Check for updates

OPEN ACCESS

EDITED BY Jonas Alex Morales Saute, Federal University of Rio Grande do Sul, Brazil

REVIEWED BY Pablo Brea Winckler, Federal University of Rio Grande do Sul, Brazil Jakub Piotr Fichna, Polish Academy of Sciences, Poland Florian Barthelemy, University of California, Los Angeles, United States André Macedo Silva, University of São Paulo, Brazil

*CORRESPONDENCE Svetlana Gorokhova, 🛙 svetlana.gorokhova@univ-amu.fr

RECEIVED 18 June 2023 ACCEPTED 29 November 2023 PUBLISHED 13 December 2023

CITATION

Morel V, Audic F, Tardy C, Verschueren A, Attarian S, Nguyen K, Salort-Campana E, Krahn M, Chabrol B and Gorokhova S (2023), Retrospective clinical and genetic analysis of COL6-RD patients with a long-term follow-up at a single French center. *Front. Genet.* 14:1242277. doi: 10.3389/fgene.2023.1242277

COPYRIGHT

© 2023 Morel, Audic, Tardy, Verschueren, Attarian, Nguyen, Salort-Campana, Krahn, Chabrol and Gorokhova. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Retrospective clinical and genetic analysis of COL6-RD patients with a long-term follow-up at a single French center

Victor Morel¹, Frédérique Audic^{2,3}, Charlotte Tardy¹, Annie Verschueren⁴, Shahram Attarian^{3,4}, Karine Nguyen^{1,3}, Emmanuelle Salort-Campana^{3,4}, Martin Krahn^{1,3}, Brigitte Chabrol^{2,3} and Svetlana Gorokhova^{1,3}*

¹Département de Génétique Médicale, Hôpital de la Timone, Marseille, Provence-Alpes-Côte d'Azur, France, ²Service de Neuropédiatrie, Centre de Référence des Maladies Neuromusculaires de l'enfant PACARARE, CHU Timone, APHM, Marseille, France, ³Inserm, U1251-MMG, Marseille Medical Genetics, Aix Marseille University, Marseille, France, ⁴Centre de Référence des Maladies Neuromusculaires et de la SLA, ERN-NMD, CHU Timone, APHM, Marseille, France

Collagen type VI-related dystrophies (COL6-RD) are rare diseases with a wide phenotypic spectrum ranging from severe Ullrich's congenital muscular dystrophy Ullrich congenital muscular dystrophy to much milder Bethlem myopathy Both dominant and recessive forms of COL6-RD are caused by pathogenic variants in three collagen VI genes (COL6A1, COL6A2 and COL6A3). The prognosis of these diseases is variable and difficult to predict during early disease stages, especially since the genotype-phenotype correlation is not always clear. For this reason, studies with long-term follow-up of patients with genetically confirmed COL6-RD are still needed. In this study, we present phenotypic and genetic data from 25 patients (22 families) diagnosed with COL6-RD and followed at a single French center, in both adult and pediatric neurology departments. We describe three novel pathogenic variants and identify COL6A2:c.1970-9G>A as the most frequent variant in our series (29%). We also observe an accelerated progression of the disease in a subgroup of patients. This large series of rare disease patients provides essential information on phenotypic variability of COL6-RD patients as well as on frequency of pathogenic COL6A gene variants in Southern France, thus contributing to the phenotypic and genetic description of Collagen type VIrelated dystrophies.

KEYWORDS

collagen type VI-related myopathies, COL6-RD, Bethlem myopathy, ullrich's congenital muscular dystrophy, COL6 genes, *COL6A1*, *COL6A2*, *COL6A3*

Introduction

Collagen VI-related Dystrophies (COL6-RDs) are caused by pathogenic variants in the *COL6A1*, *COL6A2* and *COL6A3* genes and can be inherited in an autosomal dominant or recessive manner (Foley et al., 2021). These genes encode the three alpha chains of collagen type VI (α 1, α 2 and α 3). Type VI collagen is a major component of the extracellular matrix, essential for the stability, nutrition, and development of muscle and connective tissue (Bönnemann, 2011). COL6-RDs represent a continuum of muscular disorders with large clinical spectrum consisting of three groups of COL6-RDs (Foley et al., 2021; Natera-de

Benito et al., 2021). Ullrich congenital muscular dystrophy (UCMD) corresponds to the most severe phenotype, characterized by congenital weakness, hypotonia, proximal joint contractures, distal joints laxity and kyphoscoliosis. Age of onset is neonatal or early infancy. Some affected children acquire the ability to walk independently; however, progression of the disease results in a loss of ambulation in first decade, accompanied by generalized muscle atrophy and early respiratory failure (Foley et al., 2021). Bethlem myopathy (BM) is at the other end of the phenotypic spectrum. This much milder form of COL6-RD is characterized by a combination of proximal muscle weakness and joint contractures. There is mild to no developmental motor delay and all patients achieve ambulation. Neonatal features might be noted, but age of onset is usually childhood to early adulthood with evidence of slowly progressing weakness as well as contractures of the elbows, Achilles tendons, and long finger flexors (Foley et al., 2021). The intermediate form of COL6-RD often has a congenital onset, but the prognosis for the motor and respiratory impairment is better for these patients is better comparing to that of UCMD patients (Natera-de Benito et al., 2021). Certain clinical features are common to all COL6-RD subtypes, such as signs of distal laxity, skin damage (keloid scars, follicular hyperkeratosis and normal to mildly elevated levels of serum creatine kinase (CK) (Zanoteli et al., 2020; Foley et al., 2021; Natera-de Benito et al., 2021). Specific signs on muscle imaging can also be noted (Mercuri et al., 2005).

Interpretation of variants identified by genetic testing in COL6-RD patients can be quite challenging due to variable clinical presentation as well as presence of both recessive and dominant modes of inheritance for these disorders. More studies with phenotypic and genotypic description of COL6-RD patients are therefore needed. We report a new series of 25 French patients (22 families) with genetically confirmed COL6-RD. In this retrospective study, we describe the phenotypic and genotypic characteristics of COL6-RD patients with a long-term follow-up at the University Hospital of La Timone, Marseille. We define the frequencies of pathogenic *COL6A1-3* variants in the South of France and describe a subgroup of patients with an accelerated disease progression, thus contributing to better understanding of Collagen VI-related Dystrophies.

Materials and methods

The patients were referred and followed at the University Hospital of Marseille (AP-HM, Marseille, France), in the Department of Adult Neuromuscular Disease and the Department of Pediatric Neurology. Patients F10, F14 and F15.1 were diagnosed by targeted Sanger sequencing of COL6A1 and COL6A2 genes after the diagnosis of COL6-RD was suspected due to presence of congenital hip dislocation, distal hyperlaxity or skin signs. Patients F5.2 and F15.2 were diagnosed by targeted Sanger sequencing since a pathogenic variant has been previously identified in the affected proband. The variants in the remaining 22 patients were identified by targeted exome sequencing (gene panel) using the genes on the National French Consensus Gene Lists for the diagnosis of myopathies (Krahn et al., 2019), as detailed in the Supplementary Table S1. Variants identified by high-throughput sequencing were first filtered using general population frequency, retaining only variants with <1% minor allele frequency in gnomAD v2.1.1 database. The remaining variants were then classified according to the ACMG/AMP guidelines as described in the Supplementary Material. As more information about pathogenicity of *COL6A1*, *COL6A2*, *COL6A3* variants became available over the years, initial classifications of certain variants have been modified.

All patients received standard care according to French National Protocol of Diagnostic and Care for COL6-RD established by the French Neuromuscular Disease Network (FILNEMUS, https://hassante.fr/upload/docs/application/pdf/2022-10/pnds_col6_.pdf).

The study was conducted according to the guidelines of the Declaration of Helsinki. It was registered by AP-HM (Assistance Publique, Hôpitaux de Marseille) under number GBP3T5/PADS22-157, evaluating this study as exempt from the Institutional Review Board according to the French legislation. The appropriate consent was collected from each patient as part of standard procedure during diagnostic genetic testing in our laboratory. Additional written informed consent was collected for patients if detailed phenotype description was included in the manuscript.

Results and discussion

We report 25 patients (22 families) with genetically confirmed COL6-RD. All of them have been followed at the University Hospital of Marseille (AP-HM), in the Department of Adult Neuromuscular Disease and the Department of Pediatric Neurology. Certain patients were initially followed in their childhood by the pediatric team and then transitioned to the adult care unit. The genetic diagnoses were established by the Medical Genetics Department of the same hospital for all patients except for F18. The close interactions between these three Departments as well as the unified medical record system allowed an efficient life-long follow up and care of the described COL6-RD patients, accompanied by a genetic diagnosis that in many cases became possible due to the arrival and evolution of high throughput sequencing technologies.

Summary of genetic findings in our patient series

Fourteen patients (12 probands) were diagnosed with a recessive form of COL6-RD with bi-allelic variants confirmed by molecular testing (12 patients with homozygous variants and two patients with compound heterozygous variants). Eight different variants were responsible for these cases (two nonsense/frameshift, four spliceaffecting variants and two missense variants). Eleven patients (10 probands) had a dominant COL6-RD, explained by a pathogenic or likely pathogenic heterozygous variant in one of the three COL6A genes. Eight different variants were identified in this group - five missense variants affecting Glycine residues in Gly-X-Y motifs of the triple-helical region of collagen and three in-frame deletions of one exon at the mRNA level (one large deletion and two splice-affecting single-nucleotide variants leading to exon skipping). Three of the identified pathogenic variants have never been published-one variant in the COL6A1 gene (in-frame deletion of exon 3) and two variants in the COL6A2 gene (c.1752del,

TABLE 1 Variants identified in patients with COL6-RDs.

Patient	Phenotype class	Genomic coordinates (GRCh37)	Gene	Exon intron	Variant(s)	Variant type (consequence)	Alleles
Patients with bi-allelic variants (recessive)							
F1	В	chr21:47406990	COL6A1	In 5	c.717 + 4A>G; p.(?)	splicing	hom
F2.1	В	chr21:47409010	COL6A1	Ex 9	c.817A>T; p.(Lys273*)	nonsense	hom
F2.2	В	chr21:47409010	COL6A1	Ex 9	c.817A>T; p.(Lys273*)	nonsense	hom
F3	U	chr21:47531504_47531517	COL6A2	Ex 2	c.114_115 + 12del; p.(?)	splicing	hom
F4	В	chr21:47545690	COL6A2	In 25	c.1970-9G>A; p.(Thr656fs)	splicing	hom
F5.1	В	chr21:47545690	COL6A2	In 25	c.1970-9G>A; p.(Thr656fs)	splicing	hom
F5.2	В	chr21:47545690	COL6A2	In 25	c.1970-9G>A; p.(Thr656fs)	splicing	hom
F6	В	chr21:47545690	COL6A2	In 25	c.1970-9G>A; p.(Thr656fs)	splicing	hom
F7	U	chr21:47545690	COL6A2	In 25	c.1970-9G>A; p.(Thr656fs)	splicing	hom
F8	В	chr21:47545690	COL6A2	In 25	c.1970-9G>A; p.(Thr656fs)	splicing	hom
F9	В	chr21:47545690	COL6A2	In 25	c.1970-9G>A; p.(Thr656fs)	splicing	hom
F10	В	chr21:47545690	COL6A2	In 25	c.1970-9G>A; p.(Thr656fs)	splicing	comp
		chr21:47544816		Ex 23	c.1752del; p.(Gly585Glufs*)	frameshift	net
F11	Ι	chr21:47552300	COL6A2	Ex 28	c.2894G>C; p.(Arg965Pro)	missense	hom
F12	В	chr2:238253214	COL6A3	Ex 36	c.7447A>G; p.(Lys2483Glu)	missense	comp
		chr2:238234368		Ex 43	c.9329-1G>T; p.(?)	splicing	net
Patients with mono-allelic variants (dominant)							
F13	U	chr21:(47402678_47404182) _(47404384_47406439)	COL6A1	Ex 3	c.(227 + 1_228-1)_(428 + 1_429-1)del; p.(?)	in-frame exon deletion	het
F14	В	chr21:47407552	COL6A1	Ex 8	c.788G>A; p.(Gly263Asp)	missense	het
F15.1	U	chr21:47408998	COL6A1	Ex 9	c.805G>A; p.(Gly269Arg)	missense	het
F15.2	В	chr21:47408998	COL6A1	Ex 9	c.805G>A; p.(Gly269Arg)	missense	het
F16	В	chr21:47409016	COL6A1	Ex 9	c.823G>A; p.(Gly275Arg)	missense	het
F17	U	chr21:47409043	COL6A1	Ex 9	c.850G>A; p.(Gly284Arg)	missense	het
F18	Ι	chr21:47409881	COL6A1	In 11	c.930 + 189C>T; p.(?)	in-frame pseudoexon insertion	het
F19	Ι	chr21:47410741	COL6A1	In 14	c.1056 + 1G>A; p.(?)	in-frame exon skipping	het
F20	Ι	chr21:47410741	COL6A1	In 14	c.1056 + 1G>A; p.(?)	in-frame exon skipping	het
F21	Ι	chr21:47410741	COL6A1	In 14	c.1056 + 1G>A; p.(?)	in-frame exon skipping	het
F22	Ι	chr21:47535960	COL6A2	Ex 7	c.893G>A; p.(Gly298Glu)	missense	het
Patients with highly suspected COL6-RD, carrying a VUS							
F23	U	chr21:47421932	COL6A1	Ex 31	c.2014G>A; p.(Glu672Lys)	missense	hom
F24	В	chr21:47417650	COL6A1	Ex 22	c.1498G>A; p.(Gly500Arg)	missense	het
		chr2:238253214	COL6A3	Ex 36	c.7447A>G; p.(Lys2483Glu)	missense	het

All variants in this table are pathogenic or likely pathogenic, except for variants COL6A1 NM_001848.3:c.2014G>A; p.(Glu672Lys) and COL6A1 NM_001848.3:c.1498G>A p.(Gly500Arg). See Supplementary Table S1 for the detailed evidence used for ACMG/AMP, classification of variants. F, family; In, Intron; Ex, Exon; Hom, homozygous; comp het, compound heterozygous; thet, heterozygous; U, ullrich congenital muscular dystrophy; I, intermediate form; B, bethlem myopathy; Y, yes; N, no; VUS, variant of uncertain significance. The following transcripts were used for variant nomenclature: *COL6A1* NM_001848.3, *COL6A2* NM_001849.4, *COL6A3* NM_004369.4.

Bold values indicates that families are numbered from 1 to 24 (F1-F24). .1 and .2 are used to differentiate patients within the same family.



Variants identified in patients with COL6-RDs. Pathogenic and likely pathogenic variants identified in patients with genetically confirmed COL6-RD are shown. The triple helical domains in each collagen molecule are shown in pink. The exon-intron boundaries are marked by dashed lines. The missense variants are shown in grey, truncating variants are shown in red, while in-frame exon skips and deletions as well as in-frame pseudoexon insertions are shown in lavender. The bi-allelic (recessive) variants are shown above the protein bar, while the mono-allelic (dominant) variants are shown below the protein bar. The following transcripts were used for variant nomenclature: *COL6A1* NM_001848.3, *COL6A2* NM_001849.4, *COL6A3* NM_004369.4.

p.(Gly585Glufs*11) and c.114_115 + 12del, p.(?)). The summary of all pathogenic and likely pathogenic variants identified in each patient is listed in Table 1, while detailed description of genetic evidence and ACMG/AMP codes used for variant classification is provided in the Supplementary Table S2.

The numbers of patients with dominant and recessive COL6-RD were similar in our study, which is different from most previous COL6-RD cohorts where dominant cases were more prevalent (Briñas et al., 2010; Zanoteli et al., 2020; Natera-de Benito et al., 2021; Kwong et al., 2023). This difference is due to presence of seven probands carrying COL6A2:c.1970-9G>A variant on one or both alleles (six homozygous and one compound heterozygous). Similar to other COL6-RD cohorts, the patients with dominant COL6-RD in our series carried heterozygous glycine substitutions and in-frame events (exon skipping, exon deletion or pseudo-exon insertion). As in other studies, pathogenic glycine substitutions clustered in the N-terminal part of the triple helical domain (Figure 1). Glycine residue affecting 10th to 15th Gly-X-Y motifs of the triple helical domain tend to be more severe (Butterfield et al., 2013; Lamandé and Bateman, 2018). Moreover, heterozygous variants affecting the C-terminal part of the triple helical domain are found in the general population according to gnomAD database, suggesting that these variants are recessive or benign (Lamandé and Bateman, 2018). Collagen molecules with variants in the C-terminal part of the triple helical domain are thought to be excluded from the microfibril, while molecules with the variants in the N-terminal part are incorporated, thus exerting a dominant negative effect (Pace et al., 2008; Lamandé and Bateman, 2018). Two patients in our study carried variants in the N-terminal part of the triple region—*COL6A1*:c.850G>A p.(Gly284Arg) located in the 10th repeat (F17) and *COL6A2*:c.893G>A p.(Gly298Glu) located in the 15th repeat (F22). These two patients had Ullrich and Intermediate phenotypes respectively. Homozygous variants causing premature protein truncation in the triple helix domains are generally known to lead to most severe phenotypes (Briñas et al., 2010; Natera-de Benito et al., 2021). Interestingly, the only two patients in our series with a homozygous variant in this region (*COL6A1*:c.817A>T; p.(Lys273*)) were able to walk independently, even though they lost ambulation during childhood (patient F2.1, F2.2).

Clinical findings in patients with genetically confirmed COL6-RD

The clinical features of 25 patients with genetically confirmed COL6-RD and two patients with highly suspected COL6-RD are summarized in Supplementary Table S1. Five genetically confirmed patients in our series were classified as Ullrich's congenital muscular dystrophy (UCMD) (OMIM: 254090) by the clinical team, while fifteen patients had a milder Bethlem myopathy (BM) (OMIM: 158810). The remaining patients were affected by an Intermediate form of COL6-RD. For 10 patients, certain symptoms of COL6-RD, such as hypotonia, feet deformities or congenital hip dislocation, were evident at birth. Fourteen patients had their first symptoms appearing during infancy or childhood, while three patients had a much later adult onset of symptoms. Seven patients in our series lost

the ability to walk, six of them during childhood (mean age = 8 years old). One patient had loss of ambulation at 57 years old. One patient has never acquired the ability to walk. In certain patients, a relatively mild initial disease course was followed by a phase of accelerated progression (marked as "+" in the "Accelerated Progression" column in the Supplementary Table S1). One of the patients that lost the ability to walk during childhood (at 10 years of age) and had an accelerated progression (F18) carried the COL6A1:c.930 + 189C>T deep intronic variant. Similar clinical presentation has been previously described for other carriers of this variant - these patients have a relative paucity of symptoms that then evolve rapidly to match a typical Ullrich Congenital Muscular Dystrophy with loss of ambulation around the age of nine, respiratory insufficiency and proximal contractures (Bolduc et al., 2019; Natera-de Benito et al., 2021). Another patient with an accelerated progression of symptoms and loss of ambulation during early childhood (4 years of age) carried the variant COL6A1:c.1056 + 1G>A (P21), while the other two patients with the same variant remained ambulant (F19 and F20). Interestingly, phenotypic variability has been previously described for this variant within the same family, with the mother presenting a relatively mild course of disease and the son showing an unusually fast progression with loss of ambulation during childhood (Lucioli et al., 2005). In comparison, all five patients carrying this variant in the recently published COL6-RD cohort had Bethlem myopathy (Natera-de Benito et al., 2021). Respiratory involvement was present in a third of patients (n = 8/24, data not available for one patient). Five of these patients needed bilevel positive airway pressure (BiPAP) (20.8% n = 5/24). Only two out of six UCMD patients had respiratory involvement. However, longer study is needed for the remaining four patients with normal respiratory function, as these patients are among the youngest of our series.

Intrafamilial variability in our patient series

Our series includes three families—F2, F5 and F15. Two affected members of F2 are brothers carrying homozygous truncating variant COL6A1:c.817A>T; p.(Lys273*). The phenotypes of these patients were similar, as they both presented first signs in early childhood (with weakness of the lower limbs and frequent falls at 3 years for F2.1 and abnormal walking at 4 years for F2.2). They lost the ability to walk at 4 years of age for F2.1 and 9 years of age for F2.2. Both brothers underwent spinal arthrodesis at the age of 14. They also both needed respiratory support starting from 18 years of age for F2.1 and from 33 years of age for F2.2. F2.1 died at 40 years of age of acute respiratory failure. The variant found in these patients has been reported as homozygous in a patient with neonatal onset COL6-RD, mild CK elevation (350 U/L) and reduction of collagen VI in skin biopsy (Gonzalez-Quereda et al., 2020).

Two affected members of F5 are sisters carrying homozygous variant COL6A2:c.1970-9G>A; p.(Thr656fs). While both patients had the onsets during childhood (weakness of the lower limbs for F5.1 and loss of the ability to run at the age of 12 for F5.2), the phenotype was less severe for F5.2. The patient F5.2 was followed in our neuromuscular department since the age of 48, while her sister (F5.1) was followed from the age of 14. Neither of these two patients had respiratory or skin signs.

Two affected members of F15 are mother and son, both carrying heterozygous missense variant COL6A1:c.805G>A; а p.(Gly269Arg). They had a similar onset, with first signs of the disease during the acquisition of walking, diagnosed by frequent falls. The phenotype of the son (F15.1) is somewhat more severe, as he cannot get up from the ground without assistance. Both patients have retractions and similar skin signs such as keloid scars. F15.2 has required respiratory assistance since the age of 29, while F15.1 does not have respiratory involvement. The variant COL6A1:c.805G>A; p.(Gly269Arg) has been previously reported as appearing de novo in a patient with an intermediate form of COL6-RD with a neonatal onset (Deconinck et al., 2010). Possible explanations for the observed phenotypic variability include genetic context of the variant, with coding or non-coding variants in trans or in cis, as well as epigenetic or environmental contributions to the phenotype.

Pathogenic variant frequencies observed in our patient series

Eight patients (seven unrelated families) in our series carried the variant COL6A2:c.1970-9G>A, either in a homozygous state (seven patients, F4-F9) or in a compound heterozygous state with a truncating variant c.1752del (one patient, F10), making it the most frequent variant in our series (29%). Indeed, the majority of patients with the recessive form of COL6-RD (7 out of 12 probands) carried the COL6A2:c.1970-9G>A variant. This variant introduces an aberrant splice acceptor site, leading to an insertion of seven nucleotides before exon 26 and resulting in a frame shift (Martoni et al., 2009; Foley et al., 2011). It has been identified in multiple patients affected with recessive COL6-RD, even though the frequency of this variant in published patient series is much lower than the one observed in our study (Quijano-Roy et al., 2014; Deconinck et al., 2015; Stehlíková et al., 2017). Similar to the previously described cases, the clinical presentation of patients with c.1970-9G>A was variable. For example, the age of onset ranged from birth to 35 years. Only four out of eight patients had retractions. The vast majority of these patients (7/8) were able to climb several steps without holding onto a railing. For now, none of the patients with c.1970-9G>A have lost the ability to walk.

It has been recently reported that a deep intronic variant c.930 + 189C>T inducing a pseudo-exon inclusion is a common pathogenic variant causing COL6-RD (Cummings et al., 2017; Bolduc et al., 2019). Indeed, this variant has been identified in up to 20% of patients with a suspected COL6-RD diagnosis and negative exonbased screening. Interestingly, this variant has been identified in only one patient in our series, even though the corresponding intronic region is now analyzed by our diagnostic gene panel. The frequency of the c.930 + 189C>T variant in COL6-RD patient series is thus variable depending on the population.

Patient with variants in two different COL6 genes

One patient (F24) in our series carried two missense variants in two different *COL6A* genes - COL6A1:c.1498G>A, p.(Gly500Arg) and COL6A3:c.7447A>G, p.(Lys2483Glu). The first variant has never been described and is absent from the general population. Even though this variant affects a glycine residue in one of the Gly-X-Y motifs of the triple helical domain, this variant is located more C-terminally, outside of the mutational hotspot (Butterfield et al., 2013; Lamandé and Bateman, 2018). We therefore classified this variant as variant of uncertain significance (VUS, see Supplementary Table S2 for details). Indeed, several other glycine substitutions in the proximity of p.(Gly500Arg) are classified as VUS in ClinVar. The second variant, COL6A3: c.7447A>G, p.Lys2483Glu is a well-known pathogenic variant that has only been observed in patients with recessive COL6-RD (Villar-Quiles et al., 2021). Indeed, it is present in trans with a pathogenic variant in another patient in our series (F12). Neither of the two variants identified is likely to cause disease by itself. Interestingly, another affected relative of F24 also carries both of these variants, suggesting a possible case of digenic inheritance. Further exploration of this family would be necessary in order to determine the contribution of these variants to the patient's phenotype. The digenic mode of inheritance has been previously described for a distal myopathy associated with a combination of one heterozygous variant in TIA1 gene and one in SQSTM1 gene (Niu et al., 2018). This mode of inheritance has also been suggested in a case report of a Bethlem myopathy patient carrying two putative de novo variants, one in COL6A1 and one in COL6A3 genes, though it is not clear if paternity and maternity were confirmed for the parental samples in that study (Choi et al., 2020). Exploration of the intronic regions is also necessary in order to rule out pathogenic variants that could be missed by the initial sequencing focused solely on exonic regions.

COL6-RD-like patient carrying a homozygous variant of uncertain significance

In addition to the genetically confirmed COL6-RD cases, we describe a patient with a clinical presentation compatible with COL6-RD, carrying a homozygous variant of uncertain significance (F23). This patient presented with hypotonia, club feet and congenital hip dislocation at birth. He was able to sit at 8 months of age, but never acquired the ability to walk. He had severe muscle weakness predominantly involving proximal muscles, marked hyperlaxity in distal joints as well as scoliosis. His respiratory function and serum CK were normal. In addition to the muscular presentation, the patient also developed epilepsy at the age of 11 and psoriasis at the age of 14 that are currently treated and stable. Genetic testing identified a homozygous variant of uncertain significance in COL6A1, NM_001848.3: c.2014G>A; p.(Glu672Lys). This variant changes a highly conserved residue in the C1 domain and is predicted to have a deleterious effect on protein function (CADD = 25.8; REVEL = 0.762). It is present in the general population (POPMAX filtering allele frequency is 0.0003288 in the gnomAD v2.1.1 database, 16/ 30,506 alleles in the South Asian population without any homozygous individuals). ClinVar database has an entry for this variant, classified as variant of uncertain significance. Both parents of the patient are asymptomatic heterozygous carriers of this variant.

Genetic tests used to diagnose COL6-RD patients

COL6-RDs represent a spectrum of muscular disorders from severe and more clinically recognizable Ullrich congenital muscular dystrophy (UCMD) to much milder Bethlem myopathy (BM) that can present as limb girdle muscular dystrophy (Foley et al., 2021; Natera-de Benito et al., 2021). It is for this reason that the COL6A1-3 genes have been included on three different gene panels according to the French National Consensus Gene lists: "Limb Girdle Muscular Dystrophies (LGMD)" (Exhaustive gene list of 40 genes), "Congenital Muscular Dystrophies-except alphadystroglycanopathies" (Exhaustive gene list of 17 genes) and "Retractile Myopathies" (Unique exhaustive gene list of 29 genes) (Krahn et al., 2019). All four patients in our series that were screened by LGMD gene panel were diagnosed with Bethlem myopathy (Supplementary Table S1), while six Ulrich patients were diagnosed by "Retractile Myopathy" gene panel (patients F3, F13), "Congenital Muscular Dystrophies-except alpha-dystroglycanopathies" gene panel (F7, F17, F23) and by targeted Sanger sequencing (F15.1). These results show the importance of including COL6A1-3 genes on gene panels that cover the full phenotypic spectrum of COL6-RDs. Analyzing COL6A1-3 genes is especially important since COL6-RDs represent a significant part of genetic muscular disorders. In one large cohort, pathogenic variants in COL6A1-3 genes were found to responsible for the phenotype of 97 out of 440 neuromuscular patients with confirmed genetic diagnosis (Sframeli et al., 2017). The diagnosis of COL6-RD was also established in 4% (55/1,259) and 8% (39/468) of patients with genetically confirmed LGMD (Nallamilli et al., 2018; Töpf et al., 2020).

Conclusion

In this study we present phenotypic and genetic data from 25 patients (22 families) with genetically confirmed COL6-RD that are followed at the University Hospital of Marseille (AP-HM). This large series of rare-disease patients provides key information on the frequency of pathogenic variants in *COL6A* genes in Southern France. The phenotypic data from our study also show that a certain subgroup of COL6-RD patients can present with a rapidly accelerating clinical presentation. Our results thus contribute to the genetic and phenotypic description of COL6-RD that could be useful for design of future clinical trials.

Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article/Supplementary Material.

Ethics statement

The study was registered by AP-HM (Assistance Publique, Hôpitaux de Marseille) under number GBP3T5/PADS22-157,

evaluating this study as exempt from the Institutional Review Board according to the French legislation for the studies on humans. The studies were conducted in accordance with the local legislation and institutional requirements. Written informed consent for participation was not required from the participants or the participants' legal guardians/next of kin in accordance with the national legislation and institutional requirements. The human samples used in this study were acquired from a by-product of routine care or industry.

Author contributions

VM participated in data collection, data analysis and manuscript writing, FA participated in data collection and analysis; FA, AV, SA, KN, ES-C, MK, BC, and SG followed the patients and/or were involved in identifying the genetic diagnosis as well as revised the manuscript for content; CT revised the manuscript for content; BC and SG had major roles in the conception of the study; SG had a major role in data analysis and manuscript writing. All authors contributed to the article and approved the submitted version.

Acknowledgments

We thank the patients, families and health professionals involved in patient care and diagnosis for making this study possible. We also thank Dr. Sonia Dubois for the help in data

References

Bolduc, V., Foley, A. R., Solomon-Degefa, H., Sarathy, A., Donkervoort, S., Hu, Y., et al. (2019). A recurrent COL6A1 pseudoexon insertion causes muscular dystrophy and is effectively targeted by splice-correction therapies. *JCI Insight* 4 (6), e124403. Article 6. doi:10.1172/jci.insight.124403

Bönnemann, C. G. (2011). The collagen VI-related myopathies: muscle meets its matrix. *Nat. Rev. Neurol.* 7 (7), 379–390. Article 7. doi:10.1038/nrneurol.2011.81

Briñas, L., Richard, P., Quijano-Roy, S., Gartioux, C., Ledeuil, C., Lacène, E., et al. (2010). Early onset collagen VI myopathies: genetic and clinical correlations. *Ann. Neurology* 68 (4), 511–520. doi:10.1002/ana.22087

Butterfield, R. J., Foley, A. R., Dastgir, J., Asman, S., Dunn, D. M., Zou, Y., et al. (2013). Position of glycine substitutions in the triple helix of COL6A1, COL6A2, and COL6A3 is correlated with severity and mode of inheritance in collagen VI myopathies. *Hum. Mutat.* 34 (11), 1558–1567. doi:10.1002/humu.22429

Choi, E., Shin, S., Lee, S., Lee, S. J., and Park, J. (2020). Coexistence of digenic mutations in the collagen VI genes (COL6A1 and COL6A3) leads to Bethlem myopathy. *Clin. Chimica Acta; Int. J. Clin. Chem.* 508, 28–32. doi:10.1016/j.cca.2020.05.011

Cummings, B. B., Marshall, J. L., Tukiainen, T., Lek, M., Donkervoort, S., Foley, A. R., et al. (2017). Improving genetic diagnosis in Mendelian disease with transcriptome sequencing. *Sci. Transl. Med.* 9 (386), eaal5209. Article 386. doi:10.1126/scitranslmed. aal5209

Deconinck, N., Dion, E., Ben Yaou, R., Ferreiro, A., Eymard, B., Briñas, L., et al. (2010). Differentiating Emery-Dreifuss muscular dystrophy and collagen VI-related myopathies using a specific CT scanner pattern. *Neuromuscul. Disord. NMD* 20 (8), 517–523. doi:10.1016/j.nmd.2010.04.009

Deconinck, N., Richard, P., Allamand, V., Behin, A., Lafôret, P., Ferreiro, A., et al. (2015). Bethlem myopathy: long-term follow-up identifies COL6 mutations predicting severe clinical evolution. *J. Neurology, Neurosurg. Psychiatry* 86 (12), 1337–1346. doi:10. 1136/jnnp-2013-307245

Foley, A. R., Hu, Y., Zou, Y., Yang, M., Medne, L., Leach, M., et al. (2011). Large genomic deletions: a novel cause of Ullrich congenital muscular dystrophy. *Ann. Neurology* 69 (1), 206–211. doi:10.1002/ana.22283

Foley, A. R., Mohassel, P., Donkervoort, S., Bolduc, V., and Bönnemann, C. G. (2021). "Collagen VI-related dystrophies," in *GeneReviews* (Seattle: University of Washington).

Gonzalez-Quereda, L., Rodriguez, M. J., Diaz-Manera, J., Alonso-Perez, J., Gallardo, E., Nascimento, A., et al. (2020). Targeted next-generation sequencing in a large cohort

collection as well as Prof. Carsten Bönnemann and Dr. Reghan Foley for helpful discussions.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fgene.2023.1242277/ full#supplementary-material

of genetically undiagnosed patients with neuromuscular disorders in Spain. *Genes* 11 (5), 539. doi:10.3390/genes11050539

Krahn, M., Biancalana, V., Cerino, M., Perrin, A., Michel-Calemard, L., Nectoux, J., et al. (2019). A National French consensus on gene lists for the diagnosis of myopathies using next-generation sequencing. *Eur. J. Hum. Genet.* 27 (3), 349–352. Article 3. doi:10. 1038/s41431-018-0305-1

Kwong, A. K., Zhang, Y., Ho, R. S., Gao, Y., Ling, X., Tsang, M. H., et al. (2023). Collagen VI-related myopathies: clinical variability, phenotype-genotype correlation and exploratory transcriptome study. *Neuromuscul. Disord. NMD* 33 (5), 371–381. doi:10.1016/j.nmd.2023.03.003

Lamandé, S. R., and Bateman, J. F. (2018). Collagen VI disorders: insights on form and function in the extracellular matrix and beyond. *Matrix Biol. J. Int. Soc. Matrix Biol.* 71 (72), 348–367. doi:10.1016/j.matbio.2017.12.008

Lucioli, S., Giusti, B., Mercuri, E., Vanegas, O. C., Lucarini, L., Pietroni, V., et al. (2005). Detection of common and private mutations in the COL6A1 gene of patients with Bethlem myopathy. *Neurology* 64 (11), 1931–1937. doi:10.1212/01.WNL. 0000163990.00057.66

Martoni, E., Urciuolo, A., Sabatelli, P., Fabris, M., Bovolenta, M., Neri, M., et al. (2009). Identification and characterization of novel collagen VI non-canonical splicing mutations causing Ullrich congenital muscular dystrophy. *Hum. Mutat.* 30 (5), E662–E672. Article 5. doi:10.1002/humu.21022

Mercuri, E., Lampe, A., Allsop, J., Knight, R., Pane, M., Kinali, M., et al. (2005). Muscle MRI in Ullrich congenital muscular dystrophy and Bethlem myopathy. *Neuromuscul. Disord.* 15 (4), 303–310. Article 4. doi:10.1016/j.nmd.2005.01.004

Nallamilli, B. R. R., Chakravorty, S., Kesari, A., Tanner, A., Ankala, A., Schneider, T., et al. (2018). Genetic landscape and novel disease mechanisms from a large LGMD cohort of 4656 patients. *Ann. Clin. Transl. Neurology* 5 (12), 1574–1587. doi:10.1002/acn3.649

Natera-de Benito, D., Foley, A. R., Domínguez-González, C., Ortez, C., Jain, M., Mebrahtu, A., et al. (2021). Association of initial maximal motor ability with long-term functional outcome in patients with COL6-related dystrophies. *Neurology* 96 (10), e1413–e1424. Article 10. doi:10.1212/WNL.000000000011499

Niu, Z., Pontifex, C. S., Berini, S., Hamilton, L. E., Naddaf, E., Wieben, E., et al. (2018). Myopathy with SQSTM1 and TIA1 variants: clinical and pathological features. *Front. Neurology* 9, 147. doi:10.3389/fneur.2018.00147 Pace, R. A., Peat, R. A., Baker, N. L., Zamurs, L., Mörgelin, M., Irving, M., et al. (2008). Collagen VI glycine mutations: perturbed assembly and a spectrum of clinical severity. *Ann. Neurology* 64 (3), 294–303. doi:10.1002/ana.21439

Quijano-Roy, S., Khirani, S., Colella, M., Ramirez, A., Aloui, S., Wehbi, S., et al. (2014). Diaphragmatic dysfunction in Collagen VI myopathies. *Neuromuscul. Disord. NMD* 24 (2), 125–133. doi:10.1016/j.nmd.2013.11.002

Sframeli, M., Sarkozy, A., Bertoli, M., Astrea, G., Hudson, J., Scoto, M., et al. (2017). Congenital muscular dystrophies in the UK population: clinical and molecular spectrum of a large cohort diagnosed over a 12-year period. *Neuromuscul. Disord. NMD* 27 (9), 793–803. Article 9. doi:10.1016/j.nmd.2017.06.008

Stehlíková, K., Skálová, D., Zídková, J., Haberlová, J., Voháňka, S., Mazanec, R., et al. (2017). Muscular dystrophies and myopathies: the spectrum of mutated genes in the Czech Republic. *Clin. Genet.* 91 (3), 463–469. doi:10.1111/cge.12839

Töpf, A., Johnson, K., Bates, A., Phillips, L., Chao, K. R., England, E. M., et al. (2020). Sequential targeted exome sequencing of 1001 patients affected by unexplained limbgirdle weakness. *Genet. Med. Official J. Am. Coll. Med. Genet.* 22 (9), 1478–1488. doi:10. 1038/s41436-020-0840-3

Villar-Quiles, R. N., Donkervoort, S., de Becdelièvre, A., Gartioux, C., Jobic, V., Foley, A. R., et al. (2021). Clinical and molecular spectrum associated with COL6A3 c.7447A>G p.(Lys2483Glu) variant: elucidating its role in collagen VI-related myopathies. *J. Neuromuscul. Dis.* 8 (4), 633–645. doi:10.3233/JND-200577

Zanoteli, E., Soares, P. S., Silva, A. M. S. da, Camelo, C. G., Fonseca, A. T. Q. S. M., Albuquerque, M. A. V., et al. (2020). Clinical features of collagen VI-related dystrophies: a large Brazilian cohort. *Clin. Neurology Neurosurg.* 192, 105734. doi:10.1016/j.clineuro. 2020.105734