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Soil profile N₂O efflux from a cotton field in arid Northwestern China in response to irrigation and nitrogen management

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It remains uncertain how different N inputs as synthetic fertilizer or manure and irrigation types affect nitrous oxide (N₂O) production and effluxes in the subsurface. A field trial was carried out in 2016 to evaluate the impacts of conventional urea, animal manure, and a 50/50 mix of urea and manure on N₂O production/effluxes from a cotton (*Gossypium hirsutum* L.) field under flood or drip irrigation in northwestern China. Soil N₂O concentrations were monitored at 5, 15, 30, and 60 cm depths to assess the production and diffusion rates of N₂O in the soil profile. The results showed that N₂O concentrations in 0–60 cm ranged between 221 and 532 nL L⁻¹ and averaged 344 nL L⁻¹, which was generally lower compared to other studies in the same region. Manure and flood irrigation significantly increased N₂O production at 0–5 cm and 5–15 cm, respectively. That is, the effects of nitrogen management and irrigation types on the N₂O production of the profile were reflected in the surface layers and subsurface layers, respectively. All N₂O production occurred in the 0–15 cm layer, with the 0–5 cm depth contributing 87%–100% of the surface emissions. The response discrepancy of N₂O production/diffusion to irrigation and nitrogen management in different soil depths should be fully considered in developing agricultural N₂O emission reduction measures.

KEYWORDS

profile N₂O concentration, concentration-gradient method, nitrous oxide, drip irrigation, manure

1 Introduction

Nitrous oxide (N₂O) is a powerful greenhouse gas that contributes to both global warming and stratospheric ozone destruction (Ravishankara et al., 2009). Soil N₂O emissions have rapidly increased from 6.3 Tg N₂O-N yr⁻¹ in pre-industrial times to 10.0 Tg N₂O-N yr⁻¹ in recent years, with 82% of the total increase coming from cropland (Tian et al., 2019). Application of manure and synthetic fertilizer is the main factor inducing N₂O emissions from agricultural soils (Tian et al., 2020). In addition, irrigation practice is also a crucial factor in determining N₂O emissions from agricultural ecosystems (Kuang et al., 2021).

Thus, it is essential to explore the effects of fertilization and irrigation methods on the production and emission of N_2O .

Irrigation management practices affect N_2O emissions through their impacts on the spatial and temporal distribution of soil moisture content, as well as microbial and nutrient availability (Sánchez-Martín et al., 2008; Kuang et al., 2018). Drip irrigation is an effective practice to enhance N and water use efficiency, and it is widely used in arid regions for crop production (Vázquez et al., 2006). Some studies reported that drip irrigation effectively reduced N_2O emissions from cropland, compared with traditional irrigation (Sánchez-Martín et al., 2008; Bronson et al., 2018; Li et al., 2014), whereas in others the opposite was observed (Fentabil et al., 2016; Kuang et al., 2018). A global meta-analysis showed that drip irrigation significantly reduced N_2O emissions from cropland by 32% and 46% compared to traditional flood and sprinkler irrigation, respectively (Kuang et al., 2021). Using a soil column incubation study, Kuang et al. (2019) revealed that deep-placed N fertilizers were most susceptible to denitrification under high water-filled pore space (WFPS) content but did not result in a significant surface burst of N_2O emission, suggesting soil moisture plays an essential role in determining production and consumption of N_2O across soil profiles. Understanding the effects of different management measures on soil N_2O production processes can provide a basis for optimizing agricultural management practices.

N fertilizer and manure additions are the main cause of N_2O emissions from agricultural land (Tian et al., 2020). Soil properties, including the form of N and available C can affect the biological processes of nitrification and denitrification (Velthof et al., 2003). The effects of N sources on N_2O production and consumption have highly complex regulatory mechanisms (Zhou et al., 2017). Several studies have found that manure addition increased N_2O production compared to synthetic N fertilizers by providing C substrate to denitrifiers for denitrification (Hayakawa et al., 2009; Ju et al., 2011; Forte et al., 2017; Yin et al., 2019). Other studies, on the other hand, have found that manure application reduces N_2O emissions when compared to synthetic fertilizers, by stimulating complete denitrification to N_2 (Ball et al., 2004; Mejjide et al., 2007; Tao et al., 2018). Several studies also found no differences in the use of manure and synthetic N fertilizer in terms of N_2O emissions (Meng et al., 2005; Vallejo et al., 2006). These inconsistent results reflect the need for further analysis of the effects of different N sources on N_2O production, transport, and consumption in the profile. As a result, there remains a scarcity of knowledge on the relationship between N_2O efflux underground and emissions on the surface, which are influenced by N fertilizer and manure with the drip- and flood-irrigated crops.

Surface N_2O emissions are the net result of a series of processes involving profile N_2O production, diffusion and consumption (Gao et al., 2014; Wang et al., 2018; Li et al., 2021). The rate and direction of N_2O diffusion are determined by the distribution of N_2O concentration in the profile. Depending on the concentration gradient of N_2O in the subsoil, its association with soil surface emission rates can be used to quantify the contribution of N_2O from different soil layers to surface N_2O emissions (Nan et al., 2016; Wang et al., 2018). Nan et al. (2016) reported that 99% of the total cumulative N_2O fluxes in the soil profile occurred in the 0–15 cm soil layer. According to Wang et al. (2018), soil N_2O consumption at depths of 0–5 and 5–15 cm attributed to 80.4% and 6.6% of the

surface N_2O emission, respectively. However, few studies have been conducted to compare the contribution of different soil layers to surface N_2O emissions under different irrigation and N management practices.

The objectives of this study were to 1) characterize the spatial distributions of N_2O concentrations in the soil profile with urea or manure application under drip and flood irrigation, 2) quantify the depth-dependent contributions of profile N_2O effluxes to the surface emissions, and 3) assess the impact of environmental factors on N_2O fluxes in the profile.

2 Materials and methods

2.1 Site description and soil properties

A field experiment was carried out at the Cele National Station (37°01'06"N, 80°43'48"E) in Xinjiang Uygur Autonomous Region, during the 2016 growing season. The station is situated on the southern edge of the Taklimakan Desert. The mean annual precipitation and annual potential evaporation are 42.5 mm and 2,956 mm, respectively. The average annual air temperature is 12.7°C. The soil is classified as Aridisols in the USDA ST system (USDA, 1999), and the surface soil (0–20 cm) has sand, silt, and clay content of 900, 40, and 60 g kg⁻¹, respectively. For details about the soil properties as shown in Table 1.

2.2 Experimental design and crop management

This experiment was a two-factor experiment in a randomized complete block design with two types of irrigation (drip and flood) and four N source treatments: 1) no fertilization (Control), 2) granular urea (Urea), 3) animal manure (Manure), and 4) 50% granular urea with 50% animal manure (U + M). Detailed information about the experimental design has been described in our previous study (Kuang et al., 2018). Briefly, the application rate of all fertilizer treatments was 240 kg N ha⁻¹. Granular urea (N 46%) was applied as 522 and 261 kg ha⁻¹ for Urea and U + M, animal manure was applied as 76.9 and 38.5 Mg ha⁻¹ for Manure and U + M, respectively. Under drip irrigation, 20% urea was applied at planting, with the remaining 80% applied as a topdressing six times during the growing season. Under flood irrigation, 30% of the urea was applied during planting, and the remaining 70% was top-dressed to the soil four times before irrigation. Under both irrigation systems, manure was evenly broadcast over the surface soil before sowing and immediately incorporated with the soil. For all plots, calcium phosphate (120 kg P₂O₅ ha⁻¹) and K₂SO₄ (60 kg K₂O ha⁻¹) were broadcast on the surface and mixed into soils (0–20 cm) with a rota-cultivator before planting. Each treatment had four replicated plots. In total, 32 plots were set up in our study, each plot with an area of 32 m² (10 m × 6.4 m).

Planting and crop management were described in our previous studies (Kuang et al., 2018; Li et al., 2020). Briefly, plastic film was used to cover four cotton rows with row spacing of 30–50–30 cm. For each plastic film in drip-irrigated treatments, drip tapes were installed between two cotton rows (30 cm apart) and the distance

TABLE 1 Soil properties at different layers (0–60 cm) of profiles before sowing in 2016. Values are means ± 1 standard error, $n = 4$.

Soil depth	Nitrate-N (NO_3^- -N)	Total organic C	Bulk density	pH (H_2O)
cm	mg N kg^{-1}	g kg^{-1}	g cm^{-3}	
0–10	37.2 \pm 4.40 a	5.5 \pm 0.40 a	1.46 \pm 0.01 c	6.53 \pm 0.10 c
10–20	27.3 \pm 1.70 ab	5.3 \pm 0.10 a	1.48 \pm 0.00 b	7.01 \pm 0.11 b
20–40	28.5 \pm 2.50 ab	5.2 \pm 0.10 a	1.51 \pm 0.01 b	7.22 \pm 0.13 b
40–65	20.6 \pm 1.10 b	1.6 \pm 0.10 b	1.54 \pm 0.00 a	7.65 \pm 0.02 a

For each treatment factor, means within a column followed by the same letter are not significantly different at $p < 0.05$.

between every two emitters was 10 cm. The water flow rate in emitter was 2–3 L h^{-1} . In each plot, water and fertilizer-integrated tanks were placed to record the amount of irrigation and urea application. Over the experimental period, cotton in the drip irrigation system had received 9 times of irrigation, with each irrigation providing approximately 45 mm water. In contrast, cotton in the flood irrigation system had received 7 times irrigation of approximately 140 mm water for each irrigation.

2.3 Soil N_2O gas sampling and analysis

2.3.1 Surface N_2O emissions

The static chamber method was used to monitor soil surface N_2O flux (Kuang et al., 2018). The sampling frequency was once or twice per week to make sure a sampling was done within 1–2 days after irrigation and fertilization events. The sampling time was 10:00–14:00 (GMT+8) during the day, and N_2O concentration in gas samples during this time period was used to represent the daily average.

2.3.2 Profile N_2O collection

Soil profile N_2O concentration was measured simultaneously with the surface N_2O emissions. Soil profile N_2O gas at depths of 5, 15, 30, and 60 cm were collected using an *in-situ* soil profile gas sampler (for more details, see Kuang et al. (2019)). Briefly, the gas sampler was composed of four individual silicone tubes (5.0 cm long, 36.8 mm i. d., 40.0 mm o. d.) that air but not water can go through and sealed at both ends. The silicone tube was covered by a polyethylene (PE) pipe (5.0 cm long, 40.8 mm i. d., 50.0 mm o. d.) to determine the soil N_2O gas sampling depth by the holes in the wall of the PE pipe. A hollow stainless-steel tube (0.6 mm i. d.) with a sampling port was used to collect soil N_2O gas at each depth.

In each plot, one soil profile sampler was installed between cotton rows. 35 ml gas samples from each soil depth were collected through the corresponding sampling port using the disposable airtight syringe. The gas sample was then injected into pre-evacuated 35-ml gas-tight aluminum bags (Hede Technologies, Dalian, China). In total, gas samplings were performed 13 times between 14 May and 9 November (DOY 135–314) during 2016. The N_2O concentrations were analyzed using a gas chromatography (Agilent 7890A, Agilent Technologies, Santa Clara, CA) equipped with an electron capture detector.

The effluxes of N_2O within soil profiles were calculated based on Fick's law using the following equation (Marshall, 1959).

$$q = -D_p \frac{d_c}{d_z}$$

where q is the soil N_2O efflux ($\text{g m}^{-2} \text{s}^{-1}$); D_p is the soil gas diffusion coefficient in each soil depth ($\text{m}^{-3} \text{m}^{-1} \text{s}^{-1}$); d_c is the difference of N_2O concentration in the air between two soil depths, d_z is the distance between two soil depths (m). When d_c is the difference between the soil at 5 cm depth and air N_2O concentrations in the atmosphere, it is used to assume the N_2O emission rate of the soil surface. The efflux gradient between two soil depths was used to characterize N_2O production rates at different soil layers using the following equation (Yoh et al., 1997; Kusa et al., 2010; Nan et al., 2016).

$$P_i = q_i - q_{i+1}$$

where P_i and q_i are the N_2O production rate ($\text{g m}^{-2} \text{s}^{-1}$) and efflux ($\text{g m}^{-2} \text{s}^{-1}$) of each soil layer, respectively.

Soil gas diffusion coefficient D_p was estimated using the SWLR (structure-dependent water-induced linear reduction) model (Moldrup et al., 2013).

$$D_p = D_0 \varepsilon^{(1+C_m \Phi)} \left(\frac{\varepsilon}{\Phi} \right)$$

where D_0 is the gas diffusion coefficient ($\text{m}^{-2} \text{s}^{-1}$); ε is the soil air-filled porosity ($\text{m}^3 \text{m}^{-3}$); Φ is the soil porosity ($\text{m}^3 \text{m}^{-3}$); C_m is the media complexity factor in the SWLR model, and Moldrup et al. (2013) recommended a value of 2.1 for C_m in intact soils after comparing several prediction models.

$$\Phi = 1 - \frac{\rho_b}{\rho_s}$$

$$\varepsilon = \Phi - \theta$$

where ρ_s is the average particle soil density (2.65 g m^{-3}); and θ is the soil bulk density (g m^{-3}) and soil volumetric water content of each soil layer. The diffusion coefficient D_0 was calculated based on temperature and pressure using the following equation (Campbell, 1985):

$$D_0 = D_s \left(\frac{T + 273.15}{273.15} \right)^{1.75} \left(\frac{P_0}{P} \right)$$

where T and P are the temperature ($^{\circ}\text{C}$) and air pressure (Pa), respectively; D_s is $1.43 \times 10^{-5} \text{ m}^2 \text{ s}^{-1}$, that is the diffusion coefficient of N_2O in free air at the reference temperature (273.15 K) and reference air pressure P_0 (1 atm) (Pritchard and Currie, 1982). P values for each sampling day were derived from a weather station in the field.

The growing-season cumulative N₂O emissions ($\Sigma\text{N}_2\text{O}$, g N₂O-N ha⁻¹) were calculated by summing up the daily average emissions calculated from the concentration gradient method. Linear interpolation was used to estimate the missing values where a sampling was not conducted. Similar method was used to calculate the cumulative N₂O production ($\Sigma\text{N}_2\text{O}_p$, g N₂O-N ha⁻¹) from different soil layers.

2.4 Soil sampling and analysis

In each plot, three soil samples of 0–20 cm were collected on the day of gas collection and mixed as one soil sample to measure the concentrations of NH₄⁺-N and NO₃⁻-N over the experiment. Soil temperature and volumetric water content (VWC) were measured using a sensor and data were collected using a data logger. For details about the sensor and logger see Kuang et al. (2018). The installation positions of sensors were at 5, 15, 30, and 60 cm under the drip tape in the drip irrigation treatment and the corresponding location in the flood irrigation treatment. Soil WFPS was calculated as follows:

$$\text{WFPS} = \frac{\text{VWC}}{1 - (\text{B}_D/\text{P}_D)} \times 100$$

Where, in each layer, B_D is bulk density (Mg m⁻³) and P_D is particle density (assumed 2.65 Mg m⁻³).

2.5 Statistical analysis

A two-way analysis of variance was used to test the main and interactive effects of fertilizer treatment and irrigation method on $\Sigma\text{N}_2\text{O}$ and $\Sigma\text{N}_2\text{O}_p$ (PROC MIXED). The N resource and irrigation were considered as fixed factors, while plot replicates were considered as random factors. Means of treatments were compared using the least significant differences when the main or interactive effects were significant. The relationship between N₂O concentration and temperature, WFPS, air content at 5, 15, 30, and 60 cm depths, as well as NO₃⁻-N and NH₄⁺-N of 0–20 cm top soil were examined by regression analysis. Similar regression analysis was used to examine the relationship of N₂O flux rates between the concentration-gradient method (GM) and the closed-chamber method (CM). The surface N₂O flux rates based on the closed-chamber method were previously reported by Kuang et al. (2018) and used in this study for comparison between the two methods. The normality and homogeneity of variance were checked before analysis. Differences were considered as significant at $p < 0.05$. All analysis were performed using the Statistical Analysis Software package (SAS Institute, 2011).

3 Results

3.1 Environmental and soil conditions

Irrespective of irrigation type, soil temperature at 5 cm soil depth followed a similar trend as for air temperature, gradual increased from April to July and then decreased (Figure 1). The annual total precipitation was 50 mm in 2016. In comparison, the

total water addition was 593 mm and 982 mm for drip and flood irrigation plots, respectively, which accounted for 92%–95% of the total water inputs. Soil WFPS in both drip and flood irrigation soils showed large fluctuations in response to irrigation and rainfall events, which ranged from 7.4%–43.1% and 6.4%–46.7% for drip and flood irrigation, respectively. For drip irrigation plots, soil WFPS increased with irrigation events at 5, 15, 30, and 60 cm depths, but the peaks at 5 and 15 cm depths tended to be larger than those at 30 and 60 cm. For flood irrigation plots, soil WFPS at 5, 15, 30, and 60 cm layers increased with irrigation events, with similar peaks in different depths, but the rate of water decline in deeper soils was generally slower than that in shallow soils.

3.2 Soil N₂O concentrations

Soil N₂O concentrations at different depths showed similar temporal patterns over the experimental period with two clear peaks occurred on Day 235 and 265 (Figure 2). Soil N₂O concentrations in Control, Urea, U + M, and Manure treatments under drip irrigation ranged from 290–480, 283–504, 298–479, and 298–505 nL L⁻¹, respectively. In contrast, soil N₂O concentration in Control, Urea, U + M, and Manure under flood irrigation ranged from 260–473, 293–532, 286–477, and 304–487 nL L⁻¹, respectively. The average soil N₂O concentration in the drip-irrigated plots slightly decreased with depth, being 359, 346, 340, and 333 nL L⁻¹ at 5, 15, 30, and 60 cm, respectively.

3.3 N₂O efflux rate

The soil N₂O efflux rate at each depth varied with fertilization and irrigation treatments. The peak N₂O efflux rate was higher under flood irrigation (42.91 μg N m⁻² h⁻¹) than under drip irrigation (20.63 μg N m⁻² h⁻¹) treatment, and all fertilization treatments had higher N₂O efflux peaks than the control (drip: 18.73 μg N m⁻² h⁻¹, flood: 36.13 μg N m⁻² h⁻¹) under both drip and flood irrigation conditions. Across all treatments, the 0–5 cm soil depth had the highest N₂O efflux rate, ranging from -4.60–42.91 μg N m⁻² h⁻¹ (Figure 3).

3.4 N₂O production rate

The N₂O production rates of different soil depths were calculated from the N₂O efflux rates of two adjacent layers. Overall, the N₂O production rate in the profile decreased with increasing soil depth irrespective of fertilizer and irrigation treatments, with the 0–5 cm layer having the highest N₂O production rate (Figure 4). The N₂O production rates in the 0–5 cm soil layer were 3.3, 4.2, 6.5, and 8.0 μg N m⁻² h⁻¹ for Control, Urea, U + M, and Manure treatments under drip irrigation, respectively.

Cumulative N₂O production was higher in the 0–5 cm soil layer than other depths under all treatments, and average cumulative N₂O production at 5, 15, 30, and 60 cm layers were 248, 27, -10, and -3 g N ha⁻¹, respectively (Table 2). Calculation of the contribution of cumulative N₂O production in each soil depth to

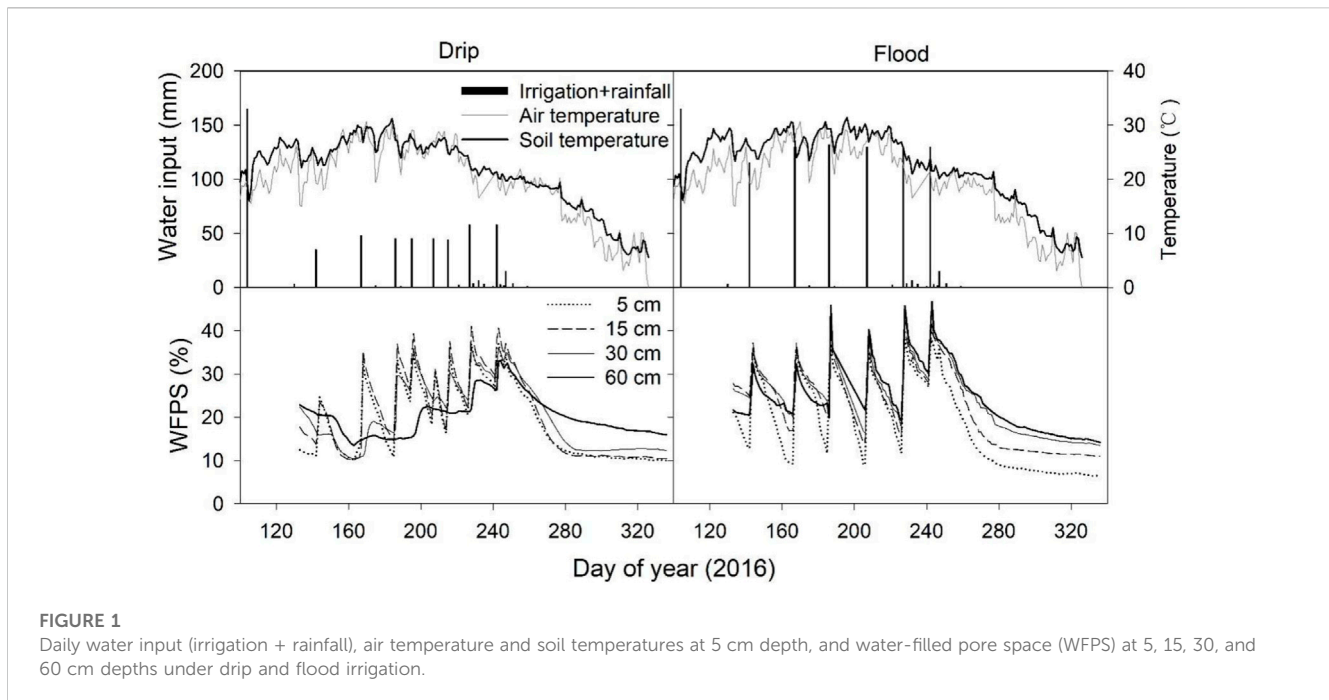


FIGURE 1

Daily water input (irrigation + rainfall), air temperature and soil temperatures at 5 cm depth, and water-filled pore space (WFPS) at 5, 15, 30, and 60 cm depths under drip and flood irrigation.

surface N_2O emissions based on the concentration-gradient method showed that the 0–5 cm and 5–15 cm depths contributed to all surface N_2O emissions, with the 0–5 cm soil depth contributing 87%–100% of the surface emissions. The contribution of cumulative N_2O production from the 0–5 cm soil layer to surface N_2O emissions was significantly affected ($p < 0.05$) by irrigation treatment, being 99% and 87% for drip and flood irrigation conditions, respectively.

3.5 Comparison of surface N_2O emission and flux between GM and CM

Using the data of surface N_2O flux measured by the closed-chamber method reported by Kuang et al. (2018), we compared the surface N_2O emission and flux between CM and GM (Table 3). Results showed a non-significant ($p = 0.128$) relationship of N_2O flux between CM and GM. Fertilizer treatments significantly affected cumulative surface N_2O emissions based on these two methods ($p < 0.05$), with a numerical trend of Control < Urea < U + M < Manure (Table 3). There was no significant difference in the effect of the irrigation method on cumulative N_2O emissions based on the concentration-gradient method, but the closed-chamber method showed cumulative N_2O emissions significantly higher under drip irrigation than flood irrigation ($p < 0.05$).

3.6 Relationship between soil profile N_2O concentration and environmental factors

Pearson correlation analysis was used to investigate the relationships of soil profile N_2O concentration with soil temperature, WFPS, soil pore air content and inorganic N content of 0–20 cm depth. Results showed a significantly

($p < 0.05$) negative relationship between soil profile N_2O concentration and soil temperature (Table 4). A significantly ($p < 0.05$) negative relationship was also shown between soil N_2O concentration at 15, 30, and 60 cm soil depth and NO_3^- -N content at 0–20 cm soil depth, however, N_2O concentration at 5 cm depth was significantly and positively correlated with NH_4^+ -N content at 0–20 cm depth ($p < 0.05$). In addition, there was a significantly ($p < 0.05$) positive correlation between N_2O concentrations in different soil layers.

4 Discussion

4.1 Low N_2O emissions from cotton fields in the arid region are due to the generally low N_2O production and the transmission of N_2O from the near-surface soil to the subsoil

In this study, soil N_2O concentrations were generally low in all soil layers under both drip and flood irrigation conditions, with average N_2O concentrations ranging from 333–359 nL L^{-1} in the profile, much lower than values reported in previous studies for other terrestrial ecosystems (Wang et al., 2013; Nan et al., 2016; Zhou et al., 2016; Wang et al., 2018; Yao et al., 2018). During the observation period, the cumulative N_2O production from different soil layers ranged from –25–306 g N ha^{-1} , which were also lower than previous reported results by Yao et al. (2018). These results indicate low N_2O concentrations and production in sandy soils, which were likely associated with the low soil moisture and C content (Kuang et al., 2018; Yin et al., 2019).

Soil surface N_2O emissions are a net gas exchange between soil and atmosphere and N_2O concentrations at different depths reflect the combined effects of N_2O processes in the soil. Several previous studies have found that N_2O concentrations increased with

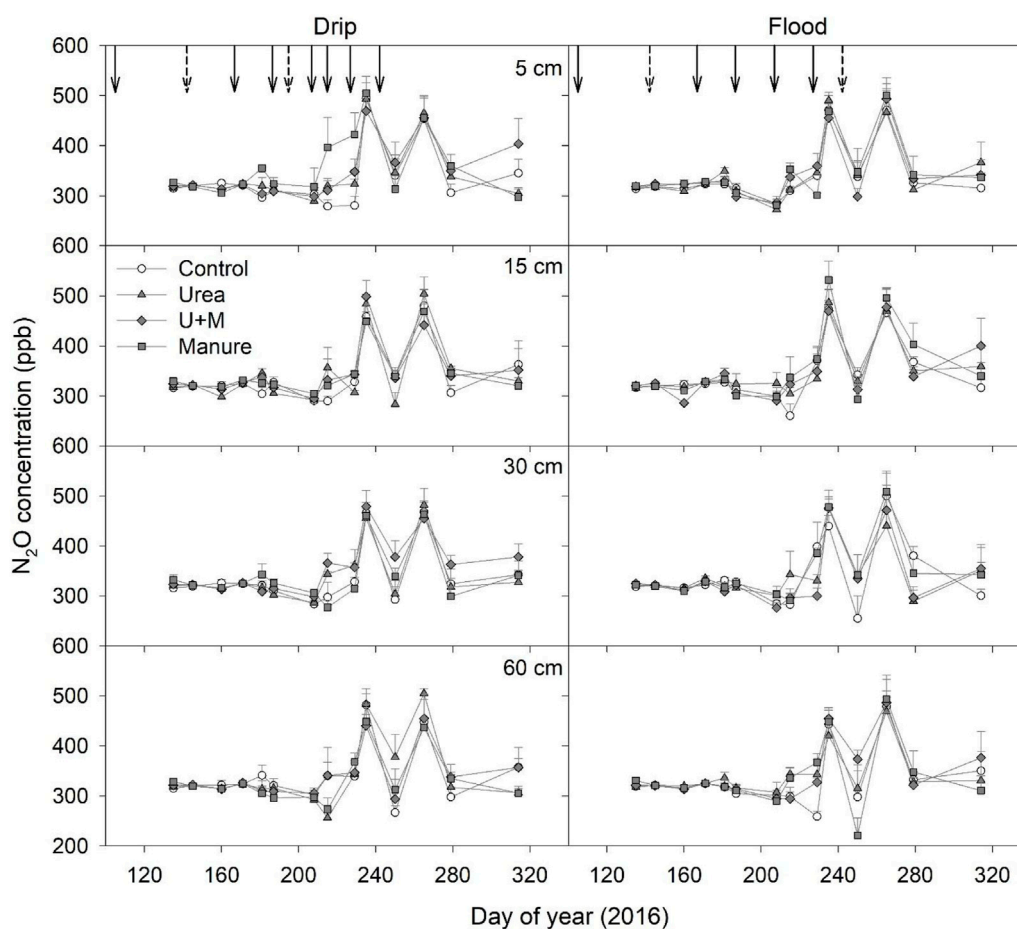


FIGURE 2

N_2O concentration at each depth as affected by fertilizer treatments under drip and flood irrigation. Dashed arrows indicate dates of irrigation only, and solid arrow indicates date of irrigation and urea application. Means \pm 1 standard error ($n = 4$) are presented.

increasing soil depth, with high N_2O concentrations in the subsoil but a low contribution to surface N_2O emissions (Wang et al., 2013; Wang et al., 2018; Yao et al., 2018), mainly because N_2O produced in the subsoil may undergo complete denitrification to N_2 during upward transport (Gao et al., 2014; Kuang et al., 2019). In contrast, the N_2O concentrations through the soil profiles in this study were all low, and the overall distribution pattern was uniform or higher in the surface soil layer than in the bottom layer. These results implies that the deep-depth soil may be a sink for these processes in cotton fields in the arid zone, and this inference can be validated by the fact that the cumulative N_2O production in the 30 and 60 cm layers was mostly negative (Table 1). Furthermore, some previous studies have also found higher N_2O concentrations in the near-surface soil than in the subsoil with a high soil moisture (Sotomayor and Rice, 1999). The low soil N_2O concentrations also indicate that sandy soils were conducive to nitrification process under the aerobic condition, and denitrification process under the anaerobic conditions. These biological processes need involvements of soil microbe and organic C, which are generally low in sandy soils as in the current study. Low water holding capacity and high hydraulic conductivity in sandy soils could have decreased WFPS to a level less conducive to denitrification. In addition, the high

permeability of sandy soil could facilitate the vertical transfer of N_2O between depths, and thus a less differentiated concentration gradient.

4.2 Topsoil is the main source of N_2O emissions from this cotton field in the arid region

The topsoil (0–15 cm) contributed to all the surface N_2O emissions in the present study, which is consistent to previous studies (Nan et al., 2016; Yao et al., 2018; Li et al., 2021). There might be two reasons for the high N_2O production rate of topsoil. The first reason is that the surface soil contains a high concentration of organic carbon and nitrogen due to the suitable temperature and humidity, which stimulate litter and root decomposition as well as the mineralization of organic fertilizer and urea. Second, the higher temperature and humidity in the topsoil promote the occurrence of microbial processes that produce N_2O (Kuang et al., 2019). According to Li et al. (2021), surface N_2O emissions were mostly associated with the topsoil of 0–15 cm under a drip irrigation cotton field. Nan et al. (2016) also reported that surface N_2O emissions

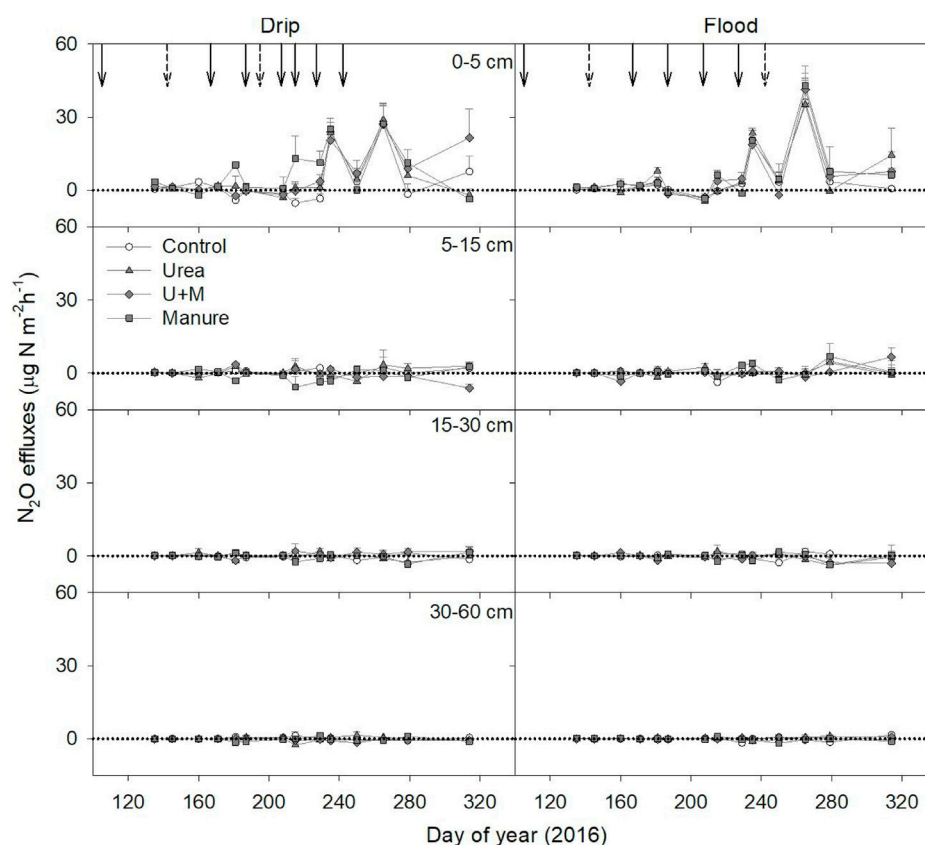


FIGURE 3

The N_2O effluxes at each soil layer as affected by fertilizer treatments under drip and flood irrigation. Dashed arrows indicate dates of irrigation only, and solid arrow indicates date of irrigation and fertigation of urea applications. Means ± 1 standard error ($n = 4$) are presented.

originated entirely from 0–30 cm layer. These may be due to the high microbial activity of soils in the 0–30 cm layer, which was dominated by N fertilizer-induced N_2O production (Wang et al., 2013). In contrast, the study's findings that subsoil may contribute negatively to surface N_2O emissions are primarily attributable to the higher gas diffusion coefficient caused by the lower water retention capacity of the sandy soil in the region. This assumption is supported by the fact that N_2O concentrations in different soil layers correlated positively with one another (Table 4). These results indicated that topsoil was the main source of surface N_2O emissions, and the subsoil played a completely different role in N_2O production and consumption in different studies. Thus, the role of the subsoil should be considered when studying the processes of N_2O emissions from the more permeable sandy soils.

Over the study period, the cumulative N_2O emissions in the topsoil of 0–5 cm contributed 87%–100% of the surface N_2O emissions. It showed a significant positive correlation ($p < 0.05$) with soil NH_4^+ -N content but a non-significant negative correlation ($p > 0.05$) with soil NO_3^- -N content at 0–20 cm depth (Table 4). These results imply that the nitrification process may be the main source of N_2O production in 0–5 cm soils, due to the low WFPS (10%–40%) of 0–5 cm soil layer and its decreased sharply after irrigation or rainfall events (Figure 1), which create a suitable condition for nitrification. Similarly, Bateman and Baggs (2005) also found that nitrification was the main pathway of N_2O

production in sandy loam soils with 35%–60% WFPS using the ^{15}N labeling method.

4.3 The key soil layers effected by nitrogen management and irrigation which on N_2O production are different

Both GM and CM results showed that the addition of manure significantly increased the N_2O flux, which was consistent with the result reported by Zhou et al. (2017). Manure addition can increase soil C availability and also the activities of associated N-cycling microbes, and consequently stimulate the nitrification and denitrification processes (Lessard et al., 1996). In addition, some studies have shown that simultaneous inputs of C and NH_4^+ into the soil, can promote the emission of N_2O more than the addition of NH_4^+ alone (Bergstrom et al., 1994). In the current study, through the N_2O production and consumption in the profile, it can be found that the addition of manure significantly increased N_2O production in the 0–5 cm layer. That is, the influence of the N source on the N_2O production of the profile is mainly manifested in the surface layer (0–5 cm). This may be related to the mixing of manure with surface soil (0–20 cm) in this study. The topsoil has higher microbial activity than the subsoil (Uchida et al., 2011). Abundant organic C compounds and available nitrogen in manure provide sufficient

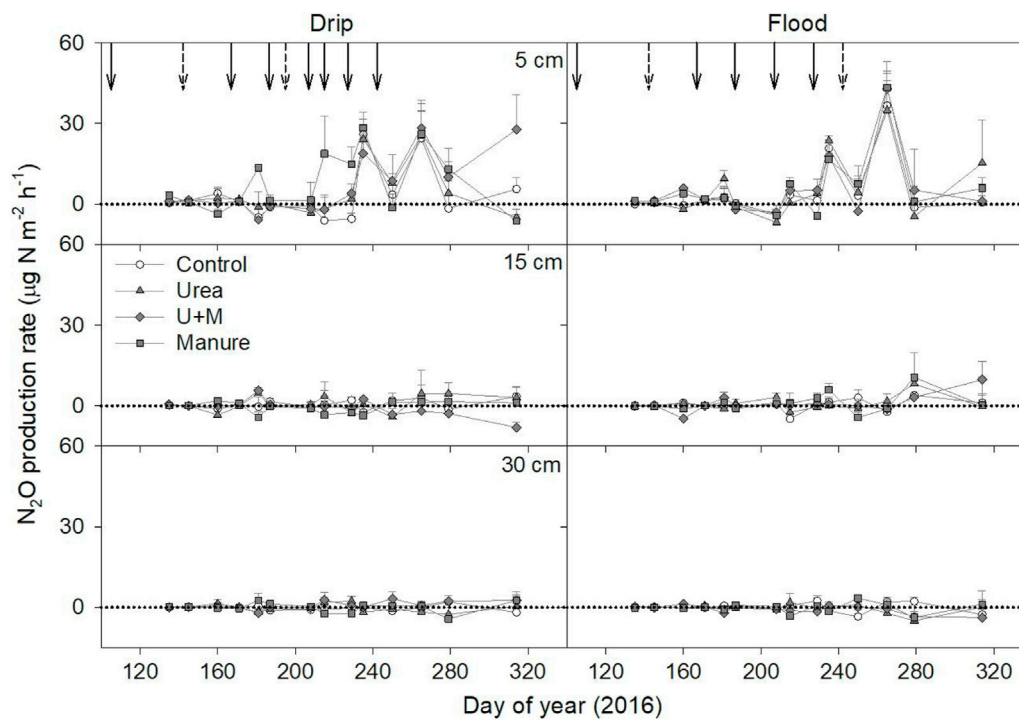


FIGURE 4
 N₂O production rate at each depth as affected by fertilizer treatments under drip and flood irrigation. Dashed arrows indicate dates of irrigation only, and solid arrow indicates date of irrigation and fertigation of urea applications. Means +1 standard error (n = 4) are presented.

TABLE 2 Cumulative N₂O production (ΣN₂O) and contribution rate of each soil layer over growing season, as affected by N addition and irrigation treatments. Values are means ± 1 standard error, n = 4.

	ΣN ₂ O (g N ha ⁻¹)				Contribution rate (%)			
	5 cm	15 cm	30 cm	60 cm	5 cm	15 cm	30 cm	60 cm
Addition								
Control	174 ± 22 c	27 ± 15 a	-4 ± 16 a	-1 ± 7 a	89 ± 6 a	11 ± 10 a	0 ± 9 a	0 ± 4 a
Urea	211 ± 34 bc	50 ± 22 a	-25 ± 21 a	2 ± 8 a	87 ± 5 a	23 ± 10 a	-10 ± 10 a	0 ± 4 a
U + M	306 ± 35 a	-1 ± 23 a	0 ± 20 a	-3 ± 6 a	100 ± 5 a	1 ± 9 a	0 ± 7 a	0 ± 3 a
Manure	297 ± 23 ab	31 ± 21 a	-11 ± 11 a	-10 ± 6 a	96 ± 5 a	10 ± 6 a	-3 ± 4 a	-3 ± 2 a
Irrigation								
Drip	254 ± 27 a	3 ± 14 b	1 ± 12 a	-6 ± 4 a	99 ± 4 a	3 ± 7 a	0 ± 5 a	-2 ± 2 a
Flood	241 ± 22 a	51 ± 13 a	-21 ± 12 a	0 ± 5 a	87 ± 3 b	19 ± 4 a	-7 ± 5 a	0 ± 2 a
ANOVA P values								
Irrigation (I)	0.619	0.014	0.199	0.451	0.028	0.066	0.403	0.678
Fertilizer (F)	0.003	0.291	0.748	0.688	0.192	0.337	0.823	0.904
I×F	0.174	0.176	0.212	0.309	0.290	0.378	0.392	0.487

For each treatment factor, means within a column followed by the same letter are not significantly different at p < 0.05.

energy and substrates for the nitrifying and denitrifying reactions of surface soil microbial biomass which was promoted N₂O emission (Lentz et al., 2014). Furthermore, the correlation between N₂O (0–5 cm) production and NH₄⁺-N, shows that the increase in N₂O flux caused by manure addition is the result of nitrification.

Using both individual experiment and meta-analysis, previously reported that flood and furrow irrigation resulted in significantly

higher N₂O emission than drip irrigation (Sánchez-Martín et al., 2008; Kuang et al., 2021). In this study, GM was used to find that flood irrigation significantly increased the cumulative production of N₂O in the 5–15 cm compared with drip irrigation, further confirming that the effect in the irrigation method on N₂O production was mainly expressed in the sub-surface layer. This may be associated with the disparities of soil profile water status

TABLE 3 Cumulative N₂O emissions (Σ N₂O) as affected by N addition and irrigation treatments based on concentration gradient and closed-chamber methods. Values are means \pm 1 standard error, n = 4.

	Σ N ₂ O (g N ha ⁻¹)	
	Concentration gradient method (GM)	Closed-chamber method (CM)
Addition		
Control	197 \pm 21 c	132 \pm 12 c
Urea	238 \pm 29 bc	221 \pm 21 b
U + M	303 \pm 28 ab	261 \pm 27 b
Manure	307 \pm 15 a	404 \pm 45 a
Irrigation		
Drip	252 \pm 21 a	293 \pm 35 a
Flood	270 \pm 18 a	216 \pm 25 b
ANOVA P values		
Irrigation (I)	0.371	0.006
Fertilizer (F)	0.002	<0.0001
I×F	0.328	0.165

For each treatment factor, means within a column followed by the same letter are not significantly different at $p < 0.05$.

TABLE 4 Pearson correlation coefficients ($n = 448$) between N₂O concentration and temperature, WFPS, air content at 5, 15, 30, and 60 cm depths, as well as NO₃⁻-N and NH₄⁺-N of 0–20 cm top soil.

N ₂ O concentration (cm)	Temperature	WFPS	Soil air content	NO ₃ ⁻ -N	NH ₄ ⁺ -N	N ₂ O concentration		
						5 cm	15 cm	30 cm
5	-0.197**	-0.039	0.038	-0.048	0.137**			
15	-0.238**	-0.072	0.071	-0.119*	0.024	0.758**		
30	-0.214**	0.070	-0.070	-0.102*	0.046	0.725**	0.743**	
60	-0.189**	0.040	-0.039	-0.112*	0.052	0.677**	0.687**	0.691**

*, ** indicate significance at $p < 0.05$ and $p < 0.01$, respectively.

caused by different irrigation methods. Compared with drip irrigation, the water content in the whole profile under flood irrigation was higher and decreased more slowly, which provided a optimal moisture condition for the production of N₂O in the profile (Kuang et al., 2019). In addition, the higher risk of N leaching under flood irrigation may have increased the nitrogen content at 5–15 cm depth, enhancing nitrification/denitrification processes and thus increasing N₂O production. In this study, no significant interaction between irrigation and N source treatments on N₂O production in each soil layer was found. Although a discrepancy exists in the N₂O emission estimated by the CM and GM, the trend of cumulative N₂O emissions under different treatments is consistent. Therefore, GM's ability to provide abundant information on profile N₂O production, reduction, and diffusion is commendable.

5 Conclusion

In the current study, soil N₂O concentrations were generally low in all soil layers under both drip and flood irrigation conditions, with average N₂O concentrations ranging from 333–359 nL L⁻¹ through the

soil profile. Such low N₂O concentration and efflux in sandy soils were mainly attributed to the generally low N₂O production due to low soil moisture and C availability. Soil N₂O concentrations decreased with increasing soil depth, and had similar seasonal patterns in different soil layers, indicating that the subsoil was a N₂O sink, which also explained the low N₂O emissions in the current study. Additionally, the topsoil (0–15 cm) contributed all the surface N₂O emissions, with a contribution of 87%–100% at a topsoil of 0–5 cm. The increase of soil moisture and C, N availability under N source and irrigation treatments were the main factors influencing the N₂O production in the 5 and 15 cm soil layers. N inputs through synthetic fertilizer or manure significantly increased N₂O concentration and production at 0–15 cm. Flood irrigation resulted in higher N₂O concentration and production compared to drip irrigation. The results confirm that soil water and nitrogen management are important drivers of N₂O production and diffusion in soil profiles of croplands in arid region. Use of nitrification inhibitor to slow N transformation process in soils can be an effective strategy to reduce N₂O production and emissions in arid regions. Future research is also needed to incorporate fertilizer into drip irrigation strategy, i.e., fertigation, to develop effective strategies for enhancing crop productivity while mitigating GHG emissions.

Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

Author contributions

XG: Conceptualization. WK and YW: Data curation. WK and XG: Funding acquisition. WK, MY, and YW: Investigation. XG: Methodology. FZ: Software, Supervision. DG: Visualization. WK and YW: Writing—original draft. WK, XG, and YW: Writing—review and editing.

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