#### Check for updates

#### OPEN ACCESS

EDITED BY Wentao Zhang, Tongji University, China

#### REVIEWED BY

Jingzeng Du, Singapore General Hospital, Singapore Isil Ezgi Eryilmaz, Bursa Uludağ University, Turkey Lei Wang, Renmin Hospital of Wuhan University, China

#### \*CORRESPONDENCE Jiling Wen,

[wenjiling@tongji.edu.cn](mailto:wenjiling@tongji.edu.cn) Mingyue Tan, [tanmoon@163.com](mailto:tanmoon@163.com) Fang Zhang, [China.medego@163.com](mailto:China.medego@163.com) Bing Shen, [urodrshenbing@shsmu.edu.cn](mailto:urodrshenbing@shsmu.edu.cn)

#### † These authors share first authorship

SPECIALTY SECTION This article was submitted to RNA,

a section of the journal Frontiers in Genetics

RECEIVED 28 October 2022 ACCEPTED 12 December 2022 PUBLISHED 04 January 2023

#### **CITATION**

Cai J, Xie H, Yan Y, Huang Z, Tang P, Cao X, Wang Z, Yang C, Wen J, Tan M, Zhang F and Shen B (2023), A novel cuproptosis-related lncRNA signature predicts prognosis and therapeutic response in bladder cancer. Front. Genet. 13:1082691. doi: [10.3389/fgene.2022.1082691](https://doi.org/10.3389/fgene.2022.1082691)

#### COPYRIGHT

© 2023 Cai, Xie, Yan, Huang, Tang, Cao, Wang, Yang, Wen, Tan, Zhang and Shen. This is an open-access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/)

[\(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# [A novel cuproptosis-related](https://www.frontiersin.org/articles/10.3389/fgene.2022.1082691/full) [lncRNA signature predicts](https://www.frontiersin.org/articles/10.3389/fgene.2022.1082691/full) [prognosis and therapeutic](https://www.frontiersin.org/articles/10.3389/fgene.2022.1082691/full) [response in bladder cancer](https://www.frontiersin.org/articles/10.3389/fgene.2022.1082691/full)

Jinming Cai<sup>1,2†</sup>, Haoran Xie<sup>3†</sup>, Yilin Yan<sup>1</sup>, Zhengnan Huang<sup>1</sup>, Pengfei Tang<sup>4</sup>, Xiangqian Cao<sup>1</sup>, Zeyi Wang<sup>4</sup>, Chenkai Yang<sup>1</sup>, Jiling Wen<sup>2\*</sup>, Mingyue Tan<sup>5\*</sup>, Fang Zhang<sup>1\*</sup> and Bing Shen<sup>1\*</sup>

1 Department of Urology, Shanghai General Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai, China, <sup>2</sup>Department of Urology, Shanghai East Hospital, Tongji University School of Medicine, Shanghai, China, <sup>3</sup>Department of Gastroenterology, Shanghai General Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai, China, <sup>4</sup> Department of Urology, Shanghai General Hospital Affiliated to Nanjing Medical University, Shanghai, China, <sup>5</sup>Department of Urology, Shuguang Hospital, Shanghai University of Traditional Chinese Medicine, Shanghai, China

Bladder cancer (BC) ranks the tenth in the incidence of global tumor epidemiology. LncRNAs and cuproptosis were discovered to regulate the cell death. Herein, we downloaded transcriptome profiling, mutational data, and clinical data on patients from The Cancer Genome Atlas (TCGA). High- and low-risk BC patients were categorized. Three CRLs (AL590428.1, AL138756.1 and GUSBP11) were taken into prognostic signature through least absolute shrinkage and selection operator (LASSO) Cox regression. Worse OS and PFS were shown in high-risk group ( $p < 0.05$ ). ROC, independent prognostic analyses, nomogram and C-index were predicted via CRLs. Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) analysis indicated IncRNAs play a biological role in BC progression. Immune-related functions showed the high-risk group received more benefit from immunotherapy and had stronger immune responses, and the overall survival was better ( $p < 0.05$ ). Finally, a more effective outcome ( $p <$ 0.05) was found from clinical immunotherapy via the TIDE algorithm and many potential anti-tumor drugs were identified. In our study, the cuproptosis-related signature provided a novel tool to predict the prognosis in BC patients accurately and provided a novel strategy for clinical immunotherapy and clinical applications.

#### KEYWORDS

cuproptosis, bladder cancer, lncRNAs, immunotherapy, survival analysis

Abbreviations: BC, bladder cancer; CRLs, cuproptosis-related lncRNAs; CRGs, Cuproptosis-related genes; TMB, tumor mutation burden.

# Introduction

One of the most common malignancies of the urogenital tract is bladder cancer (BC). According to the latest epidemic study of tumor incidence, BC ranks tenth ([Sung et al., 2021](#page-13-0)). The clinical pathology of BC contains urothelial carcinoma (90–95%), adenocarcinoma and squamous cell carcinoma ([Lenis et al., 2020\)](#page-13-1). Based on pathological classification, tumor invades the muscle or beyond, and invades the urothelium or lamina propria, which were called muscleinvasive bladder cancer (MIBC), and non–muscle-invasive bladder cancer (NMIBC), respectively ([Hurst et al., 2018](#page-12-0)). The rate of 5-year recurrence is 50–70%, with a risk of progression of 10–30%, after receiving the initial transurethral resection of bladder tumor (TURBT) ([Martinez Rodriguez et al.,](#page-13-2) [2017\)](#page-13-2). Therefore, owing to the high incidence, mortality and recurrence of BC, it is necessary to construct more prognostic models. Long non-coding RNSs (lncRNAs) can regulate the expression of oncogene or cancer related genes, which are 200 nucleotides of non-coding transcripts in length. Previous studies have indicated lncRNAs were enriched in many biological processions, such as immune responses, metabolism regulation, and cell metastasis ([Zhu et al., 2021](#page-13-3)). LncRNAs have been found to have specific relevance to the pathogenesis of BC and influence the progression of BC ([Li et al., 2020\)](#page-13-4).

Copper plays a crucial role in maintaining biological processes in various life entities. Recent researches found the copper concentration of cancer patient is significantly higher than the health in serum and cancer tissue ([Ishida et al., 2013](#page-12-1); [Blockhuys et al., 2017](#page-12-2); [Ge et al., 2022\)](#page-12-3). Given the copper disorder may lead to metabolic imbalance and cytotoxicity, copper variation of intracellular levels may influence tumor progression and development [\(Babak and Ahn, 2021\)](#page-12-4). Moreover, copper chemical materials (copper chelators, ionophores, etc) have been utilized to anticancer treatment [\(Brady et al., 2017;](#page-12-5) O'[Day et al., 2013\)](#page-13-5). A recent research uncovered a new cell death pathway, cuproptosis, in copper compounds coupled with lipoylated tricarboxylic acid (TCA) cycle components, resulting in cell death and toxic protein stress, directly ([Tsvetkov et al., 2022\)](#page-13-6). BC showed enhanced glucose utilization for glycolysis, and pathways significantly unbalanced in tumor relative to normal urothelium included tricarboxylic acid (TCA) cycle, glucose and so on ([Sahu et al., 2017](#page-13-7)). A vital role for glutamine (Gln) from TCA in promoting proliferation of Gln-dependent BC T24 cell line has been documented, which supplemented adenosine triphosphate (ATP) generation and neutralizing reactive oxygen species (ROS) for activating the STAT3 pathway [\(Sun et al., 2019a\)](#page-13-8). Several genes were identified connected with copper-induced cell death, which can offer a chance new to construct a new prognosis model.

In our study, an emerging prognostic model for BC was explored based on CRLs that can offer prognosis prediction and selection of patients for immunotherapies. The workflow was presented in [Figure 1A](#page-1-0). The prognosis model was validated via bioinformatics. Moreover, functional enrichment analysis was performed to discuss potential pathways for signaling in groups at risk. In the end, immune functions and immunotherapy were analyzed in the risk group.

<span id="page-1-0"></span>



<span id="page-2-0"></span>TABLE 1 Clinical features of BC patients in TCGA dataset.

# Materials and methods

#### Data collection and preprocessing

The expression of genes and clinical data were downloaded from TCGA database up to 29 March 2022 ([https://portal.gdc.](https://portal.gdc.cancer.gov/repository) [cancer.gov/repository\)](https://portal.gdc.cancer.gov/repository). Then, Active Perl was used to pre-process the raw data of gene expression and human configuration data

and synthetic matrices were obtained. LncRNAs data and mRNAs data were distinguished via Active Perl. Moreover, clinical data was prepared to estimate missing overall survival (OS) values. In the end, a total of 431 patients' gene expression profiles were obtained. And using the "caret" R package, train risk is divided by test risk in a 1:1 ratio. The focal-level copy number variation (CNV) values were downloaded via GISTIC2, using a "masked copy number fragment" file. [\(Mermel et al.,](#page-13-9) [2011](#page-13-9)).

## Identification of cuproptosis-related genes and lncRNAs in TCGA cohort

A previous study obtained cuproptosis-related genes (CRGs) (Supplementary Table S1) [\(Tsvetkov et al., 2022\)](#page-13-6). Using "R" (version 4.1.0) software, CRLs were distinguished via "limma" R package (|Pearson R|>0.4,  $p < 0.001$ ) [\(Supplementary Table S2\)](#page-12-6). CRGs were isolated from the whole gene expression data ([Supplementary Table S3](#page-12-6)).

#### Construction of CRLs prognostic signature

956 prognostic CRLs were identified via univariate Cox regression analysis ( $p \lt 0.05$ ) with "Survival" R package ([Eaton et al., 2020\)](#page-12-7). The gene signature was identified, which contains most like biomarkers of prognosis via "glmnet" R package [\(Friedman et al., 2010](#page-12-8)). LASSO with a tenfold crossvalidation offered a tool to establish the prognostic model. Based on the following formula:

### Risk score =  $\sum$  (Coef \* EXP),

The risk score was calculated. Coef means the coefficient and EXP means the expression level of each prognostic CRLs in the formula. Based on risk scores, High- and low-risk BC patients were categorized. To elucidate the predictive value of the CRLsbased model of prognostic signature, we performed the receiver operating characteristic (ROC) of 1-year survival in the training and testing group by "ROC" R package [\(Blanche et al., 2013](#page-12-9)). In accordance with the optimal risk cutoff value analyzed by "Survival" R package, the patients were categorized as highrisk or low-risk ([Eaton et al., 2020](#page-12-7)). The prognostic value of two-CRL signature on BC was analyzed by Kaplan-Meier. Then, the risk model was tested using analyses of Cox regressions, univariate and multivariate, to estimate if the risk model showed better predictive ability of prognosis independently associated with other clinicopathological features, such as gender, grade and pathologic staging. Moreover, a TCGA-based prognostic nomogram was developed. Clinical parameters and independent prognostic factors are included in the nomogram.



#### <span id="page-3-0"></span>FIGURE 2

Identification of the CRLs. (A).LASSO regression of prognotic CRLs. (B).The penalty parameter ( $\lambda$ ) in the LASSO regression model was selected through two cross-validation. (C).The relationship between CRLs and CRGs for the signature in the corrlation heatmap.



#### <span id="page-3-1"></span>FIGURE 3

Kaplan-Meier survival analysis of BC patients. Overall survival of BC patients in all groups (A), train groups (B), testing groups (C), respectively. (D).PFS of BC patients in all groups.



### <span id="page-4-0"></span>Construction of risk score model

Basing on median values, we divided samples of all patients into high- and low-subgroups to estimate the prognosis and the signature. Progression-free survival (PFS) and OS were calculated by "Survival" R package. We estimated independent prognostic abilities of risk prediction models using both univariate and multivariate analyses. CRLs expression and patient survival status were shown based on the risk scores via "Pheatmap" R package and pheatmap. C-Index was showed with "Survival" R package for validating the performance of the model to predict patients' survival.

## Nomogram construction and clinical feature validations

To exhibit the difference between the actual and predicted results, the nomogram was constructed for gender, grade, stage and the calibration curves were plotted. 1-, 3- and 5-year OS were predicted via a stepwise Cox regression in the TCGA dataset.

### Analysis of principle component analysis (PCA) and enrichment function

The "limma" R package and "Scatterplot3d" R package were utilized to construct PCA, which can show clear

distribution in different risk groups. The KEGG and GO enrichment of CRLs was performed with the "ClusterProlfiler" R package.

# TMB and immune-related functional analysis

The correlation between TMB and risk score was completed by "maftools" R package. The difference between patient survival status and TMB was explored via the "Survival" R package. Immune-related functions were analyzed and its differences were identified via "limma" and "GSVA" R package, which were visualized by "Pheatmap" R package.

#### Analyses of immunotherapy and potential pharmaceuticals

The Tumor Immune Dysfunction and Exclusion (TIDE) database (<http://tide.dfci.harvard.edu/>) contained tumor pretreatment expression profiles. BC's TIDE dataset was downloaded and the correlation between TMB and risk score was explored by "ggpubr" and "limma" R packages. The "pRRophetic", "ggpubr" and "ggplot2" R package were conducted to screen for potential therapeutic drugs and explore the sensitivity of drugs.

## Statistical analysis

R software and packages were used for statistical analysis. Here, based on False Discovery Rates (FDRs) < 0.05 and the p-value (<0.05), statistics were collected. Kaplan-Meier (KM) was utilized to compare OS between subgroups. Pearson correlation tests were used to identify CRLs. The Chisquare test was conducted to analyze categorical variables of groups. The differential risk scores of subgroups were compared via the Student's t-test. Moreover, the prognostic ability of risk scores and other clinical data was explored using multivariate and univariate Cox regression analyses. Herein, statistical significance was determined at  $p < 0.05$ ; \*\* $p < 0.01$ ; \*\*\* $p < 0.001$ .

# Results

## Identification of bladder cancer-specific CRLs and establishment of the prognostic signature

TCGA-BLCA dataset was downloaded, which contains 412 cancer samples and 19 normal samples. Clinical features were exhibited in the [Table 1](#page-2-0). 956 CRLs were identified from 16876 lncRNAs and 19 CRGs that met the criteria (|R|>0.4 and  $p < 0.001$ ). Sankey diagram was plotted to analyze the coexpression relationship between CRGs and CRLs [\(Figure 1B](#page-1-0)). LASSO Cox regression was utilized to identify CRLs in the training group [\(Figures 2A, B\)](#page-3-0). A total of 69 CRLs were identified through univariate Cox regression [\(Supplementary](#page-12-6) [Figure S1](#page-12-6)). Low- and high-risk lncRNAs were shown with red and green in the forest plot, respectively. Then, 3 CRLs as independent prognostic factors were identified via multivariate Cox regression ( $p < 0.05$ ). According to the formula and the expression levels of the 3 lncRNAs, the risk score of all samples was obtained. Risk score = (-0.278725960360529\* AL138756.1) + (+1.03981953291805\* AL590428.1)+(-0.454158106791872\* GUSBP11). Finally, the correlation heatmap was used to show the relationship of between CRGs and lncRNAs ([Figure 2C\)](#page-3-0).

### The correlation of survival status and genetic signature

In the light of the median risk scores as the cut-off value, lowand high-risk patients were classified. OS and PFS in high-risk group were found to be significantly shorter than that in low-risk group in three groups (all groups, training group and testing group) ( $p < 0.05$ ) [\(Figures 3A](#page-3-1)–[D\)](#page-3-1). The risk curves were visualized to exhibit the correlation between risk score and survival status in subgroups (all groups, training group and testing group), which were displayed in [Figures 4A](#page-4-0)–[C.](#page-4-0) Moreover, mortality was higher

in high-risk groups than in low-risk groups ( $p < 0.05$ ). The heatmap was utilized to show the differences of two risk subgroups for 3 lncRNAs with good consistency. AL138756.1 and GUSBP11 were overexpressed in the lowsubgroup and underexpressed in the high- subgroup, while AL590428.1 showed the opposite trend. Therefore, it was found that AL138756.1 and GUSBP11 expression are negatively correlated with risk scores. AL590428.1 showed the positive correlation with risk scores.

## Univariate and multivariatae analysis of the prognostic model

To figure out whether the signature we constructed could be utilized to be independent prognostic factors, univariate and multivariate Cox regression were conducted. The results indicated that age (HR = 1.035, 1.019-1.051,  $p < .001$ ), stage  $(HR = 1.724, 1.420 - 2.092, p < .001)$  and risk score (HR = 1.119, 1.087–1.152,  $p < .001$ ) were significantly associated with OS in univariate Cox regression [\(Figure 5A](#page-6-0)). And multivariate Cox regression suggested age (HR = 1.031, 1.014–1.047,  $p < .001$ ), stage (HR = 1.629, 1.336–1.988,  $p < .001$ ) and risk score (HR = 1.115, 1.080-1.150,  $p < .001$ ) were independently related with OS [\(Figure 5B\)](#page-6-0), which suggested the prognostic model can be regarded as an independent prognostic indicator for BC patients. Then, to estimate the predictive power of the risk score, ROCs were performed. The area under curve (AUC) of the risk score was 0.628 [\(Figure 5C](#page-6-0)). Similarly, AUCs for 1, 3, and 5 years were 0.628, 0.630, and 0.641 [\(Figure 5D\)](#page-6-0). AUCs were greater than or equal to 0.600, which suggested the prognostic model has a better diagnostic value.

## Construction and validation of the OS nomogram and principal component analysis

A nomogram, including grade, gender, age, T stage, N stage, M stage, stage and risk, was plotted and the nomogram predicted the survival of 1-, 3- and 5-year ([Figures 6A, B](#page-7-0)). Then, C-index curves were plotted to determine if there were any discrepancies in patients' survival over time [\(Supplementary Table S2\)](#page-12-6). The results indicated that the risk signature has a high predictive accuracy of survival status in BC patients and was not influenced by different clinical grades. Moreover, OS of clinical stage in subgroups was analyzed. As shown in [Figures 6C, D,](#page-7-0) OS in stages I-IV differed significantly in the low- and high-risk individuals ( $p < .05$ ). At last, the distribution of all genes, CRGs, CRLs and risk lncRNAs was analyzed via PCA ([Figure 7\)](#page-8-0), which showed a clear status of risk lncRNAs. Therefore, the results demonstrated all lncRNAs were reliably utilized to establish the signature.



<span id="page-6-0"></span>FIGURE 5

The prognostic value of the CRLs in BC patients. Univariate (A) and multivariate (B) independent prognostic analysis to explore the risk score was independently related with OS. (C).ROC curve analysis was utilized to compare the predictive accuracy between the risk model and various clinical features. (D).ROC curves for 1-, 3-, and 5-year survival prediction.

### Functional enrichment and immunerelated functional analyses

To understand the relationship between the CRLs and tumor development, GO and KEGG were performed [\(Figures 8A, B](#page-9-0)). The results of GO results showed CRLs were enriched in signaling receptor activator activity, external side of plasma membrane, and positive regulation of cell activation, which were molecular function, cellular component, and biological procession ( $p \lt .05$ ), respectively. KEGG results indicated these lncRNAs may be associated with cytokine-cytokine receptor interaction, viral protein interaction with cytokine and cytokine receptor, PI3K-Akt signaling pathway and so on  $(p < .05)$ . Circle plot showed the distribution of CRLs in GO and KEGG enrichment. Furthermore, immune-related functions were analyzed to figure out the immune status of subgroups ([Figure 8C](#page-9-0)). The results showed there was a significant increase in Type\_I\_IFN\_Response, Parainflammation, APC co-

stimulation, T cell co-stimulation, T cell co-inhibition, inflammation-promoting, cytolytic-activity and so on in the high-risk group when compared with the low-risk group ( $p < .05$ ).

## The burden of tumor mutations and drug sensitivity of tumor

Mutation of most genes was observed in the low- and highrisk groups via maftools algorithm. The mutation of TP53 (high risk, 56%; low risk 40%) and TTN (high risk, 46%; low risk, 39%) in the high-risk group was more common than that in the lowrisk one in the top-5 genes with highest mutation ( $p < .05$ ). The mutation of KMT2D (high risk, 25%; low risk 29%), MUC16 (high risk, 25%; low risk, 26%) and ARID1A (high risk, 21%; low risk, 28%) in the low-risk group was higher than that in the highrisk group in the top-5 genes with highest mutation ( $p < .05$ )



<span id="page-7-0"></span>([Figures 9A, B](#page-10-0)). Moreover, it was no significant to distinguish low-risk from high-risk groups ( $p = 0.33$ ) [\(Figure 9C](#page-10-0)). Then, to determine whether there was any association between the TMB and the BC survival status, survival curves were plotted in pure TMB and combined TMB-risk groups [\(Figures 9D, E\)](#page-10-0). There was a significant difference among subgroups ( $p < 0.05$ ). High TMB had a longer survival time than low TMB ( $p < 0.05$ ). Meanwhile, comparisons of all subgroups were significantly different ( $p < 0.05$ ). Furthermore, the TIDE algorithm was utilized to investigate the difference of sensitivity to immunotherapy in two subgroups [\(Figure 10A](#page-11-0)). The results indicated the TIDE score in the high-risk group was higher than that in the low-risk group ( $p < 0.05$ ). Finally, potential antitumor drugs were screened via the algorithm from the "pRRophetic" R package. The top 5 most significantly associated drugs were listed and an analysis of the correlation between sensitivity and risk scores was conducted ( $p < 0.05$ ), including Bexarotene, WH-4-023, Midostaurin, Cyclopamine and CGP-60474 ([Figures 10B](#page-11-0)–[F](#page-11-0)).

## **Discussion**

BC is notorious cancer with high incidence and mortality. Previous studies have found the Nuclear Matrix Protein 22 (NMP22) and cytokeratin markers (CYFRA 21-1 or UBC), which can play a role in biological markers of BC as a substitute to urinary cytology ([Sánchez-Carbayo et al., 2000\)](#page-13-10). However, no available molecular biomarker can replace cytology in sensitivity and specificity. Therefore, a reliable BC risk model was constructed, which is an aggressive need to figure out the clinical outcomes of BC patients. For the extension of multiomics data and database, optimized data mining algorithms have an important influence on tumor research [\(Angus et al., 2019;](#page-12-10) [Nacev et al., 2019](#page-13-11); [Liu et al., 2020;](#page-13-12) [Tabassum et al., 2020\)](#page-13-13). Multiple risk signatures and transcriptome profiling provided a novel insight into the prognosis of individual patients via combining the gene expression and clinical features [\(Xie et al.,](#page-13-14) [2021](#page-13-14); [Yan et al., 2021\)](#page-13-15). Due to the lack of systematic evaluation, most studies cannot be utilized to clinical practice. Hence, it is



<span id="page-8-0"></span>urgent that prognostic factors of BC patients are identified to distinguish the high-risk population.

A series of researches have reported the relationship between copper and BC (Konukoğ[lu et al., 1996](#page-13-16); [Lin et al., 2009](#page-13-17); [Mazdak](#page-13-18) [et al., 2010](#page-13-18); [Gecit et al., 2011;](#page-12-11) [Guo et al., 2012\)](#page-12-12). Moreover, copper complex [Cu<sup>II</sup> <sub>2</sub>Cu<sup>I</sup>(L)<sub>2</sub>(Br)<sub>3</sub>] have been found to succeed inducing apoptosis in pancreatic cancer, such as tolfenamic acid–Cu II) complex and Cu II) complex of ketoprofensalicylhydrazone (FPA-306) ([Hurtado et al., 2018;](#page-12-13) [Gou et al.,](#page-12-14) [2021\)](#page-12-14). Cuproptosis has only recently been identified, which was deem to be a new unique form of cell death [\(Polishchuk et al.,](#page-13-19) [2019;](#page-13-19) [Aubert et al., 2020](#page-12-15); [Dong et al., 2021;](#page-12-16) [Ren et al., 2021](#page-13-20); [Tsvetkov et al., 2022](#page-13-6)). The method of cell death leads to the gathering of acylated protein and downregulating the iron-sulfur protein via the binding of copper to lipid acylated components of the TCA. The procession results in proteotoxicity and cell death in the end [\(Koh et al., 2017](#page-13-21)). Interestingly, there were several clinical trials that have been performed with the copper ionophore micromolecular anti-tumor drugs Elesclomol ([Soma et al., 2018](#page-13-22)). The results failed to get a satisfactory outcome. LncRNAs have been widely recognized to have a deep connection with tumor progression, including BC [\(Cao](#page-12-17) [et al., 2019](#page-12-17); [Luo et al., 2019](#page-13-23)). However, rarely studies have found the regulatory role in BC.



<span id="page-9-0"></span>In our present study, 69 CRLs were identified from coexpression of lncRNAs and CRGs. With the analysis of univariate and multivariate Cox regression, 3 CRLs with prognostic value, including AL590428.1, AL138756.1 and GUSBP11, were achieved and a prognostic model was developed. The results from ROC, OS, PFS, nomogram and heatmap indicated the signature of 3 CRLs distinguished prognostic features with good accuracy in two subgroups in BC patients. Meanwhile, clinical outcomes in BC patients were predicted as independent prognostic factors. Meaningfully, AL590428.1 was investigated in cancer firstly. It is reported that AL138756.1 participated in predicting the prognosis of colon adenocarcinoma as a prognostic indicator [\(Zhou](#page-13-24) [et al., 2020](#page-13-24)). GUSBP11 has been found to regulate the progression of tumor, including triple negative breast cancer and nasopharyngeal carcinoma ([Wu et al., 2022](#page-13-25); [Zhang et al., 2022](#page-13-26)). Recent research has found that GUSBP11 was contained in a machine learning-based computational network for an indicator of immune infiltration of tumor microenvironment [\(Zhou et al.,](#page-13-27) [2021\)](#page-13-27). Herein, we further elucidated the correlation between GUSBP11 and BC. Then, signaling receptor

activator activity, external side of plasma membrane, and positive regulation of cell activation were found in the GO enrichment. The KEGG pathway analysis revealed cytokinecytokine receptor interaction, viral protein interaction with cytokine and cytokine receptor and PI3K-Akt signaling pathway. Therefore, the results suggested that disorder interactions between bioactive molecules and cellular signaling pathway severely promote the progression and generate poor clinical prognostic outcomes.

Furthermore, the correlation between TMB, immune function and risk scores was analyzed in BC. TMB was commonly regarded as an indicator for immune checkpoint blockade (ICB) in BC, lung cancer and melanoma [\(Fusco et al., 2021;](#page-12-18) [Jardim et al., 2021;](#page-12-19) [McGrail](#page-13-28) [et al., 2021](#page-13-28)). Although the lack of significant differences between two subgroups ( $p > 0.05$ ), the frequency of mutation genes in subgroups has changed a lot and survival time was significantly prolonged in low TMB ( $p <$ 0.05). We found the mutation of TP53 and TTN was increased but KMT2D, MUC16 and ARID1A was decreased in high-risk group. An increasing research has demonstrated TP53 and TTN have an important effect on the promotion of tumor ([Mogi and Kuwano, 2011;](#page-13-29) [Fu et al.,](#page-12-20)



<span id="page-10-0"></span>[2019;](#page-12-20) [Shen et al., 2020;](#page-13-30) [Siracusano et al., 2020](#page-13-31)). According to our results, the same conclusion keeps in line with previous researches. Moreover, KMT2D and ARID1A were found to be decreased in expression ([Garczyk et al., 2018;](#page-12-21) [Sun et al.,](#page-13-32) [2019b\)](#page-13-32), and MUC16 can be regarded as potential surrogate biomarker of poor prognosis and unique molecular signature ([Cotton et al., 2017](#page-12-22)). Here, low-risk group showed higher mutation levels of KMT2D and ARID1A than high-risk group. In line with previous studies, our results are also valid. We evaluated the prognosis of BC patients in immune-related functions and contribute to figure out the relationship between lncRNAs and immune functions. The analysis of immune-related functions implied all members of the risk signature were closely related to the antigenpresenting of tumor. Meanwhile, the results of the TIDE algorithm indicated a high-risk group received more benefit from immunotherapy, which keeps in line with previous conclusions. Due to the advent of Bacille Calmette Guerin (BCG), BC is one of the earliest cancers

where the concept of immunotherapy was proposed [\(Vasekar](#page-13-33) [et al., 2016\)](#page-13-33). While it is true, recent advances have revealed multiple molecular mechanism as a prevalent tumor therapy, such as CD24/Siglec-10, EMT, PD-L1/PD-1, C/EBPβ transcription factors and hypoxia/HIF-1α [\(Marigo et al.,](#page-13-34) [2010](#page-13-34); [Vaupel and Multhoff, 2018](#page-13-35); [Jiang et al., 2019](#page-12-23); [Jiang](#page-12-24) [and Zhan, 2020](#page-12-24); [Yin and Gao, 2020\)](#page-13-36). Although LncRNAs itself do not code proteins, immunotherapy and immune responses have been identified to participate [\(Jiang et al.,](#page-12-25) [2021](#page-12-25)). The interactions between lncRNAs and cancer immunotherapy involved various immune cells in tumor microenvironment ([Luo et al., 2020](#page-13-37)), which is consistent with our analyses of immune-related function. Besides, the pRRophetic algorithm was conducted to screen for potential anti-tumor drugs via the analysis of sensitivity and correlation of those drugs. Part of those have been applied to other cancers. Despite the drugs mechanisms of impact on BC need to be figure out, it provides a new insight on drawing up a therapeutic schedule.



<span id="page-11-0"></span>Immunotherapy, drug sensitivity and the relationship between the drug sensitivityand risk scores. (A). Different TIDE in the high and low groups. Observed the drug sensitivity and its relationship with risk scores in top 5 drugs: Bexarotene (B), WH-4-023 (C), Midostaurin (D), Cyclopamine (E) and CGP-60474 (F).

In brief, a CRLs signature of BC was constructed. In the light of different calculated risk scores, two subgroups were divided and the relationship was analyzed between TMB and subgroups, immunotherapy and drug sensitivity. Therefore, a novel strategy for predicting survival status and optimizing clinical therapy for BC has been developed in our study.

# Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article[/Supplementary Material](#page-12-6).

# Author contributions

The conception and design of the study: JC, HX, JW, and BS; Data collection: YY and PT; Data analysis: JC, ZH, MT, and XC; Article drafting: JC, ZW, CY, and FZ; Administrative support: BS and FZ; All authors have read and agreed to the published vision of the manuscript. Final approval of manuscript: All authors.

# Funding

This study was supported by the National Natural Science Foundation (92059112, 81972370 and 82072821), Shanghai Songjiang Municipal Science and Technology Commission Natural Science Foundation (20SJKJGG250), and Shanghai Specialized Research Fund for Integrated Chinese and Western Medicine in General Hospitals (ZHYY-ZXYJHZX-1-201705).

# Acknowledgments

We gratefully acknowledge the assistance of Zhiwen Xie debugging of the bioinformatical algorithm. We acknowledge TCGA database for providing their platforms and contributors for uploading their meaningful datasets.

# Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

# References

<span id="page-12-10"></span>Angus, L., Smid, M., Wilting, S. M., van Riet, J., Van Hoeck, A., Nguyen, L., et al. (2019). The genomic landscape of metastatic breast cancer highlights changes in mutation and signature frequencies. Nat. Genet. 51 (10), 1450–1458. doi[:10.1038/](https://doi.org/10.1038/s41588-019-0507-7) [s41588-019-0507-7](https://doi.org/10.1038/s41588-019-0507-7)

<span id="page-12-15"></span>Aubert, L., Nandagopal, N., Steinhart, Z., Lavoie, G., Nourreddine, S., Berman, J., et al. (2020). Copper bioavailability is a KRAS-specific vulnerability in colorectal cancer. Nat. Commun. 11 (1), 3701. doi:[10.1038/s41467-020-17549-y](https://doi.org/10.1038/s41467-020-17549-y)

<span id="page-12-4"></span>Babak, M. V., and Ahn, D. (2021). Modulation of intracellular copper levels as the mechanism of action of anticancer copper complexes: Clinical relevance. Biomedicines 9 (8), 852. doi[:10.3390/biomedicines9080852](https://doi.org/10.3390/biomedicines9080852)

<span id="page-12-9"></span>Blanche, P., Dartigues, J-F., and Jacqmin-Gadda, H. (2013). Estimating and comparing time-dependent areas under receiver operating characteristic curves for censored event times with competing risks. Stat. Med. 32 (30), 5381–5397. doi[:10.](https://doi.org/10.1002/sim.5958) [1002/sim.5958](https://doi.org/10.1002/sim.5958)

<span id="page-12-2"></span>Blockhuys, S., Celauro, E., Hildesjö, C., Feizi, A., Stål, O., Fierro-González, J. C., et al. (2017). Defining the human copper proteome and analysis of its expression variation in cancers. Metallomics 9 (2), 112-123. doi:[10.1039/c6mt00202a](https://doi.org/10.1039/c6mt00202a)

<span id="page-12-5"></span>Brady, D. C., Crowe, M. S., Greenberg, D. N., and Counter, C. M. (2017). Copper chelation inhibits brafv600e-driven melanomagenesis and counters resistance to BRAFV600E and MEK1/2 inhibitors. Cancer Res. 77 (22), 6240–6252. doi[:10.1158/](https://doi.org/10.1158/0008-5472.CAN-16-1190) [0008-5472.CAN-16-1190](https://doi.org/10.1158/0008-5472.CAN-16-1190)

<span id="page-12-17"></span>Cao, H. L., Liu, Z. J., Huang, P. L., Yue, Y. L., and Xi, J. N. (2019). lncRNA-RMRP promotes proliferation, migration and invasion of bladder cancer via miR-206. Eur. Rev. Med. Pharmacol. Sci. 23 (3), 1012–1021. doi:[10.26355/](https://doi.org/10.26355/eurrev_201902_16988) [eurrev\\_201902\\_16988](https://doi.org/10.26355/eurrev_201902_16988)

<span id="page-12-22"></span>Cotton, S., Azevedo, R., Gaiteiro, C., Ferreira, D., Lima, L., Peixoto, A., et al. (2017). Targeted O-glycoproteomics explored increased sialylation and identified MUC16 as a poor prognosis biomarker in advanced-stage bladder tumours. Mol. Oncol. 11 (8), 895–912. doi:[10.1002/1878-0261.12035](https://doi.org/10.1002/1878-0261.12035)

<span id="page-12-16"></span>Dong, J., Wang, X., Xu, C., Gao, M., Wang, S., Zhang, J., et al. (2021). Inhibiting NLRP3 inflammasome activation prevents copper-induced neuropathology in a murine model of Wilson's disease. Cell Death Dis. 12 (1), 87. doi:[10.1038/s41419-](https://doi.org/10.1038/s41419-021-03397-1) [021-03397-1](https://doi.org/10.1038/s41419-021-03397-1)

<span id="page-12-7"></span>Eaton, A., Therneau, T., and Le-Rademacher, J. (2020). Designing clinical trials with (restricted) mean survival time endpoint: Practical considerations. Clin. Trials 17 (3), 285–294. doi:[10.1177/1740774520905563](https://doi.org/10.1177/1740774520905563)

<span id="page-12-8"></span>Friedman, J., Hastie, T., and Tibshirani, R. (2010). Regularization paths for generalized linear models via coordinate descent. *J. Stat. Softw.* 33 (1), 1-22. doi[:10.](https://doi.org/10.18637/jss.v033.i01) [18637/jss.v033.i01](https://doi.org/10.18637/jss.v033.i01)

<span id="page-12-20"></span>Fu, D., Lu, C., Qu, X., Li, P., Chen, K., Shan, L., et al. (2019). LncRNA TTN-AS1 regulates osteosarcoma cell apoptosis and drug resistance via the miR-

The handling editor declared a shared parent affiliation with the authors JC and JW at the time of review.

## Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

# <span id="page-12-6"></span>Supplementary material

The Supplementary Material for this article can be found online at: [https://www.frontiersin.org/articles/10.3389/fgene.](https://www.frontiersin.org/articles/10.3389/fgene.2022.1082691/full#supplementary-material) [2022.1082691/full#supplementary-material](https://www.frontiersin.org/articles/10.3389/fgene.2022.1082691/full#supplementary-material)

134-5p/MBTD1 axis. Aging (Albany NY) 11 (19), 8374–8385. doi[:10.18632/](https://doi.org/10.18632/aging.102325) [aging.102325](https://doi.org/10.18632/aging.102325)

<span id="page-12-18"></span>Fusco, M. J., West, H. J., and Walko, C. M. (2021). Antibody drug conjugates for cancer treatment. JAMA Oncol. 7 (2), 1648. doi:[10.1001/jamaoncol.2019.3552](https://doi.org/10.1001/jamaoncol.2019.3552)

<span id="page-12-21"></span>Garczyk, S., Schneider, U., Lurje, I., Becker, K., Vögeli, T. A., Gaisa, N. T., et al. (2018). ARID1A-deficiency in urothelial bladder cancer: No predictive biomarker for EZH2-inhibitor treatment response? PloS One 13 (8), e0202965. doi:[10.1371/](https://doi.org/10.1371/journal.pone.0202965) [journal.pone.0202965](https://doi.org/10.1371/journal.pone.0202965)

<span id="page-12-3"></span>Ge, E. J., Bush, A. I., Casini, A., Cobine, P. A., Cross, J. R., DeNicola, G. M., et al. (2022). Connecting copper and cancer: From transition metal signalling to metalloplasia. Nat. Rev. Cancer 22 (2), 102–113. doi:[10.1038/s41568-021-00417-2](https://doi.org/10.1038/s41568-021-00417-2)

<span id="page-12-11"></span>Gecit, I., Kavak, S., Demir, H., Gunes, M., Pirincci, N., Cetin, C., et al. (2011). Serum trace element levels in patients with bladder cancer.Asian Pac J. Cancer Prev. 12 (12), 3409–3413.

<span id="page-12-14"></span>Gou, Y., Chen, M., Li, S., Deng, J., Li, J., Fang, G., et al. (2021). Dithiocarbazatecopper complexes for bioimaging and treatment of pancreatic cancer. J. Med. Chem. 64 (9), 5485–5499. doi:[10.1021/acs.jmedchem.0c01936](https://doi.org/10.1021/acs.jmedchem.0c01936)

<span id="page-12-12"></span>Guo, K-F., Zhang, Z., Wang, J-Y., Gao, S-L., Liu, J., Zhan, B., et al. (2012). Variation of urinary and serum trace elements (Ca, Zn, Cu, Se) in bladder carcinoma in China. Asian Pac J. Cancer Prev. 13 (5), 2057–2061. doi[:10.7314/apjcp.2012.13.5.2057](https://doi.org/10.7314/apjcp.2012.13.5.2057)

<span id="page-12-0"></span>Hurst, C., Rosenberg, J., and Knowles, M. (2018). SnapShot: Bladder cancer. Cancer Cell 34 (2), 350. doi:[10.1016/j.ccell.2018.07.013](https://doi.org/10.1016/j.ccell.2018.07.013)

<span id="page-12-13"></span>Hurtado, M., Sankpal, U. T., Kaba, A., Mahammad, S., Chhabra, J., Brown, D. T., et al. (2018). Novel survivin inhibitor for suppressing pancreatic cancer cells growth via downregulating Sp1 and Sp3 transcription factors. Cell Physiol. Biochem. 51 (4), 1894–1907. doi:[10.1159/000495715](https://doi.org/10.1159/000495715)

<span id="page-12-1"></span>Ishida, S., Andreux, P., Poitry-Yamate, C., Auwerx, J., and Hanahan, D. (2013). Bioavailable copper modulates oxidative phosphorylation and growth of tumors. Proc. Natl. Acad. Sci. U. S. A. 110 (48), 19507–19512. doi:[10.1073/pnas.1318431110](https://doi.org/10.1073/pnas.1318431110)

<span id="page-12-19"></span>Jardim, D. L., Goodman, A., de Melo Gagliato, D., and Kurzrock, R. (2021). The challenges of tumor mutational burden as an immunotherapy biomarker. Cancer Cell 39 (2), 154–173. doi:[10.1016/j.ccell.2020.10.001](https://doi.org/10.1016/j.ccell.2020.10.001)

<span id="page-12-25"></span>Jiang, W., Pan, S., Chen, X., Wang, Z-W., and Zhu, X. (2021). The role of lncRNAs and circRNAs in the PD-1/PD-L1 pathway in cancer immunotherapy. Mol. Cancer 20 (1), 116. doi[:10.1186/s12943-021-01406-7](https://doi.org/10.1186/s12943-021-01406-7)

<span id="page-12-23"></span>Jiang, X., Wang, J., Deng, X., Xiong, F., Ge, J., Xiang, B., et al. (2019). Role of the tumor microenvironment in PD-L1/PD-1-mediated tumor immune escape. Mol. Cancer 18 (1), 10. doi:[10.1186/s12943-018-0928-4](https://doi.org/10.1186/s12943-018-0928-4)

<span id="page-12-24"></span>Jiang, Y., and Zhan, H. (2020). Communication between EMT and PD-L1 signaling: New insights into tumor immune evasion. Cancer Lett. 468, 72–81. doi[:10.1016/j.canlet.2019.10.013](https://doi.org/10.1016/j.canlet.2019.10.013)

<span id="page-13-21"></span>Koh, E-I., Robinson, A. E., Bandara, N., Rogers, B. E., and Henderson, J. P. (2017). Copper import in Escherichia coli by the yersiniabactin metallophore system. Nat. Chem. Biol. 13 (9), 1016–1021. doi[:10.1038/nchembio.2441](https://doi.org/10.1038/nchembio.2441)

<span id="page-13-16"></span>Konukoğlu, D., Akçay, T., Celik, C., Erözenci, A., and Erozenci, A. (1996). Urinary zinc levels in patients with superficial bladder cancer. J. Basic Clin. Physiol. Pharmacol. 7 (2), 115–119. doi[:10.1515/jbcpp.1996.7.2.115](https://doi.org/10.1515/jbcpp.1996.7.2.115)

<span id="page-13-1"></span>Lenis, A. T., Lec, P. M., Chamie, K., and Mshs, M. D. (2020). Bladder cancer: A review. JAMA 324 (19), 1980–1991. doi:[10.1001/jama.2020.17598](https://doi.org/10.1001/jama.2020.17598)

<span id="page-13-4"></span>Li, Y., Li, G., Guo, X., Yao, H., Wang, G., and Li, C. (2020). Non-coding RNA in bladder cancer. Cancer Lett. 485, 38–44. doi:[10.1016/j.canlet.2020.04.023](https://doi.org/10.1016/j.canlet.2020.04.023)

<span id="page-13-17"></span>Lin, C-N., Wang, L-H., and Shen, K-H. (2009). Determining urinary trace elements (Cu, Zn, Pb, As, and Se) in patients with bladder cancer. J. Clin. Lab. Anal. 23 (3), 192–195. doi:[10.1002/jcla.20318](https://doi.org/10.1002/jcla.20318)

<span id="page-13-12"></span>Liu, J., Wang, Y., Chu, Y., Xu, R., Zhang, D., and Wang, X. (2020). Identification of a TLR-induced four-lncRNA signature as a novel prognostic biomarker in esophageal carcinoma. Front. Cell Dev. Biol. 8, 649. doi[:10.3389/fcell.2020.00649](https://doi.org/10.3389/fcell.2020.00649)

<span id="page-13-23"></span>Luo, H., Xu, C., Le, W., Ge, B., and Wang, T. (2019). lncRNA CASC11 promotes cancer cell proliferation in bladder cancer through miRNA-150. J. Cell. Biochem. 120 (8), 13487–13493. doi[:10.1002/jcb.28622](https://doi.org/10.1002/jcb.28622)

<span id="page-13-37"></span>Luo, Y., Yang, J., Yu, J., Liu, X., Yu, C., Hu, J., et al. (2020). Long non-coding RNAs: Emerging roles in the immunosuppressive tumor microenvironment. Front. Oncol. 10, 48. doi[:10.3389/fonc.2020.00048](https://doi.org/10.3389/fonc.2020.00048)

<span id="page-13-34"></span>Marigo, I., Bosio, E., Solito, S., Mesa, C., Fernandez, A., Dolcetti, L., et al. (2010). Tumor-induced tolerance and immune suppression depend on the C/EBPbeta transcription factor. Immunity 32 (6), 790–802. doi[:10.1016/j.immuni.2010.05.010](https://doi.org/10.1016/j.immuni.2010.05.010)

<span id="page-13-2"></span>Martinez Rodriguez, R. H., Buisan Rueda, O., and Ibarz, L. (2017). Bladder cancer: Present and future. Med. Clínica Engl. Ed. 149 (10), 449–455. doi:[10.1016/j.medcli.](https://doi.org/10.1016/j.medcli.2017.06.009) [2017.06.009](https://doi.org/10.1016/j.medcli.2017.06.009)

<span id="page-13-18"></span>Mazdak, H., Yazdekhasti, F., Movahedian, A., Mirkheshti, N., and Shafieian, M. (2010). The comparative study of serum iron, copper, and zinc levels between bladder cancer patients and a control group. Int. Urol. Nephrol. 42 (1), 89–93. doi[:10.1007/s11255-009-9583-4](https://doi.org/10.1007/s11255-009-9583-4)

<span id="page-13-28"></span>McGrail, D. J., Pilié, P. G., Rashid, N. U., Voorwerk, L., Slagter, M., Kok, M., et al. (2021). High tumor mutation burden fails to predict immune checkpoint blockade response across all cancer types. Ann. Oncol. 32 (5), 661–672. doi:[10.1016/j.annonc.](https://doi.org/10.1016/j.annonc.2021.02.006) [2021.02.006](https://doi.org/10.1016/j.annonc.2021.02.006)

<span id="page-13-9"></span>Mermel, C. H., Schumacher, S. E., Hill, B., Meyerson, M. L., Beroukhim, R., and Getz, G. (2011). GISTIC2.0 facilitates sensitive and confident localization of the targets of focal somatic copy-number alteration in human cancers. Genome Biol. 12 (4), R41. doi:[10.1186/gb-2011-12-4-r41](https://doi.org/10.1186/gb-2011-12-4-r41)

<span id="page-13-29"></span>Mogi, A., and Kuwano, H. (2011). TP53 mutations in nonsmall cell lung cancer. J. Biomed. Biotechnol. 2011, 583929. doi[:10.1155/2011/583929](https://doi.org/10.1155/2011/583929)

<span id="page-13-11"></span>Nacev, B. A., Feng, L., Bagert, J. D., Lemiesz, A. E., Gao, J., Soshnev, A. A., et al. (2019). The expanding landscape of 'oncohistone' mutations in human cancers. Nature 567 (7749), 473–478. doi:[10.1038/s41586-019-1038-1](https://doi.org/10.1038/s41586-019-1038-1)

<span id="page-13-5"></span>O'Day, S. J., Eggermont, A. M. M., Chiarion-Sileni, V., Kefford, R., Grob, J. J., Mortier, L., et al. (2013). Final results of phase III SYMMETRY study: Randomized, double-blind trial of elesclomol plus paclitaxel versus paclitaxel alone as treatment for chemotherapy-naive patients with advanced melanoma. J. Clin. Oncol. 31 (9), 1211–1218. doi[:10.1200/JCO.2012.44.5585](https://doi.org/10.1200/JCO.2012.44.5585)

<span id="page-13-19"></span>Polishchuk, E. V., Merolla, A., Lichtmannegger, J., Romano, A., Indrieri, A., Ilyechova, E. Y., et al. (2019). Activation of autophagy, observed in liver tissues from patients with wilson disease and from ATP7B-deficient animals, protects hepatocytes from copper-induced apoptosis. Gastroenterology 156 (4), 1173–1189. doi[:10.1053/j.gastro.2018.11.032](https://doi.org/10.1053/j.gastro.2018.11.032)

<span id="page-13-20"></span>Ren, X., Li, Y., Zhou, Y., Hu, W., Yang, C., Jing, Q., et al. (2021). Overcoming the compensatory elevation of NRF2 renders hepatocellular carcinoma cells more vulnerable to disulfiram/copper-induced ferroptosis. Redox Biol. 46, 102122. doi[:10.1016/j.redox.2021.102122](https://doi.org/10.1016/j.redox.2021.102122)

<span id="page-13-7"></span>Sahu, D., Lotan, Y., Wittmann, B., Neri, B., and Hansel, D. E. (2017). Metabolomics analysis reveals distinct profiles of nonmuscle-invasive and muscle-invasive bladder cancer. Cancer Med. 6 (9), 2106–2120. doi[:10.1002/](https://doi.org/10.1002/cam4.1109) [cam4.1109](https://doi.org/10.1002/cam4.1109)

<span id="page-13-10"></span>Sánchez-Carbayo, M., Urrutia, M., Hernández-Cerceño, M. L., González de Buitrago, J. M., and Navajo, J. A. (2000). Cytokeratins (UBC and CYFRA 21-1) and nuclear matrix proteins (NMP22) as urine tumor markers in the diagnosis of bladder cancer. Med. Clin. Barc. 114 (10), 361–366. doi[:10.1016/s0025-7753\(00\)](https://doi.org/10.1016/s0025-7753(00)71298-x) [71298-x](https://doi.org/10.1016/s0025-7753(00)71298-x)

<span id="page-13-30"></span>Shen, L., Wu, Y., Li, A., Li, L., Shen, L., Jiang, Q., et al. (2020). LncRNA TTN-AS1 promotes endometrial cancer by sponging miR-376a-3p. Oncol. Rep. 44 (4), 1343–1354. doi:[10.3892/or.2020.7691](https://doi.org/10.3892/or.2020.7691)

<span id="page-13-31"></span>Siracusano, S., Rizzetto, R., and Porcaro, A. B. (2020). Bladder cancer genomics. Urologia 87 (2), 49–56. doi[:10.1177/0391560319899011](https://doi.org/10.1177/0391560319899011)

<span id="page-13-22"></span>Soma, S., Latimer, A. J., Chun, H., Vicary, A. C., Timbalia, S. A., Boulet, A., et al. (2018). Elesclomol restores mitochondrial function in genetic models of copper deficiency. *Proc. Natl. Acad. Sci. U. S. A.* 115 (32), [1806296115](https://doi.org/10.1073/pnas.1806296115)

<span id="page-13-8"></span>Sun, N., Liang, Y., Chen, Y., Wang, L., Li, D., Liang, Z., et al. (2019). Glutamine affects T24 bladder cancer cell proliferation by activating STAT3 through ROS and glutaminolysis. Int. J. Mol. Med. 44 (6), 2189–2200. doi[:10.3892/ijmm.2019.4385](https://doi.org/10.3892/ijmm.2019.4385)

<span id="page-13-32"></span>Sun, P., Wu, T., Sun, X., Cui, Z., Zhang, H., Xia, Q., et al. (2019). KMT2D inhibits the growth and metastasis of bladder Cancer cells by maintaining the tumor suppressor genes. Biomed. Pharmacother. 115, 108924. doi:[10.1016/j.biopha.](https://doi.org/10.1016/j.biopha.2019.108924) [2019.108924](https://doi.org/10.1016/j.biopha.2019.108924)

<span id="page-13-0"></span>Sung, H., Ferlay, J., Siegel, R. L., Laversanne, M., Soerjomataram, I., Jemal, A., et al. (2021). Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J. Clin. 71 (3), 209–249. doi:[10.3322/caac.21660](https://doi.org/10.3322/caac.21660)

<span id="page-13-13"></span>Tabassum, N., Constantin, T. A., Cereser, B., and Stebbing, J. (2020). A cell-cycle signature classifier for pan-cancer analysis. Oncogene 39 (38), 6041-6042. doi[:10.](https://doi.org/10.1038/s41388-020-01426-5) [1038/s41388-020-01426-5](https://doi.org/10.1038/s41388-020-01426-5)

<span id="page-13-6"></span>Tsvetkov, P., Coy, S., Petrova, B., Dreishpoon, M., Verma, A., Abdusamad, M., et al. (2022). Copper induces cell death by targeting lipoylated TCA cycle proteins. Science 375 (6586), 1254–1261. doi:[10.1126/science.abf0529](https://doi.org/10.1126/science.abf0529)

<span id="page-13-33"></span>Vasekar, M., Degraff, D., and Joshi, M. (2016). Immunotherapy in bladder cancer. Curr. Mol. Pharmacol. 9 (3), 242–251. doi[:10.2174/1874467208666150716120945](https://doi.org/10.2174/1874467208666150716120945)

<span id="page-13-35"></span>Vaupel, P., and Multhoff, G. (2018). Hypoxia-/HIF-1α-Driven factors of the tumor microenvironment impeding antitumor immune responses and promoting malignant progression. Adv. Exp. Med. Biol. 1072, 171–175. doi:[10.1007/978-3-319-](https://doi.org/10.1007/978-3-319-91287-5_27) [91287-5\\_27](https://doi.org/10.1007/978-3-319-91287-5_27)

<span id="page-13-25"></span>Wu, G., Sun, P., and Qin, C. (2022). GUSBP11 inhibited the progression of triple negative breast cancer via targeting the miR-579-3p/SPNS2 Axis. Cell J. 24 (5), 230–238. doi:[10.22074/cellj.2022.8024](https://doi.org/10.22074/cellj.2022.8024)

<span id="page-13-14"></span>Xie, Z., Cai, J., Sun, W., Hua, S., Wang, X., Li, A., et al. (2021). Development and validation of prognostic model in transitional bladder cancer based on inflammatory response-associated genes. *Front. Oncol.* 11, 740985 [fonc.2021.740985](https://doi.org/10.3389/fonc.2021.740985)

<span id="page-13-15"></span>Yan, Y., Cai, J., Huang, Z., Cao, X., Tang, P.,Wang, Z., et al. (2021). A novel ferroptosisrelated prognostic signature reveals macrophage infiltration and EMT status in bladder cancer. Front. Cell Dev. Biol. 9, 712230. doi[:10.3389/fcell.2021.712230](https://doi.org/10.3389/fcell.2021.712230)

<span id="page-13-36"></span>Yin, S-S., and Gao, F-H. (2020). Molecular mechanism of tumor cell immune escape mediated by CD24/siglec-10. Front. Immunol. 11, 1324. doi[:10.3389/](https://doi.org/10.3389/fimmu.2020.01324)fimmu. [2020.01324](https://doi.org/10.3389/fimmu.2020.01324)

<span id="page-13-26"></span>Zhang, X., Liu, J., Ji, M., Qi, G., and Qiao, R. (2022). Long noncoding RNA GUSBP11 knockdown alleviates nasopharyngeal carcinoma via regulating miR-1226-3p/ Axis. Cancer Biother Radiopharm [Epub ahead of print].

<span id="page-13-27"></span>Zhou, M., Zhang, Z., Bao, S., Hou, P., Yan, C., Su, J., et al. (2021). Computational recognition of lncRNA signature of tumor-infiltrating B lymphocytes with potential implications in prognosis and immunotherapy of bladder cancer. Briefings Bioinforma. 22 (3), bbaa047. doi:[10.1093/bib/bbaa047](https://doi.org/10.1093/bib/bbaa047)

<span id="page-13-24"></span>Zhou, W., Zhang, S., Li, H-B., Cai, Z., Tang, S., Chen, L-X., et al. (2020). Development of prognostic indicator based on autophagy-related lncRNA analysis in colon adenocarcinoma. BioMed Res. Int. 2020, 9807918. doi[:10.1155/2020/9807918](https://doi.org/10.1155/2020/9807918)

<span id="page-13-3"></span>Zhu, Y., Wang, S., Xi, X., Zhang, M., Liu, X., Tang, W., et al. (2021). Integrative analysis of long extracellular RNAs reveals a detection panel of noncoding RNAs for liver cancer. Theranostics 11 (1), 181–193. doi:[10.7150/thno.48206](https://doi.org/10.7150/thno.48206)