these rodent models, further testing using validated assays (Silverman et al., 2010a) should be applied to fully characterize the presence of core and comorbid features in these models.

SUMMARY

The primary aim of this review was to compare endophenotypic clustering within a selection of animal models of ASD. Here we focus on models expressing at least two core ASD endophenotypes with additional endophenotypes relevant to comorbid traits reported in ASD patients.

ENDOPHENOTYPING: A NEW APPROACH

We report that models generated via environmental insult, genetic manipulation, and selective breeding processes demonstrate a number of overlapping endophenotypes (**Tables 2 and 3**) relevant to both clinical comorbid (**Table 1**) and core traits of ASD. Detailed investigation into the more subtle endophenotypes associated with these models is a relatively novel approach. Indeed, many clinical traits highlighted here have yet to be investigated in these models or should be re-examined using consistent methodological approaches. Until then any ranking of the clinical relevance of the phenotypic profiles would be premature. Interestingly however, enhanced seizure susceptibility, altered sensory function, anxiety-like behaviors, and changes in motor coordination were the most frequently reported endophenotypes across models (**Table 3**). Although not routinely investigated, several of the models also showed atypical aggressive interactions (**Tables 2 and 3**). Despite evidence for disturbed sleep and abnormal gastrointestinal function in a significant number of ASD patients (see **Table 1**), to our knowledge, circadian rhythms and gastrointestinal function have only been investigated in two models; valproate-exposed rats and BALB/c mice, respectively. As discussed, gastrointestinal motility was insensitive to serotonin antagonists in BALB/c mice in comparison to a control strain (Neal et al., 2009) and valproate-treated rats showed increased arousal during sleep compared to untreated controls (Tsujino et al., 2007; **Tables 2 and 3**).

OVERLAPPING TRAITS

The presence of both core and comorbid endophenotypes in a range of animal models suggests that at least some of these traits may be interrelated and possibly integral to the etiology of ASD. Some endophenotypes are indeed co-expressed across different model constructs (for example, seizure susceptibility is consistently increased, as are anxiety-like behaviors in examples of environmental, genetic, as well as phenotype first models;

Table 3). Both environmental models (i.e., rodents administered the fatty acids valproate and PPA) and phenotype first models show heightened seizure susceptibility and anxiety-like behaviors together with sensory and motor deficits (**Table 3**). In contrast, genetic models show varied changes in sensory and motor domains (**Table 3**) for which the underlying mechanisms are unknown.

FUTURE DIRECTIONS: POTENTIAL MECHANISMS UNDERLYING ASD ENDOPHENOTYPES

Animal models are an important tool with which to tease apart the biological mechanisms underlying ASD. Given the diverse nature of ASD, it is unlikely that a single cause is responsible for this disorder and more recent research suggests some degree of interaction between the CNS and peripheral systems. Many gene mutations identified in patients with ASD affect synaptic function (Betancur et al., 2009; Bourgeron, 2009; Betancur, 2011). This supports an emerging hypothesis that ASD is primarily a disorder of neuronal communication (Grabrucker et al., 2011; Ebert and Greenberg, 2013) and we suggest that subtle changes in neural function could underlie many of the comorbid traits described here. For example, it is well established that gene mutations coding for ion channels that result in altered synaptic function in the CNS can cause seizures in patients (Helbig et al., 2008; Goldberg and Coulter, 2013). It is also important to acknowledge, however, that many neurotransmitters and receptors that regulate neuronal communication in the CNS are of functional importance in the periphery and may thereby contribute to common comorbid traits in patient subsets. For example, in the case of gastrointestinal issues, many

of the synaptic genes associated with ASD including the Shanks, neurexins, and neuroligins are also expressed in the enteric nervous system (Huett et al., 2009; Raab et al., 2010; Zhang et al., 2013), which regulates gastrointestinal motility and secretion. It is, therefore, feasible that synaptic mutations may underlie gastrointestinal symptoms in at least a subset of patients with ASD (Gershon and Ratcliffe, 2004) in addition to altering neuronal communication in the CNS. Future research should explore potential neural mechanisms underlying endophenotypes, in particular, those that are currently understudied (such as gastrointestinal disorders and altered circadian rhythms) in animal models of ASD.

In summary, rigorous endophenotyping in animal models of ASD can assist in identifying the molecular mechanisms underlying these common comorbid traits. Such information may also contribute to the identification of putative patient subsets within this spectrum of disorders and the subsequent tailoring of potential therapies. However, in order to achieve these goals, a more consistent approach in the assessment and comparison of endophenotypes is needed.

ACKNOWLEDGMENTS

The authors thank Dr. Randal Moldrich, Professor Joel Bornstein, Dr. Emma Burrows, and Professor Terence O'Brien for the critical reading of this manuscript. Elisa L. Hill-Yardin was supported by a Department of Defense (DoD) Office of Congressionally Directed Medical Research Programs (CDMRP) Autism Research Program (ARP) Idea Development Award (AR110134) and an NHMRC Grant (1047674).

REFERENCES

- Allik, H., Larsson, J. O., and Smedje, H. (2006). Sleep patterns of school-age children with Asperger syndrome or high-functioning autism. *J. Autism Dev. Disord.* 36, 585–595. doi:10.1007/s10803-006-0099-9
- Amiet, C., Gourfinkel-An, I., Bouzamon, A., Tordjman, S., Baulac, M., Lechat, P., et al. (2008). Epilepsy in autism is associated with intellectual disability and gender: evidence from a meta-analysis. *Biol. Psychiatry* 64, 577–582. doi:10.1016/j.biopsych.2008.04.030
- Anderson, J. S., Druzgal, T. J., Froehlich, A., Dubray, M. B., Lange, N., Alexander, A. L., et al. (2011). Decreased interhemispheric functional connectivity in autism. *Cereb. Cortex* 21, 1134–1146. doi:10.1093/cercor/bhq190
- Anisman, H., Kelly, O., Hayley, S., Borowski, T., Merali, Z., and McIntyre, D. C. (2000). Acoustic startle and fear-potentiated startle in rats selectively bred for fast and slow kindling rates: relation to monoamine activity. *Eur. J. Neurosci.* 12, 4405–4416. doi:10.1046/j.0953-816X.2000.01216.x
- Anisman, H., and McIntyre, D. C. (2002). Conceptual, spatial, and cue learning in the Morris water maze in fast or slow kindling rats: attention deficit comorbidity. *J. Neurosci.* 22, 7809–7817.
- Arons, M. H., Thynne, C. J., Grabrucker, A. M., Li, D., Schoen, M., Cheyne, J. E., et al. (2012). Autism-associated mutations in ProSAP2/Shank3 impair synaptic transmission and neurexin-neuroligin-mediated transsynaptic signaling. *J. Neurosci.* 32, 14966–14978. doi:10.1523/JNEUROSCI.2215-12.2012
- Azarbar, A., McIntyre, D. C., and Gilby, K. L. (2010). Caloric restriction alters seizure disposition and behavioral profiles in seizure-prone (fast) versus seizure-resistant (slow) rats. *Behav. Neurosci.* 124, 106–114. doi:10.1037/a0018307
- Banko, M. L., Allen, K. M., Dolina, S., Neumann, P. E., and Seyfried, T. N. (1997). Genomic imprinting and audiogenic seizures in mice. *Behav. Genet.* 27, 465–475. doi:10.1023/A:1025626501148
- Baron, M. K., Boeckers, T. M., Vaida, B., Faham, S., Gingery, M., Sawaya, M. R., et al. (2006). An architectural framework that may lie at the core of the postsynaptic density. *Science* 311, 531–535. doi:10.1126/science.1118995
- Bennetto, L., Kushner, E. S., and Hyman, S. L. (2007). Olfaction and taste processing in autism. *Biol. Psychiatry* 62, 1015–1021. doi:10.1016/j.biopsych.2007.04.019
- Berkel, S., Marshall, C. R., Weiss, B., Howe, J., Roeth, R., Moog, U., et al. (2010). Mutations in the SHANK2 synaptic scaffolding gene in autism spectrum disorder and mental retardation. *Nat. Genet.* 42, 489–491. doi:10.1038/ng.589
- Betancur, C. (2011). Etiological heterogeneity in autism spectrum disorders: more than 100 genetic and genomic disorders and still counting. *Brain Res.* 1380, 42–77. doi:10.1016/j.brainres.2010.11.078
- Betancur, C., Sakurai, T., and Buxbaum, J. D. (2009). The emerging role of synaptic cell-adhesion pathways in the pathogenesis of autism spectrum disorders. *Trends Neurosci.* 32, 402–412. doi:10.1016/j.tins.2009.04.003
- Blakemore, S. J., Tavassoli, T., Calo, S., Thomas, R. M., Catmur, C., Frith, U., et al. (2006). Tactile sensitivity in Asperger syndrome. *Brain Cogn.* 61, 5–13. doi:10.1016/j.bandc.2005.12.013
- Bonaglia, M. C., Giorda, R., Mani, E., Aceti, G., Anderlid, B. M., Baroncini, A., et al. (2006). Identification of a recurrent breakpoint within the SHANK3 gene in the 22q13.3 deletion syndrome. *J. Med. Genet.* 43, 822–828. doi:10.1136/jmg.2005.038604
- Bourgeron, T. (2009). A synaptic trek to autism. *Curr. Opin. Neurobiol.* 19, 231–234. doi:10.1016/j.conb.2009.06.003
- Bozdagi, O., Sakurai, T., Papapetrou, D., Wang, X., Dickstein, D. L., Takahashi, N., et al. (2010). Haploinsufficiency of the autism-associated Shank3 gene leads to deficits in synaptic function, social interaction, and social communication. *Mol. Autism* 1, 15–15. doi:10.1186/2040-2392-1-15
- Brodin, E. S. (2007). BALB/c mice: low sociability and other phenotypes that may be relevant to autism. *Behav. Brain Res.* 176, 53–65. doi:10.1016/j.bbr.2006.06.025
- Bromley, R. L., Mawer, G., Clayton-Smith, J., and Baker, G. A. (2008). Autism spectrum disorders following in utero exposure to antiepileptic drugs. *Neurology* 71, 1923–1924. doi:10.1212/01.wnl.0000339399.64213.1a

- Buie, T., Fuchs, G. J. III, Furuta, G. T., Kooros, K., Levy, J., Lewis, J. D., et al. (2010). Recommendations for evaluation and treatment of common gastrointestinal problems in children with ASDs. *Pediatrics* 125(Suppl. 1), S19–S29. doi:10.1542/peds.2009-1878D
- Cascio, C., McGlone, F., Folger, S., Tannan, V., Baranek, G., Pelphrey, K. A., et al. (2008). Tactile perception in adults with autism: a multidimensional psychophysical study. *J. Autism Dev. Disord.* 38, 127–137. doi:10.1007/s10803-007-0370-8
- Chadman, K. K., Gong, S., Scattoni, M. L., Boltuck, S. E., Gandhi, S. U., Heintz, N., et al. (2008). Minimal aberrant behavioral phenotypes of neuroligin-3 R451C knockin mice. *Autism Res.* 1, 147–158. doi:10.1002/aur.22
- Chakrabarti, B., Dudbridge, F., Kent, L., Wheelwright, S., Hill-Cawthorne, G., Allison, C., et al. (2009). Genes related to sex steroids, neural growth, and social-emotional behavior are associated with autistic traits, empathy, and Asperger syndrome. *Autism Res.* 2, 157–177. doi:10.1002/aur.80
- Chapman, A., Keane, P. E., Meldrum, B. S., Simiand, J., and Vernieres, J. C. (1982). Mechanism of anticonvulsant action of valproate. *Prog. Neurobiol.* 19, 315–359. doi:10.1016/0301-0082(82)90010-7
- Crawley, J. N. (2007). Mouse behavioral assays relevant to the symptoms of autism. *Brain Pathol.* 17, 448–459. doi:10.1111/j.1750-3639.2007.00096.x
- Dakin, S., and Frith, U. (2005). Vagaries of visual perception in autism. *Neuron* 48, 497–507. doi:10.1016/j.neuron.2005.10.018
- Dudova, I., Vodicka, J., Havlovicova, M., Sedlacek, Z., Urbanek, T., and Hrdlicka, M. (2011). Odor detection threshold, but not odor identification, is impaired in children with autism. *Eur. Child Adolesc. Psychiatry* 20, 333–340. doi:10.1007/s00787-011-0177-1
- Durand, C. M., Betancur, C., Boeckers, T. M., Bockmann, J., Chaste, P., Fauchereau, F., et al. (2007). Mutations in the gene encoding the synaptic scaffolding protein SHANK3 are associated with autism spectrum disorders. *Nat. Genet.* 39, 25–27. doi:10.1038/ng1933
- Ebert, D. H., and Greenberg, M. E. (2013). Activity-dependent neuronal signalling and autism spectrum disorder. *Nature* 493, 327–337. doi:10.1038/nature11860
- Ekinci, O., Arman, A. R., Isik, U., Bez, Y., and Berkem, M. (2010). EEG abnormalities and epilepsy in autistic spectrum disorders: clinical and familial correlates. *Epilepsy Behav.* 17, 178–182. doi:10.1016/j.yebeh.2009.11.014
- Etherton, M., Foldy, C., Sharma, M., Tabuchi, K., Liu, X., Shaloo, M., et al. (2011). Autism-linked neuroligin-3 R451C mutation differentially alters hippocampal and cortical synaptic function. *Proc. Natl. Acad. Sci. U.S.A.* 30, 2115–2129. doi:10.1073/pnas.1111093108
- Etherton, M. R., Blaiss, C. A., Powell, C. M., and Sudhof, T. C. (2009). Mouse neuroligin-1 α deletion causes correlated electrophysiological and behavioral changes consistent with cognitive impairments. *Proc. Natl. Acad. Sci. U.S.A.* 106, 17998–18003. doi:10.1073/pnas.0910297106
- Frye, R. E., Melnyk, S., and MacFabe, D. F. (2013). Unique acyl-carnitine profiles are potential biomarkers for acquired mitochondrial disease in autism spectrum disorder. *Transl. Psychiatry* 3, e220. doi:10.1038/tp.2012.143
- Gandal, M. J., Edgar, J. C., Ehrlichman, R. S., Mehta, M., Roberts, T. P., and Siegel, S. J. (2010). Validating gamma oscillations and delayed auditory responses as translational biomarkers of autism. *Biol. Psychiatry* 68, 1100–1106. doi:10.1016/j.biopsych.2010.09.031
- Gershon, M. D., and Ratcliffe, E. M. (2004). Developmental biology of the enteric nervous system: pathogenesis of Hirschsprung's disease and other congenital dysmotilities. *Semin. Pediatr. Surg.* 13, 224–235. doi:10.1053/j.sempedsurg.2004.10.019
- Geschwind, D. H. (2009). Advances in autism. *Annu. Rev. Med.* 60, 367–380. doi:10.1146/annurev.med.60.053107.121225
- Geschwind, D. H. (2011). Genetics of autism spectrum disorders. *Trends Cogn. Sci. (Regul. Ed.)* 15, 409–416. doi:10.1016/j.tics.2011.07.003
- Gilby, K. L. (2008). A new rat model for vulnerability to epilepsy and autism spectrum disorders. *Epilepsia* 49(Suppl. 8), 108–110. doi:10.1111/j.1528-1167.2008.01851.x
- Gilby, K. L., Da Silva, A. G., and McIntyre, D. C. (2005). Differential GABA(A) subunit expression following status epilepticus in seizure-prone and seizure-resistant rats: a putative mechanism for refractory drug response. *Epilepsia* 46(Suppl. 5), 3–9. doi:10.1111/j.1528-1167.2005.01001.x
- Gilby, K. L., and O'Brien, T. J. (2013). Epilepsy, autism, and neurodevelopment: kindling a shared vulnerability? *Epilepsy Behav.* 26, 370–374. doi:10.1016/j.yebeh.2012.11.002
- Gilby, K. L., Thorne, V., Patey, A., and McIntyre, D. C. (2007). Ruling out postnatal origins to attention-deficit/hyperactivity disorder (ADHD)-like behaviors in a seizure-prone rat strain. *Behav. Neurosci.* 121, 370–379. doi:10.1037/0735-7044.121.2.370
- Goldberg, E. M., and Coulter, D. A. (2013). Mechanisms of epileptogenesis: a convergence on neural circuit dysfunction. *Nat. Rev. Neurosci.* 14, 337–349. doi:10.1038/nrn3482
- Gomot, M., Bernard, F. A., Davis, M. H., Belmonte, M. K., Ashwin, C., Bullmore, E. T., et al. (2006). Change detection in children with autism: an auditory event-related fMRI study. *Neuroimage* 29, 475–484. doi:10.1016/j.neuroimage.2005.07.027
- Grabrucker, A. M., Schmeisser, M. J., Schoen, M., and Boeckers, T. M. (2011). Postsynaptic ProSAP/Shank scaffolds in the cross-hair of synaptopathies. *Trends Cell Biol.* 21, 594–603. doi:10.1016/j.tcb.2011.07.003
- Hara, H. (2007). Autism and epilepsy: a retrospective follow-up study. *Brain Dev.* 29, 486–490. doi:10.1016/j.braindev.2006.12.012
- Hayashi, M. K., Tang, C., Verpelli, C., Narayanan, R., Stearns, M. H., Xu, R. M., et al. (2009). The postsynaptic density proteins Homer and Shank form a polymeric network structure. *Cell* 137, 159–171. doi:10.1016/j.cell.2009.01.050
- Helbig, I., Scheffer, I. E., Mulley, J. C., and Berkovic, S. F. (2008). Navigating the channels and beyond: unravelling the genetics of the epilepsies. *Lancet Neurol.* 7, 231–245. doi:10.1016/S1474-4422(08)70039-5
- Herbert, M. R. (2011). SHANK3, the synapse, and autism. *N. Engl. J. Med.* 365, 173–175. doi:10.1056/NEJMibr1104261
- Huett, A., Leong, J. M., Podolsky, D. K., and Xavier, R. J. (2009). The cytoskeletal scaffold Shank3 is recruited to pathogen-induced actin rearrangements. *Exp. Cell Res.* 315, 2001–2011. doi:10.1016/j.yexcr.2009.04.003
- Ibrahim, S. H., Voigt, R. G., Katusic, S. K., Weaver, A. L., and Barbaresi, W. J. (2009). Incidence of gastrointestinal symptoms in children with autism: a population-based study. *Pediatrics* 124, 680–686. doi:10.1542/peds.2008-2933
- Irie, M., Hata, Y., Takeuchi, M., Ichtchenko, K., Toyoda, A., Hirao, K., et al. (1997). Binding of neuroligins to PSD-95. *Science* 277, 1511–1515. doi:10.1126/science.277.5331.1511
- Jamain, S., Quach, H., Betancur, C., Rastam, M., Colineaux, C., Gillberg, I. C., et al. (2003). Mutations of the X-linked genes encoding neuroligins NLGN3 and NLGN4 are associated with autism. *Nat. Genet.* 34, 27–29. doi:10.1038/ng1136
- Jeste, S. S. (2011). The neurology of autism spectrum disorders. *Curr. Opin. Neurol.* 24, 132–139. doi:10.1097/WCO.0b013e3283446450
- Kanne, S. M., and Mazurek, M. O. (2011). Aggression in children and adolescents with ASD: prevalence and risk factors. *J. Autism Dev. Disord.* 41, 926–937. doi:10.1007/s10803-010-1118-4
- Karaki, S., Mitsui, R., Hayashi, H., Kato, I., Sugiya, H., Iwanaga, T., et al. (2006). Short-chain fatty acid receptor, GPR43, is expressed by enteroendocrine cells and mucosal mast cells in rat intestine. *Cell Tissue Res.* 324, 353–360. doi:10.1007/s00441-005-0140-x
- Kim, K. C., Kim, P., Go, H. S., Choi, C. S., Yang, S. I., Cheong, J. H., et al. (2011). The critical period of valproate exposure to induce autistic symptoms in Sprague-Dawley rats. *Toxicol. Lett.* 201, 137–142. doi:10.1016/j.toxlet.2010.12.018
- Krakiwaki, P., Goodlin-Jones, B., Hertz-Picciotto, I., Croen, L. A., and Hansen, R. L. (2008). Sleep problems in children with autism spectrum disorders, developmental delays, and typical development: a population-based study. *J. Sleep Res.* 17, 197–206. doi:10.1111/j.1365-2869.2008.00650.x
- Krueger, D. D., Tuffly, L. P., Papadopoulos, T., and Brose, N. (2012). The role of neuroligins and neuroligins in the formation, maturation, and function of vertebrate synapses. *Curr. Opin. Neurobiol.* 22, 412–422. doi:10.1016/j.conb.2012.02.012
- Kumar, R. A. (2010). SHANK2 redemption: another synaptic protein for mental retardation and autism. *Clin. Genet.* 78, 519–521. doi:10.1111/j.1399-0004.2010.01530_2.x
- Kwakye, L. D., Foss-Feig, J. H., Cascio, C. J., Stone, W. L., and Wallace, M. T. (2011). Altered auditory and multisensory temporal processing in autism spectrum disorders. *Front. Integr. Neurosci.* 4:129. doi:10.3389/fnint.2010.00129

- Latham, K., Chung, S. T., Allen, P. M., Tavassoli, T., and Baron-Cohen, S. (2013). Spatial localisation in autism: evidence for differences in early cortical visual processing. *Mol. Autism* 4, 4. doi:10.1186/2040-2392-4-4
- Leekam, S. R., Nieto, C., Libby, S. J., Wing, L., and Gould, J. (2007). Describing the sensory abnormalities of children and adults with autism. *J. Autism Dev. Disord.* 37, 894–910. doi:10.1007/s10803-006-0218-7
- Lim, C. E., Turner, L. H., and Heinrichs, S. C. (2007). Short-term social recognition memory deficit and atypical social and physiological stressor reactivity in seizure-susceptible EL mice. *Seizure* 16, 59–68. doi:10.1016/j.seizure.2006.10.006
- MacFabe, D. F., Cain, D. P., Rodriguez-Capote, K., Franklin, A. E., Hoffiman, J. E., Boon, F., et al. (2007). Neurobiological effects of intraventricular propionic acid in rats: possible role of short chain fatty acids on the pathogenesis and characteristics of autism spectrum disorders. *Behav. Brain Res.* 176, 149–169. doi:10.1016/j.bbr.2006.07.025
- MacFabe, D. F., Cain, N. E., Boon, F., Ossenkopp, K. P., and Cain, D. P. (2011). Effects of the enteric bacterial metabolic product propionic acid on object-directed behavior, social behavior, cognition, and neuroinflammation in adolescent rats: relevance to autism spectrum disorder. *Behav. Brain Res.* 217, 47–54. doi:10.1016/j.bbr.2010.10.005
- Malow, B. A., Marzec, M. L., McGrew, S. G., Wang, L., Henderson, L. M., and Stone, W. L. (2006). Characterizing sleep in children with autism spectrum disorders: a multidimensional approach. *Sleep* 29, 1563–1571.
- Markram, K., Rinaldi, T., La Mendola, D., Sandi, C., and Markram, H. (2008). Abnormal fear conditioning and amygdala processing in an animal model of autism. *Neuropsychopharmacology* 33, 901–912. doi:10.1038/sj.npp.1301453
- McFadyen-Leussis, M. P., and Heinrichs, S. C. (2005). Seizure-prone EL/Suz mice exhibit physical and motor delays and heightened locomotor activity in response to novelty during development. *Epilepsy Behav.* 6, 312–319. doi:10.1016/j.yebeh.2005.01.010
- McFarlane, H. G., Kusek, G. K., Yang, M., Phoenix, J. L., Bolivar, V. J., and Crawley, J. N. (2008). Autism-like behavioral phenotypes in BTBR T+tf/J mice. *Genes Brain Behav.* 7, 152–163. doi:10.1111/j.1601-183X.2007.00330.x
- McIntyre, D. C., Kelly, M. E., and Dufresne, C. (1999). FAST and SLOW amygdala kindling rat strains: comparison of amygdala, hippocampal, piriform and perirhinal cortex kindling. *Epilepsy Res.* 35, 197–209. doi:10.1016/S0920-1211(99)00012-1
- Meador, K. J., Baker, G. A., Finnell, R. H., Kalayjian, L. A., Liporace, J. D., Loring, D. W., et al. (2006). In utero antiepileptic drug exposure: fetal death and malformations. *Neurology* 67, 407–412. doi:10.1212/01.wnl.0000227919.81208.b2
- Meidenbauer, J. J., Mantis, J. G., and Seyfried, T. N. (2011). The EL mouse: a natural model of autism and epilepsy. *Epilepsia* 52, 347–357. doi:10.1111/j.1528-1167.2010.02898.x
- Meyer, G., Varoquaux, F., Neeb, A., Oschlies, M., and Brose, N. (2004). The complexity of PDZ domain-mediated interactions at glutamatergic synapses: a case study on neuroligin. *Neuropharmacology* 47, 724–733. doi:10.1016/j.neuropharm.2004.06.023
- Morin, C. L., Dolina, S., Robertson, R. T., and Ribak, C. E. (1994). An inbred epilepsy-prone substrain of BALB/c mice shows absence of the corpus callosum, an abnormal projection to the basal forebrain, and bilateral projections to the thalamus. *Cereb. Cortex* 4, 119–128. doi:10.1093/cercor/4.2.119
- Moy, S. S., Nadler, J. J., Poe, M. D., Nonneman, R. J., Young, N. B., Koller, B. H., et al. (2008). Development of a mouse test for repetitive, restricted behaviors: relevance to autism. *Behav. Brain Res.* 188, 178–194. doi:10.1016/j.bbr.2007.10.029
- Mulle, J. G., Sharp, W. G., and Cubells, J. F. (2013). The gut microbiome: a new frontier in autism research. *Curr. Psychiatry Rep.* 15, 337. doi:10.1007/s11920-012-0337-0
- Neal, K. B., Parry, L. J., and Bornstein, J. C. (2009). Strain-specific genetics, anatomy and function of enteric neural serotonergic pathways in inbred mice. *J. Physiol. (Lond.)* 587, 567–586. doi:10.1113/jphysiol.2008.160416
- Nesslinger, N. J., Gorski, J. L., Kurczynski, T. W., Shapira, S. K., Siegel-Bartelt, J., Dumanski, J. P., et al. (1994). Clinical, cytogenetic, and molecular characterization of seven patients with deletions of chromosome 22q13.3. *Am. J. Hum. Genet.* 54, 464–472.
- Nutt, D. J., and Lister, R. G. (1988). Strain differences in response to a benzodiazepine receptor inverse agonist (FG 7142) in mice. *Psychopharmacology (Berl.)* 94, 335–336. doi:10.1007/BF00174705
- Parmeggiani, A., Barcia, G., Posar, A., Raimondi, E., Santucci, M., and Scaduto, M. C. (2010). Epilepsy and EEG paroxysmal abnormalities in autism spectrum disorders. *Brain Dev.* 32, 783–789. doi:10.1016/j.braindev.2010.07.003
- Parracho, H. M., Bingham, M. O., Gibson, G. R., and McCartney, A. L. (2005). Differences between the gut microflora of children with autistic spectrum disorders and that of healthy children. *J. Med. Microbiol.* 54, 987–991. doi:10.1099/jmm.0.46101-0
- Pearson, B. L., Pobbe, R. L., Defensor, E. B., Oasay, L., Bolivar, V. J., Blanchard, D. C., et al. (2011). Motor and cognitive stereotypies in the BTBR T+tf/J mouse model of autism. *Genes Brain Behav.* 10, 228–235. doi:10.1111/j.1601-183X.2010.00659.x
- Peca, J., Feliciano, C., Ting, J. T., Wang, W., Wells, M. F., Venkatraman, T. N., et al. (2011). Shank3 mutant mice display autistic-like behaviours and striatal dysfunction. *Nature* 472, 437–442. doi:10.1038/nature09965
- Penagarikano, O., Abrahams, B. S., Herman, E. I., Winden, K. D., Gdalyahu, A., Dong, H., et al. (2011). Absence of CNTNAP2 leads to epilepsy, neuronal migration abnormalities, and core autism-related deficits. *Cell* 147, 235–246. doi:10.1016/j.cell.2011.08.040
- Pobbe, R. L., Defensor, E. B., Pearson, B. L., Bolivar, V. J., Blanchard, D. C., and Blanchard, R. J. (2011). General and social anxiety in the BTBR T+ tf/J mouse strain. *Behav. Brain Res.* 216, 446–451. doi:10.1016/j.bbr.2010.08.039
- Pouw, L. B., Rieffe, C., Oosterveld, P., Huskens, B., and Stockmann, L. (2013). Reactive/proactive aggression and affective/cognitive empathy in children with ASD. *Res. Dev. Disabil.* 34, 1256–1266. doi:10.1016/j.ridd.2012.12.022
- Raab, M., Woelkers, T. M., and Neuhuber, W. L. (2010). Proline-rich synapse-associated protein-1 and 2 (ProSAP1/Shank2 and ProSAP2/Shank3)-scaffolding proteins are also present in postsynaptic specializations of the peripheral nervous system. *Neuroscience* 171, 421–433. doi:10.1016/j.neuroscience.2010.08.041
- Racine, R. J., Steingard, M., and McIntyre, D. C. (1999). Development of kindling-prone and kindling-resistant rats: selective breeding and electrophysiological studies. *Epilepsy Res.* 35, 183–195. doi:10.1016/S0920-1211(99)00013-3
- Rasalam, A. D., Hailey, H., Williams, J. H., Moore, S. J., Turnpenney, P. D., Lloyd, D. J., et al. (2005). Characteristics of fetal anticonvulsant syndrome associated autistic disorder. *Dev. Med. Child Neurol.* 47, 551–555. doi:10.1017/S0012162205001076
- Reinhart, C. J., McIntyre, D. C., Metz, G. A., and Pellis, S. M. (2006). Play fighting between kindling-prone (FAST) and kindling-resistant (SLOW) rats. *J. Comp. Psychol.* 120, 19–30. doi:10.1037/0735-7036.120.1.19
- Reinhart, C. J., Pellis, S. M., and McIntyre, D. C. (2004). Development of play fighting in kindling-prone (FAST) and kindling-resistant (SLOW) rats: how does the retention of phenotypic juvenility affect the complexity of play? *Dev. Psychobiol.* 45, 83–92. doi:10.1002/dev.20016
- Rinehart, N. J., Tonge, B. J., Ijasek, R., McGinley, J., Brereton, A. V., Enticott, P. G., et al. (2006). Gait function in newly diagnosed children with autism: cerebellar and basal ganglia related motor disorder. *Dev. Med. Child Neurol.* 48, 819–824. doi:10.1111/j.1469-8749.2006.tb01229.x
- Roberts, R. R., Murphy, J. F., Young, H. M., and Bornstein, J. C. (2007). Development of colonic motility in the neonatal mouse-studies using spatiotemporal maps. *Am. J. Physiol. Gastrointest. Liver Physiol.* 292, G930–G938. doi:10.1152/ajpgi.00444.2006
- Robertson, C., Kravitz, D., Freyberg, J., Baron-Cohen, S., and Baker, C. (2013). Tunnel vision: sharper gradient of spatial attention in autism. *J. Neurosci.* 33, 6676–6681. doi:10.1523/JNEUROSCI.5120-12.2013
- Rogawski, M. A., and Loscher, W. (2004). The neurobiology of antiepileptic drugs. *Nat. Rev. Neurosci.* 5, 553–564. doi:10.1038/nrn1430
- Roulet, F. I., Wollaston, L., Decatanzaro, D., and Foster, J. A. (2010). Behavioral and molecular changes in the mouse in response to prenatal exposure to the antiepileptic drug valproic acid. *Neuroscience* 170, 514–522. doi:10.1016/j.neuroscience.2010.06.069
- Ryan, B. C., Young, N. B., Crawley, J. N., Bodfish, J. W., and Moy, S.

- S. (2010). Social deficits, stereotypy and early emergence of repetitive behavior in the C58/J inbred mouse strain. *Behav. Brain Res.* 208, 178–188. doi:10.1016/j.bbr.2009.11.031
- Sansa, G., Carlson, C., Doyle, W., Weiner, H. L., Bluvstein, J., Barr, W., et al. (2011). Medically refractory epilepsy in autism. *Epilepsia* 52, 1071–1075. doi:10.1111/j.1528-1167.2011.03069.x
- Schmeisser, M. J., Ey, E., Wegener, S., Bockmann, J., Stempel, A. V., Kuebler, A., et al. (2012). Autistic-like behaviours and hyperactivity in mice lacking ProSAP1/Shank2. *Nature* 486, 256–260. doi:10.1038/nature11015
- Schneider, T., and Przewlocki, R. (2005). Behavioral alterations in rats prenatally exposed to valproic acid: animal model of autism. *Neuropsychopharmacology* 30, 80–89. doi:10.1038/sj.npp.1300518
- Shoji, H., and Kato, K. (2009). Maternal care affects the development of maternal behavior in inbred mice. *Dev. Psychobiol.* 51, 345–357. doi:10.1002/dev.20375
- Shultz, S. R., MacFabe, D. F., Martin, S., Jackson, J., Taylor, R., Boon, F., et al. (2009). Intracerebroventricular injections of the enteric bacterial metabolic product propionic acid impair cognition and sensorimotor ability in the Long-Evans rat: further development of a rodent model of autism. *Behav. Brain Res.* 200, 33–41. doi:10.1016/j.bbr.2008.12.023
- Shultz, S. R., MacFabe, D. F., Ossenkopp, K. P., Scratch, S., Whelan, J., Taylor, R., et al. (2008). Intracerebroventricular injection of propionic acid, an enteric bacterial metabolic end-product, impairs social behavior in the rat: implications for an animal model of autism. *Neuropharmacology* 54, 901–911. doi:10.1016/j.neuropharm.2008.01.013
- Silverman, J. L., Yang, M., Lord, C., and Crawley, J. N. (2010a). Behavioural phenotyping assays for mouse models of autism. *Nat. Rev. Neurosci.* 11, 490–502. doi:10.1038/nrn2851
- Silverman, J. L., Yang, M., Turner, S. M., Katz, A. M., Bell, D. B., Koenig, J. I., et al. (2010b). Low stress reactivity and neuroendocrine factors in the BTBR T+tf/J mouse model of autism. *Neuroscience* 171, 1197–1208. doi:10.1016/j.neuroscience.2010.09.059
- Sobrian, S. K., and Nandedkar, A. K. (1986). Prenatal antiepileptic drug exposure alters seizure susceptibility in rats. *Pharmacol. Biochem. Behav.* 24, 1383–1391. doi:10.1016/0091-3057(86)90199-1
- Strauss, K. A., Puffenberger, E. G., Huentelman, M. J., Gottlieb, S., Dobrin, S. E., Parod, J. M., et al. (2006). Recessive symptomatic focal epilepsy and mutant contactin-associated protein-like 2. *N. Engl. J. Med.* 354, 1370–1377. doi:10.1056/NEJMoa052773
- Sudhof, T. C. (2008). Neuroligins and neurexins link synaptic function to cognitive disease. *Nature* 455, 903–911. doi:10.1038/nature07456
- Tabuchi, K., Blundell, J., Etherton, M. R., Hammer, R. E., Liu, X., Powell, C. M., et al. (2007). A neuroligin-3 mutation implicated in autism increases inhibitory synaptic transmission in mice. *Science* 318, 71–76. doi:10.1126/science.1146221
- Thomas, R. H., Foley, K. A., Mephram, J. R., Tichenoff, L. J., Possmayer, F., and MacFabe, D. F. (2010). Altered brain phospholipid and acylcarnitine profiles in propionic acid infused rodents: further development of a potential model of autism spectrum disorders. *J. Neurochem.* 113, 515–529. doi:10.1111/j.1471-4159.2010.06614.x
- Todorova, M. T., Burwell, T. J., and Seyfried, T. N. (1999). Environmental risk factors for multifactorial epilepsy in EL mice. *Epilepsia* 40, 1697–1707. doi:10.1111/j.1528-1157.1999.tb01586.x
- Tsujino, N., Nakatani, Y., Seki, Y., Nakasato, A., Nakamura, M., Sugawara, M., et al. (2007). Abnormality of circadian rhythm accompanied by an increase in frontal cortex serotonin in animal model of autism. *Neurosci. Res.* 57, 289–295. doi:10.1016/j.neures.2006.10.018
- Turner, L. H., Lim, C. E., and Heinrichs, S. C. (2007). Antisocial and seizure susceptibility phenotypes in an animal model of epilepsy are normalized by impairment of brain corticotropin-releasing factor. *Epilepsy Behav.* 10, 8–15. doi:10.1016/j.yebeh.2006.08.013
- Valicenti-McDermott, M., McVicar, K., Rapin, I., Wershil, B. K., Cohen, H., and Shinnar, S. (2006). Frequency of gastrointestinal symptoms in children with autistic spectrum disorders and association with family history of autoimmune disease. *J. Dev. Behav. Pediatr.* 27, S128–S136. doi:10.1097/00004703-200604002-00011
- van Steensel, F. J., Bogels, S. M., and Perrin, S. (2011). Anxiety disorders in children and adolescents with autistic spectrum disorders: a meta-analysis. *Clin. Child Fam. Psychol. Rev.* 14, 302–317. doi:10.1007/s10567-011-0097-0
- Velez, L., Sokoloff, G., Miczek, K. A., Palmer, A. A., and Dulawa, S. C. (2010). Differences in aggressive behavior and DNA copy number variants between BALB/cJ and BALB/cByJ substrains. *Behav. Genet.* 40, 201–210. doi:10.1007/s10519-009-9325-5
- Verpelli, C., and Sala, C. (2012). Molecular and synaptic defects in intellectual disability syndromes. *Curr. Opin. Neurobiol.* 22, 530–536. doi:10.1016/j.conb.2011.09.007
- Wagner, G. C., Reuhl, K. R., Cheh, M., McRae, P., and Halladay, A. K. (2006). A new neurobehavioral model of autism in mice: pre- and post-natal exposure to sodium valproate. *J. Autism Dev. Disord.* 36, 779–793. doi:10.1007/s10803-006-0117-y
- Wahlsten, D., Metten, P., and Crabbe, J. C. (2003). Survey of 21 inbred mouse strains in two laboratories reveals that BTBR T+tf/tf has severely reduced hippocampal commissure and absent corpus callosum. *Brain Res.* 971, 47–54.
- Wang, L. W., Tancredi, D. J., and Thomas, D. W. (2011a). The prevalence of gastrointestinal problems in children across the United States with autism spectrum disorders from families with multiple affected members. *J. Dev. Behav. Pediatr.* 32, 351–360. doi:10.1097/DBP.0b013e31821bd06a
- Wang, X., McCoy, P. A., Rodriguez, R. M., Pan, Y., Je, H. S., Roberts, A. C., et al. (2011b). Synaptic dysfunction and abnormal behaviors in mice lacking major isoforms of Shank3. *Hum. Mol. Genet.* 20, 3093–3108. doi:10.1093/hmg/ddr212
- Won, H., Lee, H. R., Gee, H. Y., Mah, W., Kim, J. I., Lee, J., et al. (2012). Autistic-like social behaviour in Shank2-mutant mice improved by restoring NMDA receptor function. *Nature* 486, 261–265. doi:10.1038/nature11208
- Woolfenden, S., Sarkozy, V., Ridley, G., Coory, M., and Williams, K. (2012). A systematic review of two outcomes in autism spectrum disorder—epilepsy and mortality. *Dev. Med. Child Neurol.* 54, 306–312. doi:10.1111/j.1469-8749.2012.04223.x
- Xu, B., McIntyre, D. C., Fahnestock, M., and Racine, R. J. (2004). Strain differences affect the induction of status epilepticus and seizure-induced morphological changes. *Eur. J. Neurosci.* 20, 403–418. doi:10.1111/j.1460-9568.2004.03489.x
- Zhang, Q., Wang, J., Li, A., Liu, H., Zhang, W., Cui, X., et al. (2013). Expression of neurexin and neuroligin in the enteric nervous system and their down-regulated expression levels in Hirschsprung disease. *Mol. Biol. Rep.* 40, 2969–2975. doi:10.1007/s11033-012-2368-3

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 23 April 2013; paper pending published: 05 June 2013; accepted: 12 July 2013; published online: 25 July 2013.

Citation: Argyropoulos A, Gilby KL and Hill-Yardin EL (2013) Studying autism in rodent models: reconciling endophenotypes with comorbidities. *Front. Hum. Neurosci.* 7:417. doi: 10.3389/fnhum.2013.00417

Copyright © 2013 Argyropoulos, Gilby and Hill-Yardin. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in other forums, provided the original authors and source are credited and subject to any copyright notices concerning any third-party graphics etc.