#### Check for updates

#### OPEN ACCESS

EDITED BY Wei Chen, Capital Medical University, China

#### REVIEWED BY Donghu Zhou, Capital Medical University, China Zhuolun Song, Harvard Medical School, United States

\*CORRESPONDENCE Shuangyang Mo, [202200039@sr.gxmu.edu.cn](mailto:202200039@sr.gxmu.edu.cn) Heping Qin, [850802785@qq.com](mailto:850802785@qq.com)

† These authors share first authorship

RECEIVED 22 September 2024 ACCEPTED 22 November 2024 PUBLISHED 04 December 2024

#### CITATION

Zhao Y, Xu X, Cai H, Wu W, Wang Y, Huang C, Qin H and Mo S (2024) Identification of potential biomarkers from amino acid transporter in the activation of hepatic stellate cells via bioinformatics. Front. Genet. 15:1499915. doi: [10.3389/fgene.2024.1499915](https://doi.org/10.3389/fgene.2024.1499915)

#### COPYRIGHT

© 2024 Zhao, Xu, Cai, Wu, Wang, Huang, Qin and Mo. This is an open-access article distributed under the terms of the [Creative](https://creativecommons.org/licenses/by/4.0/) [Commons Attribution License \(CC BY\).](https://creativecommons.org/licenses/by/4.0/) The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# Identifi[cation of potential](https://www.frontiersin.org/articles/10.3389/fgene.2024.1499915/full) [biomarkers from amino acid](https://www.frontiersin.org/articles/10.3389/fgene.2024.1499915/full) [transporter in the activation of](https://www.frontiersin.org/articles/10.3389/fgene.2024.1499915/full) [hepatic stellate cells via](https://www.frontiersin.org/articles/10.3389/fgene.2024.1499915/full) [bioinformatics](https://www.frontiersin.org/articles/10.3389/fgene.2024.1499915/full)

Yingying Zhao<sup>1†</sup>, Xueqing Xu<sup>2†</sup>, Huaiyang Cai<sup>2†</sup>, Wenhong Wu<sup>2</sup>, Yingwei Wang<sup>2</sup>, Cheng Huang<sup>2</sup>, Heping Qin<sup>2\*</sup> and Shuangyang Mo<sup>2\*</sup>

<sup>1</sup>Shandong University of Traditional Chinese Medicine, Jinan, China, <sup>2</sup>Liuzhou People's Hospital Affiliated to Guangxi Medical University, Liuzhou, China

**Background:** The etiopathogenesis of hepatic stellate cells (HSC) activation has yet to be completely comprehended, and there has been broad concern about the interplay between amino acid transporter and cell proliferation. This study proposed exploring the molecular mechanism from amino acid transport-related genes in HSC activation by bioinformatic methods, seeking to identify the potentially crucial biomarkers.

Methods: GSE68000, the mRNA expression profile dataset of activated HSC, was applied as the training dataset, and GSE67664 as the validation dataset. Differently expressed amino acid transport-related genes (DEAATGs), GO, DO, and KEGG analyses were utilized. We applied the protein-protein interaction analysis and machine learning of LASSO and random forests to identify the target genes. Moreover, single-gene GESA was executed to investigate the potential functions of target genes via the KEGG pathway terms. Then, a ceRNA network and a druggene interaction network were constructed. Ultimately, correlation analysis was explored between target genes and collagen alpha I (COL1A), alpha-smooth muscle actin (α-SMA), and immune checkpoints.

Results: We identified 15 DEAATGs, whose enrichment analyses indicated that they were primarily enriched in the transport and metabolic process of amino acids. Moreover, two target genes (SLC7A5 and SLC1A5) were recognized from the PPI network and machine learning, confirmed through the validation dataset. Then single-gene GESA analysis revealed that SLC7A5 and SLC1A5 had a significant positive correlation to ECM−receptor interaction, cell cycle, and TGF−β signaling pathway and negative association with retinol metabolism conversely. Furthermore, the mRNA expression of target genes was closely correlated with the COL1A and α-SMA, as well as immune checkpoints. Additionally, 12 potential therapeutic drugs were in the drug-gene interaction network, and the ceRNA network was constructed and visualized.

Conclusion: SLC7A5 and SLC1A5, with their relevant molecules, could be potentially vital biomarkers for the activation of HSC.

#### KEYWORDS

liver fibrosis, hepatic stellate cells, amino acid transporter, solute carrier family 7 member 5 (SLC7A5), solute carrier family 1 member 5 (SLC1A5), bioinformatics

# Introduction

Liver fibrosis is the pathologic reaction to persistent hepatic injury triggered by various etiological factors, which can directly lead to the ultimate consequences of liver cirrhosis and end-stage liver failure, placing a threat to public health worldwide [\(Ogaly et al., 2018\)](#page-19-0). It was reported that liver fibrosis is responsible for a large number of clinical cases and deaths annually ([Luo et al., 2021\)](#page-19-1). Hepatic stellate cells (HSC), situated in the space of Disse, are essential mesenchymal cells and perform a crucial function for liver physiology. Usually, HSC are in a quiescent state and store retinol ([Yang et al., 2019](#page-19-2)). The activation of HSC is considered the main event and promoter of liver fibrosis. The migration, proliferation, and differentiation of activated HSC (aHSC) can stimulate the excessive cumulation of extracellular matrixes (ECM) with retinol loss ([Khazali et al., 2018](#page-18-0)). However, with recent advances in the awareness of HSC activation and their dominant position in liver fibrosis, the particular molecular mechanisms of regulating these processes are unclear.

Energy metabolism is the foundation of cell proliferation, biosynthesis, and physiological activity [\(Zhang et al., 2017](#page-19-3)). Rapidly proliferating cells utilize a diversity of strategies for metabolism to fulfill the energetic demands of cell growth, biosynthesis, and karyokinesis [\(Black et al., 2020\)](#page-18-1). This shift has been defined as metabolic reprogramming, which is a hallmark of various diseases [\(Rodenfels et al., 2019](#page-19-4)). In reaction to harm factors, HSC are activated, transdifferentiating from a quiescent form to a myofibroblast state characterized by proliferation, migratory, invasive capabilities, and excessive production of ECM. HSC need to exert a diversity of metabolic energy strategies to meet the extraordinary requirements for exuberant biosynthesis and proliferation ([Gu et al., 2019\)](#page-18-2). A study demonstrated that the metabolic reprogramming of glycogen, ascorbic acid, and amino acids metabolism active remodeling of the ECM, proceeding to liver fibrogenesis in reaction to persistent impairment [\(Nault et al., 2016\)](#page-19-5). Du et al. illustrated that the hepatic glutamine uptake in liver fibrosis models of mice induced with CCl4 was significantly higher compared with control mice, and suppression of glutaminolysis was proved to prevent the accumulation of fibrogenesis [\(Chen et al., 2012\)](#page-18-3). Glutamine is an essential substrate of metabolic reprogramming in many cancer cells, which can facilitate rapid proliferation and biosynthesis [\(Du et al., 2020\)](#page-18-4). Furthermore, the Hedgehog-YAP signaling pathway may correlate closely to the HSC activation through regulating glutaminolysis [\(Du et al., 2018\)](#page-18-5). Similarly, Leucine could promote the biosynthesis of collagen alpha I (COL1A) in HSC by stimulating the regulative translation mechanisms and PI3K/Akt/mTOR signaling pathways [\(Pérez de Obanos et al., 2006\)](#page-19-6). Along these lines, regulating the cellular amino metabolism of HSC may represent a potential fresh therapeutical target for liver fibrogenesis.

Amino acids are categorized into non-essential and essential types, with the latter not being synthesized from scratch. In mammals, various amino acid transporters (AAT) located in the plasma membrane or intracellular compartments, such as the Golgi apparatus, lysosomes, and mitochondria, contribute to the regulation of amino acids transmembrane transport and promote the transmembrane exchange of other substrates [\(Zhang et al., 2020](#page-19-7)). Some studies indicated that regulating the amount or activity of particular AAT may contribute to modulating the proliferation of eukaryotes; moreover, AAT are closely correlated with cancer patients' prognosis, metastasis, and survival [\(Lin](#page-19-8) [et al., 2020](#page-19-8)). The solute carrier family 7 member 5 (SLC7A5; LAT1), which especially participates in the transport of large neutral amino acids, is extraordinarily upregulated in hepatocellular carcinoma, and suppression of SLC7A5 leads to downregulates global translation in cancer cells [\(Li et al., 2013](#page-18-6)). The amino transporter SLC6A14, regulating the transmembrane uptake of amino acids, is upregulated in many human cancers characterized by a growing requirement for amino acids ([Yazawa et al., 2015](#page-19-9)). Analogously, the expression of SLC38A1 in colorectal carcinoma is closely relevant to the clinical stage of tumor node metastasis (TNM). The SLC38A1 downregulation can restrain tumor expansion and inhibit the migration of colorectal cancer cells ([Zhou et al., 2017](#page-19-10)). The alterations of metabolism that appeared in HSC activation share various customary characteristics with cancer cells ([Gauthier-Coles et al., 2021\)](#page-18-7). Thus, we hypothesized that modulation of amino acid metabolic by AAT could have a role in aHSC operation. However, our understanding of the specific relationship between AAT and the activation of HSC is limited.

In contemporary life science research, the advancement of highthroughput sequencing and microarray technologies has positioned bioinformatics as an essential tool. It is employed to analyze differentially expressed mRNA and to predict potential therapeutic targets for specific diseases. Bioinformatic analysis is an efficacious approach to discovering biomarkers of etiopathogenesis of ailments and provides an estimable foundation for further studies ([Peng S.](#page-19-11) [et al., 2022\)](#page-19-11). Consequently, this study involved an analysis of datasets from the Gene Expression Omnibus (GEO) ([Clough and Barrett,](#page-18-8) [2016\)](#page-18-8) via bioinformatic methods to determine the molecular mechanism of amino acid transport-related genes in activating HSC and recognized critical biomarkers. Furthermore, a drug-gene interaction meshwork and ceRNA network were established.

## Material and method

#### Microarray data source

[Figure 1](#page-1-0) illustrates the analysis procedure for this research. Data series GSE68000 and GSE67664 were downloaded from the GEO

<span id="page-1-0"></span>

#### <span id="page-2-0"></span>TABLE 1 Details of the GEO data.



GEO, gene expression omnibus.

database. There were 11 samples' mRNA expression profiling of GSE68000, including 3 aHSC and 3 qHSC samples. GSE67664 contained 13 samples, including 4 aHSC and 4 qHSC samples. Further information can be found in [Table 1](#page-2-0). Dataserie GSE68000 was the training set, and data series GSE67664 was the validation set.

#### Identifying differently expressed amino acid transport-related genes

Differentially expressed genes (DEGs) between the aHSC and qHSC were identified using normalized data processed with the GEO2R tool [\(Barrett et al., 2013\)](#page-18-9), applying a threshold of | log2 Fold Change|  $>1$  and  $p < 0.05$ . The GeneCards Database was used to download 33 genes involved in the amino acid transport pathway across the plasma membrane ([https://pathcards.genecards.](https://pathcards.genecards.org/Pathway/3132) [org/Pathway/3132\)](https://pathcards.genecards.org/Pathway/3132). Altogether, identical genes in DEGs and amino acid transport across the plasma membrane pathway were defined as differently expressed amino acid transport-related Genes (DEAATGs).

#### GO, DO, and KEGG enrichment analyses

Gene ontology (GO) enrichment [included biological process (BP), cellular component (CC), and molecular function (MF)] analysis and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway analysis were applied by utilizing the R clusterProfiler package ([Yu et al., 2012](#page-19-12)). The false discovery rate (FDR) was calculated via Benjamini–Hochberg (BH) adjustment. The cutoff criterion was q-value <0.05. We utilized the R DOSE package to apply the enrichment analysis of Disease Ontology (DO) terms [\(Yu](#page-19-13) [et al., 2015\)](#page-19-13). Ultimately, the significant outcomes of these enrichment analyses were visualized using the R ggplot2 and pathview packages.

#### Investigating protein-protein interaction networks and hub genes

We predicted and constructed a protein-protein interaction (PPI) network using the STRING database by uploading DEAATGs. The network was visualized and analyzed with Cytoscape 3.9.1, and hub genes were identified using the cytohubba plugin.

#### LASSO and random forest

Subsequently, machine learning techniques, including LASSO regression and the random forests (RF) algorithm, were employed to identify feature genes from DEAATGs. The LASSO regression model

was optimized by determining the optimal parameter λ through 10-fold cross-validation, configured with "family = binomial" and "measure = deviance," while all other parameters were set to their default values ([Engebretsen and Bohlin, 2019\)](#page-18-10). Concurrently, the RF method was applied to differentiate feature genes from DEAATGs utilizing the R randomforest package [\(Pavey et al., 2017](#page-19-14)). Within the RF algorithm, which has a feature selection capability, the MeanDecreaseGini value signifies the importance of a feature. Each input gene of DEAATGs was ranked by order of importance in the classification using their MeanDecreaseGini score. Genes with a MeanDecreaseGini score not equal to zero were identified as feature genes of the RF model.

#### Target genes recognition

In this study, we identified the consistently present genes within the hub and feature gene set of the two aforementioned machinelearning models as target genes.

#### Data verification of target genes

The validation set was derived from the dataset GSE67664, comprising 4 aHSC samples and 4 qHSC samples, and was utilized to verify the reliability of these target genes.

### Gene Set Enrichment Analysis and Gene Set Variation Analysis

We investigated the roles of target genes in HSC activation by conducting single-gene Gene Set Enrichment Analysis (GSEA) using the R clusterProfiler package. Each target gene's expression level was used to categorize samples into low- and high-expression groups. GSEA was then applied to identify significantly different KEGG pathways between these groups. Then a nonparametric unsupervised method of Gene Set Variation Analysis (GSVA) was performed to demonstrate the differential enrichment KEGG pathways between the two groups similarly. In this study, the R GSVA package was utilized with the gene sets of c2.cp.kegg.symbols.gmt, downloaded from the official site. A p-value of less than 0.05 was established as the threshold for statistical significance.

#### Investigation of ceRNA network of target genes

The Prospective miRNAs associating the target genes were predicted via the miRTarBase ([Huang et al., 2022](#page-18-11)), TargetScan ([McGeary et al., 2019](#page-19-15)), and Starbase [\(Li et al., 2014](#page-18-12)), aiming at exploring the mRNA–miRNA interaction of the ceRNA network. A miRNA identified simultaneously in all three databases was enrolled, and then the potential lncRNAs targeting the miRNA were recognized from the spongeScan website [\(http://spongescan.rc.u](http://spongescan.rc.ufl.edu/)fl.edu/). Ultimately, the data visualization with a Venn diagram was performed through a web-based tool [\(https://www.bioinformatics.com.cn/](https://www.bioinformatics.com.cn/)), and the ceRNA network was visualized with Cytoscape 3.9.1 software.

## Drug–gene interaction meshwork

The DrugBank database was employed to predict current or potentially related drug substances for studying the drug-gene link ([Wishart et al., 2018](#page-19-16)). Moreover, Cytoscape software was used to construct the data visualization of the drug-gene interaction network.

#### Identification of protein subcellular localization

We used the Cell-PLoc 2.0 tool ([Chou and Shen, 2008](#page-18-13)), a package from a website, to predict the subcellular localization of proteins coded by target genes.

#### Assessment of the correlation with immune checkpoints

The correlation between the target gene and vital immune checkpoints [\(Liu, 2019](#page-19-17)), such as PD1, CTLA4, LAG3, TIGIT, HAVCR2, and PDL1, was studied with Pearson's correlation coefficient. Then the website tool ([http://www.bioinformatics.](http://www.bioinformatics.com.cn/) [com.cn/\)](http://www.bioinformatics.com.cn/) was utilized for data visualization with a scatter diagram.

#### Analysis of correlation with common key biomarkers of activated HSC

α-SMA (ACTA2) and COL1A was considered the key biomarkers for activation of HSC to a fibrogenic myofibroblast. Similarly, the correlation analyses between the target genes and these key biomarkers in HSC activation were proceeded by Pearson's correlation coefficient.

#### Statistical analysis

A Student's t-test for unpaired samples was conducted to analyze the data between the two groups, and Pearson's correlation coefficient was utilized for the correlation analysis between the two variables.  $p < 0.05$  was set as the cutoff.

# Result

## Recognition of DEGs

The mRNA expression profile dataset (GSE68000) for HSC was normalized, as depicted in Figures 2A, B. Subsequently, 3,775 DEGs

were identified from the GSE68000 dataset, comprising 2,225 upregulated and 1,565 downregulated DEGs. A volcano plot and heatmap illustrating these findings are presented in [Figures 2C, D](#page-4-0), respectively.

#### Enrichment analyses of the DEGs

The GO and DO analyses of the DEGs were conducted to elucidate their potential biological functions. Within the GO BP cluster, the DEGs are predominantly involved in the regulation of extracellular matrix organization, positive regulation of cell adhesion, amino acid transport, cellular amino acid biosynthesis process, etc. [\(Figure 2E\)](#page-4-0). The majority of DEGs are found in the membrane microdomain, cell-cell junction, and basal plasma membrane within the GO CC cluster [\(Figure 2F\)](#page-4-0). The GO MF cluster shows that mainly DEGs were enriched in actin binding, GTPase regulator activities, glycosaminoglycan binding, ECM structural constituent and growth factor binding, etc. ([Figure 2G](#page-4-0)). In the DO category, the DEGs mainly participated in the malignant tumor, liver cirrhosis, pulmonary fibrosis, etc. ([Figure 2H\)](#page-4-0). Finally, the outcomes from KEGG pathway enrichment showed that the DEGs mostly participated in cancer pathways, cell adhesion molecules, PI3K-Akt signaling pathway, and cholesterol metabolism. ([Figure 2I\)](#page-4-0).

### Recognition of DEAATGs

[Figure 3A](#page-5-0) shows the genes involved in the pathway for transporting amino acids across the plasma membrane. Then we utilized an integrated bioinformatics analysis to identify a total of 15 congruent DEAATGs, containing 8 genes consistently upregulated and 7 genes congruously downregulated ([Figures 3B,](#page-5-0) [C;](#page-5-0) [Table 2\)](#page-5-1). The heatmap for DEAATGs is exhibited in [Figure 3D.](#page-5-0)

#### Function enrichment analyses of the DEAATGs

The biology functions of DEAATGs were performed in GO and DO analyses. Expectedly, the DEAATGs were mainly located in the cell membranes and promoted the transport of various amino acids across the plasma membrane ([Figures 4A, B](#page-6-0)). In the GO MF category, the DEAATGs contribute particularly to regulating the cellular metabolic activity of plentiful amino acids. [\(Figure 4C\)](#page-6-0). The R package DOSE was utilized to conduct a comprehensive investigation into the function of DEAATGs. The findings from the DO enrichment analysis indicated that DEAATGs may be involved in amino acid metabolic disorder, inherited metabolic disorder, leiomyoma, cell type benign neoplasm, etc., which were the principal diseases [\(Figure 4D](#page-6-0)). KEGG pathway enrichment analysis indicates that DEAATGs are primarily linked to protein digestion and absorption, ferroptosis, central carbon metabolism in cancer, and the mTOR signaling pathway ([Figure 4E](#page-6-0)). These findings imply that DEAATGs primarily function in regulating cell proliferation, ferroptosis, and cellular metabolism, especially amino acid transport.

<span id="page-4-0"></span>



<span id="page-5-0"></span>Identification of DEAATGs. (A) Amino acid transport-related genes; (B) The DEAATGs of GES68000; (C) The details of each DEAATGs; (D) The heatmap of DEATTGs (Clu: Cluster).

#### <span id="page-5-1"></span>TABLE 2 The DEAATGs of GSE68000.



DEAATGs, differently expressed amino acid transport-related genes.

#### PPI network and hub gene analyses

A PPI network of the DEAATGs was built using the STRING database to investigate the relationship of each protein, consisting of 10 nodes and 15 edges. In this PPI network map, each node represented a protein, and simultaneously each edge represented an association between two proteins. Additionally, within the 10 nodes, 7 nodes were upregulated, and 3 were downregulated ([Figure 5A](#page-7-0)). As displayed in [Figure 5B](#page-7-0), The genes were ranked by target connectedness from large to small within the PPI network graph. Hub genes were identified congruously through five categories (degree, betweenness, MNC, MCC, and stress) from cytoHubba Plugin of Cytoscape software. Finally, we extracted the intersection of the top 6 hub genes and screened out 5 hub genes, SLC7A5, SLC7A8, SLC1A5, SLC3A2, and SLC3A1 might have a crucial function in the PPI network [\(Figures 5C, D;](#page-7-0) [Table 3](#page-8-0)). The outputs of DO enrichment and KEGG pathway enrichment of these 5 hub genes are shown in [Figures 5E, F.](#page-7-0) They still primarily enrich in the mTOR signaling pathway, ferroptosis, and cellular metabolism of amino acids.

#### The machine learning algorithm of LASSO and random forests

Furthermore, to screen the feature genes from DEAATGs, we trained two different machine-learning algorithms of lasso and RF.

The LASSO regression is a machine-learning technique that assumes a linear relationship and incorporates an L1 regularization penalty. Initially, LASSO regression was conducted using 10-fold crossvalidation to minimize the binomial deviance, resulting in an optimized



<span id="page-6-0"></span>λ value of 0.0325. Consequently, 6 genes with non-zero regression coefficients were chosen as feature genes for DEAATGs and included in the simplified LASSO regularization model ([Figures 6A, B](#page-8-1); [Table 4\)](#page-9-0).

The RF algorithm is a method for ensemble prediction. A random forest-supervised classification algorithm was employed to identify feature genes from DEAATGs, using the R



<span id="page-7-0"></span>randomforest package to create the RF models. The significance of each gene was assessed by computing the mean decrease in the Gini index (MeanDecreaseGini). Finally, there were 14 feature genes determined by the random forest model ([Figures 6C, D](#page-8-1); [Table 4](#page-9-0)).

# Recognition of target genes and validation

We took an intersection of the three key gene sets screened by the PPI network (hub genes), lasso model, and random forest model and having two target genes, SLC7A5 and SLC1A5 [\(Figure 6E](#page-8-1)).



#### <span id="page-8-0"></span>TABLE 3 The top five hub genes.

MCC, maximal clique centrality; MNC, maximum neighborhood component.



<span id="page-8-1"></span>The target genes were initially obtained from the training dataset GSE68000 and then validated using an independent validation dataset GSE67664. The basic information of the GSE67664 dataset is displayed in [Figure 7A](#page-10-0) and [Supplementary](#page-18-14) [Figure S1](#page-18-14). We identified 2,919 DEGs total, within which 1,592 were downregulated genes and 1,327 were upregulated genes ([Figures 7B,](#page-10-0) [C\)](#page-10-0). Analysis of enrichment and signal pathways were executed for the DEGs. Similar enrichments of GO term were observed for all the clusters of BP, CC, and MF as well ([Figure 7D](#page-10-0)). Details of the significant genes that participated in amino acid transmembrane transport and amino acid transport are given in [Supplementary](#page-18-14) [Figure S2](#page-18-14). A description of DO disease enrichment and KEGG pathway enrichment is provided in [Figures 7E, F](#page-10-0). DEGs regulate liver cirrhosis, ECM−receptor interaction, and biosynthesis of

amino acids. A study conducted using GSE67664 revealed that SLC7A5 and SLC1A5 of aHSC had significantly upregulated mRNA expression compared with the qHSC, which was consistent with the above results ([Figures 8A, B\)](#page-11-0). Therefore, our outcomes indicated that SLC7A5 and SLC1A5 might be the target genes for HSC activation.

## GSEA of target genes and GSVA

Since SLC7A5 and SLC1A5 may be pivotal in the amino acid transport of HSC and participate in the activation of HSC, and simultaneously the log2FC of them were maximum in the whole hub genes, we selected SLC7A5 and SLC1A5 for farther single-gene



<span id="page-9-0"></span>TABLE 4 The feature genes of the LASSO and random forest model.

GSEA analysis separately via KEGG. The findings from the singlegene GSEA analysis aligned with the aforementioned results.

All samples were categorized into groups with high and low SLC7A5 expression according to mRNA levels. Differential expression evaluation was performed within these two groups, and a heatmap of the top 30 upregulated and downregulated differential expression genes is shown in [Figure 9A.](#page-12-0) As [Figures](#page-12-0) [9B](#page-12-0)–[L](#page-12-0) shows, besides alcoholic liver disease, chemokine signaling pathway and cell adhesion molecules, SLC7A5 were still positively related to the glycosaminoglycan biosynthesis, cell cycle, basal transcription factors, ECM−receptor interaction, TGF−β signaling pathway, aminoacyl−tRNA biosynthesis. Inversely, SLC7A5 was negatively correlated with the retinol metabolism and Drug metabolism−cytochrome P450. Moreover, the outcomes of the SLC1A5 single-gene GSEA analysis were similar to the above, as [Figures 10A](#page-13-0)–[C](#page-13-0) shows that SLC1A5 was positively associated with glycosaminoglycan biosynthesis, biosynthesis of nucleotide sugars, cell cycle, and alcoholic liver disease.

Correspondingly, the results of GSVA analysis illustrated that the activated HSC group was closely correlated with the upregulation of the mTOR signaling pathway, cell cycle, and biosynthesis. Oppositely, the activation of HSC was negatively associated with retinol metabolism ([Figure 10D\)](#page-13-0).

#### Correlation analysis with common key biomarkers of HSC activation

As illustrated in [Figures 11A](#page-14-0)–[D,](#page-14-0) the mRNA expression of both SLC7A5 and SLC1A5 were significantly positively correlated with ACTA2(α-SMA) and COL1A, indicating that SLC7A5 and SLC1A5 may associate with the activation of HSC closely.

#### Subcellular localization and relationship with immune checkpoint

The biological functions of a particular protein are mainly associated with its different subcellular localizations. The subcellular localization of SLC7A5 and SLC1A5 was plasma membrane, predicted via Cell-PLoc 2.0. This characteristic suggests that SLC7A5 and SLC1A5 may be essential for the transmembrane transport of amino acids.

In a physiological status, immune checkpoints are necessary for immunologic tolerance, to block autoimmunity, and to preserve normal tissues from immune injury ([Wang H. et al., 2019](#page-19-18)). As presented in [Figures 12A](#page-15-0)–[F](#page-15-0), there was a significant negative correlation within SLC7A5 conventional immune checkpoints containing LAG3 and CTLA4; meanwhile, a significant negative correlation between SLC1A5 and PDCD1, LAG3, and CTLA4 was

verified. Their results revealed that SLC7A5 and SLC1A5 might further occupy an essential position in the immune response.

### mRNA–miRNA–lncRNA ceRNA network

It is well-known that miRNAs are involved in regulating gene expression at the post-transcriptional level. Moreover, the biological function of Long Non-coding RNA (lncRNA) cannot be ignored, although it does not participate in encoding proteins. The lncRNA can modulate the expression of mRNA through their interactions with miRNA in the mRNA-miRNA-lncRNA ceRNA network, and disturbance of these networks may impact diseases. Decades of miRNAs predicted to target 3′-UTR of the SLC7A5 and SLC1A5 were identified by using three miRNA target-predicted databases. The interactional miRNA of lncRNA was searched and analyzed via the spongeScan database. Finally, a ceRNA network (containing 2 target genes, 16 LncRNAs, and 42 miRNAs) was identified in [Figure 13A](#page-16-0).

## Drug-gene interplay network

The DGIdb and DrugBank databases were used to explore druggene interactions and identify existing or potential pharmaceuticals. Targeting SLC7A5 and SLC1A5 could provide a specific treatment strategy. The complete drug-gene interaction network for these genes is shown in [Figure 13B](#page-16-0). In total, 12 potential therapeutic drugs were identified.

# **Discussion**

Liver fibrosis is a common hepatic disease posing a critical threat to human health. The molecular mechanisms of liver fibrosis are complex, involving multiple molecular and signal pathway mechanisms. Although recent treatments of liver fibrosis have slight improvement and effectiveness, how to more effectively prevent and reverse it remains a significant challenge worldwide as there are still great unknowns in its direct genesis. Since HSC is the most relevant mesenchymal cell contributing to liver fibrosis, most suggested antifibrotic treatments were based on the molecular mechanisms related to HSC activation [\(Malaguarnera et al., 2015\)](#page-19-19). According to AAT, there is a promising research potential in liver fibrosis that parallels that of malignant tumors. This study was conducted to identify and characterize potential biomarkers for aHSC by examining amino acid transport-related genes through bioinformatics approaches. The focus was particularly on the accumulation of ECM and myofibroblasts, to provide novel perspectives on the underlying etiopathogenesis and possible treatments for liver fibrosis.

<span id="page-10-0"></span>



<span id="page-11-0"></span>In this study, we identified 3,775 DEGs within the aHSC mRNA-expression profile, and functional enrichment analyses of the DEGs were performed. Amino acid metabolism is essential for the rapid metabolic reprogramming of classically activated HSC ([Trivedi et al., 2021\)](#page-19-20). Our study has shown that the DEGs mostly took part in the transport and biosynthesis of amino acids according to the function enrichment analyses. These results inspired us to execute in-depth research on the correlation between amino acid transport and HSC activation. Then, a total of 15 DEAATGs were identified by crossing DEGs with genes related to amino acid transport, with 8 being upregulated and 7 downregulated. and utilized for functional gene analysis. All of the DEAATGs belong to the solute carrier family (SLC). The SLC family, a superfamily located on the eukaryotic plasma membrane, regulates the uptake and outflow of multiple solutes, such as amino acids, sugars, and drugs ([Cropp et al., 2008](#page-18-15)). The SLC family has many crucial functions in eukaryote biology and correlates with cellular protein and nucleotide biosynthesis, especially those cells in high demand for substances ([Coothankandaswamy et al., 2016\)](#page-18-16). As shown in the GO cluster, the DEAATGs mostly participated in the transport and metabolic process of amino acids. The DEAATGs are mainly involved in metabolism and signaling pathways according to the KEGG pathway, likely central carbon metabolism, protein digestion, absorption, and the mTOR signaling pathway. The Wnt signaling pathway was proven to be associated with central carbon metabolism and suppression of HSC activation through mediating the biosynthesis of glutamine synthetase and reducing ammonia levels [\(Russell and Monga, 2018](#page-19-21)). Additionally, the mTOR pathway, playing a central role in cell metabolism and growth ([Li et al., 2019](#page-18-17)), is closely correlated with HSC activation ([Zhang et al., 2019](#page-19-22)). Moreover, the result of DO further corroborates the above. These outcomes of gene function analysis for DEAATGs revealed that, to some extent, the SLC family might drive adjustments of amino acid metabolism and function to promote activation of HSC.

The interaction associations between DEAATGs-encoded proteins were investigated using the PPI and its modules. Subsequently, five hub genes were identified from it, namely, SLC7A5, SLC1A5, SLC7A8, SLC3A1, and SLC3A2. While proteins are vital for biological functions and the PPI network is crucial within the body, it does not, in isolation, offer a comprehensive representation of the intricate biological regulatory network [\(He et al., 2023](#page-18-18)). To address the limitations inherent in a singular model, we utilized additional algorithms to concurrently identify feature genes. Recently, machine learning algorithms have been extensively applied in bioinformatics analyses to filter diagnostic biomarkers and construct prognostic models [\(Fabris](#page-18-19) [et al., 2018\)](#page-18-19). Consequently, the LASSO regression and RF algorithm were employed to further assess the relative importance of feature genes, thereby facilitating dimensionality reduction and feature selection. To mitigate the risk of overfitting or selection bias, LASSO regression was validated using 10-fold cross-validation, in conjunction with the RF algorithm, to identify the signature genes. Finally, SLC7A5 and SLC1A5 genes are identified as targets based on the overlap between hub and feature gene sets. The validation dataset GSE67664 confirmed the mRNA expression of SLC7A5 and SLC1A5.



<span id="page-12-0"></span>cytochrome P450.

Although these methods yielded more accurate results compared to those based solely on PPI networks, they were not without limitations. Notably, the use of a small dataset increases the risk of the model becoming either overfitted or underfitted during training. Consequently, future research employing larger sample sizes is necessary to validate these findings.

Previous research has demonstrated that the activation of HSC can be inhibited by modulating the expression of critical genes involved in metabolic reprogramming. During liver injury, aHSC exhibits increased cell proliferation, fibrogenesis, contractility, chemotaxis, and cytokine release. The activation and functionality of HSC are contingent upon metabolic alterations. Consequently, supporting the energy metabolism of HSC may serve as a potential strategy for prevention ([Bae et al., 2022](#page-18-20)). During the activation of HSC, there is an increased demand for essential amino acids, such as leucine, as well as an enhanced uptake of glutamine.

<span id="page-13-0"></span>



<span id="page-14-0"></span>The transmembrane protein SLC7A5 is responsible for facilitating the transport of essential amino acids into cells, while SLC1A5 plays a significant role in the uptake of glutamine. In addition to facilitating cell proliferation, glutamine plays a crucial role in phagocytosis, the synthesis and secretion of pro-inflammatory cytokines, and antigen presentation. Notably, macrophages overexpressing HMBOX1 exhibited a significant reduction in glutamine concentrations, accompanied by the downregulation of the glutamine transporter SLC1A5. Furthermore, the HMBOX1/ SLC1A5-mediated reduction in glutamine uptake may represent a potential mechanism underlying the protective effects of HMBOX1 in liver inflammation ([Jiang et al., 2023\)](#page-18-21).

Consequently, targeting the transport mechanisms of these amino acids presents a potential novel therapeutic strategy.

There is a crucial role for SLC7A5 in promoting cell growth and proliferation [\(Chai et al., 2022\)](#page-18-22). In addition to nutrition, stress, and energy state, mTOR also plays a key role in intracellular signaling ([Zhang et al., 2022\)](#page-19-23). SLC7A5 transports amino acids including leucine as a result of activation of the AKT/mTOR signaling pathway [\(Chen et al., 2022](#page-18-23)). SLC7A5 knockdown reduced mTOR pathway activity and suppressed the proliferation and metastasis of tumor cells [\(Li et al., 2021](#page-18-24)). Recent reports suggest that cancer cells may shift their energy source from glucose to glutamine and manifest as a glutamine-dependent phenotype via metabolic



<span id="page-15-0"></span>reprogramming ([Sikder et al., 2020\)](#page-19-24). S. Tanaka reported that downregulated SLC1A5 expression could lead to the diminished activity of mTORC1, resulting in weakened cell proliferation ([Tanaka et al., 2018](#page-19-25)). In the experimental model of hepatic fibrosis, following hepatic cell injury, activation of the mTOR pathway in mesenchymal cells enhanced the wound healing response. In summary, overactivation of mTOR within the mesenchymal compartment exacerbated liver fibrosis induced by CCl4 ([Shan et al., 2016\)](#page-19-26). AKT and mTOR, which are pivotal components of the PI3K pathway, have the potential to regulate the activation of HSC and the progression of liver fibrosis [\(Huang](#page-18-25) [et al., 2023\)](#page-18-25). Furthermore, liver fibrosis can be mitigated through AMPK phosphorylation and inhibition of mTOR-dependent signaling cascades [\(Wang et al., 2020\)](#page-19-27). In this study, we observed that the upregulation of SLC7A5 and SLC1A5 is closely linked to enhanced activity of the mTOR pathway, corroborating previous research findings. We hypothesize that the transporters SLC7A5 and SLC1A5 could serve as novel biomarkers for the initial activation of HSC by modulating the activity of the mTOR signaling pathway.

During the progression of liver fibrosis, the TGF-β signaling pathway plays a crucial role ([Peng W. et al., 2022\)](#page-19-28). Inhibition of this pathway may lead to a reduction in hepatic fibrosis ([Kundu et al., 2023\)](#page-18-26).

Various serum markers indicative of ECM components are employed to evaluate the progression of liver fibrosis, a condition marked by the excessive accumulation of ECM [\(Wu et al., 2017](#page-19-29)). In the human body, retinol is sequestered in HSCs, which are pivotal in the fibrogenic processes of the liver [\(Saki et al., 2020\)](#page-19-30). To investigate the relationship between the target genes and principal markers of liver fibrosis, we conducted a single-gene GSEA analysis. The single-gene GSEA utilizing the KEGG pathway for SLC7A5 and SLC1A5 individually revealed a noteworthy positive correlation between the upregulation of SLC7A5 and the ECM-receptor interaction, cell cycle, and TGF-β signaling pathways. Conversely, this upregulation exhibited a significant negative association with retinol metabolism. There were similar results in single-gene GSEA analysis of SLC1A5, and both SLC1A5 and SLC7A5 were closely related to alcoholic liver disease, which is known as a common genesis of liver fibrosis around the world ([Heo et al., 2019\)](#page-18-27). The pieces of evidence that the TGF-βsignaling pathway places a central position during every step of the development of HSC activation and hepatocarcinogenesis were strong. Treatments aimed at depressing the TGF-βsignaling pathway have conspicuously prevented liver fibrosis in vivo models ([Liu et al., 2019](#page-19-31)). During HSC activation, the decreased expression of retinol acyltransferase (LRAT) may lead to a reduction of retinol significantly [\(Zhang et al., 2021\)](#page-19-32), and



<span id="page-16-0"></span>genes and the rectangle means drugs.

the deficiency of retinol is known to promote fibrosis development of the liver [\(Aguilar et al., 2009\)](#page-18-28). Consequently, utilizing single-gene GSEA, we identified that the alteration of the target genes resulted in a consistent modulation of several liver fibrosis markers. This finding further substantiates the pivotal role of the target genes in the activation of hepatic stellate cells from an alternative perspective.

Additionally, due to the limited number of data samples, constructing a nomogram and ROC curve for predicting HSC activation based on the target gene was challenging, as the area under the curve (AUC) equaled 1, indicating overfitting. Consequently, we conducted scatter plot analyses of the

correlation coefficients and performed statistical tests between the target genes and conventional diagnostic markers of liver fibrosis, specifically α-SMA and COL1A. The expression patterns of the target genes were found to be completely consistent with those of the traditional markers, according to the results. Ultimately, these results show a possibly vital role in the overexpression of SLC7A5 and SLC1A5 in the activation of HSC.

The numerous intrahepatic immunocytes play an important role in maintaining hepatic homeostasis and are the essential mechanisms for many liver illnesses [\(Liang et al., 2020a\)](#page-18-29). Recently, many studies have indicated that various intrahepatic immunocyte subcategories (containing B cells, T-cells, macrophages, and neutrophils) performed a crucial position in the progression of liver inflammation and fibrosis [\(Liang et al., 2020b\)](#page-18-30). An anomalous immune cell activation could lead to a dysfunctional immune microenvironment in the liver, inducing HSC to differentiate into myofibroblast-like cells and drive liver fibrosis ([Ikeno et al., 2020\)](#page-18-31). Immune homeostasis is crucially associated with the regulation of immune checkpoints under normal physiological conditions and inhibits irregular autoimmune damage to healthy tissues [\(Liu et al.,](#page-19-33) [2020](#page-19-33)). However, cancer cells can escape immune attack via the expression of immune checkpoints ([Tu et al., 2020](#page-19-34)). Then we speculate that immune checkpoints might correlate with HSC activation and liver fibrosis and found that SLC1A5 was significantly negatively associated with CTLA4, LAG3, and PDCD1; similarly, SLC7A5 was significantly negatively associated with CTLA4 and LAG3. CTLA4 delivers an inhibitory signal to the T-cell as a T-cell transmembrane receptor, and the antagonist of CTLA4 activates the immune system and prevents T-cell exhaustion ([Welsh et al., 2019\)](#page-19-35). The proliferation, activation, and effector function of CD4<sup>+</sup> and CD8<sup>+</sup> T-cells were negatively modulated by LAG3 expression, which has also been reported to regulate autoimmunity [\(Wang J. et al., 2019](#page-19-36)). It was found in clinical studies that anti-PD-1 and anti-CTLA-4 monotherapy could lead to a higher incidence of acute liver injury, and anti-PD-L1 antibodies such as atezolizumab may promote fast progression of liver fibrosis, triggered by acute intrahepatic immunocytes infiltration of CD4<sup>+</sup> and CD8+ T-cells principally [\(Honma et al., 2021\)](#page-18-32). Jing Xu reported that SLC7A5 performed as a checkpoint for the activation of T-cells through the mTOR pathway ([Xu et al., 2020\)](#page-19-37). Taken together, we automatically hypothesized that upregulated SLC7A5 and SLC1A5 might contribute to the development of liver fibrosis by suppressing the expression of the immune checkpoint and inducing the anomalous immunocyte activation intrahepatic. Undeniably, our knowledge of this perspective has been extraordinarily limited recently, and more evidence is imminently needed to support this hypothesis in the future.

Finally, to provide drug treatment strategies for HSC activation, we further identified 12 possible therapeutic drugs targeting SLC7A5, and one of them may have a potential therapeutical effect for SLC1A5 too. Moreover, a ceRNA network was constructed to demonstrate a potentially interactive regulation in HSC activation, which will be helpful for future studies on the posttranscriptional regulatory mechanism.

Furthermore, as far as we know, this research is the first to propose that SLC7A5 and SLC1A5 might be involved in HSC activation. Nevertheless, our research is subject to certain limitations. Notably, the GEO database offers a limited number of datasets and samples on the mRNA expression profiles of activated HSC in vitro. Moreover, the cell culture model of activated HSC may not entirely correspond with in vivo data. It is noteworthy that the expression of the target genes exhibited consistent patterns across two independent datasets, thereby minimizing the potential for batch-to-batch variation and strengthening the validity of our study. Furthermore, further research involving larger sample sizes is required to substantiate the diagnostic efficacy of these target genes in the activation of HSC, and future in vivo experiments are planned to substantiate our findings. Additionally, further investigation is required to elucidate the regulatory mechanisms of SLC7A5 and SLC1A5 within the context of HSC activation.

# **Conclusions**

We identified two target genes, SLC7A5 and SLC1A5, from amino acid transport-related genes, which mainly participated in the ECM−receptor interaction, cell cycle, TGF−β signaling pathway, and retinol metabolism, and also closely correlated with the familiar biomarkers of liver fibrosis and immune checkpoints. Therefore, SLC7A5 and SLC1A5, with their relevant molecules, might be potentially vital biomarkers for HSC activation to some extent. These results would supply a novel insight into the pathogenesis and therapeutical approaches of liver fibrosis.

# Data availability statement

Publicly available datasets were analyzed in this study. This data can be found here: [https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?](https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE67664) [acc=GSE67664](https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE67664); [https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?](https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE68000) [acc=GSE68000](https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE68000).

# Author contributions

YZ: Conceptualization, Data curation, Formal Analysis, Funding acquisition, Investigation, Methodology, Project administration, Resources, Software, Supervision, Validation, Visualization, Writing–original draft, Writing–review and editing. XX: Conceptualization, Data curation, Formal Analysis, Investigation, Methodology, Resources, Software, Supervision, Validation, Writing–review and editing. HC: Data curation, Formal Analysis, Methodology, Supervision, Writing–review and editing. WW: Formal Analysis, Project administration, Validation, Writing–review and editing. YW: Conceptualization, Formal Analysis, Investigation, Project administration, Software, Writing–review and editing. CH: Resources, Validation, Visualization, Writing–review and editing. HQ: Data curation, Investigation, Methodology, Project administration, Resources, Software, Supervision, Validation, Visualization, Writing–review and editing. SM: Conceptualization, Data curation, Formal Analysis, Funding acquisition, Investigation, Methodology, Project administration, Resources, Software, Supervision, Validation, Visualization, Writing–original draft, Writing–review and editing.

# Funding

The author(s) declare that financial support was received for the research, authorship, and/or publication of this article. This study was supported by the Scientific Research Project of Liuzhou People's Hospital affiliated with Guangxi Medical University (grant numbers: lry202309 and lry202311), the National Natural Science Foundation of China (Grant number: 82000580), and the Scientific Research Project of Liuzhou Science and Technology Bureau (grant number: 2024YB0101B005).

# Acknowledgments

We thank El Taghdouini A (Liver Cell Biology Lab, Vrije Universiteit Brussel VUB, Brussels, Belgium) and Coll M (Institut d'Investigacions Biomèdiques August Pi I Sunyer, Barcelona, Spain) for microarray profiling related to this study. We appreciate the free use of GEO, Genecards databases and other online websites.

# Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## References

<span id="page-18-28"></span>Aguilar, R. P., Genta, S., Oliveros, L., Anzulovich, A., Giménez, M. S., and Sánchez, S. S. (2009). Vitamin A deficiency injures liver parenchyma and alters the expression of hepatic extracellular matrix. J. Appl. Toxicol. JAT 29, 214–222. doi:[10.1002/jat.1399](https://doi.org/10.1002/jat.1399)

<span id="page-18-20"></span>Bae, M., Kim, M. B., and Lee, J. Y. (2022). Fucoxanthin attenuates the reprogramming of energy metabolism during the activation of hepatic stellate cells. Nutrients 14, 1902. doi[:10.3390/nu14091902](https://doi.org/10.3390/nu14091902)

<span id="page-18-9"></span>Barrett, T., Wilhite, S. E., Ledoux, P., Evangelista, C., Kim, I. F., Tomashevsky, M., et al. (2013). NCBI GEO: archive for functional genomics data sets--update. Nucleic acids Res. 41, D991–D995. doi:[10.1093/nar/gks1193](https://doi.org/10.1093/nar/gks1193)

<span id="page-18-1"></span>Black, M., Arumugam, P., Shukla, S., Pradhan, A., Ustiyan, V., Milewski, D., et al. (2020). FOXM1 nuclear transcription factor translocates into mitochondria and inhibits oxidative phosphorylation. Mol. Biol. cell 31, 1411–1424. doi[:10.1091/mbc.E19-07-0413](https://doi.org/10.1091/mbc.E19-07-0413)

<span id="page-18-22"></span>Chai, S., Sheng, Y., Sun, R., He, J., Chen, L., He, F., et al. (2022). Protective effect of miR-33-5p on the M1/M2 polarization of microglia and the underlying mechanism. Bioengineered 13, 10774–10785. doi[:10.1080/21655979.2022.2061285](https://doi.org/10.1080/21655979.2022.2061285)

<span id="page-18-23"></span>Chen, M., Zhu, Y., Li, H., Zhang, Y., and Han, M. (2022). A quantitative proteomic approach explores the possible mechanisms by which the small molecule stemazole promotes the survival of human neural stem cells. Brain Sci. 12, 690. doi:[10.3390/](https://doi.org/10.3390/brainsci12060690) [brainsci12060690](https://doi.org/10.3390/brainsci12060690)

<span id="page-18-3"></span>Chen, Y., Choi, S. S., Michelotti, G. A., Chan, I. S., Swiderska-Syn, M., Karaca, G. F., et al. (2012). Hedgehog controls hepatic stellate cell fate by regulating metabolism. Gastroenterology 143, 1319–1329.e11. doi[:10.1053/j.gastro.2012.07.115](https://doi.org/10.1053/j.gastro.2012.07.115)

<span id="page-18-13"></span>Chou, K. C., and Shen, H. B. (2008). Cell-PLoc: a package of Web servers for predicting subcellular localization of proteins in various organisms. Nat. Protoc. 3, 153–162. doi:[10.1038/nprot.2007.494](https://doi.org/10.1038/nprot.2007.494)

<span id="page-18-8"></span>Clough, E., and Barrett, T. (2016). The gene expression Omnibus database. Methods Mol. Biol. Clift. N.J. 1418, 93–110. doi:[10.1007/978-1-4939-3578-9\\_5](https://doi.org/10.1007/978-1-4939-3578-9_5)

<span id="page-18-16"></span>Coothankandaswamy, V., Cao, S., Xu, Y., Prasad, P. D., Singh, P. K., Reynolds, C. P., et al. (2016). Amino acid transporter SLC6A14 is a novel and effective drug target for pancreatic cancer. Br. J. Pharmacol. 173, 3292–3306. doi:[10.1111/bph.13616](https://doi.org/10.1111/bph.13616)

<span id="page-18-15"></span>Cropp, C. D., Komori, T., Shima, J. E., Urban, T. J., Yee, S. W., More, S. S., et al. (2008). Organic anion transporter 2 (SLC22A7) is a facilitative transporter of cGMP. Mol. Pharmacol. 73, 1151–1158. doi:[10.1124/mol.107.043117](https://doi.org/10.1124/mol.107.043117)

<span id="page-18-4"></span>Du, K., Chitneni, S. K., Suzuki, A., Wang, Y., Henao, R., Hyun, J., et al. (2020). Increased glutaminolysis marks active scarring in nonalcoholic steatohepatitis progression. Cell Mol. Gastroenterol. Hepatol. 10, 1–21. doi:[10.1016/j.jcmgh.2019.12.006](https://doi.org/10.1016/j.jcmgh.2019.12.006)

<span id="page-18-5"></span>Du, K., Hyun, J., Premont, R. T., Choi, S. S., Michelotti, G. A., Swiderska-Syn, M., et al. (2018). Hedgehog-YAP signaling pathway regulates glutaminolysis to control activation<br>of hepatic stellate cells. *Gastroenterology* 154, 1465–1479.e13. doi:[10.1053/j.gastro.2017.](https://doi.org/10.1053/j.gastro.2017.12.022) [12.022](https://doi.org/10.1053/j.gastro.2017.12.022)

<span id="page-18-10"></span>Engebretsen, S., and Bohlin, J. (2019). Statistical predictions with glmnet. Clin. epigenetics 11, 123. doi:[10.1186/s13148-019-0730-1](https://doi.org/10.1186/s13148-019-0730-1)

<span id="page-18-19"></span>Fabris, F., Doherty, A., Palmer, D., de Magalhães, J. P., and Freitas, A. A. (2018). A new approach for interpreting Random Forest models and its application to the biology of ageing. Bioinforma. Oxf. Engl. 34, 2449–2456. doi:[10.1093/bioinformatics/bty087](https://doi.org/10.1093/bioinformatics/bty087)

<span id="page-18-7"></span>Gauthier-Coles, G., Vennitti, J., Zhang, Z., Comb, W. C., Xing, S., Javed, K., et al. (2021). Quantitative modelling of amino acid transport and homeostasis in mammalian cells. Nat. Commun. 12, 5282. doi:[10.1038/s41467-021-25563-x](https://doi.org/10.1038/s41467-021-25563-x)

<span id="page-18-2"></span>Gu, L., Larson Casey, J. L., Andrabi, S. A., Lee, J. H., Meza-Perez, S., Randall, T. D., et al. (2019). Mitochondrial calcium uniporter regulates PGC-1α expression to mediate

# Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

## <span id="page-18-14"></span>Supplementary material

The Supplementary Material for this article can be found online at: [https://www.frontiersin.org/articles/10.3389/fgene.2024.1499915/](https://www.frontiersin.org/articles/10.3389/fgene.2024.1499915/full#supplementary-material) [full#supplementary-material](https://www.frontiersin.org/articles/10.3389/fgene.2024.1499915/full#supplementary-material)

metabolic reprogramming in pulmonary fibrosis. Redox Biol. 26, 101307. doi:[10.1016/j.](https://doi.org/10.1016/j.redox.2019.101307) [redox.2019.101307](https://doi.org/10.1016/j.redox.2019.101307)

<span id="page-18-18"></span>He, Z., Gao, K., Dong, L., Liu, L., Qu, X., Zou, Z., et al. (2023). Drug screening and biomarker gene investigation in cancer therapy through the human transcriptional regulatory network. Comput. Struct. Biotechnol. J. 21, 1557-1572. doi[:10.1016/j.csbj.2023.02.005](https://doi.org/10.1016/j.csbj.2023.02.005)

<span id="page-18-27"></span>Heo, M. J., Kim, T. H., You, J. S., Blaya, D., Sancho-Bru, P., and Kim, S. G. (2019). Alcohol dysregulates miR-148a in hepatocytes through FoxO1, facilitating pyroptosis via TXNIP overexpression. Gut 68, 708–720. doi:[10.1136/gutjnl-2017-315123](https://doi.org/10.1136/gutjnl-2017-315123)

<span id="page-18-32"></span>Honma, Y., Shibata, M., Gohda, T., Matsumiya, H., Kumamoto, K., Miyama, A., et al. (2021). Rapid progression of liver fibrosis induced by acute liver injury due to immunerelated adverse events of atezolizumab. Intern. Med. Tokyo, Jpn. 60, 1847–1853. doi[:10.](https://doi.org/10.2169/internalmedicine.6535-20) [2169/internalmedicine.6535-20](https://doi.org/10.2169/internalmedicine.6535-20)

<span id="page-18-11"></span>Huang, H. Y., Lin, Y. C., Cui, S., Huang, Y., Tang, Y., Xu, J., et al. (2022). miRTarBase update 2022: an informative resource for experimentally validated miRNA-target interactions. Nucleic acids Res. 50 (D1), D222–d230. doi:[10.1093/nar/gkab1079](https://doi.org/10.1093/nar/gkab1079)

<span id="page-18-25"></span>Huang, L., Yu, Q., Peng, H., and Zhen, Z. (2023). The mechanism of peach kernel and safflower herb-pair for the treatment of liver fibrosis based on network pharmacology and molecular docking technology: a review. Medicine 102, e33593. doi:[10.1097/MD.](https://doi.org/10.1097/MD.0000000000033593) [0000000000033593](https://doi.org/10.1097/MD.0000000000033593)

<span id="page-18-31"></span>Ikeno, Y., Ohara, D., Takeuchi, Y., Watanabe, H., Kondoh, G., Taura, K., et al. (2020). Foxp3+ regulatory T cells inhibit CCl(4)-induced liver inflammation and fibrosis by regulating tissue cellular immunity. Front. Immunol. 11, 584048. doi:10.3389/fi[mmu.2020.584048](https://doi.org/10.3389/fimmu.2020.584048)

<span id="page-18-21"></span>Jiang, W., Jiang, Y., Zhang, X., Mu, H., Song, Y., and Zhao, H. (2023). Metabolomic analysis reveals the influence of HMBOX1 on RAW264.7 cells proliferation based on UPLC-MS/MS. BMC genomics 24, 272. doi:[10.1186/s12864-023-09361-x](https://doi.org/10.1186/s12864-023-09361-x)

<span id="page-18-0"></span>Khazali, A. S., Clark, A. M., and Wells, A. (2018). Inflammatory cytokine IL-8/ CXCL8 promotes tumour escape from hepatocyte-induced dormancy. Br. J. cancer 118, 566–576. doi:[10.1038/bjc.2017.414](https://doi.org/10.1038/bjc.2017.414)

<span id="page-18-26"></span>Kundu, A., Gali, S., Sharma, S., Kacew, S., Yoon, S., Jeong, H. G., et al. (2023). Dendropanoxide alleviates thioacetamide-induced hepatic fibrosis via inhibition of ROS production and inflammation in BALB/(C) mice. Int. J. Biol. Sci. 19, 2630-2647. doi[:10.](https://doi.org/10.7150/ijbs.80743) [7150/ijbs.80743](https://doi.org/10.7150/ijbs.80743)

<span id="page-18-17"></span>Li, J., Cheng, D., Zhu, M., Yu, H., Pan, Z., Liu, L., et al. (2019). OTUB2 stabilizes U2AF2 to promote the Warburg effect and tumorigenesis via the AKT/mTOR signaling pathway in non-small cell lung cancer. Theranostics 9, 179–195. doi[:10.7150/thno.29545](https://doi.org/10.7150/thno.29545)

<span id="page-18-6"></span>Li, J., Qiang, J., Chen, S. F., Wang, X., Fu, J., and Chen, Y. (2013). The impact of L-type amino acid transporter 1 (LAT1) in human hepatocellular carcinoma. Tumor Biol. 34, 2977–2981. doi[:10.1007/s13277-013-0861-5](https://doi.org/10.1007/s13277-013-0861-5)

<span id="page-18-12"></span>Li, J. H., Liu, S., Zhou, H., Qu, L. H., and Yang, J. H. (2014). starBase v2.0: decoding miRNA-ceRNA, miRNA-ncRNA and protein-RNA interaction networks from largescale CLIP-Seq data. Nucleic acids Res. 42, D92–D97. doi:[10.1093/nar/gkt1248](https://doi.org/10.1093/nar/gkt1248)

<span id="page-18-24"></span>Li, Y., Wang, W., Wu, X., Ling, S., Ma, Y., and Huang, P. (2021). SLC7A5 serves as a prognostic factor of breast cancer and promotes cell proliferation through activating AKT/mTORC1 signaling pathway. Ann. Transl. Med. 9, 892. doi:[10.21037/atm-21-2247](https://doi.org/10.21037/atm-21-2247)

<span id="page-18-29"></span>Liang, Q., Hu, Y., Zhang, M., Lin, C., Zhang, W., Li, Y., et al. (2020a). The T cell receptor immune repertoire protects the liver from reconsitution. Front. Immunol. 11, 584979. doi:10.3389/fi[mmu.2020.584979](https://doi.org/10.3389/fimmu.2020.584979)

<span id="page-18-30"></span>Liang, Q., Zhang, M., Hu, Y., Zhang, W., Zhu, P., Chen, Y., et al. (2020b). Gut microbiome contributes to liver fibrosis impact on T cell receptor immune repertoire. Front. Microbiol. 11, 571847. doi[:10.3389/fmicb.2020.571847](https://doi.org/10.3389/fmicb.2020.571847)

<span id="page-19-8"></span>Lin, W., Wang, C., Liu, G., Bi, C., Wang, X., Zhou, Q., et al. (2020). SLC7A11/xCT in cancer: biological functions and therapeutic implications. Am. J. cancer Res. 10, 3106–3126.

<span id="page-19-17"></span>Liu, D. (2019). Cancer biomarkers for targeted therapy. Biomark. Res. 7, 25. doi[:10.](https://doi.org/10.1186/s40364-019-0178-7) [1186/s40364-019-0178-7](https://doi.org/10.1186/s40364-019-0178-7)

<span id="page-19-31"></span>Liu, J., Kong, D., Qiu, J., Xie, Y., Lu, Z., Zhou, C., et al. (2019). Praziquantel ameliorates CCl<sub>4</sub> -induced liver fibrosis in mice by inhibiting TGF-β/Smad<br>signalling via up-regulating Smad7 in hepatic stellate cells. Br. J. Pharmacol. 176, 4666–4680. doi[:10.1111/bph.14831](https://doi.org/10.1111/bph.14831)

<span id="page-19-33"></span>Liu, Z., Cai, C., Du, J., Liu, B., Cui, L., Fan, X., et al. (2020). TCMIO: a comprehensive database of traditional Chinese medicine on immuno-oncology. Front. Pharmacol. 11, 439. doi:[10.3389/fphar.2020.00439](https://doi.org/10.3389/fphar.2020.00439)

<span id="page-19-1"></span>Luo, N., Li, J., Chen, Y., Xu, Y., Wei, Y., Lu, J., et al. (2021). Hepatic stellate cell reprogramming via exosome-mediated CRISPR/dCas9-VP64 delivery. Drug Deliv. 28, 10–18. doi:[10.1080/10717544.2020.1850917](https://doi.org/10.1080/10717544.2020.1850917)

<span id="page-19-19"></span>Malaguarnera, M., Motta, M., Vacante, M., Malaguarnera, G., Caraci, F., Nunnari, G., et al. (2015). Silybin-vitamin E-phospholipids complex reduces liver fibrosis in patients with chronic hepatitis C treated with pegylated interferon α and ribavirin. Am. J. Transl. Res. 7, 2510–2518.

<span id="page-19-15"></span>McGeary, S. E., Lin, K. S., Shi, C. Y., Pham, T. M., Bisaria, N., Kelley, G. M., et al. (2019). The biochemical basis of microRNA targeting efficacy. Science 366, eaav1741. doi[:10.1126/science.aav1741](https://doi.org/10.1126/science.aav1741)

<span id="page-19-5"></span>Nault, R., Fader, K. A., Ammendolia, D. A., Dornbos, P., Potter, D., Sharratt, B., et al. (2016). Dose-dependent metabolic reprogramming and differential gene expression in TCDD-elicited hepatic fibrosis. Toxicol. Sci. official J. Soc. Toxicol. 154, 253–266. doi[:10.](https://doi.org/10.1093/toxsci/kfw163) [1093/toxsci/kfw163](https://doi.org/10.1093/toxsci/kfw163)

<span id="page-19-0"></span>Ogaly, H. A., Eltablawy, N. A., and Abd-Elsalam, R. M. (2018). "Antifibrogenic influence of mentha piperita L. Essential oil against CCl4-induced liver fibrosis in rats," in Oxid. Med. Cell Longev. doi:[10.1155/2018/4039753](https://doi.org/10.1155/2018/4039753)

<span id="page-19-14"></span>Pavey, T. G., Gilson, N. D., Gomersall, S. R., Clark, B., and Trost, S. G. (2017). Field evaluation of a random forest activity classifier for wrist-worn accelerometer data. J. Sci. Med. sport 20, 75–80. doi:[10.1016/j.jsams.2016.06.003](https://doi.org/10.1016/j.jsams.2016.06.003)

<span id="page-19-11"></span>Peng, S., Zhang, T., Zhang, S., Tang, Q., Yan, Y., and Feng, H. (2022a). Integrated bioinformatics and validation reveal IL1B and its related molecules as potential biomarkers in chronic spontaneous urticaria. Front. Immunol. 13, 850993. doi[:10.](https://doi.org/10.3389/fimmu.2022.850993) 3389/fi[mmu.2022.850993](https://doi.org/10.3389/fimmu.2022.850993)

<span id="page-19-28"></span>Peng, W., Zhang, S., Zhou, W., Zhao, X., Wang, K., Yue, C., et al. (2022b). Layered double hydroxides-loaded sorafenib inhibit hepatic stellate cells proliferation and activation in vitro and reduce fibrosis in vivo. Front. Bioeng. Biotechnol. 10, 873971. doi[:10.3389/fbioe.2022.873971](https://doi.org/10.3389/fbioe.2022.873971)

<span id="page-19-6"></span>Pérez de Obanos, M. P., López Zabalza, M. J., Prieto, J., Herraiz, M. T., and Iraburu, M. J. (2006). Leucine stimulates procollagen alpha1(I) translation on hepatic stellate cells<br>through ERK and PI3K/Akt/mTOR activation. *J. Cell. physiology* 209, 580–586. doi[:10.](https://doi.org/10.1002/jcp.20790) [1002/jcp.20790](https://doi.org/10.1002/jcp.20790)

<span id="page-19-4"></span>Rodenfels, J., Neugebauer, K. M., and Howard, J. (2019). Heat oscillations driven by the embryonic cell cycle reveal the energetic costs of signaling. Dev. cell 48, 646–658.e6. doi[:10.1016/j.devcel.2018.12.024](https://doi.org/10.1016/j.devcel.2018.12.024)

<span id="page-19-21"></span>Russell, J. O., and Monga, S. P. (2018). Wnt/β-Catenin signaling in liver development, homeostasis, and pathobiology. Annu. Rev. pathology 13, 351–378. doi:[10.1146/](https://doi.org/10.1146/annurev-pathol-020117-044010)<br>[annurev-pathol-020117-044010](https://doi.org/10.1146/annurev-pathol-020117-044010)

<span id="page-19-30"></span>Saki, S., Saki, N., Poustchi, H., and Malekzadeh, R. (2020). Assessment of genetic aspects of non-alcoholic fatty liver and premature cardiovascular events. Middle East J. Dig. Dis. 12, 65–88. doi[:10.34172/mejdd.2020.166](https://doi.org/10.34172/mejdd.2020.166)

<span id="page-19-26"></span>Shan, L., Ding, Y., Fu, Y., Zhou, L., Dong, X., Chen, S., et al. (2016). mTOR overactivation in mesenchymal cells aggravates CCl(4)- induced liver fibrosis. Sci. Rep. 6, 36037. doi[:10.1038/srep36037](https://doi.org/10.1038/srep36037)

<span id="page-19-24"></span>Sikder, M. O. F., Sivaprakasam, S., Brown, T. P., Thangaraju, M., Bhutia, Y. D., and Ganapathy, V. (2020). SLC6A14, a Na+/Cl--coupled amino acid transporter, functions as a tumor promoter in colon and is a target for Wnt signaling. Biochem. J. 477, 1409–1425. doi[:10.1042/bcj20200099](https://doi.org/10.1042/bcj20200099)

<span id="page-19-25"></span>Tanaka, S., Pfleger, C., Lai, J. F., Roan, F., Sun, S. C., and Ziegler, S. F. (2018). KAP1 regulates regulatory T cell function and proliferation in both foxp3 dependent and -independent manners. Cell Rep. 23, 796–807. doi:[10.1016/j.](https://doi.org/10.1016/j.celrep.2018.03.099) [celrep.2018.03.099](https://doi.org/10.1016/j.celrep.2018.03.099)

<span id="page-19-20"></span>Trivedi, P., Wang, S., and Friedman, S. L. (2021). The power of plasticity-metabolic regulation of hepatic stellate cells. Cell metab. 33, 242–257. doi[:10.1016/j.cmet.2020.](https://doi.org/10.1016/j.cmet.2020.10.026) [10.026](https://doi.org/10.1016/j.cmet.2020.10.026)

<span id="page-19-34"></span>Tu, C., Zheng, Y., Zhang, H., and Wang, J. (2020). Exploration of the personalized immune checkpoint atlas of plasma cell dyscrasias patients using high-dimensional single-cell analysis. Oncol. Rep. 44, 224–240. doi[:10.3892/or.2020.7587](https://doi.org/10.3892/or.2020.7587)

<span id="page-19-18"></span>Wang, H., Mustafa, A., Liu, S., Liu, J., Lv, D., Yang, H., et al. (2019a). Immune checkpoint inhibitor toxicity in head and neck cancer: from identification to management. Front. Pharmacol. 10, 1254. doi:[10.3389/fphar.2019.01254](https://doi.org/10.3389/fphar.2019.01254)

<span id="page-19-36"></span>Wang, J., Sanmamed, M. F., Datar, I., Su, T. T., Ji, L., Sun, J., et al. (2019b). Fibrinogenlike protein 1 is a major immune inhibitory ligand of LAG-3. Cell 176, 334–347.e12. doi[:10.1016/j.cell.2018.11.010](https://doi.org/10.1016/j.cell.2018.11.010)

<span id="page-19-27"></span>Wang, Y., Li, C., Gu, J., Chen, C., Duanmu, J., Miao, J., et al. (2020). Celastrol exerts anti-inflammatory effect in liver fibrosis via activation of AMPK-SIRT3 signalling J. Cell. Mol. Med. 24, 941-953. doi[:10.1111/jcmm.14805](https://doi.org/10.1111/jcmm.14805)

<span id="page-19-35"></span>Welsh, J. W., Tang, C., de Groot, P., Naing, A., Hess, K. R., Heymach, J. V., et al. (2019). Phase II trial of ipilimumab with stereotactic radiation therapy for metastatic disease: outcomes, toxicities, and low-dose radiation-related abscopal responses. Cancer Immunol. Res. 7, 1903–1909. doi[:10.1158/2326-6066.CIR-18-0793](https://doi.org/10.1158/2326-6066.CIR-18-0793)

<span id="page-19-16"></span>Wishart, D. S., Feunang, Y. D., Guo, A. C., Lo, E. J., Marcu, A., Grant, J. R., et al. (2018). DrugBank 5.0: a major update to the DrugBank database for 2018. Nucleic acids Res. 46, D1074–d1082. doi:[10.1093/nar/gkx1037](https://doi.org/10.1093/nar/gkx1037)

<span id="page-19-29"></span>Wu, S. P., Yang, Z., Li, F. R., Liu, X. D., Chen, H. T., and Su, D. N. (2017). Smad7 overexpressing rat BMSCs inhibit the fibrosis of hepatic stellate cells by regulating the TGF-β1/Smad signaling pathway. Exp. Ther. Med. 14, 2568–2576. doi:[10.3892/etm.](https://doi.org/10.3892/etm.2017.4836) [2017.4836](https://doi.org/10.3892/etm.2017.4836)

<span id="page-19-37"></span>Xu, J., Jiang, C., Cai, Y., Guo, Y., Wang, X., Zhang, J., et al. (2020). Intervening upregulated SLC7A5 could mitigate inflammatory mediator by mTOR-P70S6K signal in rheumatoid arthritis synoviocytes. Arthritis Res. and Ther. 22, 200. doi:[10.1186/](https://doi.org/10.1186/s13075-020-02296-8) [s13075-020-02296-8](https://doi.org/10.1186/s13075-020-02296-8)

<span id="page-19-2"></span>Yang, Y. M., Noureddin, M., Liu, C., Ohashi, K., Kim, S. Y., Ramnath, D., et al. (2019). Hyaluronan synthase 2-mediated hyaluronan production mediates Notch1 activation and liver fibrosis. Sci. Transl. Med. 11, eaat9284. doi[:10.1126/scitranslmed.aat9284](https://doi.org/10.1126/scitranslmed.aat9284)

<span id="page-19-9"></span>Yazawa, T., Shimizu, K., Kaira, K., Nagashima, T., Ohtaki, Y., Atsumi, J., et al. (2015). Clinical significance of coexpression of L-type amino acid transporter 1 (LAT1) and ASC amino acid transporter 2 (ASCT2) in lung adenocarcinoma. Am. J. Transl. Res. 7, 1126–1139.

<span id="page-19-12"></span>Yu, G., Wang, L. G., Han, Y., and He, Q. Y. (2012). clusterProfiler: an R package for comparing biological themes among gene clusters. Omics a J. Integr. Biol. 16, 284–287. doi[:10.1089/omi.2011.0118](https://doi.org/10.1089/omi.2011.0118)

<span id="page-19-13"></span>Yu, G., Wang, L. G., Yan, G. R., and He, Q. Y. (2015). DOSE: an R/Bioconductor package for disease ontology semantic and enrichment analysis. Bioinforma. Oxf. Engl. 31, 608–609. doi[:10.1093/bioinformatics/btu684](https://doi.org/10.1093/bioinformatics/btu684)

<span id="page-19-23"></span>Zhang, H. L., Chen, P., Yan, H. X., Fu, G. B., Luo, F. F., Zhang, J., et al. (2022). Targeting mTORC2/HDAC3 inhibits stemness of liver cancer cells against glutamine starvation. Adv. Sci. Weinheim, Baden-Wurttemberg, Ger. 9, e2103887. doi:[10.1002/](https://doi.org/10.1002/advs.202103887) [advs.202103887](https://doi.org/10.1002/advs.202103887)

<span id="page-19-7"></span>Zhang, J., Xu, Y., Li, D., Fu, L., Zhang, X., Bao, Y., et al. (2020). Review of the correlation of LAT1 with diseases: mechanism and treatment. Front. Chem. 8, 564809. doi[:10.3389/fchem.2020.564809](https://doi.org/10.3389/fchem.2020.564809)

<span id="page-19-32"></span>Zhang, W., Conway, S. J., Liu, Y., Snider, P., Chen, H., Gao, H., et al. (2021). Heterogeneity of hepatic stellate cells in fibrogenesis of the liver: insights from single-cell transcriptomic analysis in liver injury. Cells 10, 2129. doi:[10.3390/cells10082129](https://doi.org/10.3390/cells10082129)

<span id="page-19-22"></span>Zhang, Z., Wen, H., Weng, J., Feng, L., Liu, H., Hu, X., et al. (2019). Silencing of EPCAM suppresses hepatic fibrosis and hepatic stellate cell proliferation in mice with alcoholic hepatitis via the PI3K/Akt/mTOR signaling pathway. Cell cycleGeorget. Tex. 18, 2239–2254. doi:[10.1080/15384101.2019.1642067](https://doi.org/10.1080/15384101.2019.1642067)

<span id="page-19-3"></span>Zhang, Z., Zhao, S., Yao, Z., Wang, L., Shao, J., Chen, A., et al. (2017). Autophagy regulates turnover of lipid droplets via ROS-dependent Rab25 activation in hepatic stellate cell. Redox Biol. 11, 322–334. doi:[10.1016/j.redox.2016.12.021](https://doi.org/10.1016/j.redox.2016.12.021)

<span id="page-19-10"></span>Zhou, F. F., Xie, W., Chen, S. Q., Wang, X. K., Liu, Q., Pan, X. K., et al. (2017). SLC38A1 promotes proliferation and migration of human colorectal cancer cells. J. Huazhong Univ. Sci. Technol. Yixue Yingdewen ban 37, 30–36. doi:[10.1007/](https://doi.org/10.1007/s11596-017-1690-3) [s11596-017-1690-3](https://doi.org/10.1007/s11596-017-1690-3)