



Accelerating Detection of Variants During COVID-19 Surges by Diverse Technological and Public Health Partnerships: A Case Study From Indonesia

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OPEN ACCESS

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Specialty section:

This article was submitted to
Genetics of Common and Rare
Diseases,
a section of the journal
Frontiers in Genetics

Received: 03 November 2021

Accepted: 04 January 2022

Published: 28 January 2022

Citation:

Pradipta A, Kumaheri MA,
Wahyudi LD, Susanto AP, Agasi HI,
Shankar AH and Sudarmono P (2022)
Accelerating Detection of Variants
During COVID-19 Surges by Diverse
Technological and Public Health
Partnerships: A Case Study
From Indonesia.
Front. Genet. 13:801332.
doi: 10.3389/fgene.2022.801332

Early detection of Severe Acute Respiratory Syndrome Corona Virus 2 (SARS-CoV-2) variants and use of data for public health action requires a coordinated, rapid, and high throughput approach to whole genome sequencing (WGS). Currently, WGS output from many low- and middle-income countries (LMIC) has lagged. By fostering diverse partnerships and multiple sequencing technologies, Indonesia accelerated SARS-CoV-2 WGS uploads to GISAID from 1,210 in April 2021 to 5,791 in August 2021, an increase from 11 submissions per day between January to May, to 43 per day between June to August. Turn-around-time from specimen collection to submission decreased from 77 to 5 days, allowing for timely public health decisions. These changes were enabled by establishment of the National Genomic Surveillance Consortium, coordination between public and private sector laboratories with WGS capability, and diversification of sequencing platform technologies. Here we present how diversification on multiple levels enabled a rapid and significant increase of national WGS performance, with potentially valuable lessons for other LMICs.

Keywords: SARS-CoV-2, variant, whole-genome-sequencing, GISAID, turn-around-time, genome sequencing platform, Indonesia

INTRODUCTION

The Corona Virus Disease-2019 (COVID-19) pandemic has continued (Wang et al., 2021; World Health Organization, 2021) driven by low vaccination rates, premature reduction in physical distancing, and the emergence of variants. By early 2021, the World Health Organization (WHO) had designated at least four variants of concern (WHO Team Emergency Response, 2021) and spurred global efforts to quickly detect and track SARS-CoV-2 variants by whole genome sequencing (WGS). While high-income countries could meet WGS targets for cases and priority groups (e.g., breakthrough infections), most low- and middle-income countries (LMIC) have lagged. Many attribute this to gaps in financial resources, laboratory equipment, and staff. However,

associated with items 2) to 5) above. This overall increase was accompanied by a progressively decreasing time from specimen collection to sequence upload, with a sharp improvement following the increase in diversity of approaches.

We note that from the 3,125 specimens collected from May to August 2021, 71% (36 Alpha, 11 Beta, 2189 Delta) were variants of concern, with the Delta variant being dominant, and progressively and rapidly displacing the previously dominant B.1.466.2 variant in Indonesia. Moreover, the Delta variant was present in various sources, notably in children and rural locations. These gains in WGS rate and actionable knowledge underscore how diversification in laboratory and public health ecosystems can rapidly have broad impacts. The results translated to rapid and effective communication to authorities for COVID-19 testing and tracing efforts.

Role for Complementarity of Sequencing Platforms

We highlight that from June to August 2021, NGSC laboratories using ONT were able to submit 870 (37.8%) WGS results out of the national total of 2,301. Moreover, during this same period the average period from specimen collection to GISAID sequence upload, the turnaround time (TAT), was 8.6 days for high throughput setups. In comparison, the TAT for other laboratories was 36.5 days, although improving rapidly. We note that previous studies have shown the ability of ONT to provide sequencing data in 24 h with workflows of 8 h, allowing for same-day WGS. Other advantages include lower cost of entry for equipment, smaller batch sizes with fewer cost implications, reusable flow cells (Lipworth et al., 2020), and real-time monitoring (Bharagava et al., 2019) of sequencing reads to detect and react to suboptimal specimen processing. The ONT sequencing community provides quick access to specific workflow assistance for base-calling software (Guppy), and neural-network-based software for sequence assembly (Medaka), which are also optimized with a bioinformatics pipeline for subsequent dry analysis. Lineages can thereafter be assigned algorithmically using software such as Pangolin, which implements the Pango nomenclature system (Makalowski and Shabardina, 2020). Despite heterogeneity in specimens from diverse sources such as health centers, walk-in patients, quarantine centers, and pediatrics surveillance, most samples with ONT produced sequence reads with an average coverage of 83.75% with an average read depth of 337x, sufficient for 98% to be given a proper PANGO-based lineage for variant classification.

Other platforms using sequence-by-synthesis approaches such as Illumina or MGI (Kim et al., 2021) also offer advantages, including a wider installed base of machines, higher capacities per flow cell, and more third-party reagent options, along with excellent support. Still, other systems provide advantages for longer reads, such as Pacific Biosciences (Quail et al., 2012). To optimize speed, reliability, and resilience in serving WGS needs during the pandemic, an ecosystem with a multi-platform approach may prove advantageous compared to a single approach.

The challenges to transform WGS into a public health tool were highlighted during a surge of new cases in Indonesia that required high daily throughput and lower TAT. The increased demand

BOX 1 | Recommendations to accelerate SARS-CoV-2 whole genome sequencing in low- and middle-income countries.

- Rapidly establish a SARS-CoV-2 sequencing network.
- Deploy multiple sequencing platforms.
- Establish high throughput workflows and laboratory information systems.
- Build public health partnerships including private and public sector entities.
- Focus on coordination between interdisciplinary teams.

combined with increased number of cases inversely affected WGS output due to capacity saturation. To better understand these dynamics, we used the results (Figure 1B) to model how increased load influenced the TAT. As cases increase to over 7,000 per day, the model showed that the single platform lowest TAT would be 77 days, too long to inform appropriate public health action. However, by introducing a multi-platform approach, the TAT could be reduced to 5 days. Overall, increased surveillance efforts will be required to properly and rapidly identify populations vulnerable to variants, for example, those with decreased immune function or high exposure, such as those older than 65 years, health workers, or other yet undefined attributes at the individual and population level. As an RNA virus, SARS-CoV-2 maintains high mutation rates and, combined with unique selective pressures in each infected individual and community, can quickly give rise to novel viral populations (Korber et al., 2020; Okada et al., 2020). Efforts are ongoing to gather more accurate reports on vaccination status, comorbidity, clinical outcome, and other relevant information to uncover the possible impacts of variant-related COVID-19 surges (Ramasamy et al., 2020; Challen et al., 2021; Davies et al., 2021; Ozono et al., 2021).

CONCLUSION

The need for genomic sequencing capability dramatically increased due to the COVID-19 pandemic. In the middle of 2020, SARS-CoV-2 variants presented a significant challenge to pandemic management. Thus, the ability to detect new variants and their spread was crucial for proper public health responses. Indonesia managed to submit SARS-CoV-2 genomic sequences to GISAID in early 2021. However, the speed and amount of genomic sequencing in Indonesia were not adequate to meet the suggested minimum requirement for detection of new variants. The need for proper genomic sequencing was further highlighted by the growing number of variants of concern in early 2021.

We report that Indonesia successfully improved the total number of SARS-CoV-2 sequences submitted to GISAID and rapidly reduced the TAT. This success could be credited to an enhanced ecosystem of laboratory technology and public health partnerships. Moving forward, we recommend several additional improvements. First, substantial communication is required to ensure specimen preparation and delivery standards. Second, for scaling-up capacity, it is clear that preference would be for more mobile sequencing platforms that could reach different parts of the Indonesian archipelago, and isolated regions as seen in many LMICs. Third, there is a need for agile policy wherein requirements for minimum

sequencing and meta-data and reporting can be adjusted to better detect emergent variants. In **Box 1**, we list our conclusions and recommendations based on our case study in Indonesia.

Transition to the Delta variant was rapid, occurring within just a few weeks in the Asian megacity of Jakarta. We suggest the recommended actions are crucial for early detection of variants, such as Omicron, and enable timely decisions to mitigate the potential impact.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/Supplementary Material, further inquiries can be directed to the corresponding author.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by The Ethics Committee of the Faculty of Medicine,

REFERENCES

- Bharagava, R. N., Purchase, D., Saxena, G., and Mulla, S. I. (2019). Applications of Metagenomics in Microbial Bioremediation of Pollutants. *Microb. Divers. Genomic Era*, 459–477. Elsevier Inc. doi:10.1016/B978-0-12-814849-5.00026-5
- Challen, R., Brooks-Pollock, E., Read, J. M., Dyson, L., Tsaneva-Atanasova, K., and Danon, L. (2021). Risk of Mortality in Patients Infected with SARS-CoV-2 Variant of Concern 202012/1: Matched Cohort Study. *BMJ* 372, n579–10. doi:10.1136/bmj.n579
- Davies, N. G., Abbott, S., Barnard, R. C., Jarvis, C. I., Kucharski, A. J., Munday, J. D., et al. (2021). Estimated Transmissibility and Impact of SARS-CoV-2 Lineage B.1.1.7 in England. *Science* 372, eabg3055. doi:10.1126/science.abg3055
- European Centre for Disease Prevention and Control (2021). Technical Report : Guidance for Representative and Targeted Genomic SARS-CoV-2 Monitoring Key Messages. 1, 1–18.
- Kim, H. M., Jeon, S., Chung, O., Jun, J. H., Kim, H. S., Blazyte, A., et al. (2021). Comparative Analysis of 7 Short-Read Sequencing Platforms Using the Korean Reference Genome: MGI and Illumina Sequencing Benchmark for Whole-Genome Sequencing. *Gigascience* 10, 1–9. doi:10.1093/gigascience/giab014
- Korber, B., Fischer, W. M., Gnanakaran, S., Yoon, H., Theiler, J., Abfalterer, W., et al. (2020). Tracking Changes in SARS-CoV-2 Spike: Evidence that D614G Increases Infectivity of the COVID-19 Virus. *Cell* 182, 812e19–827. doi:10.1016/j.cell.2020.06.043
- Lipworth, S., Pickford, H., Sanderson, N., Chau, K. K., Kavanagh, J., Barker, L., et al. (2020). Optimized Use of Oxford Nanopore Flowcells for Hybrid Assemblies. *Microb. Genom* 6, 1–12. doi:10.1099/mgen.0.000453
- Makałowski, W., and Shabardina, V. (2020). Bioinformatics of Nanopore Sequencing. *J. Hum. Genet.* 65, 61–67. doi:10.1038/s10038-019-0659-4
- Okada, P., Buathong, R., Phuygun, S., Thanadachakul, T., Parnmen, S., Wongboot, W., et al. (2020). Early Transmission Patterns of Coronavirus Disease 2019 (COVID-19) in Travellers from Wuhan to Thailand, January 2020. *Euro Surveill.* 25. doi:10.2807/1560-7917.ES.2020.25.8.2000097
- Ozono, S., Zhang, Y., Ode, H., Sano, K., Tan, T. S., Imai, K., et al. (2021). SARS-CoV-2 D614G Spike Mutation Increases Entry Efficiency with Enhanced ACE2-Binding Affinity. *Nat. Commun.* 12, 848. doi:10.1038/s41467-021-21118-2

University of Indonesia–Cipto Mangunkusumo Hospital. Written informed consent to participate in this study was provided by the participants' legal guardian/next of kin.

AUTHOR CONTRIBUTIONS

AP, AS, AS, and PS contributed to the study design. MK and HA performed molecular laboratory analysis. AS, LW, and HA performed bioinformatic analysis. AP and AS contributed to the analysis of data and writing of the article. All authors read and approved the final manuscript.

FUNDING

The work was supported by Wellcome Trust Grant 222574/Z/21/Z supplement for SARS-CoV-2 genomic surveillance. The funders had no role in study design, data collection, and analysis, decision to publish, or preparation of the manuscript.

- Quail, M. A., Smith, M., Coupland, P., Otto, T. D., Harris, S. R., Connor, T. R., et al. (2012). A Tale of Three Next Generation Sequencing Platforms: Comparison of Ion Torrent, Pacific Biosciences and Illumina MiSeq Sequencers. *BMC Genomics* 13, 341. doi:10.1186/1471-2164-13-341
- Ramasamy, M. N., Minassian, A. M., Ewer, K. J., Flaxman, A. L., Folegatti, P. M., Owens, D. R., et al. (2020). Safety and Immunogenicity of ChAdOx1 nCoV-19 Vaccine Administered in a Prime-Boost Regimen in Young and Old Adults (COV002): a Single-Blind, Randomised, Controlled, Phase 2/3 Trial. *Lancet* 396, 1979–1993. doi:10.1016/S0140-6736(20)32466-1
- Wang, C., Wang, Z., Wang, G., Lau, J. Y., Zhang, K., Li, W., et al. (2021). COVID-19 in Early 2021: Current Status and Looking Forward. *Signal. Transduct. Target. Ther.* 6, 114. doi:10.1038/s41392-021-00527-1
- WHO Team Emergency Response (2021). COVID-19 Weekly Epidemiological Update no.39. WHO Emergency Situational Update, 1–32. Available at: <https://www.who.int/publications/m/item/weekly-epidemiological-update-on-covid-19-11-may-2021>.
- World Health Organization Indonesia (2021). *Coronavirus Disease 2019 (COVID-19) Situation Report - 60*, 60.

Conflict of Interest: AP, MK, LW, AS, and HA are employed by Genomik Solidaritas Indonesia (GSI) Lab. AS and PS are part of the Genomik Solidaritas Indonesia (GSI) Lab Scientific Advisory Board.

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