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Bidirectional two-sample Mendelian randomization analysis investigates causal associations between cathepsins and inflammatory bowel disease

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Background: Cathepsins, key regulators of the pathology of gastrointestinal disorders such as inflammatory bowel disease (IBD), are a target protease that has attracted much attention in recent years. IBD is a chronic and relapsing inflammatory disorder of the gut. Traditional studies have shown a correlation between cathepsin and the risk of IBD, while the causal relationship remains unclear.

Methods: This study utilized Mendelian randomization techniques to evaluate the causal relationships between eleven cathepsins and the subtypes of IBD, such as ulcerative colitis (UC) and Crohn's disease (CD). We also performed a series of sensitivity analyses to validate the primary Mendelian randomization (MR) results, including Cochran's Q test, the MR-PRESSO global test, and the MR pleiotropy test.

Results: The forward MR analyses showed no significant association between cathepsins and IBD. Reverse Mendelian randomization analyses suggested that UC might lead to elevated cathepsin G levels [inverse-variance weighted (IVW): p = 0.038, b = 9.966], and CD might cause a decrease in cathepsin B levels [IVW: p = 0.002, b = -10.525] and cathepsin L1 levels [IVW: p = 0.045, b = -4.742].

Conclusions: Our findings offer novel and comprehensive evidence on the impact of UC or CD on cathepsins, potentially providing valuable insights into the treatment and prognosis of IBD.

KEYWORDS

IBD, cathepsins, mendelian randomization, genetics, genome-wide association studies

1 Introduction

Inflammatory bowel disease (IBD) is a chronic inflammatory condition affecting the intestine, mainly defined as either ulcerative colitis (UC) or Crohn's disease (CD) (Guan, 2019; Rogler et al., 2021; Yang et al., 2021; Nóbrega et al., 2018; GBD, 2017 Inflammatory Bowel Disease Collaborators, 2020). In recent years, IBD has emerged as a significant global public health problem due to its elevated rates of morbidity and mortality, presenting a substantial threat to human health and well-being (Mak et al., 2020; Hasan et al., 2020;

Nielsen et al., 2022). Previous reports indicate that long-term IBD patients are at increased risk of gastrointestinal and extraintestinal malignancies. For instance, individuals with UC exhibit a heightened risk of developing pancreatic cancer (PC), with an odds ratio (OR) of 1.18 and a 95% confidence interval (CI) of 1.07–1.31⁹ (Huang et al., 2023). A Mendelian randomization study reported that IBD may be a risk factor in the development of oral cavity cancer (IBD: OR = 1.180, 95% CI: 1.059 to 1.316, *p* = 0.003; CD: OR = 1.112, 95% CI: 1.008 to 1.227, *p* = 0.034; UC: OR = 1.158, 95% CI: 1.041 to 1.288, *p* = 0.007) and breast cancer (IBD:OR = 1.059; 95% CI: 1.033 to 1.086; *p* < 0.0001; CD:OR = 1.029; 95% CI: 1.002 to 1.055; *p* = 0.032). Other studies have demonstrated a 17% greater overall cancer risk among IBD patients than those without the condition (Derikx et al., 2017; Singh et al., 2014; Faye et al., 2022).

Cathepsins are frequently used to catalyze the hydrolysis of unwanted proteins, play a key role in disease progression, and are important mediators of many inflammatory diseases (Reiser et al., 2010). Dysregulated cathepsins have been implicated in numerous pathologies, including arthritis, pancreatitis, and atherosclerosis (Yadati et al., 2020; Patel et al., 2018). Specific cathepsins have been identified as serological markers for certain diseases (Kyriakidi et al., 2016). In recent years, the role of cathepsins (such as cathepsin B, D, G, and L) has been widely studied in IBD (Menzel et al., 2006; Dong et al., 2022; Zhao et al., 2018). Studies reveal a significant increase in the cathepsin D protein expression in inflamed intestinal mucosa from IBD patients compared to non-inflamed mucosa (Hausmann et al., 2004). Inhibition of cathepsin D was followed by amelioration of mouse dextran sodium sulfate (DSS) colitis (Menzel et al., 2006); the level of anti-cathepsin G antibodies was significantly higher in patients with severe colitis than in those with mild or moderate colitis (p < 0.05) (Kuwana et al., 2000). Notably, Marta Dabek's study also verifies this point that cathepsin G has been found to be overexpressed in patients with UC compared to healthy controls (Dabek et al., 2009). Similar results have been found with other cathepsins, such as Zhao et al. (2018) and Zhao et al. (2016). However, our understanding of the association between other members of the cathepsin family and IBD remains limited, warranting further exploration and investigation. Meanwhile, due to the methodological constraints of observational studies, causal relationships between cathepsins and IBD cannot be conclusively established.

Genetics play a role in predisposition to IBD. The genetic risk ratio for IBD is in the range of 15-42 for CD (Monsén et al., 1991; Probert et al., 1993; Küster et al., 1989; Meucci et al., 1992; Orholm et al., 1991) and 7-17 for UC (Meucci et al., 1992; Orholm et al., 1991). Heredity as an etiological factor is stronger in Crohn's disease than in UC. Mendelian randomization (MR) analysis supplies a promising approach to exploring potential causal associations by leveraging instrumental variables (IVs) that are independent of common confounders, thus overcoming the limitations of traditional observational studies to some extent (Emdin et al., 2017; Smith and Ebrahim, 2004; Sekula et al., 2016). Recently, MR analysis has emerged as a tool for investigating relationships between IBD and various factors with the expanding size and scope of genome-wide association studies (GWAS) (Freuer et al., 2022; Zeng et al., 2023; Lund-Nielsen et al., 2018; Jones et al., 2020; Saadh et al., 2023). Long-term IBD patients have been reported to face an up to three- to five-fold increased risk of developing colitisassociated colorectal cancer. To determine whether a potential causal relationship exists between cathepsins and IBD, we conducted a bidirectional two-sample MR analysis utilizing publicly available GWAS datasets.

2 Materials and methods

2.1 Study design

MR was employed to investigate the relationships between various cathepsins and IBD. The design of our study is shown in Figure 1. Initially, we extracted genetic variants that serve as IVs for 11 cathepsins. Subsequently, we collected the summary data, including all single-nucleotide polymorphisms (SNPs) derived from GWASs of UC and CD. Next, we performed bidirectional MR analyses with five distinct MR methods, including MR-Egger, weighted median, inverse-variance weighted (IVW), simple mode, and weighted mode. Furthermore, we conducted a series of sensitivity analyses, comprising Cochran's Q test, the MR-PRESSO global test, and the MR pleiotropy test, to evaluate the heterogeneity and horizontal pleiotropy of the MR results. Because the data employed in this study were based on published studies and public databases, no additional ethical approval from an institutional review board was necessary.

2.2 Data sources

The summary-level GWAS data of 11 cathepsins (μ g/L) (including cathepsin B, D, E, F, G, H, L1, L2, O, S, and Z) and IBD (including UC and CD) were sourced from the Integrative Epidemiology Unit (IEU) open GWAS project (https://gwas.mrcieu. ac.uk/). This project, supported by the MRC IEU at the University of Bristol, collated and analyzed GWAS data from different sources, such as the UK Biobank, published articles, and the FinnGen Biobank. The approval of the Ethical Review Authority was not required because the data used in our study were public, anonymized, and de-identified. Further details regarding the exposure and outcome datasets are supplied in Table 1.

2.3 Selection of IVs

To satisfy the basic assumption of MR that IVs must be closely related to the exposure factors, SNPs that were significantly associated with various cathepsins (µg/L), including cathepsin B, D, E, F, G, H, L1, L2, O, S, and Z, at the genome-wide level ($p < 5 \times 10^{-6}$, r2 <0.001, genetic distance = 10,000 kb) were screened. The same criteria were applied for the reverse Mendelian randomization analysis related to ulcerative colitis and Crohn's disease. Subsequently, the strength of each IV was calculated by the following formula: F = R²(N-2)/1-R², in which R² represents the proportion of variability in the cathepsin explained by each IV, and N is the sample size of the GWAS for the SNP–cathepsin association. F-statistics greater than 10 were generally considered to have a strong association (Staiger and Stock, 1997; Bowden et al., 2016a).



Schematic design of a bidirectional two-sample Mendelian randomization study of cathepsins and IBD. This design assumes a bidirectional association between cathepsins and IBD. Genetic variants can influence IBD through cathepsins and *vice versa*. SNP, single-nucleotide polymorphism.

IEU GWAS id	Exposure or outcome	Identified SNPs	Participants included in the analysis	F-statistic
prot-a-718	Cathepsin B	19	3,301 individuals of European descent	34.297
ebi-a-GCST90012053	Cathepsin D	20	21,758 individuals of European descent	92.98
prot-a-720	Cathepsin E	11	3,301 individuals of European descent	21.902
prot-a-722	Cathepsin F	13	3,301 individuals of European descent	35.678
prot-a-723	Cathepsin G	15	3,301 individuals of European descent	22.807
prot-a-725	Cathepsin H	11	3,301 individuals of European descent	126.409
ebi-a-GCST90012073	Cathepsin L1	32	21,758 individuals of European descent	43.212
prot-a-728	Cathepsin L2	11	3,301 individuals of European descent	24.669
prot-a-726	Cathepsin O	13	3,301 individuals of European descent	23.267
prot-a-727	Cathepsin S	24	3,301 individuals of European descent	39.691
prot-a-729	Cathepsin Z	13	3,301 individuals of European descent	53.426
ebi-a-GCST90038684	Ulcerative colitis	46	2,515 cases and 482,083 controls	32.190
ukb-a-552	Crohn's disease	25	732 cases and 336,467 controls	26.597

TABLE 1 Information about the exposures and outcome datasets.

Abbreviation: SNPs, single-nucleotide polymorphisms; IEU, Integrative Epidemiology Unit; GWAS, Genome-wide association studies

SNPs in the MR analysis can be found in Supplementary Figure S1 and Supplementary Tables S1, S2.

2.4 Statistical analysis

First, the exposure and outcome data sets were harmonized, which ensured the reference allele (EA) remained consistently associated with the same allele across all analyses. Different MR methods (MR-Egger (Bowden et al., 2015), weighted median (Bowden et al., 2016b), IVW (Burgess et al., 2013), simple mode, and weighted mode (Hartwig et al., 2017)) were employed for the two-sample MR analysis to investigate the causal relationship between cathepsins and IBD separately. The heterogeneity of the IVW model was evaluated by Cochran's Q test, and a Q test *p*-value <0.05 indicated significant heterogeneity. Outliers identified and addressed by the MR-PRESSO method were removed immediately once detected. Then, the MR analyses were performed again. The MR-Egger method, which allows for the existence of non-zero intercepts, was employed to detect directional pleiotropy (Verbanck et al., 2018). Additionally, a leave-one-out analysis was performed to ascertain whether the removal of individual SNPs significantly impacted the results. All statistical analyses were performed using R software (version 4.3.1) with the two-sample MR package (Yavorska and Burgess, 2017).

Exposure	OR (95%CI)		P-value	Power
Cathepsin B				2.50%
MR Egger	1.000 (0.999, 1.001)		0.837	
Weighted median	1.000 (0.999, 1.001)	⊢ •	0.976	
Inverse variance weighted	1.000 (0.999, 1.000)	H	0.816	
Simple mode	1.000 (0.999, 1.001)		0.613	
Weighted mode	1.000 (0.999, 1.001)	H	0.829	
Cathepsin D				57.80%
MR Egger	1.000 (0.999, 1.001)	⊢ •−−	0.655	
Weighted median	1.001 (1.000, 1.001)	⊢ •−−1	0.322	
Inverse variance weighted	1.001 (1.000, 1.001)	ii i i i i i i i i 	0.180	
Simple mode	1.001 (0.999, 1.003)	⊢ +•−−	+ 0.241	
Weighted mode	1.000 (0.999, 1.001)	⊢ +•−−1	0.404	
Cathepsin E				2.50%
MR Egger	1.000 (0.997, 1.003) ⊢	•	0.800	
Weighted median	1.001 (0.999, 1.002)	⊢ •−−1	0.349	
Inverse variance weighted	1.000 (0.999, 1.001)		0.710	
Simple mode	1.001 (0.999, 1.002)	→	0.453	
Weighted mode	1.001 (0.999, 1.002)	⊢	0.441	
Cathepsin F				2.50%
MR Egger	0.999 (0.998, 1.001)		0.274	
Weighted median	1.000 (1.000, 1.001)	H+	0.385	
Inverse variance weighted	1.000 (0.999, 1.001)	H+	0.947	
Simple mode	1.000 (0.999, 1.001)	→	0.731	
Weighted mode	1.000 (0.999, 1.001)	H+	0.467	
athepsin G				2.50%
MR Egger	1.000 (0.998, 1.001)		0.814	
Weighted median	1.000 (0.999, 1.001)	⊢ •	0.630	
Inverse variance weighted	1.000 (0.999, 1.000)	⊢ •	0.364	
Simple mode	0.999 (0.998, 1.001)	→ →	0.422	
Weighted mode	1.000 (0.998, 1.001)		0.679	
Cathepsin H				2.50%
MR Egger	1.000 (0.999, 1.001)	H++	0.904	
Weighted median	1.000 (1.000, 1.000)	H.	0.725	
Inverse variance weighted	1.000 (1.000, 1.000)	H H	0.813	
Simple mode	1.000 (0.999, 1.001)		0.899	
Weighted mode	1.000 (1.000, 1.000)	H	0.668	
Cathepsin L1				2.50%
MR Egger	1.000 (0.998, 1.003)		0.751	
Weighted median	1.000 (0.999, 1.001)	→	0.686	
Inverse variance weighted	1.000 (0.999, 1.001)	⊢ ∔•−−1	0.355	
Simple mode	1.001 (0.999, 1.003)	· · ·	⊣ 0.365	
Weighted mode	1.000 (0.999, 1.002)	→	0.669	
athepsin L2				2.50%
MR Egger	1.000 (0.999, 1.002)	→	0.725	
Weighted median	1.000 (0.999, 1.001)		0.987	
Inverse variance weighted	1.000 (0.999, 1.001)		0.918	
Simple mode	1.000 (0.998, 1.001)		0.870	
Weighted mode	1.000 (0.998, 1.001)	H	0.921	
Cathepsin O	,			13.80%
MR Egger	0.999 (0.998.1.001)		0.562	
Weighted median	1.000 (0.999 1.001)	⊢ ↓	0.547	
Inverse variance weighted	1 001 (1 000 1 001)		0 116	
Simple mode	1.000 (0.999 1.002)		0.935	
Weighted mode	1.000 (0.998 1.002)		0.920	
Cathensin S				2.50%
MR Egger	1.000 (0.999 1.001)		0.759	
Weighted median	1 000 (0 999 1 000)	—	0.527	
Inverse variance weighted	1 000 (0 999 1 000)	H+1	0.114	
Simple mode	0.999 (0.998 1.000)		0.180	
Weighted mode	1 000 (0.990, 1.000)		0.053	
athensin Z	1.000 (0.333, 1.001)		0.933	2 50%
	1.000 (0.000, 1.001)		0.502	2.30%
Waighted median	1.000 (0.999, 1.001)		0.393	
weighted median	1.000 (0.999, 1.000)		0.200	
inverse van ance weighted	1.000 (0.999, 1.000)		0.789	
Simple mode	1.000 (0.998, 1.001)		0.378	
weighted mode	1.000 (0.999, 1.000)		0.451	

FIGURE 2

Forest plot of causal effects of cathepsin single-nucleotide polymorphisms (SNPs) on ulcerative colitis. We conducted inversevariance weighted analyses, among others, to evaluate the causal relationship between cathepsins and ulcerative colitis. (error bars represent 95% confidence intervals).

3 Results

3.1 Defining the causal relationship between various cathepsins and UC

The causal associations between eleven types of cathepsins (cathepsin B, E, F, G, H, L2, O, S, and Z) and IBD were analyzed. The forward MR analysis did not reveal any causal associations between eleven types of cathepsins and the risk of IBD (Figure 2).

Dutcome	b (95%CI)	se .	P-value	Power
American B	2 600 (17 410 22 627)	10.010	0.000	21%
MK Egger	2.009 (-17.419,22.037)	10.218	0.800	
weighted median	-0.915 (-15.021,13.191)	/.19/	0.899	
inverse variance weighted	0.303 (-8.3/8,9.104)	4.400	0.935	
Simple mode	-8./44 (-34.800,1/.3//)	13.327	0.515	
weighted mode	-3.593 (-20.917,13.731)	8.839	0.080	60.000/
And the second s	2 742 (7 508 14 004)	5 7 40	0.519	00.20%
Waighted median	2.025 (2.420 7.500)	3.740	0.516	
weighted median	2.033(-3.430,7.300)	2.788	0.400	
linverse variance weighted	0.272 (-5.725,4.270)	2.040	0.894	
Simple mode	4.330 (-0.320,14.999)	5.440	0.430	
weighted mode	5.745 (-5.050,12.542)	4.469	0.409	10.0%
MR Egger	12 822 (6 501 24 225)	10 415	0 101	100%
Weighted median	6 266 (9 729 21 470)	7 706	0.191	
Vergineer meetrali	6 204 (2 157 15 765)	1.700	0.407	
Simple mode	0.804(-2.157,15.705)	4.572	0.157	
Weighted mode	10.936 (9.594.30.362)	0.012	0.310	
weighted mode	10.830 (-8.394,30.207)	9.915	0.280	10.09/
MR Egger	4 564 (-24 017 14 000)	0.025	0.649	100%
Weighted median	4 403 (-17 720 0 022)	6 902	0.048	
weighted median	-4.403 (-17.738,8.932) 8 100 (16 681 0 483)	4 2 79	0.01/	
simula made	-6.100 (-10.081,0.482)	4.378	0.004	
Simple mode	5.890 (-25.530,31.321)	13.993	0.782	
weighted mode	-0.980 (-19.057,17.084)	9.220	0.915	10007
atnepsin G	0.500 (00.700 010)	10.007	0.002	100%
MR Egger	0.590 (-20.766,21.946)	10.896	0.957	
Weighted median	4.301 (-9.304,17.907)	6.941	0.535	
Inverse variance weighted	9.966 (0.553,19.378)	4.802	0.038	
Simple mode	13.681 (-11.879,39.241)	13.041	0.300	
Weighted mode	2.939 (-15.238,21.116)	9.274	0.753	
Cathepsin H				100%
MR Egger	-1.094 (-20.547,18.359)	9.925	0.913	
Weighted median	7.733 (-6.422,21.888)	7.222	0.284	
Inverse variance weighted	2.931 (-5.651,11.513)	4.379	0.503	
Simple mode	8.496 (-16.261,33.252)	12.631	0.505	
Weighted mode	8.943 (-8.838,26.724)	9.072	0.330	
Cathepsin L1				100%
MR Egger	2.036 (-10.113,14.185)	6.198	0.744	
Weighted median	-0.358 (-6.081,5.365)	2.920	0.902	
Inverse variance weighted	2.671 (-1.894,7.237)	2.329	0.251	
Simple mode	-2.350 (-14.339,9.638)	6.116	0.703	
Weighted mode	-3.670 (-14.308,6.969)	5.428	0.503	
athepsin L2				36.60%
MR Egger	-1.847 (-21.301,17.607)	9.926	0.853	
Weighted median	2.242 (-11.656,16.140)	7.091	0.752	
Inverse variance weighted	-0.509 (-9.091,8.073)	4.379	0.907	
Simple mode	12.179 (-14.123,38.480)	13.419	0.369	
Weighted mode	3.193 (-15.766,22.151)	9.673	0.743	
Cathepsin O				100%
MR Egger	1.919 (-20.131,23.970)	11.250	0.865	
Weighted median	12.174 (-1.346,25.694)	6.898	0.078	
Inverse variance weighted	3.638 (-5.982,13.257)	4.908	0.459	
Simple mode	10.145 (-19.539,39.829)	15.145	0.506	
Weighted mode	12.054 (-5.948,30.056)	9.185	0.196	
athepsin S				100%
MR Egger	23.952 (4.497,43.407)	9.926	0.020	
Weighted median	5.438 (-7.223,18.099)	6.460	0.400	
Inverse variance weighted	3.560 (-5.362,12.482)	4.552	0.434	
Simple mode	-18.194 (-46.680,10.291)	14.534	0.217	
Weighted mode	12.414 (-7.481,32.308)	10.150	0.228	
athepsin Z				100%
MR Egger	-16.552 (-35.993,2.889)	9.919	0.102	
Weighted median	-4.134 (-16.598,8.330)	6.359	0.516	
Inverse variance weighted	-6.788 (-15.370,1.794)	4.379	0.121	
	1 610 (20 002 27 612)	14 930	0.914	
Simple mode	-1.019 (-30.882,27.043)	14.950	0.211	
Simple mode Weighted mode	-9.031 (-28.645,10.584)	10.007	0.372	

method estimates the causal effects of ulcerative colitis on cathepsins.

We conducted reverse MR analyses to explore the possibility of reverse causality. The reverse MR analysis provided evidence that UC elevated cathepsin G levels (IVW: p = 0.038, b = 9.966) (Figures 3, 4), and the *p*-values of the Cochran's Q test, the MR-PRESSO global test, and the MR-Egger intercept showing no signs of heterogeneity and directional pleiotropy (0.167, 0.168, and 0.343, respectively). The statistical power of this analysis reached 100%. The effect estimate from MR-Egger is 0.590, while



the IVW estimate is 9.966. Although the *p*-value for the MR-Egger intercept test is 0.343, we note that the b-value from MR-Egger (0.590) is considerably smaller than its standard error

(9.966), which may suggest potential pleiotropy. No evidence supported a causal association between UC and other types of cathepsins (Figure 3).

Exposure	OR (95%CI)		P-value	Power
Cathepsin B				2.50%
MR Egger	1.000 (0.999, 1.001)		0.770	
Weighted median	1.000 (1.000, 1.001)	H +	0.956	
Inverse variance weighted	1.000 (1.000, 1.000)	H H H	0.613	
Simple mode	1.000 (0.999, 1.001)	⊢ ••−−1	0.922	
Weighted mode	1.000 (0.999, 1.001)		0.934	
Cathepsin D				2.50%
MR Egger	1.000 (0.999, 1.001)	→	0.670	
Weighted median	1.000 (0.999, 1.001)	H++	0.501	
Inverse variance weighted	1.000 (1.000, 1.001)	⊢ ♦ – I	0.527	
Simple mode	1.000 (0.998, 1.002)		0.995	
Weighted mode	1.000 (0.999, 1.001)	→	0.454	
Catheosin E	(, , , , , , , , , , , , , , , , , , ,			2.50%
MR Egger	1 000 (0 999 1 002)		0.603	
Weighted median	1 000 (0 999 1 001)		0.842	
Inverse vorignee weighted	1,000 (0,000, 1,001)		0.760	
Simple mode	1.000 (0.999, 1.001)		0.700	
Simple mode	1.000 (0.998, 1.001)		0.720	
weighted mode	1.000 (0.998, 1.001)		0.037	0.000/
amepsin r	1 000 (0 000 1 000)		0.000	2.50%
MR Egger	1.000 (0.999, 1.001)		0.986	
Weighted median	1.000 (1.000, 1.001)	H++	0.329	
Inverse variance weighted	1.000 (1.000, 1.001)	H+H	0.467	
Simple mode	1.000 (0.999, 1.001)	⊢ −	0.972	
Weighted mode	1.000 (0.999, 1.001)	⊢ •−	0.382	
Cathepsin G				2.50%
MR Egger	1.001 (0.999, 1.002)	→	0.397	
Weighted median	1.000 (0.999, 1.001)		0.704	
Inverse variance weighted	1.000 (0.999, 1.000)		0.714	
Simple mode	1 000 (0 999 1 001)		0.775	
Weighted mode	1 000 (0 999 1 001)		0.768	
Tothensin H	1.000 (0.555, 1.001)		0.700	2 50%
MP Egger	1 000 (1 000 1 000)		0.727	2.3070
Wik Egger	1.000 (1.000, 1.000)		0.737	
weighted median	1.000 (1.000, 1.000)	1	0.554	
Inverse variance weighted	1.000 (1.000, 1.000)	141	0.786	
Simple mode	1.000 (0.999, 1.000)		0.402	
Weighted mode	1.000 (1.000, 1.000)	H	0.507	
Cathepsin L1				2.50%
MR Egger	1.000 (0.998, 1.001)		0.550	
Weighted median	1.000 (0.999, 1.001)	⊢ +•−−1	0.440	
Inverse variance weighted	1.000 (0.999, 1.001)	⊢ ••−-1	0.976	
Simple mode	1.001 (0.999, 1.003)	· · · · • · · · · · · · · · · · · · · ·	0.382	
Weighted mode	1.001 (0.999, 1.003)	· · · · • · · · · · · · · · · · · · · ·	0.380	
Catheosin L2				2.50%
MR Egger	1.000 (0.998, 1.002)		0.909	
Weighted median	1 000 (1 000 1 001)	H+	0.310	
Inverse variance weighted	1 000 (0 999 1 001)		0.820	
Simple mode	1 000 (0.999, 1.001)		0.601	
Weighted mode	1.000 (0.999, 1.002)		0.562	
Tathansin O	1.000 (0.333, 1.002)		0.505	2 5 0 %
MR Farer	1 001 (0 000 1 002)		0.294	2.30%
MIK Egger	1.001 (0.999, 1.002)		0.584	
weighted median	1.000 (0.999, 1.001)		0.009	
Inverse variance weighted	1.000 (1.000, 1.001)	H.	0.616	
Simple mode	1.000 (0.998, 1.001)		0.588	
Weighted mode	1.000 (0.998, 1.001)		0.544	
Cathepsin S				2.50%
MR Egger	1.000 (1.000, 1.001)	H +	0.747	
Weighted median	1.000 (1.000, 1.001)	HH-I	0.651	
Inverse variance weighted	1.000 (1.000, 1.000)	H	0.761	
Simple mode	0.999 (0.998, 1.000)	⊢ ♦ <u></u>	0.202	
Weighted mode	1.000 (1.000, 1.001)	H + -I	0.480	
Catheosin Z				2.50%
MR Fager	1 000 (0 999 1 001)		0.896	2.0073
Weighted median	1 000 (1 000 1 001)		0.050	
Inverse variance moight- 4	1.000 (1.000, 1.001)		0.0360	
Simula made	1.000 (1.000, 1.001)		0.308	
Sumple mode	1.000 (0.999, 1.001)		0.408	
weighted mode	1.000 (1.000, 1.001)		0.794	

FIGURE 5

Forest plot of causal effects of cathepsin single-nucleotide polymorphisms (SNPs) on Crohn's disease. We conducted inversevariance weighted analyses to evaluate the causal relationship between cathepsins and Crohn's disease. (error bars represent 95% confidence intervals).

3.2 Defining the causal relationship between various cathepsins and CD

The causal associations between eleven types of cathepsins (cathepsin B, D, E, F, G, H, L1, L2, O, S, and Z) and CD were analyzed. The forward MR analysis did not reveal any causal associations between eleven types of cathepsins and CD risk (Figure 5). We conducted reverse MR analyses to explore the possibility of reverse causality. The reverse MR analysis provided evidence that CD decreased the level of cathepsin B (cathepsin B:

Outcome	b (95%CI)	se	P-value	Power
Cathepsin B				100%
MR Egger	-13.389 (-21.649,-5.129)	4.214	0.005	
Weighted median	-11.954 (-21.800,-2.108)	5.023	0.017	
Inverse variance weighted	-10.525 (-17.330,-3.721)	3.472	0.002	
Simple mode	-2.952 (-38.640,32.736)	18.208	0.873	
Weighted mode	-11.473 (-20.368,-2.578)	4.538	0.020	
Cathepsin D				83.30%
MR Egger	-1.283 (-5.836,3.270)	2.323	0.586	
Weighted median	-0.372 (-5.001,4.256)	2.361	0.875	
Inverse variance weighted	-0.447 (-4.162,3.268)	1.896	0.814	
Simple mode	-1.112 (-14.192,11.968)	6.673	0.869	
Weighted mode	-0.330 (-4.545,3.884)	2.150	0.879	
Cathepsin E				99.60%
MR Egger	-3.406 (-11.666,4.854)	4.214	0.429	
Weighted median	-2.311 (-10.340,5.719)	4.097	0.573	
Inverse variance weighted	-1.813 (-8.617,4.992)	3.472	0.602	
Simple mode	4.049 (-29.008,37.106)	16.866	0.813	
Weighted mode	-2.567 (-10.235,5.101)	3.912	0.519	
Jamepsin r	2 742 (12 001 4 510)	4.01.4	0.205	100%
Wirk Egger	-3.742 (-12.001,4.518)	4.214	0.380	
weighted median	4.481 (-12.583,3.621)	4.134	0.2/8	
Simple med	4.280 (-11.090,2.519)	3.4/2	0.217	
Simple mode	17.030 (-15.705,49.765)	10./01	0.320	
weighted mode	-4.488 (-11.774,2.798)	3.717	0.241	1009/
MR Egger	8 537 (16 071 0 104)	4 202	0.063	100%
MIK Egger	-8.537 (-10.971,-0.104)	4.303	0.002	
weighted median	-7.344 (-13.901,1.213)	4.300	0.093	
niverse vanance weighted	-4.029 (-11.384,3.327)	3.733	0.285	
Simple mode	19.840 (-18.303,38.237)	19.597	0.323	
Cothensin H	-/.4/0 (-15.089,0.757)	4.190	0.090	57 109/
VR Fager	2 062 (5 106 11 221)	4 214	0.476	57.10%
Weighted median	1 853 (5 620 9 325)	3 91 2	0.470	
Inverse verience weighted	0.830 (5.065.7.643)	3.012	0.027	
Simple mode	4 927 (27 047 27 194)	16 2 0 0	0.009	
Weighted mode	1 702 (5 521 0 117)	2 727	0.627	
Cathensin I 1	1.735 (-5.551,5.117)	3.737	0.037	100%
MR Egger	-6 306 (-12 016 -0 597)	2 913	0.041	10070
Weighted median	4 591 (-9 559 0 377)	2.535	0.070	
Inverse variance weighted	4 742 (-9 377 -0 108)	2.365	0.045	
Simple mode	-8 028 (-22 517 6 461)	7 392	0.288	
Weighted mode	-4 419 (-8 885 0 048)	2 279	0.064	
Cathensin L2	(0.000,0.010)	2.277	0.001	100%
MR Egger	4 122 (-5 226 13 471)	4 770	0.398	10070
Weighted median	3,568 (-4,231,11,366)	3.979	0.370	
Inverse variance weighted	2.816 (-4.737.10.368)	3,853	0.465	
Simple mode	-17.821 (-58.283.22.641)	20.644	0.398	
Weighted mode	3.819 (-3.283,10.920)	3.623	0.304	
Cathepsin O	(,)			100%
MR Egger	3.785 (-5.629.13.198)	4.803	0.440	
Weighted median	4.101 (-4.017.12.219)	4.142	0.322	
Inverse variance weighted	2.326 (-5.290.9.941)	3.885	0.549	
Simple mode	4.103 (-31.643.39.849)	18.238	0.824	
Weighted mode	4.103 (-3.218,11.424)	3.735	0.285	
Cathepsin S	/			32%
MR Egger	-1.190 (-9.740,7.360)	4.362	0.788	
Weighted median	-1.303 (-9.358,6.752)	4.110	0.751	
Inverse variance weighted	-0.585 (-7.460,6.291)	3.508	0.868	
Simple mode	-0.791 (-38.959,37.376)	19.473	0.968	
Weighted mode	-1.369 (-8.350,5.612)	3.562	0.705	
Cathepsin Z				3%
•	0.893 (-7.367,9.153)	4.214	0.834	
MR Egger		2 00 4	0 725	
MR Egger Weighted median	1.374 (-6.277,9.025)	3.904	0.725	
MR Egger Weighted median Inverse variance weighted	1.374 (-6.277,9.025) 0.029 (-6.776,6.833)	3.904	0.725	
MR Egger Weighted median Inverse variance weighted Simple mode	1.374 (-6.277,9.025) 0.029 (-6.776,6.833) -3.401 (-35.899,29.097)	3.472 16.581	0.723 0.993 0.840	

FIGURE 6 Two-sample Mendelian randomization study based on the MR method estimates the causal effects of Crohn's disease on cathepsins.

IVW: p = 0.002, b = -10.525) (Figures 6, 7). The *p*-values of Cochran's Q test, the MR-PRESSO global test, and the MR-Egger intercept showed no signs of heterogeneity, outliers, or directional pleiotropy (0.828, 0.709, and 0.245, respectively). The reverse MR analysis provided evidence that CD decreased the level of cathepsin L1 (IVW: p = 0.045, b = -4.742) (Figure 8), and the *p*-value of the Cochran's Q test was 0.047, suggesting the presence of slight but statistically significant heterogeneity among the genetic instruments deployed. The *p*-values of the MR-PRESSO global test and the MR-Egger intercept showed no signs of outliers or directional pleiotropy (0.147 and 0.365,



respectively). The statistical power of these analyses reached 100%. No evidence supported a causal association between CD and other types of cathepsins (Figure 6).

Overall, in primary MR analyses, IBD was statistically associated with cathepsin B and cathepsin G, suggesting the development of IBD might affect the level of these cathepsins.



4 Discussion

IBD is a prevalent chronic immune-mediated inflammatory disorder of the digestive tract that, if left untreated, can lead to

prolonged bowel damage. Recent observational analyses have suggested that intestinal pathophysiological conditions can disrupt proteolytic homeostasis, resulting in inappropriate substrate degradation and abnormal accumulation (Sekula et al., 2016). Among the key players in these processes, cathepsins have garnered significant attention from researchers. In this study, we conducted a systematic analysis of the causal relationship between eleven distinct cathepsins and IBD using genetic instruments. Our findings revealed that cathepsin levels do not significantly influence the onset or progression of IBD. Reverse Mendelian randomization analyses suggested that ulcerative colitis (UC) might lead to elevated cathepsin G levels, while Crohn's disease (CD) may cause a decrease in cathepsin B and L1 levels.

Previous observational research partially corroborates our findings regarding the association between UC and cathepsin G. Marta Dabek et al. demonstrated through RT-PCR and Western blotting that Cat-G is overexpressed in UC patients, with higher mRNA expression in biopsies than healthy subjects (0.44 ± 0.08 vs 0.24 ± 0.03 , p < 0.05) (Dabek et al., 2009). Additionally, Kuwana's study showed that anti-neutrophil cytoplasmic antibodies targeting Cat-G are present in the sera of more than half of UC patients (Kuwana et al., 2000). These findings suggest that Cat-G may play a crucial role in UC pathogenesis. However, due to the larger uncertainty in the MR-Egger estimate, we cannot entirely rule out the presence of pleiotropy. Thus, further investigation is necessary to confirm the causal relationship between UC and cathepsin G.

Interestingly, our study revealed that CD occurrence correlates with lower levels of cathepsin B and cathepsin L1, which contradicts previous observational research and clinical studies. Menzel et al. confirmed the upregulation of cathepsin B and cathepsin L in areas of tissue damage and mucosal ulceration in IBD patients using immunohistochemistry and gene expression analysis (Menzel et al., 2006). Moreover, elevated cathepsin B levels have been observed in various pathological conditions, including inflammation, infection, and cancer. Benjamin Bian et al. reported increased mRNA and activated levels of cathepsin B in human adenomas and colorectal cancers (CRCs) at all stages (Bian et al., 2016). Zhenhu Zhang's study demonstrated significantly higher expression levels of CTSL in ESCC tissues than adjacent non-cancerous tissues (Zhang et al., 2024); Shengnan Zhao's research revealed increased cathepsin B levels in colonic tissue and exacerbated dextran sodium sulfate (DSS)-induced colitis (Zhao et al., 2016). The discrepancy between our findings and previous observational studies could be attributed to bias from reverse causality or residual confounding in the latter. These divergent results underscore the need for further research to elucidate the complex relationship between cathepsins and IBD.

This study exhibits several notable strengths and limitations. First, MR leverages genetic variants to estimate the causal effects of circulating proteins on IBD, effectively overcoming the bias caused by reverse causality and confounding. Second, we performed sensitivity and pleiotropic analyses to ensure the accuracy of MR analysis. We used exposures and outcomes in European populations from different countries to minimize the potential for population stratification bias. Despite these insights, several limitations warrant consideration, such as (1) the potential overlap of participants between the exposure and outcome GWAS in the two-sample MR analyses could not be ascertained in this study; (2) the presence of pleiotropy, where instrumental SNPs may influence multiple traits, could not be entirely ruled out; however, no evidence of pleiotropy was observed in the MR analyses conducted in any of the above-mentioned MR approaches; (3) given that our study cohort comprised exclusively of individuals of European ancestry, caution should be exercised when generalizing these findings to broader populations.

In conclusion, this study's primary genetic evidence reveals a nuanced relationship between cathepsins and IBD. Cathepsins do not appear to have a direct impact on IBD. While the occurrence of UC may be associated with elevated levels of cathepsin G, CD was linked to lower levels of cathepsins B and L1. Future research should aim to definitively establish and validate these causal relationships. These insights may prove valuable in identifying biochemical markers for the prediction, screening, early diagnosis, and prognosis of IBD.

Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material; further inquiries can be directed to the corresponding author.

Author contributions

NW: writing-original draft, methodology, writing-review and editing, conceptualization, formal analysis, funding acquisition, investigation, resources, software, validation, and visualization. JL: conceptualization, investigation, validation, formal analysis, methodology, software, resources, and writing-original draft. BC: supervision, data curation, methodology, conceptualization, software, writing-original draft, investigation, writing-review and editing, and resources. JY: visualization, methodology, validation, writing-original draft, and investigation. XD: visualization, validation, writing-original draft, methodology, and investigation. QM: methodology, investigation, supervision, software, resources and writing-review and editing. XW: investigation, visualization, writing-original draft, formal analysis, resources, data curation, project administration, funding acquisition, conceptualization, supervision, validation, and writing-review and editing.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

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