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REVIEWED BY

Prashant Kumar Singh,
Mizoram University, India
Itzel Becerra-Absalón,
National Autonomous University of
Mexico, Mexico

*CORRESPONDENCE

Guilherme Scotta Hentschke,
✉ guilherme.scotta@gmail.com
Kleber Renan de Souza Santos,
✉ santoskrs@gmail.com

†These authors share first authorship

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Molecular, morphological and ecological studies of *Limnospira platensis* (Cyanobacteria), from saline and alkaline lakes, Pantanal Biome, Brazil

Kleber Renan de Souza Santos^{1,2*†},
Guilherme Scotta Hentschke^{3*†}, Graciela Ferrari⁴,
Ana Paula Dini Andreote⁵, Marli de Fátima Fiore⁵,
Vitor Vasconcelos^{3,6} and Célia Leite Sant'Anna⁷

¹Centro Universitário Leonardo da Vinci—Uniasselvi, Indaial, Brazil, ²Fundação Universidade Regional de Blumenau (FURB), Blumenau, Brazil, ³Centro Interdisciplinar de Investigação Marinha e Ambiental (CIIMAR), Blue Biotechnology and Ecotoxicology Laboratory, Matosinhos, Portugal, ⁴Laboratório Tecnológico del Uruguay (LATU), Montevideo, Uruguay, ⁵Centro de Energia Nuclear na Agricultura (CENA), Universidade de São Paulo (USP), Piracicaba, Brazil, ⁶Faculdade de Ciências, Universidade do Porto, Porto, Portugal, ⁷Instituto de Pesquisas Ambientais (IPA), São Paulo, Brazil

We studied nineteen populations of *A. platensis* (Microcoleaceae, Cyanobacteria), from Pantanal (Brazil) shallow, saline and alkaline lakes. The lakes are connected to the Paraná and Paraguay Rivers Basins, and during wet seasons, the waters flow towards La Plata River estuary. Morphology of natural populations and cultures were analyzed using optical microscope, and 16S rDNA sequences were used for the BI and ML phylogenetic analysis. The morphological analysis shows that our populations fit in the original description of *A. platensis*, but with additional aerotopes. Also, it is evident that these structures are facultative and the species is planktonic, rather than benthic without aerotopes, as originally described. The phylogenetic analysis shows our strains in the monophyletic *Limnospira* clade. Considering that, in this paper we transfer the species *A. platensis* to the genus *Limnospira*, based on phylogenetic and morphological data. This new taxonomical combination is supported also by ecological data, and indicates that the species is more related and abundant in Pantanal, than in La Plata Basin, region from where it was originally described. According to our results, *L. platensis* is planktonic or benthic and typical from saline, alkaline and warm waters.

KEYWORDS

salines, extreme environments, biodiversity, polyphasic approach, new combination

1 Introduction

Arthrospira platensis (Nodst.) Gomont is a Cyanobacteria largely used in industrial scale, as a food supplement, under the commercial name “Spirulina”. Although it is one of the most studied Cyanobacteria, due to its economic importance, its taxonomy is still controversial (Nowicka-Krawczyk et al., 2019). In an important taxonomic revision of the genus *Arthrospira* (Komárek and Lund, 1990), the authors included *Spirulina maxima* and *S. fusiformis* in the planktonic forms (with aerotopes) of *Arthrospira*, as *A. maxima* Setchell et Gardner and *A. fusiformis* (Voronichin) Komárek et Lund. Based on an exsiccate of the type

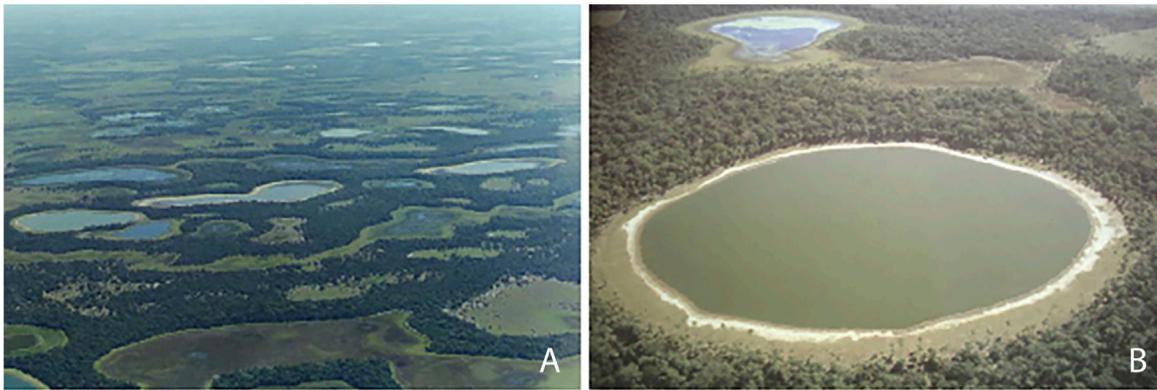


FIGURE 1
 (A) General view of Pantanal da Nhecolândia lakes, (B) Salina do Meio in Nhumirim Farm (Credits: (A). A.Y. Sakamoto 2001, (B) A. Pott, In: Pott and Pott, 2000).

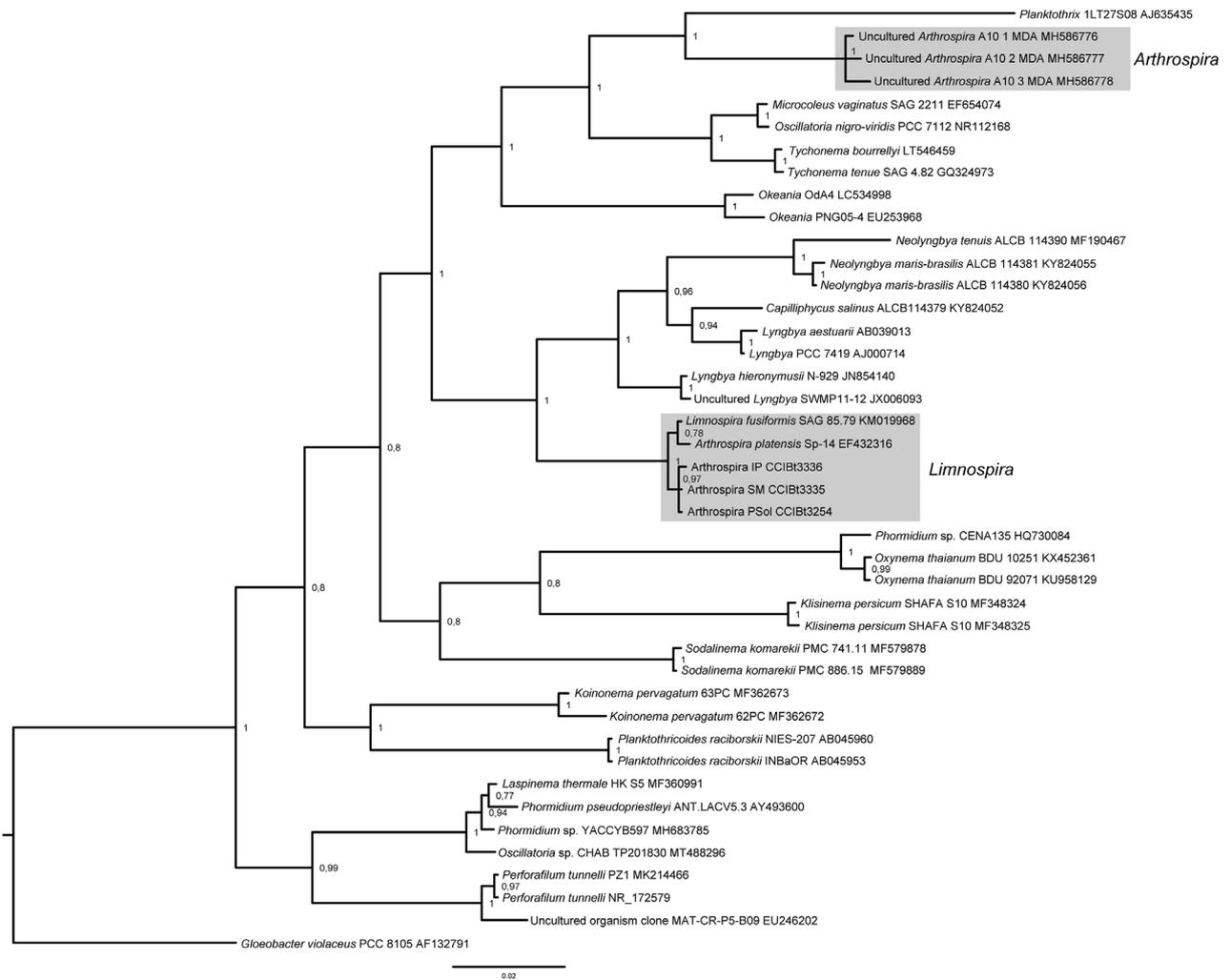
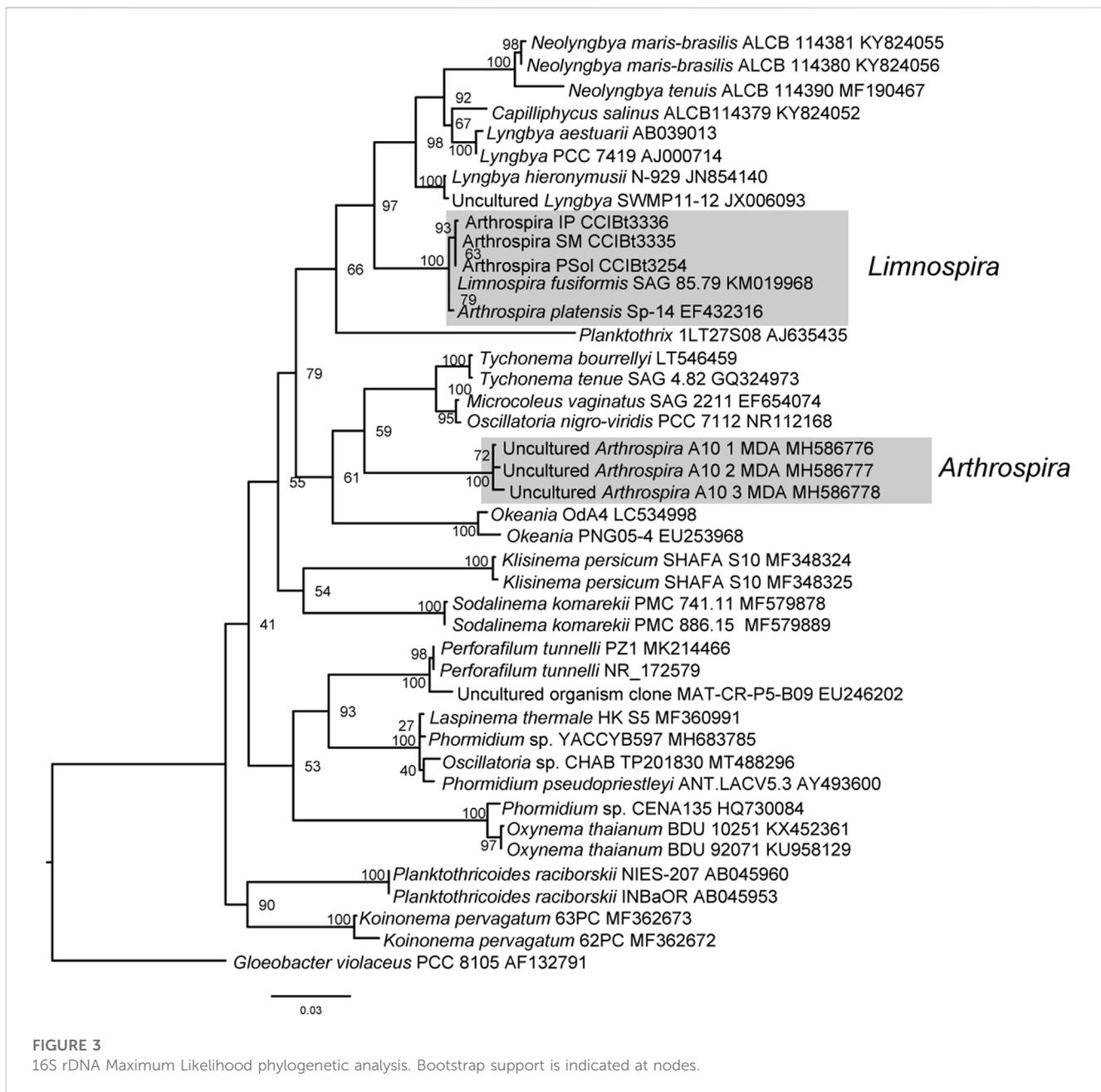


FIGURE 2
 16S rDNA Bayesian phylogenetic analysis. Posterior probabilities are indicated at nodes.



material of *A. platensis*, held at the British Museum of Natural History, the authors considered this species as benthic, without aerotopes. Some authors disagree and argue that aerotopes are lost in dry material (Tomaselli, 1997) and that the Gomont iconotype does not show all the morphological variability of the species. Because of that, *A. platensis* hypothetically could present aerotopes and be planktonic.

In this paper, we extensively studied nineteen *A. platensis* populations from saline and alkaline lakes of Pantanal da Nhecolândia, Brazil. The Pantanal Biome is located in the center of South America and is the greatest wetland in the planet (Allem and Valls, 1987). In the sub-region Pantanal da Nhecolândia, the presence of thousands of shallow lakes (maximum 2 m depth), predominantly rounded are remarkable. These lakes are commonly dominated by cyanobacterial blooms of *A. platensis* and *Anabaenopsis elenkinii* Miller.

The populations of *A. platensis* in our samples, fit in the original description of the species, but present facultative aerotopes and are phylogenetic related to the genus *Limnospira* Nowicka-Krawczyk *et al.* Consequently, based on phylogenetic, morphological and ecological data, in this paper we propose to transfer *A. platensis* to the genus *Limnospira*.

2 Materials and methods

2.1 Sampling and physicochemical water parameters

Nineteen water samples were collected in dry and wet seasons from 2004 to 2012, by immersing bottles, in “Pantanal da

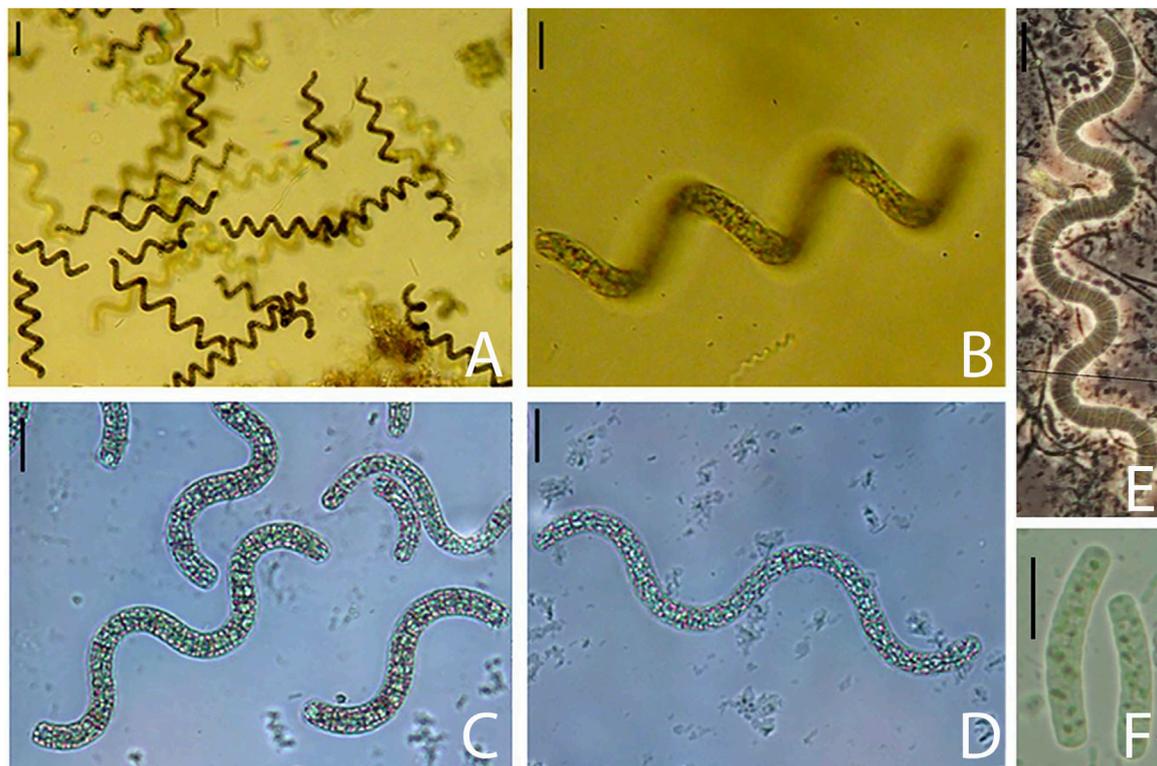


FIGURE 4
(A–D) General aspect of regularly coiled trichomes with aerotopes, (E) Trichome without aerotopes, (F) Hormogonia with granulose protoplasm and without aerotopes. Scales = 10 μm .

Nhecolândia”, Brazil, lakes. Located between Paraná and Paraguay Rivers, the region is characterized by the presence of thousands of shallow lakes (Figure 1) with alkaline (pH 9–11) and saline waters with high conductivity ($>2000 \mu\text{S cm}^{-1}$), dominated by the Cyanobacteria *A. elenkinii* and “*A. platensis*”. The lakes are part of the Paraguay River Basin, which is composed by the Paraguay-Paraná-Plata Rivers systems. During the wet seasons, the lakes are connected to this system and the waters flow towards La Plata River estuary (Santos and Sant’Anna, 2010).

For the qualitative analysis of *A. platensis* natural populations, parts of the samples were preserved with 4% formalin and later included in the collection of the State Scientific Herbarium “Maria Eneida P. Kauffman Fidalgo” (SP), in the Institute of Botany, São Paulo, Brazil. For the isolation of strains, the samples were kept under refrigeration in a Styrofoam box with ice, and then transported to the laboratory of the Institute of Botany. The isolation was made using microscope and a micropipette to transfer individuals to tubes with BG-11 (Stanier et al., 1971) liquid media, adjusted to pH 9.5 with NaOH (Santos et al., 2011). The three isolate strains were included in the Institute of Botany Culture Collection (CCIBt), São Paulo, Brazil, and kept under controlled conditions: temperature 23°C, irradiance 40–50 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$, photoperiod 14–10 h light-dark, and transferred to new flasks every 40 days. In the studied lakes, the parameters temperature, salinity, conductivity and pH were measured with a WTW 340i probe.

2.2 Morphological analysis

Morphological analysis was performed based on nature material using optical microscope. The following characters were analyzed: 1) general trichome morphology and measurements; 2) apical cells morphology and measurements; 3) cells morphology and measurements; 4) number of spirals per trichome; 5) spirals height; 6) distance between spirals; 7) presence of aerotopes; 8) presence of mucilaginous envelope using China Ink.

2.3 Molecular analysis

2.3.1 DNA extraction, PCR amplification and sequencing

DNA extraction was performed according to (Fiore et al., 2000). The 16S rDNA and the 16S-23S ITS region amplification was performed using the primers 27F1 (5'-AGAGTTTGATCC TGCTCAG-3') (Neilan et al., 1997), and 23S30R (5'-CTTCGC CTCTGTGTGCCTAGGT-3') (Lepère et al., 2000) under the following conditions: Heat 94°C/5 min; 10 cycles 94°C/45s, 57°C/45s, 72°C/2min; 25 cycles 94°C/45s, 54°C/45s, 72°C/2min and final extension 72°C/7min, and then sequenced. The inserts were cloned into “pGEM®-T Easy Vector Systems” (Promega, Madison, WI, EUA), pGEM®-T Easy Vector System (Promega,

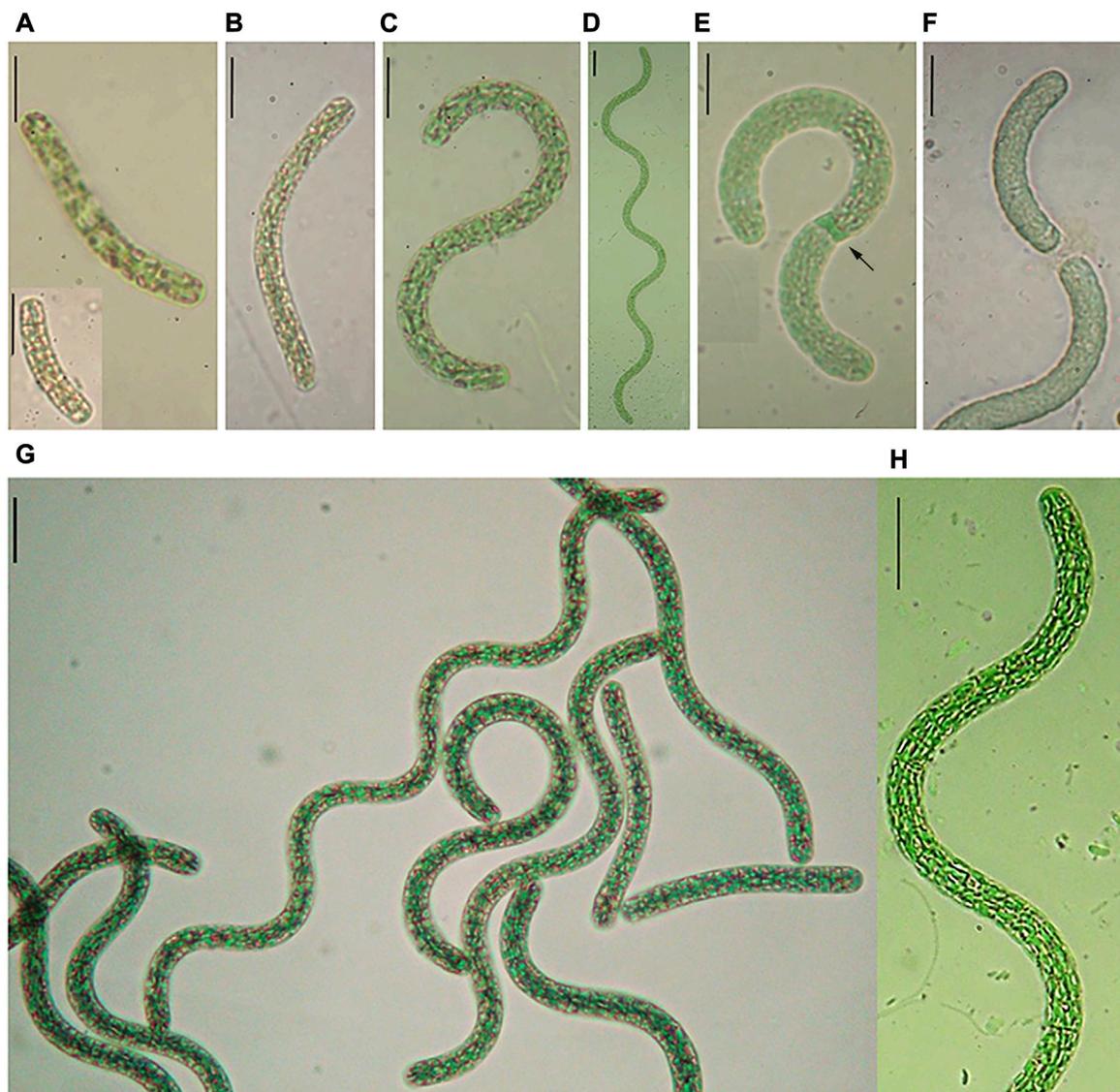
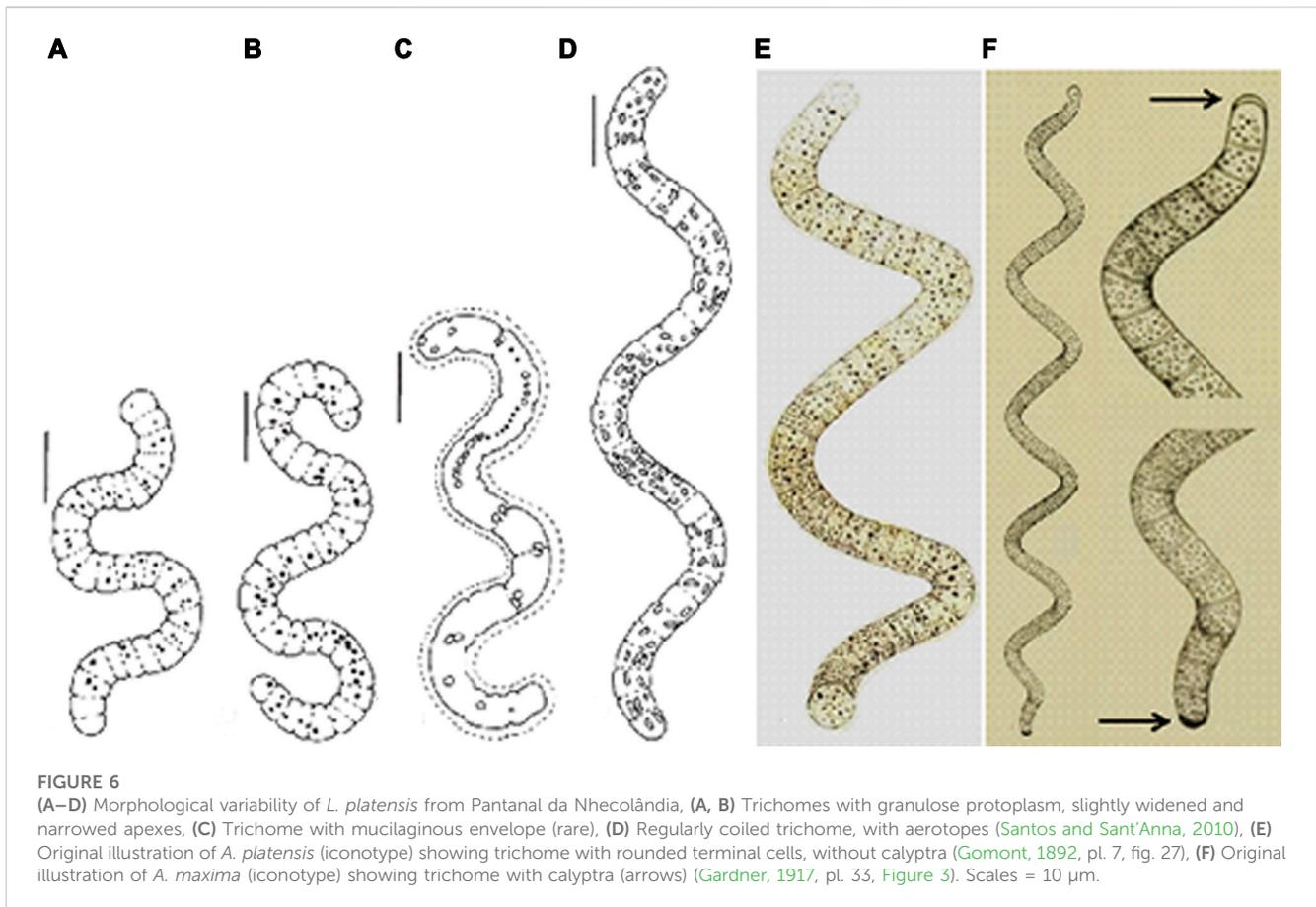


FIGURE 5
 (A–F) Life cycle of *L. platensis*, (A) Hormogonia in nature (Salina do Meio), (B, C) Hormogonia in culture, (D) Adult trichome in culture, (E) Trichome with necridium (arrow), (F) Trichome broken at necridium, (G, H) Trichome in culture.

Madison, WI, United States) according to the supplier's manual, cloned by heat-shock in *E. coli* DH5 α cells and plated for blue-white selection (Sambrook et al., 1989). Three white colonies for each strain were selected and the plasmid extractions were performed by the alkaline lysis method (Birnboim and Doly, 1979). The gene was sequenced using "Big Dye Terminator" version 3.0 (Applied Biosystems) with the plasmid primers T7 and M13 and the internal primers 357F/357R, 704F/704R and 1114F/1114R (Lane, 1991). The sequences were deposited in NCBI under the accession numbers: *Arthrospira* SM CCIBt3335 (OR142670), *Arthrospira* IP CCIBt3336 (OR142671) and *Arthrospira* PSol CCIBt3254 (OR142672).

2.3.2 Phylogenetic analysis

The 16S rDNA sequences obtained in this study were aligned using ClustalW, in MEGA11: Molecular Evolutionary Genetics Analysis version 11 (Tamura et al., 2021), with Cyanobacteria reference sequences of Microcoleaceae, Oscillatoriaceae and Desertifilaceae, retrieved from GenBank. The final dataset contained 42 sequences with 1,247 informative sites. The phylogenetic trees were built using Maximum Likelihood and Bayesian Inference analysis. GTR + G + I evolutionary model was selected by MEGA 11. The robustness of ML tree was estimated by bootstrap percentages, using 1,000 replications using IQ-Tree online version v1.6.12 (Trifinopoulos et al., 2016). Bayesian trees were constructed in two independent runs, with four chains



each, for 5×10^6 generations, burnin fraction set to 0.25, sample frequency 1,000, using MrBayes (Ronquist et al., 2012) in Cipres Gateway (Miller et al., 2010).

3 Results

3.1 Phylogenetic analysis

The Bayesian Inference Phylogeny (Figure 2) (average standard deviation of split frequencies: 0.0095), shows strong backbone support and shows our strains *L. platensis* CCIBt3335, CCIBt3336 and CCIBt3254 strongly supported (BI = 1) in the monophyletic *Limnospira* clade (Nowicka-Krawczyk et al., 2019), close related to the Microcoleaceae genera *Neolyngbya*, *Lyngbya*, *Capilliphycus* and *Limnoraphis* (BI = 1), and distant from the *Arthrospira* clade, represented by the reference strains *A. jenneri* A10 MDA (Nowicka-Krawczyk et al., 2019). The Maximum Likelihood phylogeny (Figure 3), although presents weak backbone support, corroborates with the BI phylogeny, showing Pantanal strains in the *Limnospira* clade (bootstrap 99), again related to that same Microcoleaceae genera. In both BI and ML trees *Arthrospira* clade is related to *Microcoleus*, *Tychonema* and *Okeania*.

3.2 Description of species

Limnospira platensis (Gomont) K. R. S. Santos and G. S. Hentschke comb. nov. (Figures 4–6).

Basonym: *A. platensis* Gomont 1892: 247, pl. VII: fig. 27.

Published in: Gomont, M. (1892/1893'). Monographie des Oscillariées (Nostocacées Homocystées). Deuxième partie—Lyngbyées. Annales des Sciences Naturelles, Botanique, Série 7 16: 91–264, pls 1–7.

Trichomes solitary, regularly coiled when adult, constricted or not, 35–151 (–371) μ m long, rarely with mucilaginous envelope. Spirals height 16–32.4 μ m; distance between spirals 22–40.6 μ m. Cells shorter than wide (rarely longer than wide), 1.7–5.3 (–9) μ m long and 4–7.8 μ m wide. Ratio L/W = 0.5–1.3. Apical cells rounded, slightly narrowed (rarely widened). Cell content blue-green, granule, with facultative aerotopes.

Habitat: Saline and alkaline lakes, planktonic and more rarely benthic (part of the life cycle).

Reference strain: *Arthrospira* PSol CCIBt3254 (OR142672)

Studied material: BRAZIL. Mato Grosso do Sul: Corumbá, Pantanal da Nhecolândia, Salina do Meio, 09/25/2005, K.R.S. Santos and C.F.S. Malone (SP390917), 04/22/2006, K.R.S. Santos (SP390919), 08/28/2006, K.R.S. Santos (SP390922), 05/04/2007, K.R.S. Santos (SP390927), 08/19/2009, K.R.S. Santos (SP400654),

TABLE 1 Sampling location, geographical coordinates, Herbarium identification and physicochemical parameters of Pantanal da Nhecolândia lakes.

| Lake | Coordinates | Herbarium ID | Sampling date | pH | Electrical conductivity ($\mu\text{S}/\text{cm}$) | Temp. ($^{\circ}\text{C}$) |
|----------------------|------------------------|--------------|---------------|-------|---|------------------------------|
| Salina do Meio | 18°58'29"S, 56°38'47"W | SP400654 | 19/08/2009 | 9.47 | 10,600 | 32.5 |
| Salina do Meio | 18°58'29"S, 56°38'47"W | SP427290 | 27/10/2011 | 9.89 | 4,060 | 29.7 |
| Salina do Meio | 18°58'29"S, 56°38'47"W | SP427740 | 06/05/2012 | 9.64 | 8,770 | - |
| Salina do Meio | 18°58'29"S, 56°38'47"W | SP390917 | 25/09/2005 | 9.85 | 19,020 | 23.3 |
| Salina do Meio | 18°58'29"S, 56°38'47"W | SP390919 | 22/04/2006 | 10.16 | 2,870 | 33.3 |
| Salina do Meio | 18°58'29"S, 56°38'47"W | SP390922 | 28/08/2006 | 10.09 | 12,070 | 24.7 |
| Salina do Meio | 18°58'29"S, 56°38'47"W | SP390927 | 04/05/2007 | 10.19 | 3,890 | 31.0 |
| Salina da Reserva | 18°57'35"S, 56°37'18"W | SP400842 | 09/05/2005 | - | 2,380 | 22.9 |
| Salina da Reserva | 18°57'35"S, 56°37'18"W | SP401692 | 19/08/2009 | 9.68 | 11,040 | 34.1 |
| Salina da Reserva | 18°57'35"S, 56°37'18"W | SP427741 | 06/05/2012 | 10.10 | 5,435 | - |
| Salina da Ponta | 18°58'56"S, 56°39'33"W | SP400843 | 25/09/2005 | 9.9 | 5,790 | 23.8 |
| Salina da Ponta | 18°58'56"S, 56°39'33"W | SP400845 | 22/04/2006 | 9.8 | 864 | 32.8 |
| Salina da Ponta | 18°58'56"S, 56°39'33"W | SP400487 | 28/08/2006 | 9.8 | 8,970 | 22.8 |
| Salina da Ponta | 18°58'56"S, 56°39'33"W | SP400849 | 17/11/2006 | 9.9 | 716 | 30.8 |
| Salina da Ponta | 18°58'56"S, 56°39'33"W | SP401691 | 19/08/2009 | 9.22 | 8,130 | 30.4 |
| Salina Pedra do Sol | 19°10'36"S, 56°57'44"W | SP427742 | 26/08/2006 | 10.06 | 2,100 | 28.0 |
| Salina Pedra do Sol | 19°10'36"S, 56°57'44"W | SP427743 | 16/11/2006 | 10.23 | 12,200 | 32.0 |
| Salina Pantanal | 18°55'40"S, 56°33'03"W | SP427747 | 16/08/2009 | - | - | - |
| Salitrada Campo Dora | 18°58'05"S, 56°38'58"W | SP390925 | 16/11/2006 | 8.42 | 1852 | 32.0 |

10/27/2011, K.R.S. Santos (SP427290), 05/06/2012, C.F.S. Malone and C.L. Sant'Anna (SP427740); Salina Pantanal, 08/16/2009, K.R.S. Santos (SP427747); Salina da Reserva, 08/19/2009, K.R.S. Santos (SP401692), 05/06/2012, C.F.S. Malone and C.L. Sant'Anna (SP427741); Salina da Ponta, 08/19/2009, K.R.S. Santos (SP401691); Salina Pedra do Sol, 08/26/2006, K.R.S. Santos (SP427742), 11/16/2006, K.R.S. Santos (SP427743).

Comments: *L. platensis* is morphologically different from the other species of the genus, *L. fusiformis*, *L. indica* and *L. maxima*. *L. platensis* is the only species of the genus without calyptra. Furthermore, *L. fusiformis* presents irregular spirals, while the spirals of *L. platensis* are regular. Although the presence of calyptra in *L. maxima*, this is the most similar species to *A. platensis* and the differences are discussed below. According to Komárek and Anagnostidis (2005), the relations between *L. maxima* and *L. indica* are unclear, and maybe these species are synonyms. When comparing our *L. platensis* cultures with the nature material, we did not find significant morphological differences between them.

3.3 Occurrence of *L. platensis* and physicochemical parameters

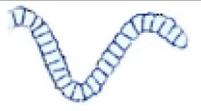
L. platensis was found forming blooms with *A. elenkinii* Miller in saline and alkaline lakes in "Pantanal da Nhecolândia". Table 1 shows all the studied samples and the

respective physicochemical parameters. The species occurred in Pantanal saline lakes with pH ranging from 9.2 to 10.2, electrical conductivity from 716 to 19020 $\mu\text{S cm}^{-1}$ and temperature from 22.8 to 33.3°C. In Lagoa Salitrada Campo Dora the species was registered only once, in the dry season (pH 8.4, electrical conductivity 1852 $\mu\text{S cm}^{-1}$ and temperature 32°C). These data evidence the dependence of the alkaline pH, high electrical conductivity and warm temperatures for the growing of *L. platensis*. These conditions also indicate that high nutrient levels are fundamental for the species.

4 Discussion

Arthrospira and *Limnospira* are monophyletic genera, morphologically distinguished from each other by the presence of aerotopes in *Limnospira* (Nowicka-Krawczyk et al., 2019). Aerotopes were commonly observed in our nature samples and isolate strains, and we also observed that this character was facultative in all or our samples and cultures. Apart of the facultative aerotopes, our specimens fit perfectly in the original description of *A. platensis* (Gomont, 1892). Figure 5 and Table 2 summarize the morphological similarity between our populations and *A. platensis*, and show the differences between this species and *L. maxima*, *A. platensis* presents smaller trichomes and the terminal cells lack calyptra, while *L. maxima* presents larger trichomes and presents calyptra at the terminal cells.

TABLE 2 Morphological characters of “*A. platensis*” and its most similar species *A. maxima* according to different authors.

| Species | Aerotopes/habit | Trichome constriction | Trichome apex | Trichome width (μm) | Cell length (μm) | Spirals height (μm) | Distance between spirals (μm) | Location and sampling date | Figure |
|---|----------------------------|---------------------------|---|---------------------|--|---------------------|-------------------------------|--|---|
| <i>A. platensis</i> (Nordstedt) Gomont (1892) original description | ?/? | Slightly constrict | Slightly or not narrowed. Rounded, without calyptra | 6–8 | 2–6 | 26–36 | 43–57 | Montevideo, Uruguay, 1884 |  |
| <i>A. platensis</i> sensu Santos and Sant’Anna (2010) and present study | Facultative/planktonic | Constrict or not | Slightly or not narrowed. Rounded without calyptra | 4–7.8 | 1.7–5.3 (9) | 16.0–32.4 | 22.0–40.6 | Saline lakes at Pantanal da Nhecolândia, Brazil; 2004–2012 |  |
| <i>A. platensis</i> sensu Komárek and Anagnostidis (2005) | Absent/periphytic, benthic | Slightly constrict | Slightly or not narrowed. Rounded with calyptra | (4) 6–7 (9?) | - | (20?) 26–36 | (24?) 30–57 | South America |  |
| <i>A. platensis</i> sensu Komárek and Lund 1990 | Absent/benthic | Slightly constrict | Slightly narrowed | (4) 6–7 (8?) | - | 26–36 | 30–57 | South America |  |
| <i>Arthrospira maxima</i> Gardner (1917) | Present/planktonic | Not constrict | Slightly narrowed with calyptra | 7–9 | 5–7 | 40–60 | 70–80 | Warm saline lakes at South California, Key Route Powerhouse, Oakland (United States of America) 1916 |  |
| <i>Arthrospira maxima</i> sensu Komárek and Komárková-Legnerová (2002) | Present/planktonic | Slightly or not constrict | Slightly or not narrowed with calyptra | 8–(12) | Shorter than wide, rarely isodiametric | (34.5)–51 | ±(34.5)–51 | Vulcanic alkaline lakes at Atlacoya (Tecuitlapa) e El Rincon de Parangueo, México; 1992–1993 |  |

? = not reported.

According to the original description, based on an herbarium exsiccate, *A. platensis* presents cylindrical trichomes, not narrowed toward the ends or slightly narrowed, 6–8 µm wide; spiral height 26–36 µm, distance between spirals 43–57 µm; cells isodiametric 2–6 µm long with granulose protoplasm. The author does not mention the occurrence of aerotopes, nor the habits benthic or planktonic for this species described from La Plata Basin. A brief indication of the species habit is described as “greenish trichomes, forming a thin layered stratum” (free translation from Latin). These morphological and habit descriptions do not exclude the possibility of the presence of aerotopes and the planktonic habit, but make these characters uncertain, considering that the specimens analyzed were from dried material. The expression “thin layered stratum” does not specify where it was sampled and also, at that time, planktonic material was not analyzed, so the author didn’t know if specimens were also growing in phytoplankton. Consequently, we consider the original description poor and encompassing only a little portion of the morphological and ecological plasticity of the species.

Komárek and Lund (1990), also based on the type material in herbarium exsiccate, considered *A. platensis* as benthic and without aerotopes. However, to consider the species benthic based on the lack of aerotopes in dry preserved specimens is questionable. Currently it is known that aerotopes collapse in dry material (Tomaselli, 1997), and probably that is the reason why these authors (and Gomont) do not mention the presence of these structures.

Considering that, all of our studied populations from nineteen lakes, fit in the original description of *A. platensis*. All populations presented facultative aerotopes, and because of that, in this paper, we add this character to the species description. This is in agreement with Jeeji-Bai (1999) genus revision, which also report aerotopes for *A. platensis*. Moreover, the benthic habit was stated by (Komárek and Lund, 1990) as a consequence of the lack of aerotopes, however, in our observations it is clear that the populations are planktonic.

Our ecological data corroborates with the morphological analysis. Although Pantanal and La Plata Basin are geographically distant, they are connected by the rivers Paraguay and Paraná. Pantanal is a corridor connecting the Amazon and La Plata Basins, and during the rainy seasons, the lakes are connected to these basins. The rivers flow to the South carrying a great diversity of species to La Plata estuary, which presents also high salinity, as observed by us in Pantanal lakes (Alho and Gonçalves, 2005; Junk and Cunha, 2005). Another important fact that can explain the presence of *A. platensis* in La Plata Basin, is that many birds migrate from this area to Pantanal every year for reproduction, and can carry the microbiota in their feathers. It is impossible to know from which environment the original described population was sampled, but we hypothesize that it could be found in temporary small pools (forming stratum), with higher salinity, pH and/or temperature. Clearly, *A. platensis* is not typical of La Plata Basin, considering that after the description of this species in 1892, even with many sampling efforts of Argentinian and Uruguayan research groups, no populations of *A. platensis* were found in this region.

The fact that *A. platensis* was never reported for La Plata Basin after the original description indicates that this species is more related to the salines of Pantanal, due to its abundance and wide distribution in this Biome. According to our results, *L. platensis* is typical from alkaline, saline, nutrient rich and warm environments

and this can explain why the species is rare in La Plata Basin. The species is reported worldwide, but its actual distribution is unknown, mainly because of misidentification of populations in Europe, America, Africa and Asia (Sili et al., 2012).

According to these statements and our phylogenetic analysis showing our strains in *Limnospira* clade, we conclude that *A. platensis* has to be combined to *L. platensis*. Also, we recommend to use *Arthrospira* PSol CCIBt3254 (OR142672) as the reference strain for the species.

Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found below: GenBank OR142670-OR142672.

Author contributions

KS (Article initiative, generated morphological data, described the species, collaborated in discussion section) GH (Article initiative, wrote the paper) GF (collaborated in discussion section) AA (generated molecular data) MF (generated molecular data) VV (collaborated in discussion section, revised the manuscript) CS’A (Author of the project, revised the manuscript). All authors contributed to the article and approved the submitted version.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

- Alho, C. J. R., and Gonçalves, H. C. (2005). *Biodiversidade do pantanal: Ecologia and conservação*. Campo Grande: UNIDERP.
- Allem, A. C., and Valls, J. F. M. (1987). *Recursos forrageiros nativos do Pantanal Mato-grossense*. Brasília: Empresa Brasileira Agropecuária/Centro Nacional de Recursos Genéticos.
- Birnboim, H. C., and Doly, J. (1979). A rapid alkaline extraction procedure for screening recombinant plasmid DNA. *Nucleic Acids Res.* 6, 1513–1523. doi:10.1093/nar/7.6.1513
- Fiore, M. F., Moon, D. H., Tsai, S. M., Lee, H., and Trevors, J. T. (2000). Miniprep DNA isolation from unicellular and filamentous cyanobacteria. *J. Microbiol. Methods* 39, 159–169. doi:10.1016/s0167-7012(99)00110-4
- Gardner, N. L. (1917). New Pacific coast marine algae I. *Univ. Calif. Publ. Bot.* 6, 377–416.
- Gomont, M. (1892). Monographie des Oscillariées (Nostocacées Homocystées). Deuxième partie. - lynchées. *Annales des Sciences Naturelles. Botanique* 7, 91–264.
- Jeeji-Bai, N. (1999). “A taxonomic revision of the genus *Arthrospira* based on certain new criteria,” in *Marine Cyanobacteria*. Editors L. Charpy and A. W. D. Larkum (Monaco: Bulletin of the Institute of Oceanography), 47–52.
- Junk, W. J., and Cunha, C. N. (2005). Pantanal: A large South American wetland at a crossroads. *Ecol. Eng.* 24, 391–401. doi:10.1016/j.ecoleng.2004.11.012
- Komárek, J., and Anagnostidis, K. (2005). “Cyanoprokaryota 2. Teil: Oscillatoriales,” in *Süßwasserflora von Mitteleuropa 19*. Editors B. Büdel, L. Krienitz, G. Gärtner, and M. Schagerl (Elsevier Spektrum Akademischer Verlag), 1–759.
- Komárek, J., and Komárková-Legnerová, J. (2002). Contribution to the knowledge of planktic cyanoprokaryotes from central Mexico. *Preslia* 74, 207–233.
- Komárek, J., and Lund, J. W. G. (1990). What is “*Spirulina platensis*” in fact? *Arch. Hydrobiol. Suppl. Algal. Stud.* 58, 1–13.
- Lane, D. J. (1991). “16S/23S rRNA sequencing,” in *Nucleic acid techniques in bacterial systematics*. Editors E. Stackebrandt and M. Goodfellow (Hoboken: Wiley and Sons), 115–175.
- Lepère, C., Wilmotte, A., and Meyer, B. (2000). Molecular diversity of *Microcystis* strains (Cyanophyceae, Chroococcales) based on 16S rDNA sequences. *Syst. Geogr. Plants* 70, 275–283. doi:10.2307/3668646
- Miller, M. A., Pfeiffer, W., and Schwartz, T. (2010). “Creating the CIPRES science gateway for inference of large phylogenetic trees,” in *Proceedings of the Gateway computing environments workshop (GCE)*, 1–8.
- Neilan, B. A., Jacobs, D., Dot, T. D., Blackall, L. L., Hawkins, P. R., Cox, P. T., et al. (1997). rRNA sequences and evolutionary relationships among toxic and nontoxic cyanobacteria of the genus *microcystis*. International union of microbiological societies. Available at: www.microbiologyresearch.org.
- Nowicka-Krawczyk, P., Mühlsteinová, R., and Hauer, T. (2019). Detailed characterization of the *Arthrospira* type species separating commercially grown taxa into the new genus *Limnospira* (Cyanobacteria). *Sci. Rep.* 9, 694. doi:10.1038/s41598-018-36831-0
- Pott, V. J., and Pott, A. (2000). *Plantas aquáticas do Pantanal*. Corumbá: EMBRAPA.
- Ronquist, F., Teslenko, M., Van Der Mark, P., Ayres, D. L., Darling, A., Höhna, S., et al. (2012). MrBayes 3.2: Efficient bayesian phylogenetic inference and model choice across a large model space. *Syst. Biol.* 61, 539–542. doi:10.1093/sysbio/sys029
- Sambrook, J., Fritsch, E. F., and Maniatis, T. (1989). *Molecular cloning: A laboratory manual*. 2nd ed. Cold Harbor: Cold Spring Harbor Laboratory Press.
- Santos, K. R. S., Jacinavicius, F. R., and Sant’Anna, C. L. (2011). Effects of the pH on growth and morphology of *Anabaenopsis elenkinii* Miller (Cyanobacteria) isolated from the alkaline shallow lake of the Brazilian Pantanal. *Fottea* 11 (1), 119–126. doi:10.5507/fot.2011.012
- Santos, K. R. S., and Sant’Anna, C. L. (2010). Cianobactérias de diferentes tipos de lagoas (“salina”, “salitrada” e “baía”) representativas do Pantanal da Nhecolândia, MS, Brasil. *Rev. Bras. Botânica* 33, 61–83. doi:10.1590/s0100-84042010000100007
- Sili, C., Giuseppe, T., and Vonshak, A. (2012). “*Arthrospira* (Spirulina),” in *Ecology of Cyanobacteria II: Their diversity in space and time*. Editor B. A. Whitton (Durham: School of Biological and Biomedical Sciences, Durham University), 677–705.
- Stanier, R. Y., Kunisawa, R., Mandel, M., and Cohen-Bazire, G. (1971). Purification and properties of unicellular blue-green algae (order Chroococcales). *Bacteriol. Rev.* 35, 171–205. doi:10.1128/br.35.2.171-205.1971
- Tamura, K., Stecher, G., and Kumar, S. (2021). MEGA11: Molecular evolutionary Genetics analysis version 11. *Mol. Biol. Evol.* 38, 3022–3027. doi:10.1093/molbev/msab120
- Tomaselli, L. (1997). “Morphology, ultrastructure and taxonomy of *Arthrospira* (Spirulina) maxima and *Arthrospira* (Spirulina) platensis,” in *Spirulina platensis (Arthrospira): Physiology, cell-biology and biotechnology*. Editor A. Vonshak (London: Taylor and Francis), 79–99.
- Trifinopoulos, J., Nguyen, L. T., Von Haeseler, A., and Minh, B. Q. (2016). W-IQ-TREE: A fast online phylogenetic tool for maximum likelihood analysis. *Nucleic Acids Res.* 44, W232–W235. doi:10.1093/nar/gkw256