Check for updates

OPEN ACCESS

EDITED BY Matthew E. R. Butchbach, Nemours Children's Health Delaware, United States

REVIEWED BY

Bruce Mao Zheng Wang, University of California, San Francisco, United States Pauline Harper, Karolinska University Laboratory, Sweden

*CORRESPONDENCE Joelle N. Chabwine, joelle.chabwine@unifr.ch

SPECIALTY SECTION This article was submitted to Genetics of Common and Rare Diseases, a section of the journal Frontiers in Genetics

RECEIVED 13 July 2022 ACCEPTED 12 October 2022 PUBLISHED 31 October 2022

CITATION

Collaud E, Wittwer L, Minder A-E, Annoni J-M, Minder El and Chabwine JN (2022), Case Report: Variegate porphyria disclosed by postgastric bypass complications and causing predominant painful sensorimotor axonal peripheral neuropathy. *Front. Genet.* 13:993453. doi: 10.3389/fgene.2022.993453

COPYRIGHT

© 2022 Collaud, Wittwer, Minder, Annoni, Minder and Chabwine. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Case Report: Variegate porphyria disclosed by post-gastric bypass complications and causing predominant painful sensorimotor axonal peripheral neuropathy

Edwige Collaud¹, Luis Wittwer^{1,2}, Anna-Elisabeth Minder³, Jean-Marie Annoni⁴, Elisabeth I. Minder³ and Joelle N. Chabwine^{1,4}*

¹Division of Neurorehabilitation, Fribourg Hospital Meyriez/Murten, Fribourg, Switzerland, ²Division of Medicine, Emmental Hospital, Burgdorf, Switzerland, ³Division of Endocrinology, Diabetology, Porphyria, Triemli Hospital, Zurich, Switzerland, ⁴Neurology Unit, Laboratory for Cognitive and Neurological Science, Department of Neuroscience and Movement Science, Faculty of Science and Medicine, University of Fribourg, Fribourg, Switzerland

Background and aims: Porphyrias constitute a group of rare genetic diseases due to various, mostly autosomal dominant mutations, causing enzymatic deficiency in heme biosynthesis. As a result, neurotoxic porphyrin precursors and light-sensitive porphyrins accumulate, while dysfunction in their targets determines the disease symptoms. Variegate porphyria (VP), one of the acute hepatic porphyrias, is caused by a protoporphyrinogen oxidase (PPOX) mutation. During acute attacks, among other factors, triggered by drugs, stressors, or fasting, an increase in urinary and fecal porphobilinogen (PBG), aminolevulinic acid (ALA), and porphyrins occurs, damaging the autonomous, peripheral, or central nervous system. The disease remains often latent or displays minimal symptoms usually overlooked, exposing undiagnosed patients to potentially serious complications in the presence of the aforementioned triggers.

Case report: This 46-year-old woman presented, some days after a bariatric surgery, with severe flaccid tetraparesis and neuropathic pain, initially misdiagnosed as a functional neurological disorder. The severe axonal sensorimotor polyneuropathy led to further investigations, disclosing high urinary porphobilinogen, ALA, and porphyrin levels due to a new PPOX mutation. Retrospectively, it appeared that the patient had had typical VP symptoms (abdominal pain, fragile skin, and dark urine episodes) for years prior to the surgery. Treated with carbohydrate load, neurorehabilitation, and

Abbreviations: AIP, acute intermittent porphyria; ALA, aminolevulinic acid; BSR, blood sedimentation rate; CMV, cytomegalovirus; CSF, cerebrospinal fluid; EBV, Epstein-Barr virus; HIV, human immunodeficiency virus; PBG, porphobilinogen; PCR, polymerase chain reaction; PPOX, protoporphyrinogen oxidase; VP, porphyria variegate.

analgesics, she slowly recovered to full mobility, with partial autonomy in her daily life activities, although fatigue and severe pain persisted, preventing her from returning to work.

Conclusion: This case documents gastric bypass surgery as a trigger of severe VP invalidating neurological symptoms and illustrates how the delayed diagnosis and post-interventional complications could have been prevented by screening for porphyria cardinal symptoms prior to the intervention. Likewise, this cost-effective screening should be performed before any treatment influencing the diet, which would dramatically improve the porphyria diagnosis rate and outcome.

KEYWORDS

variegate porphyria, pain, polyneuropathy, gastric bypass, bariatric surgery, PPOX mutation

Background

Porphyrias are a group of rare genetic diseases, caused by a deficiency in one of the eight enzymes of heme biosynthesis, which results in the accumulation of porphyrin precursors (Kauppinen, 2005). In autosomal dominant acute porphyrias, porphyrin precursors, in particular aminolevulinic acid (ALA), act toxic to the central, peripheral, and autonomic nervous system, causing acute abdominal pain and neuropathy in the context of an acute porphyria attack (Phillips, 2019). In addition to the most prevalent form of acute porphyria, the acute intermittent porphyria (AIP) variegate porphyria (VP) constitutes an autosomal dominant inherited form of porphyria caused by a mutation in the protoporphyrinogen oxidase (PPOX) gene (Tabaro et al., 2018). The latter results in the accumulation of toxic porphyrin precursors due to enzyme deficiency, especially provoking acute neurovisceral and cutaneous symptoms. The exact prevalence of VP mutations is unknown due to their low penetrance and most probably because porphyria symptoms are often overlooked (Chen et al., 2016).

Acute porphyria attacks consist of acute neurovisceral pain without peritonism, associated with pre-ileus, constipation, and symptoms of dysautonomia (tachycardia, hypertension, nausea, and vomiting). Hyponatremia, sensorimotor polyneuropathic symptoms, and red or dark urine episodes can also occur (Sassa, 2006). The acute porphyria attack is confirmed by the elevation of porphyrin precursor excretion in the urine, namely, porphobilinogen (PBG) and ALA (Pischik and Kauppinen, 2015). The most frequent triggers of a porphyria attack are various drugs, surgical procedures, fasting, smoking, and physical or psychological stressors (Sassa, 2006; Zürich and Endokrinologie, 2018). Severe porphyria attacks can be lifethreatening or result in irreversible damage, with persistent disability due to neurological deficits (Sassa, 2006). The treatment of acute porphyria attacks consists in reducing the accumulation of toxic compounds by intravenous infusion of glucose and heme inhibiting D-ALA-synthase 1. This treatment consistently prevents persistent disabilities, while eviction of triggering factors allows for avoiding porphyria attacks (Pischik and Kauppinen, 2015). Thus, porphyria can be adequately managed through well-established guidelines, under the condition that it is diagnosed.

We report in this study a patient who abruptly developed a painful flaccid tetraparesis, following gastric bypass surgery. The initial wrong diagnosis of a functional neurological disorder delayed the diagnosis of VP for several months, condemning the patient to long-term neurological complications.

Case report

This 46-year-old woman known for obesity class III (BMI 41.7kg/m²) underwent a gastric bypass surgery 2 months before admission to the rehabilitation clinic. Her medical history was otherwise remarkable for depression in a burn-out context 4 years earlier, smoking (25 p.y.), and non-specific chronic low back pain. Immediately after the gastric bypass, she displayed an abdominal wall hernia and developed severe complications, interpreted as anastomosis insufficiency, gastric ulcer, or wound abscess, leading to new operations and a prolonged hospital stay. Two weeks after discharge, she was re-admitted to the emergency department for an acute onset tetraparesis. A first diagnostic workup including a normal spinal cord MRI was inconclusive, and with respect to her prior history of depression, the tetraparesis was labeled as a functional neurological disorder. Subsequently, the patient was referred to our hospital for neurorehabilitation.

On arrival, she complained of severe fatigue, bilateral limb weakness, hypoesthesia, and pain, dysgeusia for all tastes, and dysphagia. A neurological examination disclosed bilateral facial peripheral paresis, proximally pronounced severe flaccid tetraparesis with tendon hyporeflexia, bilateral touch and pick hypoesthesia in the trigeminal area, and C1 dermatomes, as well as a glove and stocking distal hypoesthesia. The sensory

TABLE 1 Investigations over time.

Date	Investigation	Result	
Initial eva	aluations and investigations		
22.04.18	Total column MRI	Normal	
09.05.18	Test to acetylcholinesterase inhibitors (pyridostigmine 60 mg)	Negative	
11.05.18	ENMG	Signs of axonal neuropathy in the four limbs, with acute and chronic components; secondary muscular abnormalities not excluded	
15.05.18	Brain MRI	Normal	
16.05.18	CSF examinations	Normal white cell count, normal protein, normal immunologic electrophoresis, and absence of production of oligoclonal bands	
16.05.18	Blood examinations	Positive DOT for antineuronal antibodies (anti-amphiphysin)/elevated BSR (50 mm/h)/light increase of the C3 complement (1.43 g/L)/increased monocyte rate (8.9%)	
		Negative DOT for screening of autoimmune diseases (including myasthenia gravis antibodies)	
		Negative screening for the most frequent neuro-infectious diseases (syphilis, HIV, Lyme, CMV, and EBV)	
		Normal vitamins B1, B6, B12, B9, D, E, and A, but hypovitaminosis D	
		Absence of metabolic diseases (normal HbA1c and TSH) and normal metal dosage (copper and zinc)	
		Increased urinary porphyrins and porphyrin precursors	
01.07.18	Neuropsychological evaluation	Normal to slightly diminished performance in the executive functions and working memory	
Second-li	ne/follow-up evaluations and investigations		
10.07.18	Laboratory examinations	Negative DOT antineuronal antibody in the serum and in the CSF/mild BSR increase (13 mm/h)/normal C3 complement, CRP, and monocytes	
12.07.18	ENMG (evolution 1)	Stable (no change from the initial testing)	
17.07.18	Genetic testing (PPOX gene mutation)	Mutation of protoporphyrinogen oxidase (exon 4 of the PPOX gene)	
17.09.19	ENMG (evolution 2)	Improvement of the nerve conduction velocity of the right peroneus nerve, focal myelopathy of the ri median nerve suggesting a carpal tunnel syndrome, and late spinal response but not prolonged right til median nerves	
26.11.19	MOCA score	25/30	
26.11.19	Neuropsychological evaluation	Normal executive functions and working memory, some attention difficulties (treatment velocity) attributed to the newly diagnosed genetic disease and chronic pain; no other cognitive dysfunction	

Important positive results are written in bold.

abnormalities fluctuated between repeated examinations. The neuropsychological evaluation revealed no cognitive impairment, while the patient displayed anxiety and depression symptoms in response to her severe disability and the general health situation.

Diagnostic procedures

A new MRI confirmed normal findings in the brain and the spinal cord, as well as in the proximal portion of spinal nerve roots. The peripheral pattern of neurological symptoms warranted electroneuromyography, which confirmed axonal sensorimotor neuropathy involving all four limbs. A broad etiological search (Table 1, supplementary material) ruled out infectious diseases (normal serologies for syphilis, human immunodeficiency virus (HIV), cytomegalovirus (CMV), Epstein-Barr virus (EBV), and

Borrelia burgdorferi, as well as the negative cerebrospinal fluid (CSF) polymerase chain reaction (PCR) for CMV and EBV), endocrine disorders (diabetes and thyroid dysfunction), and Wilson's disease (serum and urine copper measurements). No relevant vitamin deficiency explaining the neurological findings was detected, except for low levels of vitamin D [22 nmol/l] and folate [3.4 nmol/l]) (Punchai et al., 2017). There were signs of systemic inflammation with an increased blood erythrocyte sedimentation rate (BSR) [50 mm/h], elevated serum C3c, and 8.9% of monocytes in the white blood count. Local production of oligoclonal bands in the CSF was excluded. Autoimmune disease screening identified an anti-amphiphysin antibody in the serum (no titer mentioned).

In the meantime, second-line complementary results disclosed high porphyrin precursors and porphyrin levels in the urine, with a positive plasmafluorescence scan [626 nm], as well as elevated coproporphyrin I [46.3 nmol/g] and III

TABLE 2 Value of porphyrins over time with references.

	Measured porphyrin(s)	[Urinary porphyrins/ creatinine] Reference values ^a		[Porphyrins/24h] ^b		[Feces] ^c	
28.05.18	Total porphyrins	902	[<30]	4606	[<310]		
	ALA	18.7	[<2.5]				
	Porphobilinogen	2.63	[<1.25]				
	Uroporphyrin	35.7	[<4.6]				
	Heptacarboxyporphyrin	48.7	[<0.8]				
	Hexacarboxyporphyrin	29.1	[<2.2]				
	Pentacarboxyporphyrin	145	[<1.6]				
	Coproporphyrin I	30.7	[<5.8]				
	Coproporphyrin III	553	[<21]				
05.06.18	PBG-deaminase			136	[9]		
	Plasmafluorescence scan			Peak maximum at 620	6 nm		
	Coproporphyrin I					46.3	[<20]
	Coproporphyrin III					350	[<12]
	Protoporphyrin					1444	[<80]
20.06.18	Total porphyrins	126.6	[<30]	884	[<310]		
	ALA	5.3	[<2.5]	37.1	[<50]		
	Porphobilinogen	1.12	[<1.25]	7.8	[<7.5]		
	Uroporphyrin	29.1	[<4.6]	203.3	[<60]		
	Heptacarboxyporphyrin	8.9	[<0.8]	61.9	[<10]		
	Hexacarboxyporphyrin	1.5	[<2.2]	10.5	[<21]		
	Pentacarboxyporphyrin	7.2	[<1.6]	50.4	[<16]		
	Coproporphyrin I	9.8	[<5.8]	68.2	[<100]		
	Coproporphyrin III	70.2	[<21]	430	[<210]		
28.09.18	Total porphyrins	60	[<30]				
	ALA	6.9	[<2.5]				
	Porphobilinogen	0.52	[<1.25]				
	Uroporphyrin	18.2	[<4.6]				
	Heptacarboxyporphyrin	5.3	[<0.8]				
	Hexacarboxyporphyrin	0.7	[<2.2]				
	Pentacarboxyporphyrin	7.2	[<1.6]				
	Coproporphyrin I	6.6	[<5.8]				
	Coproporphyrin III	42	[<21]				
16.01.19	Total porphyrins	133	[<30]				
	ALA	7.3	[<2.5]				
	Porphobilinogen	5	[<1.25]				
	Uroporphyrin	24.6	[<4.6]				
	Heptacarboxyporphyrin	5.8	[<0.8]				
	Hexacarboxyporphyrin	1.3	[<2.2]				
	Pentacarboxyporphyrin	9.1	[<1.6]				
	Coproporphyrin I	5.8	[<5.8]				
	Coproporphyrin III	86	[<21]				
09.03.19	Total porphyrins	189	[<30]				
	ALA	8,3	[<2.5]				
	Porphobilinogen	0.63	[<1.25]				
	Uroporphyrin	30.6	[<4.6]				

(Continued on following page)

TABLE 2 (Continued) Value of porphyrins over time with references.

	Measured porphyrin(s) Heptacarboxyporphyrin	[Urinary porphyrins/ creatinine] Reference valuesª		[Porphyrins/24h] ^b	[Feces] ^c	
		8.3	[<0.8]			
	Hexacarboxyporphyrin	2.7	[<2.2]			
	Pentacarboxyporphyrin	19.2	[<1.6]			
	Coproporphyrin I	10.6	[<5.8]			
	Coproporphyrin III	117	[<21]			
17.06.19	Porphobilinogen	2.49	[<1.25]			
12.12.19	ALA	8.3	[<2.5]			
	Porphobilinogen	3.75	[<1.25]			

^aAminolevulinic acid, porphobilinogen in µmol/mmol, and urinary porphyrins in µmol/mol. ^bAminolevulinic acid, porphobilinogen in µmol/24h, and urinary porphyrins in nmol/24h. ^cIn nmol/g dry weight.

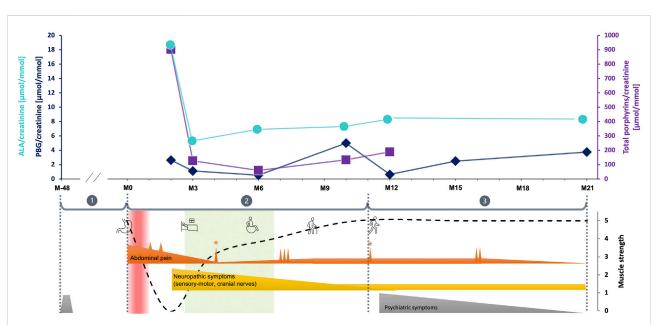


FIGURE 1

The evolution of clinical symptoms and urinary porphyrins through the follow up. The vertical red band labels the acute abdominal pain period supposedly corresponding the acute porphyria attack, while the light green is related to the stationary neurorehabilitation. The numbers within the grey circles represent the key observation periods corresponding the main evolutionary/intervention steps, as following (M month, the digit following M represents the number of months before or after the gastric bypass procedure): ① [M-48 until 01.2018 (M0)] is the retrospective period; ② [from M0 to M11 (01.2019)] corresponds to the period during which the main and most severe symptoms as well as therapeutic interventions in hospitals occurred; ③ [from M11 until the end of the follow up at M21 (12.2019)] represents the post-stationary evolution. Urinary porphyrins concentrations are labelled as following: turquoise blue dots for ALA, dark blue rhombs for PBG and violet squares for total porphyrins. Main clinical symptoms evolution over the observation period. Ordinate values are arbitrary units corresponding to a clinical estimate of symptom severity. The main symptoms were grouped in three categories represented by three different color codes: abdominal pain in orange, peripheral neuropathic signs in yellow and psychiatric symptoms in grey. The functional performance and main interventions are shown by illustrative cartoons: the stomach for bariatric surgery, the bed for the period when the patient was bed-ridden, the wheelchair and walking stick and free walking showing the improving patterns of mobility. (1) burn-out 2014 (grey pyramid); (2) acute abdominal pain with maximal intensity during the period considered to correspond to the porphyria attack, with small peaks occurring at surgical interventions (small orange spines above the baseline). The pain intensity decreased as rehabilitation and other treatments were initiated, and stabilized at very low level although 3 repetitive peaks took place at M7-M8. Of notice, two isolated peaks were recorded during appendagitis exacerbation (with * above) at M4 and M11. Neuropathic symptoms started later than abdominal pain as characteristic for porphyria. They were at their maximum from the beginning, but progressively decreased during neurorehabilitation program parallel to the overall clinical state of the patient. They finally dropped to a low, but persistent steady-state level; ③ during this post-stationary period, both abdominal pain and neuropathic symptoms remained low (except two exacerbations of abdominal pain at M16). Psychiatric symptoms including sleep disorders, fatigue, anxiety and depression reappeared, but progressively improved with appropriate medication.

TABLE 3 Symptoms and clinical signs over time.

Date	Subjective symptom	Objective sign	
12.02.18 (M0)	Brutal abdominal pain	Mimicry acute abdomen	
05.04.18	Diminished general state, asthenia, inappetence, and abdominal pain with belt-knife knock	Weak patellar reflexes, Mingazzini hardly testable, loss of superficial sensitivity discrimination to « pick and touch» and proprioception, walking	
	Pelvis paresthesia, fecal incontinence, hematuric urine, and weakness of the low limbs from distal to proximal and then upper limbs from proximal to distal	becoming impossible, sphincter tone diminished, impossibility to whistle, and suboptimal cheek blowing	
30.04.18	Bilateral loss of face sensitivity	Proximal predominant tetraparesis	
	Dysphagia	Weak reflexes	
	Weakness and pain in limbs	Hypoesthesia without a precise dermatome DD trigeminal nerve and C1 region	
	Balance and body scheme disorder (probably due to deconditioning)	Hypoesthesia to pick and touch the distal upper and lower limbs	
		Simpson's test: seems positive	
		Pigmented skin lesion behind the hands	
20.06.18 (M4)	Inconclusive abdominal pain		
28.09.18	Abdominal pain independently from eating 3-4 times a week	Strength and sensitivity improvement	
(M7)	Bad healing, skin fragility (behind the hands and face) and face hirsutismus		
16.01.19 (M11)	Low belly pain with bloating, neuropathic pain of lower limbs and body, less general strength, fatigue, skin fragility, appendagitis, and neurologic recovery	Walking without help inside and with a walking stick outside	
18.02.19	Neuropathic pain, insomnia, anxiety (due to pain and hospitalization's context), and tremor		
23.05.20	Electric discharge in all limbs and face and sensation recovery until elbows and knees	More expressive facial gestures and complete eye closing	
17.06.19 (M16)	Abdominal pain for 3 h every 2 days, especially in the evening, and sometimes stomach burning	Strength of lower limbs is still reduced but better than last time	
	Reduction of sensation and strength		
	Neuropathic pain waking up at night		
01.07.19		Sensory affect (superficial, paresthesia, and deep pain) and motor impairment of 4 limbs predominant to the LL and diminished impairment of bilateral cranial nerves V and VII	
17.09.19	Electric discharge in all limbs and muscle fatigability	Bilateral facial paresis predominant to the right side, suboptimal whistle and cheek blowing, bilateral facial paresthesia (predominant right), weak Souques' sign right, weak reflexes of 4 limbs, pallesthesia 6/8 at fingers, 5/8 at toes, and light trunk instability at Romberg testing	
29.11.19	Stability of pain and sensitivity (UL: elbows; LL: feet), without exacerbation, and insomnia	Stable neurologic state and slightly clinical improving	
12.12.19 (M21)	Total body electric pain, in the evening, caused by physical activity, fatigue, walking for 200–300 m, and sleep disorder		

[350 nmol/g], and protoporphyrin [1444 nmol/g]. Altogether, these findings, in addition to the above-detailed clinical picture, confirmed the diagnosis of VP (Hift et al., 2004) (Table 2, supplementary material). Moreover, the patient exhibited a heterozygous deletion in exon 4 of the PPOX gene (Table 1, supplementary material). To the best of our knowledge, this mutation has never been reported before.

When further retrospectively questioning the patient and more accurately examining her, it appeared that she had thin and fragile skin (prevailing as far back as she could remember in her past), however, without the typical blistering seen in VP. In addition, she repeatedly observed that her urines were dark-colored. Otherwise, she was treated for irritable bowel syndrome (metixen, pepsine, dimeticon, glutaminic acid hydrochloride, pancreatin, cellulose, and dehydrocholic acid) prior to the bariatric surgery. Her family history was unremarkable for diagnosed porphyria or evoking symptoms.

Management and follow-up

Due to the positive serum anti-amphiphysin antibody, an autoimmune origin of polyneuropathy was initially suspected, indicating an oral steroid therapy course for few weeks. The patient's mobility, dysphagia, and sensory complaints progressively improved, without further aggravation at the end of the treatment, while the anti-amphiphysin antibody and inflammation signs disappeared in the serum.

After the diagnosis of VP, the therapeutic approach was to the increase carbohydrate intake to a minimum of 250 g/day, as well as the eviction of all porphyrinogenic drugs (such as fentanyl) and other potential acute attack-triggering factors. Because the patient was not considered to be suffering an acute attack, no heme arginate treatment was applied. An intensive multidisciplinary neurorehabilitation regime and optimal symptom treatments allowed the patient to progressively gain her mobility back, with the improvement of her muscle strength, pain, somatosensory complaints, dysphagia, and dysgeusia (Figure 1). Neuropathic pain and other sensory complaints remained moderately stable despite consistent analgesic therapy (fentanyl patch replaced by tapentadol, pregabalin, and amitriptyline), while mood disorders and insomnia improved, thanks to mirtazapine. However, she pain developed acute abdominal during her neurorehabilitation stay due to appendagitis, efficiently treated by appropriate analgesia.

She was discharged after 6 weeks of stationary neurorehabilitation and continued the ambulant physical and occupational therapy, with further functional improvement (Table 3, supplementary material). After the surgical cure of abdominal hernia and appendagitis healing, abdominal pain disappeared. However, severe fatigue persisted in addition to limb neuropathic pain and preexisting low back pain, preventing the patient from returning to work. Nevertheless, she was able to care for her children and perform her daily household tasks. Family genetic counseling was carried out as proposed in porphyria guidelines (Anderson et al., 2005).

Discussion

A few weeks after undergoing a gastric bypass procedure with several postoperative complications, this middle-aged patient developed an abrupt proximal-predominant areflexia and flaccid tetraparesis associated with sensory symptoms (including neuropathic pain) involving the four limbs. After a thorough and stepwise workup excluding etiologies, her neurological symptoms were most likely caused by VP, based on typical abnormalities of urinary and fecal biomarkers (Di Pierro et al., 2021) and the finding of a new heterozygous mutation in the PPOX gene of pathological potential never reported before. Indeed, being a deletion, the observed mutation caused a frameshift and a truncated transcript most likely leading to the synthesis of a dysfunctional enzyme and subsequent toxic porphyrin precursors (Barbaro et al., 2013). The unexpected discrepancy between urinary ALA and PBG increases could be explained by their biological variations (Floderus et al., 2006) and possible differences in assay specificities (Agarwal et al., 2021), while positive plasma fluorescence and the ratio coproporphyrin III: I of ~8 further confirmed the diagnosis of porphyria. Despite the high prevalence reported in South Africa, VP is relatively frequent in Switzerland due to founder mutations (Schneider-Yin and Minder, 2006). Whether the patient was still undergoing a porphyria attack at the diagnosis time is an interesting question. The modest increase of urinary ALA alone (~7 times the reference values) and the absence of dark-colored urines were against this hypothesis. However, taking into account the decreasing kinetics of urinary markers and further stabilization thereafter, the patient's early postoperative abdominal pain episode and acute onset of sensorimotor deficits might have been caused by an acute porphyria attack, the latter being triggered by the bariatric surgery procedure.

The role of gastric bypass in triggering (or exacerbating latent) VP symptoms is also worthy of discussion. In fact, bariatric surgery encompasses several porphyria-precipitating factors (Anderson et al., 2005). First, exposure to various potentially porphyrinogenic drugs during anesthesia and the postoperative course are considered. Fentanyl prescription and its long maintenance should have deserved great caution in this patient. Not only this opiate is listed among porphyrinogenic drugs (Zürich and Endokrinologie, 2018; N APOS, 2007; Koffman et al., 2006) and could have, as such, contributed to the occurrence of porphyria symptoms but also its chronic use can induce hyperalgesia, aggravating instead of treating the patient's pain (Roeckel et al., 2016). Additionally, the combination of physical and psychological stress and a long-lasting reduction of caloric (especially carbohydrate) intake can be listed. Furthermore, in individuals with a porphyria mutation, the desired weight loss and subsequent impairment of energy metabolism in the liver trigger the metabolic cascade that results in the accumulation of neurotoxic porphyrin precursors (Danion et al., 2012). At last, the gastric bypass causes hormonal dysfunction, including insulin dysregulation (Koffman et al., 2006; Kumar, 2007; Abdelaal et al., 2017), adding up to homeostasis disruptions. Thus, the gastric bypass context cumulates risks of precipitating a latent (or poorly symptomatic) porphyria to become highly symptomatic, especially if we consider the low penetrance of related mutations (Chen et al., 2016). On the other hand, the few reports in the recent literature probably do not reflect the real (higher) incidence of such cases, of which careful documentation and reporting should raise awareness among professionals and avoid unnecessary diagnosis delay, as in the case reported in this study.

The presence of anti-amphiphysin antibodies initially questioned an inflammatory background to the axonal polyneuropathy (Dubey et al., 2019) and/or a link with the ongoing porphyria. On one hand, axonal polyneuropathy is not the most typical neurological manifestation associated with the presence of these antibodies (Moon et al., 2014), and no relapse was noticed after definitely stopping steroids. On the other hand, previous reports of concomitant AIP and autoimmune disease (e.g., systemic lupus erythematosus and Sjögren syndrome) consider the latter (or their treatments) as a precipitating rather than a causal factor (Esteve-Valverde et al., 2020; Teng et al., 2020), although there are indications that AIP could be accompanied by systemic inflammatory, but non-autoimmune, reactions (Storjord et al., 2017). Taking into account the absence of a plausible causal relationship between anti-amphiphysin antibodies and both the patient's polyneuropathy and porphyria, we finally hypothesized that the transient systemic inflammatory reaction was non-specific and consecutive to repeated surgery (with a crossed reaction to the amphiphysin antigen).

Conclusion

Overall, VP unequivocally appears to be the only possible etiology of this patient's neurological symptoms, with supporting clinical, biological, and genetic evidence. Detailed and oriented medical history revealed that she had, before undergoing bariatric surgery, symptoms (intermittently dark reddish urine and fragility of the skin when exposed to the light) that could have, if screened for, prompted the search of porphyria (by urinary ALA and PBG measurements) prior to the gastric bypass procedure. Severe abdominal crises without evident cause during the early postoperative period would have been one additional red flag justifying this investigation. The diagnosis of VP enabled the instauration of specific management measures (Anderson et al., 2005) including carbohydrate-based diet, neurorehabilitation, and optimal symptom management, which allowed significant clinical improvement up to a relative autonomy in the activities of daily life, almost a year later. Nevertheless, due to the delayed diagnostic workup and treatment onset, persisting severe fatigue and limb neuropathic pain, despite heavy analgesia, prevented the patient from returning to work.

Considering the increasing prevalence of obesity (Kumar, 2007), which already constitutes a major public health problem (Koffman et al., 2006; Abdelaal et al., 2017), the indication for bariatric surgery (one of the most frequently used therapeutic procedures) will likely raise, exposing more patients with latent porphyria to develop the disease symptoms and potentially serious complications. Therefore, learning lessons from this case, we propose a pre-operative cost-effective screening for symptoms highly evocative of porphyria, especially in obese patients intending to opt for or awaiting gastric bypass. Positive screening would prompt appropriate and timely preventive and therapeutic measures, which efficiently reduce short- and long-term adverse outcomes (Sassa, 2006).

References

Abdelaal, M., le Roux, C. W., and Docherty, N. G. (2017). Morbidity and mortality associated with obesity. *Ann. Transl. Med.* 5 (7), 161. doi:10.21037/atm.2017.03.107

Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material; further inquiries can be directed to the corresponding author.

Ethics statement

A written informed consent for publication was signed by the patient and is available by request.

Author contributions

All authors contributed to the case report conception. Data collection and analysis were performed by EC and JC. The first draft of the manuscript was written by EC and JC. A-EM, EM, and J-MA completed and revised the first manuscript draft. All authors revised and edited subsequent manuscript versions. All authors have read and approved the final version of the manuscript.

Acknowledgments

The authors are grateful to the patient for granting her consent to this case report publication.

Conflict of interest

EM is a consultant to Alnylam.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors, and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Agarwal, S., Habtemarium, B., Xu, Y., Simon, A. R., Kim, J. B., and Robbie, G. J. (2021). Normal reference ranges for urinary δ -aminolevulinic acid and porphobilinogen levels. *JIMD Rep.* 57 (1), 85–93. doi:10.1002/jmd2.12173

Anderson, K. E., Bloomer, J. R., Bonkovsky, H. L., Kushner, J. P., Pierach, C. A., Pimstone, N. R., et al. (2005). Recommendations for the diagnosis and treatment of the acute porphyrias. *Ann. Intern. Med.* 142 (6), 439–450. doi:10.7326/0003-4819-142-6-200503150-00010

Barbaro, M., Kotajarvi, M., Harper, P., and Floderus, Y. (2013). Partial protoporphyrinogen oxidase (PPOX) gene deletions, due to different Alu-

mediated mechanisms, identified by MLPA analysis in patients with variegate porphyria. Orphanet J. Rare Dis. 8, 13. doi:10.1186/1750-1172-8-13

Chen, B., Solis-Villa, C., Hakenberg, J., Qiao, W., Srinivasan, R. R., Yasuda, M., et al. (2016). Acute intermittent porphyria: Predicted pathogenicity of HMBS variants indicates extremely low penetrance of the autosomal dominant disease. *Hum. Mutat.* 37 (11), 1215–1222. doi:10.1002/humu.23067

Danion, F., Guillot, M., Castelain, V., Puy, H., Deybach, J. C., and Schneider, F. (2012). An uncommon option for surviving bariatric surgery: Regaining weight! *Am. J. Med.* 125 (11), e1-e2. doi:10.1016/j. amjmed.2012.01.028

Di Pierro, E., De Canio, M., Mercadante, R., Savino, M., Granata, F., Tavazzi, D., et al. (2021). Laboratory diagnosis of porphyria. *Diagn. (Basel)* 11 (8), 1343. doi:10. 3390/diagnostics11081343

Dubey, D., Jitprapaikulsan, J., Bi, H., Do Campo, R. V., McKeon, A., Pittock, S. J., et al. (2019). Amphiphysin-IgG autoimmune neuropathy: A recognizable clinicopathologic syndrome. *Neurology* 93 (20), e1873–e1880. doi:10.1212/WNL. 00000000008472

Esteve-Valverde, E., Tapiz-Reula, A., Ruiz, D., and Alijotas-Reig, J. (2020). Systemic lupus erythematosus and hydroxychloroquine-related acute intermittent porphyria. *Rheumatol. Int.* 40 (5), 777–783. doi:10.1007/s00296-019-04500-8

Floderus, Y., Sardh, E., Moller, C., Andersson, C., Rejkjaer, L., Andersson, D. E. H., et al. (2006). Variations in porphobilinogen and 5-aminolevulinic acid concentrations in plasma and urine from asymptomatic carriers of the acute intermittent porphyria gene with increased porphyrin precursor excretion. *Clin. Chem.* 52 (4), 701-707. doi:10.1373/clinchem.2005.058198

Hift, R. J., Davidson, B. P., van der Hooft, C., Meissner, D. M., and Meissner, P. N. (2004). Plasma fluorescence scanning and fecal porphyrin analysis for the diagnosis of variegate porphyria: Precise determination of sensitivity and specificity with detection of protoporphyrinogen oxidase mutations as a reference standard. *Clin. Chem.* 50 (5), 915–923. doi:10.1373/clinchem.2003. 025213

Kauppinen, R. (2005). 365. England, 241–252. doi:10.1016/S0140-6736(05) 17744-7PorphyriasLancet

Koffman, B. M., Greenfield, L. J., Ali, I. I., and Pirzada, N. A. (2006). Neurologic complications after surgery for obesity. *Muscle Nerve* 33 (2), 166–176. doi:10.1002/mus.20394

Kumar, N. (2007). Obesity surgery: A word of neurologic caution. *Neurology* 68 (21), E36–E38. doi:10.1212/01.wnl.0000266868.07399.8b

Moon, J., Lee, S. T., Shin, J. W., Byun, J. I., Lim, J. A., Shin, Y. W., et al. (2014). Non-stiff anti-amphiphysin syndrome: Clinical manifestations and outcome after immunotherapy. *J. Neuroimmunol.* 274 (1-2), 209–214. doi:10.1016/j.jneuroim. 2014.07.011

NAPOS (2007). Drug database for acute porphyria. Available at: $\ensuremath{\mathrm{http://www.drugs-porphyria.org/}}$

Phillips, J. D. (2019). Heme biosynthesis and the porphyrias. *Mol. Genet. Metab.* 128 (3), 164–177. doi:10.1016/j.ymgme.2019.04.008

Pischik, E., and Kauppinen, R. (2015). An update of clinical management of acute intermittent porphyria. *Appl. Clin. Genet.* 8, 201–214. doi:10.2147/TACG.S48605

Punchai, S., Hanipah, Z. N., Meister, K. M., Schauer, P. R., Brethauer, S. A., and Aminian, A. (2017). Neurologic manifestations of vitamin B deficiency after bariatric surgery. *Obes. Surg.* 27 (8), 2079–2082. doi:10.1007/s11695-017-2607-8

Roeckel, L. A., Le Coz, G. M., Gaveriaux-Ruff, C., and Simonin, F. (2016). Opioidinduced hyperalgesia: Cellular and molecular mechanisms. *Neuroscience* 338, 160–182. doi:10.1016/j.neuroscience.2016.06.029

Sassa, S. (2006)., 135. England, 281-292. doi:10.1111/j.1365-2141.2006.06289. xModern diagnosis and management of the porphyriasBr. J. Haematol.

Schneider-Yin, X., and Minder, E. I. (2006). Swiss patients with variegate porphyria have unique mutations. Swiss Med. Wkly. 136 (31-32), 515-519.

Storjord, E., Dahl, J. A., Landsem, A., Fure, H., Ludviksen, J. K., Goldbeck-Wood, S., et al. (2017). Systemic inflammation in acute intermittent porphyria: A casecontrol study. *Clin. Exp. Immunol.* 187 (3), 466–479. doi:10.1111/cei.12899

Tabaro, I., Reimondo, G., Osella, G., Aurizi, C., Caraci, P., Barbieri, L., et al. (2018). Novel mutation of PPOX gene in a patient with abdominal pain and syndrome of inappropriate antidiuresis. *Endocrine* 61 (3), 403–406. doi:10.1007/s12020-018-1569-5

Teng, Q., Ma, L., Ma, Y., Zhang, Y., Kang, N., Hu, Y., et al. (2020). The challenge of managing comorbidities: A case report of primary sjogren's syndrome in a patient with acute intermittent porphyria. *Intractable Rare Dis. Res.* 9 (3), 137–140. doi:10.5582/irdr.2020.03064

Zürich, S. T., and Endokrinologie, D. (2018). Porphyrie. Verträgliche Arzneimittel bei akuten Porphyrien. Available at: http://www.porphyria.ch/formulare/liste_sichere_arzneimittel.pdf.