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Seasonal variability of CO₂, CH₄, and N₂O content and fluxes in small agricultural reservoirs of the northern Great Plains

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Inland waters are important global sources, and occasional sinks, of CO₂, CH₄, and N₂O to the atmosphere, but relatively little is known about the contribution of GHGs of constructed waterbodies, particularly small sites in agricultural regions that receive large amounts of nutrients (carbon, nitrogen, phosphorus). Here, we quantify the magnitude and controls of diffusive CO₂, CH₄, and N₂O fluxes from 20 agricultural reservoirs on seasonal and diel timescales. All gases exhibited consistent seasonal trends, with CO₂ concentrations highest in spring and fall and lowest in mid-summer, CH₄ highest in mid-summer, and N₂O elevated in spring following ice-off. No discernible diel trends were observed for GHG content. Analyses of GHG covariance with potential regulatory factors were conducted using generalized additive models (GAMs) that revealed CO₂ concentrations were affected primarily by factors related to benthic respiration, including dissolved oxygen (DO), dissolved inorganic nitrogen (DIN), dissolved organic carbon (DOC), stratification strength, and water source (as δ¹⁸O_{water}). In contrast, variation in CH₄ content was correlated positively with factors that favoured methanogenesis, and so varied inversely with DO, soluble reactive phosphorus (SRP), and conductivity (a proxy for sulfate content), and positively with DIN, DOC, and temperature. Finally, N₂O concentrations were driven mainly by variation in reservoir mixing (as buoyancy frequency), and were correlated positively with DO, SRP, and DIN levels and negatively with pH and stratification strength. Estimates of mean CO₂-eq flux during the open-water period ranged from 5,520 mmol m⁻² year⁻¹ (using GAM-predictions) to 10,445 mmol m⁻² year⁻¹ (using interpolations of seasonal data) reflecting how extreme values were extrapolated, with true annual flux rates likely falling between these two estimates.

KEYWORDS

agricultural pond, seasonality, greenhouse gas fluxes, CO₂ equivalent (CO₂-eq), methane, nitrous oxide, carbon dioxide

1 Introduction

Inland waters are often sources of greenhouse gases (GHG) to the atmosphere as a result of net emissions of carbon dioxide (CO₂), methane (CH₄), and nitrous oxide (N₂O) (Cole et al., 2007; Tranvik et al., 2009; DelSontro et al., 2018). However, recent research, including that on hard-water prairie ecosystems, suggests that the magnitude, spatial extent, and timing of emissions varies greatly among locales (Finlay et al., 2010; Finlay et al., 2015; Webb et al., 2019a; Webb et al., 2019b; Finlay et al., 2019). Major drivers of GHGs have been shown to be linked to nutrient status (carbon, nitrogen, and phosphorus), primary productivity, surface area, temperature, and latitude (Downing 2010; Deemer et al., 2016; Holgerson and Raymond 2016; DelSontro et al., 2018). Complete accounting of the role of inland waters in the global carbon budget needs to better characterize such temporal variability in different regions and waterbody types, particularly with respect to the cumulative effects of main greenhouse gases.

Waterbodies in agricultural areas are of particular interest in greenhouse gas dynamics as they have elevated nutrient loading reflecting elevated fertilizer application and livestock densities to meet the growing demand of global human populations (Tilman 1999; Earles et al., 2012; Baumann et al., 2017). Small agricultural reservoirs (also referred to as dams, impoundments, or dugouts) appear to vary dramatically in terms of the magnitude and direction of GHG fluxes. For example, Australian farm reservoirs emit large amounts of CO₂ and CH₄ to the atmosphere, but only small quantities of N₂O (Ollivier et al., 2018; Ollivier et al., 2019). In contrast, agricultural reservoirs of the northern Great Plains of North America are often sinks for CO₂ (52% of sites) and N₂O (67%), but are consistent net sources of CH₄ (Webb et al., 2019a; Webb et al., 2019b). Unfortunately, most of these surveys are based on single samples in late-summer (August) samples (Maberly 1996; Dillon and Molot 1997; Baehr and DeGrandpre 2004; Ducharme-Riel et al., 2015; Denfeld et al., 2016; Finlay et al., 2019), raising questions of the role of seasonal variability in influencing net annual carbon sources.

Understanding the mechanistic controls of aquatic CO₂, CH₄, and N₂O concentrations is critical for upscaling efforts, predictions of future climate change effects, and management opportunities to use these systems as carbon offsets (Bastviken et al., 2004; DelSontro et al., 2016). Temporal dynamics of CO₂ are usually correlated with changes in either microbial metabolism or the supply of inorganic carbon to freshwaters (Chapin et al., 2006; Dubois et al., 2009; McDonald et al., 2013; Bogard and Giorgio 2016; Wilkinson et al., 2016). In boreal ecosystems, the main pathway of CO₂ production and consumption is through photosynthesis (*P*, consumption) and respiration (*R*, production), with CO₂ evasion occurring when *P* < *R* (Chapin et al., 2006; Bogard and Giorgio 2016). However, this metabolic control can be overridden in non-boreal systems due to substantial inputs of inorganic C (e.g., DIC in runoff or

groundwater) or changes in pH and carbonate chemistry (Dubois et al., 2009; Finlay et al., 2010; McDonald et al., 2013; Bogard and Giorgio 2016; Wilkinson et al., 2016). For example, in small prairie hardwater systems, CO₂ concentration are regulated by a complex interaction between primary production, DIC content, nutrient status, stratification intensity, groundwater fluxes, and regional edaphic conditions (Webb et al., 2019b). Together, these models suggest that seasonal variation in the interaction among control factors is likely to create significant temporal variation in net CO₂ fluxes.

Methane is produced by methanogenic bacteria through the anaerobic degradation of organic matter, most often in the anoxic or hypoxic sediments of waterbodies (Glass and Orphan 2012). This CH₄ can be released to the atmosphere through diffusion, ebullition, or plant-mediated fluxes. Rates of CH₄ production are highly dependent on temperature ($Q_{10} \sim 4$; range 1–35) (Yvon-Durocher et al., 2014; DelSontro et al., 2016), whereas CH₄ can be consumed by methane-oxidizing bacteria (“methanotrophy”) under oxic conditions (King 1992; Duc et al., 2010). Gas concentrations are also influenced by other factors including organic carbon availability and quality (Segers 1998; Bridgman et al., 2006), mineral nutrient availability, and productivity (Deemer et al., 2016; DelSontro et al., 2018; Ollivier et al., 2018; Beaulieu et al., 2019) and competitive inhibition by sulfur-reducing bacteria (Lovley and Klug 1983; Webb et al., 2019b). Landscape models of farm reservoirs suggest that lowest CH₄ concentrations occur with DO supersaturation, low sediment C:N ratios, reduced dissolved inorganic nitrogen (DIN) concentrations, intermediate water residence time, elevated groundwater inflow, and higher conductivity levels (Webb et al., 2019b). Taken together, these observations suggest that seasonal CH₄ production in agricultural ponds will reach a maximum in late summer, when temperature, productivity, thermal stratification and deepwater anoxia are greatest.

Production of N₂O arises as an intermediary product during nitrification (Quick et al., 2019), incomplete denitrification (Firestone and Davidson 1989; Glass and Orphan 2012), or dissimilatory nitrate reduction to NH₄⁺ (DNRA; Scott et al., 2008), whereas denitrification consumes N₂O (Quick et al., 2019). Previous research suggests that temporal controls of N₂O flux include DO saturation, organic carbon availability, nitrogen availability and speciation, pH, and temperature (Baulch et al., 2011; Gooding & Baulch 2017; Quick et al., 2019; Zhang et al., 2021). Spatial analysis of variation in N₂O emissions from small agricultural reservoirs reveals N₂O concentrations were lowest when thermal stratification was strong, phytoplankton biomass (as Chl-*a*) was elevated, and DO was supersaturated (Webb et al., 2019a). Although N₂O emissions from prairie wetlands are known to be high in spring (Pennock et al., 2010) when DOC and NO_x is abundant, there have been few systematic studies of the controls of seasonal variation in the regulation of N₂O fluxes.

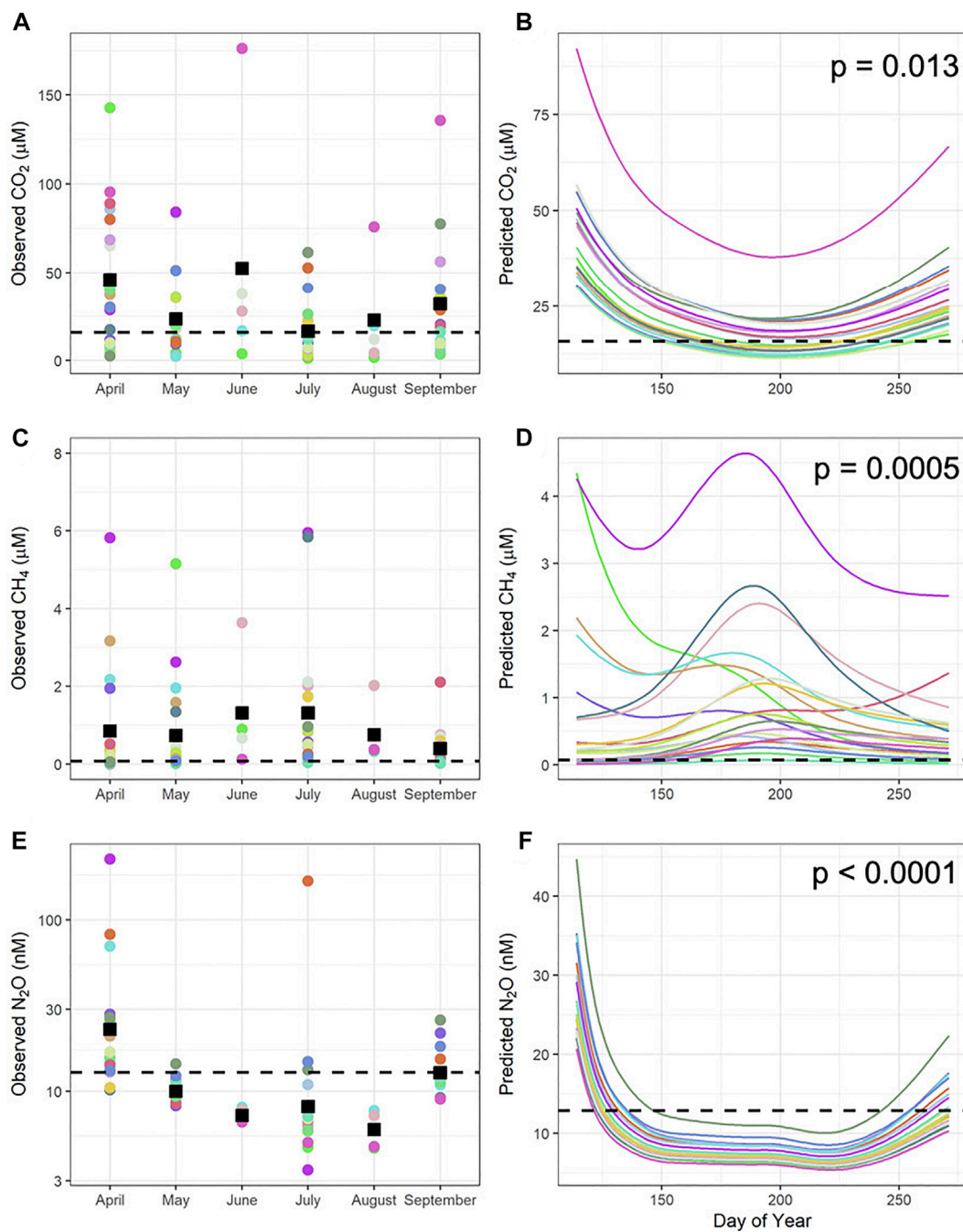


FIGURE 1

Seasonal carbon dioxide (CO_2 , (A,B)), methane (CH_4 , (C,D)), and nitrous oxide (N_2O , (E,F)) concentrations from 20 reservoirs in 2018. Left column: Raw observed GHG data for all three gases. Each coloured circle is one reservoir. The black squares represent the monthly mean. This data was used to inform the models. Right column: The modelled seasonal gas concentration from a generalized additive model (GAM) of all three gases. Each coloured line represents a single reservoir. p -values for the GAM are located at the top of each plot. Black dashed line in both panels represents the calculated (from ppm) atmospheric concentration of each gas. The colour in the left panel corresponds to the coordinating colour in the right panel. Deviance explained = 36.6% for CO_2 , 88.7% for CH_4 , and 90.2% for N_2O .

Thus, while previous research provides some mechanistic predictions for GHG fluxes from small waterbodies, it is difficult to anticipate how all three gases will sum to a CO_2 -equivalent

(CO_2 -eq) flux over the entire open water season. This study presents a seasonal and diel assessment of the magnitude of CO_2 , CH_4 , and N_2O concentration and diffusive flux, and the factors

TABLE 1 Physical and environmental parameters measured in the 20 reservoirs monthly in 2018. All parameters, excluding max depth, Secchi depth, and buoyancy frequency were measured using the YSI multi-parameter probe at 0.5 m below the water surface, excluding deep dissolved oxygen which was taken 0.5 m above the sediment. Presented as mean \pm standard deviation (SD). Superscript letters denote significant differences between months based on ANOVA and post-hoc Tukey test. DO = Dissolved Oxygen.

	Units	Month	Mean (SD)
Surface Temperature	°C	April ^a	7.98 (4.38)
		May ^b	15.6 (2.6)
		July ^c	22.7 (1.5)
		September ^a	6.99 (0.9)
Max Depth	Meters	April ^a	1.57 (1.3)
		May ^a	1.92 (0.8)
		July ^a	2.14 (0.9)
		September ^a	1.73 (0.7)
Secchi Depth	Meters	April ^a	0.481 (0.1)
		May ^a	0.591 (0.4)
		July ^a	0.824 (0.5)
		September ^a	0.556 (0.5)
Buoyancy Frequency	s ⁻²	April ^b	0.001 (0.002)
		May ^a	0.006 (0.005)
		July ^a	0.01 (0.008)
		September ^b	0 (0)
pH	Unitless	April ^a	8.29 (0.8)
		May ^{ab}	8.71 (0.5)
		July ^{ab}	8.94 (0.6)
		September ^b	8.62 (0.4)
Surface DO	%Saturation	April ^a	90.4 (30.1)
		May ^a	92.9 (20.7)
		July ^a	94.6 (44.3)
		September ^a	81.0 (22.1)
Deep DO	%Saturation	April ^a	67.2 (43.4)
		May ^{ab}	39.3 (38.6)
		July ^b	11.3 (19.0)
		September ^a	665.5 (29.1)
Conductivity	$\mu\text{S cm}^{-1}$	April ^a	737 (590.5)
		May ^{ab}	1,178 (891.2)
		July ^{ab}	1728 (1,533.4)
		September ^a	1824 (1721.9)

TABLE 2 Carbon and nutrient concentrations for each month in the 20 reservoirs in 2018. Presented as mean \pm standard deviation (SD). Superscript letters denote significant differences based on ANOVA and post-hoc Tukey test. DIC = dissolved inorganic carbon. DOC = dissolved organic carbon. TDP = total dissolved phosphorus. SRP = soluble reactive phosphorus. TDN = total dissolved nitrogen. NO_x = nitrate and nitrite concentration. NH₃ = ammonia concentration. Chl-*a* = chlorophyll *a* concentration. Dissolved inorganic nitrogen (DIN) is the sum of NO_x and NH₃.

	Units	Month	Mean (SD)
DIC	mg C L ⁻¹	April ^a	30.5 (11.8)
		May ^{ab}	40.5 (13.5)
		July ^b	47.2 (22.9)
		September ^b	56.0 (26.4)
DOC	mg C L ⁻¹	April ^a	20.1 (6.5)
		May ^{ab}	23.6 (8.0)
		July ^{ab}	26.3 (9.5)
		September ^b	31.3 (12.0)
TDP	$\mu\text{g P L}^{-1}$	April ^a	569 (816.1)
		May ^b	340 (468.3)
		July ^b	426 (860.4)
		September ^a	315 (500.0)
SRP	$\mu\text{g P L}^{-1}$	April ^a	381 (446.3)
		May ^b	293 (445.7)
		July ^b	343 (677.4)
		September ^a	265 (484.1)
TDN	$\mu\text{g N L}^{-1}$	April ^a	3,421 (3,218.0)
		May ^a	1913 (706.6)
		July ^a	2,694 (1,899.1)
		September ^a	3,197 (3,197.4)
NO _x	$\mu\text{g N L}^{-1}$	April ^a	644 (1,054.1)
		May ^b	22.7 (27.2)
		July ^{ab}	277 (675.3)
		September ^b	37.3 (49.7)
NH ₃	$\mu\text{g N L}^{-1}$	April ^a	496 (924.9)
		May ^a	118 (126.4)
		July ^a	126 (143.2)
		September ^a	544 (874.1)
Chl- <i>a</i>	$\mu\text{g L}^{-1}$	April ^a	41.9 (53.5)
		May ^a	25.5 (23.0)
		July ^a	38.4 (67.3)
		September ^a	75.1 (80.7)

controlling the seasonal concentrations of these GHGs across 20 agricultural reservoirs in the northern Great Plains, the largest farming region in Canada. This study builds on previous work which found that some regional lakes sequester CO₂ (Finlay et al., 2009; Finlay et al. 2010; Finlay et al. 2015; Finlay et al. 2019), as well as previous spatial surveys of 101 agricultural reservoirs that found highly variable fluxes of CO₂ and CH₄, but general undersaturation of N₂O (Webb et al., 2019a; Webb et al., 2019b). The goals of this study were to: 1) quantify the

change in concentrations of CO₂, CH₄, and N₂O on seasonal and diel timescales in agricultural reservoirs; 2) identify the seasonal drivers of CO₂, CH₄, and N₂O concentrations in these agricultural reservoirs, and; 3) estimate the total annual CO₂ equivalence contribution of GHG to the atmosphere from these systems. Based on earlier work, we anticipated that the reservoirs would be consistent sinks for CO₂ and N₂O, with only notable release in spring and late fall (Finlay et al., 2015; Finlay et al., 2019), that GHG release varied inversely with stratification

intensity (Webb et al., 2019a), and that CH₄ may be the main control of net CO₂-eq exchanges in these systems (Pennock et al., 2010; Webb et al., 2019b).

2 Methods

Twenty constructed agricultural reservoirs were chosen for this study (Figure 1). Agricultural reservoirs, known regionally as “dugouts”, typically have a rectangular construction, with steep sloping sides (1.5:1) and a maximum depth of 4–6 m (Webb et al., 2019b). All reservoirs are located in southern Saskatchewan, Canada, within a 150 km radius of the City of Regina (50° 27′ 10.19″ N, –104° 36′ 14.39″ W), as a previous survey of 100 regional reservoirs did not identify significant spatial component to regulation of GHG concentrations (Webb et al., 2019a; Webb et al., 2019b). Sites were selected to encompass a wide range of physical and chemical parameters (Table 1 and Table 2) based on data collected in 2017. Reservoirs were sampled four times during 2018; in late April when 75% of ice cover had melted, mid-May when sites were entirely ice-free, mid-July, and at the end of September prior to October ice formation. Five sites were also sampled in June and August to evaluate the effect of increased temporal resolution on determination of seasonal patterns in GHG content and fluxes.

2.1 Field collection

2.1.1 Seasonal sampling

During each sampling period, samples were collected within a 10-day interval, and all collections were made during daylight hours. Each site was sampled from a canoe, anchored near the center of the reservoir over the deepest water. Water temperature (°C), DO (mg L⁻¹, % saturation), conductivity (μS cm⁻¹), and pH were measured using a Yellow Springs Instrument (YSI) multi-parameter probe at ~0.5-m depth intervals from the surface to the bottom. The probe was calibrated monthly using three standard solutions for pH, whereas DO saturation was calibrated at each sampling location. Atmospheric pressure (mm Hg) was estimated at each site using the YSI probe. Depth of the reservoir was taken using a Norcross Hawkeye handheld depth finder. Water clarity was measured using a 20-cm diameter Secchi disk.

Water samples were collected using a submersible pump with an intake deployed at ~0.5-m depth. Whole water was screened through 80-μm mesh to remove large zooplankton and particulate matter and combined into a pre-rinsed cleaned carboy. Samples were transported back to the laboratory for chemical analyses.

GHG concentration of water was sampled on site using the headspace extraction method of Webb et al. (2019a). This process involves using a submersible pump to fill a 1.2-L glass serum

bottle to overflow using water from ~0.5-m depth, ensuring that there were no bubbles present, and sealing the bottle with a two-way rubber stopper. A 60-ml sample of atmospheric air was added to the bottle while 60-ml of water was removed to maintain constant pressure in the bottle. The bottle was then shaken vigorously for 2 min to equilibrate the air and water. Two replicate gas samples were extracted from the headspace using an air-tight syringe and put into 12-ml pre-evacuated Exetainer vials with double wadded caps. This process was repeated in duplicate to collect four samples per date. A sample of atmospheric air was also collected and placed in a 12-ml pre-evacuated Exetainer vial with double wadded cap. All gas samples were stored at room temperature until analysis within 2 months of collection. While methane ebullition is known to contribute significantly to methane emissions, this analysis only focuses on diffusive flux of greenhouse gases. Methane ebullition is known to be very patchy both spatially and temporally, and seasonal changes in bubble flux is currently not well constrained.

2.1.2 Diel sampling

Four reservoirs were selected for diel monitoring to assess daily variability in CO₂, CH₄, and N₂O concentrations. Samples were collected at 4-h intervals over 24 h (6 total), beginning ~10:00 h, between 26 June and 01 August 2018. Each diel cycle involved GHG sampling and YSI readings as described above. Water samples were collected at the 0-h, 12-h, and 24-h marks to determine changes in water chemistry throughout the 24-h period. Additionally, a YSI multi-parameter probe was deployed in each site for the entire 24-h period, taking readings for water temperature, DO, pH, and conductivity at 10-min intervals. Sites were selected from a set of previously sampled sites to include a wide range of physical and chemical parameters. Secondly, sites were selected based on ease of access and availability of services for the field team.

2.2 Laboratory analysis

At the laboratory, water samples were stored at 4°C and filtered within 24 h of collection. Sufficient water to visibly discolor a Whatman 1.2-μm GF/C filter was filtered was recorded, and filtrate was frozen (–10°C) in sealed darkened film canisters until analysis for chlorophyll pigments.

Greenhouse gas samples were analyzed at the Global Institute for Water Security, University of Saskatchewan. Headspace gas samples were analyzed for the dry molar fraction of CO₂, CH₄, and N₂O using a fully calibrated Scion 456 Gas Chromatograph (Bruker Ltd.) with Combipal autosampler (CTC Analytics–PAL System), using argon as the carrier gas. A flame ionization detector was used for methane samples (<100,000 ppmv). A thermal conductivity detector was used for CO₂ and high level CH₄ concentrations (>100,000 ppmv). N₂O was measured using a micro-electron capture detector and argon/methane (90/10) as

a makeup gas (injector temperature 60°C, column temperature 60°C, detector temperature 350°C). All gases were calibrated using mixed gas standards (Praxair) with the addition of a single gas N₂O standard (0.1 ppmv; Scotty).

Water chemistry and water isotope samples were analyzed at the Institute for Environmental Change and Society at the University of Regina. Water chemistry samples were analyzed for soluble reactive phosphorus (SRP), total dissolved nitrogen (TDN), nitrate and nitrite (NO_x), ammonia (NH₃), dissolved organic carbon (DOC), and dissolved inorganic carbon (DIC). DOC and DIC were analyzed using standard analytic procedures on a Shimadzu model 5000 A total carbon analyzer (Finlay et al., 2009; Webb et al., 2019b). Dissolved nitrogen (TDN, NO_x, NH₃) and dissolved phosphorus (TDP, SRP) were measured using standard procedures on a Lachat QuikChem 8,500 (APHA-AWWA/WEF 1998; Bergbusch et al., 2021). Chl-*a* concentration was standard trichromatic spectrophotometric methods (Jeffrey and Humphrey 1975; Finlay et al., 2009). Water was analyzed for δ¹⁸O-H₂O using a Picarro L2120-I cavity ring-down spectrometer (CRDS), and δ¹⁸O inflow was calculated using coupled isotope tracer methods as an estimate of water source (winter vs. summer precipitation, groundwater, Yi et al., 2008; Webb et al., 2019a; Haig et al., 2020).

2.3 Numerical analyses

The squared Brunt-Väisälä buoyancy frequency (s^{-2}) was calculated as a measure of maximum stratification strength within the water column. This approach uses the greatest density gradient from the vertical water temperature profiles taken at intervals of 0.5 m (0.25 m in shallower waterbodies) using the rLakeAnalyzer package (Read et al., 2011) in R (version 4.0.5; R Core Team, 2021) (Webb et al., 2019b).

The dry molar fractions of CO₂, CH₄, and N₂O were corrected for dilution and converted to concentrations according to the solubility coefficients and atmospheric partial pressure of each gas and the salinity of the water. Four replicate gas concentrations for each site were averaged for CO₂, CH₄, and N₂O to estimate mean concentrations for each sampling period. We removed an extreme CH₄ outlier for in one reservoir seen during April (93.5 μM) and assumed an ebullitive event that was captured in the associated water sample.

The concentration of each of the gases were used to estimate the diffusive flux for CO₂, CH₄, and N₂O. The diffusive flux (f_c) was calculated using the gas transfer velocity (k_c), gas concentration of the water (C_{water}), and the ambient air concentration (C_{air}) using the following equation (Eq. 1):

$$f_c = k_c (C_{\text{water}} - C_{\text{air}}) \quad (1)$$

The ambient air concentration of each gas was taken as the average over the sampling period at the Mauna Loa NOAA

station from April to September 2018, inclusive. The average concentrations were 408.94 μatm (or ppm) for CO₂, 1.86 μatm (or ppm) for CH₄, and 0.33 μatm (or ppm) for N₂O, and corrected for the barometric pressure for each sampling effort. The diffusive flux was calculated using the gas transfer velocity as measured on similar agricultural reservoirs in 2017 (Webb et al. 2019a; Webb et al. 2019b). We did not estimate ebullitive CH₄ fluxes, although we recognize this will lead to an underestimation of the true total CH₄ efflux.

Integrating seasonal GHG flux across all gases is challenging, owing to temporal variability and the assumptions required to extrapolate between sampling dates. Here, we calculated total CO₂-eq flux using several methods to bound true values. First, the “July only” procedure used only the daily CO₂-eq value observed in July multiplied over the total number of ice-free days to extrapolate over the entire open water period (e.g., metanalysis in DelSontro et al., 2018). Second, the “Seasonal” protocol used the CO₂-eq values measured in April, May, July, and September for the respective intervals summed over the entire open water period (e.g., Striegl and Michmerhuizen 1998; Finlay et al., 2019). Third, we calculated a “Predicted” flux using the generalized additive models (GAMs) of the smoothed seasonal splines to interpolate fluxes on each date between sampling periods. For this last method, we took both the mean smooth value and the upper 95% credible interval (“Upper CI”) to estimate mean and maximum seasonal CO₂-eq flux. Given the lack of consistent diel trend (see Section 3.4 below), we used measured flux rates from daytime determinations only. In all cases, the CO₂-eq flux was calculated by first converting the moles of diffusive flux value to grams of diffusive flux, and then multiplying by the sustained global warming potential (45 and 270 for CH₄ and N₂O, respectively) if the diffusive flux value was positive, and by the sustained global cooling potential (203 and 349 for CH₄ and N₂O, respectively) if the diffusive flux value was negative (Neubauer and Megonigal 2015).

Generalized additive models (Wood 2006; Wood et al., 2016) were used to assess the seasonal trends in CO₂, CH₄, and N₂O concentration in the reservoirs, as they are particularly useful for estimating linear, nonlinear, and nonmonotonic relationships between response variables and predictors (Webb et al., 2019a; b; Swarbrick et al., 2019). All modelling was done using the *mgcv* package (Wood 2011; Wood 2016) for R (version 4.0.5; R Core Team, 2021). GAMs were also used to link CO₂, CH₄, and N₂O concentrations to various biotic and abiotic predictive parameters to identify which factors might regulate changes in GHG concentrations in agricultural reservoirs. Covariates included those thought to control seasonal variation in GHG levels during the open-water period and were based on spatial analysis of these agricultural reservoirs (Webb et al., 2019a; Webb et al., 2019b). Variables included nutrient content (DOC, DIN,

SRP), production (Chl-*a*, DO), and stratification strength (buoyancy frequency). The CO₂ model additionally included water sources (as δ¹⁸O of inflow), specifically groundwater, rainfall, or snowmelt (Yi et al., 2008; Haig et al., 2020). Conductivity was additionally included in the CH₄ model as a proxy for ionic content and, in this region, sulfate concentrations ($r^2_{\text{cond-SO}_4} = 0.64$, $p < 0.01$). Similarly, pH was included in the N₂O model because of its known effects on N₂O production and consumption processes (Quick et al., 2019). Site identity (ID) was included as a random effect in the model, to account for the differences between individual sites.

A Gamma distribution was used with a log-link function, for positive, continuous responses of CO₂ and CH₄. We used a gammals distribution for N₂O concentrations to model the log mean and log scale parameter (or standard deviation/variance) for the probability density function. The gammals distribution was used because two sites exhibited high, outlying N₂O concentrations compared to the rest, so this allowed for grouping (high and low) of sites separately from the rest of the sites to inform a non-normal distribution. Sites were put into the high group if 25% or more of their N₂O concentrations were outliers. Basis size, dispersion of residuals, homogeneity of variance, and the relationship between the observed and predicted response were assessed in each model to ensure assumptions were not violated. Residual marginal likelihood (REML) was used for selection of smoothness parameters (Wood, 2011). To help with model selection, the double penalty approach of Marra and Wood (2011) was used. An additional penalty is applied to the perfectly smooth parts of the basis (the functions in the penalty null space) of each smooth function in the model, which allows entire smooths to be effectively penalised out of the model, while accounting for the selection procedure in the statistical tests applied to model terms. Parameters predicting CO₂, CH₄, and N₂O concentrations were considered significant at 95% confidence level ($\alpha = 0.05$) for each waterbody.

3 Results

3.1 Water quality

Physical and environmental parameters of the 20 reservoirs exhibited a wide range of values among sites and over the season (Table 1). The mean monthly max depth (mean = 1.88 m, standard deviation (SD) = 0.92), Secchi depth (0.61 ± 0.43 m), DO saturation (89.5 ± 31.2%), and Chl-*a* (44.8 ± 61.1 μg L⁻¹) of the reservoirs did not change significantly between any month (ANOVA, p -value > 0.05).

Mean surface water temperature was low in April (mean = 7.98°C, SD = 4.38), reached a maximum in July (mean = 22.7°C, SD = 1.47), and declined again in September (mean = 6.99°C, SD = 0.87, ANOVA post-hoc Tukey test, p -value < 0.001). The pH of the surface water was lowest in April (mean = 8.29, SD = 0.84), reached a maximum in July (mean = 8.94, SD = 0.61), and decreased again in September (mean = 8.62, SD = 0.44, ANOVA post-hoc Tukey test, p -value = 0.014). Conductivity increased from April (mean = 737 μS cm⁻¹, SD = 591) to September (mean = 1824 μS cm⁻¹, SD = 1722, ANOVA post-hoc Tukey test, p -value = 0.029). Salinity also increased from April (mean = 0.53 ppt, SD = 0.42) to September (mean = 1.25 ppt, SD = 1.11, ANOVA post-hoc Tukey test, p -value = 0.033).

Water chemistry parameters were highly variable both between months and among reservoirs (Table 2). Soluble reactive phosphorus (seasonal mean = 320 μg P L⁻¹, SD = 515), TDN (2,793 ± 2,141 μg N L⁻¹) and NH₃ (seasonal mean = 316 μg N L⁻¹, SD = 654) concentrations did not have a significant change between monthly means (ANOVA post-hoc Tukey test, p -value = 0.001). DOC concentrations also increased from April (mean = 20.1 mg C L⁻¹, SD = 6.54) to September (mean = 31.3 mg C L⁻¹, SD = 12.0, ANOVA post-hoc Tukey test, p -value = 0.004). The concentration of NO_x was highest initially following ice-off in April (mean = 644 μg N L⁻¹, SD = 1,054, ANOVA post-hoc Tukey test, p -value = 0.009).

3.2 Seasonal greenhouse gas concentration and flux

Temporal variation in concentrations of CO₂, CH₄, and N₂O (Figure 1A) was identified as significant seasonal trends when analyzed by GAMs for all gases (Figure 1B). The concentrations of each of the gases was highly variable between reservoirs, yet there were common trends for each gas. For example, concentrations of CO₂ were greatest immediately following ice-off in April (mean = 45.8 μM, SD = 39.3), declined during spring and summer (May–mean = 23.5 μM, SD 23.6, July–mean = 16.5 μM, SD 17.0), then increased again in fall (mean = 32.0 μM, SD 31.2, GAM, p -value = 0.013, deviance explained = 49%). In contrast, CH₄ concentration were initially low in the spring (April–mean = 0.849 μM, SD = 1.54; May–mean = 0.73 μM, SD = 1.28), but increased into a mid-summer peak (mean = 1.32 μM, SD = 1.70) before decreasing in fall (mean = 0.40 μM, SD = 0.47, GAM, p -value = 0.0005, deviance explained = 91.4%). Finally, concentration of N₂O exhibited a strong pulse during ice-off in April (mean = 35.2 nM, SD = 50.3), followed by a sharp decrease to low values in spring and summer

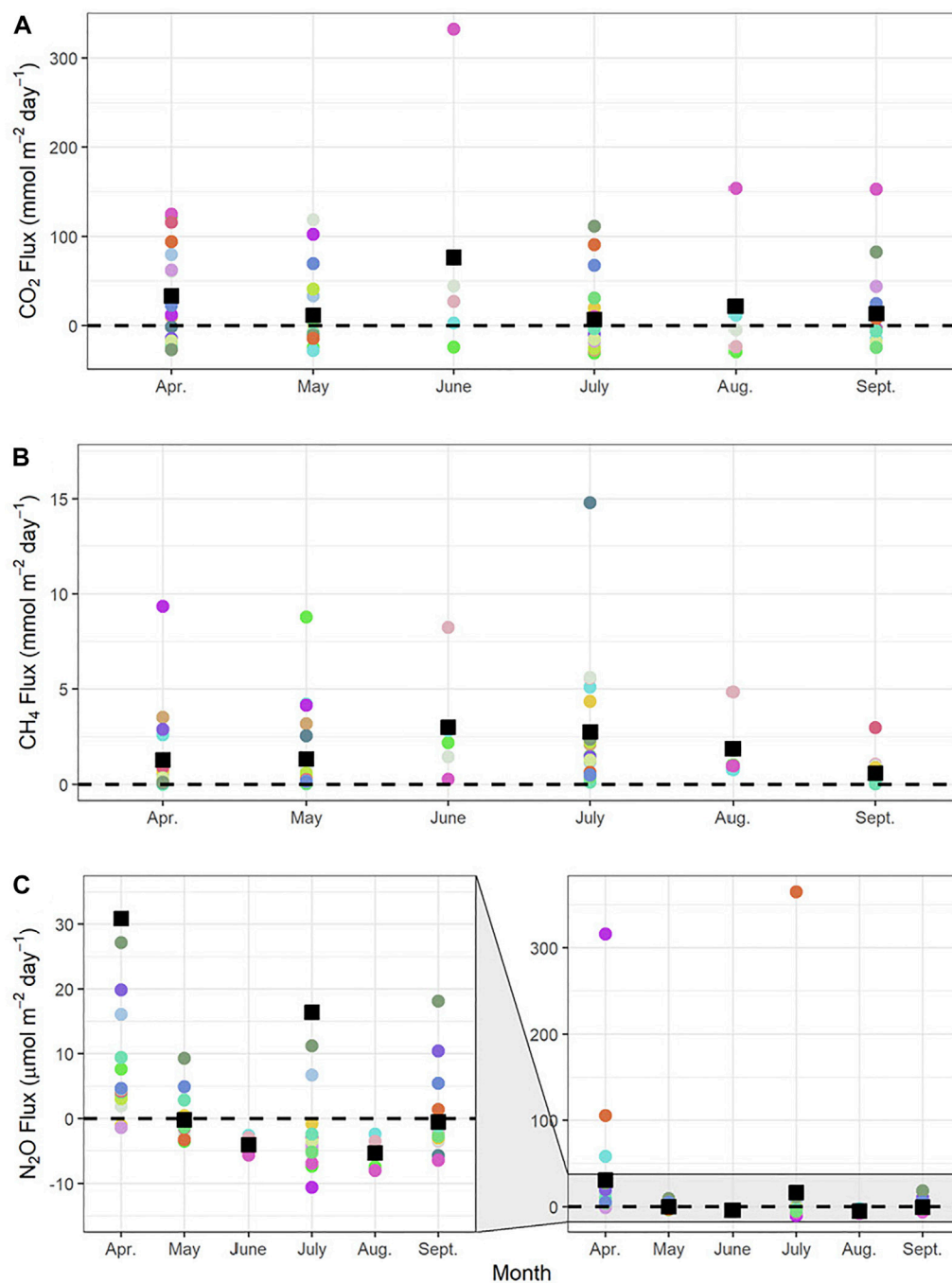


FIGURE 2

(A) Carbon Dioxide (CO₂) (B) methane (CH₄), and (C) nitrous oxide (N₂O) flux for the sampling period. Each coloured circle represents a single reservoir, while the black square represents the monthly mean. Black dashed line atmospheric equilibrium, where values above indicate loss of gas to the atmosphere and values below indicate removal of gas from the atmosphere. One outlying CH₄ value in April was omitted (value = 100 mmol m⁻² day⁻¹) as it is assumed that an ebullition event was captured.

(May–mean = 10.1 nM, SD = 1.5; July–mean = 15.4 nM, SD = 36.1), before rising slightly again prior to ice formation (mean = 13.3 nM, SD = 4.31, GAM, *p*-value < 0.0001, deviance explained = 89.5%).

Estimates of diffusive fluxes of CO₂, CH₄, N₂O were all highly variable among sites (Figure 2). Despite among-site variation, diffusive flux of CO₂ varied little between months (ANOVA, *p*-value = 0.161), with the overall mean (= SD) of 19.7 ±

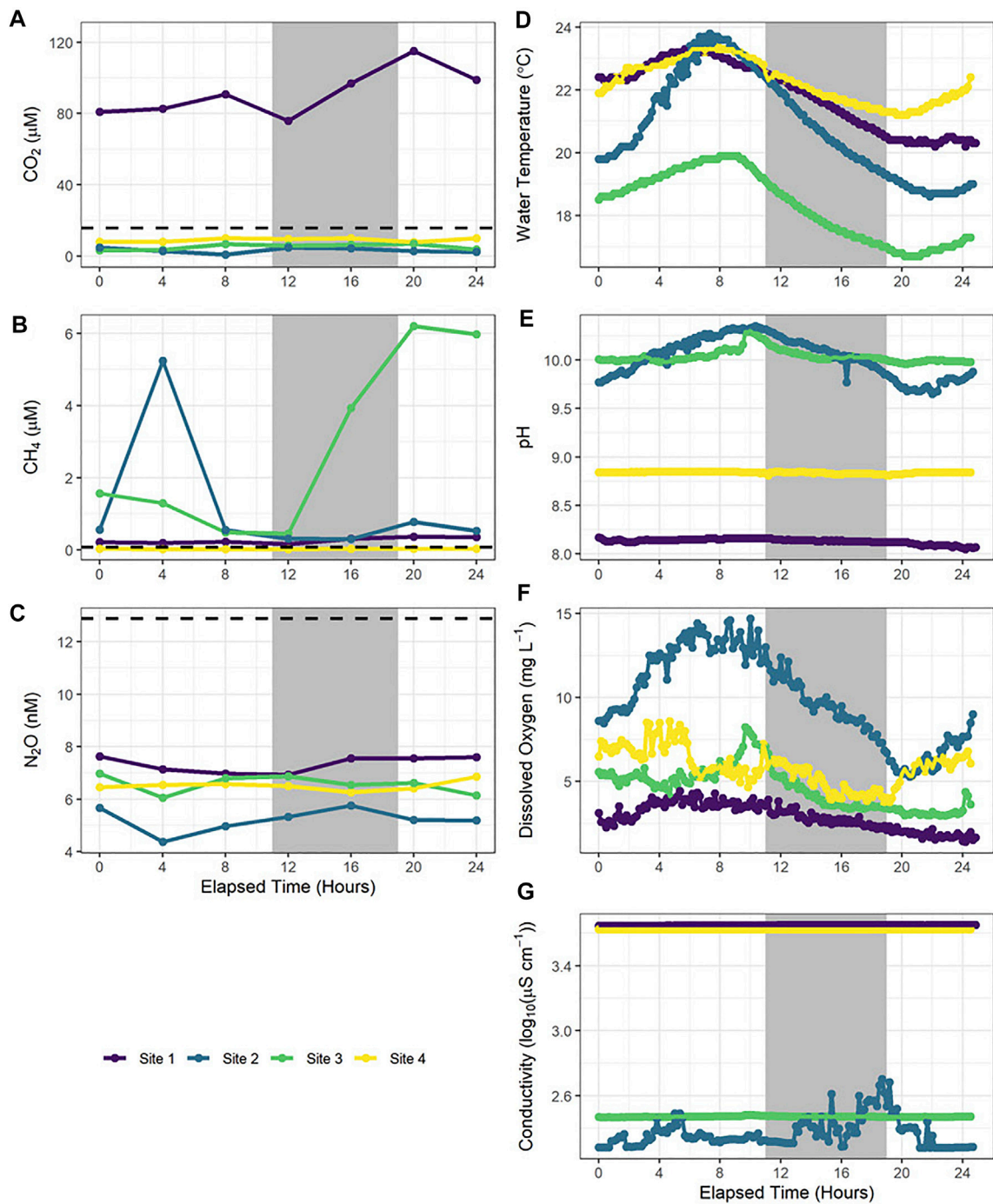
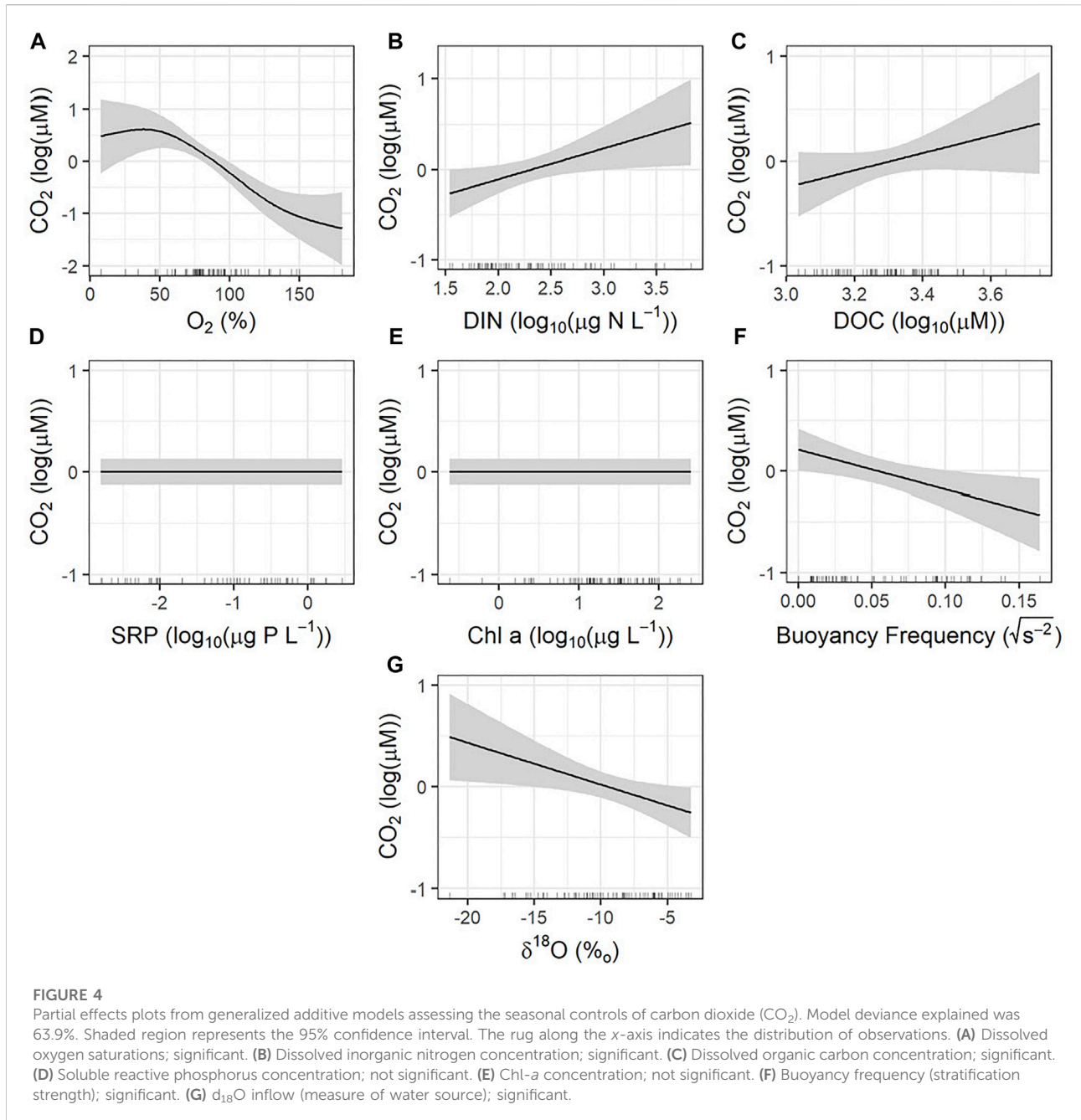


FIGURE 3

Greenhouse gas (A–C) and water quality parameters (D–G) in four reservoirs measured over a 24-h period. Each coloured line is representative of one site. The grey shaded regions are the average night-time period. Elapsed time is time from the initiation of the 24-h period. Earliest start was 09:50, while the latest start was 11:24. Temperature (D), DO (dissolved oxygen (E)), pH (F) and conductivity (G) readings were taken every 10 min, while greenhouse gas concentrations were collected every 4 h. CO₂ = carbon dioxide (A). CH₄ = methane (B). N₂O = nitrous oxide (C). See 24-h averages of nutrient, Chl-a and carbon concentrations in [Supplementary Table S1](#).

56.6 mmol m⁻² day⁻¹. Similar patterns were recorded for the diffusive flux of CH₄, with no seasonal difference between monthly means (ANOVA, *p*-value = 0.654) and an overall flux

of 2.9 ± 10.9 mmol m⁻² day⁻¹. Diffusive flux of N₂O was variable among sites but not months (ANOVA, *p*-value = 0.361), with an overall mean diffusive N₂O flux of 9.7 ± 52.9 μmol m⁻² day⁻¹.



3.3 Diel water chemistry

Most water quality parameters did not change substantially during the diel cycle (Figure 3). In general, temperature was greater during the day and lower at night, although there were up to 4°C differences among sites (Figure 3D). Similarly, while DO exhibited some high frequency variation, concentrations were generally lower at night than in the day (Figure 3E). In contrast, few trends were observed for conductivity, and pH exhibited diel cycle (elevated in day) at only one site with elevated Chl-*a* values and high diel changes in DO. Nutrients (TN, TP, SRP, DOC,

DIC) and Chl-*a* did not exhibit any observable changes during the diel cycle in any sites, while NO_x concentration and ammonia concentration only exhibited notable variation at one site with high DIN levels (Supplementary Table S1).

3.4 Diel greenhouse gas concentration and flux

Concentrations of CO_2 , CH_4 , and N_2O showed no consistent trends during a 24-h period in the four

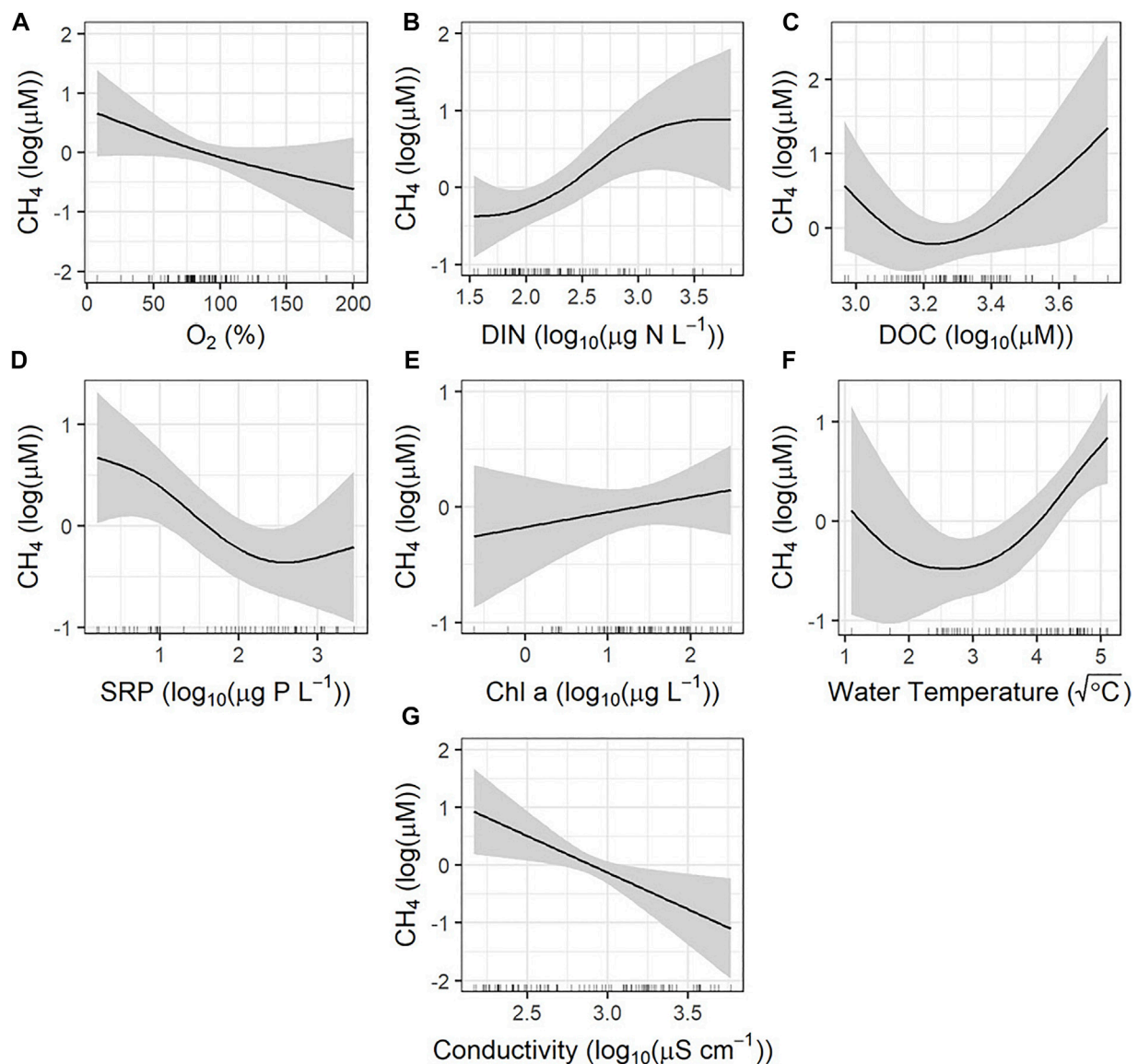


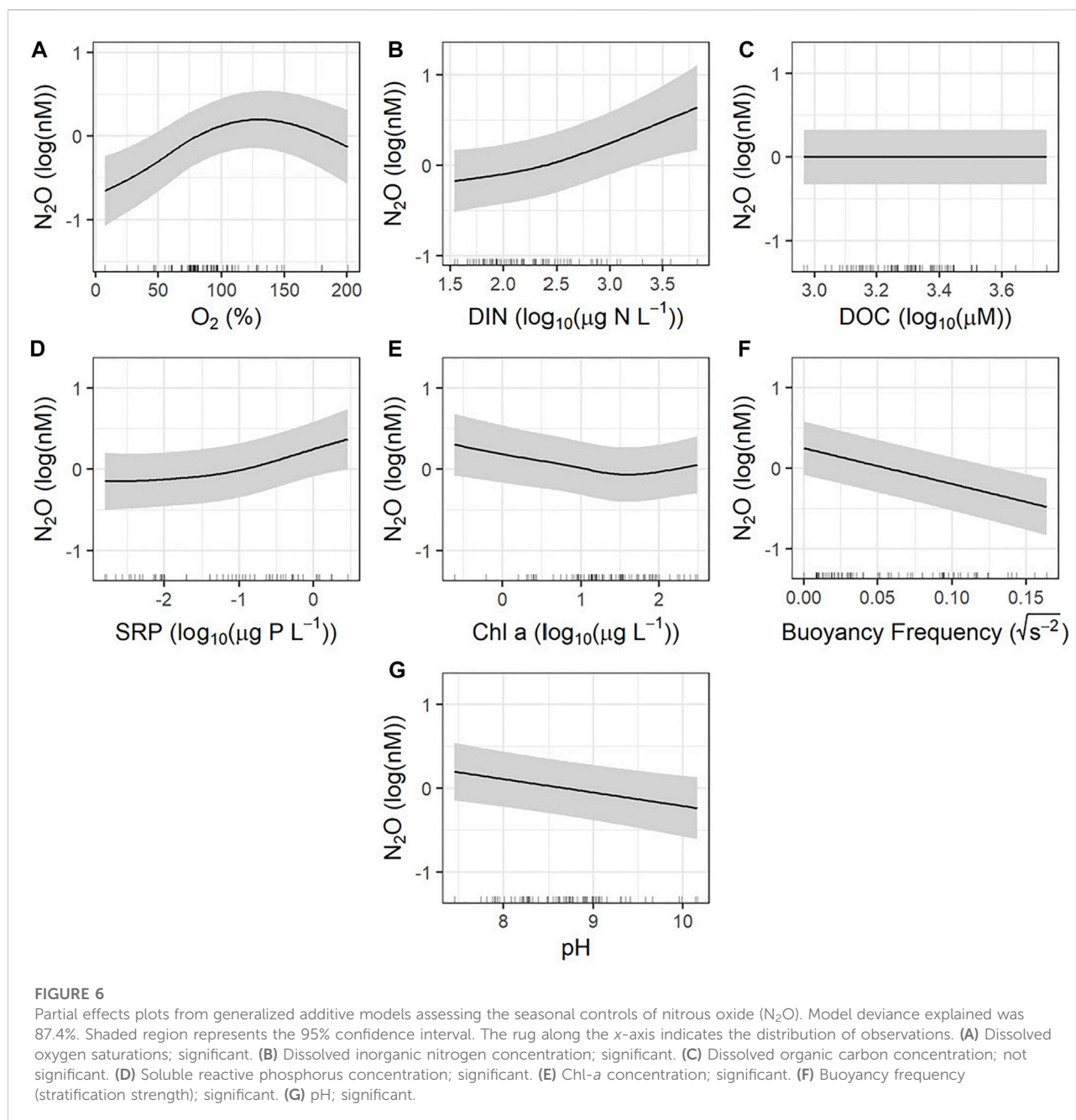
FIGURE 5

Partial effects plots from generalized additive models assessing the seasonal controls of methane (CH_4). Model deviance explained was 74.5%. Shaded region represents the 95% confidence interval. The rug along the x-axis indicates the distribution of observations. **(A)** Dissolved oxygen saturations; significant. **(B)** Dissolved inorganic nitrogen concentration; significant. **(C)** Dissolved organic carbon concentration; significant. **(D)** Soluble reactive phosphorus concentration; significant. **(E)** Chl-a concentration; not significant. **(F)** Water temperature; significant. **(G)** Conductivity; significant.

reservoirs sampled (Figure 3). At three sites, CO_2 concentrations were consistently undersaturated over the diel cycle, while values were supersaturated at the fourth site yet no more variable through time. CH_4 concentrations at two sites remained low throughout the diel cycle, while the other two basins showed inconsistent but elevated variation during the 24 h study period. Overall, N_2O concentrations were consistently low and undersaturated during the diel cycle.

3.5 Seasonal greenhouse gas controls

Seasonal changes in CO_2 concentration in the reservoirs were explained best by a GAM using the pH of surface water as the sole predictor (Supplementary Figure S2). This model explained 89.1% of deviance in CO_2 and was characterized by a strong negative relationship with CO_2 above pH of 8. When pH was not included in the model, variation in CO_2 was explained best by a combination of DO saturation (Figure 4A), DIN concentration

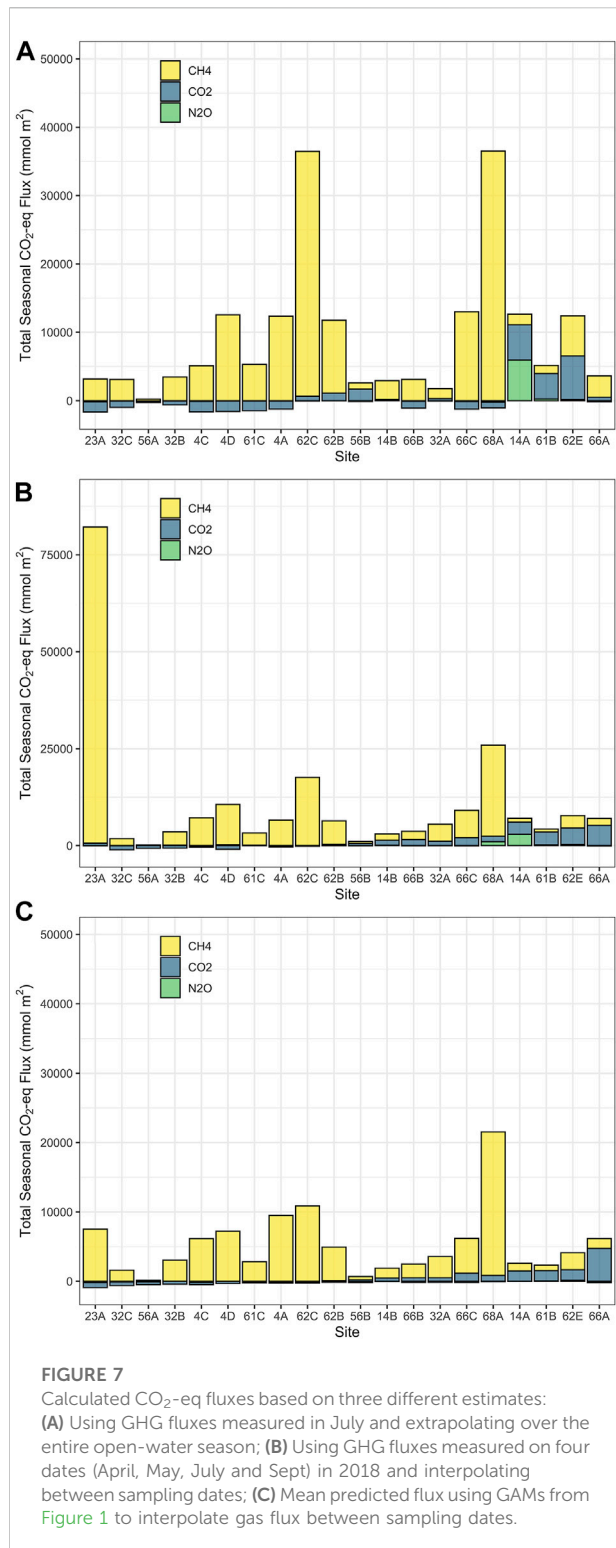


(Figure 4B), DOC levels (Figure 4C), buoyancy frequency (Figure 4F), and water source (as $\delta^{18}O$; Figure 4G). Dissolved oxygen, buoyancy frequency, and $\delta^{18}O$ exhibited significant negative relationships with variation in CO_2 concentration, while DIN and DOC had positive relationships. Overall, this model explained 63.9% of deviance in the CO_2 levels.

Seasonal changes in CH_4 concentrations were explained best by a combination of DO saturation (Figure 5A), concentrations of DIN (Figure 5B), DOC (Figure 5C), and SRP (Figure 5D), as well as surface water temperature (Figure 5F), and conductivity

(Figure 5G). Dissolved oxygen, SRP, and conductivity exhibited significant negative relationships with the seasonal CH_4 concentration, while DIN, DOC, and surface water temperature exhibited significant positive relationships with methane levels. Model deviance explained was 74.5% of variation in CH_4 levels.

Regulation of seasonal variation in N_2O concentrations was also complex, with significant effects of changes in DO saturation (Figure 6A), DIN concentration (Figure 6B), SRP concentration (Figure 6D), Chl-*a* concentration (Figure 6E), buoyancy



frequency (Figure 6F), and pH of the surface water (Figure 6G). Variation in N₂O levels were related positively to changes in DO, DIN, and SRP, and negatively to variation in Chl-*a*, buoyancy

frequency, and pH, while the model explained 87.4% of the deviance in N₂O levels.

3.6 Seasonal CO₂-eq flux

Estimates of total CO₂-eq flux over the open-water period varied dependent on the method used to estimate *in situ* GHG concentrations (Figure 7), but all were within an order of magnitude of each other. Mean seasonal CO₂-eq flux was comparable between the “July-only” (9,811 ± 10,269 mmol m⁻², Figure 7A) and “seasonal” methods (10,446 ± 17,893 mmol m⁻², Figure 7B), and were roughly twice the fluxes predicted using GAMs (5,027 ± 4,806 mmol m⁻², Figure 7C). All methods suggest that methane was the dominant GHG responsible for CO₂-eq flux, and contributed more than 50% to total GHG efflux in 16 of 20 sites (Figure 7, yellow bars). While the mean seasonal values were comparable among methods, there were some discrepancies within individual sites. For example, Site 23 A demonstrated extremely high CO₂-eq flux, attributable to one very high CH₄ measurement in April. This likely reflected an ebullition event at this time that was not captured when the “July-only” method was used, and which was constrained using the GAM-predicted method.

4 Discussion

Evaluation of temporal variability and controls of GHG in 20 agricultural reservoirs in the northern Great Plains indicated that while CO₂, CH₄, and N₂O flux exhibited few diel trends (Figures 3A–C), all GHG exhibited pronounced seasonality during the ice-free period regulated by complex interactions between microbial metabolism, allochthonous carbon inputs, and physico-chemical features (Webb et al., 2019a; Webb et al., 2019b). As predicted, all sites demonstrated peaks of CO₂ and N₂O in spring, as well as a second, lower peak of CO₂ during fall (Pennock et al., 2010; Webb et al., 2019a; Webb et al., 2019b), whereas CH₄ content was maximal in mid-summer (Figure 1), consistent with studies of Yvon-Durocher et al. (2014) and DelSontro et al. (2018). Contrary to predominantly planktonic controls identified in earlier regional GHG studies (Finlay et al., 2015; Wiik et al., 2018; Finlay et al., 2019), seasonal variation in CO₂ concentrations were correlated with temperature and organic carbon availability, indicating effects of benthic respiration (Figure 4). Similarly, CH₄ levels were correlated to changes in temperature, while a negative correlation with conductivity suggests inhibition of methanogenesis by sulfate-reducing bacteria in ion-rich waters (Figure 5). In contrast, N₂O concentrations appeared to be controlled mainly by variation in reservoir stratification or mixing (Figure 6). Taken together these findings suggest that

the response of small farm reservoirs to future climate change will depend strongly on which climatic parameters (temperature, evaporation, runoff, wind) are most affected by atmospheric heating. In particular, factors that regulate mid-summer CH₄ release appear to be paramount in determining whether small reservoirs hold potential as net CO₂-eq sources or sinks in the coming decades.

4.1 Carbon dioxide

CO₂ concentration in agricultural reservoirs was high in the spring, low throughout the summer, and began to increase again in fall (Figure 1B). This seasonal pattern was consistent with that observed in boreal and other hard-water systems, where CO₂ accumulates under the ice in the winter, causing a pulse of CO₂ released in the spring following ice off (Baehr and DeGrandpre 2002; Finlay et al., 2015; Denfeld et al., 2016). Depressed CO₂ in summer is attributed to isolation of surface waters leading to elevated photosynthetic rates that increase pH and deplete dissolved pCO₂ to values below atmospheric values (Huotari et al., 2009; Holgerson 2015). Conversely, elevated fall CO₂ concentrations appear to reflect loss of water-column stratification, and release of hypolimnetic CO₂ which had been accumulated during the summer period of stratification (Huotari et al., 2009; Holgerson 2015; Webb et al., 2019b; Finlay et al., 2019). Agricultural reservoirs in this study have a high relative depth ($5.6 \pm 3.1\%$), and thus exhibit a high resistance to vertical mixing (Wetzel and Likens 1991; Holgerson et al., 2022) during summer. As elevated primary production sinks into the hypolimnion and decomposes during summer, CO₂ is released leading to supersaturation of this gas in deep water (Huotari et al., 2009; Finlay et al., 2019).

Seasonally, pronounced negative correlations between CO₂ concentration and DO saturation (Figure 4A) and positive correlations between CO₂ and both DIN and DOC (Figure 4C) are indicative of metabolic controls of CO₂ concentrations (Holgerson 2015; Balmer and Downing 2017; Vachon et al., 2019). The metabolic stoichiometry between CO₂ and DO is well documented (Vachon et al., 2019) and represent autotrophic and heterotrophic processes occurring both in the water column or emanating from the sediments (Roulet et al., 1997; Jonsson et al., 2003; Rantakari and Kortelainen 2005; Kortelainen et al., 2006; Holgerson 2015). Here, the significant positive relationship between CO₂ concentration and DIN levels (Figure 4B), as well as DOC levels (Figure 4C), and lack of significant relationship with Chl-*a* (Figure 4E) suggests that elevated nutrient content stimulated microbial respiration rather than primary production, such as seen in other artificial waterbodies and prairie lakes (Ollivier et al., 2018; Wiik et al., 2018; Webb et al., 2019b; Peacock et al., 2019), and natural temporary wetland ponds (Holgerson 2015). As shown in mesocosm fertilization experiments, moderate

eutrophication of shallow prairie waters first elevates primary production and DO saturation, then leads to water-column anoxia due to build-up of dissolved and particulate organic matter (Donald et al., 2011; Bogard et al., 2020). Consistent with this mechanism, monitored reservoirs exhibited low ratios of dissolved N:P characteristic of N-limited regional lakes (Donald et al., 2011) (Table 2), while on-going whole-reservoir fertilization experiments show that N fertilization both increases surface water Chl-*a* levels and causes deep water supersaturation of CO₂ (C.A.C. Gushulak and P.R. Leavitt, unpublished data). At present, it is unclear whether the relationships observed in this or other studies reflect water column processes or the disproportionate effect of warm sediments in these shallow water bodies.

The negative correlation between CO₂ concentration and buoyancy frequency, a measure of stratification strength (Figure 4F), is consistent with the effects of seasonal thermal stratification in isolating deeper waters and constraining CO₂ loss to the atmosphere (Huotari et al., 2009; Holgerson 2015). Here, CO₂ concentrations are elevated both in spring after ice melting allows release of the winters' accumulation of respired CO₂ (Baehr and DeGrandpre 2002; Finlay et al., 2015; Denfeld et al., 2016), as well as in fall after when cooling air temperatures erode thermal stratification and release hypolimnetic CO₂ (Huotari et al., 2009; Holgerson 2015). Between these intervals, agricultural reservoirs can exhibit thermal stratification that isolates respired CO₂ derived from sediments and prevents accumulation in surface waters. In general, this pattern contrasts findings from a spatial survey of prairie farm reservoirs in 2017 that found a significant positive relationship between CO₂ concentration and buoyancy frequency (Webb et al., 2019b). We speculate that this difference arose because sampling for Webb et al. (2019b) took place during the summer and would have mainly captured variation in the degree of thermal stratification among sites, rather than the sharp seasonal patterns of water column mixing exhibited in this study.

CO₂ concentration and δ¹⁸O exhibited a pronounced negative relationship (Figure 4G) suggesting that elevated inflow of snowmelt or groundwater resulted in higher CO₂ concentrations in these farm reservoirs, consistent with observations in the 2017 spatial study (Webb et al., 2019b). In general, regional farmers site the reservoirs in existing lowland or wetland areas, often with direct contact to shallow aquifers, or with abundant overland runoff. In this region, snowmelt in spring is both the main source of overland flow (Pomeroy et al., 2007), and the mechanism recharging shallow aquifers (Haig et al., 2020) and, in both cases, exhibited highly depleted δ¹⁸O values relative to rainwater sources (Haig et al., 2020; Haig et al., 2021). Further, this groundwater has high concentrations of both DOC and DIC, and is frequently supersaturated with CO₂ (Macpherson 2009; Webb et al., 2019b). Seasonal monitoring of piezometers for water isotopes, CO₂ levels and dissolved C

concentrations will be required to further resolve this mechanism.

We did not observe consistent diel fluctuations of CO₂ concentrations in the four reservoirs sampled (Figure 3). Previous studies have shown that CO₂ concentrations are typically lower during the daytime when primary production exceeds respiration ($P > R$), but increase overnight due to continued respiration (Liu et al., 2016; Raymond et al., 2013; Wiik et al., 2019). The magnitude in the shifts of CO₂ concentration is often linked to the water-column productivity, with the largest diel changes observed in systems with the highest productivity (Hanson et al., 2003; Morales-Pineda et al., 2014; Shao et al., 2015; Wiik et al., 2018). Here Chl-*a* concentrations ranged from 2.9 to 296 μg L⁻¹, therefore, a strong diel change in CO₂ concentration would be expected in the more productive sites. Lack of pronounced variation through the day may reflect the high pH of these systems which would favour rapid conversion of CO₂ to bicarbonate (HCO₃⁻) and carbonate (CO₃²⁻). Furthermore, the high total alkalinity (241.3 ± 159.4 mg L⁻¹) of the systems likely resisted diel variation in pH in all but one reservoir (Figure 3D), as diel changes in CO₂ were much less than the total reservoir content of DIC (Dodds 2002; Stets et al., 2017).

4.2 Methane

CH₄ concentrations showed a marked peak during July-August (Figure 1B) similar to those seen high latitude wetlands (Pickett-Heaps et al., 2011), aquaculture ponds (Yang et al., 2015), shallow urban basins (van Bergen et al., 2019), and prairie wetlands (Bansal et al., 2016). In general, this interval corresponds to the period of greatest water temperatures and is consistent with peak CH₄ levels occurring when water temperatures are greatest in this study (Figure 5F) and the observation that metabolism of methanogenic bacteria is greater in warmer waters (Segers 1998). Such a positive relationship of CH₄ and temperature has been observed in other shallow systems including prairie wetland ponds (Bansal et al., 2016), urban ponds (van Bergen et al., 2019), natural temporary wetlands (Holgerson 2015), and sites with abundant *Phragmites* growth (Kim et al., 1998). Methane ebullition was not included in our study; however, we acknowledge that this process can contribute significantly to GHG fluxes from inland water and may also vary seasonally.

Dissolved CH₄ is expected to be more abundant under anoxic conditions (Glass and Orphan 2012), therefore, the significant negative relationship with DO saturation was expected (Figure 5A). Such a trend is common in diverse ecosystems, including boreal lakes (Kankaala et al., 2013), natural temporary wetland ponds (Holgerson 2015), and our earlier spatial survey of prairie farm reservoirs (Webb et al., 2019b).

The strong negative relationship between CH₄ concentration and conductivity (Figure 5G) was also observed in the 2017 spatial study, which is likely attributed to sulfate limiting methanogenesis (Webb et al., 2019b). Sulfate-reducing bacteria have been shown to outcompete methanogenic bacteria at sulfate concentrations as low as 60 μM in freshwater sediments in oligotrophic lakes (Lovley and Klug 1983), and as low as 200 μM in eutrophic lakes (Winfrey and Zeikus 1977; Lovley et al., 1982). A linear regression showed that conductivity and sulfate are highly related in these agricultural reservoirs ($r^2 = 0.64$, p -value < 0.01). Based on this calculation, the sulfate concentration ranges from 0.33 to 35.9 mM in these agricultural reservoirs. Surface water conductivity significantly increased over the open water season (Table 1) and is likely one of the mechanisms driving down the CH₄ concentrations in fall.

Nutrient (DOC, DIN, and SRP) levels were also important predictors of CH₄ concentrations in agricultural reservoirs (Figures 5B,C,E), although solute concentrations did not appear to be correlated to primary production as Chl-*a* (Supplementary Figure S2.1). As noted above, effects of allochthonous nutrients may be felt mainly through changes in the redox potential and dissolved oxygen content of these shallow ecosystems, rather than via augmentation of primary production. In particular, elevated production (Figure 5E) and sedimentation of labile organic matter may have favoured decomposition and anoxia in sediments, thereby improving habitat and substrate for methanogenesis (Smith et al., 1999; Anderson et al., 2002; Ollivier et al., 2019). The pronounced negative relationship between CH₄ and SRP may suggest that there is competition for substrate between methanogenesis, and denitrification at high P levels, consistent with the increase in N₂O concentration is seen at high SRP concentrations (Figure 6D). Such a mechanism has also been observed in anaerobic wetland sediment slurry incubations (Kim et al., 2015).

CH₄ concentrations did not show a consistent diel trend in the four reservoirs sampled, despite obvious variation in both temperature and DO in the surface water (Figure 3). In contrast, distinct diel patterns of CH₄ production have been seen in other systems where gas levels increase following sunrise, peak in the early afternoon, then decline quickly to a stable low level at night (Neue et al., 1997; Kim et al., 1998; Xing et al., 2004). In general, mid-day maxima were seen here only in ponds with greatly elevated CH₄ levels, possibly suggesting that the absence of strong diurnal maxima in some sites reflects a generally low abundance of methanogenic bacteria in some reservoirs. While speculative, we suggest that differences in the influx of groundwater- or sub-surface sulfate (SO₄⁼) may explain the variation in absolute and relative CH₄ levels during a 24 h cycle (see below), where the presence of sulfur-reducing bacteria can inhibit methanogenesis (Lovley and Klug 1983; Pennock et al., 2010).

4.3 Nitrous oxide

Although freshwaters are often significant emitters of N_2O (Beaulieu et al., 2008; Beaulieu et al., 2011; DelSontro et al., 2018), patterns and controls of the seasonality of N_2O concentrations are poorly understood. The seasonal trend observed here shows that following an initial pulse in N_2O production immediately following ice melt in the spring, these reservoirs were often undersaturated in N_2O for much of the ice-free period (Figure 1B). These low N_2O values may be attributed to complete denitrification of dissolved NO_3^- to di-nitrogen gas (N_2), similar to patterns seen in some lakes (Piña-Ochoa and Álvarez-Cobelas 2006), including Lake Kasumiguara, a shallow eutrophic lake that does not have an anaerobic zone (Hashimoto et al., 1993). Widespread undersaturation of N_2O was also observed in regional farm reservoirs during 2017 (Webb et al., 2019a); however, this study provides the first evidence that N_2O undersaturation last much of the summer, a finding which has not previously been recorded for nitrogen-rich agricultural landscapes. Better understanding of the mechanisms controlling this process may be important to regulating net CO_2 -eq fluxes from small water bodies in the continental interiors that represent much of the global agricultural regions.

The significant relationships seen between DO saturation, DIN ($NO_x + NH_3$), Chl-*a*, and buoyancy frequency (Figure 6) were similar to those seen spatial surveys of agricultural reservoirs (Webb et al., 2019a). This finding suggests that mechanisms relating to productivity and mixing are controlling N_2O concentrations in reservoirs both seasonally and spatially (Webb et al., 2019a). The pulse of N_2O in the spring may be explained by the build-up of N_2O under ice over the winter season, which is a common observation in prairie and boreal lakes (Soued et al., 2016; Cavaliere and Baulch, 2018). The lack of atmospheric exchange (during ice cover) combined with ongoing N_2O producing processes such as nitrification and denitrification (Cavaliere and Baulch, 2018), without the competing factor of primary productivity for DIN (NO_x was elevated in April, Table 2), may explain the higher N_2O levels. The switch to N_2O undersaturation coincides with a decline in NO_x concentration owing to elevated algal uptake, along with the onset of temporary water column stratification and a decline in the DO saturation in the deep waters (Table 1) where complete denitrification can proceed. Complete denitrification leads to the consumption of N_2O in the process of producing N_2 (Quick et al., 2019), which suggests that the conditions in agricultural reservoirs favour complete denitrification during most of the open-water period. Additionally, the significant positive relationship between N_2O concentrations and SRP may be driven by direct, or indirect effects, via impacts of SRP on algal biomass and carbon production, or via direct effects on N_2O producing microbes.

N_2O concentration in the four reservoirs sampled on a diel timescale remained under-saturated throughout the 24 h cycle. This pattern is similar to that seen in an arid zone alpine pond which remained undersaturated in N_2O for most of the diel cycle (Molina et al., 2021). Previous studies have diel variation in N_2O

concentrations, primarily in rivers and streams, although comparisons among these site exhibit little common diel pattern among habitats. For example, some studies show clear diel patterns with greater concentrations observed during nighttime relative to those during the daytime (Rosamond et al., 2011; Wu et al., 2018a; Wu et al., 2018b), while other locations show a peak during the day (Molina et al., 2021) as well as a peak during the nighttime (Rosamond et al., 2011). One study found a sharp decrease in N_2O concentration during the nighttime (Harrison et al., 2005) and others did not find a consistent diel trend (Baulch et al., 2012).

4.4 Seasonal CO_2 equivalent budget

We calculated areal CO_2 -eq flux for the farm ponds to range from an average $32.0 \text{ mmol m}^{-2} \text{ day}^{-1}$ using the GAM-predicted flux, to $66.5 \text{ mmol m}^{-2} \text{ day}^{-1}$ using the seasonal method, over the 157-days open-water season. These values are somewhat lower than the mean ($129 \text{ mmol m}^{-2} \text{ day}^{-1}$) but well within the range (-10 to $1,462 \text{ mmol m}^{-2} \text{ day}^{-1}$) of the daily flux estimates in the 2017 spatial survey (Webb et al., 2019a; Webb et al., 2019b). Similar to findings from previous studies (DelSontro et al., 2018; Ollivier et al., 2019), we observed that CH_4 was the major driver of CO_2 -eq flux (average 91.5% across all methods), while N_2O contributed the least (0.73%). The contribution of CH_4 to total CO_2 -eq flux is likely to be even higher when ebullition is also considered (DelSontro et al., 2016; Rosentreter et al., 2021).

Findings from this study show that all three GHGs exhibited significant seasonal trends, thereby suggesting that extrapolation from a single date to the entire open-water period may produce inaccurate estimates of the total CO_2 equivalent (CO_2 -eq) flux from these systems. In our study, the seasonal trends of each GHG (Figure 1), combined with the fact that CH_4 is typically the dominant gas contributing to CO_2 -eq flux (Figure 7) would suggest that extrapolating measurements in mid-summer are likely to overestimate seasonal flux. The GAM-predicted seasonal fluxes were, on average, half of that of the other two methods, likely because the modeled fluxes constrained occasional extreme effluxes GHG and avoided excessive extrapolation through time series. For example, the exceptionally high estimate of seasonal flux at site 23 A in Figure 7B reflects a pulse of CH_4 in April (observed $CH_4 = 90.7 \mu\text{M}$) which was not included in the July-only flux calculations, and was constrained in the GAM models. Depending on whether this extreme point was interpolated to the next sampling date or not, estimated CO_2 -eq flux differed during the spring period by up to $72,800 \text{ mmol m}^{-2}$ depending on whether elevated values were ($\sim 75,600 \text{ mmol m}^{-2}$) or were not ($2,800 \text{ mmol m}^{-2}$) used to estimate vernal fluxes. Although ebullition can occur due to changes in production, water depth, or meteorological conditions (DelSontro et al., 2016), observed elevated values are unlikely to persist over the 3-week spring interval. Conversely, GAM

predicted values missed the likely-real pulsed efflux, and thereby likely underestimated seasonal fluxes. Overall, the “seasonal” calculation also likely overestimates CO₂-eq flux by inflating short-term emissions, while the July-only calculation will miss seasonal trends, and the use of the GAM smooths may dampen any short-term pulses of GHG efflux and lose the contribution of these to the seasonal flux rates. Reconciling these discrepancies to provide the best estimate of total open-water GHG fluxes will require continuous monitoring of GHG with sondes, now possible for CO₂, but still limited for CH₄ and N₂O due to more limited sensor sensitivity.

This study shows that agricultural reservoirs exhibit significant seasonal trends in the concentrations of all major GHGs, and that the concentrations of these gases are regulated by a combination of physical, chemical, and biological mechanisms. Additionally, it was found that inclusion of estimates of this seasonal variability may be essential to accurately total CO₂-eq fluxes to the atmosphere. However, reconciling accurate net GHG over the annual scale will only truly be overcome through increased GHG monitoring via *in situ* sensors to capture extreme changes in water conditions (such as at ice off). The Northern Great Plains, as the largest agricultural area of Canada, is an area where agricultural water management is ongoing. Reservoirs in the landscape mediate substantive fluxes, although more work is required to reduce error in estimation of annual efflux from these often-eutrophic reservoirs. Perhaps more importantly, the Northern Great Plains are a region of extremely high variability in factors known to influence GHG efflux, and upscaling efforts aimed at understanding the role of inland waters in GHG efflux need to account for spatial variability in solutes, including sulfate.

Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found below: https://github.com/finlay4k/Jensen_dugout_seasonality.git.

Author contributions

SJ, JW, GS, PL, HB, and KF designed research; SJ performed research and drafted the paper; HB contributed new reagents/

analytic tools; JW, PL, GS, HB, and KF contributed towards ideas and data analysis; SJ developed generalized additive models. All authors revised and approved the final manuscript.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fenvs.2022.895531/full#supplementary-material>

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