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\*CORRESPONDENCE Yaxin Wang, ⊠ yxwang@shou.edu.cn

SPECIALTY SECTION This article was submitted to Toxicology, Pollution and the Environment, a section of the journal Frontiers in Environmental Science

RECEIVED 13 October 2022 ACCEPTED 16 January 2023 PUBLISHED 03 February 2023

#### CITATION

Wang Y and Chen X (2023), Microplastics in a pelagic squid (*Dosidicus gigas*) from the Eastern tropical Pacific Ocean: Characteristics, spatial variation, and preliminary risk assessment. *Front. Environ. Sci.* 11:1069124. doi: 10.3389/fenvs.2023.1069124

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# Microplastics in a pelagic squid (*Dosidicus gigas*) from the Eastern tropical Pacific Ocean: Characteristics, spatial variation, and preliminary risk assessment

### Yaxin Wang<sup>1</sup>\* and Xinjun Chen<sup>1,2,3,4</sup>

<sup>1</sup>College of Marine Sciences, Shanghai Ocean University, Shanghai, China, <sup>2</sup>Key Laboratory of Sustainable Exploitation of Oceanic Fisheries Resources, Ministry of Education, Shanghai, China, <sup>3</sup>National Engineering Research Centre for Oceanic Fisheries, Shanghai Ocean University, Shanghai, China, <sup>4</sup>Key Laboratory of Oceanic Fisheries Exploration, Ministry of Agriculture and Rural Affairs, Shanghai, China

Despite concerns about the environmental impact of microplastics (MPs), knowledge of MP contamination levels in pelagic squid is lacking. This study investigated the stomach of an ecologically and commercially important but poorly known squid species, the jumbo squid Dosidicus gigas, found in the tropical and Southern temperate Pacific. We examined the characteristics and spatial distribution of MPs in fifty D. gigas (16.0-37.8 cm mantle length) and assessed potential MP pollution risks. MPs were detected in 50.0% of specimens and had an average abundance of 0.88  $\pm$  1.12 items/individual and 0.24  $\pm$  0.36 items/ g stomach weight, and were sized 58.42-2,944.85 µm. The MPs were dominated by polyethylene terephthalate (31.82%) in polymer type, fragments (54.55%) and fibers (43.18%) in shape, and blue (59.59%) in color. All specimens could be evaluated as low risk. No statistically significant correlations were found between the MP abundances and D. gigas stomach weight, stomach fullness index, distance from the coast, and mantle length. While regression analyses showed that fuller stomachs contained larger MPs. Spatial variations in shape and polymer composition suggest that lowdensity fibrous MPs could be easily transported offshore by upwelling and/or ocean currents. These results indicate intraspecific variation in D. gigas MP contaminants and underscore the need to assess the potential site-specific ecotoxicological consequences of increased plastic pollution to pelagic squid.

### KEYWORDS

dosidicus gigas, pelagic squid, spatial variation, stomach, risk assessment, eastern tropical pacific ocean

### **1** Introduction

Since its emergence in the early 20th century, plastic products have become a widely used material based on their characteristics of durability, cheapness, and great convenience (Dehaut et al., 2019). However, the difficulties in degrading plastic have led to plastic waste causing serious environmental problems (Barnes et al., 2009). Recently, there has been growing concern about microplastics (MPs), defined as plastic particles smaller than 5 mm (Arthur et al., 2009). MPs can be divided into primary and secondary MPs. Primary MPs are formed on the microscale, such as microbeads in personal hygiene products, while secondary MPs result from the degradation of large-sized plastics through physical, chemical, biological and UV radiation (Gewert et al., 2015). Compared to large plastics, MPs are smaller, making them easier to

disperse in different environments (Zhang T et al., 2021). As a result, MPs are ubiquitous in the marine environment and are readily ingested by marine organisms (Lusher et al., 2013), either through direct capture or by feeding on contaminated prey (Nelms et al., 2018). Previous studies showed that exposure of MPs in aquatic species around the world, such as zooplankton (Collignon et al., 2014; Botterell et al., 2019), crustaceans (Cau et al., 2019; Hara et al., 2020), cephalopods (Oliveira et al., 2020; Gong et al., 2021), and fishes (Lusher et al., 2013; Zhang et al., 2019; Huang et al., 2022). Consumption of MPs can cause not only physical harm (Cole et al., 2015) but also physiological harm (Lusher et al., 2013). In addition, the high hydrophobicity of MPs can result in toxic chemicals, such as persistent organic pollutants (POPs), and trace metals, and serve as carriers for pollutant diffusion (Brennecke et al., 2016).

Because most studies have focused on crustaceans and fish, the potential presence of MPs in cephalopods (particularly in pelagic squid) has not been extensively explored (Gong et al., 2021). In the present study, we focused on a commercially important species of pelagic squid, jumbo squid Dosidicus gigas. The commercial annual catches of this species varying between 0.8 and 1.2 million tonnes in 2014-2018 (FAO yearbook. Fishery and aquaculture statistics). D. gigas is considered a voracious and adaptable pelagic predator of a wide range of prey including crustaceans, fish, and cephalopods (including cannibalism) (Nigmatullin et al., 2001; Alegre et al., 2014). Given the high detection rate of MP ingestion reported in crustaceans (Watts et al., 2014; Zhang Z et al., 2021) and fish (Boerger et al., 2010; Gassel and Rochman, 2019), there is no doubt that the digestive system of D. gigas contain MPs. In addition, differences in dietary sources could result in spatial variability in the abundance and characteristics of MPs in D. gigas. This species is also the main prey items of marine top predators, such as tunas, sharks, and whales (Ruiz-Cooley et al., 2004). They can thus act as vectors for the transfer of MPs to high-trophic-level marine predators.

The aim of the present study was to determine the characteristics and spatial distribution of MPs in the *D. gigas*, and to conduct a risk assessment for the *D. gigas* in the tropical and southern temperate Pacific. Identifying the intraspecific variation of MP pollution is essential for establishing methods to assess the potential ecological impacts of MPs in highly migratory pelagic predators. The information obtained in this study will hopefully improve the understanding of MP accumulates in pelagic squid and its availability for incorporation into the food chain is fundamental for understanding threats to high-trophic-level marine predators around the world.

### 2 Materials and methods

### 2.1 Sample collection and preparation

Dosidicus gigas specimens were collected by scientific observers during nocturnal jigging operations between February and May 2019 (Figure 1). A total of 50 individuals were collected from the main fishing grounds in the tropical and Southern temperate Pacific (see Supplementary Table S1 for detailed sampling information). Specimens were immediately frozen on board and kept stored at  $-20^{\circ}$ C until further analysis. Dorsal mantle length (ML) was recorded in the laboratory. The stomach was dissected and weighed for further analysis of MP. The stomach fullness index is assigned by visual observation (Breiby and Jobling, 1985): 0 (empty), 1 (few residues), 2 (half full), 3 (almost full), and 4 (full). The outer areas of the stomachs were rinsed with ultrapure water (Milli-Q water) to remove any adhered particles.

# 2.2 Extraction, observation, and identification of microplastics

Microplastic extraction was performed using the modified method of our previous study (Gong et al., 2021). The stomachs were transferred to Erlenmeyer flasks and treated with 10% KOH (20 mL per Gram of wet tissue), then covered with aluminum foil to avoid contamination. All Erlenmeyer flasks were placed in a constant temperature oscillator (70°C, 150 rpm) until the stomachs were completely digested. Thereafter, about 6 g NaCl per 20 mL mixture was added overnight for density separation. The suspension was vacuum filtered through a glass fiber filter (2.7  $\mu$ m pore size, 47 mm diameter, Whatman Inc.).

Each filter was placed in a clean Petri dish pending analysis. The stereomicroscope (SZX2-FOF) and a U-TV0.63XC digital camera (both from Olympus, Tokyo, Japan) were used to observe the filters. All suspect MPs (>50  $\mu$ m) were photographed and measured using ImageJ version 1.50 software. Shape and color were identified based on a visual assessment of the morphometric characteristics.

To confirm polymer composition, each suspected MP was scanned using a Micro-Fourier Transform Infrared Spectroscope (Micro-FTIR, Spotlight 400, PerkinElmer) in Attenuated Total Reflection mode. Spectra were collected over a broad spectral range ( $650-4,000 \text{ cm}^{-1}$ ) at a resolution of 4 cm<sup>-1</sup> from an average of 32 sample scans (D'Souza et al., 2020; Huang et al., 2022). The resulting spectra were compared to libraries of standard spectra. The polymer composition was only identified when the confidence level of at least a 70% match or reliable spectral match (after visual inspection) was accepted.

### 2.3 Quality control

Some measures to avoid potential contamination were implemented throughout: all the solvents (including Milli-Q water) used for sample processing and analysis were filtered over a glass fiber filter ( $2.7 \mu m$  pore size, 47 mm diameter, Whatman Inc.), all devices such as tweezers and Erlenmeyer flasks were thoroughly rinsed three times with filtered ultrapure water and dried; nitrile gloves and cotton laboratory clothing were always worn; experiments were conducted in a closed, ultra-clean laboratory to prevent potential airborne MPs. Clean Petri dishes were placed next to the working zone and checked after experiments. Reference materials such as vessel coatings and fishing gear were also collected. In the case of potential airborne contamination, the same characteristics of microparticles, according to size, shape, and color, were not included in this the results.

### 2.4 Risk assessment of microplastics

To the best of our knowledge, there is currently no standardized protocol for systematically evaluating the potential risks of MPs in pelagic squid. As a consequence, we applied MP abundance into consideration to assess the MP pollution risk in the *D. gigas*.



groups and indicated with squares, circles, and triangles for Group I ( $G_1$ ), Group II ( $G_2$ ), and Group III ( $G_3$ ), respectively.

Risk assessment was expressed as pollution load index (Tomlinson et al., 1980). The formulas employed for *PLI* were as follows:

$$CF_i = C_i / C_0$$
$$PLI_i = \sqrt{CF_i}$$

where  $PLI_i$  is the pollution load index (Tomlinson et al., 1980) for individual *i*,  $C_i$  is the MP abundance for individual *i* and  $C_0$  are defined as the baseline MP abundance, which is theoretically a reference value of the minimum average MP abundance in squid species. Given the absence of a reference value for our study area and species, we followed the approach adopted in previous studies (Kabir et al., 2021; Zhang et al., 2022) and assigned  $C_0$  to the lowest MPs abundance of contaminated *D. gigas* obtained in this study. Variation in the selected constant does not affect the relative relationships of *PLI* among individuals, only absolute values, and therefore does not influence the comparison results.

### 2.5 Statistic analysis

To assess the potential spatial variations in MP characteristics of *D. gigas*, the specimens were merged into three main groups according to the sampling sites, i.e., Group I (G<sub>1</sub>, 84°–92°W, N = 17), Group II (G<sub>2</sub>, 100°–110°W, N = 22), and Group III (G<sub>3</sub>, 114°–121°W, N = 11) (Figure 1). Comparisons among sampling groups were performed using the Kruskal-Wallis test since the assumptions of normality and homogeneity of the variances were not met. In addition, Spearman's rank correlation was used to analyze the relationships between ML, stomach weight and fullness index, and distance from coast with MP characteristics. Regarding the relationship between the stomach weight and the number of MPs per Gram of stomach weight, this correlation test was considered eliminated in the analysis. Statistically significant results were indicated with p < 0.05. All results are presented as mean ± standard deviation (SD).

### 3 Results and discussion

D gigas is widely distributed in the eastern Pacific and its population structure is complex. Several studies have reported spatial variations of foraging strategies among population units within of D. gigas (Gong et al., 2020; Gao et al., 2022). For example, fishes were the main prey items of D. gigas collected off the coast of Chile (Pardo-Gandarillas et al., 2014) while cephalopods (including cannibalism) and fishes were the main prey items of D. gigas from the Ecuadorian waters (Rosas- Luis and Chompoy-Salazar, 2016) and the waters off Peru (Alegre et al., 2014). Although the stomach content of G2 and G3 individuals were not reported, previous studies performed on trophic ecology of D. gigas using stable isotope and fatty acid analyses indicate they might have specific dietary sources (Gong et al., 2020). It is generally accepted that MPs could be ingested by pelagic predators (e.g., D. gigas), either by direct capture or by feeding contaminated prey (Gong et al., 2021; Huang et al., 2022). In this study, stomachs from 50 individuals (16.0–37.8 cm ML) were used to test the hypothesis that the D. gigas from different areas have spatial variability in MP contamination. No difference in ML was observed between D. gigas collected from three sampling groups (G1: 22.5 ± 5.3 cm, G<sub>2</sub>: 22.2 ± 4.4 cm, G<sub>3</sub>: 23.3 ± 2.5 cm, Kruskal-Wallis,  $\chi^2 = 1.45, p = 0.49$ ).

### 3.1 Microplastic abundance

The visual sorting initially identified a total of 197 suspected MPs. Of these, 44 MPs (22.34%) were confirmed to be plastic polymers by Micro-FTIR. Of all *D. gigas* analyzed, 25/50 (50.00%) specimens contained MPs and had an average abundance of 0.88 ± 1.12 items/ individual and 0.24 ± 0.36 items/g tissue wet weight. Three sampling groups had a similar detection rate of MPs, 47.06% (G<sub>1</sub>), 50.00% (G<sub>2</sub>), and 54.55% (G<sub>3</sub>), respectively. The MP abundances for sampling groups were, G<sub>1</sub>: 1.00 ± 1.32 items/individual (0.16 ± 0.23 items/g), G<sub>2</sub>: 0.68 ±



#### FIGURE 2

Violin plots of the abundance (A, B) and size (C) of microplastics in jumbo squid Dosidicus gigas collected from three areas (abbreviations as in Figure 1). The violin plot starts with a standard boxplot (with minimum, interquartile range, mean, and maximum values) and then adds rotating kernel density plots on each side of the boxplot



#### FIGURE 3

transparent (D)

0.84 items/individual (0.25  $\pm$  0.34 items/g), G<sub>3</sub>: 1.09  $\pm$  1.30 items/ individual ( $0.35 \pm 0.53$  items/g) (Figure 2). Although MP abundances varied inconsistently across the G1, G2 and G3, no difference in MP abundance was observed among sampling groups (Kruskal-Wallis, items/individual:  $\chi^2 = 0.55$ , p = 0.76; items/g:  $\chi^2 = 0.57$ , p = 0.75). For all individuals, no significant Spearman correlations (0.190.95) were found between the MP abundances and the stomach weight, stomach fullness index, distance from coast, and ML of D. gigas.

Overall, these findings suggest that all sampling groups were influenced by similar level of MPs pollution. This phenomenon possibly due to MPs could be rapidly egested by D. gigas, which has a fast digestion rate (Gao et al., 2022). Until now, no evidence of MP accumulation in large predator has been documented in other studies (Chagnon et al., 2018). Therefore, the similar MP abundance in each group maybe driven by continuous ingestion and egestion of MPs rather than prey items or distance from coast.

### 3.2 Morphology, color, and chemical composition of microplastics

The size range of MPs in all individuals was 58.42-2,944.85 µm (685.86  $\pm$  669.87  $\mu$ m, Figure 2), with the smallest MPs found in the G<sub>2</sub> and the largest found in the  $G_1$ . The average size of MPs in each group was 592.37  $\pm$  725.68  $\mu m,$  584.24  $\pm$  475.07  $\mu m,$  and 945.34  $\pm$  774.49  $\mu m,$ respectively. No difference in the size of MP was observed among sampling groups (Kruskal-Wallis,  $\chi^2 = 2.54$ , p = 0.28), possibly due to the significant positive Spearman correlations between MP size and





stomach fullness index ( $0.41 \le r_s \le 0.42$ , p < 0.04). In our study, three groups had similar body size as mentioned before and stomach fullness (Kruskal-Wallis,  $\chi^2 = 0.55$ , p = 0.76) which could result in the similar size of MP. In contrast, no significant Spearman correlations (0.21 ) were found between the MP size and the stomach weight and distance from coast of*D. gigas*.

Three shapes of MPs were recorded in all individuals, including fiber, fragment, and film (Figure 3). The most common MP type was fragment (54.55%), which differs from previous studies performed on *D.gigas* collected from the Southern waters of  $G_1$  (Gong et al., 2021). This inconsistency among studies may be linked to the D.gigas collected in this study had a relatively small body size. The fragments detected maybe came from the fragmentation of largesized plastics (Yu et al., 2022; Zhang et al., 2022). The percentage of fibers reached 43.18% in all samples and a single piece of film was found in a specimen from G<sub>2</sub>. These fibrous MPs maybe sourced from the degradation of fishing gears and ropes, which would be consistent with previous results reported for marine animals from fishing grounds (Zhang et al., 2019; Yu et al., 2022). Fibrous MPs have larger aspect ratios with the rough surface, are easier to embed in tissues, and persist longer, which can cause mechanical damage and more severe toxicological effects on organisms compared to other types of MPs (Rebelein et al., 2021; Zheng et al., 2022). However, the prevalence of fibers in the samples may often be overestimated when the collection and analysis of the samples are not conducted under strictly controlled clean air (Hermsen et al., 2017).

Different compositions of shapes were found among the sampling groups. MPs in  $G_1$  specimens had a higher proportion of fragments (76.47%), with the remaining fibers (23.53%). For  $G_2$  and  $G_3$  specimens, however, the MPs were dominated by the fibrous form (58.33% and 53.33%), with 41.67% and 40.00% MP fragments, respectively. This pattern might be due to fibrous MPs could easily be transported by ocean currents to offshore areas, which accounts for the higher detection

rate of fibers that we observed in  $G_2$  and  $G_3$ ; however, the exact mechanism of transfer of fibrous MPs in this case remains to be explored.

chloride, PDMS: polydimethylsiloxane, SBR: styrene butadiene rubber,

AC: Acrylates copolymer.

Although color distributions varied inconsistently in each group, blue was the most dominant color (59.59% of all MPs), scoring 64.71%, 46.67%, and 66.67% in  $G_1$ ,  $G_2$ , and  $G_3$ , respectively. These findings match the high detection frequency and high amount of this MP color in the seawater and other squids (Zhang et al., 2019; Daniel et al., 2021; Zhang et al., 2022). The other colors of identified MPs were black, red, yellow, and MP in transparent film form (Figure 4).

Up to eleven polymer types were determined (Supplementary Figure S1). In all samples, polyethylene terephthalate (PET) was identified most frequently at 31.82% and reached 35.29%, 40.00%, and 16.67% in G1, G2, and G<sub>3</sub>, respectively. PET is employed primarily as a textile, with applications in clothing, ropes, and fishing gears (Šaravanja et al., 2022). The other frequently polymers were cellophane (CP, 18.18%) and polystyrene (PS, 11.36%), and the remaining polymer accounted for less than 10.00% of the MPs detected (Figure 5). These polymers are ubiquitous in the marine environment (Alomar and Deudero, 2017; Li et al., 2019; Liu et al., 2020). PET (1.34 g/cm3), CP (1.42 g/cm3), and PS (1.05 g/cm<sup>3</sup>) have a higher density than seawater (1.025 g/cm<sup>3</sup>) and tend to sink into deeper water layers (Woodall et al., 2014), making themselves available to organisms living in these areas. D. gigas spends the vast majority of its time in the deep-sea hypoxic environment may be the cause for the high proportion of these high-density MPs in our study (Stewart et al., 2013). D. gigas is also considered highly migratory, undertake ontogenetic migration between the continental shelf and the open ocean (Gilly et al., 2006; Stewart et al., 2013). They can thus act as important significant bioindicators of MP contamination in a vast three-dimensional space. However, low-density polymers (e.g., polypropylene (PP), 0.90-0.92 g/cm3; polydimethylsiloxane (PDMS), 0.98 g/cm3; and styrene-butadiene rubber (SBR), 0.94 g/cm<sup>3</sup>), were also found in the D. gigas (Figure 5), possibly because of biofouling and subsequent vertical

transport (Kooi et al., 2017; Hipfner et al., 2018; Liu et al., 2020). Interestingly, the low-density polymers were only detected in *D. gigas* from  $G_2$  and  $G_3$ , which were far from the coast than  $G_1$  (Figure 1). Possibly, offshore Ekman transport off Peru could move low-density polymer MPs to relatively farther offshore, and *D. gigas* feeding in the nearshore waters (i.e.,  $G_1$ ) might be exposed to more high-density polymers than in offshore waters (i.e.,  $G_2$  and  $G_2$ ) (Thiel et al., 2007; Ory et al., 2018). It is noteworthy that the distribution of polymers in seawater of our study area has not yet been published, further efforts are needed to conclude the potential relationship between microplastic transport and Ekman dynamics.

### 3.3 Risk assessment of microplastics

Regarding wild marine organisms, there is currently no standardized method to monitor the risk of MPs, while MP abundance is a useful indicator to assess the potential risks of MPs (Xu et al., 2018; Kabir et al., 2021).

Based on the number of MPs per individual, the *PLI* values for sampling groups were 0.66  $\pm$  0.77, 0.57  $\pm$  0.61, and 0.75  $\pm$  0.76, respectively. For the number of MPs per Gram of stomach weight, the *PLI* values were 1.04  $\pm$  1.24, 1.31  $\pm$  1.49, and 1.57  $\pm$  1.77, respectively. No difference in *PLI* values was found among the sampling groups (Kruskal-Wallis, items/individual:  $\chi^2 = 0.54$ , p = 0.76; items/g:  $\chi^2 = 0.57$ , p = 0.75). These values indicated that all individuals had a very low risk in the study area (Xu et al., 2018).

Overall, this study is the first record of the intraspecific variation in MP contamination levels in pelagic squid from oceanic habitats and also provides an preliminary examination of the risk assessment of MPs. Spatial diversity of MP shape and polymer type was found. The pollution risk assessment showed that all *D. gigas* were at low risk of pollution. The site-specific MP contaminants may be used to facilitate priority pollution monitoring. The pollution risk assessment provided insights into pollution and a basis for future comprehensive environmental and human health risk assessments in the context of increased plastic pollution from pelagic squid.

### Data availability statement

The raw data supporting the conclusion of this article will be made available by the authors, without undue reservation.

### **Ethics statement**

The animal study was reviewed and approved by Institutional animal care and use Committee of Shanghai Ocean University.

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### Author contributions

YW: Conceptualization, methodology, formal analysis, writing—original draft. XC: Methodology, validation, formal analysis, resources, supervision, funding acquisition.

# Funding

This work was supported by the project on the survey and monitor-evaluation of global fishery resources sponsored by the Ministry of Agriculture and Rural Affairs.

# Acknowledgments

The authors thank all the staff at Shanghai Ocean University for their assistance with sample pretreatment. We also thank Yi Gong for assistance with data analysis and interpretation.

# **Conflict of interest**

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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# Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fenvs.2023.1069124/ full#supplementary-material

### SUPPLEMENTARY FIGURE S1

Photographs of the spectra of the FTIR analyses. A, polyvinyl chloride; B, polypropylene; C, polyacrylonitrile; D, acrylates copolymer; E, polydimethylsiloxane; F, epoxy resin.

### SUPPLEMENTARY TABLE S1

Summary information of jumbo squid Dosidicus gigas specimens sampled in the Eastern tropical Pacific Ocean.

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